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Coupled CH₄ production and oxidation support CO₂ supersaturation in a tropical flood pulse lake (Tonle Sap Lake, Cambodia)

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Carbon dioxide (CO₂) supersaturation in lakes and rivers worldwide is commonly attributed to terrestrial-aquatic transfers of organic and inorganic carbon (C) and subsequent, in situ aerobic respiration. Methane (CH₄) production and oxidation also contribute CO₂ to freshwaters, yet this remains largely unquantified. Flood pulse lakes and rivers in the tropics are hypothesized to receive large inputs of dissolved CO₂ and CH₄ from floodplains characterized by hypoxia and reducing conditions. We measured stable C isotopes of CO₂ and CH₄, aerobic respiration, and CH₄ production and oxidation during two flood stages in Tonle Sap Lake (Cambodia) to determine whether dissolved CO₂ in this tropical flood pulse ecosystem has a methanogenic origin. Mean CO2 supersaturation of 11,000 \pm 9,000 μ atm could not be explained by aerobic respiration alone. 13 C depletion of dissolved CO₂ relative to other sources of organic and inorganic C, together with corresponding ¹³C enrichment of CH₄, suggested extensive CH₄ oxidation. A stable isotope-mixing model shows that the oxidation of ¹³C depleted CH₄ to CO₂ contributes between 47 and 67% of dissolved CO_2 in Tonle Sap Lake. ¹³C depletion of dissolved CO_2 was correlated to independently measured rates of CH₄ production and oxidation within the water column and underlying lake sediments. However, mass balance indicates that most of this CH₄ production and oxidation occurs elsewhere, within inundated soils and other floodplain habitats. Seasonal inundation of floodplains is a common feature of tropical freshwaters, where high reported CO₂ supersaturation and atmospheric emissions may be explained in part by coupled CH₄ production and oxidation.

carbon dioxide supersaturation | methane oxidation | flood pulse | lake

lobally, most lakes and rivers are supersaturated with dis-Goodily, most large and inversion of superscript (CO_2) relative to the atmosphere, highlighting their outsized role in transferring and transforming terrestrial carbon (C) (1-3). Terrestrial-aquatic transfers of C can include CO₂ dissolved in terrestrial ground and surface waters (3-6), dissolved inorganic carbon (DIC) from carbonate weathering (7, 8), or organic C from various sources that is subsequently respired in lakes and rivers (9, 10). Initially, oceanic export was thought to be the only fate for terrestrial-aquatic transfers of C, but a growing body of research on sediment burial of organic C and CO₂ emissions from freshwaters prompted the "active pipe" revision to this initial set of assumptions (11). Although freshwaters are now recognized as focal points for transferring and transforming C on the landscape, most of this research has been conducted within temperate freshwaters (2, 11, 12). Few studies focus on the mechanisms of CO2 supersaturation in tropical lakes and rivers, with most conducted in just one watershed, the Amazon (4, 13-15).

 CO_2 supersaturation within tropical freshwaters is likely influenced by their unique flood pulse hydrology. The canonical flood pulse concept hypothesizes that annual flooding of riparian land will lead to organic C mobilization and respiration (16). Partial pressures of CO_2 (pCO_2) have been measured in excess of 44,000 μ atm in the Amazon River (13), 16,000 μ atm in the Congo River (17), and 12,000 μ atm in the Lukulu River (17). Richey et al. (13), Borges et al. (18), and Zuidgeest et al. (17) have each shown that that riverine pCO_2 scales with the amount of land flooded in these watersheds. Yet it was only recently that Abril and Borges (19) proposed the importance of flooded land to the "active pipe." These authors differentiate uplands that unidirectionally drain water downhill (via ground and surface water) from floodplains that bidirectionally exchange water with lakes and rivers (19). They conceptualize how floodplains combine high hydrologic connectivity, high rates of primary production, and high rates of respiration to transfer relatively large amounts of C to tropical freshwaters (19).

Methanogenesis inevitably results on floodplains after dissolved oxygen (O₂) and other electron acceptors for anaerobic respiration such as iron and sulfate are consumed (16, 19). Horizontal gradients in dissolved O₂ and reducing conditions have been observed extending from the center of lakes and rivers through their floodplains in the Mekong (20, 21),

Significance

Freshwaters inextricably link flows of carbon between the land, oceans, and atmosphere. Resulting carbon dioxide supersaturation relative to the atmosphere in most of the world's lakes and rivers has long been assumed to come from aerobic respiration. Although carbon dioxide also comes from the oxidation of anaerobically produced methane, this has been largely ignored within freshwaters. Here, we use stable carbon isotopes of carbon dioxide and methane to show that a nontrivial proportion of the total dissolved carbon dioxide in a tropical flood pulse lake comes from methane oxidation. Seasonal pulses of flooding are common in the tropics, suggesting that coupled methane production and oxidation likely contribute more broadly to flows of carbon between the land, understudied tropical freshwaters, and atmosphere.

