

Synthesis and Glycosidation of Anomeric Halides: Evolution from Early Studies to Modern Methods of the 21st Century

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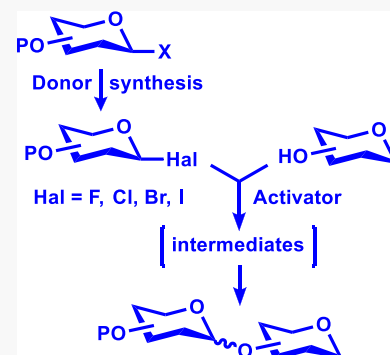


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ABSTRACT: Advances in synthetic carbohydrate chemistry have dramatically improved access to common glycans. However, many novel methods still fail to adequately address challenges associated with chemical glycosylation and glycan synthesis. Since a challenge of glycosylation has remained, scientists have been frequently returning to the traditional glycosyl donors. This review is dedicated to glycosyl halides that have played crucial roles in shaping the field of glycosciences and continue to pave the way toward our understanding of chemical glycosylation.



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1. INTRODUCTION: GLYCOSYLATION AS A CENTRAL REACTION OF TRADITIONAL AND MODERN GLYCOSCIENCES

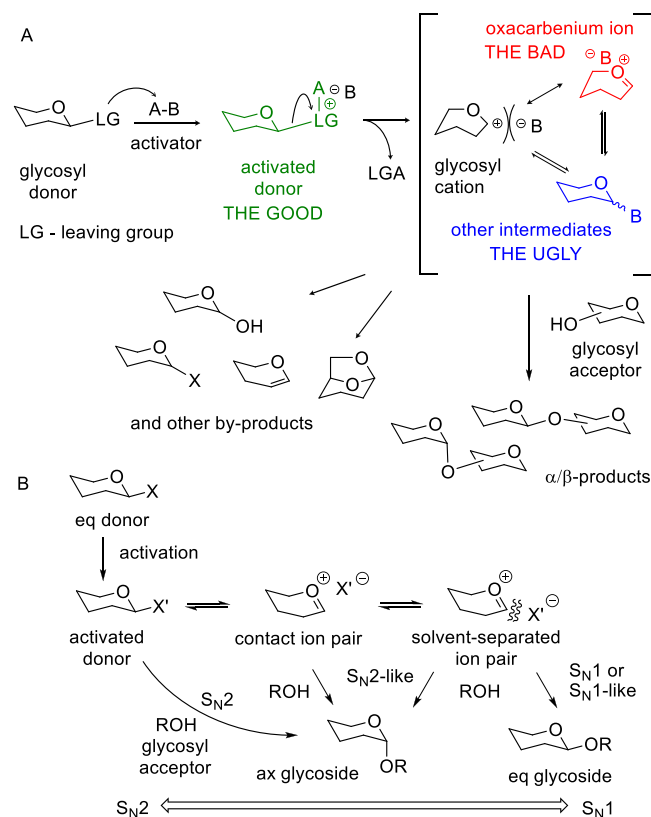
From the building blocks of nature to disease-battling pharmaceuticals, carbohydrates have had a broad impact on many scientific and industrial fields. Numerous applications of these essential biomolecules line up at the frontier of pharmaceutical, diagnostic, and functional food development. Consequently, carbohydrates have been extensively studied by biological and medical communities. Earth-abundant and renewable sources of chirally pure materials, monosaccharides remain popular building blocks for synthetic chemists. Oligomers, glycans, or oligosaccharides, wherein multiple monosaccharides are connected to each other via glycosidic linkages, have also been popular targets for chemists. However, complex carbohydrates remain challenging targets due to the requirement for elaborate protecting group manipulations and functionalization at all stages of the synthesis. Many glycosylation methods have been developed, but performing glycosylation reactions with complete chemo-, regio-, and stereoselectivity represents a notable challenge.

Nature flawlessly executes the glycosylation reaction enzymatically,^{1,2} but chemical glycosylation remains cumbersome.

³ The first chemical glycosylation reactions were performed by Michael,⁴ Fischer,^{5,6} and Koenigs/Knorr⁷ at the turn of the 20th century. With the exception of the Fischer glycosylation that relies on unprotected sugars as glycosyl donors,⁵ all other early methods relied on glycosyl halides, chlorides or bromides.^{4,6,7} Already those initial studies led to the development of glycosylation methodologies that paved the way to obtaining simple alkyl/aryl glycosides for decades. However, glycosylations of sugar acceptors were less efficient, and the synthesis of complex oligosaccharide targets was deemed practically impossible. Terminology used herein refers to “glycosylation of the acceptor with the donor” or “glycosidation of the donor with the acceptor.” One of the main directions to improve synthetic capabilities has been the investigation of leaving groups beyond the original chlorides/bromides and hemiacetals. These studies lead to the discovery of other glycosyl halides, fluorides,⁸ and iodides.⁹ However, many other classes of alternative glycosyl donors including glycosyl esters,^{10–13} thioglycosides,^{14–17} 1,2-orthoesters,^{18,19} *O*-imidates,^{20–23} thioimidates,^{24–29} alkenyl glycosides,^{30–32} sulfones,³³ thiocyanates,³⁴ diazirines,³⁵ glycols,^{36–40} sulfoxides,⁴¹ xanthates,⁴² selenium glycosides,⁴³ phosphites,^{44,45} tellurium glycosides,⁴⁶ sulfonylcarbamates,⁴⁷ heteroaryl glycosides,⁴⁸ phosphates,⁴⁹ disulfides,⁵⁰ 2-(hydroxycarbonyl)benzyl glycosides,⁵¹ and compounds equipped with alkynyl-based leaving groups^{52–61} have been developed.

Regardless of the glycosylation reaction conditions and mechanism, unimolecular S_N1-like or bimolecular S_N2-like, the following steps are typically identified (Scheme 1A).⁶² The first step involves the formation of the activated donor as a result of the interaction of the leaving group (LG) and the promoter (A-B); this step can be reversible or irreversible depending on

Scheme 1. Outline of General Glycosylation Mechanisms



the leaving group and the method of activation.⁶³ There are a few reports indicating that the glycosyl acceptor attack may be directed to the activated donor. This pure S_N2 displacement pathway would be quite desirable (THE GOOD) in terms of potential stereocontrol because it would allow for the stereospecific inversion of the leaving group: equatorial LG will produce axial glycoside whereas axial LG will lead to equatorial glycoside. However, examples of such reactions remain rare.^{64–71} Examples wherein double S_N2-like inversion would lead to the retention of the original LG configuration are also known. In a majority of cases, the second step involves the dissociation of the glycosyl donor, a typically irreversible expulsion of the activated leaving group (LGA), which is the rate-determining step (RDS) of the glycosylation reaction.

This process leads to the formation of the respective glycosyl carbocation and/or its stabilized form, oxacarbenium ion, along with the associated counteranion (B[−]). The oxacarbenium intermediate is often blamed for scrambling the stereoselectivity of the reaction (THE BAD). This is due to the existence of the oxacarbenium ion in a flattened half-chair conformation because of the sp²-hybridization of the anomeric carbon. It should be noted that other conformations of oxacarbenium intermediates are possible depending on the nature of substrates, protecting groups, and various steric and/or electronic factors. The next step involves a nucleophilic attack on the oxacarbenium ion, which is possible from either the bottom face or the top face of the ring. As a result, uncontrolled glycosylations often lead to the formation of a mixture of α/β -diastereomers. Other intermediates, the existence of which is often ignored (THE UGLY), or their impact on the reaction is underestimated, may also form at this stage with or without covalently attached B. However, there are some extended studies related to such intermediates, examples of which include tosylates, triflates, intermediates of dehydrative glycosylations among others, which can lead to unexpected side products and/or be responsible for scrambling the stereoselectivity of glycosylations. On the other hand, is the involvement of these intermediates is understood, this could lead to excellent stereocontrol how it has been proven by Schuerch,^{72,73} Crich,^{74–77} Gin,^{78–82} Woerpel,^{83–87} Bennett,^{69,88,89} and others.⁹⁰ The last step, the existence of which is also often overlooked, involves the proton transfer. This is the termination step, after which the formation of the glycosidic bond becomes irreversible.⁹¹

The simplified mechanistic outline of glycosylation presented in Scheme 1A implies that the RDS is unimolecular and is independent of the glycosyl acceptor. However, the donor–acceptor mismatch concept by Paulsen⁹² and Fraser-Reid and Lopez,^{93–97} the double stereodifferentiation phenomenon,⁹⁸ and other studies^{99–102} present a strong counterargument. In fact, a number of kinetic studies that have emerged in the past decade indicate that a majority of glycosylations follow a mixed mono- and bimolecular displacement mechanism,^{71,103–106} and the reaction order differs between various sugar series, and can even depend on the leaving group and/or covalently attached counterion type and orientation.^{107–111} As stated by Crich et al, the exact mechanism of a leaving group departure in a typical glycosylation reaction generally falls at a certain position on a continuum of mechanisms spanning from ideal S_N1 extreme to ideal S_N2 extreme (Scheme 1B).^{71,105–112}

The goal of controlling glycosylation has been pursued in many other ways with much effort dedicated to the optimization of the reaction conditions, suppressing side

reactions,^{113,114} studying stereoelectronics and conformation of the starting material and key reaction intermediates.^{62,80–84,86,87,91,115–129} Fraser-Reid's seminal work on the armed-disarmed approach showed that the building block reactivity can be modulated through the choice of protecting groups.^{130,131} The scope of the original armed-disarmed concept has been expanded and a number of efforts to quantify or even predict the reactivity of building blocks have been reported by Fraser-Reid,¹³² Ley,^{133,134} and Wong.^{135,136} Wong's study also revealed a number of building blocks that extend beyond the traditional armed-disarmed boundary. This discovery opened a new avenue for studying building block reactivity,¹³⁷ and Boons¹³⁸ and, subsequently, Demchenko¹³⁹ reported superdisarmed building blocks.

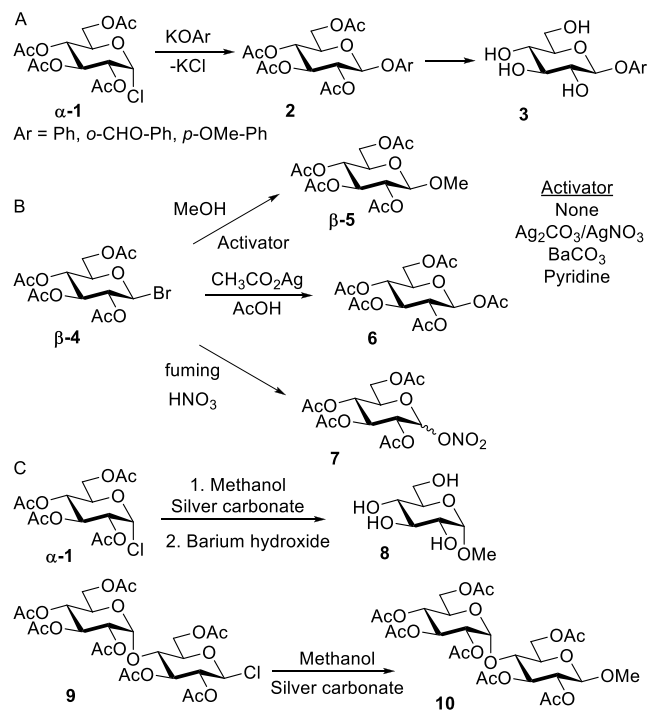
Two concepts for superarming glycosyl donors have also emerged. Bols showed that superarming can be achieved by changing the equatorial-rich ⁴C₁ conformation.^{124–127} Demchenko then reported building blocks wherein the electronic superarming was achieved via the O2/O5 cooperative effect.^{140–142} While the stereoelectronic and conformational effects on reactivity have been studied extensively, the impact on stereoselectivity (beyond building blocks equipped with 2-O-participating group) remain elusive. Although some model studies helped to establish general trends,^{83–86,115,116,128,129,143} practical applications of the stereoelectronic and conformational factors to stereocontrolling glycosylation reactions are still lacking.

Despite all these recent improvements, the challenge of glycosylation has remained, and scientists have turned their attention to reinvestigating the original glycosyl donors, hemiacetals, and halides. More recent work with hemiacetals^{69,81,144} and glycosyl halides^{70,145–147} brought these glycosylation reactions to an entirely different level of flexibility and versatility. These simple donors are typically easily accessible from a variety of precursors, can be readily activated, and offer superior atom economy. This review, dedicated to glycosyl halides, guides the reader from the first known glycosylation reactions to recent advances in the field that helped to navigate glycosciences forward. The first glycosylations performed by Michael involved glycosyl chlorides. However, in subsequent years, glycosyl chlorides were largely outshadowed by glycosyl bromides. Because of the ease of the synthesis and their higher reactivity profile, glycosyl bromides have traditionally been considered advantageous over their chloride counterparts. As a result, most of the efforts were focused on reactions of glycosyl bromides that have become prevalent glycosyl donors in the first 100 years of synthetic carbohydrate chemistry. Therefore, we will open our review with a summary of the early studies (Section 2) and then move directly to the discussion of glycosyl bromides (Section 3). Many methods developed for the activation of glycosyl bromides are also effective for the activation of glycosyl chlorides. Hence, Section 4, dedicated to glycosyl chlorides, will mainly focus on the reagent-specific or condition-specific activations of glycosyl chlorides rather than repeating the same data presented for glycosyl bromides. We will then turn our attention to glycosyl iodides (Section 5), which were outshadowed by both glycosyl bromides and glycosyl chlorides, and only recently found some notable synthetic utility. Finally, we will discuss glycosyl fluorides (Section 6), which became very prominent glycosyl donors in recent decades.

2. KEY ACCOMPLISHMENTS OF (AND LESSONS FROM) EARLY GLYCOSIDATIONS OF GLYCOSYL HALIDES AT THE TURN OF THE 20TH CENTURY

The first glycosylation was reported by Arthur Michael in 1879, about a decade before the famous addition reaction that carries his name, the Michael addition, was discovered. This reaction is depicted in Scheme 2A, wherein glycosidation of

Scheme 2. First Glycosylation Reactions Reported by Michael (A), Koenigs and Knorr (B), and Fischer (C)



chloride α -1 with alkoxide produced glycoside 2, which was deprotected under these basic reaction conditions to produce glycoside 3. In Michael's own words, the synthesis was "...starting from the interesting compound discovered by A. Colley...known under the name of acetochlorhydrose. This compound..., I have allowed to act on potassium phenate and potassium salicylate, and have obtained compounds which possess all the characteristic properties of the glucosides. After numerous experiments, I found the following conditions to yield the most satisfactory results: 27.5 gr. of acetochlorhydrose were mixed with about twice its volume of absolute alcohol, and the solution added to a cold alcoholic solution of 10 gr. of potassium phenate. After a few minutes, a crystalline precipitate began to separate from the solution, and at the same time a strong odor of acetic ether was noticed. The reaction proceeded very rapidly, and after four or five hours no further separation of the crystalline substance was observed. . . The most interesting property of this substance is its behavior towards dilute acids and emulsin. A dilute solution of sulfuric or chlorhydric acid decomposes it on gently warming very readily in glucose and phenol..."⁴

In 1901, Koenigs and Knorr introduced silver salt-promoted activation of glycosyl bromides for the synthesis of glycosides.⁷ This approach is considered as a major milestone in glycochemistry because the outcome of this reaction was much easier to control or predict than that of the Fischer glycosylation with unprotected donors. It was also shown that alcohols rather than charged nucleophiles used in the Michael

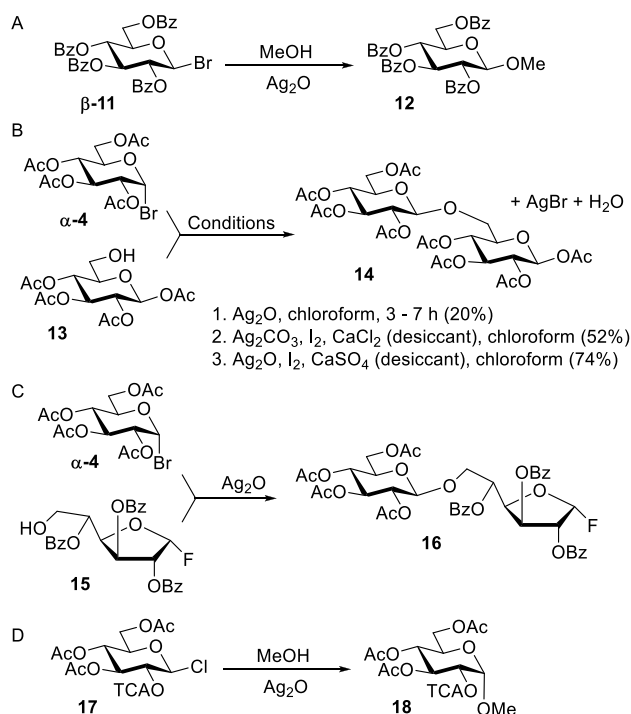
approach can be glycosylated directly. The first reactions involved glycosidation of acetylated glucosyl bromide β -4 with simple alcohols (methanol or ethanol) in the presence of silver carbonate (Ag₂CO₃) or silver nitrate (AgNO₃) as shown in Scheme 2B. It should be noted that current understanding of the reaction and its mechanisms implies that the formation of pure β -anomer of acetobromoglucose is very unlikely. It is possible that during those times, the anomeric configuration of glycosyl halides was simply unknown. To preserve integrity of the discussion, we will be using configurations that were originally proposed by the authors. Contrary to the Michael glycosylation approach⁴ wherein neutral KCl was generated, in the Koenigs–Knorr reaction the formation of HBr was inevitable. The exact role of the silver salt was not known at that moment, but it was assumed that it acted as the acid scavenger. The synthesis of methyl glucoside β -5 was also achieved by the treatment of glycosyl bromide β -4 in the presence of either barium carbonate (BaCO₃) or pyridine. It was also demonstrated that methyl glycoside can be generated in the presence of an excess of methanol without activators or additives. On the other hand, the treatment of glycosyl bromide 4 with silver acetate (CH₃COOAg) in acetic acid afforded pentaacetate 6, and a similar treatment with fuming nitric acid afforded tetra-acetylated glycosyl nitrate 7. An interesting observation was made that methyl glycoside 5 and pentaacetate 6 had the same anomeric configuration as that of the starting material, β -glycosyl bromide 4. This methodology was also extended to per-acetylated galactosyl bromides. Again, it is quite possible that the authors were dealing with α -bromides that were mistakenly presented as their β -counterparts.

Independently from Koenigs and Knorr, Fischer and co-workers also synthesized methyl glucoside from α -acetochloroglucose 1 in the presence of methanol and silver carbonate, and the product was deprotected with barium hydroxide.⁶ Differently from the results reported by Koenigs and Knorr (*vide supra*), glycosidation of α -chloride 1 resulted in the formation of methyl α -glucoside 8 (Scheme 2C). Additionally, Fischer and co-workers synthesized tetra-acetylated methyl galactoside from galactosyl chloride. Apart from monosaccharides, the authors were able to synthesize hepta-acetylated maltosyl chloride 9 that was glycosidated with methanol in the presence of silver carbonate. In this case β -linked methyl maltoside 10 was obtained.⁶ Similarly to studies by Koenigs and Knorr, Fischer also considered silver acetate AgOAc, but noticed a significant level of the acetate transfer.⁶

Following these early efforts dedicated to glycosylation of simple alcohols, Fischer and co-workers synthesized a variety of relatively complex β -glycosides including hepta-acetyl menthol maltoside.^{148,149} The authors also tried to synthesize a 1→1-linked tetrasaccharide assuming that hepta-acetyl lactosyl bromide will self-condense with the hemiacetal produced *in situ* in the presence of Ag₂CO₃.¹⁴⁸ Since the role of silver salt was considered to be as acid scavenger, Fischer thought of replacing it with an organic base, such as quinoline. This study showed unprecedented formation of significant amounts of α -phenyl glucoside when per-acetylated glycosyl bromide was heated with phenol in the presence of quinoline.¹⁵⁰ The continuation of this work led to the implementation of silver oxide as an alternate acid scavenger.¹⁵¹ In addition to acetylated bromides, glycosidation of per-benzoylated glucosyl bromide β -11 with MeOH was

proven possible in the presence of Ag_2O to afford methyl glycoside **12** (Scheme 3A).

Scheme 3. First Attempts to Enhance Utility of the Koenigs–Knorr Reaction



Helferich and co-workers were the first to synthesize a disaccharide by coupling two monosaccharide building blocks. As shown in Scheme 3B, this was achieved by treating glucosyl bromide α -4 with 1,2,3,4-tetra-*O*-acetyl- β -D-glucopyranose acceptor **13** under classical Koenigs–Knorr glycosylation conditions to afford per-acetylated β -gentiobiose **14** (conditions 1).¹⁵² However, only a modest yield of 20% was achieved, which was attributed to the presence of water in the reaction mixture. It was suggested that water molecules interfere with the reaction by continuously consuming glycosyl bromide α -4 and producing hydrolyzed byproducts. Helferich and co-workers then attempted to remove water by adding finely powdered calcium chloride as a desiccant. However, this led to a significant drop in the rate of glycosylation.¹⁵³ It was then discovered that the reaction can be accelerated when molecular iodine was added to a silver carbonate-promoted glycosylation reaction in the presence of a desiccant (conditions 2). In this case, β -gentiobiose **14** was obtained in a significantly improved yield of 52%. Reynolds and Evans also developed reaction conditions that helped to achieve complete exclusion of water, both present in the reactants from the beginning and also that generated during the reaction.¹⁵⁴ These reaction conditions involved preliminary stirring of glycosyl acceptor **13** and silver oxide with Drierite (CaSO_4) in chloroform for 1 h. Iodine was then added followed by a slow addition of a solution of glycosyl bromide α -4 in chloroform. As a result of these improvements, a very high yield of 74% for the synthesis of gentiobiose **14** was achieved (conditions 3, Scheme 3B).

Helferich and co-workers were also the first to achieve a selective activation of one leaving group over another. Some 60–70 years later, this selective activation strategy became a

very useful tool for streamlining glycan assembly because the products can be used as glycosyl donors directly, without any modification (*vide infra*).¹⁵⁵ In this application, selective activation of glycosyl bromide donor α -4 over glycosyl fluoride acceptor **15** was achieved in the presence of silver oxide to produce disaccharide **16** (Scheme 3C).¹⁵² The deprotection of acyl groups furnished gentiobiosyl fluoride which was then treated with calcium carbonate to produce a fully unprotected disaccharide.

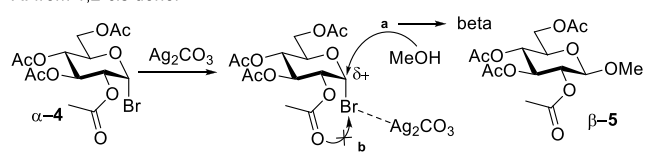
Formation of the α -phenyl glycoside in the quinoline-promoted reaction at a high temperature by Fischer (*vide supra*),¹⁵⁰ demonstrated that 1,2-*cis* glycosides can also be obtained.¹⁵⁶ Following this lead, Brigl and co-workers described the stereoselective synthesis of α -glycosides from β -chloride **17** protected with the 2-*O*-trichloroacetyl group shown in Scheme 3D.¹⁵⁷ The authors assumed that the Walden inversion occurs during the glycosylation of methanol in the presence of silver carbonate to afford α -glycoside **18**. The α/β -ratio of glycosides was found to depend on the reaction time and temperature. Similar to Brigl's investigation, Schlubach and Schroter¹⁵⁸ and also Hickinbottom¹⁵⁹ investigated the synthesis of α -glycosides from β -acetochloroglucose. Over the course of this study, the authors anticipated that the synthesis of α -glycosides from β -glycosyl chlorides can be achieved only when the experimental conditions are maintained in the way that the rate of glycosylation is greater than that of the β -chloride anomerization to its α -counterpart. The authors also anticipated that the anomerization can be suppressed by using a suitable solvent. It was further reinforced that higher α -selectivity of glycosylations is achieved at lower glycosyl acceptor concentrations. To further advance the classical Koenigs–Knorr glycosylation approach, Zemplén and co-workers switched from silver salts and showed that an efficient activation of cellobiosyl bromide can be achieved with mercuric acetate.^{160,161} Over the course of this investigation, anomeric mixtures have been obtained, and a dedicated study revealed that the ratio of α/β -glycosides mainly depends on the amount and types of alcohol used.^{160,162,163}

During those times, many reactions were hampered by the formation of side products. One of the most exciting observations was Fischer's discovery of the existence of an additional isomeric glycoside that was observed to exist besides α - and β -glycosides.¹⁶⁴ The authors coined the term of the γ -form (or third form) and suggested that it might have a different ring structure than pyranose as a result of the ring-opening, acyl migration, and subsequent ring closure during the glycosylation reaction. Also reported by Dale,¹⁶⁵ and puzzled many others,^{166–170} it was not until studies by Freudenberg^{171,172} and, independently, Haworth et al.^{173,174} who suggested the γ -form to be 1,2-orthoester **19** shown in Scheme 4. To understand mechanistic details for the formation of glycosides and orthoesters, Isbell investigated various factors affecting Koenigs and Knorr reactions and byproduct formation, such as the orientation of the substituent at C-2, solvents, temperature, and the presence of water. Apart from these, the authors conducted an expansive study of physical and chemical properties of products to understand chemical composition, structure, and conformation of products.^{175–179}

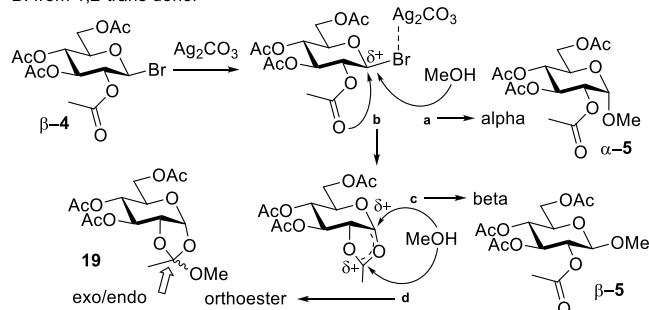
On the basis of these studies, Isbell and co-workers concluded that 1,2-*trans* glycosylation occurs when there is a participating ester group present at the C-2 position. Taking into consideration the neighboring group participation, Isbell proposed two distinct pathways for glycosidation of 1,2-*cis* and

Scheme 4. Glycosidation of 1,2-*cis* and 1,2-*trans* Glycosyl Halides and the Formation of 1,2-Orthoesters

A. from 1,2-*cis* donor



B. from 1,2-*trans* donor



1,2-*trans* glycosyl halides, α -4 and β -4, respectively.¹⁷⁶ The activation process wherein the anomeric bromide complexes with the silver salt is the same for both glycosyl donor configurations (Scheme 4). This decreases the electron density at the anomeric center, making it more susceptible to nucleophilic attack. In the case of the 1,2-*cis* glycosyl halide α -4, only the expected inversion product β -5 can be obtained (Scheme 4A, pathway a). In this case, the 1,2-orthoester does not form (pathway b) because the approach of the 2-O-acetyl group is blocked by the halide. Conversely, the 1,2-*trans* bromide β -4 yielded two anomeric glycosides α/β -5 and the orthoesters *exo/endo*-19 (Scheme 4B). Following the activation, the 1,2-*cis* product α -5 was obtained via direct nucleophilic displacement from the bottom face of the ring (pathway a). Additionally, the intramolecular attack from the adjacent carbonyl oxygen leads to the formation of a reactive acyloxonium intermediate (pathway b). Depending on the site of nucleophilic attack, the latter can produce a 1,2-*trans* glycoside β -5 (pathway c) and a 1,2-orthoester *exo/endo*-19 (pathway d). In this case, the neighboring group participation may or may not offer anchimeric assistance and hence accelerate the leaving group departure. It should be noted that although sometimes (inaccurately) used interchangeably, “neighboring group participation” and “anchimeric assistance” are not the same: while the former refers to the effect on stereoselectivity, the latter refers to the acceleration of the reaction rate. It was originally believed that the formation of orthoesters and participation is only possible when the leaving halogen (bromide) is present in the 1,2-*trans* position to the participating group on the neighboring carbon atom. However, later it was clearly demonstrated are orthoesters can also form from 1,2-*cis* glycosyl halides.^{180–190} Subsequent studies of orthoesters were extended to sugar alcohols¹⁹¹ and led to observation of diastereomeric *exo/endo*-orthoesters the existence of which was later proven by nuclear magnetic resonance (NMR).¹⁹²

Over the years, Koenigs–Knorr glycosylation reactions found broad application in glycosylation of simple alcohols. Glycosylations of sugar acceptors were less efficient and the synthesis of complex oligosaccharide targets was deemed practically impossible. One of the main directions to improve synthetic capabilities has been the investigation of different

activation conditions. It should be particularly emphasized that it is seminal studies of glycosyl halide donors discussed in this section that have enabled our understanding of many basic aspects and mechanisms behind glycosylation reactions. This, in turn, helped to resolve challenges in synthesizing many targets ranging from simple glycosides to complex glycans and glycoconjugates.

3. GLYCOSYL BROMIDES

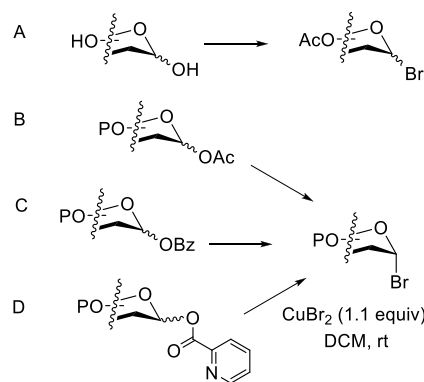
Since first silver salt-promoted activations of glucosyl bromides, innumerable reagents for the activation of glycosyl bromides have emerged. Not only glycosidation but also the synthesis of glycosyl bromides underwent significant improvements over the years with the introduction of mild reagents and developing efficient reaction conditions.

3.1. Synthesis of Glycosyl Bromides

Synthesis of glycosyl bromides is very crucial since many compounds of this class are not crystalline and their stability differs drastically based on their structure and protecting group pattern. Numerous methods have been developed for the synthesis of glycosyl bromides that are categorized below by the type of the starting material (anomeric protecting/leaving group) used for the introduction of the anomeric bromide moiety.

3.1.1. Preparation from Unprotected Sugars with Concomitant Per-acetylation. As aforementioned, synthesis of glycosyl chlorides was known since studies by Colley and then Michael. These syntheses were accomplished from the corresponding free sugars that were treated with acetyl chloride. Following this general concept, Koenigs and Knorr synthesized acetobromoglucose by treating glucose with neat acetyl bromide. The authors were able to isolate β -bromide in a good yield after crystallization (Scheme 5A).⁷ For many

Scheme 5. Synthesis of Glycosyl Bromides from Unprotected Sugars and Glycosyl Esters



years, this method remained popular for the synthesis of acetylated glycosyl bromides.^{7,148,193} Decades later, Koto and co-workers showed that the synthesis of glycosyl bromides can be achieved by treating the corresponding free sugars with acetyl bromide that was diluted with acetic acid.^{194,195} The authors demonstrated that acetic acid helps to prevent sudden evolution of toxic HBr gas compared to previously known methods for the formation of glycosyl bromide donor. Kartha and Jennings reported a one-pot synthesis of per-acetylated glycosyl bromide wherein free sugars were first treated with acetic anhydride followed by the addition of HBr in AcOH.¹⁹⁶ Another method for the synthesis of glycosyl bromides

involves the treatment of free sugars with acetic anhydride in the presence of catalytic HClO_4 in acetic acid followed by bromination with AcBr-MeOH with or without sonication.^{197,198} A reversal in the order of the reagent addition wherein sugar and acetic anhydride were added to a mixture of acetyl bromide and methanol in acetic acid also produced glycosyl bromides in excellent yields.¹⁶⁷ Lin et al. treated unprotected sugars with $\text{LiClO}_4\text{-Ac}_2\text{O}$ followed by the reaction with HBr-AcOH .¹⁹⁹ Recently, Bennett, Pohl, and co-workers developed a continuous flow platform for the synthesis of orthogonally protected monosaccharide building blocks.²⁰⁰ One relevant example includes acetylation and bromination of unprotected sugars performed in a single step. This transformation was rapidly achieved by employing acetyl bromide as acetylating and brominating reagent in a continuous flow manner. Saturated aq. NaHCO_3 was then used for in-line quench of excess HBr .

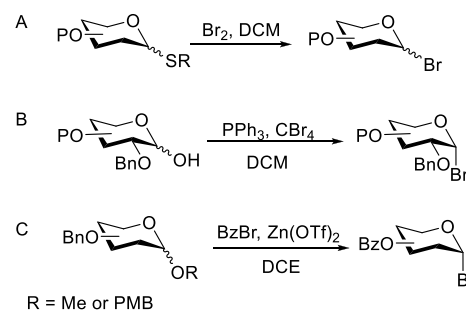
3.1.2. Preparation from Glycosyl Esters of Differentially Protected Sugars. The synthesis of acetylated glycosyl bromides can also be performed starting from presynthesized per-acetates (Scheme 5B). As described by Fischer, the treatment of glucose pentaacetate with HBr neat or in glacial acetic acid produced per-acetylated β -glucosyl bromide.¹⁴⁹ In early days, a few other methods such as red phosphorus/bromine or PBr_3 were commonly used for the synthesis of bromides.²⁰¹ Zempen and co-worker employed titanium tetrabromide for the synthesis of glycosyl bromide wherein a solution of titanium tetrabromide in chloroform was added to a solution of tetra-acetylated rhamnose in chloroform to produce acetobromo- α -L-rhamnose in an excellent yield and in crystalline form.²⁰² This protocol works well for the synthesis of 2-azido-2-deoxy bromides.²⁰³

More recent methods were predominantly focused on the development of milder reaction conditions for the formation of anomeric bromides. One such procedure for bromination involves a dilute solution of HBr in dichloromethane (DCM).^{204–206} A convenient method to generate HBr *in situ* is by reaction of MeOH and AcBr ; this protocol is particularly advantageous for the synthesis of highly reactive bromides and/or those equipped with acid-sensitive protecting groups.⁶⁴ Hunsen and co-workers utilized this protocol also for the synthesis of glycosyl bromides from the corresponding free sugars in the presence of Ac_2O .¹⁹⁷ Thiem and co-workers discovered that anomeric acetates can be efficiently converted to corresponding anomeric bromides upon treatment with trimethylsilyl bromide (TMSBr) in chloroform or benzene.²⁰⁷ The side product, trimethylsilyl acetate can be removed *in vacuo* owing to its low boiling point. Around the same time, a similar observation was independently made by Gillard and co-worker who converted different anomeric acetates to the corresponding bromides.²⁰⁸ The use of TMSBr is attractive since it eliminated the aqueous work up, which is a common operation used in many other methods. Apart from this, many common protecting groups are compatible with TMSBr . Later, Montero and co-workers reported a milder approach for halogenation by employing bismuth(III)-based reagents complexed with halosilanes.²⁰⁹ The treatment of per-acetylated sugars with $\text{BiBr}_3\text{-Me}_3\text{SiBr}$ produced corresponding glycosyl bromides in excellent yields. Mizuno and co-workers reported light-induced (352 nm, 15 W) conversion of per-acetylated sugars to the corresponding glycosyl bromides in the presence of bromine.²¹⁰

Preparation of glycosyl bromides from per-benzoylated sugars has also been reported (Scheme 5C). Fischer and Helferich treated per-benzoylated glucose with a saturated solution of HBr in glacial acetic acid.¹⁵¹ This method was further modified by Hudson and co-workers.^{211,212} Another efficient method for the synthesis of benzoylated glycosyl bromides involves the treatment of per-benzoylated sugars with AcBr-MeOH using ultrasound irradiation.¹⁹⁸ All the previous methods for the synthesis of glycosyl bromides from the corresponding 1-O-acyl derivatives rely on highly reactive, toxic, sensitive, and corrosive acids. To address these issues, Tang and co-workers described a milder reaction conditions for an efficient synthesis of glycosyl bromides.²¹³ In this approach, 1-O-picoloyl (Pico) derivatives were treated with copper(II) bromide in DCM at ambient temperature (Scheme 5D). This reaction produced the corresponding α -glycosyl bromides in good to excellent yields, and the method was found to be compatible with several protecting groups including acid-sensitive groups.

3.1.3. Preparation from Thioglycosides or Other Substrates. One of the most convenient methods for the formation of glycosyl bromides involves treatment of thioglycosides with bromine to produce the corresponding glycosyl bromides (Scheme 6A). This reaction typically

Scheme 6. Synthesis of Glycosyl Bromides from Thioglycosides (A), Hemiacetals (B), and O-Glycosides (C)



completes in minutes, does not require any work up, and the glycosyl bromides can be used immediately after solvent evaporation.^{214–217} Because of mild reaction conditions, a broad range of protecting groups including acyl, alkyl, silyl, acetal, and ketal are tolerated. A few other methods have been developed. Kobayashi and co-workers reported the conversion of 2-O-benzylated hemiacetals to corresponding glycosyl bromides by the treatment with $\text{PPh}_3\text{-CBr}_4$, commonly known as Appel conditions (Scheme 6B).²¹⁸ A one-pot conversion of benzyl-protected methyl or *p*-methoxyphenyl glycosides to per-benzoylated glycosyl bromides by treating with zinc triflate and benzoyl bromide, is another useful approach to the synthesis of per-benzoylated glycosyl bromides (Scheme 6C).²¹⁹

3.2. Glycosidation of Glycosyl Bromides

Since the first silver salt-promoted activation of glycosyl bromides (*vide supra*), innumerable reagents for the activation of glycosyl bromides have emerged. To streamline the discussion, we chose to divide the activating reagents into different categories based on their halophilic nature (Table 1). The discussion begins from silver salts that were very instrumental for understanding the metal salt involvement in splitting the anomeric C–Br bond and investigating the effects

Table 1. Activation of Glycosyl Bromide Donors

promoter	additive
	silver and copper salts
Ag ₂ CO ₃ ^{6,7}	I ₂ , ^{153,220} Lewis acid ²²¹
AgNO ₃ ^{7,151}	crown ether ^{222,223}
AgOAc ^{6,7}	
Ag ₂ O ¹⁵¹	I ₂ , ¹⁵⁴ borinic acid, ²²⁴ HOFox/Lewis acid, ^{217,225} TMSOTf, ^{146,221,226} TfOH ²²¹
AgClO ₄ ^{227–229}	
AgBF ₄ ^{227,230}	
AgPF ₆ ²³⁰	
AgOTf ²³⁰	TMU ²³¹
other organic Ag salts ^{227,232–234}	TfOH ²²¹
Ag silicate-alumina ²³⁵	
Ag zeolite ²³⁶	
Ag silica-alumina ²³⁷	
silver imidazole	ZnCl ₂ ²³⁸
Ag ₂ SO ₄	TfOH, ^{221,226}
CuI + BPhen + Xantphos ¹⁴⁷	DTBMP ¹⁴⁷
	mercury, zinc, and cadmium salts
Hg(OCOCH ₃) ₂ ^{160,161}	
Hg(CN) ₂ ²³⁹	HgBr ₂ ²³⁹
HgO ²³⁹	HgBr ₂ ²⁴⁰
HgBr ₂ ^{241,242}	
Hg(PhCOO) ₂ ²⁴³	
Hg(NpCOO) ₂ ²⁴³	
HgI ₂ ²⁴⁴	
ZnO ²³⁹	
ZnCl ₂ ^{245,246}	TrCl ₄ , ²⁴⁵ TMSCl ²⁴⁷
ZnBr ₂ ^{245,246}	TrBr ₄ , ²⁴⁵ TMSBr ^{246,247}
Zn(OTf) ₂	TMSBr ²⁴⁸
CdO ²³⁹	
CdCO ₃ ^{249–251}	
CdS ²⁴⁹	
	indium, tin, and bismuth salts
InCl ₃ ^{252,253}	
InBr ₃ ²⁵³	
InI ₃ ²⁵⁴	
In(OTf) ₃ ²⁵⁴	
In(NTf ₂) ₃ ²⁵⁴	
SnCl ₄ ²⁵⁵	
Sn(OTf) ₂	base ²⁵⁶
PbCO ₃ ²⁵¹	
	non-nucleophilic bases
pyridine ^{7,148,182,257}	
quinoline ^{150,156}	
collidine ¹⁸⁰	
phenanthroline derivatives ²⁵⁷	
2,2'-bipyridine ²⁵⁷	
	halogens or halide ions
NR ₄ Br ^{258,259}	
I ₂ ^{260,261}	DDQ ²⁶⁰
IBr ^{262,263}	DABCO ²⁶²
ICI ²⁶³	
NIS ^{261,263}	I ₂ , ²⁶³ protic acids ²⁶¹
	other methods
NR ₃ ^{264–266}	
PR ₃ ^{265,266}	
SR ₂ ^{265,266}	
solvolysis ^{7,230,267,268}	NR ₄ Br, ^{267,268} silver salts ²³⁰

of the counteranion of silver salts. We will then focus the discussion on how those studies enabled scientists to better understand the glycosylation reaction, helped develop improved methods, and how the improved methods enhanced our synthetic capabilities.

3.2.1. Activation with Silver Salts or Other Group 11 Metal Salts (Copper). As mentioned previously, glycosylation reactions in the presence of insoluble silver salts such as Ag₂O and Ag₂CO₃ proceed slowly and may result in inefficient glycosylations despite using large excess of reagents and, sometimes, excess reactants. This ultimately affects the yield of glycosides that typically fall within 30 to 70%. To improve the glycoside synthesis, several soluble silver salts were introduced as promoters for the activation of glycosyl bromides. These salts include silver acetate,^{6,7} which led to the predominant 1-O-acetylation, silver nitrate first studied by Koenigs and Knorr,⁷ and in a greater detail investigated by Knöchel et al.^{222,223} Several other soluble silver salts were explored including perchlorate,^{227,229} tetrafluoroborate,^{227,230} hexafluorophosphate,²³⁰ and trifluoromethanesulfonate (triflate).²³⁰ While being efficient activators, these silver salts required multiple equivalents of the acid scavenger to be added in these reactions. Wulff and co-workers studied glycosylation of steroidal alcohols in the presence of several silver salts of hydroxyacids and dicarboxylic acids.^{232–234,269,270} In a majority of cases, glycosides were observed as major products, but in some cases significant amounts of 1-O-acyl and 1,2-orthoester derivatives were also present. A direct correlation was paved between the formation of products and the distance between the hydroxy and carboxy groups in the organo-silver salts. As depicted in Figure 1, interactions between OH or OAg and the

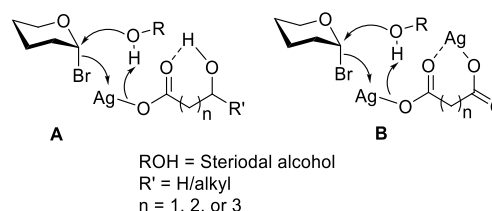


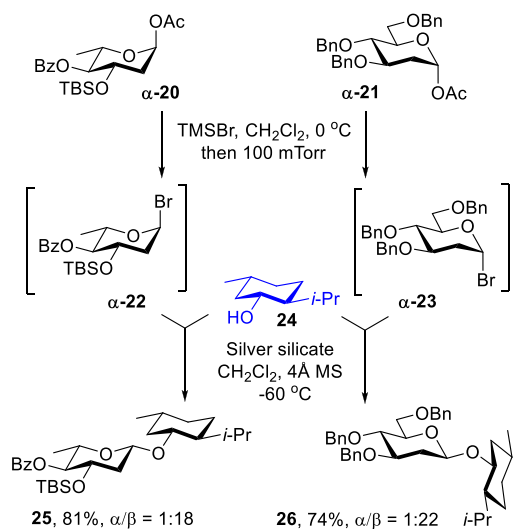
Figure 1. Mechanistic interpretation of action of hydroxyl carboxylate and dicarboxylate silver salts.

carbonyl group in 2-, 3-, and 4-hydroxycarboxylate salts (A), and 1,2-, 1,3-, 1,4-dicarboxylate salts (B) may have a role in suppressing the nucleophilic attack of the carboxylate ion. Stronger interactions help to lower the formation of 1-O-acyl derivatives. For hydroxy acids, silver 3-hydroxypentanoate and salicylate provided the highest yields, whereas disilver maleate outperformed all other diacids investigated.

Apart from the above-mentioned silver salts, numerous other silver-based promoters were reported for the activation of glycosyl bromides such as silver imidazole-ZnCl₂,²³⁸ silver silicate,²³⁵ silver silicate-alumina,²³⁵ silver zeolite,²³⁶ silver silica-alumina.²³⁷ As discovered by Paulsen and co-workers, silver silicates are very advantageous for the synthesis of β -linked glycosides.²³⁵ This study was recently extended by Herzon and co-worker to the stereoselective synthesis of 2-deoxy and 2,6-dideoxy β -glycosides, from corresponding anomeric bromides upon activation with silver silicate.²⁷¹ The authors employed in situ formation of glycosyl bromides α -22 and α -23 from the corresponding anomeric acetates in the presence of TMSBr in dichloromethane as shown in

Scheme 7. The synthesized glycosyl bromides α -22 and α -23 were then reacted with (–)-menthol acceptor **24** in the

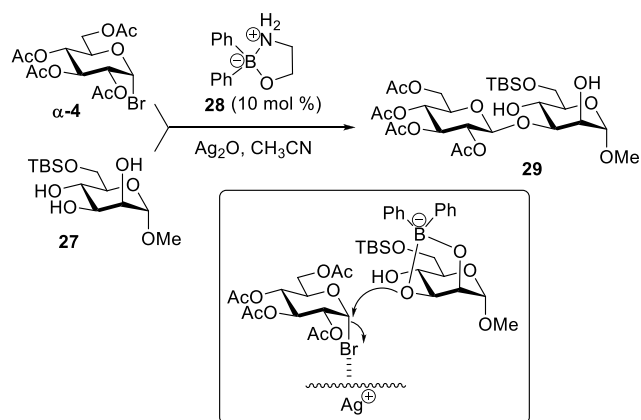
Scheme 7. Silver Silicate-Promoted Glycosidation of 2-Deoxy- and 2,6-Dideoxyglycosyl Bromides



presence of silver silicate to produce the respective β -linked glycosides in good yields and with high stereoselectivity: **25** (81%, $\alpha/\beta = 1:18$) and **26** (74%, $\alpha/\beta = 1:22$). The authors pointed out that the nature of protecting groups of the glycosyl bromide donors may play an important role in enhancing or reducing the stereoselectivity. It was also observed that acid sensitive and azide protecting groups remained unaffected during these reaction conditions.

Borinic acid-catalyzed regioselective glycosidation of glycosyl bromide (and chloride) was developed by Taylor and co-workers²²⁴ in accordance with their approach, complexation of partially protected *cis*-diol glycosyl acceptor **27** with borinic acid **28** to afford a borinate complex takes place first (Scheme 8).²⁷² The nucleophilicity of sugar hydroxyl groups is predicted by Fukui index calculation,^{273–275} which states that the boron-bound oxygen to be more nucleophilic than the free hydroxyl group. The borinate complex subsequently attacks glycosyl bromide α -4 to furnish disaccharide **29**. The outcome of the glycosylation in terms of yields and stereoselectivity mainly depends on the stereochemistry of the glycosyl halide and

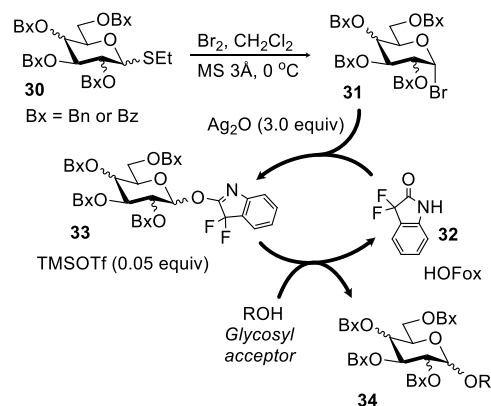
Scheme 8. Borinic Acid-Catalyzed Glycosidation of Glycosyl Bromide



reactivity of the acceptor. Exclusive 1,2-*trans* selectivity was observed in borinic acid-catalyzed glycosylation wherein α -glycosyl halides were used.

Demchenko and co-workers have developed a regenerative strategy to activate glycosyl bromides.^{217,225} As shown in Scheme 9, ethylthio glycoside **30** was first treated with a

Scheme 9. Regenerative Glycosylation



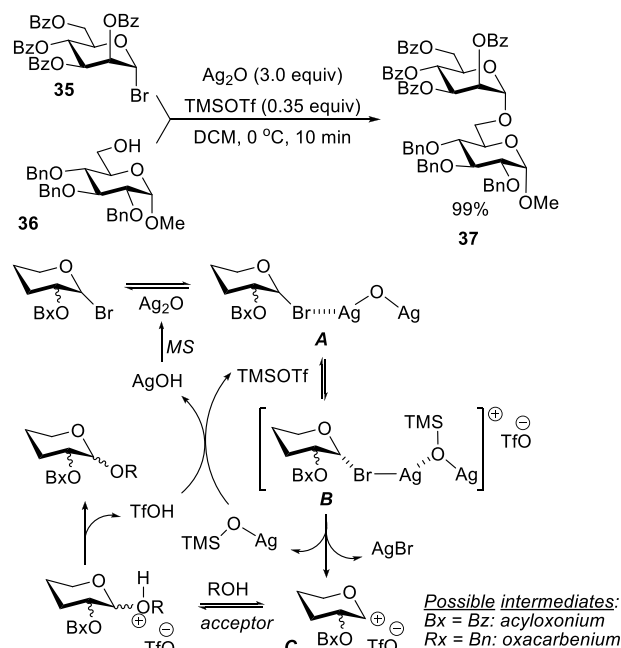
Sugar	HOFox	TMSOTf	Temp	Time	Yield
Glc	--	0.10	0 °C → rt	12 h	35%
Glc	0.25	0.10	0 °C → rt	12 h	85%
Gal	--	0.05	0 °C	1.5 h	19%
Gal	0.25	0.05	0 °C	1.5 h	90%
Man	--	0.08	0 °C	2.5 h	42%
Man	0.25	0.08	0 °C	2.5 h	98%

stoichiometric amount of bromine to afford glycosyl bromide **31**. The latter was dried *in vacuo* and used without purification for the regenerative glycosylation cycle wherein both the synthesis of the reactive OFox imide intermediate **33** and its glycosidation were performed in a catalytic, regenerative fashion. First, HOFox aglycone **32** used in catalytic amounts was reacted with glycosyl bromide **31** in the presence of silver oxide to afford OFox imide **33**.²⁷⁶ Second, glycosyl acceptor and a Lewis acid (TMSOTf) were added afford the corresponding glycosides **34**. The leaving group from the OFox intermediate departs as HOFox aglycone **32** that can be used to generate the next batch of OFox imide **33**. It was also shown that the rate of glycosylation reaction can be increased by increasing the amount of HOFox used. As illustrated in Scheme 8, only 0.25 equiv of HOFox additive was very effective at enhancing glycosidations of glycosyl bromides of the gluco, galacto, and manno series. More recently, a streamlined procedure that allows for bypassing the intermediacy of glycosyl bromides was reported.²⁷⁷

Very recently, Singh and Demchenko discovered an acid-catalyzed silver salt-promoted glycosyl halide activation. It was observed that the addition of a catalytic amount of TMSOTf to a silver oxide-promoted glycosylation dramatically accelerates the reaction and rapidly affords glycosides in very high

yields.¹⁴⁶ As depicted in Scheme 10, TMSOTf-catalyzed glycosidation of mannosyl bromide **35** with glycosyl acceptor

Scheme 10. TMSOTf-Catalyzed Koenigs–Knorr Glycosylation



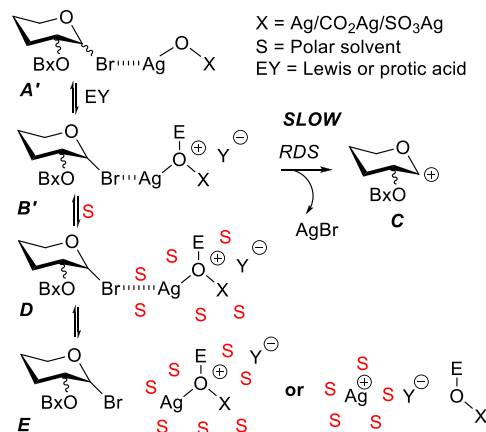
36 afforded disaccharide **37** in 99% yield in 10 min. This new reaction was applied to the synthesis of a variety of glycosides of different series. A tentative mechanism was proposed, wherein it was assumed that silver oxide coordinates with anomeric bromide (Intermediate A) and the oxide oxygen gets silylated with TMSOTf (intermediate B). This helps to shift the equilibrium of the reaction, reinforces the Ag–Br bond formation, and results in the release of AgBr that precipitates from the reaction mixture making the reaction irreversible (Scheme 10).

Depending on the C-2 protection group, either an oxacarbenium or acyloxonium ion is formed as a reactive intermediate (intermediate C), which then reacts with the glycosyl acceptor. After deprotonation and TMS exchange, TMSOTf along with silver hydroxide are formed. TMSOTf gets cycled back into the reaction, and AgOH is decomposed to silver oxide by losing a water molecule to the desiccant (molecular sieves). The authors eliminated a possibility of the *in situ* formation of AgOTf by showing that these conditions are ineffective for the activation of glycosyl STaz²⁹ and SBox^{27,28} glycosyl donors that are readily activated with AgOTf. While no further mechanistic evidence has been presented, it is certainly possible that the intermediate B may undergo an S_N2-like displacement instead of leading to the oxacarbenium ion. Modest stereoselectivity observed in reactions with glycosyl donors equipped with a nonparticipating group at C-2 is indicative of the intermediacy of the oxacarbenium ion.

From the proposed mechanistic pathway, it became clear that as little as 0.50 equiv of silver oxide (stoichiometric silver) should be sufficient for the complete consumption of glycosyl bromide donor. Further investigation of roles of different silver salts, Lewis/Bronsted acid additives, and solvents revealed further particulars of this cooperatively catalyzed reaction.²²¹

Thus, it was noted that reactions in polar solvents proceed much slower and are prone to side processes. Silver(I) oxide and acid-catalyzed glycosylation in nonpolar solvents proceed as explained in Scheme 10, and the identical reaction pathway was envisaged for both Lewis acid and protic acid cocatalysis. In polar solvents, however, it was assumed that upon addition of Lewis or Bronsted acids, strongly ionized species get solvated in polar solvents as shown for intermediate D in Scheme 11. The solvation reduces the effective interaction with

Scheme 11. Cooperatively Catalyzed Koenigs–Knorr Glycosylation in Polar and Nonpolar Solvents



the anomeric bromide E thus resulting in longer reaction times or incomplete consumption of glycosyl bromide or formation of side products. Ultimately, this investigation led to a successful activation of glycosyl bromides in the presence of 0.50 equiv of silver oxide and 0.35 equiv of TfOH in toluene. One of the most interesting aspects of the reaction was that the progress and completion of the reaction could be monitored by eye due to stark visual changes, when the glycosidation proceeds from dark brown-black appearance due the presence of Ag₂O of the reaction mixture to complete decolorization when 0.50 equiv of Ag₂O has been entirely consumed.

