

RESEARCH ARTICLE

Mechanisms of sound production in deer mice (*Peromyscus* spp.)Tobias Riede^{1,*}, Anastasiya Kobrina², Landon Bone³, Tarana Darwaiz¹ and Bret Pasch²

ABSTRACT

Rodent diversification is associated with a large diversity of species-specific social vocalizations generated by two distinct laryngeal sound production mechanisms: whistling and airflow-induced vocal fold vibration. Understanding the relative importance of each modality to context-dependent acoustic interactions requires comparative analyses among closely related species. In this study, we used light gas experiments, acoustic analyses and laryngeal morphometrics to identify the distribution of the two mechanisms among six species of deer mice (*Peromyscus* spp.). We found that high frequency vocalizations (simple and complex sweeps) produced in close-distance contexts were generated by a whistle mechanism. In contrast, lower frequency sustained vocalizations (SVs) used in longer distance communication were produced by airflow-induced vocal fold vibrations. Pup isolation calls, which resemble adult SVs, were also produced by airflow-induced vocal fold vibrations. Nonlinear phenomena (NLP) were common in adult SVs and pup isolation calls, suggesting irregular vocal fold vibration characteristics. Both vocal production mechanisms were facilitated by a characteristic laryngeal morphology, including a two-layered vocal fold lamina propria, small vocal membrane-like extensions on the free edge of the vocal fold, and a singular ventral laryngeal air pocket known as the ventral pouch. The size and composition of vocal folds (rather than total laryngeal size) appears to contribute to species-specific acoustic properties. Our findings suggest that dual modes of sound production are more widespread among rodents than previously appreciated. Additionally, the common occurrence of NLP highlights the nonlinearity of the vocal apparatus, whereby small changes in anatomy or physiology trigger large changes in behavior. Finally, consistency in mechanisms of sound production used by neonates and adults underscores the importance of considering vocal ontogeny in the diversification of species-specific acoustic signals.

KEY WORDS: Heliox, Nonlinear phenomena, Larynx, Source-filter theory, Cricetidae, Bioacoustics

INTRODUCTION

Rodents produce diverse acoustic signals over a wide spectral range using at least two different mechanisms (e.g. Fernández-Vargas et al., 2022) (Fig. 1). Vocalizations are produced by airflow-induced vocal fold vibrations at the lower end of the spectral range and a laryngeal whistle is produced at the upper spectral range (Roberts, 1975; Riede, 2011; 2013; Pasch et al., 2017). The former

mechanism is used by almost all mammals documented to date (e.g. Madsen et al., 2012; Koda et al., 2015), wherein glottal airflow draws vocal fold tissue into vibration to generate pressure fluctuations perceived as sound. In contrast, the laryngeal whistle is a unique innovation in rodents (Fernández-Vargas et al., 2022). High-frequency (or ‘ultrasonic’) whistles are produced by a glottal airstream that interacts with a rigid intralaryngeal structure, generating pressure fluctuations that resonate inside the laryngeal airway (Riede et al., 2017; Håkansson et al., 2022). One hypothesis for the origin of this innovation is to escape detection of acoustically orienting predators (Blanchard and Blanchard, 1989; Brudzynski, 2014). Regardless of the process, the distribution of whistling among rodents remains largely unknown. A better understanding of the importance of species-specific acoustic variation requires characterization of the underlying sources of such variation.

The monophyletic rodent genus *Peromyscus* (Cricetidae, Neotominae) provides a model system to study the evolution of adaptive divergence (Bedford and Hoekstra, 2015), including vocal communication (Kalcounis-Rueppell et al., 2018a). ‘Deer mice’ have evolved diverse vocal repertoires used in a variety of social contexts (Eisenberg, 1961; Miller and Engstrom, 2010, 2012; Briggs and Kalcounis-Rueppell, 2011; Kalcounis-Rueppell et al., 2006, 2010, 2018b; Pultorak et al., 2015). Three call types found in adult *Peromyscus* include sustained vocalizations (hereafter SVs), simple and complex sweeps (hereafter ‘sweeps’) and barks (sometimes referred to as screams). SVs consist of one or more syllables of short duration (~200 ms) uttered in close succession, and their fundamental frequency ranges between 10 and 25 kHz. Sweeps are vocalization of short duration (10–50 ms) with fundamental frequencies above 25 kHz (Kalcounis-Rueppell et al., 2018b). Barks are 50–100 ms vocalizations with a fundamental frequency range between 0.8 and 6 kHz. A fourth call type are pup isolation vocalizations produced by offspring during the first 3 weeks of life. Newborn deer mice produce such characteristic vocalizations when isolated from their mother (Hart and King, 1966; Smith, 1972; Johnson et al., 2017; Kalcounis-Rueppell et al., 2018c). Pup isolation calls resemble adult SVs in spectral and temporal features (Johnson et al., 2017; Kalcounis-Rueppell et al., 2018c).

Despite the extensive repertoire and radiation of *Peromyscus* (Kalcounis-Rueppell et al., 2018a), no studies have characterized mechanisms of vocal production. However, the presence of nonlinear phenomena (NLP) in the SVs of *P. californicus* (Miller and Engstrom, 2012) indicates that such vocalizations are produced by airflow-induced vocal fold vibrations (Herzel et al., 1994, 1995). NLP result from irregular vibration patterns of the vocal folds and are commonly found in human and nonhuman mammals (e.g. Wilden et al., 1998; Riede et al., 1997, 2000; Blumstein and Récapet, 2009; Titze et al., 2008). NLP may be indicative of arousal (Blumstein and Récapet, 2009), predictability (Townsend and Manser, 2011) and/or provide signatures of individual or species identity by indicating the maximum fundamental frequency at which vocal folds can perform symmetric harmonic vibrations (Riede et al., 2007).

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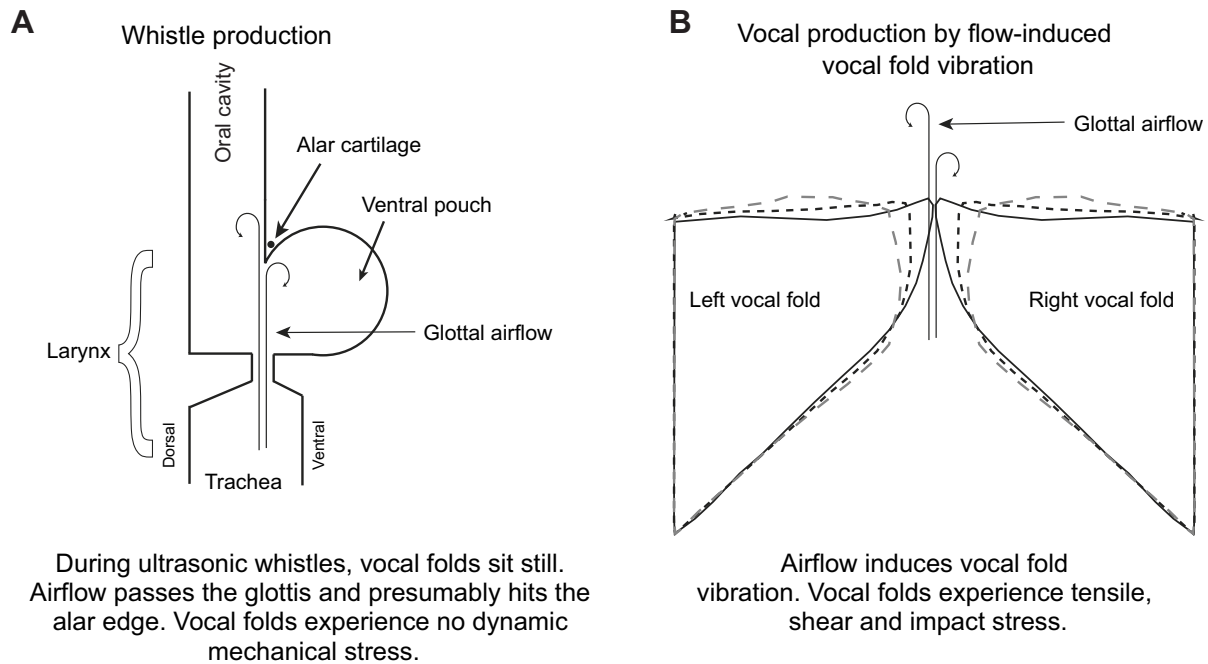


Fig. 1. Rodents employ two mechanisms of producing sound inside the larynx. (A) High frequency whistles are hypothesized to rely on a glottal airstream which interacts with an intralaryngeal structure (alar edge) generating pressure fluctuations in the ventral pouch, a small side-branch off the main laryngeal airway (Riede et al., 2017). (B) Sounds can also be produced by airflow-induced vocal fold vibrations. The glottal airflow draws vocal fold tissue into vibration. The vibrations generate pressure fluctuations perceived as sounds.

Acoustic properties are determined by vocal organ size, vocal fold composition, airway geometry, coordination of vocal organ and breathing movements (Fernández-Vargas et al., 2022). Understanding the relative contributions of these morphological and physiological traits to acoustic variation could provide insight into the evolution of species-specific vocalizations. In this study, we used light gas experiments and acoustic analyses to identify the distribution of the two mechanisms among six species of deer mice. In addition, we qualitatively described the anatomy of vocal organs to inform laryngeal biomechanics. Finally, we compared vocal organ and laryngeal airway size as well as vocal fold size and composition among the six species to better understand determinants of species-specific acoustic properties.

MATERIALS AND METHODS

Animals

A total of 74 individuals from six *Peromyscus* species were included in this study. Individuals of five species were acquired from the *Peromyscus* Genetic Stock Center and one species was wild-captured [*Peromyscus truei* (Shufeldt 1885)]. Live *Peromyscus californicus* (Gambel 1848) and *Peromyscus maniculatus* (Wagner 1845) were purchased and bred at Midwestern University, Glendale, AZ. Twelve adult animals (6/sex) from each of the two species were investigated through sound recordings, heliox experiments and anatomical analysis. Additionally, 6 pups (*P. californicus*) were investigated through sound recordings ($N=6$) and heliox experiments ($N=4$). Twelve specimens (6/sex) from each of *Peromyscus polionotus* (Wagner 1843), *Peromyscus eremicus* (Baird 1858) and *Peromyscus leucopus* (Rafinesque 1818) were purchased from the *Peromyscus* Genetic Stock Center (PGSC) for anatomical analyses.

