Guest-driven control of folding in a crown-ether-functionalized *ortho*-phenylene

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Abstract

A crown-ether-functionalized *o*-phenylene tetramer has been synthesized and coassembled with monotopic and ditopic, achiral and chiral secondary ammonium ion guests. NMR spectroscopy shows that the *o*-phenylene forms both 1:1 and 1:2 complexes with monotopic guests while remaining well-folded. Binding of an elongated ditopic guest, however, forces the *o*-phenylene to misfold by pulling the terminal rings apart. A chiral ditopic guest biases the *o*-phenylene twist sense.

Inspired by the hierarchical structure of biomolecules, researchers are interested in the design of synthetic foldamers: oligomers that favor well-defined conformations because of

noncovalent interactions (e.g., hydrogen bonding, arene-arene stacking, or backbone torsional biases).^{1,2} A variety of foldamers with peptidic^{3–5} or non-peptidic^{6–9} backbones have been reported. Folding in these systems can be influenced by interactions with guest molecules. For example, many helical aromatic foldamers have central cavities for guest binding.^{10–14} Binding sites can also be introduced to folded systems through attachment of macrocycles to their exteriors. Voyer^{15–17} and Chen¹⁸ have studied peptidic foldamers with crown ethers attached to their exteriors, which were successfully used as membrane ion channels.¹⁵ Similarly, Yashima,^{19,20} Kakuchi,^{21,22} and Nolte²³ have appended crown ether binding sites to the exteriors of helical polymers; in many cases chiral guests induce a preferred twist sense. Abe and Inouye have merged interior and exterior binding in polymeric systems, demonstrating cooperative behavior.²⁴



Chart 1: Crown-ether-functionalized *ortho*-phenylene tetramer \mathbf{oP}^4 and secondary amine guests 1-4.

The *o*-phenylenes are a simple class of aromatic foldamers, adopting helical geometries with three repeat units per turn.²⁵ A key feature of *o*-phenylenes is that their folding is easily perturbed because it is driven by (weak) arene–arene stacking interactions.²⁶ It should therefore be possible to switch *o*-phenylene conformation through suitably designed host–guest binding, a potentially useful behavior for dynamic foldamer systems.²⁷ *o*-Phenylenes lack an internal cavity that could bind guests, but their structure lends itself to incorporation of (benzo) crown ethers directly into their backbones. Here, we explore the effects of host–



Figure 1: Conformers of $\mathbf{oP^{4\prime}}$ optimized at the ω B97X-D/cc-pVDZ level (H atoms not shown).

guest binding on short *o*-phenylene tetramer foldamers functionalized with crown ethers using the compounds in Chart 1. We report the design and synthesis of dibenzo-24-crown-8-functionalized *o*-phenylene $\mathbf{oP^4}$, the changes in folding associated with the binding of monotopic and ditopic guests **1** and **2**, and chiral induction through the binding of chiral guests **3** and **4**.²⁸

Foldamer $\mathbf{oP^4}$ is expected to favor a helical conformation with one full turn, such that the two crown ethers will be stacked when folded but distant when misfolded. It has two possible backbone conformations that should exchange rapidly, shown in Figure 1 for $\mathbf{oP^{4\prime}}$, a simplified model with the crown ethers replaced with methoxy groups. These two conformers differ in the torsional angle of the central biaryl bond (approximately $\pm 55^{\circ}$ for the "A" state and $\pm 135^{\circ}$ for the "B" state).²⁵ The A conformer is a compact helix stabilized by one stacking interaction between the terminal rings and the B conformer is an extended helix without stabilizing aromatic stacking. Compound $\mathbf{oP^4}$ was prepared in a single step by Suzuki coupling of known 2,2'-dibromobiphenyl²⁹ and the appropriate dibenzo-24-crown-8 boronic acid³⁰ (Scheme S1).

We began host-guest studies by examining the interaction between guest 1 and $\mathbf{oP^4}$. It is well-known that 1 forms a 1:1 complex with dibenzo-24-crown-8, stabilized by arene-arene stacking interactions and H-bonding.³¹ The complexed and uncomplexed species are in slow exchange on the NMR timescale. Thus, both 1:1 and 1:2 complexes are expected for the $\mathbf{oP^4/1}$ system. ¹H NMR spectroscopy titration experiments were carried out in 3:2 CDCl₃/CD₃CN



Figure 2: ¹H NMR spectra of (a) $\mathbf{oP^4}$ and its (b) 1:1 (10 mM, 1 equiv $\mathbf{oP^4}$; 10 mM, 1 equiv **1**) and (c) 1:2 (10 mM, 1 equiv $\mathbf{oP^4}$; 30 mM, 3 equiv **1**) complexes with **1** (600 MHz, 3:2 CDCl₃/CD₃CN, 278 K).

