



Long-term population genetic dynamics of the invasive ascidian *Botryllus schlosseri*, lately introduced to Puget Sound (Washington, USA) marinas

Jann Zwahlen^{a,b,*}, Eitan Reem^b, Jacob Douek^b, Baruch Rinkevich^b

^a Dipartimento di Scienze e Tecnologie Biologiche ed Ambientali, Università del Salento, 73100, Lecce, Italy

^b Israel Oceanography and Limnological Research, National Institute of Oceanography, Tel Shikmona, PO Box 9753, Haifa, 3109701, Israel

ARTICLE INFO

Keywords:

Microsatellites
Alien species
Biogeography
Tunicata
Global
Newly established population

ABSTRACT

Invasive species are of increasing concern to biodiversity and the ecological functioning of a range of ecosystems, especially as the magnitude of biological invasions is increasing globally. The genetic structure of newly established populations may reveal insights into invasion processes, making population genetics an important tool for understanding current invasion pathways. Here, we studied newly established populations (non-existent < 10–20 years before the first sample) of the cosmopolitan alien ascidian *Botryllus schlosseri* in four Puget Sound marinas (Washington, USA) using eight polymorphic microsatellites. Up to seven sampling sessions over a period of 19 years revealed populations with fluctuating allelic richness ($AR = 2.693\text{--}4.417$) and expected heterozygosity ($He = 0.362\text{--}0.589$). The populations were well differentiated on spatial and temporal scales and were subject to moderate genetic drift ($F_s' = 0.027\text{--}0.071$). The significant heterozygote deficiencies that were obtained, positive inbreeding coefficients (F_{is}), and population structure measures (F_{st}) revealed that no population was in Hardy-Weinberg equilibrium. Comparing these parameters with those from two Californian sites (Moss Landing and Santa Cruz, 1200 km south, invaded by *Botryllus* during the 1940s) revealed a connection between Moss Landing and Puget Sound, whereas Santa Cruz remained isolated. On the US West Coast scale, this study revealed no significant difference in introduced population dynamics between recently established populations and those established over 60 years ago, except for fewer alleles and lower He . When comparing ten worldwide sites, only a few microsatellite loci displayed strong regional differences. Globally, the Puget Sound *Botryllus* populations exhibit genetic characteristics of recently established populations, as they have the lowest number of alleles and lowest genetic indices, further emerging as one of the youngest *B. schlosseri* populations worldwide.

1. Introduction

Biological invasions have become more frequent, driven by increasing globalization, anthropogenic activities, and climate change (Hulme, 2009; Van Kleunen et al., 2015). They cause severe negative impacts on local biota while also imposing economic consequences (Gallardo et al., 2016; Walsh et al., 2016). The footprints of biological invasions can be traced to changes in species diversity, dramatic alterations in communities and habitats, top-down and bottom-up control modifications, shifts in food chains, changes in nutrient cycling, and the attenuation of ecosystem services (Anton et al., 2019; David et al., 2017; Simberloff and Rejmánek, 2011). Similar to terrestrial ecosystems, biological invasions in the marine or oceanic realm are major drivers of ecological and evolutionary shifts, altering community structures and restructuring ecosystem functions with direct and indirect impacts on

ecosystem services (Carlton and Geller, 1993; Darling et al., 2017; Katsanevakis et al., 2014). With the increasing impact of biological invasions in marine environments, more interest is being shown in the inclusion of genetic, phylogenetic, and evolutionary aspects of research, with parameters that may improve the resolution and cost effectiveness of monitoring biological invasions (Darling et al., 2017; Rius et al., 2015) and describe changes in population genetics and adaptation properties of important invasive species in greater detail (Barrett, 2015; Tepolt, 2015).

Botryllus schlosseri (Pallas, 1766) is a common colonial ascidian species in the Mediterranean Sea and European Atlantic (Reem et al., 2022) that has been introduced to temperate zones worldwide (Freeman et al., 2016; Lin and Zhan, 2016; Lord, 2017; Reem and Rinkevich, 2014). It is currently found on all continents except Antarctica, including the coasts of Japan, New Zealand, India, South Africa, Chile,

* Corresponding author. Okinawa Institute of Science and Technology, Onna-son, Okinawa, 904-0495, Japan.

E-mail address: jann.zwahlen@gmx.net (J. Zwahlen).

<https://doi.org/10.1016/j.ecss.2022.107840>

Received 19 July 2021; Received in revised form 17 March 2022; Accepted 1 April 2022

Available online 5 April 2022

0272-7714/© 2022 Elsevier Ltd. All rights reserved.

Argentina, the USA, and Canada (Ben-Shlomo et al., 2001, 2010; Rinkevich et al., 1992; Van Name, 1945). Populations of this species were assigned to five highly divergent clades, based on cytochrome oxidase subunit I (COI) by López-Legentil et al. (2006) and then by Bock et al. (2012), with just one of the clades being cosmopolitan, which revealed significant differentiation between native and introduced populations (Lin and Zhan, 2016), while the other clades were restricted to European waters. Reem et al. (2017) pointed, however, to the possibility of admixture within two Mediterranean populations via the mixing of individuals from different clades, further revealing that the origin of *B. schlosseri*, particularly cosmopolitan clade A, is still under debate. Early hypotheses (Van Name, 1945) suggested that the species originated in European waters, a proposal supported by (Reem et al., 2017), but (Yund et al., 2015) proposed that at least one haplotype in clade A is native to the northwest Atlantic (Carlton, 2005). proposed, without supporting documentation, a possible Pacific origin.

In the United States, *B. schlosseri* was already present on the east coast in 1841 (Gould, 1841), while west coast observations (e.g., San Francisco Bay, Port Hueneme, and San Diego, CA, as well as Bremerton, WA) were recorded between 1944 and 1947 (US Navy, 1951). In the San Francisco area, it remained at low frequencies for at least a decade (Cohen and Carlton, 1995) while becoming common in the San Diego region in the early 1960s (Lambert and Lambert, 1998). In the state of Washington, *B. schlosseri* was not reported for 36 years following the first report in 1951 (US Navy, 1951), until its first validated documentation (Lambert et al., 1987). Lambert and Lambert (1998) reported an earlier anecdotal observation of *B. schlosseri* (late 1960s or early 1970s) in an oyster farm on the San Juan Island, just north of the Puget Sound. However, subsequent publications did not mention *B. schlosseri* (Kozloff, 1973, 1974, 1983; Lambert, 1969) in Washington. This species was not recorded by James T. Carlton, who surveyed the biofouling communities in Washington from 1976 to 1977, and the mention of *B. schlosseri* (Wonham and Carlton, 2005) is now considered a taxonomic error (J.T. Carlton, Williams College, Williamstown, MA, personal communication, August 2019). Furthermore, in 1987, one of the authors of this study (B. Rinkevich; unpubl.) performed a survey of marinas along the west coast of the USA and could not find *B. schlosseri* north of the Coos Bay, Oregon (an area that was still undergoing active invasion), as *B. schlosseri* failed to establish itself in the upper Coos Bay communities (Hewitt, 1993), or in several marinas surveyed in the Puget Sound. In contrast, *B. schlosseri* was very common in some marinas sampled in a subsequent survey conducted in 1999. This suggests possibly patchy early distributions of *B. schlosseri* introduced to the Seattle area. It is unknown whether the current widespread populations of *B. schlosseri* in this area (Cohen et al., 1998; Pleus et al., 2008) originated from earlier invasions reported during the 1940s and 1970s or reflect later introductions to this area.

As an introduced species, *B. schlosseri* negatively affects aquaculture organisms (Arens et al., 2011; Carver et al., 2006). With a short larval duration of no more than 2 h (Grave and Woodbridge, 1924; Grosberg, 1987; Rinkevich and Weissman, 1987), which allows only limited larval dispersal (Grosberg, 1987), *B. schlosseri* relies on floating objects for long-distance dispersal. It is commonly observed on ship hulls and submerged aquaculture infrastructure that moves between marine sites (Berrill, 1950; Lambert and Lambert, 1998; Skerman, 1960), and it also spreads to different marinas, most likely through boating activity (Lacoursière-Roussel et al., 2012; López-Legentil et al., 2015).

Long-term monitoring of population genetics (García-Navas et al., 2015; Osborne et al., 2012) may further elucidate the properties of introduced species (Kennington et al., 2012) and the roles of their genetic background in dictating the invasive potential of alien species introduced into new territories (Wellband et al., 2017). (Reem et al., 2013a) and Karahan et al. (2016) were the first to study the long-term genetic structures (13 and 12 years, respectively) of *B. schlosseri* in Santa Cruz and Moss Landing, two sites in California, USA, separated by only 20 km of shoreline, revealing two disparate population genetic structures. The Santa Cruz population sampled between 1995 and 2008

(Reem et al., 2013a) was relatively isolated, under high genetic drift, and further characterized by high mutation rates. The Moss Landing population, studied between 1996 and 2008 (Karahan et al., 2016), was affected by episodic freshwater floods and subsequent recovery.

These long-term studies on *Botryllus schlosseri* populations (Karahan et al., 2016; Reem et al., 2013a) detailed the population genetic parameters of two populations that had already been established for several decades. It is therefore of great interest to compare their parameters with those of recently established *B. schlosseri* populations, such as those on the US North Pacific coast (Puget Sound, Washington), where in the late 1980s, this species was missing or only just introduced. We studied the genetic parameters of four *B. schlosseri* populations residing in marinas in the Seattle, WA area, separated by up to 124 km coastline, during a period of up to 19 years (exact patterns of introduction are unknown). Thus, a major aim was to gain a perspective on possible fluctuations in genetic structures on a regional scale in recently introduced populations of this globally invasive species.

