

# Extremely Slow Diffusion of Gold Nanoparticles under Confinement in Nanoporous Silica

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## Zusammenfassung

We report an x-ray photon correlation spectroscopy (XPCS) study of the mobility of colloidal gold nanoparticles with diameter approximately 4 nm in glycerol under confinement within the channels of SBA-15 mesoporous silica with pore diameters of 6 nm and 12 nm. The XPCS correlation functions result from an effective heterodyne signal due to the mixing of the coherent scattering from the nanoparticles, which are dynamic, and the mesoporous silica, which is essentially static. Over the range of wave vectors and hence length scales probed, the nanoparticle dynamics are well described by one-dimensional diffusion. The nanoparticle diffusivity varies with temperature in a manner expected based on the temperature-dependent viscosity of glycerol; however, the magnitudes of the diffusion coefficients are several orders of magnitude smaller than those of the nanoparticles in bulk glycerol. We consider mechanisms that might contribute to this reduction in diffusivity including enhancement in hydrodynamic drag under confinement, effects of nanoparticle adsorption to the pore walls, and slowing structural dynamics in the glycerol due to the combination of the nanoconfinement and the presence of the nanoparticles.

## I. INTRODUCTION

The mobility of nanoscale particles suspended in fluids within confined geometries impacts a wide range of fields including biotechnology, catalysis, nanofabrication, lubrication, and environmental remediation. [1] Under nanometer-scale confinement, the Brownian diffusion that nanoparticles typically experience in a bulk fluid can be radically altered due to hydrodynamic, surface, and other interactions that become paramount when the particles are restricted to remain in close proximity to the bounding surfaces [2–4]. A number of experimental approaches have been employed to characterize nanoparticle mobility under such conditions [5–11]. In particular, recent advances in transmission electron microscopy (TEM) in liquid environments have enabled detailed single-particle-tracking experiments that have investigated nanoparticle motion in fluids under nanoscale confinement [12–18], and in several cases these studies have revealed dynamics that are orders of magnitude slower than the corresponding Brownian motion in bulk [Refs]. Such microscopy methods are well suited for tracking nanoparticles that are confined along one direction, such as between closely spaced parallel substrates. However, other important confinement geometries for nanoparticle mobility, such as within bulk nanoporous materials, present a challenge for established techniques. To address this issue, we have performed an x-ray photon correlation spectroscopy (XPCS) study of the dynamics of dilute dispersions of colloidal gold nanoparticles under confinement in the nanochannels of mesoporous SBA-15 silicate. The ability of XPCS to track the motion of nanoparticles over nanometer-scale distances makes it an ideal probe for such studies.

In XPCS, like in dynamic light scattering (DLS) with visible light, fluctuations in the coherent scattering intensity from a material provide information about the material’s dynamics. However, due to the much shorter wavelength of x-rays, XPCS can probe motions over significantly smaller distances. This advantage provides XPCS with unique access to a region of dynamic phase space at small length scales and long time scales that makes it particularly well matched for interrogating nanoparticle mobility under nanometer-scale confinement [19–22]. Another strength of XPCS stems from the relatively large penetration depth of x-rays, which enables the technique to probe the interior of optically opaque materials, a property we have exploited in probing the nanoparticle mobility within the mesoporous silicate matrices. Leveraging these advantages, a number of previous studies have

employed XPCS to interrogate nanoparticle dynamics under confinement in a number of different contexts including in polymer thin films, where the nanoparticles served as probes of the local viscosity [23–25][others], in cross-linked and entangled polymeric systems, where the nanoparticle mobility can display violations of the Stokes-Einstein relations that provide insight into the hierarchical structural dynamics of the materials [26–28][other], and in dense concentrations in porous environments, where the effects of the confinement on collective dynamics have been reported [29]. Here, we employ the technique to examine the mobility of Au nanoparticles with diameter of approximately 4 nm in glycerol confined within the straight, monodisperse pores of SBA-15 silicate templates. Templates with two pore diameters,  $D_p = 6$  nm and 12 nm, were included in the study. The measurements characterize the diffusion of the nanoparticles along the pore axes and find diffusion coefficients that depend strongly on the pore size and that are several orders of magnitude smaller than those of the nanoparticles in bulk glycerol. Nevertheless, the diffusion coefficients appear to follow a temperature dependence expected based on the viscosity of glycerol. We discuss several possible mechanisms for this strong reduction in nanoparticle mobility and propose that slowing structural dynamics of the glycerol due to the nanoconfinement in the presence of the nanoparticles could be a major contributor.

## II. METHODS

### A. Sample Synthesis

#### 1. *Synthesis of Au Nanoparticles*

To avoid the need to transfer nanoparticles from aqueous solution into glycerol, we synthesized the colloidal Au particles directly in glycerol through the reduction of gold(III) chloride trihydrate salt ( $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ ) with tert-butylamine borane (BTB) in the presence of trioctylphosphine (TOP) as a capping agent. All chemicals were purchased from Sigma-Aldrich and used as received. In a typical procedure, 0.5 mmol (0.22 ml) TOP was injected into a solution of 0.5 g of  $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$  in 20 ml glycerol at 15 °C after the reaction system had been sealed and purged with argon. 0.5 mmol BTB was solvated in 2 ml glycerol, sonicated for 10 minutes, and briefly heated with a heat gun to decrease the solvent viscosity before being injected into the gold solution. Directly after the injection of

the reducing agent, a color change from yellow to light purple was observed. The resulting solution was then heated to 30 C for 1 hour under continuous argon flow. After the end of the reaction, the solution was cooled to room temperature, and the Au nanoparticles were extracted by adding 50 mL of isopropanol as a precipitant, followed by centrifugation (10000 rpm for 10 min). The TOP-stabilized Au nanoparticles were then re-dispersed in glycerol via probe sonication. The final concentration of nanoparticles was approximately 0.25% by volume. The nanoparticles were characterized by TEM, which revealed a particle radius of about 1.5 nm. We further estimate the TOP shell adds about 0.5 nm to the nanoparticles' radius, giving the particles a hydrodynamic diameter of approximately 4 nm.