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Table 1. Mean partial pressures (μ atm) \pm 1 SD and δ^{13} C () \pm 1 SD for CO₂ and CH₄ in open water, edge, and floodplain environments of TSL during the high-water and falling-water stages of the flood pulse

	pCO_2 (μ atm)	$ ho$ CH $_4$ (μ atm)	δ ¹³ C-CO ₂ (‰)	δ ¹³ C-CH ₄ (‰) n	n
High					
All environments	13,000 ± 6,000	11,000 ± 2,000	-40 ± 7	-36 ± 2	35
Open	12,000 ± 4,000	3,000 ± 2,000	-37 ± 4	-45 ± 9	6
Edge	14,000 ± 6,000	20,000 ± 10,000	-39 ± 6	-38 ± 6	6
Floodplain	$13000 \pm 6,000$	$11,000 \pm 2,000$	-41 ± 7	-34 ± 2	23
Falling					
All environments	13,000 ± 12,000	600 ± 300	-38 ± 5	-62 ± 5	12
Open	14,000 ± 13,000	50 ± 20	-35 ± 3	-57 ± 6	6
Edge	14,000 ± 12,000	1,400 ± 800	-40 ± 6	-67 ± 7	6

Mean partial pressures (μ atm) and δ^{13} C (%) across all lake environments for the high-water and falling-water stages are also shown.

Congo (22), Pantanal (23), and Amazon watersheds (4). CH₄ production and oxidation occur along such redox gradients (4, 16, 19, 23). CH₄ is produced by acetate fermentation (Eq. 1) and carbonate reduction (Eq. 2) within freshwaters (24, 25). CH₄ production coupled with aerobic oxidation results in CO₂ (Eq. 3 and ref. 25), yet no studies have quantified the relative contribution of coupled CH₄ production and oxidation to CO₂ supersaturation within tropical freshwaters.

$$CH_3COOH \rightarrow CO_2 + CH_4,$$
 [1]

$$CO_2 + 8H^+ + 8e^- \rightarrow CH_4 + 2H_2O,$$
 [2]

$$CH_4 + 2O_2 \rightarrow CO_2 + 2H_2O.$$
 [3]

The relative contribution of coupled CH₄ production and oxidation to CO₂ supersaturation within tropical freshwaters can be traced with stable C isotopes of CO₂ and CH₄. Methanogenesis results in CH₄ that is depleted in ¹³C (δ^{13} C = -65 to -50% from acetate fermentation and -110 to -60% from carbonate reduction) compared to other potential sources of organic and inorganic C (δ^{13} C = -37 to -7.7%; see *Materials* and *Methods*) (24–26). The oxidation of this ¹³C-depleted CH₄ results in ¹³C-depleted CO₂ (24–26). At the same time, CH₄ oxidation enriches the ¹³C/¹²C of residual CH₄ as bacteria and archaea preferentially oxidize ¹²C-CH₄ (25). This means that the ¹³C/¹²C of CO₂ and CH₄ can serve as powerful tools to determine the source of CO₂ supersaturation within freshwaters.

Tonle Sap Lake (TSL) is Southeast Asia's largest lake and an understudied flood pulse ecosystem that supports a regionally important fishery (21, 22, 27). Each May through October, monsoonal rains and Himalayan snowmelt increase discharge in the Mekong River and cause one of its tributaries, the Tonle Sap River, to reverse course from southeast to northwest (21). During this course reversal, the Tonle Sap River floods TSL. The TSL flood pulse increases lake volume from 1.6 to 60 km³ and inundates 12,000 km² of floodplain for 3 to 6 mo per year (21, 27). Holtgrieve et al. (22) have shown that aerobic respiration is consistently greater than primary production in TSL (i.e., net heterotrophy), with the expectation of consistent CO_2 supersaturation. But, the partial pressures, C isotopic compositions, and ultimately the source of dissolved CO_2 in TSL remain unquantified.

To quantify CO_2 supersaturation and its origins in TSL, we measured the partial pressures of CO_2 and CH_4 and compared their C isotopic composition to other potential sources of organic and inorganic C. We carried out these measurements in distinct lake environments during the high-water and fallingwater stages of the flood pulse, hypothesizing that CH_4 production and oxidation on the TSL floodplain would support CO_2 supersaturation during the high-water stage. We found that coupled CH_4 production and oxidation account for a nontrivial proportion of the total dissolved CO_2 in all TSL environments and during both flood stages, showing that anaerobic degradation of organic C at aquatic–terrestrial transitions can support CO_2 supersaturation within tropical freshwaters.

Results

pCO₂ and pCH₄ in TSL were consistently supersaturated relative to atmospheric equilibrium. pCO₂ averaged 13,000 ± 6,000 μ atm (mean ± 1 SD) across sites during the high-water stage and 13,000 ± 12,000 μ atm during the falling-water stage (Table 1). pCH₄ was significantly greater during the high-water stage (11,000 ± 2,000 μ atm) than during the falling-water stage (600 ± 300 μ atm) (P < 0.001, d = 1.8). By contrast, pCO₂ and pCH₄ at sea level are ~400 and 1.8 μ atm, respectively.