2. Materials and methods

2.1. Sampling sites

The Puget Sound (Fig. 1) is part of the Salish Sea, an estuarine system in Washington State, USA, which is linked to the Pacific Ocean via three connections to the Strait Juan de Fuca. Four marinas were studied: three are located in the central basin (Edmonds (47°48'26"N, 122°23'34"W), Shilshole (47°40'52"N, 122°24'18"W), and Des Moines (47°24'2"N, 122°19'47"W) marinas), as well as in Shelton Yacht Club (47°12'52"N, 123°5'5"W) in the southern basin. With 36,000 recreational boats (approximately 24,000 actively cruising), the Salish Sea is a famous destination for recreational boating (City of Des Moines Marina, http://www.desmoinesmarina.com/uploads/7/2/2/4/72248139/city_of_des_moines_marina_assessment_draft_3-15-19.pdf). The Shilshole Marina has approximately 1500 mooring slips, and it is the largest marina in this study. The City of Des Moines Marina usually has anywhere between 400 and 800 boats moored, with a constant flow of 10–30 daily visitors (Tara Reilly, Office Specialist of the City of Des Moines Marina, pers. comm.). Edmonds Marina has 668 mooring slips

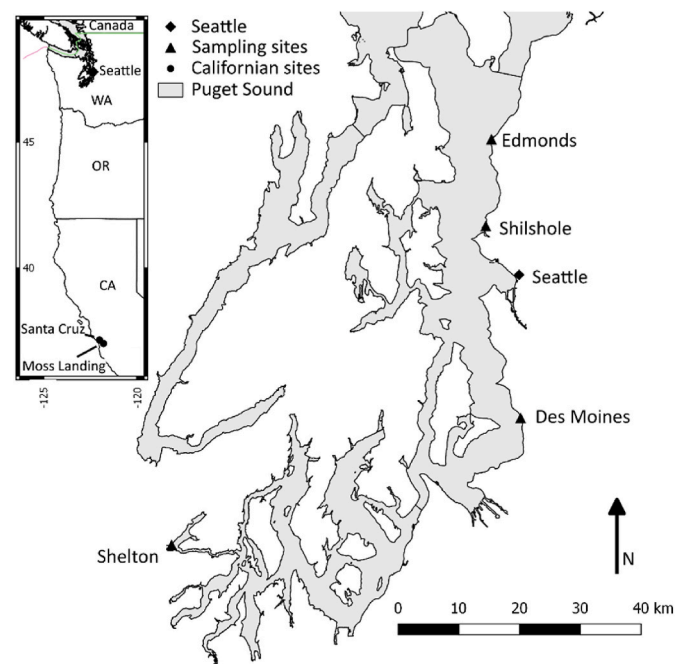


Fig. 1. Puget Sound collection sites and the two Californian reference sites on a map of the west coast of the USA. WA: Washington; OR: Oregon; CA: California.

and receives approximately 3100 visiting boats per year. The remote Shelton Yacht Club offers only 109 mooring slips and is surrounded by extensive shellfish aquaculture.

2.2. Sampling procedures, DNA extraction, and microsatellite genotyping

Botryllus schlosseri samples were collected every two years from 1999 to 2007 and in 2013 and 2018 (seven collection sessions; Suppl. Table A1). Colonies or colony fragments separated by at least 1 m were removed using single-edge razor blades and placed into separate 1.5 ml vials with 100 μ L lysis buffer. The DNA extraction was carried out according to the protocol in (Paz et al., 2003). In brief, 20 μ L NaClO₄ and 120 μ L phenol/chloroform/isoamyl alcohol (25:24:1 v:v:v) were added to the homogenized samples. After mixing and centrifugation (10,000 \times g for 5 min), the aqueous solution was transferred into another tube, and 120 μ L of chloroform-isoamyl alcohol (24:1 v/v) was added. Mixing and centrifugation were repeated, and DNA was precipitated by adding 100% ethanol and centrifuging (10,000 \times g for 15 min at 4 °C). The DNA was washed with 70% ethanol, allowed to dry, and then dissolved and diluted (1:20) with double distilled water.

Eight *B. schlosseri* microsatellites (Suppl. Table A2), BS-2, BS-8, BS-9 (Abdoulaye et al., 2010), BS-811 (Pancer et al., 1994), PB-29, PB-41, PB-49, and PBC-1 (Stoner et al., 1997), were separately amplified using polymerase chain reaction (PCR) with specific primers under the following conditions: 95 °C for 4 min; 35 \times [94 °C, 60 s; 48–56 °C, 60 s; 72 °C, 60 s]; 72 °C for 45 min; 10 °C. The PCR-mix (10 μ L) contained 5 μ L of 2 \times Taq PCR Master Mix (KT201, Tiangen Biotech, Beijing, China), 3.9 μ L double-distilled water 0.1 μ L primer mix, and 1 μ L DNA. The PCR products were run on 1.5% agar for gel electrophoresis in order to determine amplification success. When unsuccessful, PCR was repeated several times using deviant protocols with different annealing temperatures (within the range of 48–56 °C) or the protocol described in (Bock et al., 2011). Next, 1 μ L of PCR product mix of four primers was added to 0.4 μ L LIZ 500 size standard and 8.6 μ L formamide per well and analyzed using the ABI-PRISM 310 sequencer at the Technion's Biomedical Core Facility, Haifa, Israel. The length of the microsatellite alleles was evaluated using GeneMapper 5.0 (Applied Biosystems) software package. When in doubt (<5% of the cases), peak sizes were shared and discussed with other researchers, and amplifications were repeated when necessary. The majority of DNA extractions and genotyping were carried out in spring 2019 to ensure consistent amplification and genotyping success.

2.3. Data analysis

The analysis considered four geographical scales. At the local scale, each site (Des Moines, Edmonds, Shilshole, and Shelton) was analyzed separately. At the regional scale, all four sites were compared simultaneously. On the US west coast scale, the results for the Puget Sound area were compared with those of two southern sites in central California, Santa Cruz (Reem et al., 2013a) and Moss Landing (Karahana et al., 2016), both of which were subjected to the same methodology as that of the current study. Finally, allelic patterns from sites worldwide were compared at the international scale. Here, a “population” was considered to be the collected individuals from any single sampling location in a given year, unless otherwise stated.

The raw allele sizes were binned using AutoBin v 0.9 (Guichoux et al., 2011), a program that sorts alleles by size to detect relevant size changes for the proposed alleles and checks for scoring errors using Micro-Checker v. 2.2.3 (Van Oosterhout et al., 2004). The frequency of null alleles was calculated using FreeNA with the expectation maximization (EM) algorithm (Chapuis and Estoup, 2007). Null alleles might bias the outcome of the analysis; therefore, pairwise population differentiation (Fst) was calculated with and without the exclusion of null allele (ENA) correction in FreeNA using 50,000 bootstrap iterations (Aglieri et al., 2014). A two-tailed *t*-test was used to compare the two Fst

estimates.

Allele frequencies, private alleles (for each location, for each sampling year, and for the entire list of samples), observed and expected heterozygosity, population differentiation (Dest), population structure (Fst), and the inbreeding coefficient (Fis) (both using the G-statistic approach), and Nei's genetic distances were calculated and Mantel tests were performed using GenAlEx v.6.51b2 (Peakall and Smouse, 2012). For the Mantel tests, the temporal distance (years) was used for within-site analyses, and the geographical distance (km) was used for between-marina analyses. All of these were computed with both Fst and Nei's genetic distance indices, as both have been used previously (for example, Pineda et al., 2016; Reem et al., 2013a). Allelic richness was calculated in FSTAT (Goudet, 2003) using the rarefaction approach (Hurlbert, 1971) as modified by Petit et al. (1998), while HP-Rare (Kalinowski, 2005) was used to calculate private allelic richness. The significance of deviation in heterozygosity was tested using the online version of GENEPOP (<http://www.genepop.curtin.edu.au>). Pairwise Fst, a measure of population structure commonly used to show population differentiation, was calculated in Arlequin (Excoffier et al., 2005). We further grouped samples from the earlier phase (1999–2007) and then the later (2013–2018) phase of collection to reveal differences between early and more recently established populations. Bayesian analysis of population structure (BAPS) (Corander et al., 2004) and STRUCTURE v.2.1 (Pritchard et al., 2000) were used for the Bayesian structure analysis, and BAPS was used to determine the gene flow between different populations, both on a temporal and spatial scale. In BAPS, we used the clustering groups of individual options for the analysis, and in STRUCTURE, the length of the burn-in period was set to 100,000, while the number of Markov chain Monte Carlo repetitions was set to 10⁶. The number of clusters (K) was allowed to range between one and ten, and five iterations were carried out for each K. The optimal K was chosen using STRUCTURE HARVESTER (Earl and vonHoldt, 2012), and the different runs were aligned using CLUMPP (Jakobsson and Rosenberg, 2007). This step was repeated thrice to evaluate whether the number of clusters was consistent. Population clustering should be used with caution owing to the different assumptions of the programs (Sinai et al., 2019); therefore, NetStruct was used as a third method for analyzing the population structure using network theory (Greenbaum et al., 2015). After an initial run, the threshold was set between 0.2 and 0.4, and 999 permutations were carried out. The genetic drift (Fs') between different years and locations was calculated using TempoFS (Jorde and Ryman, 2007).

3. Results

Altogether, 533 samples were collected (521 analyzed) over a period of 19 years, between 1999 and 2018, in four locations in the Puget Sound area, WA, USA. The detailed allele frequencies of all eight microsatellite loci are summarized in Suppl. Table A3. A two-tailed *t*-test between the Fst values (with and without ENA correction) did not show significant results (*p* = 0.463), indicating that null alleles did not influence the results. The inbreeding coefficient (Fis) was highly significant in all populations.

3.1. Local scale

3.1.1. Des Moines marina

In total, 190 tissue samples were collected from seven sampling dates (years 1999–2018; Suppl. Table A4). All eight microsatellite loci were polymorphic, with 71 alleles in total. About one third of the total alleles (23/71) appeared in only a single sampling period: eight private alleles were found in 2018, three in each of the sampling years 1999, 2003, and 2013 and two in 2001, 2005, and 2007. Null alleles were detected at all loci, with high occurrence rates (>0.1) in PB-41 (0.284), BS-811 (0.255), PB-49 (0.229), BS-8 (0.18), PB-29 (0.175), and BS-9 (0.126). The genetic diversity indices are summarized in Table 1 and reveal

Table 1

Indices of population genetics for the Puget Sound marinas. Combined values for all eight loci. S.E.: standard error; N: mean number of samples; NA: mean number of alleles; AR: allelic richness; PAR: private allelic richness; Ho: observed heterozygosity; He: expected heterozygosity; Fis: inbreeding coefficient, calculations of average per locus. *: Ho significantly differs from He ($p < 0.05$).

