

A YARN-BASED BACTERIA-POWERED BATTERY FOR SMART TEXTILES

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ABSTRACT

We report a flexible and wearable bacteria-powered battery in which four functional yarns are placed in parallel for biological energy harvesting. A current collecting yarn is sandwiched between two conductive/hydrophilic active yarns including electricity-generating bacteria while a polymer-passivated cathodic yarn is located next to one of the active yarns to form a biological fuel cell configuration. The device uses *Shewanella oneidensis* MR-1 as a biocatalyst to produce a maximum power of $17\mu\text{W}/\text{cm}^3$ and current density $327\mu\text{A}/\text{cm}^3$, which are enough to power small-power applications. This yarn-structured biobattery can be potentially woven or knitted into an energy storage fabric to provide a higher power for smart textiles. Furthermore, sweat generated from the human body can be a potential fuel to support bacterial viability, providing the long-term operation of the battery.

INTRODUCTION

Textile-based wearable electronics have recently emerged as a technology that promises next generation, ubiquitous health monitoring [1, 2]. Even under extreme mechanical deformation, their flexible and stretchable characteristics allow conformal and reliable contact with asymmetrical and non-uniform human skin with minimum discomfort. However, there has been a significant challenge in creating a truly self-reliant and stand-alone wearable sensing system that does not rely on an external power source [3]. Recently, our group, for the first time, demonstrated a textile-based bacteria-powered biobattery and ensured its practical efficacy as a flexible and stretchable power supply [4, 5]. Bacteria-powered biobatteries are arguably the most underdeveloped for wearable electronic applications because microbial cytotoxicity may pose health concerns. However, if we consider that humans possess many bacterial cells more than human cells in their bodies, the direct use of bacterial cells as a power resource interdependently with the human body is conceivable for wearable electronics. Our previous research efforts in flexible bacteria-powered batteries focused on integrating electronic components into a single sheet of 2-D fabric through a top-down approach [4, 5].

In this work, we presented a yarn-based biobattery for flexible and wearable textile integration (Figure 1). The realization of biobattery on flexible substrates, such as paper or textile, has been mentioned in literature [4-6]. Unlike the prior arts where functional regions and parts are constructed directly on the substrate, direct functionalization of basic yarns can enable more flexible and controllable device fabrication in a smaller device footprint, enhance the versatility of the device design and improve performance per unit volume. This bacteria-powered energy harvester was built on 1-D yarns through a bottom-up approach from which various electronic components of a large textile can be woven or knitted.

MATERIALS AND EXPERIMENTAL SET-UP

Device Fabrication

The anodic compartment consisted of three yarns; a conductive thread as a current collector and two PEDOT:PSS