Twenty *P. truei* were captured near Deadman Flat, 28 km north of Flagstaff, AZ, using Sherman live-traps baited with sterilized bird seed and transferred in standard mouse cages to animal facilities at Northern

Arizona University, Flagstaff, AZ, for sound recordings. Twelve animals (6/sex) were transferred to Midwestern University, Glendale, AZ, USA, for heliox experiments and morphological analysis.

All procedures were performed in accordance with ethical standards and approval of the Institutional Animal Care and Use Committee at Midwestern University (MWU#3011) and Northern Arizona University (19-006) and guidelines of the American Society of Mammalogists (Sikes and Animal Care and Use Committee of the American Society of Mammalogists, 2016). Animals were captured with a permit from the Arizona Game and Fish Department (607608).

Heliox experiments

Recording vocal behavior in light gas atmosphere, examination of nonlinear phenomena and anatomical investigation of laryngeal tissue can be used to inform the sound production mechanism used to produce four types of vocalizations. For light gas experiments, a vocalizing animal was placed in a closed container with a gas mixture that has a lower density than normal air. The approach can differentiate between the two vocal production mechanisms. The vibration frequency of vocal folds is independent of the type of gas that surrounds them (Titze et al., 2016), i.e. the fundamental frequency of the sound does not change in light gas. However, the velocity of the sound wave is faster in the light gas and the fundamental frequency of a whistle sound therefore increases predictably (Roberts, 1975; Riede, 2011; Pasch et al., 2017; Riede and Pasch, 2020).

Acoustic recordings in a light gas atmosphere were successfully conducted in *P. californicus* (pup isolation calls, adult SVs and sweeps) and *P. maniculatus* (adult barks and sweeps). Individual mice were placed in an acrylic cage. The cage was equipped with bedding, food and water. Heliox gas (80% He, 20% O₂) was injected into the cage at flow rates between 20 and 40 l min⁻¹ through a 12 mm wide tube placed into the cage wall near the floor.

Predicted acoustic effects of light gas concentrations were estimated with a small whistle placed at the floor of the cage and connected externally by a silastic tube. The whistle was blown and recorded at regular intervals to monitor the heliox concentration. The ratio of the frequency of the whistle in air and in heliox allowed an estimation of the expected effect for any given heliox concentration.

Acoustic recordings

Heliox experiments indicated that SVs in *P. californicus* are generated by flow-induced vocal fold vibrations (see Results). Since SV calls among *Peromyscus* species show similar spectro-temporal features (Kalcounis-Rueppell et al., 2018b), we inferred that SV calls in other *Peromyscus* species were generated by the same mechanism and thus focused on the occurrence of harmonic patterns and nonlinear phenomena (NLP) that typify sounds produced by vocal fold vibration. In order to determine the occurrence of NLP in SV calls, we intensively sampled vocalizations of two species (*P. californicus* and *P. truei*). *P. californicus* vocalizations were recorded using an ultrasonic microphone (Avisoft-Bioacoustics, CM16/CMPA-5V) placed over the center of the cage. Microphone frequency range is 2 to 200 kHz and an approximate sensitivity of 500 mV Pa⁻¹. Signals were acquired through an NiDAQ 6212 acquisition device, sampled at 200 kHz, and saved as uncompressed files using Avisoft Recorder software (version 3.4.2, Avisoft-Bioacoustics, Berlin, Germany). For *P. truei*, singly housed mice in their home cage were placed in semi-anechoic coolers lined with acoustic foam. We used 1/4" (6.4 mm) microphones (Type 40BE, G.R.A.S.) connected to preamplifiers (Type 26 CB, G.R.A.S.) to obtain recordings above the center of the mouse cage. Microphone response was flat within ± 1.5 dB from 10 Hz to 50 kHz, and pre-amplifier response was flat within ± 0.2 dB from 2 Hz to 200 kHz. Microphones were

connected to a National Instruments Data Acquisition unit (USB 4431) sampling at 102.4 kHz to a desktop computer running a custom recording program in MATLAB (v. 2018a).

Micro-CT scanning and histology

Twelve adult mice per species (6/sex) were euthanized with ketamine and xylazine, and then transcardially perfused with saline solution followed by 10% buffered formalin. Larynges were dissected and placed in 10% buffered formalin phosphate (SF100-4; Fisher Scientific) for 2 days.

Larynges from eight mice (4/sex) were x-rayed at 5 μ m resolution. First, tissues were transferred from the formalin solution to 99% ethanol. Tissues were then stained in 1% phosphotungstic acid (PTA) (Sigma Aldrich, 79690) in 70% ethanol. After 5 days, the staining solution was renewed and the tissue was stained for an additional 5 days. After staining, specimens were placed in a custom-made acrylic tube and scanned in air. Micro-CT scanning was done using a Skyscan 1172 (Bruker). Reconstructed image stacks were then imported into AVIZO software (v. Lite 9.0.1). Laryngeal cartilages and the border between the airway and soft tissues of the larynx were traced manually in CT scans. This approach provided outlines of the cartilaginous framework and the airway. Derived 3D surfaces of eight specimens from each of the 6 species have been archived at Morphobank (O'Leary, Kaufman 2012), project # P4106.

Coronal histological sections of larynges from four mice (2/sex) were used to quantify vocal fold morphology (lamina propria thickness and fibrillar protein distribution). Mid-membraneous coronal sections (5 mm thick) were stained with Haematoxylin and Eosin for a general overview, Masson's Trichrome (TRI) for collagen fiber stain and Elastica-Van Gieson (EVG) for elastic fiber stain. Sections were scanned with an Aperio CS 2 slide scanner and processed with Imagescope software (v. 8.2.5.1263; Aperio Tech.).

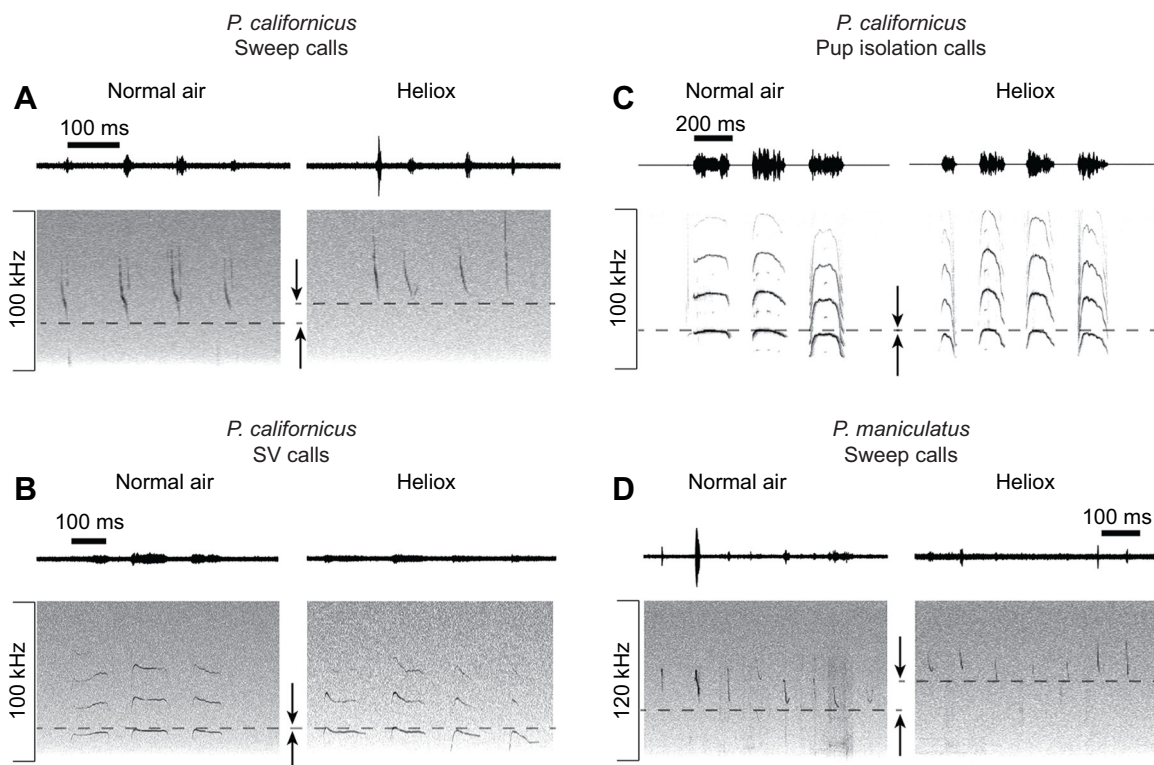


Fig. 2. *Peromyscus californicus* and *Peromyscus maniculatus* vocalizations in normal air and heliox. (A) Sweeps, (B) sustained vocalization (SV) bouts and (C) pup isolation calls produced by *P. californicus*. (D) Sweeps produced by *P. maniculatus*.

Acoustic analysis

Heliox data

Four call types were successfully recorded in heliox and normal air: sweeps, SV calls, barks and pup isolation calls. All four call types were analyzed for center, minimum ($f_{0,\min}$), maximum ($f_{0,\max}$) fundamental frequency and call/syllable duration. Fundamental frequency was quantified every 20 ms using PRAAT's pitch-tracking tool. Then, frequency values were represented as histograms (100 or 500 Hz resolution). Center fundamental frequency was calculated from the weighted median of all frequency measurements. Fundamental frequency range was calculated from the difference between $f_{0,\min}$ and $f_{0,\max}$. Acoustic differences between normal air and heliox songs were assessed with paired *t*-tests.

Nonlinear phenomena

Vocalizations were analyzed using the pitch tracking tool (1024-point Fast Fourier Transform (FFT), 75% frame size, Hann window, frequency resolution 100 Hz, temporal resolution 93.75%, 0.625 ms) in the software PRAAT (v. 5.3.80, retrieved January 2014 from <http://www.praat.org/>). Call duration, maximum fundamental frequency, and minimum fundamental frequency were manually extracted. NP were first categorized into frequency jumps (FJs), subharmonics (SHs), deterministic chaos (CH) or biphonation (BP) (Fig. 2). NP were quantitatively analyzed in *P. californicus* ($n=4/\text{sex}$) and *P. truei* ($n=6$ females, 3 males) based on visual inspection of a narrowband spectrogram of the signal (Herzel, 1993) and associated Fourier frequency spectra, following earlier studies (Riede et al., 2000, 2004; Titze et al., 2008; Zollinger et al., 2008). First, different temporal segments of an SV call were determined. Segment borders were positioned at bifurcations. A bifurcation refers to the boundaries between different regimes, such as 'no phonation', harmonic phonation, SHs, BP, CH and FJs (e.g. Riede et al., 2000, 2004). Occurrence of each NLP relative to number of SV bouts and syllables was measured. Duration of syllable and NLPs, as well as percentage occurrence was calculated.