Des Moines							
Year	N	NA	AR	PAR	Ho	He	Fis
1999	29.625	5.250	3.829	0.200	0.376*	0.544	0.303
S.E.	0.596	1.306			0.073	0.080	0.116
2001	23.625	4.375	3.724	0.180	0.230*	0.521	0.596
S.E.	0.498	0.865			0.075	0.091	0.114
2003	22.250	5.250	4.266	0.300	0.277*	0.578	0.533
S.E.	0.818	1.176			0.060	0.078	0.087
2005	26.375	4.625	3.820	0.200	0.258*	0.544	0.619
S.E.	1.194	1.017			0.096	0.095	0.147
2007	23.625	4.500	3.489	0.170	0.282*	0.479	0.514
S.E.	1.603	1.150			0.080	0.088	0.120
2013	23.375	5.125	3.864	0.230	0.340*	0.530	0.377
S.E.	0.460	1.217			0.064	0.076	0.086
2018	24.625	5.500	4.417	0.360	0.332*	0.589	0.443
S.E.	1.179	1.254			0.074	0.064	0.102
TOTAL	24.786	4.946	3.916	0.191	0.299	0.541	0.484
	0.470	0.414			0.028	0.030	0.043
Shilshole							
Year	N	NA	AR	PAR	Ho	He	Fis
2001	24.250	4.625	3.889	0.24	0.243*	0.516	0.525
S.E.	1.031	0.981			0.071	0.106	0.096
2003	22.250	4.875	3.825	0.230	0.328*	0.527	0.388
S.E.	0.773	1.246			0.097	0.097	0.128
2005	11.625	4.500	4.031	0.290	0.252*	0.473	0.451
S.E.	0.596	1.165			0.076	0.113	0.122
2007	27.250	5.000	3.798	0.310	0.264*	0.461	0.503
S.E.	0.996	1.254			0.098	0.104	0.129
2013	27.125	4.875	3.566	0.240	0.226*	0.397	0.470
S.E.	0.766	1.156			0.078	0.111	0.138
2018	27.375	5.000	3.612	0.260	0.289*	0.419	0.320
S.E.	0.944	1.195			0.085	0.109	0.128
TOTAL	23.313	4.813	3.787	0.262	0.267	0.467	0.443
	0.876	0.452			0.033	0.042	0.049
Edmonds							
Year	N	NA	AR	PAR	Ho	He	Fis
2003	16.125	3.500	3.149	0.270	0.263*	0.466	0.371
S.E.	1.008	0.732			0.085	0.102	0.143
2005	24.500	5.250	4.014	0.460	0.344*	0.574	0.345
S.E.	0.866	1.398			0.077	0.092	0.110
2007	23.000	4.250	3.539	0.280	0.268*	0.518	0.469
S.E.	0.926	1.161			0.097	0.087	0.142
2018	26.875	4.875	3.796	0.370	0.276*	0.536	0.501
S.E.	0.953	1.217			0.080	0.090	0.110
TOTAL	22.625	4.469	3.624	0.345	0.288	0.524	0.420
	0.846	0.561			0.041	0.045	0.061
Shelton							
Year	N	NA	AR	PAR	Ho	He	Fis
1999	18.000	3.125	2.693	0.310	0.236*	0.362	0.355
S.E.	0.926	0.875			0.093	0.110	0.144
2003	23.375	4.250	3.437	0.690	0.201*	0.470	0.585
S.E.	2.044	0.648			0.074	0.090	0.123
2018	26.750	4.125	3.104	0.540	0.293*	0.434	0.318
S.E.	1.221	0.972			0.105	0.097	0.162
TOTAL	22.708	3.833	3.078	0.513	0.243	0.422	0.426
	1.107	0.477			0.051	0.056	0.082
Grand Total	23.600	4.644	3.693	0.278	0.279	0.497	0.449

fluctuating allelic richness and significant ($p < 0.05$) heterozygote deficiency, indicating that the populations were not in Hardy-Weinberg equilibrium (HWE). The highest peak in allelic richness occurred in 2018, followed by another peak in 2003, both of which coincided with the peaks in private allelic richness.

The Mantel test showed no significant correlation ($p = 0.438$) between either Fst and Nei's genetic distance or the time between the different sampling periods (Suppl. Table A5). Overall, the annual Des Moines population differed significantly ($p = 0.001$) (Table 2). Interestingly, the Dest of the populations from 1999 to 2007 exceeded that of

Table 2

Changes in the Puget Sound Botryllus schlosseri population genetics between early and late sampling periods (1999–2007 vs. 2013–2018). Dest: Measure of population differentiation; Fst: Coefficient for population structure measuring the ratio of gene flow to genetic drift; Fis: Inbreeding coefficient; Fs': Genetic drift. * $p < 0.05$; ** $p \leq 0.001$. Edmonds and Shelton are included for the total period comparisons.

		Des Moines	Shilshole	Edmonds	Shelton
Dest	Total	0.053**	0.023**	0.027*	0.021*
		±0.030	±0.019	±0.016	±0.014
	99-07	0.064**	0.007 ±	0.029*	0.003 ±
	13-18	±0.042	0.016	±0.015	0.017
Fst	Total	0.041**	0.030**	0.023**	0.030**
	99-07	0.051**	0.015*	0.025*	0.008
	13-18	0.018*	0.010		
Fis	Total	0.416**	0.406**	0.409**	0.356**
	99-07	0.428**	0.430**	0.395**	0.418**
	13-18	0.385**	0.358**		
Fs'	Total	0.071	0.027	0.069	0.069
	99-07	0.076	0.024	0.08	0.050
	13-18	0.048	0.039		

the 2013–2018 populations by 0.064 ($p = 0.001$) and 0.024 ($p = 0.062$). The estimated genetic drift (Fs') was even stronger between 1999 and 2007 (0.076) than between 2013 and 2018 (0.048). Pairwise population differentiation (Fst) values showed that samples from all but three pairs (the years 2005/2013, 2007/2013, and 2005/2018) were significantly different ($p < 0.05$) (Suppl. Table A6A). Despite the lack of similarities between the years, the BAPS analysis revealed only a single cluster. However, two clusters were identified using STRUCTURE (Fig. 2A). In 1999 and 2001, cluster 1 (red in Fig. 2A) was dominant, whereas the later years were more mixed, with a slight dominance of cluster 2 (yellow in Fig. 2A) in 2007 and 2013.

3.1.2. Shilshole marina

In total, 152 tissue samples were analyzed from the Shilshole marina. Sample collections were attempted seven times from 1999 to 2018, but 1999 was not included in the analysis because only a single colony was observed (not collected). It should also be noted that the 2005 collection yielded only 13 samples, despite increased collection efforts. All microsatellites were polymorphic, representing only a single BS-8 allele in five of the sampling periods (2001, 2003, 2005, 2007, and 2018) and only one allele of BS-9 in 2005. A total of 63 microsatellite alleles were found, with 15 private alleles occurring during a single sampling period (Suppl. Table A4). Four private alleles occurred in 2018, three in 2013 and 2007, two in 2005 and 2003, and one in 2001. The null alleles were frequent (>0.1) at BS-811 (0.302), PB-41 (0.270), PB-49 (0.141), PB-29 (0.127), PBC-1 (0.126), and BS-9 (0.101). The summarized genetic indices (Table 1) revealed a gradual decline in gene diversity (He) and that the populations were not in HWE. The highest value of allelic richness was reached in 2005, which was slightly higher than that in the two preceding sampling years, whereas the peak in private allelic richness was reached in 2007. Both Fst and Nei's genetic distance indices and the time between the sampling periods were significantly correlated ($p = 0.02$ and $p = 0.041$, respectively) (Suppl. Table A5). Each run in STRUCTURE resulted in four clusters (Fig. 2C). Cluster 1 (blue in Fig. 2C) was more dominant in 2001 and 2003, cluster 2 (pink) had the same level across all years, while clusters 3 (yellow) and 4 (red) were more dominant in 2005. BAPS suggested two clusters, years 2001–2003 and 2005–2018, which is in line with the results of STRUCTURE analysis. Pairwise Fst values suggested that samples from 2001 to 2003 were

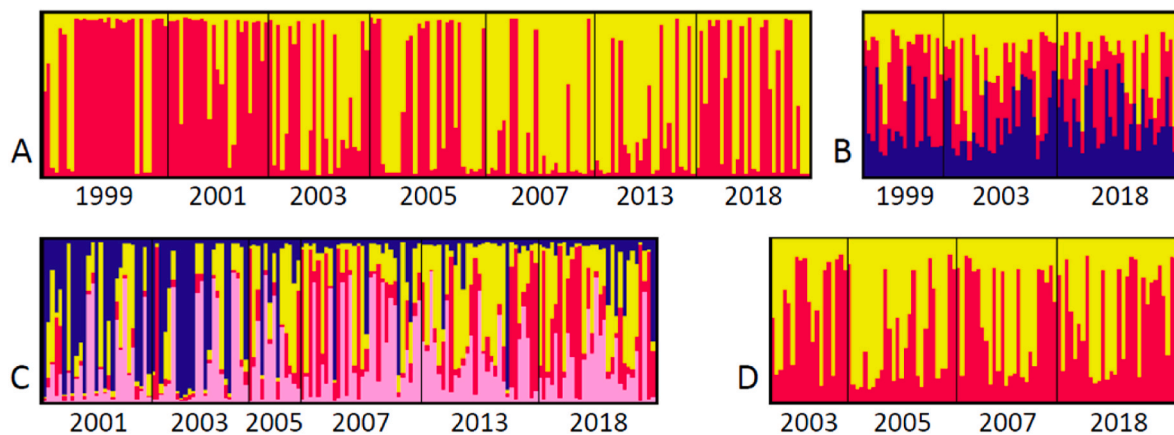


Fig. 2. Clusters obtained from STRUCTURE, visualized with Ghostscript. A: Des Moines ($K = 2$); B: Shelton ($K = 3$); C: Shilshole ($K = 4$); D: Edmonds ($K = 2$).

significantly different from those from 2007, 2013, and 2018. However, the 2007 and 2013 samples were also significantly different ($p < 0.05$) (Suppl. Table A6B). These F_{st} values further validated the clustering of 2001 and 2003. There was a weak but significant ($p \leq 0.001$) overall differentiation of the populations (Dest) and a slight genetic drift (F_s) in Shilshole (Table 2). Interestingly, F_s was an order of magnitude lower in 2001 and 2003 (0.004) than that in 2013–2018 (0.039). The lower genetic drift between 2001 and 2003 was congruent with the clustering of these two periods.

3.1.3. Edmonds marina

Four sampling periods yielded a total of 101 tissue samples. In an additional sampling session (1999), no colonies were found on the marina's hard-bottom shallow substrates. All microsatellite loci were polymorphic; however, only a single allele was found in 2003 and 2018 at locus BS-9. Among the 55 alleles (Suppl. Table A4), 17 were private alleles: two private alleles in 2003 and five in each of the other sampling years. The null alleles were frequent at PB-41 (0.351), BS-811 (0.279), PB-29 (0.172), BS-8 (0.166), and PB-49 (0.108). Genetic indices (Table 1) revealed an increase in allelic richness and gene diversity from 2003 to 2005, with a peak in allelic richness and private allelic richness in 2005. There was no correlation between Nei's genetic distance or F_{st} and the time between the sampling periods (Suppl. Table A5). Pairwise F_{st} values suggested a significant difference between the samples from the years 2003/2005, 2003/2018, and 2005/2018 ($p < 0.05$) (Suppl. Table A6C). The weak Dest was significant ($p = 0.006$), and the F_s was moderate (Table 2). The BAPS analysis suggested two clusters: one for 2003 and the second for 2005–2018. Nevertheless, the inconsistent K number suggested by STRUCTURE showed a very weak population structure. Two clusters were finally assigned owing to the constant (but not always tall) Delta K peaks at $K = 2$ (Fig. 2D). The BAPS results were confirmed using STRUCTURE, with 2003 being assigned to a single cluster, whereas the other three sampling dates failed to group into a specific cluster.

