



# Deep learning in prediction of intrinsic disorder in proteins

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## ABSTRACT

Intrinsic disorder prediction is an active area that has developed over 100 predictors. We identify and investigate a recent trend towards the development of deep neural network (DNN)-based methods. The first DNN-based method was released in 2013 and since 2019 deep learners account for majority of the new disorder predictors. We find that the 13 currently available DNN-based predictors are diverse in their topologies, sizes of their networks and the inputs that they utilize. We empirically show that the deep learners are statistically more accurate than other types of disorder predictors using the blind test dataset from the recent community assessment of intrinsic disorder predictions (CAID). We also identify several well-rounded DNN-based predictors that are accurate, fast and/or conveniently available. The popularity, favorable predictive performance and architectural flexibility suggest that deep networks are likely to fuel the development of future disordered predictors. Novel hybrid designs of deep networks could be used to adequately accommodate for diversity of types and flavors of intrinsic disorder. We also discuss scarcity of the DNN-based methods for the prediction of disordered binding regions and the need to develop more accurate methods for this prediction.

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## 1. Introduction

Intrinsic disorder in proteins is defined by lack of stable tertiary structure under physiological conditions [1–4]. Intrinsically disordered proteins (IDPs) include one or more intrinsically disordered regions (IDRs) in their sequences. Recent bioinformatics investigations conclude that IDPs are highly abundant in eukaryotic organisms [5–7] and enriched in multiple cellular compartments [8,9]. Numerous studies of IDPs reveal that they are crucial for a wide spectrum of cellular functions that include signaling, molecular recognition and assembly, cell cycle regulation, transcription, translation and phase separation [10–19]. Moreover, given their functional importance and prevalence in the human diseasesome [12,20–22], they serve as promising and currently underutilized leads for rational drug design efforts [23–27].

Experimentally characterized IDPs and IDRs can be collected from several databases, such as DisProt [28], PDB [29], IDEAL [30], DIBS [31], and MFIB [32]. However, these resources cover only a small fraction of IDPs, with the largest DisProt and PDB databases currently in-

cluding about 2 thousand and 25 thousand IDPs, respectively [28,33]. Compared to over 225 million protein sequences that are available in the newest 2021\_04 release of UniProt [34], we have a long way to go to comprehensively identify and annotate IDPs and IDRs. Computational methods that accurately predict intrinsic disorder can be used to facilitate efforts to close this huge and growing knowledge gap. Computational predictors already made large impact on the intrinsic disorder field, by powering a rapid acceleration in the research on IDPs and IDRs [35]. They are also used across many areas including rational drug design [23–26], structural genomics [36–38], and medicine [39,40].

Development of computational predictors of disorder is a long-standing research problem. A recent survey has identified 103 disorder predictors that were developed over the last four decades [41]. Current surveys point to the long history of the disorder prediction area, providing invaluable insights concerning architectures of these methods, their availability, trends in their development efforts and approaches to comparatively evaluate their predictive performance [40–48]. Moreover, users and developers benefit from empirical studies that comparatively assess predictive quality of disorder predictors [33,49–59]. These comparative studies include several community assessments, such as Critical Assessment of Structure Prediction (CASP) between CASP5 to CASP10 [53–58] and Critical Assessment of Intrinsic Protein Disorder (CAID) [52]. The community assessments involve evaluation of predictors on blind test datasets (i.e., datasets that were not available to the authors of the predictors) by independent assessors who do not take

IDP, Intrinsically disordered protein; IDR, Intrinsically disordered region; DNN, Deep neural network; CASP, Critical Assessment of Structure Prediction; CAID, Critical Assessment of Intrinsic Protein Disorder; FFNN, Feed forward neural networks; BRNN, Bidirectional recurrent neural networks; CNN, Convolutional neural networks

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part in the competitions utilizing tests and metrics that are widely accepted by the community.

The predictive architectures used to develop disorder predictors are typically divided into three categories [42,43,46,47]: (1) sequence scoring functions; (2) machine learning models; and (3) *meta*-predictors. The first category uses additive and/or weighted functions, some of which are grounded in physical principles governing protein folding, to process the input protein sequence and sequence-derived structural and evolutionary information. Representative disorder predictors that fall into this category include FoldIndex [60], IUPred [61, 62], and IUPred3 [63]. The machine learning predictors apply models that are trained from data using a variety of machine learning algorithms, such as support vector machines [64–66], regression [67], conditional random fields [68–70], radial basis function networks [71], and shallow neural networks [36,72–76]. Example popular machine learning predictors include DisEMBL [36], DISOPRED [75,76], PONDR [73], and PrDOS [64]. The *meta*-predictors use multiple disorder predictions as inputs to re-predict disorder. The underlying rationale was to exploit potential complementarity among the input disorder predictions to generate a new prediction that would improve over the inputs. These efforts were also fueled by the availability of diverse sequence-scoring and machine learning predictors and studies that empirically show that well-designed *meta* predictors indeed produce predictions that outperform their inputs [77–79]. Representative example *meta*-predictors of disorder include metaPrDOS [80], MFDp [65,81,82], Cspritz [83], disCoP [77,84], and MobiDB-lite [78]. We observe that some *meta*-predictors use machine learning algorithms (e.g., metaPrDOS [80] and MFDp [65]), which means that they can be cross-listed in both categories.

Results of CASP10, the most recent CASP community assessment that covers disorder prediction (i.e., subsequent CASP experiments do not include disorder predictions), reveal that the top three predictors belong to the machine learning (PrDOS and DISOPRED) and *meta*-predictor (MFDp) categories [58]. However, a recent survey notes a rapid influx of a new subfamily of machine learning methods that relies on deep neural networks (DNNs) after the first DNN-based method was released in 2013 [41]. DNNs differ from shallow neural networks, which were commonly used to implement disorder predictors in early 2000 s [36,72–76], by use of multiple hidden layers and more sophisticated types of neurons and connections. The shift to the deep network models is motivated by their favorable levels of predictive performance when compared with the other types of disorder predictors. In particular, we observe that the best performing methods from the just completed CAID experiment [85], which include fIDPnn [86], SPOT-Disorder2 [87], RawMSA [88] and AUCpred [89], rely on DNNs. Motivated by their growing numbers and success, we provide the first review of the DNN-based disorder predictors. We identify and summarize 13 DNN-based disorder predictors that were developed since 2013. We analyze trends in the development of these predictors and empirically compare predictive quality produced by the deep learners against the other types of disorder predictors based on results produced on blind test dataset from the CAID experiment. We also comment on future prospects in the development of the DNN-based disorder predictors.

## 2. Prediction of intrinsic disorder using deep learning

Nowadays, deep learning is widely used to develop methods that predict protein structure and function. Perhaps the most obvious example is protein structure prediction where deep learning models, such as AlphaFold, have deservedly dominated over other types of methods [90–93]. Moreover, deep learning is utilized to predict other structural aspects of proteins, such as contacts [94], secondary structure [95] and torsional angles [96]. DNNs are also successfully applied to predict protein function [97–99], protein-drug interactions [100,101], and functional sites [102–104].

The intrinsic disorder prediction field was not immune to the infusion of the deep learning-based approaches. The first DNN-based disorder predictor, DNDisorder [105], was published in 2013. Table 1 summarizes a comprehensive list of 36 disorder predictors that were published since that time. This list contextualizes the efforts to develop deep learning predictors in a broader setting of the entire disorder prediction field. We identify the 36 predictors using a wide-ranging list of sources including databases of disorder predictions: MobiDB [122], D2p2 [123] and DescribePROT [124]; community assessments and surveys that were published on or after 2013 [33,41–43,46,47,49,50,52, 58,59], and a manual search of relevant articles from PubMed that we collect using the “(disorder[Title]) AND (prediction[Title]) AND protein” query. Table 1 reveals that 13 out of the 36 recent disorder predictors use deep learning models. We find that it took two more years for the second DNN-based predictor, DeepCNF-D, to be published in 2015 [112]. The following three years include similarly low numbers of new deep learning tools, with two methods published in 2016, one in 2017, and one more in 2018. Year 2019 marks a turning point in the efforts to develop DNN-based disorder predictors, with two tools published in 2019, two in 2020, and four in 2021. Fig. 1 conveniently summarizes the corresponding trends. It highlights the gradual shift to developing predictors that rely on deep networks and the fact that these methods constitute majority (58%) of the predictors that were published over the last three years (green line in Fig. 1). We also note that the consistent levels of the release of new methods that range between 11 and 13 per every three-years long interval.

