



A Global Ocean Oxygen Database and Atlas for Assessing and Predicting Deoxygenation and Ocean Health in the Open and Coastal Ocean

Marilaure Grégoire^{1*}, Véronique Garçon^{2*}, Hernan Garcia³, Denise Breitburg⁴, Kirsten Isensee⁵, Andreas Oschlies⁶, Maciej Telszewski⁷, Alexander Barth⁸, Henry C. Bittig⁹, Jacob Carstensen¹⁰, Thierry Carval¹¹, Fei Chai¹², Francisco Chavez¹³, Daniel Conley¹⁴, Laurent Coppola¹⁵, Sean Crowe¹⁶, Kim Currie¹⁷, Minhan Dai¹⁸, Bruno Deflandre¹⁹, Boris Dewitte^{20,21,22}, Robert Diaz²³, Emilio Garcia-Robledo²⁴, Denis Gilbert²⁵, Alessandra Giorgetti²⁶, Ronnie Glud²⁷, Dimitri Gutierrez²⁸, Shigeki Hosoda²⁹, Masao Ishii³⁰, Gil Jacinto³¹, Chris Langdon³², Siv K. Lauvset³³, Lisa A. Levin³⁴, Karin E. Limburg³⁵, Hela Mehrten⁶, Ivonne Montes³⁶, Wajih Naqvi³⁷, Aurélien Paulmier², Benjamin Pfeil³⁸, Grant Pitcher³⁹, Sylvie Pouliquen¹¹, Nancy Rabalais⁴⁰, Christophe Rabouille⁴¹, Virginie Recape¹¹, Michaël Roman⁴², Kenneth Rose⁴³, Daniel Rudnick³⁴, Jodie Rummer⁴³, Catherine Schmechtig¹⁵, Sunke Schmidtko⁶, Brad Seibel⁴⁴, Caroline Slomp⁴⁵, U. Rashid Sumalia⁴⁶, Toste Tanhua⁶, Virginie Thierry⁴⁷, Hiroshi Uchida²⁹, Rik Wanninkhof⁴⁸ and Moriaki Yasuhara⁴⁹

OPEN ACCESS

Edited by:

Oscar Schofield,
Rutgers, The State University
of New Jersey, United States

Reviewed by:

Antoine De Ramon N'Yeurt,
University of the South Pacific, Fiji
Richard Kelvin Dewey,
University of Victoria, Canada
David Paul Nicholson,
Woods Hole Oceanographic
Institution, United States

***Correspondence:**

Marilaure Grégoire
mgregoire@ulliege.be
Véronique Garçon
veronique.garcon@legos.obs-mip.fr

Specialty section:

*This article was submitted to
Ocean Observation,
a section of the journal
Frontiers in Marine Science*

Received: 14 June 2021

Accepted: 14 October 2021

Published: 21 December 2021

¹ *Department of Astrophysics, MAST-FOCUS, Geophysics and Oceanography, University of Liège, Liège, Belgium,*

² Laboratoire d'Études en Géophysique et Océanographie Spatiales, CNRS/IRD/UPS/CNES, Toulouse, France, ³ National Centers for Environmental Information, National Oceanic and Atmospheric Administration, Maryland, MD, United States,

⁴ Smithsonian Environmental Research Center, Edgewater, MD, United States, ⁵ Intergovernmental Oceanographic Commission of UNESCO, Paris, France, ⁶ GEOMAR Helmholtz Centre for Ocean Research Kiel, Kiel, Germany.

⁷ International Ocean Carbon Coordination Project, Institute of Oceanology of Polish Academy of Sciences, Sopot, Poland,

⁸ Department of Astrophysics, GHER-FOCUS, Geophysics and Oceanography, University of Liège, Liège, Belgium, ⁹ Leibniz Institute for Baltic Sea Research Warnemünde, Rostock, Germany, ¹⁰ Department of Bioscience, Aarhus University, Roskilde,

¹¹ Coriolis, IFREMER, Brest, France, ¹² School of Marine Sciences, University of Maine, Orono, ME, United States,

¹³ Monterey Bay Aquarium Research Institute, Moss Landing, CA, United States. ¹⁴ Department of Geology, Lund University,

Lund, Sweden. ¹⁵ CNRS, Laboratoire d'Océanographie de Villefranche, LOV, Sorbonne Université, Villefranche-sur-Mer.

France. ¹⁶ Laboratory of Microbiology and Immunology, Life Sciences Centre, Vancouver, BC, Canada. ¹⁷ NIWA, Auckland.

New-Zealand, ¹⁸ Department of Oceanography, Xiamen University, Xiamen, China, ¹⁹ CNRS, EPOC, EPHE, UMR 5805,

Université de Bordeaux, Bordeaux, France. ²⁰ Centro de Estudios Avanzado en Zonas Áridas, La Serena, Chile.

²¹ Departamento de Biología, Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Chile. ²² CECI,

Departamento de Biología, Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Chile, ²²CEOR, Université de Toulouse, CERFACS/CNRS, Toulouse, France, ²³Department of Biological Sciences, University of Virginia.

University of Toulouse, CERIMCO/INRS, Toulouse, France, ²³ Department of Biological Sciences, University of Virginia, Virginia, VA, United States, ²⁴ Department of Biology, University of Cadiz, Cadiz, Spain, ²⁵ Maurice-Lamontagne Institute,

Virginia, VA, United States, Department of Biology, University of Cadiz, Cadiz, Spain, Maurice-Lamontagne Institute, Fisheries and Oceans Canada, Mont-Joli, QC, Canada. ²⁶ Istituto Nazionale di Oceanografia e di Geofisica Sperimentale.

Trieste, Italy. ²⁷ Department of Biology, Danish Institute for Advanced Study, Odense, Denmark. ²⁸ Dirección General

29. Japan Agency for Marine-Earth

Science and Technology, Yokosuka, Japan. ³⁰ Department of Climate and Geochemistry, Meteorological Research Institute,

Ibaraki, Japan. ³¹ The Marine Science Institute, University of the Philippines, Quezon City, Philippines. ³² Department of Marine

Biology and Ecology, Rosenstiel School of Marine and Atmospheric Science, University of Miami, Miami, FL, United States.

³³ NORCE Norwegian Research Centre, Bjerknes Centre for Climate Research, Bergen, Norway. ³⁴ Integrative Oceanography

*NORCE Norwegian Research Centre, Bjerknes Centre for Climate Research, Bergen, Norway, Integrative Oceanography Division, Scripps Institution of Oceanography, University of California, San Diego, La Jolla, CA, United States.*³⁵ State

University of New York College of Environmental Science and Forestry, Syracuse, NY, United States. ³⁶ Instituto Geofísico del

University of New York College of Environmental Science and Forestry, Syracuse, NY, United States, ³⁷ Instituto Geológico del Perú, Lima, Perú, ³⁸ Council of Scientific and Industrial Research, New Delhi, India ³⁹ Geophysical Research, Bjerknes Centre

Peru, Lima, Peru, -- Council of Scientific and Industrial Research, New Delhi, India, -- Geophysical Research, Bjerknes Centre for Climate Research, University of Bergen, Bergen, Norway.³⁹ Department of Biological Sciences, University of Cape Town

⁴⁰ Department of Oceanography and Coastal Sciences, Louisiana State University, Baton Rouge, Louisiana, USA.

LA, United States.⁴¹ Laboratoire des Sciences du Climat et de l'Environnement, CEA-CNRS-UVSQ, Gif-sur-Yvette, France.

⁴² Horn Point Laboratory, University of Maryland Center for Environmental Science, Cambridge, MD, United States. ⁴³ ABC

Centre of Excellence for Coral Reef Studies, James Cook University, Townsville, QLD, Australia, ⁴⁴ College of Marine Science, University of South Florida, St. Petersburg, FL, United States, ⁴⁵ Faculty of Geosciences, Utrecht University, Utrecht, Netherlands, ⁴⁶ Fisheries Economics Research Unit, University of British Columbia, Vancouver, BC, Canada, ⁴⁷ Ifremer, CNRS, IRD, LOPS, University of Brest, Plouzané, France, ⁴⁸ Atlantic Oceanographic and Meteorological Laboratory, NOAA, Miami, FL, United States, ⁴⁹ School of Biological Sciences, Division for Ecology and Biodiversity, Swire Institute of Marine Science, and State Key Laboratory of Marine Pollution, The University of Hong Kong, Hong Kong, Hong Kong SAR, China

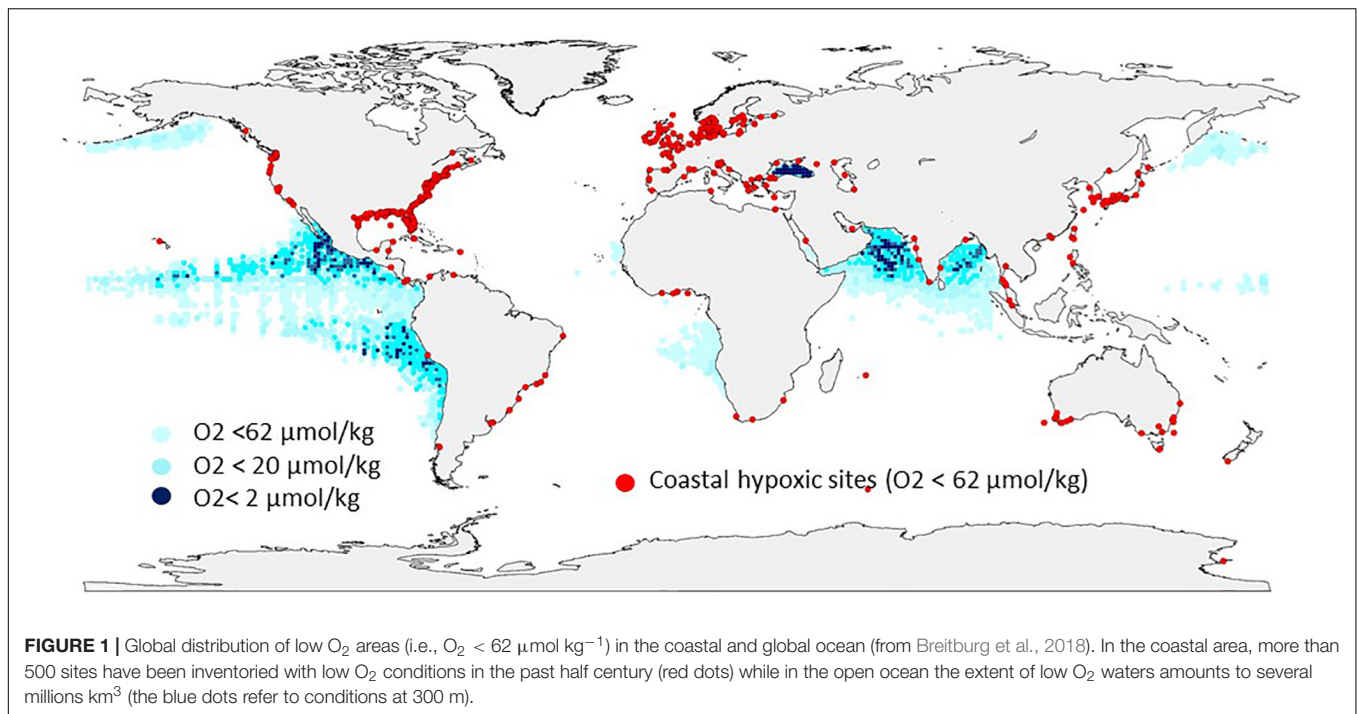
In this paper, we outline the need for a coordinated international effort toward the building of an open-access Global Ocean Oxygen Database and ATLAS (GO₂DAT) complying with the FAIR principles (Findable, Accessible, Interoperable, and Reusable). GO₂DAT will combine data from the coastal and open ocean, as measured by the chemical Winkler titration method or by sensors (e.g., optodes, electrodes) from Eulerian and Lagrangian platforms (e.g., ships, moorings, profiling floats, gliders, ships of opportunities, marine mammals, cabled observatories). GO₂DAT will further adopt a community-agreed, fully documented metadata format and a consistent quality control (QC) procedure and quality flagging (QF) system. GO₂DAT will serve to support the development of advanced data analysis and biogeochemical models for improving our mapping, understanding and forecasting capabilities for ocean O₂ changes and deoxygenation trends. It will offer the opportunity to develop quality-controlled data synthesis products with unprecedented spatial (vertical and horizontal) and temporal (sub-seasonal to multi-decadal) resolution. These products will support model assessment, improvement and evaluation as well as the development of climate and ocean health indicators. They will further support the decision-making processes associated with the emerging blue economy, the conservation of marine resources and their associated ecosystem services and the development of management tools required by a diverse community of users (e.g., environmental agencies, aquaculture, and fishing sectors). A better knowledge base of the spatial and temporal variations of marine O₂ will improve our understanding of the ocean O₂ budget, and allow better quantification of the Earth's carbon and heat budgets. With the ever-increasing need to protect and sustainably manage ocean services, GO₂DAT will allow scientists to fully harness the increasing volumes of O₂ data already delivered by the expanding global ocean observing system and enable smooth incorporation of much higher quantities of data from autonomous platforms in the open ocean and coastal areas into comprehensive data products in the years to come. This paper aims at engaging the community (e.g., scientists, data managers, policy makers, service users) toward the development of GO₂DAT within the framework of the UN Global Ocean Oxygen Decade (GOOD) program recently endorsed by IOC-UNESCO. A roadmap toward GO₂DAT is proposed highlighting the efforts needed (e.g., in terms of human resources).

Keywords: oxygen, atlas, database, observing, mapping, data-products, open and coastal ocean, deoxygenation

INTRODUCTION

Current evidence indicates that the coastal (i.e., most directly influenced by land) and open ocean is losing oxygen (O₂) since the middle of the last century (**Figure 1**), with consequences for living organisms and biogeochemical cycles that are not yet fully understood (e.g., Keeling et al., 2010; Breitburg et al., 2018). In the open ocean the O₂ inventory has decreased by a few

percent (i.e., 0.5–3%) and the Oxygen Minimum Zones (OMZs) are expanding. Although observations and model simulations document a net decline of the global ocean O₂ inventory, there is disagreement among analyses (**Figure 2**) resulting in inconsistent regional estimates of the rate of O₂ loss (i.e., deoxygenation) (e.g., Oschlies et al., 2018). Uncertainties and differences between estimates could be attributable to the scarcity of accessible data, the use of different datasets (e.g., volume



and quality of data) and the employment of different mapping techniques and models.

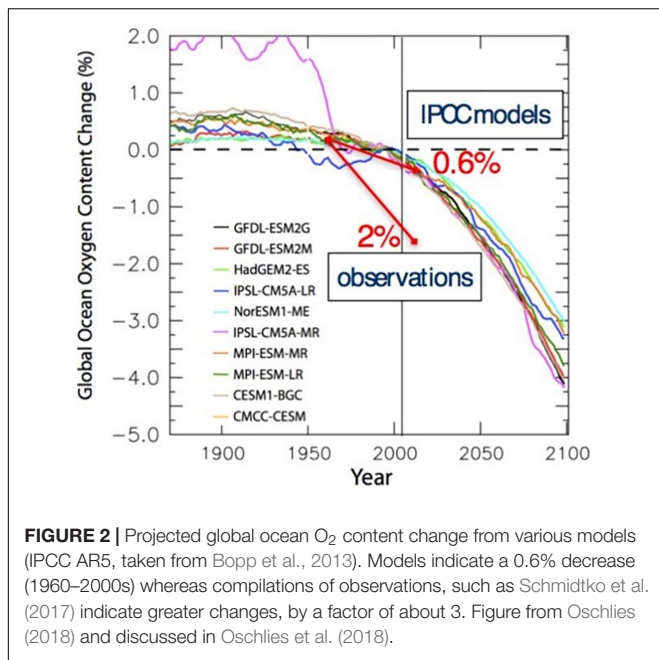
In the global coastal ocean, O_2 deficient zones are mostly intermittent features that essentially appear in the benthic boundary layer (BBL, i.e., the layer overlying the seabed where processes are influenced by the presence of the bottom) when the O_2 consumption by respiration outpaces the O_2 supply by ventilation and bottom photosynthesis (e.g., Pitcher et al., 2021). The number of coastal hypoxic sites has increased in response to worldwide eutrophication; yet a precise quantification of the O_2 trend in the global coastal zone is still debated. The availability of O_2 observations in international databases does not allow a comprehensive quantitative assessment of the severity of hypoxia (i.e., $O_2 < 60 \mu\text{mol kg}^{-1}$) at seasonal and interannual scales. This issue is particularly critical in regions with limited institutional infrastructure where the connection between National Oceanographic Data Centers (NODCs) and international oceanographic databases is not always well-established, or NODCs do not exist. Short-term (i.e., in terms of hours) and small-scale (i.e., in the BBL) variability and amplification of non-linearity in the coastal ocean require data of higher temporal and spatial resolution to characterize O_2 dynamics and events of depletion. The reference distribution of hypoxic sites assembled by Diaz and Rosenberg (2008) (Figure 1) highlighted for the first time the worldwide extent of the coastal hypoxia phenomenon. This effort has been valuable but could be updated and amended with the large volume of (sometimes disparate) quantitative information on coastal O_2 concentrations, including inventories of the frequency, timing, duration, intensity and spatial extension of the hypoxic events, and links to the original data contained in a globally accessible database. Tracking of the severity and evolution of coastal hypoxia requires access to

coherent observational O_2 datasets for the global coastal ocean that are amenable to global analysis.

Since the community white paper by Gruber et al. (2010) that called for the addition of O_2 sensors on Argo floats, the number of Argo O_2 profiles has increased considerably, up to 220,880 O_2 profiles as of June 8th 2021. Profiling biogeochemical (BGC) Argo floats provide unprecedented vertical (i.e., at least one order of magnitude larger than the 12–36 points offered by Niskin bottles) and temporal ($\sim$ each 5–10 days) resolution and coverage at regional and global scales down to a depth of 2000 m. Currently $\sim$ 10% of the active Argo floats (i.e., 396 over 3828 floats) have O_2 sensors¹. This percentage increases to more than 25% (i.e., 41 over 152 floats) when considering the Deep Argo floats only. This spurs initiatives like the GOOS strategy for BGC variables and the international BGC Argo program (Johnson and Claustre, 2016; Roemmich et al., 2019). This new program aims at operating a global array of 1000 BGC-floats that measure vertical profiles of all six core BGC and environmental variables including O_2 . In this context, the U.S. National Science Foundation has recently decided to add 500 BGC floats (all measuring O_2) to the global Argo network in the next five years, and several other nations are increasing their contributions, which will boost the array to meet 50% of its target for global BGC sampling of the ocean.

The increasing number of observation platforms will improve our capabilities to monitor O_2 in the open ocean and would remedy the chronic lack of O_2 data compared to physical data for mapping and modeling marine systems. However, these several hundred thousand O_2 profiles collected by profiling floats and gliders are not currently incorporated in a global compilation of data. Current gridded products and assessment of climate trends

¹<https://fleetmonitoring.euro-argo.eu/dashboard>



(e.g., Garcia et al., 2005, 2013, 2019; Stramma et al., 2008; Lauvset et al., 2016; Schmidt et al., 2017) are generally based on Winkler and CTD data. The lack of a community accepted data standard treatment (e.g., quality checks, adjustment procedures) and information to the users (e.g., quality flagging) across databases and platforms challenges the development of uniform-quality data-synthesis products that combine different data sources. The integration of sensor and Winkler data from different databases would offer the possibility to advance data-synthesis products at an increased resolution. It would also support the development of an O₂ gridded product for the global coastal ocean facilitating a regular assessment of the evolution of coastal hypoxia in a warming climate.

This paper aims at outlining the need, establishing and proposing a roadmap toward the building of a Global Ocean Oxygen Database and Atlas (GO₂DAT) for regional seas, coastal zones and open ocean. GO₂DAT will integrate O₂ data from both Eulerian and Lagrangian observations (i.e., Winkler titrations and O₂ sensor measurements performed on conductivity-temperature-depth-O₂ sondes, fixed moorings, autonomous platforms, and other emergent platforms). It will adopt a fully documented metadata format with traceable quality control (QC) procedures and assess uncertainty including quality flagging (QF), that will be agreed upon by the scientific community. The GO₂DAT concept is aligned with the principles of the Framework for Ocean Observing (FOO, UNESCO, 2012). GO₂DAT will comply with the FAIR² principles (Findable, Accessible, Interoperable, and Reusable, Wilkinson et al., 2016), be available free of charge and contribute to the motto “measure once-use many times.” GO₂DAT datasets and products are envisaged to support ocean research and societal needs and will be made available through a web-platform for scientific analysis,

²FAIR principles: <https://www.go-fair.org/fair-principles/>

decision making and education (**Figure 3**). In this paper, we aim at engaging the community (e.g., scientists, data managers, policy makers, service users) toward the development of GO₂DAT.

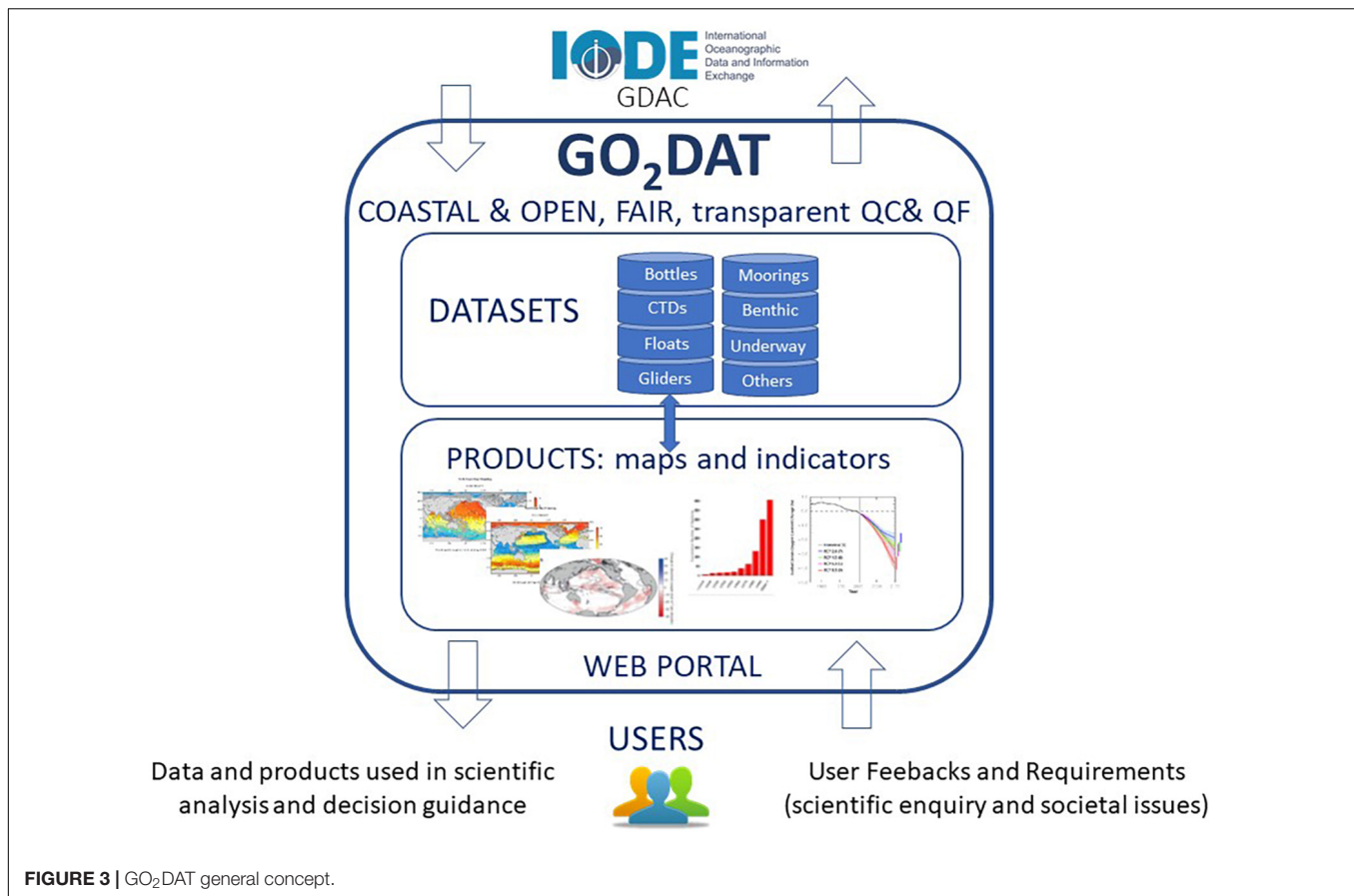
This paper is organized along the three pillars of the FOO value chain of O₂ data from requirement setting, through data collection to product delivery. We first demonstrate the need for GO₂DAT in support of scientific analyses and decision making and underline the requirements on the quality of O₂ data to assess O₂ state and variability. Next, we review the methodologies for observing ocean O₂ concentrations. The basic principles underlying the most used techniques to measure O₂, known technical challenges and possible solutions are described. For each observing platform, we synthesize the evolution of O₂ measurements, calibration procedures and the existence of Data Assembly Centers (DACs) that centralize data processing according to an established protocol vetted by the oceanography community. The adequacy of current O₂ data to assess variability at climate and weather scales and calibration procedures applied to collected O₂ data (i.e., post-calibration or second order calibration) are discussed. Finally, a roadmap for achieving GO₂DAT is presented.

