

# A Highly Scalable Approach to Perform Ecological Surveys of Selfing *Caenorhabditis* Nematodes

Timothy A. Crombie<sup>1</sup>, Robyn E. Tanny<sup>1</sup>, Claire M. Buchanan<sup>1</sup>, Nicole M. Roberto<sup>1</sup>, Erik C. Andersen<sup>1</sup>

<sup>1</sup> Department of Molecular Biosciences, Northwestern University

## Corresponding Author

Erik C. Andersen  
erik.andersen@gmail.com

## Citation

Crombie, T.A., Tanny, R.E., Buchanan, C.M., Roberto, N.M., Andersen, E.C. A Highly Scalable Approach to Perform Ecological Surveys of Selfing *Caenorhabditis* Nematodes. *J. Vis. Exp.* (), e63486, doi:10.3791/63486 (2021).

## Date Published

December 15, 2021

## DOI

10.3791/63486

## URL

jove.com/video/63486

## Abstract

*Caenorhabditis elegans* is one of the major model organisms in biology, but only recently have researchers focused on its natural ecology. The relative sparsity of information about *C. elegans* in its natural context comes from the challenges involved in the identification of the small nematode in nature. Despite these challenges, an increasing focus on the ecology of *C. elegans* has caused a wealth of new information regarding its life outside of the laboratory. The intensified search for *C. elegans* in nature has contributed to the discovery of many new *Caenorhabditis* species and revealed that congeneric nematodes frequently cohabitate in the wild, where they feed on microbial blooms associated with rotting plant material. The identification of new species has also revealed that the androdioecious mating system of males and self-fertilizing hermaphrodites has evolved three times independently within *Caenorhabditis*. The other two selfing species, *C. briggsae* and *C. tropicalis*, share the experimental advantages of *C. elegans* and have enabled comparative studies into the mechanistic basis of important traits, including self-fertilization. Despite these advances, much remains to be learned about the ecology and natural diversity of these important species. For example, we still lack functional information for many of their genes, which might only be attained through an understanding of their natural ecology. To facilitate ecological research of selfing *Caenorhabditis* nematodes, we developed a highly scalable method to collect nematodes from the wild. Our method makes use of mobile data collection platforms, cloud-based databases, and the R software environment to enhance researchers' ability to collect nematodes from the wild, record associated ecological data, and identify wild nematodes using molecular barcodes.

## Introduction

The last two decades have brought an increased interest in the ecology of *Caenorhabditis* nematodes. From these

studies, we know that the free-living *Caenorhabditis* species can be isolated from ephemeral micro-habitats in both temperate and tropical regions, where they feed on microbial blooms associated with decomposing plant

material, sometimes in sympatry<sup>1,2,3,4,5,6,7,8</sup>. We have also learned that convergent evolution of self-fertilization has occurred in the genus three times, and selfing is the dominant mode of reproduction for *C. briggsae*, *C. elegans*, and *C. tropicalis*<sup>9,10</sup>. Among these selfers, *C. elegans* is one of the most widely studied animals on Earth and has been used by researchers to make critical advances in biology. Importantly, the other selfing *Caenorhabditis* species share many of *C. elegans* experimental advantages and are rapidly advancing comparative studies in the genus. However, the cryptic nature of these nematodes in the wild makes it difficult to study their ecology and natural diversity, which is critical for understanding the biological functions of their genes and the ways in which evolution has shaped genetic diversity among the species<sup>10,11</sup>.

The greatest challenge to studying the ecology of selfing *Caenorhabditis* nematodes in the wild is their small size; adult nematodes are often 1 mm in length or less. This challenge requires that researchers sample substrates from the wild and attempt to separate nematodes of interest from the substrates in the laboratory without the ability to observe animals in the wild. Because even trained experts find it difficult to discriminate selfing *Caenorhabditis* nematodes from other free-living nematodes under the microscope, nematodes are typically removed from the substrate, isolated, and left to proliferate before they are identified by sequence identity using established molecular barcodes<sup>3,12,13,14</sup>. The time and effort required to process each nematode in this way present an organizational challenge, as researchers must be able to trace the identity of each nematode isolated in the laboratory back to the exact substrate and associated ecological data sampled in the field. Here, we describe a step-by-step process to efficiently collect and identify selfing *Caenorhabditis* nematodes from the field and faithfully link

these isolates with their associated spatial and ecological data at a high scale.

This collection method increases the scale and accuracy of ecological surveys by using mobile data collection platforms, cloud-based databases, and the R software environment. Fulcrum is a customizable data-collection platform that works with most mobile devices and allows users to build custom applications to gather and organize location-based data (<https://www.fulcrumapp.com>). This protocol provides detailed instructions on how to use customized data-collection applications to organize spatially explicit ecological data from the field and accurately link those data with the identity of nematodes isolated in the laboratory. The protocol also explains how to efficiently identify selfing *Caenorhabditis* nematodes using established molecular barcodes. The data from these methods can be processed simply and reproducibly with the accompanying R software package *easyFulcrum*<sup>15</sup> to explore the ecology and genetic diversity of natural *Caenorhabditis* populations.

## Protocol

### 1. Collection preparation

1. Identify a location to survey *Caenorhabditis* nematodes.  
**NOTE:** In most temperate regions, *C. elegans* and *C. briggsae* can be isolated easily from human-associated habitats like agricultural fields or rural and urban gardens<sup>1</sup>. In subtropical and tropical regions, *C. briggsae*, *C. elegans*, and *C. tropicalis* can all be found in the human-associated habitats listed above, sometimes in close proximity to one another. However, *C. elegans* seems to prefer cooler, drier habitats than the other species in tropical habitats<sup>7,8</sup>. Each of the species can

also be isolated from wild habitats that are not associated with humans, but these habitats are sampled less often.

2. Create a Fulcrum project to organize the collection and isolation data with the mobile data-collection applications.
  1. Create an account with Fulcrum online using a no-cost educational agreement<sup>16</sup>. Add the **Nematode Field Sampling** application to a Fulcrum account by clicking on **ADD APP** button<sup>17</sup>.
  2. Add the **Nematode Isolation** application to an account by clicking on **ADD APP** button<sup>18</sup>.
3. Add users to the Fulcrum account to grant them access to the collection project. Ensure that each user downloads the Fulcrum mobile application to participate in the project.
4. Print a set of QR-code labels to track the collections (C-labels) and the nematode isolations (S-labels) with the mobile application. Attach the C-labels to zip-lock plastic bags, roll the labeled bags into groups of 25, and wrap them with a rubber band for packing. Keep the set of S-labels for use in the laboratory.

**NOTE:** Throughout this protocol, the collections (substrates from the field) are contained in bags or on plates and are labeled with C-labels. The isolated nematodes are labeled with S-labels. The C-labels are used to identify unique collections, and the S-labels are used to identify unique nematode isolates. These two types of labels are used to make the connection between a particular collection (C-label) and the nematodes

isolated from that collection (S-labels) in the Fulcrum database. Print twice the number of S-labels as C-labels for a collection project because, on average, two nematodes are isolated per collection. More S-labels can be printed later if needed. 2,500 unique C-labels (**Supplemental File 1**) and 5,000 unique S-labels (**Supplemental File 2**) are provided in the supplement.

5. Prepare 10 cm NGMA plates for collections and 3.5 cm NGMA plates for isolating nematodes. Make one 10 cm plate and at least two 3.5 cm plates per collection<sup>21</sup>. These plates are seeded with *Escherichia coli* strain OP50 following established protocols. Store the plates prior to use at 4 °C for no more than 1 month.