Table 1 provides a few additional insights. We manually check websites of the corresponding methods and find that 23 out of 36 predictors (over 60%) are available to the end users as either standalone software (5 methods), webserver (10 methods) or in both modalities (10 methods). Interestingly, all DNN-based predictors that were published after 2016, except for fIDPlr, are among the publicly available tools. This rate of availability is substantially better compared to related areas including prediction of protein-binding and RNA-binding residues where the availability is at around 40% [103,125]. The web-servers are a convenient option to less programming savvy end users, such as some biochemists or structural biologists. In this case, predictions are performed on the webserver end and users are not required to install and run the software on their hardware. However, the main drawbacks of web-servers are that they depend on the uninterrupted availability of Internet, limit the size of individual jobs (i.e., number of proteins can be predicted), and their results could be delayed when their workload is heavy. On the other hand, the standalone software option is best suited for skilled programmers and bioinformaticians. The software must be installed and executed locally. This facilitates running larger jobs and allows embedding a given disorder predictor into other bioinformatics pipelines. For instance, putative disorder generated by the popular IUPred [61,62,120] was used to predict DNA-binding residues [126], B-cell epitopes [127], and quality of protein structures [128].

Table 2 details the 13 deep learning-based disorder predictors. We summarize inputs, topologies, predictive performance, and run-time of these methods. The inputs cover a broad range of relevant information including the input sequence itself and several sequence-derived characteristics, such as evolutionary information (e.g., position-specific scoring matrix (PSSM) and residue-level conservation), putative structural features (e.g., secondary structure and solvent accessibility), and physiochemical characteristics that are typically quantified at the amino acid level (e.g., polarizability, hydrophobicity, and isoelectric point). We define topologies based on two key aspects: type of the deep network and its size/depth. The network types include classical deep feed forward neural networks (FFNNs) and more sophisticated restricted Boltzmann machines (RBM), convolutional neural networks (CNNs) and bidirectional recurrent neural networks (BRNNs). We grade the network sizes by the number of

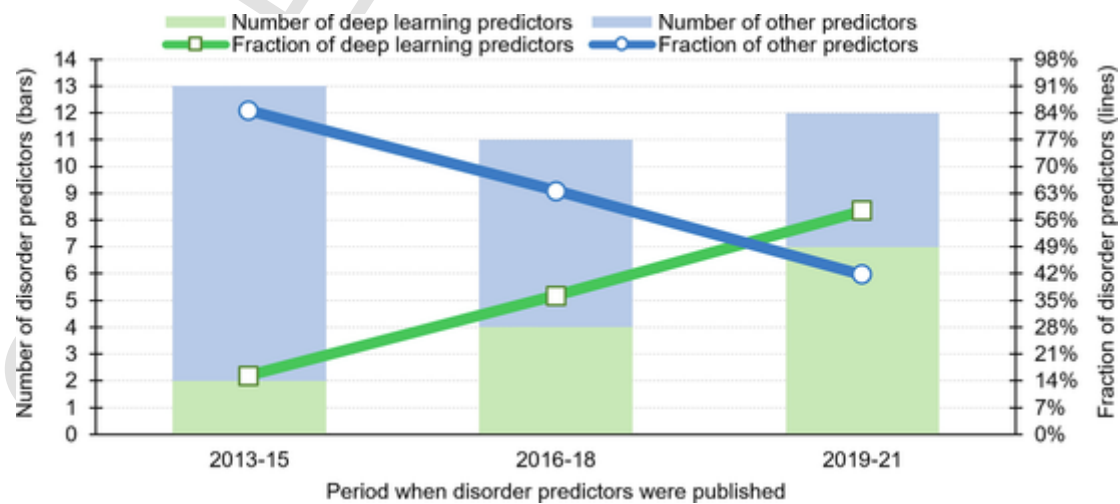
**Table 1**

Summary of intrinsic disorder predictors that were developed since 2013 when the first deep learning-based method was released. The predictors are sorted in the chronological order of their year of publications. “\*” denotes predictors that are used in Fig. 3.

Predictor name	Year published	Reference <sup>1</sup>	Applies DNN	Availability <sup>2</sup>	URL
MFDp2	2013	[81]	No	WS	<a href="https://biomine.cs.vcu.edu/servers/MFDp2/">https://biomine.cs.vcu.edu/servers/MFDp2/</a>
DNdisorder	2013	[105]	Yes	N/A	N/A
preDNdisorder	2013	[105]	No	N/A	N/A
Ulg-GIGA	2013	[106]	No	N/A	N/A
DisMeta	2014	[107]	No	WS	<a href="https://montelionelab.chem.rpi.edu/dismeta/">https://montelionelab.chem.rpi.edu/dismeta/</a>
disCoP	2014	[77,84]	No	WS	<a href="https://biomine.cs.vcu.edu/servers/disCoP/">https://biomine.cs.vcu.edu/servers/disCoP/</a>
DynaMine	2014	[67,108]	No	SP + WS	<a href="https://dynamine.ibsquare.be/">https://dynamine.ibsquare.be/</a>
PON-Diso	2014	[109]	No	WS	<a href="https://structure.bme.lu.se/PON-Diso">https://structure.bme.lu.se/PON-Diso</a>
DISOPRED3*	2015	[75]	No	SP + WS	<a href="https://bioinf.cs.ucl.ac.uk/psipred/">https://bioinf.cs.ucl.ac.uk/psipred/</a>
s2D-1	2015	[110]	No	No	N/A
s2D-2*	2015	[110]	No	No	N/A
DisoMCS	2015	[111]	No	N/A	N/A
DeepCNF-D	2015	[112]	Yes	SP	<a href="https://home.ttic.edu/~wangsheng/software.html">https://home.ttic.edu/~wangsheng/software.html</a>
AUCpred*	2016	[89]	Yes	N/A	N/A
AUCpred-np*	2016	[89]	Yes	N/A	N/A
DisPredict (DisPredict2)*	2016	[66]	No	SP	<a href="https://github.com/tamjidul/DisPredict2_PSEE">https://github.com/tamjidul/DisPredict2_PSEE</a>
MobiDB-lite*	2017	[78]	No	WS	<a href="https://mobidb.bio.unipd.it/">https://mobidb.bio.unipd.it/</a>
SPOT-Disorder*	2017	[113]	Yes	SP + WS	<a href="https://sparks-lab.org/server/spot-disorder/">https://sparks-lab.org/server/spot-disorder/</a>
IUpred2A-long*	2018	[114]	No	SP + WS	<a href="https://iupred2a.elte.hu/">https://iupred2a.elte.hu/</a>
IUpred2A-short*	2018	[114]	No	SP + WS	<a href="https://iupred2a.elte.hu/">https://iupred2a.elte.hu/</a>
pyHCA*	2018	No	No	SP	<a href="https://github.com/T-B-F/pyHCA">https://github.com/T-B-F/pyHCA</a>
SPOT-Disorder-Single*	2018	[115]	Yes	SP + WS	<a href="https://sparks-lab.org/server/spot-disorder-single/">https://sparks-lab.org/server/spot-disorder-single/</a>
Predictor by Zhao and Xue	2018	[116]	No	No	N/A
IDP-CRF	2018	[69]	No	No	N/A
rawMSA*	2019	[88]	Yes	SP	<a href="https://bitbucket.org/clami66/rawmsa/src/master/">https://bitbucket.org/clami66/rawmsa/src/master/</a>
SPOT-Disorder2*	2019	[87]	Yes	SP + WS	<a href="https://sparks-lab.org/server/spot-disorder2/">https://sparks-lab.org/server/spot-disorder2/</a>
Spark-IDPP	2019	[117]	No	No	N/A
IDP-FSP	2019	[70]	No	No	N/A
DisoMine*	2020	No	Yes	WS	<a href="https://www.bio2byte.be/b2btools/disomine/">https://www.bio2byte.be/b2btools/disomine/</a>
ODiNPred	2020	[118]	No	WS	<a href="https://st-protein.chem.au.dk/odinpred">https://st-protein.chem.au.dk/odinpred</a>
IDP-Seq2Seq*	2020	[119]	Yes	WS	<a href="https://bliulab.net/IDP-Seq2Seq/">https://bliulab.net/IDP-Seq2Seq/</a>
fIDPnn*	2021	[86]	Yes	SP + WS	<a href="https://biomine.cs.vcu.edu/servers/fIDPnn/">https://biomine.cs.vcu.edu/servers/fIDPnn/</a>
fIDPlr*	2021	[86]	No	No	N/A
IUPred3	2021	[63]	No	SP + WS	<a href="https://iupred3.elte.hu/">https://iupred3.elte.hu/</a>
RFPR-IDP*	2021	[120]	Yes	WS	<a href="https://bliulab.net/RFPR-IDP/server">https://bliulab.net/RFPR-IDP/server</a>
Metapredict*	2021	[121]	Yes	SP + WS	<a href="https://github.com/idptools/metapredict">https://github.com/idptools/metapredict</a>

<sup>1</sup> “No” means that a given predictor was not published in a peer-reviewed journal but was included based on participation in the CASP and/or CAID assessment.