3.1.4. Shelton marina

Shelton marina was visited during only three sessions, yielding 78 tissue samples. All microsatellite loci were polymorphic, although just a single allele was represented each in 1999 at loci BS-8, BS-9, and PB-41, in 2003 at locus BS-9, and in 2018 at locus BS-8. Forty-six alleles were recorded, of which 19 were private (e.g., found on a single sampling date) (Suppl. Table A4). Most private alleles occurred in 2003 ($n = 9$), followed by 2018 ($n = 7$) and 1999 ($n = 3$). Null alleles were frequent at four loci (PB-41, PB-49, BS-811, and PB-29), ranging from 0.220 to 0.268. The genetic indices (Table 1) revealed peaks in allelic richness and private allelic richness in 2003. F_{st} and Nei's genetic distance indices were not correlated with the time elapsed between the sampling

events (Suppl. Table A5). STRUCTURE suggested three clusters (Fig. 2B); however, no meaningful trend was detected. BAPS, on the other hand, grouped the years 1999 and 2003 into one cluster and 2018 as the second. The different populations were weakly ($p = 0.014$) separated according to Dest (Table 2), and F_s was moderate. The pairwise F_{st} showed significant differentiation between the samples from 1999/2018 and 2003/2018 (Suppl. Table A6D), which is in line with the clusters suggested by BAPS.

3.2. Regional scale - Puget Sound

3.2.1. Overall analysis

At a regional scale, 104 alleles were recorded during the 19-year sampling period. Nineteen private alleles were found in Des Moines (26.8% of local alleles), eight (17.4%) in Shelton, nine (14.5%) in Shilshole, and eight (14.5%) in Edmonds. Analyses of the three most frequent alleles per locus (Fig. 3) revealed that only a single allele at loci BS-8 and BS-9 (181 bp and 194 bp, respectively) was dominant every year in each location, whereas loci BS-811, PB-29, and PBC-1 showed variable allele frequencies. In Edmonds, the third most frequent allele at locus BS-811 was frequent in 2003 and 2018 but absent in the years between. Only eight alleles were present in every sampled population, alleles 186 bp and 189 bp at BS-2, alleles 152 bp and 156 bp at PB-29, and alleles 199 bp, 202 bp, 205 bp, and 210 bp at PBC-1. Some alleles reflected site-specific distributions. For example, an allele of 178 bp at microsatellite BS-2 was present in all Shilshole and Shelton populations but was absent in Des Moines and Edmonds populations. In contrast, allele 192 bp at microsatellite BS-8 was present during every sampling period in Des Moines and Edmonds populations but completely absent in Shilshole and Shelton populations.

The Des Moines populations were almost twice as differentiated (Dest) as the populations of the other three marinas (Table 2), a result further supported by the population structure F_{st} , revealing the highest values for Des Moines. For both the indices, the differences between the four locations (Dest and F_{st}) were higher during the earlier period (1999–2007) than between 2013 and 2018. For the inbreeding (F_{is}) index, only Shelton showed remarkably low values. The overall genetic drift values were similar in Des Moines, Edmonds, and Shelton and less than half in Shilshole. For allelic richness, the highest numbers were assigned to Des Moines (Table 1), followed by Shilshole, Edmonds, and Shelton. Des Moines also showed the highest H_e , which is also a measure of the evenness of allelic frequencies, similar to the number of effective alleles (Brown and Weir, 1983). These high levels were followed by those of Edmonds, Shilshole, and Shelton. The physical distance between marinas and Nei's genetic distance or F_{st} between the sites was not correlated (Suppl. Table A5).

Analysis of all 20 populations simultaneously resulted in four BAPS

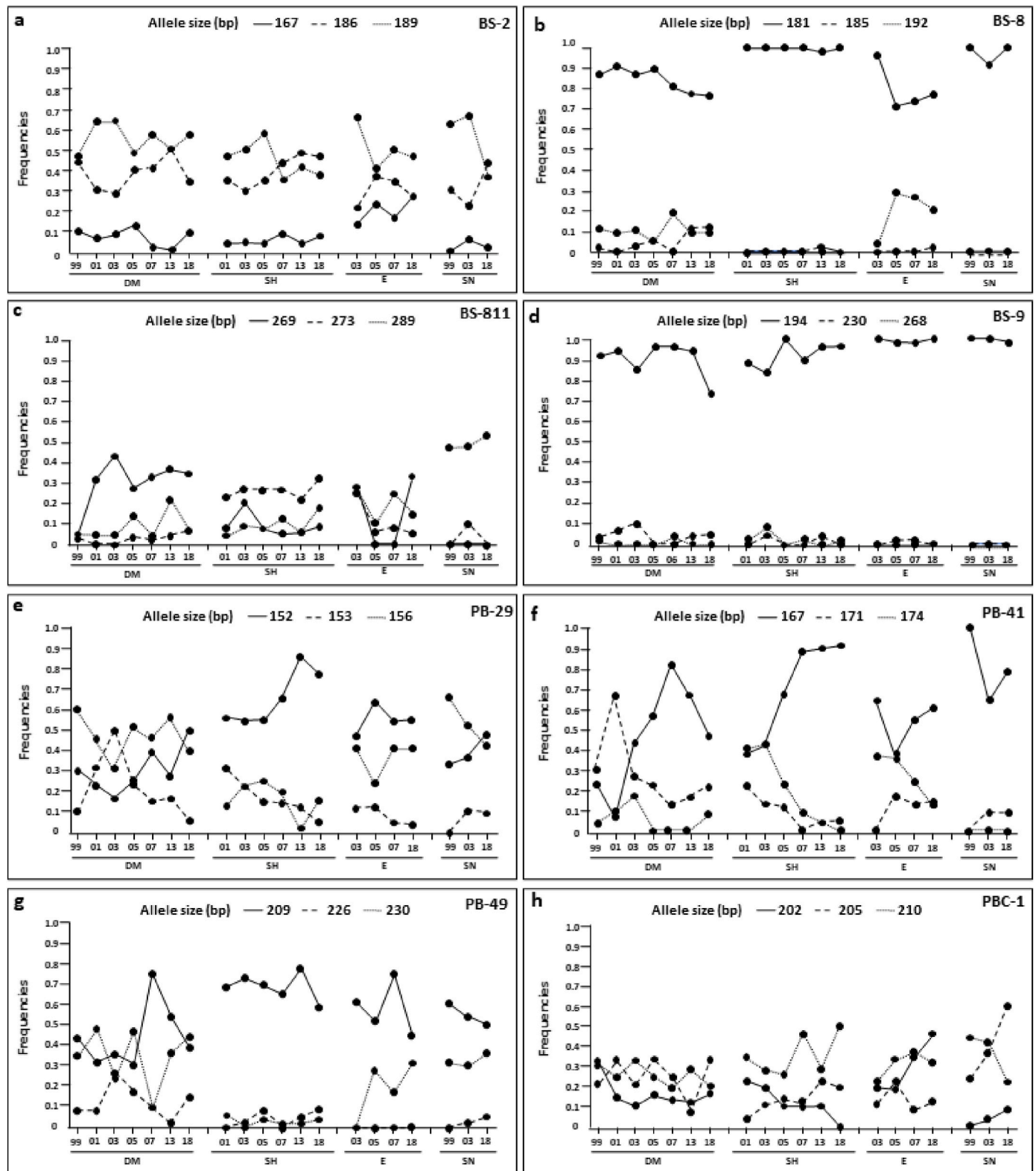


Fig. 3. Frequencies of the three most frequent alleles for each locus.

individual clusters, where each location was assigned to a separate cluster (Suppl. Fig. A1). STRUCTURE, however, suggested two clusters (Fig. 4A). The Des Moines samples were mainly assigned to cluster 1 (yellow in Fig. 4A), whereas the Shilshole samples belonged primarily to cluster 2 (red in Fig. 4A), with individuals from Shelton and Edmonds mixed together. Netstruc suggested three highly significant clusters

(Fig. 4B), with Des Moines belonging mainly to cluster one (blue in Fig. 4B), Shilshole to cluster two (red in Fig. 4B), and Shelton to cluster three (green in Fig. 4B), whereas Edmonds appeared to be well mixed.

Gene flow between sites during the study period was somewhat restricted (Suppl. Fig. A2), as Shelton received gene flow just from Shilshole and thus emerged as the most isolated site, with only 0.02 of

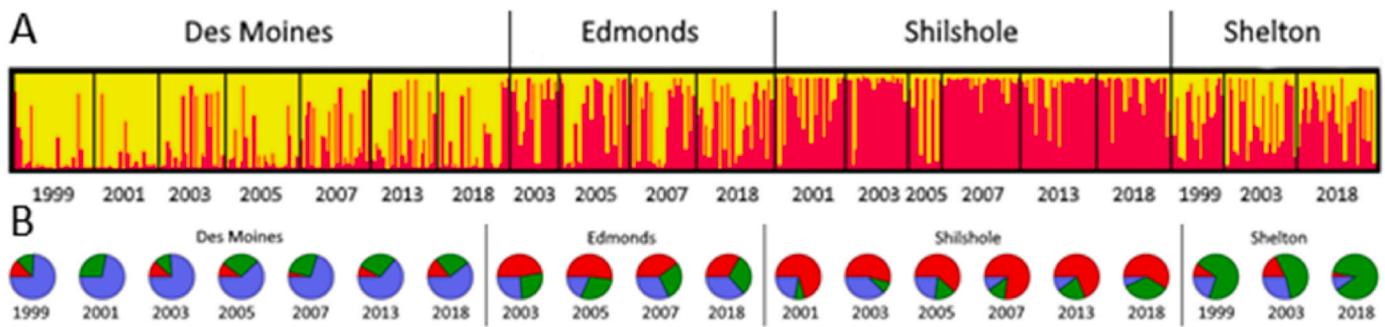


Fig. 4. Clusters of Puget Sound populations obtained for all locations and years simultaneously. A: STRUCTURE ($K = 2$), visualized with Ghostscript; B: NetStruc with threshold 0.256 and modularity 0.3 ($K = 3$).

gene flow from other locations, compared to 0.06 at Edmonds and Shilshole and 0.07 in Des Moines. The strongest gene flow was from Shilshole to Edmonds (0.031), followed by that from Des Moines to Shilshole (0.023). Edmonds and Shelton were not connected at all, while there was a unidirectional connection from Shelton to Des Moines (0.022) and bidirectional exchange with Shilshole. Comparing only the sites without yearly divisions showed a highly significant pairwise F_{st} for all pairs (Suppl. Table A6E).