## 2. Field collection

**NOTE:** *Caenorhabditis* nematodes are most often isolated from rotting vegetable material, including fruits, nuts, seeds, pods, flowers, stems, vegetal litter, and compost<sup>1,5,6,8</sup>. The best substrates are rotten and almost unrecognizable as fruits or flowers; avoid substrates that are too dry or wet (**Figure 1**). Substrates are most efficiently collected from the field by working in pairs. The individual with the non-contact infrared thermometer will select a substrate for collection and collect the sample while their partner uses the Nematode Collection Fulcrum application to record the collection data. The pair of collectors will repeat this process until the desired number of samples is collected. The list of materials required for fieldwork is found in (**Supplemental Table 1**).

1. Open the Fulcrum mobile app, select **Nematode Field Sampling** from the drop-down menu. Press **+** to start a new record in the project (**Figure 2A**). Take a photo of the substrate.

2. Click on the box in the top center to select the correct collection project made in step 1.2 (**Figure 2B**). Tap on the **C-label** field at the bottom of the collection record and choose Scan when the prompt appears. **Scan** the barcode on the collection bag using the mobile device camera, then tap on **Done** in the upper right of the screen.
3. Tap on the **Substrate** field and select a substrate type from the drop-down menu. Add notes about the substrate by tapping the **Substrate Notes** field and manually entering notes.
4. Choose a landscape from the drop-down menu. Pick the landscape that best represents the sampling site.
5. Choose a sky view. When choosing sky view, describe the sky visibility at the sampling site (e.g., A full-sky view without obstructed views from trees or other structures = full).
6. Measure the surface temperature of the substrate using the non-contact thermometer and record the value in the substrate temperature field.  
**NOTE:** Hold the non-contact thermometer no more than 14 inches from the substrate while recording the temperature.
7. Measure the ambient temperature and humidity with the handheld device and record these data in the proper fields.  
**NOTE:** Check that the ambient temperature and humidity device are not on **Hold**. The measurement unit will change when the button is released. Keep the device in an outside pocket to avoid irregular readings.
8. Save the record in Fulcrum by tapping on **Save** in the upper left of the screen.
9. Collect about a tablespoon of the substrate without sticks or other hard pieces by inverting the collection bag to use it as a "glove" to pick up the substrate, then seal the bag. Put a paper towel in the bag if the sample is particularly moist.  
**NOTE:** In hot climates, place the bags in soft coolers with cooler packs to keep the collections cool.
10. After all the samples have been collected for the day, clean the collection equipment, take batteries out of probes, recharge batteries, re-freeze freezer packs. Sync the Fulcrum collection data by tapping the Sync button on the top left of the **Nematode Field Sampling** application.  
**NOTE:** The uploads can take several minutes without a strong cellular connection so it might be best to wait for WiFi access. The data will remain on mobile devices and will be synchronized to the cloud.
11. Ship the samples to a home institution by placing them in an overnight shipping box. Minimize the time the samples are exposed to temperatures less than 11 °C or greater than 25 °C by shipping packages on days when cargo is transported.  
**NOTE:** Most shipping facilities do not ship cargo overnight on weekends in remote locations.

### 3. Plating out field collections in the laboratory

**NOTE:** This section details how to organize the transfer of samples from labeled collection bags to labeled plates. The samples may arrive from an overnight shipment or directly from the field.

1. Receive the shipment of collections and inspect for broken bags or other evidence of damage. If bags are broken, discard the material and clean the unbroken collection bags with 70% ethanol; avoid the C-label on

the bag with the ethanol as it will discolor the label and make it difficult to read.

2. For each zip-lock bag, note the C-label on the bag and attach a matching C-label to the lid of a 10 cm plate spotted with OP50 bacteria.

**NOTE:** The labeled 10 cm plates are referred to as 'C-plates' for the rest of the protocol. The easiest way to organize the samples is to place the collection bags on a lab bench with the matching C-plate on top (**Figure 3**).

3. For each collection, transfer about one tablespoon of sample from the collection bag to the C-plate using a clean plastic spoon. Add the sample around the bacterial lawn in a crescent or ring shape, do not completely cover the bacterial lawn (**Figure 4**).

**NOTE:** Keep the tablespoon clean by placing it in a beaker of 95% ethanol when it is not in use. Use a paper towel to dry the tablespoon before transferring additional samples.

4. Record the time the collections were transferred from collection bags to C-plates and keep the C-plates at room temperature (RT) for at least 24 h before attempting to isolate nematodes in section 4.

#### 4. Isolating nematodes from collections

1. Open the **Fulcrum** application on the mobile device and choose **Nematode Isolation** from the application menu (**Figure 5A**). Make a new isolation record by tapping the + icon in the lower right (**Figure 5B**).
2. In the new isolation record screen, confirm the correct collection project by checking the project name displayed in the box at the top center. If the wrong project is displayed, tap the project name to switch to the correct project (**Figure 5C**).

3. Tap on the **Select** button under the **C-label** field to find the C-label associated with the sample from which nematodes are being isolated (**Figure 5D**). Tap on the **Search** icon, then tap on the **Scan** icon to scan the C-label QR code on the C-plate with the device camera. Once the QR code is scanned, a C-label record will appear in the **C-label** field.

4. Tap on the **Camera** icon in the **Photos** field to open the device camera and use it to take a photo of the sample on the C-plate with the QR code visible (**Figure 5E**). Tap on **Done** to return to the Isolation screen.

**NOTE:** These isolation record photos can be used to explore specific attributes of the substrate at a later time.

5. Use a dissecting microscope to look for nematodes on the C-plate. Tap on the **Worms on Sample** field to record the presence of nematodes on the sample (**Figure 5F**). Tap on **Yes** if nematodes are present on the C-plate and tap on **No** if no nematodes are present.

6. Tap on **Tracks** if only nematode tracks are present. If no nematodes are present, parafilm the C-plate and dispose of it in a biohazard bin.

**NOTE:** Invert the C-plate over the biohazard waste bin and gently tap the back of the plate to dislodge all of the sampled substrates. This step makes it easier to find and isolate nematodes that can be under the substrate on the C-plate.

7. If nematodes are present, isolate up to five nematodes from the C-plate. To isolate a nematode, transfer one nematode from the C-plate to an S-plate using a platinum wire pick. Isolate healthy, gravid adults if possible. However, isolate other stages if adults are not found.

**NOTE:** After isolation, up to five S-plates will each have a single nematode on them. Keep these S-plate(s) with

isolated nematodes from the same C-plate organized together in a neat stack away from other S-plates until they are entered into Fulcrum.

8. Tap on the **S-labeled Plates** field to enter the S-plate(s) used for this isolation. Tap on the **+** in the lower right. Tap on **S-label** and then click on **Scan** to open the device camera. Use the device camera to scan the S-label QR code on the S-plate.

**NOTE:** Ensure that the S-label code matches the code on the plate. If it matches, tap on **Done**. If it does not, tap on **Cancel** and rescan until it matches, then click on **Done**. Sometimes QR codes of nearby plates are accidentally scanned.

9. After entering each S-plate, save the entry with the **Save** button on top right. The entry will be lost if it is not saved. Tap on the **+** in the lower right to add more S-labeled plates if necessary until all nematodes isolated from the C-plate are entered. After adding all the S-labeled plates for the isolation record, tap on the **<** button on the upper left to go back to the isolation record screen.

**NOTE:** To cancel an isolation record because mistakes cannot be resolved, click on **Cancel** in the upper left. This step will open a dialog asking whether the record can be discarded without saving. If desired, click on **Yes, Discard**.

10. Tap on the **Save** button on the upper right once the isolation record has all information added correctly. Then parafilm the S-plates with isolated nematodes and set them aside in an area designated to hold S-plates with nematodes.
11. Parafilm the C-plate and discard it in the biohazard bin. Tap on the **Sync** icon to upload all the data to Fulcrum.

