

Review

To Fight or to Grow: The Balancing Role of Ethylene in Plant Abiotic Stress Responses

Hao Chen, David A. Bullock, Jr., Jose M. Alonso and Anna N. Stepanova * 

Program in Genetics, Department of Plant and Microbial Biology, North Carolina State University, Raleigh, NC 27695, USA; hchen22@ncsu.edu (H.C.); dabullo2@ncsu.edu (D.A.B.J.); jmalonso@ncsu.edu (J.M.A.)

* Correspondence: atstepan@ncsu.edu

Abstract: Plants often live in adverse environmental conditions and are exposed to various stresses, such as heat, cold, heavy metals, salt, radiation, poor lighting, nutrient deficiency, drought, or flooding. To adapt to unfavorable environments, plants have evolved specialized molecular mechanisms that serve to balance the trade-off between abiotic stress responses and growth. These mechanisms enable plants to continue to develop and reproduce even under adverse conditions. Ethylene, as a key growth regulator, is leveraged by plants to mitigate the negative effects of some of these stresses on plant development and growth. By cooperating with other hormones, such as jasmonic acid (JA), abscisic acid (ABA), brassinosteroids (BR), auxin, gibberellic acid (GA), salicylic acid (SA), and cytokinin (CK), ethylene triggers defense and survival mechanisms thereby coordinating plant growth and development in response to abiotic stresses. This review describes the crosstalk between ethylene and other plant hormones in tipping the balance between plant growth and abiotic stress responses.

Keywords: ethylene; abiotic stress; hormone crosstalk; growth and defense tradeoff



Citation: Chen, H.; Bullock, D.A., Jr.; Alonso, J.M.; Stepanova, A.N. To Fight or to Grow: The Balancing Role of Ethylene in Plant Abiotic Stress Responses. *Plants* **2022**, *11*, 33. <https://doi.org/10.3390/plants11010033>

Academic Editor: Irina Vaseva

Received: 2 December 2021

Accepted: 19 December 2021

Published: 23 December 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Abiotic stresses such as salinity, heat, cold, drought, and flooding have detrimental effects on plant yield and can lead to complete crop failure. In nature, various plants have evolved an array of fascinating mechanisms to rapidly detect changing conditions and respond to the challenge by altering their growth and development, thus enabling adaptation to the unstable environment. The severe phenotypic defects in plant growth and fecundity commonly observed in unfavorable environmental conditions are the consequence of altered developmental programs that plants adopt in an attempt to survive and reproduce under these suboptimal conditions [1]. Stress adaptation usually involves trade-offs between optimal growth and maximal stress tolerance, with stress-tolerant plant varieties typically displaying lower growth rates and productivity under favorable conditions [2,3]. Thus, the focus of modern breeding efforts and biotechnological innovations on minimizing the growth/defense trade-offs and leveraging those natural stress response mechanisms without eliciting a yield penalty will be key to securing a stable future food supply under the ever-changing climate.

One promising factor that may prove crucial to developing effective solutions for uncoupling plant growth from defense is ethylene. Ethylene is an endogenous, gaseous plant hormone that is involved not only in a variety of physiological and developmental processes, from regulating organ growth to inducing fruit ripening, but also in multiple stress responses. In fact, endogenous ethylene production in plants is influenced by several biotic and abiotic factors that affect many of the same physiological and developmental processes that ethylene is known to regulate [4], raising the possibility of rationally manipulating ethylene biosynthesis, signaling, or response to trigger plant adaptation to stress without eliciting growth arrest.

The biological function of ethylene was first characterized in dark-grown pea seedlings that, in the presence of this hormone, display a characteristic set of morphological changes

known as the triple response [4]. In the model system *Arabidopsis thaliana* that proved instrumental in the identification of the molecular machinery required for ethylene biosynthesis, perception, and signaling, the ethylene-triggered triple response consists of hypocotyl swelling, exaggeration of the apical hook curvature, and the inhibition of hypocotyl and root elongation in dark-grown seedlings [5]. This robust triple response phenotype has served as the basis for ethylene mutant identification that ultimately led to the cloning and characterization of critical ethylene-related genes [5–7].

Ethylene biosynthesis in plants involves three enzyme-catalyzed steps (Figure 1A). Methionine is first converted to S-adenosyl-methionine (SAM) by SAM synthetases, then to 1-aminocyclopropane-1-carboxylic acid (ACC) by ACC synthases (ACS) and, finally, to ethylene by ACC oxidases (ACO) [8–11]. Ethylene biosynthesis is not specific to any one plant tissue or organ, and it is believed that most, if not all, plant cells can produce ethylene [6]. Under favorable growth conditions, overall ethylene production is kept at a low level, but during specific developmental processes and in response to certain abiotic stresses, ethylene biosynthesis can be rapidly induced [12,13].

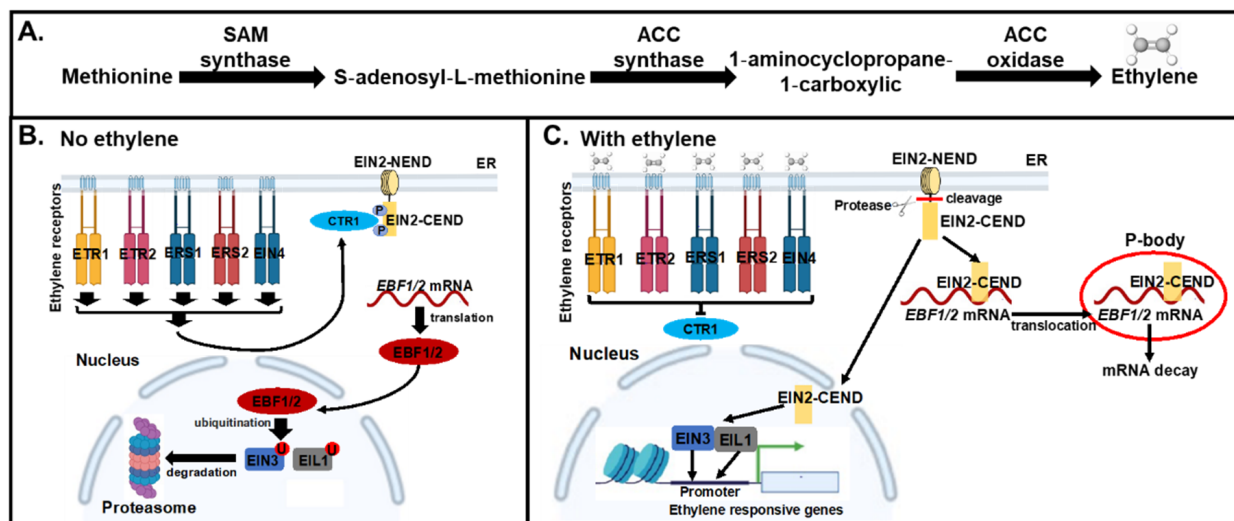


Figure 1. Schematic overview of ethylene biosynthesis and signaling in *Arabidopsis*. (A) Ethylene biosynthetic pathway. Amino acid methionine is converted into hormone ethylene by three families of enzymes, SAM synthetases that produce S-adenosyl-methionine out of methionine, ACC synthases that make 1-aminocyclopropane-1-carboxylic acid, and ACC oxidases that generate ethylene. Ethylene then diffuses out of and into plant cells. (B) Ethylene signaling pathway in the absence of ethylene. Ethylene receptors ETR1, ETR2, ERS1, ERS2, and EIN4 localized in the ER membrane activate CTR1 kinase, which in turn phosphorylates the C-terminal end of EIN2 (EIN2-CEND) and turns EIN2 off. The mRNAs for F-box proteins EBF1 and EBF2 are translated and target the master transcriptional regulators of ethylene signaling, EIN3 and EIL1, to proteasomes for protein turnover, thus preventing ethylene responses. (C) Ethylene signaling pathway in the presence of ethylene. Ethylene binding to the receptors shuts them off, CTR1 is inactivated, EIN2 is dephosphorylated and is cleaved by an unknown protease, thus releasing EIN2-CEND that functions in the cytoplasm and in the nucleus. In the cytosol, EIN2-CEND recruits *EBF1* and *EBF2* mRNAs to the P-bodies and inhibits their translation. In the nucleus, EIN2-CEND directly or indirectly potentiates the activity of EIN3 and EIL1. Ethylene-triggered stabilization of the EIN3/EIL1 transcription factors leads to the transcriptional regulation of multiple target genes, including induction of several members of the *ERF* gene family that encode transcription factors of the second tier of the ethylene response that propagate the EIN3/EIL1-triggered transcriptional programs. Positive interactions such as activation, production, and stabilization are represented with lines that end in an arrowhead →. Negative interactions such as inactivation and repression are represented by lines that end in a hammerhead ⊥. Images were created with BioRender.

Ethylene can freely diffuse through plant membranes reaching the endoplasmic reticulum (ER), where perception takes place and triggers a signaling cascade that ends in the transcriptional regulation of ethylene-responsive genes in the nucleus [14]; Figure 1B,C. The ethylene receptors ETHYLENE RESPONSE SENSOR1 (ERS1), ERS2, ETHYLENE RESPONSE1 (ETR1), ETR2 and ETHYLENE INSENSITIVE4 (EIN4) in Arabidopsis are negative regulators of the pathway that, in the absence of ethylene, activate the CONSTITUTIVE TRIPLE RESPONSE1 (CTR1) protein [15,16]. CTR1 is a kinase that phosphorylates and inhibits the positive regulator of ethylene signaling, EIN2, an ER-localized membrane protein [16]. In the presence of ethylene, upon ethylene binding, the receptors and CTR1 become inactivated, EIN2 gets dephosphorylated and cleaved, and its released C-terminus (CEND) enters the nucleus as well as moves to P-bodies [17,18]. In the nucleus, EIN2-CEND directly or indirectly promotes the activity of EIN3 and EIN3-LIKE1 (EIL1), the transcriptional master-regulators of ethylene signaling. In the cytosol, the EIN2-CEND represses the translation of *EIN3 BINDING F-BOX PROTEIN1* (*EBF1*) and *EBF2* transcripts by directly or indirectly binding to their 3'-untranslated regions (3'-UTRs) and moving along with these transcripts to the P-bodies [17,18]. *EBF1* and *EBF2* are F-box proteins that target EIN3/EIL1 for proteasomal degradation [19,20]. In the presence of ethylene, the translation of *EBF1* and *EBF2* is inhibited, and EIN3 and EIL1 are stabilized and regulate the transcription of ethylene-responsive target genes, such as *ETHYLENE RESPONSE FACTORS* (*ERFs*) [21,22].

ERFs are AP2-domain-containing transcription factors (TFs) that regulate genes involved in diverse biological processes such as growth, development, hormone, and stress responses through several mechanisms, including transcriptional and post-translational control [23–25]. Different subclasses of ERFs have specific DNA binding preferences [26]. For example, a subset of related ERFs recognizes Dehydration-Responsive or C-Repeat Element (DRE/CRT) with A/GCCGAC core sequence found in stress-responsive genes to regulate drought, cold and heat abiotic stress responses. Through direct binding to target gene promoters, ERFs can either activate or repress target gene expression [27,28].

In Arabidopsis, ethylene has been shown to inhibit hypocotyl and root elongation in dark-grown seedlings, but to promote hypocotyl growth in light-grown plants [29]. Thus, by cooperating with different environmental signals, ethylene can differentially regulate growth to mediate plant adaptation to variable environments [30]. Not surprisingly, endogenous ethylene levels and the transcript abundance of ethylene signaling genes are rapidly upregulated in response to multiple abiotic stresses. For example, flooding, heat, shade, heavy metals, salt, osmotic, drought, and cold stress conditions have all been described to trigger ethylene synthesis and increased expression of ethylene pathway genes [31–37]; Table 1.

Correspondingly, ethylene presence affects the plant tolerance to these abiotic stresses [56] through multiple levels of regulation (Figure 2). For example, ethylene increases the plant tolerance to salt stress by altering the physiological and developmental activities of plants, e.g., by promoting adventitious root formation, regulating stem and petiole growth, and controlling stomatal aperture [57]. In response to flooding and heavy metal stresses, ethylene cooperates with the reactive oxygen species (ROS) response pathway to modulate plant metabolism and antioxidant system to promote plant survival [58]. Furthermore, ethylene balances photosynthesis and abiotic stress response to fine-tune plant growth and development under unfavorable conditions [59,60]; Figure 2.

To help plants deal with abiotic stress, ethylene engages in a sophisticated crosstalk with multiple other phytohormones to enable plant adaptation [1,31,60,61]. With the explosion in the volume of research studies published in the field of plant biology in the past 30 years, it comes as no surprise that different plant hormones have been found to engage and cooperate in multiple physiological processes, including responses to abiotic stresses [61,62]. Some of this hormone-hormone crosstalk occurs at the level of transcription and involves the coordinated action of regulators for several hormones. For instance, in Arabidopsis exposed to cold stress, JA and GA pathway regulators JASMONATE ZIM-DOMAIN (JAZ) and DELLA proteins physically interact with EIN3 to activate C-

REPEAT/DEHYDRATION-RESPONSIVE ELEMENT BINDING FACTOR (CBF) TF genes to enhance cold tolerance, but CBFs can also be induced by ABA [63,64]. This review describes the current state of knowledge on the interactions of ethylene with other plant hormones in response to abiotic stress to balance the trade-off between investing in growth and protecting the plant against stress-induced damage.

Table 1. Effects of different abiotic stress factors on transcript abundance of ethylene biosynthetic and signaling genes in Arabidopsis.

Stress Type	Regulation	Ethylene Biosynthetic Genes	Ethylene Signaling Genes
Drought stress	Up	ACS8, 12 [37,38]	
	Down	ACO2, 4, 5, ACS10, 11 [38,39]	
Flooding/hypoxia stress	Up	ACS2, 6, 7, 9, 11 [40,41]	EIN3 [42]
	Down		
Osmotic stress	Up	ACS7, ACO2 [43,44]	ETR2, ERS1, CTR1, EIN3/EIL1, EBF1/2 [44,45]
	Down		ETR1, EIN2 [46,47]
Salt stress	Up	ACS2, 6, 7, 11, ACO2, 4 [47,48]	EIN3 [49]
	Down		ETR1, EIN2 [45,46]
Heat stress	Up	ACS6, 7, 8, 10, 11, 12, ACO2, 4 [43,50]	ERS2, ETR2, EIN3 [50]
	Down	ACS2, 4, 5, ACO1, 3 [50]	ERS1, CTR1, EIN2 [50]
Cold stress	Up	ACS2, 11, ACO2 [41,47]	EIN3 [51]
	Down		ETR1, EIN4, EIN2, EBF1/2 [51]
Heavy metal stress	Up	ACS2, 5, 6, ACO1, 2 [52–54]	EIN2 [55]
	Down		

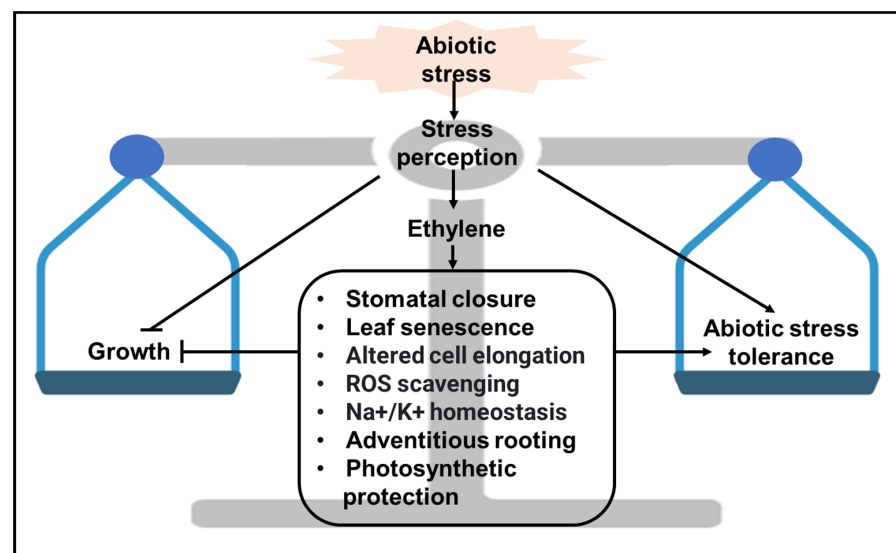


Figure 2. Simplified overview of the role of ethylene in mediating the tradeoff between abiotic stress and growth in plants. Abiotic stress induces ethylene biosynthesis and triggers ethylene accumulation in plants. Ethylene perception and signaling result in multiple physiological responses that not only inhibit plant growth, but also confer stress tolerance that maximizes plant survival in adverse conditions. Positive interactions such as activation, production, and stabilization are represented with lines that end in an arrowhead →. Negative interactions such as inactivation and repression are represented by lines that end in a hammerhead ⊣. The background image of balance was created with BioRender.