SCIENTIFIC RATIONALE

Physical and Biogeochemical Controls of Ocean O₂ Dynamics

The dynamics of O₂ in the ocean is governed by both physical, biogeochemical and biological processes. The ocean gains O₂ in the upper layer (i.e., ~0–100 m) due to photosynthesis by autotrophic organisms and dissolution from the atmosphere in undersaturated waters near the surface. Conversely, O₂ is lost due to the diffusion of O₂ to the atmosphere in over-saturated surface waters, and from the respiration of aerobic organisms and oxidation of chemical species in the water column and sediments. The surface mixed layer is well oxygenated in most of the ocean, but below the sunlit surface layer, there is no photosynthesis and, the renewal of the O₂ consumed requires a physical mechanism (i.e., currents) that transports relatively well-oxygenated waters to regions of lower O₂. This mechanism, called ocean ventilation, is responsible for the oxygenation of the ocean interior and modifies the distribution of O₂ within the ocean. The distribution of O₂ in the ocean interior thus offers information on physical and biogeochemical mechanisms in the ocean. For instance, physical oceanographers investigate the circulation and mixing of water masses from its distribution (e.g., Körtzinger et al., 2004; Coppola et al., 2017; Racapé et al., 2019; Li and Tanhua, 2020; Ulses et al., 2021), marine biogeochemists infer the balance of biological and chemical processes from its variability (e.g., Emerson et al., 1997; Keeling and Manning, 2014; Resplandy et al., 2018) and the O₂ content is an indicator of ocean health.

In some subsurface regions where the renewal of O₂ by physical transport is low, the O₂ concentration can reach levels that are too low to support the survival of many aerobically respiring organisms. Organisms that are found in these areas can exhibit reduced growth and reproduction, avoidance, and, in extreme cases and for non-mobile species, experience low-O₂



induced mortality (e.g., Levin, 2003; Levin et al., 2009). In the open ocean, OMZs are mainly found in the Eastern boundary upwelling systems in the tropical Pacific and Atlantic, and in the northern Indian Ocean (**Figure 1**) and extend from ~100 to 1000 m (e.g., Helly and Levin, 2004; Paulmier and Ruiz-Pino, 2009). In these OMZs, the degradation of the organic material sinking from the surface takes place through denitrification leading to a consumption of fixed nitrogen and production of N₂ and potentially N₂O (**Figure 4**). Observational analysis and model results indicate that OMZs have expanded during recent decades (e.g., Stramma et al., 2012; Schmidtke et al., 2017).

O₂ dynamics in the coastal ocean can be highly variable over small spatial and temporal scales, from diel cycles with nighttime hypoxia developing in productive coastal systems (Tyler et al., 2009) to episodic events developing over days, to seasonal and multiannual scales (Rabalais et al., 2002; Conley et al., 2009; Carstensen and Conley, 2019). Coastal hypoxia occurs when O₂ consumption outpaces its supply, (e.g., during the period of stratification), driving concentrations below thresholds critical for living aerobic communities that cannot escape and have not developed adaptation process. Since the middle of the last century, the worldwide eutrophication³ process has increased the level of primary production in the coastal

zone, promoting bottom O₂ depletion with consequences for living marine communities (e.g., Conley et al., 2009; Rabalais et al., 2010; Yasuhara et al., 2012, 2017; Breitburg et al., 2018). Scientific papers and technical reports document the widespread occurrence of coastal hypoxia around the world (Diaz and Rosenberg, 2008; Breitburg et al., 2018; Pitcher et al., 2021), although some regions are still poorly investigated (Altieri et al., 2017).

Why Is a Global O₂ Database and Atlas Needed?

The need for a standardized, well-documented, high-quality, and comprehensive database integrating O₂ data and other environmental variables from multiple sources is growing rapidly with the increasing threat of ocean deoxygenation for marine ecosystems and delivery of their associated services to society.

The decision-making process aimed at sustaining a healthy, productive and resilient ocean as prioritized by international policies and initiatives (e.g., UN SDG 14, the EU MFS, the UN Decade of Ocean Science for Sustainable Development) requires sound environmental mapping and modeling capabilities of known quality.

³The enrichment of water by nutrients causing an accelerated growth of algae and higher forms of plant life to produce undesirable disturbance to the balance of

organisms present in the water and to the quality of the water concerned (OSPAR definition).

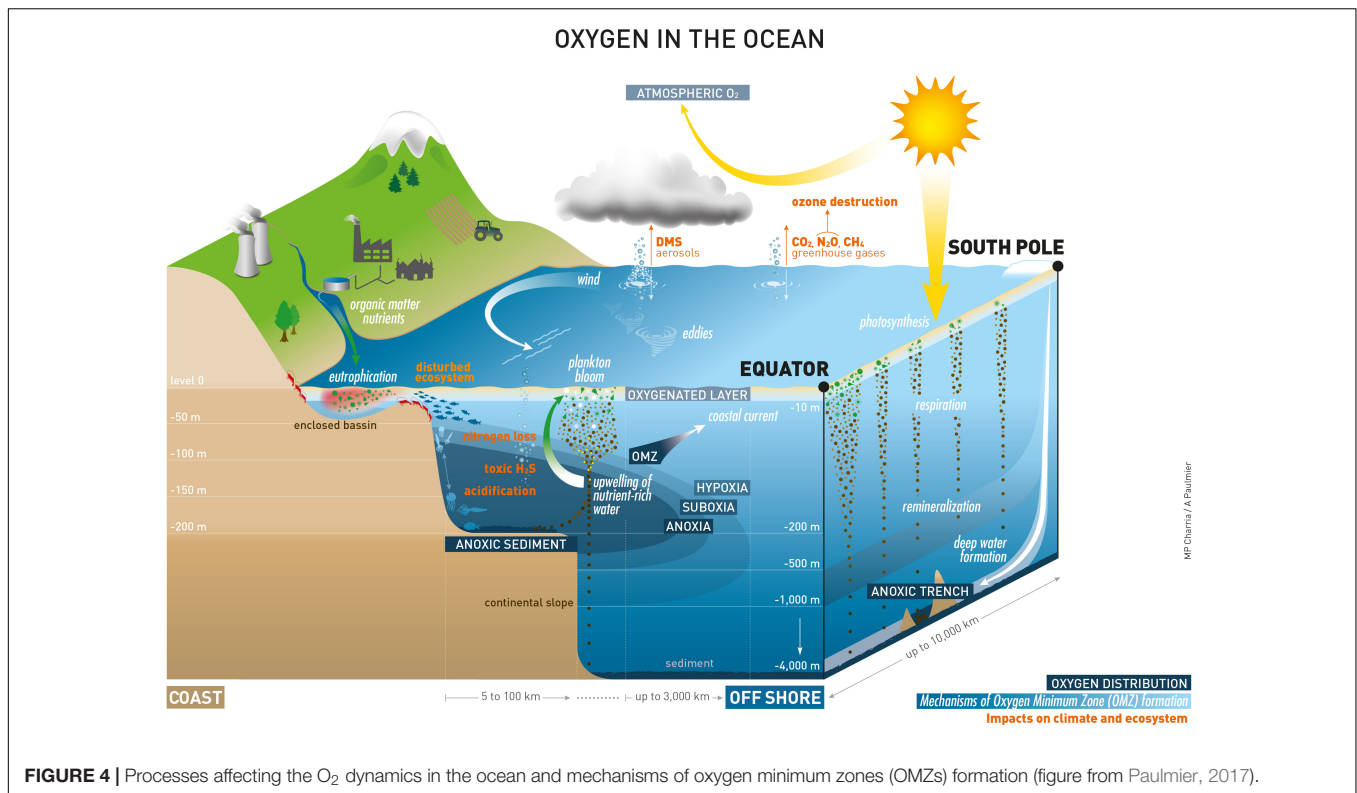


FIGURE 4 | Processes affecting the O₂ dynamics in the ocean and mechanisms of oxygen minimum zones (OMZs) formation (figure from Paulmier, 2017).

O₂ data and derived products (e.g., indicators, maps) can be used by international policy makers (see **Table 1** for a definition of acronyms) and the science that supports policy in the context of climate change (UNFCCC, IPCC), water quality and pollution (e.g., IMO, GESAMP), biodiversity (e.g., CBD, IPBES); international legally binding instruments UNCLOS-BBNJ, seabed mining (e.g., LTC-ISA), fisheries (FAO, RFMOs) and the UN General Assembly. Marine spatial planning and significant, special, vulnerable and protected area designations can benefit from O₂ predictions (e.g., Ecologically or Biologically Significant Marine Areas, Vulnerable Marine Ecosystems). GO₂DAT can promote achievement of Sustainable Development Goal (SDG) 14 on the oceans, and specifically SDG14.1 on pollution, SDG14.5 on conserving 10% of the ocean, SDG 14.7 on Small Island Developing States (SIDS) sustainable use of marine resources, including through sustainable management of fisheries, aquaculture and tourism, and SDG 14.a on scientific knowledge, research capacity and marine technology.

Following the call for international programs contributing to the success of the UN Ocean Science Decade (IOC, 2019), GO₂NE in collaboration with partners submitted the “Global Ocean Oxygen Decade” (GOOD) program which has been endorsed. The activities and actions planned will raise global awareness about ocean deoxygenation, provide knowledge for action and develop hypoxia mitigation and adaptation measures through local, regional and global efforts, including intensified monitoring, transdisciplinary research, bi-directional knowledge transfer among stakeholders and scientists, innovative outreach and ocean education and literacy. The high-level

objective of GOOD is to provide data, knowledge and best practices to enable society, stakeholders, and scientists to co-design and develop measures that can mitigate the drivers and impacts of ocean deoxygenation and provide appropriate adaptation measures where mitigation is not possible. GOOD will be implemented through several projects carried out by different consortia in different regions of the world ocean. The development of GO₂DAT is proposed as one of the outcomes of the GOOD program.

GO₂DAT will be a key element, a tool to support assessments and decision-making leading to mitigation strategies for climate change and its impacts, maintaining ocean health and biodiversity preservation, and sustainable fisheries management (Garçon et al., 2019; Laffoley and Baxter, 2019; Limburg et al., 2020). Declining O₂ is a stressor that covaries with warming and ocean acidification (Zhai et al., 2009; Gobler and Baumann, 2016; Breitburg et al., 2018) and now there is opportunity to ready the scientific and management communities for the next set of analyses and applications. When combined with socioeconomic data such as those reported in Dyck and Sumaila (2010), Swartz et al. (2013), Pauly and Zeller (2016), and Sumaila et al. (2019), GO₂DAT will support the decision-making processes associated with the emerging blue economy.

Below, we illustrate the many uses of O₂ data in coastal sciences and oceanography using selected examples which relate to the societal outcomes stated as part of the UN Ocean Decade. Our review is not comprehensive but rather serves to show the diverse uses of O₂ in analyses and modeling and how GO₂DAT would provide data to many applications.

TABLE 1 | Examples of stakeholders, international conventions, directives and programs that would benefit from GO₂DAT.

International bodies	
CBD	Convention on Biological Diversity
EEA	European Environmental Agency
FAO	Food and Agriculture Organization of the United Nations
GOOS	Global Ocean Observing System
GESAMP	Group of Experts on the Scientific Aspects of Marine Environmental Protection
IMO	International Maritime Organization
IOC-UNESCO	Intergovernmental Oceanographic Commission of UNESCO
IOCCP	International Ocean Carbon Coordination Project
IPBES	Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services
IPCC	Intergovernmental Panel on Climate Change
LTC-ISA	The Legal and Technical Commission of the International Seabed Authority
RFMO	Regional Fisheries Management Organizations and advisory bodies
WMO	World Meteorological Organization
International conventions, directives	
EU MSFD	EUropean Marine Strategy Framework Directive
UNCLOS-BBNJ	United Nations Convention on the Law of the Sea for marine Biodiversity of areas Beyond National Jurisdiction
UNFCCC	United Nations Framework Convention on Climate change
International programs, projects, networks and initiatives	
CMEMS	Copernicus Marine Environment and Monitoring Service
DOSI	Deep-Ocean Stewardship Initiative
EMODnet	European Marine Observation and Data Network
GLODAP	Global Ocean Data Analysis Project
GOOD	Global Ocean Oxygen Decade
IMBeR	Integrated Marine Biosphere Research
IODE	International Oceanographic Data and Information Exchange program
SOLAS	Surface Ocean Lower Atmosphere Study
UN Decade	UN Decade of Ocean Science for Sustainable Development

A Clean Ocean

Increasingly, O₂ factors into management-related analyses of ecological responses to nutrient inputs into coastal zones (Rabalais et al., 2007a; Kemp et al., 2009), ensuring sustainable fisheries (Breitburg et al., 2009; Dulvy et al., 2011; Eero et al., 2015; Rabalais, 2015), ocean management under climate change (Queirós et al., 2016; Lu et al., 2018; Levin et al., 2020), and as indicators of the progress of large-scale restoration efforts (e.g., Rabalais et al., 2010; Andersen et al., 2011; Zhang et al., 2018). Tomasetti and Gobler (2020) argue that environmental regulations and water quality standards have not kept pace with the rising importance and increasing scientific understanding of coastal hypoxia and acidification. Although some coastal zones are well characterized, many are not; for example, it has been found that corals in the tropics may be at risk from both warming, eutrophication and hypoxia (Altieri et al., 2017; Hughes et al., 2020).

A Healthy and Resilient Ocean

The O₂ content is an indicator of ocean health. The development and extension of low O₂ areas degrade the living conditions and vertically contract the metabolically viable habitat for a large number of pelagic, mesopelagic, and benthic organisms. At 100% saturation, the amount of O₂ available in one liter of water is ~35 times less than that in an equivalent volume of air. The effects on individuals exposed to low O₂ can cascade and result in altered foodweb structure, including the diversion of organic matter flow to low O₂ tolerant species like microbes (Breitburg et al., 2018). O₂ data are important for assessing the habitat quality of many organisms within the coastal-shelf-open ocean environments. Examples include field-based determination of effects of O₂ on zooplankton (Wishner et al., 2018; Roman et al., 2019), benthos (Levin, 2003; Rabalais et al., 2007b; Levin et al., 2009; Yasuhara et al., 2012, 2017; Gammal et al., 2017), fish and shellfish (Keister et al., 2000; Breitburg, 2002; Pollock et al., 2007; Thomas and Rahman, 2010; Keller et al., 2017; Limburg and Casini, 2019; Roman et al., 2019), deep-sea coral along seamounts (e.g., Anderson et al., 2016), and fish habitat (e.g., Stramma et al., 2012; Sánchez-Velasco et al., 2019; Gallo et al., 2020). The effects of low O₂ on the physiological conditions of organisms have been investigated in many laboratory and field studies (e.g., Vaquer-Sunyer and Duarte, 2008; Wu, 2009; Neilan and Rose, 2014; Limburg and Casini, 2018, 2019) and are summarized by the metabolic index (ability to meet temperature-dependent O₂ demands) and related indices (P_{crit}) for species in a variety of systems (Chu and Gale, 2017; Deutsch et al., 2020; Howard et al., 2020; Seibel et al., 2021). The tight link between thermal tolerance (TT) and O₂ and capacity limitation (OCL) is proposed to govern body size, species biogeographical range distributions and their response to climate change (Pörtner, 2021).

A Productive Ocean

Evidence is growing that O₂ is an important driver of fish population and food web dynamics (Noone et al., 2013; de Mutsert et al., 2017; Townhill et al., 2017; Cheung, 2018; Rose et al., 2018) and its relationship with fishing activities (vessels and fishers) has been documented for multiple fisheries (Druon et al., 2017; Keller et al., 2017; Purcell et al., 2017). Understanding O₂ conditions in the current and future climate has important consequences for management decisions in fisheries and other sectors. For example, Asch et al. (2018) used a habitat-based approach and showed that maximum catch potential under climate change (2100 under business-as-usual RCP 8.5 scenario), which included a decrease in O₂ concentrations, would be reduced by more than 50% in some areas for the Pacific Island countries and territories. Rose et al. (2019) recently reviewed, using case studies, how low O₂ in coastal systems and OMZs can affect the modeling analyses used to inform fisheries management decisions; situations arise that result in decisions (management advice) that likely underestimate the risk of overfishing.

Beyond fisheries, climate-sensitive industries such as aquaculture and tourism may rely on O₂ data for a historical perspective, for spatial planning, and potentially for the provision of an early warning. Management of living resources and tourism can benefit from improved hypoxia modeling, understanding

changes in habitat quality, links to HABs, and resilience. The same is true for environmental management of ocean industries extracting non-living resources such as minerals or oil (Levin et al., 2020).

A Predicted Ocean

A challenge going forward is the use of O₂ data to understand and mitigate climate risks. This will involve cross-system comparisons and extensive development and testing of biogeochemical models. Presently, O₂ data are used to provide inventories, budgets, and spatio-temporal trends analyses (Fennel and Testa, 2019; Mavropoulou et al., 2020) on regional (Long et al., 2016; Coppola et al., 2017; Levin, 2018; Sasano et al., 2018; Savchuk, 2018) and global scales (Beaty et al., 2017; Schmidtke et al., 2017; Claustre et al., 2020).

O₂ data are frequently used to calibrate, validate, initialize and force regional (Montes et al., 2014; Vergara et al., 2016; Duteil et al., 2018) and earth system (global) models (Ridgwell et al., 2007; Andrews et al., 2017; Kriest et al., 2017; Oschlies et al., 2018). O₂, along with other water quality variables, has been incorporated into data assimilation (Dickey, 2003; Brasseur et al., 2009) to improve the forecasting skill of biogeochemical, water quality, and nutrient-phytoplankton-zooplankton models (Pastres et al., 2003; Gharamti et al., 2017). Further analyses and model development to project future ocean O₂ content and distribution in the open and coastal ocean will be greatly enhanced by the development of GO₂DAT that will offer a high-quality reference dataset for constraining formulation, parameterization, assimilation and validation of models.

A Safe Ocean

GO₂DAT will offer the information to support a better understanding and forecasting of hypoxic and anoxic events, which in the extreme cases are hazards to human health. Deoxygenation can lead to the production of H₂S, a toxic gas that can eventually escape to the atmosphere, and consumption of spoiled seafood killed by low-oxygen events can lead to illness in people dependent on these resources for health and livelihoods.

An Accessible Ocean

With some exceptions such as the World Ocean Database (WOD, Boyer et al., 2018), GLODAP (Global Ocean Data Analysis Project, Key et al., 2004; Olsen et al., 2016, 2020), the Copernicus Marine Environment and Monitoring Service (CMEMS) and the European Marine Observation and Data Network (EMODnet) – Chemistry (Vinci et al., 2017; Martín Míguez et al., 2019), a large portion of observational O₂ data are not openly accessible, nor are they usable online without restrictions and with well-documented uncertainty estimates. For this reason, most analyses of O₂ data use only a subset of all that are available and the quality and quantity of the data vary across analyses. Each system has multiple case studies that use different, often independent O₂ datasets, making cross-systems analyses a challenge. In some cases, published information (rather than the underlying data) has been used to characterize O₂ conditions in coastal and ocean areas (Diaz and Rosenberg, 2008; Gilbert et al., 2010; Levin, 2018). Additionally, to improve the overall quality of individual datasets,

researchers often perform empirical adjustments (e.g., data bias adjustments) and specific quality control procedures that are often not described in detail. Transparency of quality control protocols and resulting adjustments is paramount in order to achieve reproducibility of data analyses (e.g., trend estimate) and reusability for data synthesis. It is difficult to replicate independently the findings of most studies that assess the trend of ocean O₂ content based on observations. The GO₂DAT will offer a single unified database to enable more rigorous and comparative analyses, and better integration of results across studies within a system and across systems.

An Inspiring and Engaging Ocean

GO₂DAT can also support educational activities from K-12 through to underpinning Ph.D. theses, provide a resource for journalists and other communication media, facilitate capacity building and provide a foundation for technology transfer. GO₂DAT will contribute to the building of a digital twin of the ocean for O₂ making available to a wide variety of stakeholders a quality control information on ocean O₂ in the three space directions and in time.

Quality Requirements of O₂ Data to Assess Mean State, Weather and Climate Variability

The required accuracy (i.e., the difference between the measurement and a true value, the Certified Reference Material, CRMs, which is not directly defined for O₂ but indirectly through e.g., Winkler, atmospheric pO₂) and precision (i.e., the repeatability between measurements of a same sample) of the O₂ measurements differ depending on the scientific questions to be answered, and the signal amplitude to be detected which varies from a few tenths up to a few $\mu\text{mol kg}^{-1}$ for the climate and weather changes, respectively (Table 2). Quantifying global ocean deoxygenation trend (i.e., $0.075 \mu\text{mol kg}^{-1} \text{yr}^{-1}$ or $\sim 4.5 \mu\text{mol kg}^{-1}$ over the last 60 years) imposes stringent requirements on the quality of the data, while for estimating the typical O₂ levels in a region or the occurrence of hypoxia, even far less accurate data can be of scientific value (e.g., accuracy better than $5 \mu\text{mol kg}^{-1}$).

The quantification of the Net Community Production (i.e., NCP, the net difference between community photosynthesis and respiration) from the high frequency variation of O₂ is very sensitive to the presence of bias and noise in the data. A 0.5% ($\approx 1 \mu\text{mol kg}^{-1}$) error in O₂ measurements significantly degrades the quality of the retrieved NCP (Emerson et al., 2008; Soetaert and Grégoire, 2011). In OMZs, because, nanomolar levels O₂ inhibit denitrification and anaerobic ammonium oxidation, studies of these processes require sensors with detection limits of a few nanomoles (Tiano et al., 2014).

