



Draft Genome Sequence of *Salegentibacter* sp. Strain BDJ18, a Plankton-Associated Bacterium in the Northeast Atlantic Ocean

Emily J. McDermith,^a Alexa R. Sterling,^a Matthew J. Bertin,^b Bethany D. Jenkins^{a,c}

^aDepartment of Cell and Molecular Biology, University of Rhode Island, Kingston, Rhode Island, USA

^bDepartment of Biomedical and Pharmaceutical Sciences, College of Pharmacy, University of Rhode Island, Kingston, Rhode Island, USA

^cGraduate School of Oceanography, University of Rhode Island, Narragansett, Rhode Island, USA

ABSTRACT *Salegentibacter* sp. strain BDJ18 was isolated from a plankton-associated seawater sample from the northeast Atlantic Ocean. We report its draft genome assembly, which includes genes potentially important for microbial interactions in the marine environment.

Salegentibacter spp., Gram-negative bacteria within *Flavobacteriia* that require oxygen and salt for growth, are known to associate with marine phytoplankton (1, 2) and have been isolated from saline habitats, including hypersaline lakes, ocean sediments, and marine animals (1). We report a *Salegentibacter* sp. isolate from the water column plankton community, indicating its potential role in marine microbial interactions.

Salegentibacter sp. strain BDJ18 was isolated from seawater that had been collected at the deep chlorophyll maximum (55 m; salinity, 36.8 practical salinity units [PSU]; 23°C) in the northeast Atlantic Ocean (36.96294°N, –71.21921°W) onboard the R/V *Neil Armstrong* in May 2017 (cruise AR16). Plankton-associated bacteria were grown by filtering seawater onto a 5-μm-pore-size filter and stamping it onto an F/2 agar plate (1% agar with filtered seawater, tryptone, yeast extract, and F/2 nutrients [3]). Colonies were assayed on chrome azurol S plates (4), and those with halos indicating siderophore production were restreaked and maintained on F/2 plates at 25°C. The *Salegentibacter* sp. strain BDJ18 colonies were yellow and were stored in 30% glycerol at –80°C in June 2017.

For genomic DNA isolation, BDJ18 was revived from a glycerol stock and grown in the aforementioned F/2 medium without agar. DNA was purified with the NucleoSpin DNA RapidLyse kit (Macherey-Nagel, Düren, Germany), quantified with a Qubit fluorometer (Invitrogen, Waltham, MA, USA), and sheared with an ultrasonicator (Covaris, Inc., Woburn, MA, USA). Sanger sequencing of the PCR-amplified 16S rRNA gene was used to identify the isolate as *Salegentibacter* sp. The sequence library was prepared by the Rhode Island Genomics and Sequencing Center (Kingston, RI, USA) using an Apollo next-generation sequencing (NGS) library preparation system with the PrepX DNA library kit (TaKaRa Bio USA, Inc., Mountain View, CA, USA), run on a Bioanalyzer DNA high-sensitivity chip (Agilent Technologies, Inc., Santa Clara, CA, USA), and quantified by quantitative PCR (qPCR) in a LightCycler 480 system (Roche Molecular Systems, Inc., Pleasanton, CA, USA) with an Illumina kit (KAPA Biosystems, Woburn, MA, USA). Samples were sequenced (2 × 300 bp) with the 600-cycle reagent kit on a MiSeq system (Illumina, Inc., San Diego, CA, USA), yielding 2,614,628 paired-end reads. Paired-end reads were uploaded to the open-source U.S. Department of Energy Systems Biology Knowledgebase (Kbase) (<http://kbase.us>) (5), where Trimmomatic v1.2.14 (6) was used to remove NexteraPE-PE adapters (2 seed mismatches, 30 palindrome clip, and 10 simple clip) and to perform quality filtering (4-bp sliding window with 15 minimum quality, 20 leading and trailing minimum quality, and 20 bp minimum). SPAdes v1.2.4 (7) with default settings was used to assemble contigs, with coverage ranging from 19× to 774×, after removal of contigs with <1,000 bp or with zero coverage. The PATRIC

Citation McDermith EJ, Sterling AR, Bertin MJ, Jenkins BD. 2021. Draft genome sequence of *Salegentibacter* sp. strain BDJ18, a plankton-associated bacterium in the northeast Atlantic Ocean. Microbiol Resour Announc 10:e00628-21. <https://doi.org/10.1128/MRA.00628-21>.

Editor Frank J. Stewart, Montana State University

Copyright © 2021 McDermith et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

Address correspondence to Bethany D. Jenkins, bdjenkins@uri.edu.

Received 17 June 2021

Accepted 9 August 2021

Published 9 September 2021

v3.6.8 Similar Genome Finder (8) identified *Salegentibacter* sp. strain T436 (GenBank accession number [PRJNA297197](#)) as a good reference genome at a distance of 0.3937, supporting the designation of BDJ18 as *Salegentibacter*. The MAUVE Contig Mover (9) in Geneious Prime v2021.0.1 ordered the BDJ18 contigs against *Salegentibacter* sp. strain T436. In Kbase, *Salegentibacter* sp. strain BDJ18 was assessed with CheckM v1.4.0 (10) and QUAST v0.0.6 (11). Annotations were performed using web-based RASTtk (<https://rast.nmpdr.org/>; February 2021) with automatic error correction, as *Salegentibacter* sp. (NCBI taxonomy identifier 903072) (12–14). FeGenie v1 (15) identified potential iron-related genes, and antiSMASH v5.0 (16) predicted specialized metabolites.