Laryngeal morphology and vocal fold histology

Laryngeal anatomy was investigated by focusing on three aspects of the vocal organ. First, to test whether overall organ size served as a proxy for laryngeal valve function, we quantified thyroid cartilage (whole organ) centroid size and vocal fold length (laryngeal valve). Second, previous work suggested that a small pocket (i.e. the ventral pouch), plays an important role in ultrasonic whistle production (Riede et al., 2017). Therefore, laryngeal airway shape was described qualitatively. Thirdly, shape and composition of vocal folds determine their biomechanical properties and vibration characteristics (Titze et al., 2016). In a related genus (*Onychomys* spp.), a heterogeneous lamina propria and presence of vocal membranes support the production of long-distance low frequency calls (Pasch et al., 2017). Therefore, we studied lamina propria heterogeneity and presence of vocal membranes located near the free edge of the vocal folds.

Size was described by centroid size and vocal fold length in 48 specimens (8 per species and 4 per sex; *P. californicus*, *P. maniculatus*, *P. leucopus*, *P. eremicus*, *P. polionotus*, *P. truei*). Geometric morphometric methods were developed previously and are outlined in detail in Borgard et al. (2020) and Riede et al. (2020). Briefly, the Geomorph package (v. 3.0.5.; <https://CRAN.R-project.org/package=geomorph>) for R (<https://www.r-project.org/>) was used to measure thyroid cartilage centroid size as a proxy of overall larynx size. Landmarks (24 curve landmarks and 100 semi-landmarks) were placed on 3D surface renderings of the thyroid cartilage. Centroid size was calculated as the square root of the sum of squared distances of each landmark from the center of the cartilage (Zelditch et al., 2004). Vocal fold length was measured between the most ventral tip of the vocal process of the arytenoid cartilage and the midline thyroid cartilage near its caudal edge. Body size was estimated through body mass and left femur length. Body mass and femur length were found to be strongly positively correlated (Pearson correlation, $N=72$; $r=0.81$; $P<0.001$). Body mass ($F_{2,69}=9.3$; $P<0.001$) but not femur length ($F_{2,69}=2.1$; $P=0.13$)

Table 1. Center fundamental frequency, fundamental frequency range and duration of adult California mouse (*Peromyscus californicus*) sweep calls, SV syllables, barks calls and pup isolation calls (means $\pm$ s.d.)

	Normal air				Expected f_0 increase	Helium/oxygen mix			
	f_0 center (kHz)	f_0 range (kHz)	f_0 mean (kHz)	Duration (ms)		f_0 center (kHz)	f_0 range (kHz)	f_0 mean (kHz)	Duration (ms)
Sweeps									
3A (2 males)	46	31–70	–	14.8±5.7	1.4	59	51–88	–	13.6±3.0
3B (2 males)	68	47–81	–	13.7±11.2	1.2	71	50–88	–	10.6±6.1
3D (pair 1)	54	27–80	–	10.8±3.5	1.3	61	41–91	–	12.1±4.3
3E (pair 2)	50	32–80	–	13.8±3.8	1.3	59	42–85	–	15.4±8.2
3F (pair 3)	41	32–78	–	20.0±5.5	1.4	56	46–88	–	19.0±6.8
3G (pair 4)	47	31–79	–	13.0±5.7	1.5	68	50–87	–	12.3±2.7
3H (pair 5)	42	30–69	–	10.6±2.4	1.5	64	50–86	–	10.0±3.0
4A (pair 6)	53	38–73	–	11.5±3.4	1.2	59	43–89	–	14.1±4.6
SV syllables									
3D (pair 1)	–	–	18.1±1.8	194.1±32.6	1.3	–	–	17.5±1.7	171.7±19.3
3E (pair 2)	–	–	17.2±1.7	327.0±87.1	1.3	–	–	18.0±1.2	355.7±61.7
3G (pair 4)	–	–	18.7±1.9	250.0±96.2	1.5	–	–	16.7±3.3	210.3±41.5
3H (pair 5)	–	–	11.8±1.7	123.7±42.4	1.5	–	–	14.0±2.2	144.0±9.4
Barks									
4A (pair 6)	–	–	10.1±1.6	24.2±9.4	1.2	–	–	10.9±0.8	17.1±2.5
Pup isolation calls									
Pup 1 (PND 6)	–	–	22.1±2.2	174.5±21.3	1.4	–	–	19.2±3.0	115.6±28.9
Pup 2 (PND 5)	–	–	19.2±1.4	166.6±19.7	1.5	–	–	18.8±1.6	133.3±17.3
Pup 3 (PND 5)	–	–	19.8±1.6	112.0±23.6	1.4	–	–	17.8±0.9	97.6±8.8
Pup 4 (PND 4)	–	–	17.9±1.9	129.0±32.7	1.5	–	–	17.1±2.6	138.0±37.0

PND, postnatal day.

Table 2. Center fundamental frequency and fundamental frequency range of adult deer mouse (*P. maniculatus*) sweeps and barks

	Normal air				Expected f_0 increase	Helium/oxygen mix			
	f_0 center (kHz)	f_0 range (kHz)	f_0 mean (kHz)	Duration (ms)		f_0 center (kHz)	f_0 range (kHz)	f_0 mean (kHz)	Duration (ms)
Sweeps									
2A (pair 1)	60	37–79	—	—	1.2	69	45–89	—	—
2D (pair 2)	67	53–78	—	—	1.5	92	87–105	—	—
4A (pair 3)	44	32–76	—	—	1.4	66	46–85	—	—
4C (pair 4)	48	39–71	—	—	1.5	72	57–87	—	—
5D (pair 5)	48	31–74	—	—	1.5	70	58–90	—	—
Barks									
2C (pair 6)	—	—	6.2±0.4	61.0±52.5	1.3	—	—	5.4±0.4	46.4±14.5
4A (pair 3)	—	—	7.2±1.2	21.3±10.6	1.4	—	—	6.7±1.0	10.0±5.3
4C (pair 4)	—	—	6.8±0.9	22.4±9.9	1.5	—	—	6.9±0.4	11.8±1.3

was different between males and females. Therefore, males and females were combined for analyses and femur length was used as body size estimate.

In 24 specimens (4/species; 2/sex), lamina propria thickness was measured and averaged across 3 locations positioned equidistant under the non-ciliated epithelium. In 18 (out of 24) specimens, we found vocal membrane-like structures. The height and width of vocal membranes were measured. All measurements were taken in mid-membranous coronal sections using software ImageJ. We used multiple regression to assess whether anatomical measures could be predicted from body size and/or species identity.

Collagen and elastin content of the lamina propria was quantified by digitally isolating the lamina propria of the free edge of the vocal fold. First, we drew an imaginary line bisecting the lamina propria into superficial (medial) and deep (lateral) halves. In trichrome stains, the blue-staining collagen fiber pixels within the blue range were selected using the color threshold tool in ImageJ. The image was then converted into binary mode that converted blue pixels into black and all other pixels into white. The proportion of black pixels was counted in each of five transects placed into the superficial and the deep lamina propria. Care was taken so that transects would not overlap between deep and superficial lamina propria or reach into the epithelial tissue. Black-staining elastin fibers were similarly quantified using the brightness slider in ImageJ.

RESULTS

Heliox experiments

In order to determine the mechanism of sound production in *Peromyscus* spp., we recorded adult SVs, sweeps, barks and pup isolation calls in air and in light gas atmosphere. Fig. 2 shows spectrographic representations of sweeps and SV bouts produced in air and in heliox. In adult *P. californicus*, fundamental frequency of sweep calls increased in heliox (paired t -tests, $f_{0, \text{center}}$: $t_7 = -4.86$, $P < 0.001$; $f_{0, \text{min}}$: $t_7 = -5.75$, $P < 0.01$; $f_{0, \text{max}}$: $t = -6.6$, $P < 0.001$)

compared with normal air (Table 1). However, sweep call duration did not change in heliox ($t_7 = 0.21$, $P = 0.84$) compared with normal air (Table 1; Fig. 2D). In contrast, fundamental frequency of SV syllables did not change in heliox (paired t -tests, $f_{0, \text{mean}}$: $t_3 = -0.34$, $P = 0.75$) compared with normal air (Table 2). Similarly, SV duration also did not change in heliox ($t_3 = 0.33$, $P = 0.76$) compared with normal air (Table 2).

Similarly to adult SVs, fundamental frequency of pup isolation calls did not change in heliox (paired t -tests, $f_{0, \text{mean}}$: $t_3 = 2.67$, $P = 0.076$) compared with normal air (Table 3), nor did their duration ($t_3 = 1.70$, $P = 0.19$; Table 3).

In *P. maniculatus*, we similarly found that fundamental frequency of sweep calls increased in heliox (paired t -tests, $f_{0, \text{center}}$: $t_3 = -7.01$, $P < 0.01$; $f_{0, \text{min}}$: $t_3 = -4.36$, $P < 0.05$; $f_{0, \text{max}}$: $t_3 = -4.87$, $P < 0.01$) (Table 2; Fig. 2E), but their duration did not ($t_3 = 2.02$, $P = 0.11$; Table 2). Fundamental frequency of barks did not change in heliox (paired t -tests, $f_{0, \text{mean}}$: $t_2 = 1.51$, $P = 0.27$) compared with normal air (Table 2). Barks were shorter in heliox than in normal air ($t_2 = 9.8$, $P < 0.05$) (Table 2).

In sum, the data suggest that both adult *P. californicus* and *P. maniculatus* produce sweeps by a whistle mechanism. Barks, SVs and pup isolation calls, however, are produced by airflow-induced vocal fold vibration.