2. Ethylene and JA

JA is a fatty-acid-derived hormone that was named after *Jasminum grandiflorum*'s jasmine oil [65]. JA was identified for its stress-related functions and the ability to regulate plant growth and development, with its activity being partially dependent on JA interactions with ethylene [66]. JA biosynthesis has recently been reviewed in detail [67,68]. In brief, JAs are predominantly synthesized through two chloroplasts-based pathways: the octadecane route that uses linolenic acid to produce 12-oxo-phytodienoic acid (OPDA) and the hexadecane pathway that converts hexadecatrienoic acid (16:3) to dinor-oxo-phytodienoic acid (dn-OPDA) [69]. These OPDAs are subsequently oxidized into JA in peroxisomes, and the JA is then conjugated with isoleucine (Ile) to form JA-Ile in the cytoplasm. JA-Ile is the bioactive form of JA which regulates plant development and environmental adaptation. JA-Ile binds to the CORONATINE-INSENSITIVE1 (COI1) receptor, inducing the degradation of the JAZ proteins and releasing the MYC2 TF from the JAZ-MYC2 complex. MYC2 then triggers the expression of JA-responsive genes [69].

One of the best-understood points of crosstalk between ethylene and abiotic stress-induced signaling is the transcriptional regulation of stress-related genes by EIN3/EIL1 master-regulators that directly bind to and alter the expression of many TF genes implicated in abiotic stress tolerance [70]. Some examples of such TF genes in Arabidopsis include *ERF1*, *SALT-INDUCED AND EIN3/EIL1-DEPENDENT1*, and *ETHYLENE AND SALT INDUCIBLE1* involved in salt stress response [71], *RELATED TO AP2.2 (RAP2.2)/ERF73* and *RAP2.12* associated with flooding stress tolerance [72], *MEDIATOR COMPLEX SUBUNIT16 (MED16)* and *MED25* participating in responses to nutrient deficiency [73], *ERF95* and *ERF97* regulating heat stress tolerance [74], and *CBF1*, 2 and 3 controlling cold response [51]. Importantly, the transcription of some of the EIN3-regulated downstream TF genes is also controlled by JA signaling. For example, *ERF1* induction in Arabidopsis requires JA under several abiotic stresses, such as drought, salt, and heat stress [75,76]. Through a ubiquitin-mediated de-repression mechanism, JA signaling also regulates EIN3/EIL1 activities. In light-grown Arabidopsis seedlings, JAZ proteins (JAZ1, 3, and 9) can physically interact with EIN3/EIL1 and enhance EIN3/EIL1 binding to HISTONE DEACETYLASE6 (HDA6), a histone tail modification enzyme that blocks transcription [77,78]. The resulting complex that forms in the absence of JA inhibits EIN3/EIL1-mediated transcription [78,79]. Upon JA treatment, JAZ degradation attenuates HDA6-EIN3/EIL1 association, thus activating EIN3/EIL1 [78]. Multiple abiotic stresses enhance endogenous JA production, thereby boosting JA levels in cells and promoting the release of EIN3/EIL1 from JAZ-HDA6 hijacking [78]. Further studies show that EIN3/EIL1 positively regulate JA-dependent root hair development and thus enhance drought tolerance in Arabidopsis [80], whereas JAZ1 and HDA6 are known to, vice versa, inhibit root hair growth and thus attenuate plant drought tolerance in Arabidopsis and rice [81–84], which is consistent with the cooperative roles of ethylene and JA in regulating plant drought tolerance and root hair development [85].

In addition to the collaborative effects of ethylene and JA on the activity of EIN3/EIL1 and their targets in Arabidopsis and rice seedlings, JA and ethylene can antagonistically regulate many abiotic stress genes in some contexts [86]. In etiolated Arabidopsis seedlings, JA-activated TFs MYC2, MYC3, and MYC4 can physically interact with EIN3/EIL1 and repress their DNA binding ability, thereby reducing the expression of EIN3/EIL1-inducible target genes [86,87]. For example, through this mechanism, the expression of EIN3-regulated flooding stress genes is inhibited by MYC 2/3/4 [88]. Reciprocally, EIN3 can repress the transcription of MYC2-controlled wounding-responsive genes [86]. This mutual inhibition of MYC2 and EIN3 in Arabidopsis modulates the antagonistic effects of JA and ethylene, thereby fine-tuning the expression of many stress-related *ERFs* [75,89,90]. One classical example of the direct target of MYC2 and EIN3 that mediates responses to multiple abiotic stresses is the TF gene *ERF1* [75,89,90] antagonistically regulated by JA and ethylene at the transcriptional level. Arabidopsis *ERF1* controls the expression of multiple suites of genes activated by JA, drought, salt, and heat [75,89]. In fact, *ERF1*-overexpressing plants have been shown to have increased tolerance to drought, heat, and salt stress [89].

Besides this layer of transcriptional regulation, JA-triggered inhibition of the EIN3 protein function has also been described to take place at the post-transcriptional level in Arabidopsis apical hook formation [86]. JA treatment decreases EIN3 and EIL1 protein abundance in wild-type Arabidopsis plants, but not in the *ebf1 ebf2* background [86]. Further mechanistic inquiry revealed that JA enhances the transcription of *EBF1*, and this upregulation is diminished in the *myc2* mutant [86]. Taken together, these observations suggest that in Arabidopsis apical hooks, JA triggers EIN3/EIL1 degradation via MYC2-mediated transcriptional induction of *EBF1* [86]. However, it remains unknown if these antagonistic actions of ethylene and JA signaling at the post-transcriptional level also take place in response to abiotic stresses. Furthermore, the stability of EBF1/EBF2 in Arabidopsis is dependent on the RING-type E3 ligase SALT- AND DROUGHT-INDUCED RING FINGER1 (SDIR1), which directly targets EBF1/EBF2 for ubiquitination and proteasome-dependent degradation [91]. Accordingly, SDIR1 promotes drought tolerance and salt sensitivity in Arabidopsis, and the SDIR1-mediated regulation of EBF1/EBF2 is fine-tuned by temperature fluctuations [91,92].

As mentioned above, the cooperative and antagonistic regulation of ethylene and JA crosstalk is largely dependent on the interaction of EIN3, JAZs, HDA, MYCs, and EBFs, whose abundance is tightly controlled by endogenous ethylene and JA. For example, the abundance of EIN3 in Arabidopsis is enhanced by high endogenous levels of ethylene and repressed by high endogenous levels of JA [21,86]. The ethylene and JA production is, in turn, controlled by several abiotic stress factors, with cold stress, for example, reducing the levels of ethylene and enhancing the levels of JA [79]. Accordingly, the abundance and interaction of the aforementioned proteins are altered in response to stress-induced hormone level changes [81–87]. The relative concentrations of various protein complexes differentially affect plant growth and stress tolerance, a phenomenon illustrated by the JA-ethylene crosstalk mediated by JAZ, HDA, and EIN3 interactions in balancing root hair development and drought tolerance [78,81]. In the future, building the JA- and ethylene-mediated gene regulatory networks from protein-DNA and protein-protein interactions with spatial and temporal resolution in response to specific stress factors would be critical for a comprehensive understanding of the role of ethylene and JA crosstalk in abiotic stress tolerance and plant growth and development.

3. Ethylene and ABA

ABA, whose name comes from “abscisin II” [93], is an isoprenoid plant hormone synthesized through the plastidial 2-C-methyl-D-erythritol-4-phosphate pathway. ABA can be produced in nearly all plastid-containing cells and is transported by ABA transporters [94]. In the presence of ABA, soluble PYRABACTIN RESISTANCE1 (PYR1)/PYR1-LIKE/REGULATORY COMPONENT OF ABA RECEPTOR proteins bind the hormone, leading to the inhibition of ABI1-INSENSITIVE1 (ABI1) phosphatase. The ABA-PYR complex causes the accumulation of phosphorylated SNF1-RELATED PROTEIN KINASE2 (SnRK2) family, which in turn phosphorylates various target proteins, including ABA-RESPONSIVE ELEMENT-BINDING FACTORS, to trigger cellular responses [94]. In the absence of ABA, ABI1 inhibits the action of SnRKs, thus switching off the ABA signaling [94].

Abscicic acid is well known as a stress hormone for its role in alleviating abiotic stress in plants [95]. Ethylene and ABA have been described to have antagonistic interactions, influencing each other’s biosynthetic and signaling pathways [96]. Genetic evidence in Arabidopsis suggests that the knockouts of ethylene biosynthesis and signaling genes result in altered plant sensitivity to ABA and, accordingly, affect the ABA-dependent abiotic stress tolerance in several developmental processes [43,46,97,98]. For example, the Arabidopsis *ACS7* loss-of-function mutant with reduced endogenous ethylene levels is hypersensitive to ABA, accumulates higher levels of endogenous ABA, and displays enhanced salt tolerance during seed germination and seedling growth [43,98]. However, the interactions between ethylene and ABA in the regulation of salt tolerance are more complex and developmental

processes dependent. For instance, in *Arabidopsis* leaf development, the loss-of-function mutations of *EIN2* and *EIN3* and gain-of-function mutations of *ETR1* and *EIN4* have been shown to confer salt sensitivity, with these mutants accumulating higher endogenous ABA levels [46,99–102], whereas the inactivation of *CTR1* and *EBF1/EBF2* leads to enhanced salt tolerance and reduced ABA levels in *Arabidopsis* [102].

Another abiotic stress-sensitive process affected by the ethylene-ABA crosstalk is stomatal opening and closure (Figure 3). ABA acts as the key regulator of stomatal closure to ameliorate the effect of abiotic stress, such as drought [103]. Exogenous ethylene treatment of different plant species, including *Arabidopsis*, tomato, carnation, and sunflower, inhibits ABA- and stress-induced stomatal closure [104–107]. The inhibitory effect of ethylene on ABA-regulated stomata activity stems from flavonol accumulation [108–110]. Flavonol build-up under stress conditions is an *EIN2*-dependent process that represses ABA-induced ROS production and stomatal closure in *Arabidopsis* [108–110]. Controversially, some other studies found that ethylene can cause stomatal closure, resulting from the NADPH oxidase *AtRbohF*-dependent H_2O_2 production through the activation of the $G\alpha$ protein in *Arabidopsis* guard cells [109,110], Figure 3. The contradictory results for the effect of ethylene on stomatal opening and closure may be due to differences in experimental conditions, such as stress duration, stress strength, or the measured time point. In an attempt to reconcile these findings and decipher the perplexing control of stomatal closure by the ethylene-ABA crosstalk, a mathematical model that integrates all relevant signaling components in *Arabidopsis* was constructed for guard cells using a continuous logical modeling framework [111]. This model suggests that an increase in either ethylene or ABA alone results in stomatal closure, whereas the presence of both hormones diminishes stomatal closure.

The complexity in the regulation of stomatal closure via the reciprocal regulation of ABA and ethylene biosynthesis has also been described. Plants overexpressing ethylene-inducible *ERFs* (*AtERF1* in *Arabidopsis*, *JERF1* in tobacco, *TSRF1* in rice, and *TaERF1* in *Triticum aestivum*) show enhanced expression of ABA biosynthetic genes and greater endogenous ABA content, and thus result in stomatal closure and improved abiotic stress tolerance [75,112–115]. Reciprocally, high ABA levels in *Arabidopsis* induce the expression of the TF gene *ABI4*, and *ABI4* protein in turn negatively regulates ethylene production by repressing the transcription of ethylene biosynthesis genes *ACS4* and *ACS8* [116], Figure 3. Consequently, high concentrations of ABA limit ethylene production in guard cells and induce stomata closure [117]. The negative feedback loop coordinating the ethylene and ABA levels could partially explain the early and late response of stomata actions in response to abiotic stresses [111], Figure 3.

The antagonistic roles of ethylene and ABA have also been discovered in flooding stress responses. To date, the role of ethylene in flooding stress tolerance has been well-studied in rice and Polygonaceae species whose root systems grow under water submergence [118]. In rice, high concentrations of ABA inhibit shoot elongation [118]. Flooding stress promotes the accumulation of ethylene in rice shoots and that extra ethylene inhibits ABA biosynthesis as well as further decreases ABA concentration by inducing the breakdown of ABA into phaseic acid [119–121]. With the degradation of ABA, rapid shoot elongation is promoted [120] and takes plants out of water, thus enabling them to escape from flooding stress.

So far, we have shown that the antagonistic roles of ABA and ethylene are observed in multiple abiotic stress responses. Likewise, ABA and ethylene interact antagonistically in many processes of plant development and growth. For example, ABA negatively regulates seed germination, seedling development, petiole angle, root quiescence center cell division and differentiation, and lateral root emergence, whereas ethylene positively regulates all of these ABA-inhibited processes [122–127]. The studies highlight some common downstream regulatory modules, such as TFs, that are antagonistically controlled by ethylene and ABA, and future experimental inquiries would further clarify the nature of this crosstalk and the underlying mechanisms of regulation in balancing plant growth and abiotic stress tolerance.

The rich genetic resources available for interrogating ethylene and ABA pathways in model plants *Arabidopsis* and rice, together with the clear transcriptional roadmaps downstream of ethylene and ABA [128,129], will continue to fuel these important efforts.

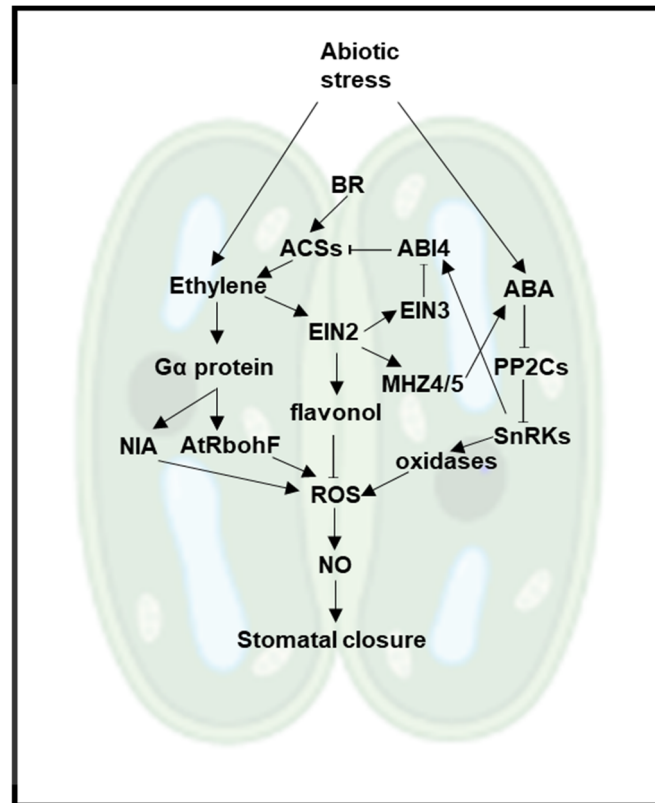


Figure 3. A model of stomatal closure regulation by crosstalk between ABA, BR, and ethylene in *Arabidopsis*. Endogenous ABA and ethylene accumulate in response to abiotic stress. The biosynthesis of ethylene is upregulated by BR but repressed by ABA due to the transcriptional regulation of *ACS* genes. The ABA biosynthesis is upregulated by ethylene through *MHZ4/5*. ABA induces *ABI4* transcript levels and the *ABI4* protein, in turn, represses the transcription of the *ACS* genes. Ethylene also regulates stomatal closure via two ROS-dependent pathways. One pathway is mediated by the ethylene-triggered activation of the $G\alpha$ protein, which in turn promotes H_2O_2 production by activating the NADPH oxidase gene *AtRbohF* and by inducing NO accumulation via the transcriptional upregulation of the nitrate reductase gene *NIA1*. The increased ROS (H_2O_2 and NO) results in stomatal closure. The second pathway, vice versa, links ethylene with reduced ROS levels via EIN2-dependent synthesis of flavonols, secondary metabolites that serve as ROS scavengers. Conversely, high levels of ABA signaling lead to elevated ROS production and stomatal closure. Positive interactions such as activation, production, and stabilization are represented with lines that end in an arrowhead $\rightarrow$. Negative interactions such as inactivation and repression are represented by lines that end in a hammerhead $\dashv$. The background image of stomata was created with BioRender.

4. Ethylene and BR

BRs, which were first discovered in an organic extract of rapeseed (*Brassica napus*), are a class of polyhydroxy steroid plant hormones [130]. BRs are synthesized by multiple and complex routes, all of which proceed through triterpenoid pathways from campesterol in the ER [130,131]. BRs can be produced by most plant organs and act locally. BRs bind to the plasma membrane-localized receptor, BR-INSENSITIVE1 (BRI1) in *Arabidopsis*, enabling BRI1 to interact with its coreceptor BRI1-ASSOCIATED KINASE1 (BAK1). The BRI1-BAK1 complex deactivates another kinase, BRASSINOSTEROID INSENSITIVE2 (BIN2), via a phosphorylation cascade. BIN2 phosphorylates and inhibits multiple TFs, such as BRASSINAZOLE-RESISTANT1 and BRI1-EMS-SUPPRESSOR1. The inactivation of

BIN2 by BRs releases and potentiates these TFs to trigger the BR-mediated transcriptional cascade [131].

