



# Complete Genome Sequences of Eight *Streptococcus equi* subsp. *zooepidemicus* Strains Isolated from Mares in Estrus with Endometritis

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**ABSTRACT** Eight isolates of *Streptococcus equi* subsp. *zooepidemicus* were isolated from mares with clinical cases of endometritis. *S. equi* subsp. *zooepidemicus* strains were chosen for sequencing based on differing levels of biofilm production *in vitro*. Using Illumina short-read sequencing in conjunction with MinION sequencing, we report the genomes of eight isolates.

Infectious endometritis, one of the leading causes of reduced fertility in mares, can be caused by fungal pathogens or bacterial pathogens such as *Streptococcus equi* subsp. *zooepidemicus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae* (1–4). A total of eight *S. equi* subsp. *zooepidemicus* isolates were chosen on the basis of their *in vitro* biofilm production using the microtiter dish biofilm formation assay (5). Three of the strains produced very little or no biofilm, two strains were medium biofilm producers, and three strains were high biofilm producers (Table 1).

*S. equi* subsp. *zooepidemicus* isolates were collected from the uterus of mares (age range, 4 to 13 years) in estrus using a sterile, double-guarded uterine swab during the breeding season of March 2017 to May 2018. Isolates of *S. equi* subsp. *zooepidemicus* were streaked for purity on tryptic soy agar (TSA) with 5% sheep blood plates. A single colony from each sample was grown overnight in Todd-Hewitt broth at 37°C in 5% CO<sub>2</sub>. Cryopreserved isolates were used for identification using matrix-assisted laser desorption/ionization–time of flight mass spectrometry (MALDI-TOF MS) (6). *S. equi* subsp. *zooepidemicus* isolates were grown statically overnight in 2 ml Todd-Hewitt broth with 0.5% Tween 80 at 37°C in 5% CO<sub>2</sub>. Genomic DNA was isolated from the bacterial cell pellet using the Qiagen DNeasy blood and tissue kit, following the manufacturer's instructions for Gram-positive bacteria.

For DNA sequencing and bioinformatic processing, a hybrid assembly approach utilized Illumina short reads and Oxford Nanopore Technologies (ONT) long reads obtained by MinION sequencing, which produced a single contig per genome or plasmid. Genomic DNA was sequenced on an Illumina NextSeq 550 sequencer from a 2 × 150-bp paired-end library that was prepared at the Microbial Genome Sequencing Center (MiGS) (University of Pittsburgh) using the Nextera DNA library preparation kit with modifications that included the substitution of KAPA HiFi PCR mix and a modified PCR protocol that used 7 cycles to attach indices followed by an additional 5 cycles after the addition of P5/P7 Illumina primers, as described by Baym et al. (7). Long-read sequencing was performed at Colorado State University using the ONT MinION platform with an R9 MinION flow cell using the rapid barcoding kit (P/N SQK-RBK004). The DNA was fragmented during tagmentation but was not size selected. The ONT read N<sub>50</sub> value (total for all data sets) was 6,618 bp. Base calling was performed using Guppy (v3.3.0). FastQ files from Illumina sequencing were assessed for quality using FastQC (v0.11.8) (8) and were trimmed using Cutadapt (v2.5) (9) to remove

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**TABLE 1** Characteristics and accession numbers of *Streptococcus equi* subsp. *zooepidemicus* genomes

| Isolate | Age of mare (yr) | Biofilm production level | No. of Illumina reads | SRA accession no. for Illumina reads | No. of MinION reads | SRA accession no. for MinION reads | Plasmid size (bp) | GenBank accession no. plasmid | Assembly size (bp) | No. of coding sequences | G+C content (%) | GenBank accession no. |
|---------|------------------|--------------------------|-----------------------|--------------------------------------|---------------------|------------------------------------|-------------------|-------------------------------|--------------------|-------------------------|-----------------|-----------------------|
| SEZ13   | 13               | Low                      | 3,631,288             | SRR12988984                          | 15,448              | SRR12995728                        | 4,723             | CP065055                      | 2,150,055          | 1,950                   | 41.6            | CP065054              |
| SEZ14   | 9                | Low                      | 2,598,319             | SRR12988983                          | 127,002             | SRR12995727                        | 4,652             | CP065057                      | 2,147,596          | 2,003                   | 41.6            | CP065056              |
| SEZ18   | 10               | Medium                   | 2,183,355             | SRR12988982                          | 92,263              | SRR12995726                        | NA <sup>a</sup>   |                               | 2,082,352          | 1,890                   | 41.6            | CP065058              |
| SEZ25   | 10               | High                     | 2,147,048             | SRR12988981                          | 80,784              | SRR12995725                        | NA                |                               | 2,098,084          | 1,949                   | 41.6            | CP065059              |
| SEZ28   | 12               | Low                      | 1,988,827             | SRR12988980                          | 77,047              | SRR12995724                        | NA                |                               | 2,044,877          | 1,841                   | 41.7            | CP065060              |
| SEZ33   | 4                | High                     | 1,577,737             | SRR12988979                          | 79,480              | SRR12995723                        | NA                |                               | 2,040,445          | 1,825                   | 41.7            | CP065061              |
| SEZ36   | 10               | High                     | 3,550,098             | SRR12988978                          | 115,841             | SRR12995722                        | NA                |                               | 2,140,969          | 2,001                   | 41.7            | CP065190              |
| SEZ46   | 14               | Medium                   | 2,350,585             | SRR12988977                          | 43,338              | SRR12995721                        | NA                |                               | 2,154,650          | 1,996                   | 41.4            | CP065191              |

<sup>a</sup>NA, not applicable.

sequences that were less than 80 bp and those that were below a minimum quality score of 30 on the Phred scale. MinION reads and trimmed Illumina reads were assembled using the SPAdes hybrid assembler (v3.13.1) (10) and Unicycler (v0.4.8) (11). *S. equi* subsp. *zooepidemicus* genomes were annotated with Prokka (v1.14.0) (12). Prokka annotations for all genomes can be found in Table 1. To determine the start position within each genome, *dnaA* from the *S. equi* subsp. *zooepidemicus* H70 genome (GenBank accession number [FM204884.1](https://www.ncbi.nlm.nih.gov/nuclseq/FM204884.1)) (13) was aligned to the chromosomal assemblies using the Burrows-Wheeler Aligner (v0.7.17) (14). Using an in-house Python script (<https://github.com/lakinsm/strep-equi-mra>), the chromosomal assemblies were aligned, reverse complemented, and circularly shifted so that all chromosomal assemblies had the same orientation and start position. Visualization and additional analyses were performed using Geneious Prime (v2019.2.1). Default parameters were used except where otherwise noted.

*S. equi* subsp. *zooepidemicus* genomes ranged from 2,040,445 to 2,154,650 bp (Table 1). Two of the strains, SEZ13 and SEZ14 (both low biofilm producers), maintained plasmids of 4,723 bp and 4,652 bp, respectively.

**Data availability.** All eight *S. equi* subsp. *zooepidemicus* genomes are associated with BioProject [PRJNA673199](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA673199). Illumina and MinION raw reads are available under the accession numbers listed in Table 1.

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