Acoustic analysis

Fig. 3 shows examples of four types of NLP in SV bouts produced by *P. californicus* and *P. truei*. In *P. californicus*, the percentage of SVs containing at least one type of NLP within individuals ranged between 1.3 and 48.3% (Table 3). Subharmonics were present in the SVs of 6 out of 7 mice, and the number of calls containing subharmonic segments ranged widely among individuals (0 to 44.8%) and appeared to be individual specific (Table 3). Cumulatively across all *P. californicus*, out of 785 SVs that exhibited NLPs, 34 SVs (4.3%) displayed one or

Table 3. Rate of occurrence of nonlinear phenomena in *P. californicus* SV bouts (mean±s.d.)

Mouse ID	SV bouts	SV syllables	No. SV syllables bout ⁻¹	Syllable duration (s)	NLP (%)	NLP duration (%)	FJ	SH	BP	CH
Male pair 1	60	212	3.3±0.78	0.26±0.06	14.6	28.5±16.3	6	23	2	0
Female pair 1	45	78	2.36±1.0	0.18±0.04	29.5	40.3±15.5	10	3	0	0
Male pair 2	40	58	1.65±0.6	0.35±0.08	48.3	39.4±18.3	2	26	0	0
Female pair 2	13	25	2.77±0.9	0.17±0.02	32.0	26.3±1.8	2	6	0	0
Male pair 3	23	51	2.21±0.5	0.33±0.07	39.2	30.7±11.8	7	16	1	0
Female pair 3	59	230	3.9±0.8	0.18±0.03	1.3	0	3	0	0	0
Male pair 4	40	131	3.35±0.5	0.25±0.07	27.5	48.3±20.1	4	21	11	0
Female pair 4	0	0	0	0	0	0	0	0	0	0

All SVs were produced by pair-bonded males and females while separated for 24 h in two cages with visual, olfactory and auditory contact.

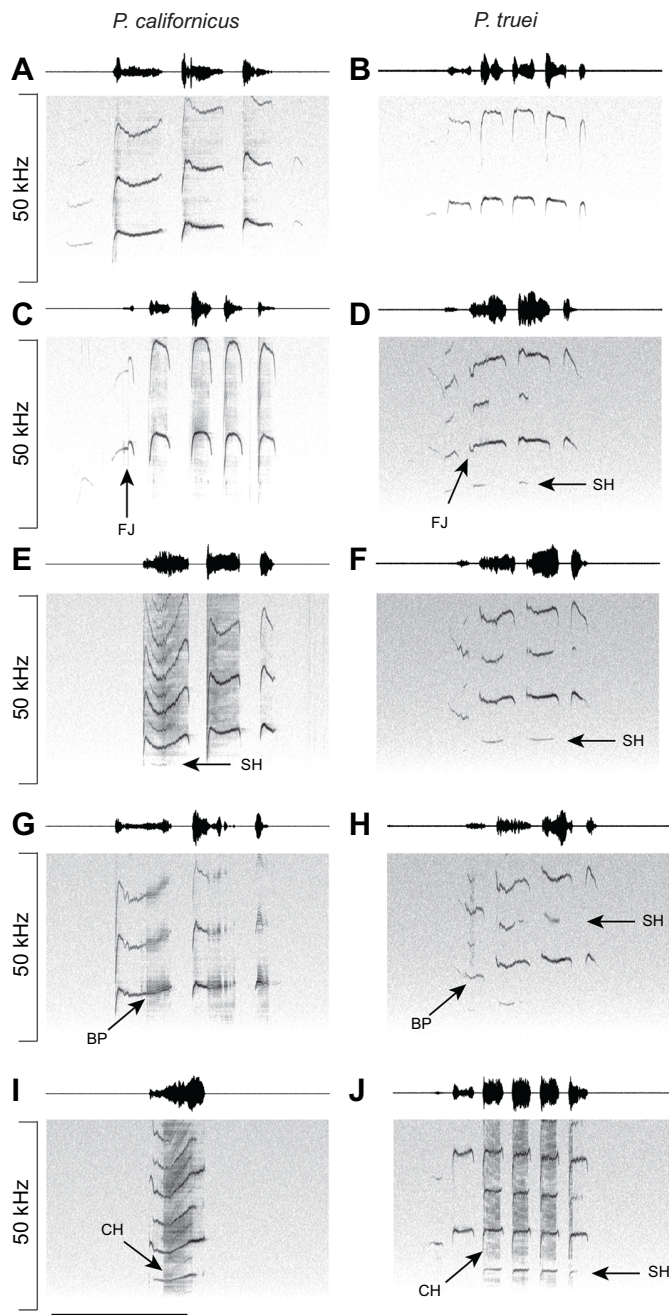


Fig. 3. Nonlinear phenomena in *Peromyscus californicus* and *Peromyscus truei* sustained vocalization calls. (A,B) The syllables do not contain any nonlinear phenomena. (C,D) Frequency jumps (FJs) are sudden upward or downward changes in fundamental frequency. (E,F) Several syllables contain subharmonics (SHs) which appear as sudden additional lines between harmonics representing integer fractional values of an identifiable fundamental frequency (e.g. $f_{0/2}$, $f_{0/3}$ and so on) and as harmonics of these values. (G,H) Biphonation (BP) refers to two simultaneous independent fundamental frequencies. The example in the first syllable shows two lines moving in opposite directions above and below the fundamental frequency. (I,J) Deterministic chaos (CH) represent nonrandom noise which is characterized by a sudden onset and the occurrence of harmonic windows and traces of subharmonics. The reference bar in I is 1 s and applies to all panels.

more frequency jumps, 95 SVs (12.1%) contained one or more subharmonic segments, 3 SVs (0.4%) had one or more chaotic segments and 14 SVs (1.8%) exhibited biphonation. We found 8 calls (1.0%) that exhibited different combinations

of two types of NLP. NLP duration ranged between 26 and 48% of call duration (Table 3), indicating substantial variation in calls among mice. We also screened 20 calls from each of six 2-day-old pups and found that a proportion of 0 to 90% of calls contained NLP.

In *P. truei*, females vocalized extensively during social isolation, while males produces very few vocalizations (Table 4). The percentage of SVs containing at least one type of NLP within individuals ranged between 0.7 to 48% (Table 4). Similarly to *P. californicus*, *P. truei* produced more subharmonics than other NLP types, with a wide range of within-individual variability (0 to 40.7%, Table 4). Cumulatively across all *P. truei*, out of 3052 SV syllables that contained NLP, 242 SVs (7.9%) had frequency jumps, 1410 SVs (46.2%) contained one or more subharmonic segments, 279 SVs (9.1%) had one or more chaotic segments, and 14 SVs (0.5%) exhibited biphonation. Interestingly, *P. truei* produced more calls (650 calls, 21.3%) that contained >2 NLP types relative to *P. californicus*. Lastly, we discovered that duration of NLPs varied widely among individuals (3.9 to 97%).

Larynx anatomy

Thyroid cartilage centroid size was correlated with body size among and within species ($F_{2,45}=100.5$; $P<0.001$). Vocal fold length (measured as distance between vocal process of the arytenoid cartilage and attachment to the interior of the thyroid cartilage) ranged between 595 and 741 μm in the largest of the six species (*P. californicus*) and between 619 and 817 μm in the smallest of six species (*P. polionotus*) (Table 5). Vocal fold length was not predicted by body size neither within nor among species ($F_{2,45}=0.587$; $P=0.56$).

A ventral pouch was present in all 48 individuals investigated by micro-CT imaging (Fig. 4). The pouch is positioned medially and rostral from the vocal folds. The air pocket is separated from the main laryngeal airway by a constriction consisting of alar cartilage. The ventral pouch is surrounded by the thyroid cartilage (Fig. 4). Histological images suggest that the alar cartilage is connected to a branch of the thyroarytenoid muscle which regulates the distance between the glottis and alar edge.

Vocal fold consisted of the thyroarytenoid muscle, lamina propria and epithelium. The thickness of the lamina propria measured between 79 and 133 μm (Table 5) in the six species and was not associated with body size within or among species ($F_{2,45}=0.339$; $P=0.717$). Collagen and elastin fibers were present but not homogeneously distributed in the lamina propria (Fig. 5). Protein density was measured within transects positioned either deep (more laterally) or superficial (in the medial lamina propria). A higher density for both proteins was found in the superficial layer. The ratios between superficial and deep lamina propria for each protein were greater than zero (Fig. 5C,D).

Vocal membranes were present in all four individuals of *P. californicus*, *P. leucopus* and *P. maniculatus*, but only in 2 of 4 *P. eremicus*, 1 of 4 *P. polionotus*, and 3 of 4 *P. truei* (Fig. 5E). In all cases, vocal membranes were positioned symmetrically on both vocal folds. Vocal membranes consisted of lamina propria and an epithelial layer that were single-lobed or consisted of two or more lobes. For example, Fig. 5B shows single lobes for *P. californicus*, *P. eremicus*, *P. leucopus* and *P. maniculatus*, multiple small lobes for *P. polionotus* and two lobes for *P. truei*.

Height and width of vocal membrane-like structures among six species ranged between 29 and 55 μm (height) and from 17 to 37 μm (width) (Fig. 5F,G). We tested whether vocal membrane size scaled with body size within or among species. Height was

Table 4. Rate of occurrence of nonlinear phenomena in *P. truei* SV bouts

Mouse ID	SV bouts	SV syllables	No. SV syllables bout ⁻¹	Syllable duration (s)	NLP (%)	NLP duration (%)	FJ	SH	BP	CH
Female PT02	22	157	7.14±2.3	0.10±0.03	18.5	37.4±22.3	4	24	1	0
Female PT03	1311	6980	5.32±1.7	0.13±0.04	15.3	31.2±25.4	156	908	5	233
Female PT04	115	693	6.02±2.0	0.11±0.04	0.72	16.2±9.9	0	5	0	0
Female PT10	25	141	5.64±1.7	0.14±0.05	19.9	40.7±22.9	0	28	0	3
Female PT14	2	9	4.50±1.7	0.12±0.09	11.1	5.5	0	1	0	0
Female PT16	254	1092	4.30±1.3	0.16±0.08	48.9	54.4±53.7	82	444	8	43
Male PT01	5	13	2.6±1.2	0.06±0.02	0	0	0	0	0	0
Male PT05	1	3	3.0	0.29±0.19	0	0	0	0	0	0
Male PT18	1	3	3.0	0.24±0.14	0	0	0	0	0	0

Values are means±s.d. All SVs were produced by singly housed mice over the course of 3–7 days. NLP, nonlinear phenomena; FJ, frequency jump; SH, subharmonic; BP, biphonation; CH, deterministic chaos.

associated with body size among but not within species ($F_{2,45}=6.71$; $P<0.01$). Width was not associated with body size either among or within species ($F_{2,45}=3.244$; $P=0.067$).