An unusual reactivity trend has been unveiled in these cooperatively catalyzed glycosylation conditions where benzoylated α -bromides **35** and **11** turned out to be much more reactive compared to their benzylated counterparts **39** and **38** (Figure 2). The higher reactivity of benzoylated α -bromides compared to their benzylated counterparts strikingly contradicts the armed-disarmed theory proposed by Fraser-Reid.²⁷⁸

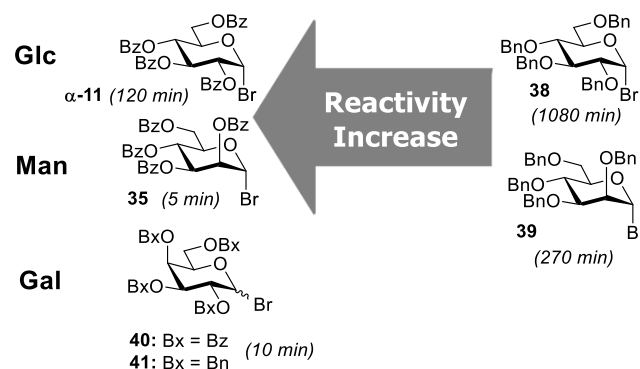
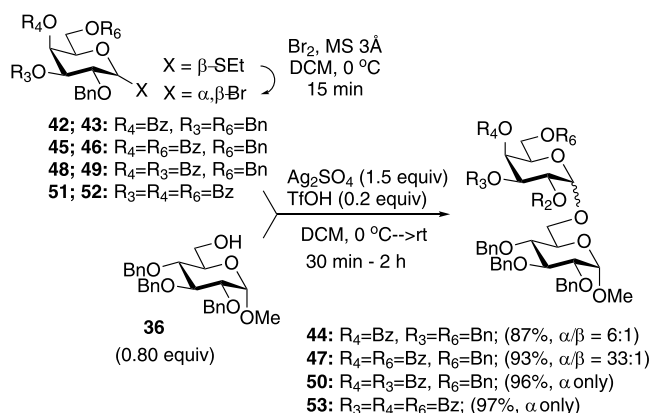


Figure 2. Contradicting reactivity trends in the cooperatively catalyzed Koenigs–Knorr glycosylations.

This was found to be consistent irrespective of silver salts, acids, and sugar series employed with a less pronounced effect in highly reactive galactosyl bromide donors **40** and **41**.

The glycosidation of glycosyl bromide donors bearing a nonparticipating (benzyl) group at C-2 under cooperative catalysis produce glycosides in good yield albeit no stereo-selectivity. To address this issue, Demchenko and co-workers performed a preliminary investigation of 1,2-*cis* selective galactosylation reaction by optimizing the reaction conditions in combination with switching the location of common protecting groups OBn and OBz across different position of galactosyl bromides.²²⁶ For this purpose, galactosyl bromide **43**, obtained from bromination thioglycoside **42**, was used as glycosyl donor and methyl 2,3,4-tri-*O*-benzyl- α -D-glucopyranoside acceptor **36** in the presence of Ag₂SO₄ and TfOH (Scheme 12). Disaccharide **44** was obtained in 87% yield with

Scheme 12. Acyl Group-Directed α -Stereoselective Galactosylation in Cooperatively Catalyzed Glycosylations



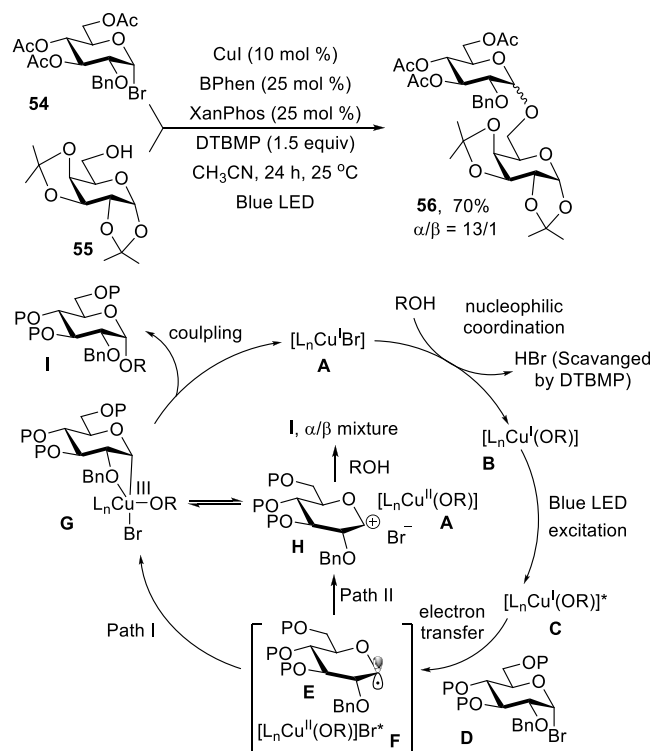
predominant α -selectivity (α/β = 6.0:1). From this study, the authors concluded that the 4-*O*-acyl group is strictly necessary, as previously discovered by Boons,²⁷⁹ but it may be insufficient to drive α -selective galactosylation under cooperative catalysis of Ag₂SO₄ (1.5 equiv) and TfOH (0.20 equiv) in dichloromethane.

Subsequently, the authors achieved an excellent 1,2-*cis* α -galactosylation from galactosyl bromide **46** having 4- and 6-OBz groups. The bromide donor **46** was obtained by bromination of thiogalactoside **45**. Glycosidation of donor **46** with glycosyl acceptor **36** produced disaccharide **47** in 93% yield (α/β = 33.0:1) as shown in Scheme 12. Following this general line of thought, thiogalactoside **48** having 3- and 4-OBz groups and **51** having 3-, 4-, and 6-OBz groups were synthesized, which were later brominated to produce galactosyl bromides **49** and **52**, respectively. Glycosidation of both galactosyl bromides **49** and **52** with glycosyl acceptor **36** produced the respective disaccharides **50** and **53** in excellent yields and with exclusive α -selectivity. Construction of different types of 1,2-*cis* galactosidic linkages was then extended to a variety of differently protected glycosyl acceptors.

Very recently, Nguyen and co-workers developed a method for copper-catalyzed activation of glycosyl bromides in the presence of visible light.¹⁴⁷ Reaction optimization revealed that copper iodide (CuI), 4,5-bis(diphenylphosphino)-9,9-dimethylxanthene (XantPhos), 4,7-diphenyl-1,10-phenanthroline (BPhen), and di-*tert*-butylmethylpyridine (DTBMP) are

strictly necessary to induce an efficient coupling between glycosyl bromide **54** and glycosyl acceptor **55** in acetonitrile affording disaccharide **56** in 72% yield (Scheme 13). The

Scheme 13. Visible Light-Mediated Cu(I)-Catalyzed 1,2-*cis* α -Selective Glycosylation



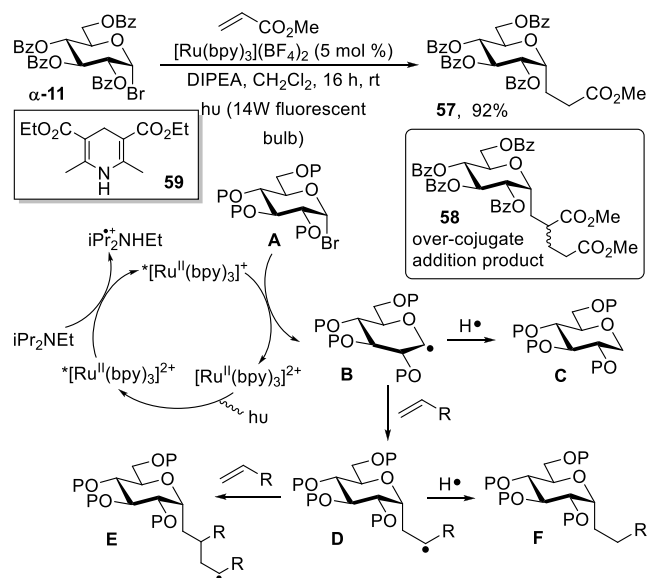
comparison of chemical shift value (³¹P NMR, δ = -11.99 ppm) obtained from in situ generated complex was identical with the isolated complex [Cu(BPhen)(Xantphos)]BF₄ (³¹P NMR, δ = -11.87 ppm) and the glycosylation outcome from the isolated catalyst, [Cu(BPhen)(Xantphos)]BF₄ was similar to the aforementioned conditions, indicating [Cu(BPhen)(Xantphos)]⁺ as an active catalyst for this transformation. The optimized reaction conditions facilitated the construction of glycosidic linkages using coupling partners of different sugar series.

In addition to the outcome of glycosylations, data from absorption and emission spectroscopies and electrochemistry experiments suggested the following mechanistic pathway for the visible light-mediated copper-catalyzed activation of glycosyl bromide. The initial step includes the reaction of Cu(I) catalyst A with alcohol to produce Cu(I)-oxygen complex B (Scheme 13).^{280,281} Upon photoirradiation (blue light) the latter is converted to excited Cu(I)-complex C that enables electron transfer to glycosyl bromide D to afford glycosyl radical E and Cu(II) complex F. This can potentially lead to either Cu(III) complex G²⁸²⁻²⁸⁵ via Path I or oxocarbenium ion intermediate H via Path II.²⁸⁶ Control experiments were performed by replacing oxygen at C-2 with fluorine/hydrogen atom. Fluorine is known to interact with the copper center;²⁸⁷ indeed, 2-fluoro-2-deoxy-derivative favored α -1,2-*cis* selectivity. Conversely, the 2-deoxy derivative led to the formation of an α/β mixture, which could be due to the formation of a 2-deoxy cation. These observations strongly suggest that α -1,2-*cis* selectivity does not arise from the oxocarbenium ion intermediate H generated either via Path II

or from the collapse of Cu(III) complex G. This concludes the Path I is more likely to be operative wherein reductive elimination of Cu(III) complex G affords glycoside I and regenerates Cu(I) catalysts to endure the catalytic glycosylation cycle.

Herein, we are describing a representative example to construct C-glycosidic linkages by activating glycosyl bromides with a combined effect of visible light and $[\text{Ru}(\text{bpy})_3]^{2+}$.²⁸⁸ Thus, as described by Gagné and co-workers, these reactions produce a glycosyl radical from glycosyl bromide, which gets intermolecularly trapped with electron-deficient alkene to afford C-glycosides in a stereoselective manner. For example, per-benzoylated glycosyl bromide α -11 produced the corresponding glycosyl radical upon visible light-mediated activation in the presence of $[\text{Ru}(\text{bpy})_3](\text{BF}_4)_2$ (5 mol %) and a stoichiometric amount of *N,N*-diisopropylethylamine (DIPEA) in DCM. Once formed, the glycosyl radical gets immediately consumed by methyl acrylate to produce the corresponding α -C-glycoside 57 (Scheme 14). The authors observed the

Scheme 14. Visible Light-Mediated Intermolecular Addition of Glycosyl Bromide to Methyl Acrylate

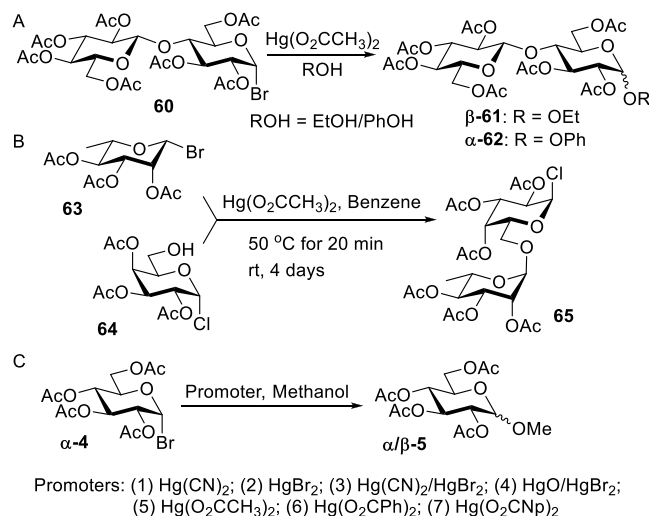


formation of an overconjugate addition product 58. This side reaction could be suppressed by the addition of Hantzsch ester 59, which also was found to accelerate the reaction. This observation allowed to obtain α -C-glycoside 57 in a very impressive 92% yield and led to an improved understanding of the mechanistic pathway. As shown in Scheme 14, as electron transfer occurs from the photogenerated species $[\text{Ru}(\text{bpy})_3]^+$ to anomeric bromide A producing glycosyl radical B. The latter can get reduced to produce C or can be trapped by an electron-deficient alkene to produce C-glycoside radical D. Subsequent step depends on relative rates for the formation of overconjugate product E versus that of the formation of expected C-glycoside F. As it was mentioned, Hantzsch ester 59 accelerates the reduction of D into F.

3.2.2. Activation with Mercury and Other Group 12 Metal Salts (Zinc or Cadmium). As aforementioned, Zemlen and co-workers showed an efficient activation of cellobiosyl bromide 60 in the presence of aluminum metal and mercuric acetate $\text{Hg}(\text{OAc})_2$. However, it was later realized that aluminum did not play an active role in splitting the C—Br

bond.^{160,161} The anomeric distribution of glucosides was difficult to predict, and a subsequent dedicated study revealed that the ratio of α/β -linked glycosides mainly depends on the amount and types of alcohol used.^{160,162,163} For example, as shown in Scheme 15A, the treatment of glycosyl bromide 60

Scheme 15. Mercury Salt-Promoted Glycosidations of Glycosyl Bromides



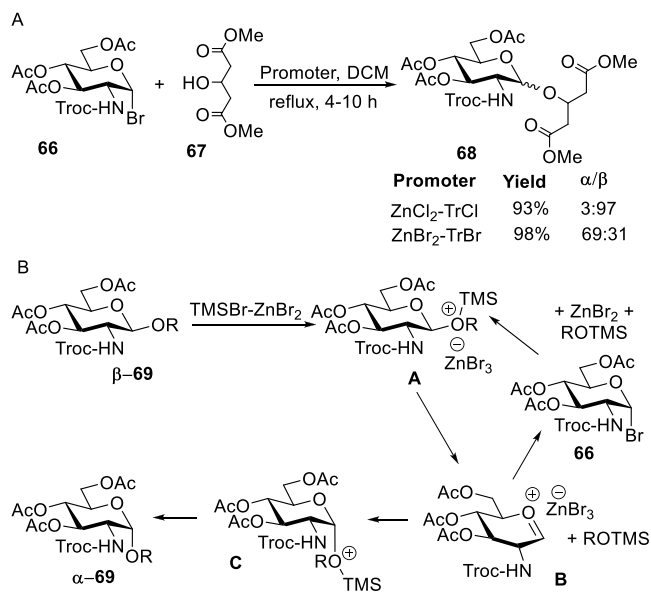
with ethanol or phenol in the presence of $\text{Hg}(\text{OAc})_2$ produces the respective β -glycoside 61 and α -glycoside 62. The α -anomer formation was favored when the reaction was performed with an excess of alcohol. Zemlen and co-workers applied this methodology to the synthesis of several disaccharides wherein selective activation of the leaving group was employed. For example, selective glycosidation of acetobromorhamnose 63 with partially protected galactosyl chloride 64 was performed in the presence of $\text{Hg}(\text{OAc})_2$ to produce disaccharide chloride 65 (Scheme 15B).²⁸⁹ Several other disaccharide chlorides were synthesized employing this selective activation of glycosyl bromides over chlorides in the presence of $\text{Hg}(\text{OAc})_2$.²⁹⁰ The selective activation approach helps to streamline oligosaccharide assembly because the products are already equipped with a leaving group, chloride in this case, and hence can be used as glycosyl donors in subsequent glycosylations directly.

Helferich and co-workers showed that a very fast activation of glycosyl bromide α -4 can take place in the presence of mercuric cyanide. This powerful improvement of the basic methodology earned a name of the Helferich modification of the classical Koenigs–Knorr method.²³⁹ By varying the amount of alcohol and mercuric cyanide, a rapid formation (30–100 min) of methyl β -glycoside 5 from glycosyl bromide α -4 and methanol has been achieved in good yields (up to 83%, Scheme 15C). It was also found that the addition of mercuric bromide to a mercuric cyanide-promoted glycosylation, further increases the rate of the reaction, whereas mercuric bromide used by itself did not produce any glycoside product. However, later it was shown that mercuric bromide can activate glycosyl bromide donors in the presence of MS 4 Å in refluxing methylene chloride. These reactions produced α -glycosides.^{241,242} Several other mercuric salts were tested (Scheme 15C), and some aryl mercuric salts such as phenyl mercury acetate, and naphthyl mercury acetate turned out to be effective promoters due to their better solubility in organic

solvents.²⁴³ Green and co-workers have shown that mercuric oxide in combination with a small amount of mercuric bromide is also capable of an effective activation of glycosyl bromides.²⁴⁰ Meldal and Bock showed that mercuric iodide alone is capable of activating glycosyl bromides.²⁴⁴ Mercuric salt-catalyzed glycosylation method was employed for the synthesis of different types of biologically relevant glycosides and oligosaccharides.^{181,242,291,292}

In other attempts to replace silver salts in glycosylations, Helferich and co-workers investigated zinc salts as activators of glycosyl bromides.²³⁹ For example, when glycosidation of bromide α -4 with methanol was conducted in the presence of zinc oxide methyl β -glucoside, **5** was obtained in 37% yield. Different primary alcohols were tested, and the corresponding glycosides were obtained in good yields (41–76%). Increasing the amount of zinc oxide was detrimental for the glycosylation reaction and led to decreased yields of glycosides. However, changing the reaction solvent from nonpolar to polar showed a positive effect. A slower activation was observed when zinc acetate was used instead under otherwise identical reaction conditions. Kusama and co-workers showed that glycosyl bromide **66** can be activated with equimolar amounts of zinc chloride and trityl chloride. Glycosylation of acceptor **67** produced glucoside **68** in a high yield with excellent β -stereoselectivity (Scheme 16A).²⁴⁵

Scheme 16. Glycosylation and Anomerization in Zinc Salt-Promoted Glycosylations

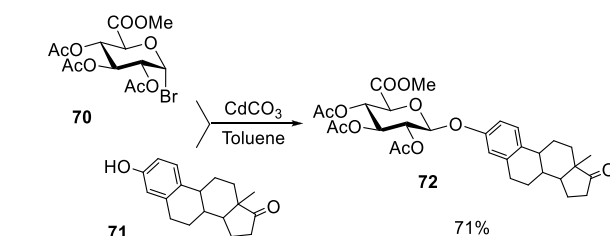


A similar activation was also observed with ZnBr₂ and trityl bromide (TrBr), but a notable loss of stereoselectivity was observed (Scheme 16A). In this case, α -glucoside **68** was obtained as the major product.²⁴⁷ Several other alcohols were glycosylated, and a similar stereoselectivity trend was observed. It is well-known that Lewis acids such as TiCl₄, AlCl₃, ZnCl₂, SnCl₂, and BF₃-Et₂O promote anomerization of glycosides.^{293–299} Analysis of the reaction mixture revealed that in this case the formation of β -glucoside takes place first, which is then followed by anomerization as the reaction progresses. A dedicated study confirmed that HBr generated during glycosylation induces the anomerization of the kinetic β -glucoside product to the thermodynamic α -glucoside. A

plausible mechanistic pathway for anomerization in the presence of ZnBr₂ and TMSBr was proposed (Scheme 16B).^{247,248} Reaction of β -glycoside β -69 in the presence of TMSBr-ZnBr₂ produced salt A, which collapsed to oxacarbenium ion B and trimethylsilyl ether. Nucleophilic displacement by bromide ion (ZnBr₃[−]) can produce glycosyl bromide **66**. Conversely, a more thermodynamically stable product C can be formed by the nucleophilic attack of trimethylsilyl ether to form α -69. It was later discovered that adding 1.0–2.0 equiv of a glycosyl acceptor can drastically decrease the formation of glycosyl bromide and afford predominantly α -linked products. This methodology was applied to the synthesis of 1→6-linked glycolipid derivatives.^{300,301} Surprisingly, only β -disaccharides were obtained irrespective of the promoter system (TrCl-Zn salts or zinc salts alone). When β -linked disaccharides were exposed to the anomerization conditions (TMSBr-ZnBr₂), only the starting material was recovered. The stability of β -linked disaccharide lipid derivatives was attributed to the inductive effect of the oxygen functional group at the reducing end. Shibakami and co-workers presented a similar activation protocol for per-benzoylated glycosyl bromide wherein NBS and catalytic ZnI₂ or ZnBr₂ were employed.²⁴⁶ In the case of ZnI₂, the reaction was found to proceed via intermediacy of the respective glycosyl iodide formed *in situ*.

Helferich and co-workers reported a cadmium salt-promoted activation of glycosyl bromide donors.²³⁹ The authors demonstrated that reaction of per-acetylated glycosyl bromides with MeOH in the presence of cadmium oxide afforded the corresponding methyl β -glucoside in a moderate yield. Bernstein and Conrow investigated the Cd salt-promoted glycosylation in a greater detail by employing glycosyl halides of different sugar series.²⁴⁹ For example, glycosidation of glycosyl uronide donor **70** with estrone acceptor **71** in the presence of CdCO₃ produced glucuronide **72** in 71% yield (Scheme 17). Cadmium sulfide was also investigated for the

Scheme 17. Cadmium Salt-Promoted Glycosylation

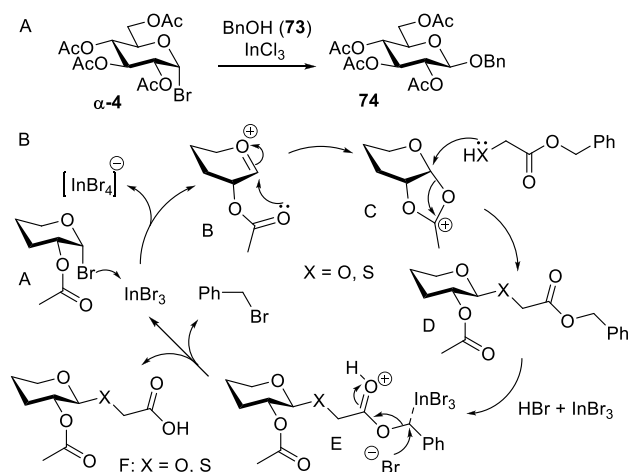


activation of donor **70**; however, the yield of product **72** was only 20%. Several other steroidal phenolic glucuronides, glycosides, and acetyl glucosaminides were successfully synthesized using similar glycosylation conditions.^{250,302,303} In addition to β -glycosides, small amounts of the corresponding α -glucosides and/or C-glycosides were also isolated, and the ratio of products was found to depend on the coupling partners and the reaction conditions employed. It was suggested that the generated cadmium bromide was the actual activator, but cadmium bromide by itself was ineffective, similar to Helferich's observations made with mercuric cyanide activation.²³⁹ Later, it was also confirmed that the product composition depends on the surface area of cadmium carbonate.²⁵¹

3.2.3. Activation with Group 13–15 Post-transition Metal Salts (Indium, Tin, Lead, or Bismuth). Chowdhury

and co-workers developed indium chloride-promoted activation of glycosyl bromides (Scheme 18A).²⁵² A catalytic amount

Scheme 18. Indium Salt-Promoted Glycosidations of Glycosyl Bromides

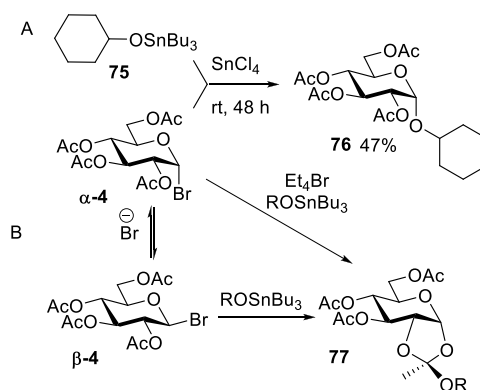


of indium chloride without any additive was sufficient to promote the glycosylation. For example, when tetra-acetylated glucosyl bromide α -4 was treated with benzyl alcohol (73, 1.0 equiv) in the presence of InCl_3 (0.4 equiv), glucoside 74 was obtained in 68% yield. Several other glycosides and disaccharides were successfully synthesized in good yields (62–90%). Indium chloride-catalyzed C-glycosylations of pyrrole and indoles were also described.³⁰⁴ In addition, various O- and S-glycosyl esters were synthesized by activation of glycosyl bromides with catalytic InCl_3 .²⁵³

It was also found that glycosylation and removal of benzyl esters can be achieved in one pot in the presence of indium bromide (InBr_3). A tentative mechanism involving coordination of the anomeric bromide A with indium tribromide to afford the reactive intermediate, oxacarbenium ion B, which gets stabilized as the acyloxonium ion C has been proposed (Scheme 18B). Following the nucleophilic attack and proton exchange, InBr_3 and HBr were generated. The latter is capable of protonating benzyl ester D to produce intermediate E, which undergoes the nucleophilic attack by the bromide ion to produce O- or S-glycosyl acids F and benzyl bromide as a side product. To broaden the scope of indium salt-catalyzed glycosylations, Xue and co-workers investigated several other indium salts. The authors found that as low as 0.05–0.15 equiv of $\text{In}(\text{NTf}_2)_3$ without any external additive can catalyze glycosidations of glycosyl bromides with simple alcohols.²⁵⁴ To achieve efficient glycosylation of sugar alcohols, 2.0 equiv of glycosyl acceptors was needed.

Matsui and Ogawa showed that glycosidation of glycosyl bromides with trialkylstannyl alkoxide acceptors can be achieved in the presence of and tin(IV) chloride.²⁵⁵ Different alkoxide acceptors produced either α - or β -glycosides albeit with fair stereoselectivity. For example, when tetra-acetylated glucosyl bromide α -4 was reacted with tributylstannylated cyclohexyloxide 75 in the presence of SnCl_4 . The corresponding α -glucoside 76 was obtained in 47% yield, as shown in Scheme 19A. When similar reaction conditions were applied to glycosylation of the tributylstannyl benzyloxide acceptor, the corresponding β -glucoside was obtained in 52% yield. By employing the halide ion-catalyzed anomerization procedure in

Scheme 19. Tin Salt-Mediated Glycosidation of Glycosyl Bromides



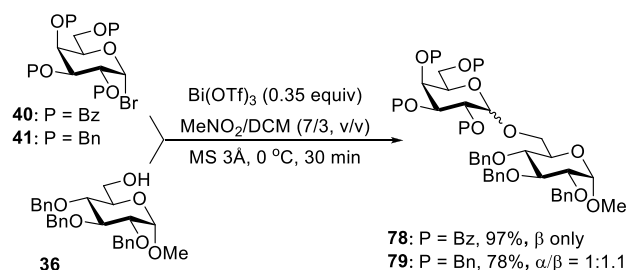
the presence of Et_4NBr ,³⁰⁵ several trialkylstannyl alkoxide acceptors were converted to 1,2-orthoesters 77 (Scheme 19B).¹⁸³

Malleron and Lubineau developed β -stereoselective glycosidations of glycosyl bromides by employing tin triflate as an activator.²⁵⁶ One equivalent of a base as an acid scavenger was necessary to produce glycosides in good yields. The authors observed a competitive transesterification side reaction when primary alcohols were used as glycosyl acceptors. This led to compromised yields, for example, methyl glycoside was obtained in 33% yield. Secondary alcohols, however, did not seem to encounter this side reaction and produced glycosides in better yields. Several β -glycosides, β -disaccharides, β -aminosugars,³⁰⁶ and N-linked- β -disaccharides³⁰⁷ were successfully synthesized. In another study, the authors successfully applied tin triflate-promoted activation of glycosyl chloride donors (*vide infra*).³⁰⁶ Dick has reported activation of glycosyl bromides in the presence of lead(II) carbonate, wherein a moderate yield (37%) for the synthesis of tetra-acetylated α,β -phenyl glucosides was obtained when per-acetylated glucosyl bromide was treated with PhOH in the presence of PbCO_3 .²⁵¹

Recently, Demchenko and co-workers have shown an efficient activation of glycosyl halides in the presence of $\text{Bi}(\text{OTf})_3$.³⁰⁸ Compared to most common metal salt activators of glycosyl halides wherein stoichiometric amount of activators is needed, a rapid activation of glycosyl bromides was achieved in the presence of 0.35 equiv of $\text{Bi}(\text{OTf})_3$. Due to the detrimental effect of strongly acidic TfOH produced as a byproduct, the authors have increased the amount of molecular sieves and identified nitromethane-DCM as the preferred reaction solvent. As a result, glycosidation of benzoylated and benzylated galactosyl bromides 40 and 41 with glycosyl acceptor 36 in the presence of $\text{Bi}(\text{OTf})_2$ (0.35 equiv) afforded respective galactosides 78 and 79 in good yields (Scheme 20). An effective glycosylation of a variety of glycosyl acceptors was achieved for sugars of the gluco, manno, rhamno, and glucosamino series equipped with different protecting groups.³⁰⁸

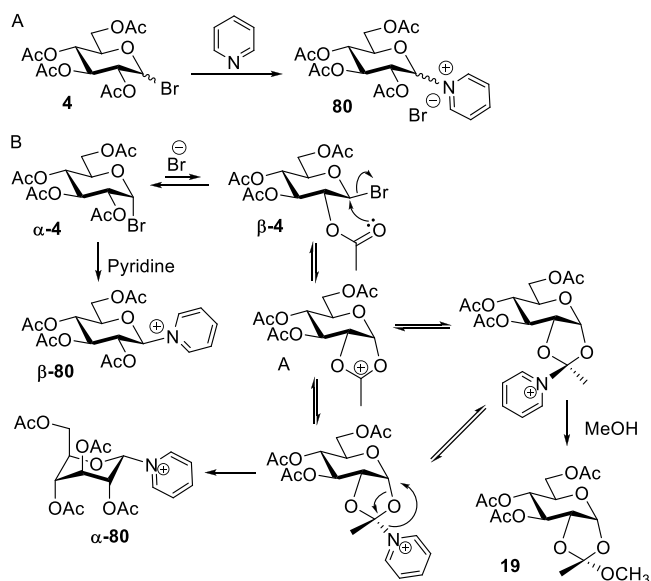
3.2.4. Activation with Non-nucleophilic Organic Bases. Koenigs and Knorr were the first to report non-metal-based activation of glycosyl bromides, wherein glycosyl bromide was treated with methanol in the presence of pyridine that afforded methyl β -glycoside.⁷ The exact mechanistic action of pyridine was not discussed or known, rather it was assumed that pyridine acts as hydrogen bromide scavenger. A few years later, Fischer and co-workers showed the formation

Scheme 20. Bismuth(III) Salt-Promoted Activation of Glycosyl Halides



of anomeric mixtures of pyridinium salts α/β -80 when tetra-*O*-acetylated glucosyl bromide α/β -4 was treated with pyridine in the absence of an alcohol (Scheme 21A).¹⁴⁸ This undoubtedly excluded the possibility of pyridine purpose solely as the hydrogen halide scavenger.

Scheme 21. Reaction of Glycosyl Bromide 4 in the Presence of Pyridine



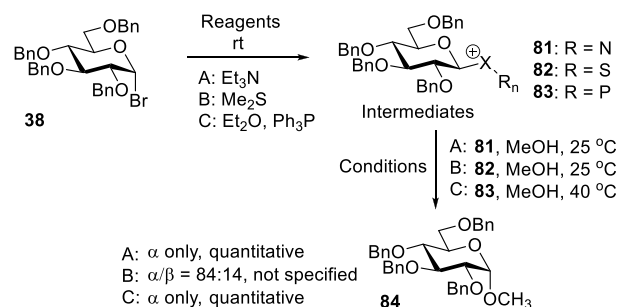
Fischer also investigated glycosylations in the presence of quinoline.¹⁵⁰ Lemieux and Morgan also observed the formation of anomeric pyridinium salts.¹⁸² To gain mechanistic insights for this reaction, the authors performed an extensive concentration-dependence study depicted in Scheme 21B. It was found that the anomeric distribution of α - and β -pyridinium salts 80 depends on the concentration of glycosyl bromide α -4 in pyridine. At a low concentration, the formation of β -80 predominates, but at higher concentrations both α - and β -80 are produced and the ratio of anomers was found to depend on the concentration of glycosyl bromide α -4. When the equimolar amount of tetra-*n*-butyl ammonium bromide was added to the reaction, only α -pyridinium salt 80 was obtained, whereas reactions in the presence of tetra-*n*-butylammonium perchlorate produced the anomeric mixture.

These results were indicative of the intermediacy of β -bromide β -4 and acyloxonium intermediate A. In the presence of methanol this reaction led to a significant amount of orthoester 19. NMR spectroscopy studies suggested that α -pyridinium salt α -80 adopts the ¹C₄ conformation, as shown in Scheme 21B.³⁰⁹ Further experiments were conducted to

support the routes for the formation of α -80 and its β -linked counterpart,^{183,310} which ultimately led to appreciation that charged intermediates can serve as glycosyl donors. A variety of other approaches to obtain charged intermediates followed by their glycosidation have been explored. Micheel and Micheel examined the formation of positively charged anomeric salts from triethylamine. However, the authors never used these salts as a glycosyl donors.^{264,311} Hess and Heumann treated a glycosyl chloride with trimethylamine in the presence of ethyl alcohol and observed the formation of ethyl α -glycosides in addition to the anticipated anomeric salts.³¹²

Expanding upon these findings, Schuerch and co-workers performed a dedicated study of the formation and glycosidation of positively charged intermediates that led to the development of a highly stereocontrolled α -glycosylation.^{265,266} It was shown that positively charged intermediates 81–83 depicted in Scheme 22 can be synthesized by treating

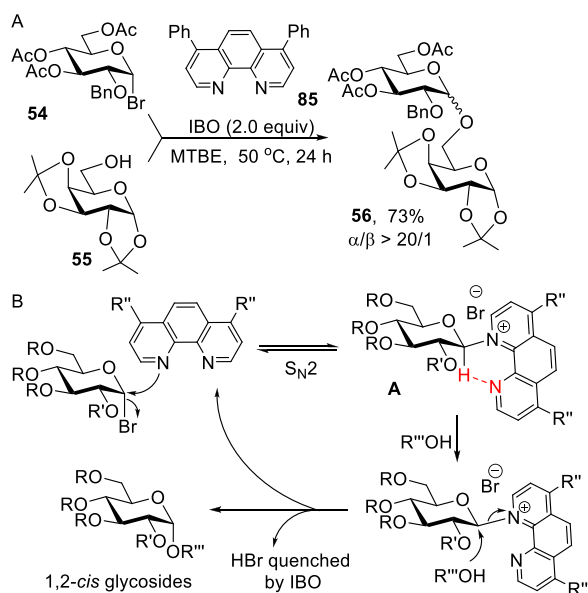
Scheme 22. Synthesis of α -Glycoside via Positively Charged Glycosyl Donors



glycosyl bromide 38 with triethylamine, dimethyl sulfide, or triphenylphosphine, respectively. These intermediates can be isolated and purified. The positively charged anomeric substituents would have a strong propensity to adopt the equatorial position due to the reverse anomeric effect.³⁰⁹ As a result, these positively charged equatorially placed leaving groups would undergo the nucleophilic attack from the α -face to give axial glycoside products.

Thus, glycosidation of ammonium 81 and phosphonium 83 salts produced α -glycosides 84 exclusively, whereas glycosidation of sulfonium 82 salt showed somewhat relaxed stereoselectivity (Scheme 22).²⁶⁵ An attempt to synthesize an α -1 $\rightarrow$ 3-linked disaccharide has failed, and this outcome was attributed to low reactivity of the sugar alcohol. Upon subsequent optimization of the reaction conditions wherein reaction solvents and temperatures were refined, a successful synthesis of an α -1 $\rightarrow$ 6-linked disaccharide has been achieved.³¹³

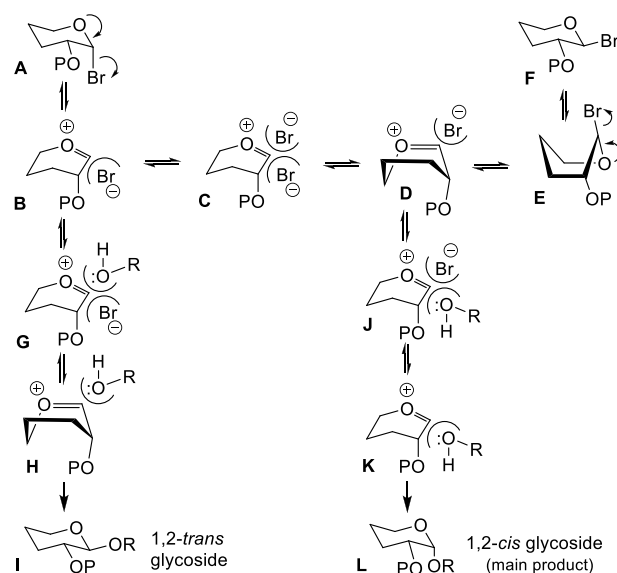
Very recently, Nguyen and co-workers presented an improved approach to achieve stereoselective activation of glycosyl bromides bearing a nonparticipating group at C-2.²⁵⁷ It was discovered that the activation of glycosyl bromide 54 with phenanthroline catalyst 85 in the presence of isobutylene oxide (as hydrogen bromide scavenger) preferentially afforded 1,2-*cis* glycosides. For example, glycosylation of glycosyl acceptor 55 produced disaccharide 56 in 73% with excellent α -stereoselectivity, as shown in Scheme 23A. The mechanistic pathway involves double S_N2-like displacement wherein the first step involves the formation of β -phenanthrolium intermediate A depicted in Scheme 23B. The stability of intermediate A was attributed to noncovalent interaction of

Scheme 23. 1,2-*cis* Glycosylation via β -Phenanthrolium Intermediates

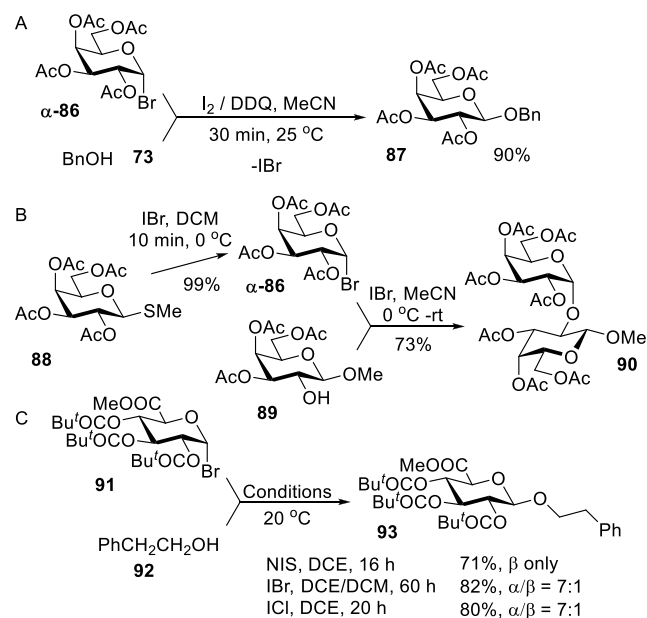
anomeric α -hydrogen with nitrogen of phenanthroline. Subsequently, intermediate A reacts with a glycosyl acceptor to form an α -linked product. This stereoselective glycosylation methodology was utilized to synthesize many glycosidic linkages as well as a series of α -glycans (*vide infra*).

3.2.5. Activation with Halogens, Halide Ions, or Halonium Ions. Lemieux and co-workers performed an extensive study of a halide ion-catalyzed glycosylation with glycosyl halides. This approach has led to effective 1,2-*cis* stereoselective syntheses of glycosides and oligosaccharides.²⁵⁸ The early attempts to address the issues of stereoselectivity were primarily directed on the development of new catalytic systems, as well as optimization of the reaction conditions (solvent, temperature, pressure).³¹⁴ Many valuable developments for the synthesis of 1,2-*cis* glycosides have been published by Schuerch and co-workers,^{72,230,265,315–318} However, a major breakthrough in the understanding of the principles of the α -glycosidic bond formation emerged with the discovery and thorough elaboration of the *in situ* anomerization concept, so-called “halide ion catalyzed glycosidation reactions” by Lemieux and co-workers (Scheme 24).²⁵⁸ Thus, it was observed that a rapid equilibrium could be established between a relatively stable α -halide A and its far more reactive β -counterpart F by the addition of tetraalkyl ammonium bromide (Et_4NBr). Therefore, a glycosyl acceptor (ROH) would attack the more reactive intermediate in an overall $\text{S}_{\text{N}}2$ fashion (via D), providing α -glycoside L. More detailed analysis of the glycosylation process showed that the energy barrier for a nucleophilic substitution F to L (formation of α -glycosides from the highly reactive β -bromide F) is somewhat lower than that for the reaction from A to I (formation of β -glycosides from the less reactive α -bromide A).

Indeed, α -glycoside L is formed faster than its β -linked counterpart I, which in combination with higher thermodynamic stability of the α -anomer makes this glycosylation process very favorable overall. If the difference in the energy barrier had been sufficient, it would be possible to direct the reaction toward the formation of α -anomers with complete stereoselectivity. Therefore, in order to achieve high stereo-

Scheme 24. Formation of 1,2-*cis* Glycosides via *In Situ* Anomerization

selectivity, the entire glycosylation process has to be performed in a highly controlled manner. In this particular case the control is achieved by the use of extremely mild catalyst (Et_4NBr), although very reactive substrates and prolonged reaction times are thus required. A series of per-benzylated glycosyl halides of the gluco, galacto, and fuco series were tested. These reactions produced good yields of glycosides with excellent α -stereoselectivity. This method found practical application in synthesizing blood group determinants and other biologically relevant oligosaccharides having α -glycosidic linkages (*vide infra*).^{259,319} In another effort to find a substitute to heavy metal promoters, Field and co-workers studied the activation of per-acetylated glycosyl bromides by utilizing I_2 alone or with DDQ as an additive. As shown in Scheme 25A, when galactosyl bromide α -86 was reacted with benzyl alcohol

Scheme 25. Iodine or Iodine Monobromide/Chloride-Promoted Glycosylations

73 in the presence of I_2 and DDQ at rt, an excellent yield of galactoside **87** was achieved within 30 min.²⁶⁰ It was proposed that iodine acts as a halophilic reagent which, following the interaction with the anomeric bromide, fragments to give iodine monobromide and oxacarbenium or acyloxonium ion. Subsequently, the authors realized that iodine monobromide (IBr) could also be used as an efficient activator for glycosyl bromides. Indeed, IBr alone or with DABCO additive could activate glycosyl bromides to produce different disaccharides in 35–73% yields. The lower yields in this series were generally attributed to side reactions rather than promoter problems. All glycosyl bromide donors were synthesized by treating the corresponding thioglycosides with IBr.

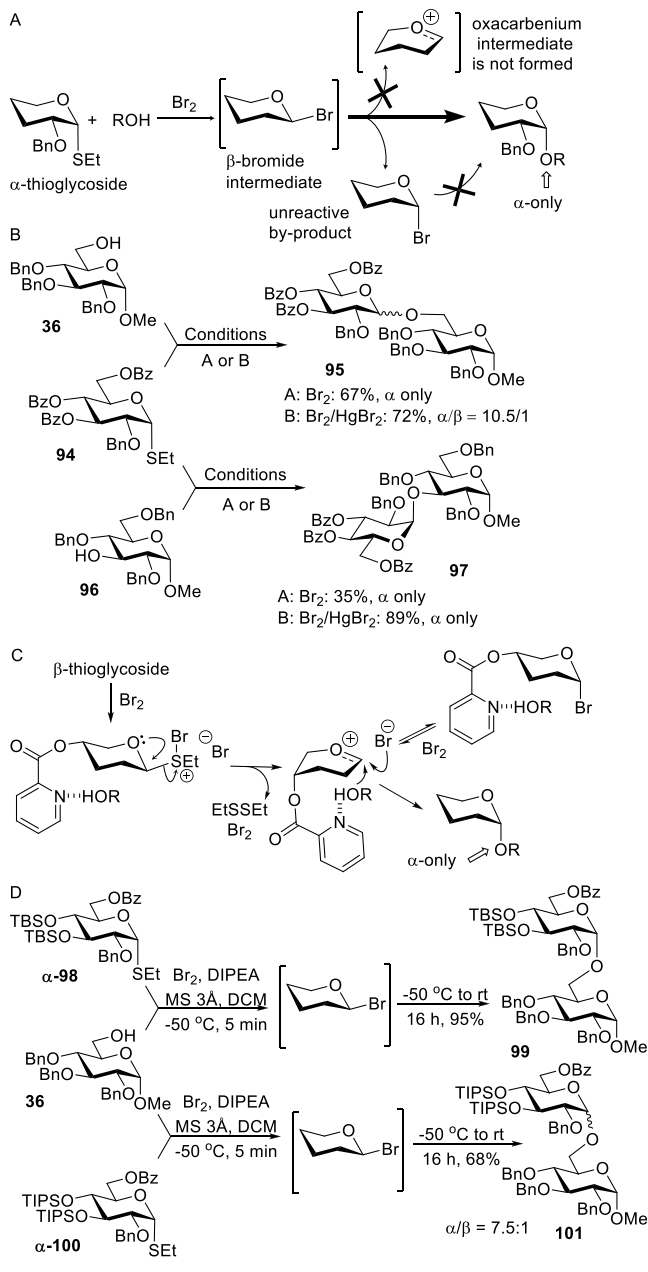
One such example is the synthesis of galactosyl bromide α -**86** from the corresponding methylthio galactoside **88** in the presence of IBr depicted in Scheme 25B. The bromide donor α -**86** was later glycosidated with secondary galactosyl acceptor **89** in the presence of IBr that produced disaccharide **90** in 73% yield.²⁶² Glycosyl chloride donors were also activated under identical reaction conditions. Besides glycosyl halide activation, thioglycoside activation was also shown to occur in the presence of IBr. Iodine-promoted glycosidation of glycosyl halide was further extended to amino acid acceptors.³²⁰ Anhydrous potassium carbonate was used as a scavenger for a hydrogen iodide byproduct.

Stachulski employed *N*-iodosuccinimide as a source of the iodonium ion for the activation of glycosyl bromides.²⁶³ An efficient activation of pivaloyl protected bromide **91** was achieved in the presence of NIS, IBr or ICl at 20 °C. For example, glycosylation of acceptor **92** produced glucuronide **93** in good yields of 71–82% as depicted in Scheme 25C. Excellent to good yields of other glucuronides were achieved with several primary and secondary alcohols. However, the stereoselectivity was found to differ drastically depending on the type of promoter used. Thus, NIS produced the product in high β -stereoselectivity, whereas IBr- or ICl-promoted reactions were α -stereoselective.

Demchenko developed reaction conditions wherein bromine was used both as the reagent to convert thioglycosides into bromides and to activate the latter for glycosylation.³²¹ To achieve highly stereocontrolled 1,2-*cis* glycosylation, it was deemed necessary to form reactive β -glycosyl bromides. It was assumed that α -thioglycoside upon bromination can produce the reactive β -bromide as shown in Scheme 26A. This reaction was monitored by NMR, showing that β -bromide is indeed the reactive intermediate, which can undergo a rapid anomerization into the α -linked counterpart. Once formed, the α -bromide was found to be totally unreactive under the established reaction conditions. The glycosylations of primary acceptors were smooth and stereoselective. For example, glycosidation of thioglycoside **94** with 6-OH glycosyl acceptor **36** in the presence of Br_2 produced disaccharide **95** in 67% yield with complete 1,2-*cis* stereoselectivity (Scheme 26B).

However, slower glycosylations of secondary acceptors were less efficient due to the competing $\beta \rightarrow \alpha$ halide anomerization, although all reactions were still α -stereoselective. For example, glycosidation of thioglycoside **94** with 3-OH glycosyl acceptor **96** in the presence of Br_2 produced disaccharide **97** in 35% yield with complete 1,2-*cis* stereoselectivity. It was also shown that the α -bromide can be reactivated in the presence of mercury(II) additive. This pathway was found to be very beneficial for the glycosylation of secondary alcohols. For example, glycosidation of thioglycoside **94** with 3-OH glycosyl

Scheme 26. Bromine-Promoted α -Selective Glycosylations



acceptor **96** in the presence of Br_2 and $HgBr_2$ produced disaccharide **97** in a significantly improved yield of 89% yield with complete 1,2-*cis* stereoselectivity. However, when applied to glycosylation of highly reactive primary acceptors, these reaction conditions can compromise α -selectivity. Thus, glycosidation of thioglycoside **94** with 6-OH glycosyl acceptor **36** in the presence of Br_2 and $HgBr_2$ produced disaccharide **95** in a slightly improved yield of 72% yield albeit reduced stereoselectivity ($\alpha/\beta = 10.5/1$, Scheme 26B).

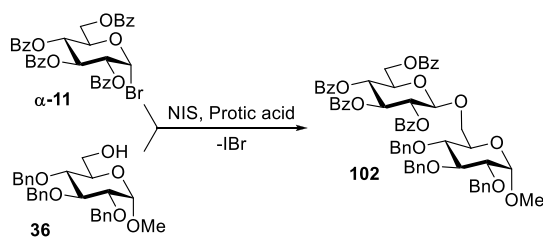
Bromine has also been used as a promoter in glycosylations by means of an H-bond-mediated Aglycone Delivery (HAD).³²² Glucosyl donors bearing the 4-*O*-picoloyl group exhibited excellent stereoselectivity with a number of different glycosyl and aliphatic acceptors.³²³ Furthermore, bromine was also shown to activate a wide variety of leaving groups including thioglycosides, imidates, and thioimides. A low temperature NMR monitoring of the reaction showed direct

and rapid formation of the α -bromide. Differently from the benzoylated bromides (*vide supra*), a highly benzylated α -bromide was reactive under these reaction conditions. Upon departure of the bromide leaving group, the 4-O-picoloyl mediated HAD ensures that the nucleophile is delivered exclusively from the bottom face of the resulting oxacarbenium intermediate (Scheme 26C). Mercuric bromide could also be added to this reaction to improve the yield and shorten the reaction time.

Further efforts were made to apply the bromine-promoted glycosylation reaction to conformationally superarmed glycosyl donors.^{124,125} Dedicated NMR experiments and computational studies revealed that introduction TBS and TIPS groups at C-3 and C-4 positions of β - or α -thioglycosides **98** and **100** distort the ring conformation.¹⁴³ Reactive β -glycosyl bromide formation was seen only with α -**98**, whereas β -**98** afforded an anomeric mixture of bromides. Glycosidation of α -**98** with primary glycosyl acceptor **36** in the presence of Br₂ and DIPEA as an additive provided 1,2-*cis* disaccharide **99** in an excellent yield of 95% (α only) as shown in Scheme 26D. The reactivity trend of the corresponding TIPS protected glycosyl donor **100** was similar; however, the yield and stereoselectivity both decreased. For example, when glycosyl donor α -**100** was glycosidated with acceptor **36** disaccharide **101** was produced in 68% yield with reduced stereoselectivity ($\alpha/\beta = 7.5:1$). This reduced yield was attributed to competing silyl group cleavage occurring under these reaction conditions.

Madsen and co-workers used protic acid in combination with N-iodosuccinimide to achieve a facile release of the iodonium ion.²⁶¹ An efficient glycosidation of per-benzoylated glucosyl bromide α -**11** with reactive glycosyl acceptor **36** was achieved in the presence of NIS/TfOH to afford disaccharide **102** in yields up to 88% (Scheme 27). However, changing the sugar series and protecting groups of glycosyl donors led to reduced yields (60–80%) even in reactions with reactive glycosyl acceptors.

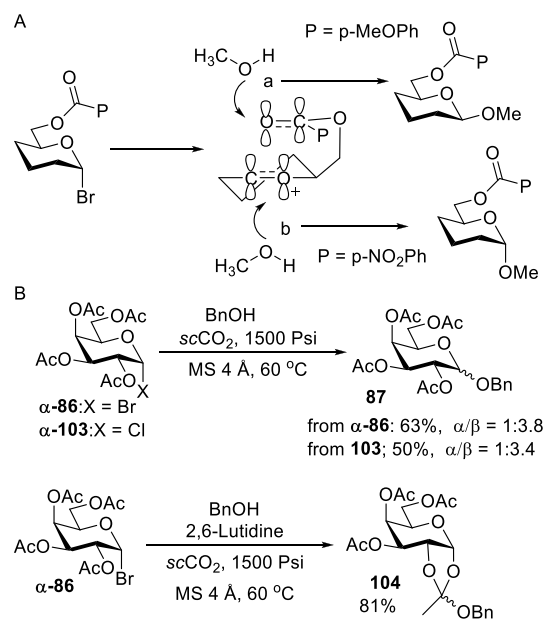
Scheme 27. NIS-TfOH-Promoted Glycosidation of Glycosyl Bromides



3.2.6. Solvolysis. Methanolysis of glycosyl bromides was studied both in the presence and the absence of various additives such as tetrabutylammonium bromide²⁶⁷ or silver salts.²³⁰ To understand the mechanistic details of solvolysis reactions, several experiments were conducted and it was determined that the concentration of alcohol plays a significant role on reaction rates and stereoselectivity.²³⁰ At lower concentrations of methanol, less reactive α -glycosyl bromides are in anomeric equilibrium with the more reactive β -counterpart, which subsequently gives methyl α -glycoside as the major product.^{230,267} At high concentrations of methanol, α -glycosyl bromides directly react with MeOH to preferentially produce methyl β -glycoside. A study by Frechet and Schuerch revealed variations in the rate of methanolysis or anomeriza-

tion caused by steric and/or electronic effects of substituents at C-6 of glycosyl donors.²⁶⁸ The study concluded that the anomeric distribution of methyl glycosides from methanolysis (solvolysis) of glycosyl bromides strongly depends on various factors such as structure of glycosyl halides, concentration of methanol, and reaction conditions. Several glycosyl bromides having different acyl groups at C-6 have been investigated revealing useful trends.²⁶⁸ By changing the electronics of the functional groups at C-6, methyl glycosides were formed from 90% of α -anomer to over 90% of its β -counterpart. It was proposed that it is the interaction of the oxacarbenium ion with the carbonyl group of the substituent at C-6 that plays a significant role in deciding the stereochemistry of the product. For the instance, electron rich *p*-methoxybenzoyl group increases the electron density on the carbonyl group, stabilizes the positive charge on C-1, and facilitates the top face attack of the glycosyl acceptor (pathway a, Scheme 28A). The hydrogen

Scheme 28. Examples of Alcoholysis Reactions



bonding with acceptor could also be possible due to electron-rich carbonyl oxygen at C-6, which would also favor the formation of the β -linked product. Conversely, the *p*-nitrobenzoyl group with the electron deficient carbonyl group is unable to stabilize the positive charge on C-1. As a result, the attack of the glycosyl acceptor will take place from the less shielded bottom face resulting in the corresponding α -glycoside. The anomeric distribution of methyl glycosides produced from glycosyl bromides equipped with differentially *p*-substituted 6-O-benzoates was shown to follow the σ Hammett substituent constant value. Frechet and Schuerch extended the alcoholysis (methanolysis) of glycosyl bromide to solid phase synthesis wherein glycosyl bromides were coupled with polystyrene resin-immobilized glycosyl acceptors.^{324,325}

Recently, Csuk and co-workers synthesized several β -glycosides by treating acetylated glycopyranosyl bromides with excess of *n*-alkyl alcohol without any additives.³²⁶ Glycosides were produced on gram scale in good to moderate yields in 4–7 days at room temperature. Matheu and co-workers discovered a green approach to construct glycosidic linkages.³²⁷ For that, glycosylation was conducted employing

glycosyl halide donors in super critical CO₂ without any volatile organic solvent and metal catalysts. The authors demonstrated an efficient activation of tetra-acetylated galactosyl bromide **α-86** with benzyl alcohol **73** in the presence of scCO₂ at 1500 Psi and 60 °C to afford galactoside **87** in 63% yield ($\alpha/\beta = 1:3.8$, Scheme 28B). To broaden the scope, differently protected galactosyl bromide donors in combination with various glycosyl acceptors were tested to afford the corresponding galactosides in moderate to good yields. Interestingly, pivaloyl-protected galactosyl bromide donor produced far better results in terms of yields and stereoselectivity in comparison to those achieved with its acetyl-, benzoyl-, or benzyl-protected counterparts. Reactions performed in the presence of 2,6-lutidine led to the formation of orthoester **104** in a good yield (81%, Scheme 28B). Glycosyl chlorides were investigated alongside providing comparable results; however, the activation of glycosyl chloride turned out to be slower and required high temperatures (90 °C).

