# Soft Matter



PAPER View Article Online
View Journal | View Issue



**Cite this**: *Soft Matter*, 2021, **17**, 4738

Received 2nd January 2021, Accepted 5th April 2021

DOI: 10.1039/d1sm00003a

rsc.li/soft-matter-journal

# Mechanochemical induction of wrinkling morphogenesis on elastic shells

Andrei Zakharov \*\* and Kinjal Dasbiswas \*\* \*\*

Morphogenetic dynamics of tissue sheets require coordinated cell shape changes regulated by global patterning of mechanical forces. Inspired by such biological phenomena, we propose a minimal mechanochemical model based on the notion that cell shape changes are induced by diffusible biomolecules that influence tissue contractility in a concentration-dependent manner – and whose concentration is in turn affected by the macroscopic tissue shape. We perform computational simulations of thin shell elastic dynamics to reveal propagating chemical and three-dimensional deformation patterns arising due to a sequence of buckling instabilities. Depending on the concentration threshold that actuates cell shape change, we find qualitatively different patterns. The mechanochemically coupled patterning dynamics are distinct from those driven by purely mechanical or purely chemical factors, and emerge even without diffusion. Using numerical simulations and theoretical arguments, we analyze the elastic instabilities that result from our model and provide simple scaling laws to identify wrinkling morphologies.

#### 1 Introduction

Morphogenetic events during embryo development involve tissue shape changes that are driven by mechanical forces.<sup>1,2</sup> A prototypical example is the folding of sheets of epithelial cells through the constriction and shape change of individual cells by actively contractile forces generated by myosin molecular motors in the cytoskeletal network on the cell surface.<sup>3,4</sup> The myosin motor activity is in turn triggered by complex chemical signaling cascades secreted by the cells themselves which create a spatiotemporal pattern of active mechanical forces in the tissue sheets.<sup>5</sup> The chemical signaling itself can be affected by mechanical cues such as forces<sup>6</sup> and deformations.<sup>7,8</sup> Inspired by the natural spatiotemporal control of shape change in biological tissue sheets, we are motivated to ask how such bio-inspired feedback can be used to realize self-actuated patterning of soft materials.

Thin elastic plates and shells constitute a fundamental class of soft matter that exhibit sensitive response to stimulii changes because of the geometric nonlinearity of their mechanical properties. Tissue morphogenesis can inspire the design of such slender structures that change shape in a programmable way. Desired shapes can be attained by pre-patterning the internal structure, for example *via* metric change, 10-12 controlled intrinsic curvature, 13,14 pre-stress, 15 or be self-organized, for example, through a propagating chemical reaction coupled with mechanical

Department of Physics, University of California, Merced, Merced, CA 95343, USA. E-mail: azakharov@ucmerced.edu, kdasbiswas@ucmerced.edu

deformations.<sup>16–21</sup> The latter approach provides reconfigurable conformations in contrast with frozen-in patterns, and resembles the shape changes during morphogenesis with anisotropic deformations including different types of mechanical instabilities, such as creases, wrinkles, folds, and ridges.<sup>22</sup>

Instabilities in spherical shells are of special interest because of their diverse applications and non-trivial behavior from a mathematical perspective. Examples of thin shells that exhibit wrinkling morphology abound in nature and range from pollen grains and viral capsids to organoids and organs such as brain and developing embryo at the "blastula" stage<sup>2</sup> (monolayer of cells arranged in a shell surrounding a fluid-filled lumen). Several studies have examined buckling behaviour of spherical shells due to external pressure, <sup>23,24</sup> confinement, <sup>25</sup> differential growth, <sup>26,27</sup> and demonstrated that patterns arising on curved surfaces are quite different from those on planar surfaces. Curvature-controlled structure formation encompasses a broad class of phenomena in physical, <sup>28,29</sup> biological <sup>30</sup> and chemical systems. <sup>31</sup>

In comparison to synthetic materials, living matter can show more complex cycles of feedback and control where chemical and mechanical signalling are tightly coupled. Inspired by this inherently mechanochemical basis of pattern formation in biological tissue, <sup>32</sup> we consider the dynamics and steady state of shapes induced by chemical gradients in thin elastic shells, where the chemical is itself affected by the sheet curvature. In analogy with tissue patterning by gradients of morphogens, diffusible biomolecules that induce cell fate changes slowly in a concentration-dependent manner during embryogenesis, <sup>33</sup>

**Paper** Soft Matter

we have posited "mechanogens" as biochemical agents that affect the cell mechanical state.34,35 They can do so by enhancing or relaxing the cell cytoskeletal contractility, which is a more physical change in comparison to the usual genetic changes wrought by morphogen gradients. Candidate mechanogens could be chemical factors, such as Ca<sup>2+</sup> ions, proteins, ATP or drugs, that regulate actomyosin contractility and cytoskeletal remodeling. While disentangling the various chemical signals and their interactions with mechanics is challenging in vivo, recent experiments in vitro with reconstituted cytoskeletal gels that exhibit molecular motor activitydriven buckling and wrinkling36-38 realize such active elastic processes in a controlled setting. These biological materials, along with synthetic materials such as liquid crystal elastomers<sup>39</sup> and gels,<sup>40</sup> in principle, allow for the spatial control of mechanical deformation and shape actuation through external chemical, electrical or optical stimuli.41,88

In this work, we explore how in-plane stresses induced in a thin elastic sheet by gradients of chemical signals ("mechanogens") can be relaxed by energetically less costly out-of-plane deformations. We thus seek to demonstrate how a short range chemical activation leads to long range elastic response and subsequent pattern formation. Unlike models for tissue folding that are based on differential apical-basal constrictions that lead to wedge-shape of the constituent cells, 42,43 our model is based on a spontaneous curvature arising due to in-plane incompatibility between domains of different tension - an effect that is similar to differential growth and also occurs in principle in contractile biological tissue. We show that depending on key parameters, such as a threshold of activation of cell mechanical response by the chemical gradient and the thickness of the elastic sheet, we can access qualitatively different patterns such as ridges and spots. Further, the mechanical feedback on the chemical gradient results in pattern propagation from an initial local region of activation. While inspired by tissue morphogenesis, these results may also be applicable to synthetic gels provided such a feedback can be set up.

The structure of the paper is organized as follows. We first define governing equations for a model involving mechanochemical interactions and give an estimation of the time scales that allow simplifying assumptions. In Section 3, we demonstrate patterns arising due to feedback between chemical production and elastic instabilities, and explore the parameter space identifying qualitatively different stationary shapes. In Section 4, we discuss the buckling and wrinkling instabilities, and provide simple scaling laws for the observed patterning. In the Appendix, we describe the discretized elastic energy, the numerical methods applied to perform simulations of model dynamics, and provide supplementary simulation results obtained for parameter values different from those in main text.

## 2 Mechanochemical model for tissue shape change

Inspired by the biochemical patterning of mechanical forces in thin layers of tissue, we introduce a model of chemical-induced pattern formation in thin elastic sheets. We consider the

situation of a thin monolayer of cells tightly adhered to each other and surrounded by extracellular fluid, as shown in the schematic in Fig. 1a. The cells change their shape in response to diffusible chemical signals (which we term mechanogens) in the extracellular fluid, that are secreted by the cells themselves, and that bind to receptors on the cells apical (top) surfaces and trigger changes in the contractile tension of their actomyosin cytoskeleton. For concreteness, we consider mechanogens that relax contractility and increase cell apicobasal surface area while reducing their lateral surface area to conserve volume. The resulting spatially inhomogeneous in-plane expansion of the tissue sheet causes the region exposed to higher chemical signal to buckle out of plane. We then explore the role of mechanochemical feedback (Fig. 1b), that is, the chemical concentration is in turn affected by the tissue mechanical state, specifically its curvature.

The cell monolayer, that could be freely suspended or adhered to a thin basal substrate, is modeled as a continuum elastic shell with uniform properties throughout its thickness. We assume that over the short timescales of interest, the mechanical response of the sheet to induced stresses is elastic and ignore the viscous dissipation and fluid flows that could result from cell rearrangements and motility. Then the mid-surface of the thin shell can be represented as a two-dimensional surface embedded in 3D space that allows a convenient description for large, out-of-plane deformations and calculation of the deformed 3D shape. The mechanical free energy of deformed elastic shells is determined

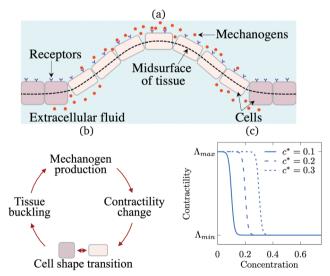


Fig. 1 (a) Schematic of buckling of a tissue sheet in response to chemical signals (mechanogens) that bind to receptors on the apical (upper) surface of cells and trigger change in cell contractility, and therefore cell shape (in this case, the aspect ratios as a result of force balance<sup>42</sup>). Outward tissue curvature in turn stimulates the release of mechanogens into the extracellular fluid, where they can diffuse and influence cell contractility. In simulations, the tissue layer is represented by the mid-surface (black dashed line) that reproduces macroscopic deformations. (b) Schematic of the mechanochemical feedback considered in our model. (c) Thresholded dependence of cell contractility on mechanogen concentration,  $\Lambda(c)$ , assumed in eqn (2) at b = 100, which gives rise to domains of differential tension and buckling of the tissue sheet.