<sup>2</sup> Availability: released as “SP” (standalone program), “WS” (web server). “No” not released as either SP (standalone program) or WS (web server), and “N/A” (not available) SP and/or WS were released at the time of publication (i.e. URL was provided in the original article) but they were not available as of February 2022 when the access was tested.



**Fig. 1.** Development of disorder predictors since 2013 when the first deep learning-based predictor was released. The left/right y-axis gives the number/fraction of predictors in a given time period. The predictors are color-coded where green represents deep neural network-based methods and blue represents other types of predictors.

**Table 2**

Summary of intrinsic disorder predictors that use deep neural network models. The predictors are sorted in the chronological order of their year of publications. X marks inputs that are used by a given predictor. “\*” denotes predictors that are used in Fig. 3.

Predictor name	Year published	Inputs				Network architecture		AUC	Runtime <sup>7</sup>
		Sequence <sup>1</sup>	Evolutionary features <sup>2</sup>	Predicted structural feature <sup>3</sup>	Physicochemical properties <sup>4</sup>	Type <sup>5</sup>	Size <sup>6</sup>		
DNDISORDER	2013		X	X		RBM	Moderately deep	N/A	N/A
DeepCNF-D	2015		X	X	X	CNN	Moderately deep	N/A	N/A
AUCpred*	2016	X	X	X	X	CNN	Moderately deep	0.757	7.0
AUCpred-np*	2016	X		X	X	CNN	Moderately deep	0.751	<0.5
SPOT-Disorder*	2017		X	X	X	BRNN	Moderately deep	0.744	5.0
SPOT-Disorder-Single*	2018	X	X		X	BRNN + CNN	Deep	0.757	0.8–1.0
rawMSA*	2019		X	X		BRNN + CNN	Very deep	0.780	>10.0
SPOT-Disorder2*	2019	X	X	X		BRNN + CNN	Very deep	0.760	>10.0
DisoMine*	2020			X		BRNN	Moderately deep	0.765	<0.5
IDP-Seq2Seq*	2020		X	X	X	BRNN	Very deep	0.754	12.0
fIDPnn*	2021		X	X	X	FFNN	Moderately deep	0.814	0.5–1.0
RFPR-IDP*	2021		X		X	BRNN + CNN	Moderately deep	0.722	<0.5
Metapredict*	2021			X		BRNN	Moderately deep	0.746	<0.5

<sup>1</sup> The input sequence was encoded and directly used as predictive input.

<sup>2</sup> Evolutional features computed from the input sequence including position-specific scoring matrix (PSSM), entropy-based conservation, and multiple sequence alignment.

<sup>3</sup> Structural features predicted from the input sequence, such as putative secondary structure, solvent accessibility, and half-sphere exposures.

<sup>4</sup> Physicochemical properties of the amino acids in the input sequence including polarizability, hydrophobicity, and isoelectric point.

<sup>5</sup> Type of the deep learning neural network used: “RBM” (Restricted Boltzmann Machine); “CNN” (Convolutional Neural Network); “BRNN” (Bidirectional Recurrent Neural Network); and “FFNN” (Feed Forward Neural Network).

<sup>6</sup> The number of hidden layers: moderately deep with 2 to 3 layers; deep with 4 to 5 layers; and very deep with over 5 layers.

<sup>7</sup> The average runtime in minutes to predict one amino acid sequence. N/A denotes that the results could not be collected since a working implementation of the corresponding predictor is not available.

hidden layers into three categories: moderately deep with between 2 and 3 hidden layers; deep with 4 to 5 hidden layers; and very deep with over 5 hidden layers. We observe a few interesting patterns. First, majority of the predictors rely on multiple input types, with the two most popular options being evolutionary and putative structural data. These methods take advantages of the deep neural network’s ability to combine diverse types of inputs including numeric data, such as conservation and relative solvent accessibility, nominal data, such as secondary structure, and binary data, such as one-hot encoding of amino acid types, to produce high-quality latent feature space. Second, these disorder predictors rely on a diverse collection of network types, including hybrid designs that combine convolutional and bidirectional recurrent topologies. Third, they utilize designs with widely varying network sizes including nine moderately deep, one deep and three very deep networks. Altogether, this analysis reveals that the current designs broadly explore the input and network topology spaces.

The recently completed CAID experiment reveals that some of the DNN-based solutions provide favorable predictive performance when compared to other types of disorder predictors [52]. This conclusion is perhaps best captured with the following quote: “*The SPOT-Disorder2 and fIDPnn, followed by RawMSA and AUCpred, are consistently good. However, fIDPnn is at least an order of magnitude faster than its competitors, and it succeeded on all sequences, whereas SPOT-Disorder2 skipped 5% of sequences as a result of a length limitation.*” [85]. While these four best predictors rely on deep learning, they implement the underlying predictive models using very different designs. More specifically, fIDPnn relies on moderately deep FFNN architecture [86], SPOT-Disorder2 and RawMSA are very deep hybrids of CNN and BRNN [87,

88], while AUCpred utilizes moderately deep CNN topology [89]. This observation suggests that accurate disorder prediction can be accomplished using different types of deep learners.

We provide a wider comparison of the predictive performance of deep learners. We cover 11 DNN-based methods that exclude only the two oldest methods, DNDISORDER and DeepCNF-D. DNDISORDER is not available to the end users (Table 1) while the standalone version of DeepCNF-D requires specific feature encoding of the sequence that we could not reproduce. We compare predictive performance of the remaining 11 deep learners using the annotated CAID dataset from <https://idpcentral.org/caid/data/1/> and <https://idpcentral.org/caid/data/1/reference/disprot-disorder.txt>. This dataset includes 652 protein sequences and 337,908 amino acids, with 838 disordered regions and 54,820 disordered residues. For the 8 of the 11 predictors that were evaluated in CAID (i.e., AUCpred [89], AUCpred-np [89], DisoMine [129], fIDPnn [86], rawMSA [88], SPOT-Disorder [113], SPOT-Disorder-Single [115] and SPOT-Disorder2 [87]), we parse their CAID predictions from <https://idpcentral.org/caid/data/1/predictions/>. We collect results for the other three methods (IDP-Seq2Seq [119], RFPR-IDP [120], and Metapredict [121]) using the web servers and standalone programs provided by the authors. Table 2 shows that the predictive quality of deep learners measured with the area under the ROC curve (AUC) ranges between 0.722 for RFPR-IDP and 0.814 for fIDPnn.

We further evaluate whether differences in the AUCs of the 11 predictors are robust across different datasets by comparing results across 20 randomly selected disjoint sets of 5% of proteins from the CAID dataset. We assess significance of differences in AUCs between the best-performing fIDPnn and the other methods. We use the *t*-test if the underlying data are normal; otherwise, we use the Wilcoxon signed-rank



test; we test normality with the Anderson-Darling test at the 0.05 significance. We find that fIDPnn and RawMSA are not statistically different ( $p$ -value  $\geq 0.05$ ) but fIDPnn is statistically better than the other 9 methods ( $p$ -value  $< 0.05$ ). We similarly quantify significance of differences between RFPR-IDP that has the lowest AUC and the other 10 predictors. This analysis reveals that SPOT-Disorder, Metapredict, AUCpred-np and IDP-Seq2Seq produce predictions that are not statistically better than RFPR-IDP ( $p$ -value  $\geq 0.05$ ). The remaining 4 predictors that include AUCpred, SPOT-Disorder-Single, SPOT-Disorder2, and DisoMine are significantly better than RFPR-IDP ( $p$ -value  $< 0.05$ ) and significantly worse than fIDPnn ( $p$ -value  $< 0.05$ ). Correspondingly, we identify 3 groups of the DNN-based predictors: 1) fIDPnn and RawMSA that secure the best results (AUC  $> 0.78$ ); AUCpred, SPOT-Disorder-Single, SPOT-Disorder2, and DisoMine that obtain the second-best performance ( $0.755 < \text{AUC} < 0.78$ ); and RFPR-IDP, SPOT-Disorder, Metapredict, AUCpred-np and IDP-Seq2Seq that provide more modest levels of predictive quality ( $0.720 < \text{AUC} < 0.755$ ).