3.3. Glycosyl Bromides in Glycan and Glycoconjugate Synthesis

3.3.1. Linear Synthesis. After understanding the mechanistic details for the formation of side products, strategies were developed for the synthesis of higher oligosaccharides. As depicted in Figure 3A, linear oligosaccharide synthesis consists of the glycosylation of a monosaccharide donor with a monosaccharide acceptor in the presence of a promoter to give the desired disaccharide. Most commonly, the latter is then converted into the second-generation glycosyl acceptor via liberation a specific hydroxyl group. The acceptor is then allowed to react with a glycosyl donor, resulting in the

formation of trisaccharide, and this deprotection–glycosylation sequence can be then reiterated to yield a glycan of the desired length.

Alternatively, the intermediate oligosaccharides can be converted into a glycosyl donor via the introduction of a suitable leaving group. Many glycans have been synthesized using linear approaches; the only approach that was known in early days of oligosaccharide synthesis. Successful stepwise linear syntheses of gentiotriose **105** and gentiotetraose **106** were successfully demonstrated employing the Koenigs–Knorr glycosylation method in the presence of silver salt, iodine, and Drierite (Figure 3B).^{220,228} Following the same strategy, Takiura et al. synthesized gentio-oligosaccharides **105**–**108**.³²⁸ The halide ion-catalyzed glycosylation method was applied to the synthesis of blood group determinants **109**–**111** containing challenging α -glycosidic linkages (Figure 3C).^{259,319}

3.3.2. Convergent Building Block Assembly. Pioneering studies by Zen,³²⁹ Paulsen,³³⁰ and Ogawa³³¹ paved the way to what is currently known as a convergent building-block strategy for oligosaccharide synthesis. According to this strategy, two di- (or oligo) saccharides, a glycosyl donor and a glycosyl acceptor, are constructed individually and then coupled together (converged) to afford a larger oligosaccharide. One benefit of such block synthesis is the decreased number of steps required for the overall assembly. One relevant example is a 2+3+2 assembly of a biantennary heptasaccharide **117** as depicted in Scheme 29.³³² According to the developed strategy, lactosamine bromide donor **112** was glycosidated with trimannosyl acceptor **113** in the presence of AgOTf and 2,6-lutidine to produce pentasaccharide **114** in

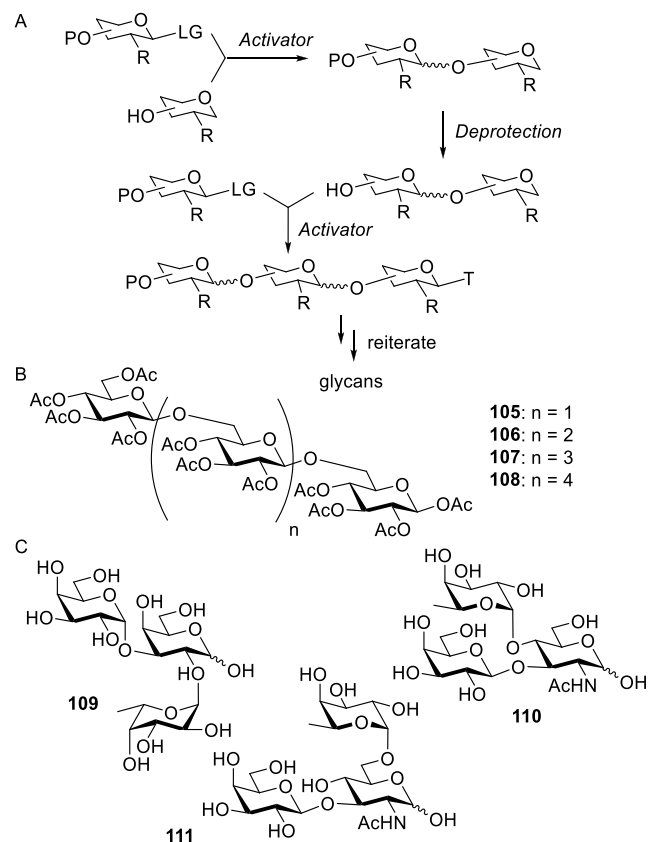
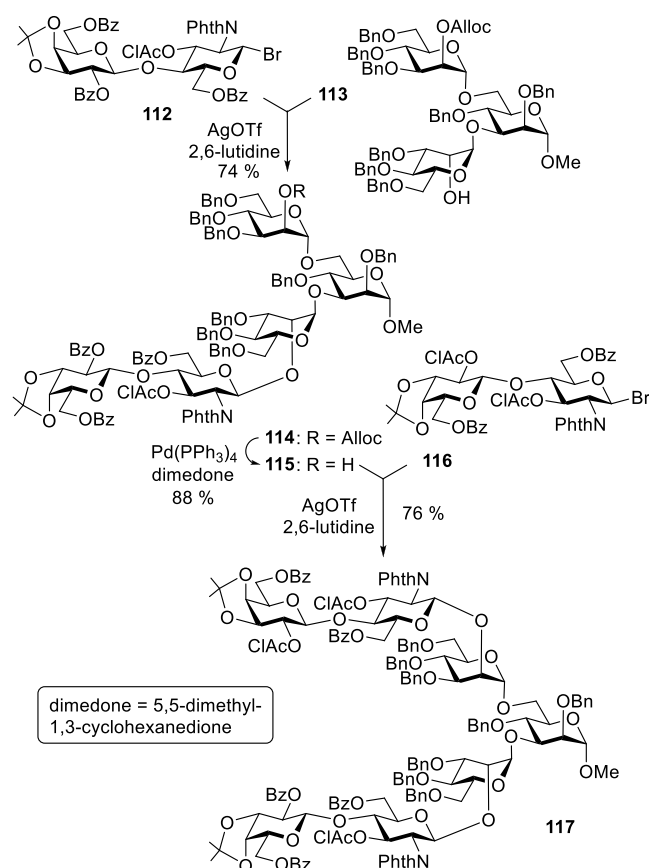


Figure 3. Linear oligosaccharides syntheses using Koenigs–Knorr (A) and halide-catalyzed (B) approaches

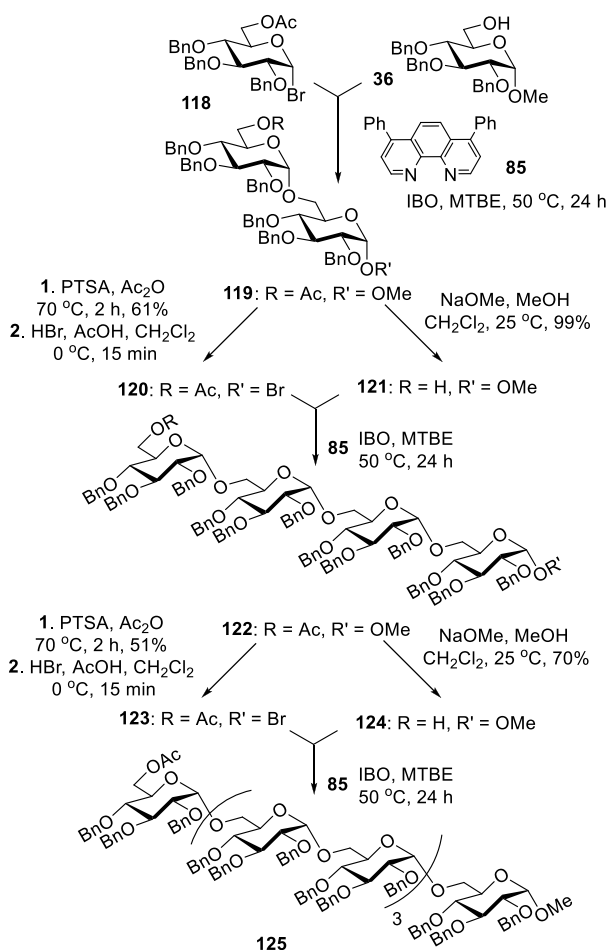
Scheme 29. Convergent Synthesis of Heptasaccharide 117



71% yield. The latter was subjected to the allyloxycarbonyl (Alloc) group removal with $\text{Pd}(\text{PPh}_3)_4$ to provide glycosyl acceptor **115** in 88% yield, which was then glycosylated with glycosyl donor **116** in the presence of AgOTf and 2,6-lutidine to produce the heptasaccharide **117** in 76% yield.

A conceptually related convergent approach has been proven particularly advantageous if the synthesis of two or more repeating units is required. In this case, an intermediate di- (or oligo) saccharide is divided into two portions: one portion is converted into a glycosyl donor and another into a glycosyl acceptor. Subsequently, the two reactants are coupled to afford a dimer. This manipulation can be then repeated for the synthesis of larger structures. A relevant recent example of such an approach was described by Nguyen and co-workers. The authors employed their phenanthroline-catalyzed glycosylation (*vide supra*) to assemble 1,2-*cis*-linked oligosaccharides as shown in Scheme 30.

Scheme 30. Phenanthroline-Catalyzed Synthesis of 1,2-*cis* Glycans

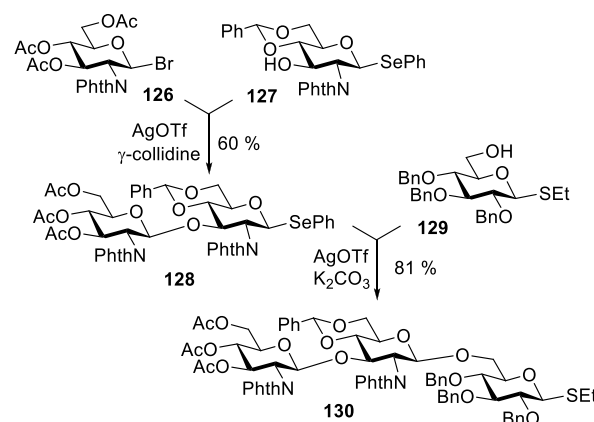


Glycosyl bromide **118** was glycosidated with glycosyl acceptor **29** in the presence of 5% of phenanthroline **85** to obtain disaccharide **119** in 89% yield ($\alpha/\beta > 20:1$). The latter was converted to glycosyl bromide donor **120** via sequential acetolysis-bromination. Intermediate **119** was also converted to glycosyl acceptor **121** by removing acetyl in the presence of sodium methoxide in methanol. Further, glycosyl bromide donor **120** was glycosidated with glycosyl acceptor **121** in the presence of phenanthroline **85** to obtain tetrasaccharide **122** in

86% yield ($\alpha/\beta > 20:1$). The latter was converted into tetrasaccharide bromide donor **123** and tetrasaccharide acceptor **124**, and the resulting counterparts were coupled in the presence of phenanthroline **85** to produce octasaccharide **125** in a good yield and excellent stereocontrol (77%, $\alpha/\beta > 20:1$).

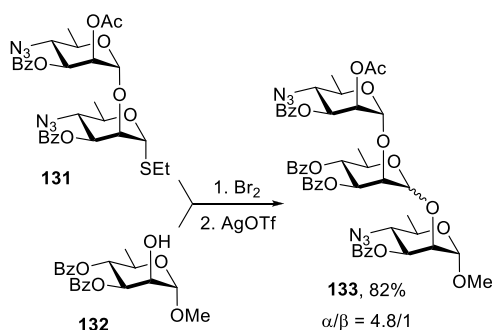
3.3.3. Selective Activation. When the arsenal of the glycosylation techniques was limited to Fischer and Koenigs–Knorr approaches (or their variations), selective activation of one leaving group over another was rare. A few relevant examples wherein more reactive glycosyl bromides were activated over less reactive glycosyl halides (chlorides, fluorides) that could act as glycosyl acceptors were discussed in previous subsections. However, when stable glycosyl donors such as thio- or selenoglycosides have emerged, the ability to activate one leaving group over another dramatically enhanced. Pioneering research of Zen,³²⁹ Nicolaou,^{333,334} Lonn,^{335,336} Garegg,^{216,337} and others^{338,339} involved the activation of alkyl halides over S-alkyl/aryl glycosides. In addition to a few relevant examples discussed previously, an example of such a synthesis is presented in Scheme 31.³⁴⁰ First, glycosyl bromide

Scheme 31. Selective Activation Utilizing Three Leaving Groups



donor **126** was activated over the phenylselenide acceptor **127** in the presence of AgOTf/γ -collidine to give the desired disaccharide **128** in 60% yield. The latter was then directly activated over the SET acceptor **129** in the presence of $\text{AgOTf}/\text{K}_2\text{CO}_3$ to produce the desired trisaccharide **130** in 81% yield. Apparently, since the latter is equipped with the anomeric SET leaving group, it can be directly activated for subsequent glycosylations under appropriate reaction conditions. A similar activation sequence was reported by the Ley group.³⁴¹

3.3.4. Two-Step Activation and Regenerative Glycosylation Concept. Two-step activation of thioglycosides with bromine via glycosyl bromides has several potential applications for oligosaccharide synthesis. It can range from simple glycosylation to sophisticated iterative approaches. As shown in Scheme 32, the initial step involves the treatment of thioglycoside precursor **131** with bromine. The resulting glycosyl bromide is then directly reacted with glycosyl acceptor **132** in the presence of silver triflate to afford trisaccharide **133**.²³¹ Since the acceptor is not carrying a leaving group, it can be present in the reaction mixture from the beginning. The two-stage activation approach can also be applied to situations when both glycosyl donor and glycosyl acceptor initially bear the same type of a leaving group (thioglycoside).³²⁹ In this

Scheme 32. In Situ Activation of Thioglycosides via Bromides

case, thioglycoside precursor of the donor is first converted to glycosyl bromide. The acceptor equipped with the SR leaving group is then added, and the glycosyl bromide is selectively activated in the presence of a suitable activator. Since the resulting product is a thioglycoside, this two-step activation sequence can be reiterated.

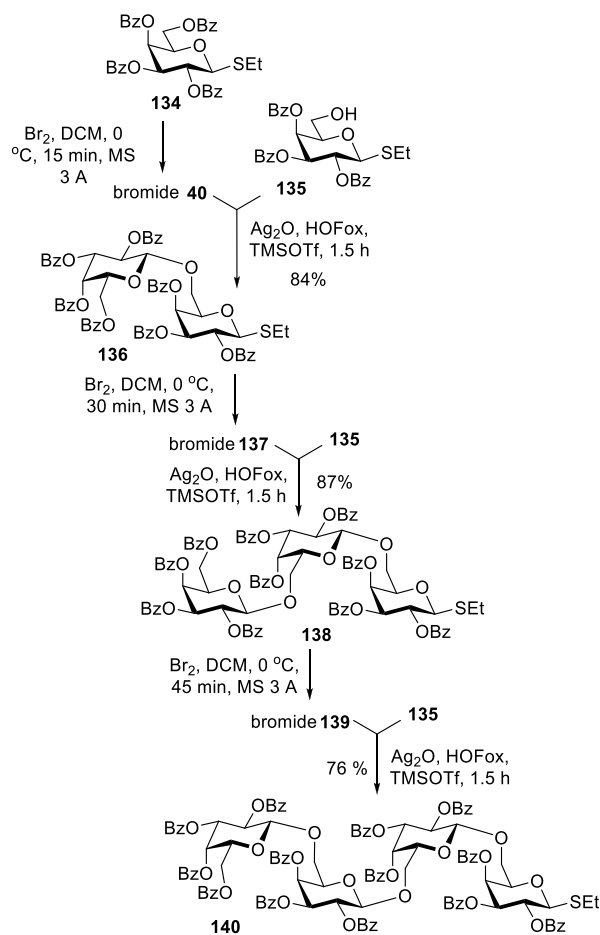
Demchenko and co-workers have developed a regenerative strategy to activate glycosyl bromides (*vide supra*).^{217,225} In accordance with this approach, a thioglycosides precursor is first treated with bromine to afford the corresponding glycosyl bromide. The latter is then activated with HOFox/TMSOTf for the regenerative glycosylation cycle wherein both the synthesis of the reactive OFox imidate intermediate and its glycosidation is performed in a catalytic, regenerative fashion. Further, the regenerative glycosylation was applied to oligosaccharide synthesis. For this purpose, per-benzoylated galactosyl bromide **40**, generated from the corresponding thiogalactoside **134**, was glycosidated with thiogalactoside acceptor **135** in the presence of Ag₂O, HOFox, and TMSOTf, as depicted in Scheme 33. As a result, disaccharide **136** was obtained in 84% yield. Subsequently, thioglycoside **136** was converted to bromide donor **137** and coupled with acceptor **135** under the regenerative reaction conditions to produce trisaccharide **138** in 87% yield. Finally, the latter was converted into bromide **139**, which was then coupled with acceptor **135** to afford tetrasaccharide **140** in 76% yield.

4. GLYCOSYL CHLORIDES

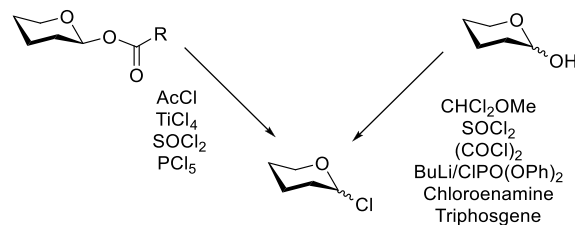
First glycosylations performed by Michael (*vide supra*) involved glycosyl chlorides.⁴ However, in subsequent years, glycosyl chlorides were largely overshadowed by glycosyl bromides that have traditionally been considered advantageous over their chloride counterparts. After the thorough discussion of glycosyl bromides (Section 3), we note that many methods developed for the activation of glycosyl bromides are also effective for the activation of glycosyl chlorides. Hence, this section will mainly focus on the reagent-specific or condition-specific synthesis, activation, and application of glycosyl chlorides rather than repeating the same data presented for glycosyl bromides.

4.1. Synthesis of Glycosyl Chlorides

For the first time, glycosyl chlorides were synthesized from free glucose by treatment with acetyl chloride by Colley in 1870.³⁴² Expanding on this early work, others have also used AcCl for the synthesis of glycosyl chlorides of other sugar series.^{343,344} Subsequently, more efficient methods, surveyed in this Section, have been developed for the synthesis of glycosyl chlorides from a variety of precursors.

Scheme 33. Oligosaccharide Synthesis via Regenerative Glycosylation

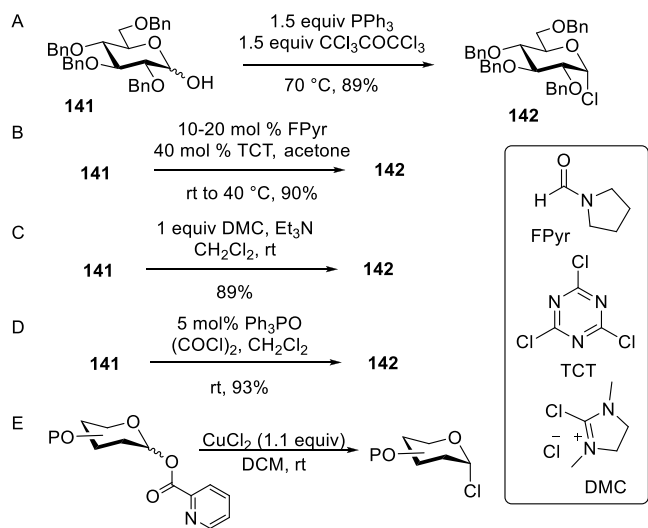
4.1.1. Preparation from Hemiacetals or Glycosyl Esters. Most commonly, glycosyl chlorides are prepared from two anomeric groups, either an ester such as acetate or a hemiacetal (Scheme 34). Selected reagents suitable for

Scheme 34. Synthesis of Glycosyl Chlorides from Ethers or Hemiacetals

converting anomeric esters into glycosyl chlorides include TiCl₄,³⁴⁵ SOCl₂,³⁴⁶ and PCl₅.³⁴⁷ Representative reagents used in the synthesis of chlorides from hemiacetals include CHCl₂OMe,^{348,349} SOCl₂,³⁵⁰ oxalyl chloride,^{351–353} *n*-BuLi and ClPO(OPh)₂,³⁵⁴ chloroamine,³⁵⁵ and triphosgene.³⁵⁶ Evidently, many if not all of these reactions use harsh conditions and/or toxic reagents. Finding new methods to avoid some of these harsh conditions, toxic, or heavy metal reagents has been a vibrant area of study in recent years.

In 2017, Iadonisi developed a solvent-free method for the synthesis of a variety of glycosyl chlorides (Scheme 35A).³⁵⁷

Scheme 35. New Methods for the Synthesis of Glycosyl Chlorides



Using triphenyl phosphine and hexachloroacetone at 70 °C, glycosyl chloride **142** was obtained from the hemiacetal precursor **141** in 45 min. These conditions were applied to various sugar series including mannose, galactose, and fucose giving high yields (80%+) in most cases. This method was less effective when applied to nitrogen-containing sugar series. For example, the glycosyl chloride from glucosamine was obtained in only 44% yield.

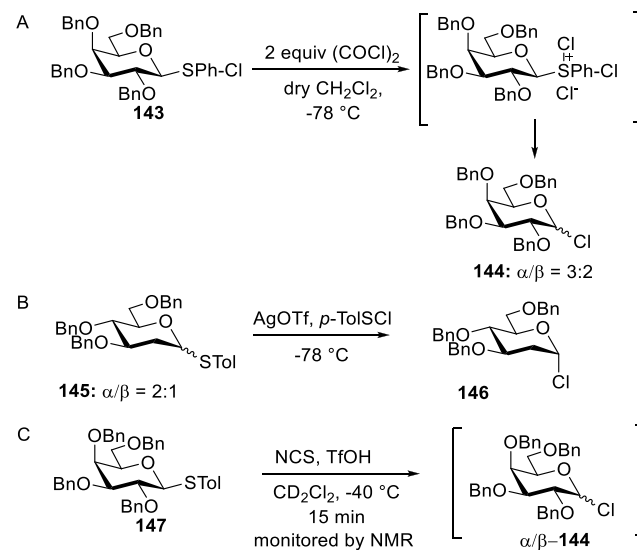
Huy and Filbrich³⁵⁸ have recently developed a method for the synthesis of glycosyl chlorides from the corresponding hemiacetal derivatives using as little as 34 mol % of trichlorotriazine (TCT) as a source of the stoichiometric chlorine. Thus, reaction of hemiacetal **141** with TCT in the presence of 10–20 mol % of *N*-formylpyrrolidine (FPyr) at 40 °C produced glycosyl chloride **142** in 90% yield (Scheme 35B). These conditions were shown to work both with sugar substrates such as glucose and fructose and with aliphatic alcohols. In 2018, Judeh et al.³⁵⁹ introduced a chlorinating reagent 2-chloro-1,3-dimethylimidazolinium chloride (DMC) that was applied to the synthesis of glycosyl chlorides. Using stoichiometric DMC in the presence of triethylamine, hemiacetal **141** was converted into the corresponding glycosyl halide **142** in 15–30 min in 89% yield (Scheme 35C). This reaction worked well for a variety of sugars (glucose, mannose, galactose) giving 80–95% yield in most cases. The developed conditions were found to be compatible with many commonly used protecting groups such as acetates, silyl ethers, and acetals. McGarrigle et al.³⁶⁰ found that catalytic Appel conditions using 5 mol % Ph_3PO and 1.5 equiv $(\text{COCl})_2$ were also capable of chlorinating hemiacetals (Scheme 35D). Using these conditions, chloride **142** was synthesized from hemiacetal **141** in 93% yield. While this protocol worked well for glucose, most other sugars such as mannose, galactose, and 2-deoxy glucose gave lower yields between 67 and 79%. Glucosamine and galactosamine derivatives performed the worst under these reaction conditions, giving mixtures of products.

Recently, Tang and co-workers developed a milder reaction conditions for the synthesis of glycosyl chloride donors.²¹³ In accordance with their approach, 1-*O*-picoloyl (Pico) derivatives were treated with copper(II) chloride in DCM at ambient

temperature (Scheme 35E). An interesting insight of this method is that 1-*O*-picoloyl (Pico) derivatives exclusively produced the corresponding α -glycosyl chlorides, and the method was found to be compatible with many commonly used protecting groups.

4.1.2. Preparation from Thioglycosides. Thioglycosides have many advantages such as being stable, allowing for protecting group manipulations, and can be stored for long periods of time. Hence, most modern building block syntheses today involve thioglycosides. Conversion from thioglycosides to glycosyl chlorides typically involved a two-step protocol with the first step being hydrolysis of the anomeric thioglycoside to the hemiacetal. This represented a notable disadvantage, and the effort has been made to develop methods for the direct conversion of thioglycosides to glycosyl chlorides. The first method was reported by Sugiyama and Diakur,³⁶¹ wherein 4-chlorophenylthio glycoside **143** was treated with oxalyl chloride in dichloromethane. The reaction was found to proceed through the intermediacy of a glycosyl halosulfonium salt shown in Scheme 36A. The latter is

Scheme 36. Direct Synthesis of Glycosyl Chlorides from Thioglycosides



unstable and falls apart to form the corresponding glycosyl chloride **144**. The resulting glycosyl chlorides were isolated in high yields (90%+) or crude chlorides could be used for subsequent glycosylations directly. While this protocol allowed for the one-step conversion, it still used harsh reaction conditions and the scope of this reaction was limited to 4-chlorophenylthio glycosides.

Verma and Wang³⁶² employed stoichiometric *p*-TolSCl in the presence of catalytic AgOTf to successfully convert *p*-tolylthio (STol) 2-deoxyglycoside **145** into the corresponding chloride **146** (Scheme 36B). Following this conversion, the corresponding chloride could be used for glycosylation without further purification. Subsequently, the authors discovered that the formation of anomeric chlorides can be accompanied by anomeric triflates, and the product distribution was dependent on the relative reactivity value (RRV)¹³⁵ of the tolylthio glycoside precursor. For example, when armed tolylthio galactoside **147** (RRV = 17 000) was treated with *N*-chlorosuccinimide NCS (1.0 equiv) and TfOH (0.3 equiv) in CD_2Cl_2 at -40 °C, exclusive formation of anomeric chloride

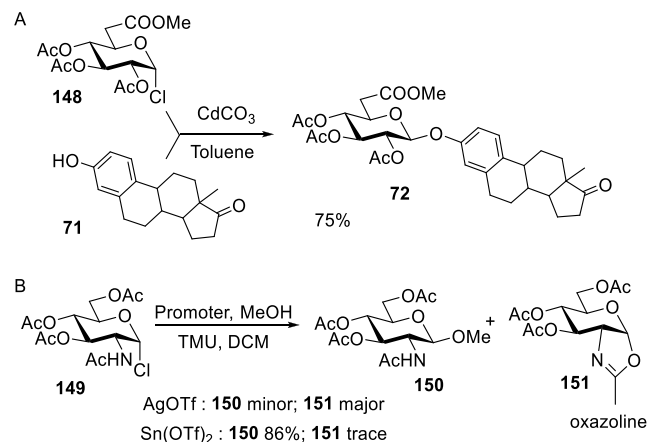
144 was observed by means of the NMR spectroscopy monitoring (Scheme 36C).³⁶³ Regardless of the reaction intermediates, tolylthio glycosides could be glycosidated in the presence of a suitable glycosyl acceptor, NCS, and triflic acid. Reactions proceeding via the intermediacy of glycosyl chlorides were highly α -stereoselective.

4.2. Glycosidation of Glycosyl Chlorides

Michael in 1879 was the first to perform a glycosylation reaction with glycosyl chlorides (*vide supra*).⁴ Per-acetylated glycosyl chloride was reacted with potassium phenoxide giving phenyl glucoside as the product. Then in 1901, the activation of glycosyl chlorides was performed by Koenigs and Knorr.⁷ These reactions used insoluble silver(I) salts such as silver oxide or silver carbonate which were thought to act as an acid scavenger. Little was done to improve the activation of chlorides until 1949 when Helferich³⁶⁴ introduced mercury salts as active promoters. While these methods can be used to synthesize oligosaccharides,^{365,366} mercury salts are very toxic and the environmental considerations of the 21st century called for further quest of better activation conditions. Nevertheless, both the Koenigs-Knorr method and the Helferich modification have found broad utility in the synthesis of simple glycosides to date.

Some early work included the activation of glycosyl chlorides with cadmium(II) salts.²⁴⁹ For example, glycosidation of glycosyl uronide donor 148 with estrone acceptor 71 in the presence of CdCO₃ produced glucuronide 72 in 71% yield (Scheme 37A). Cadmium sulfide was also investigated as the

Scheme 37. Cadmium or Tin Salt-Promoted Glycosidations of Glycosyl Chlorides



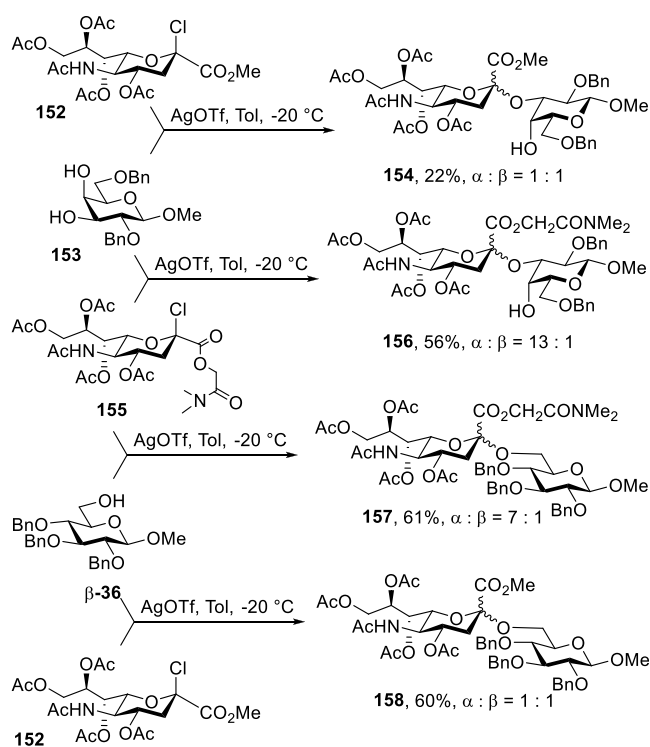
activator for the glycosidation of the anomeric chloride; however, the yield of the glucuronide was below the preparative value. Several other steroidal phenolic glucuronides, glycosides, and acetyl glucosaminides were successfully synthesized.^{250,302,303} In addition to β -glycosides, a small amount of the corresponding α -glucosides and/or C-glycosides were also isolated, and the ratio of reagents was found to depend on coupling partners and reaction conditions employed. An interesting aspect of these reactions was the development of color as the reaction progresses. It was suggested that the generated cadmium chloride was the actual catalyst of activation. However, cadmium chloride by itself was ineffective, similar to Helferich's explanation for mercuric cyanide activation.²³⁹ As discussed in Section 3.2.2, the

product composition also in this case depends on the surface area of cadmium carbonate.²⁵¹

In another study, the authors successfully applied the stannous triflate-promoted glycosylation for the activation of glycosyl chloride donors.³⁰⁶ When glycosyl chloride 149 bearing a 2-acetamido (NHAc) group was employed as glycosyl donor in the presence of AgOTf, oxazoline 151 was observed as a major product (Scheme 37B). Changing AgOTf with Sn(OTf)₂ under otherwise identical reaction conditions suppressed the formation of oxazoline 151 while producing glycoside 150 as the major product. The control experiments showed that oxazoline does not serve as a reaction intermediate under these reaction conditions.

4.2.1. Activation with Silver Salts. Being partially soluble, silver(I) triflate is typically an effective promoter for the activation of glycosyl chlorides. However, these conditions were somewhat ineffective in cases of relatively unreactive glycosyl donors such as sialic acid chlorides.^{367,368} Thus, glycosidation of chloride 152 with glycosyl acceptor 153 in the presence of AgOTf produced glycoside 154 in a poor yield of 22% without any preferred stereoselectivity (Scheme 38).

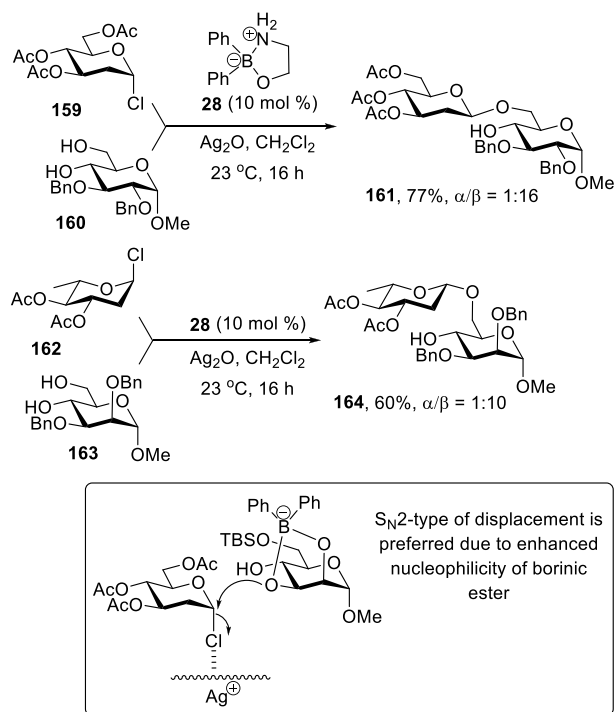
Scheme 38. Silver Triflate-Promoted Activation of Sialic Acid Chlorides



While investigating a participating moiety at C-1, Gin and co-workers introduced sialyl chloride donor 155 bearing an *N,N*-dimethylglycolamido ester functionality.³⁶⁹ A commendable result was obtained with galactosyl acceptor 153 for the formation of disaccharide 156 in 56% yield and with high α -stereoselectivity $\alpha/\beta = 13/1$. Glycosylation of the primary glycosyl acceptor β -36 was nearly as effective with both sialyl donors 152 and 155 in terms of the yields of the respective products 158 and 157 that were isolated in 60–61% yields. However, the activation of the standard sialyl chloride donor 152 was nonstereoselective, and disaccharide 158 was isolated as a racemic mixture.

As described in Section 3.2.1, Taylor and co-workers developed borinic acid-catalyzed regioselective activation of glycosyl bromides.^{224,272} A similar approach was later applied by the authors to the β -selective synthesis of 2-deoxy and 2,6-dideoxyglycosides.³⁷⁰ Anomeric chlorides as glycosyl donors and partially protected cis-1,2- and 1,3-diols as glycosyl acceptors were found to be very effective. As shown in Scheme 39, per-acetylated 2-deoxyglycosyl chloride 159,

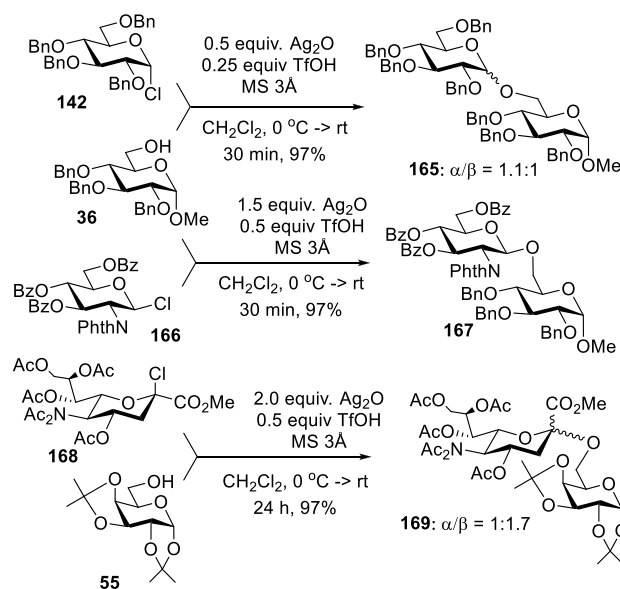
Scheme 39. Organoboron-Catalyzed Glycosidation of 2-Deoxy and 2,6-Dideoxy Glycosyl Chlorides



synthesized by treating with BCl_3 , was reacted with glycosyl acceptor 160 in the presence of aminoethyl diphenylborinate 28 and silver oxide to produce 2-deoxy disaccharide 161 in 77% yield with high β -selectivity ($\alpha/\beta = 1:16$). The authors also synthesized challenging β -2,6-dideoxy glycosides commonly found in bioactive natural products. For example, reaction of 2-deoxy-L-rhamnopyranosyl chloride 162 with glycosyl acceptor 163 under similar reaction conditions produced disaccharide 164 in a good yield of 60% with high β -selectivity ($\alpha/\beta = 1:10$). Changing the protecting group from OAc to OBn turned out to be detrimental for both β -selectivity and yields. Similarly to that discussed in Section 3.2.1, the enhanced nucleophilicity of the hydroxyl group, as predicted by Fukui index calculation, in the borinate complex resulted in an $\text{S}_{\text{N}}2$ -like attack on the glycosyl chloride donor.

Following the initial success with catalytic glycosidation of glycosyl chlorides, the Demchenko group investigated acid-catalyzed silver(I) oxide-mediated glycosylations that were initially developed for glycosyl bromides (*vide supra*).²²¹ Using Ag_2O combined with TfOH allows one to drastically increase the glycosylation reaction rate while reducing the amount of silver needed to 0.50 equiv. This reaction worked well with benzylated glucosyl chloride 142 and disaccharide 165 was obtained in 97% yield in the presence of just 0.50 equiv of Ag_2O and 0.25 equiv of TfOH in 30 min (Scheme 40).³⁷¹ Glycosylations of secondary glycosyl acceptors provided

Scheme 40. Glycosyl Chloride Activation Using Ag_2O and TfOH Promoter System



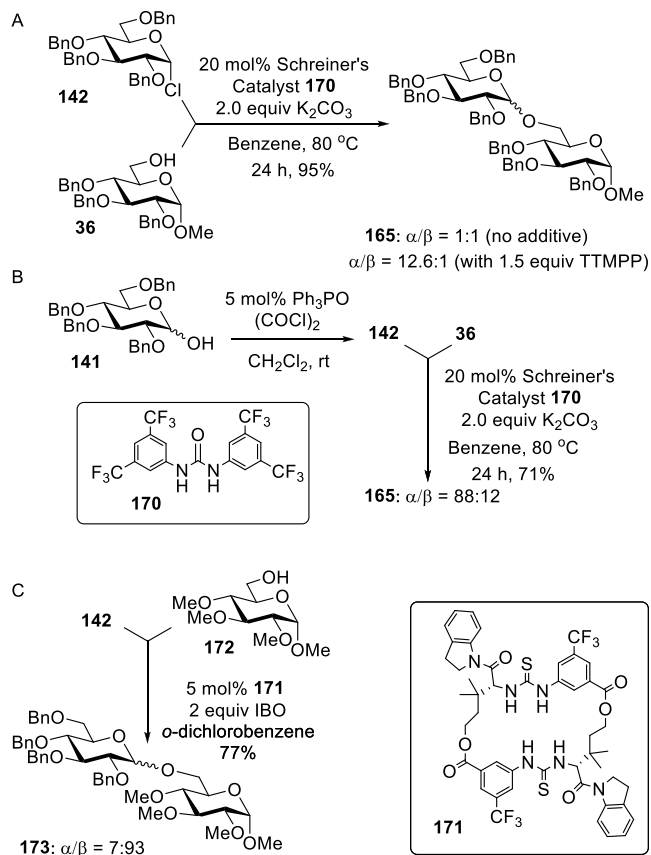
similar results. Benzylated mannosyl and galactosyl chlorides required a higher amount of TfOH (0.50 equiv). These reaction conditions provided high yields (98%+) and fast reaction times.

Nitrogen-containing sugar derivatives such as glucosamine and sialic acid could also be readily activated under these reaction conditions, but required higher amounts of activators. Thus, primary glycosyl acceptor 36 was coupled with phthalimide-protected glycosyl chloride donor 166 in the presence of 1.5 equiv of Ag_2O and 0.5 equiv of TfOH afforded disaccharide 167 in 97% yield. However, upon switching to less reactive secondary acceptors the yields dropped to 68–76%. Sialic acid chloride 168 was glycosidated with the primary glycosyl acceptor 55 allowing disaccharide 169 in an excellent yield of 97% yield, albeit with poor stereoselectivity.

4.2.2. Organocatalytic Activation. These classical methods were state-of-the-art until recently, when new methods for the activation of glycosyl chlorides have emerged. The first new method was introduced by the Ye group in 2016.¹⁴⁵ Using a variety of benzylated glycosyl chlorides, 20 mol % of Schreiner's catalyst 170 and 2.0 equiv K_2CO_3 in benzene at 80 °C the respective disaccharides were produced in high yields (80%+). At first, these reactions were rather slow (24 h), and the stereoselectivity was poor. Thus, glycosylation between glycosyl chloride donor 142 and primary acceptor 36 gave disaccharide 165 in 95% yield as an anomeric mixture ($\alpha/\beta = 1:1$, Scheme 41A). When the same reaction was carried out in the presence of 1.5 equiv of tris(2,4,6-trimethoxyphenyl)-phosphine (TTMPP) as an additive, the anomeric stereoselectivity could be improved to a commendable $\alpha/\beta = 12.6:1$. The improvement of stereoselectivity with TTMPP was seen throughout a series of glycosyl acceptors. On the basis of the NMR data, the authors theorized that TTMPP noncovalently interacted with the anomeric carbon from the β -face. This interaction directed the acceptor attack from the opposite face giving rise to α -linked glycosides.

In 2019, McGarrigle³⁶⁰ applied the Appel conditions (*vide supra*) to the synthesis of glycosyl chlorides followed by their glycosidation in one pot. The treatment of hemiacetal 141 with

Scheme 41. Glycosyl Chloride Activation Using Thioureas

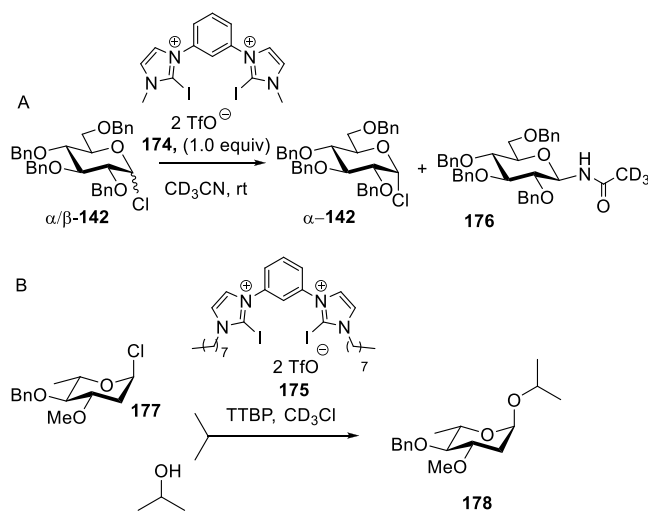


Ph_3PO and oxalyl chloride in dichloromethane produced glycosyl chloride **142**. Then glycosyl acceptor **36** was added *in situ* along with 20 mol % of Schreiner's catalyst **170**, 2.2 equiv of K_2CO_3 , and 20 mol % of TTMP (Ye's conditions, Scheme 41B). This one-pot approach led to a decrease in yield of disaccharide **165** that was obtained in 71% yield (vs 95% reported by Ye) with an anomeric ratio of $\alpha/\beta = 88:12$.

The Jacobsen group developed thiourea catalyst **171**,⁷⁰ which cooperatively activates both the glycosyl chloride donor and the glycosyl acceptor. The glycosyl chloride donor hydrogen bonds with the thiourea portion of catalyst **171**, which enhances its leaving group ability. At the same time, the incoming nucleophile is also activated by the catalyst via Lewis basic interactions with the carbonyl oxygen of the amide of the catalyst. The combination of these two activations by the catalyst leads to an S_N2 -like displacement. These reactions were conducted in the presence of 5 mol % of **171**, 2 equiv of isobutylene oxide (IBO), which acts as an electrophilic trap for the released HCl, in *o*-dichlorobenzene. Reacting donor **142** with acceptor **172** using the conditions above gave disaccharide **173** in 77% yield ($\alpha/\beta = 7:93$, Scheme 41C). Other sugars showed similar yields and stereoselectivity. To prove that these glycosylations undergo an S_N2 -like displacement, the authors also studied the glycosyl chloride configuration at the anomeric center. It was determined that an α -chloride leaving group gave primarily the β -linked product whereas a β -chloride gave predominantly the α -linked product. This trend was seen throughout a series of glycosyl chlorides, showing that these reactions follow an S_N2 -like displacement of the leaving group, without the formation of an oxacarbenium ion intermediate.

Codee and co-workers developed a halogen bond-mediated activation method for glycosyl chlorides. For that purpose iodoimidazolium compounds **174** and **175** was employed as halogen-bond donors.³⁷² NMR experiments were performed to establish the activation of 2,3,4,6-tetra-*O*-benzyl α,β -glucosyl chloride **142** that was treated with bis(iodoimidazolium) compound **174** (1.0 equiv) in CD_3CN in an NMR tube (Scheme 42A). Based on NMR signals and LC-MS data it was

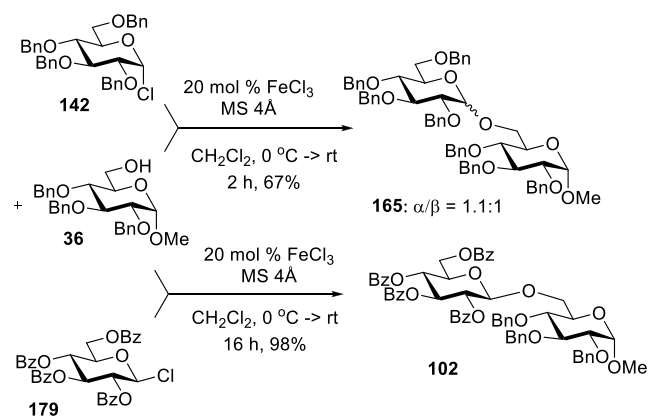
Scheme 42. Halogen Bond-Mediated Activation of Glycosyl Chloride



suggested that more reactive β -glycosyl chloride was completely consumed and furnished anomeric acetamide **176** by a Ritter type process³⁷³ while the α -anomer did not react. Later, it was realized that the stable α -anomer requires longer duration (several weeks) for activation under these reaction conditions. Several control experiments were performed to confirm that halogen-bond activation was essential for activating the anomeric C–Cl bond. In a separate NMR experiment, it was shown that glucosyl bromide is more reactive than corresponding chloride however, its use is restricted by lower stability and spontaneous decomposition. To explore the ability of the dicationic halogen-bond donors to promote the synthesis of *O*-glycosidic linkages, several attempts were made. When the reactive *L*-oleandrosyl chloride **177** was treated with isopropyl alcohol in the presence of a lipophilic halogen bond activator **175** and 2,4,6-tri-*tert*-butyl pyrimidine as an acid scavenger a satisfactory conversion to corresponding glycoside **178** was achieved as shown in Scheme 42B. This method was largely limited to highly reactive glycosyl chlorides.

4.2.3. Catalytic Activation with Iron, Bismuth, or Palladium Salts. In 2018, Geringer and Demchenko introduced the first catalytic glycosidation of glycosyl chlorides.³⁷⁴ Thus, benzylated glucosyl chlorides could be readily glycosidated in the presence of only 20 mol % of iron(III) chloride ($FeCl_3$) and molecular sieves (4 Å) in dichloromethane. These glycosylations led to the corresponding disaccharides in moderate to good yields (47–80%). For example, glycosidation of chloride **142** with primary acceptor **36** afforded disaccharide **165** in 67% yield ($\alpha/\beta = 1.1:1$, Scheme 43). Somewhat modest yields in these glycosylations were partially attributed to the formation of an undesirable side product, 1,6-anhydro-2,3,4-tri-*O*-benzyl- β -D-glucopyranose.

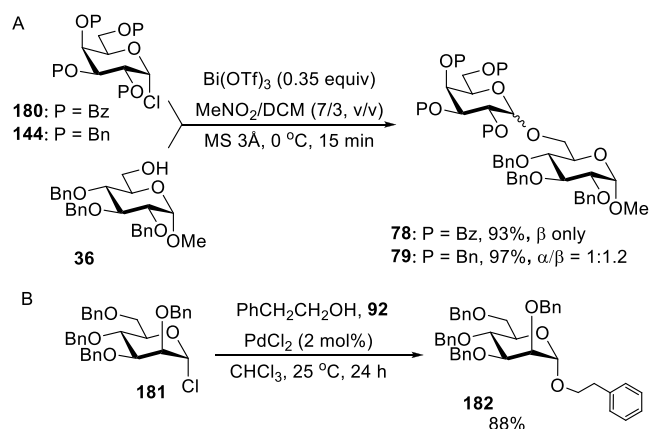
Scheme 43. Glycosyl Chloride Activation Using Catalytic FeCl_3



Notably, switching to benzoylated glucosyl chlorides or either mannosyl or galactosyl chlorides, resulted in an increase in yields to 52–98% depending on the acceptor. For example, the reaction of benzoylated chloride 179 with acceptor 36 afforded disaccharide 102 in 98% yield as shown in Scheme 43. In these cases, no 1,6-anhydro derivative formation was noticed. The proposed reaction mechanism follows a traditional Lewis acid-mediated reaction according to which FeCl_3 associates with the corresponding glycosyl chloride, causing the activated glycosyl donor to form. The chloride then leaves resulting in the formation of the oxacarbenium ion, which was the cause of the poor stereoselectivity observed in most glycosylations with benzylated glycosyl bromides due to the ability of the acceptor to attack from either side of the flattened chair. Besides being catalytic, the ability to glycosylate the disarmed, benzoylated sugars was advantageous over previously described methods.

In 2021, Demchenko and co-workers have developed very mild reaction conditions for the activation glycosyl chlorides in the presence of bismuth(III) triflate.³⁰⁸ This reaction worked similarly to that of glycosylation with glycosyl bromides. Tetra-benzoylated and tetra-benzylated galactosyl chlorides 180 and 144 were glycosidated with glycosyl acceptor 36 in the presence of $\text{Bi}(\text{OTf})_3$ (0.35 equiv) to produce galactosides 78 and 79, respectively (Scheme 44A). An interesting observation was made when activation of glycosyl chlorides and bromides were compared. The activation of galactosyl chloride 180 was

Scheme 44. Bismuth(III)- and Palladium(II)-Catalyzed Activations of Glycosyl Chlorides



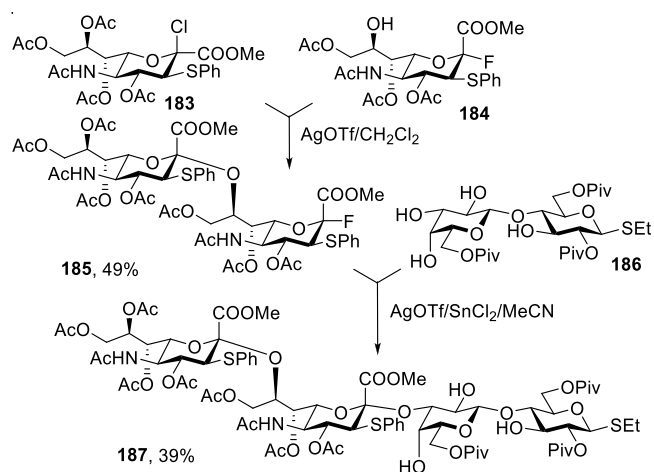
faster (15 min) when compared with the activation of a similarly protected galactosyl bromide 40 (30 min). Benzylated galactosyl chloride 144 produced galactoside 79 in a better yield (97%, $\alpha/\beta = 1/1.2$) when compared to results from benzylated galactosyl bromide 34 (78%, $\alpha/\beta = 1/1.1$).

Very recently, Chen and co-workers developed a new protocol to activate glycosyl chloride donors to synthesize O-glycosides using palladium(II) reagents.³⁷⁵ On the basis of their previous work of palladium-catalyzed C-aryl glycoside synthesis, the authors established that palladium acetate can serve as a Lewis acid that will interact with the chloride leaving group thereby activating the anomeric C–Cl bond.³⁷⁶ To broaden the scope of this approach to the synthesis of O-glycosides, 2,3,4,6-tetrabenzyl mannosyl bromide 181 was treated with 2-phenylethanol 92 in the presence of 2 mol % of $\text{Pd}(\text{OAc})_2$ in chloroform at room temperature (Scheme 44B). The glycosylation produced glycoside 182 in 88% yield with exclusive α -stereoselectivity. By tuning the reaction temperature (25 to 110 °C) and amount of $\text{Pd}(\text{OAc})_2$ (2 to 5 mol %), various glycosyl acceptors were glycosylated producing the respective products in good to excellent yields with exclusive α -stereoselectivity. Glycosyl donors of different sugar series including gluco, galacto, rhamno, ribofurano, and mannofurano were also investigated. Glycoside products were obtained in good yields; however, the stereoselectivity was difficult to predict.