by contributions from the stretching energy  $\mathcal{U}_s$ , which is proportional to the shell thickness, h (here, determined by cell size), and arises due to in-plane compression or extension of the shell, and the bending energy  $\mathcal{U}_b$ , which is a result of curvature and scales as h.<sup>344</sup> Since thickness is usually small compared to shell size ( $h \ll R$ ), the stretching energy cost dominates that of bending, meaning that it is more favorable to bending than to stretching. For an isotropic and linear elastic material, the stretching and bending elastic energy of a deformed elastic shell is given in terms of the strain tensor  $\varepsilon_{ii}$  and curvature  $\kappa_{ii}$  respectively, as,<sup>44</sup>

$$\begin{split} \mathscr{U} &= \mathscr{U}_{s} + \mathscr{U}_{b} = \frac{1}{2} \int \! \mathrm{d}A \left( \lambda \varepsilon_{ii}^{2} + 2\mu \varepsilon_{ik}^{2} \right) \\ &+ \frac{1}{2} \left[ \mathrm{d}A \, B \left[ \kappa_{ii}^{2} - 2(1 - \nu) \det \left( \kappa_{ij} \right) \right], \end{split} \tag{1}$$

where  $\lambda$ ,  $\mu$  are the two-dimensional Lamé coefficients related with a two-dimensional Young's modulus  $Y = \frac{4\mu(\mu + \lambda)}{2\mu + \lambda}$ , which in turn depends on the 3D elastic modulus of the material, E, and thickness of the shell as Y = Eh. The in-plane strain  $\varepsilon_{ik}$  depends on the local contraction or expansion, which in living tissues is caused by actomyosin contractility with an effective contractile tension,  $\Lambda$ . For simplicity, we assume an inverse dependence of in-plane expansion on contractility, so that, the shell expands in-plane when contractility decreases. The second term in eqn (1) corresponds to the bending energy of the shell where  $B = Eh^3/[12(1 - \nu^2)]$  is the bending stiffness with  $\nu$  as the Poisson ratio of the material.  $\kappa_{ii}$  is the curvature strain including deviation from any spontaneous curvature. This accounts for out-of-plane displacements which arise as a result of the tendency to reduce stretching. Summation over repeating indices is assumed and det() is the determinant. We describe eqn(1) with a discretized model based on an irregular triangular mesh suited to our numeric simulations. This is described in detail in the Appendix A. The discretization of the bending energy corresponds to the general form given in the second term in eqn (1). The in-plane stretching is modeled by associated energy costs of deforming the triangles and their edges in the mesh, which effectively captures both the in-plane shear and area deformations contained in the first term in eqn (1). Although this type of discretization corresponds to an unspecified choice of the relative values of the two 2D Lamé coefficients in the corresponding continuum medium, it adequately captures the competition of bending and stretching and the resulting pattern is expected to be robust to the choice of relative weights of in-plane shear and area deformation modulii. We note that both the stretching and bending stiffness, Y and B, can be spatially inhomogeneous if the tissue shell does not maintain uniform thickness. We treat both possibilities in our model: where h is uniform, and where h depends on local in-plane area changes to keep the tissue volume constant.

To relate the elastic shell model to chemical signals, we note that several recent studies have identified that a change in the concentration of signaling agents ("mechanogens") induces change in tissue contractility. 45 Thus, it is natural to consider a dependence on concentration c of the tissue surface contractility  $\Lambda(c)$ , that

develops in the actomyosin belt and tends to constrict the cell apical surface, and plays the role of a physical "line tension" in determining cell shape. The contractility change in response to the chemical signal could be either positive or negative depending on the specific chemical.

Additionally, developing tissue is usually subdivided into discrete regions of different cells according to thresholds in concentration of morphogen gradients, 46 which lead to the classic "French flag" pattern of gene expression induced by morphogens.47 Although such compartmentalization is not general in physical systems, it is natural and common in living organisms. Inspired by patterns in biological tissues, we assume non-linear coupling between chemical pattern and mechanical stress, and impose the response in contractility strength using a sigmoid function,

$$\Lambda(c) = (\Lambda_{\text{max}} e^{c^*b} + \Lambda_{\text{min}} e^{bc})/(e^{c^*b} + e^{bc})$$
 (2)

where  $c^*$  defines the concentration threshold at which contractility changes from its maximal to minimal value,  $\Lambda_{\text{max}}$  and  $\Lambda_{\text{min}}$ , respectively, and b is the steepness that prescribes the width of transitional zone. This gives non-linear dependence of contractility on concentration leading to sharp interfaces between regions of different contractility (Fig. 1c). The mathematical form of this sigmoid dependence of  $\Lambda(c)$  is a specific choice we make to demonstrate our model predictions. A different choice that leads to a similar sharp transition in contractility is expected to lead to qualitatively similar patterns, because mechanical buckling phenomena are generic and do not rely on specific biomolecules. What is ultimately important for the mechanical deformations in our model, which follow from the general theory of thin elastic shells, is the existence of domains of unequal contraction, which can arise in practice through different biophysical mechanisms including cell surface area changes as well as cell divisioninduced expansion. Further, differential expansion can arise even if  $\Lambda$  changes linearly with concentration, c, (as opposed to the assumed sigmoid dependence) because cell shape was shown to be a bistable function of  $\Lambda$  in ref. 42.

Cell shape in living tissue is determined by a balance of mechanical forces arising from actomyosin contractility at both cell apical and lateral surfaces as well as cell-cell and cellsubstrate adhesion,42 all of which can in principle be affected by mechanogens. Following previous works that assume incompressibility, that is the conservation of cell volume, the local area expansion caused by the chemical is accompanied by a reduction of the thickness. The case of coupling between concentration and lateral, instead of apical tension, has similar but reverse effect, causing thickness increase and surface area contraction with c, due to volume conservation. Dependencies of cell shape on chemical concentration alternative to eqn (2) can therefore occur. However, even in scenarios where the cell volume can change or if the tissue retains its thickness and expands in-plane through cell division, the surface contractility,  $\Lambda$ , remains a convenient effective parameter that allows in-plane area changes required for tissue buckling. Henceforth, we omit microscopic details of individual cell shapes and their interactions, but, rather, consider the tissue Paper

sheet as a continuum and model the consequences of differential contractility along its plane.

On the other hand, it was shown experimentally that production rate of the signaling regulating developmental gene expression is coupled to mechanical stress. 48-50 Similar observations indicate the higher expression of morphogens at regions of high curvature. 51 We close the mechano-chemical feedback loop by assuming that production of chemicals is coupled to the local curvature. Though such a dependency is written on general grounds, we provide as an example a plausible mechanism for the curvature-dependent release of chemicals shown in Fig. 1a. Within this assumed picture, gaps can open between cells as the tissue bends and a curved shape (nonzero deviation of mean curvature from its intrinsic value) allowing the release of mechanogens into the extracellular fluid.<sup>52</sup> It was shown that even the interstitial flow is small, it still biases the local gradients around cells.<sup>53</sup> Since epithelial cells have apicobasal polarity,<sup>54</sup> meaning that the upper and lower sides of the tissue sheet are not chemically the same, we consider that only upward bending (positive curvature) is effective in mechanogen production. For example, if receptors are localized on the apical (upper) surface, then concentration only on this surface needs to be considered. Recent experiments<sup>55</sup> have demonstrated a similar scenario for morphogen gradients arising in regions of high curvature and spreading only on one side of the epithelial layer due to the difference in morphogen-receptor interactions on different sides. We also assume the usual uptake of mechanogens which leads to their removal from the extracellular space leading to a linear degradation rate. The mechanogens can be transmitted along the surface of tissue by diffusion in the extracellular fluid, forming concentration gradients. The time evolution of the chemical concentration on the upper side of the shell is then governed by the equation

$$\partial_t c = D\nabla^2 c + \frac{\Theta(\tilde{H})w\tilde{H}}{1 + \tilde{H}/H_s} - \beta c, \tag{3}$$

where D is the diffusivity,  $\tilde{H} = H - H_0$  is the effective mean curvature accounting for deviations of geometric curvature, H, from the intrinsic curvature of the undeformed shell,  $H_0$ , and  $\Theta(\tilde{H})$  is the Heaviside function such that production takes place only when the deviations are outward ( $\tilde{H} > 0$ ). This assumption is aimed to guarantee a stable pattern<sup>17,43</sup> and consistent with experiments.  $^{48}$  Thus,  $H_{\rm s}$  is a characteristic curvature at which the chemical production saturates to a rate given by  $wH_s$ , and  $\beta$  is the rate of degradation. The maximum local steady state concentration at a region of high curvature,  $\tilde{H} \gg H_{\rm s}$ , is then given by  $c_{\text{max}} = wH_{\text{s}}/\beta$ .

The chemical kinetics given by the diffusion, production and degradation rates defined in eqn (3) are typically slower than elastic stress propagation. For constant production rate, the timescale to reach a steady state in concentration is given by  $\beta^{-1}$ , which for morphogens in developing tissue has been measured to be of the order of hours.<sup>56</sup> This is at least one order of magnitude slower than the time scale for contractility and tissue length remodeling in epithelial cell sheets, which is of the order of minutes.<sup>57–59</sup> The strain relaxation by out-of-plane deformations is assumed to be the fastest process in the mechano-chemical loop we consider. We assume that the elastic energy relaxes through the overdamped dynamics given by,  $y\partial u/\partial t = -\delta \mathcal{U}/\delta u$ , where u is the material displacement. We consider viscous damping by the surrounding fluid and neglect any possible viscous remodeling of the tissue material.60 For a sheet of characteristic size, R, immersed in a fluid of viscosity,  $\eta$ , the frictional drag on the sheet goes as  $\gamma \sim \eta R$ . A small out-ofplane deformation  $\delta x$  leads to a bending elastic restoring force that can be estimated from eqn (1) to be  $E \cdot hR^2 \cdot h^2 (\delta x/R^2)^2 \cdot \delta x^{-1} \sim$  $Eh^3\delta x/R^2$ . By balancing this against a viscous drag force of  $\eta R\delta x/R^2$  $\tau_{\rm el}$ , we can estimate a characteristic timescale for the relaxation of the elastic deformation energy,  $\tau_{\rm el} \sim (\eta/E) \cdot (R/h)^3$ . For the typical material properties of a suspended epithelial monolayer<sup>61</sup>  $(E \sim 10 \text{ kPa}, \eta \sim 1 \text{ Pa s}, R/h \sim 10^2)$ , we get  $\tau_{\rm el} \sim 10^1 - 10^2 \text{ s}$ .