Similar to ethylene, BR is a hormone that can improve stress tolerance at the expense of altered growth and development. In contrast to the antagonistic role of ethylene and ABA in abiotic stress response, the synergistic interactions between ethylene and BR have been demonstrated in several aspects of plant stress responses in *Arabidopsis* and tomato [132]. For example, salt stress inhibits seed germination, whereas either exogenous ethylene or BR treatment increases germination rate by ameliorating the inhibitory effect of salt stress in *Arabidopsis* [133]. Further studies in *Arabidopsis* found that salt stress reduces ethylene production by repressing *ACO* expression in seed germination, whereas BR reverses the drop in ethylene biosynthesis under salt stress by recovering *ACO* transcript levels [133]. The stimulating effect of BR on ethylene evolution was also observed in tomato seedlings exposed to salt stress [118]. In this system, BR promotes ethylene biosynthesis and signaling by enhancing ACS activity and stabilizing the EILs [134]. The synergistic effect of BR and ethylene then leads to H_2O_2 accumulation and improved tomato seedling salt stress tolerance due to an increase in antioxidant enzyme activities [134]. Furthermore, inhibition of ethylene synthesis in cucumber blocks BR-relieved oxidative damage and thus reduces plant tolerance to salt, osmotic, and cold stress, suggesting that ethylene may also be required for BR-mediated abiotic stress tolerance [135]. Likewise, in rice, upon submergence in water, flooding stress induces the expression of an *ERF* gene *SUBMERGENCE1A* (*SUB1A*) which in turn increases BR levels by upregulating the transcription of BR biosynthetic genes. BR and the accumulated ethylene then work together to suppress rice shoot elongation, keeping the plants in a quiescent state and enabling plant survival [136], Figure 4.

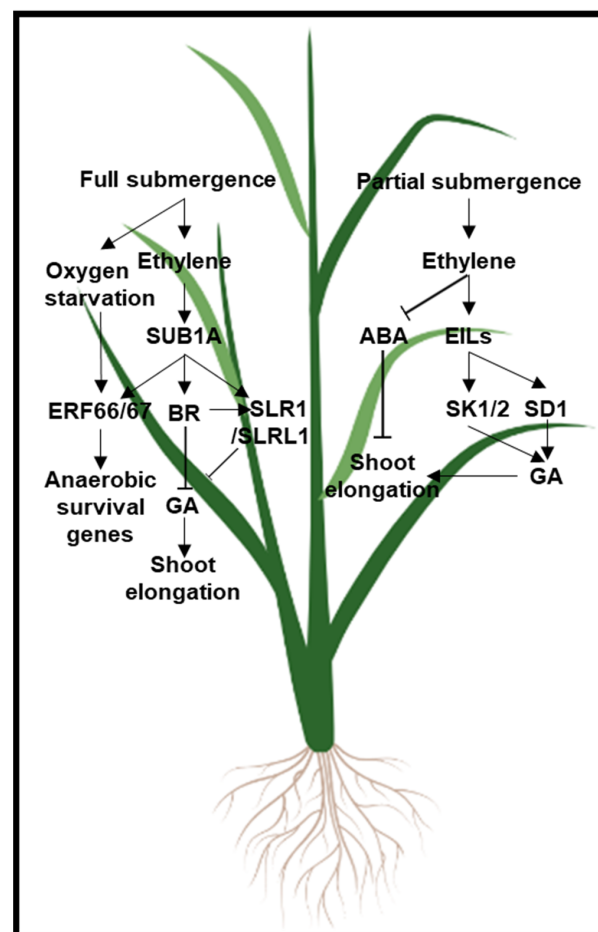


Figure 4. A model of rice adaptation to partial and complete water submergence regulated by GA,

ABA, BR, and ethylene. Upon full submergence of rice plants under water (left box), ethylene activates transcription of an ERF TF gene *SUB1A*, and the SUB1A protein then up-regulates anaerobic survival genes via the transcriptional induction and protein stabilization of ERF66/67 upon hypoxia stress. In addition, ethylene suppresses GA-mediated stem elongation by directly activating the transcription of DELLA protein genes, *SLR1* and *SLRL1*, as well as by inhibiting BR signaling. Upon partial submergence under water (right box), ethylene acts via the EIL proteins to induce the transcript levels of *ERF* genes *SK1* and *SK2* bringing about the GA signaling-mediated stem elongation in rice. In deepwater wild rice, EIL-mediated signaling triggered by the hormone ethylene directly enhances GA production and induces shoot elongation by transcriptionally activating a GA biosynthesis gene *SD1* that encodes a GA20-oxidase. Furthermore, ethylene promotes shoot elongation by relieving ABA-imposed shoot growth inhibition. Positive interactions such as activation, production, and stabilization are represented with lines that end in an arrowhead →. Negative interactions such as inactivation and repression are represented by lines that end in a hammerhead ⊥. The background image of a rice plant was created with BioRender.

Another interesting example that illustrates the synergistic regulatory effects of ethylene and BR is the process of stomatal closure. In *Arabidopsis*, BR induces stomatal closure by triggering H_2O_2 and NO production [110], Figure 3. BR-stimulated H_2O_2 and NO accumulation is significantly decreased in the dominant ethylene-insensitive mutant *etr1* but is strengthened in the ethylene overproducing *ACS5* mutant *ethylene overproducer1-1* [110], suggesting that BR-mediated stomatal closure is dependent on ethylene signaling.

Importantly, the synergistic effects of BR and ethylene have been observed not only in abiotic stress tolerance-related processes, but also in normal plant development, such as in root growth or fruit ripening [132]. However, most of the developmental and abiotic stress-related inquiries that explored the BR and ethylene crosstalk in vegetable species lack critical genetic evidence [134,135]. Looking ahead, the systematic analysis of mutants in key hormonal genes in both model and crop species would be necessary to further elucidate the underlying mechanisms of BR and ethylene crosstalk to shed light on the mechanisms that balance plant growth and stress tolerance at the phenotypic and molecular level.

5. Ethylene and Auxin

Auxin, whose name originates from the Greek “auxein” meaning “to grow/increase”, was first described to play roles in plant development and growth by a Dutch biologist Frits Warmolt Went [137]. The best-studied type of auxin, indole-3-acetic acid (IAA), is synthesized from the amino acid tryptophan through a two-step pathway. Tryptophan is converted to indole-3-pyruvic acid (IPyA) by the TRYPTOPHAN AMINOTRANSFERASE OF ARABIDOPSIS1 (TAA1)/TAA1-RELATED family, and IPyA is then metabolized to IAA by flavin-containing monooxygenases, YUCCAs (YUCs) [138]. IAA is perceived in the nucleus where it binds to the TRANSPORT INHIBITOR-RESISTANT1 (TIR1)/AUXIN SIGNALING F-BOX (AFB) family of auxin receptors. This hormone-binding enables the TIR1/AFB receptors to interact with AUXIN/INDOLE-3-ACETIC ACID (Aux/IAA) proteins, triggering Aux/IAA degradation and the release of AUXIN RESPONSE FACTORS (ARFs) which can now initiate transcriptional cascades of auxin-response genes [139].

The crosstalk between ethylene and auxin has been well characterized in recent years in many developmental processes, especially in root elongation, root hair formation, and adventitious root growth [140–142]. In most abiotic stress conditions, including high salinity and water deficit, the developmental plasticity of the plant root is regulated by the auxin-ethylene interaction, and the stress-induced remodeling of root architecture confers stress tolerance [143]. Excess aluminum (Al), one of the major toxic elements in acidic soils, alters the accumulation, polar transport, and the asymmetric distribution of auxin in *Arabidopsis* roots, leading to reduced root elongation and enhanced Al stress tolerance [144–147], Figure 5. These shifts in auxin abundance and spatial distribution in response to Al stress arise from changes in the TAA1- and YUC3/5/7/8/9-mediated local auxin biosynthesis [146,147] and PIN-FORMED2 (PIN2)- and AUXIN RESISTANT1 (AUX1)-mediated polar auxin transport in roots [53]. These processes, in turn, depend on

proper ethylene biosynthesis and signaling [53,146,147]. Ethylene signaling is thought to act upstream and synergistically with auxin to mediate Al-induced root growth inhibition in *Arabidopsis*. Furthermore, EIN3 directly activates the expression of *TAA1*, *YUC9*, and *AUX1* to induce auxin accumulation in root transition zones, resulting in the inhibition of root growth [21,53,146–148], Figure 5.

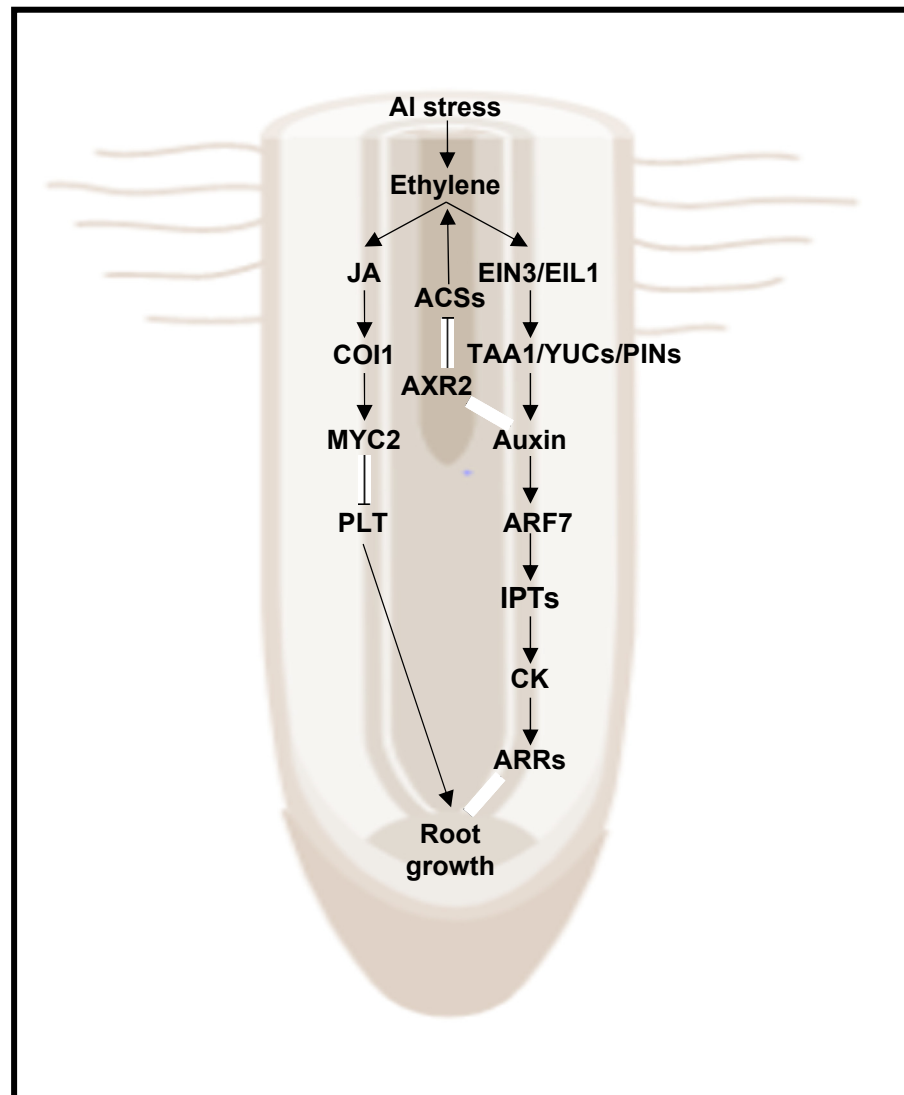


Figure 5. A model of root growth regulation in *Arabidopsis* triggered by Al stress and mediated by the crosstalk between JA, CK, auxin, and ethylene. Under Al stress, enhanced ethylene production upregulates the local transcript levels of auxin biosynthesis genes *TAA1* and *YUCs* promoted by the transcriptional upregulation of *EIN3* and *EIL1*. The increased protein abundance of *TAA1* and *YUCs* results in local auxin accumulation within the roots, suppressing primary root growth. Downstream of auxin, local accumulation of CK triggered by ARF7-mediated induction of the CK biosynthesis genes *IPTs* activates the CK response, which also contributes to root growth inhibition in response to Al stress. In parallel, COI1-mediated JA signaling downstream of ethylene is involved in Al stress-induced root growth inhibition through the transcriptional repression of *PLT* genes encoding AP2 TFs. Positive interactions such as activation, production, and stabilization are represented with lines that end in an arrowhead →. Negative interactions such as inactivation and repression are represented by lines that end in a hammerhead ⊥. The background image of an *Arabidopsis* root was created with BioRender.

Besides Al, other toxic metals characteristic of acidic soils, such as dichromate, cadmium, or excess iron, and alkaline soils have also been described to enhance ethylene production in *Arabidopsis* roots [148–152]. Plant responses to these toxic metals lead to root growth inhibition and involve enhanced ethylene production and signaling, which in turn induce auxin accumulation and polar auxin transport by upregulating the expression of *AUX1* and/or *PIN2* in *Arabidopsis* [148–153], Figure 5. The resulting root architecture changes triggered by these abiotic stresses in the soil and mediated by the ethylene-auxin crosstalk are thought to increase plant tolerance to these unfavorable conditions [143,152,153].

The auxin- and ethylene-mediated root architecture remodeling also drives plant tolerance in response to nutrient deficiencies [154]. To mitigate the magnesium deficiency stress, plants enhance root hair initiation and growth through the mutual activation of ethylene and auxin signaling [155]. Magnesium deficiency in *Arabidopsis* induces ethylene production that promotes auxin accumulation by activating *AUX1*, *PIN1*, and *PIN2* transcription in roots. The extra auxin stimulates further ethylene production by inducing the expression of *ACO* and *ACS* enzyme genes [155]. Knockout mutant analyses show that the roles of ethylene in regulating magnesium deficiency-induced root hair development require auxin signaling [155], suggesting that auxin acts primarily downstream of ethylene to regulate root hair morphogenesis. Similarly, ethylene signaling and ethylene-triggered downstream auxin and ROS signaling act together in the inhibition of root hair growth caused by boron deficiency, as evidenced by genetic and chemical treatment analyses [156,157]. Moreover, auxin, ethylene, and ROS also cooperate in regulating root architecture in response to mechanical impedance stress [158].

Another classical example of the ethylene and auxin crosstalk mediating abiotic stress tolerance and leading to major plant architecture and growth pattern changes is the response to flooding stress [159]. Ethylene and auxin interactions play important roles in adventitious root formation, which helps to relieve the flooding stress plants face [159]. In waterlogged *Rumex palustris*, the adventitious root formation is mediated by ethylene accumulation that enhances the sensitivity of root-forming tissues to auxin [160]. In tomato, waterlogging-induced adventitious root formation requires ethylene perception, as revealed by the study of an ethylene receptor mutant, *Never ripe* [161]. Flooding stress promotes ethylene production and, hence, ethylene signaling that, in turn, stimulates auxin transport to submerged stems to induce adventitious root growth [161]. In cucumber, ethylene signal transduction and ethylene-triggered auxin signaling act through the ROS pathway to induce adventitious root growth in response to flooding stress [162]. Together, these studies suggest that auxin signaling works downstream of ethylene to promote flooding stress-induced adventitious root formation in several species.

Besides making adventitious roots, submerged crop species, such as rice and *R. palustris*, often develop aerenchymatous tissue in both roots and shoots to facilitate gas diffusion to the roots to help cope with flooding-induced anoxia [163]. The rice mutant *iaa13* with suppressed auxin signaling shows reduced ethylene-induced aerenchyma formation under O₂-deprived conditions, along with the decreased expression of ethylene biosynthesis genes [164], supporting the notion that intricate ethylene-auxin interactions are involved in regulating aerenchyma formation in response to flooding/waterlogging stress.

The ethylene and auxin crosstalk is not limited to roots, but also plays a role in hypocotyl responses to heat stress. In *Arabidopsis* grown at normal temperatures in the light, ethylene-activated EIN3 promotes hypocotyl elongation, in part by stimulating *PIF3* expression [165]. Under elevated temperatures, EIN3 attenuates auxin-stimulated thermomorphogenic hypocotyl elongation through the inhibition of the apoplastic acidification progression by transcriptionally repressing the *APD7* protein phosphatase gene [166], suggesting antagonistic roles of auxin and ethylene in regulating hypocotyl response to heat stress. In contrast, auxin and ethylene show synergistic effects upon heat stress in roots. Elevated temperatures enhance TAA1-dependent auxin biosynthesis through ETR1-mediated

ethylene signaling, and the accumulated auxin and subsequent auxin transport are critical for maintaining normal root growth and gravity sensing under heat stress [167].

Taken together, the studies described above highlight the importance of altered root architecture controlled by ethylene-auxin interactions for conferring tolerance to water deficiency, heavy metals, high salinity, nutrient deprivation, and flooding. To further elucidate the mechanisms behind different plant strategies for root architecture remodeling in response to abiotic stress, mathematical models incorporating primary root initiation and lateral root growth governed by the auxin and ethylene crosstalk are required. By quantitatively evaluating the effects of the biosynthetic and signaling components of the individual auxin and ethylene pathways and the genes involved in the crosstalk between these two signals in response to different abiotic stresses, we can gain insights as to how to design and genetically modify plants with enhanced tolerance to these abiotic stress factors while minimizing the tradeoff in plant growth and development.