4.3. Glycosyl Chlorides in Glycan and Glycoconjugate Synthesis

2-Chloro and 2-fluoro sialic acid derivatives 183 and 184, both bearing a participating 3-S-phenyl auxiliary, have been employed for the preparation of tetrasaccharide 187 using a selective activation strategy.^{367,377} Glycosyl fluorides are stable toward the reaction conditions required for the activation of glycosyl chlorides. Thus, coupling of sialyl chloride donor 183 with 8-OH sialyl fluoride acceptor 184 in the presence of AgOTf gave (2→8)-linked dimer 185 in 49% yield with complete α -stereoselectivity (Scheme 45). The resulting dimer 185 was used in the subsequent glycosylation of thiolactosyl acceptor 186 in the presence of the $\text{AgOTf}/\text{SnCl}_2$ promoter system. The resulting tetrasaccharide 187 was obtained in 39% yield and its anomeric thioglycoside moiety can be activated for subsequent glycosylations directly.

Scheme 45. Selective Activation of Sialyl Chloride 142 over Sialyl Fluoride 167



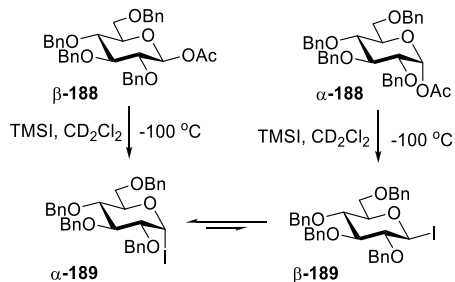
5. GLYCOSYL IODIDES

5.1. Synthesis of Glycosyl Iodides

The first synthesis of glycosyl iodides was reported by Fischer, who synthesized 2,3,4,6-tetra-*O*-acetyl- α -D-glucopyranosyl iodide by reaction of per-*O*-acetylated glucose with HI.¹⁴⁸ Fischer also noted that the glycosyl iodide quickly reacted with methanol in the presence of silver carbonate to afford the methyl glycoside. The field did not have much growth until 1974 when Kronzer and Schuerch³¹⁵ discovered that the glycosidation of benzylated glucosyl bromides could be promoted by the addition of sodium iodide. These glycosylation reactions were performed under metal-free conditions and presumed to occur through the intermediacy of glycosyl iodides.

Shortly thereafter, Thiem and Meyer²⁰⁷ reported that glycosyl iodides could be synthesized from a variety of precursors such as anhydrosugars, methyl glycosides, and per-acetylated hexoses using TMSI. This discovery allowed for the synthesis of many glycosyl iodide donors that for the first time became readily available. However, only acetylated glycosyl iodides were sufficiently stable to be fully characterized. Benzylated iodides were deemed too unstable and had to be synthesized and used in subsequent glycosylations *in situ*.⁵¹ This was the case until Gervay et al. devised a technique to fully characterize benzylated glycosyl iodides by monitoring their formation in the presence of TMSI in CD₂Cl₂ at $-100\text{ }^{\circ}\text{C}$ in an NMR spectrometer.³⁷⁸ The authors found that the anomeric peaks appeared as a doublet at either 6.68 ppm for α -glycosyl iodide **189** or at 5.61 ppm for β -glycosyl iodide. The ratios of these products depended on the configuration of the anomeric acetate in the substrate, α - or β -acetate **188**. Regardless of the initial configuration, the iodide displaced the acetates in an S_N2-like manner. Following the displacement, anomerization of the β -iodide rapidly occurs to the thermodynamically stable α -iodide (Scheme 46). Following these first major mechanistic studies, glycosyl iodides found a much broader application in synthetic chemistry.

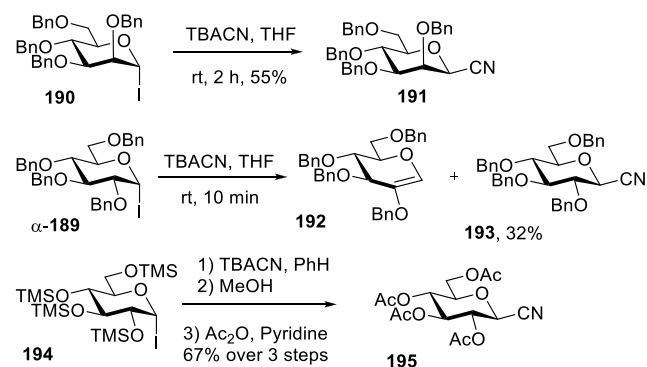
Scheme 46. Anomerization of Glycosyl Iodides



5.2. Glycosidation of Glycosyl Iodides

Among the first applications of glycosyl iodides as donors was the synthesis of C-glycosides that were shown to form in an S_N2-like manner with direct displacement of the glycosyl iodide with the nucleophile. Using tetrabutylammonium cyanide (TBACN) in tetrahydrofuran (THF) and α -mannosyl iodide **190**, β -cyanoglycoside **191** was obtained in 55% yield (Scheme 47).³⁷⁹ While this reaction worked well with the mannose iodide, glucosyl iodide α -189 produced β -cyanoglycoside **193** in a modest yield of 32% along with the major side product oxyglycal **192** resulting from the competing E2

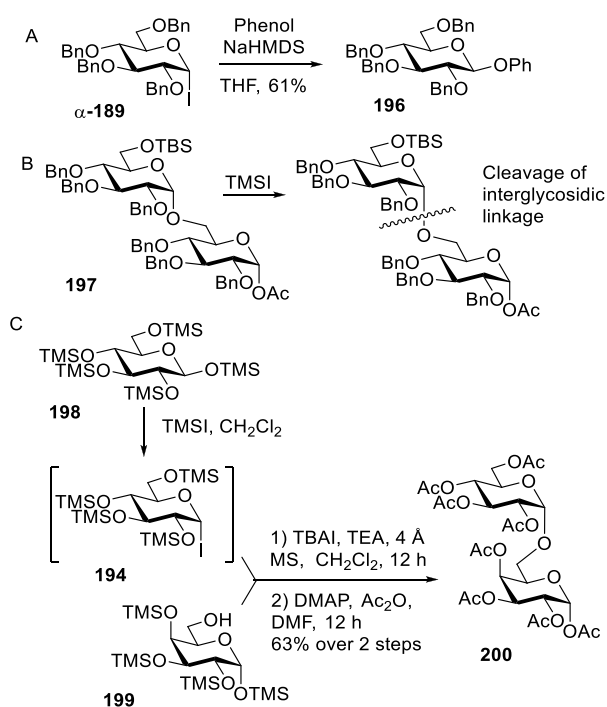
Scheme 47. Synthesis of C-Glycosides Using Glycosyl Iodides



elimination reaction. Switching to glucosyl iodide **194** equipped with TMS ethers allowed for a much more successful synthesis of the corresponding β -cyanoglycoside. This was accomplished using TBACN in toluene, followed by the cleavage of the TMS groups that was affected by addition of MeOH, and subsequent acetylation using Ac₂O in pyridine to give **195** in an overall yield of 67%. Similar C-glycoside formation was accomplished by using Grignard reagents in the synthesis of the glycolipid BbGL2, as reported by Kulkarni and Gervay-Hague.³⁸⁰

The synthesis of O-glycosides has also been approached from glycosyl iodides. When small nucleophiles were used such as phenol with NaHMDS in THF, glycosyl iodide α -189 gave phenol glycoside **196** in 61% yield (Scheme 48A). Other small nucleophiles such as sodium acetate or sodium tert-butoxide worked well giving complete β -stereoselectivity following direct displacement of the α -iodide.^{9,381} Synthesis of the disaccharide has also proven to be straightforward, however, converting the disaccharide into the second-generation

Scheme 48. Synthesis of O-Glycosides Using Glycosyl Iodides



glycosyl donor was troublesome. During displacement of the *O*-acetyl anomeric group from anomeric acetate **197** to the iodide donor, cleavage of the interglycosidic bond has occurred (Scheme 48B). This problem was solved by Lam and Gervay-Hague³⁸² by a simple addition of an acetate (or other electron-withdrawing) group at the C-6 position of the glycosyl donor. This modification allowed for the synthesis of oligosaccharide derivatives using both solid phase and solution phase strategies using TBAI as a promoter system. Other promoter systems used for the synthesis of oligosaccharides include AgOTf,³⁸² tetrabutylammonium bromide/Na₂CO₃,³⁸³ AgNO₃,³⁸⁴ ZnI,³⁸⁵ and TBAI/DIPEA.³⁸⁶

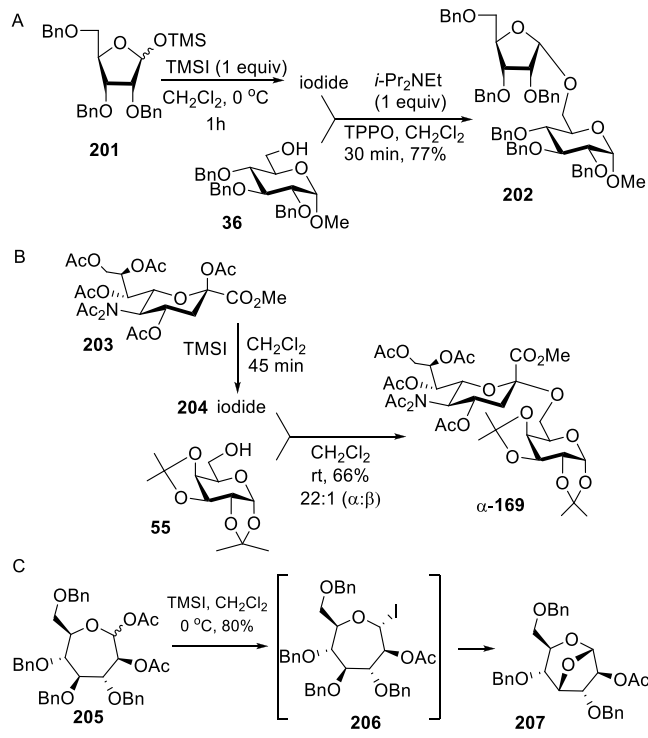
Another method to help combat the interglycosidic bond cleavage employs fully trimethylsilyl protected substrates. Introduced by Gervay-Hague and co-workers,³⁸⁷ this approach allowed to achieve high yields in glycosidations of per-*O*-silylated galactosyl iodides. This approach was successfully applied to α -stereoselective synthesis of glycolipids, however some decline in yields was seen due to the formation of silylated acceptors as side products. The reaction was not perfected until later when Gervay-Hague³⁸⁸ found that the formation of silylated side products can be suppressed by reducing the amount of TBAI to 1.5 equiv as opposed to 3.0 equiv used previously. Since this discovery, many research groups have used per-*O*-silylated sugars to synthesize a variety of natural products.^{389–391} Glycosyl iodides have been used in a variety of ways that were comprehensively discussed in previous reviews by Kulkarni,³⁹² Lowary,³⁹³ and Gervay-Hague.³⁹⁴

More recently, Zhang and co-workers³⁹⁵ expanded the scope of per-silylated glycosyl donors such as **198** and improved the outcome of the reaction by supplementing TBAI-promoted glycosylations with triethylamine (Scheme 48C). Under these conditions, glycosyl donor **198** was reacted with glycosyl acceptor **199** to form disaccharide **200** in 63% yield over two steps after subsequent acetylation. These reactions worked well with a variety of sugar series such as glucosyl, galactosyl, and fucosyl donors.

Glycosyl iodides were also used in α -stereoselective ribofuranosylation of alcohols.³⁹⁶ Ribofuranosyl iodide could be generated using TMSI from precursor **201** (Scheme 49A). Following the addition of *i*-Pr₂NEt and triphenylphosphine oxide, which acts as an additive to improve α -stereoselectivity, and glycosyl acceptor **36**, ribofuranosylation would occur. As a result, disaccharide **202** was obtained in 77% yield with complete α -stereoselectivity. These conditions worked well for a variety of glycosyl acceptors ranging from primary aliphatic alcohols to hindered sugar alcohols with yields of 75% or higher. Ribosylation was also studied by Houston and Koreeda³⁹⁷ using *i*PrOH as the glycosyl acceptor in the presence of I₂ as a promoter in THF. This reaction gave the corresponding β -ribosides in high yields. The authors also found that ribosylations performed in the presence of acetone led to the formation of a 1,2-*O*-isopropylidene derivative instead.

The most recent advancement in the application of glycosyl iodides was reported by Park and Gervay-Hague.³⁹⁸ The authors achieved the first, promoter-free sialylations with sialyl iodides, which was applied to the synthesis of steryl β -sialosides. These glycosylations only worked with C-5 modified sialic acid donors, whereas traditional *N*-acetamido sialic acids underwent 2,3-elimination upon the attempt to obtain a sialyl iodide donor. However, when 5-*N*-acetylacetamido precursor

Scheme 49. Glycosidation of Iodides of Different Series



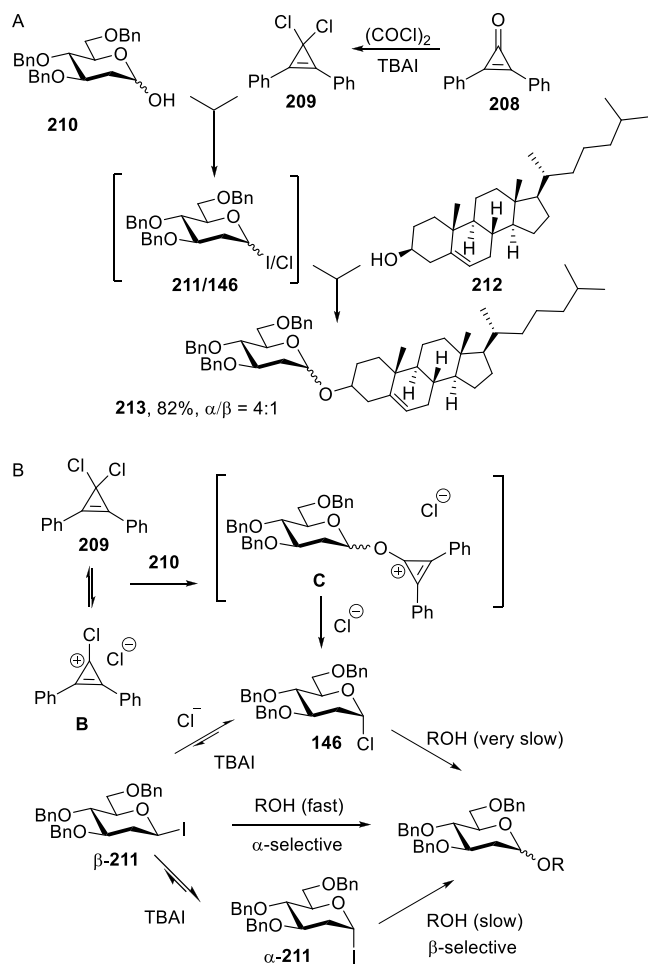
203 was investigated, the corresponding α -iodide donor **204** was smoothly produced (Scheme 49B). Sialylation could then be performed in a one-pot manner at rt; for example, reaction of the primary glycosyl acceptor **55** gave disaccharide **169** in 66% yield with excellent α -stereoselectivity (α : β = 22:1). Cholesterol-based acceptors provided respectable yields ranging from 52% to 85% giving sialosides with complete β -stereoselectivity.

Applications of glycosyl iodides in synthesis span beyond their use as glycosyl donors. For instance, glycosyl iodides were used as precursors in the formation of 1,4-anhydroseptanoses.³⁹⁹ As depicted in Scheme 49C, septanose **205** was reacted with TMSI to form septanosyl iodide **206** that readily rearranged to form 1,4-anhydroseptanose **207** in 30 min in 80% yield. This rapid cyclization was occurring in septanoses derived from glucose, mannose, xylose, and galactose.

Bennett and co-workers reported a dehydrative glycosidation of 2-deoxy and 2,6-dideoxy-sugars in the presence of 3,3-dichloro-1,2-diphenylcyclopropene **209**, tetrabutylammonium iodide (TBAI), and *N,N*-diisopropylethylamine (DIPEA).⁴⁰⁰ The authors assumed that the treatment of benzyl protected 2-deoxy hemiacetal **210** under these reaction conditions will lead to the *in situ* formation of the corresponding glycosyl iodide **211**. The latter will then be trapped with cholesterol **212** to produce glycoside **213** in 72% yield with preferential α -selectivity (α / β = 4:1, Scheme 50A). Increasing the amount of TBAI to 5.0 equiv increased the yield of **213** to 82%, however, there was no apparent change in stereoselectivity. Similarly, an α -selective formation 2,6-dideoxyglycosides was achieved.

The authors proposed a stepwise mechanistic conversion. 3,3-Dichloro-1,2-diphenylcyclopropene **209** is postulated to exist in equilibrium with intermediate B (Scheme 50B). The latter assists conversion of hemiacetal **210** to the corresponding anomeric chloride **146** through a reactive intermediate C. As evident from NMR experiments, anomeric chloride **146**

Scheme 50. Dehydrative Glycosidation of Hemiacetals

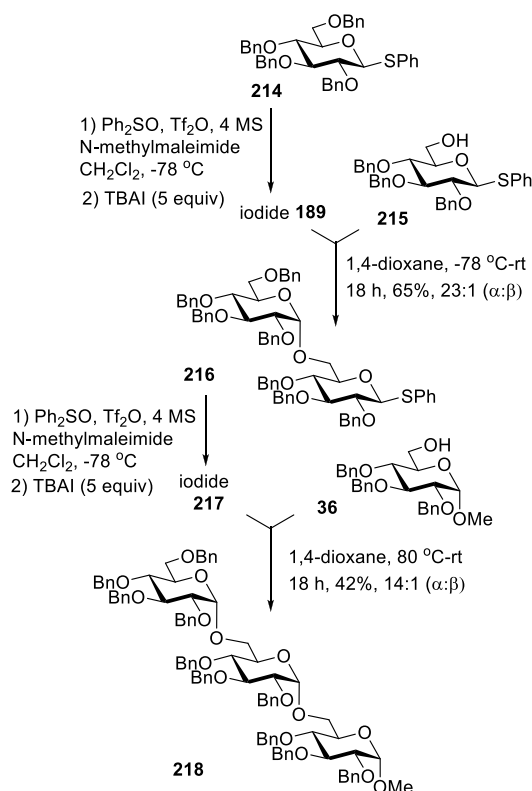


exists in equilibrium with another species that is postulated to be either α -211 or β -211. Excess iodide (TBAI) in reaction mixture promotes equilibrium between stable α -211 and its reactive counterpart β -211. The fact that preferential α -selective was observed for the products suggests that glycosyl acceptor might be approaching β -iodide intermediate 211 in an S_N2 -like fashion.

5.3. Glycosyl Iodides in Glycan and Glycoconjugate Synthesis

Bennett used a glycosyl iodide generated *in situ* for the synthesis of α -glycosides without directing groups.⁸⁸ Starting from stable thioglycoside 214, the corresponding anomeric triflate was generated in the presence of Ph_2SO , Tf_2O , and 4 Å molecular sieves in dichloromethane at -78°C (Scheme 51). Following generation of the glycosyl triflate *in situ*, 5 equiv of TBAI was added to produce glycosyl iodide 189. 1,4-Dioxane was then added to improve α -stereoselectivity along with glycosyl acceptor 215. As a result, disaccharide 216 was synthesized in 65% yield in excellent α -stereoselectivity ($\alpha \rightarrow \beta = 23/1$). This reaction sequence was reiterated with *in situ* generated iodide 217 and glycosyl acceptor 36 allowing for the synthesis of trisaccharide 218. However, a modest yield of 42% was observed since a sterically bulky glycosyl donor was used at this stage.

Scheme 51. Iterative Synthesis of Trisaccharide 196 Using Glycosyl Iodides



6. GLYCOSYL FLUORIDES

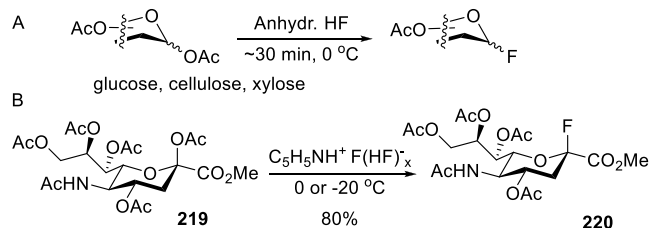
The early expansion of synthetic carbohydrate chemistry was made possible due to the improved understanding of the mechanistic aspects of glycosylation with glycosyl halides (mainly chlorides and bromides) as donors. The synthesis of glycosyl fluorides has been known for a long time, but their glycosidation under Koenigs-Knorr conditions is impossible due to their higher stability. This demanded further investigation, and Mukaiyama and co-workers were the first to show the activation of glycosyl fluorides in the presence of Lewis acids such as tin(II) chloride (SnCl_2). The activation protocol was subsequently improved by using silver salts as additives. These early attempts led to the formation of 1,2-*cis* glycosides, which are difficult to synthesize. Following these early discoveries, glycosyl fluorides have become increasingly popular glycosyl donors in synthetic glycochemistry due to a unique combination of high stability and relatively mild Lewis acidic conditions required for their glycosidation.

6.1. Synthesis of Glycosyl Fluorides

Numerous methods for the synthesis of glycosyl fluorides from a variety of precursors have been developed. In the following, all known methods have been categorized by the type of the starting material used for the introduction of the anomeric fluoride moiety.

6.1.1. Preparation from Anomeric Acetates. The very first synthesis of glycosyl fluorides comprised the treatment of per-acetylated sugars with anhydrous hydrofluoric acid. The acid was generated by heating dry potassium hydrogen fluoride allowing for various glycosyl fluorides to be synthesized in 30 min (Scheme 52A).⁴⁰¹ Over the years, milder protocols that are compatible with acid-sensitive functional and protecting groups have been developed. One popular protocol involves

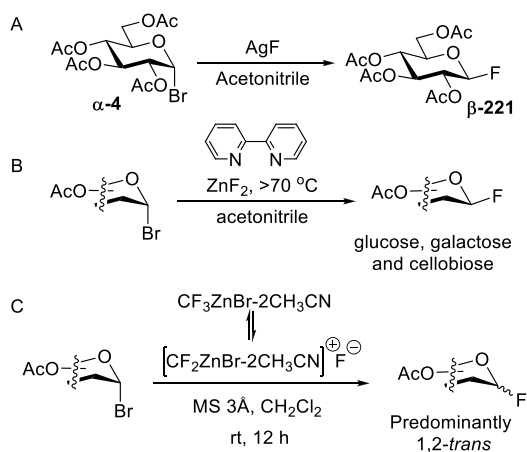
Scheme 52. Synthesis of Glycosyl Fluorides from Acetates



Py-HF complex as a source of HF. Thus, Sharma and Eby employed the fluorination procedure developed by Olah et al.⁴⁰² to convert sialic acid **219** to the corresponding β -fluoride **220** (Scheme 52B).⁴⁰³ Noyori and co-workers extended this approach to different sugar series for the synthesis of anomeric fluorides.^{404,405} It was shown that the treatment of a solution of benzyl or acetyl protected glycosyl acetates in toluene with 50 or 70% hydrogen fluoride-pyridine mixture readily produced the corresponding fluorides in good to excellent yields. This method turned out to be advantageous for the synthesis of thermodynamically favored α -fluorides. Differentially protected hemiacetals are also suitable substrates (*vide infra*), but their conversion to fluorides under these conditions was not as efficient as that from the anomeric acetates.⁴⁰⁶

6.1.2. Preparation from Glycosyl Chlorides and Bromides. Helferich and Gootz performed a halogen exchange reaction to synthesize glucosyl fluorides from the corresponding glucosyl bromides.⁴⁰⁷ For example, when 2,3,4,6-tetra-*O*-acetyl- α -D-glucosyl bromide α -4 was treated with silver fluoride (AgF) in acetonitrile the corresponding fluoride β -221 was afforded in 54% yield after recrystallization (Scheme 53A). Klemer and Micheel extended this approach to

Scheme 53. Synthesis of Glycosyl Fluorides from Other Halides



the synthesis of anomeric fluorides of other sugar series from the corresponding glycosyl bromides and chlorides.^{408,409} Hall and co-workers employed a similar approach to obtain glycosyl fluorides to study their conformational properties.⁴¹⁰ Igarashi and co-workers reported the synthesis of glycosyl fluorides from the corresponding chlorides in the presence of AgBF₄ in ether or toluene.⁴¹¹

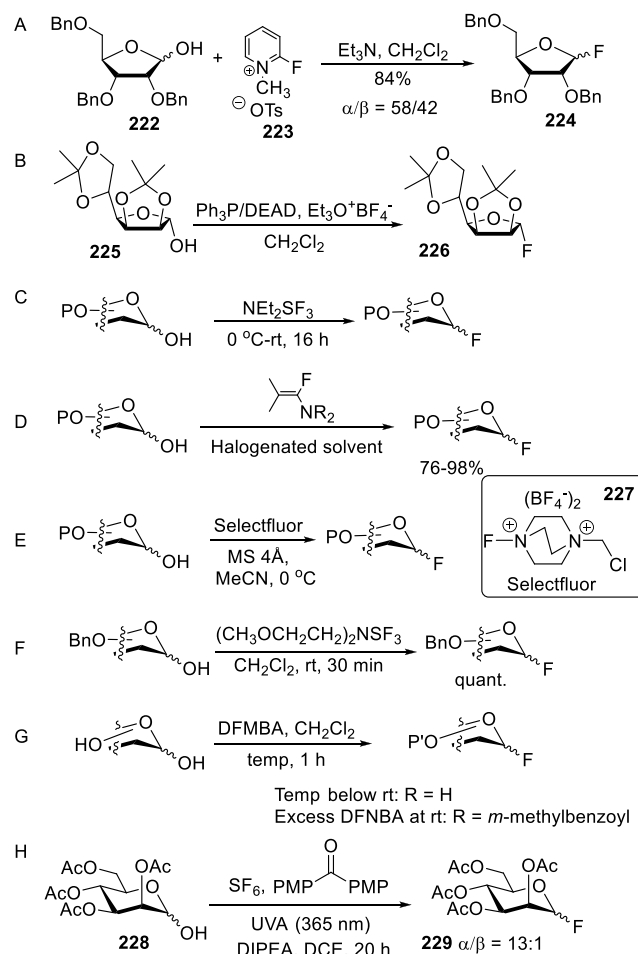
Walinsky and co-workers also investigated a halogen exchange procedure for the synthesis of glycosyl fluorides.⁴¹² For that, they chose zinc fluoride and the conversion of glycosyl bromides to the corresponding fluorides was

successfully conducted in the presence of ZnF₂ alone or in combination with 2,2'-bipyridine (Scheme 53B). Because of the poor solubility of zinc fluoride, these reactions worked best at higher temperatures in acetonitrile. Reactions in the presence of 2,2'-bipyridine were slower, but these conditions led to increased yields of glycosyl fluoride products.

Trifluoromethylzinc bromide (CF₃ZnBr·2CH₃CN) was known as a reagent for difluoromethylation reactions.^{413–415} Naumann et al. suggested that it can also act as a potential nucleophilic fluorinating reagent.⁴¹⁶ Miethchen and co-workers successfully fluorinated α -bromides of per-acetylated glucose, galactose, mannose, lyxose, and rhamnose in DCM with CF₃ZnBr·2CH₃CN in DCM (Scheme 53C).⁴¹⁷ Predominantly, 1,2-*trans* fluorides were obtained in these reactions. This protocol also worked with hemiacetals (*vide infra*), and the yield could be enhanced with TiF₄ additive. Miethchen and co-workers also achieved bromine–fluorine exchange in the presence of two-phase system Et₃N·3HF/CCl₄.^{418,419} This protocol also worked with hemiacetals as starting materials.

6.1.3. Preparation from Hemiacetals. Mukaiyama and co-workers reported the synthesis of glycosyl fluorides from the corresponding hemiacetals.⁴²⁰ Synthesis of 2,3,5-tri-*O*-benzyl- α/β -D-ribofuranosyl fluoride **224** was achieved by treating the corresponding hemiacetal **222** with 2-fluoro-L-methylpyridinium tosylate **223** in the presence of triethylamine as shown in Scheme 54A. The anomeric fluorides were

Scheme 54. Synthesis of Glycosyl Fluorides from Hemiacetals



separated by column chromatography, and the α -anomer was shown to equilibrate to β -anomer in the presence of boron trifluoride etherate. Kunz and Sager applied a modified Mitsunobu reaction^{421,422} to the synthesis of glycosyl fluorides equipped with acid-sensitive protecting groups.⁴²³ For example, isopropylidene protected mannofuranose **225** was treated with triphenylphosphine (Ph_3P), diethyl azodicarboxylate (DEAD), and triethyloxonium tetrafluoroborate to afford fluoride **226** in 54% yield (Scheme 54B).

Rosenbrook and co-workers successfully applied diethylaminosulfur trifluoride (DAST), a well-known reagent for the direct conversion of alcohols to fluorides⁴²⁴ to sugar hemiacetals. Thus, when hemiacetals were treated with neat DAST at 0 °C followed by warming to rt, the corresponding anomeric fluorides were produced in good yields (60–91%, Scheme 54C).⁴²⁵ Concomitantly, Posner and Haines reported DAST-mediated fluorination reactions that were surprisingly fast (20 min) when conducted in THF as the reaction solvent.⁴²⁶

Ernst and Winkler extended the fluorinating property of α -haloenamines, known reagents for the synthesis of acyl⁴²⁷ and alkyl halides,^{428,429} to the synthesis of glycosyl fluorides.³⁵⁵ Thus, when hemiacetals of pyranose or furanose sugars were treated with 1-fluoro-*N,N*,2-trimethylprop-1-en-1-amine or a similar reagent, glycosyl fluorides were produced in 76–98% yield (Scheme 54D). The mild nature of α -haloenamines along with the neutral reaction conditions were found to be compatible with various protecting groups including acetyl, benzoyl, isopropylidene, benzyl, and silyl. 1-Chloromethyl-4-fluoro-1,4-diazoniabicyclo[2.2.2]octane bis(tetrafluoroborate) **227**,⁴³⁰ commonly known as Selectfluor, was found to be an efficient reagent for converting hemiacetals to the corresponding fluorides (Scheme 54E).⁴³¹ This electrophilic fluorinating reagent was also found to be effective for converting glycals into 2-deoxyglycosyl fluorides. In the presence of SMe_2 additive, Selectfluor also reacted with thioglycosides to produce glycosyl fluorides (*vide infra*).

Thermal instability of DAST hampered its application in large-scale syntheses or reactions at high temperatures.⁴³² Lal and co-workers developed the bis(2-methoxyethyl)aminosulfur trifluoride (Deoxo-Fluor) reagent, as a thermally stable analogue of DAST.⁴³³ Deoxo-Fluor is also a more efficient fluorination reagent compared to DAST as evidenced by a quantitative conversion of hemiacetal derivatives to the corresponding anomeric fluorides (Scheme 54F). Hara and co-workers reported the synthesis of glycosyl fluorides by treating hemiacetal derivatives with *N,N*-diethyl- α,α -difluoro-(*m*-methylbenzyl)amine (DFMBA) as shown in Scheme 54G.^{434,435} It was noticed that hydroxyl groups at nonanomeric positions were not affected at low temperatures. However, the reactions performed in the presence of excess DFMBA would lead to *m*-methylbenzoylation along with the anomeric fluorination.

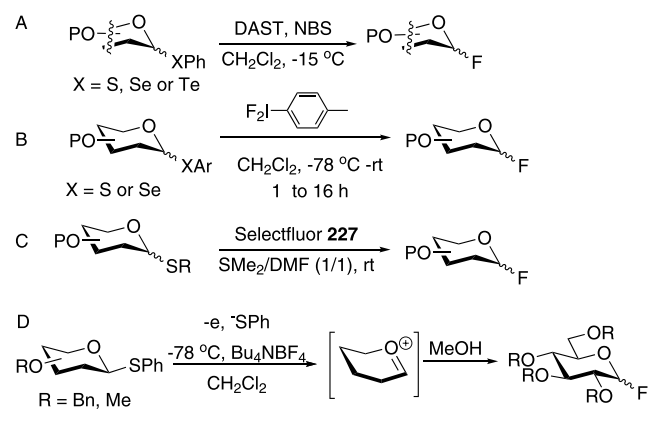
Nagorny and co-workers developed photochemical reaction conditions for the synthesis of glycosyl fluorides from the corresponding hemiacetals.⁴³⁶ For that, the authors employed sulfur(VI) hexafluoride, which is an inexpensive and mild fluorinating reagent, along with a photocatalyst. As shown in Scheme 54H, fluorination of 2,3,4,6-tetra-*O*-acetyl- β -mannose **228** with SF_6 (gas, 1 atm) in the presence of photocatalyst 4,4'-dimethoxybenzophenone (30 mol %) using a UV-A LED source ($\lambda_{\text{max}} = 365 \text{ nm}$) produced the corresponding mannosyl fluoride **229** in 70% yield ($\alpha/\beta = 13:1$). Different sugar series

with a variety of protecting groups were investigated, and glycosyl fluorides were obtained in yields of 43–97%. Further, the authors successfully showed gram scale formation of glycosyl fluorides in continuous flow systems and using electrochemical synthesis.⁴³⁷

6.1.4. Preparation from Thio- and Selenoglycosides.

Nicolaou and co-workers reported the direct conversion of phenylthio glycosides to glycosyl fluorides.³³³ Phenylthio glycosides were first treated with DAST followed by *N*-bromosuccinimide (NBS) to afford the corresponding glycosyl fluorides (Scheme 55A). This method is very effective, and

Scheme 55. Synthesis of Glycosyl Fluorides from Chalcone Glycosides



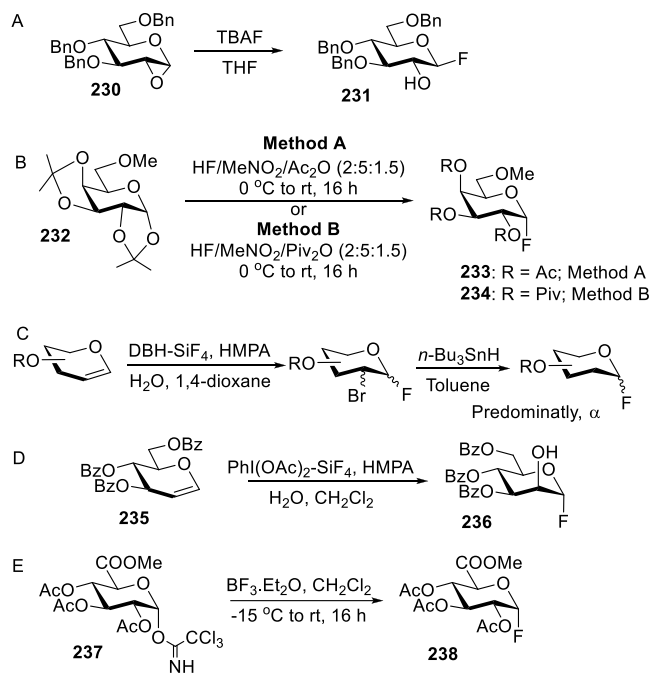
several common protecting groups and *O*-glycosidic linkages were tolerated effortlessly. This method has created a basis for developing a two-stage activation and orthogonal strategies for glycan assembly (*vide infra*). Because of the similarity between thioglycosides and other chalcogen glycosides, Horne and Mackei converted phenylseleno and phenyltelluro glycosides to the corresponding glycosyl fluorides in the presence of DAST and NBS/NIS.⁴³⁸ The stereochemistry of obtained glycosyl fluorides was found to depend on various factors such as substituents at C-2, the nature of solvents, and the stereochemistry of starting materials.

Synthesis of several glucosyl fluorides was also achieved by treating arylthio glycosides with iodoarene difluoride reagents. Originally developed for noncarbohydrate substrates,⁴³⁹ Motherwell and co-workers then extended their studies to the synthesis of glycosyl fluorides from thioglycosides⁴⁴⁰ and selenoglycosides (Scheme 55B).⁴⁴¹ As aforementioned, Wong's approach to synthesizing fluorides with Selectfluor could be applied to hemiacetals, glycals, and thioglycosides as substrates.⁴³¹ The latter required the use of dimethyl sulfide along with Selectfluor (Scheme 55C). Yoshida and co-workers attempted to isolate glycosyl cation intermediate by the "cation pool" method.⁴⁴² Although this attempt was unsuccessful, the formation of glycosyl fluorides was achieved with the right combination of electrolytes.⁴⁴³ A low-temperature electrolysis reaction was conducted in the presence of Bu_4NBF_4 in CH_2Cl_2 . Subsequent addition of methanol did not produce methyl glycosides, but rather afforded corresponding glycosyl fluorides in good yields (Scheme 55D). Subsequently, the authors figured out the one-pot glycosylation method by replacing Bu_4NBF_4 with other suitable electrolytes.

6.1.5. Preparation from Other Substrates. Danishefsky and Gordon reported the synthesis of 3,4,6-tri-*O*-benzyl- β -D-

glucopyranosyl fluoride **231** by treating 1,2-anhydro-3,4,6-tri-*O*-benzyl- α -D-glucopyranose **230** with tetrabutylammonium fluoride (TBAF, Scheme 56A).⁴⁴⁴ Though the yield was

Scheme 56. Synthesis of Glycosyl Fluorides from Other Starting Materials



moderate (53%), the method is uniquely suited to produce the β -anomer only. Miethchen and co-workers prepared a fluorinating reagent by mixing anhydrous HF with acylating reagents to perform glycosyl fluoride synthesis and protection in one pot.⁴⁴⁵ Thus, a homogeneous mixture of HF/MeNO₂/Ac₂O (2/5/1.5, Method A) or HF/MeNO₂/Piv₂O (2/5/1.5, Method B) was found to transform 1,2:3,4-di-*O*-isopropylidene-6-*O*-methyl- α -D-galactopyranose **232** and other similar derivatives to the corresponding tri-*O*-acylated fluorides such as **233** and **234** as depicted in Scheme 56B. Protecting groups that are compatible with these conditions are alkyl, acyl, alkyl sulfone, etc.

As aforementioned, Selectfluor is another efficient method for converting glycals to the corresponding 2-deoxy-2-fluoro derivatives.⁴³¹ Shimizu and co-workers developed a protocol for bromofluorination of olefins with silicon tetrafluoride and 1,3-dibromo-5,5-dimethylhydantoin (DBH),⁴⁴⁶ which was later applied to the synthesis of 2-deoxyglycosyl fluorides.⁴⁴⁷ This was accomplished via sequential bromofluorination of glycals followed by debromination as shown in Scheme 56C. First, bromofluorination of glycals derived from glucose, galactose, rhamnose, and fucose was conducted with SiF₄, DBH, hexamethylphosphoric triamide (HMPA) in water/1,4-dioxane to produce the corresponding 2-bromoglycosyl fluoride derivatives. The latter were then subjected to debromination with tributyltin hydride (*n*-Bu₃SnH) to afford the corresponding 2-deoxyglycosyl fluorides. The same authors also described hydroxyfluorination of glycals in the presence of PhI(OAc)₂, SiF₄, and HMPA. As shown in Scheme 56D, under these reaction conditions, α -mannosyl fluoride **236** was obtained from glucal precursor **235** in 73% yield.

Von Itzstein and co-workers observed the formation of glycosyl fluorides from *O*-imidates.⁴⁴⁸ As shown in Scheme 56E, the treatment of glycosyl imidate **237** with BF₃·Et₂O (1.0 equiv) resulted in the formation of α -fluoride **238** in 24%. This reaction was accompanied by the formation of a hemiacetal derivative because of competing hydrolysis. Jones and co-workers reported the synthesis of 1-fluorocellobiosyl fluoride by treating the corresponding diazine derivative with xenon difluoride (XeF₂).⁴⁴⁹

6.2. Glycosidation of Glycosyl Fluorides

Many different reagents and cooperative systems for the activation of glycosyl fluorides have been developed. To streamline the discussion, we chose to divide the activating reagents into different categories based on their fluorophilic nature (Table 2). The discussion begins from tin salts that were very instrumental for understanding the metal salt involvement in splitting the anomeric C–F bond and investigating the effects of different additives and the counteranions. We will then focus the discussion on how those studies enabled scientists to develop improved methods, and how the improved methods enhanced our synthetic capabilities. Recent studies dedicated to conformational analysis of glycosyl cations generated from glycosyl fluorides significantly enhanced our understanding of processes behind typical glycosylation reactions.⁴⁵⁰

6.2.1. Activation with Group 14-Based Reagents (Tin and Silicon). Mukaiyama and co-workers discovered that glycosyl fluorides can be activated by a cooperative effect of tin(II) chloride (SnCl₂, stannous chloride) and silver perchlorate (AgClO₄) to afford glycosides in excellent yields.⁸ The glycosylation under these reaction conditions often proceeds in a stereoselective manner affording 1,2-*cis* α -glycosides predominantly. As shown in Scheme 57A, glycosidation of 2,3,4,6-tetra-*O*-benzyl- β -D-glucopyranosyl fluoride **239** with various aliphatic and sugar acceptors provided the respective products in good yields of 76–96% and respectable stereoselectivity. A similar approach was extended to the synthesis of 1,2-*cis* glycofuranosides.⁴²⁰ However, these reactions were predominantly 1,2-*trans* stereoselective. Interestingly, replacing silver perchlorate with trityl perchlorate (TrClO₄) additive allowed to obtain the respective products with predominant 1,2-*cis* stereoselectivity (Scheme 57B).

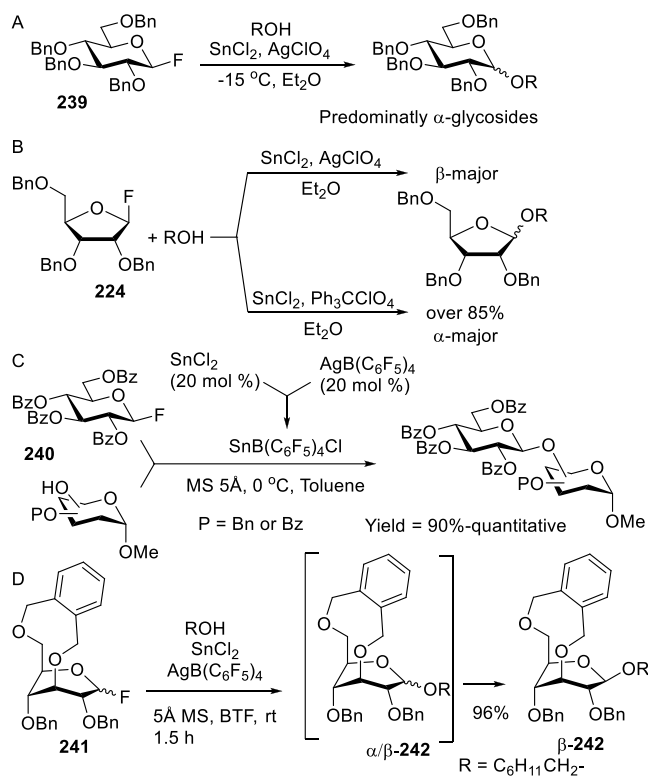
Subsequently, Mukaiyama and co-workers extended their study to SnCl₂ in combination with AgB(C₆F₅)₄ as a stable and useful reagent for generating the active catalyst.⁴⁵¹ This study also revealed that MS 5 Å additive (3 g/mmol) is necessary to achieve efficient activations. The cooperative catalytic activation was achieved by the combined use of SnCl₂ and AgB(C₆F₅)₄, 20 mol % each, in toluene. It was suggested that SnB(C₆F₅)₄Cl is an active catalyst for the activation of glycosyl fluoride **240** (Scheme 57C). An important aspect of this cooperative catalyst is that it does not work well in polar solvents, which was attributed to deactivation of Lewis acid by coordination to stannous cation. This cooperative system led to a successful installation of different linked disaccharides. For example, Yamada and co-workers activated the 3,6-*O*-bridged glucosyl fluoride donor **241** under these reaction conditions to obtain β -linked products, for example **242**, with complete stereoselectivity (Scheme 57D).⁴⁵⁴ The key to this exclusive β -stereoselectivity was due to the following two factors. First, the ability of the bridging positions C-3 and C-6 to force the sugar substituents to lock into the axial positions. Second, isomer-

Table 2. Activation of Glycosyl Fluoride Donors

promoter	additive
tin- and silicon-based reagents	
SnCl_2 ^{8,420}	AgClO_4 ^{8,451} , TrClO_4 ⁴²⁰ , AgOTf ^{451,452}
SnCl_4 ⁴⁵⁵	$\text{AgB}(\text{C}_6\text{F}_5)_4$ ^{451,453,454}
SnF_4 ⁴⁵⁶	$\text{AgB}(\text{C}_6\text{F}_5)_4$ ⁴⁵³
R_3SnCl or R_2SnCl_2 ⁴⁵⁷	AgClO_4 ⁴⁵⁷
$\text{Sn}(\text{OTf})_2$ ⁴⁵⁸	$\text{BF}_3\cdot\text{Et}_2\text{O}$, TiCl_4 , $\text{La}(\text{OTf})_3$, $\text{Yb}(\text{OTf})_3$, or $\text{La}(\text{ClO}_4)_3\cdot n\text{H}_2\text{O}$ ⁴⁵⁹
SiF_4 ⁴⁰⁴	
$\text{CF}_3\text{SO}_3\text{SiMe}_3$ ^{404,456,460}	
SiCl_4 or Ph_3SiCl ⁴⁵³	$\text{AgB}(\text{C}_6\text{F}_5)_4$ ⁴⁵³
boron-, aluminum-, and gallium-based reagents	
$\text{BF}_3\cdot\text{Et}_2\text{O}$ ^{423,455,461}	
$\text{TrB}(\text{C}_6\text{F}_5)_4$ ^{462,463}	
$\text{HB}(\text{C}_6\text{F}_5)_4$ ^{464,465}	
$\text{B}(\text{C}_6\text{F}_5)_3$ ⁴⁶⁶	
AlMe_3 ⁴⁵⁵	
Me_2GaCl , Me_2GaOTf or $\text{MeGa}(\text{OTf})_2$ ⁴⁶⁷	
main group metal salts	
$\text{MgBr}_2\cdot\text{Et}_2\text{O}$ ⁴⁵⁵	
LiClO_4 ^{468,469}	CsF ^{468,469}
CaBr_2 or $\text{Ca}(\text{NO}_3)_2$ ⁴⁷⁰	NMe_3 ⁴⁷⁰
$\text{Ca}(\text{OTf})_2$ ⁴⁷⁰	NMe_3 ^{470,471}
$\text{Ca}(\text{OH})_2$ ⁴⁷¹	
transition metal salts	
TiF_4 ^{456,460}	AgClO_4 ⁴⁷² , AgBF_4 ⁴⁷³
Cp_2TiCl_2 ⁴⁷⁴	AgClO_4 ⁴⁷⁴ , $\text{AgB}(\text{C}_6\text{F}_5)_4$ ⁴⁵³
TiCl_2 ⁴⁵³	$\text{AgB}(\text{C}_6\text{F}_5)_4$ ⁴⁵³
Cp_2ZrCl_2 ⁴⁷⁴	AgClO_4 ^{474,475} , AgOTf , AgBF_4 , AgPF_6 , or AgSbF_6 ⁴⁷⁵ , $\text{AgB}(\text{C}_6\text{F}_5)_4$ ⁴⁵³
SO_4/ZrO_2 ⁴⁷⁶	
Cp_2HfCl_2 ⁴⁷⁴	AgClO_4 ^{474,477,478} , $\text{AgB}(\text{C}_6\text{F}_5)_4$ ⁴⁵³
$\text{Hf}(\text{OTf})_4$ ⁴⁷⁹	
$\text{Cu}(\text{OTf})_2$ ⁴⁸⁰	
lanthanide metal salts	
$\text{Yb}(\text{OTf})_3$ ⁴⁸¹	K_2CO_3 ^{481,482} , CaCO_3 , $\text{K}_2\text{CO}_3/\text{ZnCl}_2$, or $\text{K}_2\text{CO}_3/\text{Ba}(\text{ClO}_4)_2$ ⁴⁸¹
YbCl_3 ⁴⁸¹	CaCO_3 ⁴⁸¹
Yb-Amberlyst-15 ⁴⁸³	
$\text{Yb}[\text{N}(\text{O}_2\text{SC}_4\text{F}_9)_2]_3$ ⁴⁸⁴	
$\text{La}(\text{ClO}_4)_3\cdot 7\text{H}_2\text{O}$ ^{481,482}	K_2CO_3 ^{481,482}
$\text{La}(\text{ClO}_4)_3\cdot n\text{H}_2\text{O}$, $\text{Pr}(\text{ClO}_4)_3\cdot n\text{H}_2\text{O}$, or $\text{Eu}(\text{ClO}_4)_3\cdot n\text{H}_2\text{O}$ ⁴⁸²	K_2CO_3 ⁴⁸²
$\text{Ce}(\text{ClO}_4)_3\cdot n\text{H}_2\text{O}$ ^{482,485}	K_2CO_3 ^{482,485}
anhydrides, protic acids, and other reagents	
Ti_2O ⁴⁶⁰	
TiOH ^{465,486–488}	
HClO_4 , $\text{HOSO}_2\text{C}_4\text{F}_9$, HNTf_2 or HSbF_6 ^{464,465}	
$\text{RN}^+\text{B}(\text{C}_6\text{F}_5)_4$, RN^+OTf , RN^+SbF_6 , RN^+BF_4 or RN^+ClO_4 ⁴⁸⁹	
solvolytic ⁴⁹⁰	

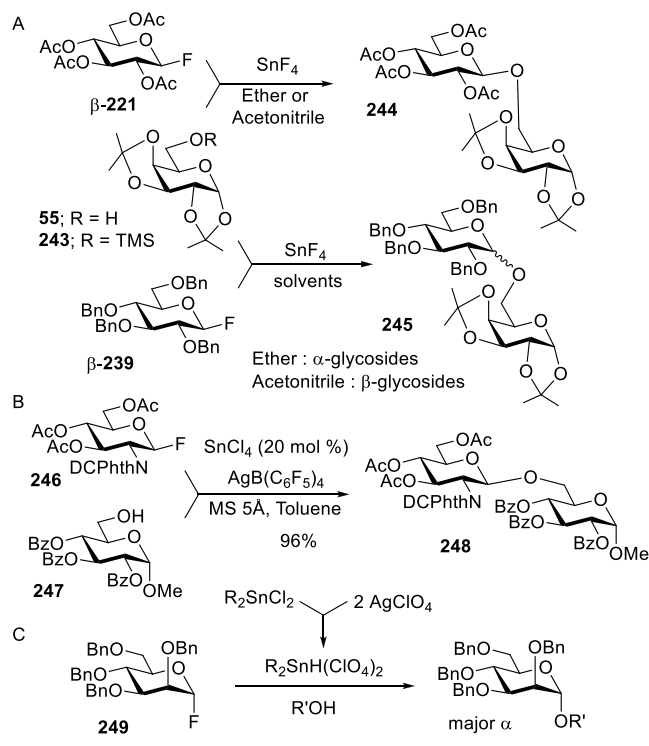
ization propensity to produce thermodynamically stable anomer. Upon glycosylation, α/β -glycosides, such as **242**, are produced along with $\text{HB}(\text{C}_6\text{F}_5)_4$, and the latter can catalyze the anomerization cycle of the α - to β -glycosides and, along with SnCl_4 , induce the regeneration of $\text{SnB}(\text{C}_6\text{F}_5)_4\text{Cl}$. Mechanistic investigation confirmed that $\text{SnB}(\text{C}_6\text{F}_5)_4\text{Cl}$ is the active catalyst for glycosylation.

Scheme S7. Glycosidation of Glycosyl Fluoride in the Presence of Tin(II)-Based Reagents



Beyond initial studies by Mukaiyama and co-workers wherein the activation was achieved with SnCl_2 ,⁴²⁰ several other tin salts are investigated such as SnF_2 , SnBr_2 , $\text{Sn}(\text{OAc})_2$. However, none of the salts provided satisfactory results without a copromoter.^{8,420} Similar observations were made by Nicolaou for SnCl_4 ⁴⁵⁵ and by Shibasaki for $\text{Sn}(\text{OSO}_2\text{CF}_3)_2$.⁴⁵⁸ Thiem and co-workers reported the activation of glycosyl fluorides with tin(IV) fluoride (stannic fluoride, SnF_4) without any additive.⁴⁵⁶ Glycosidation of acylated glycosyl fluoride β -221 having 2-participating group with glycosyl acceptor **55** or its 6-O-TMS-protected counterpart **243** in the presence of SnF_4 produced 1,2-*trans* glycoside **244** (Scheme 58A).

However, the yields of glycosides were below the preparative value (42–48%). Conversely, glycosidation of the benzylated glycosyl fluoride β -239 was more efficient and produced glycoside **245** in up to 93% yield, and the distribution of the products was found to be solvent-dependent, α -linked in ether or β -linked disaccharides in acetonitrile. It was then discovered that SnCl_4 is able to efficiently activate glycosyl fluorides in the presence of $\text{AgB}(\text{C}_6\text{F}_5)_4$.⁴⁵³ Surprisingly, SnCl_2 in combination with $\text{AgB}(\text{C}_6\text{F}_5)_4$ (20 mol % each), only produced the expected disaccharide **248** in trace amounts when glycosyl donor **246** was glycosidated with acceptor **247**. In comparison, SnCl_4 in combination with $\text{AgB}(\text{C}_6\text{F}_5)_4$ (20 mol % each) produced disaccharide **248** in an excellent yield (Scheme 58B). Therefore, SnCl_4 was employed for further optimization of the amount of $\text{AgB}(\text{C}_6\text{F}_5)_4$ that helped to establish 20 mol % of SnCl_4 and 40 mol % of $\text{AgB}(\text{C}_6\text{F}_5)_4$ as the most effective catalytic system for activating glycosyl donor **246** in toluene at -20°C . Finally, the optimized reaction conditions were extended to several disarmed glycosyl donors as well as

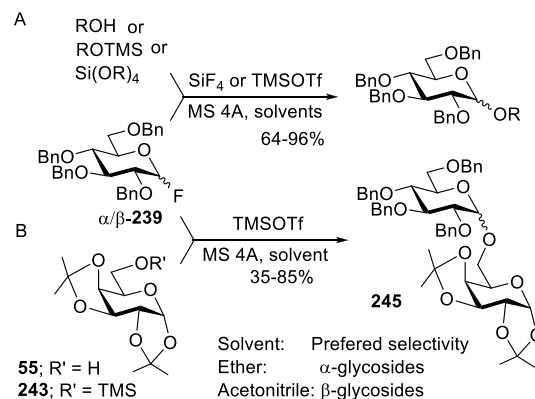
Scheme 58. Glycosidation of Glycosyl Fluoride in the Presence of Other Tin-Based Reagents

acceptors to synthesize the desired glycosides in excellent yields.

A metallocene-Ag salt-catalyzed approach was extended to organotin(IV) compounds to generate new activators for glycosyl fluoride donors.⁴⁵⁷ Two types of organotin compounds, R_2SnCl_2 and R_3SnCl (R = aryl or alkyl), were used in the presence of AgClO_4 . Whereas glycosidation of fluoride donor 249 in the presence of R_2SnCl_2 and AgClO_4 was almost instantaneous, reactions in the presence of R_3SnCl and AgClO_4 proceed slower. $\text{R}_2\text{Sn}(\text{ClO}_4)_2$ was proposed as the active catalyst for these reactions (Scheme 58C), and the stereo-selectivity was found to depend on the nature of the alkyl/aryl ligands of tin.