12. Sort all the S-plates into alphanumeric order, then place the S-plates into cardboard boxes. Make sure the S-plates are lid-side down and parafilm. Stack up to four S-plates into one position in the box and label the cardboard box with the project name, date, time, and a unique box number.
13. Store the labeled boxes at RT. These isolates will be checked for proliferation at 48 h and again at 168 h if necessary.

## 5. Exporting S-plates from Fulcrum

**NOTE:** This section details how to export S-labels used in the isolation process from the Fulcrum project database. These S-labels will be used to track proliferating isofemale lines while they are being identified by sequence identity in sections 6-9.

1. Sign in to the Fulcrum website and select the **Nematode Isolation** application. Click on **Exporter** from the left-hand side of the screen. Click to select the desired project, and check the **Nematode Isolation** box. Click **Next** to download a .zip file that contains the 'nematode\_isolation\_s\_labeled\_plates.csv' file.
2. Open the 'nematode\_isolation\_s\_labeled\_plates.csv' file and sort it by the 'S-label' column in ascending order (the smallest S-label will be on top). Select all the S-labels and copy them from the spreadsheet.
3. Navigate to the wild isolate genotyping template google sheet (wild\_isolate\_genotyping\_template) using a web browser.
  1. Make a copy of this google sheet by right-clicking on the **Genotyping Template** tab then selecting the **Copy to New Spreadsheet** option. Select **Open Spreadsheet** to view the new google sheet.

2. Name this new sheet with the fulcrum project name followed by 'wild\_isolate\_genotyping', e.g., '2020FebruaryAustralia\_wild\_isolate\_genotyping.'

**NOTE:** This sheet is referred to as the 'genotyping sheet' throughout the rest of the protocol.

4. Paste the S-labels copied from the 'nematode\_isolation\_s\_labeled\_plates.csv' 's\_label' column into the genotyping sheet column titled 's\_label'. Check the 's\_label\_repeat\_error' column for '1's. A value of '1' in this column means the S-label is duplicated somewhere on the genotyping sheet. If duplications are discovered, investigate and correct them before moving forward.
5. Fill in the genotyping sheet 'isolation\_box\_number' column for all S-labels.

## 6. Check for proliferation on S-plates

1. Check for proliferating animals on S-plates 48 h after isolation (use the date and time of last isolation on the box from step 4.11 to guide timing).  
**NOTE:** Proliferating nematodes are characterized by offspring on the S-plate.
2. If an S-plate is proliferating, enter '1' in the proliferation\_48 column on the genotyping sheet, then move the S-plate to a box labeled '48 h proliferation, box 1'. Place a maximum of 88 S-plates in a proliferation box, then start filling a new box labeled '48 h proliferation, box 2'. Ensure the S-labels are organized in alphanumeric order in the 48 h proliferation boxes.  
**NOTE:** Do not dispose of the non-proliferating S-plates; these plates will be checked again at 168 h post-isolation. If desired, consolidate these S-plates in numeric order in boxes labeled '48 h non-proliferating, box X', but

remember to record when the 168 h check must occur on the new box.

3. After identifying all the proliferating S-labels at 48 h, move on to section 7 for S-plates with proliferation at 48 h.
4. Check the S-plates that were not proliferating at 48 h post-isolation again at 168 h post-isolation.
  1. If an S-plate is now proliferating, enter '1' in the proliferation\_168 column on the genotyping sheet and then move the S-plate to a box labeled '168 h proliferation, box 1'.
  2. Place a maximum of 88 S-plates in a proliferation box, then start filling a new box labeled '168 h proliferation, box 2'. Be sure to organize S-labels in alphanumeric order in the 168 h proliferation boxes.
5. Discard the S-plates that have no proliferation after 168 h. Move on to section 7 for S-plates with proliferation at 168 h.

## 7. Lysis of isofemale lines

**NOTE:** This step will use the data filter tool in google sheets to help print lysis worksheets for the S-plates in the proliferation boxes. The purpose of the lysis worksheets is to provide personnel with the correct positions for S-labels in lysis strip tubes at the bench.

1. Open the genotyping sheet for the desired project and select all cells by typing **Cmd+A**. Click on **Data > Create a Filter** to add a filter button to each column header. Use the **Filter** buttons to display only the S-plates that will be genotyped. For example, if all the S-plates with proliferation at 48 h are to be lysed: Click on the Filter button in the 'proliferation\_48' column and select '1'.

2. Once the genotyping google sheet has been filtered, review the list of S-labels displayed to ensure they are the S-labels to be printed on the worksheet.

1. In the 'strip\_tube\_number' column of the genotyping google sheet, enter a unique number every 11 rows.
2. Enter the strip tube numbers for a project in successive order starting at 1 and never duplicated. In the 'strip\_tube\_position', enter 2 through 12 for each strip tube number.

**NOTE:** Use 12-tube strip tubes for lysis. The first position (strip\_tube\_position 1) will be control, but the controls are not added to the lysis worksheets (only the strip\_tube\_positions are added, 2-12). At the time of lysis, the positive control strain 'N2' will be added to position 1 of every even-numbered strip tube as a positive control. No worms are added to position 1 of every odd-numbered strip tube as a negative control.

3. Filter the genotyping google sheet further to include just the S-labels in one proliferation box that are to be lysed, then select the columns 's\_label' through 'lysis\_notes'. Print a lysis worksheet for each proliferation box to be lysed.

4. Click on the drop-down menu in the **Print** field and select **Selected Cells**. Click on **Next** in the upper right, then use the dialogue to print the lysis worksheet for the proliferation box.

5. Repeat steps 7.3-7.5 to print a lysis worksheet for each proliferation box.

**NOTE:** Each proliferation box holds up to 88 S-plates, which corresponds with eight 12-well strip tubes.

6. Prepare 12-well strip tubes for all the samples that will be lysed. Label a strip tube with a unique

'strip\_tube\_number' assigned in the lysis worksheet. This label must be written on the cap strip and the strip tube to avoid confusion if they are separated. The EVEN strip tubes have a positive control (N2 worms) in position 1. The ODD strip tubes have a negative control (no worms) in position 1.

7. Make up enough lysis buffer (100 mM KCl, 20 mM Tris pH 8.2, 5 mM MgCl<sub>2</sub>, 0.9% IGEPAL, 0.9% Tween 20, 0.02% gelatin with proteinase K added to a final concentration of 0.4 mg/mL) for all of the samples and add 5% extra for pipette error. Scale as necessary.

**NOTE:** The lysis buffer is best prepared by combining all ingredients except for proteinase K and freezing in 10-50 mL aliquots at -20 °C. Thaw aliquots and keep at 4 °C prior to use; immediately before use, add proteinase K and mix thoroughly. Keep the lysis buffer on ice while working.

8. Arrange the S-plates for a particular strip tube in order using the printed lysis worksheet as a guide.

9. Uncap one strip tube and add 8 µL of lysis buffer to each cap with a repeat pipettor. Add the lysis buffer to one strip of caps at a time because the lysis buffer will evaporate if left at RT and uncovered. Pick 3-5 animals from the source plates (S-plate or N2 stock plate for positive controls) into the appropriate cap positions indicated on the lysis worksheet. Record notes for any S-plate with fewer than 5 worms picked to the lysis in the lysis\_notes section of the lysis worksheet.

10. After loading nematodes into each position of the strip tube, place the cap strip back on the strip tube. Match the marked cap (position 1) with the marked tube (position 1). Once capped, centrifuge the strip tube briefly until the nematodes are at the bottom of the tube.