We also analyze an average per-protein runtime for the predictors from Table 2. Similar to the analysis of the predictive performance, we could not perform this analysis for DNDisorder and DeepCNF-D that do not provide working implementations. We extract the runtime data from the CAID results for the eight methods that participated in this experiment [52], and we estimate it for the other three methods (IDP-Seq2Seq, RFPR-IDP and Metapredict) based on the implementations provided by the authors. We find that the runtime of the 11 predictors varies widely (Table 2), with the fastest predictors that produce results in several seconds and the slowest that require over 10 min for the same task.

Using the above analysis, Fig. 2 compares the 11 available predictors based on three key characteristics: predictive performance quantified with AUC, speed measured with runtime, and mode of availability. We score each characteristic in the 0 to 2 range where higher number is associated with darker shade and indicates better quality, i.e., higher AUC, lower runtime and more ways to access a given predictor. The

most well-rounded predictors include fIDPnn (total score of 6), SPOT-Disorder-Single (score of 5), DisoMine (score of 4) and Metapredict (score of 4). When analyzing individual dimensions, the fastest methods (i.e., per-protein runtime  $< 1$  min) include AUCpred-np, SPOT-Disorder-Single, DisoMine, fIDPnn, RFPR-IDP and Metapredict. The most accurate methods are fIDPnn and rawMSA and methods that are available in two modes (webserver and standalone) include SPOT-Disorder, SPOT-Disorder-Single, SPOT-Disorder2, fIDPnn and Metapredict.

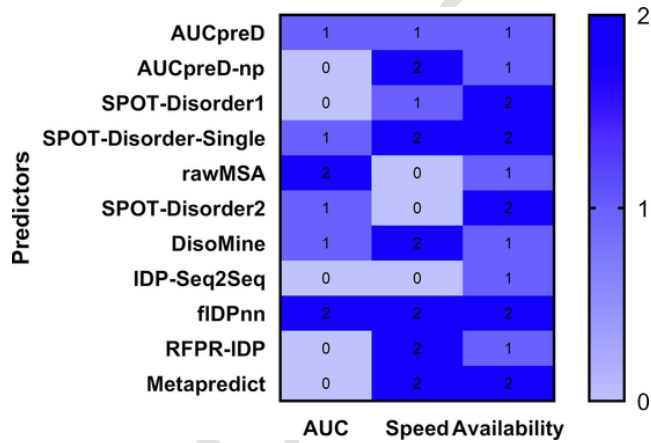
### 3. Deep learning methods outperform other predictors of intrinsic disorder

Motivated by the finding that the top performing predictors in CAID are deep learners [52,85], we investigate whether this result can be extended more broadly to other DNN-based methods. More specifically, we compare the results for the 11 available deep learning-based disorder predictors from Table 2 against the results of other types of methods that we collect using the same CAID data. This analysis covers a comprehensive set of 29 disorder predictors including 11 deep learners that are annotated with \* in Table 2 and 18 methods that use the other types of models. The latter group includes 12 machine learning predictors (DisEMBL-465 [36], DisEMBL-HL [36], DISOPRED3 [75], DisPredict2 [66], Espritz-D [130], Espritz-N [130], Espritz-X [130], fIDPlr [86], PONDR VSL2B [131], PreDisorder [74], RONN [132], and s2D-2 [110]); 5 sequence scoring function-based methods (FoldUnfold [133], IsUnstruct [134], IUPred2A-long [114], IUPred2A-short [114], and pyHCA [135]) and one meta-predictor (MobiDB-lite [78]). We mark these methods with \* in Table 1, except for DisEMBL-465, DisEMBL-HL, JRONN, FoldUnfold, PONDR VSL2B, PreDisorder, IsUnstruct, Espritz-D, Espritz-N, and Espritz-X that were published before 2013. We quantify the predictive performance using four popular metrics that are consistent with the measures used in the most recent community assessments [52,58], including AUC, area under the precision-recall curve (AUPR), F1 and Matthews correlation coefficient (MCC). Finally, we quantify statistical significance of differences in the predictive performance between the results of the 11 deep learners and the 18 other methods. We test normality of the measured scores with the Anderson-Darling test and we apply the student  $t$ -test for normal data and the Wilcoxon test otherwise.

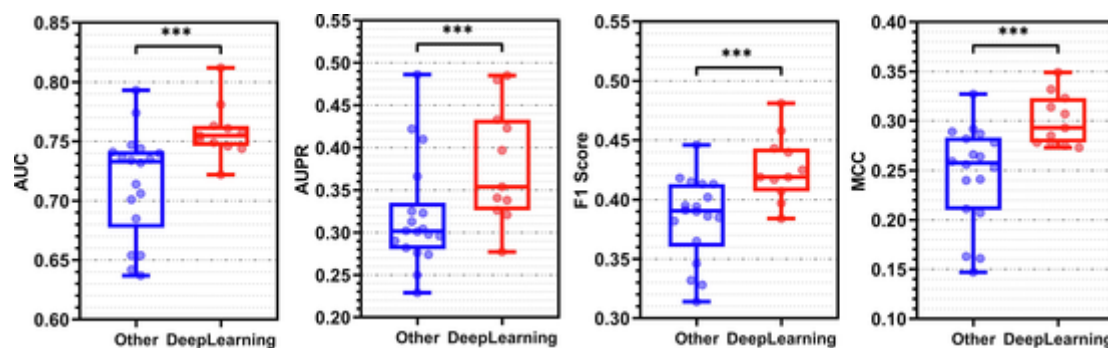
Fig. 3 summarizes the corresponding empirical results. The median AUC of the deep learners is 0.76 vs. 0.73 for the other tools. We observe similarly substantial magnitude of differences for the other metrics, with median AUPR of 0.35 vs. 0.31, median F1 of 0.42 vs. 0.39 and median MCC of 0.29 vs. 0.26. The statistical analysis reveals that the DNN-based methods outperform the other disorder predictors by a statistically significant margin across the four metrics ( $p$ -value  $< 0.05$ ). This consistent and statistically significant trend suggests that the deep neural networks are the best choice to develop accurate disorder predictors.

### 4. Summary and outlook

Disorder prediction is an active and well-establish research area with over 40 years of history. The first DNN-based disorder predictor was published in 2013 and 12 more deep learners were published since. We find that majority of the disorder predictors that were developed in the last three years utilize deep neural networks. The popularity of this design is motivated by several factors. First, these models can be molded into many different architectures that are flexible to use diverse types of inputs. Our analysis of the 13 DNN-based disorder predictors reveals that they rely on very diverse designs that explore different inputs, topologies and sizes. Second, our empirical results reveal that the DNN-based predictors are in general statistically better when directly compared against a representative collection of the other types of pre-



**Fig. 2.** Heatmap that compares 11 available deep learners based on three key characteristics: predictive performance quantified with AUC, speed measured with runtime, and mode of availability. The predictors are sorted in the chronological order of their year of publications. The color-coded scores represent quality where 2 (dark blue) is best, 1 (blue) is intermediate, and 0 (light blue) is worst. The AUC values are categorized into three groups using statistical test that measures robustness of differences between predictors over different protein sets; details are described in the text. Methods with AUCs that are not statistically different ( $p$ -value  $\geq 0.05$ ) from the best (worst) performing fIDPnn (RFPR-IDP) are labeled with 2 (0), while the remaining predictors are labeled with 1. The runtime is divided into three ranges:  $< 1$  min (score of 2); between 1 and 10 min (score of 1); and  $\geq 10$  min (score of 0). The availability score counts the number of modes where 2 means that both SP (standalone program) or WS (web server) are available and 1 that either SP or WS are available.