Noyori co-workers presented the synthesis of glycosides by coupling glycosyl fluorides and trimethylsilyl ether-protected glycosyl acceptors in the presence of tetrafluorosilane (SiF_4) or trimethylsilyl triflate (TMSOTf).⁴⁰⁴ Glycosidation of α - and β -glycosyl fluoride 239 was successfully achieved and the selectivity of glycoside products was found to depend on the type of solvents employed (Scheme 59A). Expectedly, acetonitrile predominantly produced 1,2-*trans* glycosides whereas ether favored 1,2-*cis* glycosides irrespective of anomeric conformation of glycosyl fluorides.

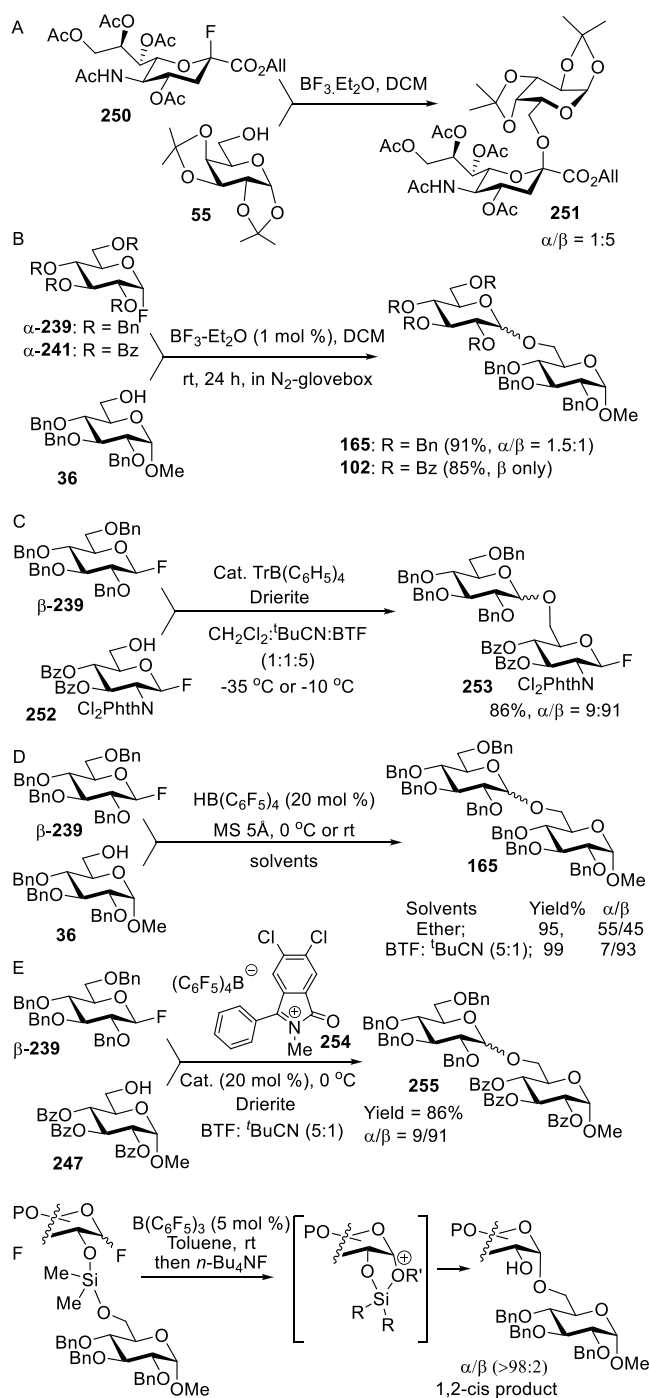
Further, it was confirmed that glycosides do not undergo anomericization over the course of the reaction. The efficiency of glycosylation reaction with glycosyl fluorides depends on the affinity of silicon atom to fluorine. Similarly, Thiem and co-workers also reported the activation of per-benzylated glycosyl fluorides in the presence of TMSOTf .⁴⁵⁶ For example, glycosidation of donor 239 with glycosyl acceptors 55 or 243 produced disaccharide 245. The yield of 245 could fall below preparative standards and the stereoselectivity of glycosides was also solvent dependent, as shown in Scheme 59B. An efficient activation was observed when glycosylation

Scheme 59. Glycosidation of Glycosyl Fluoride in the Presence of Silicon-Based Reagents

was conducted in the presence of SiCl_4 in combination with $\text{AgB}(\text{C}_6\text{F}_5)_4$ (20 mol % each). However, only moderate efficiency was achieved when the same glycosylation was conducted in the presence of Ph_3SiCl .⁴⁵³

6.2.2. Activation with Group 13-Based Reagents (Boron, Aluminum, and Gallium). Nicolaou and co-workers described the synthesis of O-, S-, and N-glycosides employing glycosyl fluoride donors in the presence of $\text{BF}_3\cdot\text{Et}_2\text{O}$ (0.3 equiv).⁴⁵⁵ This activator was found to be effective in most cases, however, there were specific cases where other metal promoters were found to be more effective (*vide infra*). Following essentially the same principle, Kunz and Waldman synthesized a variety of pyranosides and furanosides. Among these, coupling β -2-fluoro-neuraminic acid donor 250 with diacetone galactose acceptor 55 in the presence of $\text{BF}_3\cdot\text{Et}_2\text{O}$ afforded disaccharide 251 in 44% yield ($\alpha/\beta = 1:5$, Scheme 60A).^{423,461} Recently, $\text{BF}_3\cdot\text{Et}_2\text{O}$ -mediated glycosylation was revisited by Fukase and co-workers wherein it was discovered that an efficient activation of glycosyl fluorides can be achieved in the presence of only 1 mol % of $\text{BF}_3\cdot\text{Et}_2\text{O}$ in a nitrogen-filled glovebox.⁴⁹¹ An interesting feature of this reaction is that it does not require any dehydrating reagent compared to most glycosylation reactions. Armed per-benzylated glycosyl fluoride 239 and disarmed per-benzoylated glycosyl fluoride 240 both were successfully glycosidated with glycosyl acceptor 36 in the presence of 1 mol % of $\text{BF}_3\cdot\text{Et}_2\text{O}$ in nitrogen-filled glovebox and to produce disaccharides 165 in 91% yield ($\alpha/\beta = 1:5/1$) and 102 in 85% yield (β only), respectively (Scheme 60B). When the reaction was conducted in a polytetrafluoroethylene vessel, the yield has drastically declined suggesting the involvement of a glass vessel in glycosylation reaction. Earlier, a similar observation of glass vessel's involvement was made by Pedersen and co-workers wherein the formation of SiF_4 was suggested to occur from HF and glass vessel.⁴⁹² On the basis of the observations, the authors proposed a tentative mechanism according to which the anomeric activation is initiated by $\text{BF}_3\cdot\text{Et}_2\text{O}$ that produces HF. The latter then reacts with glass and produces SiF_4 , which acts as the activator of the anomeric fluoride.

Mukaiyama and Takeuchi reported a stereocontrolled catalytic activation of glycosyl fluorides in the presence trityl tetrakis(pentafluorophenyl)borate.⁴⁶² Optimization of the reaction conditions such as catalyst loading, solvents, and temperature led to the establishment of a reaction conditions, which favored 1,2-*trans* β -glycosides. Differently linked disaccharides were synthesized in presence of 10–20 mol %

Scheme 60. Glycosidation of Glycosyl Fluoride in the Presence of Boron-Based Reagents


of trityl tetrakis(pentafluorophenyl) borate and Drierite in pivalonitrile-benzotrifluoride (1:5) at -10 to 0°C . Similarly, β -stereoselective glycosylation was observed with tetrabenzylated galactosyl fluoride and various glycosyl acceptors.⁴⁸⁶ Further, $\text{TrB}(\text{C}_6\text{F}_5)_4$ -catalyzed glycosylation was successfully applied to an armed-disarmed glycosylation concept, wherein armed glycosyl fluoride donor β -239 was coupled with disarmed glycosyl fluoride acceptor 252 to afford the corresponding disaccharide 253 in a good yield (Scheme 60C).⁴⁸⁶ Several other disarmed fluoride acceptors were successfully glycosylated with armed glycosyl donor β -239.

Tetrakis(pentafluorophenyl)boric acid-catalyzed activation was also investigated. As shown in Scheme 60D, reaction of glycosyl fluoride donor β -239 with acceptor 36 produced disaccharide 165 with practically no stereoselectivity.⁴⁶⁴ Solvent investigation studies showed that $\text{HB}(\text{C}_6\text{F}_5)_4$ -catalyzed glycosylation in a mixture of solvent, i.e. benzotrifluoride/*tert*-butyl nitrile (5:1), produced disaccharides with excellent β -selectivity. The above observations suggested that the counteranion together with solvents play a crucial role in directing the stereoselectivity of the reaction. β -Stereoselective glycosidation of glycosyl fluoride β -239 was extended with a variety of glycosyl acceptors. In all cases disaccharides were obtained in high yields and with excellent β -selectivity (up to $\alpha/\beta = 2/98$). $\text{HB}(\text{C}_6\text{F}_5)_4$ -catalyzed α -stereoselective glycosylation was also achieved when a glycosyl fluoride having diethylthiocarbamoyl group at C-6 was used as the glycosyl donor and diethyl ether was used as the reaction solvent.⁴⁹³

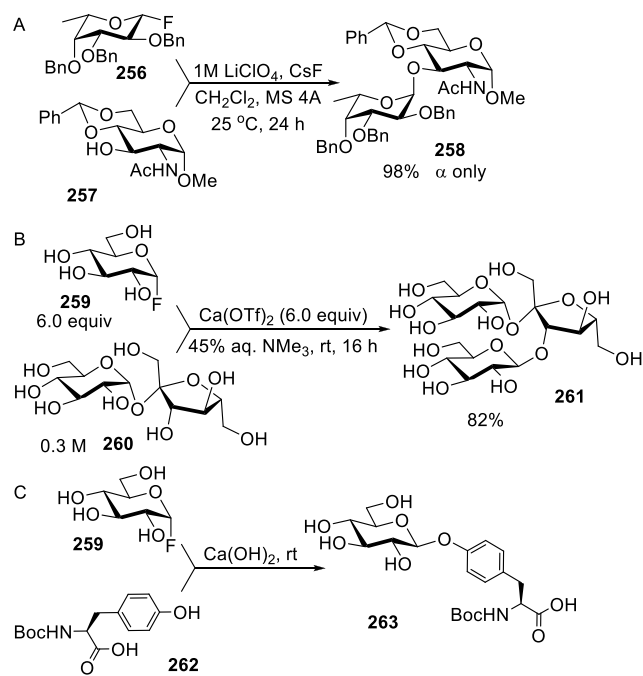
Mukaiyama and co-workers investigated the activation of glycosyl fluorides in the presence of catalytic carbocationic species paired with counterion such as tetrakis (pentafluorophenyl) borate or other anions.⁴⁸⁹ Glycosidation of glycosyl fluoride β -239 with glycosyl acceptor 247 in ${}^t\text{BuCN}$ in the presence of a cationic catalyst 254 produced disaccharide 255 in 86% yield with excellent β -selectivity (Scheme 60E). Boron-based catalytic activation of glycosyl fluorides was extended by Montgomery and co-workers for silyl ethers as glycosyl acceptors.⁴⁶⁶ The Frustrated Lewis Pair (FLP) characteristics of $\text{B}(\text{C}_6\text{F}_5)_3$ ^{494,495} and $\text{B}(\text{C}_6\text{F}_5)_3$ -catalyzed trifluoromethylation via fluoride-rebound mechanism^{496,497} inspired the idea for the coupling glycosyl fluoride with silyl ethers in the presence of $\text{B}(\text{C}_6\text{F}_5)_3$.

Efficient coupling of numerous glycosyl fluorides equipped with a 2-O-acyl group with silyl ether acceptors in the presence of 5 mol % of $\text{B}(\text{C}_6\text{F}_5)_3$ rapidly provided the corresponding 1,2-*trans*-linked glycosides in good to excellent yields at rt. Exploitation of silicon-tethered intramolecular aglycone delivery (IAD) method was employed to construct 1,2-*cis* linkages. As shown in Scheme 60F, the glycosidation of the tethered donor-acceptor pair led to glycosides with excellent stereoselectivity. It was assumed that the reaction proceeds via the key five-membered intermediate that collapsed to produce 1,2-*cis*-linked products with concomitant removal of the silicon tether.

Nicolaou and co-workers described the synthesis of O-, S-, and N-glycosides employing glycosyl fluoride donors in the presence of AlMe_3 .⁴⁵⁵ Kobayashi and co-workers utilized different methylgallium chlorides and triflates that were found to activate glycosyl fluorides.⁴⁶⁷ Simple aliphatic alcohols produced glycosides in excellent yields (77% to quantitative) with moderate stereoselectivity; however, switching to hindered or sugar alcohols led to decrease in yields.

6.2.3. Activation with Main Group Metal Salts (Magnesium, Lithium, and Calcium). Nicolaou and co-workers investigated the synthesis of O-, S-, and N-glycosides employing glycosyl fluoride donors in the presence of $\text{MgBr}_2 \cdot \text{Et}_2\text{O}$.⁴⁵⁵ Waldmann and Bohm discovered neutral reaction conditions for the activation of glycosyl fluorides.⁴⁶⁸ As shown in Scheme 61A, fluoride donor 256 was reacted with glycosyl acceptor 257 in a 1 M solution of LiClO_4 in the presence of CsF as the proton scavenger to afford disaccharide 258 in 98% yield with exclusive α -stereoselectivity. Different types of glycosyl acceptors produced both glycosides with predominant α -stereoselectivity. This methodology was later applied to

Scheme 61. Glycosyl Fluoride Activation with Main Group Metal Salts



glycosidation of benzylated glucosyl fluorides.⁴⁹⁸ The glucosides were obtained in moderate yields, but increasing the concentration of LiClO_4 promoted the formation of the respective 1,6-anhydro derivative. Further, acylated glucosyl fluorides were tested; while pivaloyl-protected fluoride produced 1,2-*trans*-linked products, acetylated fluoride was unreactive.⁴⁶⁹

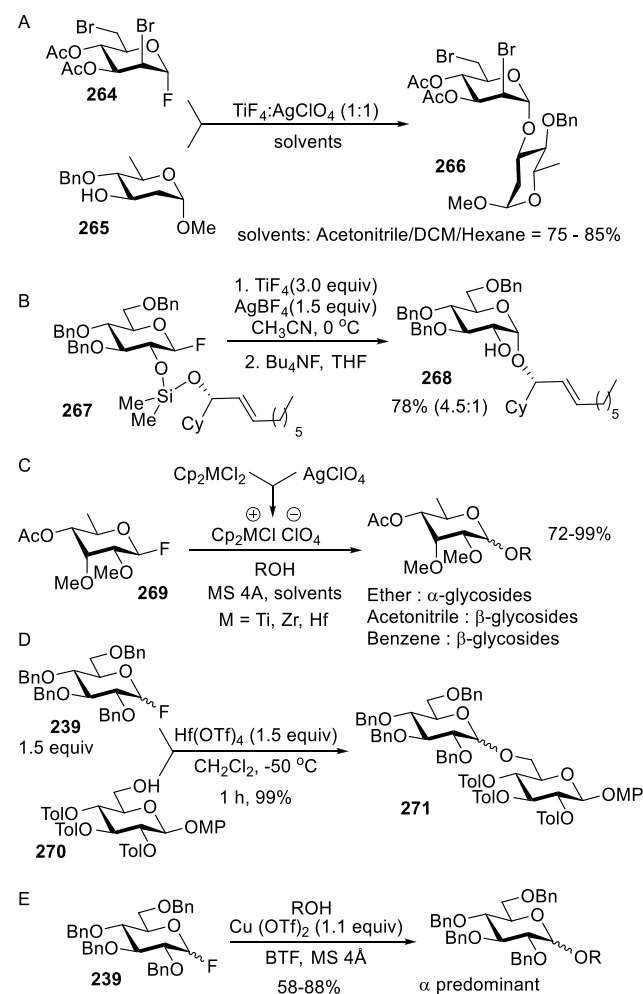
Glycosylations with unprotected sugars in aqueous solutions are challenging.^{499–501} Encouraged by the seminal work of Jencks^{502–504} and Bennett,^{490,505} Miller and co-workers described the activation of glycosyl fluorides in the aqueous phase to produce glycosides.⁴⁷⁰ After investigation of a number of metal salts and careful optimization of the reaction conditions, glycosidation of glycosyl fluoride **259** (6.0 equiv) with sucrose **260** (1.0 equiv, 0.3 M) was accomplished in the presence of $\text{Ca}(\text{OTf})_2$ (6.0 equiv) in aqueous trimethylamine to produce trisaccharide **261** in a good yield of 82% (Scheme 61B). The unique feature of this method is its β -stereoselectivity and regioselectivity for the 3'-position of sucrose. Extended glycosylation and NMR studies, similar to those described by Davies^{506–508} and by O'Leary,^{509–512} revealed that the unprecedented regioselectivity and reactivity of sucrose and its derivatives could be related to the hydrogen-bonding network. This network could have a significant impact on nucleophilicity of different hydroxyls of sugars and, hence, influence their interaction with Ca^{2+} during glycosylation.

In another report, Miller and co-workers extended the aqueous phase glycosidation of glycosyl fluorides to phenolic derivatives of biological relevance.⁴⁷¹ Glycosylation of amino acids in aqueous phase remains an immense task,^{513–518} and calcium(II)-mediated aqueous phase glycosylation in the presence of $\text{Ca}(\text{OH})_2$ worked very well. As depicted in Scheme 61C, glycosylation between glycosyl fluoride **259** (3.0 equiv) and *N*-Boc-protected tyrosine derivative **262** (1.0 equiv) 1.0 M in H_2O in the presence of $\text{Ca}(\text{OH})_2$ (3.0 equiv) produced glycoside **263** with complete β -stereoselectivity. The

concentration dependence of the reaction rate offered strong evidence of the $\text{S}_{\text{N}}2$ -like mechanism.^{109,503,505,519} Subsequently, the standardized glycosylation conditions were applied to various phenolic derivatives wherein electron-rich (less acidic) phenolic derivatives turned out to perform better compared to the electron-deficient ones.

6.2.4. Activation with Transition Metal Salts (Titanium, Zirconium, Hafnium, and Copper). Thiem and co-workers showed the activation of glycosyl fluorides with titanium fluoride (TiF_4).⁴⁵⁶ Per-acetylated, per-benzylated, and 2-deoxyglycosyl fluorides were investigated. These studies evolved into investigation of the cooperative catalyst comprising titanium and silver salts.⁴⁷² To enhance the stereoselectivity, the authors investigated brominated glycosyl fluoride donors depicted in Scheme 62A. Glycosidation of

Scheme 62. Glycosyl Fluoride Activation with Transition Metals Salts



mannosyl fluoride in the presence of TiF_4 and AgClO_4 produced α -disaccharides irrespective of the solvents employed. When mannosyl fluoride **264** was reacted with glycosyl acceptor **265** in acetonitrile, diethyl ether, or hexane α -disaccharide **266** was obtained in a good yield and exclusive stereoselectivity. Montgomery and co-workers reported facile construction of 1,2-*cis* glycosidic linkages by means of silicon-directed intramolecular aglycone delivery (IAD) method (*vide supra*) in the presence of TiF_4 and AgBF_4 .⁴⁷³ As depicted in Scheme 62B, silyl-tethered precursor **267** was subjected to

intramolecular glycosylation to give the corresponding glycoside **268** in 78% yield. Functional groups such as ketones, hydroxy groups, silyl ethers, and esters were shown to remain unaffected during these transformations.

Suzuki and co-workers discovered a new cooperative promoter system $\text{Cp}_2\text{MCl}_2\text{-AgClO}_4$ ($\text{M} = \text{Zr}, \text{Hf}, \text{Ti}$) for glycosyl fluoride donors, which enabled very promising glycosylations (Scheme 62C).⁴⁷⁴ A controlled experiment showed that Cp_2MCl_2 alone is practically inert in glycosylations, and it was suggested that Cp_2MCl_2 and AgClO_4 are required in molar equivalent to drive the reaction to completion. A rough order of reactivity of $\text{Zr} \geq \text{Hf} \gg \text{Ti}$ was estimated by glycosylation between glycosyl fluoride **269** and cyclohexyl methanol. Later, the authors explored the activation of anomeric fluoride of amino sugars with $\text{Cp}_2\text{MCl}_2\text{-AgClO}_4$ promoter systems.⁴⁷⁷ In this application, $\text{Cp}_2\text{HfCl}_2\text{-AgClO}_4$ was superior to its Zr counterpart. Subsequently, the cooperative catalyst $\text{Cp}_2\text{MCl}_2\text{-AgClO}_4$ ($\text{M} = \text{Zr}, \text{Hf}$) was utilized for the total synthesis of mycinamicin IV by the stereoselective glycosidation of D-mycinosyl and D-desosaminyl fluorides with mycinamicin VII.⁵²⁰

Further investigations revealed that $\text{Cp}_2\text{MCl}_2\text{-AgClO}_4$ ($\text{M} = \text{Hf}, \text{Zr}$) in a ratio of 1:2 is more efficient for the activation of glycosyl fluorides.^{478,521} The authors suggested that the activation of glycosyl fluorides depends on the fluorophilicity of early transition metals and their ability to form the reactive promoter shown in Scheme 62C. The driving force for the glycosylation reaction is believed to be due to the formation of a strong M-F bond. To enhance the efficiency of glycosylation of hindered alcohols, several silver salts (AgBF_4 , AgOTf , AgPF_6 , and AgSbF_6) in combination with Cp_2ZrCl_2 were examined, and AgBF_4 was found to be superior in this application, both in terms of reactivity and stereoselectivity.⁴⁷⁵

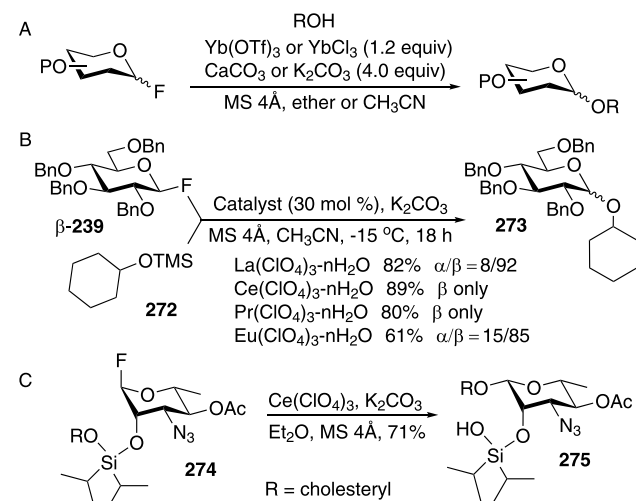
Matsumura and co-workers reported the stereocontrolled mannosylation in the presence of sulfated zirconia (SO_4/ZrO_2) and MS 5 Å.⁴⁷⁶ The advantage of solid acid-promoted glycosylation is that the acid can be recovered by filtration and then reused. Also, 2-deoxy- α -D-glucopyranosyl fluoride was investigated in the presence of sulfated zirconia.⁵²²

Ito and Manabe investigated hafnium(IV) reagents such as $\text{Hf}(\text{OTf})_4$, a well-known Lewis acid used for many synthetic transformations,^{523–526} for the activation of glycosyl fluorides.⁴⁷⁹ Optimization of the reaction conditions led to an effective glycosylation reaction between glycosyl donor **239** and acceptor **270** in CH_2Cl_2 to afford disaccharide **271** in 99% yield ($\alpha/\beta = 20:80$, Scheme 62D). Participating solvents were found to influence the stereochemistry of glycosylated products,^{404,527,528} a mixture of dioxane/toluene provided mainly α -glycosides, and β -selectivity was observed in acetonitrile. Glycosyl fluorides of other sugar series with different protection patterns were glycosylated with primary and secondary glycosyl acceptors under the identical reaction conditions and the corresponding glycosides were obtained in good to excellent yields (58–94%). The method was then applied for solid-phase oligosaccharide synthesis (*vide infra*).

Yamada and Hayashi reported activation of glycosyl fluorides in the presence of $\text{Cu}(\text{OTf})_2$.⁴⁸⁰ As depicted in Scheme 62E, glycosyl fluoride **239** was reacted with various alcohols in the presence of the stoichiometric amount of $\text{Cu}(\text{OTf})_2$ in benzonitrile at temperatures above 60 °C to afford the corresponding glycosides in good yields with predominant α -stereoselectivity.

6.2.5. Activation with Lanthanide Metal Salts (Lanthanum, Cerium, Ytterbium, etc.). Relying on a high dissociation energy of the Ln–F bond,^{529,530} Shibasaki and co-workers employed rare earth metal salts for the activation of glycosyl fluorides.⁴⁸¹ Ytterbium(III) triflate or chloride were found to be very effective for the activation of different glycosyl fluorides (Scheme 63A). The authors suggested that the

Scheme 63. Glycosyl Fluoride Activation with Lanthanide Metals Salts



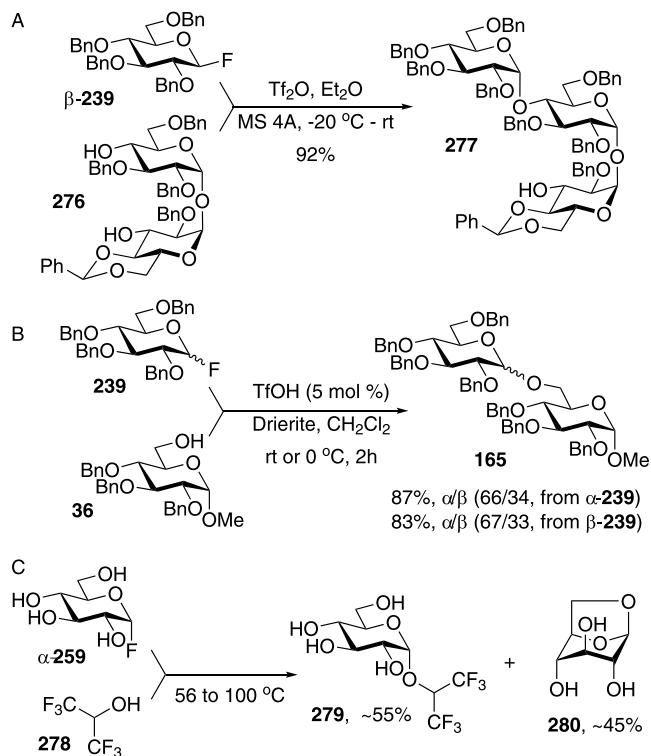
reaction proceeds through the formation of the oxacarbenium ion intermediate since the stereoselectivity of glycosides was reaction solvent dependent. Predominant 1,2-*trans* stereoselective glycosylation was observed in the presence of $\text{Yb}(\text{OTf})_3$ and K_2CO_3 in acetonitrile, whereas 1,2-*cis* stereoselectivity was achieved in a similar reaction conducted in the presence of $\text{Yb}(\text{OTf})_3$ and CaCO_3 in ether. The authors also showed that the addition of Lewis acid such as ZnCl_2 or $\text{Ba}(\text{ClO}_4)_2$ to ytterbium(III)-promoted glycosylation enhances the rate and improves the yield of glycosides.⁴⁸¹ Wang and co-workers immobilized lanthanide ions on ion exchange resins and utilized these catalysts in several chemical modifications.⁴⁸³ Glycosylation reaction between glycosyl fluorides and methanol was shown to be promoted by Yb-Amberlyst-15. Interestingly, complete inversion at the anomeric center was observed in these reactions. Inazu and co-workers showed the activation of glycosyl fluorides in the presence of catalytic amounts of ytterbium(III) tris[bis(perfluorobutylsulfonyl) amide] ($\text{Yb}[\text{N}(\text{O}_2\text{SC}_4\text{F}_9)_2]_3$, 5 mol %).⁴⁸⁴

Other rare earth metals were found to promote the glycosidation of glycosyl fluorides.^{481,482} Detailed investigation led to the development of a catalytic activation of glycosyl fluorides with trimethylsilyl ether acceptors.⁴⁸² Thus, coupling of glycosyl fluoride β -**239** with trimethylsilyl hexyl ether **272** in the presence of rare earth metal perchlorate hydrates in acetonitrile produced product **273** in good yields and with high β -stereoselectivity (Scheme 63B). Catalytic activation of glycosyl fluorides with rare earth metal perchlorates was attributed to the increased fluorophilicity of metal cations.^{531–534} Further, ^{19}F NMR experiments confirmed the formation of trimethylsilyl fluoride, which suggested the formation of oxacarbenium ion resulted from the interaction of naked metal cation (perchlorate) with anomeric fluoride which was then attacked by trimethylsilyl ether to produce

glycosides, trimethyl fluoride and regenerates metal perchlorates to run the catalytic cycle. It was also suggested that glycosylation occurred with a certain range of ionic radii, it was supported by the failed glycosylation by metal perchlorate hydrates of gadolinium, holmium, ytterbium, and yttrium. Differently protected glycosyl fluorides were couples with several alcohols in the presence of $\text{La}(\text{ClO}_4)_3 \cdot n\text{H}_2\text{O}$ to obtain the respective glycosides in good to excellent yields.⁵³⁵ Packard and Rychnovsky investigated the activation of anomeric fluorides in the presence of $\text{Ce}(\text{ClO}_4)_3$,⁴⁸⁵ wherein β -stereoselective glycosidation of anomeric fluoride derivative of D-mycosamine **274** was conducted via the IAD.^{536,537} This reaction afforded cholesteryl β -glycoside **275** in 71% yield as shown in Scheme 63C.

6.2.6. Activation with Anhydrides, Protic Acids, and Other Reagents. Wessel introduced triflic anhydride as a new activator for glycosyl fluorides.⁴⁶⁰ The author proposed that the activation of glycosyl fluorides occurred due to a higher affinity of CF_3SO_2^+ to fluorine than oxygen. As depicted in Scheme 64A, glycosidation of fluoride β -**239** with glycosyl

Scheme 64. Other Methods for the Activation of Glycosyl Fluorides



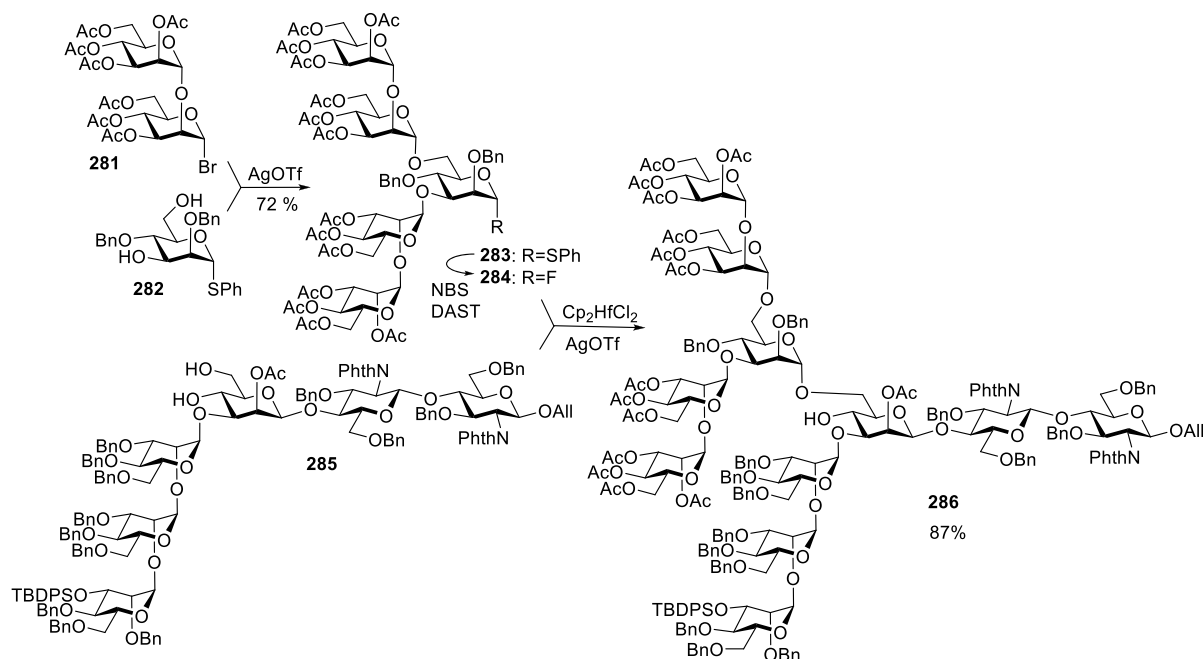
acceptor **276** afforded α -glycoside **277** in 92% yield. Further, this promoter was applied for the synthesis of differently linked disaccharides.⁵³⁸ Interestingly, it was discovered that low and moderately reactive alcohols successfully delivered products, whereas reactive alcohols provided fair yields due to the competing sulfonation of the hydroxyl group. Mukaiyama and co-workers discovered the activation of glycosyl fluorides with cat. trifluoromethanesulfonic acid (TfOH).^{465,486,487} Glycosidation of glycosyl fluoride α - or β -**239** with glycosyl acceptor **36** in dichloromethane in the presence of a catalytic amount of TfOH (5 mol %) produced disaccharide **165** within 2 h in a good yield (Scheme 64B).

TfOH -catalyzed activation was extended to reactions of differently protected glycosyl fluoride donors with various glycosyl acceptors. Without a nonparticipating group at C-2, the stereochemical outcome was difficult to predict. To address the problem associated with stereoselectivity, a thorough investigation for optimal reaction conditions was conducted. It was established that TfOH -catalyzed glycosylations produce good to excellent 1,2-*cis* selectivity in ethereal solvents in the presence of MS 5 Å or Drierite.⁴⁸⁸ Further investigation revealed that efficient activation of glycosyl fluoride donors can also be achieved in the presence of other protic acids such as HClO_4 , $\text{HOSO}_2\text{C}_4\text{F}_9$, HNTf_2 , HSbF_6 , and $\text{HB}(\text{C}_6\text{F}_5)_4$.⁴⁶⁴ Some of the strong acids were generated *in situ* in a similar way to the procedures described by Kevill⁵³⁹ or Kato.⁵⁴⁰ The diastereomeric ratio of glycoside products was found to be a subject of reaction conditions such as the nature of solvents, counteranions, and protecting groups.⁴⁹³ As previously discussed, Mukaiyama and co-workers performed the activation of glycosyl fluorides in the presence of catalytic carbocationic species paired with counterion such as tetrakis-(pentafluorophenyl)borate.⁴⁸⁹ Accordingly, trifluoromethanesulfonate and perchlorate were found to be effective in activating glycosyl fluoride donors.

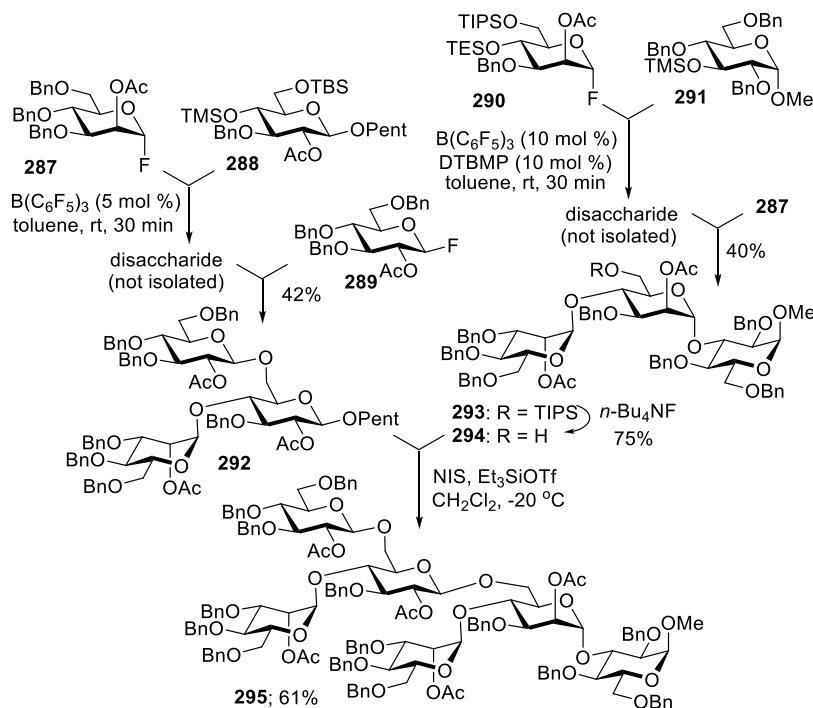
Chan and Bennett described the solvolysis of glycosyl fluorides in weak nucleophilic solvents such as hexafluoro-2-propanol **278** (HFIP).⁴⁹⁰ The formation of the retained solvolysis product, 1,1,1,3,3,3-hexafluoropropan-2-yl α -D-glucopyranoside **279**, and the inverted product, 1,6-anhydro- β -D-glucopyranose **280** from glucosyl fluoride α -**259** in HFIP or HFIP-*d* was observed (Scheme 64C). To understand the product distribution and pathway of the formation, the solvolysis of fluoride α -**259** in HFIP was conducted at different temperatures and the activation parameters were calculated from Eyring plot. The obtained results suggested that solvolysis occurs via a highly ordered transition state, which resulted from solvation of the fluoride ion by the HFIP solvent.⁵⁴¹ The expansive experimental and computed KIE measurements suggest solvolysis of fluorides α -**259** in HFIP solvent involves cleavage of the C–F bond as the rate-determining step, wherein proton transfer occurs from solvating HFIP molecule to the leaving group fluoride to afford the solvent-separated ion pair. Concurrently, the formation of solvolysis product **279** proceeds via the collapse of the solvent-separated ion pair in the $\text{S}_{\text{N}}1$ reaction, whereas the formation of inverted product **280** proceeds via the intramolecular capture of the solvent-equilibrated glycosyl cation generated from the dissociation of the solvent-separated ion pair. The authors also showed that an $\text{S}_{\text{N}}2$ -like nucleophilic displacement of anomeric fluoride of unprotected fluoride donor α -**259** with an aqueous solution of sodium azide produces corresponding β -glycosyl azide,^{505,541} wherein the anomeric center displayed that the reaction proceeds through a loose (exploded) transition state.⁵⁰⁵

6.3. Glycosyl Fluorides in Glycan and Glycoconjugate Synthesis

6.3.1. Convergent Building Block Assembly. Early examples of impressive convergent syntheses employing fluorides as glycosyl donors involve synthesis of α -cyclodextrin Ogawa and Takahashi wherein glycosylations were promoted with $\text{SnCl}_2 \cdot \text{AgOTf}$.⁵⁴² Nicolaou and co-workers employed the cooperative catalytic systems SnCl_2 -Ag salts and also Cp_2MgCl_2 -Ag salts for the total synthesis of the tumor-associated Le^x glycosphingolipids and sialyl dimeric Le^x.^{543,544} A representa-

Scheme 65. Convergent Assembly of High Mannose Type *N*-Glycan

Scheme 66. Borane-Catalyzed Convergent Oligosaccharides Synthesis



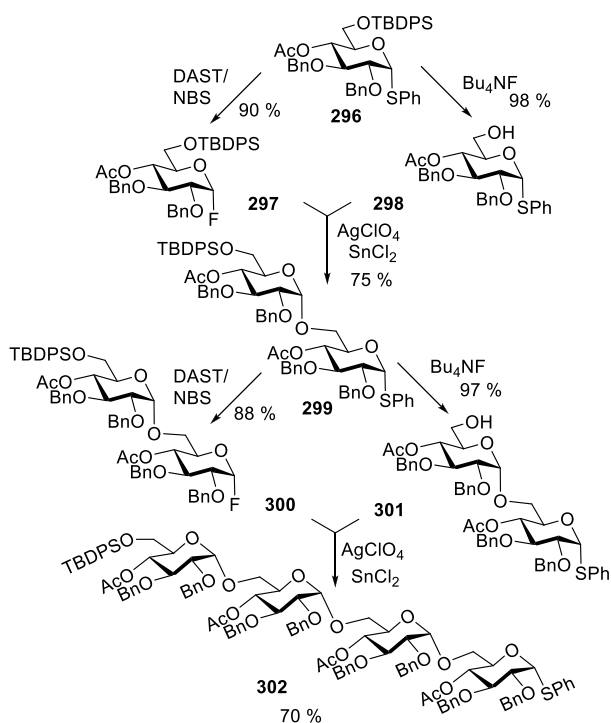
tive example of such an approach is the synthesis of the high mannose-type *N*-glycan **286** illustrated in Scheme 65.⁵⁴⁵ Glycosyl acceptor diol **282** was bis-glycosylated with the bromide donor **281** in the presence of AgOTf to afford the pentasaccharide **283** in 72% yield. The latter was then converted into a glycosyl fluoride donor **284** by reaction with DAST in the presence of NBS. The presynthesized hexasaccharide acceptor **285** was then regioselectively glycosylated with fluoride donor **284** in the presence of a $\text{Cp}_2\text{HfCl}_2/\text{AgOTf}$ promoter system to afford oligosaccharide **286** in an excellent yield of 87%.

A recent convergent assembly reported by Montgomery and co-workers comprises elements of one-pot synthesis.⁴⁶⁶ A three-component coupling was performed between monosaccharides **287**–**291** as shown in Scheme 66. The coupling of fluoride **287** with glycosyl acceptor **288** proceeded regioselectively at the C-4 position protected with TMS ether. Glycosyl fluoride donor **289** was then added to the reaction mixtures to produce the corresponding trisaccharide **292** in a good yield. In this reaction, TBS-protected C-6 position was glycosylated. Applying a similar approach for controlling the relative reactivity of silylated hydroxyl groups (TMS vs

triethylsilyl vs triisopropylsilyl) trisaccharide **293** was synthesized. For this purpose, mannosyl fluoride donor **290** was coupled with glucosyl acceptor **291** to produce the corresponding disaccharide that was then reacted with mannosyl donor **287**. Trisaccharide **293** was converted to glycosyl acceptor **294** by TIPS group removal with *n*-tetrabutylammonium fluoride (*n*-Bu₄NF). Finally, *n*-pentenyl leaving group of trisaccharide donor **292** was activated for reaction with acceptor **294** in the presence of NIS/Et₃SiOTf to afford hexasaccharide **295** in 61% yield.

6.3.2. Two-Step Activation. According to the two-stage activation approach, both glycosyl donor and glycosyl acceptor initially bear the same type of leaving group. However, to couple these two reactants, the LG of the potential glycosyl donor unit is first converted into a different (more reactive) LG, which is then selectively activated. This two-step activation sequence can be reiterated. This concept was discovered by Zen for *S*-ethyl and bromo groups³²⁹ and further expanded by Nicolaou for *S*-phenyl and fluoro groups.^{333,546} For a relevant example, see the synthesis of **302** depicted in Scheme 67.³³³ First thioglycoside **296** was converted into

Scheme 67. Two-Step Activation with Fluorides and Thioglycosides



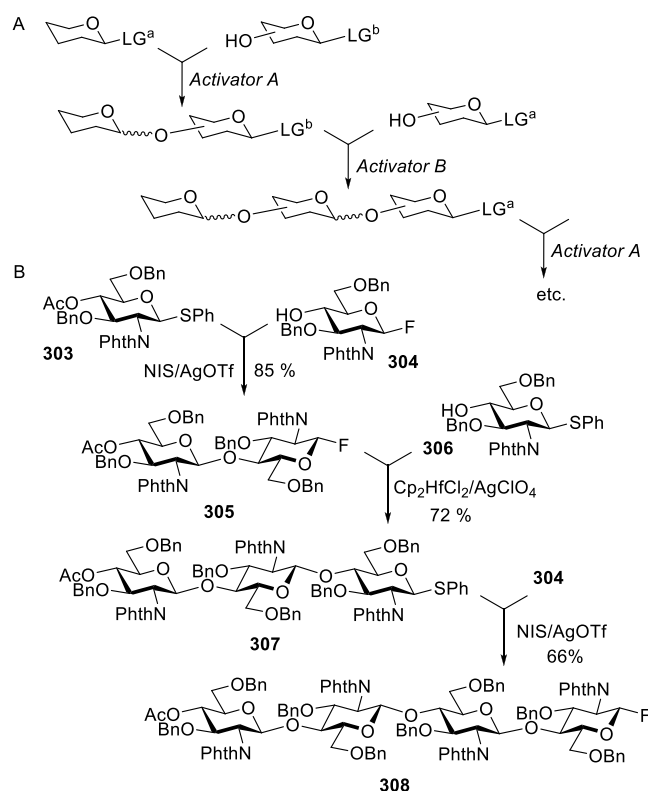
glycosyl fluoride **297** by the treatment with NBS/DAST. The latter was then coupled with a thioglycoside acceptor **298**, which was generated from **296** by the treatment with TBAF, in the presence of SnCl₂/AgClO₄ to provide disaccharide **299** in 75% yield. This sequence is then repeated to furnish the desired fluoride donor **300** and thioglycoside acceptor **301**, followed by glycosylation in a 2+2 fashion to provide tetrasaccharide **302**. This approach combines advantages offered by the two-step activation and convergent strategies.

6.3.3. Selective and Orthogonal Activation. Another general concept to expedite oligosaccharide synthesis is to achieve selective activation of different leaving groups. In a typical application, a glycosyl fluoride donor is activated over a

thioglycoside acceptor in the presence of a suitable promoter. The thio-leaving group of the resulting disaccharide is then activated directly to afford a trisaccharide. Examples of these applications include TfOH as the promoter for the first step, and NIS added for the second step.⁴⁸⁸ Another example involves HB(C₆F₅)₄ and NIS as activators for the activation of fluoride and thioglycoside, respectively.⁴⁶⁵ Demchenko and co-workers synthesized a hexasaccharide in only five steps via sequential selective activation of five leaving groups, including fluorides.⁵⁴⁷ As a part of their strategy, *S*-benzoxazolyl (SBox) trisaccharide donor was coupled with fluoride acceptor in the presence of MeOTf, and the resulting tetrasaccharide was then activated in the presence of AgClO₄/Cp₂ZrCl₂ to afford a pentasaccharide in 84% yield.

The combination of two chemically distinct glycosylation reactions, in which one of the leaving groups is activated while the other one remains intact, and *vice versa*, has led to the discovery of the orthogonal strategy for oligosaccharide synthesis.⁵⁴⁸ It requires the use of two orthogonal classes of glycosyl donors.^{549,550} As with the selective activation strategy, at the first step the glycosyl donor bearing LG^a is activated over the glycosyl acceptor bearing LG^b. Uniquely to the orthogonal strategy, the LG^b is then activated over LG^a of the new glycosyl acceptor (Scheme 68A). This activation sequence can then be

Scheme 68. Orthogonal Activation Was Discovered with Fluorides and Thioglycosides

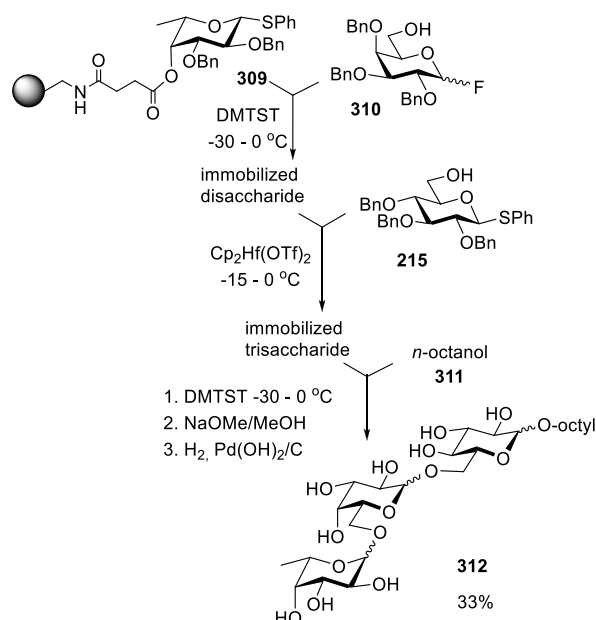


reiterated to give straightforward access to larger oligosaccharides. Specifically, this unique strategy was discovered with glycosyl fluorides, and the classic variation of the orthogonal activation route also involves building blocks bearing the *S*-phenyl leaving group. Thus, Ogawa and co-workers illustrated that phenylthio glycoside **303** can be selectively activated over glycosyl fluoride **304** in the presence of NIS/AgOTf to afford

disaccharide **305** (Scheme 68B).⁵⁴⁸ The fluoro leaving group of disaccharide donor **305** can then be activated over the SPh moiety of glycosyl acceptor **306** in the presence Cp_2HfCl_2 and AgClO_4 to afford trisaccharide **307** in 72% yield. Phenylthio glycosyl donor **307** was then activated over fluoride acceptor **304** to provide tetrasaccharide **308** in 66% yield. These reactions have been performed both in solution^{548,551} and on the polymer support (*vide infra*).^{551,552}

6.3.4. Polymer- and Tag-Supported Synthesis. Among major breakthroughs that have emerged in the area of synthetic chemistry is the development of organic synthesis on the solid phase.^{553–556} As a consequence, the past two decades have witnessed dramatic improvements in the area of solid phase-supported oligosaccharide synthesis,^{557–561} particularly in the context of automation.^{3,562–566} Polymer-supported synthesis allows for rapid synthesis of oligosaccharide sequences without the necessity of purifying and characterizing the intermediates. Another advantage of oligosaccharide synthesis on solid-phase support is the ease of excess reagent removal (by filtration). Among multiple methods and approaches that have been developed to date, two main strategies for solid-phase saccharide synthesis that differ in the type of the attachment can be identified. In the most common glycosyl acceptor bound approach excess of the glycosyl donor and promoter are present in the solution phase. In the rarer glycosyl donor-bound approach, glycosyl acceptor and promoters are present in the solution phase. Application of the orthogonal strategy is a rare example of a donor-bound approach in polymer supported synthesis.⁵⁵¹ As reported Kanie et al.,^{552,567} polymer-bound donor **309** was activated selectively over the solution-phase glycosyl fluoride acceptor **310** in the presence of dimethyl(methylthio)sulfonium trifluoromethanesulfonate (DMTST) shown in Scheme 69. The immobilized disaccharide fluoride was then activated over *S*-phenyl acceptor **215** in the presence of $\text{Cp}_2\text{Hf}(\text{OTf})_2$. Finally, the immobilized *S*-phenyl trisaccharide was glycosidated with octanol **311** in the presence of DMTST. The resulting oligosaccharide **312** was cleaved off the polymer support.⁵⁶⁷ The $\text{Hf}(\text{OTf})_4$ -promoted

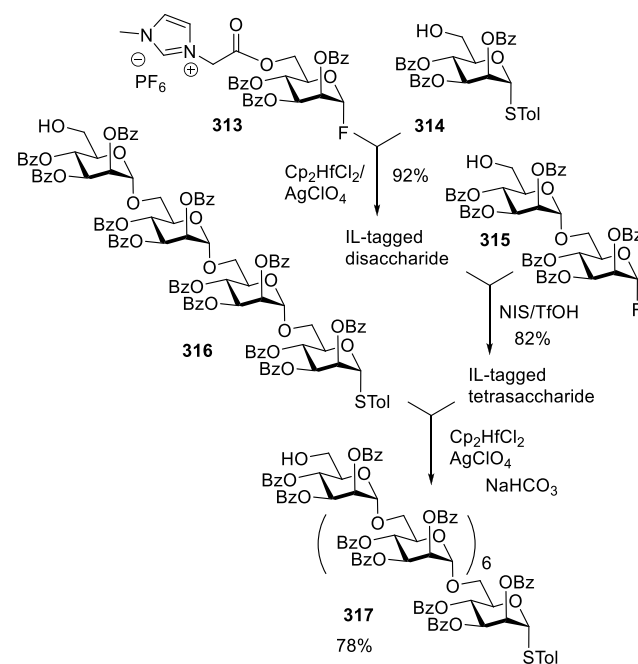
Scheme 69. Orthogonal Synthesis on Solid Phase



activation of solution phase mannosyl fluoride donor was applied to glycosylation of an acceptor immobilized on TentaGel.⁵⁶⁸

Another promising technique for supported oligosaccharide synthesis makes use of an ionic-liquid tag.^{569,570} Ionic liquid-supported assembly also expedites oligosaccharide synthesis by eliminating the need for chromatographic purification of the intermediates. After the desired reaction of the tagged compound has been completed, the reaction mixture is concentrated. The excess of organic reagents is removed by extraction with low polarity solvents in which the tagged compounds are insoluble. This approach is illustrated by a synthesis that incorporates elements of an orthogonal strategy making use of alternating activations of STol and F leaving groups and the convergent approach.^{571,572} A (1-methylimidazoliumhexafluorophospho) acetyl ionic liquid (IL) tag was introduced via the corresponding 6-chloroacetylated starting material by reaction with *N*-methylimidazole and potassium hexafluorophosphate. As depicted in Scheme 70, the tagged

Scheme 70. Ionic Liquid-Tagged Synthesis of Mannans

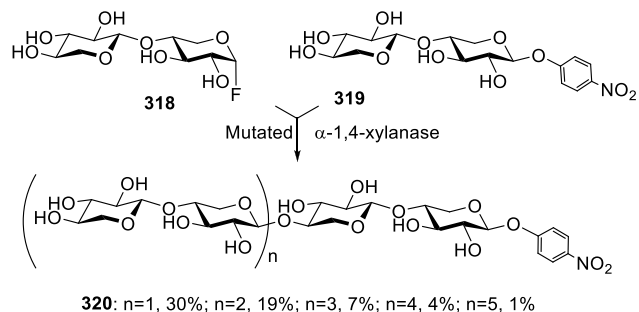


mannosyl fluoride donor **313** was glycosidated with thioglycoside acceptor **314** to afford the IL-tagged disaccharide. Meanwhile, disaccharide **315** was produced using tagged thioglycoside as the donor and a fluoride acceptor. Each disaccharide was split into portions, and the tag was removed from one portion. Coupling of the two disaccharides in the presence of NIS/TfOH produced a tagged tetrasaccharide. The latter was then coupled with tetrasaccharide **316**, which was prepared in a similar fashion. The resulting glycan was then cleaved off from the IL support to afford mannan **317**.