DISCUSSION

Here, we investigated the biology of sound production mechanisms used in acoustic interactions among closely related species of deer mice. Like other cricetid rodents, we found that deer mice use two distinct production mechanisms: whistling to produce high frequency vocalizations in close-distance contexts, and airflow-induced vocal fold vibrations to produce SVs and isolation calls used in longer distance communication. The common occurrence of NLP in adult SVs and in pup isolation calls support the finding that sounds are generated by flow-induced vocal fold vibrations and that such vibrations may be irregular. As in other murid rodents, a characteristic ventral pouch likely facilitates whistle production, and small vocal membranes arising from a two-layered lamina propria presumably underlies SV production. Species differences in vocal fold size and composition may contribute to species-specific acoustic properties. We discuss our findings in relation to functional, ontogenetic and evolutionary factors that may influence the diversification of rodent acoustic signals.

Sound production mechanisms and social context

The dual sound production mechanisms found herein correspond to the functional context of vocalizations. Pup isolation calls and adult SVs both function to advertise the sender's presence to conspecifics over distances greater than a body length, either to absent parents (Rieger et al., 2019) or potential mates or rivals (Kobrina et al., 2022; Pultorak et al., 2017; Rieger and Marler, 2018). Employment of flow-induced vocal fold vibration

facilitates production of sounds across a wide frequency range at high amplitudes, both acoustic features exhibit reduced environmental attenuation (Wahlberg and Larsen, 2017). In contrast, all *Peromyscus* spp. simple and complex sweeps were produced by a whistle mechanism. Such low amplitude, high frequency vocalizations are often produced in close-distance (<body length) social contexts where environmental attenuation is less important. Our results correspond to findings in grasshopper mice (*Onychomys* spp.; Pasch et al., 2017), the sister taxon to *Peromyscus*, indicating that such dual production mechanisms may accommodate similar social contexts in many murid rodents.

These findings also underscore the utility of cricetid rodents (e.g. *Onychomys* spp. and *Peromyscus* spp.) in providing new avenues to explore vocal fold form and function, the relationship between physiological and environmental factors in vocal diversification, and commonalities with human speech. Unlike traditional rodent models (*Mus* spp. and *Rattus* spp.) that whistle when they vocalize (Roberts, 1975; Riede, 2011; Riede et al., 2017; Håkansson et al., 2022; Fernández-Vargas et al., 2022; Fig. 1), human studies of flow-induced vocal fold vibration highlight the integration of precise motor control, somatosensory feedback and tissue properties in speech production (e.g. Titze, 1988; Steinecke and Herzel, 1995). In particular, the mechanical demands (linear, shear and impact stress) acting on vocal fold epithelium and lamina propria during speech (Riede et al., 2011; Titze et al., 2016) indicate that strong inferences will require further study of vocal fold use and aging.

NLP

Heliox data indicated that *Peromyscus* spp. SV calls are produced by flow-induced vocal fold vibration. A characteristic feature of such vocalizations are NLP (e.g. Herzel et al., 1994; Tokuda, 2017). NLP were frequently present in SVs and pup isolation calls of

Table 5. Measurements of body and vocal organ size

	Body mass (g) (N=12/species)	Femur length (mm) (N=12/species)	VFL (μm) (N=4/species)	LP depth (μm) (N=4/species)	LP heterogeneity (a.u.) (N=4/species)	Vocal membrane H×W (μm) (N out of 4)
<i>P. californicus</i>	48.6±12.0	19.6±0.8	668±62	53.3±10.6	C: 1.9±0.8 E: 2.9±1.5	33.3×36.7 N=4
<i>P. leucopus</i>	20.2±3.7	15.8±0.5	605±53	39.1±11.6	C: 2.1±1.1 E: 1.5±1.1	34.3×23.0 N=4
<i>P. eremicus</i>	23.1±3.2	16.5±0.9	634±95	50.1±15.2	C: 3.0±1.3 E: 2.6±1.9	38.6×37.1 N=2
<i>P. truei</i>	29.9±5.9	19.0±0.5	654±74	35.9±20.6	C: 2.7±0.4 E: 1.5±0.7	55.2×17.7 N=3
<i>P. maniculatus</i>	20.7±5.5	14.4±1.4	644±77	37.5±3.9	C: 2.5±0.6 E: 1.6±0.3	43.3×23.9 N=4
<i>P. polionotus</i>	15.1±1.4	14.3±0.3	698±64	62.1±21.0	C: 1.7±0.4 E: 1.4±0.5	28.9×21.4 N=1

Values are means±s.d. C, collagen; E, elastin; VFL, vocal fold length; LP, lamina propria.

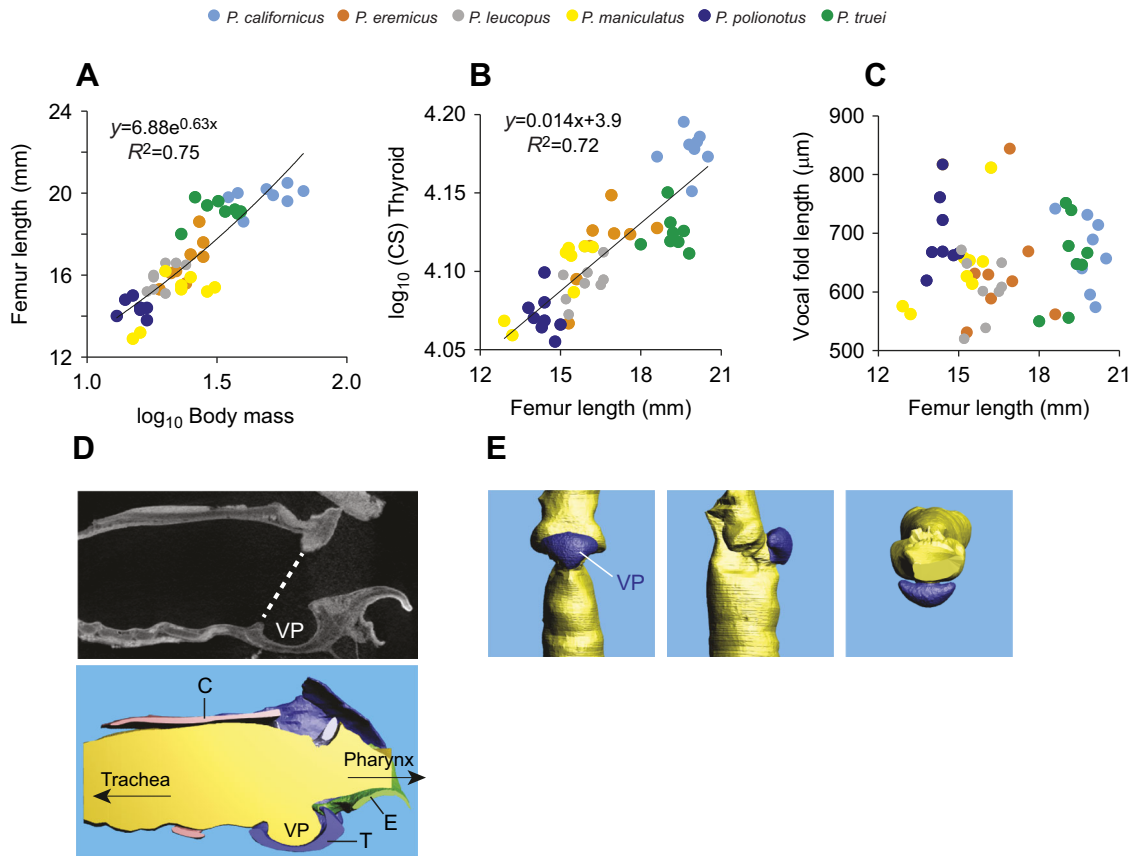


Fig. 4. Vocal organ and glottis size in six *Peromyscus* species. (A) Body mass (g) and femur length are positively correlated. (B) The size of the thyroid cartilage (centroid size, CS) scales allometrically with body size. (C) Vocal fold length does not scale with body size. (D,E) The laryngeal airway in all six species is characterized by the presence of a singular supraglottal positioned ventral pouch (VP). Vocal fold position is indicated by the white dashed line in D (C, cricoid cartilage; T, thyroid cartilage; E, epiglottis). The 3D reconstruction (left to right: ventral view, right lateral view, top view) of the main airway (yellow) and the ventral pouch (blue) for a male *P. californicus* is shown in E.

P. californicus and in SVs of *P. truei*, sometimes occurring in over 50% of vocalizations produced by some individuals. Mechanistically, NLP can arise from asymmetries in vocal fold size (e.g. Tokuda et al., 2007) or nonlinear interactions between the sound source and vocal tract filter resonance (e.g. Titze et al., 2008). Additionally, vocal membrane-like structures (below) may contribute to the nonlinear dynamics of vocal production (Mergell et al., 1999). Mergell et al. (1999) described the vocal membrane as an additional reed-like plate fixed to the vocal fold. Neubauer (2004) updated Mergell's model by allowing the vocal membrane to vibrate independently from the vocal fold. In both models, the addition facilitates higher fundamental frequencies (Mergell et al., 1999; Neubauer, 2004). Such high vibration rates may also promote irregular vibrations that characterize NLP. Experimental manipulation of vocal membrane presence, size and/or symmetry would provide strong inference for their contribution to NLP.

Numerous functional hypotheses have been proposed to explain the presence of nonlinear phenomena in vocalizations. In rodent pups, NLP could serve as an honest signal of distress used to recruit older conspecifics to fend off predators (Blumstein et al., 2008). In adults, NLP may facilitate receiver arousal and fear, consequently increasing predator vigilance and decreasing habituation to alarm calls (i.e. unpredictability hypothesis; Blumstein and Récapet, 2009). NLP may also be associated with personality (Lee et al., 2021) and play a role in individual recognition and discrimination (e.g. Wilden et al., 1998; Volodina et al., 2006). Conversely, NLP

may represent non-functional side-effects of vocal disorders (e.g. Tokuda et al., 2001.), overuse (e.g. Vilkman, 2004) or age (e.g. Baken, 2005; Marx et al., 2021). Given increased documentation of NLP in rodent vocalizations (Blumstein et al., 2008; Miller and Engstrom, 2010, 2012), the significance of NLP awaits further experimentation.