Finally, the feedback loop has a coupling between tissue deformations and the chemical production rate. Since the production rate is associated with opening interstitial gaps, we assume a very small delay in this process, but the following chemical rearrangements are slowed down due to limited diffusion discussed above. Thus, slow modes of chemical rearrangements evolve on a longer time scale, while fast modes of mechanical conformations follow them quasi-statically, accordingly to Haken's slaving principle: "fast modes are slaved by slow modes".62 This separation of timescales allows us to reproduce the tissue morphing dynamics in simulation by implementing an iterative procedure. Starting from the reference equilibrium configuration, a small perturbation in concentration is introduced, which leads to a local contractility change. The stress is updated according to the altered contractility, and since the mechanical response is fast, we find actual three-dimensional shapes by minimizing the elastic energy eqn (1). As the system reaches a mechanical equilibrium, the concentration profile is updated by a small time step taking into account the new shape that prescribes a new production rate. The iterative process continues till the change in concentration profile, which is coupled with reshaping, vanishes. Since the pattern develops on a domain of finite size, the dynamics slows down and eventually becomes stationary.

We also tested the emergent dynamics and patterns assuming comparable timescales for mechanical remodeling and chemical rearrangements. In this case we performed a steady-state dynamics by sequential computations of mechanical and chemical equilibrium states. Iteratively repeating the update, the system dynamics eventually converged. This approach is more computationally time-consuming and does not allow to reproduce dynamics with a time scale related to a measurable parameter. However, it still captures the same resulting configurations as in computations when we assume the separation of time scales described earlier. This demonstrates that even if our assumption of separation of time scales is not true in a given experimental system, the approach we use captures the final steady states in the system we consider. However in general, the mechanical response might be slowed, for example due to a delay between signal and contractility change or as a result of viscous drag in elastic relaxation, in which cases it requires simultaneous updates.

## 3 Results

**Soft Matter** 

Although eqn (1)-(3) are in principle valid for elastic shells of any shape, we now proceed with a shell of spherical geometry for the sake of concreteness. Consider a spherical shell of uniform radius R and small thickness  $h \ll R$  in the undeformed state, representing a developing tissue sheet. Assume the tissue remains at a spontaneous curvature,  $H_0 = 1/R$ , without any residual, in-plane stress. Thus, without any further perturbation, the total elastic energy of the spherical shell is vanishing and it remains in a stationary, mechanical equilibrium state. Then we introduce an initial small point perturbation of the chemical profile defined by a Gaussian function with the spot size comparable to the shell thickness, d = h, and magnitude of the order of the upper concentration limit  $c = c_{\text{max}}$  (Fig. 3 at t = 0), causing a local point-like inhomogeneity in contractility. The shell now has two domains: an outer one with a higher initial contractility ( $\Lambda_{max}$ ), and a smaller, inner one with higher chemical concentration and reduced contractility ( $\Lambda_{\min}$ ) that is associated with in-plane expansion and tendency to develop larger surface area. Thus, being constrained by the outer domain, the inner region is under compressive stress. With the shell experiencing in-plane incompatibility, it deforms to reduce stretching and is prone to bending. The macroscopic manifestation of this incompatibility is out-of-plane deformations of the shell due to buckling instability. The dynamics of deformations depends on mechanical properties of the sheet, coupling between concentration and contractility, and the diffusivity of signaling species. A full list of the parameters we use in our simulations is recapitulated in Table 1. We find that the resulting pattern is sensitive to varying thickness, h, and the concentration threshold,  $c^*$ , at which contractility transitions to its lower value. The threshold c\* determines the nonlinear mechanical response that links chemical inputs to shape change actuation. Varying this parameter leads to qualitatively different shapes.

We first aim to explore the feedback between curvature and local chemical production as a cue for spatial patterning. To see this, we assume vanishing diffusion D=0, such that production is balanced by linear degradation locally. The simulated spheres at R=30 reveal that, even in the absence of diffusion, coupled mechanical deformations and chemical production result in propagating patterns, as shown in Fig. 2. We find that when thickness  $h \geq 1$  and threshold  $c^* \leq 0.3$ , the initial

Table 1 Parameters used in simulations

Parameter	Description
$E = 10^6$	Young's modulus
$\nu = 0.5$	Poisson ratio
$\Lambda_{\text{max}} = 1$	Maximal contractility
$\Lambda_{\min} = 0.65$	Minimal contractility
b = 100	Width of transitional zone between
	domains with $\Lambda_{\max}$ and $\Lambda_{\min}$
$H_{\rm s} = 0.05$	Characteristic curvature at which the
	chemical production saturates
w = 15	Production rate
$\beta = 1$	Degradation rate
$\gamma = 100$	Friction coefficient

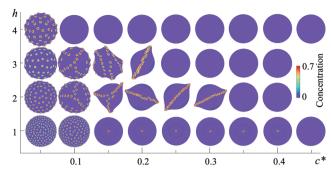


Fig. 2 Steady states of a spherical shell colored according to the local concentration at different threshold  $c^*$  and the initial thickness h. Patterns emerge as a consequence of a small initial chemical perturbation at a pole of the sphere (the central point in the top views). Higher concentration regions are associated with greater positive (outward) curvature. Simulations performed at D=0 and the initial radius R=30, other parameters are listed in Table 1.

excitation makes the state unstable and the shape becomes distorted via buckling instability. At small h, the shell can develop larger amplitude and smaller wavelength out-of-plane deflections for the same bending energy cost, <sup>63</sup> since curvature goes as  $\kappa \sim A/\lambda^2$ . In this case (lower row in Fig. 2), only a single localized bulge develops in the inner domain of lower contractility, while the outer region with the larger contractility retains its shape. Increasing thickness, h, leads to smaller out-of-plane deflections at a given compressive load leading to a smaller curvature and longer length scale deformations that extend out of the initial perturbation spot. <sup>63</sup> In this case, the positive mechanochemical feedback allows the pattern to propagate because the deflected region has higher curvature which leads to chemical production, which in turn drives the incompatibility responsible for out-of-plane deflection.

For lower concentration threshold,  $c^* = 0.05$ , there is more mechanical stress and incompatibility since the inner region over which the shell relaxes its contractility is larger. In this case, the shell adopts a pattern of almost uniformly distributed spots of high concentration, where each spot locally deforms the shell to a bulged shape. The spot size and separation between spots is seen to increase with h. The scaling law and underlying mechanism is discussed in Section 4. At intermediate  $c^*$  and thickness lower than h = 4, the emergent bulges form ridge-like lines that propagate in random directions if the initial perturbation is symmetric, but demonstrate a preferred direction when the initial spot is elongated due to non-axisymmetric perturbation. The transition between spotted and ridge structures is continuous, for example at  $0.1 < c^* < 0.2$ , mixed regimes occur with more than one ridge or consisting both types of patterning. In case of a high  $c^*$  and h, the shell is stable to the initial perturbation because the buckling-induced curvature developed at a given thickness is small and does not cause enough chemical production required for contractility change. In this case, the chemical concentration level eventually decreases at steady state due to linear degradation in eqn (3), making contractility uniform over the shell.

The time series for patterns of two representative cases are depicted in Fig. 3. A typical evolution in both cases starts with a

**Paper** 

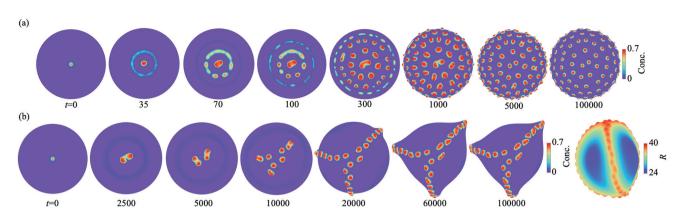


Fig. 3 Evolution of propagating chemical patterns and equilibrium shapes of a spherical shell at diffusion D = 0, the initial thickness h = 2.0, radius R = 30, and different thresholds (a)  $C^* = 0.05$ , and (b)  $C^* = 0.15$ . Concentration fields are shown in top view while the right-most panel shows the shape of the deformed shell in side view as a color map indicating the local radius (the distance to the center of mass). Simulation snapshots demonstrate occurring elastic instabilities: buckling to a bulge at the initial perturbation resulting in increased concentration and widening the spot ((a) t = 35), bulge splitting ((a) t = 75 and (b) t = 2500), buckling at secondary ring ((a) t = 70) and wrinkling along the ring ((a) t = 100). The ridge-like propagating pattern (b) arises at higher c\* when curvature at the secondary ring does not cause enough production to exceed the threshold of contractility change, whereas it becomes spotted (a) at lower c\* by means of wrinkling at the secondary ring.

small perturbation in the concentration that leads to a local decrease in contractility followed by area expansion and buckling. As curvature increases with buckling, it causes production enhancement, and as a result the size of the initial bulge grows. Then a ring of higher concentration is formed at a short distance from the initial spot (t = 35 and t = 2500 in Fig. 3a and b, respectively), which at low threshold  $c^* = 0.05$  breaks into multiple bulges along the ring (t = 70 in Fig. 3a). Each high concentration spot becomes wider following the same dynamics as the initial bulge, and creates another row of bulges (t = 100 in Fig. 3a). The process eventually slows down and terminates when the pattern occupies the entire spherical shell, with uniformly distributed spots ( $t = 100\,000$  in Fig. 3a), except at the site of the initial imperfection.