CO₂ supersaturation exceeded dissolved O₂ deficits, indicating sources of dissolved CO₂ other than aerobic respiration (Fig. 1 A and B). CO₂ supersaturation is expected to vary with dissolved O₂ deficits in a -1/1 O₂:CO₂ ratio as one μ micromole of dissolved O₂ is consumed for each μ micromole of dissolved CO₂ produced. Instead, ratios of -0.1/1 were observed during both the high-water and falling-water stages. During the high-water stage, the greatest CO₂ supersaturation occurred under the most hypoxic conditions (Fig. 1A).

The intercept of the relationship between 1/CO₂ and δ^{13} C-CO₂ can be used to determine the source of dissolved CO₂ (Keeling Intercepts; *SI Appendix*, Table S1) (28, 29). In TSL, the inverse of *p*CO₂ was strongly correlated with ¹³C depletion of CO₂. The intercept of δ^{13} C-CO₂ was as low as $-51\%_{00}$ during the high-water stage and $-43\%_{00}$ during the falling-water stage. This indicates a ¹³C-depleted source of dissolved CO₂ relative to the other potential sources of organic and inorganic C measured, which ranged from -37 to $-7.7\%_{00}$ (Fig. 24). Observed ¹³C depletion of dissolved CO₂ coincided with ¹³C enrichment of dissolved CH₄ (Table 1 and Fig. 3 *A* and *B*). Acetate fermentation produces δ^{13} C-CH₄ ranging from -65 to $-50\%_{00}$ and carbonate reduction produces δ^{13} C-CH₄ ranging from -110 to $-60\%_{00}$ (24–26). By contrast, δ^{13} C-CH₄ averaged $-36 \pm 2\%_{00}$ during the high-water stage. During this flood stage, dissolved CO₂ became more ¹³C depleted, and dissolved CH₄ became more ¹³C enriched from open water environments (δ^{13} C-CO₂ = $-37 \pm 4\%_{00}$, δ^{13} C-CH₄ = $-45 \pm 9\%_{00}$) to edge environments (δ^{13} C-CO₂ = $-39 \pm 6\%_{00}$, δ^{13} C-CH₄ = $-38 \pm 6\%_{00}$) to floodplain environments (δ^{13} C-CO₂ = $-41 \pm 7\%_{00}$, δ^{13} C-CH₄ (simply, δ^{13} C-CO₂ – δ^{13} C-CH₄) of typically <10\%_{00} in TSL indicates substantial CH₄ oxidation (25) (Fig. 3 *A* and *B*).

A two-source, stable isotope-mixing model for δ^{13} C-CO₂ was used to estimate fractional contributions to dissolved CO₂ by CH₄ oxidation, compared with other potential sources of organic and inorganic C (Fig. 24). Assuming oxidation of CH₄



Fig. 1. Dissolved O_2 deficit and CO_2 supersaturation, relative to atmospheric equilibrium in open water, edge, and floodplain environments of TSL (A) during the high-water and falling-water stages of the flood pulse (B). Dissolved O_2 deficit and CO_2 supersaturation are calculated as the difference between atmospheric equilibrium, according to Henry's Law. Orange lines show atmospheric equilibrium at a dissolved O_2 deficit and CO_2 supersaturation of 0 μ mol \cdot L⁻¹. A slope (m) of -1.0 represents the equimolar consumption of dissolved O_2 and production of dissolved CO_2 expected during aerobic respiration (black dashed line). Instead, a slope of -0.1 was observed during both the high-water and falling-water stages. O_2 deficits were strongly correlated to CO_2 supersaturation during the high-water stage, but there was no such correlation during the falling-water stage.

produced by acetate fermentation only, the fractional contributions by CH₄ oxidation to dissolved CO₂ range from 63 to 85% across the distinct lake environments and flood stages of TSL (*SI Appendix*, Table S3). Assuming oxidation of CH₄ produced by both acetate fermentation and carbonate reduction, these contributions by CH₄ oxidation fall to a more conservative 47 to 67%. Apparent fractionation between δ^{13} C-CO₂ and δ^{13} C-CH₄ (simply, δ^{13} C-CO₂/ δ^{13} C-CH₄) of typically <1.055 in TSL indicate substantial CH₄ production by acetate fermentation with some carbonate reduction (24, 25) (Fig. 2*B*).

 δ^{13} C-CO₂ was strongly correlated to independent measurements of net CH₄ oxidation in the water column during the high-water stage (Fig. 3 *C* and *D*). The same significant relationship was observed between δ^{13} C-CO₂ and gross CH₄ production within the sediments (Fig. 3 *E* and *F*). Despite these relationships, CO₂ mass balance indicates that CH₄ production and oxidation within the water column and underlying sediments contribute at most 9% to dissolved CO₂ in TSL (*SI Appendix*, Table S4). Of these two processes, CH₄ production contributes one to two orders of magnitude more CO₂ than CH₄ oxidation. Other processing of C within the water column

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and underlying sediments, such as aerobic respiration, also contribute a relatively small share of total dissolved CO_2 (13 ± 8%).