Anatomical mechanisms

Peromyscus spp. vocal folds possess a narrow lamina propria with a characteristic organization of fibrillary proteins, suggesting differentiation into a superficial and a deep layer. The lamina propria together with the epithelium forms vocal membranes near the free edge of the vocal fold. Flow-induced vocal fold vibration is characterized by phase differences in tissue movement between the upper (cranial) and lower (caudal) portions of the lamina propria. In *Peromyscus* spp. (this study) as well as in another cricetid rodent genera, *Onychomys* (Pasch et al., 2017), vocal membrane-like structures likely support such cranial-caudal phase difference owing to their position in the laryngeal lumen that creates a concave-shaped vocal fold surface in the coronal plane (Thomson et al., 2005). Vocal fold design is critical for multiple aspects of voice characteristics. Investigations of a nonhuman primate larynx suggested that vocal membranes facilitate sound production at higher efficiency, i.e. greater loudness for a given lung pressure (Zhang et al., 2019). Species-specific lamina propria morphology also defines a characteristic fundamental frequency range

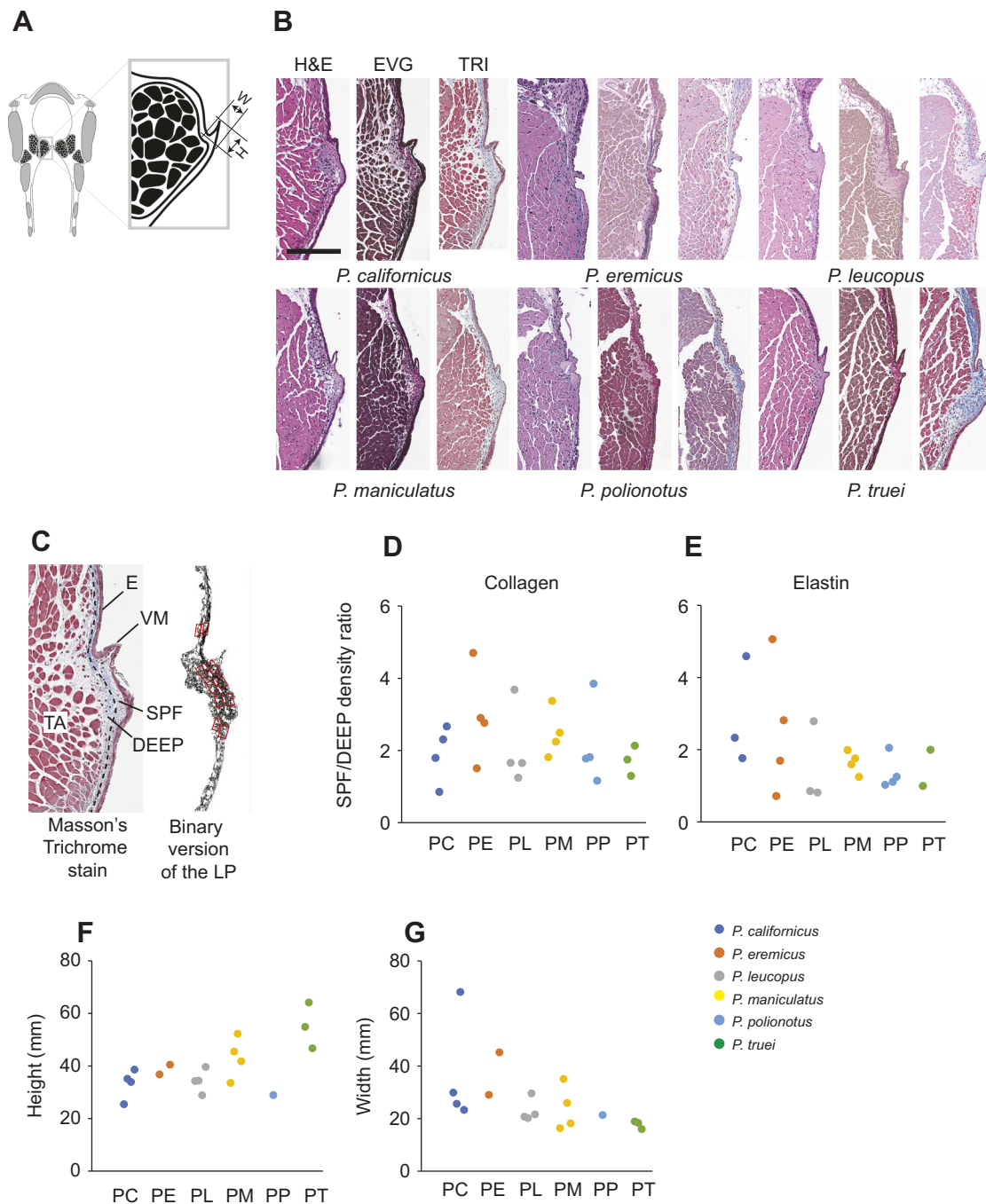


Fig. 5. Heterogeneous lamina propria design and size of vocal membranes in six *Peromyscus* species. (A) Schematic of a coronal larynx section. The location of the coronal vocal fold sections in B are indicated by the small box. (B) Coronal left vocal fold sections of six *Peromyscus* species. Black bar reference indicates 200 μm and applies to all panels in B. The three images for each species are successive sections stained with Hematoxylin and Eosin (H&E), Elastic-van-Gieson (EVG) and Masson's Trichrome (TRI). Elastin fibers stain black with EVG and collagen fibers appear as deep red staining, notably cranially to the free edge. In the TRI stain, blue-staining collagen fibers are organized superficially in the lamina propria (LP), most concentrated just below the epithelium. (C) In order to quantify collagen and elastin concentration in the lamina propria, first a binary version of the lamina propria was generated in ImageJ tracking all blue (collagen in TRI stain) or black (elastin in EVG stain) pixels, respectively. Then 10 transects (red rectangles) were placed in the superficial and deep lamina propria, respectively. The ratio in protein concentration between the superficial and deep lamina propria for collagen (D) and elastin (E) were all larger than 1, which suggests that fibers of both proteins are more densely packed in the superficial lamina propria. Vocal membranes were measured in height (F) and width (G). See A for an explanation of height (H) and width (W). TA, thyroarytenoid muscle; E, epithelium; VM, vocal membrane; SFP, superficial (or medial) lamina propria; DEEP, deep (or lateral) lamina propria.

determined by compressional (lateral) and tensile (along the length) stiffness of the collagen and elastin fibers in the superficial layer (Titze et al., 2016). Indeed, our data provide preliminary support for a relationship among fundamental frequency, vocal fold size and

collagen elastin consistency, since all vary among species (Table 1, Fig. 5), after controlling for body size (Fig. 4). Formal analysis of causal relationships among these variables is currently under investigation.

Interestingly, vocal membranes were not present in every adult investigated in this study (Table 5). Could such features be artifacts of faulty tissue removal, fixation, tissue processing, embedding, microtomy, staining and mounting procedures? Both historical and more recent reviews of common histological artifacts did not include structures that resemble small folds protruding from the lamina propria and epithelium if the underlying tissue is fully intact (i.e. not torn or cut) (e.g. Mehregan and Pinkus, 1966; Kumar et al., 2012; Taqi et al., 2018). Therefore, until *in vivo* observations of mouse vocal folds become available, we infer that histological images of vocal membranes represent the *in vivo* situation of the free edge of the mouse vocal fold. It is unclear whether vocal membranes are normal variations of vocal fold anatomy or a consequence of stresses and strains associated with use. Vocal fold nodules in humans, like vocal membrane-like structures in *Peromyscus* spp., are bilateral, symmetrical structures (e.g. Hirano et al., 1990; Ford et al., 1996; Glanz et al., 1997; Giovanni et al., 2007). Humans with nodules and other lesions may experience voice irregularities, which are nonlinear phenomena (Baken, 2005). Although vocal fold lesions remain an idiopathic disease, i.e. a disease with unknown cause, they tend to be more common among people using their voice professionally (teachers, actors, singers etc.) (e.g. Vilkman, 2004). In *Peromyscus* spp., future studies that characterize the ontogeny of acoustic properties coincident with developmental changes of vocal folds will elucidate the functional morphology of their vocal folds.

Social origin of vocalization is paralleled by vocal production mechanisms

Isolation calls that solicit maternal attention and care (Wöhr and Schwarting, 2008) often serve as precursors to adult vocalizations used in other social contexts (e.g. Oller et al., 2016; Pistorio et al., 2006; Matrosova et al., 2007). For example, adult *Mus* spp. and *Rattus* spp. vocal repertoires used in mating contexts likely emerge from pup vocalizations (Hofer, 2010; Brudzynski, 2014) based on spectro-temporal similarities (Wöhr and Schwarting, 2008; Hofer, 2010). Similarly, spectro-temporal similarities occur between pup isolation vocalizations and adult SVs in *P. californicus* (Johnson et al., 2017). Both consist of bouts of 2–4 syllables with fundamental frequency ranges between 25 and 30 kHz, only slightly above the 15–24 kHz range for adult SV calls. Our findings confirm this observation by noting similarities in the duration of individual syllables in pup isolation calls (50–200 ms; Johnson et al., 2017; Kalcounis-Rueppell et al., 2018c) and adult SVs (this study; Table 1).

Importantly, we found consistency in the sound production mechanism between pup calls and adult SVs in *Peromyscus* spp.: both were produced using flow-induced vocal fold vibration. Stability in production also occurs in *Mus* spp. and *Rattus* spp. pup and adult vocalizations, albeit using a whistle mechanism (Roberts, 1975; Riede, 2011). Such developmental stability may constrain the evolution of rodent vocalizations, both contextually and in acoustic content. For example, while *Mus* spp. and *Rattus* spp. rely almost exclusively on ultrasonic whistling for social communication, many cricetids appear to whistle only as adults (although *Peromyscus* spp. pups occasionally produce high frequency whistles). Similarly, many pup isolation calls show surprising spectral overlap with adult vocalizations, which is puzzling because size-dependent spectral properties would dictate a more dramatic decrease in fundamental frequency. Together, our finding highlights the need for further comparative studies that specify the ontogeny and mechanisms of vocal repertoires, including the origins of whistling. At the very least, our results challenge the predator escape hypothesis given that

altricial *Peromyscus* spp. pups produce relatively low frequency calls audible to predators at their most vulnerable life stage.