In the case,  $c^* = 0.15$ , the ring formed around the initial perturbation does not generate a contractility change in that region. For a given thickness, the concentration at the secondary ring is constant at different  $c^*$ , but spots emerge only at a low threshold. Thus, when the ring does not break into multiple spots, an initial bulge continues instead to split into two separate bulges (t = 2500 in Fig. 3b) and this division propagates along the shell creating ridge-like deformations. The lines of bulges retain their position and strongly distort the spherical shell shape when compared with the uniformly distributed spots at a lower  $c^*$ .

Although the bulges can be organized into very different patterns, the common feature of both types of patterns is the formation of separate bulges. The reason for bulge formation is the high mean curvature when the shell develops cap-like shapes with positive curvature in two orthogonal directions coupled to higher local production of chemical at these bulges. Wrinkles or folds with curvature only in one direction, provide a weaker feedback and do not cause contractility change, thus bulges become eventually round-shaped even if they were elongated at earlier stages of evolution (t = 1000 in Fig. 3a).

The assumption of cell volume conservation causes the shell to become thinner in regions where in-plane contractility

is lower. We now show that even if cell volume is not conserved and the shell remains at uniform thickness, similar spotted patterns can occur. In Fig. 4a, we display stationary shapes for

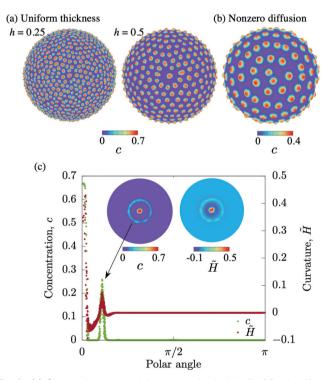


Fig. 4 (a) Chemical patterns arising on a spherical shell of fixed uniform thickness h at D = 0,  $c^* = 0.1$ , R = 30. Shells obeying the volume conservation condition at increased diffusion (b) lead to larger spot size D = 1.5, h = 2,  $c^* = 0.05$ , R = 30, than seen in Fig. 3a. (c) Concentration and curvature at nodes of mesh on sphere depending on polar angle with the initial perturbation at the pole. The state corresponds to t=35 in Fig. 3a. The secondary peak at a nonzero polar angle represents the secondary ring that forms as a result of the buckling instability around the initial perturbation at the pole.

spheres with  $c^*$  = 0.1 and of uniform thickness (shown for two different shells with varying thickness, h). We notice that the pattern can now be propagated at smaller thickness for the same contractility change. This is because the inner region with  $\Lambda_{\min}$  remains at the initial thickness and is therefore thicker now than the corresponding volume-conserved case. This leads to smaller out-of-plane deflection and curvature and the critical initial shell thickness at which propagating patterns result is lower than for the volume-conserved shells presented before.

Soft Matter

So far in this paper, we have considered pattern formation at vanishing diffusion rate valid for slow chemical spreading. Now we allow the spreading of chemicals in a thin fluid layer surrounding the shell and with thickness comparable to that of the shell to examine the effect of diffusion. For a single-cell layer of epithelium tissue, the typical thickness is  $h \sim 10 \, \mu \text{m}$ , which corresponds to a unit of length in our simulations. The typical degradation rate for morphogens  $\sim 0.01 \text{ s}^{-156}$  that gives a characteristic time in our simulations  $1/\beta$ . Thus, a unit modelling diffusion (D = 1 in simulation units) corresponds to 1 μm<sup>2</sup> s<sup>-1</sup>, which is close to the experimentally measured diffusion of morphogens in extracellular fluid.<sup>56</sup> By performing simulations at a different D and keeping other parameters constant, we found that the patterns, which appear at D > 0, show only quantitative change compared to D = 0. In Fig. 4b we demonstrate the concentration field and 3D shape at large diffusion D = 1.5, which significantly exceeds the diffusion rate in real biological systems. One can see that spot size is enlarged even though the maximum concentration decreases due to spreading. However, an increased diffusion D > 0.4 leads to a propagating pattern with multiple spots at h = 1,  $c^* = 0.15$ , while it remains in the single bulge equilibrium state at D = 0(bottom line in Fig. 2), thus, providing a qualitatively different shape. As expected, diffusion helps to scale up spot size and propagate the pattern by spreading the chemical more uniformly over the surface.

Note that without coupling between deformations and chemical production, the initial perturbation does not lead to a propagating pattern. The feedback is clearly seen in Fig. 4c, where higher curvature,  $\tilde{H}$ , (red data points) is followed by increased concentration (green data points). Eqn (3) at D = 0 in a steady state (production is the same as degradation) defines the relation between curvature and concentration  $c = w\tilde{H}/\beta(1 + \tilde{H}/H_s)$ . It allows to find concentration at given  $\beta$ , w,  $H_s$  (Table 1) and curvature, and see if it is higher than the threshold  $c^*$ . If  $\tilde{H}$  is small, or  $c^*$  is high, not enough chemical is produced to change contractility and pattern does not develop spots at the secondary ring and propagate as a ridge. In Fig. 4c one can see that concentration significantly exceeds the threshold  $c^* = 0.05$  in the vicinity of the initial perturbation at small polar angle and then decreases as curvature  $\tilde{H}$  becomes negative. A small increase in  $\tilde{H}$  at a larger angle leads to the secondary peak in c and the formation of bulges at small  $c^*$  (the inset to Fig. 4c). However, this secondary peak does not cause a contractility change at higher threshold, e.g.  $c^* = 0.15$ , for which  $c \ll c^*$ , as seen in Fig. 3b. If the production term in eqn (3) is independent of the shape change, the concentration will decrease with time

due to a linear degradation, followed by decreasing domain of low contractility, and eventually the initial buckling will disappear. Also, in the case of a constant source of chemicals without feedback, the system will approach a steady state resting at a balance between production and degradation. This last scenario will also give rise to a pattern with wrinkles in the expanded domain. The wrinkles will decay and will have increasing wavelength away from the interface, leading to a qualitatively different type of pattern.<sup>64</sup>

## 4 Analysis of elastic instabilities

We now seek insight into the mechanism of formation of the patterns shown in Fig. 2 and its dependence on concentration threshold and shell thickness. In particular, we would like to understand why the spotted pattern arises at small  $c^*$  whereas the initial bulge subdivides into two separate bulges forming the linear ridge pattern at intermediate  $c^*$ .

We distinguish three different elastic instabilities that occur successively in the shell. First, the initial perturbation in chemical field associated with the contractility change leads to buckling and formation of the initial bulge that attains different shapes depending on  $c^*$  (t=35 in Fig. 3a and t=2500 in Fig. 3b). At the same time, a ring of larger curvature and concentration appears at a short distance from the initial bulge. Finally, for the spotted patterns occurring at low  $c^*$ , this secondary ring breaks into multiple equally separated bulges (t=70 in Fig. 3a).

Let us examine the instability that occurs first, which causes the initial bulge on the shell. This is also similar to the third instability in the sequence described above, where the secondary ring develops wrinkles. For simplicity and in order to focus on the mechanical deformations alone, we numerically test the elastic instabilities on a piece of the spherical shell assumed to be nearly flat, which can be considered as a disk of radius  $R_1$  and initial thickness h. While the disk in an undeformed, reference state has initial uniform contractility  $\Lambda_{\text{max}}$ , we consider the deformations induced by an inner, circular domain of radius  $R_2 < R_1$  whose contractility is lowered to  $\Lambda_{\min}$ , as depicted in Fig. 5a, mimicking the effect of a higher chemical concentration in the initial spot. Similar to simulations on a sphere with natural curvature 1/R, we assume that the sheet is flat in the initial state  $H_0 = 0$ , the boundary is constrained against deflections to avoid boundary effects on the pattern but it is allowed for in-plane displacements, elastic properties are isotropic, and, finally, the contractility is constant in time. By prescribing values of  $\Lambda_{\text{max}}$  and  $\Lambda_{\text{min}}$  we avoid solving the chemical concentration updates in eqn (3), and perform minimization of the elastic energy eqn (1) to investigate the buckling processes involved in the formation of the initial pattern. The inner circle expands laterally in both radial and circumferential direction due to the lower contractility  $\Lambda_{\min}$ , as it tries to reach an optimal radius  $\bar{R}_2 = R_2 (\Lambda_{\text{max}} / \Lambda_{\text{min}})$ , and decreases its thickness to a value  $\bar{h}$  due to the volume conservation. It is also confined by the outer region that does not change its area and remains at the initial thickness. The biaxial compression caused

**Paper** 

(a) (b) \_0.4 0.2 (c) (d)

Fig. 5 (a) Scheme of buckling instabilities for the initial bulge and the secondary ring. (b) Wrinkling instability in a disk of contractility  $\Lambda = 15$  with the inner circle of lower contractility  $\Lambda$  = 10 and decreased thickness demonstrating different number of wrinkles depending on the initial thickness and size of the inner circle at (b) h = 2,  $R_2 = 2$ , (c) h = 2,  $R_2 = 3$ , and (d) h = 0.25,  $R_2 = 3$ . (e) The profile of the shell middle line in the vicinity of inner circle (only one half is shown due to symmetry) along the radial direction depicted by the white dashed line in (b) where the difference between deflections in two domains is comparatively small. Dotted line is the initial planar shape, while it develops a spherical cup shape in region with low contractility (red line) and small out-of-plane displacements in the outer region (purple line), which bring about the appearance of the secondary ring of radius  $R_r$ .

by this constraint can be fully or partly removed by buckling. The buckled shape depends on the elastic and geometric properties of the shell and in an equilibrium state satisfies the minimum of elastic energy eqn (1).