Discussion

Contributions of CH₄ production and oxidation to CO₂ supersaturation are understudied within tropical freshwaters, where extensive flooding, dissolved O₂ deficits, and reducing conditions at aquatic–terrestrial transitions make such contributions likely. The subtropics and tropics are home to many high-order flood pulse rivers, such as the Amazon, Orinoco, Congo, Zambezi, and Mekong, which are collectively responsible for over 30% of global mean annual discharge (30). Along this tropical "active pipe" lays 52% of the world's floodplains, transferring and transforming C at relatively high rates (20, 31). Using a combination of isotopic tracers and mass balance, we show that a substantial fraction this transfer and transformation of C occurs through coupled CH₄ production and oxidation in TSL.

A majority of our measured δ^{13} C-CO₂ fell between the ¹³C-depleted CO₂ known to result from CH₄ oxidation and



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flood pulse lake (Tonle Sap Lake, Cambodia)



Fig. 2. (*A*) Measured δ^{13} C-CO₂ (blue) relative to other potential sources of organic and inorganic C during the high-water and falling-water stages of the flood pulse. "Other" potential sources of organic and inorganic C measured by this study in TSL include macrophytes, terrestrial C3 vegetation, periphyton, phytoplankton, and DIC. Emergent aquatic C4 grasses, measured by Hedges et al. (48) in the Amazon, and atmospheric CO₂ in equilibrium with water (47) are also included. Isotopic values quantified by this study are in blue and white, and those quantified by other studies (24–26, 47, 48) are in gray. (*B*) Apparent fractionation between δ^{13} C-CO₂ and δ^{13} C-CO₄ (α_{app}) indicated substantial CH₄ production through acetate fermentation with some carbonate reduction in TSL (24, 25). Therefore, a two-source, isotope-mixing model was created using 1) a continuous uniform distribution of δ^{13} C-CO₂ from DIC and the aerobic respiration of potential organic C sources (white box).



Fig. 3. δ^{13} C-CO₂ and δ^{13} C-CH₄ in open-water, edge, and floodplain environments of TSL during the high-water (A) and falling-water stages of the flood pulse, modified from Whiticar (25) (*B*). Zones of CH₄ production by acetate fermentation, CH₄ production by carbonate reduction, and CH₄ oxidation based on apparent fractionation between δ^{13} C-CO₂ and δ^{13} C-CH₄ (ϵ_c) are shaded in gray. ¹³C depletion of CO₂ was strongly correlated to independent measurements of net CH₄ oxidation in the water column during the high-water stage (*C*), though not during the falling-water stage (*D*). ¹³C depletion of CO₂ was also strongly correlated to gross CH₄ production within sediments during the high-water stage (*E*), though not during the falling-water stage (*F*).

the relatively more ¹³C-enriched phytoplankton, periphyton, macrophytes, and terrestrial C3 vegetation measured in TSL (Fig. 2A). Because there is little fractionation during aerobic respiration of organic C, measured δ^{13} C-CO₂ in lakes can be expected to fall inside the range of δ^{13} C observed for commonly considered sources of organic and inorganic C (31, 32). Instead, our observed δ^{13} C-CO₂ fell outside of this range. Potential sources of organic and inorganic C in TSL ranged from $\delta^{13}C = -37\%_{00}$ for macrophytes to $\delta^{13}C = -7.7\%_{00}$ for atmospheric CO₂ in equilibrium with water. De Kluiver and others (33, 34) have reported relatively ¹³C-depleted phytoplankton ($\delta^{13}C = -41\%$) SI Appendix, Table S2). However, the net heterotrophy and CO₂ supersaturation consistently observed in TSL and other lakes (34) makes substantial contributions to dissolved CO₂ from aquatic primary producers such as phytoplankton unlikely, because these ecosystems are inferred to receive greater inputs of terrestrial organic C than aquatic organic C. Accordingly, the δ^{13} C of dissolved CO₂ measured in the same study by De Kluiver et al. (34) ranges from -21 to -9%, suggesting that the aerobic respiration of relatively ¹³C-depleted phytoplankton in net heterotrophic lakes does not substantially impact δ^{13} C-CO₂. Furthermore, our most ¹³C-depleted dissolved CO_2 was sampled on the TSL floodplain, where the water column and underlying sediments are largely shaded by macrophytes and other emergent vegetation, limiting phytoplankton production (20). Ultimately, methanogenesis is the only possible source of the ¹³C-depleted, dissolved CO₂ observed in TSL. We can therefore use a twosource, stable isotope-mixing model to estimate relative contributions to dissolved CO₂ by 1) potential sources of organic and inorganic C and 2) CH₄ oxidation. This mixing model shows that CH₄ oxidation contributes between 47 and 67% of dissolved C-CO₂ across the distinct lake environments and flood stages of TSL, which is unprecedented in the aquatic C-cycling literature.