## 2. *Synthesis of SBA-15 Mesoporous Materials*

SBA-15 is a mesoporous silica sieve containing pores with a narrow pore size distribution that are arranged in highly ordered, uniform hexagonal lattice [30]. SBA-15 matrices with two pore sizes were synthesized. Both syntheses used micelles of the nonionic triblock copolymer Pluronic P123 as the template. For the smaller pore diameter (6 nm), we followed the procedure reported by Zhao *et al.* [31] with modifications to the thermal treatments during synthesis based on the study by Brodie-Linder *et al.* [32]. Tetraethylorthosilicate (TEOS) was used as the silica source and 38% HCl was used as the acid catalyst for the silica condensation reaction. The Pluronic (4.0 g) was dissolved in a solution of distilled water and concentrated HCl 38% (125 and 25 g, respectively). After stirring for 3 hr at 30 C, the TEOS (8.6 g) was added, and the solution was vigorously stirred for an additional 10 minutes. Then, the flask was sealed and the solution was kept for 24 hr at the same temperature. At this point, the temperature was adjusted to 100 C, and the mixture was again kept for 24 hr under reflux in static conditions. The resulting white solid was passed through a Buchner filter while being washed with 300 ml of water to eliminate the acid and excess reactants. It was then dried at room temperature for 24 hr, and then placed in a furnace where it was heated under air at 550 C for 18 hr. A white powder of SBA-15 was then collected. The resulting SBA-15 matrices, containing straight and highly parallel, monodisperse pores, had a porosity of approximately  $1 \text{ cm}^3\text{g}^{-1}$ , as determined by adsorption isotherms. (The porosity is the total pore volume and can be deduced from the amount of adsorbate at a relative pressure of 0.95 in adsorption isotherms.) The pore diameter of the

SBA-15,  $D_p \approx 6$  nm, was obtained using the Barrett-Joyner-Halenda (BJH) method [33].

To obtain SBA-15 with larger pore size, we followed the synthesis reported by Kruk *et al.* [34], in which hexane was used to expand the Pluronic micelles. The synthesis proceeded by dissolving 0.027 g  $\text{NH}_4\text{F}$  and 2.4 g P123 in 84 mL 1.3 M HCL solution at room temperature. The mixture was then moved to a water bath at 15 C and kept there for 1 hr, after which a mixture of 5.17 g TEOS and 8.45 g hexane was added. The reaction mixture was kept at 15 C under continuous stirring for an additional 24 hr. The mixture was then transferred into a teflon-lined autoclave and heated to 130 C for 24 hr. The resulting powder was washed twice with ethanol and water, dried in a vacuum oven, and then calcined at 550 C under air to remove the surfactant. The white SBA-15 powder was then recovered and characterized. Using adsorption isotherms, the porosity was found to be approximately  $1.12 \text{ cm}^3 \text{ g}^{-1}$ . The pore diameter was determined by the improved Kruk-Jaroniec-Sayari method [35], which is appropriate for larger-pore SBA-15 molecular sieves, and was found to be  $D_p \approx 12$  nm.

## B. XPCS

XPCS experiments were performed at the CHX beamline at the NSLS-II synchrotron facility at Brookhaven National Laboratory. Samples were contained in sealed stainless steel holders with ??? mm thickness and polyimide windows for transmission x-ray scattering measurements. The samples were prepared directly in the holders by combining appropriate quantities of the SBA-15 matrix and nanoparticle suspension to achieve 90% filling of the total available pore volume. Such fill has been found to cancel fully the Bragg peaks of the empty matrix in contrast-matching neutron scattering while leaving no observable fluid outside the pores [36]. As the glycerol strongly wets the SBA-15 surfaces, the suspension was taken up quickly by the matrix through capillary action. After the holders were filled, the samples were left overnight at 40 C to ensure complete filling of the channels and equilibration of the system.

The sample holders were positioned in a temperature-control stage on the beam line. A partially coherent x-ray beam of energy 9 keV and size  $10 \times 10 \text{ } \mu\text{m}^2$  was incident on the sample. An area detector XXX m after the sample measured the scattering intensity at a frame rate of 500 fps over the range  $0.02 \text{ nm}^{-1} < q < 0.65 \text{ nm}^{-1}$ . All measurements

were conducted at least one hour after reaching each temperature to ensure its stabilization. The primary quantity obtained in the XPCS measurements was the time auto-correlation function of the scattered intensity,

$$g^{(2)}(\mathbf{q}, t) = \frac{\langle I(\mathbf{q}, t')I(\mathbf{q}, t' + t) \rangle}{\langle I(\mathbf{q}, t') \rangle^2} \quad (1)$$

where  $I(\mathbf{q}, t')$  is the intensity measured at wave vector  $\mathbf{q}$  and measurement time  $t'$ , and the averages are over  $t'$  and over detector pixels within a small vicinity of  $\mathbf{q}$ . At large delay times  $t$ ,  $g^{(2)}(\mathbf{q}, t)$  approaches unity if the system is ergodic. At  $t \rightarrow 0$ , it approaches  $1 + \beta$ , where  $\beta$ , known as the optical contrast, is the variance of the fluctuations in intensity divided by  $\langle I \rangle^2$ . The optical contrast depends on instrumental factors, such as the coherence of the incident beam, the solid angle subtended by a detector pixel, and the sample thickness [37]. In the measurements on 6-nm SBA-15,  $\beta \approx 0.11$ , while in the measurements on 12-nm SBA-15,  $\beta \approx 0.15$  due some modifications to the x-ray optics between the two sets of measurements.