11. Place the strip in the -80 °C freezer until completely frozen (at least 10 min). Repeat steps 7.9 to 7.11 until all strips have nematodes added for lysis. Organize the tube strips in numeric order.
12. Remove the sets of strip tubes and run the lysis program in a thermocycler: 1 h at 60 °C, 15 min at 95 °C, hold at 12 °C. When the lysis program is done, spin down the samples at 300 x g for 15 s at RT and store the lysates at -80 °C for up to 1 week.
13. Organize the tube strips in numeric order using 96-well plate holders and include a label with a proliferation box number, strip tube number range, date, and the initials of the researcher. Update the genotyping sheet columns 'lysis\_date' and 'lysis\_notes' with information from the lysis worksheet.

## 8. PCR of SSU and ITS2 sequences

**NOTE:** This section will provide instructions on how to perform two separate PCRs for each lysed S-plate. The first primer set amplifies a 500-bp fragment of the 18S rDNA small subunit gene (SSU); oECA1271 = forward primer TACAATGGAAGGCAGCAGGC, oECA1272 = reverse primer CCTCTGACTTTCGTTCTTGATTAA<sup>12</sup>. This PCR is used to check the quality of the template DNA. The PCR amplifies the SSU region for nearly all nematode species. If the SSU PCR fails to amplify, this result suggests that the lysis quality is poor and the lysis must be repeated for this S-plate. The second primer set amplifies a 2,000-bp fragment of the internal transcribed spacer region between the 5.8S and 28S rDNA genes (ITS2); oECA1687 = forward primer CTGCGTTACTTACCACGAATTGACARAC, oECA202 = reverse primer GCGGTATTTGCTACTACCAYYAMGATCTGC<sup>3</sup>. The ITS2

PCR product is Sanger sequenced and the sequence is used to identify nematodes in the *Caenorhabditis* genus to the species level by sequence similarity.

1. Use the filtering tool in the genotyping sheet to view only the S-labels to be used for PCR.
  1. Update the pcr\_plate\_number, and pcr\_well columns in the genotyping sheet. To prevent degradation of lysis material, the SSU and ITS2 PCRs are run at the same time.
  2. Use the same pcr\_plate\_number for the ITS2 and SSU PCRs even though these are separate reactions in separate plates. They will be distinguished with 'SSU' or 'ITS2' labels.
2. Assign a pcr\_plate\_number to eight or fewer strip tubes (one strip tube per row of the 96-well PCR plate, arranged in ascending order, e.g., lowest strip tube number on top). Then assign a pcr\_plate\_well to each S-label in the strip tubes.
 

**NOTE:** The strip tubes are arranged in ascending order, with the lowest strip tube number assigned to row A and the highest number in row H. Position 1 of all strip tubes is assigned to column 1. Therefore, strip tube number 1, position 1 will be assigned to PCR plate number 1, well A01.
3. Label 96-well PCR plate(s) to accommodate the samples that will be used for PCR. Label each PCR plate with the following information: project name, PCR type, PCR plate number, and date of PCR (e.g., 2020FebruaryAustralia\_SSU\_1\_20200304). Also, label the plate with the strip tube numbers that will be loaded into each row.

4. Remove the lysis material from the -80 °C freezer and thaw the strip tubes containing the lysis material on ice. While the lysis material is thawing, prepare ITS2 and SSU master mixes in separate tubes on ice. The SSU and ITS2 PCR recipes are found in **Supplemental Table 2**.  
**NOTE:** Prepare 100 reactions of PCR master mix for each 96-well plate to allow for pipetting error. Use a 15 mL or 50 mL conical to hold the master mix if large volumes are to be used.
5. Vortex the master mix gently until Taq is distributed throughout the mix. Once mixed, aliquot 38 µL of the master mix to the appropriate wells of the PCR plates on ice. Use sterile single-use v-bottom troughs and a 12-well multi-channel pipette to transfer the master mix to the PCR plates.
6. Spin down the thawed lysis strip tubes to remove lysis material from the caps. Carefully remove the lids of all the strip tubes that will be loaded into the first PCR plate. Use a low-volume multi-channel pipette (either 12-well or 8-well) to add 2 µL of lysate to the appropriate well in the PCR plate. Gently pipette the lysate up and down once before removing the 2 µL.  
**NOTE:** Check the tips to ensure they contain the lysis before the transfer. Remember to change tips between rows or columns.
7. Cover the PCR plate with PCR adhesive foil and use a roller to create a tight seal. After the foil is applied, briefly spin down the PCR plates in a centrifuge. Keep the plate on ice until it is ready to run in the thermocycler.
8. Run the PCRs with the appropriate thermocycler program. Refer to **Supplemental Table 2** for the details of the SSU and ITS2 PCR programs.
9. Repeat steps 8.4- 8.8 until all the PCRs are run.
10. While the PCR reactions are running, pour a 100 mL 1.5% agarose gel. Each gel will hold samples or a single PCR plate.
  1. Add 1.5 g of agarose to a 500 mL flask, then add 100 mL of 1x TAE buffer (**Supplemental Table 3**) and swirl to mix. Microwave to dissolve and cool the gel.
  2. Once the solution is cooled, add 5 µL of 10 mg/mL Ethidium bromide solution and mix to combine. Pour the solution into a casting tray with four 25-well combs so that the gel can accommodate 96 samples plus a ladder for each row in the gel.  
**NOTE:** Ethidium bromide is a potent mutagen. When handling ethidium bromide, use a lab coat, chemical resistant gloves, and chemical safety goggles.
11. Just before the PCR is finished, add 6x loading dye to a disposable trough and use a multi-channel pipette to add 2 µL of 6x loading dye to each well of a new 96-well PCR plate. This plate will be used to load the samples into the gel. Make enough of these plates to accommodate all the samples.
12. When the PCRs are done, remove the PCR plates and briefly centrifuge them at 300 x g for 15 s at RT. Store the PCR plates on ice until the PCR products can be run out on a gel.
  1. To run the products out on a gel, use a 12-well multi-channel pipette to add 5 µL of each sample to the appropriate well of a 96-well plate containing 2 µL of 6x loading dye.
  2. Then load 6 µL of this mixture into each well of a recently cast gel. Load 6 µL of 1 KB plus ladder into the first well of each row of the gel.

**NOTE:** To fill the wells of the gel, it may be necessary to intersperse Row A and Row B from the PCR plate in the first row of the gel. To avoid confusion, record the gel\_number and gel\_position in the genotyping sheet for each PCR sample.

13. Place a new foil lid on the remaining PCR in the plate(s) and store them at 4 °C. These reaction products will be used for sequencing in step 9.
14. Run the PCR products out on the gel at 120 V for 20 min. Image the gel and record which S-labels yield ITS2 and/or SSU PCR products in the 'pcr\_product\_its2' and 'pcr\_product\_ssu' columns of the genotyping sheet. Mark the presence of a band with a '1'; mark a '0' for no band.

## 9. Identifying nematodes with Sanger sequencing and sequence BLAST

**NOTE:** This section provides instructions for sequencing the ITS2 amplicons from the S-labels, aligning those sequences to the National Center for Biotechnology Information (NCBI) database using the BLAST algorithm, and parsing the BLAST results to identify the nematodes on the S-plates.

1. For each sample that is ITS2-positive, use the remaining ITS2 PCR product for Sanger sequencing using the forward primer oECA306 (CACTTTCAAGCAACCCGAC). Arrange for the sequencing output files to be easily linked to an S-label by recording the 'sequencing\_plate' and 'sequencing\_well' columns of each S-label in the genotyping sheet.
2. Obtain the .seq output files for each S-label from the sequencing platform. Arrange the .seq files for a project in a single directory with .seq files for each batch of sequencing located in subdirectories.