For ecological analyses, the required data quality depends on the specific organisms, biological processes, and questions (hypotheses) being addressed. Common topics include detecting, and if possible, quantifying, the O₂ effects on individual organisms in terms of: (1) presence, absence or abundance, (2) energetics-related physiology, and (3) behavior and vital process

TABLE 2 | Examples of scientific topics, characteristics of the signal, recommended quality, applications.

Scientific topics	Signal characteristics to capture	Recommended quality of O ₂ measurement	Applications
Mean state determination			
Order of magnitude of the O ₂ content Gridded fields Coastal hypoxic area	227.4 ± 1.1 petamoles (10 ¹⁵ mol) ⁽¹⁾	Accuracy better than 5 μmol kg ⁻¹	Model initialization, validation
OMZs (O ₂ < 45 μmol kg ⁻¹) Extension, Oxycline AMZs (O ₂ < a few nmol kg ⁻¹) Extension	High vertical gradient, up to 20 μmol kg ⁻¹ m ⁻¹ ⁽³⁾ Detection of zero value	Accuracy better than 2.5 μmol kg ⁻¹ ⁽²⁾ High ratio of the sampling frequency over the response time (a few sec) and/or low vertical sampling velocity ⁽⁴⁾ Zero detection a few nmol kg ⁻¹ ⁽³⁾	Ocean health, environmental Management Ocean health Global denitrification, model parameterization in anoxic conditions
Climate variability			
Global ocean deoxygenation trend	Global decrease: 4.38 ± 1.98 μmol kg ⁻¹ since 1960 ⁽¹⁾	Accuracy better than 0.5 μmol kg ⁻¹	Climate application Carbon budget
Weather variability			
Air-Sea seasonal fluxes	Open ocean 5–10 μmol kg ⁻¹ in oligotrophic waters, up to 37.5 μmol kg ⁻¹ for well-mixed regions ⁽⁵⁾ . Coastal vegetated ecosystem 37.5–50 μmol kg ⁻¹ ⁽⁶⁾ .	Accuracy: ~1–5 μmol kg ⁻¹ Precision: ~ 1–5 μmol kg ⁻¹	
Processes estimation			
Net Community Production	Open ocean: from 1 to 6 molO ₂ m ⁻² yr ⁻¹ ⁽⁵⁾ . Coastal vegetated ecosystem: 23.2 ± 8.2 molO ₂ m ⁻² yr ⁻¹ ⁽⁶⁾ . > 20 mol m ⁻² yr ⁻¹ (during the months of convection) ^(9,10)	Very sensitive to the presence of noise (e.g., an error of 0.5% degrades the estimation of the rate) ^(7,8) . Smoothing is recommended. Deep waters: 1–2 μmol kg ⁻¹ Intermediate waters: few μmol kg ⁻¹	Understanding ocean physical and biogeochemical functioning
Ventilation/convection			

Recommended quality of O₂ measurement is done through either referring to existing studies or considering that the measurement's accuracy should be better than 1% of the expected signal. (1) Schmidtko et al., 2017; (2) 1% of saturation; (3) Tiano et al., 2014; (4) Bittig et al., 2014; (5) Emerson et al., 2008; (6) Champenois and Borges, 2019; (7) Emerson et al., 2008; (8) Soetaert and Grégoire, 2011; (9) Körtzinger et al., 2004; (10) Coppola et al., 2017.

rates of growth, mortality, and reproduction (Batiuk et al., 2009). Analyses are also performed to quantify O₂ effects on groups of individuals, such as spatial distributions, and on scaled-up responses, such as population-level (reduced annual abundance), community (e.g., diversity), and food web (e.g., changes in predator-prey interactions) (Wu, 2002). In some cases, very high precision and accuracy will be needed because responses are ultra-sensitive to small changes in O₂ concentrations (Wishner et al., 2018), while in other situations precision and accuracy can be relatively low because the taxa or responses of interest show low sensitivity to variation or the O₂ concentrations themselves vary greatly across the comparisons.

METHODOLOGY FOR O₂ OBSERVATION

Methods for Measuring O₂

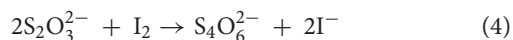
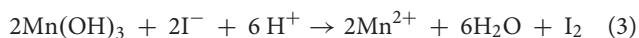
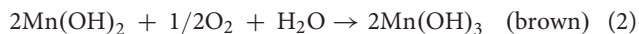
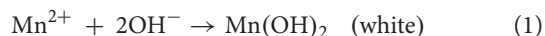
There are three major methods used for determining the amount of O₂ in seawater. These are (a) the iodometric titration (the

so-called Winkler titration); (b) electrochemical approaches (e.g., potentiometry, amperometry); and (c) optical (fluorescence quenching) sensors method. Each method has its specific merits, applications and quality.

Iodometric Methods: Winkler Titration

Winkler titration (Winkler, 1888) is a reference method used for 140 years to measure O₂. Winkler O₂ determination during oceanographic cruises requires a discrete sample commonly collected from a Niskin bottle into a glass bottle (Winkler bottle). The Winkler chemical titration is based on the oxidation of Mn-(II) by O₂ in alkaline conditions (Equations 1 and 2) and the reaction of the precipitated Mn-(III) with iodide in acidic conditions to form iodine and tri-iodine in a second step (Equation 3). The lumped amount of iodine and tri-iodine is quantified by a titration with thiosulfate (Equation 4; Carpenter, 1965; Langdon, 2010) or spectrophotometrically (Labasque et al., 2004) and from that the concentration of O₂ is determined. The Winkler O₂ measurement is a commonly used reference to

calibrate the data provided by O₂ sensors.



Sensors

The electrochemical and optical methods for measuring O₂ are simple, fast and automated. They are now widely used on platforms such as CTD-O₂ rosette samplers, autonomous platforms (e.g., floats, gliders), drifting buoys and fixed-point moorings. Their use on autonomous platforms offers continuous monitoring of O₂ in space and time throughout the ocean. The number of sensor-based O₂ measurements in the water column collected in the past decade alone is now comparable to the Winkler O₂ data collected since 1900 (**Figure 5**).

Electrochemical Sensors

Electrochemical detection is the most widely used method, within which the polarographic O₂ sensor is the most common application (see the review by Wei et al., 2019). The working principle of polarographic sensors is based on the measurement of a current that is proportional to the rate of O₂ diffusing through a permeable membrane and being reduced on a metal surface. In Clark-type O₂ sensors (Clark et al., 1953), the O₂ diffuses through the membrane and dissolves in the electrolyte before reacting with the cathode. The amount of O₂ that diffuses through the membrane depends on the external O₂ partial pressure (pO₂) (Glud et al., 2000). At the cathode, O₂ is reduced

and gives an electric signal that depends on the amount of O₂. This implies that the sensor signal is dependent on the transport of the analyte from the surrounding medium, the so-called stirring sensitivity (Glud et al., 2000). A robust conversion of the electric signal into a pO₂ requires a steady flux of O₂ into the sensor tip. If there is no steady-state O₂ diffusion across the membrane and cathode, the conversion is not robust. There is thus a trade-off between the sensor's stirring sensitivity and its response time: sensors with a high stirring sensitivity have short response time but the O₂ flux is much more variable. Low stirring sensitivity microsensors can be constructed while macro-sensors are more affected by this effect.

The modifications of Clark-type O₂ microsensors led to the development of the STOX (Switchable Trace amount OXygen) sensor, based on Clark-type microsensors, where O₂ can be determined at low (nM) concentrations (Revsbech et al., 2009). The small dimensions of the tip in O₂ microsensors make them able to achieve response times of 0.1 s and are thus recommended for high-resolution measurements. However, O₂ microsensors are more affected by aging and sensitive to drifts than macrosensors due to their reduced quantity of electrolytes and the smaller size of their cathodes.

Optical Sensors

Optical O₂ sensors (optodes) have become attractive in the past four decades because there is no O₂ consumed during the measurement (i.e., no stirring sensitivity as with polarographic sensors), they offer good precision and accuracy and they can measure O₂ in both gas and liquid phase. Bittig et al. (2018, 2019) provide a thorough review of the most widely used optodes. Optical O₂ sensors used in oceanography are mostly

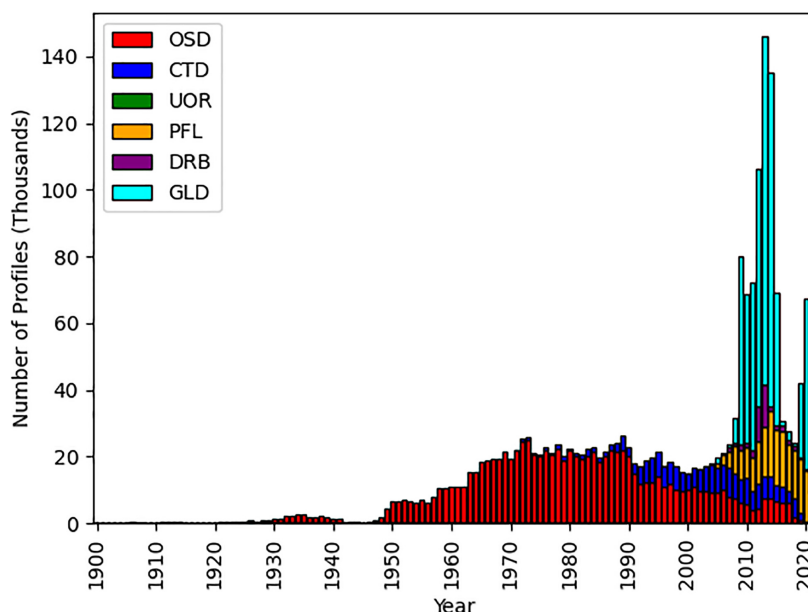


FIGURE 5 | Number of dissolved O₂ profiles available in the WOD 2018 (WOD18, Boyer et al., 2018) by observing system/instrument. OSD: chemical Winkler data from shipboard bottle Ocean Station Data; CTD, sensor data from Conductivity-Temperature-Depth-Oxygen (~200 000 profiles); PFL, Profiling Float data (~160 000 profiles); DBD, Drifting Buoy Data (~40 000 profiles); GLD, GLider Data (~450 000 profiles); UOR, Undulating Oceanographic Recorder (~360 profiles).

based on the principle of luminescence quenching (Lakowicz, 2006). A luminescent substance is embedded in a sensing foil or coating and placed on an optical window. The sensing foil of the optode is permeable only to gases, i.e., the sensors respond to the gases' partial pressure, in this case pO_2 . By modification of the luminescent substance, oxygen optodes can be used to measure trace amounts of oxygen down to a detection limit of 0.5 nM (Lehner et al., 2015), though only available so far for laboratory measurements.

Platforms

We provide an overview of the Eulerian and Lagrangian platforms from which O_2 is measured using the Winkler titration method or optical and electrochemical sensors. **Table 3** provides a synthesis, for each O_2 observing platform, of information on the existence of best practices documents, a DAC that centralizes data processing, community-agreed calibration, correction (secondary QC) and QF as well as level of accessibility, in particular, in Real Time (RT) and Delayed Mode (DM) mode. This across platforms analysis allows to understand how the different platforms are currently organized in terms of calibration, QC and QF and can be coordinated toward the building of GO₂DAT.

Ship-Based CTD- O_2 Observations

Over the past decades, there have been significant advancements in O_2 sensor instrumentation mounted on CTD rosette samplers (e.g., Wang et al., 2019) and probes. Historically, polarographic sensors were used on ship-based CTDs while now optical sensors are also used. The polarographic sensors used on CTD casts are mostly based on the Clark-type polarographic membrane-covered electrodes. In order to avoid the stirring effect (Gust et al., 1987) without compromising the time response of the sensors in rosette samplers, an external water pump supplying a constant water flow to the sensor is used in the standard CTD configurations. In areas with sharp O_2 gradients such as in OMZs and low O_2 coastal waters the cast speed should be adjusted to adequately sample the oxycline.

Irrespective of present-day O_2 sensor type, calibration at sea or in delayed mode of O_2 sensors is often performed using simultaneous water sampling at one or preferably more selected depths (e.g., at all the Niskin sampling depths) together with routine O_2 analysis using Winkler titrations (Langdon, 2010; Uchida et al., 2010). This calibration is particularly important in oceanic regions with large spatial O_2 concentration gradients. International research programs such as the World Ocean Circulation Experiment (WOCE), Climate and Ocean:

TABLE 3 | Characteristics of the last generation of ocean O_2 observing systems: (1) Bittig and Körtzinger, 2015; (2) Thierry et al., 2018; (3) Thierry and Bittig, 2018; (4) Bittig et al., 2019; (5) BGC-Argo data are freely available from the Global Data Assembly Centres (GDACs: <ftp://ftp.ifremer.fr/ifremer/argo/> and <ftp://usgoda.org/pub/outgoing/argo/>); (6) Testor et al., 2019; (7) oceangliders (<https://www.oceangliders.org/>); (8) Uchida et al., 2010; (9) WOCE and GO-SHIP protocols; (10) WOCE and GO-SHIP data are freely available on CLIVAR and Carbon Hydrographic Data Office; (11) Coppola et al., 2016; (12) Best Practices EMSO/OceanSites (OBPS); (13) National Data Buoy Center (NDBC) in the United States and Coriolis at IFREMER; (14) in delayed mode (see GO-SHIP method); (15) under development; (16) based on Winkler or 100%-0% calibration of sensors; (17) no DAC but Stratmann et al., 2019.

Observing platform	Methods	Spatial and temporal scales resolved	Spatial and temporal coverage	Community-agreed calibration and best practices	Community-agreed QF	Secondary community-agreed correction	GDAC	(N)RT/DM FAIR	
Profiling floats	Sensors	Horizontal: 10^{-2} km Vertical: 10^{-3} km 5–10 days	10^2 – 10^3 km 3–5 years	With the atmosphere ⁽¹⁾	Yes ^(2,3)	Yes <i>In situ</i> Drift ⁽⁴⁾	Yes ⁽⁵⁾	NRT and DM	Yes
Gliders	Sensors	Horizontal: 10^{-1} –1 km Vertical: 10^{-3} km Hourly-daily	1– 10^2 km Several days-weeks	In progress ⁽⁶⁾	No	No	Yes ⁽⁷⁾	NRT and DM	Yes
Ship-based Repeat Hydrography	Winkler + Sensors	Horizontal: 10 km Vertical: 10^{-3} km Decadal	10^2 – 10^3 km Multi-decadal	At sea or in delayed mode using Winkler	Yes ⁽⁸⁾	Yes ⁽⁹⁾	Yes ⁽¹⁰⁾	DM	Yes
Moored fixed point observatories	Mainly optical sensors	Hourly	Multi-year	Yes ⁽¹¹⁾	Yes ⁽¹²⁾	No	Yes ⁽¹³⁾	NRT & DM	No
Ship-based Fixed-point Observatories	Winkler + Sensors	Monthly	Multi-year	No ⁽¹⁴⁾	Yes ⁽⁸⁾	Yes	Yes ⁽¹⁰⁾	DM	Yes
Ship-based underway observations	Optical sensors	Sub-weekly to monthly	10 – 10^2 km	No ⁽¹⁵⁾	No	No	No	DM	No
Coastal/benthic	Winkler + Sensors	Vertical: 10^{-5} – 10^{-4} km Temporal: punctual	10 – 10^2 km	Yes ⁽¹⁶⁾	No	No	No ⁽¹⁷⁾	DM	No

Table adapted from the Global Ocean Observing System (GOOS) Panel-Biogeochemistry-01-EOV-Oxygen Essential Ocean Variables (EOV) version 2.0 (August, 2017) see here https://www.jcomm.info/index.php?option=com_oce&task=viewDoclistRecord&doclistID=168.

Variability, Predictability and Change (CLIVAR) and Global Ocean Ship-based Hydrographic Investigations Program (GO-SHIP) adopt the precision of the “best-laboratory” O₂ data. For example, WOCE and GO-SHIP adopted an O₂ target precision of 0.08% ($\sim \pm 0.2 \mu\text{mol kg}^{-1}$) of the highest concentration found in the ocean (Joyce, 1991; Hood et al., 2010).

Biogeochemical Argo

O₂ was the first BGC variable to be measured by Argo (Körtzinger et al., 2004; Gruber et al., 2010) and continues to be the most common BGC variable measured. **Figure 6** illustrates how Argo floats can be used to investigate the mesoscale O₂ dynamics at the oxic-anoxic transitional layer in the Black Sea where the O₂ level varies from saturation to zero.

Since 2002, BGC Argo floats have been equipped with different categories of O₂ sensors (electrochemical, optical, multipoint-calibrated, air-calibrated). **Table A1** in Appendix summarizes information on the types of sensors used, known issues, adjustments performed and data streams. BGC-Argo has two data streams: “RT” and “DM.” The RT stream, targeted at operational users, is subject to automated, RT QC (e.g., automated search for outliers) and may receive an automated, initial adjustment (“A” data). The DM stream, targeted at science users, provides the best quality data.

Some floats include regular in-air measurements as recommended by the SCOR WG 142 (Bittig et al., 2015). This provides an accurate (i.e., 1%), easily accessible reference for correction (Bittig and Körtzinger, 2015; Johnson et al., 2015; Bushinsky et al., 2016) of the drift between manufacturer calibration and deployment (Johnson et al., 2017; Bittig et al., 2018). For older floats where in-air measurements were not performed, such data can be mimicked by using the WOA

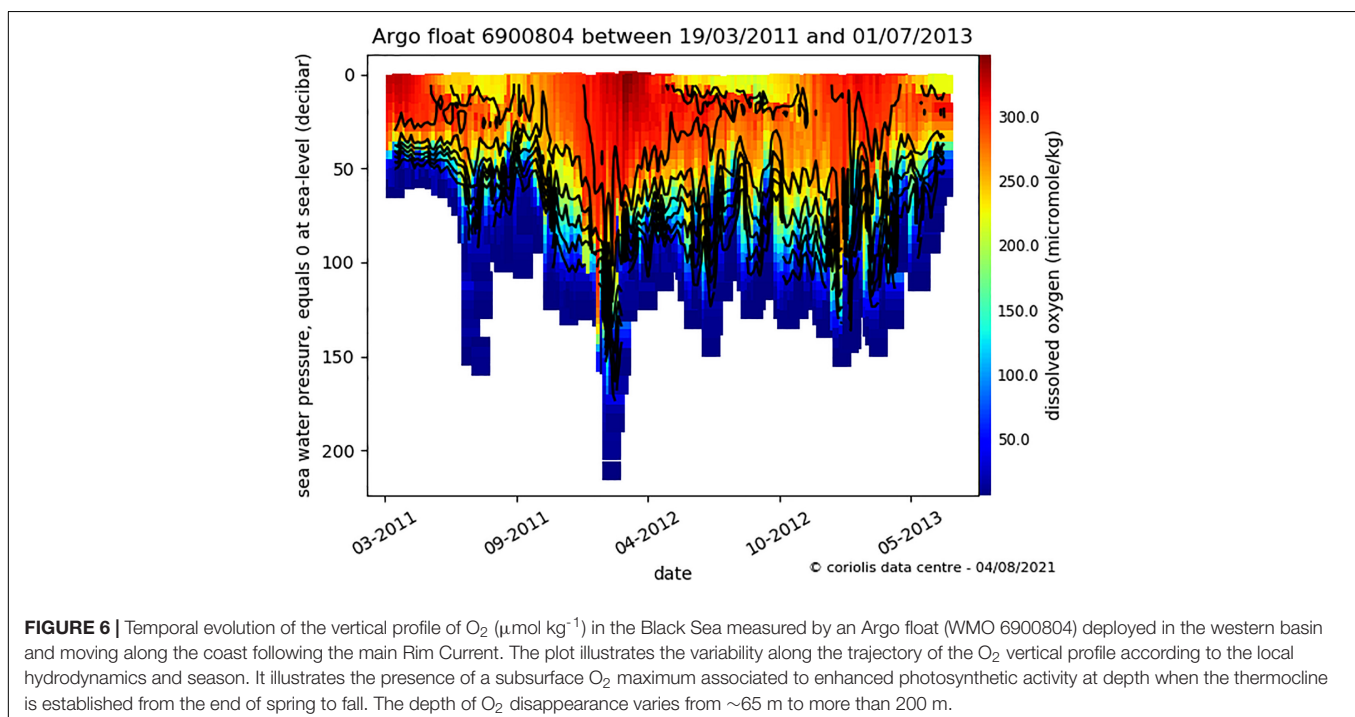
surface O₂ saturation field and converting O₂ saturation to surface partial pressure (pO₂) using the float’s temperature and salinity (Thierry and Bittig, 2018).

When available, a laboratory calibration using standard O₂ water (Winkler) before the launching of the float and corrections based on frequent ship observations after the float launching can be used to check the accuracy of measurements. If multiple check and correction methods are available before and after launching Argo O₂ floats, the integrated calibration procedure will be useful to assess and improve accuracies for the O₂ and the long-term drift of O₂ values.

Gliders

Underwater gliders may be thought of as profiling floats that are able to control their horizontal position (Rudnick, 2016a). This control over position makes gliders especially useful for regional oceanography, complementing the global footprint of Argo. The same suite of sensors as in floats is used, including Aanderaa O₂ optodes (Nicholson et al., 2008; Adams et al., 2016), and Sea-Bird SBE 43 (Ohman et al., 2013) and 63 (Takeshita et al., 2021). Gliders are used to investigate the (sub)mesoscale O₂ variability (**Figure 7**).

Because gliders are deployed and recovered, there is opportunity for pre- and post-mission laboratory calibrations and drift correction. In some applications of Sea-Bird instruments, the sensor is plumbed in line with the CTD, taking advantage of a constant flow and the presence of tributyltin antifouling measures. The requirement for plumbing in this case and the need to minimize drag on gliders make it challenging to measure in air for *in situ* calibration. However, Nicholson and Feen (2017) used a mount forward of a glider’s tail fin to achieve this calibration at sea using an Aanderaa optode.



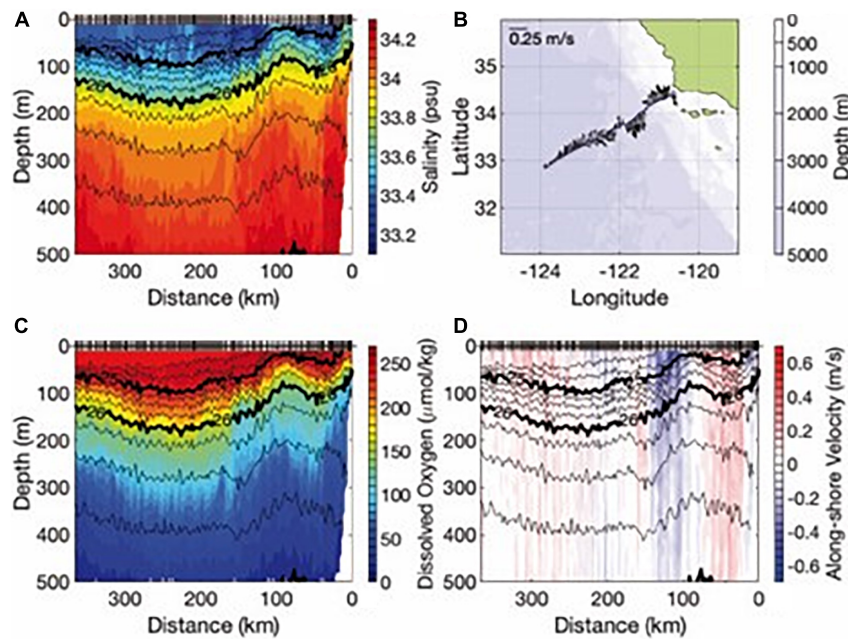


FIGURE 7 | Data from the California Underwater Glider Network (Rudnick, 2016b; Rudnick et al., 2017). Shown are (B) the track and 500-m depth average velocity, and sections of (A) salinity, (C) dissolved O₂, and (D) alongshore velocity. Nearshore upwelling brings dense low O₂ water to the surface. Further offshore a strong cyclonic feature has upward doming isopycnals with associated high salinity and low O₂. The separation between glider profiles is 3 km allowing resolution of these structures with sizes of tens of kilometers.