### III. RESULTS

Figure 1 shows the x-ray scattering intensity  $I(q)$  from the SBA-15 matrix with 12 nm pores when filled with pure glycerol and when filled with the Au nanoparticle suspension. Both profiles show a pronounced peak at  $q \approx 0.45 \text{ nm}^{-1}$  resulting from spatial correlations among the hexagonally packed SBA-15 pores, which have a characteristic spacing  $2\pi/0.45 \approx 14 \text{ nm}$ . For comparison, the curves have been normalized with respect to the height of this peak. Above  $q \approx 0.3 \text{ nm}^{-1}$ , the two profiles are very similar, indicating that the mesoporous silica dominates the scattering at high  $q$ . At lower  $q$ , the matrix containing the nanoparticle suspension shows enhanced scattering intensity reflecting the contribution from the Au nanoparticles. Analysis of the XPCS results to characterize the nanoparticle dynamics was hence restricted to wave vectors below a maximum,  $q_{max} = 0.25 \text{ nm}^{-1}$ . The size of the contribution to  $I(q)$  from the nanoparticles varied with location in the sample probed by the microbeam, suggesting spatial inhomogeneity in the concentration of nanoparticles. However, for all measurements, the matrix contributed substantially to the scattering intensity even at low  $q$ , influencing our interpretation of the XPCS results described below.

Figure 2(a) shows the XPCS autocorrelation function  $g_2(q, t)$  at  $q = 0.04 \text{ nm}^{-1}$  measured on the filled SBA-15 with 12 nm pores at temperatures ranging from 310 K to 380 K.

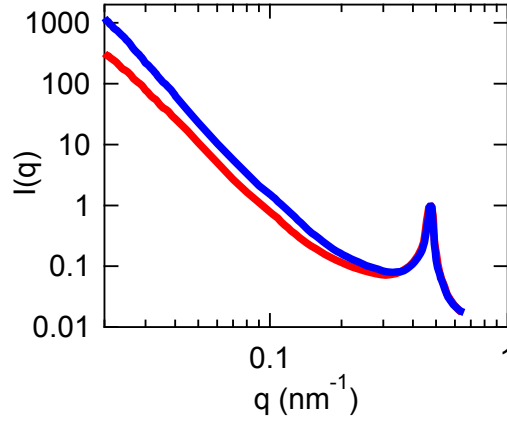


Abbildung 1. X-ray scattering intensity  $I(q)$  measured on the SBA-15 matrix with 12 nm pores when filled with pure glycerol (red) and when filled with the Au nanoparticle suspension (blue). Both profiles show a pronounced peak at  $q \approx 0.45 \text{ nm}^{-1}$  resulting from spatial correlations among the SBA-15 pores, which have a characteristic spacing  $2\pi/0.45 \approx 14 \text{ nm}$ . To aid comparison, the curves have been normalized with respect to the height of this peak.

Figure 2(b) shows correlation functions at  $q = 0.10 \text{ nm}^{-1}$  for the filled SBA-15 with 6 nm pores between 403 K to 483 K. In both cases, the correlation functions display a partial decay that moves to shorter delay times with increasing temperature. We associate this decay with motion of the Au nanoparticles within the pores. We further interpret the fact that the system is not ergodic (*i.e.*,  $g_2(q, t)$  does not decay fully to one) as a consequence of the contribution to the scattering from the mesoporous silica, which is effectively static. This static contribution is larger for the 6 nm pore sample, reflecting the larger contribution to the scattering in this case. The dynamics of the nanoparticles in the 6 nm pores is also significantly slower than in the 12 nm pores, as illustrated by the fact that the decays in  $g_2(q, t)$  in Fig. 2(b) occur at larger  $t$  than in Fig. 2(a) despite the higher measurement temperatures. In both cases, these dynamics are far slower than expected based on the diffusivity of the Au nanoparticles in bulk glycerol. This difference is illustrated by the result for  $g_2(q, t)$  at  $q = 0.04 \text{ nm}^{-1}$  measured on the bulk nanoparticle solution at 263 K, which is shown by the open squares in Fig. 2(a). The dashed line through bulk data shows the result of a fit using an exponential decay,

$$g_2(q, t) = 1 + b \exp(-t/\tau) \quad (2)$$

from which the diffusion coefficient can be extracted through the relation  $\tau = 1/Dq^2 = \frac{6\pi\eta R}{k_B T q^2}$ , where  $\eta$  is the glycerol viscosity. Despite the viscosity of glycerol decreasing approximately 2 to 3 orders of magnitude between 263 K and the temperature range of the measurements in the 12 nm pores (310 K to 380 K) [38, 39], the decay times are similar.