**Fig. 3.** Comparison of predictive performance between disorder predictors that utilize deep neural networks (in red) and the other disorder predictors (in blue). The predictive performance is quantified with AUC, AUPR, F1 and MCC. Results of individual predictors are denoted by dots. Distributions of these values are summarized with the box plots. \*\*\* means that the predictive performance of the deep learners is significantly higher than the performance of the other methods ( $p$ -value < 0.05).

dictive models. This conclusion is in line with the results of the recent CAID experiment where the top four predictors are deep learners [52, 85]. Third, our multifaceted comparison of the deep learners provides useful clues for the end users by identifying methods that are accurate, fast and widely available. We identify several well-rounded predictors that include fIDPnn (very accurate, very fast, and available in multiple ways), SPOT-Disorder-Single (accurate, very fast, and available in multiple ways), DisoMine (accurate and very fast) and Metapredict (very fast and available in multiple ways). These results and accolades support conclusions of the a recent article that say “*deep-learning-based methods will likely continue to show the greatest potential for future improvement*” [85].

Our analysis finds that the architectures of the current deep learners are considerably diverse. This suggests that the optimal architecture is yet to be identified. We reason that this should be a hybrid design to accommodate for the underlying variety of different types/flavors of disorder [136–138]. For instance, IDRs cover a wide spectrum of sizes, from short regions that are frequently localized at the sequence termini to very long regions that span the entire protein sequence [139,140]. IDRs also vary in their conformational space, which is signified by their classification into the native coils, native pre-molten globules and native molten globules [4,141]. Moreover, IDRs carry out many different functions, and some of them are multifunctional (moonlighting) [142, 143], which results in many different biases in their sequences [4,137]. Interestingly, design of the recently published and well-rounded fIDPnn suggests that predictive quality can be improved by innovating inputs that are fed into the deep networks [86]. The authors point to multiple options including development of extended sequences profiles that cover relevant sequence-derived protein characteristics beyond the commonly-used inputs listed in Table 2, and construction of aggregate features that quantify sequence bias at the region or whole sequence level. These two future directions go hand in hand given the fact that the hybrid deep learners are inherently capable of handling diverse and large inputs.

While most of recently released predictors of intrinsic disorder utilize DNNs, this is not necessarily the case for the methods that predict binding IDRs. There are close to 20 predictors of disordered protein-binding regions [144] and several methods that predict IDRs that interact with nucleic acids and lipids [42,145]. Examples of the recently published tools include FLIPPER [146], SPOT-MoRF [147], OPAL+ [148], DisoLipPred [149] and DeepDISObind [150]. The CAID experiment evaluated close to a dozen of these predictors and concluded that “*disordered binding regions remain hard to predict*” [52], motivating further efforts in this area. One of the potential reasons for the low predictive performance of these tools is a relatively low utilization of the deep learning architectures. We identify only a handful of DNN-based predictors of binding IDRs including SPOT-MoRF [147], MoRFPred\_en [151], en\_DCNNMoRF [152], DeepDISObind [150], and DisoLipPred [149]. A

similar situation is true in the context of prediction of disordered linker regions where neither of the two currently available methods, DFLpred [153] and APOD [154], applies deep learning and their predictive performance is relatively limited. Given the success of DNNs in the disorder prediction, we believe that this technology could be successfully applied to strengthen the quality of the predictors of binding IDRs and disordered linkers.

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## CRediT authorship contribution statement

**Bi Zhao:** Formal analysis, Data curation, Investigation, Validation, Writing – original draft, Writing – review & editing. **Lukasz Kurgan:** Conceptualization, Formal analysis, Validation, Writing – original draft, Writing – review & editing, Project administration, Supervision, Funding acquisition.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## References

- [1] Lieutaud P, Ferron F, Uversky AV, et al. How disordered is my protein and what is its disorder for? A guide through the “dark side” of the protein universe. *Intrinsically Disord Proteins* 2016;4(1):e1259708.
- [2] Habchi J, Tompa P, Longhi S, et al. Introducing protein intrinsic disorder. *Chem Rev* 2014;114(13):6561–88.
- [3] Dunker AK, Babu MM, Barbar E, et al. What's in a name? Why these proteins are intrinsically disordered: Why these proteins are intrinsically disordered. *Intrinsically Disord Proteins*. 2013 Jan-Dec;1(1):e24157.
- [4] Oldfield CJ, Uversky VN, Dunker AK, et al. Introduction to intrinsically disordered proteins and regions. In: Salvi N, editor. *Intrinsically Disordered Proteins*. Academic Press; 2019. p. 1–34.
- [5] Ward JJ, Sodhi JS, McGuffin LJ, et al. Prediction and functional analysis of native disorder in proteins from the three kingdoms of life. *J Mol Biol* 2004;337(3):635–45.
- [6] Peng Z, Yan J, Fan X, et al. Exceptionally abundant exceptions: comprehensive characterization of intrinsic disorder in all domains of life. *Cell Mol Life Sci* 2015; 72(1):137–51.
- [7] Xue B, Dunker AK, Uversky VN. Orderly order in protein intrinsic disorder distribution: disorder in 3500 proteomes from viruses and the three domains of life. *J Biomol Struct Dyn* 2012;30(2):137–49.
- [8] Zhao B, Katuwawala A, Uversky VN, et al. IDPology of the living cell: intrinsic disorder in the subcellular compartments of the human cell. *Cell Mol Life Sci* 2020.
- [9] Meng F, Na I, Kurgan L, et al. Compartmentalization and functionality of