The OceanGliders program (Testor et al., 2019) provides an international umbrella in GOOS for ocean observations using underwater gliders. In common with many regional observing assets, the support for glider programs usually comes from the countries where the observations are undertaken. For this reason, the international glider data system is not as advanced as other more mature programs like Argo. Data management is accomplished through regional DAC, with established centers in the United States, Australia, and Europe. An ongoing initiative is to coalesce around a single international underwater glider data format. This format, which will be compliant with Climate Forecasting conventions, is likely to be adopted within the next 2 years.

Moorings

Although any type of O₂ sensor can be used on fixed-point mooring systems, optical sensors are more common due to their long-term stability compared to electrochemical sensors. In general, the response time of optical sensors is slower than that of electrochemical sensors; however, the effect of slow response time is reduced on fixed-point mooring systems as the surrounding environment (O₂ concentration, temperature and pressure) does not change rapidly. But, if O₂ sensors with slow response times are used on moored profilers, data quality may be reduced particularly in environments with strong vertical gradients.

Drift in O₂ measurements over time is the main issue for long-term deployments (e.g., 1-year). The drift is principally due to: (1) degradation, (2) marine biofouling, and (3) time-dependent pressure hysteresis of the sensing element (only on

very deep moorings >1000 m). In particular, biofouling may rapidly degrade the quality of O₂ and CTD data collected at shallow depths (<200 m).

Due to the efforts of the FIXEd-point Open Ocean Observatories (FIXO³) and European Multidisciplinary Seafloor and Water-Column Observatory (EMSO) European Research Infrastructure Consortium (ERIC) communities some recommendations have been proposed as best practices for O₂ sensors installed on fixed moorings (Coppola et al., 2016). The recommendations include conducting an intercomparison at sea with Winkler data after mooring line recovery. This intercomparison allows O₂ data from the mooring to be adjusted using slope and offset coefficients.

Benthic Platforms

Numerous benthic O₂ measurements have been performed over the last 40 years. They were generally based on single sampling or deployments in the water column and sediment conducted during cruises (Tengberg et al., 1995). Benthic observation platforms were originally based on landers which carry instrumentation for measuring exchange fluxes between the sediment and the water column by benthic chambers or O₂ distribution in sediment porewaters using micro-electrodes. The basic principle of these benthic chambers is the enclosure of a known surface area of sediment and its incubation over a short time to estimate exchange fluxes (Jahnke and Christiansen, 1989; Tengberg et al., 1995; Berelson et al., 2019). O₂ concentration measurements in the benthic chambers are nowadays mostly based on optode measurements in the chamber (see Hall et al.,

2017; Staudinger et al., 2018). These exchange fluxes offer a unique method to describe the metabolic activity in sediments.

Observation of O₂ in BBL and sediments was achieved by measuring O₂ distribution across the water-sediment interface using *in situ* microsensors (Tengberg et al., 1995; Wenzhöfer and Glud, 2002; Glud, 2008; Cathalot et al., 2010). The microsensors measured O₂ concentration in the BBL as a forcing variable or external control, but very few measurement campaigns were dedicated to the measurement of O₂ in the BBL except specific instruments such as the benthic boundary sampler (Holtappels et al., 2011). Recently, nanolanders for *in situ* monitoring of O₂ in the BBL and benthic fauna behavior in O₂-limited zones (Gallo et al., 2020) were developed. Among other things, these new types of observation allow a better understanding of temporal O₂ dynamics in hypoxic bottom waters and its impact on benthic fauna.

In addition, a new observation system has been developed during the last decade based on eddy covariance techniques (Berg and Hüttl, 2008; Reimers et al., 2012). In this technique, joint high frequency measurement of O₂ (>10 Hz) and of vertical currents at a given point in BBL by an acoustic doppler velocimeter (ADV) enables continuous estimates of the O₂ exchange flux and O₂ dynamics within the BBL. The technique is also applicable in habitats with hard and complex substrate in coastal or deep-ocean settings (Berg et al., 2009; Rovelli et al., 2015; Attard et al., 2019; Polsenaere et al., 2021).

More recently, mobile and fixed platforms were developed in order to capture temporal variations linked to intense events (deep mixing by convection, storms and floods in the coastal ocean). Single-point benthic observatories measure O₂ concentration and various other environmental characteristics in the BBL (e.g., S, T, turbidity) in the deep-sea (EMSO, NEPTUNE) or the coastal sea (Pairaud et al., 2016). At these coastal or deep-sea observatories, time series of sediment O₂ distributions based on micro-electrode profiles are acquired by benthic stations in the coastal ocean (Toussaint et al., 2014; Rigaud et al., 2018). Furthermore, mobile platforms called crawlers or rovers (Wenzhöfer et al., 2016; Smith et al., 2017; Lemburg et al., 2018) were recently developed and deployed to explore the spatial heterogeneity inherent in the benthic realm and to study its temporal variation. This enables investigations of the spatio-temporal O₂ dynamics in the BBL and porewaters for one year (or more) (Purser et al., 2013).

CTDs with O₂ sensors are routinely mounted on remotely operated vehicles (ROVs) and human occupied vehicles (HOVs) conducting benthic research. Single dives typically provide vertical profiles made during descent and ascent and extended measurements within 2 m of the sea floor during bottom operations. The data may be accessed through operating agencies (e.g., NOAA⁴) or through PIs, but are not always accessible or in databases.

The availability in databases of O₂ concentrations in the BBL and in sediments is limited and not optimally organized. Pioneering papers (CARS, 2009; Jahnke, 1996) proposed a synthesis of the limited dataset of benthic O₂ demand collected

at that time which contains indications of O₂ concentration in bottom waters. Some regional syntheses were conducted using basin scale compilations of O₂ demands (e.g., South Atlantic; Wenzhöfer and Glud, 2002). More recently, a compilation based on 230 papers containing a collection of total and diffusive O₂ uptakes was achieved and made publicly available (Stratmann et al., 2019) from published and unpublished data (Pangaea data publisher⁵). None of these data collections, however, comprises an exhaustive compilation of O₂ in the BBL and in the sediments. This work remains to be completed.

Concerning common (or best) practices, most calibrations ultimately rely on sensor readings in 100 and 0% O₂ solutions prior to deployment most often combined with Winkler O₂ titration on recovered water samples, some long-term moorings rely on measurements of O₂ concentration using calibrated optodes throughout the deployment, but no consensus on best practices has been established for the bottom part of the ocean including sediments.

At a first stage, GO₂DAT will incorporate O₂ concentration data from the water column including the BBL. Accurate O₂ data in the BBL are crucial for the mapping of bottom water hypoxia. CTDs usually stop ~2 m before the bottom and do not sample the BBL. Benthic O₂ fluxes and oxygen concentrations in sediment porewaters will be integrated at a later stage.

Underway Systems

Underway systems are typically ship-based, but can also include instrumented marine mammals. The potential for collecting high temporal resolution data from such platforms is high, but currently there is no real community for underway O₂ measurements (on any platform) and no consensus as to which sensors should be used and what quality control measures should be taken. Ships Of Opportunity (SOOPs) of the European Integrated Carbon Observation System (ICOS) ERIC, which measure surface pCO₂ and contribute to the Surface Ocean CO₂ Observing NETwork (SOCONET, Wanninkhof et al., 2019), are required to perform concurrent surface O₂ measurements for scientific use to be labeled as “Class 1” station (Steinhoff et al., 2019), which currently 11 out of 12 ICOS-Ocean SOOP lines have. They typically use O₂ optodes in a flow through system. However, data processing and quality control is not yet standardized by SOOPs. To this end, Becker et al. (2021) present a setup that allows regular in-air measurements in such underway installations on SOOPs, thus allowing continuous drift correction and drastically improving data accuracy. From a preliminary assessment with only a few transects, they estimate an accuracy around 3% with an uncertainty (i.e., the standard deviation of the measured O₂ concentration compared to a standard reference value) of 0.26%. Bailleul et al. (2015) added an optode sensor 4330F to Argos CTD-SRDL (i.e., Depth-Satellite Relayed Data Loggers) tags deployed on female elephant seals in the Southern Ocean as a way to both increase the O₂ database in the Southern Ocean and to investigate the relationship between air-breathing marine predators and other components of marine ecosystems. O₂ pop-up satellite archival tags (DO-PATs) have been deployed

⁴<https://www.ncei.noaa.gov/maps/oer-digital-atlas/mapsOE.htm>

⁵<https://www.pangaea.de>

on bluntnose six gill sharks to understand their vertical and horizontal distributions (Coffey and Holland, 2015). A MEOP (Marine Mammals exploring the ocean pole to pole) data base⁶ exists but presently only includes pressure, temperature and salinity profiles.

Quality and Calibration of O₂ Data

Data Quality of Winkler O₂ Measurements

Winkler O₂ measurements are the reference for the major range of O₂ concentrations in the oxygenated ocean. Historical (pre-1970) Winkler O₂ measurements have an overall precision of about $\pm 1 \mu\text{mol kg}^{-1}$. In recent years, the use of potentiometric, amperometric, spectro-photometric and photometric O₂ end-point detection methods have improved the quality of the titrations because of greater automation. Winkler O₂ measurements have an estimated accuracy of 0.1% (Carpenter, 1965); equivalent to approximately $\pm 0.25 \mu\text{mol kg}^{-1}$ and a precision of $0.15 \mu\text{mol kg}^{-1}$ (Saunders, 1986; Langdon, 2010).

However, some compounds commonly present in seawater can modify the Winkler-estimated O₂ concentration. In fully oxic conditions, the most relevant interference is iodate that increases the measured concentration by 1.5 $\mu\text{moles of O}_2$ per $\mu\text{mol of iodate}$. The concentration of iodate in the oceans is relatively stable with values of 0.35–0.45 $\mu\text{mol kg}^{-1}$ in the surface and deep waters, respectively, and lower values in coastal areas (Wong and Cheng, 1998; Chance et al., 2014). Considering these typical iodate concentrations, Winkler measurements overestimate the real O₂ concentration by 0.53–0.68 $\mu\text{mol O}_2 \text{ kg}^{-1}$, which could be a significant error in OMZs, if one wants to estimate the air-sea O₂ fluxes (Wong and Li, 2009) or determine NCP from O₂ trends (Yang et al., 2017). In addition, samples can be contaminated by exposure to the atmosphere during sample handling and from polymers, present even in the widely used Niskin bottle (Broenkow and Cline, 1969; Garcia-Robledo et al., 2021). Therefore, the quality of Winkler measurement degrades and overestimations of the *in situ* O₂ concentrations might be produced which can be of significance in OMZ (see **Box 1**). For the building of GO₂DAT, full information on the data collection protocol is required (e.g., type of bottles, atmospheric contamination). This is particularly critical in OMZs where details on the level of NO₂ that can potentially interfere with the measurement of O₂ is also needed.

Data Quality of Sensor O₂ Measurements

Although O₂ sensor technology has advanced further compared to some other biogeochemical sensors, the quality of sensor-based *in situ* O₂ measurements is still heterogeneous and differs, depending mainly on the calibration type, calculation method (see **Box 2** for an example), and possibility of frequent recalibration. Proper use, calibration, response time and drift remain challenges.

The uncertainty (i.e., the standard deviation of the measured O₂ concentration compared to a standard reference value) on O₂ sensor-based data differs depending mainly on the calibration type, calculation method and possibility of frequent recalibration

through comparison with Winkler measurements or in-air calibration (e.g., BGC Argo). **Table 4** shows that the uncertainty associated to the last generation of optical sensors that uses a community-adopted calibration and calculation method has been reduced by a factor of $\sim 2\text{--}3$ compared to the previous generation.

O₂ sensors are generally affected by three main types of drift: (1) an O₂ sensitivity drift when not deployed and stored in air (i.e., the “storage” drift), (2) an “*in situ*” drift when submerged in the water and (3) a drift in the pressure correction visible on deep moorings ($>1000 \text{ m}$). The storage regime drift is the most problematic with a loss of O₂ sensitivity that can reach $\sim 5\% \text{ yr}^{-1}$ and requires a re-calibration of the sensor before its deployment (see **Box 2**). The *in situ* drift amounts $\sim 0.5\% \text{ yr}^{-1}$ and requires frequent recalibrations and adjustments. For instance, best practices for electrochemical sensors (e.g., SBE43) recommend calibration of the sensor daily with Winkler O₂ data acquired from surface to deep waters (Coppola et al., 2016). A least-square method (Uchida et al., 2010) is then used to adjust the coefficients of the empirical equation relating O₂ voltage and temperature to O₂ concentrations. These frequent recalibrations are particularly critical for autonomous platforms with long time series and/or without recovery like Argo floats. In that case, climatological surface O₂ data can be employed for crude correction (Takeshita et al., 2013), and the advent of optodes with in-air measurements provides a robust approach to address *in situ* drift correction, offering a routine calibration of the sensors (Bittig and Körtzinger, 2015). In addition, oxygen optodes can show a time-dependent lag in response to a change in O₂, which affects data accuracy when sensors experience O₂ gradients, e.g., on profiling platforms. Methods to characterize the response time and to correct for the sensor lag have been described in the literature (Bittig et al., 2014; Bittig and Körtzinger, 2017; Gordon et al., 2020). The time response of the sensors is a function of the membrane thickness, temperature, and flow. For instance, for the most common O₂ sensor used in CTDs, the Sea-Bird SBE-43, it may vary from a few to several tens of second. The effect of time-dependent pressure hysteresis is serious for O₂ sensors deployed at deeper depths as long-term change in the O₂ concentration in the deep ocean might be smaller than the magnitude of the pressure hysteresis (e.g., Uchida et al., 2010). The Scientific Committee on Oceanic Research (SCOR) 142 Working Group on “Quality Control Procedures for O₂ and Other Biogeochemical Sensors on Floats and Gliders” produced well-documented guidelines for the calibration of O₂ optodes (e.g., Bittig et al., 2018) that serve notably for the calibration of BGC Argo (e.g., Thierry and Bittig, 2018).

The quality flagging of O₂ data collected by sensors requires the full specification of the sensors, calibration method and equation as well as information on potential drift corrections by the data owner. This will be recommended for the building of GO₂DAT (see section “Roadmap to GO₂DAT”).

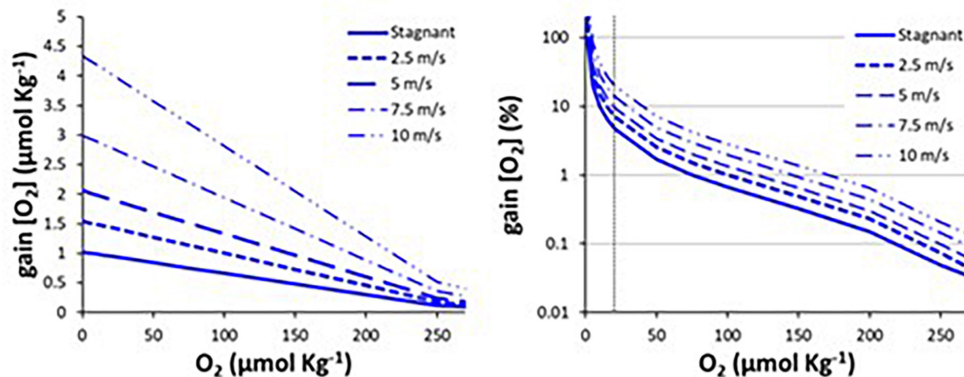
Adequation of O₂ Measurements to Assess Mean State, Weather and Climate Variability

The uncertainty associated with the last generation of O₂ sensors that uses the best calibration and calculation methods amounts, in the best case at $\sim 2 \mu\text{mol kg}^{-1}$ (**Table 4**) which is adequate

⁶<http://www.meop.net/database/>

BOX 1 | Challenges of measuring low O_2 values.

Anoxic samples collected in the core of the OMZs and subpycnocline waters of some coastal systems have the maximum concentration difference with the atmosphere or with O_2 permeable materials such as many polymers commonly used in scientific equipment (e.g., silicone, PVC, rubber, nylon). The contact of the sample with these compounds results in the increase of the O_2 concentration from the initially anoxic sample to values up to $1 \mu\text{mol kg}^{-1}$ just by the Niskin collection (García-Robledo et al., 2021). In addition, samples are exposed to the atmosphere during the filling process and the waiting time until reagents are added and the bottle is finally closed, allowing certain equilibration with the atmosphere. As a consequence, in under-saturated waters, atmospheric O_2 dissolves in the sample and increases the measured concentration up to a few $\mu\text{mol kg}^{-1}$.



The above figure shows the estimation of the O_2 contamination from the atmosphere calculated as a function of the initial sample concentration (x-axis) and wind speed on deck (or sailing speed if there is no wind, in m/s), considering an oceanic sample at 10°C being collected on a 160 ml bottle with an opening diameter of 2.4 cm. The extra O_2 dissolved in the sample is also used to calculate the gain in percentage of the initial O_2 in the sample (right panel). A dotted line at $20 \mu\text{mol kg}^{-1}$ is drawn to illustrate the O_2 range in the OMZs. Note the logarithmic scale in the right panel. The contamination of O_2 introduced only by the atmosphere could be as high as ca. $4.5 \mu\text{mol kg}^{-1}$.

Some compounds interfere with the Winkler titration. At low O_2 concentrations, these interferences gain relevance due to the lower O_2 measured and the increase in some of these compounds. Iodate is consumed in anoxic environments, but at low O_2 conditions such as the oxyclines of OMZs, it can increase the measured concentrations by $0.5\text{--}0.6 \mu\text{mol kg}^{-1}$. Below the oxycline, nitrite usually accumulates in truly anoxic conditions such as the anoxic core of the Pacific OMZs. However, nitrite also interferes with Winkler titration, increasing the apparent O_2 concentration by $0.5 \mu\text{moles per } \mu\text{mole of nitrite}$. Considering that nitrite concentrations up to $10 \mu\text{mol kg}^{-1}$ can be measured in the Pacific OMZs, the deviation from anoxic values can be as high as $5 \mu\text{mol kg}^{-1}$ excluding other sources of error. If the contamination with atmospheric O_2 is taken into account, a variable apparent O_2 concentration of ca. $10 \mu\text{mol kg}^{-1}$ could be measured in samples collected in anoxic environments. Although extreme precautions can be taken to avoid atmospheric O_2 contamination (Broenkow and Cline, 1969), other chemical compounds must be also quantified to correct the final O_2 concentration estimated by Winkler titration. Due to the complexity of such corrections and the likely lack of complementary data from historical databases, Winkler determinations should not be used to correct *in situ* sensors in waters with $[O_2] < 15\text{--}20 \mu\text{mol kg}^{-1}$ (estimated error $\sim 10\%$). In such cases a zero-calibration is recommended.

to answer questions on the mean state and weather variability but is still too large to resolve climate variability (Table 2). On the other hand, Winkler O_2 data have the level of accuracy and precision needed to assess the long-term climate deoxygenation signal. Yet, these data do not have the quality, spatial and temporal coverage needed to infer the long-term trend in OMZs spatial extension and to concurrently correct sensor data in these regions. A *posteriori* correction or calibration of historical O_2 data requires information on the sampling (e.g., contamination, concentrations of other compounds, calibration procedures, sampling protocol) that is very often not available. The rapid evolution over the last decades of sensor technology and calibration methods makes the application of a correction scheme derived from current instruments to past measurements difficult.

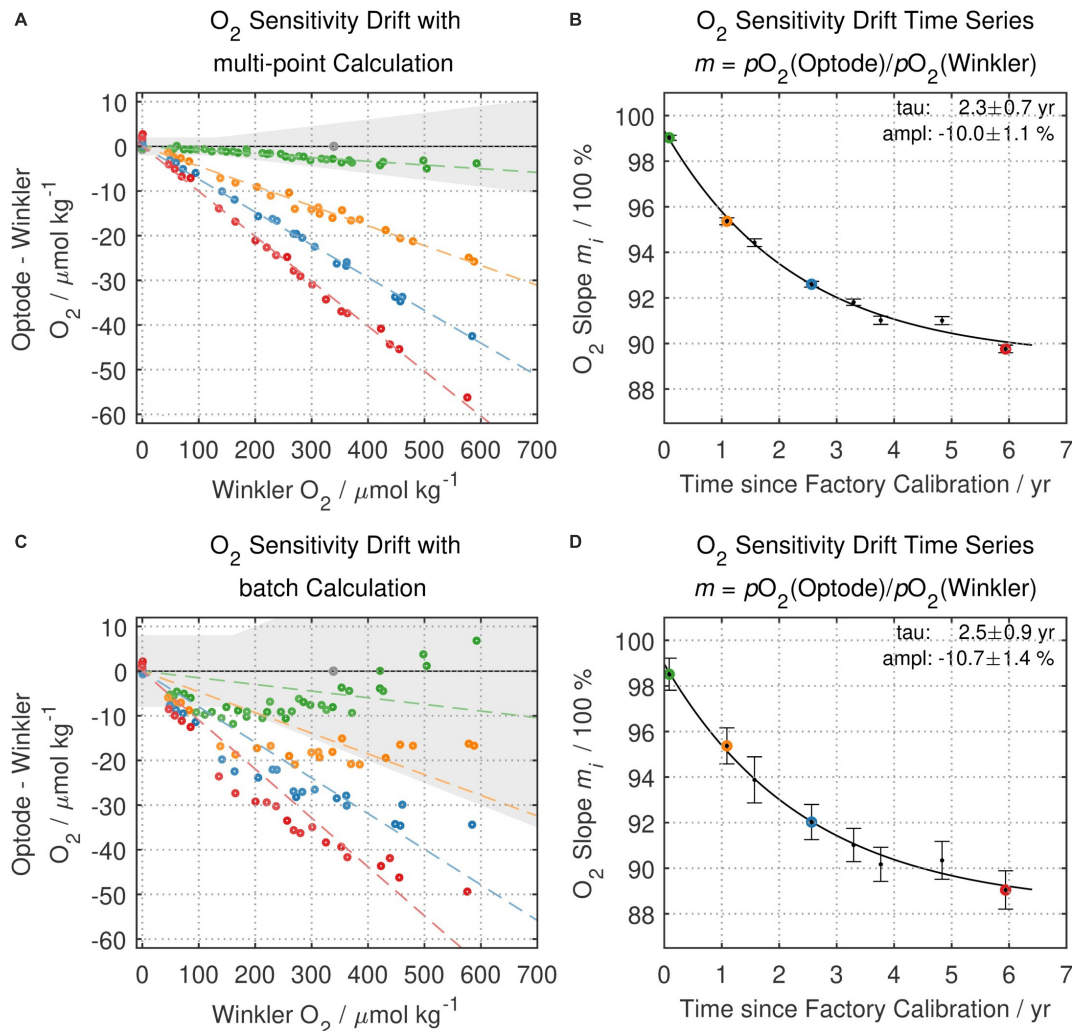
Second Order Adjustment of O_2 Data

A *posteriori* correction can also be performed to increase the internal consistency of diverse datasets collected over several decades by different methodologies and laboratories. This secondary correction will not improve the accuracy of the data but rather is expected to increase the internal consistency of

the datasets (e.g., Key et al., 2004) for e.g., the building of data synthesis products. In the deep ocean (~ 2000 m), a common procedure consists of correcting sensors' offsets by comparing their measured values with those collected by Winkler (e.g., Winkler-based climatology, data taken years apart), assuming that the level of O_2 remains unchanged in that part of the ocean. For instance, GLODAP (Key et al., 2004; Olsen et al., 2016) uses a crossover analysis to determine cruise-by-cruise offsets for discrete bottle data (Winkler) collected since 1972. The comparison is made irrespective of the year and season of data collection. Identified offsets larger than 1% are then adjusted after expert evaluation of all available information, including known climate trends in O_2 and water mass structure (Tanhua et al., 2010; Lauvset and Tanhua, 2015; Olsen et al., 2020). Significant O_2 and temperature warming trends have been detected even below 2000 m depth (e.g., Desbruyères et al., 2016; Schmidtke et al., 2017). For instance, Desbruyères et al. (2016) reported long-term (1991–2010) global warming trends at all pressure levels below 2000 m with the strongest warming rates in the abyssal layer (4000–6000 m). Even if the changes in O_2 associated with this warming is likely less than 1%, it requires differentiating sensor drift from climate variability.