Deformations caused by confinement in the radial direction alone can be described as the classical Euler buckling of a plate in response to an in-plane stress in one direction. In this case a plate buckles to a spherical shell, when compressive stress exceeds the critical value<sup>63</sup>  $\sigma_{cr} = K^2 B / (\bar{R}_2^2 \bar{h})$ , where K = 2.3 is a geometric constant for the first buckling mode when ends of the plate are not allowed to rotate. Using  $R_2 = 1$ , h = 2 and the relation between biaxial stress and strain, we obtain a critical strain  $\varepsilon_{\rm cr} \approx 0.1$ , which is much smaller than the strain  $\varepsilon = \Lambda_{\rm max}/$  $\Lambda_{\min}$  – 1 = 0.5 exerted due to the contractility change in an unconstrained shell, and thus, the inner circle exhibits buckling. However, it was demonstrated that the buckling of a circular plate under biaxial compressive load leads to more complicated morphologies due to compression in circumferential direction. 65,66 Several wrinkles arise along the interface between two domains, demonstrating non-axisymmetric buckling modes as  $\sigma_{\rm cr}$  increases, while it remains spherical at the lowest eigenmode with smooth deflections and nearly circular boundaries at low  $\sigma_{\rm cr}$  value.

The characteristic wrinkling wavelength,  $\lambda$ , of an axially compressed, suspended, thin sheet is given by the scaling

relation,  $\lambda \sim \sqrt{h}/\varepsilon^{1/4}$ , that arises from the constrained minimization of elastic energy of plates under tension. <sup>67</sup> To examine how the wrinkling within the expanded inner region in our setup scales with its size and the thickness of the disk, we study the equilibrium shape of disks with varying  $R_2$  and h. Simulated disks of initial thickness h = 2 with an inner circle of reduced contractility which leads to biaxial expansion by  $\varepsilon = 0.5$ , and a smaller thickness  $h/(\varepsilon + 1)^2$  to satisfy the volume conservation condition, develop the first wrinkling instability in the azimuthal direction at  $R_2$  = 1.95 with  $\lambda \sim 6.12$ . By varying  $R_2$ , we find that the subsequent transitions to three and four wrinkles occur at  $R_2$  = 2.9 and  $R_2$  = 3.9, respectively, corresponding to the same characteristic wrinkling wavelength, λ. This is consistent with the theory expression from ref. 67 and with our simulations on a spherical shell, reproducing the same bulge shape (t = 750in Fig. 3a). Two typical examples of buckling instability resulting in different number of wrinkles with increasing  $R_2$  are depicted in Fig. 5b and c. Decreasing h at constant  $R_2 = 3.0$  leads to increased number of wrinkles with wavelength scaling as  $\lambda \sim h^{1/2}$  in agreement with ref. 67. To illustrate this, we show the disk shape at a smaller h = 0.25 in Fig. 5d, where eight wrinkles of a larger curvature appear near the boundary of the inner circle that relax toward the center. At the parameters we use to obtain the results reported on a spherical shell in Fig. 3, two wrinkles tend to form within the initial central bulge. Since the initial perturbation in simulations on a sphere is chosen to be small with width of order h, this first leads to buckling with no azimuthal wrinkles, and than two wrinkles appear when the bulge increases in radius. Each wrinkle generates increasing concentration due to higher curvature along it, which in turn causes bulge division by developing positive Gaussian curvature in two spots at the wrinkle tips instead of the single initial one.

Buckling and transition to the bulge shape causes propagating deformations in the outer region generating curvature of both signs (Fig. 5e). Near the interface, the shell has inward deflections to accommodate the gentle slope in the transition region between the thin bulge region and thick planar domain, and it also has a small positive jump in curvature exhibiting oscillatory behavior along the radial direction on a disk or along the polar direction on a sphere (Fig. 4c). Unlike the simple Euler buckling of a slender column, the out-of-plane deflections must be small because they are associated with stretching or compression of the shell in the azimuthal direction, and are thus constrained and oscillate about the initial state leading to the secondary ring. The position of the secondary ring,  $R_r$  does not define the final spot separation but slightly disturbs the spot distribution around the initial perturbation, as seen in Fig. 2 at  $c^* = 0.1$ .

The region of positive curvature forming a ring in the outer domain leads to a higher production of chemical and subsequent contractility change if  $c > c^*$ , which can be satisfied only at small  $c^*$ . The resulting local expansion of the sheet causes, in turn, another shape instability due to a local biaxial expansion at the secondary ring. Thus, multiple wrinkles appear along the ring to satisfy the incompatibility and forming the next ring at a larger polar angle. These wrinkles arising within the expanded ring break into bulges that are seen as spots in the final pattern.

For this spotted pattern (that occurs, for example, at  $c^* = 0.05$ ), we estimate both the spot size and separation, and show how the pattern arising from the buckling instability scales with changing thickness. Assuming the shape of spots is close to circular, we calculate the average spot radius as  $\sqrt{A_l/\pi N_{\rm sp}}$ , where  $A_l=$  $\sum A_j$ ,  $\Lambda(A_j) < \Lambda_{\text{max}}$  is the total surface area of regions of the shell with low contractility, and  $N_{\rm sp}$  is the total number of spots. The results for simulated spheres of different thicknesses are depicted in Fig. 6a along with linear and square root dependencies. The dependence of average spot radius on the initial shell thickness demonstrates the same  $\lambda \sim h^{1/2}$  scaling, both in the case of shells that maintain uniform thickness as well as those which conserve volume. The data is offset by a constant for the volume conservation cases, because the actual shell thickness is reduced at the spots. The pattern does not propagate when h < 0.125 and h < 1.3 for the uniform and nonuniform thickness, respectively, but the shells do exhibit the buckling instability at the location of initial perturbation. On the other hand, the shell is stable to small perturbations at large h and returns to the initial spherical shape of uniform radius.

To obtain a characteristic wavelength of the pattern, we estimate the distance between spots (in Fig. 6), which is computed as  $2(\sqrt{A_{\rm tot}/\pi N_{\rm sp}} - \sqrt{A_{\rm l}/\pi N_{\rm sp}})$ , where  $A_{\rm tot}$  is the total shell area in deformed state. One can see that the spot separation in both cases is in a good agreement with<sup>67</sup> scaling as  $\lambda \sim h^{1/2}$ . Since the domain of high contractility remains at the initial thickness in both the cases considered, they show the same dependence on h.

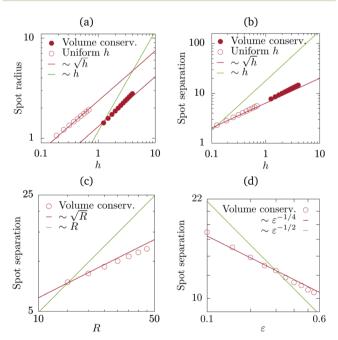


Fig. 6 Dependence of (a) spot size and (b) spot separation on the shell thickness, h, for the volume conserved (h decreases with in-plane expansion) case and for uniform h all over the shell. The plots are with respect to the initial undeformed value of h in the volume conserved cases. (c and d) Spot separation as a function of the system size R and strain  $\varepsilon$  for the volume conserved case. (a-d) Parameters are set to D=0,  $c^*=0.05$ , R=30,  $\varepsilon=0.5$ , h = 2, otherwise correspond to the horizontal axis

The simulated spheres reveal that spot separation and spot size also depend on the sphere radius R. Fig. 6c depicts the numerical results of spot separation together with different power law dependencies. It shows a satisfactory agreement with prediction of characteristic deformation length on shells scaling<sup>44</sup> as  $\sim \sqrt{R}$ , unlike deformations in plates that do not depend on the system size.

So far, the contractility change  $\Lambda_{\text{max}}/\Lambda_{\text{min}}$  associated with biaxial expansion  $\varepsilon$  was kept constant. However,  $\varepsilon$  also determines the characteristic wavelength of wrinkles. Simulations at various  $\varepsilon$ show that a decreased ratio leads to a much larger spot size and separation, obeying the scaling  $\lambda \sim \varepsilon^{-1/4}$  (Fig. 6d), and thus, demonstrating the robustness of patterns to varying model parameters. This dependence is different from wrinkling of a plate attached to a soft substrate, where wavelength depends only on thickness and the ratio between layer stiffness. 68 We provide additional computational results for patterning with large bulges in the Appendix C.

Since the ridge pattern does not occur over an extensive range of parameter space, it is difficult to establish a reliable scaling unlike the spotted patterning. Comparison of results at constant  $c^* = 0.2$  and  $2 \le h \le 3$  shows a non-linear increase in the total area with lower contractility.

We also note that the global structure of the spotted pattern is constrained by topology in analogy with the packing and crystallization of particles on curved surfaces. 69-71 The wrinkling pattern can be considered as a triangular lattice tessellation of a spherical surface with bulges at the vertices, which allows only finite number of configurations with equi-spaced vertices as defined by the Euler characteristic. 72 Any defect in this tessellation, for example a bulge with a different number of neighbors that makes the lattice irregular, causes an increase of elastic energy. Such a bulge changes its shape (the initial bulge in Fig. 3a), or eventually merges with another bulge (small bulges at the secondary ring in Fig. 3a), to reduce the elastic stress and accommodate the characteristic wavelength of spot separation. On the other hand, we note that the ridge-like structures (Fig. 3b) resemble scars on spherical crystals,73 but arise from a completely different physical mechanism. The scars are high-angle grain boundaries which develop as a row of alternating disclinations associated with positive and negative Gaussian curvature, whereas our linear rows of bulges do not require any crystalline order.