- nuclear disorder: intrinsic disorder and protein-protein interactions in intra-nuclear compartments. *Int J Mol Sci* 2015;17(1).
- [10] V.N. Uversky C.J. Oldfield A.K. Dunker Showing your ID: intrinsic disorder as an ID for recognition, regulation and cell signaling *J Mol Recognit*. 2005 Sep-Oct; 18(5):343–384.
- [11] Peng Z, Oldfield CJ, Xue B, et al. A creature with a hundred waggly tails: intrinsically disordered proteins in the ribosome. *Cell Mol Life Sci CMLS* 2014;71(8):1477–504.
- [12] Babu MM. The contribution of intrinsically disordered regions to protein function, cellular complexity, and human disease. *Biochem Soc Trans* 2016;44(5): 1185–200.
- [13] Peng ZL, Mizianty MJ, Xue B, et al. More than just tails: intrinsic disorder in histone proteins. *Mol Biosyst* 2012;8(7):1886–901.
- [14] Zhou JH, Zhao SW, Dunker AK. Intrinsically disordered proteins link alternative splicing and post-translational modifications to complex cell signaling and regulation. *J Mol Biol* 2018;430(16):2342–59.
- [15] Hahn S. Phase separation, protein disorder, and enhancer function. *Cell* 2018; 175(7):1723–5.
- [16] Staby L, O'Shea C, Willemoes M, et al. Eukaryotic transcription factors: paradigms of protein intrinsic disorder. *Biochem J* 2017;474(15):2509–32.
- [17] Gruszka DT, Mendonca CA, Paci E, et al. Disorder drives cooperative folding in a multidomain protein. *Proc Natl Acad Sci U S A* 2016;113(42):11841–6.
- [18] Peng Z, Xue B, Kurgan L, et al. Resilience of death: intrinsic disorder in proteins involved in the programmed cell death. *Cell Death Differ* 2013;20(9):1257–67.
- [19] Fuxreiter M, Toth-Petroczy A, Kraut DA, et al. Disordered proteinaceous machines. *Chem Rev* 2014;114(13):6806–43.
- [20] Midic U, Oldfield CJ, Dunker AK, et al. Protein disorder in the human diseasesome: unfoldomics of human genetic diseases. *BMC Genomics* 2009;10 (Suppl 1):S12.
- [21] Uversky VN. Intrinsic Disorder, Protein-Protein Interactions, and Disease. *Adv Protein Chem Struct Biol* 2018;110:85–121.
- [22] Uversky VN, Dave V, Iakoucheva LM, et al. Pathological unfoldomics of uncontrolled chaos: intrinsically disordered proteins and human diseases. *Chem Rev* 2014;114(13):6844–79.
- [23] Hu G, Wu Z, Wang K, et al. Untapped Potential of Disordered Proteins in Current Druggable Human Proteome. *Curr Drug Targets* 2016;17(10):1198–205.
- [24] Hosoya Y, Ohkanda J. Intrinsically Disordered Proteins as Regulators of Transient Biological Processes and as Untapped Drug Targets. *Molecules* 2021. Apr 7;26(8).
- [25] Biesaga M, Frigole-Vivas M, Salvatella X. Intrinsically disordered proteins and biomolecular condensates as drug targets. *Curr Opin Chem Biol* 2021;62:90–100.
- [26] Ambadipudi S, Zweckstetter M. Targeting intrinsically disordered proteins in rational drug discovery. *Expert Opin Drug Discov* 2016;11(1):65–77.
- [27] Santofimia-Castano P, Rizzuti B, Xia Y, et al. Targeting intrinsically disordered proteins involved in cancer. *Cell Mol Life Sci* 2020;77(9):1695–707.
- [28] Hatos A, Hajdu-Soltesz B, Monzon AM, et al. DisProt: intrinsic protein disorder annotation in 2020. *Nucleic Acids Res* 2020;48(D1):D269–76.
- [29] Le Gall T, Romero PR, Cortese MS, et al. Intrinsic disorder in the Protein Data Bank. *J Biomol Struct Dyn* 2007;24(4):325–42.
- [30] Fukuchi S, Amemiya T, Sakamoto S, et al. IDEAL in 2014 illustrates interaction networks composed of intrinsically disordered proteins and their binding partners. *Nucleic Acids Res* 2014;42(D1):D320–5.
- [31] Schad E, Ficho E, Pancsa R, et al. DIBS: a repository of disordered binding sites mediating interactions with ordered proteins. *Bioinformatics* 2018;34(3):535–7.
- [32] Ficho E, Remenyi I, Simon I, et al. MFIB: a repository of protein complexes with mutual folding induced by binding. *Bioinformatics* 2017;33(22):3682–4.
- [33] Walsh I, Giollo M, Di Domenico T, et al. Comprehensive large-scale assessment of intrinsic protein disorder. *Bioinformatics* 2015;31(2):201–8.
- [34] UniProt C. UniProt: the universal protein knowledgebase in 2021. *Nucleic Acids Res* 2021;49(D1):D480–9.
- [35] Kurgan L, Radivojac P, Sussman JL, et al. On the Importance of Computational Biology and Bioinformatics to the Origins and Rapid Progression of the Intrinsically Disordered Proteins Field. *Biocomputing 2020* 2020. p. 149–158.
- [36] Lindner R, Jensen LJ, Diella F, et al. Protein disorder prediction: implications for structural proteomics. *Structure* 2003;11(11):1453–9.
- [37] Hu G, Wang K, Song J, et al. Taxonomic landscape of the dark proteomes: whole-proteome scale interplay between structural darkness, intrinsic disorder, and crystallization propensity. *Proteomics* 2018;10:e1800243.
- [38] Oldfield CJ, Xue B, Van YY, et al. Utilization of protein intrinsic disorder knowledge in structural proteomics. *Biochim Biophys Acta* 2013;1834(2):487–98.
- [39] Deng X, Gumm J, Karki S, et al. An overview of practical applications of protein disorder prediction and drive for faster, more accurate predictions. *Int J Mol Sci* 2015;16(7):15384–404.
- [40] Kurgan L, Li M, Li Y. The methods and tools for intrinsic disorder prediction and their application to systems medicine. In: Wolkenhauer O, editor. *Systems Medicine*. Oxford: Academic Press; 2021. p. 159–69.
- [41] Zhao B, Kurgan L. Surveying over 100 predictors of intrinsic disorder in proteins. *Expert Rev Proteomics* 2021.
- [42] Meng F, Uversky VN, Kurgan L. Comprehensive review of methods for prediction of intrinsic disorder and its molecular functions. *Cell Mol Life Sci* 2017; 74(17):3069–90.
- [43] Liu Y, Wang X, Liu B. A comprehensive review and comparison of existing computational methods for intrinsically disordered protein and region prediction. *Briefings Bioinf* 2019;20(1):330–46.
- [44] Deng X, Eickholt J, Cheng J. A comprehensive overview of computational protein disorder prediction methods. *Mol Biosyst* 2012;8(1):114–21.
- [45] He B, Wang K, Liu Y, et al. Predicting intrinsic disorder in proteins: an overview. *Cell Res* 2009;19(8):929–49.
- [46] Meng F, Uversky V, Kurgan L. Computational Prediction of Intrinsic Disorder in Proteins. *Curr Protoc Protein Sci*. 2017 Apr 3;88:2 16 1-2 16 14.
- [47] Li J, Feng Y, Wang X, et al. An overview of predictors for intrinsically disordered proteins over 2010–2014. *Int J Mol Sci* 2015;16(10):23446–62.
- [48] Dosztanyi Z, Meszaros B, Simon I. Bioinformatical approaches to characterize intrinsically disordered/unstructured proteins. *Briefings Bioinf* 2010;11(2): 225–43.
- [49] Katuwawala A, Kurgan L. Comparative assessment of intrinsic disorder predictions with a focus on protein and nucleic acid-binding proteins. *Biomolecules* 2020;10(12).
- [50] Necci M, Piovesan D, Dosztanyi Z, et al. A comprehensive assessment of long intrinsic protein disorder from the DisProt database. *Bioinformatics* 2018;34(3): 445–52.
- [51] Peng ZL, Kurgan L. Comprehensive comparative assessment of in-silico predictors of disordered regions. *Curr Protein Pept Sci* 2012;13(1):6–18.
- [52] Necci M, Piovesan D, Predictors C, et al. Critical assessment of protein intrinsic disorder prediction. *Nat Methods* 2021;18(5):472–81.
- [53] Jin Y, Dunbrack Jr RL. Assessment of disorder predictions in CASP6. *Proteins* 2005;61 (Suppl 7):167–75.
- [54] Bordoli L, Kiefer F, Schwede T. Assessment of disorder predictions in CASP7. *Proteins* 2007;69(Suppl 8):129–36.
- [55] Noivirt-Brik O, Prilusky J, Sussman JL. Assessment of disorder predictions in CASP8. *Proteins* 2009;77(Suppl 9):210–6.
- [56] Monastyrskyy B, Fidelis K, Mout J, et al. Evaluation of disorder predictions in CASP9. *Proteins* 2011;79(Suppl 10):107–18.
- [57] Melamud E, Mout J. Evaluation of disorder predictions in CASP5. *Proteins* 2003;53(Suppl 6):561–5.
- [58] Monastyrskyy B, Kryshchakovich A, Mout J, et al. Assessment of protein disorder region predictions in CASP10. *Proteins* 2014;82(Suppl 2):127–37.
- [59] Katuwawala A, Oldfield CJ, Kurgan L. Accuracy of protein-level disorder predictions. *Briefings Bioinf* 2020;21(5):1509–22.
- [60] Prilusky J, Felder CE, Zeev-Ben-Mordehai T, et al. FoldIndex: a simple tool to predict whether a given protein sequence is intrinsically unfolded. *Bioinformatics* 2005;21(16):3435–8.
- [61] Dosztanyi Z, Csizmek V, Tompa P, et al. The pairwise energy content estimated from amino acid composition discriminates between folded and intrinsically unstructured proteins. *J Mol Biol* 2005;347(4):827–39.
- [62] Dosztanyi Z, Csizmek V, Tompa P, et al. IUPred: web server for the prediction of intrinsically unstructured regions of proteins based on estimated energy content. *Bioinformatics* 2005;21(16):3433–4.
- [63] Erdos G, Pajkos M, Dosztanyi Z. IUPred3: prediction of protein disorder enhanced with unambiguous experimental annotation and visualization of evolutionary conservation. *Nucleic Acids Res* 2021;49(W1):W297–303.
- [64] Ishida T, Kinoshita K. PrDOS: prediction of disordered protein regions from amino acid sequence. *Nucleic Acids Res*. 2007 Jul;35(Web Server issue):W460–4.
- [65] Mizianty MJ, Stach W, Chen K, et al. Improved sequence-based prediction of disordered regions with multilayer fusion of multiple information sources. *Bioinformatics* 2010;26(18):i489–96.
- [66] Iqbal S, Hoque MT. DisPredict: a predictor of disordered protein using optimized RBF Kernel. *PLoS ONE* 2015;10(10):e0141551.
- [67] Cilia E, Pancsa R, Tompa P, et al. From protein sequence to dynamics and disorder with DynaMine. *Nat Commun* 2013;4:2741.
- [68] Wang L, Sauer UH. OnD-CRF: predicting order and disorder in proteins using [corrected] conditional random fields. *Bioinformatics* 2008;24(11):1401–2.
- [69] Liu YM, Wang XL, Liu B. IDP-CRF: Intrinsically Disordered Protein/Region Identification Based on Conditional Random Fields. *International Journal of Molecular Sciences*. 2018 Sep;19(9).
- [70] Liu Y, Chen S, Wang X, et al. Identification of intrinsically disordered proteins and regions by length-dependent predictors based on conditional random fields. *Mol Ther Nucleic Acids* 2019;6(17):396–404.
- [71] Su CT, Chen CY, Hsu CM. iPDA: integrated protein disorder analyzer. *Nucleic Acids Res* 2007;35:W465–72.
- [72] Xue B, Dunbrack RL, Williams RW, et al. PONDR-FIT: a meta-predictor of intrinsically disordered amino acids. *Biochim Biophys Acta* 2010;1804(4): 996–1010.
- [73] Peng K, Vucetic S, Radivojac P, et al. Optimizing long intrinsic disorder predictors with protein evolutionary information. *J Bioinform Comput Biol* 2005; 3(1):35–60.
- [74] Deng X, Eickholt J, Cheng J. PreDisorder: ab initio sequence-based prediction of protein disordered regions. *BMC Bioinf* 2009;21(10):436.
- [75] Jones DT, Cozzetto D. DISOPRED3: precise disordered region predictions with annotated protein-binding activity. *Bioinformatics* 2015;31(6):857–63.
- [76] Jones DT, Ward JJ. Prediction of disordered regions in proteins from position specific score matrices. *Proteins* 2003;53(Suppl 6):573–8.
- [77] Fan X, Kurgan L. Accurate prediction of disorder in protein chains with a comprehensive and empirically designed consensus. *J Biomol Struct Dyn* 2014;32 (3):448–64.
- [78] Necci M, Piovesan D, Dosztanyi Z, et al. MobiDB-lite: fast and highly specific consensus prediction of intrinsic disorder in proteins. *Bioinformatics* 2017;33(9): 1402–4.
- [79] Peng Z, Kurgan L. On the complementarity of the consensus-based disorder prediction. *Proc Symp Biocomput* 2012;176–87.
- [80] Ishida T, Kinoshita K. Prediction of disordered regions in proteins based on the meta approach. *Bioinformatics* 2008;24(11):1344–8.