3. Open the command-line interface tool and navigate to the top directory containing the .seq files by entering the command: `cd <path_to_directory>`. If it does not already exist, create a merged FASTA for all the .seq files by entering the following command: `for dir in */; do cd $dir; for file in *.seq; do echo ">"$file; cat $file; done >>../all_seqs.fa; cd ..; done.`

**NOTE:** This code will create a merged FASTA file named 'all\_seqs.fa' from all the .seq files in the project directory. This file can be used in the NCBI online nucleotide BLAST tool to rapidly align the ITS2 sequence of each S-label to NCBI's sequence database.

4. In a web browser, navigate to the NCBI BLAST website<sup>19</sup> and click on the **Choose File** button. Select the all\_seqs.fa file that was just created, then click on the button **Somewhat Similar Sequences (BLASTn)**. Click on the **BLAST** button at the bottom of the page to begin the BLAST search.
5. Update the genotyping sheet with the BLAST results for each S-label. Use the filter tool to make updating the genotyping google sheet easier. Click on **Data > Create a Filter** to add a filter button to each column header. Filter the sequencing\_plate column to select the sequencing plates that are to be updated with BLAST results.
6. Use the drop-down menu on the NCBI BLAST results page to check the results for each S-plate ITS2 sequence (**Figure 6**).
7. Check for no BLAST hits. A sequence ID in the drop-down prefixed with \* has no blast hits. For these S-labels, enter 'no hit <current date>' in the manual\_blast\_notes column of the genotyping sheet.
8. Check for a possible new *Caenorhabditis* species. Click the link on the top hit to visualize the alignment (**Figure**

- 6). If the top hit is (1) a *Caenorhabditis* species, (2) the alignment contains more than five mismatches in the center of the sequence, and (3) the query coverage is greater than 50%, this result suggests the isolate may be a new *Caenorhabditis* species (**Figure 7**). For these S-plates enter, the species of the top BLAST hit in the 'species\_id' column, enter a 1 in the 'possible\_new\_caeno\_sp' column, and 'possible new Caeno sp.' into the 'manual\_blast\_notes' column along with percent identity, (e.g., 'possible new Caeno sp. 89% identity').
9. For S-plate sequences that BLAST to a *Caenorhabditis* species, enter the full genus and species name of the top BLAST hit in the 'species\_id' column. For example, '*Caenorhabditis elegans*'.
10. For S-plate sequences that BLAST to a non-*Caenorhabditis* species, enter only the genus of the top BLAST hit followed by 'sp.' in the 'species\_id' column. This notation means the isolate is an unknown species within the named genus. For example, '*Oscieus* sp.'.
- NOTE:** The ITS2 sequence can not be used to reliably identify isolates to the species -level outside of the *Caenorhabditis* genus<sup>3,13</sup>.
11. Enter 1 in the 'make\_strain\_name' column of the genotyping sheet if 'species\_id' = '*Caenorhabditis elegans*', '*Caenorhabditis briggsae*', or '*Caenorhabditis tropicalis*', OR 'possible\_new\_caeno\_sp' = 1.
12. Name the strains with unique names following *Caenorhabditis* nomenclature conventions, i.e., a unique laboratory designation consisting of 2-3 uppercase letters followed by a number for each unique strain<sup>22</sup>. Enter the strain names in the 'strain\_name' column.

13. After strains are named, they can be cryopreserved using established protocols<sup>23</sup>.

## 10. Processing the collection data with the easyFulcrum package in R

**NOTE:** This step describes how to link the collection data (C-labels) and the nematode isolation data (S-labels) together using the easyFulcrum R package. The software contains functions that will further join the Fulcrum data with the genotyping data from the genotyping sheet so that S-label species identities and strain names are organized in a single data frame.

1. Create a new directory named for the collection project. Arrange the folder structure within the directory to match the requirements described in the R package easyFulcrum<sup>15</sup>.
2. Navigate to the Fulcrum website and sign in. Export the raw project data from the Fulcrum database using the Fulcrum website's data export tool on the left and selecting the following checkboxes: project, include photos, include GPS data, field sampling, and isolation.
- NOTE:** The Fulcrum data for the project will be exported as five comma-separated value (.csv) files. The complete project data will be joined together into a single data frame using the easyFulcrum package in R.
3. Move the five .csv files exported from Fulcrum into the project directory created in step 10.1 as instructed in the easyFulcrum vignette<sup>20</sup>.
4. Open an Rstudio session and install easyfulcrum package in R by entering the following commands in the R console 'install.packages("devtools")' and 'devtools::install\_github("AndersenLab/easyfulcrum")'.

5. Open a new R script and follow the directions in the easyfulcrum vignette to process the collection data<sup>20</sup>.

## Representative Results

This protocol has been used to collect *Caenorhabditis* nematodes from multiple locations, including Hawaii and California. The isolation success rate for *Caenorhabditis* nematodes varies with collection location, climate, sampling experience, and substrate types sampled. The protocol has been used to extensively sample the Hawaiian Islands, where nine collection projects have been conducted over multiple years and seasons. The isolation success rates for selfing *Caenorhabditis* species are nearly identical for *C. briggsae* (162 of 4,506 samples, 3.6%) and *C. elegans* (163 of 4,506 samples, 3.6%), and much lower for *C. tropicalis* (26 of 4,506 samples, 0.58%)<sup>8</sup>. Each of the selfing species is enriched on rotting fruit and flower substrates relative to the other substrate categories. Sample rotting fruit and flower substrates if the researcher is attempting to maximize the success rate rather than characterize substrate preferences. However, the success rate varies with the quality of the substrate selected. For example, among fruit and flower substrates, those substrates that are too dry, wet, or fresh will likely not yield *Caenorhabditis* nematodes.

The scalability of this collection protocol is evident from the number of collections a single pair of researchers can collect from the wild. For example, in October of 2018, a pair of researchers using this collection protocol was able to collect a total of over 1,000 samples in 7 days from multiple locations on two Hawaiian Islands. This field team shipped the samples overnight to the laboratory, where a team of eight researchers isolated over 2,000 nematodes from the samples as they arrived. A key advantage of this protocol is that it minimizes the cost associated with sampling in

remote locations by reducing the equipment and personnel required in the field. Using this protocol, a small field team can focus on sampling while the isolation team can process the samples at their home institution using fragile and heavy equipment like dissecting microscopes and agar plates for isolating nematodes. Moreover, the implementation of the mobile data-collection application allows all the field data associated with the samples to be linked directly to the C-label, which enables the isolation team to work independently from the field team while processing samples.