6.3.5. Oligosaccharide Synthesis with Hydrolases. In the natural environment, these enzymes hydrolyze glycosidic bonds and therefore are responsible for degradation of oligosaccharides. However, the reverse hydrolytic activity of hydrolases can also be exploited for the glycosidic bond-formation process.^{573,574} Glycosyl hydrolases are much more readily available than glycosyltransferases. They also are typically less regioselective, and the transformation yields are

lower. There are two main catalytic mechanisms for hydrolases: one leading to inversion of the anomeric configuration and the other leading to retention.^{575,576} The easiest way to employ this approach is to perform the glycosylation under thermodynamically controlled conditions, where the reverse hydrolysis is achieved at equilibrium, but the yields can be low. A significantly improved outcome can be achieved under kinetically controlled glycosylation conditions. In this case, activated glycosyl fluoride donors can be employed. Kinetically controlled glycosylations employing glycosidases (often called “transglycosylation” reactions) have been employed for the synthesis of a variety of oligosaccharides.⁵⁷⁴ As shown in Scheme 71, the application of glycosynthase, a synthetic

Scheme 71. Synthesis of Xylans 320 Using an Engineered Xylanase



enzyme derived from a retaining glycosidase, allowed irreversible glycosidation of glycosyl fluoride donor **318** with disaccharide acceptor **319**.⁵⁷⁷ Application of the glycosynthase derived from the β -1,4-xylanase of *Cellulomonas fimi* resulted in a very efficient synthesis of a series of xylans. The (1 $\rightarrow$ 4)-linked xylooligosaccharides **320**, ranging from tetra- to dodeca-saccharides, have been obtained regio- and stereoselectively in over 60% combined yield.

7. CONCLUSIONS AND OUTLOOK

To keep pace with the expanding areas of glycosciences, it is critical to make glycans more accessible to the general chemical, biomedical, and industrial audiences. The advancement of glycosylation methods and strategies and their broader adoption is crucial to meeting this need. While new developments are required both in the generalization of the strategies and in the optimization of methods for glycoside synthesis all existing methods of carbohydrate chemistry require further tuning of reactivity levels and reaction conditions. The goal of optimizing glycosylation has been pursued in many ways. This significantly enhanced our understanding and ability to refine the reaction conditions, suppress side reactions, understand stereoelectronics and conformation of the starting material and key reaction intermediates, and predict the outcomes of many glycosylations. Despite all these recent improvements, the challenge of glycosylation has remained, and many scientists have turned their attention to reinvestigating glycosyl halides. These simple donors are typically easily accessible from a variety of precursors, fairly stable, can be readily activated, and offer superior atom economy. Recent work has demonstrated that many glycosyl halides can be activated with less toxic promoters. These studies with glycosyl chlorides and bromides brought these glycosylation reactions to an entirely different

level of flexibility and versatility. Conversely, glycosyl fluorides, once prominent glycosyl donors, have been used more and more rarely in the past decade. The authors of this review believe that the full potential of glycosyl fluorides is yet to be revealed, and the glycosynthetic audience will witness further improvements of this promising method already in the next decade.

With further improvement of our synthetic capabilities, new effective approaches to stereoselective chemical glycosylation that will be broadly applicable to a wide range of substrates and synthetic targets will emerge. We anticipate that exploring glycosyl halide donors in combination with other methods and approaches will contribute significantly to the general methodological field by enhancing our ability to monitor all steps of the glycosylation and study the key reaction intermediates. This, in turn, will open exciting opportunities for studying and understanding general aspects of glycosylation, reveal and solve problem spots, and develop reliable strategies for troubleshooting and sidetracking, all of which will improve our ability to obtain glycosides with high stereo-control. As a result, the methodological advances made in the synthetic field will boost innovations in the related fields of glycosciences. It is our belief that a dedicated study of the methods and mechanisms of the glycosylation process will strategically advance the field such that both one-step glycosylation and multistep syntheses of glycans will be considered standard.

Many current methods for chemical glycosylation remain highly sophisticated, operationally complex, and require significant user know-how. By contrast, automation of glycosylation reactions showcases a highly accessible method of synthesis because it offers an idea of operational simplicity and reproducibility. At this stage it is still unclear whether a dedicated glycosylation reaction needs to be developed or existing methods used in solution can simply be repurposed to fulfill the requirements of automated synthesis. Upon achieving a reliable and simple platform for completely automated glycosylation, anybody should be able to perform automated synthesis of glycans. Machine-assisted synthesis will help to eliminate variability, and to accurately reproduce experiments multiple times by different users.^{3,200,562–564,566,578–596} Synthesis of glycans using user-friendly automated platforms will accelerate discovery of new carbohydrate-based or carbohydrate-containing diagnostics,^{597–612} pharmaceuticals,^{597,613–624} and vaccines.^{625–635} This will lead to innovations in many scientific disciplines and can significantly impact technology, society, the economy, and public health.

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Notes

The authors declare no competing financial interest.

Biographies

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Scott Geringer graduated from the Southern Illinois University–Edwardsville (SIUE) with a Bachelor of Science (B.S.) in Chemistry in 2013. He continued his education at SIUE and graduated with a Master of Science (M.Sc.) in Chemistry in 2015. He worked with Dr. Cristina De Meo. He completed his thesis studying the effects of the picoloyl protecting group on sialic acid glycosylation as well as studying the conformational equilibrium of the oxacarbenium ion intermediate. In August of 2015, Scott joined Dr. Alexei Demchenko's lab at University of Missouri–St. Louis (UMSL) as a Ph.D. student. He successfully graduated from UMSL in May of 2020 with a Doctor of Philosophy in Chemistry with an emphasis on Organic Chemistry. Here Scott worked on developing new catalytic reactions in carbohydrate chemistry. These methods include developing a method for catalytic glycosylation of glycosyl chlorides using iron(III) chloride, a new method for chemoselective picoloyl cleavage using catalytic iron(III) chloride, and activation of glycosyl chlorides using the cooperative silver(I) oxide-triflic acid catalyst system. Dr. Geringer currently works as a senior scientist at MilliporeSigma.

Alexei Demchenko graduated from the Mendeleev University of Chemical Technology of Russia with a Diploma (M.S.) in Chemical Engineering (1988) before joining the laboratory of the late Professor Kochetkov at the Zelinsky Institute of Organic Chemistry in Moscow. In 1993, he was awarded a Ph.D. in Organic Chemistry by the Russian Academy of Sciences for his work on the development of thiocyanate methodology for glycosylation. After two postdoctoral years under Kochetkov, he joined Professor Boons' group at the University of Birmingham (UK) as a BBSRC postdoctoral research fellow. In 1998, he moved to the Complex Carbohydrate Research Center, University of Georgia (USA) as a research associate. In 2001, he joined the

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REFERENCES

- (1) Hehre, E. J. Glycosyl transfer: a history of the concept's development and view of its major contributions to biochemistry. *Carbohydr. Res.* **2001**, *331*, 347–368.
- (2) Muthana, S.; Cao, H.; Chen, X. Recent progress in chemical and chemoenzymatic synthesis of carbohydrates. *Curr. Opin. Chem. Biol.* **2009**, *13*, 573–581.
- (3) Panza, M.; Pistorio, S. G.; Stine, K. J.; Demchenko, A. V. Automated chemical oligosaccharide synthesis: novel approach to traditional challenges. *Chem. Rev.* **2018**, *118*, 8105–8150.
- (4) Michael, A. On the synthesis of helicin and phenolglucoside. *Am. Chem. J.* **1879**, *1*, 305–312.
- (5) Fischer, E. Über die glucoside der alkohole. *Ber. Dtsch. Chem. Ges.* **1893**, *26*, 2400–2412.
- (6) Fischer, E.; Armstrong, E. F. Over isomers of the aceto-halogen derivative of grape sugar and the synthesis of the glucosides. *Ber. Dtsch. Chem. Ges.* **1901**, *34*, 2885–2900.
- (7) Koenigs, W.; Knorr, E. Über einige derivate des traubenzuckers und der galactose. *Ber. Dtsch. Chem. Ges.* **1901**, *34*, 957–981.
- (8) Mukaiyama, T.; Murai, Y.; Shoda, S.-i. An efficient method for glucosylation of hydroxy compounds using glucopyranosyl fluoride. *Chem. Lett.* **1981**, *10*, 431–432.
- (9) Gervay, J.; Hadd, M. J. Anionic additions to glycosyl iodides: highly stereoselective syntheses of β -C-, N-, and O-glycosides. *J. Org. Chem.* **1997**, *62*, 6961–6967.
- (10) Helferich, B.; Schmitz-Hillebrecht, E. A new method for the synthesis of glucosides of the phenols. *Ber. Dtsch. Chem. Ges.* **1933**, *66B*, 378–383.
- (11) Boursier, M.; Descotes, G. Activation of the anomeric carbon of sugars by the carbonate group and applications in glycosidic synthesis. *C. R. Acad. Sci. Ser.* **2 1989**, *308*, 919–921.
- (12) Koide, K.; Ohno, M.; Kobayashi, S. A new glycosylation reaction based on a "remote activation concept": glycosyl 2-pyridinecarboxylate as a novel glycosyl donor. *Tetrahedron Lett.* **1991**, *32*, 7065–7068.
- (13) Kunz, H.; Zimmer, J. Glycoside synthesis via electrophile-induced activation of N-allyl carbamates. *Tetrahedron Lett.* **1993**, *34*, 2907–2910.
- (14) Ferrier, R. J.; Hay, R. W.; Vethaviasar, N. A potentially versatile synthesis of glycosides. *Carbohydr. Res.* **1973**, *27*, 55–61.
- (15) Nicolaou, K. C.; Seitz, S. P.; Papahatjis, D. P. A mild and general method for the synthesis of O-glycosides. *J. Am. Chem. Soc.* **1983**, *105*, 2430–2434.
- (16) Garegg, P. J.; Henrichson, C.; Norberg, T. A reinvestigation of glycosidation reactions using 1-thioglycosides as glycosyl donors and thiophilic cations as promoters. *Carbohydr. Res.* **1983**, *116*, 162–165.
- (17) Oscarson, S. General synthetic methods: Reactions at the anomeric center: S-glycosylation. In *Glycoscience: Chemistry and*

Chemical Biology; Fraser-Reid, B.; Tatsuta, K.; Thiem, J., Eds.; Springer: Berlin - Heidelberg - New York, 2001; Vol. 1; pp 643–671.

(18) Bochkov, A. F.; Kochetkov, N. K. A new approach for the synthesis of oligosaccharides. *Carbohydr. Res.* **1975**, *39*, 355–357.

(19) Kochetkov, N. K.; Backinowsky, L. V.; Tsvetkov, Y. E. Sugar thio orthoesters as glycosylating agents. *Tetrahedron Lett.* **1977**, *18*, 3681–3684.

(20) Pougny, J. R.; Jacquinet, J. C.; Nassr, M.; Duchet, D.; Milat, M. L.; Sinay, P. A novel synthesis of 1,2-cis-disaccharides. *J. Am. Chem. Soc.* **1977**, *99*, 6762–6763.

(21) Schmidt, R. R.; Michel, J. Facile synthesis of α - and β -O-glycosyl imidates; Preparation of glycosides and disaccharides. *Angew. Chem., Int. Ed. Engl.* **1980**, *19*, 731–732.

(22) Adinolfi, M.; Barone, G.; Iadonisi, A.; Schiattarella, M. Iodine/triethylsilane as a convenient promoter system for the activation of disarmed glycosyl trichloro- and N-(phenyl)trifluoroacetimidates. *Synlett* **2002**, *2002*, 269–270.

(23) Adinolfi, M.; Barone, G.; Iadonisi, A.; Schiattarella, M. Efficient activation of glycosyl N-(phenyl)trifluoroacetimidate donors with ytterbium(III) triflate in the glycosylation reaction. *Tetrahedron Lett.* **2002**, *43*, 5573–5577.

(24) Mukaiyama, T.; Nakatsuka, T.; Shoda, S. I. An efficient glycosylation of alcohol using 1-thioglycoside derivative. *Chem. Lett.* **1979**, *8*, 487–490.

(25) Hanessian, S.; Bacquet, C.; Lehong, N. Chemistry of the glycosidic linkage. Exceptionally fast and efficient formation of glycosides by remote activation. *Carbohydr. Res.* **1980**, *80*, c17–c22.

(26) Woodward, R. B.; Logusch, E.; Nambiar, K. P.; Sakan, K.; Ward, D. E.; Au-Yeung, B. W.; Balaran, P.; Browne, L. J.; Card, P. J.; Chen, C. H. Asymmetric total synthesis of erythromycin. 3. Total synthesis of erythromycin. *J. Am. Chem. Soc.* **1981**, *103*, 3215–3217.

(27) Demchenko, A. V.; Malysheva, N. N.; De Meo, C. S-Benzoxazolyl (SBox) glycosides as novel, versatile glycosyl donors for stereoselective 1,2-cis glycosylation. *Org. Lett.* **2003**, *5*, 455–458.

(28) Demchenko, A. V.; Kamat, M. N.; De Meo, C. S-Benzoxazolyl (SBox) glycosides in oligosaccharide synthesis: novel glycosylation approach to the synthesis of β -D-glucosides, β -D-galactosides, and α -D-mannosides. *Synlett* **2003**, *2003*, 1287–1290.

(29) Demchenko, A. V.; Pornsuriyasak, P.; De Meo, C.; Malysheva, N. N. Potent, versatile, and stable: thiazolyl thioglycosides as glycosyl donors. *Angew. Chem., Int. Ed.* **2004**, *43*, 3069–3072.

(30) Fraser-Reid, B.; Konradsson, P.; Mootoo, D. R.; Udodong, U. Direct elaboration of pent-4-enyl glycosides into disaccharides. *J. Chem. Soc., Chem. Commun.* **1988**, 823–825.

(31) Marra, A.; Esnault, J.; Veyrieres, A.; Sinay, P. Isopropenyl glycosides and congeners as novel classes of glycosyl donors: theme and variations. *J. Am. Chem. Soc.* **1992**, *114*, 6354–6360.

(32) Boons, G. J.; Isles, S. Vinyl glycosides in oligosaccharide synthesis. Part 1: A new latent-active glycosylation strategy. *Tetrahedron Lett.* **1994**, *35*, 3593–3596.

(33) Brown, D. S.; Ley, S. V.; Vile, S. Preparation of cyclic ether acetals from 2-benzenesulphonyl derivatives: a new mild glycosidation procedure. *Tetrahedron Lett.* **1988**, *29*, 4873–4876.

(34) Kochetkov, N. K.; Klimov, E. M.; Malysheva, N. N. Novel highly stereospecific method of 1,2-cis-glycosylation. Synthesis of α -D-glycosyl-D-glucoses. *Tetrahedron Lett.* **1989**, *30*, 5459–5462.

(35) Briner, K.; Vasella, A. Glycosylidene carbenes a new approach to glycoside synthesis. Part 1. Preparation of glycosylidene-derived diaziridines and diazirines. *Helv. Chim. Acta* **1989**, *72*, 1371–1382.

(36) Friesen, R. W.; Danishefsky, S. J. On the controlled oxidative coupling of glycals: A new strategy for the rapid assembly of oligosaccharides. *J. Am. Chem. Soc.* **1989**, *111*, 6656–6660.

(37) Halcomb, R. L.; Danishefsky, S. J. On the direct epoxidation of glycals: application of a reiterative strategy for the synthesis of β -linked oligosaccharides. *J. Am. Chem. Soc.* **1989**, *111*, 6661–6666.

(38) Balmond, E. I.; Coe, D. M.; Galan, M. C.; McGarrigle, E. M. α -Selective organocatalytic synthesis of 2-deoxygalactosides. *Angew. Chem., Int. Ed.* **2012**, *51*, 9152–9155.

(39) Palo-Nieto, C.; Sau, A.; Galan, M. C. Gold(I)-catalyzed direct stereoselective synthesis of deoxyglycosides from glycals. *J. Am. Chem. Soc.* **2017**, *139*, 14041–14044.

(40) Kinfe, H. H. Versatility of glycals in synthetic organic chemistry: coupling reactions, diversity oriented synthesis and natural product synthesis. *Org. Biomol. Chem.* **2019**, *17*, 4153–4182.

(41) Kahne, D.; Walker, S.; Cheng, Y.; van Engen, D. Glycosylation of unreactive substrates. *J. Am. Chem. Soc.* **1989**, *111*, 6881–6882.

(42) Marra, A.; Sinay, P. A novel stereoselective synthesis of N-acetyl- α -neuraminosyl-galactose disaccharide derivatives, using anomeric S-glycosyl xanthates. *Carbohydr. Res.* **1990**, *195*, 303–308.

(43) Mehta, S.; Pinto, B. M. Phenylselenoglycosides as novel, versatile glycosyl donors. selective activation over thioglycosides. *Tetrahedron Lett.* **1991**, *32*, 4435–4438.

(44) Kondo, H.; Ichikawa, Y.; Wong, C. H. β -Sialyl phosphite and phosphoramidite: synthesis and application to the chemoenzymatic synthesis of CMP-sialic acid and sialyl oligosaccharides. *J. Am. Chem. Soc.* **1992**, *114*, 8748–8750.

(45) Martin, T. J.; Schmidt, R. R. Efficient sialylation with phosphite as leaving group. *Tetrahedron Lett.* **1992**, *33*, 6123–6126.

(46) Stick, R. V.; Tilbrook, D. M. G.; Williams, S. J. The selective activation of telluro- over seleno- β -D-glucopyranosides as glycosyl donors: a reactivity scale for various telluro, seleno and thio sugars. *Aust. J. Chem.* **1997**, *50*, 237–240.

(47) Hinklin, R. J.; Kiessling, L. L. Glycosyl sulfonylcarbamates: new glycosyl donor with tunable reactivity. *J. Am. Chem. Soc.* **2001**, *123*, 3379–3380.

(48) Huchel, U.; Schmidt, C.; Schmidt, R. R. Synthesis of hetaryl glycosides and their glycosyl donor properties. *Eur. J. Org. Chem.* **1998**, *1998*, 1353–1360 and references therein.

(49) Plante, O. J.; Andrade, R. B.; Seeberger, P. H. Synthesis and use of glycosyl phosphates as glycosyl donors. *Org. Lett.* **1999**, *1*, 211–214.

(50) Davis, B. G.; Ward, S. J.; Rendle, P. M. Glycosyldisulfides: a new class of solution and solid phase glycosyl donors. *Chem. Commun.* **2001**, 189–190.

(51) Kim, K. S.; Kim, J. H.; Lee, Y. J.; Lee, Y. J.; Park, J. 2-(Hydroxycarbonyl)benzyl glycosides: a novel type of glycosyl donors for highly efficient β -mannopyranosylation and oligosaccharide synthesis by latent-active glycosylation. *J. Am. Chem. Soc.* **2001**, *123*, 8477–8481.

(52) Li, Y.; Yang, Y.; Yu, B. An efficient glycosylation protocol with glycosyl ortho-alkynylbenzoates as donors under the catalysis of Ph₃PAuOTf. *Tetrahedron Lett.* **2008**, *49*, 3604–3608.

(53) Mamidyala, S. K.; Finn, M. G. Glycosylation using unprotected alkynyl donors. *J. Org. Chem.* **2009**, *74*, 8417–8420.

(54) Li, Y.; Yang, X.; Liu, Y.; Zhu, C.; Yang, Y.; Yu, B. Gold(I)-catalyzed glycosylation with glycosyl ortho-alkynylbenzoates as donors: general scope and application in the synthesis of a cyclic triterpene saponin. *Chem.—Eur. J.* **2010**, *16*, 1871–1882.

(55) Yang, W.; Sun, J.; Lu, W.; Li, Y.; Shan, L.; Han, W.; Zhang, W.; Yu, B. Synthesis of kaempferol 3-O-(3",6"-di-O-E-p-coumaroyl)- β -D-glucopyranoside, efficient glycosylation of flavonol 3-OH with glycosyl o-alkynylbenzoates as donors. *J. Org. Chem.* **2010**, *75*, 6879–6888.

(56) Li, Y.; Sun, J.; Yu, B. Efficient synthesis of lupane-type saponins via gold(I)-catalyzed glycosylation with glycosyl ortho-alkynylbenzoates as donors. *Org. Lett.* **2011**, *13*, 5508–5511.

(57) Ma, Y.; Li, Z.; Shi, H.; Zhang, J.; Yu, B. Assembly of Digitoxin by gold(I)-catalyzed glycosidation of glycosyl o-alkynylbenzoates. *J. Org. Chem.* **2011**, *76*, 9748–9756.

(58) Zhu, Y.; Yu, B. Characterization of the Isochromen-4-yl-gold(I) intermediate in the gold(I)-catalyzed glycosidation of glycosyl ortho-alkynylbenzoates and enhancement of the catalytic efficiency thereof. *Angew. Chem., Int. Ed.* **2011**, *50*, 8329–8332.

(59) Yang, F.; Wang, Q.; Yu, B. ortho-Alkynylphenyl thioglycosides as a new type of glycosylation donors under the catalysis of Au(I) complexes. *Tetrahedron Lett.* **2012**, *53*, 5231–5234.

- (60) Tang, Y.; Li, J.; Zhu, Y.; Li, Y.; Yu, B. Mechanistic insights into the gold(I)-catalyzed activation of glycosyl ortho-alkynylbenzoates for glycosylation. *J. Am. Chem. Soc.* **2013**, *135*, 18396–18405.
- (61) Mishra, B.; Neralkar, M.; Hotha, S. Stable alkynyl glycosyl carbonates: catalytic anomeric activation and synthesis of a tridecasaccharide reminiscent of *Mycobacterium tuberculosis* cell wall lipoarabinomannan. *Angew. Chem., Int. Ed.* **2016**, *128*, 7917–7922.
- (62) Nukada, T.; Berces, A.; Whitfield, D. M. Can the stereochemical outcome of glycosylation reactions be controlled by the conformational preferences of the glycosyl donor? *Carbohydr. Res.* **2002**, *337*, 765–774.
- (63) Ranade, S. C.; Demchenko, A. V. Mechanism of chemical glycosylation: focus on the mode of activation and departure of anomeric leaving groups. *J. Carbohydr. Chem.* **2013**, *32*, 1–43.
- (64) Kochetkov, N. K.; Klimov, E. M.; Malysheva, N. N.; Demchenko, A. V. A new stereospecific method for 1,2-cis-glycosylation. *Carbohydr. Res.* **1991**, *212*, 77–91.
- (65) Kochetkov, N. K.; Klimov, E. M.; Malysheva, N. N.; Demchenko, A. V. Stereospecific 1,2-cis-glycosylation: a modified thiocyanate method. *Carbohydr. Res.* **1992**, *232*, C1–C5.
- (66) Cato, D.; Buskas, T.; Boons, G.-J. Highly efficient stereospecific preparation of Tn and TF building blocks using thioglycosyl donors and the Ph₂SO/Tf₂O promotor system. *J. Carbohydr. Chem.* **2005**, *24*, 503–516.
- (67) Mydock, L. K.; Kamat, M. N.; Demchenko, A. V. Direct synthesis of diastereomerically pure glycosyl sulfonium salts. *Org. Lett.* **2011**, *13*, 2928–2931.
- (68) Hoang, K. L. M.; Liu, X.-W. The intriguing dual-directing effect of 2-cyanobenzyl ether for a highly stereospecific glycosylation reaction. *Nat. Commun.* **2014**, *5*, 5051.
- (69) Issa, J. P.; Bennett, C. S. A reagent-controlled SN₂-glycosylation for the direct synthesis of beta-linked 2-deoxy-sugars. *J. Am. Chem. Soc.* **2014**, *136*, 5740–5744.
- (70) Park, Y.; Harper, K. C.; Kuhl, N.; Kwan, E. E.; Liu, R. Y.; Jacobsen, E. N. Macrocyclic bis-thioureas catalyze stereospecific glycosylation reactions. *Science* **2017**, *355*, 162–166.
- (71) Adero, P. O.; Amarasekara, H.; Wen, P.; Bohe, L.; Crich, D. The experimental evidence in support of glycosylation mechanisms at the SN₁-SN₂ Interface. *Chem. Rev.* **2018**, *118*, 8242–8284.
- (72) Eby, R.; Schuerch, C. The use of 1-O-tosyl-D-glucopyranose derivatives in α -D-glucoside synthesis. *Carbohydr. Res.* **1974**, *34*, 79–90 and references therein.
- (73) Marousek, V.; Lucas, T. J.; Wheat, P. E.; Schuerch, C. The influence of reactant structure and solvent on galactoside synthesis from galactosyl sulfonates. *Carbohydr. Res.* **1978**, *60*, 85–96.
- (74) Crich, D.; Sun, S. Are glycosyl triflates intermediates in the sulfoxide glycosylation method? a chemical and ¹H, ¹³C, and ¹⁹F NMR spectroscopic investigation. *J. Am. Chem. Soc.* **1997**, *119*, 11217–11223.
- (75) Crich, D.; Sun, S. Direct chemical synthesis of β -mannopyranosides and other glycosides via glycosyl triflates. *Tetrahedron* **1998**, *54*, 8321–8348.
- (76) Crich, D. Chemistry of glycosyl triflates: synthesis of β -mannopyranosides (Reprinted from *Glycochemistry: Principles, Synthesis, and Applications*, pp 53–75, 2001). *J. Carbohydr. Chem.* **2002**, *21*, 667–690.
- (77) Nokami, T.; Shibuya, A.; Tsuyama, H.; Suga, S.; Bowers, A. A.; Crich, D.; Yoshida, J. Electrochemical generation of glycosyl triflate pools. *J. Am. Chem. Soc.* **2007**, *129*, 10922–10928.
- (78) Garcia, B. A.; Poole, J. L.; Gin, D. Y. Direct glycosylations with 1-hydroxy glycosyl donors using trifluoromethanesulfonic anhydride and diphenyl sulfoxide. *J. Am. Chem. Soc.* **1997**, *119*, 7597–7598.
- (79) Garcia, B. A.; Gin, D. Y. Dehydrative glycosylation with activated diphenyl sulfonium reagents. Scope, mode of C(1)-hemiacetal activation, and detection of reactive glycosyl intermediates. *J. Am. Chem. Soc.* **2000**, *122*, 4269–4279.
- (80) Nguyen, H. M.; Chen, Y. N.; Duron, S. G.; Gin, D. Y. Sulfide-mediated dehydrative glycosylation. *J. Am. Chem. Soc.* **2001**, *123*, 8766–8772.
- (81) Gin, D. Dehydrative glycosylation with 1-hydroxy donors (reprinted from *Glycochemistry: Principles, Synthesis, and Applications*, pg 33–52, 2001). *J. Carbohydr. Chem.* **2002**, *21*, 645–665.
- (82) Boebel, T. A.; Gin, D. Y. Probing the mechanism of sulfoxide-catalyzed hemiacetal activation in dehydrative glycosylation. *J. Org. Chem.* **2005**, *70*, 5818–5826.
- (83) Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. Stereochemical reversal of nucleophilic substitution reactions depending upon substituent: reactions of heteroatom-substituted six-membered-ring oxocarbenium ions through pseudoaxial conformers. *J. Am. Chem. Soc.* **2000**, *122*, 168–169.
- (84) Ayala, L.; Lucero, C. G.; Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. Stereochemistry of nucleophilic substitution reactions depending upon substituent: evidence for electrostatic stabilization of pseudoaxial conformers of oxocarbenium ions by heteroatom substitution. *J. Am. Chem. Soc.* **2003**, *125*, 15521–15528.
- (85) Shenoy, S. R.; Woerpel, K. A. Investigations into the role of ion pairing in reactions of heteroatom-substituted cyclic oxocarbenium ions. *Org. Lett.* **2005**, *7*, 1157–1160.
- (86) Billings, S. B.; Woerpel, K. A. Nucleophilic substitution reactions of sulfur-substituted cyclohexanone acetals: an analysis of the factors controlling stereoselectivity. *J. Org. Chem.* **2006**, *71*, 5171–5178.
- (87) Beaver, M. G.; Billings, S. B.; Woerpel, K. A. C-Glycosylation reactions of sulfur-substituted glycosyl donors: Evidence against the role of neighboring-group participation. *J. Am. Chem. Soc.* **2008**, *130*, 2082–2086.
- (88) Chu, A.-H. A.; Nguyen, S. H.; Sisel, J. A.; Minciunescu, A.; Bennett, C. S. Selective synthesis of 1,2-cis- α -glycosides without directing groups. Application to iterative oligosaccharide synthesis. *Org. Lett.* **2013**, *15*, 2566–2569.
- (89) Soliman, S. E.; Bennett, C. S. Reagent-controlled synthesis of the branched trisaccharide fragment of the antibiotic Saccharomicin B. *Org. Lett.* **2018**, *20*, 3413–3417.
- (90) Frihed, T. G.; Bols, M.; Pedersen, C. M. Mechanisms of glycosylation reactions studied by low-temperature nuclear magnetic resonance. *Chem. Rev.* **2015**, *115*, 4963–5013.
- (91) Whitfield, D. M. Computational studies of the role of glycopyranosyl oxocarbenium ions in glycobiology and glycochemistry. *Adv. Carbohydr. Chem. Biochem.* **2009**, *62*, 83–159.
- (92) Paulsen, H. Selectivity and reactivity in oligosaccharide synthesis. In *Selectivity - a Goal for Synthetic Efficiency*; Bartmann, W.; Trost, B. M., Eds.; Verlag Chemie: Weinheim, 1984; pp 169–190.
- (93) Uriel, C.; Gomez, A. M.; Lopez, J. C.; Fraser-Reid, B. Reciprocal donor-acceptor selectivity: the influence of the donor O-2 substituent in the regioselective mannosylation of myo-inositol orthopentanoate. *Eur. J. Org. Chem.* **2009**, *2009*, 403–411.
- (94) Jayaprakash, K. N.; Lu, J.; Fraser-Reid, B. Synthesis of a lipomannan component of the cell-wall complex of *Mycobacterium tuberculosis* is based on Paulsen's concept of donor/acceptor "match". *Angew. Chem., Int. Ed.* **2005**, *44*, 5894–5898.
- (95) Uriel, C.; Gomez, A. M.; Lopez, J. C.; Fraser-Reid, B. Further insight into 'matching' of donors and acceptors via reciprocal donor acceptor selectivity (RDAS) studies. *Synlett* **2003**, *2003*, 2203–2207.
- (96) Mach, M.; Schlueter, U.; Mathew, F.; Fraser-Reid, B.; Hazen, K. C. Comparing n-pentenyl orthoesters and n-pentenyl glycosides as alternative glycosyl donors. *Tetrahedron* **2002**, *58*, 7345–7354.
- (97) Fraser-Reid, B.; Lopez, J. C.; Radhakrishnan, K. V.; Nandakumar, M. V.; Gomez, A. M.; Uriel, C. One pot/two donors /one diol give one differentiated trisaccharide: powerful evidence for reciprocal donor-acceptor selectivity (RDAS). *Chem. Commun.* **2002**, 2104–2105.
- (98) Spijker, N. M.; van Boeckel, C. A. A. Double stereo-differentiation in carbohydrate coupling reactions: the mismatched Interaction of donor and deceptor as an unprecedented factor

governing the α/β -ratio of glycoside formation. *Angew. Chem., Int. Ed. Engl.* **1991**, *30*, 180–183.

(99) Zhang, Q.; van Rijssel, E. R.; Walvoort, M. T. C.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Acceptor Reactivity in the Total Synthesis of Alginate Fragments Containing α -L-Guluronic Acid and β -D-Mannuronic Acid. *Angew. Chem. Int. Ed.* **2015**, *54*, 7670–7673.

(100) van der Vorm, S.; Hansen, T.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. The influence of acceptor nucleophilicity on the glycosylation reaction mechanism. *Chem. Sci.* **2017**, *8*, 1867–1875.

(101) van der Vorm, S.; van Hengst, J. M. A.; Bakker, M.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Mapping the relationship between glycosyl acceptor reactivity and glycosylation stereoselectivity. *Angew. Chem., Int. Ed. Engl.* **2018**, *57*, 8240–8244.

(102) van der Vorm, S.; Hansen, T.; van Hengst, J. M. A.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Acceptor reactivity in glycosylation reactions. *Chem. Soc. Rev.* **2019**, *48*, 4688–4706.

(103) Crich, D.; Chandrasekera, N. S. Mechanism of 4,6-O-benzylidene-directed β -mannosylation as determined by α -deuterium kinetic isotope effects. *Angew. Chem., Int. Ed.* **2004**, *43*, 5386–5389 and references therein.

(104) El-Badri, M. H.; Willenbring, D.; Tantillo, D. J.; Gervay-Hague, J. Mechanistic Studies on the Stereoselective Formation of beta-Mannosides from Mannosyl Iodides Using α -Deuterium Kinetic Isotope Effects. *J. Org. Chem.* **2007**, *72*, 4663–4672.

(105) Huang, M.; Garrett, G. E.; Birlirakis, N.; Bohe, L.; Pratt, D. A.; Crich, D. Dissecting the mechanisms of a class of chemical glycosylation using primary ^{13}C kinetic isotope effects. *Nat. Chem.* **2012**, *4*, 663–667.

(106) Adero, P. O.; Furukawa, T.; Huang, M.; Mukherjee, D.; Retailleau, P.; Bohe, L.; Crich, D. Cation clock reactions for the determination of relative reaction kinetics in glycosylation reactions: Applications to gluco- and mannopyranosyl sulfoxide and trichloroacetimidate type donors. *J. Am. Chem. Soc.* **2015**, *137*, 10336–10345.

(107) Crich, D. Mechanism of a chemical glycosylation reaction. *Acc. Chem. Res.* **2010**, *43*, 1144–1153.

(108) Huang, M.; Retailleau, P.; Bohe, L.; Crich, D. Cation clock permits distinction between the mechanisms of α - and β -O- and β -C-glycosylation in the mannopyranose series: evidence for the existence of a mannopyranosyl oxocarbenium ion. *J. Am. Chem. Soc.* **2012**, *134*, 14746–14749.

(109) Bohe, L.; Crich, D. A propos of glycosyl cations and the mechanism of chemical glycosylation; the current state of the art. *Carbohydr. Res.* **2015**, *403*, 48–59.

(110) Andreana, P. R.; Crich, D. Guidelines for O-glycoside formation from first principles. *ACS Cent. Sci.* **2021**, *7*, 1454–1462.

(111) Crich, D. En route to the transformation of glycoscience: A chemist's perspective on internal and external crossroads in glycochemistry. *J. Am. Chem. Soc.* **2021**, *143*, 17–34.

(112) Crich, D. Methodology development and physical organic chemistry: a powerful combination for the advancement of glycochemistry. *J. Org. Chem.* **2011**, *76*, 9193–9209.

(113) Demchenko, A. V. General aspects of the glycosidic bond formation. In *Handbook of Chemical Glycosylation*; Demchenko, A. V., Ed.; Wiley-VCH: Weinheim, Germany, 2008; pp 1–27.

(114) Christensen, H. M.; Oscarson, S.; Jensen, H. H. Common side reactions of the glycosyl donor in chemical glycosylation. *Carbohydr. Res.* **2015**, *408*, 51–95.

(115) Baghdasarian, G.; Woerpel, K. A. Electrostatic effects on the reactions of cyclohexanone oxocarbenium ions. *J. Org. Chem.* **2006**, *71*, 6851–6858.

(116) Yang, M. T.; Woerpel, K. A. The effect of electrostatic interactions on conformational equilibria of multiply substituted tetrahydropyran oxocarbenium ions. *J. Org. Chem.* **2009**, *74*, 545–553.

(117) Bérces, A.; Enright, G.; Nukada, T.; Whitfield, D. M. The conformational origin of the barrier to the formation of neighboring

group assistance in glycosylation reactions: a dynamical density functional theory study. *J. Am. Chem. Soc.* **2001**, *123*, 5460–5464.

(118) Nukada, T.; Bérces, A.; Wang, L.; Zgierski, M. Z.; Whitfield, D. M. The two-conformer hypothesis: 2,3,4,6-tetra-O-methyl-mannopyranosyl and -glucopyranosyl oxocarbenium ions. *Carbohydr. Res.* **2005**, *340*, 841–852.

(119) Whitfield, D. M. Plausible transition states for glycosylation reactions. *Carbohydr. Res.* **2012**, *356*, 180–190.

(120) Jensen, H. H.; Lyngbye, L.; Bols, M. A free-energy relationship between the rate of acidic hydrolysis of glycosides and the pK_a of isofagomines. *Angew. Chem., Int. Ed.* **2001**, *113*, 3555–3557.

(121) Jensen, H. H.; Bols, M. Steric effects are not the cause of the rate difference in hydrolysis of stereoisomeric glycosides. *Org. Lett.* **2003**, *5*, 3419–3421.

(122) Jensen, H. H.; Nordstrom, L. U.; Bols, M. The disarming effect of the 4,6-acetal group on glycoside reactivity: torsional or electronic? *J. Am. Chem. Soc.* **2004**, *126*, 9205–9213.

(123) Jensen, H. H.; Bols, M. Stereoelectronic substituent effects. *Acc. Chem. Res.* **2006**, *39*, 259–265.

(124) Jensen, H. H.; Pedersen, C. M.; Bols, M. Going to extremes: “super” armed glycosyl donors in glycosylation chemistry. *Chem.—Eur. J.* **2007**, *13*, 7576–7582.

(125) Pedersen, C. M.; Nordstrom, L. U.; Bols, M. Super armed” glycosyl donors: conformational arming of thioglycosides by silylation. *J. Am. Chem. Soc.* **2007**, *129*, 9222–9235.

(126) Pedersen, C. M.; Marinescu, L. G.; Bols, M. Conformationally armed glycosyl donors: reactivity quantification, new donors and one pot reactions. *Chem. Commun.* **2008**, 2465–2467.

(127) Heuckendorff, M.; Pedersen, C. M.; Bols, M. Quantifying electronic effects of common carbohydrate protecting groups in a piperidine model system. *Chem.—Eur. J.* **2010**, *16*, 13982–13994.

(128) Beaver, M. G.; Woerpel, K. A. Erosion of stereochemical control with increasing nucleophilicity: O-glycosylation at the diffusion limit. *J. Org. Chem.* **2010**, *75*, 1107–1118.

(129) Garcia, A.; Sanzone, J. R.; Woerpel, K. A. Participation of alkoxy groups in reactions of acetals: violation of the reactivity/selectivity principle in a Curtin–Hammett kinetic scenario. *Angew. Chem., Int. Ed. Engl.* **2015**, *54*, 12087–12090.

(130) Mootoo, D. R.; Konradsson, P.; Udodong, U.; Fraser-Reid, B. “Armed” and “disarmed” n-pentenyl glycosides in saccharide couplings leading to oligosaccharides. *J. Am. Chem. Soc.* **1988**, *110*, 5583–5584.

(131) Fraser-Reid, B.; Udodong, U. E.; Wu, Z. F.; Ottosson, H.; Merritt, J. R.; Rao, C. S.; Roberts, C.; Madsen, R. n-Pentenyl glycosides in organic chemistry: a contemporary example of serendipity. *Synlett* **1992**, 1992, 927–942 and references therein.

(132) Fraser-Reid, B.; Wu, Z.; Andrews, C. W.; Skowronski, E.; Bowen, J. P. Torsional effects in glycoside reactivity: saccharide couplings mediated by acetal protecting groups. *J. Am. Chem. Soc.* **1991**, *113*, 1434–1435.

(133) Wilson, B. G.; Fraser-Reid, B. n-Pentenyl glycoside based methodology for determining the relative reactivities of variously protected pairs of glycosides. *J. Org. Chem.* **1995**, *60*, 317–320.

(134) Douglas, N. L.; Ley, S. V.; Lucking, U.; Warriner, S. L. Tuning glycoside reactivity: new tool for efficient oligosaccharides synthesis. *J. Chem. Soc., Perkin Trans. 1* **1998**, 51–65.

(135) Zhang, Z.; Ollmann, I. R.; Ye, X. S.; Wischnat, R.; Baasov, T.; Wong, C. H. Programmable one-pot oligosaccharide synthesis. *J. Am. Chem. Soc.* **1999**, *121*, 734–753.

(136) Fridman, M.; Solomon, D.; Yogev, S.; Baasov, T. One-pot synthesis of glucosamine oligosaccharides. *Org. Lett.* **2002**, *4*, 281–283.

(137) Bandara, M. D.; Yasomane, J. P.; Demchenko, A. V. Application of armed, disarmed, superarmed and superdisarmed building blocks in stereocontrolled glycosylation and expeditious oligosaccharide synthesis. In *Selective Glycosylations: Synthetic Methods and Catalysts*; Bennett, C. S., Ed.; Wiley, 2017; pp 29–58.

- (138) Zhu, T.; Boons, G. J. Thioglycosides protected as trans-2,3-cyclic carbonates in chemoselective glycosylations. *Org. Lett.* **2001**, *3*, 4201–4203.
- (139) Kamat, M. N.; Demchenko, A. V. Revisiting the armed-disarmed concept rationale: chemoselective activation of the S-benzoxazolyl glycosides in oligosaccharide synthesis. *Org. Lett.* **2005**, *7*, 3215–3218.
- (140) Mydock, L. K.; Demchenko, A. V. Superarming the S-benzoxazolyl glycosyl donors by simple 2-O-benzoyl-3,4,6-tri-O-benzyl protection. *Org. Lett.* **2008**, *10*, 2103–2106.
- (141) Mydock, L. K.; Demchenko, A. V. Application of the superarmed glycosyl donor to chemoselective oligosaccharide synthesis. *Org. Lett.* **2008**, *10*, 2107–2110.
- (142) Premathilake, H. D.; Mydock, L. K.; Demchenko, A. V. Superarming common glycosyl donors by simple 2-O-benzoyl-3,4,6-tri-O-benzyl protection. *J. Org. Chem.* **2010**, *75*, 1095–1100.
- (143) Panza, M.; Civera, M.; Yasomane, J. P.; Belvisi, L.; Demchenko, A. V. Bromine-promoted glycosidation of conformationally superarmed thioglycosides. *Chem.—Eur. J.* **2019**, *25*, 11831–11836.
- (144) Nogueira, J. M.; Issa, J. P.; Chu, A.-H. A.; Sisel, J. A.; Schum, R. S.; Bennett, C. S. Halide effects on cyclopropenium cation promoted glycosylation with deoxy sugars: highly α -selective glycosylations using a 3,3-dibromo-1,2-diphenylcyclopropene promoter. *Eur. J. Org. Chem.* **2012**, *2012*, 4927–4930.
- (145) Sun, L.; Wu, X.; Xiong, D. C.; Ye, X. S. Stereoselective Koenigs-Knorr glycosylation catalyzed by urea. *Angew. Chem., Int. Ed.* **2016**, *55*, 8041–8044.
- (146) Singh, Y.; Demchenko, A. V. Koenigs-Knorr glycosylation reaction catalyzed by trimethylsilyl trifluoromethanesulfonate. *Chem.—Eur. J.* **2019**, *25*, 1461–1465.
- (147) Yu, F.; Dickson, J. L.; Loka, R. S.; Xu, H.; Schaugard, R. N.; Schlegel, H. B.; Luo, L.; Nguyen, H. M. Diastereoselective sp³ CO bond formation via visible light-induced, copper-catalyzed cross couplings of glycosyl bromides with aliphatic alcohols. *ACS Catal.* **2020**, *10*, 5990–6001.
- (148) Fischer, E.; Fischer, H. Über einige Derivate des Milchzuckers und der Maltose und über zwei neue Glucoside. *Ber. Dtsch. Chem. Ges.* **1910**, *43*, 2521–2536.
- (149) Fischer, E. Acetohalogen Glucoses and Maltose and Melibiose p-Bromophenylsazones. *Ber. Dtsch. Chem. Ges.* **1911**, *44*, 1898–1904.
- (150) Fischer, E.; von Mechel, L. Synthesis of phenyl glucosides. *Ber. Dtsch. Chem. Ges.* **1916**, *49*, 2813–2820.
- (151) Fischer, E.; Helferich, B. Über neue synthetische glucoside. *Ann. Chem.* **1911**, *383*, 68–91.
- (152) Helferich, B.; Klein, W. Disaccharides. IV. Two tetraacetyl-b-D-glucoses. *Liebigs Ann.* **1926**, *450*, 219–229.
- (153) Helferich, B.; Bohn, E.; Winkler, S. Unsaturated derivatives of gentiobiose and cellobiose. *Ber. Dtsch. Chem. Ges.* **1930**, *63B*, 989–998.
- (154) Reynolds, D. D.; Evans, W. L. The preparation of α - and β -gentiobiose octaacetates. *J. Am. Chem. Soc.* **1938**, *60*, 2559–2561.
- (155) Kaothip, S.; Demchenko, A. V. Expeditious oligosaccharide synthesis via selective and orthogonal activation. *Carbohydr. Res.* **2011**, *346*, 1371–1388.
- (156) Helferich, B.; Brederick, H. Zuckersynthesen. VIII. *Liebigs Ann. Chem.* **1928**, *465*, 166–184.
- (157) Brigl, P.; Keppler, H. Carbohydrates. IV. Synthesis of α -glucosides. *Ber. Dtsch. Chem. Ges.* **1926**, *59*, 1588–1591.
- (158) Schlubach, H. H.; Schroter, G. A. Die Gewinnung von Glucosiden der α -Reihe: Krystallisiertes α -Methyl-fructosid. *Ber.* **1928**, *61*, 1216–1222.
- (159) Hickinbottom, W. J. Glucosides. Part II. The preparation of α -glucosides from β -glucosyl chlorides. *J. Chem. Soc.* **1929**, *0*, 1676–1687.
- (160) Zemplén, G. Einwirkung von Aluminiummetall und Quecksilbersalzen auf Aceto-halogen-zucker, I.: Synthese von α -Biosiden. *Ber. Dtsch. Chem. Ges.* **1929**, *62*, 990–993.
- (161) Zemplén, G.; Nagy, Z. S. Einwirkung von Quecksilbersalzen auf Aceto-halogenzucker. II. Mitteil.: Bildungs-Bedingungen des α -Phenyl-cellobiosids. *Ber. Dtsch. Chem. Ges.* **1930**, *63*, 368–368.
- (162) Zemplén, G.; Gerecs, A. Action of mercury salts on acetohalogenosugars. IV. Direct preparation of alkyl biosides of the α -series. *Ber. Dtsch. Chem. Ges.* **1930**, *63B*, 2720–2729.
- (163) Zemplén, G. Einwirkung von Quecksilbersalzen auf Acetohalogenzucker, III. Mitteil.: Synthese der achtfach methylierten Cellobiose. *Ber. Dtsch. Chem. Ges.* **1930**, *63*, 1820–1823.
- (164) Fischer, E.; Bergmann, M.; Rabe, A. Acetobromorhanose and its use for the synthesis of rhamnosides. *Ber. Dtsch. Chem. Ges. B* **1920**, *53B*, 2362–2388.
- (165) Dale, J. K. Three isomeric crystalline tetra-acetyl-methyl-D-mannosides. *J. Am. Chem. Soc.* **1924**, *46*, 1046–1051.
- (166) Levene, P. A.; Wolfrom, M. L. Acetylmannosides. V. The rates of hydrolysis of tetraacetylmethylmannosides and of triacetylmethylxylosides. *J. Biol. Chem.* **1928**, *79*, 471–474.
- (167) Haworth, W. N.; Hirst, E. L.; Miller, E. J. Development of a novel form of stereoisomerism in the sugar series. I. The third variety of triacetylmethylrhamnoside. *J. Chem. Soc.* **1929**, *0*, 2469–2479.
- (168) Hudson, C. Relations between rotatory power and structure in the sugar group. XXV. The ring structures of various monosaccharides. *J. Am. Chem. Soc.* **1930**, *52*, 1680–1700.
- (169) Isbell, H. The structures of the acetylmethylmannosides. *J. Am. Chem. Soc.* **1930**, *52*, 5298–5298.
- (170) Haworth, W. N.; Hirst, E. L.; Streight, H. R. L.; Thomas, H. A.; Webb, J. I. CCCXLIX.—The structure of carbohydrates and their optical rotatory power. Part II. 4-Glucosido- α -mannose and its derivatives. *J. Chem. Soc.* **1930**, *0*, 2636–2644.
- (171) Freudenberg, K.; Braun, E. A new isomerism in the sugar group. *Naturwissenschaften* **1930**, *18*, 393.
- (172) Freudenberg, K.; Scholz, H. Acetonesugars and other compounds of the carbohydrates. XXII. Cyclic acetates in the sugar group. *Ber. Dtsch. Chem. Ges. B* **1930**, *63B*, 1969–1972.
- (173) Bott, H. G.; Haworth, W. N.; Hirst, E. L. Development of a novel form of isomerism in the sugar series. II. The third variety of tetraacetylmethylmannoside. *J. Chem. Soc.* **1930**, *0*, 1395–1405.
- (174) Haworth, W. N.; Hirst, E. L.; Stacey, M. Walden inversion in the α -glucoheptose series. Preparation of new derivatives and the determination of the structure of methyl α -glucoheptoside. *J. Chem. Soc.* **1931**, *0*, 2864–2872.
- (175) Isbell, H. S. Derivatives of 4-glucosidomannose. *Bur. Stand. J. Res.* **1931**, *7*, 1115–1131.
- (176) Isbell, H. S. Chemistry of the Carbohydrates and Glycosides. *Annu. Rev. Biochem.* **1940**, *9*, 65–92.
- (177) Frush, H. L.; Isbell, H. S. Sugar acetates, acetylglycosyl halides and orthoacetates in relation to the Walden inversion. *J. Res. Natl. Bur. Stand. (U. S.)* **1941**, *27*, 413–428.
- (178) Frush, H. L.; Isbell, H. Acetyl derivatives of certain heptoses, of gulose, and of lactulose. *J. Res. Natl. Bur. Stds.* **1945**, *34*, 111–116.
- (179) Isbell, H. S.; Frush, H. L. Mechanisms for the formation of acetylglycosides and orthoesters from acetylglycosyl halides. *J. Res. Natl. Bur. Stand.* **1949**, *43*, 161–171.
- (180) Helferich, B.; Doppstadt, A.; Gottschlich, A. Synthesen von Glykosiden im homogenen Medium. *Naturwissenschaften* **1953**, *40*, 441–442.
- (181) Helferich, B.; Weis, K. Zur Synthese von Glucosiden und von nicht-reduzierenden Disacchariden. *Chem. Ber.* **1956**, *89*, 314–321.
- (182) Lemieux, R.; Morgan, A. The mechanism for the formation of 1, 2-cis-pyridine nucleosides from 1, 2-cis-acetohalogenosugars. A novel rearrangement. *J. Am. Chem. Soc.* **1963**, *85*, 1889–1890.
- (183) Lemieux, G. A.; Morgan, A. R. The preparation and configurations of tri-O-acetyl- α -D-glucopyranose 1,2-(orthoesters). *Can. J. Chem.* **1965**, *43*, 2198–2204.
- (184) Kochetkov, N. K.; Khorlin, A. Y.; Bochkov, A. F. A new method of glycosylation. *Tetrahedron* **1967**, *23*, 693–707.
- (185) Schulz, M.; Steinmaus, H. 1,2-Orthoacetate aus 1,2-cis-Acetohalogenosen. *Zeitschrift für Naturforschung B* **1964**, *19*, 263–263.