6. Ethylene and GA

GA, whose name originates from the fungus *Gibberella fujikuroi* that induces rapid growth in rice shoots, was first discovered in plants as a growth-regulating hormone [168]. GAs are derived from trans-geranylgeranyl diphosphate, which is first converted to entkaurene and then to GA₁₂, a nonbioactive GA. GA₁₂ is then transformed into bioactive GAs, such as GA₁, GA₃, GA₄, and GA₇ by the GA₂₀- and GA₃-OXIDASE family enzymes [169]. The bioactive GAs in the nucleus bind to the soluble GA receptor GIBBERELLIN-INSENSITIVE DWARF1 (GID1) that interacts with DELLA proteins. When GA-GID1-DELLA complex forms, DELLAs undergo structural changes that enable their recognition by GID2/SLEEPY1 F-box proteins and targeting for proteasomal degradation. With DELLAs degraded, the TFs deactivated by DELLA are released and can initiate the GA-mediated transcriptional cascade [170].

The role of ethylene in plant responses to flooding has been studied in several species. Two distinct ethylene-and-GA crosstalk mechanisms in plants coping with different types of flooding stress have been elucidated to date. In rice, flooding rapidly promotes ethylene accumulation [159]. Upon partial submergence in water, ethylene activates the transcription of class VII ERFs *SNORKEL1* (SK1) and SK2, and in turn, SK1 and SK2 induce GA signaling-mediated stem elongation in rice [170,171], Figure 4. Some deep-water wild rice varieties elongate their stems through the OsEIL1-mediated induction of a GA biosynthetic gene *SEMI-DWARF1* (SD1), the gene whose loss-of-function allele in domesticated rice has been at the heart of the Green Revolution [172]. The stem elongation enables rice to escape out from the water and perform photosynthesis to alleviate the flooding stress. With full submergence under water, flooding stress becomes more detrimental for plants, primarily because of limited gas exchange under water, which leads to an energy and carbohydrate deficit. In coping with full submergence under water, rice deprioritizes growth to reach increased hypoxia tolerance. In this way, another rice ERF, SUB1A, with high protein similarity to SKs, is transcriptionally induced by full submergence flooding stress through ethylene signaling, not only dampening ethylene biosynthesis [173,174], but also suppressing GA-mediated stem elongation by directly activating the transcription of DELLA protein genes *SLENDER RICE1* (SLR1) and *SLENDER RICE-LIKE1* (SLRL1) [175], Figure 4. Furthermore, SUB1A-1 directly activates the transcription of *ERF66* and *ERF67*, whose overexpression activates the expression of anaerobic survival genes and whose protein stability is enhanced under hypoxia stress upon full plant submergence in water [176]. In line with these molecular studies, rice varieties possessing the *SUB1A-1* gene are tolerant to prolonged full flooding [173,174,177]. These two opposite plant responses to partial submergence and full submergences, which rely on different ethylene-induced ERFs and their crosstalk with GA (Figure 4), are exciting examples of how plants can differentially adapt to variable environments to maximize plant survival.

Two contrasting regulatory effects of the ethylene-GA crosstalk on plant growth have also been described in response to osmotic stress. Upon exposure to mannitol, *Arabidopsis*

plants synthesize and accumulate ethylene in actively dividing leaf cells, ultimately leading to the induction of ethylene signaling genes *ETR2*, *ERS1*, *CTR1*, *EIN3*, *EIL1*, *EBF1*, and *EBF2* and several *ERF* genes including *ERF6* and *ERF11* [31]. Studies show that overexpression of *ERF6*, a direct transcriptional target of EIN3 in Arabidopsis [128], not only results in the increased expression of *GA2-OXIDASE6* that degrades GA, but also in the enhanced accumulation of DELLAs due to the stabilization of these regulatory proteins, which in turn inhibit the shoot and leaf growth [177]. Conversely, another mannitol-induced ERF, *ERF11*, promotes internode elongation and leaf growth by directly increasing GA biosynthesis and, therefore, DELLA degradation and indirectly inhibiting ethylene biosynthesis via transcriptional repression of ACS genes [178,179]. Interestingly, both *ERF6*-overexpression lines and *erf11* loss-of-function mutants are more tolerant to mannitol [178,179]. Taken together, these studies demonstrate that *ERF6* and *ERF11* in Arabidopsis seedlings have opposing roles in regulating the tradeoff between growth and osmotic stress tolerance. Follow-up work has shown that *ERF6* acts as an upstream regulator of *ERF11* through direct transcriptional activation [178,180], suggesting that the effect of ethylene and GA interaction may quickly shift in response to abiotic stress.

To sum up, the interaction between GA and ethylene in abiotic stress tolerance is primarily elucidated at the transcriptional level, with the two hormones affecting the same set of TFs. As in the case of JA-ethylene synergy in etiolated Arabidopsis seedlings, GA was also discovered to enhance apical hook curvature, at least in part via a release of EIN3 from the DELLA-EIN3 repressive complex [21,181]. Given that the JA signaling components JAZs and MYCs form repressive complexes with EIN3/EIL1 [78], constructing a transcriptional regulatory network incorporating JA, GA, and ethylene TFs under different types of abiotic stresses would be important to understand how plants cope with adverse environmental conditions by fine-tuning the transcriptional cascade through leveraging the hormone crosstalk.

7. Ethylene and SA

SA, whose name comes from the Latin word *salix* (willow), is a phenolic compound produced by plants in response to pathogen exposure and was first identified as a plant immunity-related hormone [182]. SA is synthesized from chorismate through two independent pathways, the isochorismate, and the phenylalanine ammonia-lyase (PAL) pathways. SA binds to NON-EXPRESSOR OF PATHOGENESIS-RELATED1 (NPR1) oligomers and causes the complex to dissociate into monomers that migrate to the nucleus. In the nucleus, NPR1 interacts with the TGACG-BINDING FACTOR family of TFs to induce the expression of *PATHOGENESIS-RELATED* genes [183].

The rapidly induced ethylene production triggered by environmental stress is thought to modulate oxidative stress, which in turn affects plant growth [184]. SA has been reported to inhibit the ethylene production induced by several types of abiotic stresses [185,186]. For example, exogenous SA alleviates heat stress by increasing proline-metabolism and restricting ethylene biosynthesis in heat-stressed plants by inhibiting the ACS activity in wheat [187]. Similarly, the SA-mediated ACS activity inhibition enhances plant tolerance to salt and drought stress in mustard, mung bean, and sweet pepper [188–190]. Additionally, exogenous SA has been described to block the conversion of ACC to ethylene, possibly by inhibiting the ACO activity as demonstrated in pear suspension cells [191]. On the contrary, ethylene and SA act in concert to regulate cell death in Arabidopsis and tobacco leaves under ozone (O₃) stress [192,193]. O₃-induced ethylene promotes SA biosynthesis by up-regulating the expression of the SA biosynthetic genes encoding CHORISMATE MUTASE and PAL [193]. Inhibition of either ethylene biosynthesis or SA biosynthesis in Arabidopsis rescues the O₃-induced hypersensitive response cell death phenotype, whereas overproduction of ethylene aggravates cell death [192]. On the other hand, in Arabidopsis, chemical or genetic perturbation of SA signaling blocks ethylene production in response to O₃, suggesting that SA is required for O₃-induced ethylene biosynthesis [192].

In summary, many characterized SA-ethylene interactions in abiotic stress responses involve the mutual regulation of these hormones' biosynthetic pathways. The synergistic effects of ethylene and SA in abiotic stress tolerance propose that SA and ethylene may leverage some common machinery to regulate downstream genes, making this an intriguing possibility to investigate in future studies. Furthermore, given the synergistic nature of the JA-ethylene crosstalk at the transcriptional level and the antagonistic crosstalk between the SA and JA signaling pathways [194], elucidation of the genetic interplay between SA, ethylene, and JA would be another interesting opportunity to explore in the future.

8. Ethylene and CK

CKs, whose effects were first uncovered by studying plant shoot induction in tissue culture in response to coconut milk, are involved in the regulation of cell growth and differentiation [195]. CKs are adenine-derived hormones that are synthesized via multiple-step reactions involving ISOPENTENYL TRANSFERASE (IPT), CYTOCHROME P450 FAMILY 735A, and LONELY GUY enzymes [196]. In cells, CKs bind to the receptors ARABIDOPSIS HISTIDINE KINASEs (AHKs) and induce their autophosphorylation. AHKs also phosphorylate ARABIDOPSIS HISTIDINE PHOSPHOTRANSMITTER (AHP) proteins that in turn activate type-B ARABIDOPSIS RESPONSE REGULATOR (ARR) TFs to initiate CK-mediated transcription [195].

Just like in the case of stress-triggered ethylene production, abiotic factors can rapidly modulate CK levels in plants by altering the expression of CK biosynthetic genes [197]. In Arabidopsis, a high level of CK has been described to stabilize ACS5 and ACS9 [198–200], thus leading to ethylene accumulation. Reciprocally, Al stress-induced ethylene is known to enhance local CK biosynthesis by activating the expression of CK biosynthetic genes encoding IPTs, leading to root growth inhibition under Al stress [148,201], Figure 5. Not only do ethylene and CK mutually regulate each other's biosynthetic pathways, but also ethylene interferes with the CK signaling output at the level of transcriptional regulation [51]. Specifically, Arabidopsis EIN3 directly suppresses the expression of the CK signaling repressor genes, type-A ARRs ARR5, ARR7, and ARR15, by directly binding to their promoters [51]. Overexpression of these three ARRs confers enhanced freezing tolerance [51], suggesting that these genes serve as mediators of the crosstalk between ethylene and CK in the cold stress response [51].

Thus, the bulk of current studies focusing on CK and ethylene interactions in abiotic stress are consistent with the model where CK induces ethylene biosynthesis and, vice versa, ethylene induces CK biosynthesis and signaling. However, these two hormones also intersect in normal plant development. For example, in root growth regulation, the ethylene receptor ETR1 upon binding ethylene was shown to directly activate a multistep phosphorelay CK signal transduction cascade (AHK-AHP-ARRs), integrating ethylene and CK in the control of root apical meristem size [202,203]. It will be interesting to test if the ETR1-mediated CK signaling also plays a role in abiotic stress responses.

9. Concluding Remarks and Future Perspectives

In this review, we summarized several lines of evidence supporting the notion that the growth-stress tolerance trade-off is in part regulated by the active crosstalk between ethylene and other plant hormones. The pertinent knowledge acquired in the past few decades about ethylene signaling and its interactions with other hormones will guide scientists in their efforts to engineer better-performing crops in which stress responses are uncoupled from the growth penalty, and high yields are maintained under suboptimal environmental conditions. As described above, some hormone interactions enable plants to enhance the growth of certain plant organs or tissues in response to specific stresses, and such selective growth helps the overall plant survival and ultimate productivity by reducing the adverse effects of stress. The rational strategy would then be to equip crops with such specialized tissue-specific responses that permit plant adaptation to the corresponding types of stress. For example, by modulating the auxin-ethylene crosstalk in rice roots,

aerenchymatous tissue formation could be induced to improve crop growth and yield under oxygen-deficient conditions [164,204–206]. Similarly, crops with enhanced adventitious roots, lateral roots, and root hairs can be obtained by engineering JA, auxin, or ethylene crosstalk, and these root-boosted plants would have better growth and yield in nutrient-poor soils and under water-scarce conditions, as has been demonstrated in multiple studies, including those performed on tomato [161] and maize [207–209]. Alternatively, under some abiotic stress conditions, rational manipulation of hormone crosstalk can deprioritize the plant growth but trigger specialized metabolism and antioxidant systems to enhance plant stress tolerance and survival. Yet, in other scenarios, the hormone crosstalk can maximize organ growth to enable the plant to escape from unfavorable conditions. The seemingly contradictory approach is well-illustrated by the aforementioned ethylene-GA crosstalk-mediated flooding stress response in rice, with opposite strategies adopted in full versus partial submergence conditions (Figure 4). In fact, both strategies have been implemented in transgenic plants to enhance flooding stress tolerance [170,174,210].

Given the great potential of hormone engineering in improving plant growth and crop yield under unfavorable environmental conditions, future efforts should also be directed at bettering our understanding of hormone interactions at the basic mechanistic level. At present, many gaps remain in our comprehension of how the hormone crosstalk is modified by the environment and serves to balance plant growth with abiotic stress responses. For example, while we know that the endogenous levels of ethylene and other hormones in plants are rapidly altered in response to abiotic stresses, the mechanisms that detect and then translate the environmental signals into changes in the biosynthesis or catabolism of these growth regulators are only starting to get unraveled. Hypoxia or flooding stress, for example, could, in theory, be directly sensed by the loss of oxidation of the N termini of proteins, as may be the case for the ACO1 enzyme involved ethylene production and a subgroup of ERFs involved in ethylene responses, which ultimately modulate the plant tolerance to these types of stresses [211–215]. Likewise, heat stress may be sensed by the red/far-red light receptor PHYTOCHROME B [216,217], osmotic/drought stress is perceived by the calcium channel REDUCED HYPEROSMOLARITY-INDUCED $[Ca^{2+}]$ INCREASE1 (OSCA1) and its paralogs [218], cold stress is detected by the OPEN STOMATA1 pathway [219], and salt stress is discerned via binding of monovalent cations to the glucuronic acid of the glycosyl inositol phosphoryl ceramide sphingolipids [219]. However, besides the aforementioned flooding stress-induced hypoxia, it remains elusive how this initial perception is mechanistically linked to the changes in hormone biosynthesis and/or signaling and how the coordination between the respective hormonal pathways is achieved. Furthermore, plants in the real world are often exposed to various combinations of abiotic stresses, and it remains to be deciphered how ethylene and other hormones are leveraged by plants to integrate the inputs from multiple stress signals and at what level the signaling cascades intersect and feedback onto one another. With the development of sensitive analytical and genetic methods of hormone quantification and monitoring, as well as various high-throughput omics approaches and mathematical models, complex relationships can now be explored at multiple levels, from protein-protein or protein-DNA interactions to intricate transcriptional regulatory networks. These research directions are expected to provide new insights into the mechanisms of adaptive responses co-regulated by multiple hormones at once.

Another interesting and important aspect of the signal crosstalk that needs to be explored at a much greater depth is the spatiotemporal regulation of hormone interactions. The response of plants to any stress is dynamic and often tissue-specific [220]. Therefore, monitoring the temporal evolution of hormone crosstalk in specific tissues or developmental stages under specific conditions (e.g., in the dynamic regulation of stomatal opening and closure in leaves under drought stress, or time-series documenting the process of lateral root initiation and outgrowth in response to nutrient deprivation) is an important endeavor critical for the future of plant trait improvement efforts. Perhaps, a combination of single-cell omics approaches with the development of new, more sensitive versions of

hormone reporters and biosensors, calcium, and ROS sensors, etc. and stacking of these detection devices in model plants to monitor the activity of multiple hormones and second messengers at once will be instrumental for reconstructing the sequence of events from the moment the stress is perceived to the phenotypic outcomes. Translating the gained knowledge to non-model crop plants would greatly impact agriculture.

In conclusion, the network of hormone interactions orchestrates both normal plant development and responses to biotic and abiotic stress. How a handful of growth regulators are leveraged by plants to interpret a combination of internal and environmental inputs and tip the balance between resource allocation towards growth versus defense and survival is a fascinating long-term question that will be key to our ability to maintain and further improve agricultural production under deteriorating climates.

Author Contributions: Conceptualization, H.C., A.N.S. and J.M.A.; writing—original draft preparation, H.C. and A.N.S.; writing—review and editing, H.C., A.N.S., J.M.A. and D.A.B.J.; preparation of Figures and Table, H.C., A.N.S. and J.M.A.; funding assistance, A.N.S. and J.M.A. All authors have read and agreed to the published version of the manuscript.

Funding: The research in the Alonso-Stepanova lab is supported by the National Science Foundation grants 1650139, 1444561, and 1940829 to A.N.S. and J.M.A., and 1750006 to A.N.S.

Data Availability Statement: Not applicable.