- [81] M.J. Mizianty Z. Peng L. Kurgan MFDp2: Accurate predictor of disorder in proteins by fusion of disorder probabilities, content and profiles. *Intrinsically Disordered Proteins*. 2013 Jan-Dec;(11):e24428.
- [82] Mizianty MJ, Uversky V, Kurgan L. Prediction of intrinsic disorder in proteins using MFDp2. *Methods Mol Biol* 2014;1137:147–62.
- [83] Walsh I, Martin AJ, Di Domenico T, et al. CSpritz: accurate prediction of protein disorder segments with annotation for homology, secondary structure and linear motifs. *Nucleic Acids Res*. 2011 Jul;39(Web Server issue):W190–6.
- [84] Oldfield CJ, Fan X, Wang C, et al. Computational Prediction of Intrinsic Disorder in Protein Sequences with the disCoP Meta-predictor. *Methods Mol. Biol. (Clifton, NJ)* 2020;2141:21–35.
- [85] Lang B, Babu MM. A community effort to bring structure to disorder. *Nat Methods* 2021;18(5):454–5.
- [86] Hu G, Katuwawala A, Wang K, et al. fIDPnn: Accurate intrinsic disorder prediction with putative propensities of disorder functions. *Nat Commun* 2021;12(1):4438.
- [87] Hanson J, Paliwal KK, Litfin T, et al. SPOT-Disorder 2: improved protein intrinsic disorder prediction by ensembled deep learning. *Genom. Proteom. Bioinform*. 2019;17(6):645–56.
- [88] Mirabello C, Wallner B. rawMSA: end-to-end deep learning using raw multiple sequence alignments. *PLoS ONE* 2019;14(8):e0220182.
- [89] Wang S, Ma J, Xu J. AUCpred: proteome-level protein disorder prediction by AUC-maximized deep convolutional neural fields. *Bioinformatics* 2016;32(17):i672–9.
- [90] Torrisi M, Pollastri G, Le Q. Deep learning methods in protein structure prediction. *Comput Struct Biotechnol J* 2020;18:1301–10.
- [91] AlQuraishi M. AlphaFold at CASP13. *Bioinformatics* 2019;35(22):4862–5.
- [92] Jumper J, Evans R, Pritzel A, et al. Highly accurate protein structure prediction with AlphaFold. *Nature* 2021;596(7873):583–9.
- [93] Suh D, Lee JW, Choi S, et al. Recent applications of deep learning methods on evolution- and contact-based protein structure prediction. *Int J Mol Sci* 2021;22(11).
- [94] Schaarschmidt J, Monastyrskyy B, Kryshchukovych A, et al. Assessment of contact predictions in CASP12: Co-evolution and deep learning coming of age. *Proteins* 2018;86(Suppl 1):51–66.
- [95] Guo ZY, Hou J, Cheng JL. DNSS2: Improved ab initio protein secondary structure prediction using advanced deep learning architectures. *Proteins-Struct. Funct. Bioinform*. 2021;89(2):207–17.
- [96] Li H, Hou J, Adhikari B, et al. Deep learning methods for protein torsion angle prediction. *BMC Bioinf* 2017;18(1):417.
- [97] Zhang F, Song H, Zeng M, et al. DeepFunc: a deep learning framework for accurate prediction of protein functions from protein sequences and interactions. *Proteomics* 2019;19(12):e1900019.
- [98] Kulmanov M, Khan MA, Hoehndorf R, et al. DeepGO: predicting protein functions from sequence and interactions using a deep ontology-aware classifier. *Bioinformatics* 2018;34(4):660–8.
- [99] Littmann M, Heinzinger M, Dallago C, et al. Embeddings from deep learning transfer GO annotations beyond homology. *Sci Rep* 2021;11(1):1160.
- [100] Muller C, Rabal O, Diaz GC. Artificial intelligence, machine learning, and deep learning in real-life drug design cases. *Methods Mol Biol* 2022;2390:383–407.
- [101] Kim J, Park S, Min D, et al. Comprehensive survey of recent drug discovery using deep learning. *Int J Mol Sci* 2021;22(18).
- [102] Li F, Chen J, Leier A, et al. DeepCleave: a deep learning predictor for caspase and matrix metalloprotease substrates and cleavage sites. *Bioinformatics* 2020;36(4):1057–65.
- [103] Wang K, Hu G, Wu Z, et al. Comprehensive survey and comparative assessment of RNA-binding residue predictions with analysis by RNA type. *Int J Mol Sci* 2020;21(18):6879.
- [104] Zhang J, Ghadermarzi S, Kurgan L. Prediction of protein-binding residues: dichotomy of sequence-based methods developed using structured complexes versus disordered proteins. *Bioinformatics* 2020;36(18):4729–38.
- [105] Eickholt J, Cheng J. DNDisorder: predicting protein disorder using boosting and deep networks. *BMC Bioinf* 2013;6(14):88.
- [106] Becker J, Maes F, Wehenkel L. On the encoding of proteins for disordered regions prediction. *PLoS ONE* 2013;8(12):e82252.
- [107] Huang YJ, Acton TB, Montelione GT. DisMeta: a meta server for construct design and optimization. *Methods Mol. Biol. (Clifton, NJ)* 2014;1091:3–16.
- [108] Cilia E, Pancsa R, Tompa P, et al. The DynaMine webserver: predicting protein dynamics from sequence. *Nucleic Acids Res*. 2014 Jul;42(Web Server issue):W264–70.
- [109] Ali H, Urolagin S, Gursarslan O, et al. Performance of protein disorder prediction programs on amino acid substitutions. *Hum Mutat* 2014;35(7):794–804.
- [110] Sormanni P, Camilloni C, Fariselli P, et al. The s2D method: simultaneous sequence-based prediction of the statistical populations of ordered and disordered regions in proteins. *J Mol Biol* 2015;427(4):982–96.
- [111] Wang Z, Yang Q, Li T, et al. DisoMCS: accurately predicting protein intrinsically disordered regions using a multi-class conservative score approach. *PLoS ONE* 2015;10(6):e0128334.
- [112] Wang S, Weng S, Ma J, et al. DeepCNF-D: predicting protein order/disorder regions by weighted deep convolutional neural fields. *Int J Mol Sci* 2015;16(8):17315–30.
- [113] Hanson J, Yang Y, Paliwal K, et al. Improving protein disorder prediction by deep bidirectional long short-term memory recurrent neural networks. *Bioinformatics* 2017;33(5):685–92.
- [114] Meszaros B, Erdos G, Dosztanyi Z. iUPred2A: context-dependent prediction of protein disorder as a function of reduced state and protein binding. *Nucleic Acids Res* 2018;46(W1):W329–37.
- [115] Hanson J, Paliwal K, Zhou Y. Accurate single-sequence prediction of protein intrinsic disorder by an ensemble of deep recurrent and convolutional architectures. *J Chem Inf Model* 2018;58(11):2369–76.
- [116] Zhao B, Xue B. Decision-tree based meta-strategy improved accuracy of disorder prediction and identified novel disordered residues inside binding motifs. *Int J Mol Sci* 2018;19(10).
- [117] Malysiak-Mrozek B, Baron T, Mrozek D. Spark-IDPP: high-throughput and scalable prediction of intrinsically disordered protein regions with Spark clusters on the Cloud. *Cluster Comput* 2019;22(2):487–508.
- [118] Dass R, Mulder FAA, Nielsen JT. ODINPred: comprehensive prediction of protein order and disorder. *Sci Rep* 2020;10(1):14780.
- [119] Tang YJ, Pang YH, Liu B. IDP-Seq2Seq: identification of intrinsically disordered regions based on sequence to sequence learning. *Bioinformatics* 2021;36(21):5177–86.
- [120] Liu Y, Wang X, Liu B. RFPR-IDP: reduce the false positive rates for intrinsically disordered protein and region prediction by incorporating both fully ordered proteins and disordered proteins. *Briefings Bioinf* 2021;22(2):2000–11.
- [121] Emenecker RJ, Griffith D, Holehouse AS. Metapredict: a fast, accurate, and easy-to-use predictor of consensus disorder and structure. *Biophys J* 2021;120(20):4312–9.
- [122] Piovesan D, Necci M, Escobedo N, et al. MobiDB: intrinsically disordered proteins in 2021. *Nucleic Acids Res* 2021;49(D1):D361–7.
- [123] Oates ME, Romero P, Ishida T, et al. D(2)P(2): database of disordered protein predictions. *Nucleic Acids Res*. 2013 Jan;41(Database issue):D508–16.
- [124] Zhao B, Katuwawala A, Oldfield CJ, et al. DescribePROT: database of amino acid-level protein structure and function predictions. *Nucleic Acids Res* 2021;49(D1):D298–308.
- [125] Zhang J, Kurgan L. Review and comparative assessment of sequence-based predictors of protein-binding residues. *Brief Bioinform* 2018;19(5):821–37.
- [126] Zhang J, Ghadermarzi S, Katuwawala A, et al. DNAgenie: accurate prediction of DNA-type-specific binding residues in protein sequences. *Brief Bioinform* 2021;22(6).
- [127] da Silva BM, Myung Y, Ascher DB, et al. epitope3D: a machine learning method for conformational B-cell epitope prediction. *Brief Bioinform* 2021.
- [128] Ghadermarzi S, Krawczyk B, Song J, et al. XRRpred: accurate predictor of crystal structure quality from protein sequence. *Bioinformatics* 2021.
- [129] Orlando G, Raimondi D, Codice F, et al. Prediction of disordered regions in proteins with recurrent Neural Networks and protein dynamics. *bioRxiv*. 2020: 2020.05.25.115253.
- [130] Walsh I, Martin AJ, Di Domenico T, et al. ESpritz: accurate and fast prediction of protein disorder. *Bioinformatics* 2012;28(4):503–9.
- [131] Peng K, Radivojac P, Vucetic S, et al. Length-dependent prediction of protein intrinsic disorder. *BMC Bioinf* 2006;7:208.
- [132] Yang ZR, Thomson R, McNeil P, et al. RONN: the bio-basis function neural network technique applied to the detection of natively disordered regions in proteins. *Bioinformatics* 2005;21(16):3369–76.
- [133] Galzitskaya OV, Garbuzynskiy SO, Lobanov MY. FoldUnfold: web server for the prediction of disordered regions in protein chain. *Bioinformatics* 2006;22(23):2948–9.
- [134] Lobanov MY, Galzitskaya OV. The Ising model for prediction of disordered residues from protein sequence alone. *Phys Biol* 2011;8(3):035004.
- [135] Bitard-Feidel T, Callebaut I. HCAtk and pyHCA: A Toolkit and Python API for the Hydrophobic Cluster Analysis of Protein Sequences. *bioRxiv*. 2018:249995.
- [136] Necci M, Piovesan D, Tosatto SC. Large-scale analysis of intrinsic disorder flavors and associated functions in the protein sequence universe. *Protein Sci* 2016;25(12):2164–74.
- [137] Deiana A, Forcelloni S, Porrello A, et al. Intrinsically disordered proteins and structured proteins with intrinsically disordered regions have different functional roles in the cell. *PLoS ONE* 2019;14(8):e0217889.
- [138] Howell M, Green R, Killeen A, et al. Not that rigid midgits and not so flexible giants: on the abundance and roles of intrinsic disorder in short and long proteins. *J Biol Syst* 2012;20(4):471–511.
- [139] Uversky VN. The most important thing is the tail: multitudinous functionalities of intrinsically disordered protein termini. *FEBS Lett* 2013;587(13):1891–901.
- [140] Nielsen JT, Mulder FA. There is diversity in disorder-“in all chaos there is a cosmos, in all disorder a secret order”. *Front Mol Biosci* 2016;3:4.
- [141] Uversky VN. Unusual biophysics of intrinsically disordered proteins. *Biochim Biophys Acta* 2013;1834(5):932–51.
- [142] Meng F, Kurgan L. High-throughput prediction of disordered moonlighting regions in protein sequences. *Proteins* 2018;86(10):1097–110.
- [143] Sluchanko NN, Bustos DM. Intrinsic disorder associated with 14-3-3 proteins and their partners. *Prog Mol Biol Transl Sci* 2019;166:19–61.
- [144] Katuwawala A, Peng Z, Yang J, et al. Computational prediction of MoRFs, short disorder-to-order transitioning protein binding regions. *Comput Struct Biotechnol J* 2019;17:454–62.
- [145] Katuwawala A, Ghadermarzi S, Kurgan L. Computational prediction of functions of intrinsically disordered regions. *Prog Mol Biol Transl Sci* 2019;166:341–69.
- [146] Monzon AM, Bonato P, Necci M, et al. FLIPPER: predicting and characterizing linear interacting peptides in the protein data bank. *J Mol Biol* 2021;433(9):166900.
- [147] Hanson J, Litfin T, Paliwal K, et al. Identifying molecular recognition features in intrinsically disordered regions of proteins by transfer learning. *Bioinformatics* 2020;36(4):1107–13.
- [148] Sharma R, Sharma A, Raicar G, et al. OPAL+: length-specific MoRF prediction