3.2.2. Yearly analyses

Des Moines and Shilshole exhibited a similar number of private alleles on most sampling dates and generally the highest numbers (Table 3). However, in 2005, Edmonds had more private alleles than Des Moines and Shilshole. Interestingly, Shelton had fewer private alleles than Des Moines in 1999 and all other sites in 2018. On a yearly basis, pairwise F_{st} between years for the whole Puget Sound area showed that samples from all combinations except 2001/2003, 2003/2005, and 2007/2013 were significantly differentiated (Suppl. Table A6F). In both 2003 and 2018, the geographical distance was not correlated with either Nei's genetic distance or F_{st} between the marinas (Suppl. Table A5).

3.3. US west coast scale

The analyses included the Puget Sound sites, and two Californian sites, Santa Cruz (Reem et al., 2013a) and Moss Landing (Karahan et al., 2016), involving the five shared microsatellite loci (BS-811, PB-29, PB-41, PB-49, and PBC-1) used for analyses in these locations.

3.3.1. Overview

Allelic richness and expected heterozygosity in the Puget Sound populations were lower than those in the two Californian populations (Table 4). Despite similar allelic richness and expected heterozygosity, the Moss Landing and Santa Cruz populations were clustered into two different groups (orange and red in Fig. 5A) in STRUCTURE, and all four Puget Sound populations formed a third cluster (yellow in Fig. 5A). BAPS created three distinct clusters when the four Puget Sound sites were considered as a single location and no gene flow between any of the sites was discernible (Suppl. Fig. A4). Three highly significant clusters

were selected using Netstruct for the west coast analysis (Fig. 5B). Strikingly, the Santa Cruz populations clustered together as a single entity, whereas the Moss Landing populations were mostly associated with the remote Seattle populations, showing minimal similarities to the closer Santa Cruz cluster.

3.3.2. Separate years

During four sampling years (1999, 2001, 2005, and 2007), DNA samples were taken in at least one Californian and two Puget Sound sites. Analyses for each year separately revealed only a single genetic connection between California and Puget Sound (2007) and a substantial gene flow (0.039) from Des Moines to Moss Landing, whereas Santa Cruz, Shilshole, and Edmonds remained isolated (Fig. 6).

3.4. Global scale

For global scale analyses, we used ten sites (Table 4) and five shared microsatellite loci (BS-811, PB-29, PB-41, PB-49, and PBC-1). Locus BS-811 had the highest number of alleles at all the locations (20–65; Table 4). When considering the total number of individuals analyzed, Puget Sound emerged as the region with the lowest number of alleles per individual at either locus and at the combined loci. This places the whole Mediterranean area (Reem et al., 2017), Moss Landing (Karahan et al., 2016), and South America (Ben-Shlomo et al., 2010) as containing the highest numbers of allele per 100 colonies ($N = 57$ –66). The number of alleles in the Puget Sound was notoriously low ($N = 16$), primarily at locus PBC-1, presenting only 61.5% of alleles as compared to those of the second lowest site, Moss Landing. The European and Mediterranean locations (without Scandinavia) showed the highest number of alleles at PB-29, PB-41, and PBC-1 and were also leading in the number of alleles among the other two loci. The average expected heterozygosity among the Puget Sound populations was by far the lowest across the globe, suggesting a limited evenness of allele frequencies. Furthermore, allelic richness was among the lowest in Puget Sound, whereas the highest values were observed in Israel and Santa Cruz.

Analysis of frequent alleles (>0.1 in at least a single population; Suppl. Table A7) revealed some disparities among sites worldwide. For example, all the frequent alleles on locus PB-29 on the west coast were

Table 3

Private alleles (PA) obtained in the yearly analyses. # Samples: Total number of samples per site; Total PA: Sum of private alleles across all loci in a specific year; PA/sample: Sum of private alleles/total number of samples per site; DM: Des Moines; SH: Shilshole; E: Edmonds; SN: Shelton.

SITE	# SAMPLES	1999	2001	2003	2005	2007	2013	2018	Total PA	PA/Sample
DM	190	23	11	10	5	7	13	14	83	0.44
SH	152	–	13	9	5	10	11	8	56	0.37
E	101	–	–	3	11	6	–	6	26	0.26
SN	78	6	–	7	–	–	–	3	16	0.21
	Total PA	29	24	29	21	23	24	31	181	
	Total samples	48	48	84	63	74	51	106		
	PA/Sample	0.60	0.50	0.35	0.33	0.31	0.47	0.29		

Table 4

A summary for five microsatellite loci in 10 assigned populations worldwide. Area: Location of the study; N: Number of studied samples; BS-811, PB-29, PB-41, PB-49, PBC-1: number of alleles found at each locus (number alleles/N*100); Total: number of alleles over all loci; NA/100 specimens: total number of alleles per 100 specimens; AR: allelic richness (where provided); He: expected heterozygosity; Sources: 1: current study; 2: Karahan et al. (2016); 3: Reem et al. (2013a); 4: Stoner et al. (2002); 5: Reem et al. (2017); 6: Paz et al. (2003); 7: Ben-Shlomo et al. (2006); 8: Reem et al. (2013b); 9: Ben-Shlomo et al. (2010); 10: Ben-Shlomo et al. (2001).

Region	Area	N	BS-811	PB-29	PB-41	PB-49	PBC-1	Total	NA/100 specimens	AR	He
N. America	(1) Puget Sound	521	43 (8.3)	5 (1.0)	12 (2.3)	13 (2.5)	8 (1.5)	81	16	5.355	0.627
	(2) Moss Landing	141	36 (25.5)	5 (3.5)	13 (9.2)	13 (9.2)	13 (9.2)	80	57	6.05	0.745
	(3) Santa Cruz	278	56 (20.1)	6 (2.2)	11 (4.0)	32 (11.5)	22 (7.9)	127	46	8.596	0.762
	(4) US East Coast	99	32 (32.3)	4 (4.0)	3 (3.0)	6 (6.1)	–	45	45	–	–
Europe/Israel	(5) Mediterranean	288	65 (22.6)	12 (4.2)	27 (9.4)	37 (12.8)	41 (14.2)	182	63	9.683	0.875
	(6) Israel (Mediterranean)	323	36 (11.1)	14 (4.3)	23 (7.1)	25 (7.7)	31 (9.6)	129	40	–	0.898
	(7) Atlantic Europe	360	45 (12.5)	11 (3.1)	25 (6.9)	22 (6.1)	–	103	29	–	0.644
	(8) Scandinavia	319	49 (15.4)	7 (2.2)	12 (3.8)	20 (6.3)	20 (6.3)	108	34	6.642	0.62
Others	(9) South America	130	32 (24.6)	5 (3.8)	11 (8.5)	23 (17.7)	15 (11.5)	86	66	–	0.724
	(10) New Zealand	195	20 (10.3)	4 (2.1)	8 (4.1)	8 (4.1)	15 (7.7)	55	28	–	0.675

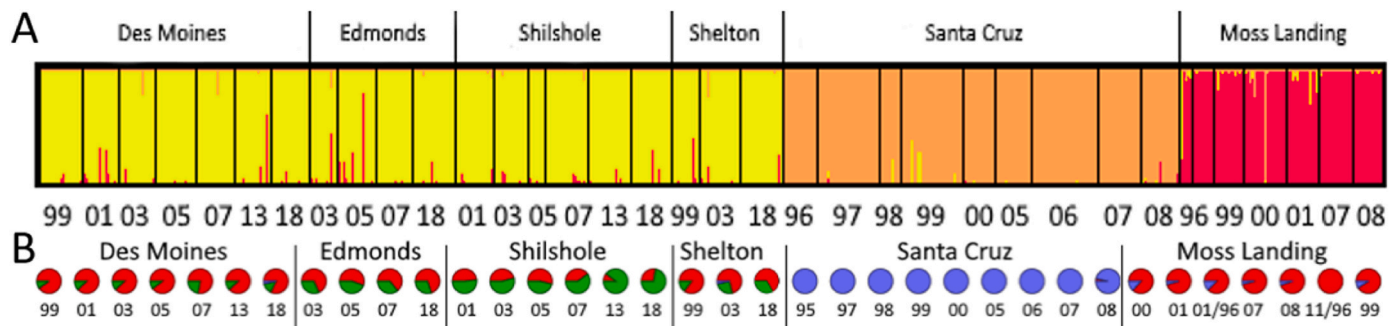


Fig. 5. Clusters obtained for the US west coast populations. A: STRUCTURE analysis, visualized with Ghostscript; B: Netstruct analysis with threshold 0.25 and modularity 0.34 ($K = 3$); Numbers show the year of collection (01/96 and 11/96 for January and November 1996, respectively).

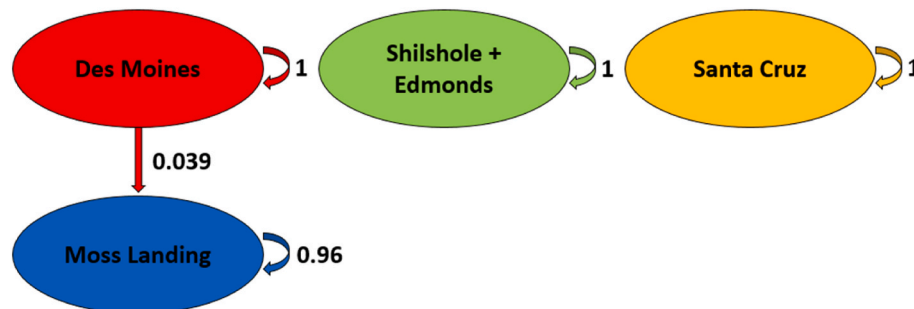


Fig. 6. Possible gene flow patterns computed by BAPS between Puget Sound and California populations in 2007. Cluster 1: Des Moines; Cluster 2: Shilshole/Edmonds; Cluster 3: Moss Landing; Cluster 4: Santa Cruz.

<160 bp, while in the other regions, at least one allele was >160 bp. At the most polymorphic locus, BS-811, most of the European, South American, and New Zealand frequent alleles were <260 bp in contrast to the North American alleles (<300 + bp). Locus PB-41 did not display different patterns among the locations, except for Israel (Paz et al., 2003). At most sites, the majority of frequent alleles were <180 bp; however, most alleles in the Israeli populations were >180 bp. For locus PB-49, allele sizes >240 bp were just found in Santa Cruz (Reem et al., 2013a), the US east coast, (Stoner et al., 2002), and the Mediterranean Sea (Reem et al., 2017), whereas in Israel (Paz et al., 2003), Scandinavia, (Reem et al., 2013b), and South America (Ben-Shlomo et al., 2010), the majority of frequent allele sizes were <220 bp. Locus PBC-1 contained alleles >210 bp everywhere except in the Puget Sound (this study), South America (Ben-Shlomo et al., 2010), and New Zealand (Ben-Shlomo et al., 2001). In the Mediterranean Sea (Reem et al., 2017), the Atlantic coasts of Europe (Ben-Shlomo et al., 2006), and South America (Ben-Shlomo et al., 2010) they were shorter, <190 bp. In New Zealand (Ben-Shlomo et al., 2001), most PBC-1 alleles were between 190 and 200

bp, while in the Puget Sound (this study), Moss Landing (Karahan et al., 2016), and Israel (Paz et al., 2003), sizes of 201–210 bp were most commonly observed.