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Competing interests

The authors declare no competing or financial interests.

Author contributions

Conceptualization: T.R., B.P.; Methodology: T.R.; Software: T.R.; Validation: T.R.; Formal analysis: T.R., A.K., L.B., T.D., B.P.; Investigation: T.R.; Resources: T.R.; Data curation: T.R., A.K., L.B., T.D., B.P.; Writing - original draft: T.R., B.P.; Writing - review & editing: T.R., A.K., B.P.; Visualization: T.R.; Supervision: T.R., B.P.; Project administration: T.R., B.P.; Funding acquisition: T.R., B.P.

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Data availability

Derived 3D surfaces of airways have been archived at Morphobank (project #P4106): https://morphobank.org/index.php/Projects/ProjectOverview/project_id/4106.

References

- Baken, R. J. (2005). The aged voice: a new hypothesis. *J. Voice* **19**, 317–325. doi:10.1016/j.jvoice.2004.07.005
- Bedford, N. L. and Hoekstra, H. E. (2015). The natural history of model organisms: *Peromyscus* mice as a model for studying natural variation. *elife* **4**, e06813. doi:10.7554/eLife.06813
- Blanchard, R. J. and Blanchard, D. C. (1989). Antipredator defensive behaviors in a visible burrow system. *J. Comp. Psychol.* **103**, 70–82. doi:10.1037/0735-7036.103.1.70
- Blumstein, D. T. and Récapet, C. (2009). The sound of arousal: the addition of novel non-linearities increases responsiveness in marmot alarm calls. *Ethology* **115**, 1074–1081. doi:10.1111/j.1439-0310.2009.01691.x
- Blumstein, D. T., Richardson, D. T., Cooley, L., Winternitz, J. and Daniel, J. C. (2008). The structure, meaning and function of yellow-bellied marmot pup screams. *Anim. Behav.* **76**, 1055–1064. doi:10.1016/j.anbehav.2008.06.002
- Borgard, H. L., Baab, K., Pasch, B. and Riede, T. (2020). The shape of sound: a geometric morphometrics approach to laryngeal functional morphology. *J. Mammal. Evol.* **27**, 577–590. doi:10.1007/s10914-019-09466-9
- Briggs, J. R. and Kalcounis-Rueppell, M. C. (2011). Similar acoustic structure and behavioural context of vocalizations produced by male and female California mice in the wild. *Anim. Behav.* **82**, 1263–1273. doi:10.1016/j.anbehav.2011.09.003
- Brudzynski, S. (2014). Social origin of vocal communication in rodents. In *Biocommunication of Animals* (ed. G. Witzani), pp. 63–79. Springer Science.
- Eisenberg, J. F. (1961). Studies on the behavior of *Peromyscus maniculatus gambelii* and *Peromyscus californicus* parasiticus. *Behaviour* **19**, 177–207. doi:10.1163/156853962X00014
- Fernández-Vargas, M., Riede, T. and Pasch, B. (2022). Mechanisms and constraints underlying acoustic variation in rodents. *Animal Behavior*. **184**, 135–147. doi:10.1016/j.anbehav.2021.07.011
- Ford, C. N., Inagi, K., Khidr, A., Bless, D. M. and Gilchrist, K. W. (1996). Sulcus vocalis: a rational analytical approach to diagnosis and management. *Ann. Otol. Rhinol. Laryngol.* **105**, 189–200. doi:10.1177/000348949610500304
- Giovanni, A., Chanteret, C. and Lagier, A. (2007). Sulcus vocalis: a review. *Eur. Arch. Otorhinolaryngol.* **264**, 337. doi:10.1007/s00405-006-0230-8
- Glanz, H., Schulz, A., Kleinsasser, O., Schulze, W., Dreyer, T. and Arens, C. (1997). Benign lesions of the larynx: basic clinical and histopathological data. In *Advances in Laryngology in Europe* (ed. O. Kleinsasser, H. Glanz and J. Olofson), pp. 3–14. Elsevier Science.
- Håkansson, J., Jiang, W., Xue, Q., Zheng, X., Ding, M., Agarwal, A. A. and Elemans, C. P. (2022). Aerodynamics and motor control of ultrasonic vocalizations for social communication in mice and rats. *BMC Biol.* **20**, 1–15. doi:10.1186/s12915-021-01185-z
- Hart, F. M. and King, J. A. (1966). Distress vocalizations of young in two subspecies of *Peromyscus maniculatus*. *J. Mammal.* **47**, 287–293. doi:10.2307/1378125
- Herzel, H. (1993). Bifurcation and chaos in voice signals. *Appl. Mech. Rev.* **46**, 399–413. doi:10.1115/1.3120369