at 278 K at a constant concentration of $\mathbf{oP^4}$ (10 mM), varying the concentration of **1** (5–40 mM) (Figure S6). Representative ¹H NMR spectra are shown in Figure 2. New signals appear with increasing **1** that were assigned to the 1:1 and 1:2 complexes. All ¹H signals for the major species were assigned by using standard 2D NMR experiments (COSY, HSQC, HMBC) (Table S1). Signals corresponding to H_{1b} (Chart 1) are particularly useful in distinguishing the different species as they are well-separated from the rest of the spectrum. Integration shows that the 1:1 and 1:2 complexes are the major species at 1:1 and 1:3 ratios of $\mathbf{oP^4/1}$, respectively, although signals from other species are always present.

The association constant for dibenzo-24-crown-8 and **1** has been previously determined to be on the order of 500 M⁻¹.³² The apparent binding constants from the NMR titration experiments (Figures S45, S46) are concentration-dependent, as expected,³² but of the same order ($K_1 \approx 500 \text{ M}^{-1}$ for the 1:1 complex and $K_2 \approx 100 \text{ M}^{-1}$ for subsequent binding in the 1:2 complex). We conclude that binding of monotopic guest **1** to host **oP**⁴ is comparable to the parent system and is non-cooperative (since K_1 is approximately $4K_2$).³³

NMR chemical shifts provide important insight into the folding state of *o*-phenylenes in solution²⁵ because geometry changes tend to move protons in and out of the shielding zones of nearby aromatic rings. In both the 1:1 and 1:2 complexes, the signals for H_{1b} , H_{1c} , and H_{1e} of **oP**⁴ are shifted upfield, which is likely due to the shielding effect of **1**. Protons on the



Figure 3: ¹H NMR spectra of (a) $\mathbf{oP^4}$ and (b) its 1:1 (2 mM, 1 equiv $\mathbf{oP^4}$; 2 mM, 1 equiv 2) complex with 2 (500 MHz, 3:2 CDCl₃/CD₃CN, 278 K) (protons labeled * originate on 2, † on the distal arenes of $\mathbf{oP^4}$). Inset: UV–vis titration monitoring absorbance at 310 nm.

internal rings $(H_{2b}-H_{2e})$ are far from the binding sites, but should be sensitive to changes in conformation. Their chemical shifts exhibit only small differences (<0.1 ppm) between the uncomplexed and complexed states. Thus, the binding of guest **1** to the crown ether functionalized tetramer in the 1:1 and 1:2 complexes does not substantially affect the folding of the *o*-phenylene.

We then conducted host-guest studies of host $\mathbf{oP^4}$ with $\mathbf{2}$, which comprises two secondary ammonium ion centers. The 1:2 complex of ditopic guest $\mathbf{2}$ with dibenzo-24-crown-8 has been reported.³¹ Since the $\mathbf{oP^4}$ and $\mathbf{2}$ are both ditopic, we anticipated a twofold-symmetrical complex with the two ammonium ions in $\mathbf{2}$ simultaneously binding to the two crown-ethers in $\mathbf{oP^4}$. This expectation was confirmed by ¹H NMR titration experiments that were carried out with varying concentration of guest (1–7 mM) and constant host concentration (2 mM) at 278 K in 3:2 CDCl₃/CD₃CN (Figure S17). Figure 3 shows a representative spectrum. At sub-stoichiometric equivalents of $\mathbf{2}$, new signals corresponding to a major new species appear that were assigned to the 1:1 complex. With stoichiometric (or excess) $\mathbf{2}$, signals assigned to the 1:1 complex predominate along with signals from free $\mathbf{2}$ (some additional small signals suggest that further complexation may be possible, albeit unfavorable).

Binding in this system was clearly much stronger than with guest **1**. To estimate the association constant, UV–vis titration experiments were performed at constant concentration

of host (0.1 mM) with varying concentration of guest (0.01–0.35 mM) at 278 K in 3:2 $CHCl_3/CH_3CN$ (inset Figure 3 and Figure S43). As the concentration of **2** increases, the absorbance peak assigned to $\mathbf{oP^4}$ at 310 nm gradually decreases, with an isosbestic point at 300 nm (Figure S42). The overall association constant for the 1:1 complex of host $\mathbf{oP^4}$ and guest **2** was estimated as $K = (4.7 \pm 1.4) \times 10^4 \text{ M}^{-1}$ (278 K). Thus, binding between $\mathbf{oP^4}$ and **2** is approximately two orders of magnitude stronger than that between $\mathbf{oP^4}$ and **1** because of the chelate effect.