Acknowledgments: We are thankful to Chengsong Zhao, Mario Fenech, and Anna Yaschenko for the critical reading of this manuscript.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Zhang, H.; Zhu, J.; Gong, Z.; Zhu, J. Abiotic stress responses in plants. *Nat. Rev. Genet.* **2021**, *21*, 1–16. [[CrossRef](#)] [[PubMed](#)]
2. Chapin, F.S. Integrated Responses of Plants to Stress. *BioScience* **1991**, *41*, 29–36. [[CrossRef](#)]
3. Heil, M.; Walters, D.; Newton, A.; Lyon, G. Trade-offs associated with induced resistance. In *Induced Resistance for Plant Defense*; John Wiley & Sons: Chichester, UK, 2014; pp. 171–192.
4. Neljubow, D. Über die horizontale Nutation der Stengel von Pisum Sativum und einiger anderen Pflanzen. *Bot. Centralbl. Beih.* **1901**, *10*, 128–139.
5. Merchante, C.; Stepanova, A.N. The Triple Response Assay and Its Use to Characterize Ethylene Mutants in Arabidopsis. *Ethyl. Signal.* **2017**, *1573*, 163–209. [[CrossRef](#)]
6. Bleecker, A.B.; Kende, H. Ethylene: A Gaseous Signal Molecule in Plants. *Annu. Rev. Cell Dev. Biol.* **2000**, *16*, 1–18. [[CrossRef](#)]
7. Binder, B.M. Ethylene signaling in plants. *J. Biol. Chem.* **2020**, *295*, 7710–7725. [[CrossRef](#)]
8. Adams, D.O.; Yang, S.F. Ethylene biosynthesis: Identification of 1-aminocyclopropane-1-carboxylic acid as an intermediate in the conversion of methionine to ethylene. *Proc. Natl. Acad. Sci. USA* **1979**, *76*, 170–174. [[CrossRef](#)]
9. Boller, T.; Herner, R.C.; Kende, H. Assay for and enzymatic formation of an ethylene precursor, 1-aminocyclopropane-1-carboxylic acid. *Planta* **1979**, *145*, 293–303. [[CrossRef](#)]
10. Hamilton, A.J.; Bouzayen, M.; Grierson, D. Identification of a tomato gene for the ethylene-forming enzyme by expression in yeast. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 7434–7437. [[CrossRef](#)]
11. Verweridis, P.; John, P. Complete recovery in vitro of ethylene-forming enzyme activity. *Phytochemistry* **1991**, *30*, 725–727. [[CrossRef](#)]
12. Kende, H. Ethylene biosynthesis. *Annu. Rev. Plant Biol.* **1993**, *44*, 283–307. [[CrossRef](#)]
13. Xu, J.; Zhang, S. Ethylene Biosynthesis and Regulation in Plants. In *Ethylene in Plants*; Wen, C.-K., Ed.; Springer: Dordrecht, The Netherlands, 2015; pp. 1–25. ISBN 978-94-017-9483-1.
14. Alonso, J.M.; Stepanova, A.N. The Ethylene Signaling Pathway. *Science* **2004**, *306*, 1513–1515. [[CrossRef](#)] [[PubMed](#)]
15. Lacey, R.F.; Binder, B.M. How plants sense ethylene gas—The ethylene receptors. *J. Inorg. Biochem.* **2014**, *133*, 58–62. [[CrossRef](#)] [[PubMed](#)]
16. Shakeel, S.N.; Gao, Z.; Amir, M.; Chen, Y.-F.; Rai, M.I.; Haq, N.U.; Schaller, G.E. Ethylene Regulates Levels of Ethylene Receptor/CTR1 Signaling Complexes in Arabidopsis thaliana. *J. Biol. Chem.* **2015**, *290*, 12415–12424. [[CrossRef](#)]
17. Li, W.; Ma, M.; Feng, Y.; Li, H.; Wang, Y.; Ma, Y.; Li, M.; An, F.; Guo, H. EIN2-Directed Translational Regulation of Ethylene Signaling in Arabidopsis. *Cell* **2015**, *163*, 670–683. [[CrossRef](#)] [[PubMed](#)]
18. Merchante, C.; Brumos, J.; Yun, J.; Hu, Q.; Spencer, K.R.; Enríquez, P.; Binder, B.M.; Heber, S.; Stepanova, A.N.; Alonso, J.M. Gene-Specific Translation Regulation Mediated by the Hormone-Signaling Molecule EIN2. *Cell* **2015**, *163*, 684–697. [[CrossRef](#)]

19. Guo, H.; Ecker, J.R. Plant responses to ethylene gas are mediated by SCFEBF1/EBF2-dependent proteolysis of EIN3 transcription factor. *Cell* **2003**, *115*, 667–677. [\[CrossRef\]](#)
20. Potuschak, T.; Lechner, E.; Parmentier, Y.; Yanagisawa, S.; Grava, S.; Koncz, C.; Genschik, P. EIN3-Dependent Regulation of Plant Ethylene Hormone Signaling by Two Arabidopsis F Box Proteins: EBF1 and EBF2. *Cell* **2003**, *115*, 679–689. [\[CrossRef\]](#)
21. Dolgikh, V.A.; Pukhovaya, E.M.; Zemlyanskaya, E.V. Shaping ethylene response: The role of EIN3/EIL1 transcription factors. *Front. Plant Sci.* **2019**, *10*, 1030. [\[CrossRef\]](#)
22. Debbarma, J.; Sarki, Y.N.; Saikia, B.; Boruah, H.P.D.; Singha, D.L.; Chikkaputtaiah, C. Ethylene Response Factor (ERF) Family Proteins in Abiotic Stresses and CRISPR-Cas9 Genome Editing of ERFs for Multiple Abiotic Stress Tolerance in Crop Plants: A Review. *Mol. Biotechnol.* **2019**, *61*, 153–172. [\[CrossRef\]](#)
23. Dietz, K.; Vogel, M.O.; Viehhauser, A. AP2/EREBP transcription factors are part of gene regulatory networks and integrate metabolic, hormonal and environmental signals in stress acclimation and retrograde signalling. *Protoplasma* **2010**, *245*, 3–14. [\[CrossRef\]](#)
24. Mizoi, J.; Shinozaki, K.; Yamaguchi-Shinozaki, K. AP2/ERF family transcription factors in plant abiotic stress responses. *Biochim. Biophys. Acta Gene Regul. Mech.* **2012**, *1819*, 86–96. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Chandler, J.W. Class VIIIb APETALA2 Ethylene Response Factors in Plant Development. *Trends Plant. Sci.* **2018**, *23*, 151–162. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Nakano, T.; Suzuki, K.; Fujimura, T.; Shinshi, H. Genome-Wide Analysis of the ERF Gene Family in Arabidopsis and Rice. *Plant. Physiol.* **2006**, *140*, 411–432. [\[CrossRef\]](#) [\[PubMed\]](#)
27. Kagale, S.; Rozwadowski, K. EAR motif-mediated transcriptional repression in plants: An underlying mechanism for epigenetic regulation of gene expression. *Epigenetics* **2011**, *6*, 141–146. [\[CrossRef\]](#) [\[PubMed\]](#)
28. Tiwari, S.B.; Belachew, A.; Ma, S.F.; Young, M.; Ade, J.; Shen, Y.; Marion, C.M.; Holtan, H.E.; Bailey, A.; Stone, J.K. The EDLL motif: A potent plant transcriptional activation domain from AP2/ERF transcription factors. *Plant J.* **2012**, *70*, 855–865. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Harkey, A.; Yoon, G.M.; Seo, D.H.; Delong, A.; Muday, G.K. Light Modulates Ethylene Synthesis, Signaling, and Downstream Transcriptional Networks to Control Plant Development. *Front. Plant. Sci.* **2019**, *10*, 1094. [\[CrossRef\]](#) [\[PubMed\]](#)
30. Pierik, R.; Sasidharan, R.; Voesenek, L.A.C.J. Growth Control by Ethylene: Adjusting Phenotypes to the Environment. *J. Plant. Growth Regul.* **2007**, *26*, 188–200. [\[CrossRef\]](#)
31. Kazan, K. Diverse roles of jasmonates and ethylene in abiotic stress tolerance. *Trends Plant. Sci.* **2015**, *20*, 219–229. [\[CrossRef\]](#)
32. Kurepin, L.V.; Yeung, E.C.; Reid, D.M.; Pharis, R.P. Light Signaling Regulates Tulip Organ Growth and Ethylene Production in a Tissue-Specific Manner. *Int. J. Plant. Sci.* **2016**, *177*, 339–346. [\[CrossRef\]](#)
33. Maksymiec, W. Signaling responses in plants to heavy metal stress. *Acta Physiol. Plant.* **2007**, *29*, 177–187. [\[CrossRef\]](#)
34. Argueso, C.T.; Hansen, M.; Kieber, J.J. Regulation of Ethylene Biosynthesis. *J. Plant. Growth Regul.* **2007**, *26*, 92–105. [\[CrossRef\]](#)
35. Zhao, M.; Liu, W.; Xia, X.; Wang, T.; Zhang, W. Cold acclimation-induced freezing tolerance of Medicago truncatula seedlings is negatively regulated by ethylene. *Physiol. Plant.* **2014**, *152*, 115–129. [\[CrossRef\]](#)
36. Zhang, M.; Smith, J.A.C.; Harberd, N.P.; Jiang, C. The regulatory roles of ethylene and reactive oxygen species (ROS) in plant salt stress responses. *Plant Mol. Biol.* **2016**, *91*, 651–659. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Dubois, M.; Claeys, H.; Broeck, L.V.D.; Inzé, D. Time of day determines Arabidopsis transcriptome and growth dynamics under mild drought. *Plant Cell Environ.* **2017**, *40*, 180–189. [\[CrossRef\]](#)
38. Baerenfaller, K.; Massonnet, C.; Walsh, S.; Baginsky, S.; Bühlmann, P.; Hennig, L.; Hirsch-Hoffmann, M.; Howell, K.; Kahlau, S.; Radziejewski, A.; et al. Systems-based analysis of Arabidopsis leaf growth reveals adaptation to water deficit. *Mol. Syst. Biol.* **2012**, *8*, 606. [\[CrossRef\]](#)
39. Clauw, P.; Coppens, F.; De Beuf, K.; Dhondt, S.; Van Daele, T.; Maleux, K.; Storme, V.; Clement, L.; Gonzalez, N.; Inzé, D. Leaf Responses to Mild Drought Stress in Natural Variants of Arabidopsis. *Plant Physiol.* **2015**, *167*, 800–816. [\[CrossRef\]](#) [\[PubMed\]](#)
40. Peng, H.-P.; Lin, T.-Y.; Wang, N.N.; Shih, M.-C. Differential expression of genes encoding 1-aminocyclopropane-1-carboxylate synthase in Arabidopsis during hypoxia. *Plant Mol. Biol.* **2005**, *58*, 15–25. [\[CrossRef\]](#)
41. Eun, H.-D.; Ali, S.; Jung, H.; Kim, K.; Kim, W.-C. Profiling of ACC synthase gene (ACS11) expression in Arabidopsis induced by abiotic stresses. *Appl. Biol. Chem.* **2019**, *62*, 42. [\[CrossRef\]](#)
42. Wang, X.; Ma, Q.; Wang, R.; Wang, P.; Liu, Y.; Mao, T. Submergence stress-induced hypocotyl elongation through ethylene signaling-mediated regulation of cortical microtubules in Arabidopsis. *J. Exp. Bot.* **2019**, *71*, 1067–1077. [\[CrossRef\]](#)
43. Dong, H.; Zhen, Z.; Peng, J.; Chang, L.; Gong, Q.; Wang, N.N. Loss of ACS7 confers abiotic stress tolerance by modulating ABA sensitivity and accumulation in Arabidopsis. *J. Exp. Bot.* **2011**, *62*, 4875–4887. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Skirycz, A.; Claeys, H.; De Bodt, S.; Oikawa, A.; Shinoda, S.; Andriankaja, M.; Maleux, K.; Eloy, N.; Coppens, F.; Yoo, S.-D.; et al. Pause-and-Stop: The Effects of Osmotic Stress on Cell Proliferation during Early Leaf Development in Arabidopsis and a Role for Ethylene Signaling in Cell Cycle Arrest. *Plant Cell* **2011**, *23*, 1876–1888. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Zhao, X.-C.; Schaller, G. Effect of salt and osmotic stress upon expression of the ethylene receptor ETR1 in Arabidopsis thaliana. *FEBS Lett.* **2004**, *562*, 189–192. [\[CrossRef\]](#)
46. Wang, Y.; Liu, C.; Li, K.; Sun, F.; Hu, H.; Li, X.; Zhao, Y.; Han, C.; Zhang, W.; Duan, Y.; et al. Arabidopsis EIN2 modulates stress response through abscisic acid response pathway. *Plant Mol. Biol.* **2007**, *64*, 633–644. [\[CrossRef\]](#)

47. Ahmadizadeh, M.; Chen, J.-T.; Hasanzadeh, S.; Ahmar, S.; Heidari, P. Insights into the genes involved in the ethylene biosynthesis pathway in *Arabidopsis thaliana* and *Oryza sativa*. *J. Genet. Eng. Biotechnol.* **2020**, *18*, 62. [\[CrossRef\]](#)
48. Yan, Z.; Wang, J.; Wang, F.; Xie, C.; Lv, B.; Yu, Z.; Dai, S.; Liu, X.; Xia, G.; Tian, H. MPK3/6-induced degradation of ARR1/10/12 promotes salt tolerance in *Arabidopsis*. *EMBO Rep.* **2021**, *22*, e52457. [\[CrossRef\]](#) [\[PubMed\]](#)
49. Peng, J.; Li, Z.; Wen, X.; Li, W.; Shi, H.; Yang, L.; Zhu, H.; Guo, H. Salt-induced stabilization of EIN3/EIL1 confers salinity tolerance by deterring ROS accumulation in *Arabidopsis*. *PLoS Genet.* **2014**, *10*, e1004664. [\[CrossRef\]](#)
50. Sun, L.; Jing, Y.; Liu, X.; Li, Q.; Xue, Z.; Cheng, Z.; Wang, D.; He, H.; Qian, W. Heat stress-induced transposon activation correlates with 3D chromatin organization rearrangement in *Arabidopsis*. *Nat. Commun.* **2020**, *11*, 1886. [\[CrossRef\]](#)
51. Shi, Y.; Tian, S.; Hou, L.; Huang, X.; Zhang, X.; Guo, H.; Yang, S. Ethylene Signaling Negatively Regulates Freezing Tolerance by Repressing Expression of CBF and Type-A ARR Genes in *Arabidopsis*. *Plant Cell* **2012**, *24*, 2578–2595. [\[CrossRef\]](#)
52. Schellingen, K.; Van Der Straeten, D.; Vandenbussche, F.; Prinsen, E.; Remans, T.; Vangronsveld, J.; Cuypers, A. Cadmium-induced ethylene production and responses in *Arabidopsis thaliana* rely on ACS2 and ACS6 gene expression. *BMC Plant Biol.* **2014**, *14*, 214. [\[CrossRef\]](#)
53. Sun, P.; Tian, Q.-Y.; Chen, J.; Zhang, W.-H. Aluminium-induced inhibition of root elongation in *Arabidopsis* is mediated by ethylene and auxin. *J. Exp. Bot.* **2009**, *61*, 347–356. [\[CrossRef\]](#)
54. Liang, X.; Shen, N.F.; Theologis, A. Li+-regulated 1-aminocyclopropane-1-carboxylate synthase gene expression in *Arabidopsis thaliana*. *Plant J.* **1996**, *10*, 1027–1036. [\[CrossRef\]](#)
55. Cao, S.; Chen, Z.; Liu, G.; Jiang, L.; Yuan, H.; Ren, G.; Bian, X.; Jian, H.; Ma, X. The *Arabidopsis* Ethylene-Insensitive 2 gene is required for lead resistance. *Plant Physiol. Biochem.* **2009**, *47*, 308–312. [\[CrossRef\]](#)
56. Husain, T.; Fatima, A.; Suhel, M.; Singh, S.; Sharma, A.; Prasad, S.M.; Singh, V.P. A brief appraisal of ethylene signaling under abiotic stress in plants. *Plant Signal. Behav.* **2020**, *15*, 1782051. [\[CrossRef\]](#)
57. Druege, U. Ethylene and Plant Responses to Abiotic Stress. In *Ethylene Action in Plants*; Springer: Berlin/Heidelberg, Germany, 2006; pp. 81–118.
58. Steffens, B. The role of ethylene and ROS in salinity, heavy metal, and flooding responses in rice. *Front. Plant Sci.* **2014**, *5*, 685. [\[CrossRef\]](#) [\[PubMed\]](#)
59. Nazar, R.; Khan, I.R.; Iqbal, N.; Masood, A.; Khan, N.A. Involvement of ethylene in reversal of salt-inhibited photosynthesis by sulfur in mustard. *Physiol. Plant.* **2014**, *152*, 331–344. [\[CrossRef\]](#)
60. Sharma, A.; Kumar, V.; Sidhu, G.P.S.; Kumar, R.; Kohli, S.K.; Yadav, P.; Kapoor, D.; Bali, A.S.; Shahzad, B.; Khanna, K. Abiotic stress management in plants: Role of ethylene. In *Molecular Plant Abiotic Stress: Biology and Biotechnology*; Wiley: Hoboken, NJ, USA, 2019; pp. 185–208.
61. Ku, Y.-S.; Sintaha, M.; Cheung, M.-Y.; Lam, H.-M. Plant Hormone Signaling Crosstalks between Biotic and Abiotic Stress Responses. *Int. J. Mol. Sci.* **2018**, *19*, 3206. [\[CrossRef\]](#)
62. Devireddy, A.R.; Zandalinas, S.I.; Fichman, Y.; Mittler, R. Integration of reactive oxygen species and hormone signaling during abiotic stress. *Plant J.* **2020**, *105*, 459–476. [\[CrossRef\]](#)
63. Knight, H.; Zarka, D.G.; Okamoto, H.; Thomashow, M.F.; Knight, M.R. Absciscic Acid Induces CBF Gene Transcription and Subsequent Induction of Cold-Regulated Genes via the CRT Promoter Element. *Plant Physiol.* **2004**, *135*, 1710–1717. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Yang, J.; Duan, G.; Li, C.; Liu, L.; Han, G.; Zhang, Y.; Wang, C. The Crosstalks Between Jasmonic Acid and Other Plant Hormone Signaling Highlight the Involvement of Jasmonic Acid as a Core Component in Plant Response to Biotic and Abiotic Stresses. *Front. Plant Sci.* **2019**, *10*, 1349. [\[CrossRef\]](#)
65. Demole, E.; Lederer, E.; Mercier, D. Isolement et détermination de la structure du jasmonate de méthyle, constituant odorant caractéristique de l'essence de jasmin. *Helv. Chim. Acta* **1962**, *45*, 675–685. [\[CrossRef\]](#)
66. Iqbal, N.; Khan, N.A.; Ferrante, A.; Trivellini, A.; Francini, A.; Khan, M.I.R. Ethylene Role in Plant Growth, Development and Senescence: Interaction with Other Phytohormones. *Front. Plant Sci.* **2017**, *8*, 475. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Wasternack, C.; Hause, B. Jasmonates: Biosynthesis, perception, signal transduction and action in plant stress response, growth and development. An update to the 2007 review in *Annals of Botany*. *Ann. Bot.* **2013**, *111*, 1021–1058. [\[CrossRef\]](#)
68. Ruan, J.; Zhou, Y.; Zhou, M.; Yan, J.; Khurshid, M.; Weng, W.; Cheng, J.; Zhang, K. Jasmonic Acid Signaling Pathway in Plants. *Int. J. Mol. Sci.* **2019**, *20*, 2479. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Chini, A.; Monte, I.; Zamarreño, A.M.; Hamberg, M.; Lassueur, S.; Reymond, P.; Weiss, S.; Stintzi, A.; Schaller, A.; Porzel, A.; et al. An OPR3-independent pathway uses 4,5-didehydrojasmonate for jasmonate synthesis. *Nat. Chem. Biol.* **2018**, *14*, 171–178. [\[CrossRef\]](#) [\[PubMed\]](#)
70. Xie, Z.; Nolan, T.M.; Jiang, H.; Yin, Y. AP2/ERF transcription factor regulatory networks in hormone and abiotic stress responses in *Arabidopsis*. *Front. Plant Sci.* **2019**, *10*, 228. [\[CrossRef\]](#) [\[PubMed\]](#)
71. Riyazuddin, R.; Verma, R.; Singh, K.; Nisha, N.; Keisham, M.; Bhati, K.K.; Kim, S.T.; Gupta, R. Ethylene: A Master Regulator of Salinity Stress Tolerance in Plants. *Biomolecules* **2020**, *10*, 959. [\[CrossRef\]](#)
72. Voesenek, L.A.C.J.; Sasidharan, R.; Visser, E.J.; Bailey-Serres, J. Flooding stress signaling through perturbations in oxygen, ethylene, nitric oxide and light. *New Phytol.* **2015**, *209*, 39–43. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Yang, Y.; Ou, B.; Zhang, J.; Si, W.; Gu, H.; Qin, G.; Qu, L. The *Arabidopsis* Mediator subunit MED16 regulates iron homeostasis by associating with EIN3/EIL1 through subunit MED25. *Plant J.* **2014**, *77*, 838–851. [\[CrossRef\]](#)