- (186) Wulff, G.; Krüger, W. Untersuchungen Glykosidsynthese: III. Teil. Eine neue darstellungsmethode für 1, 2-D-glucose-orthoester. *Carbohydr. Res.* **1971**, *19*, 139–142.
- (187) Hanessian, S.; Banoub, J. Chemistry of the glycosidic linkage. A rapid and efficient synthesis of carbohydrate 1, 2-orthoesters. *Carbohydr. Res.* **1975**, *44*, C14–C17.
- (188) Wulff, G.; Schröder, U.; Wilchellhaus, J. On the stereochemical control of glycosylation reactions by the addition of tetrahydrofuran. *Carbohydr. Res.* **1979**, *72*, 280–284.
- (189) Wang, W.; Kong, F. New synthetic methodology for regio- and stereoselective synthesis of oligosaccharides via sugar ortho ester intermediates. *J. Org. Chem.* **1998**, *63*, 5744–5745.
- (190) Kong, F. Recent studies on reaction pathways and applications of sugar orthoesters in synthesis of oligosaccharides. *Carbohydr. Res.* **2007**, *342*, 345–373.
- (191) Talley, E. A.; Reynolds, D. D.; Evans, W. L. The synthesis of certain disaccharide acetates in the mannose series. *J. Am. Chem. Soc.* **1943**, *65*, 575–582.
- (192) Perlin, A. Formation of diastereomeric orthoacetates of D-mannose: N.M.R. spectral evidence. *Can. J. Chem.* **1963**, *41*, 399–406.
- (193) Ditmar, R. Über Methylglucoside und andere Derivate des Milchsuckers. *Monatsh. Chem.* **1902**, *23*, 865–876.
- (194) Koto, S.; Yoshida, T.; Takenaka, K.; Zen, S. A Through-process for the preparation of methyl per-O-acetyl 1-thio-glycosides from aldoses. *Bull. Chem. Soc. Jpn.* **1982**, *55*, 3667–3668.
- (195) Koto, S.; Morishima, N.; Shichi, S.; Haigoh, H.; Hirooka, M.; Okamoto, M.; Higuchi, T.; Shimizu, K.; Hashimoto, Y.; Irisawa, T.; Kawasaki, H.; Takahashi, Y.; Yamazaki, M.; Mori, Y.; Kudo, K.; Ikegaki, T.; Suzuki, S.; Zen, S. Dehydrative glycosylation using heptabenzyl derivatives of glucobioses and lactose. *Bull. Chem. Soc. Jpn.* **1992**, *65*, 3257–3274.
- (196) Kartha, K. P. R.; Jennings, H. J. A simplified, one-pot preparation of acetobromosugars from reducing sugars. *J. Carbohydr. Chem.* **1990**, *9*, 777–781.
- (197) Hunsen, M.; Long, D. A.; D'Ardenne, C. R.; Smith, A. L. Mild one-pot preparation of glycosyl bromides. *Carbohydr. Res.* **2005**, *340*, 2670–2674.
- (198) Zhao, J.-z.; Zhang, X.-q.; Wu, X.; Xing, Z.-b.; Yue, A.-q.; Shao, H.-w. Rapid synthesis of glycosyl bromides by ultrasound irradiation. *Chem. Res. Chin. Univ.* **2013**, *29*, 71–75.
- (199) Huang, L. C.; Liang, P. H.; Liu, C. Y.; Lin, C. C. Large-scale synthesis of per-O-acetylated saccharides and their sequential transformation to glycosyl bromides and thioglycosides. *J. Carbohydr. Chem.* **2006**, *25*, 303–313.
- (200) Yalamanchili, S.; Nguyen, T. A.; Zsikla, A.; Stamper, G.; DeYong, A. E.; Florek, J.; Vasquez, O.; Pohl, N. L. B.; Bennett, C. S. Automated, Multistep Continuous-Flow Synthesis of 2,6-Dideoxy and 3-Amino-2,3,6-trideoxy Monosaccharide Building Blocks. *Angew. Chem., Int. Ed. Engl.* **2021**, *60*, 23171–23175.
- (201) Brederick, H.; Hamsch, E. About alkali metal compounds of 2,3,4,6-tetramethyl- α -D-glucopyranose and 1,3,4,6-tetramethyl- α -D-fructofuranose and their use in glycoside and disaccharide syntheses. *Chem. Ber.* **1954**, *87*, 38–45.
- (202) Zemlén, G.; Gerecs, Á. Einwirkung von Quecksilbersalzen auf Aceto-halogen-zucker, IX. Mitteil.: Synthese von Derivaten der β -1-Rhamnosido-6-d-glykose. *Ber. Dtsch. Chem. Ges.* **1934**, *67*, 2049–2051.
- (203) Kochetkov, N. K.; Klimov, E. M.; Malysheva, N. N.; Demchenko, A. V. Stereospecific synthesis of 1,2-cis-glycosides of 2-amino sugars. *Carbohydr. Res.* **1993**, *242*, C7–C10.
- (204) Barker, R.; Fletcher, H. G., Jr. 2,3,5-Tri-O-benzyl-D-ribose and -L-arabinose Bromides. *J. Org. Chem.* **1961**, *26*, 4605–4609.
- (205) Lemieux, R. U.; Kondo, T. Benzyl trifluoromethanesulfonate. Preparation of tri-O-acetyl-2-O-benzyl- α -D-galactopyranosyl bromide from 1,3,4,6-tetra-O-acetyl- α -D-galactopyranose. *Carbohydr. Res.* **1974**, *35*, C4–C6.
- (206) Excoffier, G.; Gagnaire, D. Y.; Vignon, M. R. Synthesis of oligosaccharides on a polymeric support. VII. Stereospecificity of glycosylation reactions: synthesis of isomaltose. *Carbohydr. Res.* **1976**, *46*, 215–226.
- (207) Thiem, J.; Meyer, B. Synthesen mit Iod- und Bromtrimethylsilan in der Saccharidchemie. *Chem. Ber.* **1980**, *113*, 3075–3085.
- (208) Gillard, J. W.; Israel, M. Trimethylsilyl bromide as a mild, stereoselective anomeric brominating agent. *Tetrahedron Lett.* **1981**, *22*, 513–516.
- (209) Montero, J.-L.; Winum, J.-Y.; Leydet, A.; Kamal, M.; Pavia, A. A.; Roque, J.-P. A convenient synthesis of peracetylated glycosyl halides using bismuth (III) halides as catalysts. *Carbohydr. Res.* **1997**, *297*, 175–180.
- (210) Tojino, M.; Hirose, Y.; Mizuno, M. Convenient synthesis of glycosyl bromide from 1-O-acetyl sugars by photo-irradiative phase-vanishing reaction of molecular bromine. *Tetrahedron Lett.* **2013**, *54*, 7124–7126.
- (211) Fletcher, H. G., Jr.; Hudson, C. 1, 5-Anhydro-xylitol. *J. Am. Chem. Soc.* **1947**, *69*, 921–924.
- (212) Ness, R. K.; Fletcher Jr, H. G.; Hudson, C. The reaction of 2, 3, 4, 6-tetrabenzoyl- α -D-glucopyranosyl bromide and 2, 3, 4, 6-tetrabenzoyl- α -D-mannopyranosyl bromide with methanol. Certain benzoylated derivatives of D-Glucose and D-Mannose. *J. Am. Chem. Soc.* **1950**, *72*, 2200–2205.
- (213) Wen, P.; Simmons, C. J.; Ma, Z.-x.; Blaszczyk, S. A.; Balzer, P. G.; Ye, W.; Duan, X.; Wang, H.-Y.; Yin, D.; Stevens, C. M.; Tang, W. Synthesis of glycosyl chlorides and bromides by chelation assisted activation of picolinic esters under mild neutral conditions. *Org. Lett.* **2020**, *22*, 1495–1498.
- (214) Weygand, F.; Ziemann, H.; Bestmann, H. J. Eine neue methode zur darstellung von acetobromzuckern. *Chem. Ber.* **1958**, *91*, 2534–2537.
- (215) Weygand, F.; Ziemann, H. Glycosyl bromides from ethyl 1-thioglycosides. II. *Ann.* **1962**, *657*, 179–198.
- (216) Fugedi, P.; Garegg, P. J.; Lonn, H.; Norberg, T. Thioglycosides as glycosylating agents in oligosaccharide synthesis. *Glycoconjugate J.* **1987**, *4*, 97–108.
- (217) Singh, Y.; Wang, T.; Geringer, S. A.; Stine, K. J.; Demchenko, A. V. Regenerative glycosylation. *J. Org. Chem.* **2018**, *83*, 374–381.
- (218) Shingu, Y.; Nishida, Y.; Dohi, H.; Matsuda, K.; Kobayashi, K. Convenient access to halide ion-catalyzed α -glycosylation free from noxious fumes at the donor synthesis. *J. Carbohydr. Chem.* **2002**, *21*, 605–611.
- (219) Polat, T.; Linhardt, R. J. Zinc triflate-benzoyl bromide: a versatile reagent for the conversion of ether into benzoate protecting groups and ether glycosides into glycosyl bromides. *Carbohydr. Res.* **2003**, *338*, 447–449.
- (220) Helferich, B.; Gootz, R. Synthesis of a tetrasaccharide acetate. *Ber. Dtsch. Chem. Ges.* **1931**, *64B*, 109–114.
- (221) Singh, Y.; Demchenko, A. V. Defining the scope of the acid-catalyzed glycosidation of glycosyl bromides. *Chem.—Eur. J.* **2020**, *26*, 1042–1051.
- (222) Knöchel, A.; Rudolph, G. Anionenaktivierung I: Anionenaktivierung durch makrocyclische Polyether und ihr Einfluß auf nucleophile Substitutionsreaktionen. *Tetrahedron Lett.* **1974**, *15*, 3739–3740.
- (223) Knöchel, A.; Rudolph, G.; Thiem, J. Homogene koenigs-knorr-reaktion mit hilfe zyklischer polyäther. *Tetrahedron Lett.* **1974**, *15*, 551–552.
- (224) Gouliaras, C.; Lee, D.; Chan, L.; Taylor, M. S. Regioselective activation of glycosyl acceptors by a diarylborinic acid-derived catalyst. *J. Am. Chem. Soc.* **2011**, *133*, 13926–13929.
- (225) Nigudkar, S. S.; Stine, K. J.; Demchenko, A. V. Regenerative glycosylation under nucleophilic catalysis. *J. Am. Chem. Soc.* **2014**, *136*, 921–923.
- (226) Shadrack, M.; Singh, Y.; Demchenko, A. V. Stereocontrolled α -galactosylation under cooperative catalysis. *J. Org. Chem.* **2020**, *85*, 15936–15944.
- (227) Brederick, H.; Wagner, A.; Faber, G.; Ott, H.; Rauther, J. Eine vereinfachte Oligosaccharid-Synthese. *Chem. Ber.* **1959**, *92*, 1135–1139.

- (228) Bredereck, H.; Wagner, A.; Kuhn, H.; Ott, H. Oligosaccharidsynthesen. II. Synthesen von Di- und Trisacchariden des Gentio-biosetyps. *Chem. Ber.* **1960**, *93*, 1201–1206.
- (229) Khorlin, A. Y.; Nesmeyanov, V. A.; Zurabyan, S. E. Glycosylation of sugar 2, 3-diphenyl-2-cyclopropen-1-yl ethers. A new route to oligosaccharides. *Carbohydr. Res.* **1975**, *43*, 69–77.
- (230) Kronzer, F. J.; Schuerch, C. The methanolysis of some derivatives of 2,3,4-tri-O-benzyl- α -D-glucopyranosyl bromide in the presence and absence of silver salts. *Carbohydr. Res.* **1973**, *27*, 379–390.
- (231) Kihlberg, J. O.; Leigh, D. A.; Bundle, D. R. The in situ activation of thioglycosides with bromine: an improved glycosylation method. *J. Org. Chem.* **1990**, *55*, 2860–2863.
- (232) Wulff, G.; Röhle, G.; Krüger, W. New methods for preparation of glycosides. *Angew. Chem., Int. Ed.* **1970**, *9*, 455–456.
- (233) Wulff, G.; Röhle, G.; Krueger, W. Glycoside synthesis. IV. Novel silver salts in glycoside synthesis. *Chem. Ber.* **1972**, *105*, 1097–1110.
- (234) Wulff, G.; Röhle, G. Results and problems of O-glycoside synthesis. *Angew. Chem., Int. Ed. Engl.* **1974**, *13*, 157–170.
- (235) Paulsen, H.; Lockhoff, O. Building units for oligosaccharides. XXX. New effective β -glycoside synthesis of mannose glycosides. Syntheses of mannose containing oligosaccharides. *Chem. Ber.* **1981**, *114*, 3102–3114.
- (236) Garegg, P. J.; Ossowski, P. Silver zeolite as promoter in glycoside synthesis. The synthesis of β -D-mannopyranosides. *Acta Chem. Scand., Ser. B* **1983**, *37b*, 249–250.
- (237) Van Boeckel, C.; Beetz, T.; Kock-van Dalen, A.; Van Bekkum, H. A note on the use of porous silver silicates as promoter in carbohydrate coupling reactions. *Recl. Trav. Chim. Pays-Bas* **1987**, *106*, 596–598.
- (238) Garegg, P. J.; Johansson, R.; Samuelsson, B. Silver imidazolate-assisted glycosidations. Part 6. Synthesis of 1,2-trans-linked disaccharides. *Acta Chem. Scand., Ser. B* **1982**, *36b*, 249–250.
- (239) Helferich, B.; Wedemeyer, K. F. Preparation of glucosides from acetobromoglucose. *Ann.* **1949**, *563*, 139–145.
- (240) Schroeder, L. R.; Green, J. W. Koenigs-Knorr syntheses with mercuric salts. *J. Chem. Soc. C: Organic* **1966**, *0*, 530–531.
- (241) Sinay, P. Recent advances in glycosylation reactions. *Pure Appl. Chem.* **1978**, *50*, 1437–1452.
- (242) Paulsen, H.; Kolář, Č. Bausteine von Oligosacchariden, XX. Synthese der Oligosaccharid-Determinanten der Blutgruppensubstanzen der Type 2 des ABH-Systems. Diskussion der α -Glycosid-Synthese. *Chem. Ber.* **1981**, *114*, 306–321.
- (243) Helferich, B.; Wedemeyer, K. F. Zur Darstellung von Glykosiden aus Acetobromhalogenosen. *Chem. Ber.* **1950**, *83*, 538–540.
- (244) Bock, K.; Meldal, M. Mercury iodide as a catalyst in oligosaccharide synthesis. *Acta Chem. Scand.* **1983**, *37b*, 775–783.
- (245) Higashi, K.; Nakayama, K.; Soga, T.; Shioya, E.; Uoto, K.; Kusama, T. Novel stereoselective glycosidation by the combined use of trityl halide and Lewis acid. *Chem. Pharm. Bull.* **1990**, *38*, 3280–3282.
- (246) Murakami, T.; Sato, Y.; Shibakami, M. Stereoselective glycosylations using benzoylated glucosyl halides with inexpensive promoters. *Carbohydr. Res.* **2008**, *343*, 1297–1308.
- (247) Higashi, K.; Nakayama, K.; Uoto, K.; Shioya, E.; Kusama, T. A novel glycoside anomerization catalyzed by trimethylsilyl bromide and zinc bromide in combination. *Chem. Pharm. Bull.* **1991**, *39*, 590–592.
- (248) Higashi, K.; Nakayama, K.; Shioya, E.; Kusama, T. Direct transformation of O-glycosides into other O-glycosides using trimethylsilyl bromide and zinc bromide. *Chem. Pharm. Bull.* **1992**, *40*, 1042–1043.
- (249) Bernstein, S.; Conrow, R. Steroid conjugates. VI. Improved Koenigs-Knorr synthesis of aryl glucuronides using cadmium carbonate, a new and effective catalyst. *J. Org. Chem.* **1971**, *36*, 863–870.
- (250) Wimmer, Z.; Pechová, L.; Saman, D. Koenigs-Knorr synthesis of cycloalkyl glycosides. *Molecules* **2004**, *9*, 902–912.
- (251) Dick Jr, W. E. Efficiency of cadmium carbonate as an aryl glycosidation catalyst: effects of lot variations on product compositions. *Carbohydr. Res.* **1979**, *70*, 313–318.
- (252) Mukherjee, D.; Kumar Ray, P.; Sankar Chowdhury, U. Synthesis of glycosides via indium (III) chloride mediated activation of glycosyl halide in neutral condition. *Tetrahedron* **2001**, *57*, 7701–7704.
- (253) Chandra, S.; Yadav, R. N.; Paniagua, A.; Banik, B. K. Indium salts-catalyzed O and S-glycosylation of bromo sugar with benzyl glycolate: an unprecedented hydrogenolysis. *Tetrahedron Lett.* **2016**, *57*, 1425–1429.
- (254) Li, C.; Liang, H.; Zhang, Z.-x.; Wang, Z.; Yu, L.; Liu, H.; An, F.; Wang, S.; Ma, L.; Xue, W. A catalytic Koenigs-Knorr glycosylation based on acceptor activation with In(NTf₂)₃. *Tetrahedron* **2018**, *74*, 3963–3970.
- (255) Ogawa, T.; Matsui, M. An approach to synthesis of glycosides: enhancement of nucleophilicity of hydroxyl groups by trialkylstannylation. *Carbohydr. Res.* **1976**, *51*, C13–C18.
- (256) Lubineau, A.; Malleron, A. Stannous triflate mediated glycosidations. A stereoselective synthesis of β -D-glucosides. *Tetrahedron Lett.* **1985**, *26*, 1713–1716.
- (257) Yu, F.; Li, J.; DeMent, P.; Tu, Y.; Schlegel, H.; Nguyen, H. Phenanthroline-catalyzed stereoretentive glycosylations. *Angew. Chem., Int. Ed.* **2019**, *58*, 6957–6961.
- (258) Lemieux, R. U.; Hendriks, K. B.; Stick, R. V.; James, K. Halide ion catalyzed glycosylation reactions. Syntheses of α -linked disaccharides. *J. Am. Chem. Soc.* **1975**, *97*, 4056–4062 and references therein.
- (259) Lemieux, R. U.; Driguez, H. Chemical synthesis of 2-acetamido-2-deoxy-4-O-(α -L-fucopyranosyl)-3-O-(β -D-galactopyranosyl)-D-glucose. Lewis a blood-group antigenic determinant. *J. Am. Chem. Soc.* **1975**, *97*, 4063–4069.
- (260) Kartha, K. P. R.; Aloui, M.; Field, R. A. Iodine: a versatile reagent in carbohydrate chemistry III. Efficient activation of glycosyl halides in combination with DDQ. *Tetrahedron Lett.* **1996**, *37*, 8807–8810.
- (261) Lanz, G.; Madsen, R. Glycosylation with disarmed glycosyl bromides promoted by iodonium ions. *Eur. J. Org. Chem.* **2016**, *2016*, 3119–3125.
- (262) Kartha, K. R.; Field, R. A. Glycosylation chemistry promoted by iodine monobromide: Efficient synthesis of glycosyl bromides from thioglycosides, and O-glycosides from 'disarmed' thioglycosides and glycosyl bromides. *Tetrahedron Lett.* **1997**, *38*, 8233–8236.
- (263) Stachulski, A. V. Glucuronidation of alcohols using the bromosugar-iodonium reagent method. *Tetrahedron Lett.* **2001**, *42*, 6611–6613.
- (264) Micheel, F.; Micheel, H. Configuration of the α - and β -forms in the sugar series. *Ber. Dtsch. Chem. Ges. B* **1930**, *63B*, 386–393.
- (265) West, A. C.; Schuerch, C. The reverse anomeric effect and the synthesis of α -glycosides. *J. Am. Chem. Soc.* **1973**, *95*, 1333–1335.
- (266) Kronzer, F. J.; Schuerch, C. Use of positively charged, leaving groups in the synthesis of α -D-linked galactosides. Attempted synthesis of 3-O- α -(D-(galactopyranosyl)-D-galactose. *Carbohydr. Res.* **1974**, *33*, 273–280.
- (267) Ishikawa, T.; Fletcher, H. G. The synthesis and solvolysis of some D-glucopyranosyl bromides having a benzyl group at C-2. *J. Org. Chem.* **1969**, *34*, 563–571.
- (268) Frechet, J. M.; Schuerch, C. Solid-phase synthesis of oligosaccharides. II. Steric control by C-6 substituents in glucoside syntheses. *J. Am. Chem. Soc.* **1972**, *94*, 604–609.
- (269) Wulff, G.; Röhle, G. Untersuchungen zur Glykosidsynthese, VI. Kinetische Untersuchungen zum Mechanismus der Koenigs-Knorr-Reaktion. *Chem. Ber.* **1972**, *105*, 1122–1132.
- (270) Wulff, G.; Röhle, G.; Schmidt, U. Untersuchungen zur Glykosidsynthese, V. Reaktionsprodukte und Stereospezifität der Glucosylierung in Gegenwart unlöslicher Silberalze in Diäthyläther. *Chem. Ber.* **1972**, *105*, 1111–1121.
- (271) Kaneko, M.; Herzon, S. B. Scope and limitations of 2-deoxy- and 2,6-dideoxyglycosyl bromides as donors for the synthesis of beta-

- 2-deoxy- and beta-2,6-dideoxyglycosides. *Org. Lett.* **2014**, *16*, 2776–2779.
- (272) Lee, D.; Williamson, C. L.; Chan, L.; Taylor, M. S. Regioselective, borinic acid-catalyzed monoacylation, sulfonylation and alkylation of diols and carbohydrates: expansion of substrate scope and mechanistic studies. *J. Am. Chem. Soc.* **2012**, *134*, 8260–8267.
- (273) Lee, D.; Taylor, M. S. Borinic acid-catalyzed regioselective acylation of carbohydrate derivatives. *J. Am. Chem. Soc.* **2011**, *133*, 3724–3727.
- (274) Chan, L.; Taylor, M. S. Regioselective alkylation of carbohydrate derivatives catalyzed by a diarylborinic acid derivative. *Org. Lett.* **2011**, *13*, 3090–3093.
- (275) Yang, W.; Mortier, W. J. The use of global and local molecular parameters for the analysis of the gas-phase basicity of amines. *J. Am. Chem. Soc.* **1986**, *108*, 5708–5711.
- (276) Nigudkar, S. S.; Wang, T.; Pistorio, S. G.; Yasomanee, J. P.; Stine, K. J.; Demchenko, A. V. OFox imidates as versatile glycosyl donors for chemical glycosylation. *Org. Biomol. Chem.* **2017**, *15*, 348–359.
- (277) Escopy, S.; Singh, Y.; Stine, K. J.; Demchenko, A. V. A Streamlined regenerative glycosylation reaction: direct, acid-free activation of thioglycosides. *Chem.—Eur. J.* **2021**, *27*, 354–361.
- (278) Fraser-Reid, B.; Wu, Z.; Udodong, U. E.; Ottosson, H. Armed/disarmed effects in glycosyl donors: rationalization and sidetracking. *J. Org. Chem.* **1990**, *55*, 6068–6070.
- (279) Demchenko, A. V.; Rousson, E.; Boons, G. J. Stereoselective 1,2-cis-galactosylation assisted by remote neighboring group participation and solvent effects. *Tetrahedron Lett.* **1999**, *40*, 6523–6526.
- (280) Creutz, S. E.; Lotito, K. J.; Fu, G. C.; Peters, J. C. Photoinduced Ullmann C–N coupling: demonstrating the viability of a radical pathway. *Science* **2012**, *338*, 647–651.
- (281) Ahn, J. M.; Ratani, T. S.; Hannoun, K. I.; Fu, G. C.; Peters, J. C. Photoinduced, copper-catalyzed alkylation of amines: a mechanistic study of the cross-coupling of carbazole with alkyl bromides. *J. Am. Chem. Soc.* **2017**, *139*, 12716–12723.
- (282) Kochi, J.; Subramanian, R. Kinetics of Electron-Transfer Oxidation of Alkyl Radicals by Copper (II) Complexes. *J. Am. Chem. Soc.* **1965**, *87*, 4855–4866.
- (283) Kochi, J. K. Mechanisms of organic oxidation and reduction by metal complexes. *Science* **1967**, *155*, 415–424.
- (284) Kochi, J.; Bemis, A.; Jenkins, C. Mechanism of electron transfer oxidation of alkyl radicals by copper (II) complexes. *J. Am. Chem. Soc.* **1968**, *90*, 4616–4625.
- (285) De La Mare, H.; Kochi, J.; Rust, F. The oxidation of free radicals by metal ions. *J. Am. Chem. Soc.* **1961**, *83*, 2013–2015.
- (286) Wayner, D. D. M.; McPhee, D. J.; Griller, D. Oxidation and reduction potentials of transient free radicals. *J. Am. Chem. Soc.* **1988**, *110*, 132–137.
- (287) Zhang, Z.; Wang, F.; Mu, X.; Chen, P.; Liu, G. Copper-catalyzed regioselective fluorination of allylic halides. *Angew. Chem., Int. Ed.* **2013**, *52*, 7549–7553.
- (288) Andrews, R. S.; Becker, J. J.; Gagne, M. R. Intermolecular addition of glycosyl halides to alkenes mediated by visible light. *Angew. Chem., Int. Ed. Engl.* **2010**, *49*, 7274–7276.
- (289) Zemplen, G.; Gerecs, A.; Flesch, H. Action of mercury salts on acetohalosugars. XI. Synthesis of some derivatives of β -1-l-rhamnosido-6-d-galactose. *Ber. Dtsch. Chem. Ges. B* **1938**, *71B*, 774–776.
- (290) Zemplen, G.; Bogner, R. Action of mercury salts on acetohalosugars. XIII. Synthesis of isoprimeverose (6- α -d-xylosido-d-glucose), the α -isomer of primeverose, and its derivatives. *Ber. Dtsch. Chem. Ges. B* **1939**, *72B*, 1160–1167.
- (291) Helferich, B.; Berger, A. Über die Synthese von Glucuroniden. *Chem. Ber.* **1957**, *90*, 2492–2498.
- (292) Helferich, B.; Zirner, J. Synthesis of tetra-O-acetylhexoses with a free 2-hydroxyl group. Synthesis of disaccharides. *Chem. Ber.* **1962**, *95*, 2604–2611.
- (293) Lemieux, R. U. Some implications in carbohydrate chemistry of theories relating to the mechanisms of replacement reactions. *Adv. Carbohydr. Chem. Biochem.* **1954**, *9*, 1–57 and references therein.
- (294) Lemieux, R.; Brice, C. A comparison of the properties of pentaacetates and methyl 1, 2-orthoacetates of glucose and mannose. *Can. J. Chem.* **1955**, *33*, 109–119.
- (295) Guthrie, R. D.; McCarthy, J. F. Acetolysis. *Adv. Carbohydr. Chem.* **1967**, *22*, 11–23.
- (296) Conchie, J.; Levvy, G.; Marsh, C. Methyl and phenyl glycosides of the common sugars. *Adv. Carbohydr. Chem.* **1957**, *12*, 157–187.
- (297) Bishop, C.; Cooper, F. Glycosidation of sugars: I. Formation of methyl-D-xylosides. *Can. J. Chem.* **1962**, *40*, 224–232.
- (298) Bishop, C.; Cooper, F. Glycosidation of sugars: II. Methanolysis of d-xylose, d-arabinose, d-lyxose, and d-ribose. *Can. J. Chem.* **1963**, *41*, 2743–2758.
- (299) Schmidt, R. R. New methods for the synthesis of glycosides and oligosaccharides - are there alternatives to the Koenigs-Knorr method. *Angew. Chem., Int. Ed. Engl.* **1986**, *25*, 212–235.
- (300) Nakayama, K.; Higashi, K.; Soga, T.; Uoto, K.; Kusama, T. Zinc triflate-promoted glycosidation: synthesis of lipid A disaccharide intermediates. *Chem. Pharm. Bull.* **1992**, *40*, 2909–2912.
- (301) Kusama, T.; Soga, T.; Ono, Y.; Kumazawa, E.; Shioya, E.; Nakayama, K.; Uoto, K.; Osada, Y. Synthesis and biological activities of lipid A analogs: modification of a glycosidically bound group with chemically stable polar acidic groups and lipophilic groups on the disaccharide backbone with tetradecanoyl or N-dodecanoylglycyl groups. *Chem. Pharm. Bull.* **1991**, *39*, 3244–3253.
- (302) Bernstein, S.; Joseph, J. P.; Dusza, J. P. Steroid conjugates. VII. Preparation of N-acetylglucosaminides of 17 α - and 17 β -estradiol. *Biochemistry* **1971**, *10*, 2941–2947.
- (303) Iida, T.; Tazawa, S.; Ohshima, Y.; Niwa, T.; Goto, J.; Nambara, T. Analysis of conjugated bile acids in human biological fluids. Synthesis of hyodeoxycholic acid 3- and 6-glycosides and related compounds. *Chem. Pharm. Bull.* **1994**, *42*, 1479–1484.
- (304) Mukherjee, D.; Sarkar, S. K.; Chowdhury, U. S.; Taneja, S. C. A rapid stereoselective C-glycosidation of indoles and pyrrole via indium trichloride promoted reactions of glycosyl halides. *Tetrahedron Lett.* **2007**, *48*, 663–667.
- (305) Lemieux, R.; Hayami, J.-i. The mechanism of the anomerization of the tetra-O-acetyl-D-glucopyranosyl chlorides. *Can. J. Chem.* **1965**, *43*, 2162–2173.
- (306) Lubineau, A.; Le Gallic, J.; Malleron, A. Stannous triflate mediated glycosidations. A stereoselective synthesis of 2-amino-2-deoxy- β -D-glucopyranosides directly with the natural N-acetyl protecting group. *Tetrahedron Lett.* **1987**, *28*, 5041–5044.
- (307) Renaudet, O.; Dumy, P. Synthesis of glycosylated- β (1–4)-amino (methoxy) and-oxyamino carbohydrate analogues. *Tetrahedron* **2002**, *58*, 2127–2135.
- (308) Steber, H. B.; Singh, Y.; Demchenko, A. V. Bismuth(III) triflate as a novel and efficient activator for glycosyl halides. *Org. Biomol. Chem.* **2021**, *19*, 3220–3233.
- (309) Lemieux, R. U.; Morgan, A. R. The abnormal conformations of pyridinium α -glycopyranosides. *Can. J. Chem.* **1965**, *43*, 2205–2213.
- (310) Lemieux, R. U.; Morgan, A. R. Mechanisms for the reactions of the anomeric tetra-O-acetyl-D-glucopyranosyl bromides with pyridine and 2-pyridone. *Can. J. Chem.* **1965**, *43*, 2214–2221.
- (311) Micheel, F.; Micheel, H. Configuration of the α - and β -forms in the sugar series. II. Configuration of glucosamine. *Ber. Dtsch. Chem. Ges. B* **1932**, *65B*, 253–258.
- (312) Hess, K.; Heumann, K. E. The question of the determination of configuration at the glucosidic carbon atom in sugars by ammonium salt formation. *Ber. Dtsch. Chem. Ges. B* **1939**, *72B*, 1495–1499.
- (313) Eby, R.; Schuerch, C. The use of positively charged leaving-groups in the synthesis of α -D-linked glucosides. Synthesis of methyl 2,3,4-tri-O-benzyl-6-O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)- α -D-glucopyranoside. *Carbohydr. Res.* **1975**, *39*, 33–38.

- (314) Igarashi, K. The Koenigs-Knorr reaction. *Adv. Carbohydr. Chem. Biochem.* **1977**, *34*, 243–283.
- (315) Kronzer, F. J.; Schuerch, C. The use of 2,3,4,6-tetra-O-benzyl- α -D-glycopyranosyl iodides in α -glycoside synthesis. *Carbohydr. Res.* **1974**, *34*, 71–78 and references therein.
- (316) Lucas, T. J.; Schuerch, C. Methanolysis as a model reaction for oligosaccharide synthesis of some 6-substituted 2,3,4-tri-O-benzyl-D-galactopyranosyl derivatives. *Carbohydr. Res.* **1975**, *39*, 39–45.
- (317) Srivastava, V. K.; Schuerch, C. Synthesis of β -D-mannopyranosides and β -L-rhamnopyranosides by glycosidation at C-1. *J. Org. Chem.* **1981**, *46*, 1121–1126.
- (318) Eby, R.; Schuerch, C. The synthesis of α - and β -(1 $\rightarrow$ 3)-linked glucopyranose disaccharides and their protein conjugates. *Carbohydr. Res.* **1982**, *102*, 131–138 and references therein.
- (319) Lemieux, R. U.; Driguez, H. The chemical synthesis of 2-O-(α -L-fucopyranosyl)-3-O-(α -D-galactopyranosyl)-D-galactose. The terminal structure of blood-group B antigenic determinant. *J. Am. Chem. Soc.* **1975**, *97*, 4069–4075.
- (320) Kartha, K. R.; Ballell, L.; Bilke, J.; McNeil, M.; Field, R. A. Iodine and its interhalogen compounds: versatile reagents in carbohydrate chemistry. XIV. Glycosylated amino acid synthesis. *J. Chem. Soc. Perkin Trans. 1* **2001**, 770–772.
- (321) Kaeothip, S.; Yasomane, J. P.; Demchenko, A. V. Glycosidation of thioglycosides in the presence of bromine: mechanism, reactivity, and stereoselectivity. *J. Org. Chem.* **2012**, *77*, 291–299.
- (322) Mannino, M. P.; Demchenko, A. V. Hydrogen-bond-mediated Aglycone Delivery (HAD) and related methods in carbohydrate chemistry. In *Carbohydrate Chemistry: Chemical and Biological Approaches (Specialist Periodical Reports)*; Rauter, A. P., Lindhorst, T. K., Queneau, Y., Eds.; RSC, 2021; Vol. 44, pp 93–116.
- (323) Yasomane, J. P.; Demchenko, A. V. Hydrogen bond-mediated aglycone delivery (HAD): a highly stereoselective synthesis of 1,2-cis α -D-glucosides from common glycosyl donors in the presence of bromine. *Chem.—Eur. J.* **2015**, *21*, 6572–6581.
- (324) Frechet, J. M.; Schuerch, C. Solid-phase synthesis of oligosaccharides. I. Preparation of the solid support. Poly[p-(1-propen-3-ol-1-yl)styrene]. *J. Am. Chem. Soc.* **1971**, *93*, 492–496.
- (325) Frechet, J. M.; Schuerch, C. Solid-phase synthesis of oligosaccharides. III. Preparation of some derivatives of di- and tri-saccharides via a simple alcoholysis reaction. *Carbohydr. Res.* **1972**, *22*, 399–412.
- (326) Serbian, I.; Prell, E.; Al-Harrasi, A.; Csuk, R. Stereoselective synthesis of alkyl pyranosides on a laboratory scale. *Mediterranean J. Chem.* **2020**, *10*, 269–276.
- (327) Cardona, A.; Boutureira, O.; Castillón, S.; Díaz, Y.; Matheu, M. I. Metal-free and VOC-free O-glycosylation in supercritical CO₂. *Green Chem.* **2017**, *19*, 2687–2694.
- (328) Takiura, K.; Honda, S.; Endo, T.; Kakehi, K. Studies of oligosaccharides. IX. Synthesis of gentiooligosaccharides by bock condensation. *Chem. Pharm. Bull.* **1972**, *20*, 438–442.
- (329) Koto, S.; Uchida, T.; Zen, S. Syntheses of isomaltose, isomaltotetraose, and isomaltoctaose. *Bull. Chem. Soc. Jpn.* **1973**, *46*, 2520–2523.
- (330) Paulsen, H. Advances in selective chemical syntheses of complex oligosaccharides. *Angew. Chem., Int. Ed. Engl.* **1982**, *21*, 155–173.
- (331) Ogawa, T.; Yamamoto, H.; Nukada, T.; Kitajima, T.; Sugimoto, M. Synthetic approach to glycan chains of a glycoprotein and proteoglycan. *Pure Appl. Chem.* **1984**, *56*, 779–795.
- (332) Depre, D.; Duffels, A.; Green, L. G.; Lenz, R.; Ley, S. V.; Wong, C. H. Synthesis of glycans from the glycodefins: two undeca-, two deca-, three nona-, an octa- and a heptasaccharide. *Chem.—Eur. J.* **1999**, *5*, 3326–3340.
- (333) Nicolaou, K. C.; Dolle, R. E.; Papahatjis, D. P.; Randall, J. L. Practical synthesis of oligosaccharides. Partial synthesis of avermectin B1a. *J. Am. Chem. Soc.* **1984**, *106*, 4189–4192.
- (334) Randall, J. L.; Nicolaou, K. C. Preparation and reactions of glycosyl fluorides. *ACS Symp. Ser.* **1988**, *374*, 13–28.
- (335) Lonn, H. Synthesis of a tri- and a heptasaccharide which contain α -L-fucopyranosyl groups and are part of the complex type of carbohydrate moiety of glycoproteins. *Carbohydr. Res.* **1985**, *139*, 105–113.
- (336) Lonn, H. Synthesis of a tetra- and a nona-saccharide which contain α -L-fucopyranosyl groups and are part of the complex type of carbohydrate moiety of glycoproteins. *Carbohydr. Res.* **1985**, *139*, 115–121.
- (337) Garegg, P. J.; Oscarson, S. A synthesis of 8-methoxycarbo-nyloct-1-yl O- α -D-galactopyranosyl-(1 $\rightarrow$ 3)-O- β -D-galactopyranosyl-(1 $\rightarrow$ 4)-2-acetamido-2-deoxy- β -D-glucopyranoside. *Carbohydr. Res.* **1985**, *136*, 207–213.
- (338) Leontein, K.; Nilsson, M.; Norberg, T. Synthesis of the methyl and 1-octyl glycosides of the P-antigen tetrasaccharide (globotetraose). *Carbohydr. Res.* **1985**, *144*, 231–240.
- (339) Paulsen, H.; Heume, M.; Nurnberger, H. Synthese der verzweigten Nonsaccharid-Sequenz der "bisected" Struktur von N-Glycoproteinen. *Carbohydr. Res.* **1990**, *200*, 127–166.
- (340) Mehta, S.; Pinto, B. M. Novel glycosidation methodology. The use of phenylselenoglycosides as glycosyl donors and acceptors in oligosaccharide synthesis. *J. Org. Chem.* **1993**, *58*, 3269–3276.
- (341) Baeschlin, D. K.; Chaperon, A. R.; Charbonneau, V.; Green, L. G.; Ley, S. V.; Lucking, U.; Walther, E. Rapid assembly of oligosaccharides: Total synthesis of a glycosylphosphatidylinositol anchor of trypanosoma brucei. *Angew. Chem., Int. Ed.* **1998**, *37*, 3423–3428.
- (342) Colley, M. A. Action des haloides libres et de quelques chlorures sur la glucose. *Ann. Chem. Phys.* **1870**, *21*, 363–377.
- (343) Kononov, L. O.; Magnusson, G. Synthesis of methyl and allyl α -glycosides of N-acetylneuraminic acid in the absence of added promoter. *Acta Chem. Scand.* **1998**, *52*, 141–144.
- (344) Shpirt, A. M.; Kononov, L. O.; Torgov, V. I.; Shibaev, V. N. Conversion of N-acetylneuraminic acid glycosyl chloride into dibenzyl glycosyl phosphate: O-glycosylation in the absence of a promoter. *Russ. Chem. Bull.* **2004**, *53*, 717–719.
- (345) Bonomi, P.; Attieh, M. D.; Gonzato, C.; Haupt, K. A New Versatile Water-Soluble Iniferter Platform for the Preparation of Molecularly Imprinted Nanoparticles by Photopolymerisation in Aqueous Media. *Chem.—Eur. J.* **2016**, *22*, 10150–10154.
- (346) Singhamahapatra, A.; Sahoo, L.; Paul, K. J. V.; Varghese, B.; Loganathan, D. Improved synthesis of per-O-acetylated C1 hydroxyglycopyranose and structural study as non-covalent organic framework. *Tetrahedron Lett.* **2013**, *54*, 6121–6124.
- (347) Ibatullin, F. M.; Selivanov, S. I. Reaction of 1,2-trans-glycosyl acetates with phosphorus pentachloride: new efficient approach to 1,2-trans-glycosyl chlorides. *Tetrahedron Lett.* **2002**, *43*, 9577–9580.
- (348) Bednarczyk, D.; Walczewska, A.; Grzywacz, D.; Sikorski, A.; Liberek, B.; Myska, H. Differently N-protected 3,4,6-tri-O-acetyl-2-amino-2-deoxy-D-glucopyranosyl chlorides and their application in the synthesis of diosgenyl 2-amino-2-deoxy- β -D-glucopyranoside. *Carbohydr. Res.* **2013**, *367*, 10–17.
- (349) Ziegler, T.; Seidl, U. Preparation of some Amygdalin-Derived Gentibiosyl Donors and Acceptors for Oligosaccharide Syntheses. *J. Carbohydr. Chem.* **1991**, *10*, 813–831.
- (350) Watt, G. M.; Boons, G.-J. A convergent strategy for the preparation of N-glycan core di-, tri-, and pentasaccharide thioaldoses for the site-specific glycosylation of peptides and proteins bearing free cysteines. *Carbohydr. Res.* **2004**, *339*, 181–193.
- (351) Iversen, T.; Bundle, D. R. Antigenic determinants of *Salmonella* serogroups A and D₁. Synthesis of trisaccharide glycosides for use as artificial antigens. *Carbohydr. Res.* **1982**, *103*, 29–40.
- (352) Encinas, L.; Chiara, J. L. Polymer-assisted solution-phase synthesis of glycosyl chlorides and bromides using a supported dialkylformamide as catalyst. *J. Comb. Chem.* **2008**, *10*, 361–363.
- (353) Gómez, A. M.; Pedregosa, A.; Casillas, M.; Uriel, C.; López, J. C. Synthesis of C-1 alkyl and aryl glycals from pyranosyl or furanosyl chlorides by treatment with organolithium reagents. *Eur. J. Org. Chem.* **2009**, *2009*, 3579–3588.

- (354) Hung, S. C.; Wong, C. H. Synthesis of glycosyl chlorides with acid-labile protecting groups. *Tetrahedron Lett.* **1996**, *37*, 4903–4906.
- (355) Ernst, B.; Winkler, T. Preparation of glycosyl halides under neutral conditions. *Tetrahedron Lett.* **1989**, *30*, 3081–3084.
- (356) Cicchillo, R. M.; Norris, P. A convenient synthesis of glycosyl chlorides from sugar hemiacetals using triphosgene as the chlorine source. *Carbohydr. Res.* **2000**, *328*, 431–434.
- (357) Traboni, S.; Liccardo, F.; Bedini, E.; Giordano, M.; Iadonisi, A. Solvent-free synthesis of glycosyl chlorides based on the triphenyl phosphine/hexachloroacetone system. *Tetrahedron Lett.* **2017**, *58*, 1762–1764.
- (358) Huy, P. H.; Filbrich, I. A General Catalytic Method for Highly Cost- and Atom-Efficient Nucleophilic Substitutions. *Chem.—Eur. J.* **2018**, *24*, 7410–7416.
- (359) Tatina, M. B.; Khong, D. T.; Judeh, Z. M. A. Efficient synthesis of α -glycosyl chlorides using 2-chloro-1,3-dimethylimidazolinium chloride: A convenient protocol for quick one-pot glycosylation. *Eur. J. Org. Chem.* **2018**, *2018*, 2208–2213.
- (360) Pongener, I.; Nikitin, K.; McGarrigle, E. M. Synthesis of glycosyl chlorides using catalytic Appel conditions. *Org. Biomol. Chem.* **2019**, *17*, 7531–7535.
- (361) Sugiyama, S.; Diakur, J. M. A convenient preparation of glycosyl chlorides from aryl/alkyl thioglycosides. *Org. Lett.* **2000**, *2*, 2713–2715.
- (362) Verma, V. P.; Wang, C.-C. Highly Stereoselective Glycosyl-Chloride-Mediated Synthesis of 2-Deoxyglucosides. *Chem.—Eur. J.* **2013**, *19*, 846–851.
- (363) Chang, C. W.; Wu, C. H.; Lin, M. H.; Liao, P. H.; Chang, C. C.; Chuang, H. H.; Lin, S. C.; Lam, S.; Verma, V. P.; Hsu, C. P.; Wang, C. C. Establishment of guidelines for the control of glycosylation reactions and intermediates by quantitative assessment of reactivity. *Angew. Chem., Int. Ed. Engl.* **2019**, *58*, 16775–16779.
- (364) Helferich, B.; Wedemeyer, K. F. Zur Darstellung von Glucosiden aus Acetobromglucose. *Justus Liebigs Ann. Chem.* **1949**, *563*, 139–145.
- (365) Chauvin, A. L.; Nepogodiev, S. A.; Field, R. A. Synthesis of an apiose-containing disaccharide fragment of rhamnogalacturonan-II and some analogues. *Carbohydr. Res.* **2004**, *339*, 21–27.
- (366) Ito, Y.; Ogawa, T. Highly stereoselective glycosylation of sialic acid aided by stereocontrolling auxiliaries. *Tetrahedron* **1990**, *46*, 89–102.
- (367) Kondo, T.; Tomoo, T.; Abe, H.; Isobe, M.; Goto, T. Simple construction of Neu5Ac(α 2–8)Neu5Ac and total synthesis of ganglioside GD₃. *J. Carbohydr. Chem.* **1996**, *15*, 857–878.
- (368) Chernyak, A.; Oscarson, S.; Turek, D. Synthesis of the Lewis b hexasaccharide and squarate acid-HSA conjugates thereof with various saccharide loadings. *Carbohydr. Res.* **2000**, *329*, 309–316.
- (369) Haberman, J. M.; Gin, D. Y. A new C(1)-auxiliary for anomeric stereocontrol in the synthesis of α -sialyl glycosides. *Org. Lett.* **2001**, *3*, 1665–1668.
- (370) Beale, T. M.; Moon, P. J.; Taylor, M. S. Organoboron-catalyzed regio- and stereoselective formation of beta-2-deoxyglycosidic linkages. *Org. Lett.* **2014**, *16*, 3604–3607.
- (371) Geringer, S. A.; Singh, Y.; Hoard, D. J.; Demchenko, A. V. A highly efficient glycosidation of glycosyl chlorides using cooperative silver(I) oxide - triflic acid catalysis. *Chem.—Eur. J.* **2020**, *26*, 8053–8063.
- (372) Castelli, R.; Schindler, S.; Walter, S. M.; Kniep, F.; Overkleef, H. S.; Van der Marel, G. A.; Huber, S. M.; Codee, J. D. C. Activation of glycosyl halides by halogen bonding. *Chem.—Asian J.* **2014**, *9*, 2095–2098.
- (373) Walter, S. M.; Kniep, F.; Herdtweck, E.; Huber, S. M. Halogen-bond-induced activation of a carbon-heteroatom bond. *Angew. Chem., Int. Ed.* **2011**, *50*, 7187–7191.
- (374) Geringer, S. A.; Demchenko, A. V. Iron(III) chloride-catalyzed activation of glycosyl chlorides. *Org. Biomol. Chem.* **2018**, *16*, 9133–9137.
- (375) An, S.; Wang, Q.; Zhu, W.; Sun, Q.; He, G.; Chen, G. Palladium-catalyzed O- and N-glycosylation with glycosyl chlorides. *CCS Chemistry* **2021**, *3*, 1821–1829.
- (376) Wang, Q.; An, S.; Deng, Z.; Zhu, W.; Huang, Z.; He, G.; Chen, G. Palladium-catalyzed C-H glycosylation for synthesis of C-aryl glycosides. *Nat. Catal.* **2019**, *2*, 793–800.
- (377) Kondo, T.; Tomoo, T.; Abe, H.; Isobe, M.; Goto, T. Total synthesis of GD₃, a ganglioside. *Chem. Lett.* **1996**, *25*, 337–338.
- (378) Gervay, J.; Nguyen, T. N.; Hadd, M. J. Mechanistic studies on the stereoselective formation of glycosyl iodides: first characterization of glycosyl iodides. *Carbohydr. Res.* **1997**, *300*, 119–125.
- (379) Bhat, A. S.; Gervay-Hague, J. Efficient Syntheses of β -Cyanosugars Using Glycosyl Iodides Derived from Per-O-silylated Mono- and Disaccharides. *Org. Lett.* **2001**, *3*, 2081–2084.
- (380) Kulkarni, S. S.; Gervay-Hague, J. Efficient Synthesis of a C-Analogue of the Immunogenic Bacterial Glycolipid BbGL2. *Org. Lett.* **2006**, *8*, 5765–5768.
- (381) Lam, S. N.; Gervay-Hague, J. Efficient Route to 2-Deoxy β -O-Aryl-d-Glycosides via Direct Displacement of Glycosyl Iodides. *Org. Lett.* **2003**, *5*, 4219–4222.
- (382) Lam, S. N.; Gervay-Hague, J. Efficient Synthesis of Man₂, Man₃, and Man₅ Oligosaccharides, Using Mannosyl Iodide Donors 1. *J. Org. Chem.* **2005**, *70*, 8772–8779.
- (383) Salvadó, M.; Amgarten, B.; Castellón, S.; Bernardes, G. J. L.; Boutureira, O. Synthesis of Fluorosugar Reagents for the Construction of Well-Defined Fluoroglycoproteins. *Org. Lett.* **2015**, *17*, 2836–2839.
- (384) Zhang, W.; Luo, X.; Wang, Z.; Zhang, J. One-pot synthesis of β -2,6-dideoxyglycosides via glycosyl iodide intermediates. *J. Carbohydr. Chem.* **2016**, *35*, 315–325.
- (385) Baldoni, L.; Marino, C. Synthetic tools for the characterization of galactofuranosyl transferases: glycosylations via acylated glycosyl iodides. *Carbohydr. Res.* **2013**, *374*, 75–81.
- (386) Walejko, P.; Baj, A. The synthesis of vitamin E sugar 1,2-orthoesters. *Monatsh. Chem.* **2019**, *150*, 275–282.
- (387) Du, W.; Kulkarni, S. S.; Gervay-Hague, J. Efficient, one-pot syntheses of biologically active α -linked glycolipids. *Chem. Commun.* **2007**, 2336–2338.
- (388) Schombs, M.; Park, F. E.; Du, W.; Kulkarni, S. S.; Gervay-Hague, J. One-Pot Syntheses of Immunostimulatory Glycolipids. *J. Org. Chem.* **2010**, *75*, 4891–4898.
- (389) Jervis, P. J.; Veerapen, N.; Bricard, G.; Cox, L. R.; Porcelli, S. A.; Besra, G. S. Synthesis and biological activity of alpha-glucosyl C24:0 and C20:2 ceramides. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 3475–3478.
- (390) Veerapen, N.; Reddington, F.; Bricard, G.; Porcelli, S. A.; Besra, G. S. Synthesis and biological activity of α -l-fucosyl ceramides, analogues of the potent agonist, α -d-galactosyl ceramide KRN7000. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 3223–3226.
- (391) Gu, X.; Chen, L.; Wang, X.; Liu, X.; You, Q.; Xi, W.; Gao, L.; Chen, G.; Chen, Y.-L.; Xiong, B.; Shen, J. Direct Glycosylation of Bioactive Small Molecules with Glycosyl Iodide and Strained Olefin as Acid Scavenger. *J. Org. Chem.* **2014**, *79*, 1100–1110.
- (392) Kulkarni, S. S.; Gervay-Hague, J. Glycosyl chlorides, bromides and iodides. In *Handbook of Chemical Glycosylation*; Demchenko, A. V., Ed.; Wiley-VCH: Weinheim, Germany, 2008; pp 59–93.
- (393) Meloncelli, P. J.; Martin, A. D.; Lowary, T. L. Glycosyl iodides. History and recent advances. *Carbohydr. Res.* **2009**, *344*, 1110–1122.
- (394) Gervay-Hague, J. Taming the reactivity of glycosyl iodides to achieve stereoselective glycosidation. *Acc. Chem. Res.* **2016**, *49*, 35–47.
- (395) Wang, H.; Cui, Y.; Zou, R.; Cheng, Z.; Yao, W.; Mao, Y.; Zhang, Y. Synthesis of oligosaccharides using per-O-trimethylsilyl-glycosyl iodides as glycosyl donor. *Carbohydr. Res.* **2016**, *427*, 1–5.
- (396) Oka, N.; Kajino, R.; Takeuchi, K.; Nagakawa, H.; Ando, K. α -Selective Ribofuranosylation of Alcohols with Ribofuranosyl Iodides and Triphenylphosphine Oxide. *J. Org. Chem.* **2014**, *79*, 7656–7664.

- (397) Houston, T. A.; Koreeda, M. Iodine-promoted ribosylation leads to a facile acetonide-forming reaction. *Carbohydr. Res.* **2009**, *344*, 2240–2244.
- (398) Park, S. S.; Gervay-Hague, J. Step-economy synthesis of β -steryl sialosides using a sialyl iodide donor. *J. Antibiot.* **2019**, *72*, 449–460.
- (399) Vannam, R.; Pote, A. R.; Pecuh, M. W. Formation and selective rupture of 1,4-anhydrosepanoses. *Tetrahedron* **2017**, *73*, 418–425.
- (400) Nogueira, J. M.; Nguyen, S. H.; Bennett, C. S. Cyclopropenium cation promoted dehydrative glycosylations using 2-deoxy- and 2,6-dideoxy-sugar donors. *Org. Lett.* **2011**, *13*, 2814–2817.
- (401) Brauns, D. Fluoro-acetyl derivatives of sugars. I. *J. Am. Chem. Soc.* **1923**, *45*, 833–835.
- (402) Olah, G. A.; Nojima, M.; Kerekes, I. Synthetic methods and reactions II. Hydrofluorination of alkenes, cyclopropane and alkynes with poly-hydrogen fluoride/pyridine (trialkylamine) reagents. *Synthesis* **1973**, *1973*, 779–780.
- (403) Sharma, M. N.; Eby, R. Synthesis and conformational studies of 2- β -chloro, 2- α -fluoro, and 2- β -fluoro derivatives of 2-deoxy-N-acetylneuraminic acid. *Carbohydr. Res.* **1984**, *127*, 201–210.
- (404) Hashimoto, S.; Hayashi, M.; Noyori, R. Glycosylation using glucopyranosyl fluorides and silicon-based catalysts. Solvent dependency of the stereoselection. *Tetrahedron Lett.* **1984**, *25*, 1379–1382.
- (405) Hayashi, M.; Hashimoto, S.-i.; Noyori, R. Simple synthesis of glycosyl fluorides. *Chem. Lett.* **1984**, *13*, 1747–1750.
- (406) Szarek, W. A.; Gryniewicz, G.; Doboszewski, B.; Hay, G. W. The synthesis of glycosyl fluorides using pyridinium poly (hydrogen fluoride). *Chem. Lett.* **1984**, *13*, 1751–1754.
- (407) Helferich, B.; Gootz, R. Über Fluor-Derivate von Kohlehydraten. *Ber. Dtsch. Chem. Ges.* **1929**, *62*, 2505–2507.
- (408) Micheel, F.; Klemer, A. Über den Reaktionsmechanismus der Glykosidbildung aus α - und β -1-Fluor-Derivaten der d-Glucose und d-Mannose. *Chem. Ber.* **1958**, *91*, 663–667.
- (409) Micheel, F.; Klemer, A. Glycosyl fluorides and azides. *Adv. Carbohydr. Chem.* **1962**, *16*, 85–103.
- (410) Hall, L.; Manville, J.; Bhacca, N. Studies of specifically fluorinated carbohydrates. Part I. Nuclear magnetic resonance studies of hexopyranosyl fluoride derivatives. *Can. J. Chem.* **1969**, *47*, 1–17.
- (411) Igarashi, K.; Honma, T.; Irisawa, J. Reaction of glycosyl chlorides with silver tetrafluoroborate. *Carbohydr. Res.* **1969**, *11*, 577–578.
- (412) Goggin, K. D.; Lambert, J. F.; Walinsky, S. W. Synthesis of peraceto- β -D-glycosyl fluorides using zinc fluoride. *Synlett* **1994**, *1994*, 162–164.
- (413) Naumann, D.; Möckel, R.; Tyrre, W. Difluoromethyl compounds of chalcogens: A NMR study of the reactions of Cd (CF₃)₂· 2MeCN and ZnBr (CF₃)₂· 2MeCN with dialkyl chalcogenides and boron trifluoride. *Angew. Chem., Int. Ed.* **1994**, *33*, 323–325.
- (414) Miethchen, R.; Hein, M.; Naumann, D.; Tyrre, W. Organofluorine compounds and fluorinating agents, 15. O-difluoromethylations of monosaccharides by trifluoromethylzinc bromide. *Liebigs Ann.* **1995**, *1995*, 1717–1719.
- (415) Tyrre, W.; Naumann, D.; Pasenok, S. V.; Yagupolskii, Y. L. Carbenoid reactions of trifluoromethylelement compounds. Part 4. Reactions of trifluoromethylzinc bromide with enamines and methylene bases. *J. Fluorine Chem.* **1995**, *70*, 181–185.
- (416) Tyrre, W.; Naumann, D. Trifluormethylzinkbromid - ein bequemes handhabbares Trifluor- und Difluormethylierungsmittel. *Journal für Praktische Chemie/Chemiker-Zeitung* **1996**, *338*, 283–286.
- (417) Miethchen, R.; Hager, C.; Hein, M. Organofluorine compounds and fluorinating agents; 18: trifluoromethylzinc bromide as a reagent for the preparation of glycosyl fluorides. *Synthesis* **1997**, *1997*, 159–161.
- (418) Miethchen, R.; Kolp, G.; Peters, D.; Holz, J. Reactions with and in water-free hydrogen fluoride. Part 3. Selective syntheses of glycosyl fluorides. *Z. Chem.* **1990**, *2* (30), 56.
- (419) Miethchen, R.; Kolp, G. Reactions with and in anhydrous hydrogen fluoride systems. Part 8. Triethylamine trishydrofluoride—a convenient reagent for the stereoselective synthesis of glycosyl fluorides. *J. Fluorine Chem.* **1993**, *60*, 49–55.
- (420) Mukaiyama, T.; Hashimoto, Y.; Shoda, S. Stereoselective synthesis of 1,2-cis-glycofuranosides using glycofuranosyl fluorides. *Chem. Lett.* **1983**, *12*, 935–938.
- (421) Mitsunobu, O. The use of diethyl azodicarboxylate and triphenylphosphine in synthesis and transformation of natural products. *Synthesis* **1981**, *1981*, 1–28.
- (422) Loibner, H.; Zbiral, E. Reaktionen mit phosphororganischen Verbindungen. XLI [1]. Neuartige synthetische Aspekte des Systems Triphenylphosphin-Azodicarbonsäureester-Hydroxyverbindung. *Helv. Chim. Acta* **1976**, *59*, 2100–2113.
- (423) Kunz, H.; Sager, W. Stereoselective glycosylation of alcohols and silyl ethers using glycosyl fluorides and boron trifluoride etherate. *Helv. Chim. Acta* **1985**, *68*, 283–287.
- (424) Penglis, A. A. E. Fluorinated carbohydrates. *Adv. Carbohydr. Chem. Biochem.* **1981**, *38*, 195–285.
- (425) Rosenbrook, W., Jr.; Riley, D. A.; Lartey, P. A. A new method for the synthesis of glycosyl fluorides. *Tetrahedron Lett.* **1985**, *26*, 3–4.
- (426) Posner, G. H.; Haines, S. R. A convenient, one-step, high-yield replacement of an anomeric hydroxyl group by a fluorine atom using DAST. Preparation of glycosyl fluorides. *Tetrahedron Lett.* **1985**, *26*, 5–8.
- (427) Devos, A.; Remion, J.; Frisque-Hesbain, A.-M.; Colens, A.; Ghosez, L. Synthesis of acyl halides under very mild conditions. *J. Chem. Soc., Chem. Commun.* **1979**, 1180–1181.
- (428) Schmidt, U.; Lieberknecht, A.; Griesser, H.; Utz, R.; Beuttler, T.; Bartkowiak, F. Amino acids and peptides. 55. Synthesis of biologically active cyclopeptides. 7. Total synthesis of chlamydocin. *Synthesis* **1986**, *1986*, 361–366.
- (429) Ernst, B.; Wagner, B. Preliminary communication. *Helv. Chim. Acta* **1989**, *72*, 165–171.
- (430) Abdul-Ghani, M.; Banks, E. R.; Besheesh, M. K.; Sharif, I.; Syvret, R. G. N-Halogeno compounds. Part 14. “Transfer fluorination” of quinuclidine using F-TEDA-BF₃ (Selectfluor™ reagent): laboratory synthesis of N-fluoroquinuclidinium salts not requiring the use of elemental fluorine. *J. Fluorine Chem.* **1995**, *73*, 255–257.
- (431) Burkart, M. D.; Zhang, Z.; Hung, S.-C.; Wong, C.-H. A new method for the synthesis of fluoro-carbohydrates and glycosides using selectfluor. *J. Am. Chem. Soc.* **1997**, *119*, 11743–11746.
- (432) Middleton, W. J. Explosive hazards with DAST (diethylaminosulfur trifluoride). *Chem. Eng. News* **1979**, *57*, 43.
- (433) Lal, G. S.; Pez, G. P.; Pesaresi, R. J.; Prozone, F. M.; Cheng, H. Bis (2-methoxyethyl) aminosulfur trifluoride: a new broad-spectrum deoxofluorinating agent with enhanced thermal stability. *J. Org. Chem.* **1999**, *64*, 7048–7054.
- (434) Kobayashi, S.; Yoneda, A.; Fukuhara, T.; Hara, S. Selective synthesis of fluorinated carbohydrates using N, N-diethyl- α , α -difluoro-(m-methylbenzyl) amine. *Tetrahedron Lett.* **2004**, *45*, 1287–1289.
- (435) Kobayashi, S.; Yoneda, A.; Fukuhara, T.; Hara, S. Deoxyfluorination of alcohols using N, N-diethyl- α , α -difluoro-(m-methylbenzyl) amine. *Tetrahedron* **2004**, *60*, 6923–6930.
- (436) Kim, S.; Khomutnyk, Y.; Bannykh, A.; Nagorny, P. Synthesis of glycosyl fluorides by photochemical fluorination with sulfur(VI) hexafluoride. *Org. Lett.* **2021**, *23*, 190–194.
- (437) Kim, S.; Nagorny, P. Electrochemical synthesis of glycosyl fluorides using sulfur(VI) hexafluoride as the fluorinating agent. *Org. Lett.* **2022**, *24*, 2294–2298.
- (438) Horne, G.; Mackie, W. Preparation of glycosyl fluorides from phenyl 1-seleno- and phenyl 1-telluroglycosides. *Tetrahedron Lett.* **1999**, *40*, 8697–8700.
- (439) Edmunds, J. J.; Motherwell, W. B. Observations on the reaction of some electron rich dienes with hypervalent iodoarene difluorides: a novel mechanism for fluorination. *J. Chem. Soc., Chem. Commun.* **1989**, 881–883.