The ¹H NMR signals for the 1:1 complex could be assigned by using standard 2D NMR experiments (COSY, HSQC, HMBC) (Table S2 in SI). As shown in Figure 3, the chemical shifts of the protons on the internal rings (H_{2b}-H_{2e}) are significantly different in the complex compared to in the free host $\mathbf{oP^4}$, strongly suggesting that the conformation of the *o*-phenylene in the complex has changed. To confirm that NMR chemical shift changes are consistent with the expected changes in geometry, that is, correspond to a switch from the A to B states in Figure 1, isotropic shieldings were calculated for the A and B conformers of $\mathbf{oP^{4'}}$ using DFT (see Supporting Information). The resulting predicted chemical shift differences between the A and B states of $\mathbf{oP^{4'}}$ ($\delta^{\text{B}}_{calc} - \delta^{\text{A}}_{calc}$) were compared with the experimental chemical shifts differences between the complex and free $\mathbf{oP^4}$ ($\delta^{\text{complex}}_{exp} - \delta^{\text{free host}}_{exp}$). The good linear correlation suggests that the host $\mathbf{oP^4}$ is likely forced into the B conformer in the complex (Figure S23). That is, binding to 2 forces the outer rings of the *o*-phenylene apart. This is reasonable given the relative lengths of the species: the distance between the two N atoms in 2 is 7.5 Å, longer than the approximately 5 Å separation between terminal rings in $\mathbf{oP^4}$ in the A state.³⁴

After investigating the effect of *o*-phenylene folding with achiral guests 1 and 2, we examined twist-sense-induction from the binding of chiral guest 3. ¹H NMR experiments were carried out with guest 3 and host oP^4 at 278 K in 3:2 CDCl₃/CD₃CN (Figure S24). Very small signals in the spectra show that there is weak binding, although the complexity of the NMR spectrum did not allow straightforward estimation of binding constants. Similar



Figure 4: CD spectra of a 1:1 ratio of $\mathbf{oP^4/4}$ (1 mM each) and 4 (3 mM) alone (3:2 CHCl₃/CH₃CN, 278 K). The dotted line represents the predicted CD spectrum for (*M*)- $\mathbf{oP^{4'}}$ in its B state (arbitrary units, TD/PCM(CHCl₃/CAM-B3LYP/6-31G(d,p)// ω B97X-D/cc-pVDZ). Inset: CD titration monitoring at 290 nm.

results were observed for dibenzo-24-crown-8 with **3** (Figure S25), where K was determined to be approximately 40 M⁻¹, about an order of magnitude less than the binding to **1**.

Given the much stronger binding of $\mathbf{oP^4}$ to $\mathbf{2}$ compared to $\mathbf{1}$, we shifted our attention to ditopic chiral guest $\mathbf{4}$ (Chart 1).³⁵ NMR spectroscopy indicates that dibenzo-24-crown-8 binds to $\mathbf{4}$ (Figure S34).³⁶ The formation of a 1:1 complex between $\mathbf{oP^4}$ and $\mathbf{4}$ was confirmed by ¹H NMR titration experiments with varying concentration of $\mathbf{4}$ (0.5–4 mM) at a constant concentration of $\mathbf{oP^4}$ (1 mM) at 278 K in 3:2 CDCl₃/CD₃CN (Figure S26). The resulting ¹H NMR spectra are complex because of the two diastereomeric 1:1 complexes that can be formed between $\mathbf{oP^4}$ and $\mathbf{4}$ (i.e., the two possible twist senses of $\mathbf{oP^4}$ matched with the single enantiomer of $\mathbf{4}$). The signals for one of the complexes could be assigned by using standard 2D NMR experiments (COSY, HSQC, HMBC) (Table S3). The observed changes in chemical shifts compared to free $\mathbf{oP^4}$ are consistent with those observed for binding to $\mathbf{2}$; that is, comparison of the chemical shifts to isotropic shielding calculations suggest that the *o*-phenylene must unfold on binding (Figure S32).

As shown in Figure 4, the addition of chiral guest 4 to \mathbf{oP}^4 induces a negative Cotton effect at 290 nm, a region where the *o*-phenylene absorbs but not 4. CD spectroscopy of comparable mixtures of \mathbf{oP}^4 and 3 show the induction of a similar but much weaker Cotton effect (Figure S47). The incremental addition of chiral guest 4 to a solution of *o*-phenylene tetramer $\mathbf{oP^4}$ results in the increase and eventual saturation of the CD signal. This is due to the formation of the 1:1 complex as the dominant species as seen with the achiral guest 2, consistent with the NMR results. The overall association constant for the 1:1 complex of host $\mathbf{oP^4}$ and guest 4 determined in the CD titration is comparable to the binding of $\mathbf{oP^4}$ to 2, but with significant uncertainty.