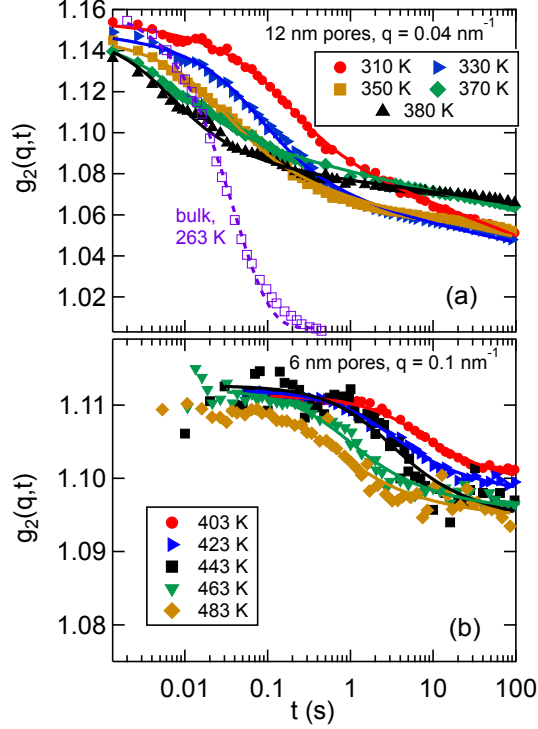


Abbildung 2. XPCS intensity autocorrelation function  $g_2(q, t)$  measured on matrices with SBA-15 with (a) 12 nm and (b) 6 nm pores filled with Au nanoparticle suspensions at several temperature as indicated in the legends. The solid lines show the results of fits using a model of heterodyne scattering as described in the text. The open squares in (a) show  $g_2(q, t)$  measured on the bulk Au suspension at 263 K, and the dashed line shows the result of a fit using an exponential decay.

#### IV. ANALYSIS

As described above, both the Au nanoparticles and SBA-15 matrix contribute appreciably to the scattering below  $q_{max} \approx 0.25 \text{ nm}^{-1}$ . In this wave-vector range the XPCS measurements hence detected a heterodyne signal, wherein the scattering from the matrix and from



the Au nanoparticles mixed coherently [40]. For such heterodyne scattering, the intensity autocorrelation function  $g_2(q, t)$  can be expressed as

$$g_2(\mathbf{q}, t) = 1 + \beta \left( X^2 |g_{1,\text{Au}}(\mathbf{q}, t)|^2 + 2X(1-X) \text{Re} [g_{1,\text{Au}}(\mathbf{q}, t) g_{1,\text{SBA}}^*(\mathbf{q}, t)] + (1-X)^2 |g_{1,\text{SBA}}(\mathbf{q}, t)|^2 \right) \quad (3)$$

where  $g_{1,\text{Au}}(\mathbf{q}, t)$  and  $g_{1,\text{SBA}}(\mathbf{q}, t)$  are the intermediate scattering functions of the Au nanoparticles and the SBA-15 matrix, respectively,  $X$  is the mixing ratio that is a measure of the fractional contribution to the scattering intensity from the Au nanoparticles.

Since the scattering from the Au nanoparticles contributed appreciably to the total scattering only below  $q_{\text{max}} \approx 0.25 \text{ nm}^{-1}$ , the measurements probed the nanoparticle motion over a range of length scales above a lower limit of approximately  $q_{\text{max}}^{-1} \approx 4 \text{ nm}$ . Further, since the particles had a hydrodynamic diameter of about 4 nm, their motion within the 12-nm pores in the directions transverse to the pore axes was restricted to displacements less than about 8 nm, and within the 6-nm pores the transverse displacements were restricted to roughly 2 nm. Therefore, the measurements accessed primarily the nanoparticle motion along the direction of the pore axes. To interpret the XPCS results under this condition, consider a pore that is oriented along a direction  $\hat{\mathbf{x}}$  at an angle  $\theta$  to the scattering wave vector direction  $\hat{\mathbf{q}}$ , as depicted in the schematics in Fig. 3. We model particle motion down the pore as simple diffusion in one dimension wherein the particles have a probability distribution of displacements  $\Delta x$  parallel to the pore axis in a time duration  $t$  that is Gaussian,  $p(\Delta x, t) = \exp(-\Delta x^2/4Dt)/\sqrt{4\pi Dt}$  and a mean squared displacement given by  $\langle \Delta x^2(t) \rangle = 2Dt$ , where  $D$  is the diffusion coefficient. To understand how these dynamics affect  $g_{1,\text{Au}}(\mathbf{q}, t)$ , we assume the nanoparticles are sufficiently dilute that their dynamics are independent, so that the intermediate scattering function contains only self terms,

$$g_{1,\text{Au}}(\mathbf{q}, t) = \left\langle \frac{1}{N} \sum_{j=1}^N \exp [i\mathbf{q} \cdot (\mathbf{r}_j(t+t') - \mathbf{r}_j(t'))] \right\rangle_{t'} \quad (4)$$

where  $\mathbf{r}_j(t')$  is the position of particle  $j$  at time  $t'$ . That is,  $g_{1,\text{Au}}(\mathbf{q}, t)$  depends on the particle displacements projected onto  $\hat{\mathbf{q}}$ . For diffusion along the pore axis, that projected motion is similarly simple diffusion except with an effective diffusion coefficient  $D_{\text{eff}} = D \cos^2 \theta$ . For such diffusion, the intermediate scattering function decays as a simple exponential,

$$g_{1,\text{Au}}^\theta(q, t) = \exp(-q^2 D_{\text{eff}} t) = \exp(-q^2 D t \cos^2 \theta) \quad (5)$$

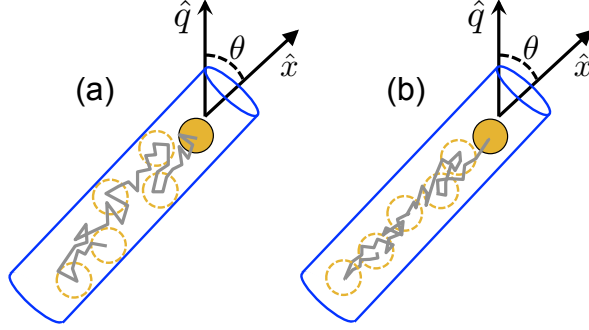


Abbildung 3. Schematics depicting a nanoparticle diffusing in glycerol within an SBA-15 nanopore. In (a) the particle's trajectory is punctuated by repeated adsorption to the pore walls. In (b), the particle is restricted to central region of the pore, for instance because of an immobile layer of glycerol adjacent to the walls.