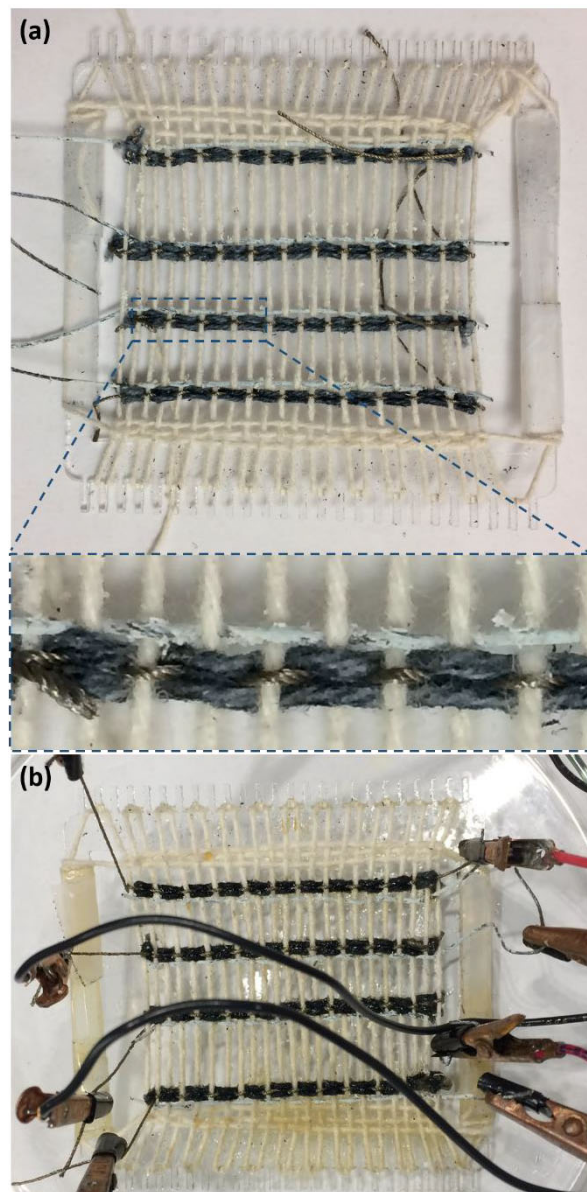


Figure 1. (a) Photo-images showing the textile weaving process of four yarn-based biobatteries and the enlarged view of the interlaced structure of one biobattery with four functional yarns. (b) Operating biobatteries.

(poly(3,4-ethylenedioxythiophene) polystyrene sulfonate) functionalized cotton yarns (or active yarns) were placed in parallel shown in (Figure 1 & Figure 2). The cathodic yarn was based on the same conductive thread as the anode. The silver-based cathodic paste was first applied the cathodic yarn and a microporous polytetrafluoroethylene (PTFE) membrane as a proton exchange membrane was coated over the cathode yarn.