High-CO₂ supersaturation and an imbalance with dissolved O_2 such as we observed in TSL (Fig. 1 *A* and *B*) have previously been attributed to autotrophic and heterotrophic respiration of macrophytes and other emergent aquatic vegetation on flooded land (15, 16, 17, 35). Macrophytes and other emergent aquatic vegetation fix primarily atmospheric CO₂, acting more as terrestrial primary producers than aquatic primary producers. Melack and Engle (35) have shown that floating macrophytes dominate primary production and provide the bulk of organic C to an Amazon floodplain lake. Abril et al. (15) have further suggested that floodplain and riparian wetland

vegetation in the Amazon could export fully half of its primary production on an annual basis. Data from TSL supports a more nuanced interpretation. The most ¹³C-depleted source of organic C in TSL was an individual macrophyte ($\delta^{13}C = -37\%_{oo}$, mean $\delta^{13}C = -33 \pm 4\%_{oo}$). Even so, 70% of our dissolved CO₂ measurements were depleted in ¹³C below $-37\%_{oo}$. As confirmed by our stable isotope-mixing model, this means that aerobic respiration of macrophytes can contribute to but not explain the C isotopic depletion of dissolved CO₂ observed in TSL.

Corresponding ¹³C enrichment of dissolved CH₄ indicated a fractionating loss process, further supporting the interpretation that CH₄ oxidation supports CO₂ supersaturation in TSL. Acetate fermentation within tropical lake sediments from the Amazon and Pantanal has been shown to produce δ^{13} C-CH₄ values ranging from -86 to -61% (36, 37). The same studies showed concurrent carbonate reduction producing CH₄ even more depleted in ¹³C (36, 37). By contrast, we measured an overall mean δ^{13} C-CH₄ of -43 ± 9% in TSL, with some values as high as -11% (Table 1 and Fig. 3 *A* and *B*). Similar values were measured by Barbosa et al. (38) on Amazon River floodplains (δ^{13} C-CH₄ = -70.1 to -14.8%). Independently measured rates of CH₄ production and oxidation in TSL support this conclusion. Both net CH₄ oxidation in the water column of TSL (Fig. 3 *B* and *C*) and gross CH₄ production within the sediments (Fig. 3 *D* and *E*) were strongly correlated to δ^{13} C-CO₂.

Despite these relationships, CH_4 production and oxidation and aerobic respiration within the water column and underlying sediments typically contribute less than 15% of dissolved CO₂ in TSL (*SI Appendix*, Table S4). In our mass balance, we solve for CO₂ advected from elsewhere, within inundated soils and other floodplain habitats, and infer that this is a far greater contributor to CO₂ supersaturation. This was initially hypothesized by Junk et al. (18) and later combined with the "active pipe" by Abril and Borges (20). Yet it has been empirically tested using dissolved CO₂ and CH₄ in only two other locations (15, 17) and never with the C isotopic composition of these dissolved gases.

never with the C isotopic composition of these dissolved gases. The ¹³C depletion of CO₂, ¹³C enrichment of CH₄, and their correlations to independently measured rates of CH₄ production and oxidation suggest that these coupled processes support CO₂ supersaturation in TSL. By extension, coupled CH₄ production and oxidation are disproportionately responsible for CO_2 emissions from TSL. Lauerwald et al. (13) estimate that >50% of global riverine CO₂ emissions occur in the tropics, emphasizing the importance of tropical "active pipes." Data on the stable \breve{C} isotopes of both CO_2 and CH_4 are rarely reported for freshwaters, though ¹³C-enriched dissolved CH_4 (>-50_{\lowed{c}}) reported in tropical and temperate lakes, wetlands, peatlands, and the Amazon River implies widespread oxidation of CH4 to CO₂ (SI Appendix, Table S5). Coupled CH₄ production and oxidation have thus been understudied but may support CO₂ supersaturation and CO₂ emissions from other tropical freshwaters with large amounts of seasonally or perennially flooded land. The extent of this flooding will most likely change under the twin stressors of hydropower development and climate change in the tropics (21), impacting the future role of floodplains in the transfer and transformation of C from terrestrial to aquatic ecosystems.

Materials and Methods

Field Sampling. Field sampling was conducted during the high-water and falling-water stages of the annual flood pulse in October 2015 and March 2016, respectively, representing the typical hydrological range in TSL. Flood stages were assessed using historical data from a gauging station at Kampong Luong (*SI Appendix*, Fig. S1) (21). Sampling focused on three locations in the southwest (Kampong Preah), central (Anlang Reang), and northwest (Prek Konteil) basins of TSL. Transects designed to capture horizontal gradients in dissolved O₂ and reducing conditions were established at each location. These transects consisted of six points extending through the distinct open water (Transect Point 1), edge (Transect Point 2), and floodplain environments of TSL (Transect Points 4 to 6). The edge environments were characterized by a transition from open water environments to emergent, permanently rooted floodplain vegetation.