BOX 2 | O₂ response drift in optical and electrochemical sensors.

An O₂ sensitivity drift has been reported frequently between calibration by the manufacturer and deployment in the field (i.e., the “storage” drift, order $-5\% \text{ yr}^{-1}$, Bittig et al., 2018). This leads to a significant deviation of the stability properties compared to those found during the factory/laboratory calibrations (e.g., Bittig et al., 2012; Takeshita et al., 2013). Bittig and Körtzinger (2015) describe the nature of the storage drift as systematic, linear with O₂ level and towards underestimation of actual O₂.



The above figure shows the characterization of the O₂ sensitivity storage drift using as an example a subset of calibration data from an optode (Aanderaa optode model 3835) with factory two-point/batch calibration (in November 2010), multi-point calibrated directly afterward (December 2010) and, thereafter, multi-point recalibrated in approximately yearly intervals. All data courtesy of Craig Neill (CSIRO Oceans & Atmosphere, Hobart). **(A)** Difference between optode data and Winkler reference data using the multi-point calculation, with the factory two-point data (gray circles) as initial state. Dashed lines are the slope regression forced through the origin and the different colors correspond to different time since factory calibration shown in panel **(B)**. Only a subset of recalibrations (colored circles) is shown for clarity. **(B)** Temporal evolution of the O₂ slope [i.e., $p\text{O}_2(\text{Optode})/p\text{O}_2(\text{Winkler})$] with colors corresponding to calibrations shown in panel **(A)**. **(C)** Same data as in **(A)** but treated with the batch-foil/2-point adjustment calibration calculation. The linear O₂ sensitivity drift appears non-linear with the batch-foil calculation. A slope O₂ regression (dashed lines) still shows a drift towards lower O₂, however, slopes are dependent on at which O₂ levels reference data are available. **(D)** Same as **(B)** but for batch-foil calculation. The above figure illustrates the potentially biasing effect of calibration method on the appearance of storage drift. **(A)** Shows that in the case of the multi-point calibration the O₂ sensitive bias can be modelled (through regression) at each calibration time (i.e., from 0 to 7 years since factory calibration) as a linear function of the O₂ going through the origin with a negative slope. In the case of using the same, multi-point data but with the batch foil calibration approach with a two-point correction (e.g., 0 and 100 % O₂ saturation), the O₂ sensitivity drift appears non-linear but still a regression line indicates a drift towards lower O₂ **(C)**. However, slopes are dependent on which O₂ levels reference data are available. This is where a large portion of the added uncertainty on batch-foil calibrated optodes originates (Table 4).

Bittig et al. (2018) propose a parameterization of the drift as a slope effect (i.e., the loss of O₂ sensitivity defined as $p\text{O}_2(\text{optode})/p\text{O}_2(\text{Winkler})$) resulting from reduced O₂ sensitivity with a small positive offset coming from a reduced luminophore lifetime. **(B)** Shows that the storage drift rate (i.e., the slope effect) decreases with age of the sensor, being strongest in new optodes, of the order of $5\% \text{ yr}^{-1}$, and decreasing in more mature optodes. Therefore, *in-situ* calibration before or during deployment is crucial and can be done either using Winkler-O₂ determination on a cast taken at the same time and place of the sensor's deployment or, in the case of Argo floats equipped with optode in-air measurements, using near surface and in-air measurements (Bittig and Körtzinger, 2015; Johnson et al., 2015).

TABLE 4 | Calibration type and calculation method for sensors and platforms.

Calibration type and calculation method	Examples for sensors	Examples for platforms	Uncertainty
Electrochemical sensors with 2-point calibration	SBE43	Shipboard CTD, 1st category of sensors used on Argo Floats, Moorings, Benthic Observations, Gliders	2–10 $\mu\text{mol kg}^{-1}$
Batch-foil calibration; polynomial with 2-point adjustment	Aanderaa 3830/3835, all Aanderaa 4330/4831 produced until 2013, some Aanderaa 4330/4831 produced 2013–2018	Moorings, Benthic Observations, Gliders, 2nd category of sensors used on Argo Floats.	6–25 $\mu\text{mol kg}^{-1}$
Multi-point calibration; SVU/Uchida et al., 2010-like equation	Aanderaa 4330/4831 since 2018, SBE63, RINKO ARO-FT	Moorings, Benthic Observations, Gliders, 3rd category of sensors used on Argo Floats.	2–10 $\mu\text{mol kg}^{-1}$

Measurement uncertainty is defined here as the standard deviation of the measured O_2 concentration compared to a standard reference value (mostly Winkler).

Shallower coastal regions are subject to higher variability than deep, open-ocean regions, which further complicates the application of adjustments. Deep reference values cannot be used for data calibration in the shallower coastal areas, and thus, targeted data calibration approaches and QC procedures are needed there.

Mapping O_2 : Technical Challenges

Some of the most common objective mapping procedures used in ocean sciences are the Barnes scheme (Barnes, 1964) which is used to create WOA18 and GLODAPv1.1, and variational inverse analysis (such as DIVA – data interpolating variational analysis; Troupin et al., 2012; Barth et al., 2014), which is used to create GLODAPv2. There are many objective mapping methods, and each has advantages and disadvantages depending on the data product requirements, the availability of high-quality data, behavior of the mapping method around topographic features (e.g., islands, trenches, seamounts), the ability to take ocean currents into account, and other factors. Mapping is commonly done on depth/pressure surfaces, but mapping on isopycnal (density) surfaces is also useful where possible. There are several technical challenges for creating mapped O_2 data products, including the need to merge several highly diverse data sources with different sampling frequencies in time and space. Additionally, there are issues specifically related to ocean biogeochemical variables and, in particular, O_2 , such as the heterogeneous data distribution in time and space, data quality, and the existence of Anoxic Marine Zones (AMZs) (i.e., $\text{O}_2 = 0 \mu\text{mol kg}^{-1}$) and OMZs, which present several challenges including the scarcity of data, high gradients and ultra-low O_2 values (e.g., Bianchi et al., 2012). The global coastal ocean presents another challenge given its relatively higher variability when compared to the open ocean. It is therefore likely that mapped data products for the global open and coastal ocean will have to sometimes rely on auxiliary data (such as temperature and salinity) and machine learning methods (such as neural networks, e.g., Krasnopolsky et al., 2016; Barth et al., 2020).

Existing and commonly used mapped data products include GLODAPv1.1 (Key et al., 2004), GLODAPv2 (Lauvset et al., 2016), WOA18 (Garcia et al., 2019), CSIRO Atlas of Regional

Seas (CARS, 2009), and Global Nutrients Dataset 2013 (GND13; Aoyama, 2020). All these data products are examples of global three-dimensional (longitude, latitude, and depth) fields of O_2 and other variables, with varying temporal resolution (monthly to annual averages), created using various forms of objective mapping procedures. In addition, there exist several regional mapped data products such as EMODNet Chemistry.

ROADMAP TO GO_2DAT

The WOD (Garcia et al., 2005, 2019), CMEMS (Le Traon et al., 2019), GLODAP (Key et al., 2004; Olsen et al., 2016, 2020), and EMODnet (Giorgetti et al., 2018) are excellent foundations to support the development of GO_2DAT . Data collected by Argo, gliders and moorings have their own GDACs, which offers a certain level of consistency in terms of metadata specifications, QC, QF, computation models of sensors response (i.e., from raw data to O_2 value), calibration and adjustment procedures. Best practice documents are available for sensor calibration (e.g., see Table 4) and the data are delivered according to the FAIR principles in RT and DM streams. However, these different databases and GDACs have their specific standards and, inside a given platform, these standards may differ across regional DACs. A GO_2DAT steering committee will be established and will work with existing GDACs and the IOC UNESCO International Oceanographic Data and Information Exchange program (IODE) towards the definition of common best practices and an alignment between GDACs of metadata structure, QC and QF procedures as concerns O_2 . These community agreed best practices will be communicated to data providers and repositories via the OceanTeacher network. GO_2DAT will have its own O_2 GDAC (i.e., GO_2DAT -GDAC), approved by IODE and complying with the terms of references of a, IODE-approved GDAC (see https://www.iode.org/index.php?option=com_content&view=article&id=372&Itemid=100088). For instance, GO_2DAT -GDAC will receive and assemble marine O_2 data and metadata from the data streams described in section “ GO_2DAT data flow,” check their consistency, identify duplicates, make sure that the data are quality controlled according to the GO_2DAT -GDAC standards and methods, provide feedbacks

to the source of data regarding quality issues, make data accessible and metadata available through the GO₂DAT data portal and to IODE.

GO₂DAT Data Flow

In order to enable a sustainable data submission system that will integrate and make use of existing infrastructures, it is crucial to align it with overarching data flow and the structural elements of IODE. IODE will offer a seamless connection with its structural elements such as National Oceanographic Data Centres (NODCs), Associated Data Units (ADUs), and GDACS.

The proposed data submission will be two-fold: a centralized data submission to existing GDACS and a bottom-up data flow via NODCs (Figure 8). For platforms that already have their own GDAC (see Table 3), the submission process will remain unchanged (i.e., data will be submitted to the corresponding GDAC, e.g., Argo, gliders, moorings). Otherwise, submission to NODCs will be encouraged. Data providers will nevertheless still have the possibility to directly submit their data to regional hubs (e.g., EMODnet, CMEMS-TAC) and, in that case, data will be instantly exchanged with NODCs where PIDs will be assigned, and data get archived.

At the end, all data will be delivered and archived in the GO₂DAT GDAC. The adjustments performed on each data set at each level should be traceable and well documented. During the whole process, best practices and OceanTeacher will guide data providers in the submission process.

Acknowledgment of data providers will be provided through the assignment of a PID (e.g., DOI) where appropriate credit is given not only to the data provider but also to the project, funding

agency and hosting agencies/institutes. The data usage license should be free and open but requires citation (e.g., Creative Commons BY). Data providers, NODCs, and Associated Data Units (ADUs) should be informed of download statistics.

Annual release of the GO₂DAT data will be publicized and interconnection with international initiatives (e.g., CMEMS, SeaDataNet, GLODAP, WOD) and other related GDACS will be recommended. Data will be received and made available through the World Data Service for Oceanography of the World Data System.

Table 5 summarizes the fundamental principles of GO₂DAT.

Definition of Community-Approved GO₂DAT Standards Metadata Specifications

Understanding O₂ variability and change requires information on the environmental conditions associated with each measurement. For instance, temperature and salinity are essential variables to estimate the density and O₂ solubility of a water parcel allowing differentiation of physical and biogeochemical contributions to the O₂ dynamics and trends. Temperature and salinity are also needed to convert observed O₂ concentration into partial pressure (pO₂), a meaningful quantity when assessing the impact on marine life (e.g., Seibel and Deutsch, 2020). We recommend that raw O₂ data be provided with information on temperature, salinity and pressure. Data will be delivered on a per mass basis in order to avoid the temperature and pressure effect on the volume. A clear and comprehensive definition of metadata specifications needs to be agreed upon by the scientific community. It will be recommended that NODCs, regional

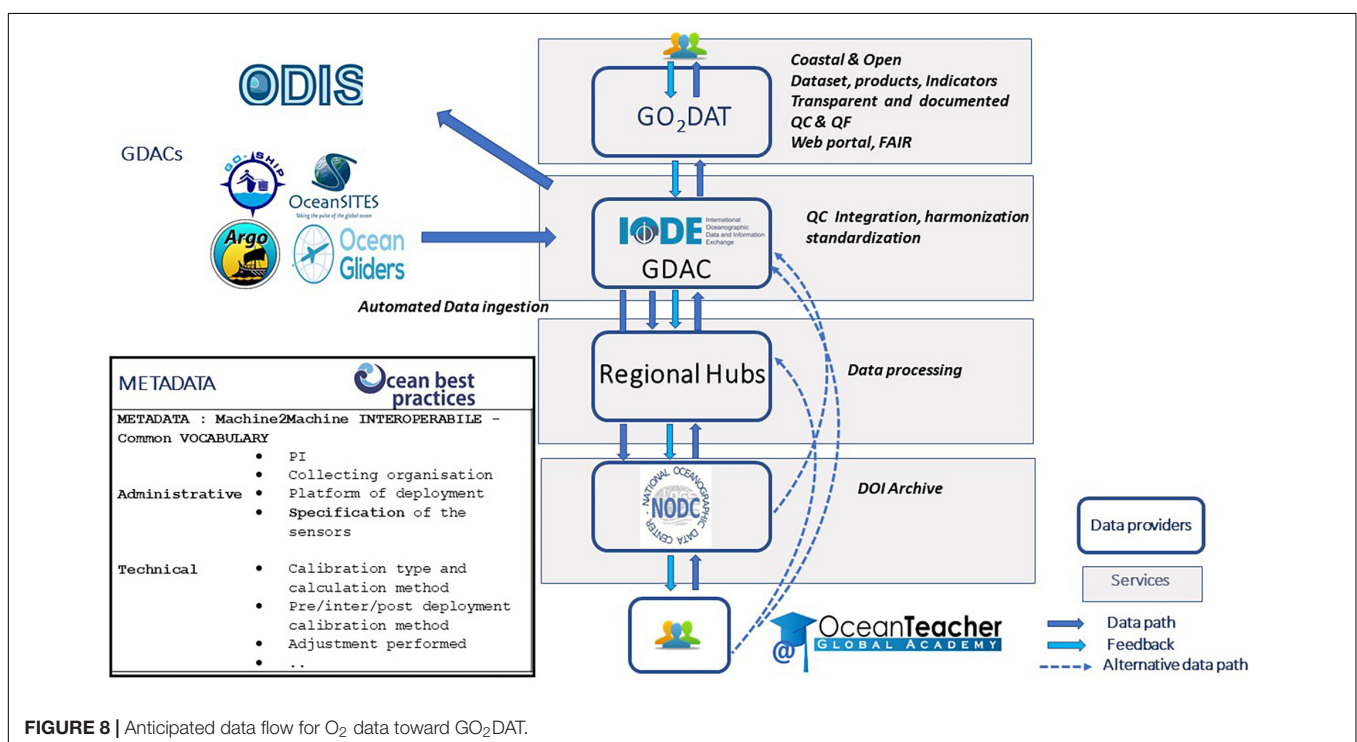


TABLE 5 | GO₂DAT principles.

Attribute for raw data	Requirements
Access	Open-access without data use restrictions under Creative Commons.
Cost to use data and products	Free
Data and product attribution	PID, DOI
Archival of primary reference	All data must be available at an international long-term archive with long term funding (e.g., World Data Service for Oceanography)
Sustainability	Long-term to ensure reproducibility of the products
Data versioning	Metadata best practices
Reference QC and QF	Community consensus
Database structure	Common digital format (e.g., netCDF, ASCII), units, and searchable/download online.
Data products	Available online without restriction in common format (e.g., netCDF)
Variables	EOV, (ECV, EBV)
Reference	Depth (pressure), density
Other...	

hubs, GDACs map their existing standards to the agreed GO₂DAT standard.

A Fully Documented and Transparent Quality Flagging System

GO₂DAT will ensure that data in each flagging level are assigned an uncertainty, and that sufficient metadata to interpret this uncertainty are included. This will facilitate understanding of data uncertainty by the users allowing them to decide whether these data fit their specific requirements. Data correction factors and flagging should be fully traceable, consistent and based on a community agreement between groups of experts defined for the different sensors and platforms. Fully documented, transparent flagging procedure will allow any user to understand the details of a particular QF assigned to a data value and to have sufficient information to assess the suitability of the data for a particular purpose (e.g., mean state, variability, climate trend assessment). The questions listed in **Table 2** can serve as a base to define the QF system. In the coastal ocean, the quality of data may vary greatly from very accurate data taken according to the best protocols to low quality data taken with hand-held, air-calibrated meters (e.g., YSI O₂ meter). Even if of lower quality, such datasets should not be rejected from GO₂DAT, but should be appropriately flagged. Some important applications of O₂ data in coastal systems (e.g., protection of fisheries species) do not require the accuracy and precision required in open-ocean climate studies.

Integrating Coastal Ocean Data in GO₂DAT

Whereas open data sharing is generally accepted for open-ocean monitoring, this concept needs further support in coastal communities and, particularly, from NODCs and national and local organizations funding the monitoring programs. The compilation of data for the coastal ocean will be facilitated by the establishment of regional hubs adhering to the GO₂DAT- best

practices. The connection of these regional hubs and GO₂DAT will be realized through international and regional initiatives. For instance, the Chesapeake Bay Program⁷ and, the Baltic Environmental Database⁸ have been instrumental in data sharing, thereby supporting studies of hypoxia at a larger scale (e.g., Hagy et al., 2004; Conley et al., 2009; Carstensen et al., 2014). In the Pacific region, connection with WESPAC-O₂NE (WESTern PACific-Oxygen Network) will promote the sharing of data in a region where the risks of eutrophication and hypoxia are likely to continue to intensify due to the projected increase of discharges of inorganic nitrogen and phosphorus over the next decades (Seitzinger et al., 2010). In the North Pacific, GO₂DAT will benefit from the efforts led by the North Pacific Marine Science Organization (PICES) that is planning to construct a list of coastal O₂ monitoring stations among their member countries. In the Eastern Pacific, existing initiatives like the California Cooperative Oceanic Fisheries Investigation (CalCOFI) and the Tonga Bay (off Chile) observing system⁹ will provide open access, long-term O₂ data that have stimulated numerous mechanistic studies of regional O₂ trends (e.g., Evans et al., 2020). The building of the research infrastructure JERICO-RI for the observation of the coastal ocean at the EU level will probably provide a regular flow of coastal O₂ data to GO₂DAT.

Despite the existing services to facilitate and streamline the ingestion of data by providers not yet connected to marine data management infrastructures, the success of such services depends on the willingness of data holders to share their data. The existence of GO₂DAT, with rules on publication and attribution of data, may encourage individual researchers as well as government agencies responsible for data collection in coastal systems to participate and increase access to their data.

GO₂DAT Products

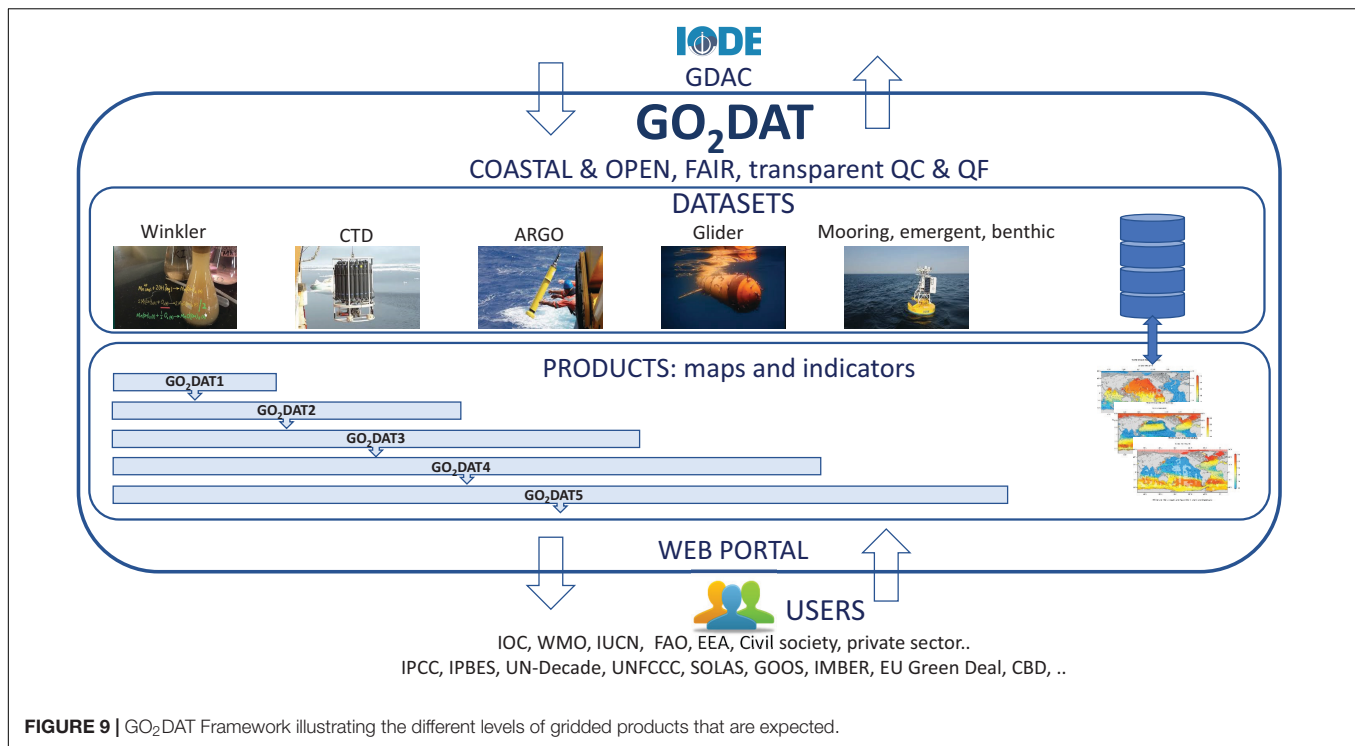
GO₂DAT gridded data products (hereafter referred to as “mapped data”) will be two- or three-dimensional fields of O₂ concentration, pO₂ and degree of saturation for the coastal and global ocean in support of scientific and management purposes. Each GO₂DAT product will be provided with a documented QF and with information about the quality, source and treatment of the data used for their production.