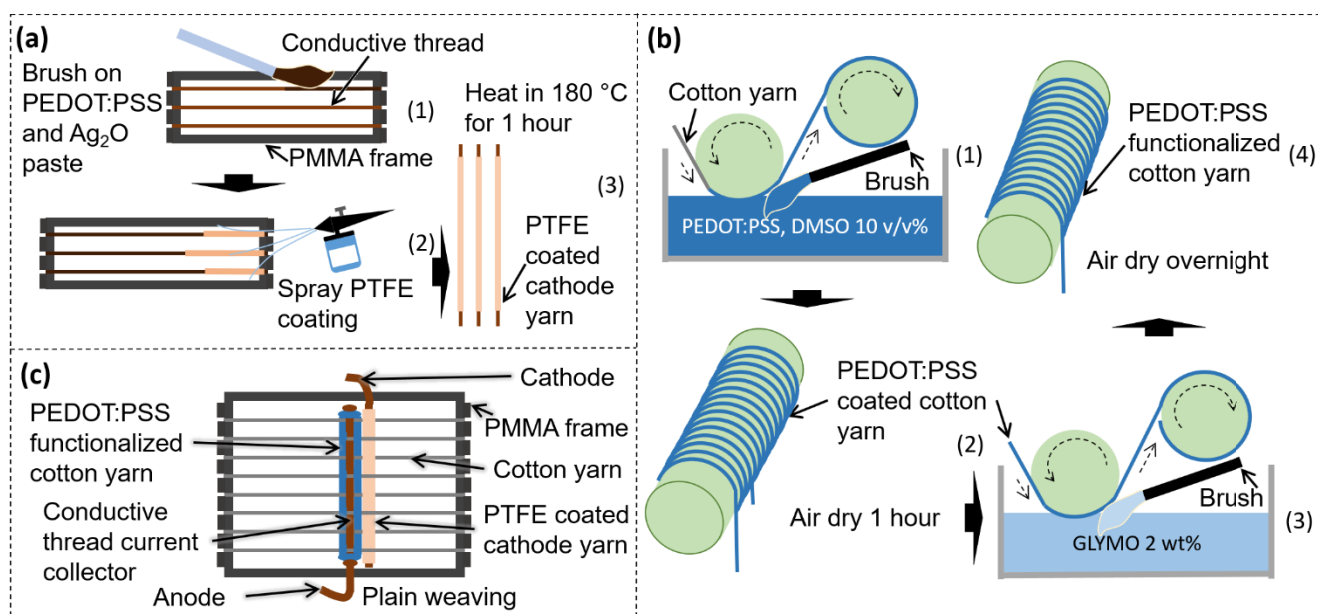


Figure 2: Schematic illustration of the processes employed to fabricate yarn-based biobattery, (a) cathodic yarns, (b) anodic yarns and (c) woven textile with anode and cathode in weft direction.

Four functional yarns were woven into a plain weaving pattern (Figure 2) as the prototype yarn-based biobattery (Figure 3).

Cathodic yarn fabrication

Commercial cotton yarns were first sterilized by immersing in 70 v/v % ethanol with sonication for 30min, followed by drying in a ventilated oven at 50°C. The silver-based cathodic paste was prepared by mixing silver (I) oxide (Sigma-Aldrich) with 1wt% PEDOT:PSS. The PEDOT:PSS weight percentage increased from 1 wt% to 2 wt% by further heating and stirring processes on a hot plate stirrer (60°C with 110 rpm). After the pre-cleaned conductive threads were stretched on the poly(methyl methacrylate) (PMMA) frame, the cathodic paste was brushed onto the thread and air-dried (Figure 2a). To passivate the treated cathodic yarn, 60 wt% PTFE water emulsion was mixed with isopropyl alcohol and sonicated for 30 min, yielding a uniform 40 wt% PTFE emulsion. The emulsion was sprayed onto the conductive cathodic thread and dried at 180°C for 1 h.

Anodic yarn fabrication

Conductive ink was prepared by mixing 1 wt% PEDOT:PSS (Clevios™ PH1000, Heraeus) with dimethyl sulfoxide (DMSO, Sigma-Aldrich) at a volume ratio of 9:1. The cleaned cotton yarns were pulled through PEDOT:PSS conductive ink via a customized reeling setup (Figure 2b). A brush was pressed on the cotton yarns to facilitate the conductive ink penetration. The conductive cotton yarns were air-dried for 1 h to allow the solvent to evaporate. Then, the yarns were treated with 2 wt% 3-glycidoxypentyltrimethoxysilane (GLYMO, Sigma-Aldrich) to improve its hydrophilic property. The functionalized yarns were further air-dried overnight.

Weaving of biobatteries

The cleaned non-conductive threads were used as warp yarns and held stationary in the PMMA frame (Figure 2c). The anodic

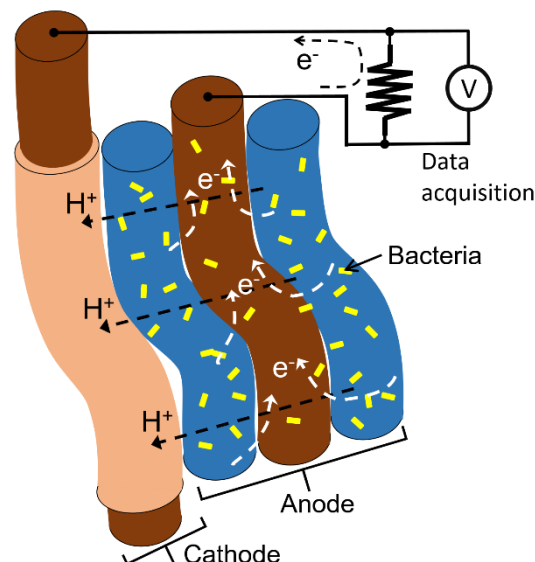


Figure 3: Schematic diagram of the yarn-based biobattery.

and cathodic yarns were weaved through the warps in a plain pattern (Figure 3).