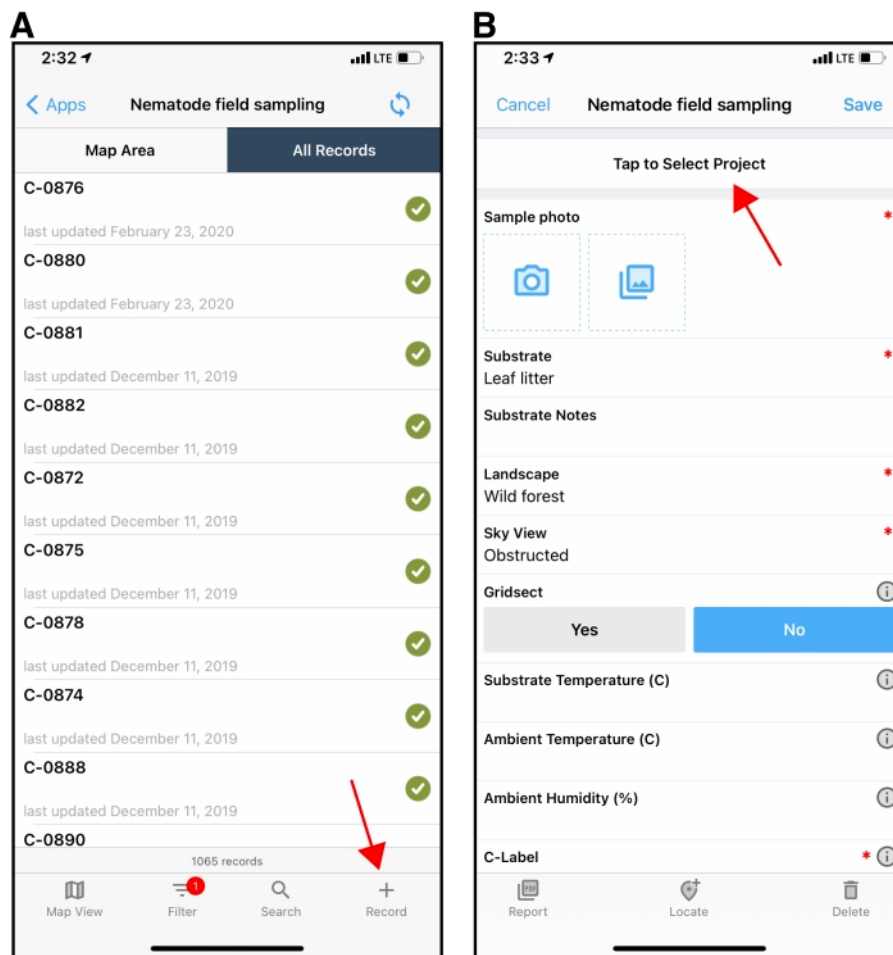
Researchers that use this collection protocol must consider the effort that is required to isolate nematodes prior to a collection project. The isolation and identification steps are rate-limiting, and a small collection team can quickly overwhelm isolators with samples. Moreover, the laboratory space required to process many collections can interfere with ongoing research (**Figure 3**). Additionally, some isolated nematodes require additional effort to genotype. For example, approximately 2% of isolates fail to amplify with the SSU PCR primer set after the first lysis attempt and must be re-lysed to ensure that the lysis material is suitable for amplification with the ITS2 primer set (**Figure 8**). Furthermore, approximately 3% of isolates fail to produce quality sequences after an initial round of Sanger sequencing. For these isolates, another round of lysis, ITS2 PCR, and Sanger sequencing is often required, which can increase handling time for the isolation team. Importantly, sequence identity alone is not sufficient evidence to justify a new *Caenorhabditis* species (**Figure 7**). To properly justify raising an isolate as a new *Caenorhabditis* species, additional effort must be made to perform mating experiments and establish a typed specimen<sup>13</sup>. A formal morphological description of the typed specimen is also preferred but not required<sup>3</sup>. Together, these considerations suggest that researchers adopting this collection protocol will

benefit from trial tests of the isolation and identification steps to ensure resources are properly allocated before a collection project begins. Importantly, even small collection projects

can benefit from this protocol because the process is highly reproducible, and the data can easily be audited for quality control purposes across laboratory groups.



**Figure 1: Substrate examples.** (A) An ideal rotting fruit is shown in the center of the image (1), the fruit is almost unrecognizable. Less rotted fruit is shown nearby; avoid sampling freshly fallen fruits (2). (B) An ideally decomposed flower is shown at the top (3). Avoid sampling freshly fallen flowers (4). (C) The dark leaf litter under the top layer of dry leaves is ideal when sampling for selfing *Caenorhabditis* nematodes (5). Avoid sampling dry leaf litter (6). [Please click here to view a larger version of this figure.](#)



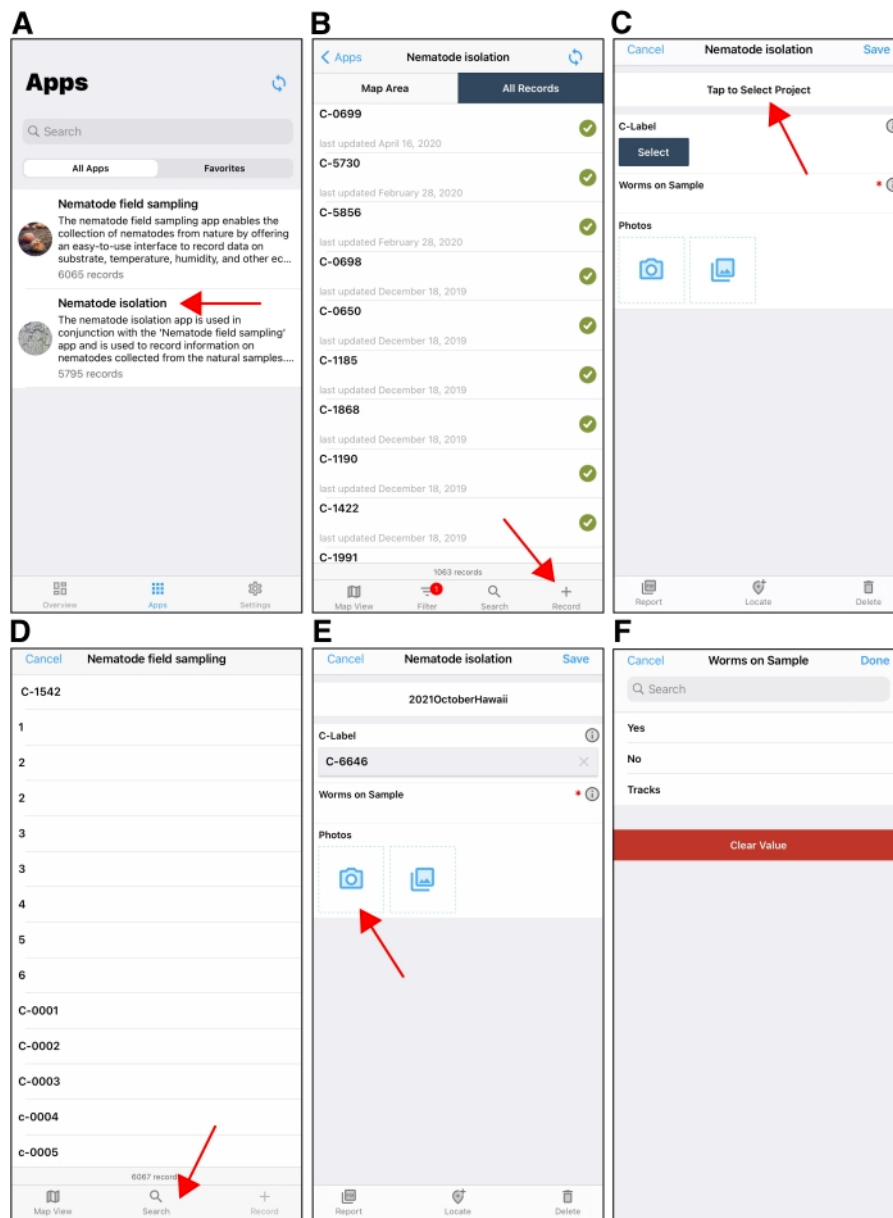
**Figure 2: The Nematode Field Sampling mobile application. (A)** The initial screen after opening the **Nematode Field Sampling** application on an Apple device in Fulcrum. The red arrow in the lower right points to the + button used to create a new collection record. **(B)** An example of a new collection record shown on an Apple device. The red arrow points to the 'Project' field at the top of the collection record screen. Be sure to select the correct project when sampling in the field. The project field will default to the last project used when creating subsequent collection records. [Please click here to view a larger version of this figure.](#)



**Figure 3: Collection bags and collection plates organized prior to plating out samples.** This figure shows the samples in C-labeled collection bags on the left. Each collection bag has a matching C-labeled 10 cm plate on top of it. On the right are 10 cm collection plates that contain sample material after it was transferred from the collection bags. [Please click here to view a larger version of this figure.](#)



**Figure 4: A collection plate (C-plate) with properly a transferred sample.** A 10 cm C-plate with decomposing fruit placed on the edge of the bacterial lawn. The C-label is attached to the plate lid. [Please click here to view a larger version of this figure.](#)