We anticipate providing several choices of mapped O₂ products at different spatial and temporal scales to give users the choice to select the most appropriate for their application. In addition, we will strive to provide information about which methods are appropriate and relevant for the user application. The building of GO₂DAT mapped O₂ products will progress in different iterative steps, as illustrated in **Figure 9**. At the first level, GO₂DAT mapped products will integrate O₂ Winkler observation from the different databases (e.g., WOD, EMODnet, CMEMS) using the consistent community-agreed QC procedure. The second level of mapped O₂ products will include the CTD-O₂ sensor data, after an appropriate QC procedure. The third and fourth levels entail inclusion of O₂ data from Argo-floats

⁷<https://www.chesapeakebay.net/>

⁸<http://nest.su.se/bed/>

⁹<http://www.cdom.cl/equipamiento.php>



and gliders, respectively. The fifth and final level is inclusion of data from moorings and underway systems, as well as BBL data and other emergent data platforms. GO₂DAT will thus make the development of a range of data synthesis products at different depth and density levels possible.

Users will be able to download separate synthesis products for each level, or, when completed, a combined product with all levels included. It is envisioned that the synthesis product will be scalable to accommodate new observing systems and to facilitate continuous updates as new data become available. This is an important and required characteristic to enable frequent updates of products and to serve a wide range of users. GO₂DAT will collaborate closely with the CMEMS *in situ* Thematic Centre, NOAA NCEI and EMODnet Chemistry to avoid duplication of efforts.

While GO₂DAT will focus on providing mapped data products, it will also aid the users wanting to use GO₂DAT datasets for building their own mapped products and develop user-specific tools.

An interactive web-platform where GO₂DAT data and products can be visualized will enhance interactions with the users and general public. This GO₂DAT web interface will be optimized in order to obtain feedback on the interface, datasets and products.

GO₂DAT Human Resources

The development of GO₂DAT will require human resources and engagement from the community including scientists, data managers, data providers and data users. The following tasks have been identified for the building of GO₂DAT to which human resources need to be dedicated.

Definition and documentation in the Ocean Best Practices System of community-agreed GO₂DAT standards for metadata specification, sensors calibration, data quality check and flagging.

Mapping existing data standards (e.g., in NODCs, GDACs) to that agreed in GO₂DAT.

Integration (preferably interoperable) in GO₂DAT GDAC of existing datasets, search for duplicates, application of the agreed GO₂DAT standards.

Building regular updates (e.g., yearly) and analysis of the GO₂DAT mapped products and indicators.

Building of the web interface and interactions with the users.

CONCLUSION

Since the community white paper by Gruber et al. (2010) that called for the addition of O₂ sensors on Argo floats and more generally on autonomous platforms, the number of O₂ sensors has dramatically increased offering unprecedented capabilities to monitor O₂ state and variability in the open, regional and coastal ocean. As the number of profiles increased, the quality of the sensor measurements has also improved. The uncertainty associated to the last generation of sensors has been reduced by a factor of ~3 when the best calibration and calculation method are used. This offers new opportunities to combine sensors and Winkler data in consistent data-synthesis products. These products will be used to understand the O₂ dynamics and the deoxygenation process in the open and global coastal ocean in response to climate change.

The building of GO₂DAT is a timely initiative that can build on WOD, EMODnet, CMEMS, GLODAP. The integration of sensors and Winkler data from eulerian and lagrangian platforms from the different databases would offer data-synthesis products at an increased resolution. For the coastal ocean, the necessity to support the decision-making process aiming at preserving ocean health requires a coordination of NODCs, (inter)national and local organizations via the establishment of regional hubs. GO₂DAT will offer the opportunity to build an O₂ gridded product for the global coastal ocean with regular updates to monitor the multiannual evolution of coastal hypoxia in a warming climate.

The building of GO₂DAT would allow to fully harness the potential of the boost in O₂ profiles that is expected to quadruple in the frame of the future GOOS strategy. The alignment of well-documented and consistent QC and QF procedures proposed in GO₂DAT will allow the user to make an informed choice on which data that are fit for purpose. GO₂DAT will facilitate the dissemination of information on ocean deoxygenation and its impact for marine life to a wide community of stakeholders (e.g., policymakers, industries, see **Table 1**) and contribute to the education of the young generation and general public.

The development of GO₂DAT requires human resources and engagement from the scientific community, data providers, data managers and end-users. Engagement around GO₂DAT will be promoted by the UN Decade GOOD program.

AUTHOR CONTRIBUTIONS

MG, VG, HG, DB, KI, AO, and MT organized the workshops. MG, VG, and HG designed and wrote the manuscript with contributions from all co-authors. All the authors discussed and reviewed the manuscript.

REFERENCES

- Adams, K. A., Barth, J. A., and Shearman, R. K. (2016). Intraseasonal cross-shelf variability of hypoxia along the newport, oregon, hydrographic line. *J. Phys. Oceanogr.* 46, 2219–2238. doi: 10.1175/JPO-D-15-0119.1
- Altieri, A. H., Harrison, S. B., Seemann, J., Collin, R., Diaz, R. J., and Knowlton, N. (2017). Tropical dead zones and mass mortalities on coral reefs. *Proc. Natl. Acad. Sci.* 114, 3660–3665. doi: 10.1073/pnas.1621517114
- Andersen, J. H., Axe, P., Backer, H., Carstensen, J., Claussen, U., Fleming-Lehtinen, V., et al. (2011). Getting the measure of eutrophication in the Baltic Sea: towards improved assessment principles and methods. *Biogeochemistry* 106, 137–156. doi: 10.1007/s10533-010-9508-4
- Anderson, O. F., Guinotte, J. M., Rowden, A. A., Clark, M. R., Mormede, S., Davies, A. J., et al. (2016). Field validation of habitat suitability models for vulnerable marine ecosystems in the South Pacific Ocean: implications for the use of broad-scale models in fisheries management. *Ocean Coast. Manag.* 120, 110–126. doi: 10.1016/j.ocecoaman.2015.11.025
- Andrews, O., Buitenhuis, E., Le Quéré, C., and Suntharalingam, P. (2017). Biogeochemical modelling of dissolved oxygen in a changing ocean. *Philos. Trans. Royal Soc. A* 375:20160328. doi: 10.1098/rsta.2016.0328

FUNDING

All authors would like to thank IOC-UNESCO, International Ocean Carbon Coordination Project (IOCCP), NOAA, and the German SFB754. MG is funded by the Fonds National de la Recherche Scientifique (FRS-FNRS) and received fundings from the FNRS BENTHOX program grant T.1009.15, the Copernicus Marine Service (CMEMS), and the European Union's Horizon 2020 BRIDGE-BS project under grant agreement No. 101000240. MG, VG, KI, and BDew are supported by the Project CE2COAST funded by ANR (FR), BELSPO (BE), FCT (PT), IZM (LV), MI (IE), MIUR (IT), Rannis (IS), and RCN (NO) through the 2019 “Joint Transnational Call on Next Generation Climate Science in Europe for Oceans” initiated by JPI Climate and JPI Oceans. MT, KC, and VG acknowledge support from the United States National Science Foundation grant OCE-1840868 to the Scientific Committee on Oceanic Research (SCOR, United States). BoD also acknowledges support from ANID grants R20F0008-CEAZA and 1190276. This research (through VG, AP and BoD) received fundings from the European Union's Horizon 2020 Research and Innovation Programme under grant agreement No. 869300 (FutureMARES). CB, AP, VG, LC, BrD, VR, VT, and CS acknowledge support of the French CES ODATIS Oxygen through INSU funding. SKL acknowledges support from the Research Council of Norway (Grant No. 269753). This manuscript is a contribution to the UN Decade Global Ocean Oxygen (GOOD) Program.

ACKNOWLEDGMENTS

This manuscript has been produced as an outcome of a 2-day workshop organized in Sopot in November 2019 by the IOC-UNESCO Global Ocean Oxygen Network (GO₂NE) and sponsored by IOC-UNESCO, the International Ocean Carbon Coordination Project (IOCCP), U.S. NOAA and German SFB754 and of a 2-day virtual workshop in November 2020.

- Aoyama, M. (2020). Global certified-reference-material- or reference-material-scaled nutrient gridded dataset GND13. *Earth Syst. Sci. Data* 12, 487–449. doi: 10.5194/essd-12-487-2020
- Asch, R. G., Cheung, W. W., and Reygondeau, G. (2018). Future marine ecosystem drivers, biodiversity, and fisheries maximum catch potential in Pacific Island countries and territories under climate change. *Mar. Pol.* 88, 285–294. doi: 10.1016/j.marpol.2017.08.015
- Attard, K. M., Rodil, I. F., Glud, R., Berg, P., Norkko, J., and Norkko, A. (2019). Seasonal ecosystem metabolism across shallow benthic habitats measured by aquatic eddy covariance. *Limnol. Oceanogr. Lett.* 4, 79–86. doi: 10.1002/lol2.10107
- Bailleul, F., Vacquie-Garcia, J., and Guinet, C. (2015). Dissolved oxygen sensor in animal-borne instruments: An innovation for monitoring the health of oceans and investigating the functioning of marine ecosystems. *PLoS One* 10:e0132681. doi: 10.1371/journal.pone.0132681
- Barnes, S. L. (1964). A technique for maximizing details in numerical weather map analysis. *J. App. Meteor.* 3, 396–409. doi: 10.1175/1520-0450(1964)003<0396:ATFMDI>2.0.CO;2
- Barth, A., Alvera-Azcárate, A., Licer, M., and Beckers, J.-M. (2020). DINCAE 1.0: a convolutional neural network with error estimates to reconstruct sea

- surface temperature satellite observations. *Geosci. Model. Dev.* 13, 1609–1622. doi: 10.5194/gmd-13-1609-2020
- Barth, A., Beckers, J.-M., Troupin, C., Alvera-Azcárate, A., and Vandenbulcke, L. (2014). Divand-1.0: n-dimensional variational data analysis for ocean observations. *Geosci. Mod. Dev.* 7, 225–241. doi: 10.5194/gmd-7-225-2014
- Batiuk, R. A., Breitburg, D. L., Diaz, R. J., Cronin, T. M., Secor, D. H., and Thursby, G. (2009). Derivation of habitat-specific dissolved oxygen criteria for Chesapeake Bay and its tidal tributaries. *J. Exper. Mar. Biol. Ecol.* 381, S204–S215. doi: 10.1016/j.jembe.2009.07.023
- Beatty, T., Heinze, C., Hughlett, T., and Winguth, A. M. (2017). Response of export production and dissolved oxygen concentrations in oxygen minimum zones to pCO₂ and temperature stabilization scenarios in the biogeochemical model HAMOCC 2.0. *Biogeosciences* 14, 781–797. doi: 10.5194/bg-14-781-2017
- Becker, M., Olsen, A., and Reverdin, G. (2021). In-air one-point calibration of oxygen optodes in underway systems. *Limnol. Oceanogr. Methods* 19, 293–302. doi: 10.1002/lom3.10423
- Berelson, W., McManus, J., Severmann, S., and Rollin, N. (2019). Benthic fluxes from hypoxia-influenced Gulf of Mexico sediments: Impact on bottom water acidification. *Mar. Chem.* 209, 94–106. doi: 10.1016/j.marchem.2019.01.004
- Berg, P., and Hüttel, M. (2008). Monitoring the seafloor using the non-invasive Eddy Correlation technique: Integrated benthic exchange dynamics. *Oceanography* 21, 164–167. doi: 10.5670/oceanog.2008.13
- Berg, P., Glud, R. N., Hume, A., Stahl, H., Oguri, K., Meyer, V., et al. (2009). Eddy correlation measurements of oxygen uptake in deep ocean sediments. *Limnol. Oceanogr.* 7, 576–584. doi: 10.4319/lom.2009.7.576
- Bianchi, D., Dunne, J. P., Sarmiento, J. L., and Galbraith, E. D. (2012). Data-based estimates of suboxia, denitrification, and N₂O production in the ocean and their sensitivities to dissolved O₂. *Glob. Biogeochem. Cycles* 26:2009. doi: 10.1029/2011GB004209
- Bittig, H. C., and Körtzinger, A. (2015). Tackling oxygen optode drift: Near-surface and in-air oxygen optode measurements on a float provide an accurate in situ reference. *J. Atmos. Ocean. Technol.* 32, 1536–1543. doi: 10.1175/JTECH-D-14-00162.1
- Bittig, H. C., and Körtzinger, A. (2017). Technical note: Update on response times, in-air measurements, and in situ drift for oxygen optodes on profiling platforms. *Ocean Sci.* 13, 1–11. doi: 10.5194/os-13-1-2017
- Bittig, H. C., Körtzinger, A., Neill, C., van Ooijen, E., Plant, J. N., Hahn, J., et al. (2018). Oxygen optode sensors: principle, characterization, calibration, and application in the ocean. *Front. Mar. Sci.* 4:429. doi: 10.3389/fmars.2017.00429
- Bittig, H., Fiedler, B., Scholz, R., Krahmann, G., and Körtzinger, A. (2014). Time response of oxygen optodes on profiling platforms and its dependence on flow speed and temperature. *Limnol. Oceanogr. Methods* 12, 617–636. doi: 10.4319/lom.2014.12.617
- Bittig, H., Fiedler, B., Steinhoff, T., and Körtzinger, A. (2012). A novel electrochemical calibration setup for oxygen sensors and its use for the stability assessment of Aanderaa optodes. *Limnol. Oceanogr. Methods* 10, 921–933. doi: 10.4319/lom.2012.10.921
- Bittig, H., Körtzinger, A., Johnson, K., Claustre, H., Emerson, S., Fennel, K., et al. (2015). SCOR WG 142: Quality Control Procedures for Oxygen and Other Biogeochemical Sensors on Floats and Gliders. Recommendation for Oxygen Measurements From Argo Floats, Implementation of In-Air-Measurement Routine to Assure Highest Long-Term Accuracy. Plouzane, France: IFREMER.
- Bittig, H., Maurer, T., Plant, J., Schmechtig, C., Wong, A., Claustre, H., et al. (2019). A BGC-argo guide: planning, deployment, data handling and usage. *Front. Mar. Sci.* 6:502. doi: 10.3389/fmars.2019.00502
- Boyer, T. P., Baranova, O. K., Coleman, C., Garcia, H. E., Grodsky, A., Locarnini, R. A., et al. (2018). “World ocean database 2018,” in NOAA Atlas NESDIS, ed. A. V. Mishonov (Silver Spring, MD: NOAA), 87.
- Bopp, L., Resplandy, L., Orr, J. C., Doney, S. C., Dunne, J. P., Gehlen, M., et al. (2013). Multiple stressors of ocean ecosystems in the 21st century: projections with CMIP5 models. *Biogeosciences* 10, 6225–6245.
- Brasseur, P., Gruber, N., Barciela, R., Brander, K., Doron, M., Elmoussaoui, A., et al. (2009). Integrating biogeochemistry and ecology into ocean data assimilation systems. *Oceanography* 22, 206–215. doi: 10.5670/oceanog.2009.80
- Breitburg, D. (2002). Effects of hypoxia, and the balance between hypoxia and enrichment, on coastal fishes and fisheries. *Estuaries* 25, 767–781. doi: 10.1007/BF02804904
- Breitburg, D. L., Craig, J. K., Fulford, R. S., Rose, K. A., Boynton, W. R., Brady, D. C., et al. (2009). Nutrient enrichment and fisheries exploitation: interactive effects on estuarine living resources and their management. *Hydrobiologia* 629, 31–47. doi: 10.1007/s10750-009-9762-4
- Breitburg, D., Levin, L. A., Oschlies, A., Grégoire, M., Chavez, F. P., Conley, D. J., et al. (2018). Declining oxygen in the global ocean and coastal waters. *Science* 359:6371. doi: 10.1126/science.aam7240
- Broenkow, W. W., and Cline, J. D. (1969). Colorimetric determination of dissolved oxygen at low concentrations. *Limnol. Oceanogr.* 4, 450–454. doi: 10.4319/lo.1969.14.3.0450
- Bushinsky, S., Emerson, S., Riser, S., and Swift, D. (2016). Accurate oxygen measurements on modified Argo floats using in situ air calibrations. *Limnol. Oceanogr. Methods* 14, 491–505. doi: 10.1002/lom3.10107
- Cai, W.-J., and Sayles, F. (1996). Oxygen penetration depths and fluxes in marine sediments. *Mar. Chem.* 52, 123–131.
- Carpenter, J. H. (1965). The accuracy of the winkler method for dissolved oxygen analysis. *Limnol. Oceanogr.* 10, 135–140. doi: 10.4319/lo.1965.10.1.0135
- CARS (2009). CSIRO Atlas of Regional Seas – World Monthly. Battery Point, TAS: Integrated Marine Observing System.
- Carstensen, J., and Conley, D. J. (2019). Baltic Sea hypoxia takes many shapes and sizes. *Bul. Limnol. Oceanogr.* 28, 125–129. doi: 10.1002/lob.10350
- Carstensen, J., Andersen, J. H., Gustafsson, B. G., and Conley, D. J. (2014). Deoxygenation of the Baltic Sea during the last century. *Proc. Natl. Acad. Sci.* 111, 5628–5633. doi: 10.1073/pnas.1323156111
- Cathalot, C., Rabouille, C., Pastor, L., Deflandre, B., Viollier, E., Buscail, R., et al. (2010). Temporal variability of carbon recycling in coastal sediments influenced by rivers: assessing the impact of flood inputs in the Rhone River prodelta. *Biogeosciences* 7, 1187–1205. doi: 10.5194/bg-7-1187-2010
- Champanois, W., and Borges, A. (2019). Inter-annual variations over a decade of primary production of the seagrass *Posidonia oceanica*. *Limnol. Oceanogr.* 64, 32–45. doi: 10.1002/lno.11017
- Chance, R., Baker, A. R., Carpenter, L., and Jickells, T. (2014). The distribution of iodide at the sea surface. *Environ. Sci. Process Impacts* 16, 1841–1859. doi: 10.1039/C4EM00139G
- Cheung, W. W. L. (2018). The future of fishes and fisheries in the changing oceans. *J. Fish Biol.* 92, 790–803. doi: 10.1111/jfb.13558
- Chu, J. W., and Gale, K. S. (2017). Ecophysiological limits to aerobic metabolism in hypoxia determine epibenthic distributions and energy sequestration in the northeast Pacific Ocean. *Limnol. Oceanogr.* 62, 59–74. doi: 10.1002/lno.10370
- Clark, L. C. Jr., Wolf, R., Granger, D., and Taylor, Z. (1953). Continuous recording of blood oxygen tensions by polarography. *J. Appl. Physiol.* 6, 189–193. doi: 10.1152/jappl.1953.6.3.189
- Claustre, H., Johnson, K. S., and Takeshita, Y. (2020). Observing the global ocean with biogeochemical-Argo. *Annu. Rev. Mar. Sci.* 12, 23–48. doi: 10.1146/annurev-marine-010419-010956
- Coffey, D. M., and Holland, K. N. (2015). First autonomous recording of in situ dissolved oxygen from free-ranging fish. *Anim. Biotelem.* 3, 47. doi: 10.1186/s40317-015-0088-x
- Conley, D. J., Carstensen, J., Vaquer-Sunyer, R., and Duarte, C. M. (2009). Ecosystem thresholds with hypoxia. *Hydrobiologia* 629, 21–29. doi: 10.1007/s10750-009-9764-2
- Coppola, L., Ntoulas, M., Bozzano, R., Bensi, M., Hartman, S. E., Charcos Llorens, M., et al. (2016). Handbook of Best Practices for Open Ocean Fixed Observatories. Report of the FixO3 Project. Brussels, Belgium: European Commission, 127.
- Coppola, L., Prieur, L., Taupier-Letage, I., Estournel, C., Testor, P., Lefevre, D., et al. (2017). Observation of oxygen ventilation into deep waters through targeted deployment of multiple Argo-O₂ floats in the north-western Mediterranean Sea in 2013. *J. Geophys. Res. Oceans* 122, 6325–6341. doi: 10.1002/2016JC012594
- de Mutsert, K., Steenbeek, J., Cowan, J. H., and Christensen, V. (2017). “Using ecosystem modeling to determine hypoxia effects on fish and fisheries,” in *Modeling Coastal Hypoxia*, eds J. Dubravko, K. Rose, R. Hetland, and K. Fennel (Cham: Springer), 377–400. doi: 10.1007/978-3-319-54571-4_14
- Desbruyères, D. G., Purkey, S. G., McDonagh, E. L., Johnson, G. C., and King, B. A. (2016). Deep and abyssal ocean warming from 35 years of repeat hydrography. *Geo. Phys. Res. Lett.* 43, 10356–10365. doi: 10.1002/2016GL070413
- Deutsch, C., Penn, J. L., and Seibel, B. (2020). Metabolic trait diversity shapes marine biogeography. *Nature* 585, 557–562. doi: 10.1038/s41586-020-2721-y