Since the SBA-15 samples are powders, they contain grains with a uniform distribution of orientations. Therefore, at any given scattering wave vector  $\mathbf{q}$ , the intermediate scattering function will correspond to an average over pore orientations,

$$g_{1,\text{Au}}(q, t) = \langle g_{1,\text{Au}}^\theta(q, t) \rangle_\theta = \frac{1}{2} \int_{-1}^1 \exp(-q^2 D t \cos^2 \theta) d(\cos \theta). \quad (6)$$

Evaluating this average leads to

$$g_{1,\text{Au}}(q, t) = \frac{\sqrt{\pi}}{2} \frac{\text{erf}(\sqrt{\Gamma t})}{\sqrt{\Gamma t}} \quad (7)$$

where  $\Gamma = q^2 D$ , and  $\text{erf}()$  is the error function.

Regarding  $g_{1,\text{SBA}}(q, t)$ , the intermediate scattering function of the SBA-15, since the structure of the porous silica is nominally static, in principle  $g_{1,\text{SBA}}(q, t) \approx 1$ . However, the XPCS measurements appear to capture some slow dynamics in the SBA-15, as evidenced from the fact that the  $g_2(q, t)$  curves in Fig. 2 do not plateau to a constant value but show a slight decay at large  $t$ . We believe these slow dynamics might result from slow stress relaxation in the matrix like that observed with XPCS in other nanoporous solids [37, 41]. Hence, to describe these dynamics we employ a compressed exponential lineshape that has been shown to describe accurately the correlation function in these other cases,

$$g_{1,\text{SBA}}(q, t) = \exp \left[ -(t/\tau_S)^{1.5} \right] \quad (8)$$

We thus fit the results for  $g_2(q, t)$  using Eq. (3) with  $g_{1,\text{Au}}(q, t)$  and  $g_{1,\text{SBA}}(q, t)$  given by Eq. (7) and Eq. (8), respectively. The solid lines in Fig. 2 display results of such fits. The parameters included in the fits were  $X$ ,  $\Gamma$ , and  $\tau_S$ . Typically, the fits found  $\tau_S > 1000$  s with significant uncertainty, reflecting the slow decay of  $g_2(q, t)$  that is only partially captured in the data. The values of  $X$  obtained from the fits, which tend to decrease with increasing  $q$ , vary from approximately 0.3 to 0.6 for the 12-nm pores and 0.1 to 0.3 for the 6-nm pores, consistent with the range expected based on the ratio  $I(q)$  for the silica matrices filled with nanoparticle suspension and with pure glycerol.

As the agreement between the fits and data in Fig. 2 demonstrates, the initial decay of  $g_2(q, t)$  is accurately captured by the model of one-dimensional diffusion of the nanoparticles within the pores described above. Figures 4(a) and (b) show the decay rates  $\Gamma$  obtained from the fits as a function of  $q^2$  at several temperatures for the 12-nm pores and 6-nm pores, respectively. At all temperatures,  $\Gamma$  varies linearly with  $q^2$  as expected for diffusive dynamics. The solid lines in Figs. 4(a) and (b) show the results of fits using the form  $\Gamma = q^2 D$ , from which we obtain the temperature-dependent diffusion coefficients  $D$  of the nanoparticles.

Figure 5 shows the nanoparticle diffusion coefficients as a function of inverse temperature. As noted above, the mobility of the nanoparticles confined to the pores is highly suppressed with respect to that in bulk glycerol. Surprisingly, however, the temperature dependence of the diffusivity appears to track, at least approximately, that of the inverse of the viscosity of glycerol. Specifically, the red dashed line in Fig. 5 through the diffusion coefficients in the 12-nm pores shows the diffusion coefficient expected for the nanoparticles based on the Stokes-Einstein relation,  $D = \frac{k_B T}{6\pi\eta R}$ , if they were in a fluid with viscosity  $\eta$  equal to 132 times that of glycerol's temperature-dependent viscosity [38, 39]. The line tracks the measured diffusion coefficients of the nanoparticles. Similarly, the blue dashed line, which tracks  $D$  of the particles in the 6-nm pores, shows the diffusion coefficients expected for the nanoparticles in a fluid with a viscosity equal to 820,000 times that of glycerol. From these comparisons we hence conclude that the Au nanoparticles in glycerol confined to nanoporous SBA-15 undergo diffusion that is extremely slowed compared with that in bulk glycerol but that nevertheless varies with temperature approximately as expected based on the viscosity of glycerol.

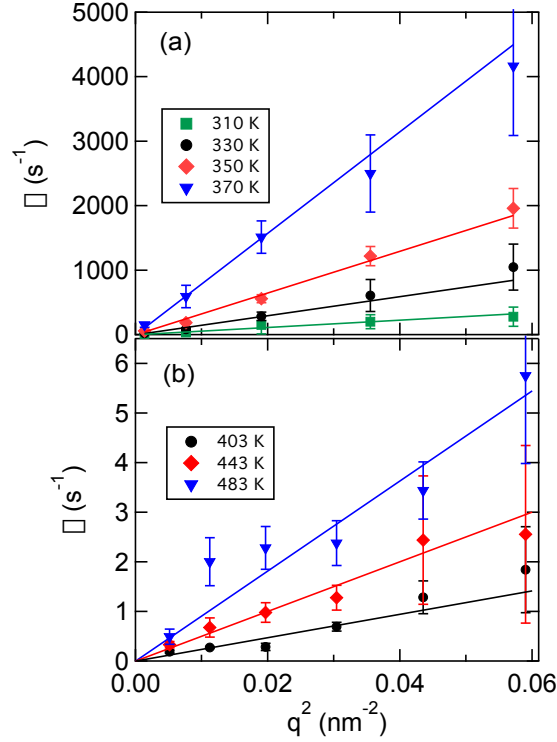


Abbildung 4. Decay rate  $\Gamma$  obtained from fits using Eq. (7) as a function of wave vector squared at various temperatures indicated in the legend for (a) 12-nm pores and (b) 6-nm pores. The lines display the results of linear fits from which the nanoparticle diffusion coefficient is obtained.