**Figure 5: The Nematode isolation mobile application.** (A) The application selection screen in the Fulcrum mobile application. The red arrow points to the **Nematode Isolation** application. (B) The initial screen after opening the **Nematode Isolation** application on an Apple device in Fulcrum. The red arrow in the lower right points to the **+** button used to create a new isolation record. (C) An example of a new isolation record shown on an Apple device. The red arrow points to the 'Project' field at the top of the isolation record screen. Be sure to select the correct project when isolating. The project field will default to the last project used when creating subsequent isolation records. (D) After tapping the **Select** field under C-label, users will tap the search button (red arrow) to find the C-label from which they are isolating nematodes. (E) After the C-label is selected, users will photograph the C-plate using the device camera. (F) Users then input whether there are

nematodes on the C-plate or not. S-labels are added to the isolation record if there are nematodes to be isolated. [Please click here to view a larger version of this figure.](#)

Job Title: S-05002\_oECA306\_A03.seq  
 RID: DFKMH75701R Search expires on 06-05-00-23 am Download All  
 Results for: 37:iclQuery\_44458 S-05554\_oECA306\_D03.seq(1024bp)  
 Program: BLASTN Citation  
 Database: nt See details  
 Query ID: iclQuery\_44458  
 Description: S-05554\_oECA306\_D03.seq  
 Molecule type: dna  
 Query Length: 1024  
 Other reports: Distance tree of results

Filter Results  
 Organism: only top 20 will appear exclude  
 Type common name, binomial, taxid or group name  
 + Add organism  
 Percent Identity: to E value: to Query Coverage: to  
 Filter Reset

Descriptions Graphic Summary Alignments Taxonomy

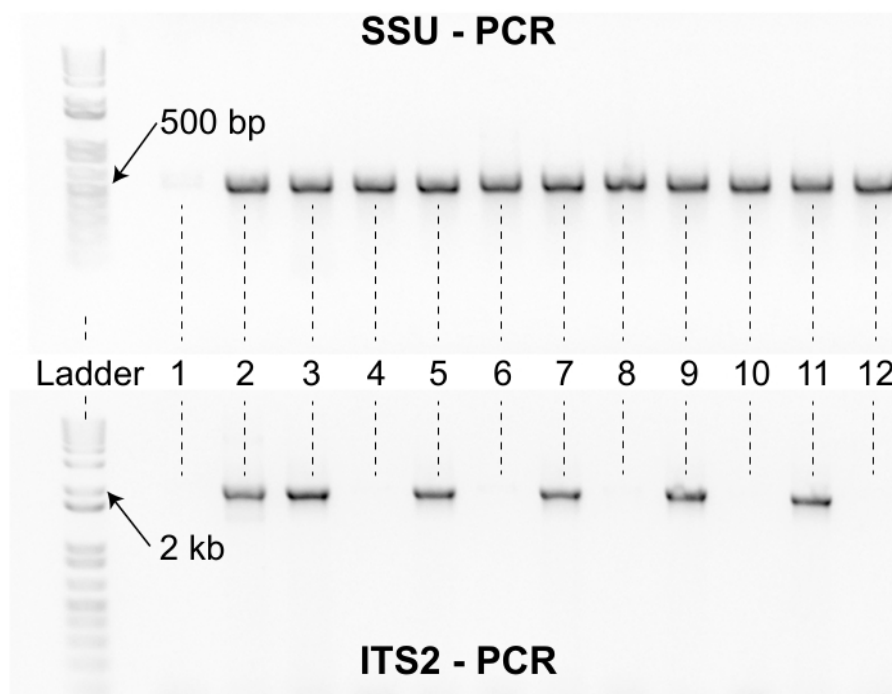
Sequences producing significant alignments Download Manage Columns Show 100

select all 100 sequences selected GenBank Graphics Distance tree of results

| Description                                                                                                                              | Max Score | Total Score | Query Cover | E value | Per. Ident | Accession  |
|------------------------------------------------------------------------------------------------------------------------------------------|-----------|-------------|-------------|---------|------------|------------|
| Caenorhabditis briggsae 18S ribosomal RNA gene, internal transcribed spacer 1, 5.8S ribosomal RNA gene, internal transcribed spacer 2    | 1288      | 1288        | 68%         | 0.0     | 99.43%     | JN636061.1 |
| Caenorhabditis briggsae strain AF16 18S ribosomal RNA gene, internal transcribed spacer 1, 5.8S ribosomal RNA gene, internal transcribed | 1284      | 1284        | 68%         | 0.0     | 99.43%     | MN519141.1 |

**Figure 6: NCBI BLAST results page.** (1) The drop-down menu used to view the BLAST results for all sequences. (2) The description of the current sequence selected from the drop-down. In this case the results for S-label S-05554 are shown. (3) The top BLAST hit for S-05554 is shown. The purple text indicates the link to visualize this alignment has been clicked. Please be sure to inspect the alignments by eye to identify possible new *Caenorhabditis* species, see step 9.8 above. [Please click here to view a larger version of this figure.](#)

Copyright © 2021 JoVE Journal of Visualized Experiments [jove.com](https://www.jove.com) December 2021 • e63486 • Page 20 of 24



**Figure 8: SSU and ITS2 PCR products.** The top gel shows PCR products generated with the SSU primer set for 12 representative samples. A DNA ladder is included on the left as a reference. The SSU PCR products for *Caenorhabditis* nematodes are approximately 500 bp in length. Samples 2-12 amplified with the SSU primer set but sample one did not. The absence of a 500 bp SSU amplicon for sample one suggests that the lysis material was of poor quality and the sample must be re-lysed. The bottom gel shows PCR products generated with the ITS2 primer set for the same 12 Samples shown in the top gel. The ladder and samples are in the same orientation for both gels. Six of the 12 samples did not amplify with the ITS2 primer set. The samples with SSU and ITS2 bands are Sanger sequenced and identified by sequence similarity using the NCBI BLAST algorithm. [Please click here to view a larger version of this figure.](#)

**Supplemental File 1: C-labels.** A PDF file containing 2500 unique C-labels. [Please click here to download this File.](#)

**Supplemental File 2: S-labels.** A PDF file containing 5000 unique S-labels. [Please click here to download this File.](#)

**Supplemental Table 1: Field Materials.** A packing list of materials used in the field to sample nematodes. [Please click here to download this Table.](#)

**Supplemental Table 2: PCR recipes and thermocycler conditions.** A table of PCR recipes and thermocycler conditions for the ITS2 and SSU PCRs. [Please click here to download this Table.](#)

**Supplemental Table 3: Electrophoresis buffer recipes.** A recipe for 0.5 M pH 8.0 Ethylenediaminetetraacetic acid solution (EDTA) and the TRIS-acetate-EDTA (TAE) buffer solution. [Please click here to download this Table.](#)

## Discussion

This protocol contains critical steps that must be executed with caution. For example, it is important that the field and isolation teams are careful to select the correct collection project in the application prior to collecting samples from the field or isolating nematodes from the samples in the laboratory. In the event that the wrong collection project is selected, the errant data records are best corrected in the Fulcrum database using the record editing tools online. This process can be tedious for many misplaced records. However, the database retains any changes to records so that a complete auditing of collection and isolation records is possible. The other critical steps in this protocol involve the handling of samples from the field and the nematodes isolated from those samples. To ensure *Caenorhabditis* nematodes survive the sampling and shipment steps, the temperature of samples should be maintained between 4 °C and 25 °C. Temperatures above 25 °C can cause sterility in *C. elegans*<sup>14</sup>. Ensure samples are transferred from collection bags to collection plates within five days whenever possible to minimize the loss to nematodes. After nematodes are isolated, it is critical that they are genotyped and cryopreserved before they perish. It is difficult to find living nematodes on S-plates that are more than two to three weeks old because fungal and bacterial contamination can make the S-plates inhospitable.