- Diaz, R. J., and Rosenberg, R. (2008). Spreading dead zones and consequences for marine ecosystems. *Science* 321, 926–929. doi: 10.1126/science.1156401
- Dickey, T. D. (2003). Emerging ocean observations for interdisciplinary data assimilation systems. *J. Mar. Syst.* 40, 5–48. doi: 10.1016/S0924-7963(03)00111-3
- Druon, J. N., Chassot, E., Murua, H., and Lopez, J. (2017). Skipjack tuna availability for purse seine fisheries is driven by suitable feeding habitat dynamics in the Atlantic and Indian Oceans. *Front. Mar. Sci.* 4:315. doi: 10.3389/fmars.2017.00315
- Dulvy, N., Reynolds, J. D., Pilling, G. M., Pinnegar, J. K., Scutt Phillips, J., Allison, E. H., et al. (2011). *Fisheries management and governance challenges in a climate change*, in *The Economics of Adapting Fisheries to Climate Change*. Paris: OECD Publishing, 31–88. doi: 10.1787/9789264090415-4-en
- Duteil, O., Oeschlies, A., and Böning, C. W. (2018). Pacific Decadal Oscillation and recent oxygen decline in the eastern tropical Pacific Ocean. *Biogeosciences* 15, 7111–7126. doi: 10.5194/bg-15-7111-2018
- Dyck, A. J., and Sumaila, U. R. (2010). Economic impact of ocean fish populations in the global fishery. *J. Bioecon.* 12, 227–243. doi: 10.1007/s10818-010-9088-3
- Eero, M., Hjelm, J., Behrens, J., Buchmann, K., Cardinale, M., Casini, M., et al. (2015). Eastern Baltic cod in distress: biological changes and challenges for stock assessment. *ICES J. Mar. Sci.* 72, 2180–2186. doi: 10.1093/icesjms/fsv109
- Emerson, S., Quay, P., Karl, D. M., Winn, C., and Tupas, L. M. (1997). Experimental determination of the organic carbon flux from open-ocean surface waters. *Nature* 389, 951–954. doi: 10.1038/40111
- Emerson, S., Stump, C., and Nicholson, D. (2008). Net biological oxygen production in the ocean: Remote in situ measurements of O₂ and N₂ in surface waters. *Glob. Biogeochem. Cycles* 22:GB3023. doi: 10.1029/2007GB003095
- Evans, N., Schroeder, I. D., Pozo Buil, M., Jacox, M. G., and Bograd, S. J. (2020). Drivers of subsurface deoxygenation in the southern California Current System. *Geophys. Res. Lett.* 46:e2020GL089274. doi: 10.1029/2020GL089274
- Fennel, K., and Testa, J. M. (2019). Biogeochemical controls on coastal hypoxia. *Annu. Rev. Mar. Sci.* 11, 105–130. doi: 10.1146/annurev-marine-010318-095138
- Gallo, N., Wei, C.-L., Levin, L., Kuhn, L., and Barry, J. (2020). Dissolved oxygen and temperature best predict deep-sea fish community structure in the Gulf of California with implications for climate change. *Mar. Ecol. Prog. Ser.* 637, 159–180. doi: 10.3354/meps13240
- Gammal, J., Norkko, J., Pilditch, C. A., and Norkko, A. (2017). Coastal hypoxia and the importance of benthic macrofauna communities for ecosystem functioning. *Estuaries Coasts* 40, 457–468. doi: 10.1007/s12237-016-0152-7
- Garcia, H. E., Boyer, T. P., Levitus, S., Locarnini, R. A., and Antonov, J. (2005). On the variability of dissolved oxygen and apparent oxygen utilization content for the upper world ocean: 1955 to 1998. *Geophys. Res. Lett.* 32:L09604. doi: 10.1029/2004GL022286
- Garcia, H. E., Locarnini, R. A., Boyer, T. P., Antonov, J. I., Baranova, O. K., Zweng, M. M., et al. (2013). “World ocean atlas 2013, volume 3: dissolved oxygen, apparent oxygen utilization, and oxygen saturation,” in *NOAA Atlas NESDIS*, Vol. 75, eds S. Levitus and A. Mishonov (Silver Spring, MD: NOAA).
- Garcia, H. E., Weathers, K., Paver, C. R., Smolyar, I., Boyer, T., Locarnini, R., et al. (2019). “World Ocean Atlas 2018, Volume 3: Dissolved Oxygen, Apparent Oxygen Utilization, and Oxygen Saturation,” in *NOAA Atlas NESDIS*, Vol. 83, ed. A. Mishonov (Silver Spring, MD: NOAA).
- Garcia-Robledo, E., Paulmier, A., Borisov, S. M., and Revsbech, N. P. (2021). Sampling in low oxygen aquatic environments: The deviation from anoxic conditions. *Limnol. Oceanogr. Methods* 2021:10457. doi: 10.1002/lom3.10457
- Garçon, V. C., Karstensen, J., Palacz, A., Telszewski, M., Aparco Lara, T., Breitburg, D., et al. (2019). Multidisciplinary observing in the World Ocean’s oxygen minimum zone regions: from climate to fish-the VOICE initiative. *Front. Mar. Sci.* 6:722. doi: 10.3389/fmars.2019.00722
- Gharant, M. E., Tjiputra, J., Bethke, I., Samuelsen, A., Skjelvan, I., Bentsen, M., et al. (2017). Ensemble data assimilation for ocean biogeochemical state and parameter estimation at different sites. *Ocean Model.* 112, 65–89. doi: 10.1016/j.ocemod.2017.02.006
- Gilbert, D., Rabalais, N. N., Diaz, R. J., and Zhang, J. (2010). Evidence for greater oxygen decline rates in the coastal ocean than in the open ocean. *Biogeosciences* 7:2283. doi: 10.5194/bg-7-2283-2010
- Giorgetti, A. E., Partescano, E., Barth, A., Buga, L., Gatti, J., Giorgi, G., et al. (2018). EMODnet chemistry spatial data infrastructure for marine observations and related information. *Ocean Coast. Manag.* 166, 9–17. doi: 10.1016/j.ocecoaman.2018.03.016
- Glud, R. N. (2008). Oxygen dynamics of marine sediments. *Mar. Biol. Res.* 4, 243–289. doi: 10.1080/17451000801888726
- Glud, R. N., Gundersen, J. K., Ramsing, N. B., Buffle, J., and Horvai, G. (2000). *Electrochemical and optical oxygen microsenors for in situ measurements: In situ monitoring of aquatic systems: Chemical analysis and speciation*. Chichester, UK: John Wiley and Sons, 20–73.
- Gobler, C. J., and Baumann, H. (2016). Hypoxia and acidification in ocean ecosystems: coupled dynamics and effects on marine life. *Biol. Lett.* 12:20150976. doi: 10.1098/rsbl.2015.0976
- Gordon, C., Fennel, K., Richards, C., Shay, L. K., and Brewster, J. K. (2020). Can ocean community production and respiration be determined by measuring high-frequency oxygen profiles from autonomous floats? *Biogeosciences* 17, 4119–4134. doi: 10.5194/bg-17-4119-2020
- Gruber, N., Doney, S. C., Emerson, S. R., Gilbert, D., Kobayashi, T., Körtzinger, A., et al. (2010). “Adding oxygen to argo: Developing a global in situ observatory for ocean deoxygenation and biogeochemistry,” in *Proceedings of Ocean Obs ’09: Sustained Ocean Observations and Information for Society*, eds J. Hall, D. E. Harrison, and D. Stammer (New Zealand: ESA Publication), 12. doi: 10.5270/OceanObs09.cwp.39
- Gust, G., Boeij, K., Helder, W., and Sundby, B. (1987). On the velocity sensitivity (stirring effect) of polarographic oxygen microelectrodes. *Neth. J. Sea Res.* 21, 255–263. doi: 10.1016/0077-7579(87)90001-9
- Hagy, J., Boynton, W., Keefe, C., and Wood, K. (2004). Hypoxia in Chesapeake Bay, 1950–2001: Long-term change in relation to Nutrient loading and river flow. *Estuaries* 27, 634–658. doi: 10.1007/BF02907650
- Hall, P. O. J., Almroth-Rosell, E., Bonaglia, S., Dale, A. W., Hylen, A., Kononets, M., et al. (2017). Influence of natural oxygenation of baltic proper deep water on benthic recycling and removal of phosphorus, nitrogen, silicon and carbon. *Front. Mar. Sci.* 4:27. doi: 10.3389/fmars.2017.00027
- Helly, J., and Levin, L. A. (2004). Global distribution of naturally occurring marine hypoxia on continental margins. *Deep Sea Res.* 51, 1159–1168. doi: 10.1016/j.dsr.2004.03.009
- Holtappels, M., Kuypers, M. M. M., Schlüter, M., and Bruchert, V. (2011). Measurement and interpretation of solute concentration gradients in the benthic boundary layer. *Limnol. Oceanogr. Meth.* 9, 1–13. doi: 10.4319/lom.2011.9.1
- Hood, E. M., Sabine, C. L., and Sloyan, B. M. (eds) (2010). *The GO-SHIP Repeat Hydrography Manual: A Collection of Expert Reports and Guidelines, Version 1. (IOCCP Report 14), (ICPO Publication Series 134)*. Available online at: <http://www.go-ship.org/HydroMan.html>. doi: 10.25607/OBP-1341
- Howard, E. M., Penn, J. L., Frenzel, H., Seibel, B. A., Bianchi, D., Renault, L., et al. (2020). Climate-driven aerobic habitat loss in the California Current System. *Sci. Adv.* 6:eay3188. doi: 10.1126/sciadv.aay3188
- Hughes, D. J., Alderdice, R., Cooney, C., Kühl, M., Pernice, M., Voolstra, C. R., et al. (2020). Coral reef survival under accelerating ocean deoxygenation. *Nat. Climate Change* 10, 296–307. doi: 10.1038/s41558-020-0737-9
- IOC (2019). *The Science We Need for the Ocean We Want: The United Nations Decade of Ocean Science for Sustainable Development (2021–2030)*. Paris: IOC, 24.
- Jahnke, R. A. (1996). The global ocean flux of particulate organic carbon: Areal distribution and magnitude. *Glob. Biogeochem. Cycles* 10, 71–88. doi: 10.1029/95GB03525
- Jahnke, R. A., and Christiansen, M. B. (1989). A free vehicle benthic chamber instrument for sea floor studies. *Deep-sea Res.* 32, 885–897. doi: 10.1016/0198-0149(89)90011-3
- Johnson, K. S., and Claustre, H. (2016). *Bringing Biogeochemistry Into the Argo age*. Washington, DC: EOS, 97. doi: 10.1029/2016EO062427
- Johnson, K. S., Plant, J. N., Riser, S. C., and Gilbert, D. (2015). Air oxygen calibration of oxygen optodes on a profiling float array. *J. Atmos. Ocean. Technol.* 32, 2160–2172. doi: 10.1175/JTECH-D-15-0101.1
- Johnson, K. S., Plant, J., Coletti, L., Jannasch, H., Sakamoto, C., Riser, S., et al. (2017). Biogeochemical sensor performance in the SOCCOM profiling float array. *J. Geophys. Res. Oceans* 122, 6416–6436. doi: 10.1002/2017JC012838

- Joyce, T. M. (1991). *Introduction to the Collection of Expert Reports Compiled for the WHP Programme. WHP Operations and Methods—July 1991*. Southampton: WOCE Hydrographic Programme Office, 4.
- Keeling, R. F., and Manning, A. C. (2014). *Studies of Recent Changes in Atmospheric O₂ Content. Treatise on Geochemistry*, Second Edn, Vol. 5. Amsterdam: Elsevier, 385–404. doi: 10.1016/B978-0-08-095975-7.00420-4
- Keeling, R., Körtzinger, A., and Gruber, N. (2010). Ocean deoxygenation in a warming world. *Annu. Rev. Mar. Sci.* 2, 199–229. doi: 10.1146/annurev.marine.010908.163855
- Keister, J. E., Houde, E. D., and Breitburg, D. L. (2000). Effects of bottom-layer hypoxia on abundances and depth distributions of organisms in Patuxent River, Chesapeake Bay. *Mar. Ecol. Prog. Ser.* 205, 43–59. doi: 10.3354/meps205043
- Keller, A. A., Ciannelli, L., Wakefield, W. W., Simon, V., Barth, J. A., and Pierce, S. D. (2017). Species-specific responses of demersal fishes to near-bottom oxygen levels within the California Current large marine ecosystem. *Mar. Ecol. Prog. Ser.* 568, 151–173. doi: 10.3354/meps12066
- Kemp, W. M., Testa, J. M., Conley, D. J., Gilbert, D., and Hagy, J. D. (2009). Coastal hypoxia responses to remediation. *Biogeosciences* 6, 2985–3008. doi: 10.5194/bg-6-2985-2009
- Key, R. M., Kozyr, A., Sabine, C. L., Lee, K., Wanninkhof, R., Bullister, J. L., et al. (2004). A global ocean carbon climatology: Results from Global Data Analysis Project (GLODAP). *Glob. Biogeochem. Cycles* 18, 1–23. doi: 10.1029/2004GB002247
- Körtzinger, A., Schimanski, J., Send, U., and Wallace, D. (2004). The ocean takes a deep breath. *Science* 306:1337. doi: 10.1126/science.1102557
- Krasnopolsky, V., Nadiga, S., Mehra, A., Bayler, E., and Behringer, D. (2016). Neural networks technique for filling gaps in satellite measurements: application to ocean color observations. *Comput. Intel. Neurosci.* 2016:6156513. doi: 10.1155/2016/6156513
- Kriest, I., Sauerland, V., Khatriwala, S., Srivastav, A., and Oschlies, A. (2017). Calibrating a global three-dimensional biogeochemical ocean model (MOPS-1.0). *Geosci. Model Dev.* 10, 127–154. doi: 10.5194/gmd-10-127-2017
- Labasque, T., Chaumery, C., Aminot, A., and Kergoat, G. (2004). Spectrophotometric winkler determination of dissolved oxygen: Re-examination of critical factors and reliability. *Mar. Chem.* 88, 53–60. doi: 10.1016/j.marchem.2004.03.004
- Laffoley, D., and Baxter, J. M. (2019). *Ocean Deoxygenation: Everyone's Problem - Causes, Impacts, Consequences and Solutions*. Gland, Switzerland: IUCN, 580. doi: 10.2305/IUCN.CH.2019.13.en
- Lakowicz, J. (2006). *Principles of Fluorescence Spectroscopy*, 3rd Edn. Amsterdam: Elsevier, 954. doi: 10.1007/978-0-387-46312-4
- Langdon, C. (2010). "Determination of dissolved oxygen in seawater by Winkler titration using Amperometric Technique," in *The GO-SHIP Repeat Hydrography Manual: A Collection of Expert Reports and Guidelines. Version 1 (IOCCP Report Number 14; ICPO Publication Series Number 134)*, eds E. M. Hood, C. L. Sabine and B. M. Sloyan (Lyon: ICPO), 18. doi: 10.25607/OBP-1350
- Lauvset, S. K., and Tanhua, T. (2015). A toolbox for secondary quality control on ocean chemistry and hydrographic data. *Limnol. Oceanogr. Meth.* 13, 601–608. doi: 10.1002/lom3.10050
- Lauvset, S. K., Key, R. M., Olsen, A., van Heuven, S., Velo, A., Lin, X., et al. (2016). A new global interior ocean mapped climatology: the 1°x1° GLODAP version 2. *Earth Syst. Sci. Data* 8, 325–340. doi: 10.5194/essd-8-325-2016
- Le Traon, P. Y., Reppucci, A., Fanjul, E. A., Aouf, L., Behrens, A., Belmonte, M., et al. (2019). From observation to information and users: The copernicus marine service perspective. *Front. Mar. Sci.* 6:234. doi: 10.3389/fmars.2019.00234
- Lehner, P., Larndorfer, C., Garcia-Robledo, E., Larsen, M., Borisov, S. M., Revsbech, N. P., et al. (2015). Lumos - a sensitive and reliable optode system for measuring dissolved oxygen in the nanomolar range. *PLoS One* 10:e0128125. doi: 10.1371/journal.pone.0128125
- Lemburg, J., Wenzhöfer, F., Hofbauer, M., Färber, P., and Meyer, V. (2018). "Benthic crawler NOMAD," in *Proceedings of the 2018 OCEANS - MTS/IEEE Kobe Techno-Oceans (OTO)*, Kobe, 1–7. doi: 10.1109/OCEANSKOBE.2018.8559446
- Levin, L. A. (2003). Oxygen minimum zone benthos: Adaptation and community response to hypoxia. *Oceanogr. Mar. Biol.* 41, 1–45.
- Levin, L. A. (2018). Manifestation, drivers, and emergence of open ocean deoxygenation. *Annu. Rev. Mar. Sci.* 10, 229–260. doi: 10.1146/annurev-marine-121916-063359
- Levin, L. A., Ekau, W., Gooday, A. J., Jorissen, F., Middelburg, J. J., Naqvi, S. W. A., et al. (2009). Effects of natural and human-induced hypoxia on coastal benthos. *Biogeosciences* 6, 2063–2098. doi: 10.5194/bg-6-2063-2009
- Levin, L. A., Wei, C. L., Dunn, D. C., Amon, D., Ashford, O., Cheung, W., et al. (2020). Climate change considerations are fundamental to management of deep-sea resource extraction. *Glob. Change Biol.* 26, 4664–4678. doi: 10.1111/gcb.15223
- Li, P., and Tanhua, T. (2020). Recent changes in deep ventilation of the mediterranean sea; evidence from long-term transient tracer observations. *Front. Mar. Sci.* 7:594. doi: 10.3389/fmars.2020.00594
- Limborg, K. E., and Casini, M. (2018). Effect of marine hypoxia on Baltic Sea cod *Gadus morhua*: evidence from otolith chemical proxies. *Front. Mar. Sci.* 5:482. doi: 10.3389/fmars.2018.00482
- Limborg, K. E., and Casini, M. (2019). Otolith chemistry indicates recent worsened Baltic cod condition is linked to hypoxia exposure. *Biol. Lett.* 15:20190352. doi: 10.1098/rsbl.2019.0352
- Limborg, K. E., Breitburg, D., Swaney, D., and Jacinto, G. (2020). Ocean deoxygenation: a primer. *One Earth* 2, 24–29. doi: 10.1016/j.oneear.2020.01.001
- Long, M. C., Deutsch, C., and Ito, T. (2016). Finding forced trends in oceanic oxygen. *Glob. Biogeochem. Cycles* 30, 381–397. doi: 10.1002/2015GB005310
- Lu, Y., Yuan, J., Lu, X., Su, C., Zhang, Y., Wang, C., et al. (2018). Major threats of pollution and climate change to global coastal ecosystems and enhanced management for sustainability. *Environ. Pollut.* 239, 670–680. doi: 10.1016/j.envpol.2018.04.016
- Martin Míguez, B., Novellino, A., Vinci, M., Claus, S., van Calewaert, J.-B., Vallius, H., et al. (2019). The European Marine Observation and Data Network (EMODnet): visions and roles of the gateway to marine data in Europe. *Front. Mar. Sci.* 6:313. doi: 10.3389/fmars.2019.00313
- Mavropoulos, A. M., Vervatis, V., and Sofianos, S. (2020). Dissolved oxygen variability in the Mediterranean Sea. *J. Mar. Syst.* 2020:103348. doi: 10.1016/j.jmarsys.2020.103348
- Montes, I., Dewitte, B., Gutknecht, E., Paulmier, A., Dadou, I., Oschlies, A., et al. (2014). High-resolution modeling of the eastern Tropical Pacific oxygen minimum zone: Sensitivity to the tropical oceanic circulation. *J. Geophys. Res. Oceans* 119, 5515–5532. doi: 10.1002/2014JC009858
- Neilan, R. M., and Rose, K. (2014). Simulating the effects of fluctuating dissolved oxygen on growth, reproduction, and survival of fish and shrimp. *J. Theor. Bio.* 343, 54–68. doi: 10.1016/j.jtbi.2013.11.004
- Nicholson, D. P., and Feen, M. L. (2017). Air calibration of an oxygen optode on an underwater glider. *Limnol. Oceanogr. Meth.* 15, 495–502. doi: 10.1002/lom3.10177
- Nicholson, D., Emerson, S., and Eriksen, C. C. (2008). Net community production in the deep euphotic zone of the subtropical North Pacific gyre from glider surveys. *Limnol. Oceanogr.* 53, 2226–2236. doi: 10.4319/lo.2008.53.5_part_2.2226
- Noone, K. J., Sumaila, U. R., and Diaz, R. J. (2013). *Managing Ocean Environments in a Changing Climate: Sustainability and Economic Perspectives*. London, UK: Elsevier, 359.
- Ohman, M. D., Rudnick, D. L., Chekalyuk, A., Davis, R. E., Feely, R. A., Kahru, M., et al. (2013). Autonomous ocean measurements in the California current ecosystem. *Oceanography* 26, 18–25. doi: 10.5670/oceanog.2013.41
- Olsen, A., Key, R. M., van Heuven, S., Lauvset, S. K., Velo, A., Lin, X., et al. (2016). The Global Ocean Data Analysis Project version 2 (GLODAPv2) – an internally consistent data product for the world ocean. *Earth Syst. Sci. Data* 8, 297–323. doi: 10.5194/essd-8-297-2016
- Olsen, A., Lange, N., Key, R. M., Tanhua, T., Bittig, H., and Kozyr, A. (2020). GLODAPv2.2020 – the second update of GLODAPv2. *Earth Syst. Sci. Data* 2020:165. doi: 10.5194/essd-2020-165
- Oschlies, A. (2018). "Reconciling systematic differences between observed and simulated ocean deoxygenation," in *Proceedings of the 4th International Symposium on the Effects of Climate Change on the World's Oceans 4 (ECCWO) Meeting*, Washington, DC.