Inoculum

Cryostock *Shewanella oneidensis* MR-1 was cultured in 3 wt% soy broth media for 24 h, and diluted with fresh media. Growth was monitored by measuring the optical density at 600nm (OD_{600}), and the culture reached an OD_{600} of 1.6.

Operating principle and measurement setup

The yarn-based biobatteries were connected to a National Instrument (NI) data acquisition module (USB-6212) with a customized user interface for voltage measurements. To activate the biobattery, 260μL of bacterial inoculum was pipetted to each

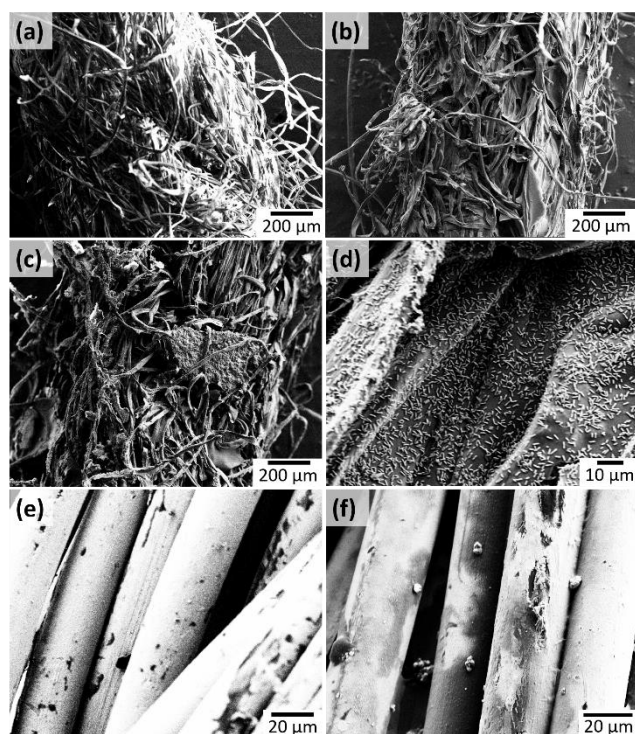


Figure 4: SEM images of (a) the cotton yarn without treatment, (b) the PEDOT:PSS functionalized anodic cotton yarn, (c & d) the anodic yarns after inoculation, (e) the conductive cathodic yarn and (f) the cathodic yarn with PTFE coating.

device. Resistive loads were connected to the biobatteries, and the output current and power were calculated via Ohm's law.

RESULTS AND DISCUSSION

The cotton yarn (Figure 4a) was chosen because of its good hydrophilicity, resistance to organic solvents, fabric integrity at elevated temperatures and low cost [7]. Adding the conductive polymer PEDOT:PSS to the cotton yarn created a conductive matrix with the high specific surface, improving bacterial attachment and their respiration (Figure 4b, 4c & 4d) [5, 8]. Therefore, the treatment for the conductive yarn engineering can reduce the anodic and cathodic resistance and minimize the ohmic loss of the device, increasing the performance (Figure 4e) [4]. Our prior arts demonstrated great potential of the silver (I) oxide with PEDOT:PSS as a stable- and high-preforming cathodic material for flexible biobatteries [5]. To encapsulate the silver-based cathode and promote ion-exchange between the anode and the cathode, a PTFE was selected to form a microporous proton exchange membrane [9]. The cathodic yarn was passivated with the homogenous PTFE membrane at 180 °C (Figure 4f).