## V. DISCUSSION

Several possible mechanisms might contribute to the very small diffusion coefficients of the Au nanoparticles within the SBA-15 and their temperature dependence [cite Zhdanov and Shen?]. One contribution is the enhancement to the hydrodynamic drag on the nanoparticles due of the confinement. The corrections to Stokes drag on a sphere translating in a cylindrical channel have been the subject of detailed fluid-dynamics calculations and experiments. [42, 43]. The Stokes drag on a 4-nm sphere is enhanced over that in bulk by a factor of roughly 20 in a 6-nm pore and by a factor of roughly 3 in 12-nm pore, depending of the position of the sphere relative to the central axis of the pore. While not insignificant, these increases are too small to account fully for the large suppression in diffusivity observed in the experiments.

A second possible contribution involves surface interactions that lead to adsorption and desorption of the particles at the internal silica surfaces of the pores. Reduced mobility dicta-

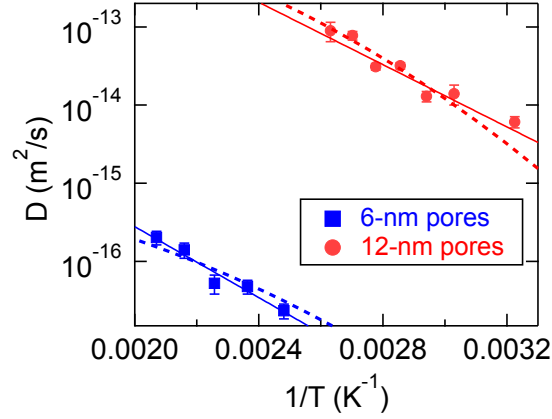


Abbildung 5. Diffusion coefficients  $D$  of the Au nanoparticles in the 12 nm (red circles) and 6 nm (blue squares) pores as a function of inverse temperature. The solid lines display the results of fits using an Arrhenius form for  $D$ . The dashed lines are the diffusion coefficients for Au nanoparticles with hydrodynamic radius of 2 nm in a fluid with a viscosity proportional to the temperature-dependent viscosity of glycerol. Specifically, the red dashed line is the diffusion coefficient the particles in a fluid with a temperature-dependent viscosity equal to 132 times that of glycerol, and the blue dashed line is in a fluid with a viscosity 820,000 times that of glycerol.

ted by adsorption and desorption has been observed for nanoparticles in planer confinement [Refs] and for fluorescent probe molecules in nanoporous membranes [44]. We hypothesize that the nanoparticles within the pores might follow diffusive trajectories like that depicted schematically in Fig. 3(a), in which the nanoparticles repeatedly adsorb to the pore walls, where they remain for an average residence time  $\tau_r$ . Upon desorption, the particles diffuse such that they travel along the pore axis  $\hat{x}$  a typical distance  $L$  before re-adsorbing. To estimate  $L$ , we assume the particles traverse along the pore axis a distance approximately equal to their lateral confinement, so  $L \approx 8$  nm in the 12-nm pores and  $L \approx 2$  nm in the 6-nm pores. The diffusion coefficient would hence be  $D = L^2/\tau_r$ . Since the desorption should be thermally activated, the diffusion coefficient can be expected to follow an Arrhenius form,

$$D = L^2 f_0 \exp(-E_A/k_B T) \quad (9)$$

Where  $E_A$  is the activation energy, and  $f_0$  is an attempt frequency. The red and blue solid lines in Fig. 5 show the results of fits to  $D$  using Eq. (9) for the 12-nm and 6-nm pores,

respectively. The fits for the two pores sizes, which describe the temperature dependence accurately, find very similar activation energies,  $(4600 \pm 300)k_B$  for the 12-nm pores and  $(5200 \pm 600)k_B$  for the 6-nm pores. The large difference in diffusion coefficients between the pore sizes is instead captured by  $f_0$ , which is  $(2.0 \pm 0.1) \times 10^8 \text{ s}^{-1}$  for the 12-nm pores but only  $(2.5 \pm 0.3) \times 10^6 \text{ s}^{-1}$  for the 6-nm pores. Such a large difference in attempt frequencies is difficult to interpret. Naively, one would expect that the difference in  $D$  between the pore sizes would be due to differing activation energies, particularly since the smaller radius of curvature in the 6-nm pore should lead to larger effective contact area between the particles and pore walls, raising questions about whether adsorption and desorption indeed plays a major role in dictating nanoparticle mobility in the pores.

A third, intriguing possible contribution to the suppressed nanoparticle diffusivity in the SBA-15 is slowed structural dynamics in the glycerol induced by the nanoconfinement. Nanoconfinement is known to alter the structural dynamics and hence viscosity of liquids and to change the glass transition temperature  $T_g$  of glass-forming liquids like glycerol [45, 46]. In the case of glycerol, experiments have reported that nanoconfinement can both increase and decrease  $T_g$ , which is approximately 190 K in bulk [47–50]. This variation in response of the structural dynamics to confinement is attributed to the different influences of the finite size effects versus the surface interactions between the liquid molecules and confining matrix, which depend on the specific surface chemistry of the matrix [51]. Surface interactions can be expected to be particularly strong between SBA-15 and glycerol since surface hydroxyl groups (silanols) on the amorphous silica can form hydrogen bonds with the glycerol. To account for surface interactions, researchers have proposed models in which a confined fluid contains two regions, a relatively immobile, near-surface layer close to the confining walls and a more mobile internal volume. Indeed, Trofymuk *et al.* employed FTIR spectroscopy to investigate the structural dynamics of glycerol in SBA-15 and identified an interfacial layer with an estimated thickness of 0.7 nm [48]. Trofymuk *et al.* further employed calorimetry to show a upward shift in  $T_g$  of about 5 K in 6-nm pores, indicating slowing structural dynamics due to the confinement. Levchenko *et al.* further used  $^{13}\text{C}$  NMR measured directly the reorientational dynamics of the glycerol in the mesoporous silica matrix MCM-41 with 2.2 nm pores and found the dynamics were slowed by more than 3 orders of magnitude at room temperature due to the confinement [52]. We note these previous studies investigated neat glycerol under nanoconfinement and that the addition of nanoparticles within the nanopores