- Oschlies, A., Brandt, P., Stramma, L., and Schmidtko, S. (2018). Drivers and mechanisms of ocean deoxygenation. *Nat. Geosci.* 11, 467–473. doi: 10.1038/s41561-018-0152-2
- Pairaud, I., Repecaud, M., Ravel, C., Fuchs, R., Arnaud, M., Champelovier, A., et al. (2016). “MesuRhô. Plateforme instrumentée de suivi des paramètres environnementaux à l’embouchure du Rhône,” in *Mesures à haute résolution dans l’environnement marin côtier*, eds F. G. Schmitt and A. Lefebvre (Paris: CNRS), 73–87.
- Pastres, R., Ciavatta, S., and Solidoro, C. (2003). The Extended Kalman Filter (EKF) as a tool for the assimilation of high frequency water quality data. *Ecol. Model.* 170, 227–235. doi: 10.1016/S0304-3800(03)00230-8
- Paulmier, A. (2017). “Oxygen and the ocean,” in *The Ocean revealed*, eds A. Euzen, F. Gaill, D. Lacroix, and P. Cury (Paris: CNRS), 48–49.
- Paulmier, A., and Ruiz-Pino, D. (2009). Oxygen minimum zones (OMZs) in the modern ocean. *Prog. Oceanogr.* 80, 113–128. doi: 10.1016/j.pocean.2008.08.001
- Pauly, D., and Zeller, D. (2016). Catch reconstructions reveal that global marine fisheries catches are higher than reported and declining. *Nat. Commun.* 7, 1–9. doi: 10.1038/ncomms10244
- Pitcher, G. C., Aguirre, A., Breitburg, D., Cardich, J., Carstensen, J., Conley, D. J., et al. (2021). System controls of coastal and open ocean oxygen depletion. *Prog. Oceanogr.* 197:102613. doi: 10.1016/j.pocean.2021.102613
- Pollock, M. S., Clarke, L. M. J., and Dubé, M. G. (2007). The effects of hypoxia on fishes: from ecological relevance to physiological effects. *Environ. Rev.* 5, 1–14. doi: 10.1139/a06-006
- Polsenaere, P., Deflandre, B., Thouzeau, G., Rigaud, S., Cox, T., Amice, E., et al. (2021). Comparison of benthic oxygen exchange measured by aquatic Eddy Covariance and Benthic Chambers in two contrasting coastal biotopes (Bay of Brest, France). *Regional Stud. Mar. Sci.* 43:101668. doi: 10.1016/j.rsma.2021.101668
- Pörtner, H. O. (2021). Climate impacts on organisms, ecosystems and human societies: integrating OCLTT into a wider context. *J. Exper. Biol.* 224:jeb238360. doi: 10.1242/jeb.238360
- Purcell, K. M., Craig, J. K., Nance, J. M., Smith, M. D., and Benneer, L. S. (2017). Fleet behavior is responsive to a large-scale environmental disturbance: Hypoxia effects on the spatial dynamics of the northern Gulf of Mexico shrimp fishery. *PLoS One* 12:e0183032. doi: 10.1371/journal.pone.0183032
- Purser, A., Thomsen, L., Barnes, C., Best, M., Chapman, R., Hofbauer, M., et al. (2013). Temporal and spatial benthic data collection via an internet operated deep sea crawler. *Meth. Oceanogr.* 5, 1–18. doi: 10.1016/j.mio.2013.07.001
- Queirós, A. M., Huebert, K. B., Keyl, F., Fernandes, J. A., Stolte, W., Maar, M., et al. (2016). Solutions for ecosystem-level protection of ocean systems under climate change. *Glob. Change Biol.* 22, 3927–3936. doi: 10.1111/gcb.13423
- Rabalais, N. N. (2015). Human impacts on fisheries across the land–sea interface. *Proc. Natl. Acad. Sci.* 112, 7892–7893. doi: 10.1073/pnas.1508766112
- Rabalais, N. N., Diaz, R. J., Levin, L. A., Turner, R. E., Gilbert, D., and Zhang, J. (2010). Dynamics and distribution of natural and human-caused hypoxia. *Biogeosciences* 7:585. doi: 10.5194/bg-7-585-2010
- Rabalais, N. N., Turner, R. E., and Wiseman, W. J. (2002). Gulf of Mexico hypoxia, a.k.a. “the dead zone”. *Annu. Rev. Ecol. Syst.* 33, 235–263. doi: 10.1146/annurev.ecolsys.33.010802.150513
- Rabalais, N. N., Turner, R. E., Gupta, B. S., Boesch, D. F., Chapman, P., and Murrell, M. C. (2007a). Hypoxia in the northern Gulf of Mexico: Does the science support the plan to reduce, mitigate, and control hypoxia? *Estuaries Coasts* 30, 753–772. doi: 10.1007/BF02841332
- Rabalais, N. N., Turner, R. E., Sen Gupta, B. K., Platon, E., and Parsons, M. L. (2007b). Sediments tell the history of eutrophication and hypoxia in the northern Gulf of Mexico. *Ecol. Appl.* 17, S129–S143. doi: 10.1890/06-0644.1
- Racapé, V., Thierry, V., Mercier, H., and Cabanes, C. (2019). ISOW spreading and mixing as revealed by deep-argo floats launched in the charlie gibbs fracture zone. *J. Geophys. Res.* 124, 6787–6808. doi: 10.1029/2019JC015040
- Reimers, C. E., Özkan-Haller, H. T., Berg, P., Devol, A., McCann-Grosvenor, K., and Sanders, R. D. (2012). Benthic oxygen consumption rates during hypoxic conditions on the Oregon continental shelf: Evaluation of the eddy correlation method. *J. Geophys. Res.* 117:C02021. doi: 10.1029/2011JC007564
- Resplandy, L., Keeling, M., Edehbar, Y., Brooks, M. K., Wang, R., Bopp, L., et al. (2018). Quantification of ocean heat uptake from changes in atmospheric O₂ and CO₂ composition. *Nature* 563, 105–108. doi: 10.1038/s41586-018-0651-8
- Revsbech, N. P., Larsen, L. H., Gundersen, J. K., Dalsgaard, T., Ulloa, O., Thamdrup, B., et al. (2009). Determination of ultra-low oxygen concentrations in oxygen minimum zones by the stox sensor. *Limnol. Oceanogr. Meth.* 7, 371–381. doi: 10.4319/lom.2009.7.371
- Ridgwell, A., Hargreaves, J. C., Edwards, N. R., Annan, J. D., Lenton, T. M., Marsh, R., et al. (2007). Marine geochemical data assimilation in an efficient earth system model of global biogeochemical cycling. *Biogeosciences* 4, 87–104. doi: 10.5194/bg-4-87-2007
- Rigaud, S., Deflandre, B., Maire, O., Bernard, G., Duchêne, J. C., Poirier, D., et al. (2018). Transient biogeochemistry in intertidal sediments: New insights from tidal pools in *Zostera noltei* meadows of Arcachon Bay (France). *Mar. Chem.* 200, 1–13. doi: 10.1016/j.marchem.2018.02.002
- Roemmich, D., Alford, M. H., Claustre, H., Johnson, K., King, B., Moum, J., et al. (2019). On the future of Argo: An enhanced global array of physical and biogeochemical sensing floats. *Front. Mar. Sci.* 6:439. doi: 10.3389/fmars.2019.00439
- Roman, M. R., Brandt, S. B., Houde, E. D., and Pierson, J. J. (2019). Interactive effects of hypoxia and temperature on coastal pelagic zooplankton and fish. *Front. Mar. Sci.* 6:139. doi: 10.3389/fmars.2019.00139
- Rose, K. A., Creekmore, S., Thomas, P., Craig, J. K., Rahman, M. S., and Neilan, R. M. (2018). Modeling the population effects of hypoxia on Atlantic croaker (*Micropogonias undulatus*) in the northwestern Gulf of Mexico: part 1—model description and idealized hypoxia. *Estuaries Coasts* 41, 233–254. doi: 10.1007/s12237-017-0266-6
- Rose, K. A., Gutierrez Aguilar, D., Breitburg, D., Conley, D., Craig, J. K., Froehlich, H. E., et al. (2019). “Impacts of ocean deoxygenation on fisheries,” in *Ocean Deoxygenation: Everyone’s Problem - Causes, impacts, consequences and solutions*, eds D. Laffoley and J. M. Baxter (Gland, Switzerland: International Union for Conservation of Nature and Natural Resources (IUCN)), 519–544.
- Rovelli, L., Attard, K. M., Bryant, L. D., Flögel, S., Stahl, H., Roberts, J. M., et al. (2015). Benthic O₂ uptake of two cold-water coral communities estimated with the non-invasive eddy-correlation technique. *Mar. Ecol. Prog. Ser.* 525, 97–104. doi: 10.3354/meps11211
- Rudnick, D. L. (2016b). *Instrument Development Group, Scripps Institution of Oceanography*. La Jolla, CA: California Underwater Glider Network.
- Rudnick, D. L. (2016a). Ocean research enabled by underwater gliders. *Annu. Rev. Mar. Sci.* 8, 519–541. doi: 10.1146/annurev-marine-122414-033913
- Rudnick, D. L., Zaba, K. D., Todd, R. E., and Davis, R. E. (2017). A climatology of the California current system from a network of underwater gliders. *Prog. Oceanogr.* 154, 64–106. doi: 10.1016/j.pocean.2017.03.002
- Sánchez-Velasco, L., Godínez, V. M., Ruvalcaba-Aroche, E. D., Márquez-Artavia, A., Beier, E., Barton, E. D., et al. (2019). Larval fish habitats and deoxygenation in the northern limit of the oxygen minimum zone off Mexico. *J. Geophys. Res. Oceans* 124, 9690–9705. doi: 10.1029/2019JC015414
- Sasano, D., Takatani, Y., Kosugi, N., Nakano, T., Midorikawa, T., and Ishii, M. (2018). Decline and biennial oscillations of dissolved oxygen in the Oyashio region and their propagation to the western North Pacific. *Glob. Biogeochem. Cycles* 32, 909–931. doi: 10.1029/2017GB005876
- Saunders, P. M. (1986). The accuracy of salinity, oxygen in the deep ocean. *J. Phys. Ocean.* 16, 189–195. doi: 10.1175/1520-0485(1986)016<0189:TAOMOS>2.0.CO;2
- Savchuk, O. P. (2018). Large-scale nutrient dynamics in the Baltic Sea, 1970–2016. *Front. Mar. Sci.* 5:95. doi: 10.3389/fmars.2018.00095
- Schmidtko, S., Stramma, L., and Visbeck, M. (2017). Decline in global oceanic oxygen content during the past five decades. *Nature* 542, 335–339. doi: 10.1038/nature21399
- Seibel, B. A., and Deutsch, C. (2020). Oxygen supply capacity in animals evolves to meet maximum demand at the current oxygen partial pressure regardless of size or temperature. *J. Exp. Biol.* 223:jeb210492. doi: 10.1242/jeb.210492
- Seibel, B. A., Andres, A., Birk, M. A., Shaw, C. T., Timpe, A., and Welsh, C. (2021). Oxygen supply capacity breathes new life into the critical oxygen partial pressure (Pcrit). *J. Exper. Biol.* 224:jeb242210. doi: 10.1242/jeb.242210
- Seitzinger, S. P., Mayorga, E., Bouwman, A. F., Kroeze, C., Beusen, A. H. W., Billen, G., et al. (2010). Global river nutrient export: A scenario analysis of past and future trends. *Glob. Biogeochem. Cycles* 24:GB0A08. doi: 10.1029/2009GB003587

- Smith, K. L. Jr., Sherman, A. D., McGill, P. R., Henthorn, R. G., Ferreira, J., and Huffard, C. L. (2017). Evolution of monitoring an abyssal time-series station in the northeast Pacific over 28 years. *Oceanography* 30, 72–81. doi: 10.5670/oceanog.2017.425
- Soetaert, K., and Grégoire, M. (2011). Estimating marine biogeochemical rates of the carbonate pH system—A Kalman filter tested. *Ecol. Modell.* 222, 1929–1942. doi: 10.1016/j.ecolmodel.2011.03.012
- Staudinger, C., Strobl, M., Fischer, J. P., Thar, R., Mayr, T., Aigner, D., et al. (2018). A versatile optode system for oxygen, carbon dioxide, and pH measurements in seawater with integrated battery and logger. *Limnol. Oceanogr. Meth.* 16, 459–473. doi: 10.1002/lom3.10260
- Steinhoff, T., Gkritzalis, T., Lauvset, S. K., Jones, S., Schuster, U., Olsen, A., et al. (2019). Constraining the oceanic uptake and fluxes of greenhouse gases by building an ocean network of certified stations: The Ocean Component of the Integrated Carbon Observation System, ICOS-Oceans. *Front. Mar. Sci.* 6:544. doi: 10.3389/fmars.2019.00544
- Stramma, L., Johnson, G. C., Sprintall, J., and Mohrholz, V. (2008). Expanding oxygen-minimum zones in the tropical oceans. *Science* 320, 655–658. doi: 10.1126/science.1153847
- Stramma, L., Prince, E. D., Schmidtko, S., Luo, J., Hoolihan, J. P., Visbeck, M., et al. (2012). Expansion of oxygen minimum zones may reduce available habitat for tropical pelagic fishes. *Nat. Climate Change* 2, 33–37. doi: 10.1038/nclimate1304
- Stratmann, T., Soetaert, K., Wei, C.-L., Lin, Y.-S., and van Oevelen, D. (2019). The SCOC database, a large, open, and global database with sediment community oxygen consumption rates. *Sci. Data* 6:242. doi: 10.1038/s41597-019-0259-3
- Sumaila, U. R., Ebrahim, N., Schuhbauer, A., Skerritt, D., Li, Y., Kim, H., et al. (2019). Updated estimates and analysis of global fisheries subsidies. *Mar. Policy* 109:103695. doi: 10.1016/j.marpol.2019.103695
- Swartz, W., Sumaila, U. R., and Watson, R. (2013). Global Ex-Vessel Fish Price Database revisited: a new approach for estimating 'missing' prices. *Environ. Resour. Econ.* 56, 467–480. doi: 10.1007/s10640-012-9611-1
- Takeshita, Y., Jones, B., Johnson, K. S., Chavez, F. P., Rudnick, D., Blum, M., et al. (2021). Accurate pH and O₂ measurements from Spray underwater gliders. *J. Atmos. Ocean. Technol.* 38, 181–195. doi: 10.1175/JTECH-D-20-0095.1
- Takeshita, Y., Martz, T. R., Johnson, K. S., Plant, J. N., Gilbert, D., Riser, S. C., et al. (2013). A climatology-based quality control procedure for profiling float oxygen data. *J. Geophys. Res. Oceans* 118, 5640–5650. doi: 10.1002/jgrc.20399
- Tanhua, T., van Heuven, S., Key, R., Velo, A., Olsen, A., and Schirnick, C. (2010). Quality control procedures and methods of the CARINA database. *Earth Syst. Sci. Data* 2, 35–49. doi: 10.5194/essd-2-35-2010
- Tengberg, A., De Bovee, F., Hall, P., Berelson, W., Cicceri, G., Crassous, P., et al. (1995). Benthic chamber and profile landers in oceanography - A review of design, technical solutions and functioning. *Prog. Oceanogr.* 35, 253–294. doi: 10.1016/0079-6611(95)00009-6
- Testor, P., de Young, B., Rudnick, D. L., Glenn, S., Hayes, D., Lee, C. L., et al. (2019). OceanGliders: A component of the integrated GOOS. *Front. Mar. Sci.* 6:422. doi: 10.3389/fmars.2019.00422
- Thierry, V., and Bittig, H. (2018). *Argo Quality Control Manual for Dissolved Oxygen Concentration*, v2.0. France: IFREMER for Argo BGC Group.
- Thierry, V., Gilbert, D., Kobayashi, T., and Schmid, C. (2018). *Processing argo oxygen data at the DAC level*, v2.3.1. Brest, France: IFREMER.
- Thomas, P., and Rahman, M. S. (2010). Region-wide impairment of Atlantic croaker testicular development and sperm production in the northern Gulf of Mexico hypoxic dead zone. *Mar. Env. Res.* 69, S59–S62. doi: 10.1016/j.marenvres.2009.10.017
- Tiano, L., Garcia-Robledo, E., Dalsgaard, T., Devol, A. H., Ward, B. B., Ulloa, O., et al. (2014). Oxygen distribution and aerobic respiration in the north and south eastern tropical Pacific oxygen minimum zones. *Deep-Sea Res Part I-Oceanogr. Res. Pap.* 94, 173–183. doi: 10.1016/j.dsr.2014.10.001
- Tomasetti, S. J., and Gobler, C. J. (2020). Dissolved oxygen and pH criteria leave fisheries at risk. *Science* 368, 372–373. doi: 10.1126/science.aba4896
- Toussaint, F., Rabouille, C., Cathalot, C., Bombled, B., Abchiche, A., Aouji, O., et al. (2014). A new device to follow temporal variations of oxygen demand in deltaic sediments: the LSCE benthic station. *Limnol. Oceanogr. Methods* 12, 729–741. doi: 10.4319/lom.2014.12.729
- Townhill, B. L., van der Molen, J., Metcalfe, J. D., Simpson, S. D., Farcas, A., and Pinnegar, J. K. (2017). Consequences of climate-induced low oxygen conditions for commercially important fish. *Mar. Ecol. Prog. Ser.* 580, 191–204. doi: 10.3354/meps12291
- Troupin, C., Barth, A., Sirjacobs, D., Ouberdous, M., Brankart, J.-M., Brasseur, P., et al. (2012). Generation of analysis and consistent error fields using the Data Interpolating Variational Analysis (DIVA). *Ocean Model.* 52–53, 90–101. doi: 10.1016/j.ocemod.2012.05.002
- Tyler, R., Brady, D. C., and Targett, T. E. (2009). Temporal and spatial dynamics of diel-cycling hypoxia in estuarine tributaries. *Estuaries Coasts* 32:145. doi: 10.1007/s12237-008-9108-x
- Uchida, H., Johnson, G. C., and McTaggart, K. E. (2010). "CTD oxygen sensor calibration procedures," in *The GO-SHIP Repeat Hydrography Manual: A Collection of Expert Reports and Guidelines*, eds E. M. Hood, C. L. Sabine, and B. M. Sloyan (Brest, France: ICPO Publication).
- Ulses, C., Estournel, C., Fourrier, M., Coppola, L., Kessouri, F., Lefèvre, D., et al. (2021). Oxygen budget of the north-western Mediterranean deep- convection region. *Biogeosciences* 18, 937–960. doi: 10.5194/bg-18-937-2021
- UNESCO (2012). *FOO: Framework for Ocean Observing By the Task Team for an Integrated Framework for Sustained Ocean Observing, UNESCO 2012, IOC/INF-1284*. Paris: UNESCO.
- Vaquier-Sunyer, R., and Duarte, C. M. (2008). Thresholds of hypoxia for marine biodiversity. *Proc. Natl. Acad. Sci.* 105, 15452–15457. doi: 10.1073/pnas.0803833105
- Vergara, O., Dewitte, B., Montes Torres, I., Garçon, V., Ramos, M., Paulmier, A., et al. (2016). Seasonal variability of the oxygen minimum zone off Peru in a high-resolution regional coupled model. *Biogeosciences* 13, 4389–4410. doi: 10.5194/bg-13-4389-2016
- Vinci, M., Giorgetti, A., and Lipizer, M. (2017). The role of EMODnet chemistry in the European challenge for good environmental status. *Nat. Hazards Earth Syst. Sci.* 17, 197–204. doi: 10.5194/nhess-17-197-2017
- Wang, Z. A., Moustahfid, H., Mueller, A. V., Michel, A. P. M., Mowlem, M., Glazer, B. T., et al. (2019). Advancing observation of ocean biogeochemistry, biology, and ecosystems with cost-effective in-situ sensing technologies. *Front. Mar. Sci.* 6:519. doi: 10.3389/fmars.2019.00519
- Wanninkhof, R., Pickers, P. A., Omar, A. M., Sutton, A., Murata, A., Olsen, A., et al. (2019). A surface ocean CO₂ reference network, SOCONET and associated marine boundary layer CO₂ measurements. *Front. Mar. Sci.* 6:400. doi: 10.3389/fmars.2019.00400
- Wei, Y., Jiao, Y., An, D., Li, D., Li, W., and Wei, Q. (2019). Review of dissolved oxygen detection technology: from laboratory analysis to online intelligent detection. *Sensors* 19:3995. doi: 10.3390/s19183995
- Wenzhöfer, F., and Glud, R. N. (2002). Benthic carbon mineralization in the Atlantic: A synthesis based on in situ data from the last decade. *Deep-Sea Res. (I)* 49, 1255–1279. doi: 10.1016/S0967-0637(02)00025-0
- Wenzhöfer, F., Oguri, K., Middelboe, M., Turnewitsch, R., Toyofuku, T., Kitazato, H., et al. (2016). Benthic carbon mineralization in hadal trenches: Assessment by in situ O₂ microprofile measurements. *Deep-Sea Res. I* 116, 276–286. doi: 10.1016/j.dsr.2016.08.013
- Wilkinson, M., Dumontier, M., Aalbersberg, I., Appleton, G., Axton, M., Baak, A., et al. (2016). The FAIR guiding principles for scientific data management and stewardship. *Sci. Data* 3:160018. doi: 10.1038/sdata.2016.18
- Winkler, L. W. (1888). Die Bestimmung des im Wasser gelösten Sauerstoffes. *Chem. Ber.* 21, 2843–2855. doi: 10.1002/cber.18880210212
- Wishner, K. F., Seibel, B. A., Roman, C., Deutsch, C., Outram, D., Shaw, C. T., et al. (2018). Ocean deoxygenation and zooplankton: Very small oxygen differences matter. *Sci. Adv.* 4:eaau5180. doi: 10.1126/sciadv.aau5180
- Wong, G. T. F., and Cheng, X.-H. (1998). Dissolved organic iodine in marine waters: Determination, occurrence and analytical implications. *Mar. Chem.* 59, 271–281. doi: 10.1016/S0304-4203(97)00078-9
- Wong, G. T. F., and Li, K.-Y. (2009). Winkler's method overestimates dissolved oxygen in seawater: Iodate interference and its oceanographic implications. *Mar. Chem.* 115, 86–91. doi: 10.1016/j.marchem.2009.06.008
- Wu, R. S. (2002). Hypoxia: from molecular responses to ecosystem responses. *Mar. Pollut. Bull.* 45, 35–45. doi: 10.1016/S0025-326X(02)00061-9
- Wu, R. S. (2009). "Effects of hypoxia on fish reproduction and development," in *Fish physiology*, Vol. 27, eds J. G. Richards, A. P. Farrell, and C. J. Brauner (Cambridge: Academic Press), 79–141. doi: 10.1016/S1546-5098(08)00003-4
- Yang, B., Emerson, S. R., and Bushinsky, S. M. (2017). Annual net community production in the subtropical Pacific Ocean from in situ oxygen measurements

- on profiling floats. *Glob. Biogeochem. Cycles* 31, 728–744. doi: 10.1002/2016GB005545
- Yasuhara, M., Hunt, G., Breitburg, D., Tsujimoto, A., and Katsuki, K. (2012). Human-induced marine ecological degradation: micropaleontological perspectives. *Ecol. Evol.* 2, 3242–3268. doi: 10.1002/ece3.425
- Yasuhara, M., Tittensor, D. P., Hillebrand, H., and Worm, B. (2017). Combining marine macroecology and palaeoecology in understanding biodiversity: microfossils as a model. *Biol. Rev.* 92, 199–215. doi: 10.1111/brv.12223
- Zhai, W. D., Dai, M., and Cai, W. J. (2009). Coupling of surface pCO₂ and dissolved oxygen in the northern South China Sea: impacts of contrasting coastal processes. *Biogeosciences* 6, 2589–2598. doi: 10.5194/bg-6-2589-2009
- Zhang, Q., Tango, P. J., Murphy, R. R., Forsyth, M. K., Tian, R., Keisman, J., et al. (2018). Chesapeake Bay dissolved oxygen criterion attainment deficit: three decades of temporal and spatial patterns. *Front. Mar. Sci.* 5:422. doi: 10.3389/fmars.2018.00422
- Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.
- Citation:** Grégoire M, Garçon V, Garcia H, Breitburg D, Isensee K, Oschlies A, Telszewski M, Barth A, Bittig HC, Carstensen J, Carval T, Chai F, Chavez F, Conley D, Coppola L, Crowe S, Currie K, Dai M, Deflandre B, Dewitte B, Diaz R, Garcia-Robledo E, Gilbert D, Giorgetti A, Glud R, Gutierrez D, Hosoda S, Ishii M, Jacinto G, Langdon C, Lauvset SK, Levin LA, Limburg KE, Mehrrens H, Montes I, Naqvi W, Paulmier A, Pfeil B, Pitcher G, Pouliquen S, Rabalais N, Rabouille C, Recape V, Roman M, Rose K, Rudnick D, Rummer J, Schmechtig C, Schmidtke S, Seibel B, Slomp C, Sumalia UR, Tanhua T, Thierry V, Uchida H, Wanninkhof R and Yasuhara M (2021) A Global Ocean Oxygen Database and Atlas for Assessing and Predicting Deoxygenation and Ocean Health in the Open and Coastal Ocean. *Front. Mar. Sci.* 8:724913. doi: 10.3389/fmars.2021.724913
- Copyright © 2021 Grégoire, Garçon, Garcia, Breitburg, Isensee, Oschlies, Telszewski, Barth, Bittig, Carstensen, Carval, Chai, Chavez, Conley, Coppola, Crowe, Currie, Dai, Deflandre, Dewitte, Diaz, Garcia-Robledo, Gilbert, Giorgetti, Glud, Gutierrez, Hosoda, Ishii, Jacinto, Langdon, Lauvset, Levin, Limburg, Mertens, Montes, Naqvi, Paulmier, Pfeil, Pitcher, Pouliquen, Rabalais, Rabouille, Recape, Roman, Rose, Rudnick, Rummer, Schmechtig, Schmidtke, Seibel, Slomp, Sumalia, Tanhua, Thierry, Uchida, Wanninkhof and Yasuhara. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

APPENDIX

TABLE A1 | Characteristics of each generation of floats: type of sensors, known issues, drift correction, number of floats equipped with O₂ sensors, number of profiles (in June 2020) and data streams.

Generation of floats	Sensors type	Known issues	Drift correction	# floats	# profiles	# RT (1)	# A (2)	# DM (3)
First Generation	SBE43 sensors	Sensor stability and drift ⁽⁴⁾	A gain and offset formulation established in stable deep waters ⁽⁵⁾	160	26921	9972	0	16949
Second Generation	Aanderaa 3830/3835 4330/4831	Inadequate temperature compensation and/or distorted, incorrect O ₂ response especially at low to intermediate O ₂ levels ⁽⁶⁾	Either a gain and offset formulation established in stable deep waters ⁽⁵⁾ or a temperature-dependent O ₂ gain ⁽⁶⁾	598	111520	41929	2730	66861
Third Generation	Aanderaa 4330/4831-SBE63, RINKO, ARO-FT	Linear O ₂ sensitivity drift ⁽⁶⁾	Drift correction by an O ₂ gain factor using the surface p O ₂ estimated either from in-air measurement or estimated from climatologies at the ARGO-measured Temperature and salinity ^(6,7)	498	50464	22187	4883	23394

⁽¹⁾Real-Time, available 24 h after profiling; ⁽²⁾Adjusted, RT data after automated QC; ⁽³⁾Delayed-Mode, best quality data including realistic error estimates, sophisticated data adjustments and QC procedures, manual inspection by either the float's PI or pre-identified Delayed Mode (DMQC) expert (e.g. Bittig et al., 2019); ⁽⁴⁾Gruber et al., 2010; ⁽⁵⁾Takeshita et al., 2013; ⁽⁶⁾Bittig et al., 2018; ⁽⁷⁾Thierry and Bittig, 2018.