- (440) Caddick, S.; Motherwell, W. B.; Wilkinson, J. A. A concise method for the preparation of glycosyl fluorides via displacement reactions of 1-arylthioglycosides with 4-methyl (difluoroiodo) benzene. *J. Chem. Soc., Chem. Commun.* **1991**, 674–675.
- (441) Caddick, S.; Gazzard, L.; Motherwell, W.; Wilkinson, J. Preparation of 1-fluoroglycosides from 1-arylthio and 1-arylselenoglycosides using 4-methyl (difluoroiodo) benzene. *Tetrahedron* **1996**, *52*, 149–156.
- (442) Yoshida, J. i.; Suga, S. Basic concepts of “cation pool” and “cation flow” methods and their applications in conventional and combinatorial organic synthesis. *Chem.—Eur. J.* **2002**, *8*, 2650–2658.
- (443) Suzuki, S.; Matsumoto, K.; Kawamura, K.; Suga, S.; Yoshida, J. i. Generation of alkoxy-carbenium ion pools from thioacetals and applications to glycosylation chemistry. *Org. Lett.* **2004**, *6*, 3755–3758.
- (444) Gordon, D. M.; Danishefsky, S. J. Displacement reactions of a 1, 2-anhydro- α -D-hexopyranose: installation of useful functionality at the anomeric carbon. *Carbohydr. Res.* **1990**, *206*, 361–366.
- (445) Miethchen, R.; Gabriel, T.; Kolp, G. Reaktionen mit und in wasserfreiem Fluorwasserstoff 7.1 Von Isopropylidenzuckern zu acylierten Pyranosyl- oder Furanosylfluoriden in einem Schritt. *Synthesis* **1991**, 1991, 885–888.
- (446) Shimizu, M.; Nakahara, Y.; Yoshioka, H. Selective bromofluorination of alkenes with 1,3-dibromo-5,5-dimethylhydantoin and silicon tetrafluoride. *J. Chem. Soc., Chem. Commun.* **1989**, 1881–1882.
- (447) Shimizu, M.; Nakahara, Y.; Yoshioka, H. Stereocontrolled halofluorination of glycals with silicon tetrafluoride, leading to a facile synthesis of glycosyl fluorides. *J. Fluorine Chem.* **1999**, *97*, 57–60.
- (448) Pearson, A. G.; Kiefel, M. J.; Ferro, V.; von Itzstein, M. Towards the synthesis of aryl glucuronides as potential heparanase probes. An interesting outcome in the glycosidation of glucuronic acid with 4-hydroxycinnamic acid. *Carbohydr. Res.* **2005**, *340*, 2077–2085.
- (449) Koivula, A.; Ruohonen, L.; Wohlfahrt, G.; Reinikainen, T.; Teeri, T. T.; Piens, K.; Claeyssens, M.; Weber, M.; Vasella, A.; Becker, D.; Sinnott, M. L.; Zou, J.-y.; Kleywegt, G. J.; Szardenings, M.; Ståhlberg, J.; Jones, T. A. The active site of cellobiohydrolase cel6A from *Trichoderma reesei*: The roles of aspartic acids D221 and D175. *J. Am. Chem. Soc.* **2002**, *124*, 10015–10024.
- (450) Martin, A.; Arda, A.; Désiré, J.; Martin-Mingot, A.; Probst, N.; Sinaÿ, P.; Jiménez-Barbero, J.; Thibaudeau, S.; Blériot, Y. Catching elusive glycosyl cations in a condensed phase with HF/SbF₅ superacid. *Nat. Chem.* **2016**, *8*, 186.
- (451) Mukaiyama, T.; Maeshima, H.; Jona, H. Catalytic and stereoselective glycosylation with disarmed glycosyl fluoride by using a combination of stannous (II) chloride (SnCl₂) and silver tetrakis (pentafluorophenyl) borate [AgB (C₆F₅)₄] as a catalyst. *Chem. Lett.* **2001**, *30*, 388–389.
- (452) Ogawa, T.; Sugimoto, M. Synthesis of α -Neu5Acp-(2 $\rightarrow$ 3)-D-Gal and α -Neu5Acp-(2 $\rightarrow$ 3)- β -D-Galp-(1 $\rightarrow$ 4)-D-Glc. *Carbohydr. Res.* **1985**, *135*, c5–c9.
- (453) Jona, H.; Maeshima, H.; Mukaiyama, T. Catalytic and stereoselective glycosylation with disarmed glycosyl fluoride having phthaloyl or dichlorophthaloyl protected amino function using a 1:2 combination of stannic chloride and silver tetrakis (pentafluorophenyl) borate. *Chem. Lett.* **2001**, *30*, 726–727.
- (454) Okada, Y.; Asakura, N.; Bando, M.; Ashikaga, Y.; Yamada, H. Completely β -selective glycosylation using 3,6-O-(*o*-xylylene)-bridged axial-rich glycosyl fluoride. *J. Am. Chem. Soc.* **2012**, *134*, 6940–6943.
- (455) Nicolaou, K.; Chucholowski, A.; Dolle, R. E.; Randall, J. L. Reactions of glycosyl fluorides. Synthesis of O-, S-, and N-glycosides. *J. Chem. Soc., Chem. Commun.* **1984**, 1155–1156.
- (456) Kreuzer, M.; Thiem, J. Synthesis of oligosaccharides with glycosyl fluorides under Lewis acid catalysis. *Carbohydr. Res.* **1986**, *149*, 347–361.
- (457) Maeta, H.; Matsumoto, T.; Suzuki, K. “Dibutyltin diperchlorate” for activation of glycosyl fluoride. *Carbohydr. Res.* **1993**, *249*, 49–56.
- (458) Kim, W.-S.; Sasai, H.; Shibasaki, M. β -Selective glycosylation with α -mannosyl fluorides using tin (II) triflate and lanthanum perchlorate. *Tetrahedron Lett.* **1996**, *37*, 7797–7800.
- (459) Kandasamy, J.; Hurevich, M.; Seeberger, P. H. Automated solid phase synthesis of oligoarabinofuranosides. *Chem. Commun.* **2013**, *49*, 4453–4455.
- (460) Wessel, H. P. Comparison of catalysts in α -glucosylation reactions and identification of triflic anhydride as a new reactive promoter. *Tetrahedron Lett.* **1990**, *31*, 6863–6866.
- (461) Kunz, H.; Waldmann, H. Directed stereoselective synthesis of α - and β -N-acetyl neuraminic acid-galactose disaccharides using 2-chloro and 2-fluoro derivatives of neuraminic acid allyl ester. *J. Chem. Soc., Chem. Commun.* **1985**, 638–640.
- (462) Takeuchi, K.; Mukaiyama, T. Trityl tetrakis (pentafluorophenyl) borate catalyzed stereoselective glycosylation using glycopyranosyl fluoride as a glycosyl donor. *Chem. Lett.* **1998**, *27*, 555–556.
- (463) Mukaiyama, T.; Yanagisawa, M.; Iida, D.; Hachiya, I. A novel carbocationic species paired with tetrakis (pentafluorophenyl) borate anion in catalytic Aldol reaction. *Chem. Lett.* **2000**, *29*, 606–607.
- (464) Jona, H.; Mandai, H.; Mukaiyama, T. A catalytic and stereoselective glycosylation with glucopyranosyl fluoride by using various protic acids. *Chem. Lett.* **2001**, *30*, 426–427.
- (465) Jona, H.; Mandai, H.; Chavasiri, W.; Takeuchi, K.; Mukaiyama, T. Protic acid catalyzed stereoselective glycosylation using glycosyl fluorides. *Bull. Chem. Soc. Jpn.* **2002**, *75*, 291–309 and references therein.
- (466) Sati, G.; Martin, J.; Xu, Y.; Malakar, T.; Zimmerman, P. M.; Montgomery, J. Fluoride migration catalysis enables simple, stereoselective, and iterative glycosylation. *J. Am. Chem. Soc.* **2020**, *142*, 7235–7242.
- (467) Kobayashi, S.; Koide, K.; Ohno, M. Gallium reagents in organic synthesis: Dimethylgallium chloride and triflate as activators in glycosidation using glycopyranosyl fluorides. *Tetrahedron Lett.* **1990**, *31*, 2435–2438.
- (468) Bohm, G.; Waldmann, H. Synthesis of glycosides of fucose under neutral conditions in solutions of LiClO₄ in organic solvents. *Tetrahedron Lett.* **1995**, *36*, 3843–3846.
- (469) Böhm, G.; Waldmann, H. O-Glycoside Synthesis under Neutral Conditions in Concentrated Solutions of LiClO₄ in Organic Solvents Employing O-Acyl-Protected Glycosyl Donors. *Liebigs Ann.* **1996**, 1996, 621–625.
- (470) Pelletier, G.; Zwicker, A.; Allen, C. L.; Schepartz, A.; Miller, S. J. Aqueous glycosylation of unprotected sucrose employing glycosyl fluorides in the presence of calcium ion and trimethylamine. *J. Am. Chem. Soc.* **2016**, *138*, 3175–3182.
- (471) Wadzinski, T. J.; Steinauer, A.; Hie, L.; Pelletier, G.; Schepartz, A.; Miller, S. J. Rapid phenolic O-glycosylation of small molecules and complex unprotected peptides in aqueous solvent. *Nat. Chem.* **2018**, *10*, 644–652.
- (472) Junnemann, J.; Lundt, I.; Thiem, J. 2-Deoxyglycosyl fluorides in oligosaccharide synthesis. *Liebigs Ann. Chem.* **1991**, 1991, 759–764.
- (473) Partridge, K. M.; Bader, S. J.; Buchan, Z. A.; Taylor, C. E.; Montgomery, J. A streamlined strategy for aglycone assembly and glycosylation. *Angew. Chem., Int. Ed.* **2013**, *52*, 13647–13650.
- (474) Matsumoto, T.; Maeta, H.; Suzuki, K.; Tsuchihashi, G. I. New glycosidation reaction 1. Combination use of Cp₂ZrCl₂-AgClO₄ for activation of glycosyl fluorides and application to highly β -selective glycosidation of D-mycinoose. *Tetrahedron Lett.* **1988**, *29*, 3567–3570.
- (475) Suzuki, K.; Maeta, H.; Suzuki, T.; Matsumoto, T. Cp₂ZrCl₂-AgBF₄ in benzene: a new reagent system for rapid and highly selective α -mannoside synthesis from tetra-O-benzyl-D-mannosyl fluoride. *Tetrahedron Lett.* **1989**, *30*, 6879–6882.
- (476) Toshima, K.; Kasumi, K.; Matsumura, S. Novel stereocontrolled glycosidations using a solid acid, SO₄/ZrO₂, for direct syntheses of α - and β -mannopyranosides. *Synlett* **1998**, 1998, 643–645.
- (477) Suzuki, K.; Maeta, H.; Matsumoto, T.; Tsuchihashi, G. I. New glycosidation reaction 2. Preparation of 1-fluoro-D-desosamine

derivative and its efficient glycosidation by the use of $\text{Cp}_2\text{HfCl}_2\text{-AgClO}_4$ as the activator. *Tetrahedron Lett.* **1988**, 29, 3571–3574.

(478) Suzuki, K.; Maeta, H.; Matsumoto, T. An improved procedure for metallocene-promoted glycosidation. Enhanced reactivity by employing 1:2-ratio of $\text{Cp}_2\text{HfCl}_2\text{-AgClO}_4$. *Tetrahedron Lett.* **1989**, 30, 4853–4856.

(479) Manabe, S.; Ito, Y. Hafnium (IV) tetratriflate as a glycosyl fluoride activation reagent. *J. Org. Chem.* **2013**, 78, 4568–4572.

(480) Yamada, H.; Hayashi, T. A substrate-unspecific glycosylation reaction promoted by copper (II) trifluoromethanesulfonate in benzotrifluoride. *Carbohydr. Res.* **2002**, 337, S81–S85.

(481) Hosono, S.; Kim, W. S.; Sasai, H.; Shibasaki, M. A new glycosylation procedure utilizing rare earth salts and glycosyl fluorides, with or without the requirement of Lewis acid. *J. Org. Chem.* **1995**, 60, 4–5.

(482) Kim, W. S.; Hosono, S.; Sasai, H.; Shibasaki, M. Rare earth perchlorate catalyzed glycosidation of glycosyl fluorides with trimethylsilyl ethers. *Tetrahedron Lett.* **1995**, 36, 4443–4446.

(483) Yu, L.; Chen, D.; Li, J.; Wang, P. G. Preparation, characterization, and synthetic uses of lanthanide (III) catalysts supported on ion exchange resins. *J. Org. Chem.* **1997**, 62, 3575–3581.

(484) Yamanai, T.; Nagayama, S.; Ishida, H.-k.; Nishikido, J.; Inazu, T. Ytterbium (III) tris {bis (perfluoro-butylsulfonyl) amide} as a Lewis acid for glycosylations. *Synth. Commun.* **2001**, 31, 899–903.

(485) Packard, G. K.; Rychnovsky, S. D. β -Selective glycosylations with masked D-mycosamine precursors. *Org. Lett.* **2001**, 3, 3393–3396.

(486) Mukaiyama, T.; Takeuchi, K.; Jona, H.; Maeshima, H.; Saitoh, T. A catalytic and stereoselective glycosylation with β -glycosyl fluorides. *Helv. Chim. Acta* **2000**, 83, 1901–1918.

(487) Mukaiyama, T.; Jona, H.; Takeuchi, K. Trifluoromethanesulfonic acid (TfOH)-catalyzed stereoselective glycosylation using glycosyl fluoride. *Chem. Lett.* **2000**, 29, 696–697.

(488) Jona, H.; Takeuchi, K.; Mukaiyama, T. 1,2-Cis selective glycosylation with glycosyl fluoride by using a catalytic amount of trifluoromethanesulfonic acid (TfOH) in the coexistence of molecular sieve 5A(MSSA). *Chem. Lett.* **2000**, 29, 1278–1279.

(489) Yanagisawa, M.; Mukaiyama, T. Catalytic and stereoselective glycosylation with glycosyl fluoride using active carbocationic species paired with tetrakis(pentafluorophenyl)borate or trifluoromethanesulfonate. *Chem. Lett.* **2001**, 30, 224–225.

(490) Chan, J.; Tang, A.; Bennet, A. J. A stepwise solvent-promoted $\text{S}_{\text{N}}\text{i}$ reaction of α -D-glucopyranosyl fluoride: Mechanistic implications for retaining glycosyltransferases. *J. Am. Chem. Soc.* **2012**, 134, 1212–1220.

(491) Manabe, Y.; Matsumoto, T.; Ikinaga, Y.; Tsutsui, Y.; Sasaya, S.; Kadanaga, Y.; Konishi, A.; Yasuda, M.; Uto, T.; Dai, C.; Yano, K.; Shimoyama, A.; Matsuda, A.; Fukase, K. Revisiting glycosylations using glycosyl fluoride by $\text{BF}_3\text{-Et}_2\text{O}$: activation of disarmed glycosyl fluorides with high catalytic turnover. *Org. Lett.* **2022**, 24, 6–10.

(492) Nielsen, M. M.; Qiao, Y.; Wang, Y.; Pedersen, C. M. Vessel effect in C-F bond activation prompts revised mechanism and reveals an autocatalytic glycosylation. *Eur. J. Org. Chem.* **2020**, 2020, 140–144.

(493) Mukaiyama, T.; Suenaga, M.; Chiba, H.; Jona, H. Highly α -selective glycosylation with glycopyranosyl fluorides having diethylthiocarbamoyl group. *Chem. Lett.* **2002**, 31, 56–57.

(494) Stephan, D. W. The broadening reach of frustrated Lewis pair chemistry. *Science* **2016**, 354, aaf7229.

(495) Caputo, C. B.; Stephan, D. W. Activation of alkyl C-F bonds by B (C6F5) 3: stoichiometric and catalytic transformations. *Organometallics* **2012**, 31, 27–30.

(496) Levin, M. D.; Chen, T. Q.; Neubig, M. E.; Hong, C. M.; Theulier, C. A.; Kobylanski, I. J.; Janabi, M.; O'Neil, J. P.; Toste, F. D. A catalytic fluoride-rebound mechanism for C (sp³)-CF₃ bond formation. *Science* **2017**, 356, 1272–1276.

(497) Chitnis, S. S.; LaFortune, J. H.; Cummings, H.; Liu, L. L.; Andrews, R.; Stephan, D. W. Phosphorus coordination chemistry in

catalysis: air stable P (III)-dications as Lewis acid catalysts for the allylation of C-F bonds. *Organometallics* **2018**, 37, 4540–4544.

(498) Böhm, G.; Waldmann, H. O-Glycoside Synthesis under Neutral Conditions in Concentrated Solutions of LiClO_4 in Organic Solvents Employing Benzyl-Protected Glycosyl Donors. *Liebigs Ann.* **1996**, 1996, 613–619.

(499) Delacroix, S.; Bonnet, J.-P.; Courty, M.; Postel, D.; Van Nhien, A. N. Glycosylation mediated—BAIL in aqueous solution. *Carbohydr. Res.* **2013**, 381, 12–18.

(500) Yoshida, N.; Noguchi, M.; Tanaka, T.; Matsumoto, T.; Aida, N.; Ishihara, M.; Kobayashi, A.; Shoda, S. i. Direct dehydrative pyridylthio-glycosidation of unprotected sugars in aqueous media using 2-chloro-1, 3-dimethylimidazolium chloride as a condensing agent. *Chem.—Asian J.* **2011**, 6, 1876–1885.

(501) Sato, S.; Naito, Y.; Aoki, K. Scandium cation-exchanged montmorillonite catalyzed direct C-glycosylation of a 1, 3-diketone, dimedone, with unprotected sugars in aqueous solution. *Carbohydr. Res.* **2007**, 342, 913–918.

(502) Banait, N. S.; Jencks, W. P. Reactions of anionic nucleophiles with α -D-glucopyranosyl fluoride in aqueous solution through a concerted, $\text{A}_{\text{N}}\text{D}_{\text{N}}$ ($\text{S}_{\text{N}}2$) mechanism. *J. Am. Chem. Soc.* **1991**, 113, 7951–7958.

(503) Banait, N. S.; Jencks, W. P. General-acid and general-base catalysis of the cleavage of α -D-glucopyranosyl fluoride. *J. Am. Chem. Soc.* **1991**, 113, 7958–7963.

(504) Sinnott, M. L.; Jencks, W. P. Solvolysis of D-glucopyranosyl derivatives in mixtures of ethanol and 2, 2, 2-trifluoroethanol. *J. Am. Chem. Soc.* **1980**, 102, 2026–2032.

(505) Chan, J.; Sannikova, N.; Tang, A.; Bennet, A. J. Transition-state structure for the quintessential $\text{S}_{\text{N}}2$ reaction of a carbohydrate: Reaction of α -glucopyranosyl fluoride with azide ion in water. *J. Am. Chem. Soc.* **2014**, 136, 12225–12228.

(506) Christofides, J. C.; Davies, D. B.; Martin, J. A.; Rathbone, E. B. Intramolecular hydrogen bonding in 1'-sucrose derivatives determined by SIMPLE proton NMR spectroscopy. *J. Am. Chem. Soc.* **1986**, 108, 5738–5743.

(507) Christofides, J. C.; Davies, D. B. Co-operative and competitive hydrogen bonding in sucrose determined by SIMPLE 1 H nmr spectroscopy. *J. Chem. Soc., Chem. Commun.* **1985**, 1533–1534.

(508) Christofides, J. C.; Davies, D. B. Secondary isotope multiplet NMR spectroscopy of partially labeled entities. Carbon-13 SIMPLE NMR of carbohydrates. *J. Am. Chem. Soc.* **1983**, 105, 5099–5105.

(509) Anderson, C. E.; Pickrell, A. J.; Sperry, S. L.; Vasquez Jr, T. E.; Custer, T. G.; Fierman, M. B.; Lazar, D. C.; Brown, Z. W.; Iskenderian, W. S.; Hickstein, D. D. NMR detection of intramolecular OH/OH hydrogen bond networks: an approach using isotopic perturbation and hydrogen bond mediated OH . . . OH J-coupling. *Heterocycles* **2007**, 72, 469–495.

(510) O'Leary, D. J.; Hickstein, D. D.; Hansen, B. K.; Hansen, P. E. Theoretical and NMR studies of deuterium isotopic perturbation of hydrogen bonding in symmetrical dihydroxy compounds. *J. Org. Chem.* **2010**, 75, 1331–1342.

(511) Craig, B. N.; Janssen, M. U.; Wickersham, B. M.; Rabb, D. M.; Chang, P. S.; O'Leary, D. J. Isotopic perturbation of intramolecular hydrogen bonds in rigid 1, 3-diols: NMR studies reveal unusually large equilibrium isotope effects. *J. Org. Chem.* **1996**, 61, 9610–9613.

(512) Vasquez, T. E.; Bergset, J. M.; Fierman, M. B.; Nelson, A.; Roth, J.; Khan, S. I.; O'Leary, D. J. Using equilibrium isotope effects to detect intramolecular OH/OH hydrogen bonds: Structural and solvent effects. *J. Am. Chem. Soc.* **2002**, 124, 2931–2938.

(513) Boutureira, O.; Bernardes, G. J.; D'Hooge, F.; Davis, B. G. Direct radiolabelling of proteins at cysteine using [18 F]-fluorosugars. *Chem. Commun.* **2011**, 47, 10010–10012.

(514) Adamo, R.; Nilo, A.; Castagner, B.; Boutureira, O.; Berti, F.; Bernardes, G. J. Synthetically defined glycoprotein vaccines: current status and future directions. *Chem. Sci.* **2013**, 4, 2995–3008.

(515) Boutureira, O.; Bernardes, G. J. L.; Fernández-González, M.; Anthony, D. C.; Davis, B. G. Selenenylsulfide-Linked Homogeneous

Glycopeptides and Glycoproteins: Synthesis of Human "Hepatic Se Metabolite A. *Angew. Chem., Int. Ed.* **2012**, *51*, 1432–1436.

(516) Ribeiro Morais, G.; Falconer, R. A.; Santos, I. Carbohydrate-based molecules for molecular imaging in nuclear medicine. *Eur. J. Org. Chem.* **2013**, *2013*, 1401–1414.

(517) Grayson, E. J.; Bernardes, G. J.; Chalker, J. M.; Boutureira, O.; Koeppe, J. R.; Davis, B. G. A coordinated synthesis and conjugation strategy for the preparation of homogeneous glycoconjugate vaccine candidates. *Angew. Chem., Int. Ed.* **2011**, *50*, 4127–4132.

(518) Gamblin, D. P.; Scanlan, E. M.; Davis, B. G. Glycoprotein synthesis: an update. *Chem. Rev.* **2009**, *109*, 131–163.

(519) Stubbs, J. M.; Marx, D. Glycosidic bond formation in aqueous solution: on the oxocarbenium intermediate. *J. Am. Chem. Soc.* **2003**, *125*, 10960–10962.

(520) Matsumoto, T.; Maeta, H.; Suzuki, K. First total synthesis of mycinamicin IV and VII.: Successful application of new glycosidation reaction. *Tetrahedron Lett.* **1988**, *29*, 3575–3578.

(521) Suzuki, K.; Matsumoto, T. New methods in glycoside synthesis by using early transition metal-derived reagent. *J. Synth. Org. Chem. Jpn.* **1993**, *51*, 718–732.

(522) Toshima, K.; Kasumi, K.-i.; Matsumura, S. Novel stereocontrolled glycosidations of 2-deoxyglucopyranosyl fluoride using a heterogeneous solid acid, sulfated zirconia (SO₄/ZrO₂). *Synlett* **1999**, *1999*, 813–815.

(523) Hachiya, I.; Moriaki, M.; Kobayashi, S. Catalytic Friedel-Crafts acylation reactions using hafnium triflate as a catalyst in lithium perchlorate-nitromethane. *Tetrahedron Lett.* **1995**, *36*, 409–412.

(524) Waller, F. J.; Barrett, A. G.; Braddock, D. C.; Ramprasad, D. Hafnium (IV) and zirconium (IV) triflates as superior recyclable catalysts for the atom economic nitration of o-nitrotoluene. *Tetrahedron Lett.* **1998**, *39*, 1641–1642.

(525) Noji, M.; Konno, Y.; Ishii, K. Metal triflate-catalyzed cationic benzylation and allylation of 1, 3-dicarbonyl compounds. *J. Org. Chem.* **2007**, *72*, 5161–5167.

(526) Wu, Y.-C.; Zhu, J. Hafnium trifluoromethanesulfonate (hafnium triflate) as a highly efficient catalyst for chemoselective thioacetalization and transthioacetalization of carbonyl compounds. *J. Org. Chem.* **2008**, *73*, 9522–9524.

(527) Braccini, I.; Derouet, C.; Esnault, J.; de Penhoat, C.; Mallet, J.-M.; Michon, V.; Sinaÿ, P. Conformational analysis of nitrilium intermediates in glycosylation reactions. *Carbohydr. Res.* **1993**, *246*, 23–41.

(528) Satoh, H.; Hansen, H. S.; Manabe, S.; van Gunsteren, W. F.; Hunenberger, P. H. Theoretical investigation of solvent effects on glycosylation reactions: stereoselectivity controlled by preferential conformations of the intermediate oxocarbenium-counterion complex. *J. Chem. Theory Comput.* **2010**, *6*, 1783–1797.

(529) Kleinschmidt, P.; Lau, K.; Hildenbrand, D. Thermochemistry of the gaseous fluorides of samarium, europium, and thulium. *J. Chem. Phys.* **1981**, *74*, 653–660.

(530) Zmbov, K. F.; Margrave, J. L. Mass spectrometric studies at high temperatures. XII. Stabilities of dysprosium, holmium, and erbium subfluorides. *J. Phys. Chem.* **1966**, *70*, 3379–3382.

(531) Di Bernardo, P.; Choppin, G. R.; Portanova, R.; Zanonato, P. L. Lanthanide (III) trifluoromethanesulfonate complexes in anhydrous acetonitrile. *Inorg. Chim. Acta* **1993**, *207*, 85–91.

(532) Favier, F.; Pascal, J.-L. Synthesis and structural analysis of a homogeneous series of anhydrous rare-earth-metal perchlorates. *J. Chem. Soc., Dalton Trans.* **1992**, *1992*, 1997–2002.

(533) Bünzli, J.-C. G.; Kasperek, V. FT-IR and fluorometric investigation of rare-earth and metal ion solvation. Part 10. Inner-sphere interaction between perchlorate and lanthanide ions in anhydrous acetonitrile. *Inorg. Chim. Acta* **1991**, *182*, 101–107.

(534) Buenzli, J. C. G.; Yersin, J. R.; Mabillard, C. FT IR and fluorometric investigation of rare earth and metallic ion solvation. 1. Europium perchlorate in anhydrous acetonitrile. *Inorg. Chem.* **1982**, *21*, 1471–1476.

(535) Qiu, D.; Schmidt, R. R. Glycosyl imidates, 52. Synthesis of globotriaosylceramide (Gb3) and isoglobotriaosylceramide (isoGb3). *Liebigs Ann. Chem.* **1992**, *1992*, 217–224.

(536) Stork, G.; La Clair, J. J. Stereoselective synthesis of β -mannopyranosides via the temporary silicon connection method. *J. Am. Chem. Soc.* **1996**, *118*, 247–248.

(537) Bols, M. Stereocontrolled synthesis of α -glucosides by intramolecular glycosidation. *J. Chem. Soc., Chem. Commun.* **1992**, *1992*, 913–914.

(538) Wessel, H. P.; Ruiz, N. α -Glucosylation reactions with 2,3,4,6-tetra-O-benzyl-D-glucopyranosyl fluoride and triflic anhydride as promoter. *J. Carbohydr. Chem.* **1991**, *10*, 901–910.

(539) Kevill, D. N.; Sutthoff, R. F. Kinetics of the reactions of t-butyl chloride and [2 H 9] t-butyl chloride with silver salts in acetonitrile. *J. Chem. Soc., Perkin Trans. 2* **1977**, *201*–206.

(540) Kato, T.; Tanaka, M.; Hoshikawa, M.; Yagi, M. An efficient construction of trinevitane and kempene skeletons from the common intermediate. *Tetrahedron Lett.* **1998**, *39*, 7553–7556.

(541) Zhang, Y.; Bommuswamy, J.; Sinnott, M. L. Kinetic isotope effect study of transition states for the hydrolyses of α - and β -glucopyranosyl fluorides. *J. Am. Chem. Soc.* **1994**, *116*, 7557–7563.

(542) Ogawa, T.; Takahashi, Y. Total synthesis of α -cyclodextrin. *Carbohydr. Res.* **1985**, *138*, c5–c9.

(543) Nicolaou, K.; Caulfield, T.; Kataoka, H.; Stylianides, N. Total synthesis of the tumor-associated Lex family of glycosphingolipids. *J. Am. Chem. Soc.* **1990**, *112*, 3693–3695.

(544) Nicolaou, K. C.; Hummel, C. W.; Iwabuchi, Y. Total synthesis of sialyl dimeric Le^x. *J. Am. Chem. Soc.* **1992**, *114*, 3126–3128.

(545) Matsuo, I.; Totani, K.; Tatami, A.; Ito, Y. Comprehensive synthesis of ER related high-mannose-type sugar chains by convergent strategy. *Tetrahedron* **2006**, *62*, 8262–8277.

(546) Nicolaou, K. C.; Caulfield, T.; Kataoka, H.; Kumazawa, T. A practical and enantioselective synthesis of glycosphingolipids and related compounds. Total synthesis of globotriaosylceramide (Gb₃). *J. Am. Chem. Soc.* **1988**, *110*, 7910–7912.

(547) Kaeothip, S.; Demchenko, A. V. On orthogonal and selective activation of glycosyl thioimides and thioglycosides: application to oligosaccharide assembly. *J. Org. Chem.* **2011**, *76*, 7388–7398.

(548) Kanie, O.; Ito, Y.; Ogawa, T. Orthogonal glycosylation strategy in oligosaccharide synthesis. *J. Am. Chem. Soc.* **1994**, *116*, 12073–12074.

(549) Paulsen, H. Progress in oligosaccharide synthesis through a new orthogonal glycosylation strategy. *Angew. Chem., Int. Ed. Engl.* **1995**, *34*, 1432–1434.

(550) Kanie, O. Orthogonal strategy in oligosaccharide synthesis. In *Carbohydrates in Chemistry and Biology*; Ernst, B., Hart, G. W., Sinay, P., Eds.; Wiley-VCH: Weinheim, Germany, 2000; Vol. 1, pp 407–426.

(551) Ito, Y.; Kanie, O.; Ogawa, T. Orthogonal glycosylation strategy for rapid assembly of oligosaccharides on a polymer support. *Angew. Chem., Int. Ed. Engl.* **1996**, *35*, 2510–2512.

(552) Kanie, O.; Ohtsuka, I.; Ako, T.; Daikoku, S.; Kanie, Y.; Kato, R. Orthogonal glycosylation reactions on solid phase and synthesis of a library consisting of a complete set of fucosyl galactose isomers. *Angew. Chem., Int. Ed.* **2006**, *45*, 3851–3854.

(553) Winter, M. Supports for solid-phase organic synthesis. In *Combinatorial peptide and nonpeptide libraries: a handbook*; Jung, G., Ed.; Wiley-VCH: Weinheim, Germany, 1996; pp 465–510.

(554) Fruchtel, J. S.; Jung, G. Organic chemistry on solid supports. *Angew. Chem., Int. Ed. Engl.* **1996**, *35*, 17–42.

(555) Hermkens, P. H. H.; Ottenheijm, H. C. J.; Rees, D. Solid-phase organic reactions: a review of the recent literature. *Tetrahedron* **1996**, *52*, 4527–4554.

(556) Brown, R. C. D. Recent developments in solid-phase organic synthesis. *J. Chem. Soc., Perkin Trans. 1* **1998**, *3293*–3320.

(557) Osborn, H. M. I.; Khan, T. H. Recent developments in polymer supported syntheses of oligosaccharides and glycopeptides. *Tetrahedron* **1999**, *55*, 1807–1850.

- (558) Krepinsky, J. J.; Douglas, S. P. Polymer-supported synthesis of oligosaccharides. In *Carbohydrates in Chemistry and Biology*; Ernst, B., Hart, G. W., Sinay, P., Eds.; Wiley-VCH: Weinheim, Germany, 2000; Vol. 1, pp 239–265.
- (559) Seeberger, P. H.; Haase, W. C. Solid-phase oligosaccharide synthesis and combinatorial carbohydrate libraries. *Chem. Rev.* **2000**, *100*, 4349–4393.
- (560) Fukase, K. Oligosaccharides: Synthesis: Combinatorial and solid phase methods in oligosaccharide synthesis. In *Glycoscience: Chemistry and Chemical Biology*; Fraser-Reid, B., Tatsuta, K., Thiem, J., Eds.; Springer: Berlin - Heidelberg - New York, 2001; Vol. 2, pp 1621–1660.
- (561) Seeberger, P. H. Solid phase oligosaccharide synthesis (reprinted from *Glycochemistry: principles, synthesis, and applications*, pg 1–32, 2001). *J. Carbohydr. Chem.* **2002**, *21*, 613–643.
- (562) Seeberger, P. H. The logic of automated glycan assembly. *Acc. Chem. Res.* **2015**, *48*, 1450–1463.
- (563) Hahm, H. S.; Schlegel, M. K.; Hurevich, M.; Eller, S.; Schuhmacher, F.; Hofmann, J.; Pagel, K.; Seeberger, P. H. Automated glycan assembly using the Glycoteer 2.1 synthesizer. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114*, E3385–E3389.
- (564) Wen, L.; Edmunds, G.; Gibbons, C.; Zhang, J.; Gadi, M. R.; Zhu, H.; Fang, J.; Liu, X.; Kong, Y.; Wang, P. G. Toward automated enzymatic synthesis of oligosaccharides. *Chem. Rev.* **2018**, *118*, 8151–8187.
- (565) Guberman, M.; Seeberger, P. H. Automated Glycan Assembly: A Perspective. *J. Am. Chem. Soc.* **2019**, *141*, 5581–5592.
- (566) Shadrack, M.; Panza, M.; Vijaya Ganesh, N.; Pistorio, S. G.; Stine, K. J.; Demchenko, A. V. HPLC-Based automated oligosaccharide synthesis. *Comprehensive Glycoscience* **2021**, 623–636.
- (567) Ako, T.; Daikoku, S.; Ohtsuka, I.; Kato, R.; Kanie, O. A method of orthogonal oligosaccharide synthesis leading to a combinatorial library based on stationary solid-phase reaction. *Chem.—Asian J.* **2006**, *1*, 798–813.
- (568) Manabe, S.; Ishii, K.; Ito, Y. N-Benzyl-2,3-trans-carbamate-bearing glycosyl donors for 1,2-cis-selective glycosylation reactions. *Eur. J. Org. Chem.* **2011**, *2011*, 497–516.
- (569) Huo, C.; Chan, T. H. A novel liquid-phase strategy for organic synthesis using organic ions as soluble supports. *Chem. Soc. Rev.* **2010**, *39*, 2977–3006.
- (570) Galan, M. C.; Corfield, A. P. Ionic liquids in oligosaccharide synthesis: Towards mucin-type glycan probes. *Biochem. Soc. Trans.* **2010**, *38*, 1368–1373.
- (571) Pathak, A. K.; Yerneni, C. K.; Young, Z.; Pathak, V. Oligomannan synthesis using ionic liquid supported glycosylation. *Org. Lett.* **2008**, *10*, 145–148.
- (572) Yerneni, C. K.; Pathak, V.; Pathak, A. K. Imidazolium cation supported solution-phase assembly of homolinear $\alpha(1\rightarrow6)$ -linked octamannoside: an efficient alternate approach for oligosaccharide synthesis. *J. Org. Chem.* **2009**, *74*, 6307–6310.
- (573) Scigelova, M.; Singh, S.; Crout, D. H. G. Glycosidases - a great synthetic tool. *J. Mol. Catal. B-Enzym.* **1999**, *6*, 483–494.
- (574) Voadlo, D. J.; Withers, S. G. Glycosidase-catalyzed oligosaccharide synthesis. In *Carbohydrates in Chemistry and Biology*; Ernst, B., Hart, G. W., Sinay, P., Eds.; Wiley-VCH: Weinheim, Germany, 2000; Vol. 1, pp 723–844.
- (575) Kren, V.; Thiem, J. Glycosylation employing bio-systems: from enzymes to whole cells. *Chem. Soc. Rev.* **1997**, *26*, 463–473.
- (576) Hehre, E. J. A fresh understanding of the stereochemical behavior of glycosylases: structural distinction of “inverting” (2-MCO-type) versus “retaining” (1-MCO-type) enzymes. *Adv. Carbohydr. Chem. Biochem.* **2000**, *55*, 265–310.
- (577) Kim, Y. W.; Fox, D. T.; Hekmat, O.; Kantner, T.; McIntosh, L. P.; Warren, R. A. J.; Withers, S. G. Glycosynthase-based synthesis of xylo-oligosaccharides using an engineered retaining xylanase from *Cellulomonas fimi*. *Org. Biomol. Chem.* **2006**, *4*, 2025–2032.
- (578) Plante, O. J.; Palmacci, E. R.; Seeberger, P. H. Automated solid-phase synthesis of oligosaccharides. *Science* **2001**, *291*, 1523–1527.
- (579) Sears, P.; Wong, C. H. Toward automated synthesis of oligosaccharides and glycoproteins. *Science* **2001**, *291*, 2344–2350.
- (580) Seeberger, P. H.; Werz, D. B. Automated synthesis of oligosaccharides as a basis for drug discovery. *Nat. Rev. Drug Discovery* **2005**, *4*, 751–763.
- (581) Tanaka, H.; Matoba, N.; Tsukamoto, H.; Takimoto, H.; Yamada, H.; Takahashi, T. Automated parallel synthesis of a protected oligosaccharide library based upon the structure of dimeric Lewis X by one-pot sequential glycosylation. *Synlett* **2005**, *2005*, 824–828.
- (582) Codée, J. D. C.; Kröck, L.; Castagner, B.; Seeberger, P. H. Automated solid-phase synthesis of protected oligosaccharides containing β -mannosidic linkages. *Chem.—Eur. J.* **2008**, *14*, 3987–3994.
- (583) Seeberger, P. H. Automated oligosaccharide synthesis. *Chem. Soc. Rev.* **2008**, *37*, 19–28.
- (584) Machida, K.; Hirose, Y.; Fuse, S.; Sugawara, T.; Takahashi, T. Development and application of a solution-phase automated synthesizer, ‘ChemKonzert’. *Chem. Pharm. Bull.* **2010**, *58*, 87–93.
- (585) Hsu, C. H.; Hung, S. C.; Wu, C. Y.; Wong, C. H. Toward automated oligosaccharide synthesis. *Angew. Chem., Int. Ed.* **2011**, *50*, 11872–11923.
- (586) Krock, L.; Esposito, D.; Castagner, B.; Wang, C.-C.; Bindschadler, P.; Seeberger, P. H. Streamlined access to conjugation-ready glycans by automated synthesis. *Chem. Sci.* **2012**, *3*, 1617–1622.
- (587) Vijaya Ganesh, N.; Fujikawa, K.; Tan, Y. H.; Stine, K. J.; Demchenko, A. V. HPLC-assisted automated oligosaccharide synthesis. *Org. Lett.* **2012**, *14*, 3036–3039.
- (588) Nokami, T.; Isoda, Y.; Sasaki, N.; Takaiso, A.; Hayase, S.; Itoh, T.; Hayashi, R.; Shimizu, A.; Yoshida, J.-i. Automated electrochemical assembly of the protected potential TMG-chitotriomycin precursor based on rational optimization of the carbohydrate building block. *Org. Lett.* **2015**, *17*, 1525–1528.
- (589) Tang, S.-L.; Pohl, N. L. B. Automated solution-phase synthesis of β -1,4-mannuronate and β -1,4-mannan. *Org. Lett.* **2015**, *17*, 2642–2645.
- (590) Pistorio, S. G.; Nigudkar, S. S.; Stine, K. J.; Demchenko, A. V. HPLC-assisted automated oligosaccharide synthesis: the implementation of the autosampler as a mode of the reagent delivery. *J. Org. Chem.* **2016**, *81*, 8796–8805.
- (591) Trobe, M.; Burke, M. D. The molecular industrial revolution: automated synthesis of small molecules. *Angew. Chem., Int. Ed.* **2018**, *57*, 4192–4214.
- (592) Li, T.; Liu, L.; Wei, N.; Yang, J.-Y.; Chapla, D. G.; Moremen, K. W.; Boons, G.-J. An automated platform for the enzyme-mediated assembly of complex oligosaccharides. *Nat. Chem.* **2019**, *11*, 229–236.
- (593) Mattes, D. S.; Jung, N.; Weber, L. K.; Brase, S.; Breiting, F. Miniaturized and Automated Synthesis of Biomolecules-Overview and Perspectives. *Adv. Mater.* **2019**, *31*, e1806656.
- (594) Chatterjee, S.; Guidi, M.; Seeberger, P. H.; Gilmore, K. Automated radial synthesis of organic molecules. *Nature* **2020**, *579*, 379–384.
- (595) Kern, M. K.; Pohl, N. L. B. Automated Solution-Phase Synthesis of S-Glycosides for the Production of Oligomannopyranoside Derivatives. *Org. Lett.* **2020**, *22*, 4156–4159.
- (596) Gromski, P. S.; Granda, J. M.; Cronin, L. Universal chemical synthesis and discovery with ‘The Chemputer’. *Trends Chem.* **2020**, *2*, 4–12.
- (597) Dube, D. H.; Bertozzi, C. R. Glycans in cancer and inflammation-potential for therapeutics and diagnostics. *Nat. Rev. Drug Discovery* **2005**, *4*, 477–488.
- (598) Lebrilla, C. B.; An, H. J. The prospects of glycan biomarkers for the diagnosis of diseases. *Mol. Biosystems* **2009**, *5*, 17–20.
- (599) Cheng, Y.; Li, M.; Wang, S.; Peng, H.; Reid, S.; Ni, N.; Fang, H.; Xu, W.; Wang, B. Carbohydrate biomarkers for future disease detection and treatment. *Sci. China Chem.* **2010**, *53*, 3–20.
- (600) Chaubard, J. L.; Krishnamurthy, C.; Yi, W.; Smith, D. F.; Hsieh-Wilson, L. C. Chemoenzymatic probes for detecting and

- imaging fucose- α (1–2)-galactose glycan biomarkers. *J. Am. Chem. Soc.* **2012**, *134*, 4489–4492.
- (601) Konforte, D.; Diamandis, E. P. Is early detection of cancer with circulating biomarkers feasible? *Clin. Chem.* **2013**, *59*, 35.
- (602) Mahon, E.; Mouline, Z.; Sillion, M.; Gilles, A.; Pinteala, M.; Barboiu, M. Multilayer lectin-glyconanoparticles architectures for QCM enhanced detection of sugar-protein interaction. *Chem. Commun.* **2013**, *49*, 3004–3006.
- (603) Ruhaak, L. R.; Miyamoto, S.; Lebrilla, C. B. Developments in the identification of glycan biomarkers for the detection of cancer. *Mol. Cell. Proteomics* **2013**, *12*, 846–855.
- (604) Hu, X.-L.; Jin, H.-Y.; He, X.-P.; James, T. D.; Chen, G.-R.; Long, Y.-T. Colorimetric and Plasmonic Detection of Lectins Using Core-Shell Gold Glyconanoparticles Prepared by Copper-Free Click Chemistry. *ACS Appl. Mater. Interfaces* **2015**, *7*, 1874–1878.
- (605) Oyelaran, O.; Gildersleeve, J. C. Glycan arrays: recent advances and future challenges. *Curr. Opin. Chem. Biol.* **2009**, *13*, 406–413.
- (606) Rillahan, C. D.; Paulson, J. C. Glycan microarrays for decoding the glycome. *Annu. Rev. Biochem.* **2011**, *80*, 797–823.
- (607) Laurent, N.; Haddoub, R.; Voglmeir, J.; Flitsch, S. L. MALDI-ToF MS analysis of glycosyltransferase activities on gold surface arrays. *Carbohydrate Microarrays*, in *Methods in Molecular Biology* **2012**, *808*, 269–284.
- (608) Song, X.; Heimbürg-Molinari, J.; Smith, D. F.; Cummings, R. D. Glycan microarrays. *Methods Mol. Biol.* **2012**, *800*, 163–171.
- (609) Park, S.; Gildersleeve, J. C.; Blixt, O.; Shin, I. Carbohydrate microarrays. *Chem. Soc. Rev.* **2013**, *42*, 4310–4326.
- (610) Smith, D. F.; Cummings, R. D. Investigating virus-glycan interactions using glycan microarrays. *Curr. Opin. Virol.* **2014**, *7*, 79–87.
- (611) Wang, L.; Cummings, R. D.; Smith, D. F.; Huflejt, M.; Campbell, C. T.; Gildersleeve, J. C.; Gerlach, J. Q.; Kilcoyne, M.; Joshi, L.; Serna, S.; Reichardt, N. C.; Parera Pera, N.; Pieters, R. J.; Eng, W.; Mahal, L. K. Cross-platform comparison of glycan microarray formats. *Glycobiology* **2014**, *24*, 507–517.
- (612) Li, Z.; Feizi, T. The neoglycolipid (NGL) technology-based microarrays and future prospects. *FEBS letters* **2018**, *592*, 3976–3991.
- (613) Wennekes, T.; van den Berg, R. J. B. H. N.; Boot, R. G.; Van der Marel, G. A.; Overkleef, H. S.; Aerts, J. M. F. G. Glycosphingolipids - nature, function, and pharmacological modulation. *Angew. Chem., Int. Ed.* **2009**, *48*, 8848–8869.
- (614) *Carbohydrate-based vaccines and immunotherapies*; Guo, Z., Boons, G. J., Eds.; Wiley: Hoboken, 2009.
- (615) Cipolla, L.; Arajo, A. C.; Bini, D.; Gabrielli, L.; Russo, L.; Shaikh, N. Discovery and design of carbohydrate-based therapeutics. *Expert Opin. Drug Discovery* **2010**, *5*, 721–737.
- (616) Kaeothip, S.; Paranjape, G.; Terrill, S. E.; Bongat, A. F. G.; Udan, M. L. D.; Kamkhachorn, T.; Johnson, H. L.; Nichols, M. R.; Demchenko, A. V. Development of LPS antagonistic therapeutics: synthesis and evaluation of glucopyranoside-spacer-amino acid motifs. *RSC Adv.* **2011**, *1*, 83–92.
- (617) Liu, L.; Zha, J.; DiGiandomenico, A.; McAllister, D.; Stover, C. K.; Wang, Q.; Boons, G. J. Synthetic Enterobacterial Common Antigen (ECA) for the Development of a Universal Immunotherapy for Drug-Resistant Enterobacteriaceae. *Angew. Chem., Int. Ed. Engl.* **2015**, *54*, 10953–10957.
- (618) *Carbohydrate-based drug discovery*; Wong, C. H., Ed.; Wiley-VCH: Weinheim, Germany, 2003.
- (619) *Carbohydrate drug design*; Klyosov, A. A.; Witczak, Z. J., Platt, D., Eds.; American Chemical Society: Washington, DC, 2006; Vol. 932.
- (620) Ernst, B.; Magnani, J. L. From carbohydrate leads to glycomimetic drugs. *Nat. Rev. Drug Discovery* **2009**, *8*, 661–677.
- (621) Galan, M. C.; Benito-Alifonso, D.; Watt, G. M. Carbohydrate chemistry in drug discovery. *Org. Biomol. Chem.* **2011**, *9*, 3598–3610.
- (622) Klyosov, A. A. Carbohydrates and Drug Design. In *Glycobiology and Drug Design*; ACS Symposium Series 1102; American Chemical Society: Washington, DC, 2012; Vol. 1102, pp 3–22.
- (623) Hsieh-Wilson, L. C.; Griffin, M. E. Improving Biologic Drugs via Total Chemical Synthesis. *Science* **2013**, *342*, 1332–1333.
- (624) Wong, C. F. Flexible receptor docking for drug discovery. *Expert Opin. Drug Discovery* **2015**, *10*, 1189–1200.
- (625) Guo, Z.; Wang, Q. Recent development in carbohydrate-based cancer vaccines. *Curr. Opinion in Chem. Biology* **2009**, *13*, 608–617.
- (626) Daum, R. S.; Spellberg, B. Progress Toward a Staphylococcus aureus Vaccine. *Clin. Infect. Dis.* **2012**, *54*, 560–567.
- (627) Yin, Z.; Huang, X. Recent Development in Carbohydrate Based Anticancer Vaccines. *J. Carbohydr. Chem.* **2012**, *31*, 143–186.
- (628) Huang, Y. L.; Hung, J. T.; Cheung, S. K.; Lee, H. Y.; Chu, K. C.; Li, S. T.; Lin, Y. C.; Ren, C. T.; Cheng, T. J.; Hsu, T. L.; Yu, A. L.; Wu, C. Y.; Wong, C. H. Carbohydrate-based vaccines with a glycolipid adjuvant for breast cancer. *Proc. Nat. Acad. Sci.* **2013**, *110*, 2517–2522.
- (629) Peri, F. Clustered Carbohydrates in Synthetic Vaccines. *Chem. Soc. Rev.* **2013**, *42*, 4543–4556.
- (630) Khatun, F.; Stephenson, R. J.; Toth, I. An Overview of Structural Features of Antibacterial Glycoconjugate Vaccines That Influence Their Immunogenicity. *Chem.—Eur. J.* **2017**, *23*, 4233–4254.
- (631) Gouttefangeas, C.; Rammensee, H. G. Personalized cancer vaccines: adjuvants are important, too. *Cancer Immunol. Immunother.* **2018**, *67*, 1911–1918.
- (632) Li, Q.; Guo, Z. Recent advances in Toll like receptor-targeting glycoconjugate vaccines. *Molecules* **2018**, *23*, 1583.
- (633) Behera, A.; Rai, D.; Kulkarni, S. S. Total Syntheses of Conjugation-Ready Trisaccharide Repeating Units of Pseudomonas aeruginosa O11 and Staphylococcus aureus Type 5 Capsular Polysaccharide for Vaccine Development. *J. Am. Chem. Soc.* **2020**, *142*, 456–467.
- (634) Wang, P.; Huo, C. X.; Lang, S.; Caution, K.; Nick, S. T.; Dubey, P.; Deora, R.; Huang, X. Chemical Synthesis and Immunological Evaluation of a Pentasaccharide Bearing Multiple Rare Sugars as a Potential Anti-pertussis Vaccine. *Angew. Chem., Int. Ed. Engl.* **2020**, *59*, 6451–6458.
- (635) Seeberger, P. H. Discovery of Semi- and Fully-Synthetic Carbohydrate Vaccines Against Bacterial Infections Using a Medicinal Chemistry Approach. *Chem. Rev.* **2021**, *121*, 3598–3626.