makes the issues of finite size effects and surface interactions more complicated since both pore walls and particles provide confining surfaces. For instance, in the 6-nm pores the 4-nm nanoparticles are never more than 1 nm from a pore wall. Thus, strong slowing of the glycerol structural dynamics like that reported by Levchenko *et al.* could potentially be a major contributor to the large reduction in nanoparticle diffusion that we observe.

We note that the presence of a relatively immobile surface layer could restrict the nanoparticles to the center of the pores, leading to diffusive trajectories like that depicted in Fig. 3(b). Such restriction to the pore centers would help explain the success of Eq. (7) to wave vectors as large as  $q_{max} = 0.25 \text{ nm}^{-1}$ . Specifically, since the nanoparticle motion is expected to transition from 3D-like to 1D-like at a length scale set by the lateral confinement,  $g_{1,Au}(q, t)$  can be expected to change from Eq. (7) in the limit of low  $q$  to something closer to a simple exponential decay at high  $q$ . Based on an estimate of 8 nm for the extent of lateral motion of the nanoparticles in the 12-nm pores, one might expect to see evidence of this crossover in the lineshape below  $q_{max}$ ; however, further confinement due to an interfacial layer would push the crossover to higher  $q$ .

## VI. CONCLUSION

In conclusion, we have observed that gold nanoparticles in glycerol confined to nanometer-scale pores undergo diffusion that is dramatically slower than that of the nanoparticles in bulk glycerol. We consider several mechanisms that might contribute to this reduction in diffusivity and hypothesize that suppression of the structural dynamics of the glycerol solvent due to the confinement imposed by the pore surfaces in conjunction with the presence of the nanoparticles could be a major contributor. Further studies, such as those employing SBA-15 with silanized surfaces that suppress hydrogen bonding with the glycerol or employing water instead of glycerol as the solvent, would make for interesting comparisons with these results. More broadly these experiments have shown how XPCS can be an effective method to probe nanoparticle dynamics under nanoconfinement in matrices where alternative techniques are unfeasible. Future XPCS studies that explore this capability further would be valuable.

## VII. ACKNOWLEDGEMENTS

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- [1] I. L. Molnar, W. P. Johnson, J. I. Gerhard, C. S. Willson, and D. M. O’Carroll, *Water Resour. Res.* **51**, 6804 (2015).
  - [2] O. A. Vasilyev, S. Dietrich, and S. Kondrat, *Soft Matter* **14**, 586 (2018).
  - [3] F. Babayekhorasani, D. E. Dunstan, R. Krishnamoorti, and J. C. Conrad, *Soft Matter* **12**, 8407 (2016).
  - [4] P. S. Burada, P. Hänggi, F. Marchesoni, G. Schmid, and P. Talkner, *ChemPhysChem* **10**, 45 (2009).
  - [5] S. Faez, Y. Lahini, S. Weidlich, R. F. Garmann, K. Wondraczek, M. Zeisberger, M. A. Schmidt, M. Orrit, and V. N. Manoharan, *ACS Nano* **9**, 12349 (2015).
  - [6] R. Raccis, A. Nikoubashman, M. Retsch, U. Jonas, K. Koynov, H.-J. Butt, C. N. Likos, and G. Fytas, *ACS Nano* **5**, 4607 (2011).
  - [7] J. D. C. Jacob, K. He, S. T. Retterer, R. Krishnamoorti, and J. C. Conrad, *Soft Matter* **11**, 7515 (2015).
  - [8] M. J. Skaug, L. Wang, Y. Ding, and D. K. Schwartz, *ACS Nano* **9**, 2148 (2015).
  - [9] S. Fringes, F. Holzner, and A. W. Knoll, *Beilstein J. Nanotechnol.* **9**, 301 (2018).
  - [10] C. Giraudet, M. S. G. Knoll, Y. Galvan, S. Süß, D. Segets, N. Vogel, M. H. Rausch, and A. P. Fröba, *Transp. Porous Media* **131**, 723 (2020).
  - [11] H. Wu and D. K. Schwartz, *Acc. Chem. Res.* **53**, 2130 (2020).
  - [12] T. J. Woehl and T. Prozorov, *J. Phys. Chem. C* **119**, 21261 (2015).
  - [13] A. Verch, M. Pfaff, and N. de Jonge, *Langmuir* **31**, 6956 (2015).
  - [14] S. W. Chee, Z. Baraissov, N. D. Loh, P. T. Matsudaira, and U. Mirsaidov, *J. Phys. Chem. C* **120**, 20462 (1985).
  - [15] A. S. Powers, H.-G. Liao, S. N. Raja, N. D. Bronstein, A. P. Alivisatos, and H. Zheng, *Nano Lett.* **17**, 15 (2017).
  - [16] X. Tian, H. Zheng, and U. Mirsaidov, *Nanoscale* **9**, 10044 (2017).