74. Huang, J.; Zhao, X.; Bürger, M.; Wang, Y.; Chory, J. Two interacting ethylene response factors regulate heat stress response. *Plant Cell* **2020**, *33*, 338–357. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Cheng, M.-C.; Liao, P.-M.; Kuo, W.-W.; Lin, T.-P. The Arabidopsis ETHYLENE RESPONSE FACTOR1 Regulates Abiotic Stress-Responsive Gene Expression by Binding to Different cis-Acting Elements in Response to Different Stress Signals. *Plant Physiol.* **2013**, *162*, 1566–1582. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Chini, A.; Gimenez-Ibanez, S.; Goossens, A.; Solano, R. Redundancy and specificity in jasmonate signalling. *Curr. Opin. Plant Biol.* **2016**, *33*, 147–156. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Earley, K.; Lawrence, R.J.; Pontes, O.; Reuther, R.; Enciso, A.J.; Silva, M.; Neves, N.; Gross, M.; Viegas, W.; Pikaard, C.S. Erasure of histone acetylation by Arabidopsis I6 mediates large-scale gene silencing in nucleolar dominance. *Genes Dev.* **2006**, *20*, 1283–1293. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Zhu, Z.; An, F.; Feng, Y.; Li, P.; Xue, L.A.M.; Jiang, Z.; Kim, J.M.; To, T.K.; Li, W.; Zhang, X.; et al. Derepression of ethylene-stabilized transcription factors (EIN3/EIL1) mediates jasmonate and ethylene signaling synergy in Arabidopsis. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 12539–12544. [\[CrossRef\]](#)
79. Zhu, Z.; Lee, B. Friends or Foes: New Insights in Jasmonate and Ethylene Co-Actions. *Plant Cell Physiol.* **2014**, *56*, 414–420. [\[CrossRef\]](#)
80. Cui, M.; Lin, Y.; Zu, Y.; Efferth, T.; Li, D.; Tang, Z. Ethylene increases accumulation of compatible solutes and decreases oxidative stress to improve plant tolerance to water stress in Arabidopsis. *J. Plant Biol.* **2015**, *58*, 193–201. [\[CrossRef\]](#)
81. Kim, J.; To, T.K.; Matsui, A.; Tanoi, K.; Kobayashi, N.I.; Matsuda, F.; Habu, Y.; Ogawa, D.; Sakamoto, T.; Matsunaga, S. Acetate-mediated novel survival strategy against drought in plants. *Nat. Plants* **2017**, *3*, 17097. [\[CrossRef\]](#)
82. Fu, J.; Wu, H.; Ma, S.; Xiang, D.; Liu, R.; Xiong, L. OsJAZ1 Attenuates Drought Resistance by Regulating JA and ABA Signaling in Rice. *Front. Plant Sci.* **2017**, *8*, 2108. [\[CrossRef\]](#)
83. Jin, J.; Duan, J.; Shan, C.; Mei, Z.; Chen, H.; Feng, H.; Zhu, J.; Cai, W. Ethylene insensitive3-like2 (OsEIL2) confers stress sensitivity by regulating OsBURP16, the β subunit of polygalacturonase (PG1 β -like) subfamily gene in rice. *Plant Sci.* **2019**, *292*, 110353. [\[CrossRef\]](#)
84. Riemann, M.; Dhakarey, R.; Hazman, M.; Miro, B.; Kohli, A.; Nick, P. Exploring jasmonates in the hormonal network of drought and salinity responses. *Front. Plant Sci.* **2015**, *6*, 1077. [\[CrossRef\]](#)
85. Zhu, C.; Gan, L.; Shen, Z.; Xia, K. Interactions between jasmonates and ethylene in the regulation of root hair development in Arabidopsis. *J. Exp. Bot.* **2006**, *57*, 1299–1308. [\[CrossRef\]](#)
86. Zhang, X.; Zhu, Z.; An, F.; Hao, D.; Li, P.; Song, J.; Yi, C.; Guo, H. Jasmonate-Activated MYC2 Represses ETHYLENE INSENSITIVE3 Activity to Antagonize Ethylene-Promoted Apical Hook Formation in Arabidopsis. *Plant Cell* **2014**, *26*, 1105–1117. [\[CrossRef\]](#)
87. Song, S.; Huang, H.; Gao, H.; Wang, J.; Wu, D.; Liu, X.; Yang, S.; Zhai, Q.; Li, C.; Qi, T.; et al. Interaction between MYC2 and ETHYLENE INSENSITIVE3 Modulates Antagonism between Jasmonate and Ethylene Signaling in Arabidopsis. *Plant Cell* **2014**, *26*, 263–279. [\[CrossRef\]](#) [\[PubMed\]](#)
88. Van Moerkercke, A.; Duncan, O.; Zander, M.; Simura, J.; Broda, M.; Vanden Bossche, R.; Lewsey, M.G.; Lama, S.; Singh, K.B.; Jung, K.; et al. A MYC2/MYC3/MYC4-dependent transcription factor network regulates water spray-responsive gene expression and jasmonate levels. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 23345–23356. [\[CrossRef\]](#)
89. Lorenzo, O.; Piqueras, R.; Sánchez-serrano, J.J.; Solano, R. ETHYLENE RESPONSE FACTOR1 integrates signals from ethylene and jasmonate pathways in plant defense. *Plant Cell* **2003**, *15*, 165–178. [\[CrossRef\]](#)
90. Zander, M.; Lewsey, M.G.; Clark, N.M.; Yin, L.; Bartlett, A.; Guzmán, J.P.S.; Hann, E.; Langford, A.E.; Jow, B.; Wise, A.; et al. Integrated multi-omics framework of the plant response to jasmonic acid. *Nat. Plants* **2020**, *6*, 290–302. [\[CrossRef\]](#)
91. Hao, D.; Jin, L.; Wen, X.; Yu, F.; Xie, Q.; Guo, H. The RING E3 ligase SDIR1 destabilizes EBF1/EBF2 and modulates the ethylene response to ambient temperature fluctuations in Arabidopsis. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2024592118. [\[CrossRef\]](#)
92. Zhang, Y.; Yang, C.; Li, Y.; Zheng, N.; Chen, H.; Zhao, Q.; Gao, T.; Guo, H.; Xie, Q. SDIR1 Is a RING Finger E3 Ligase That Positively Regulates Stress-Responsive Absciscic Acid Signaling in Arabidopsis. *Plant Cell* **2007**, *19*, 1912–1929. [\[CrossRef\]](#) [\[PubMed\]](#)
93. Dar, N.A.; Amin, I.; Wani, W.; Wani, S.A.; Shikari, A.B.; Wani, S.H.; Masoodi, K.Z. Absciscic acid: A key regulator of abiotic stress tolerance in plants. *Plant Gene* **2017**, *11*, 106–111. [\[CrossRef\]](#)
94. Emenecker, R.J.; Strader, L.C. Auxin-Absciscic Acid Interactions in Plant Growth and Development. *Biomolecules* **2020**, *10*, 281. [\[CrossRef\]](#)
95. Sah, S.K.; Reddy, K.R.; Li, J. Absciscic acid and abiotic stress tolerance in crop plants. *Front. Plant Sci.* **2016**, *7*, 571. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Müller, M. Foes or Friends: ABA and Ethylene Interaction under Abiotic Stress. *Plants* **2021**, *10*, 448. [\[CrossRef\]](#) [\[PubMed\]](#)
97. Kumar, D.; Hazra, S.; Datta, R.; Chattopadhyay, S. Transcriptome analysis of Arabidopsis mutants suggests a crosstalk between ABA, ethylene and GSH against combined cold and osmotic stress. *Sci. Rep.* **2016**, *6*, 1–13. [\[CrossRef\]](#)
98. Marczak, M.; Ciesla, A.; Janicki, M.; Kasproicz-Maluski, A.; Kubiak, P.; Ludwikow, A. Protein Phosphatases Type 2C Group A Interact with and Regulate the Stability of ACC Synthase 7 in Arabidopsis. *Cells* **2020**, *9*, 978. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Ueda, H.; Ito, T.; Inoue, R.; Masuda, Y.; Nagashima, Y.; Kozuka, T.; Kusaba, M. Genetic Interaction Among Phytochrome, Ethylene and Absciscic Acid Signaling During Dark-Induced Senescence in Arabidopsis Thaliana. *Front. Plant Sci.* **2020**, *11*, 564. [\[CrossRef\]](#)

100. Lei, G.; Shen, M.; Li, Z.-G.; Zhang, B.; Duan, K.-X.; Wang, N.; Cao, Y.-R.; Zhang, W.-K.; Ma, B.; Ling, H.-Q.; et al. EIN2 regulates salt stress response and interacts with a MA3 domain-containing protein ECIPI1 in Arabidopsis. *Plant Cell Environ.* **2011**, *34*, 1678–1692. [\[CrossRef\]](#)
101. Cao, W.-H.; Liu, J.; He, X.-J.; Mu, R.-L.; Zhou, H.-L.; Chen, S.-Y.; Zhang, J.-S. Modulation of Ethylene Responses Affects Plant Salt-Stress Responses. *Plant Physiol.* **2006**, *143*, 707–719. [\[CrossRef\]](#)
102. Achard, P.; Chen, D.; De Grauwe, L.; Decat, J.; Schoutteten, H.; Moritz, T.; Van Der Straeten, D.; Peng, J.; Harberd, N.P. Integration of Plant Responses to Environmentally Activated Phytohormonal Signals. *Science* **2006**, *311*, 91–94. [\[CrossRef\]](#)
103. Munemasa, S.; Hauser, F.; Park, J.; Waadt, R.; Brandt, B.; Schroeder, J.I. Mechanisms of abscisic acid-mediated control of stomatal aperture. *Curr. Opin. Plant Biol.* **2015**, *28*, 154–162. [\[CrossRef\]](#)
104. Madhavan, S.; Chrominski, A.; Smith, B.N. Effect of Ethylene on Stomatal Opening in Tomato and Carnation Leaves. *Plant Cell Physiol.* **1983**, *24*. [\[CrossRef\]](#)
105. Tanaka, Y.; Sano, T.; Tamaoki, M.; Nakajima, N.; Kondo, N.; Hasezawa, S. Ethylene Inhibits Absciscic Acid-Induced Stomatal Closure in Arabidopsis. *Plant Physiol.* **2005**, *138*, 2337–2343. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Benlloch-González, M.; Arquero, O.; Fournier, J.M.; Barranco, D.; Benlloch, M. K⁺ starvation inhibits water-stress-induced stomatal closure. *J. Plant Physiol.* **2008**, *165*, 623–630. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Iqbal, N.; Nazar, R.; Syeed, S.; Masood, A.; Khan, N.A. Exogenously-sourced ethylene increases stomatal conductance, photosynthesis, and growth under optimal and deficient nitrogen fertilization in mustard. *J. Exp. Bot.* **2011**, *62*, 4955–4963. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Watkins, J.M.; Hechler, P.J.; Muday, G.K. Ethylene-Induced Flavonol Accumulation in Guard Cells Suppresses Reactive Oxygen Species and Moderates Stomatal Aperture. *Plant Physiol.* **2014**, *164*, 1707–1717. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Desikan, R.; Last, K.; Harrett-Williams, R.; Tagliavia, C.; Harter, K.; Hooley, R.; Hancock, J.T.; Neill, S.J. Ethylene-induced stomatal closure in Arabidopsis occurs via AtrbohF-mediated hydrogen peroxide synthesis. *Plant J.* **2006**, *47*, 907–916. [\[CrossRef\]](#)
110. Shi, C.; Qi, C.; Ren, H.; Huang, A.; Hei, S.; She, X. Ethylene mediates brassinosteroid-induced stomatal closure via Gα protein-activated hydrogen peroxide and nitric oxide production in Arabidopsis. *Plant J.* **2015**, *82*, 280–301. [\[CrossRef\]](#) [\[PubMed\]](#)
111. Nazareno, A.L.; Hernandez, B. A mathematical model of the interaction of abscisic acid, ethylene and methyl jasmonate on stomatal closure in plants. *PLoS ONE* **2017**, *12*, e0171065. [\[CrossRef\]](#)
112. Xu, Z.-S.; Xia, L.-Q.; Chen, M.; Cheng, X.-G.; Zhang, R.-Y.; Li, L.-C.; Zhao, Y.-X.; Lu, Y.; Ni, Z.-Y.; Liu, L.; et al. Isolation and molecular characterization of the Triticum aestivum L. ethylene-responsive factor 1 (TaERF1) that increases multiple stress tolerance. *Plant Mol. Biol.* **2007**, *65*, 719–732. [\[CrossRef\]](#)
113. Xing, L.; Di, Z.; Yang, W.; Liu, J.; Li, M.; Wang, X.; Cui, C.; Wang, X.; Zhang, R.; Xiao, J. Overexpression of ERF1-V from *Haynaldia villosa* can enhance the resistance of wheat to powdery mildew and increase the tolerance to salt and drought stresses. *Front. Plant Sci.* **2017**, *8*. [\[CrossRef\]](#)
114. Wu, L.; Chen, X.; Ren, H.; Zhang, Z.; Zhang, H.; Wang, J.; Wang, X.-C.; Huang, R. ERF protein JERF1 that transcriptionally modulates the expression of abscisic acid biosynthesis-related gene enhances the tolerance under salinity and cold in tobacco. *Planta* **2007**, *226*, 815–825. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Quan, R.; Hu, S.; Zhang, Z.; Zhang, H.; Zhang, Z.; Huang, R. Overexpression of an ERF transcription factorTSRF1improves rice drought tolerance. *Plant Biotechnol. J.* **2010**, *8*, 476–488. [\[CrossRef\]](#)
116. Dong, Z.; Yu, Y.; Li, S.; Wang, J.; Tang, S.; Huang, R. Absciscic Acid Antagonizes Ethylene Production through the ABI4-Mediated Transcriptional Repression of ACS4 and ACS8 in Arabidopsis. *Mol. Plant* **2015**, *9*, 126–135. [\[CrossRef\]](#)
117. Agurla, S.; Gahir, S.; Munemasa, S.; Murata, Y.; Raghavendra, A.S. Mechanism of stomatal closure in plants exposed to drought and cold stress. In *Survival Strategies in Extreme Cold and Desiccation*; Springer: Singapore, 2018; pp. 215–232.
118. Khan, M.I.R.; Trivellini, A.; Chhillar, H.; Chopra, P.; Ferrante, A.; Khan, N.A.; Ismail, A.M. The significance and functions of ethylene in flooding stress tolerance in plants. *Environ. Exp. Bot.* **2020**, *179*, 104188. [\[CrossRef\]](#)
119. Benschop, J.J.; Jackson, M.B.; Gühl, K.; Vreeburg, R.A.M.; Croker, S.J.; Peeters, A.J.M.; Voesenek, L.A.C.J. Contrasting interactions between ethylene and abscisic acid in Rumex species differing in submergence tolerance. *Plant J.* **2005**, *44*, 756–768. [\[CrossRef\]](#) [\[PubMed\]](#)
120. Benschop, J.J.; Bou, J.; Peeters, A.J.; Wagemaker, N.; Gühl, K.; Ward, D.; Hedden, P.; Moritz, T.; Voesenek, L.A. Long-Term Submergence-Induced Elongation in Rumex palustris Requires Absciscic Acid-Dependent Biosynthesis of Gibberellin1. *Plant Physiol.* **2006**, *141*, 1644–1652. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Saika, H.; Okamoto, M.; Miyoshi, K.; Kushiro, T.; Shinoda, S.; Jikumaru, Y.; Fujimoto, M.; Arikawa, T.; Takahashi, H.; Ando, M.; et al. Ethylene Promotes Submergence-Induced Expression of OsABA8ox1, a Gene that Encodes ABA 8'-Hydroxylase in Rice. *Plant Cell Physiol.* **2006**, *48*, 287–298. [\[CrossRef\]](#)
122. Arroyo, A.; Bossi, F.; Finkelstein, R.R.; León, P. Three Genes That Affect Sugar Sensing (Absciscic Acid Insensitive 4, Absciscic Acid Insensitive 5, and Constitutive Triple Response 1) Are Differentially Regulated by Glucose in Arabidopsis. *Plant Physiol.* **2003**, *133*, 231–242. [\[CrossRef\]](#)
123. Benschop, J.J.; Millenaar, F.F.; Smeets, M.E.; Van Zanten, M.; Voesenek, L.; Peeters, A.J. Absciscic Acid Antagonizes Ethylene-Induced Hyponastic Growth in Arabidopsis. *Plant Physiol.* **2006**, *143*, 1013–1023. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Ortega-Martínez, O.; Pernas, M.; Carol, R.J.; Dolan, L. Ethylene Modulates Stem Cell Division in the Arabidopsis thaliana Root. *Science* **2007**, *317*, 507–510. [\[CrossRef\]](#)