Partial Pressures of CO₂ and CH₄. Partial pressures of CO₂ and CH₄ at each transect point and flood stage in TSL were quantified as the average of three duplicates collected at 0.1 m below the water surface and at 0.5 m above the lake bottom where water depth exceeded 0.5 m (n = 143 duplicates, n = 47 replicates). Water was collected into 74-mL gas-tight serum bottles using a van Dorn sampler, preserved in the field with 74 μ L of 50% mass/volume zinc chloride solution, and placed on ice for transport to the Royal University of Phnom Penh, where they were stored at 4°C until analysis. For analysis, samples were displaced with helium to roughly equal parts headspace and water, left to equilibrate for 12 h, and analyzed for headspace pCO₂ and pCH₄ using gas chromatography (SRI 8610c GC) by referencing to certified standards of known concentrations.

Stable C Isotopes of CO₂ and CH₄. Following analysis for partial pressures, samples were resealed with Apiezon grease, inverted, placed on ice, and transported to the University of Washington for C isotopic analysis (n = 47). A 20-mL headspace sample was analyzed for the ¹³C/¹²C of CO₂ and CH₄ simultaneously using a cavity ring-down spectrometer (Picarro G2201*i*) with a small sample introduction module (Picarro A0314 SSIM). Following Malowany et al. (39), a column of reduced copper shavings was installed on the small sample introduction module to eliminate interference by hydrogen sulfide with isotopic measurements. Samples exceeding 300 μ atm CH₄ were diluted with ultra-high purity nitrogen to further eliminate interference by high concentrations of this gas with isotopic measurements. Stable C isotopes of CO₂ and CH₄ are each expressed in delta (δ) notation relative to Vienna Pee Dee Belemnite by referencing to certified CO₂ and CH₄ standards of known concentrations and ¹³C/¹²C.

Stable C Isotopes of Organic and Inorganic C. Grab samples of floating macrophytes (Eichhornia species), terrestrial C3 vegetation, periphyton, and phytoplankton were collected across the distinct lake environments and flood stages of TSL, combined, and considered a single, lake-wide sample with a minimum of four replicates. Phytoplankton were collected using a Wisconsin net sampler (Wildco 40-A50), and periphyton was scraped from the benthos and the surfaces of floating macrophytes and emergent, aquatic vegetation. Macrophytes ($\delta^{13}C = -33 \pm 4_{00}^{\circ}$, n = 4), terrestrial C3 vegetation ($\delta^{13}C = -29 \pm 10^{10}$ 2_{00}° , n = 7), periphyton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18), and phytoplankton ($\delta^{13}C = -28 \pm 4_{00}^{\circ}$, n = 18, $-24 \pm 4^{\circ}_{00}$, n = 6) in TSL were freeze dried, ground, and analyzed for bulk ¹³C/¹²C using an elemental analyzer (CE Instruments 2500 NA) interfaced with an isotope ratio mass spectrometer (DeltaV IRMS). Laboratory working standards were glutamic acid 1 (δ^{13} C = -28.3% versus VPDB), glutamic acid 2 (δ^{13} C = -13.7%), and sockeye salmon ($\delta^{13}C = -21.3$ %). DIC ($\delta^{13}C = -13.8 \pm 0.4$ %, n =98) samples from another sampling effort across the same lake environments and flood stages were acidified, displaced with a helium headspace, analyzed on a DeltaV IRMS, and considered a lake-wide sample, as described previously.

Depth-Integrated Gross Primary Production and Aerobic Respiration. Gross primary production (GPP) and aerobic respiration were modeled across the distinct lake environments and flood stages of TSL using diel-dissolved O2 data in the "LakeMetabolizer" R package (n = 16) (40, 41). Model inputs include hourly dissolved oxygen (millimoles per liter), hourly water temperature (degrees Celsius), and hourly photon flux for photosynthetically active radiation (PAR; microeinsteins per second · per square meter). Continuously logging dissolved O2 and water temperature sensors were deployed for a minimum of 20 h (Precision Measurement Engineering miniDO₂T Logger, accuracy \pm 0.16 mg, O₂ L⁻¹, and \pm 0.1 °C). Accuracy of dissolved O₂ sensors was verified prior to field deployment using the Winkler titration method. PAR was not measured directly but calculated from full-spectrum irradiance based on latitude, longitude, aspect, slope, transmissivity data, and the "astrocalc4r" function in the "fishmethods" R package (42). GPP and aerobic respiration were converted to millimoles of CO2 per cubic meter per day using an assimilation efficiency of 1.2 for photosynthesis (43, 44) and a conversion efficiency of 1.0 for respiration.

Volumetric rates were multiplied by mixing depths to obtain areal rates in terms of millimoles of CO₂ per square meter per day. Mixing depths were evaluated with dissolved O₂ profiles at each site using a multiparameter sonde calibrated just prior to deployment with water-saturated air (YSI 6920). Dissolved O₂ data were plotted over depth (m), smoothed using a loess-spanning function of 0.2, and interrogated for inflection points in R (41). The depth of these inflection points at each transect was considered the mixing depth.