4. Discussion

Botryllus schlosseri is a marine invertebrate (Berrill, 1950; López-Legentil et al., 2006; Paz et al., 2003; Reem et al., 2017) found in the Mediterranean sea and European Atlantic shallow waters (down to 200 m depth) (Berrill, 1950; López-Legentil et al., 2006; Paz et al., 2003; Reem et al., 2017) that has become a cosmopolitan alien species on man-made submerged hard-bottom substrates, primarily inhabiting marinas and harbors all over the temperate zones of the northern and southern hemispheres (Berrill, 1950; Ben-Shlomo et al., 2001, 2010; Lambert, 2001; Reem et al., 2013a,b, 2017). This species is also considered to be a fouling pest that is associated with economic costs in aquaculture (Arens et al., 2011; Carver et al., 2006). Traits such as fast adaptation to man-made environmental conditions (Lambert and

Lambert, 2003; Lambert, 2001) and enhanced mutation rates that increase genetic variability in newly established populations (Reem et al., 2013a) further contribute to *B. schlosseri*'s invasiveness in a way that, upon the establishment of pioneering colonies, this species quickly spreads to become one of the most common species in the hard-bottom invertebrate consortia (Lambert and Lambert, 1998, 2003; Locke et al., 2009; Martin et al., 2011). Many *B. schlosseri* populations beyond the Mediterranean and European Atlantic coasts have been established for more than five decades (e.g., in Africa (Millar, 1955) and South America (Orensanz et al., 2002)) and up to a century and more, as in the US east coast (Gould, 1841), California, (Van Name, 1945), and New Zealand (Van Name, 1945). Puget Sound populations are likely among the youngest regional populations of this species. Despite early mentions of *B. schlosseri* in a naval shipyard in the Seattle area (US Navy, 1951), this species has not been recorded in civil marinas for several decades. Indeed, the partial absence in our 1999 survey may indicate a very recent establishment and coincides with the finding of *B. schlosseri* in Des Moines and Shelton (Cohen et al., 1998), but not in Edmonds. Thus, Puget Sound provides a unique opportunity to study relatively recently established (ca. three decades ago) *B. schlosseri* populations.

The results of the present study reveal significantly differentiated populations on both a spatial and temporal scale. Moderate genetic drift (ranging from 0.027 to 0.071) within populations and limited gene flow (up to 0.031) between populations from different locations were also observed. Population clustering showed that the Des Moines and Shilshole populations were assigned to different clusters, whereas the Edmonds and Shelton populations represented a mix of the clusters. The low number of microsatellite alleles found in the Puget Sound populations compared to that of other sites worldwide is remarkable. These results differed from a study on European harbor populations of the solitary ascidian *Styela plicata*, which revealed differences only on a spatial scale but not on a temporal scale (Pineda et al., 2016).

The absence of *B. schlosseri* in two surveys conducted in Edmonds marina in 1998 (Cohen et al., 1998) and 1999 (this study) made this marina a candidate for the most recently established site (out of the four studied) for the *B. schlosseri* Puget Sound populations. Four years later (2003), the allelic indices of this newly established population significantly differed from those of the populations from the subsequent sampling period (two years later). In contrast, the Shilshole populations showed fewer fluctuations in allelic indices between the sampling points, despite only a single colony being found in 1999. This indication of a stable population is further supported by genetic drift, which was much lower at Shilshole than at the other three sites. Genetic indices showed diverse behaviors in the marinas studied. While the allelic richness (AR) and gene diversity (He) were highly fluctuating in Des Moines, Edmonds, and Shelton, these two indices slowly declined over time in the Shilshole marina. The two northernmost marinas, Edmonds and Shilshole, showed a peak in allelic richness during the 2005 sampling period, whereas in Des Moines and Shelton, the highest numbers were observed in 2018 and 2003, respectively. Only in Shilshole did the peaks of He (2003) and private allelic richness (2007) not coincide with those of the AR. The differences in genetic indices suggest that even close populations exhibit distinct genetic patterns. However, Shelton had by far the lowest values in genetic indices, all indicating its remoteness.

Heterozygote deficiency and significant inbreeding coefficients (Fis) found in all Puget Sound populations were in line with those of previous studies on the US west coast (Karahan et al., 2016; Reem et al., 2013a; Stoner et al., 2002) and worldwide populations (Ben-Shlomo et al., 2010; Lacoursière-Roussel et al., 2012; Paz et al., 2003; Reem et al., 2013b, 2017). Temporal fluctuations in allelic richness seem to be characteristic of *B. schlosseri* populations (this study) (Karahan et al., 2016; Reem et al., 2013a) and differ from long-term trends in isolated populations of other organisms, where a decreasing (García-Navas et al., 2015) or increasing (Jason Kennington et al., 2012) allelic richness was observed.

On the US west coast scale, the four Puget Sound populations were clustered together, despite more than 120 km separating the two furthest populations. In contrast, the Moss Landing (Karahan et al., 2016) and Santa Cruz (Reem et al., 2013a) populations in California were always assigned to separate clusters, despite being only 20 km apart. This indicates that, despite significant Fst and Dest values and different allelic richness peak timings, the Puget Sound populations at the west coast level remained genetically close. It is also of interest to find that Moss Landing populations are genetically closer to the Puget Sound populations than to the Santa Cruz populations, as revealed by the Netstruct clusters and gene flow charts, a result supported by (Reem et al., 2013a) with the assumption that Santa Cruz populations are isolated. Additionally, null alleles were frequent in both Puget Sound and Moss Landing populations (Karahan et al., 2016) but not in Santa Cruz populations (Reem et al., 2013a). The year 2007 was marked by the highest gene flow between the Puget Sound and Moss Landing populations, a year following a severe flood occurring in Moss Landing, where *B. schlosseri* was temporarily eradicated from that marina (Karahan et al., 2016). Thus, the genetic flow originating from the Puget Sound might have a footprint in the recolonization of the new population in the Moss Landing marina.

In the global comparison, Puget Sound's *B. schlosseri* populations had one of the lowest allelic richness and gene diversity values. These low numbers, even when compared to those of remote populations, such as of New Zealand (Ben-Shlomo et al., 2001) and South America (Ben-Shlomo et al., 2010), further illustrate the late establishment of the Puget Sound populations and can be explained by the limited time for the accumulation of mutations and inflow of genetic material from distant populations. The low allelic richness in Puget Sound could also reflect a low number of founding individuals. Furthermore, *B. schlosseri* microsatellite loci exhibited disparate repertoires of allele sizes and richness in different populations worldwide. While BS-811 had numerous alleles in all locations, other loci, such as PBC-1, revealed fewer alleles in regions recently invaded by *B. schlosseri* colonies. In addition, the amplification success of *B. schlosseri* microsatellites showed local differences even when following the same protocols. In the Puget Sound populations, the amplification success of BS-8, BS-9, and PB-41 was relatively low (up to 30% missing data), whereas the amplification of these loci in samples from the Israeli coast resulted in no failures (Tamir et al., 2022). Moreover, the native European populations (Reem et al., 2017; Reem and Rinkevich, 2014) revealed more alleles per microsatellite locus, further supporting the Mediterranean/Eastern Atlantic origin of *B. schlosseri*.

Studying newly established *B. schlosseri* populations over a period of 19 years in the Puget Sound revealed highly fluctuating patterns in genetic indices, such as allelic richness (AR), gene diversity (He), inbreeding coefficients (Fis), and population structures (Fst), showing a significant deviation from the HWE in all populations. Owing to these fluctuations, no temporal trend could be observed, and it is suggested that despite remarkable variations between the different sites, the Puget Sound populations remain isolated and still closely related. A comparison at a worldwide level revealed a considerably lower number of alleles in the Puget Sound population, supporting the recent introduction hypothesis.

CRedit authorship contribution statement

Jann Zwahlen: Data curation, Formal analysis, Writing – original draft, Writing – review & editing. **Eitan Reem:** Conceptualization, Data curation, Formal analysis. **Jacob Douek:** Conceptualization, Data curation, Formal analysis. **Baruch Rinkevich:** Conceptualization, Data curation, Funding acquisition, Writing – original draft, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

We would like to thank James T. Carlton for his valuable insights into the *B. schlosseri* invasion history in the Puget Sound area and G. Paz for creating Fig. 3. This study was part of Jann Zwahlen's Master's thesis. This work was supported by a grant from the Israeli Science Foundation (ISF) and by a grant from the United States-Israel Binational Science Foundation (BSF), Israel, as part of a joint program with the National Science Foundation (NSF), USA (NSF/BSF no. 2021650).

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ecss.2022.107840>.