- [17] M. N. Yesibolati, K. I. Mortensen, H. Sun, A. Brostrøm, S. Tidemand-Lichtenberg, and K. Mølhave, *Nano Lett.* **20**, 7108 (2020).
- [18] V. Jamali, C. Hargus, A. Ben-Moshe, A. Aghazadeh, H. D. Ha, K. K. Mandadapu, and A. P. Alivisatos, *Proc. Natl. Acad. Sci. U. S. A.* **118**, e2017616118 (2021).
- [19] R. L. Leheny, *Curr. Opin. Colloid Interface Sci.* **17**, 3 (2012).
- [20] N. Jiang, M. K. Endoh, and T. Koga, *Polym. J.* **45**, 26 (2013).
- [21] L. Cristofolini, *Curr. Opin. Colloid Interface Sci.* **19**, 228 (2014).
- [22] B. Ruta, O. Czakkel, Y. Chushkin, F. Pignon, R. Nervo, F. Zontone, and M. Rinaudo, *Soft Matter* **10**, 4547 (2014).
- [23] T. Koga, C. Li, M. K. Endoh, J. Koo, M. Rafailovich, S. Narayanan, D. R. Lee, L. B. Lurio, and S. K. Sinha, *Phys. Rev. Lett.* **104**, 066101 (2010).
- [24] T. Koga, N. Jiang, P. Gin, M. K. Endoh, S. Narayanan, L. B. Lurio, and S. K. Sinha, *Phys. Rev. Lett.* **107**, 225901 (2011).
- [25] K. J. Johnson, E. Glynos, S.-D. Maroulas, S. Narayanan, G. Sakellariou, and P. F. Green, *Macromolecules* **50**, 7241 (2017).
- [26] A. Grein-Iankovski, I. C. Riegel-Vidotti, F. F. Simas-Tosin, S. Narayanan, R. L. Leheny, and A. R. Sandy, *Soft Matter* **12**, 9321 (2016).
- [27] J. Lee, A. Grein-Iankovski, S. Narayanan, and R. L. Leheny, *Macromolecules* **50**, 406 (2017).
- [28] H. Guo, G. Bourret, R. B. Lennox, M. Sutton, J. L. Harden, and R. L. Leheny, *Phys. Rev. Lett.* **109**, 055901 (2012).
- [29] K. Nygård, J. Buitenhuis, M. Kagias, K. Jefimovs, F. Zontone, and Y. Chushkin, *Phys. Rev. Lett.* **116**, 167801 (2016).
- [30] V. Chaudhary and S. Sharma, *J. Porous Mater.* **24**, 741 (2017).
- [31] D. Zhao, J. Feng, Q. Huo, N. Melosh, G. H. Fredrickson, B. F. Chmelka, and G. D. Stucky, *Science* **279**, 548 (1998).
- [32] N. Brodie-Linder, G. Dosseh, C. Alba-Simonesco, F. Audonnet, and M. Impérator-Clerc, *Mater. Chem. Phys.* **108**, 73 (2008).
- [33] R. Bardestani, G. S. Patience, and S. Kaliaguine, *Can. J. Chem. Eng.* **97**, 2781 (2019).
- [34] M. Kruk and L. Cao, *Langmuir* **23**, 7247 (2007).
- [35] M. Jaroniec and L. A. Solovyov, *Langmuir* **22**, 6757 (2006).

- [36] R. Mhanna, A. Hamid, S. Dutta, R. Lefort, L. Noirez, B. Frick, and D. Morineau, J. Chem. Phys. **146**, 024501 (2017).
- [37] A. Madsen, R. L. Leheny, H. Guo, M. Sprung, and O. Czakkel, New J. Phys. **12**, 055001 (2010).
- [38] W. M. Slie and W. M. Madigosky, J. Chem. Phys. **48**, 2810 (1968).
- [39] A. G. Ferreira, A. P. Egas, I. M. Fonseca, A. C. Costa, D. C. Abreu, and L. Q. Lobo, J. Chem. Thermodyn. **113**, 162 (2017).
- [40] F. Livet, F. Bley, F. Ehrburger-Dolle, I. Morfin, E. Geissler, and M. Sutton, J. Synchrotron Radiat. **13**, 453 (2006).
- [41] R. Hernández, A. Nogales, M. Sprung, C. Mijangos, and T. A. Ezquerra, J. Chem. Phys. **140**, 024909 (2014).
- [42] A. Ambari, B. Gauthier-Manuel, and E. Guyon, Phys. Fluids **28**, 1559 (1985).
- [43] J. Happel and H. Brenner, *Low Reynolds number hydrodynamics* (Springer Netherlands, 1983).
- [44] J. Hohlbein, M. Steinhart, C. Schiene-Fischer, A. Benda, M. Hof, and C. Hübner, Small **3**, 380 (2007).
- [45] M. Alcoutlabi and G. B. McKenna, J. Phys.: Condens. Matter **17**, R461 (2005).
- [46] R. Richert, Annu. Rev. Phys. Chem. **62**, 65 (2011).
- [47] R. Busselez, R. Lefort, Q. Ji, F. Affouard, and D. Morineau, Phys. Chem. Chem. Phys. **11**, 11127 (2009).
- [48] O. Trofymuk, A. A. Levchenko, and A. Navrotsky, J. Chem. Phys. **123**, 194509 (2005).
- [49] W. Zheng and S. L. Simon, J. Chem. Phys. **127**, 194501 (2007).
- [50] D. Kilburn, P. E. Sokol, V. García Sakai, and M. Ashraf Alam, Appl. Phys. Lett. **92**, 033109 (2008).
- [51] D. Morineau, Y. Xia, and C. Alba-Simionesco, J. Chem. Phys. **117**, 8966 (2002).
- [52] A. A. Levchenko, P. Jain, O. Trofymuk, P. Yu, A. Navrotsky, and S. Sen, J. Phys. Chem. B **114**, 3070 (2010).