When *S. oneidensis* MR-1 inoculum were introduced to the anodic yarns (Figure 3), the capillary force of the hydrophilic yarns rapidly wicked the anolyte to fill the anodic yarn of the biobattery. While the conductive PEDOT:PSS functionalized anodic yarns harvested electrons from bacterial respiration, the anodic conductive thread provided an efficient electrical pathway through which electrons flowed to the external circuit and reached the cathode. The protons produced by bacterial metabolism

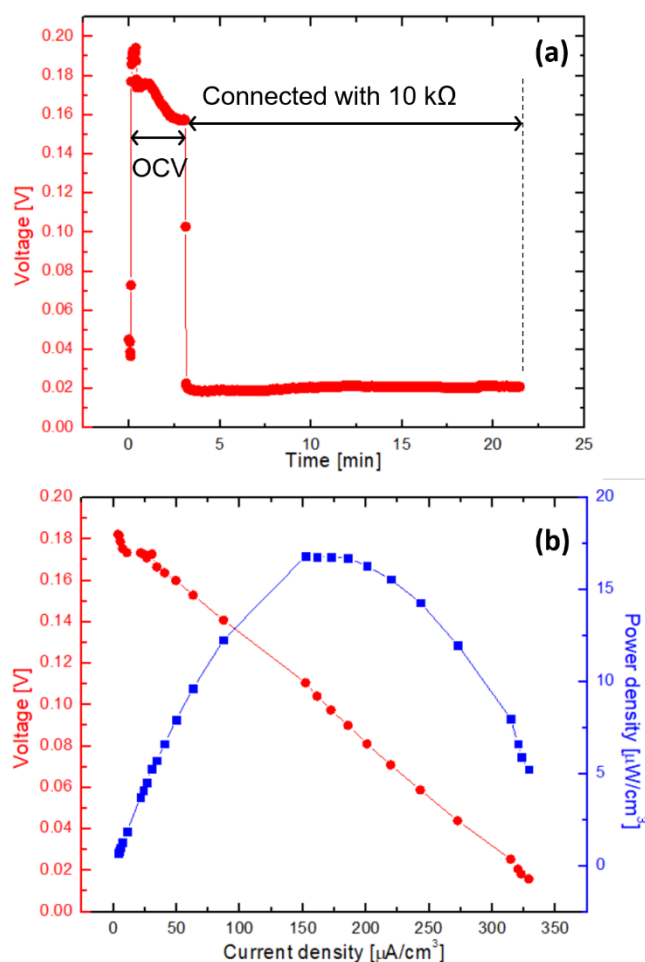


Figure 5: (a) Output voltage measured from a single yarn-based biobattery with or without the external resistor and (b) polarization curve/output power as a function of current.

traveled to the cathodic yarns through the passivated PTFE membrane. The electrons and protons moved from the anodic yarns finally reduced Ag_2O to Ag at the cathodic yarn.

The open circuit voltage (OCV) of 168 mV on average was obtained from the yarn-based biobatteries. A stable current of 2.2 μA was produced with a 10 kΩ resistor between the anode and cathode, providing a proof-of-concept demonstration of the bacterial battery (Figure 5a). The low OCV could be addressed by serially connecting the individual cells with various easy-to-configure threading or weaving patterns. A cell polarization curve and power output of the battery were measured and obtained based on the saturated current value at a given external resistance (Figure 5b). Slight bending of the textile did not affect the device performance. However, further detailed stretching and bending tests are needed to optimize the device fabrication and configuration. After the device operation, some parts of the PTFE membrane were slightly peeled off from the cathodic yarn (Figure 4f). This exposure of cathode would cause potential releasing of hazardous antibacterial silver ions, cathode biofouling, or short-circuit between anode and cathode, all of which would negatively affect the biobattery performance. Fabrication improvement of the PTFE membrane would be necessary, and further studies regarding mechanical strength, flexibility, and porosity of the

membrane are required. Our yarn-based biobattery will be suitable for wearable sensors, such as wrist health monitors [10], textile integrated electrochemical sensors [11], flexible and economical point-of-care diagnostic devices [12].

CONCLUSION

This work demonstrated the first integration of the bacterial-powered biobattery into yarns with a simple, scalable and fabrication-friendly fashion. The biobattery was built on 1-D yarns through a bottom-up approach, which can be potentially woven into a large energy storage fabric to ultimately deliver high power for practical, flexible applications.

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