- in intrinsically disordered protein sequences. *Proteomics* 2019;19(6):e1800058.
- [149] Katuwawala A, Zhao B, Kurgan L. DisoLipPred: accurate prediction of disordered lipid binding residues in protein sequences with deep recurrent networks and transfer learning. *Bioinformatics* 2021.
- [150] Zhang F, Zhao B, Shi W, et al. DeepDISOBind: accurate prediction of RNA-, DNA- and protein-binding intrinsically disordered residues with deep multi-task learning. *Brief Bioinform.* 2021 Dec 15.
- [151] Fang C, Moriwaki Y, Li C, et al. MoRFPred\_en: Sequence-based prediction of MoRFs using an ensemble learning strategy. *J Bioinform Comput Biol* 2019;17(6): 1940015.
- [152] Fang C, Moriwaki Y, Tian A, et al. Identifying short disorder-to-order binding regions in disordered proteins with a deep convolutional neural network method. *J Bioinform Comput Biol* 2019;17(1):1950004.
- [153] Meng F, Kurgan L. DFLpred: High-throughput prediction of disordered flexible linker regions in protein sequences. *Bioinformatics* 2016;32(12):i341–50.
- [154] Peng Z, Xing Q, Kurgan L. APOD: accurate sequence-based predictor of disordered flexible linkers. *Bioinformatics* 2020;36(Supplement\_2):i754–61.