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Depth-Integrated CH₄ Production and Oxidation. Gross CH₄ production within lake sediments was guantified as the average of three duplicate sediment incubations. At each transect point and flood stage in TSL, sediment cores were taken with a stainless-steel corer. The upper 1 cm³ of each core was sealed inside a 74-mL gas-tight serum bottle (n = 72 duplicates, n = 24 replicates). The remaining volume of the bottle was filled with bottom water collected 0.5 m above the sediments. Three additional bottles were filled with bottom water only and three with water collected 0.1 m below the water surface. All bottles were incubated at ambient air temperatures (25 to 33 4 °C), which were typically <4 °C different from water temperatures in TSL and sampled daily from a helium headspace for 7 d. pCH₄ was analyzed as described previously and corrected for progressively decreasing headspace:water ratios. Net CH₄ oxidation in surface waters was multiplied by mixing depths to obtain areal rates as in Depth Integrated Gross Primary Production and Aerobic Respiration. Net oxidation in bottom waters was added to net CH₄ production measured in the bottles containing a combination of sediment cores and bottom water and considered gross CH₄ production. Following incubation, each sediment core was dried at 100 °C for 3 h and weighed. Gross CH₄ production rates were then corrected for sediment core weight and scaled to nanomoles of CH₄ per cubic meter per day. Previously published studies of CH₄ production in lake sediment cores show that rates measured at the sediment-water interface are consistent to a sediment depth of 0.1 m (25, 45). Volumetric rates of CH₄ production were thus multiplied by 0.1 m to obtain areal rates in terms of nanomoles of CH₄ per square meter per day. Because one mole of CO₂ is produced for each mole of CH₄ produced during acetate fermentation, presumed to be dominant in TSL (Fig. 2B) and within freshwaters more broadly (24, 25), rates were also considered in terms of nanomoles of CO2 per square meter per day. Each transect sampled included negative control incubations amended with a 74 μ L of 50% mass/volume zinc chloride solution.

Mass Balance. A mass balance for dissolved CO_2 in TSL was created from processes resulting in a gain or loss of CO_2 :

 $\mathsf{CO}_{2,\mathsf{Measured}} = \mathsf{CO}_{2,\mathsf{Advected}} + \mathsf{CO}_{2,\mathsf{Respiration}} - \mathsf{CO}_{2,\mathsf{GPP}} + \mathsf{CO}_{2,\mathsf{MProd}} + \mathsf{CO}_{2,\mathsf{MOx}},$

where CO_{2,Respiration} is the CO₂ gained from modeled aerobic respiration, CO_{2,GPP} is the CO₂ lost from modeled GPP, CO_{2,MProd} is the CO₂ gained from measured gross CH₄ production within sediments, and CO_{2,MOx} is the CO₂ gained from measured net CH₂ oxidation in the water column, each in millimoles per square meter per day. CO_{2,Measured} is the pCO₂ measured within the water column of TSL and multiplied by a temperature dependent Henry's constant and mixing depth to yield millimoles of dissolved CO2 per square meter on the day of sampling. Diffusion of CO₂ from TSL to the atmosphere was modeled using CO_{2.Measured} following Cole and Caraco (46). CO2 diffusion reflects an atmospheric loss subsequent to CO2, Measured and was ultimately excluded from the mass balance. CO2, Advected is the remaining CO_2 in the mass balance assumed to result from aerobic respiration and anaerobic degradation of organic C elsewhere, within inundated soils and other floodplain habitats under steady-state conditions (millimoles per square meter per day). Mean daily $CO_{2,Advected} \pm 1$ SE was quantified using normal distributions-based on sample size, mean, and SD-of other terms in the mass balance over 10,000 Monte Carlo simulations in R (41).

Stable, Isotope-Mixing Model. The C isotopic composition of CO₂ measured in TSL fell between the ${}^{13}C{}^{12}C$ produced by 1) the oxidation of ${}^{13}C{}^{-depleted} CH_4$ to CO₂ (-110 to -50‰) and 2) that of other potential organic and inorganic sources of CO₂ (-37 to -7.7‰). Here, the sole concern is the fraction of CO₂ derived from CH₄ oxidation. Thus, a two-source ("Methane" versus "Other"), stable, isotope-mixing model was deemed appropriate. The model also accounted for CO₂ losses from primary production and atmospheric diffusion and took the form:

$$\begin{split} \delta^{13}\text{CO}_2 &= \left(\delta^{13}\text{C}_{\text{Methane}} \cdot \textbf{f}_{\text{Methane}} \right) + \left(\delta^{13}\text{C}_{\text{Other}} \cdot \textbf{f}_{\text{Other}} \right) \\ &- \left(\left(\delta^{13}\text{CO}_{2,\text{Measured}} + \epsilon_{\text{GPP}} \right) \cdot \textbf{f}_{\text{GPP}} \right) \\ &- \left(\left(\delta^{13}\text{CO}_{2,\text{Measured}} + \epsilon_{\text{Diffusion}} \right) \cdot \textbf{f}_{\text{Diffusion}} \right) \end{split}$$

where $\Sigma f_i = 1.00. \delta^{13} C_{Methane}$ was modeled as a continuous, uniform distribution of $\delta^{13}C$ -CH₄ values produced by methanogenesis, ranging from -110 to -50‰ (Fig. 2A) (24–26). f_{Methane} is the fraction of CO₂ resulting from CH₄