- Herzel, H., Berry, D., Titze, I. R. and Saleh, S. (1994). Analysis of vocal disorders with methods from nonlinear dynamics. *J. Speech Hear. Res.* **37**, 1008-1019. doi:10.1044/jshr.3705.1008
- Herzel, H., Berry, D., Titze, I. and Steinecke, I. (1995). Nonlinear dynamics of the voice - signal analysis and biomechanical modeling. *Chaos* **5**, 30-34. doi:10.1063/1.166078
- Hirano, M., Yoshida, T., Tanaka, S. and Hibi, S. (1990). Sulcus vocalis: functional aspects. *Ann. Oto. Rhino. Laryngol.* **99**, 679-683. doi:10.1177/000348949009900901
- Hofer, M. A. (2010). Evolution of the infant separation call: rodent ultrasonic vocalization. In *Handbook of Mammalian Vocalization: An Integrative Neuroscience Approach*, 1st edn (ed. S. M. Brudzynski), pp. 29-35. Amsterdam: Academic/Elsevier.
- Johnson, S. A., Painter, M. S., Javurek, A. B., Murphy, C. R., Howald, E. C., Khan, Z. Z., Conard, C. M., Gant, K. L., Ellersieck, M. R., Hoffmann, F. et al. (2017). Characterization of vocalizations emitted in isolation by California mouse (*Peromyscus californicus*) pups throughout the postnatal period. *J. Comp. Psychol.* **131**, 30-39. doi:10.1037/com0000057
- Kalcounis-Rueppell, M. C., Metheny, J. D. and Vonnhof, M. J. (2006). Production of ultrasonic vocalizations by *Peromyscus* mice in the wild. *Frontiers Zool.* **3**, 1-12. doi:10.1186/1742-9994-3-3
- Kalcounis-Rueppell, M. C., Petric, R., Briggs, J. R., Carney, C. and Marshall, M. M. (2010). Differences in ultrasonic vocalizations between wild and laboratory California mice (*Peromyscus californicus*). *PLoS one* **5**, e9705. doi:10.1371/journal.pone.0009705
- Kalcounis-Rueppell, M. C., Petric, R. and Marler, C. A. (2018a). The bold, silent type: predictors of ultrasonic vocalizations in the genus *Peromyscus*. *Front. Ecol. Evol.* **6**, 198. doi:10.3389/fevo.2018.00198
- Kalcounis-Rueppell, M. C., Pultorak, J. D. and Marler, C. A. (2018b). Ultrasonic vocalizations of mice in the genus *Peromyscus*. *Handbook Behav. Neurosci.* **25**, 227-235. doi:10.1016/B978-0-12-809600-0.00022-6
- Kalcounis-Rueppell, M. C., Pultorak, J. D., Blake, B. H. and Marler, C. A. (2018c). Ultrasonic vocalizations of young mice in the genus *Peromyscus*. *Handbook Behav. Neurosci.* **25**, 149-156. doi:10.1016/B978-0-12-809600-0.00014-7
- Kobrina, A., Letowt, M. E. and Pasch, B. (2022). The influence of social context on pinyon mouse (*Peromyscus truei*) vocalizations. *J. Mammal.* **103**, 275-286. doi:10.1093/jmammal/gyab127
- Koda, H., Tokuda, I. T., Wakita, M., Ito, T. and Nishimura, T. (2015). The source-filter theory of whistle-like calls in marmosets: acoustic analysis and simulation of helium-modulated voices. *J. Acoust. Soc. Am.* **137**, 3068. doi:10.1121/1.4921607
- Kumar, K., Shetty, D. C. and Dua, M. (2012). Biopsy and tissue processing artifacts in oral mucosal tissues. *Int. J. Head Neck Surg.* **3**, 92-98. doi:10.5005/jp-journals-10001-1102
- Lee, J. M., Roy, N., Park, A., Muntz, H., Meier, J., Skirko, J. and Smith, M. (2021). Personality in children with vocal fold nodules: a multitrait analysis. *J. Speech Lang. Hear. Res.* **64**, 3742-3758. doi:10.1044/2021_JSLHR-21-00144
- Madsen, P. T., Jensen, F. H., Carder, D. and Ridgway, S. (2012). Dolphin whistles: a functional misnomer revealed by heliox breathing. *Biol. Lett.* **8**, 211-213. doi:10.1098/rsbl.2011.0701
- Marx, A., Lenkei, R. and Pérez Fraga, P. (2021). Occurrences of non-linear phenomena and vocal harshness in dog whines as indicators of stress and ageing. *Sci. Rep.* **11**, 4468. doi:10.1038/s41598-021-83614-1
- Matrosova, V. A., Volodin, I. A., Volodina, E. V. and Babitsky, A. F. (2007). Pups crying bass: vocal adaptation for avoidance of age-dependent predation risk in ground squirrels? *Behav. Ecol. Sociobiol.* **62**, 181-191. doi:10.1007/s00265-007-0452-9
- Mehregan, A. H. and Pinkus, H. (1966). Artifacts in dermal histopathology. *Arch. Dermatol.* **94**, 218-225. doi:10.1001/archderm.1966.01600260110018
- Mergell, P., Fitch, W. T. and Herzel, H. (1999). Modeling the role of nonhuman vocal membranes in phonation. *J. Acoust. Soc. Am.* **105**, 2020-2028. doi:10.1121/1.426735
- Miller, J. R. and Engstrom, M. D. (2010). Stereotypic vocalizations in harvest mice (Reithrodontomys): Harmonic structure contains prominent and distinctive audible, ultrasonic, and non-linear elements. *J. Acoust. Soc. Am.* **128**, 1501-1510. doi:10.1121/1.3455855
- Miller, J. R. and Engstrom, M. D. (2012). Vocal stereotypy in the rodent genera *Peromyscus* and *Onychomys* (Neotominae): Taxonomic signature and call design. *Bioacoustics* **21**, 193-213. doi:10.1080/09524622.2012.675176
- Neubauer, J. (2004). Nonlinear dynamics of the voice: Bifurcations and mode analysis of complex spatio-temporal signals. *PhD Thesis*, Humboldt University of Berlin.
- Oller, D. K., Griebel, U. and Warlaumont, A. S. (2016). Vocal development as a guide to modeling the evolution of language. *Topics* **8**, 382-392.
- Pasch, B., Tokuda, I. T. and Riede, T. (2017). Grasshopper mice employ distinct vocal production mechanisms in different social contexts. *Proc. R. Soc. B Biol. Sci.* **284**, 20171158. doi:10.1098/rspb.2017.1158
- Pistorio, A. L., Vintch, B. and Wang, X. (2006). Acoustic analysis of vocal development in a New World primate, the common marmoset (*Callithrix jacchus*). *J. Acoust. Soc. Am.* **120**, 1655-1670. doi:10.1121/1.2225899
- Pultorak, J. D., Fuxjager, M. J., Kalcounis-Rueppell, M. C. and Marler, C. A. (2015). Male fidelity expressed through rapid testosterone suppression of ultrasonic vocalizations to novel females in the monogamous California mouse. *Horm. Behav.* **70**, 47-56. doi:10.1016/j.yhbeh.2015.02.003
- Pultorak, J. D., Matusinec, K. R., Miller, Z. K. and Marler, C. A. (2017). Ultrasonic vocalization production and playback predicts intrapair and extrapair social behaviour in a monogamous mouse. *Anim. Behav.* **125**, 13-23. doi:10.1016/j.anbehav.2016.12.023
- Riede, T. (2011). Subglottal pressure, tracheal airflow, and intrinsic laryngeal muscle activity during rat ultrasound vocalization. *J. Neurophysiol.* **106**, 2580-2592. doi:10.1152/jn.00478.2011
- Riede, T. (2013). Stereotypic laryngeal and respiratory motor patterns generate different call types in rat ultrasound vocalization. *J. Exp. Zool. A Ecol. Genet. Physiol.* **319**, 213-224. doi:10.1002/jez.1785
- Riede, T. and Pasch, B. (2020). Pygmy mouse songs reveal anatomical innovations underlying acoustic signal elaboration in rodents. *J. Exp. Biol.* **223**, 223925. doi:10.1242/jeb.223925
- Riede, T., Wilden, I. and Tembrock, G. (1997). Subharmonics, biphonations, and frequency jumps-common components of mammal vocalization or indicators for disorders? *Zeitschrift für Säugetierkunde* **62**, 198-203.
- Riede, T., Herzel, H., Mehwald, D., Seidner, W., Trumler, E., Böhme, G. and Tembrock, G. (2000). Nonlinear phenomena in the natural howling of a dog-wolf mix. *J. Acoust. Soc. Am.* **108**, 1435-1442. doi:10.1121/1.1289208
- Riede, T., Owren, M. J. and Arcadi, A. C. (2004). Nonlinear acoustics in pant hoots of common chimpanzees (*Pan troglodytes*): frequency jumps, subharmonics, biphonation, and deterministic chaos. *Am. J. Primatol.* **64**, 277-291. doi:10.1002/ajp.20078
- Riede, T., Arcadi, A. C. and Owren, M. J. (2007). Nonlinear acoustics in the pant hoots of common chimpanzees (*Pan troglodytes*): vocalizing at the edge. *J. Acoust. Soc. Am.* **121**, 1758-1767. doi:10.1121/1.2427115
- Riede, T., York, A., Furst, S., Müller, R. and Seebeck, S. (2011). Elasticity and stress relaxation of a very small vocal fold. *J. Biomech.* **44**, 1936-1940. doi:10.1016/j.jbiomech.2011.04.024
- Riede, T., Borgard, H. L. and Pasch, B. (2017). Laryngeal airway reconstruction indicates that rodent ultrasonic vocalizations are produced by an edge-tone mechanism. *R. Soc. Open Sci.* **4**, 170976. doi:10.1098/rsos.170976
- Riede, T., Coyne, M., Tafaya, B. and Baab, K. L. (2020). Postnatal development of the mouse larynx: negative allometry, age-dependent shape changes, morphological integration, and a size-dependent spectral feature. *J. Speech Lang. Hear. Res.* **63**, 2680-2694. doi:10.1044/2020_JSLHR-20-00070
- Rieger, N. S. and Marler, C. A. (2018). The function of ultrasonic vocalizations during territorial defence by pair-bonded male and female California mice. *Anim. Behav.* **135**, 97-108. doi:10.1016/j.anbehav.2017.11.008
- Rieger, N. S., Stanton, E. H. and Marler, C. A. (2019). Division of labour in territorial defence and pup retrieval by pair-bonded California mice, *Peromyscus californicus*. *Anim. Behav.* **156**, 67-78. doi:10.1016/j.anbehav.2019.05.023
- Roberts, L. H. (1975). The rodent ultrasound production mechanism. *Ultrasonics* **13**, 83-88. doi:10.1016/0041-624X(75)90052-9
- Sikes, R. S. and Animal Care and Use Committee of the American Society of Mammalogists (2016). 2016 Guidelines of the American Society of Mammalogists for the use of wild mammals in research and education. *J. Mammal.* **97**, 663-688. doi:10.1093/jmammal/gyw078
- Smith, J. C. (1972). Sound production by infant *Peromyscus maniculatus* (Rodentia: Myomorph). *J. Zool. Lond.* **168**, 369-379. doi:10.1111/j.1469-7998.1972.tb01352.x
- Steinecke, I. and Herzel, H. (1995). Bifurcations in an asymmetric vocal-fold model. *J. Acoust. Soc. Am.* **97**, 1874-1884. doi:10.1121/1.412061
- Taqi, S. A., Sami, S. A., Sami, L. B. and Zaki, S. A. (2018). A review of artifacts in histopathology. *J. Oral Maxillofac. Pathol.* **22**, 279. doi:10.4103/jomp.JOMFP_125_15
- Thomson, S. L., Mongeau, L. and Frankel, S. H. (2005). Aerodynamic transfer of energy to the vocal folds. *J. Acoust. Soc. Am.* **118**, 1689-1700. doi:10.1121/1.2000787
- Titze, I. R. (1988). The physics of small-amplitude oscillation of the vocal folds. *J. Acoust. Soc. Am.* **83**, 1536-1552. doi:10.1121/1.395910
- Titze, I. R., Riede, T. and Popolo, P. (2008). Nonlinear source-filter coupling in phonation: Vocal exercises. *J. Acoust. Soc. Am.* **123**, 1902-1915. doi:10.1121/1.2832339
- Titze, I., Riede, T. and Mau, T. (2016). Predicting achievable fundamental frequency ranges in vocalization across species. *PLoS Comput. Biol.* **12**, e1004907. doi:10.1371/journal.pcbi.1004907
- Tokuda, I. T. (2017). *Nonlinear Dynamics and Temporal Analysis: Comparative Bioacoustics: an Overview*. Oak Park: Bentham Science.
- Tokuda, I. T., Riede, T., Neubauer, J., Owren, M. J. and Herzel, H. (2001). Nonlinear analysis of irregular animal vocalizations. *J. Acoust. Soc. Am.* **111**, 2908-2919. doi:10.1121/1.1474440
- Tokuda, I. T., Horáček, J., Švec, J. G. and Herzel, H. (2007). Comparison of biomechanical modeling of register transitions and voice instabilities with excised larynx experiments. *J. Acoust. Soc. Am.* **122**, 519-531. doi:10.1121/1.2741210

- Townsend, S. W. and Manser, M. B.** (2011). The function of nonlinear phenomena in meerkat alarm calls. *Biol. Lett.* **7**, 47–49. doi:10.1098/rsbl.2010.0537
- Vilkman, E.** (2004). Occupational safety and health aspects of voice and speech professions. *Folia Phoniatr. Logop.* **56**, 220–253.
- Volodina, E. V., Volodin, I. A., Isaeva, I. V. and Unck, C.** (2006). Biphonation may function to enhance individual recognition in the dhole, *Cuon alpinus*. *Ethology* **112**, 815–825. doi:10.1111/j.1439-0310.2006.01231.x
- Wahlberg, M. and Larsen, O. N.** (2017). Propagation of sound. In *Comparative Bioacoustics: An Overview*, pp. 61–120. Bentham Science Publishers.
- Wilden, I., Herzel, H., Peters, G. and Tembrock, G.** (1998). Subharmonics, biphonation, and deterministic chaos in mammal vocalization. *Bioacoustics* **9**, 171–196. doi:10.1080/09524622.1998.9753394
- Wöhr, M. and Schwarting, R. K.** (2008). Maternal care, isolation-induced infant ultrasonic calling, and their relations to adult anxiety-related behavior in rats. *Behav. Neurosci.* **122**, 310–330. doi:10.1037/0735-7044.122.2.310
- Zelditch, M. L., Lundrigan, B. L. and Garland, T., Jr** (2004). Developmental regulation of skull morphology. I. Ontogenetic dynamics of variance. *Evol. Dev.* **6**, 194–206. doi:10.1111/j.1525-142X.2004.04025.x
- Zhang, Y. S., Takahashi, D. Y., Liao, D. A., Ghazanfar, A. A. and Elemans, C. P.** (2019). Vocal state change through laryngeal development. *Nat. Commun.* **10**, 1–12. doi:10.1038/s41467-018-07882-8
- Zollinger, S. A., Riede, T. and Suthers, R. A.** (2008). Two-voice complexity from a single side of the syrinx in northern mockingbird *Mimus polyglottos* vocalizations. *J. Exp. Biol.* **211**, 1978–1991. doi:10.1242/jeb.014092