This protocol can be modified easily to accommodate different types of data that researchers may want to collect while in the field. For example, it is easy to customize the 'Nematode field sampling' application with new data entry fields using Fulcrum's online GUI for application editing. Moreover, the data analysis package, *easyFulcrum*, can accommodate these edits when processing the new data<sup>15</sup>. Another modification users may find appealing is to use a different

sampling method in the field. Rather than sampling discrete substrates, researchers may desire to sample larger areas containing multiple substrate types. These larger samples are best processed in the laboratory using the Baermann funnel or tray extraction methods<sup>13</sup>. Importantly, the use of C-labels and S-labels still apply for these techniques and are therefore compatible with mobile applications.

The primary limitations of this protocol relate to the handling time of nematodes prior to isolation in the laboratory. First, the lag time between sample collection and nematode isolation makes it impossible to record the developmental stages of nematodes on a given sample at the time of collection. Second, the frequency of males and outcrossing in nature are key evolutionary questions for selfing *Caenorhabditis* nematodes<sup>10</sup>. This method is not well suited to addressing these questions because nematodes are likely to have gone through multiple generations prior to isolation. Delayed isolation means that direct evidence of male frequency in nature is impossible. Furthermore, the multi-generational delay during the genotyping steps means that genomic evidence of outcrossing (heterozygosity) will be eroded before a nematode strain can be sequenced. To identify heterozygosity in nature, the offspring produced by a nematode directly isolated from nature are used for sequencing<sup>2</sup>. Another potential limitation of this protocol is that it is biased toward the identification of selfing *Caenorhabditis*. This is because isolated nematodes of selfing species have a greater chance of proliferating than obligate outcrossers, which will only proliferate if a fertilized female is isolated.

This collection method is based on existing collection protocols<sup>13,14</sup>. The major advancement of this technique is the use of mobile technology and customized software

to facilitate the organization of vast quantities of ecological and molecular data associated with large-scale collection projects. The ecological data generated using this collection protocol can be used to address outstanding questions for natural populations of *Caenorhabditis* species. For example, data generated with this method have been used to discover niche preferences for the species across the Hawaiian Islands. Moreover, by sequencing the genomes of cryopreserved nematodes, researchers can investigate how patterns of genetic variation are correlated with ecological data. Research of this kind can uncover signatures of local adaptation in *Caenorhabditis* populations and provide important insights into the relevance of genetic variation in natural contexts<sup>8</sup>. To gain a functional understanding of many genes in *Caenorhabditis* nematodes, ecological studies are likely required<sup>11</sup>. Even for *C. elegans* a large fraction of genes lacks functional annotations, despite it being the first multicellular animal to be sequenced and one of the most thoroughly studied animals on Earth. This collection protocol was developed to help address this knowledge gap by facilitating the collection of wild *Caenorhabditis* nematodes and the study of their ecology and natural genetic diversity.

## Disclosures

The authors report no conflicts of interest.

## Acknowledgments

This research was supported by start-up funds from Northwestern University and a National Science Foundation CAREER Award (IOS-1751035), both granted to E.C.A.

## References

1. Frézal, L., Félix, M. -A. *C. elegans* outside the Petri dish. *eLife*. **4**, eLife.05849 (2015).
2. Sivasundar, A., Hey, J. Sampling from natural populations with RNAi reveals high outcrossing and population structure in *Caenorhabditis elegans*. *Current Biology: CB*. **15** (17), 1598-1602 (2005).
3. Kiontke, K. C. et al. A phylogeny and molecular barcodes for *Caenorhabditis*, with numerous new species from rotting fruits. *BMC Evolutionary Biology*. **11**, 339 (2011).
4. Petersen, C. et al. Ten years of life in compost: temporal and spatial variation of North German *Caenorhabditis elegans* populations. *Ecology and Evolution*. **5** (16), 3250-3263 (2015).
5. Richaud, A., Zhang, G., Lee, D., Lee, J., Félix, M. - A. The local coexistence pattern of selfing genotypes in *Caenorhabditis elegans* natural metapopulations. *Genetics*. **208** (2), 807-821 (2018).
6. Ferrari, C., Salle, R., Callemeyn-Torre, N., Jovelín, R., Cutter, A. D., Braendle, C. Ephemeral-habitat colonization and neotropical species richness of *Caenorhabditis* nematodes. *BMC Ecology*. **17** (1), 43 (2017).
7. Crombie, T. A. et al. Deep sampling of Hawaiian *Caenorhabditis elegans* reveals high genetic diversity and admixture with global populations. *eLife*. **8**, e50465 (2019).
8. Crombie, T. A. et al. Local adaptation and spatiotemporal patterns of genetic diversity revealed by repeated sampling of *Caenorhabditis elegans* across the Hawaiian Islands. *bioRxiv*. 2021.10.11.463952 (2021).

9. Noble, L. M. et al. Selfing is the safest sex for *Caenorhabditis tropicalis*. *eLife*. **10**, eLife.62587 (2021).
10. Cutter, A. D., Morran, L. T., Phillips, P. C. Males, Outcrossing, and Sexual Selection in *Caenorhabditis Nematodes*. *Genetics*. **213** (1), 27-57 (2019).
11. Petersen, C., Dirksen, P., Schulenburg, H. Why we need more ecology for genetic models such as *C. elegans*. *Trends in Genetics: TIG*. **31** (3), 120-127 (2015).
12. Haber, M., Schüngel, M., Putz, A., Müller, S., Hasert, B., Schulenburg, H. Evolutionary history of *Caenorhabditis elegans* inferred from microsatellites: evidence for spatial and temporal genetic differentiation and the occurrence of outbreeding. *Molecular Biology and Evolution*. **22** (1), 160-173 (2005).
13. Barrière, A., Félix, M. -A. *WormBook: Isolation of C. elegans and Related Nematodes*. at <<https://www.ncbi.nlm.nih.gov/books/NBK19764/>> (2018).
14. Pouillet, N., Braendle, C. Sampling and isolation of *C. elegans* from the natural habitat. *Methods in Molecular Biology*. **1327**, 221-229 (2015).
15. Di Bernardo, M., Crombie, T. A., Cook, D. E., Andersen, E. C. easyFulcrum: An R package to process and analyze ecological sampling data generated using the Fulcrum mobile application. *PloS one*. **16** (10), e0254293 (2021).
16. Mobile data collection & workflow automation in NCAP at <<https://www.fulcrumapp.com/>> (2021).
17. Nematode Isolation Application at <<https://www.fulcrumapp.com/apps/nematode-isolation>> (2021).
18. Nematode Field Sampling Application at <<https://www.fulcrumapp.com/apps/nematode-field-sampling>> (2021).
19. Nucleotide BLAST: Search nucleotide databases using a nucleotide query at <[https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE\\_TYPE=BlastSearch](https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE_TYPE=BlastSearch)> (2021).
20. easyFulcrum at <<http://andersenlab.org/easyfulcrum/articles/easyFulcrum.html>> (2021).
21. Andersen, E. C., Bloom, J. S., Gerke, J. P., Kruglyak, L. A variant in the neuropeptide receptor npr-1 is a major determinant of *Caenorhabditis elegans* growth and physiology. *PLoS Genetics*. **10** (2), e1004156 (2014).
22. Tuli, M. A., Daul, A., Schedl, T. *WormBook: Caenorhabditis Nomenclature*. at <<https://www.ncbi.nlm.nih.gov/books/NBK535515/>> (2018).
23. Stiernagle, T. *WormBook: Maintenance of C. elegans*. at <<https://www.ncbi.nlm.nih.gov/books/NBK19649/>> (2006).