TD-DFT predictions of the CD spectra of $\mathbf{oP^{4'}}$ in its B state (dotted line in Figure 4) indicate that the binding to 4 likely favors the M (left-handed) twist sense of the *o*-phenylene helix. Unfortunately, we could not identify NMR signals for the two *o*-phenylene twist senses that were sufficiently well-resolved to determine the diastereometric excess (de) of the complex.

In summary, we have explored the effect of guest binding on *o*-phenylene folding. Binding of monotopic guest **1** to host $\mathbf{oP^4}$ has little effect on the oligomer's conformation, which remains well-folded in its compact helical A state in both 1:1 and 1:2 complexes. Achiral ditopic guest **2** forms a much stronger 1:1 complex and forces the *o*-phenylene tetramer to misfold into an extended helical B state because of a mismatch between the separation of the ammonium groups in **2** and the crown ethers in $\mathbf{oP^4}$. Chiral ditopic guest **4** forms an analogous 1:1 complex and generates an excess of the *M* twist sense.

Experimental

Synthesis

Unless otherwise noted, all starting materials, reagents, and solvents were purchased from commercial sources and used without further purification. Melting points were determined using a Thermal Analysis Q20 differential scanning calorimeter at a heating rate of 10 °C/min. NMR spectra were measured for CDCl₃ and CD₃CN solutions using Bruker Avance 600 or 500 MHz NMR spectrometers. Structural assignments were made with additional information from gCOSY, gHSQC, and gHMBC experiments. The guest cations $1, {}^{31}$ $2, {}^{31}$ $3, {}^{37}$ and $\mathbf{4}^{35}$ were synthesized according to literature procedures, as was 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-dibenzo-24-crown-8.³⁰

o-Phenylene tetramer o P^4

An oven-dried Schlenk vacuum tube was charged with 2,2'-dibromobiphenyl²⁹ (0.14 g, 0.45 mmol), 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-dibenzo-24-crown-8³⁰ (0.644 g, 1.12 mmol), Pd(OAc)₂ (0.010 g, 0.045 mmol), SPhos (0.022 g, 0.053 mmol), and K₃PO₄ (0.285 g, 1.35 mmol), then evacuated and back-filled with argon $(3\times)$. To the tube was added degassed THF (12 mL) and deionized water (3 mL). The mixture was degassed by three freeze-pump-thaw cycles and then filled with argon. The reaction mixture was heated at 85–90 °C for 24 h (silicone oil bath), then cooled to room temperature and diluted with water (10 mL) and DCM (5 mL). The organic layer was separated and the aqueous layer extracted with DCM $(3 \times 5 \text{ mL})$. The combined organic layers were washed with brine, dried over MgSO₄, filtered, and concentrated. Purification by flash chromatography on basic alumina (1:9 acetone:DCM), recrystallization (MeOH), and chromatography on a C-18 column (MeOH) gave compound oP^4 as a white solid (0.132 g, 28%): mp 118.86 °C; ¹H NMR (600 MHz 3:2 $CDCl_3/CD_3CN$, 278 K) δ 7.20 (d, J = 7.6 Hz, 2H), 7.14 (t, J = 7.5 Hz, 2H), 7.08 (t, J =8.3 Hz, 2H), 6.89 (d, J = 8.9 Hz, 2H), 6.66 (m, 8H), 6.29 (d, J = 8.3 Hz, 2H), 5.86 (d, J = 8.3 Hz, 2H), 5. = 8.3 Hz, 2H), 5.83 (s, 2H), 3.90–3.78 (m, 12H), 3.65–3.61 (m, 12H), 3.54–3.43 (m, 22H), 3.35–3.30 (m, 2H); ${}^{13}C{}^{1}H$ NMR (126 MHz, 3:2 CDCl₃/CD₃CN, 278 K) δ 147.8, 146.5, 146.0, 139.6, 139.2, 132.8, 130.5, 128.6, 126.7, 126.1, 120.7, 120.4, 113.3, 112.8, 111.7, 70.12, 70.09, 70.0, 68.9, 68.8, 68.6, 68.02, 68.00, 67.4; HRMS (ESI-Orbitrap) m/z: [M+Na⁺] Calcd for $C_{60}H_{70}NaO_{16}^{+}$ 1069.4556; Found 1069.4513.

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Supporting Information

Synthetic scheme; NMR spectra; UV–vis and CD spectra; determination of association constants; computational chemistry details (PDF)

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