References

- Abdoulaye, D., Acevedo, I., Adebayo, A.A., Behrmann-Godel, J., Benjamin, R.C., Bock, D.G., et al., 2010. Permanent genetic resources added to molecular ecology resources database 1 August 2009–30 September 2009. *Mol. Ecol. Resour.* 10, 232–236. <https://doi.org/10.1111/j.1755-0998.2009.02796.x>.
- Aglieri, G., Papetti, C., Zane, L., Milisenda, G., Boero, F., Piraino, S., 2014. First evidence of inbreeding, relatedness and chaotic genetic patchiness in the holoplanktonic jellyfish *Pelagia noctiluca* (Scyphozoa, Cnidaria). *PLoS One* 9. <https://doi.org/10.1371/journal.pone.0099647>.
- Anton, A., Gerdali, N.R., Lovelock, C.E., Apostolaki, E.T., Bennett, S., Cebrian, J., et al., 2019. Global ecological impacts of marine exotic species. *Nat. Ecol. Evol.* 3, 787–800. <https://doi.org/10.1038/s41559-019-0851-0>.
- Arens, C.J., Christine Paetzold, S., Ramsay, A., Davidson, J., 2011. Pressurized seawater as an antifouling treatment against the colonial tunicates *Botrylloides violaceus* and *Botryllus schlosseri* in mussel aquaculture. *Aquat. Invasions* 6, 465–476. <https://doi.org/10.3391/ai.2011.6.4.12>.
- Barrett, S.C.H., 2015. Foundations of invasion genetics: the Baker and Stebbins legacy. *Mol. Ecol.* <https://doi.org/10.1111/mec.13014>.
- Ben-Shlomo, R., Douek, J., Rinkevich, B., 2001. Heterozygote deficiency and chimerism in remote populations of a colonial ascidian from New Zealand. *Mar. Ecol. Prog. Ser.* 209, 109–117. <https://doi.org/10.3354/meps209109>.
- Ben-Shlomo, R., Paz, G., Rinkevich, B., 2006. Postglacial-period and recent invasions shape the population genetics of botryllid ascidians along European Atlantic coasts. *Ecosystems* 9, 1118–1127. <https://doi.org/10.1007/s10021-006-0141-y>.
- Ben-Shlomo, R., Reem, E., Douek, J., Rinkevich, B., 2010. Population genetics of the invasive ascidian *Botryllus schlosseri* from South American coasts. *Mar. Ecol. Prog. Ser.* 412, 85–92. <https://doi.org/10.3354/meps08688>.
- Berrill, N.J., 1950. *The Tunicata*. Bernard Quaritch, London.
- Bock, D.G., Zhan, A., Lejeune, C., MacIsaac, H.J., Cristescu, M.E., 2011. Looking at both sides of the invasion: patterns of colonization in the violet tunicate *Botrylloides violaceus*. *Mol. Ecol.* 20, 503–516. <https://doi.org/10.1111/j.1365-294X.2010.04971.x>.
- Bock, D.G., MacIsaac, H.J., Cristescu, M.E., 2012. Multilocus genetic analyses differentiate between widespread and spatially restricted cryptic species in a model ascidian. *Proc. Biol. Sci.* 279, 2377–2385. <https://doi.org/10.1098/rspb.2011.2610>.
- Brown, A.H.D., Weir, B., 1983. Measuring genetic variation in plant populations. In: *Isozymes in Plant Genetics and Breeding, Part A*, pp. 219–239.
- Carlton, 2005. Setting ascidian invasions on the global stage. In: *International Invasive Sea Squirt Conference*. Woods Hole Oceanic Institution Massachusetts, USA.
- Carlton, J.T., Geller, J.B., 1993. Ecological Roulette: the global transport of nonindigenous marine organisms. *Science* 261, 78–82. <https://doi.org/10.1126/science.261.5117.78>.
- Carver, C., Mallet, A.L., Vercaemer, B., 2006. *Biological Synopsis of the Colonial Tunicates, Botryllus Schlosseri and Botrylloides Violaceus*. Canadian Manuscript Report of Fisheries and Aquatic Sciences No. p. 2747.
- Chapuis, M.P., Estoup, A., 2007. Microsatellite null alleles and estimation of population differentiation. *Mol. Biol. Evol.* 24, 621–631. <https://doi.org/10.1093/molbev/msl191>.
- Cohen, A.N., Carlton, J.T., 1995. *Biological Study: Nonindigenous Aquatic Species in a United States Estuary: a Case Study of the Biological Invasions of the San Francisco Bay and Delta*. Springfield, Virginia, USA.
- Cohen, A., Mills, C., Berry, H., Wotham, M., Bingham, B., Bockheim, B., Carlton, J., et al., 1998. Puget Sound Expedition. A Rapid Assessment Survey of Non-indigenous Species in the Shallow Waters of Puget Sound, ... of Puget Sound. Olympia, Washington.
- Corander, J., Waldmann, P., Marttinen, P., Sillanpää, M.J., 2004. BAPS 2: enhanced possibilities for the analysis of genetic population structure. *Bioinformatics* 20, 2363–2369. <https://doi.org/10.1093/bioinformatics/bth250>.
- Darling, J.A., Galil, B.S., Carvalho, G.R., Rius, M., Viard, F., Piraino, S., 2017. Recommendations for developing and applying genetic tools to assess and manage biological invasions in marine ecosystems. *Mar. Pol.* 85, 54–64. <https://doi.org/10.1016/j.marpol.2017.08.014>.
- David, P., Thébaud, E., Anneville, O., Duyck, P.F., Chapuis, E., Loeuille, N., 2017. Impacts of invasive species on food webs: a review of empirical data. *Adv. Ecol. Res.* 56, 1–60. <https://doi.org/10.1016/bs.aecr.2016.10.001>.
- Earl, D.A., vonHoldt, B.M., 2012. Structure HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conserv. Genet. Resour.* 4, 359–361. <https://doi.org/10.1007/s12686-011-9548-7>.
- Excoffier, L., Laval, G., Schneider, S., 2005. Arlequin (version 3.0): an integrated software package for population genetics data analysis. *Evol. Bioinform. Online* 1, 117693430500100. <https://doi.org/10.1177/117693430500100003>.
- Freeman, A.S., Frischeisen, A., Blakeslee, A.M.H., 2016. Estuarine fouling communities are dominated by nonindigenous species in the presence of an invasive crab. *Biol. Invasions* 18, 1653–1665. <https://doi.org/10.1007/s10530-016-1108-3>.
- Gallardo, B., Clavero, M., Sánchez, M.I., Vilà, M., 2016. Global ecological impacts of invasive species in aquatic ecosystems. *Global Change Biol.* 22, 151–163. <https://doi.org/10.1111/gcb.13004>.
- García-Navas, V., Bonnet, T., Waldvogel, D., Wandeler, P., Camenisch, G., Postma, E., 2015. Gene flow counteracts the effect of drift in a Swiss population of snow voles fluctuating in size. *Biol. Conserv.* 191, 168–177. <https://doi.org/10.1016/j.biocon.2015.06.021>.
- Goudet, J., 2003. FSTAT (ver. 2.9.4), A Program To Estimate And Test Population Genetics Parameters. Available from. <http://www.unil.ch/izea/software/fstat.html>. Updated from Goudet 1995.
- Gould, A.A., 1841. *Report on the Invertebrata of Massachusetts, Comprising the Mollusca, Crustacea, Annelida, and Radiata*. Folsom, Wells, and ThursV.
- Grave, Caswell, Woodbridge, H., 1924. *Botryllus schlosseri* (Pallas): the behavior and morphology of the free-swimming larva. *J. Morphol. Physiol.* 39, 207–247. <https://doi.org/10.1002/jmor.1050390107>.
- Greenbaum, G., Templeton, A.R., Bar-David, S., 2015. Inference and Analysis of Population Structure Using Genetic Data and Network Theory. <https://doi.org/10.1101/024042> bioRxiv.
- Grosberg, R.K., 1987. Limited dispersal and proximity-dependent mating success in the colonial ascidian *Botryllus schlosseri*. *Evolution* 41, 372–384. <https://doi.org/10.2307/2409145>.
- Guichoux, E., Lagache, L., Wagner, S., Chaumeil, P., Léger, P., Lepais, O., Lepoittevin, C., Malaua, T., Revardel, E., Salin, F., Petit, R.J., 2011. Current trends in microsatellite genotyping. *Mol. Ecol. Resour.* 11, 591–611. <https://doi.org/10.1111/j.1755-0998.2011.03014.x>.
- Hewitt, C.L., 1993. Marine Biological Invasions: the Distributional Ecology and Interactions between Native and Introduced Encrusting Organisms. University of Oregon. <https://doi.org/10.1017/CBO9781107415324.004>.
- Hulme, P.E., 2009. Trade, transport and trouble: managing invasive species pathways in an era of globalization. *J. Appl. Ecol.* 46, 10–18. <https://doi.org/10.1111/j.1365-2664.2008.01600.x>.
- Hurlbert, S.H., 1971. The nonconcept of species diversity: a critique and alternative parameters. *Ecology* 52, 577–586. <https://doi.org/10.2307/1934145>.
- Jakobsson, M., Rosenberg, N.A., 2007. CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. *Bioinformatics* 23, 1801–1806. <https://doi.org/10.1093/bioinformatics/btm233>.
- Jason Kennington, W., Hevroy, T.H., Johnson, M.S., 2012. Long-term genetic monitoring reveals contrasting changes in the genetic composition of newly established populations of the intertidal snail *Bembicium vittatum*. *Mol. Ecol.* 21, 3489–3500. <https://doi.org/10.1111/j.1365-294X.2012.05636.x>.
- Jorde, P.E., Ryman, N., 2007. Unbiased estimator for genetic drift and effective population size. *Genetics* 177, 927–935. <https://doi.org/10.1534/genetics.107.075481>.
- Kalinowski, S.T., 2005. HP-RARE 1.0: a computer program for performing rarefaction on measures of allelic richness. *Mol. Ecol. Notes* 5, 187–189. <https://doi.org/10.1111/j.1471-8286.2004.00845.x>.
- Karahan, A., Douek, J., Paz, G., Rinkevich, B., 2016. Population genetics features for persistent, but transient, *Botryllus schlosseri* (Urochordata) congregations in a central Californian marina. *Mol. Phylogenet. Evol.* 101, 19–31. <https://doi.org/10.1016/j.ympev.2016.05.005>.
- Katsanevakis, S., Wallentinus, I., Zenetos, A., Leppäkoski, E., Çınar, M.E., Öztürk, B., Grabowski, M., Golani, D., Cardoso, A.C., 2014. Impacts of invasive alien marine species on ecosystem services and biodiversity: a pan-European review. *Aquat. Invasions* 9, 391–423. <https://doi.org/10.3391/ai.2014.9.4.01>.
- Kozloff, E.N., 1973. *Seashore Life of Puget Sound, the Strait of Georgia, and the San Juan Archipelago*. University of Washington Press, Seattle and London.
- Kozloff, E.N., 1974. *Keys to the Marine Invertebrates of Puget Sound, the San Juan Archipelago, and Adjacent Regions*. University of Washington Press, Seattle and London.
- Kozloff, E.N., 1983. *Seashore Life of the Northern Pacific Coast. An Illustrated Guide to Northern California, Oregon, Washington, and British Columbia*. University of Washington Press, Seattle and London.
- Lacoursière-Roussel, E., Bock, D.G., Cristescu, M.E., Guichard, F., Girard, P., Legendre, P., McKindsey, C.W., 2012. Disentangling invasion processes in a dynamic shipping-boating network. *Mol. Ecol.* 21, 4227–4241. <https://doi.org/10.1111/j.1365-294X.2012.05702.x>.