125. Linkies, A.; Müller, K.; Morris, K.; Turečková, V.; Wenk, M.; Cadman, C.S.; Corbineau, F.; Strnad, M.; Lynn, J.R.; Finch-Savage, W.E.; et al. Ethylene Interacts with Absciscic Acid to Regulate Endosperm Rupture during Germination: A Comparative Approach Using *Lepidium sativum* and *Arabidopsis thaliana*. *Plant Cell* **2009**, *21*, 3803–3822. [\[CrossRef\]](#)
126. Zhang, H.; Han, W.; De Smet, I.; Talboys, P.; Loya, R.; Hassan, A.; Rong, H.; Jürgens, G.; Knox, J.P.; Wang, M.-H. ABA promotes quiescence of the quiescent centre and suppresses stem cell differentiation in the *Arabidopsis* primary root meristem. *Plant J.* **2010**, *64*, 764–774. [\[CrossRef\]](#) [\[PubMed\]](#)
127. Jung, J.K.H.M.; McCouch, S.R.M. Getting to the roots of it: Genetic and hormonal control of root architecture. *Front. Plant Sci.* **2013**, *4*, 186. [\[CrossRef\]](#)
128. Chang, K.N.; Zhong, S.; Weirauch, M.T.; Hon, G.; Pelizzola, M.; Li, H.; Huang, S.-S.C.; Schmitz, R.; Urich, M.A.; Kuo, D.; et al. Temporal transcriptional response to ethylene gas drives growth hormone cross-regulation in *Arabidopsis*. *eLife* **2013**, *2*, e00675. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Song, L.; Huang, S.-S.C.; Wise, A.; Castanon, R.; Nery, J.R.; Chen, H.; Watanabe, M.; Thomas, J.; Bar-Joseph, Z.; Ecker, J.R. A transcription factor hierarchy defines an environmental stress response network. *Science* **2016**, *354*, aag1550. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Planas-Riverola, A.; Gupta, A.; Betegón-Putze, I.; Bosch, N.; Ibañes, M.; Caño-Delgado, A.I. Brassinosteroid signaling in plant development and adaptation to stress. *Development* **2019**, *146*, dev151894. [\[CrossRef\]](#)
131. Nolan, T.M.; Vukašinović, N.; Liu, D.; Russinova, E.; Yin, Y. Brassinosteroids: Multidimensional Regulators of Plant Growth, Development, and Stress Responses. *Plant Cell* **2019**, *32*, 295–318. [\[CrossRef\]](#)
132. Jiroutova, P.; Oklestkova, J.; Strnad, M. Crosstalk between Brassinosteroids and Ethylene during Plant Growth and under Abiotic Stress Conditions. *Int. J. Mol. Sci.* **2018**, *19*, 3283. [\[CrossRef\]](#) [\[PubMed\]](#)
133. Wang, B.; Zhang, J.; Xia, X.; Zhang, W.-H. Ameliorative effect of brassinosteroid and ethylene on germination of cucumber seeds in the presence of sodium chloride. *Plant Growth Regul.* **2011**, *65*, 407–413. [\[CrossRef\]](#)
134. Zhu, T.; Deng, X.; Zhou, X.; Zhu, L.; Zou, L.; Li, P.; Zhang, D.; Lin, H. Ethylene and hydrogen peroxide are involved in brassinosteroid-induced salt tolerance in tomato. *Sci. Rep.* **2016**, *6*, 35392. [\[CrossRef\]](#)
135. Wei, L.-J.; Deng, X.-G.; Zhu, T.; Zheng, T.; Li, P.-X.; Wu, J.-Q.; Zhang, D.-W.; Lin, H.-H. Ethylene is Involved in Brassinosteroids Induced Alternative Respiratory Pathway in Cucumber (*Cucumis sativus* L.) Seedlings Response to Abiotic Stress. *Front. Plant Sci.* **2015**, *6*, 982. [\[CrossRef\]](#)
136. Schmitz, A.; Folsom, J.J.; Jikamaru, Y.; Ronald, P.; Walia, H. SUB 1 A-mediated submergence tolerance response in rice involves differential regulation of the brassinosteroid pathway. *New Phytol.* **2013**, *198*, 1060–1070. [\[CrossRef\]](#) [\[PubMed\]](#)
137. Lavy, M.; Estelle, M. Mechanisms of auxin signaling. *Development* **2016**, *143*, 3226–3229. [\[CrossRef\]](#)
138. Zhao, Y. Auxin biosynthesis. *Arab. Book* **2014**, *12*, e0173. [\[CrossRef\]](#)
139. Enders, T.A.; Strader, L.C. Auxin activity: Past, present, and future. *Am. J. Bot.* **2015**, *102*, 180–196. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Muday, G.K.; Rahman, A.; Binder, B.M. Auxin and ethylene: Collaborators or competitors? *Trends Plant Sci.* **2012**, *17*, 181–195. [\[CrossRef\]](#) [\[PubMed\]](#)
141. Stepanova, A.N.; Alonso, J.M. From Ethylene-Auxin Interactions to Auxin Biosynthesis and Signal Integration. *Plant Cell* **2019**, *31*, 1393–1394. [\[CrossRef\]](#) [\[PubMed\]](#)
142. Mazzoni-Putman, S.M.; Brumos, J.; Zhao, C.; Alonso, J.M.; Stepanova, A.N. Auxin Interactions with Other Hormones in Plant Development. *Cold Spring Harb. Perspect. Biol.* **2021**, *13*, a039990. [\[CrossRef\]](#)
143. Koevoets, I.T.; Venema, J.H.; Elzenga, J.T.M.; Testerink, C. Roots Withstanding their Environment: Exploiting Root System Architecture Responses to Abiotic Stress to Improve Crop Tolerance. *Front. Plant Sci.* **2016**, *7*, 1335. [\[CrossRef\]](#)
144. Bojórquez-Quintal, E.; Escalante-Magaña, C.; Echevarría-Machado, I.; Martínez-Estévez, M. Aluminum, a Friend or Foe of Higher Plants in Acid Soils. *Front. Plant Sci.* **2017**, *8*, 1767. [\[CrossRef\]](#)
145. Shen, H.; Hou, N.; Schlicht, M.; Wan, Y.; Mancuso, S.; Baluska, F. Aluminium toxicity targets PIN2 in *Arabidopsis* root apices: Effects on PIN2 endocytosis, vesicular recycling, and polar auxin transport. *Sci. Bull.* **2008**, *53*, 2480–2487. [\[CrossRef\]](#)
146. Yang, Z.B.; Geng, X.Y.; He, C.M.; Zhang, F.; Wang, R.; Horst, W.J.; Ding, Z.J. TAA1-Regulated Local Auxin Biosynthesis in the Root-Apex Transition Zone Mediates the Aluminum-Induced Inhibition of Root Growth in *Arabidopsis*. *Plant Cell* **2014**, *26*, 2889–2904. [\[CrossRef\]](#) [\[PubMed\]](#)
147. Liu, G.; Gao, S.; Tian, H.; Wu, W.; Robert, H.S.; Ding, Z. Local Transcriptional Control of YUCCA Regulates Auxin Promoted Root-Growth Inhibition in Response to Aluminium Stress in *Arabidopsis*. *PLoS Genet.* **2016**, *12*, e1006360. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Yang, Z.; Liu, G.; Liu, J.; Zhang, B.; Meng, W.; Müller, B.; Hayashi, K.; Zhang, X.; Zhao, Z.; De Smet, I.; et al. Synergistic action of auxin and cytokinin mediates aluminum—Induced root growth inhibition in *Arabidopsis*. *EMBO Rep.* **2017**, *18*, 1213–1230. [\[CrossRef\]](#)
149. Wakeel, A.; Ali, I.; Upreti, S.; Azizullah, A.; Liu, B.; Khan, A.R.; Huang, L.; Wu, M.; Gan, Y. Ethylene mediates dichromate-induced inhibition of primary root growth by altering AUX1 expression and auxin accumulation in *Arabidopsis thaliana*. *Plant Cell Environ.* **2018**, *41*, 1453–1467. [\[CrossRef\]](#)
150. Wakeel, A.; Gan, Y. A model for the ethylene-mediated auxin distribution under Cr(VI) stress in *Arabidopsis thaliana*. *Plant Signal. Behav.* **2018**, *13*, e1473685. [\[CrossRef\]](#)
151. Li, G.; Song, H.; Li, B.; Kronzucker, H.J.; Shi, W. Auxin resistant1 and PIN-FORMED2 protect lateral root formation in *Arabidopsis* under iron stress. *Plant Physiol.* **2015**, *169*, 2608–2623.

152. Li, J.; Xu, H.-H.; Liu, W.-C.; Zhang, X.-W.; Lu, Y.-T. Ethylene Inhibits Root Elongation during Alkaline Stress through AUXIN1 and Associated Changes in Auxin Accumulation. *Plant Physiol.* **2015**, *168*, 1777–1791. [[CrossRef](#)] [[PubMed](#)]
153. Grato, P.; Monteiro, C.C.; Carvalho, R.F.; Tezotto, T.; Piotto, F.; Peres, L.; Azevedo, R.A. Biochemical dissection of diageotropica and Never ripe tomato mutants to Cd-stressful conditions. *Plant Physiol. Biochem.* **2012**, *56*, 79–96. [[CrossRef](#)]
154. Gruber, B.D.; Giehl, R.F.; Friedel, S.; von Wirén, N. Plasticity of the Arabidopsis root system under nutrient deficiencies. *Plant Physiol.* **2013**, *163*, 161–179. [[CrossRef](#)]
155. Liu, M.; Zhang, H.; Fang, X.; Zhang, Y.; Jin, C. Auxin acts downstream of ethylene and nitric oxide to regulate magnesium deficiency-induced root hair development in *Arabidopsis thaliana*. *Plant Cell Physiol.* **2018**, *59*, 1452–1465. [[CrossRef](#)]
156. Martín-Rejano, E.M.; Camacho-Cristóbal, J.J.; Herrera-Rodríguez, M.B.; Rexach, J.; Navarro-Gochicoa, M.T.; Gonzalez-Fontes, A. Auxin and ethylene are involved in the responses of root system architecture to low boron supply in Arabidopsis seedlings. *Physiol. Plant.* **2011**, *142*, 170–178. [[CrossRef](#)] [[PubMed](#)]
157. Camacho-Cristóbal, J.J.; Martín-Rejano, E.M.; Herrera-Rodríguez, M.B.; Navarro-Gochicoa, M.T.; Rexach, J.; González-Fontes, A. Boron deficiency inhibits root cell elongation via an ethylene/auxin/ROS-dependent pathway in Arabidopsis seedlings. *J. Exp. Bot.* **2015**, *66*, 3831–3840. [[CrossRef](#)] [[PubMed](#)]
158. Jacobsen, A.G.R.; Jervis, G.; Xu, J.; Topping, J.F.; Lindsey, K. Root growth responses to mechanical impedance are regulated by a network of ROS, ethylene and auxin signalling in Arabidopsis. *New Phytol.* **2021**, *231*, 225–242. [[CrossRef](#)] [[PubMed](#)]
159. Sasidharan, R.; Voesenek, L.A.C.J. Ethylene-Mediated Acclimations to Flooding Stress. *Plant Physiol.* **2015**, *169*, 3–12. [[CrossRef](#)] [[PubMed](#)]
160. Visser, E.; Cohen, J.; Barendse, G.; Blom, C.; Voesenek, L. An Ethylene-Mediated Increase in Sensitivity to Auxin Induces Adventitious Root Formation in Flooded *Rumex palustris* Sm. *Plant Physiol.* **1996**, *112*, 1687–1692. [[CrossRef](#)] [[PubMed](#)]
161. Vidoz, M.L.; Loreti, E.; Mensuali, A.; Alpi, A.; Perata, P. Hormonal interplay during adventitious root formation in flooded tomato plants. *Plant J.* **2010**, *63*, 551–562. [[CrossRef](#)] [[PubMed](#)]
162. Qi, X.; Li, Q.; Ma, X.; Qian, C.; Wang, H.; Ren, N.; Shen, C.; Huang, S.; Xu, X.; Xu, Q.; et al. Waterlogging—Induced adventitious root formation in cucumber is regulated by ethylene and auxin through reactive oxygen species signalling. *Plant Cell Environ.* **2018**, *42*, 1458–1470. [[CrossRef](#)]
163. Yamauchi, T.; Shimamura, S.; Nakazono, M.; Mochizuki, T. Aerenchyma formation in crop species: A review. *Field Crop. Res.* **2013**, *152*, 8–16. [[CrossRef](#)]
164. Yamauchi, T.; Tanaka, A.; Tsutsumi, N.; Inukai, Y.; Nakazono, M. A Role for Auxin in Ethylene-Dependent Inducible Aerenchyma Formation in Rice Roots. *Plants* **2020**, *9*, 610. [[CrossRef](#)]
165. Zhong, S.; Shi, H.; Xue, C.; Wang, L.; Xi, Y.; Li, J.; Quail, P.H.; Deng, X.W.; Guo, H. A Molecular Framework of Light-Controlled Phytohormone Action in Arabidopsis. *Curr. Biol.* **2012**, *22*, 1530–1535. [[CrossRef](#)]
166. Kim, J.Y.; Park, Y.-J.; Lee, J.-H.; Kim, Z.H.; Park, C.-M. EIN3-Mediated Ethylene Signaling Attenuates Auxin Response during Hypocotyl Thermomorphogenesis. *Plant Cell Physiol.* **2021**, *62*, 708–720. [[CrossRef](#)]
167. Fei, Q.; Wei, S.; Zhou, Z.; Gao, H.; Li, X. Adaptation of root growth to increased ambient temperature requires auxin and ethylene coordination in Arabidopsis. *Plant Cell Rep.* **2017**, *36*, 1507–1518. [[CrossRef](#)] [[PubMed](#)]
168. Binenbaum, J.; Weinstain, R.; Shani, E. Gibberellin Localization and Transport in Plants. *Trends Plant Sci.* **2018**, *23*, 410–421. [[CrossRef](#)] [[PubMed](#)]
169. Hernández-García, J.; Briones-Moreno, A.; Blázquez, M.A. Origin and evolution of gibberellin signaling and metabolism in plants. *Semin. Cell Dev. Biol.* **2020**, *109*, 46–54. [[CrossRef](#)] [[PubMed](#)]
170. Hattori, Y.; Nagai, K.; Furukawa, S.; Song, X.-J.; Kawano, R.; Sakakibara, H.; Wu, J.; Matsumoto, T.; Yoshimura, A.; Kitano, H.; et al. The ethylene response factors SNORKEL1 and SNORKEL2 allow rice to adapt to deep water. *Nature* **2009**, *460*, 1026–1030. [[CrossRef](#)] [[PubMed](#)]
171. Bailey-Serres, J.; Fukao, T.; Gibbs, D.; Holdsworth, M.; Lee, S.C.; Licausi, F.; Perata, P.; Voesenek, L.; van Dongen, J. Making sense of low oxygen sensing. *Trends Plant Sci.* **2012**, *17*, 129–138. [[CrossRef](#)]
172. Kuroha, T.; Nagai, K.; Gamuyao, R.; Wang, D.R.; Furuta, T.; Nakamori, M.; Kitaoka, T.; Adachi, K.; Minami, A.; Mori, Y.; et al. Ethylene-gibberellin signaling underlies adaptation of rice to periodic flooding. *Science* **2018**, *361*, 181–186. [[CrossRef](#)]
173. Fukao, T.; Xu, K.; Ronald, P.C.; Bailey-Serres, J. A Variable Cluster of Ethylene Response Factor-Like Genes Regulates Metabolic and Developmental Acclimation Responses to Submergence in Rice. *Plant Cell* **2006**, *18*, 2021–2034. [[CrossRef](#)]
174. Xu, K.; Xu, X.; Fukao, T.; Canlas, P.; Maghirang-Rodriguez, R.; Heuer, S.; Ismail, A.M.; Bailey-Serres, J.; Ronald, P.C.; Mackill, D.J. Sub1A is an ethylene-response-factor-like gene that confers submergence tolerance to rice. *Nat. Cell Biol.* **2006**, *442*, 705–708. [[CrossRef](#)]
175. Fukao, T.; Bailey-Serres, J. Submergence tolerance conferred by Sub1A is mediated by SLR1 and SLRL1 restriction of gibberellin responses in rice. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 16814–16819. [[CrossRef](#)]
176. Lin, C.-C.; Chao, Y.-T.; Chen, W.-C.; Ho, H.-Y.; Chou, M.-Y.; Li, Y.-R.; Wu, Y.-L.; Yang, H.-A.; Hsieh, H.; Lin, C.-S.; et al. Regulatory cascade involving transcriptional and N-end rule pathways in rice under submergence. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 3300–3309. [[CrossRef](#)]
177. Dubois, M.; Skirycz, A.; Claeys, H.; Maleux, K.; Dhondt, S.; De Bodt, S.; Bossche, R.V.; De Milde, L.; Yoshizumi, T.; Matsui, M.; et al. Ethylene response factor6 Acts as a Central Regulator of Leaf Growth under Water-Limiting Conditions in Arabidopsis. *Plant Physiol.* **2013**, *162*, 319–332. [[CrossRef](#)] [[PubMed](#)]