We have thus shown that the pattern formation can be understood from the mechanics of elastic instabilities. Although, these are driven by local expansile stresses that are in turn induced by a propagating chemical concentration, considerable insight into the kinetic pathway and final patterns can be obtained by analyzing the buckling and wrinkling transitions in terms of thin shell elastic theory for a prescribed differential tension created by a fixed chemical gradient.

#### 5 Discussion

Morphogenetic folding in biology encompass a wide variety of buckling mechanisms ranging from those driven by individual cell shape changes in local regions to those stemming from

**Paper** Soft Matter

differential growth rates in neighboring tissue regions.<sup>74</sup> While the details differ, all these processes require a spatial patterning of the tension or the growth rate in the tissue, which is typically accomplished by biochemical gradients and resulting gene expression in biology.

It is conceivable that there are multiple biophysical pathways through which mechanics can affect the chemical profile that imprints mechanical gradients (and which we therefore term "mechanogens"), such as through in-plane stresses that affect cell shape and membrane tension and therefore, the uptake and production of mechanogens.<sup>34</sup> Furthermore, a mechanogen can also in principle stimulate or suppress cell contractility, and thus lead to local contractions and expansions. In the absence of direct experimental evidence of such a candidate mechanogen and its effect on cell mechanics in a specific biological setting, we explore, here one such plausible mechanism in a specific, simplified setting as proof-of-concept.

In our model system, wrinkling results from differential changes in the in-plane area of the cells in response to mechanogens whose production is stimulated by tissue curvature. This positive mechanochemical feedback drives a propagating pattern of wrinkles on the surface of a thin elastic shell that are associated with both high curvature and high mechanogen concentration. We note that while this realizes a new pathway for obtaining a spotted pattern morphology on a spherical surface compared to those realized through purely mechanical means, such as by buckling of pressurized shells,24,75 or by purely chemical means, such as through phase separating chemical factors on the surface of shells modeling pollen grains, 76 the linear ridge-like morphology is very different. Unlike ridges appearing in an elastic material that is attached to a compliant substrate and subjected to a large compression, 77-79 we have shown that similar but high-aspectratio patterns can be developed in a single-layer unconstrained system due to dynamic localized expansion. Such patterns are of particular interest due to potential applications in tissue engineering and fabrication of functional surfaces, for instance hydrophobic coatings.80 This study also provides a possible pathway for diverse surface patterning in naturally occurring shells, for example the spikes, pores and ridges on pollen grains and viral capsids, <sup>70,76,81</sup> and may also have implications for deformations of cell membranes which are fluid in-plane but are endowed with curvature elasticity.82

Our mechanochemical model realizes an excitable medium, where a local initial stimulus is propagated through the medium because a localized chemical fluctuation results in a global mechanical response. This is therefore a complementary proposal to that of Turing patterns which arise from the chemical interactions of a fast and a slow diffusing chemical species. We have shown that the mechanochemical feedback between one chemical species which acts as a local activator, and the global mechanical response of elastic shell, can result in propagating and tunable pattern formation that does not require interactions between multiple chemical species with very different diffusivity. Moreover, we show that buckling deformations provide long-ranging interactions that cause spontaneous propagation of chemical signalling patterns without requiring any diffusion.

Given the complexity of biological tissue, we propose that a first experimental exploration of these ideas should be in an in vitro context, such as in organoids (collections of cultured cells in vitro that mimic organs), where recent progress has been made in studying mechanical wrinkling83 or in cell cytoskeletal extracts such as actin gels embedded with contractile myosin motors, 36 which can be combined with chemical gradients that induce differential mechanical stresses. The details of the mechanochemical feedback are likely to be different from that assumed here, but our modeling approach is general and could be easily modified to capture these scenarios.

### Conflicts of interest

There are no conflicts to declare.

## Appendix A: details of the discretized model

The elastic energy defined by eqn (1) is discretized on a triangular mesh with each node denoted by i and triangles denoted by  $\alpha$ . The length of a bond connecting the *i*th and *j*th nodes is written as  $l_{ii}$ . Each bond is spring-like and resists any change in its length, whereas each triangle has area elasticity. Both changes in bond length and area are penalized by an elastic energy cost given by the local 2D stiffness which in turn depends on the local thickness. The total stretching energy in a discrete form is then written as,

$$\mathscr{U}_{s}^{d} = \frac{1}{2} \sum_{\langle jk \rangle} \langle A \rangle_{jk} E \langle h \rangle_{jk} \left( \frac{l_{jk}}{\overline{l}_{jk}} - 1 \right)^{2} + \frac{1}{2} \sum_{a} A_{\alpha} E h_{\alpha} \left( \frac{A_{\alpha}}{\overline{A}_{\alpha}} - 1 \right)^{2},$$

$$(4)$$

where stretching energy reconstitutes the continuum limit up to particular choice of the Lamé coefficients assuming  $Y \sim Eh$ . The first term in eqn (4) gives the elastic energy cost of stretching of the bonds where  $\langle jk \rangle$  denotes a sum over all bonds. The actual thickness and area at bonds are calculated as an approximate average over the two triangles that share the bond:  $\langle h \rangle_{jk}$  =  $(h_{\alpha} + h_{\beta})/2$ , and  $\langle A \rangle_{ik} = (A_{\alpha} + A_{\beta})/3$ , where  $A_{\alpha}$  and  $A_{\beta}$  are the areas of the two triangles that share the bond, and 1/3 appears because each triangle contributes to three sides. The second term represents the area elasticity of the triangles, where  $A_{\alpha}$  and  $h_{\alpha}$  are the actual area and the actual thickness associated with the  $\alpha$ th triangle.

In our simulations, the preferred bond lengths,  $\bar{l}_{ik}$  and, as a consequence, the preferred triangle areas  $\bar{A}_{\alpha}$  are modified by the nonuniform in-plane contractility  $\Lambda(c)$ , which is prescribed by eqn (2). For simplicity, we assume an inverse proportionality between contractility and length change, and define this by  $\bar{l}_{ik} = l_{ik}^0(\Lambda^0/\Lambda(c))$ , where  $l_{ik}^0$  is the length at the initial contractility  $\Lambda^0$  (that is chosen to be  $\Lambda_{\rm max}$  in the simulations). The preferred area changes as  $\bar{A}_{\alpha} = A_{\alpha}^{0} (\Lambda^{0}/\Lambda(c))^{2}$  relatively to the initial value  $A_{\alpha}^{0}$ . In the case of nearly incompressible materials, a change in the optimal area is expected to induce a corresponding change in the optimal thickness in accordance with the Poisson's ratio,  $\nu$ ,

of the material. For very thin shells, such changes are negligible and we can work with the uniform thickness approximation. In our computations, we keep triangle thickness  $h_{\alpha}$  at the initial thickness  $h^0$  to model the case of uniform thickness, or we assume volume conservation at  $\nu = 1/2$  that implies an inverse dependence of thickness change on the in-plane area change. In this latter case, we calculate the actual triangle thickness as  $h_{\alpha} = h^0 A_{\alpha}^0 / A_{\alpha}$ , where  $h^0$ ,  $A_{\alpha}^0$  are the initial undeformed thickness and triangle areas, respectively. Not all triangles are equal in area in a non-uniform mesh, but the thickness is assumed uniform in the undeformed state.

The bending energy corresponds exactly to the second term in eqn (1) and reads as

$$\mathcal{U}_{b}^{d} = \frac{1}{2} \sum_{\text{nodes},i} \langle A \rangle_{i} B_{i} \Big( 4(H_{i} - H_{0})^{2} - 2(1 - \nu) \big( K_{i} - 2H_{i}H_{0} + H_{0}^{2} \big) \Big),$$
(5)

where the local bending stiffness,  $B_i$  depends on the local thickness,  $h_i$ , which is computed by averaging over the triangles sharing the node. The energy vanishes in planar configuration if there is no spontaneous curvature of the sheet,  $H_0 = 0$ , and at  $H_0 = 1/R$  if the undeformed shape is a spherical shell of radius R. The local curvature at each node is computed representing curvature strain via the mean and Gaussian curvatures. The discrete Gaussian curvature  $K_i = \left(2\pi - \sum_{\alpha} \rho_i^{\alpha}\right) / \langle A \rangle_i$  is expressed through the sum of all angles,  $\rho_i^{\alpha}$ , subtended at the node i by all triangles that share this node,  $\langle A \rangle_i = \sum A_{\alpha}/3$  is the approximated area associated with the node. The mean curvature  $H_i$  is calculated taking half of the mean curvature normal operator  $H_i = \sum \left( (x_i - x_j) \left( \cot \theta_{ij}^1 + \cot \theta_{ij}^2 \right) \right) / (2\langle A \rangle_i)$  defined over adjacent nodes, where  $\theta_{ii}^k(k=1,2)$  are the two angles opposite to the bond  $l_{ii}$  in the two triangles sharing the bond <sup>84</sup> (Fig. 7). The sign of the mean curvature coincides with the sign of  $H_i$ :  $x_i$ , being positive when  $H_i$  oriented outward of the spherical shell.