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oxidation. Because $\delta^{13}C_{\text{Methane}}$ encompasses the range of $\delta^{13}C$ values produced by both acetate fermentation (-65 to -50‰)—presumed to be dominant in TSL and within freshwaters more broadly (24, 25)—and carbonate reduction (-110 to -60‰), the model results in a conservative estimate of f_{Methane} for this freshwater lake (Fig. 2*B*).

 $δ^{13}C_{Other}$ was also modeled as a continuous, uniform distribution of $\delta^{13}C$ values encompassing other potential sources of organic and inorganic C (*SI* Appendix, Fig. S2). This distribution ranges from the most ${}^{13}C$ -depleted source of organic C measured in TSL, macrophytes ($\delta^{13}C = -37\%_0$), to the most ${}^{13}C$ -enriched source of inorganic C, atmospheric CO₂ in equilibrium with water ($\delta^{13}C = -7.7\%_0$) (47). $\delta^{13}C_{Other}$ therefore encompasses the $\delta^{13}C$ of terrestrial C3 vegetation, periphyton, phytoplankton, and DIC measured in TSL and the $\delta^{13}C$ of emergent, aquatic C4 grasses ($\delta^{13}C = -12.2 \pm 0.3\%_0$) measured by Hedges et al. (48) in the Amazon. With multiple sources of organic and inorganic C that overlap in $\delta^{13}C$ and no prior information on the relative importance of each, the most parsimonious option was to treat them as a group with equal probability across the full range of $\delta^{13}C$ values. However, multiple alternative models were also tested (*SI Appendix*, Table S3).

 $\delta^{13} C_{\text{CO}_2,\text{Measured}}$ and mass-dependent fractionations for photosynthesis (f_{GPP}) and diffusion to the atmosphere (f_Diffusion) in the model were quantified by this study and its mass balance (*SI Appendix*, Table S4). The kinetic fractionation factors for photosynthesis and diffusion, ϵ_{GPP} and $\epsilon_{\text{Diffusion}}$, are -19 and $-1.1_{\text{oor}}^{\prime\prime}$ respectively (47). Following the IsoSource-mixing model by Phillips et al. (49), f_Methane and f_Other were assigned possible values between 0.00 and 1.00 by 0.05, and was solved for iteratively in R (41). If the resulting $\delta^{13}\text{CO}_2 = \delta^{13}\text{CO}_2,\text{Measured} \pm 1_{\text{oo}}^{\prime\prime}$ and f_Other > 0.15 (allowing a minimum f_Other of 20%), then f_Methane was saved. Sensitivities of the continuous, uniform distributions generated by the model were quantified over 10,000 Monte Carlo simulations in R (41). The mean of all saved f_Methane values was then reported as the fraction of CO_2 resulting from CH4 production and oxidation. Variance around these saved f_Methane values is based on different, continuous uniform distributions generated at random by the mixing model and was ultimately not reported.

Statistical Analyses. Normality in the data were assessed using quantilequantile plots and Shapiro-Wilk tests. Homogeneity of variance in the data were assessed using Levene's tests. pCH_4 and $\delta^{13}C-CO_2$ followed nonnormal distributions and were log transformed for parametric comparisons along with $p\text{CO}_2$ and $\delta^{13}\text{C-CH}_4$ across the distinct lake environments and flood stages of TSL using ANOVA. Multiple pairwise comparisons between means in the open water, edge, and floodplain environments during the high-water and falling-water stages were carried out subsequently using Tukey Honest Significant Differences. Our Bonferroni-corrected, critical alpha value for multiple pairwise comparisons was 0.025 (for linear regression, our critical alpha value remained 0.050). To assess whether differences between means were independent of sample size, we also calculated effect sizes using Cohen's d, where d = 0.2 to 0.4 corresponds to a small effect and low support for differences between means, d = 0.5 to 0.7 corresponds to a medium effect, and d > 0.9corresponds to a large effect and high support for differences (50). All statistical analyses were conducted using R (41).

Data Availability. All data are freely available and can be accessed at the publically accessible repository GitHub, https://github.com/blm8/PNAS_Tonle-Sap-Carbon-Dioxide-Supersaturation. Code data have been deposited in GitHub, https://github.com/blm8/PNAS_Tonle-Sap-Carbon-Dioxide-Supersaturation. All other study data are included in the article and/or *SI Appendix*.

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