178. Dubois, M.; Broeck, L.V.D.; Claeys, H.; Van Vlierberghe, K.; Matsui, M.; Inzé, D. The Ethylene Response factors ERF6 and ERF11 Antagonistically Regulate Mannitol-Induced Growth Inhibition in Arabidopsis. *Plant Physiol.* **2015**, *169*, 166–179. [[CrossRef](#)] [[PubMed](#)]
179. Zhou, X.; Zhang, Z.-L.; Park, J.; Tyler, L.; Yusuke, J.; Qiu, K.; Nam, E.A.; Lumba, S.; Desveaux, D.; McCourt, P.; et al. The ERF11 Transcription Factor Promotes Internode Elongation by Activating Gibberellin Biosynthesis and Signaling. *Plant Physiol.* **2016**, *171*, 2760–2770. [[CrossRef](#)] [[PubMed](#)]
180. Broeck, L.V.D.; Dubois, M.; Vermeersch, M.; Storme, V.; Matsui, M.; Inzé, D. From network to phenotype: The dynamic wiring of an Arabidopsis transcriptional network induced by osmotic stress. *Mol. Syst. Biol.* **2017**, *13*, 961. [[CrossRef](#)]
181. An, F.; Zhang, X.; Zhu, Z.; Ji, Y.; He, W.; Jiang, Z.; Li, M.; Guo, H. Coordinated regulation of apical hook development by gibberellins and ethylene in etiolated Arabidopsis seedlings. *Cell Res.* **2012**, *22*, 915–927. [[CrossRef](#)]
182. Lefevere, H.; Bauters, L.; Gheysen, G. Salicylic Acid Biosynthesis in Plants. *Front. Plant Sci.* **2020**, *11*, 338. [[CrossRef](#)]
183. Fu, Z.Q.; Yan, S.; Saleh, A.; Wang, W.; Ruble, J.; Oka, N.; Mohan, R.; Spoel, S.; Tada, Y.; Zheng, N.; et al. NPR3 and NPR4 are receptors for the immune signal salicylic acid in plants. *Nature* **2012**, *486*, 228–232. [[CrossRef](#)]
184. Iqbal, N.; Masood, A.; Khan, N.A. Phytohormones in salinity tolerance: Ethylene and gibberellins cross talk. In *Phytohormones and Abiotic Stress Tolerance in Plants*; Springer: Berlin/Heidelberg, Germany, 2012; pp. 77–98.
185. Khan, M.I.R.; Fatma, M.; Per, T.S.; Anjum, N.A.; Khan, N.A. Salicylic acid-induced abiotic stress tolerance and underlying mechanisms in plants. *Front. Plant Sci.* **2015**, *6*, 462. [[CrossRef](#)]
186. Peng, Y.; Yang, J.; Li, X.; Zhang, Y. Salicylic Acid: Biosynthesis and Signaling. *Annu. Rev. Plant Biol.* **2021**, *72*, 761–791. [[CrossRef](#)]
187. Khan, M.I.R.; Iqbal, N.; Masood, A.; Per, T.S.; Khan, N.A. Salicylic acid alleviates adverse effects of heat stress on photosynthesis through changes in proline production and ethylene formation. *Plant Signal. Behav.* **2013**, *8*, e26374. [[CrossRef](#)] [[PubMed](#)]
188. Khan, M.I.R.; Asgher, M.; Khan, N.A. Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (*Vigna radiata* L.). *Plant Physiol. Biochem.* **2014**, *80*, 67–74. [[CrossRef](#)] [[PubMed](#)]
189. Ahmed, W.; Imran, M.; Yaseen, M.; Haq, T.U.; Jamshaid, M.U.; Rukh, S.; Ikram, R.M.; Ali, M.; Ali, A.; Maqbool, M.; et al. Role of salicylic acid in regulating ethylene and physiological characteristics for alleviating salinity stress on germination, growth and yield of sweet pepper. *PeerJ* **2020**, *8*, e8475. [[CrossRef](#)] [[PubMed](#)]
190. Nazar, R.; Umar, S.; Khan, N.; Sareer, O. Salicylic acid supplementation improves photosynthesis and growth in mustard through changes in proline accumulation and ethylene formation under drought stress. *S. Afr. J. Bot.* **2015**, *98*, 84–94. [[CrossRef](#)]
191. Leslie, C.A.; Romani, R.J. Salicylic acid: A new inhibitor of ethylene biosynthesis. *Plant Cell Rep.* **1986**, *5*, 144–146. [[CrossRef](#)]
192. Rao, M.V.; Lee, H.-I.; Davis, K.R. Ozone-induced ethylene production is dependent on salicylic acid, and both salicylic acid and ethylene act in concert to regulate ozone-induced cell death. *Plant J.* **2002**, *32*, 447–456. [[CrossRef](#)]
193. Ogawa, D.; Nakajima, N.; Sano, T.; Tamaoki, M.; Aono, M.; Kubo, A.; Kanna, M.; Ioki, M.; Kamada, H.; Saji, H. Salicylic Acid Accumulation Under O₃ Exposure is Regulated by Ethylene in Tobacco Plants. *Plant Cell Physiol.* **2005**, *46*, 1062–1072. [[CrossRef](#)]
194. Thaler, J.S.; Humphrey, P.T.; Whiteman, N.K. Evolution of jasmonate and salicylate signal crosstalk. *Trends Plant Sci.* **2012**, *17*, 260–270. [[CrossRef](#)]
195. Kieber, J.J.; Schaller, G.E. Cytokinin signaling in plant development. *Development* **2018**, *145*, dev149344. [[CrossRef](#)]
196. Hutchison, C.E.; Li, J.; Argueso, C.; Gonzalez, M.; Lee, E.; Lewis, M.W.; Maxwell, B.B.; Perdue, T.D.; Schaller, G.E.; Alonso, J.M.; et al. The Arabidopsis Histidine Phosphotransfer Proteins Are Redundant Positive Regulators of Cytokinin Signaling. *Plant Cell* **2006**, *18*, 3073–3087. [[CrossRef](#)]
197. Zwack, P.J.; Rashotte, A.M. Interactions between cytokinin signalling and abiotic stress responses. *J. Exp. Bot.* **2015**, *66*, 4863–4871. [[CrossRef](#)] [[PubMed](#)]
198. Vogel, J.; Woeste, K.E.; Theologis, A.; Kieber, J.J. Recessive and dominant mutations in the ethylene biosynthetic gene ACS5 of Arabidopsis confer cytokinin insensitivity and ethylene overproduction, respectively. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 4766–4771. [[CrossRef](#)] [[PubMed](#)]
199. Chae, H.S.; Faure, F.; Kieber, J.J. The eto1, eto2, and eto3 Mutations and Cytokinin Treatment Increase Ethylene Biosynthesis in Arabidopsis by Increasing the Stability of ACS Protein. *Plant Cell* **2003**, *15*, 545–559. [[CrossRef](#)] [[PubMed](#)]
200. Hansen, M.; Chae, H.S.; Kieber, J.J. Regulation of ACS protein stability by cytokinin and brassinosteroid. *Plant J.* **2009**, *57*, 606–614. [[CrossRef](#)] [[PubMed](#)]
201. Jalmi, S.K.; Bhagat, P.K.; Verma, D.; Noryang, S.; Tayyeba, S.; Singh, K.; Sharma, D.; Sinha, A.K. Traversing the Links between Heavy Metal Stress and Plant Signaling. *Front. Plant Sci.* **2018**, *9*, 12. [[CrossRef](#)] [[PubMed](#)]
202. Artner, C.; Benkova, E. Ethylene and Cytokinin: Partners in Root Growth Regulation. *Mol. Plant* **2019**, *12*, 1312–1314. [[CrossRef](#)]
203. Skalak, J.; Nicolas, K.L.; Vankova, R.; Hejatko, J. Signal Integration in Plant Abiotic Stress Responses via Multistep Phosphorelay Signaling. *Front. Plant Sci.* **2021**, *12*. [[CrossRef](#)]
204. Yamauchi, T.; Yoshioka, M.; Fukazawa, A.; Mori, H.; Nishizawa, N.K.; Tsutsumi, N.; Yoshioka, H.; Nakazono, M. An NADPH Oxidase RBOH Functions in Rice Roots during Lysigenous Aerenchyma Formation under Oxygen-Deficient Conditions. *Plant Cell* **2017**, *29*, 775–790. [[CrossRef](#)]
205. Yamauchi, T.; Tanaka, A.; Inahashi, H.; Nishizawa, N.K.; Tsutsumi, N.; Inukai, Y.; Nakazono, M. Fine control of aerenchyma and lateral root development through AUX/IAA- and ARF-dependent auxin signaling. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 20770–20775. [[CrossRef](#)]

206. Tavares, E.Q.P.; Grandis, A.; Lembke, C.G.; Souza, G.M.; Purgatto, E.; De Souza, A.P.; Buckeridge, M.S. Roles of auxin and ethylene in aerenchyma formation in sugarcane roots. *Plant Signal. Behav.* **2018**, *13*, e1422464. [[CrossRef](#)]
207. Ponce, G.; Barlow, P.W.; Feldman, L.J.; Cassab, G.I. Auxin and ethylene interactions control mitotic activity of the quiescent centre, root cap size, and pattern of cap cell differentiation in maize. *Plant Cell Environ.* **2005**, *28*, 719–732. [[CrossRef](#)]
208. Habben, J.E.; Bao, X.; Bate, N.J.; DeBruin, J.L.; Dolan, D.; Hasegawa, D.; Helentjaris, T.G.; Lafitte, R.H.; Lovan, N.; Mo, H.; et al. Transgenic alteration of ethylene biosynthesis increases grain yield in maize under field drought-stress conditions. *Plant Biotechnol. J.* **2014**, *12*, 685–693. [[CrossRef](#)]
209. Li, Z.; Zhang, X.; Zhao, Y.; Li, Y.; Zhang, G.; Peng, Z.; Zhang, J. Enhancing auxin accumulation in maize root tips improves root growth and dwarfs plant height. *Plant Biotechnol. J.* **2017**, *16*, 86–99. [[CrossRef](#)] [[PubMed](#)]
210. Emerick, K.; Ronald, P.C. Sub1 Rice: Engineering Rice for Climate Change. *Cold Spring Harb. Perspect. Biol.* **2019**, *11*, a034637. [[CrossRef](#)] [[PubMed](#)]
211. Licausi, F.; Kosmacz, M.; Weits, D.A.; Giuntoli, B.; Giorgi, F.M.; Voesenek, L.A.C.J.; Perata, P.; van Dongen, J.T. Oxygen sensing in plants is mediated by an N-end rule pathway for protein destabilization. *Nature* **2011**, *479*, 419–422. [[CrossRef](#)]
212. Gasch, P.; Funderinger, M.; Müller, J.T.; Lee, T.; Bailey-Serres, J.; Mustroph, A. Redundant ERF-VII Transcription Factors Bind to an Evolutionarily Conserved cis-Motif to Regulate Hypoxia-Responsive Gene Expression in Arabidopsis. *Plant Cell* **2015**, *28*, 160–180. [[CrossRef](#)] [[PubMed](#)]
213. Perata, P. Ethylene Signaling Controls Fast Oxygen Sensing in Plants. *Trends Plant Sci.* **2019**, *25*, 3–6. [[CrossRef](#)] [[PubMed](#)]
214. Lamers, J.; van der Meer, T.; Testerink, C. How Plants Sense and Respond to Stressful Environments. *Plant Physiol.* **2020**, *182*, 1624–1635. [[CrossRef](#)] [[PubMed](#)]
215. Jung, J.-H.; Domijan, M.; Klose, C.; Biswas, S.; Ezer, D.; Gao, M.; Khattak, A.K.; Box, M.S.; Charoensawan, V.; Cortijo, S.; et al. Phytochromes function as thermosensors in Arabidopsis. *Science* **2016**, *354*, 886–889. [[CrossRef](#)]
216. Legris, M.; Klose, C.; Burgie, E.S.; Rojas, C.C.R.; Neme, M.; Hiltbrunner, A.; Wigge, P.A.; Schäfer, E.; Vierstra, R.D.; Casal, J.J. Phytochrome B integrates light and temperature signals in Arabidopsis. *Science* **2016**, *354*, 897–900. [[CrossRef](#)]
217. Yuan, F.; Yang, H.; Xue, Y.; Kong, D.; Ye, R.; Li, C.; Zhang, J.; Theprungsirikul, J.; Shrift, T.; Krichilsky, B.; et al. OSCA1 mediates osmotic-stress-evoked Ca²⁺ increases vital for osmosensing in Arabidopsis. *Nature* **2014**, *514*, 367–371. [[CrossRef](#)] [[PubMed](#)]
218. Ding, Y.; Lv, J.; Shi, Y.; Gao, J.; Hua, J.; Song, C.; Gong, Z.; Yang, S. EGR 2 phosphatase regulates OST 1 kinase activity and freezing tolerance in Arabidopsis. *EMBO J.* **2018**, *38*, e99819. [[CrossRef](#)]
219. Jiang, Z.; Zhou, X.; Tao, M.; Yuan, F.; Liu, L.; Wu, F.; Wu, X.; Xiang, Y.; Niu, Y.; Liu, F.; et al. Plant cell-surface GIPC sphingolipids sense salt to trigger Ca²⁺ influx. *Nature* **2019**, *572*, 341–346. [[CrossRef](#)] [[PubMed](#)]
220. Huot, B.; Yao, J.; Montgomery, B.L.; He, S.Y. Growth–Defense Tradeoffs in Plants: A Balancing Act to Optimize Fitness. *Mol. Plant* **2014**, *7*, 1267–1287. [[CrossRef](#)] [[PubMed](#)]