The *Salegentibacter* sp. strain BDJ18 genome is 3,847,815 bp, with 41 contigs and a GC content of 36.87%. It is 99.4% complete, with an N_{50} value of 177,704 bp (10, 11). It has 3,510 coding sequences and 45 RNAs across 264 subsystems. Potential genes include those for resistance to the antibiotic fluoroquinolone and transport of the siderophore enterobactin. Bacterium-phytoplankton interaction genes include those for potential auxin biosynthesis, which may increase phytoplankton growth (17), and those for mitigation of oxidative stress, providing possible protection from phytoplankton-derived reactive oxygen species (18). Only three potential biosynthetic gene clusters (a type III polyketide synthase [PKS] system, arylpolyene, and terpene) were identified, suggesting a limited number of modular biosynthetic systems. This draft genome increases knowledge of how marine bacteria are equipped to interact with other microbes.

Data availability. This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number [JAFLOX000000000](#). The version described in this paper is version [JAFLOX010000000](#). The associated raw sequencing reads have been deposited under the SRA accession number [SRR13857245](#) under the BioProject accession number [PRJNA706513](#).

ACKNOWLEDGMENTS

Thanks go to the AR16 chief scientist Ben VanMooy, science party, and crew. Thanks go to Janet Atoyan at the Rhode Island Genomics and Sequencing Center for helpful sequencing advice.

This material is based on work conducted at a Rhode Island National Science Foundation (NSF) EPSCoR research facility, the Genomics and Sequencing Center, which is supported in part by NSF EPSCoR cooperative agreement OIA-1655221. This project was conducted with support from NSF grant OCE-1558490 to B.D.J. and a (URI)² undergraduate research grant to E.J.M.

REFERENCES

- Bowman JP. 2016. *Salegentibacter*. In Trujillo ME, Dedysh S, DeVos P, Hedlund B, Kämpfer P, Rainey FA, Whitman WB (ed), *Bergey's manual of systematics of archaea and bacteria*. Wiley, Hoboken, NJ. <https://doi.org/10.1002/9781118960608.gbm00338.pub2>.
- Buchan A, LeCleir GR, Gulvik CA, González JM. 2014. Master recyclers: features and functions of bacteria associated with phytoplankton blooms. *Nat Rev Microbiol* 12:686–698. <https://doi.org/10.1038/nrmicro3326>.
- Guillard RRL. 1975. Culture of phytoplankton for feeding marine invertebrates, p 29–60. In Smith WL, Chanley MH (ed), *Culture of marine invertebrate animals*. Plenum Press, New York, NY.
- Schwyn B, Neillands J. 1987. Universal chemical assay for the detection and determination of siderophores. *Anal Biochem* 160:47–56. [https://doi.org/10.1016/0003-2697\(87\)90612-9](https://doi.org/10.1016/0003-2697(87)90612-9).
- Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettn TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia JM, Chia JM, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devold S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, Joachimiak MP, Keegan KP, Kondo S, Kumar V, Land ML, Meyer F, Mills M, Novichkov PS, Oh T, Olsen GJ, Olson R, Parrello B, Pasternak S, Pearson E, Poon SS, Price GA, Ramakrishnan S, Ranjan P, Ronald PC, Schatz MC, Seaver SMD, Shukla M, Sutormin RA, Syed MH, Thomason J, Tintle NL, Wang D, Xia F, Yoo H, Yoo S, Yu D. 2018. KBase: the United States Department of Energy Systems Biology Knowledgebase. *Nat Biotechnol* 36:566–569. <https://doi.org/10.1038/nbt.4163>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Prijbelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using SPAdes de novo assembler. *Curr Protoc Bioinformatics* 70:e102. <https://doi.org/10.1002/cpbi.102>.
- Davis JJ, Wattam AR, Aziz RK, Brettn T, Butler R, Butler RM, Chlenski P, Conrad N, Dickerman A, Dietrich EM, Gabbard JL, Gerdes S, Guard A, Kenyon RW, Machi D, Mao C, Murphy-Olson D, Nguyen M, Nordberg EK, Olsen GJ, Olson RD, Overbeek JC, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomas C, Vanoeffelen M, Vonstein V, Warren AS, Xia F, Xie D, Yoo H, Stevens R. 2020. The PATRIC Bioinformatics Resource Center: expanding data and analysis capabilities. *Nucleic Acids Res* 48:D606–D612. <https://doi.org/10.1093/nar/gkz943>.
- Darling ACE, Mau B, Blattner FR, Perna NT. 2004. Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res* 14:1394–1403. <https://doi.org/10.1101/gr.2289704>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.

11. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
12. Aziz RK, Bartels D, Best A, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
13. Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
14. Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
15. Garber AI, Nealson KH, Okamoto A, McAllister SM, Chan CS, Barco RA, Merino N. 2020. FeGenie: a comprehensive tool for the identification of iron genes and iron gene neighborhoods in genome and metagenome assemblies. *Front Microbiol* 11:37. <https://doi.org/10.3389/fmicb.2020.00037>.
16. Blin K, Shaw S, Steinke K, Villebro R, Ziemert N, Lee SY, Medema MH, Weber T. 2019. antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline. *Nucleic Acids Res* 47:W81–W87. <https://doi.org/10.1093/nar/gkz310>.
17. Amin SA, Hmelo LR, van Tol HM, Durham BP, Carlson LT, Heal KR, Morales RL, Berthiaume CT, Parker MS, Djunaedi B, Ingalls AE, Parsek MR, Moran MA, Armbrust EV. 2015. Interaction and signalling between a cosmopolitan phytoplankton and associated bacteria. *Nature* 522:98–101. <https://doi.org/10.1038/nature14488>.
18. Diaz JM, Plummer S. 2018. Production of extracellular reactive oxygen species by phytoplankton: past and future directions. *J Plankton Res* 40: 655–666. <https://doi.org/10.1093/plankt/fby039>.