We assume overdamped dynamics and the positions of nodes  $x_i$  in the discretized shell are found by minimizing the elastic energy, &, defined in the main text, by following the pseudo-time evolution equations:  $\gamma \partial x_i / \partial t = -\delta \mathcal{U}_d / \delta x_i$ . Here,  $\gamma$  is the friction coefficient associated with energy dissipation during

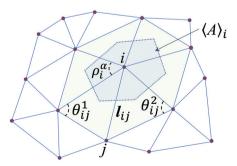


Fig. 7 Geometry of discretized shell that is constituted as a collection of nodes with positions  $x_i$  connected by links  $l_{ii}$ , where each node spans an

elastic deformation, which can be caused by the surrounding fluid as well as viscous remodeling of the tissue material.

## Appendix B: numerical methods

We numerically investigate out-of-equilibrium dynamics of a deformed tissue sheet by solving eqn (2)-(5) using spatially unstructured finite-volume discretizations. The medium of uniform thickness in initial state is triangulated on an irregular mesh consisting of 25 000 nodes satisfying the Delaunay condition using the mesh generator Distmesh.85 The number of nodes remains constant and no remeshing takes place. The discretization allows to treat three-dimensional deformations following the physical displacement of the nodes constituting the vertices of triangles. The elastic energy (4,5) is minimized over the positions of all nodes using the conjugate gradient algorithm with tolerance chosen as 10<sup>-4</sup>% of energy gain by executing a minimization step. The elastic energy gradients are calculated by sequential virtual displacement of nodes along each coordinate and then we perform iterative simultaneous update of all node positions. Once the system approaches its mechanical equilibrium, a small time step (dt = 0.01) using an explicit Euler forward scheme is performed in the dynamic eqn (3) leading to new concentrations. Then using eqn (2) contractility at each node is computed that allows to find new optimal triangle side lengths respectively to given contractility.

The spatial derivatives in diffusion eqn (3) are calculated based on local approximation using the divergence theorem (Green-Gauss) gradient scheme.86 We apply the node based method to find concentration at nodes. The concentration at node i is calculated as an average over concentration in adjacent

triangles  $c_i = \frac{\sum c_\alpha/d}{\sum 1/d}$  which is weighted according to the distance d between node and the center of the triangle  $\alpha$  (Fig. 7). The

concentration values  $c_{\alpha}$  at triangles are governed by eqn (3). To find the gradients at the center of a triangle, the Green-Gauss theorem is used, which states  $\int cndS = \int \nabla cdV$ , where c is the concentration scalar field. Then the integrals over the volume V and surface S enclosing the volume can be replaced by an approximation of gradients at triangle accounting for fluxes trough the sides of triangle<sup>87</sup> given by  $(\nabla c)_{\alpha} = \frac{1}{V_{\alpha}} \sum c_{\rm f} \mathbf{n}_{\rm f} s_{\rm f}$ , where

 $c_f$  is the concentration in the center of each triangle's side that is an average over two node values,  $n_{\rm f}$  is the normal to the triangle side,  $s_f = l_{ii}h_{ii}$  is the side area, and summation is assumed over the three triangle's sides. For chemicals diffusing along the sheet's surface in a layer of unit thickness,  $h_{ij}$  can be chosen of unit length and thus  $V_{\alpha} = A_{\alpha}$  for 2D diffusion.

## Appendix C: patterning with large spots

In addition to the patterning on a spherical shell considered in the main part of this Communication, we explored the parameter space to demonstrate how the system can be tuned to produce target patterns and to study robustness of the mechanochemical

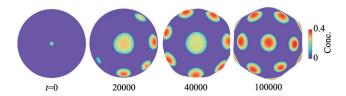


Fig. 8 Simulation snapshots of spontaneous pattern formation on a spherical shell at h = 3, R = 30, D = 0, decreased contractility change  $\Lambda_{\rm max}/\Lambda_{\rm min}$  = 1.1 and concentration threshold  $c^*$  = 0.05. The evolution reveals a fast phase of a spotted pattern formation followed by a slow coarsening. At the steady state (t = 100000), only a few spots of high concentration with large radius remain, consistent with the theoretically predicted, <sup>67</sup>  $\lambda \sim \sqrt{Rh}/\varepsilon^{1/4}$ , scaling.

feedback. Since the wavelength of wrinkling instability depends on both thickness and contractility change, the spot size and separation increase by reducing  $\Lambda_{\text{max}}/\Lambda_{\text{min}}$  and increasing h. However, larger wavelength is associated with smaller amplitude of deformations, and so to make the pattern propagating, the threshold  $c^*$  has to be low to compensate for the decreased curvature in eqn (3). Starting with a single small point-like perturbation in concentration, the resulting patterns demonstrate only a few uniformly distributed large spots of circular shape (Fig. 6). Unlike patterning with smaller structures, here the time evolution is much slower, but demonstrate similar coarsening that results in only several large, equi-spaced bulges on the shell surface. The global structure is constrained by topology and defined by the Euler characteristic.<sup>72</sup> Thus, the sphere with large characteristic spot separation accommodates a finite number of spots at given sphere radius. At  $\Lambda_{\text{max}}/\Lambda_{\text{min}} = 1.1$ , h = 3 and R = 30, the spot separation is of the order of the sphere radius and the shell develops a regular dodecahedral shape with 20 bulges at vertices. Simulations at various  $\Lambda_{\text{max}}/\Lambda_{\text{min}}$  demonstrate spot separation and radius that follow  $\lambda \sim \sqrt{Rh}/\epsilon^{1/4}$  scaling. For instance, at  $\Lambda_{\rm max}/\Lambda_{\rm min}$  = 1.15 and  $\Lambda_{\text{max}}/\Lambda_{\text{min}}$  = 1.2 the sphere demonstrates average spot separation  $\sim$  0.9R and  $\sim$  0.85R, respectively. Note that concentration saturation level is lower than in Fig. 2-4 because the developed curvature is smaller and as a result production decreased (Fig. 8).

# Acknowledgements

This work was supported by funding from the National Science Foundation: NSF-CREST: Center for Cellular and Biomolecular Machines (CCBM) at the University of California, Merced: NSF-HRD-1547848. We gratefully acknowledge computing time on the Multi-Environment Computer for Exploration and Discovery (MERCED) cluster at UC Merced, which was funded by National Science Foundation Grant No. ACI-1429783. We acknowledge insightful discussions with Stephanie Höhn, Nir Gov and Daniel Beller. We also gratefully acknowledge the anonymous reviewers for their valuable comments on an earlier version of the manuscript.

#### References

1 D. W. Thompson, On Growth and Form, Cambridge University Press, 1992.

- 2 G. Forgacs and S. A. Newman, Biological Physics of the Developing Embryo, Cambridge University Press, 2005.
- 3 T. Lecuit and P.-F. Lenne, Nat. Rev. Mol. Cell Biol., 2007, 8, 633 EP.
- 4 D. Gilmour, M. Rembold and M. Leptin, Nature, 2017, 541, 311-320.
- 5 J. M. Sawyer, J. R. Harrell, G. Shemer, J. Sullivan-Brown, M. Roh-Johnson and B. Goldstein, Dev. Biol., 2010, 341, 5 - 19.
- 6 Forces and Tension in Development, ed. M. Labouesse, Academic Press, 1st edn, 2011, vol. 95.
- 7 E. Farge, Forces and Tension in Development, Academic Press, 2011, vol. 95, pp. 243-265.
- 8 M.-E. Fernandez-Sanchez, T. Brunet, J.-C. Röper and E. Farge, Annu. Rev. Cell Dev. Biol., 2015, 31, 373-397.
- 9 D. P. Holmes, Curr. Opin. Colloid Interface Sci., 2019, 40,
- 10 J. Kim, J. A. Hanna, M. Byun, C. D. Santangelo and R. C. Hayward, Science, 2012, 335, 1201-1205.
- 11 H. Aharoni, Y. Xia, X. Zhang, R. D. Kamien and S. Yang, Proc. Natl. Acad. Sci. U. S. A, 2018, 115, 7206-7211.
- 12 Y. Klein, E. Efrati and E. Sharon, Science, 2007, 315, 1116-1120.
- 13 A. S. Gladman, E. A. Matsumoto, R. G. Nuzzo, L. Mahadevan and J. A. Lewis, Nat. Mater., 2016, 15, 413-418.
- 14 A. Zakharov and L. Pismen, Phys. Rev. Mater., 2019, 3, 055603.
- 15 T. Xie, Nature, 2010, 464, 267-270.
- 16 S. Okuda, T. Miura, Y. Inoue, T. Adachi and M. Eiraku, Sci. Rep., 2018, 8, 1-15.
- 17 M. Mercker, D. Hartmann and A. Marciniak-Czochra, PLoS One, 2013, 8, e82617.
- 18 A. Mietke, F. Jülicher and I. F. Sbalzarini, Proc. Natl. Acad. Sci. U. S. A., 2019, 116, 29-34.
- 19 R. Yoshida, Adv. Mater., 2010, 22, 3463-3483.
- 20 P. W. Miller and J. Dunkel, Soft Matter, 2020, 16, 3991-3999.
- 21 I. Levin, R. Deegan and E. Sharon, Phys. Rev. Lett., 2020, 125, 178001.
- 22 B. Li, Y.-P. Cao, X.-Q. Feng and H. Gao, Soft Matter, 2012, 8, 5728-5745.
- 23 J. W. Hutchinson, Proc. R. Soc. A, 2016, 472, 20160577.
- 24 J. Paulose and D. R. Nelson, Soft Matter, 2013, 9, 8227-8245.
- 25 A. Trushko, I. Di Meglio, A. Merzouki, C. Blanch-Mercader, S. Abuhattum, J. Guck, K. Alessandri, P. Nassoy, K. Kruse and B. Chopard, et al., BioRxiv, 2020, 513119.
- 26 B. Li, F. Jia, Y.-P. Cao, X.-Q. Feng and H. Gao, Phys. Rev. Lett., 2011, 106, 234301.
- 27 S. Höhn, A. R. Honerkamp-Smith, P. A. Haas, P. K. Trong and R. E. Goldstein, Phys. Rev. Lett., 2015, 114, 178101.
- 28 N. Stoop, R. Lagrange, D. Terwagne, P. M. Reis and J. Dunkel, Nat. Mater., 2015, 14, 337-342.
- 29 G. Cao, X. Chen, C. Li, A. Ji and Z. Cao, Phys. Rev. Lett., 2008, 100, 036102.
- 30 D. P. Richman, R. M. Stewart, J. W. Hutchinson and V. S. Caviness, Science, 1975, 189, 18-21.
- 31 J. Yin, X. Han, Y. Cao and C. Lu, Sci. Rep., 2014, 4, 5710.
- 32 J. Howard, S. W. Grill and J. S. Bois, Nat. Rev. Mol. Cell Biol., 2011, 12, 392-398.