- Lambert, C.C., 1969. Urochordata. A key prepared by richard Snyder (1951) enlarged and revised by Charles Lambert. In: *Marine Fauna of the San Juan Archipelago*. University of Washington Friary Harbor Laboratories, p. 5.
- Lambert, G., 2001. A global overview of ascidian introductions and their possible impact on the endemic Fauna. *Biol. Ascidians* 249–257. https://doi.org/10.1007/978-4-431-66982-1_40.
- Lambert, C.C., Lambert, G., 1998. Non-indigenous ascidians in southern California harbors and marinas. *Mar. Biol.* 130, 675–688. <https://doi.org/10.1007/s002270050289>.
- Lambert, C.C., Lambert, G., 2003. Persistence and differential distribution of nonindigenous ascidians in harbors of the Southern California Bight. *Mar. Ecol. Prog. Ser.* 259, 145–161. <https://doi.org/10.3354/meps259145>.
- Lambert, Charles C., Lambert, Gretchen, Kozloff, E.N., 1987. *Phylum Urochordata*. In: *Marine Invertebrates of the Pacific Northwest*. University of Washington Press, Seattle and London, pp. 467–479.
- Lin, Y., Zhan, A., 2016. Population genetic structure and identification of loci under selection in the invasive tunicate, *Botryllus schlosseri*, using newly developed EST-SSRs. *Biochem. Systemat. Ecol.* 66, 331–336. <https://doi.org/10.1016/j.bse.2016.05.007>.
- Locke, A., Hanson, J.M., MacNair, N.G., Smith, A.H., 2009. Rapid response to non-indigenous species. 2. Case studies of invasive tunicates in Prince Edward Island. *Aquat. Invasions* 4, 249–258. <https://doi.org/10.3391/ai.2009.4.1.25>.
- López-Legentil, S., Turon, X., Planes, S., 2006. Genetic structure of the star sea squirt, *Botryllus schlosseri*, introduced in southern European harbours. *Mol. Ecol.* 15, 3957–3967. <https://doi.org/10.1111/j.1365-294X.2006.03087.x>.
- López-Legentil, S., Legentil, M.L., Erwin, P.M., Turon, X., 2015. Harbor networks as introduction gateways: contrasting distribution patterns of native and introduced ascidians. *Biol. Invasions* 17. <https://doi.org/10.1007/s10530-014-0821-z>.
- Lord, J.P., 2017. Impact of seawater temperature on growth and recruitment of invasive fouling species at the global scale. *Mar. Ecol.* 38, 1–10. <https://doi.org/10.1111/maec.12404>.
- Martin, J.L., LeGresley, M.M., Thorpe, B., McCurdy, P., 2011. Non-indigenous tunicates in the Bay of Fundy, eastern Canada (2006–2009). *Aquat. Invasions* 6, 405–412. <https://doi.org/10.3391/ai.2011.6.4.05>.
- Millar, R.H., 1955. On a collection of ascidians from South Africa. *Proc. Zool. Soc. Lond.* 125, 169–221. <https://doi.org/10.1111/j.1096-3642.1955.tb00597.x>.
- Navy, U.S., 1951. Report on Marine Borers and Fouling Organisms in 56 Important Harbors and Tabular Summaries of Marine Borer Data from 160 Widespread Locations. Bureau of Yards and Docks, Department of the Navy, Washington, DC.
- Orensanz, J.M., Schwindt, E., Pastorino, G., Bortolus, A., Casas, G., Darrigran, G., Elías, R., López Gappa, J.J., Obenat, S., Pascual, M., Penchaszadeh, P., Piriz, M.L., Scarabino, F., Spivak, E.D., Vallarino, E.A., 2002. No longer the pristine confines of the world ocean: a survey of exotic marine species in the southwestern Atlantic. *Biol. Invasions* 4, 115–143. <https://doi.org/10.1023/A:1020596916153>.
- Osborne, M.J., Carson, E.W., Turner, T.F., 2012. Genetic monitoring and complex population dynamics: insights from a 12-year study of the rio grande silvery minnow. *Evolut. Appl.* 5, 553–574. <https://doi.org/10.1111/j.1752-4571.2011.00235.x>.
- Pallas, P.S., 1766. No Title. In: *Elenchi Zoophytorum Hagae-Comitum*, pp. 352–357.
- Pancer, Z., Gershon, H., Rinkevich, B., 1994. Direct typing of polymorphic microsatellites in the colonial tunicate *Botryllus schlosseri* (Ascidacea). *Biochem. Biophys. Res. Commun.* <https://doi.org/10.1006/bbrc.1994.2231>.
- Paz, G., Douek, J., Mo, C., Goren, M., Rinkevich, B., 2003. Genetic structure of *Botryllus schlosseri* (Tunicata) populations from the Mediterranean coast of Israel. *Mar. Ecol. Prog. Ser.* 250, 153–162. <https://doi.org/10.3354/meps250153>.
- Peakall, R., Smouse, P.E., 2012. GenALEX 6.5: genetic analysis in Excel. Population genetic software for teaching and research-an update. *Bioinformatics* 28, 2537–2539. <https://doi.org/10.1093/bioinformatics/bts460>.
- Petit, R., Mousadik, A.E.L., Ponst, O., 1998. Identifying basis of populations markers for conservation on the genetic. *Conserv. Biol.* 12, 844–855. <https://doi.org/10.1111/j.1523-1739.1998.96489.x>.
- Pineda, M.C., Lorente, B., López-Legentil, S., Palacín, C., Turon, X., 2016. Stochasticity in space, persistence in time: genetic heterogeneity in harbour populations of the introduced ascidian *Styela plicata*. *PeerJ*. <https://doi.org/10.7717/peerj.2158>, 2016.
- Pleus, A., Leclair, L., Schultz, J., 2008. 2007–09 Tunicate Management Plan, pp. 1–65.
- Pritchard, J.K., Stephens, M., Donnelly, P., 2000. Inference of population structure using multilocus genotype data. *Genetics* 155, 945–959.
- Reem, E., Rinkevich, B., 2014. *Botryllus schlosseri* and *Botrylloides leachii* (Chordata, Ascidacea) have not been recorded in the Red Sea. *Mar. Biodivers.* 44, 585–587. <https://doi.org/10.1007/s12526-014-0211-x>.
- Reem, E., Douek, J., Katzir, G., Rinkevich, B., 2013a. Long-term population genetic structure of an invasive urochordate: the ascidian *Botryllus schlosseri*. *Biol. Invasions* 15, 225–241. <https://doi.org/10.1007/s10530-012-0281-2>.
- Reem, E., Mohanty, I., Katzir, G., Rinkevich, B., 2013b. Population genetic structure and modes of dispersal for the colonial ascidian *Botryllus schlosseri* along the Scandinavian Atlantic coasts. *Mar. Ecol. Prog. Ser.* 485, 143–154. <https://doi.org/10.3354/meps10313>.
- Reem, E., Douek, J., Paz, G., Katzir, G., Rinkevich, B., 2017. Phylogenetics, biogeography and population genetics of the ascidian *Botryllus schlosseri* in the Mediterranean Sea and beyond. *Mol. Phylogenet. Evol.* 107, 221–231. <https://doi.org/10.1016/j.ympev.2016.10.005>.
- Reem, E., Douek, J., Rinkevich, B., 2022. A critical deliberation of the ‘species complex’ status of the globally spread colonial ascidian *Botryllus schlosseri*. *J. Mar. Biol. Assoc. U. K.* 1–14. <https://doi.org/10.1017/s0025315422000029>.
- Rinkevich, B., Weissman, I.L., 1987. The fate of *Botryllus* (Ascidacea) larvae cosettled with parental colonies: beneficial or deleterious consequences? *Biol. Bull.* 173, 474–488. <https://doi.org/10.2307/1541694>.
- Rinkevich, B., Shapira, M., Weissman, I.L., Saito, Y., 1992. Allopatric responses between three remote populations of the cosmopolitan ascidian *Botryllus schlosseri*. *Zool. Sci.* 9, 989–994.
- Rius, M., Turon, X., Bernardi, G., Volckaert, F.A.M., Viard, F., 2015. Marine invasion genetics: from spatio-temporal patterns to evolutionary outcomes. *Biol. Invasions* 17, 869–885. <https://doi.org/10.1007/s10530-014-0792-0>.
- Simberloff, D., Rejmánek, M. (Eds.), 2011. *Encyclopedia of Biological Invasions*. University of California Press, Berkeley.
- Sinai, I., Segev, O., Weil, G., Oron, T., Merilä, J., Templeton, A.R., Blaustein, L., Greenbaum, G., Blank, L., 2019. The role of landscape and history on the genetic structure of peripheral populations of the Near Eastern fire salamander, *Salamandra atra*, in Northern Israel. *Conserv. Genet.* 20, 875–889. <https://doi.org/10.1007/s10592-019-01181-5>.
- Skerman, T.M., 1960. Ship-fouling in New Zealand waters: a survey of marine fouling organisms from vessels of the coastal and overseas trades. *N. Z. J. Sci.* 3, 620–648.
- Stoner, D.S., Quattro, J.M., Weissman, I.L., 1997. Highly polymorphic microsatellite loci in the colonial ascidian *Botryllus schlosseri*. *Mol. Mar. Biol. Biotechnol.* 6, 163–171.
- Stoner, D.S., Ben-Shlomo, R., Rinkevich, B., Weissman, I.L., 2002. Genetic variability of *Botryllus schlosseri* invasions to the east and west coasts of the USA. *Mar. Ecol. Prog. Ser.* 243, 93–100. <https://doi.org/10.3354/meps243093>.
- Tamir, S., Reem, E., Paz, G., Tikochinski, Y., Rinkevich, B., 2022. The ancient Levantine *Botryllus schlosseri* (Tunicata): population genetics landscape under frequent natural disturbances. *Mediterr. Mar. Sci.* <https://doi.org/10.12681/mms.28622>.
- Tepolt, C.K., 2015. Adaptation in marine invasion: a genetic perspective. *Biol. Invasions* 17, 887–903. <https://doi.org/10.1007/s10530-014-0825-8>.
- Van Kleunen, M., Dawson, W., Essl, F., Pergl, J., Winter, M., Weber, E., et al., 2015. Global exchange and accumulation of non - native plants. *Nature* 525, 100–103. <https://doi.org/10.1038/nature14910>.
- Van Name, W.G., 1945. The North and south American ascidians. *Bull. Am. Mus. Nat. Hist.* 84.
- Van Oosterhout, C., Hutchinson, W.F., Wills, D.P.M., Shipley, P., 2004. MICRO-CHECKER: software for identifying and correcting genotyping errors in microsatellite data. *Mol. Ecol. Notes* 4, 535–538. <https://doi.org/10.1111/j.1471-8286.2004.00684.x>.
- Walsh, J.R., Carpenter, S.R., Van Der Zanden, M.J., 2016. Invasive species triggers a massive loss of ecosystem services through a trophic cascade. *Proc. Natl. Acad. Sci. U.S.A.* 113, 4081–4085. <https://doi.org/10.1073/pnas.1600366113>.
- Wellband, K.W., Pettitt-Wade, H., Fisk, A.T., Heath, D.D., 2017. Differential invasion success in aquatic invasive species: the role of within- and among-population genetic diversity. *Biol. Invasions* 19, 2609–2621. <https://doi.org/10.1007/s10530-017-1471-8>.
- Wonham, M.J., Carlton, J.T., 2005. Trends in marine biological invasions at local and regional scales: the Northeast Pacific Ocean as a model system. *Biol. Invasions* 7, 369–392. <https://doi.org/10.1007/s10530-004-2581-7>.
- Yund, P.O., Collins, C., Johnson, S.L., 2015. Evidence of a native northwest Atlantic COI haplotype clade in the cryptogenic colonial ascidian *Botryllus schlosseri*. *Biol. Bull.* 228, 201–216. <https://doi.org/10.1086/BBLv228n3p201>.