- 33 L. Wolpert, J. Theor. Biol., 1969, 25, 1.
- 34 K. Dasbiswas, E. Alster and S. Safran, Sci. Rep., 2016, 6, 27692.
- 35 K. Dasbiswas, E. Hannezo and N. S. Gov, *Biophys. J.*, 2018, **114**, 968–977.
- 36 Y. Ideses, V. Erukhimovitch, R. Brand, D. Jourdain, J. S. Hernandez, U. R. Gabinet, S. A. Safran, K. Kruse and A. Bernheim-Groswasser, *Nat. Commun.*, 2018, **9**, 2461.
- 37 A. Senoussi, S. Kashida, R. Voituriez, J.-C. Galas, A. Maitra and A. Estevez-Torres, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, 116, 22464–22470.
- 38 T. Strübing, A. Khosravanizadeh, A. Vilfan, E. Bodenschatz, R. Golestanian and I. Guido, *Nano Lett.*, 2020, 20(9), 6281–6288.
- 39 T. J. White and D. J. Broer, Nat. Mater., 2015, 14, 1087-1098.
- 40 L. Ionov, Mater. Today, 2014, 17, 494-503.
- 41 T. Bruegmann, D. Malan, M. Hesse, T. Beiert, C. J. Fuegemann, B. K. Fleischmann and P. Sasse, *Nat. Methods*, 2010, 7, 897–900.
- 42 E. Hannezo, J. Prost and J.-F. Joanny, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 27–32.
- 43 F. Brinkmann, M. Mercker, T. Richter and A. Marciniak-Czochra, *PLoS Comput. Biol.*, 2018, **14**, e1006259.
- 44 L. D. Landau and E. M. Lifshits, *Theory of Elasticity*, Pergamon Press, New York, 1986.
- 45 D. Gilmour, M. Rembold and M. Leptin, *Nature*, 2017, 541, 311–320.
- 46 B. Goodwin, *How the leopard changed its spots: The evolution of complexity*, Princeton University Press, 2020, vol. 113.
- 47 L. Wolpert, J. Theor. Biol., 1969, 25, 1-47.
- 48 N. Desprat, W. Supatto, P.-A. Pouille, E. Beaurepaire and E. Farge, *Dev. Cell*, 2008, **15**, 470–477.
- 49 E. Brouzés and E. Farge, Curr. Opin. Genet. Dev., 2004, 14, 367-374.
- 50 T. J. Kirby and J. Lammerding, Nat. Mater., 2016, 15, 1227-1229.
- 51 B. Hobmayer, F. Rentzsch, K. Kuhn, C. M. Happel, C. C. von Laue, P. Snyder, U. Rothbächer and T. W. Holstein, *Nature*, 2000, 407, 186–189.
- 52 P. C. Benias, R. G. Wells, B. Sackey-Aboagye, H. Klavan, J. Reidy, D. Buonocore, M. Miranda, S. Kornacki, M. Wayne and D. L. Carr-Locke, *et al.*, *Sci. Rep.*, 2018, **8**, 1–8.
- 53 C.-L. E. Helm, M. E. Fleury, A. H. Zisch, F. Boschetti and M. A. Swartz, *Proc. Natl. Acad. Sci. U. S. A.*, 2005, **102**, 15779–15784.
- 54 T. J. Harris and M. Peifer, J. Cell Biol., 2005, 170, 813-823.
- 55 A. E. Shyer, T. R. Huycke, C. Lee, L. Mahadevan and C. J. Tabin, *Cell*, 2015, **161**, 569–580.
- 56 A. Kicheva, T. Bollenbach, O. Wartlick, F. Jülicher and M. Gonzalez-Gaitan, *Curr. Opin. Genet. Dev.*, 2012, 22, 527–532.
- 57 K. E. Cavanaugh, M. F. Staddon, E. Munro, S. Banerjee and M. L. Gardel, *Dev. Cell*, 2020, 52, 152–166.
- 58 A. C. Martin, M. Kaschube and E. F. Wieschaus, *Nature*, 2009, 457, 495–499.
- 59 W. Choi, B. R. Acharya, G. Peyret, M.-A. Fardin, R.-M. Mège, B. Ladoux, A. S. Fanning and M. Peifer, *et al.*, *J. Cell Biol.*, 2016, 213, 243–260.
- 60 D. A. Matoz-Fernandez, F. A. Davidson, N. R. Stanley-Wall and R. Sknepnek, *Phys. Rev. Res.*, 2020, 2, 013165.
- 61 A. R. Harris, L. Peter, J. Bellis, B. Baum, A. J. Kabla and G. T. Charras, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 16449–16454.

- 62 H. Haken, Chem. Biol., 1983, 206-216.
- 63 S. P. Timoshenko and J. M. Gere, *Theory of elastic stability*, Courier Corporation, 2009.
- 64 H. Vandeparre, M. Piñeirua, F. Brau, B. Roman, J. Bico, C. Gay, W. Bao, C. N. Lau, P. M. Reis and P. Damman, *Phys. Rev. Lett.*, 2011, **106**, 224301.
- 65 M. Ortiz and G. Gioia, J. Mech. Phys. Solids, 1994, 42, 531-559.
- 66 D. P. Holmes, M. Ursiny and A. J. Crosby, *Soft Matter*, 2008, 4, 82–85.
- 67 E. Cerda and L. Mahadevan, Phys. Rev. Lett., 2003, 90, 074302.
- 68 J. D. Paulsen, E. Hohlfeld, H. King, J. Huang, Z. Qiu, T. P. Russell, N. Menon, D. Vella and B. Davidovitch, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, 113, 1144–1149.
- 69 M. Brojan, D. Terwagne, R. Lagrange and P. M. Reis, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, 112, 14–19.
- 70 J. Lidmar, L. Mirny and D. R. Nelson, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2003, **68**, 051910.
- 71 S. Li, P. Roy, A. Travesset and R. Zandi, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 10971–10976.
- 72 H. S. M. Coxeter, *Introduction to geometry*, New York, London, 1961.
- 73 A. Bausch, M. J. Bowick, A. Cacciuto, A. Dinsmore, M. Hsu, D. Nelson, M. Nikolaides, A. Travesset and D. Weitz, *Science*, 2003, 299, 1716–1718.
- 74 M. Tozluolu and Y. Mao, *Philos. Trans. R. Soc.*, B, 2020, 375, 20190564.
- 75 N. Stoop, R. Lagrange, D. Terwagne, P. M. Reis and J. Dunkel, *Nat. Mater.*, 2015, **14**, 337–342.
- 76 M. O. Lavrentovich, E. M. Horsley, A. Radja, A. M. Sweeney and R. D. Kamien, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, 113, 5189–5194.
- 77 J. Zang, X. Zhao, Y. Cao and J. W. Hutchinson, *J. Mech. Phys. Solids*, 2012, **60**, 1265–1279.
- 78 C. Cao, H. F. Chan, J. Zang, K. W. Leong and X. Zhao, *Adv. Mater.*, 2014, **26**, 1763–1770.
- 79 C. Fei, S. Mao, J. Yan, R. Alert, H. A. Stone, B. L. Bassler, N. S. Wingreen and A. Košmrlj, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, 117, 7622–7632.
- 80 L. Mammen, K. Bley, P. Papadopoulos, F. Schellenberger, N. Encinas, H.-J. Butt, C. K. Weiss and D. Vollmer, *Soft Matter*, 2015, 11, 506–515.
- 81 S. Dharmavaram, F. Xie, W. Klug, J. Rudnick and R. Bruinsma, *Phys. Rev. E*, 2017, **95**, 062402.
- 82 N. Tamemoto and H. Noguchi, Sci. Rep., 2020, 10, 1-10.
- 83 E. Karzbrun, A. Kshirsagar, S. R. Cohen, J. H. Hanna and O. Reiner, *Nat. Phys.*, 2018, **14**, 515–522.
- 84 M. Meyer, M. Desbrun, P. Schröder and A. H. Barr, *Visualization and mathematics III*, Springer, 2003, pp. 35–57.
- 85 P.-O. Persson and G. Strang, SIAM Rev., 2004, 46, 329-345.
- 86 J. H. Ferziger, M. Perić and R. L. Street, *Computational methods for fluid dynamics*, Springer, 2002, vol. 3.
- 87 M. Deka, S. Brahmachary, R. Thirumalaisamy, A. Dalal and G. Natarajan, *J. Comput. Phys.*, 2018, 108325.
- 88 S. Li, D. A. Matoz-Fernandez, A. Aggarwal and M. Olvera de la Cruz, Proc. Natl. Acad. Sci. U. S. A., 2021, 118(10), e2025717118.