

RESEARCH ARTICLE

Diet mediates thermal performance traits: implications for marine ectotherms

Emily A. Hardison*, Krista Kraskura, Jacey Van Wert, Tina Nguyen and Erika J. Eliason

ABSTRACT

Thermal acclimation is a key process enabling ectotherms to cope with temperature change. To undergo a successful acclimation response, ectotherms require energy and nutritional building blocks obtained from their diet. However, diet is often overlooked as a factor that can alter acclimation responses. Using a temperate omnivorous fish, opaleye (*Girella nigricans*), as a model system, we tested the hypotheses that (1) diet can impact the magnitude of thermal acclimation responses and (2) traits vary in their sensitivity to both temperature acclimation and diet. We fed opaleye a simple omnivorous diet (*ad libitum* *Artemia* sp. and *Ulva* sp.) or a carnivorous diet (*ad libitum* *Artemia* sp.) at two ecologically relevant temperatures (12 and 20°C) and measured a suite of whole-animal (growth, sprint speed, metabolism), organ (cardiac thermal tolerance) and cellular-level traits (oxidative stress, glycolytic capacity). When opaleye were offered two diet options compared with one, they had reduced cardiovascular thermal performance and higher standard metabolic rate under conditions representative of the maximal seasonal temperature the population experiences (20°C). Further, sprint speed and absolute aerobic scope were insensitive to diet and temperature, while growth was highly sensitive to temperature but not diet, and standard metabolic rate and maximum heart rate were sensitive to both diet and temperature. Our results reveal that diet influences thermal performance in trait-specific ways, which could create diet trade-offs for generalist ectotherms living in thermally variable environments. Ectotherms that alter their diet may be able to regulate their performance at different environmental temperatures.

KEY WORDS: Temperature, Thermal acclimation, Fish, Thermal limits, *Girella nigricans*, Omnivore

INTRODUCTION

Understanding the full range and maximum capacity of ectotherm physiological responses to environmental change is essential to predict species' vulnerability to global climate change (Huey et al., 2012; Somero, 2011; Stillman, 2003). Temperature is a critical environmental factor governing ectotherm physiology, behavior and ecology (Brett, 1971). The current paradigm suggests that ectotherms have three options when faced with unfavorable temperatures: they can move to a more suitable habitat, adapt

over multiple generations or acclimate to the new conditions (Daufresne et al., 2009; Glanville and Seebacher, 2006; Hofmann and Todgham, 2010; Somero, 2011). Thermal acclimation is an essential survival mechanism for ectotherms living in variable environments and a critical coping mechanism against global climate change (Bernhardt and Leslie, 2013; Jackson et al., 2021; Seebacher et al., 2015). During thermal acclimation, ectotherms undergo reversible phenotypic changes that improve their performance at a given temperature (Fig. 1; e.g. enzyme activity, membrane composition, mitochondrial density, oxygen transport, organ morphology and function; Anttila et al., 2014; Chung and Schulte, 2020; Ekström et al., 2016; Little et al., 2020a; Seebacher et al., 2015). It is often assumed that ectotherms will achieve the same level of performance after repeated exposures to a temperature, as long as all other environmental conditions (i.e. salinity, pH, dissolved oxygen) are held the same (Sinclair et al., 2016). To undergo a successful acclimation response, ectotherms require energy and nutritional building blocks obtained from their diet. Diets vary considerably in nutritional and energetic content, which suggests that different diets may mediate distinct thermal acclimation responses (Fig. 1).

Food quality and availability can change seasonally, with global climate change and across habitats (Alton et al., 2020; Arnold et al., 2010; Birnie-Gauvin et al., 2017; Ho et al., 2010). Many ectotherms are also generalists and vary their diet to meet their nutritional requirements or maximize energy-use efficiency (Jobling, 2016; Johnson et al., 2017; Kaiser and Hughes, 1993; Raubenheimer et al., 2005; Rubio et al., 2003, 2009; Sánchez-Vázquez et al., 1998). Some ectotherms also change their diet with temperature (Boersma et al., 2016a; Carreira et al., 2016; Jang et al., 2015; Rho and Lee, 2017; Schmitz and Rosenblatt, 2017; Vejříková et al., 2016). For example, multiple omnivorous fishes consume higher proportions of algae as water temperatures increase (e.g. Behrens and Lafferty, 2012; Emde et al., 2016; González-Bergonzoni et al., 2016; Guinan et al., 2015; Prejs, 1984). The exact reasons for these diet shifts are unknown, with some suggesting that the optimal dietary protein to carbohydrate ratio for ectotherms differs across temperatures (Lee et al., 2015; Rho and Lee, 2017; Zhang et al., 2020), or that cold temperature constrains the digestive physiology of herbivores and omnivores (Floeter et al., 2005; González-Bergonzoni et al., 2012). These proposed explanations hint at a broader hypothesis: that omnivores consume different proportions of plants and animals to regulate their physiological responses to changing temperatures. More broadly, any changes in an ectotherm's diet that coincide with a change in environmental temperature (through differences in dietary preference, availability or nutrient composition) might alter its thermal performance.

To understand the interaction between diet and temperature, we must consider how traits critical to survival may be differentially affected (Fig. 1). Measuring thermal limits in conjunction with vital biological rates provides comprehensive insight into ectotherm

Department of Ecology, Evolution and Marine Biology, University of California, Santa Barbara, Santa Barbara, CA 93106, USA.

*Author for correspondence (emilyhardison@ucsb.edu)

© E.A.H., 0000-0002-3668-5035; K.K., 0000-0001-9225-3037; J.V., 0000-0001-7262-9420; E.J.E., 0000-0002-0120-7498

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.

Received 10 May 2021; Accepted 1 October 2021

List of symbols and abbreviations

ABT	Arrhenius breakpoint temperature
CT _{max}	critical thermal maxima
CT _{min}	critical thermal minima
f_H	heart rate
$f_{H,max}$	maximum overall heart rate
$f_{H,min}$	minimum overall heart rate
MMR	maximum metabolic rate
SMR	standard metabolic rate
T_{AB}	breakpoint temperature on the heart
T_{arr}	temperature at the first cardiac arrhythmia
T_{peak}	temperature corresponding to maximum heart rate
TL	total length
TPC	thermal performance curve

thermal biology in variable and changing environments (Magozzi and Calosi, 2015). A common assumption in thermal biology is that biological rates have the same thermal sensitivity and that aerobic capacity and baseline metabolism can be used as proxies for many performance traits (Fry, 1947; Brett, 1971; Claireaux and Lefrançois, 2007; Pörtner, 2001, 2010; for a critique, see Clark et al., 2013; Schulte, 2015). However, there is growing support for a multiple-performance, multiple-optima model, which states that thermal sensitivity differs across biological rates (i.e. absolute aerobic scope, standard metabolic rate, growth rate, sprint speed) and is not always predictable based on aerobic capacity or baseline metabolism (Clark et al., 2013; Dell et al., 2011; Kellermann et al., 2019; Seebacher et al., 2015). For example, Healy and Schulte (2012) demonstrated that specific growth rate was negative at temperatures where absolute aerobic scope (i.e. maximum–standard metabolic rate) was maximal in killifish. This model has been challenging to test empirically as few performance traits are usually

measured per study and these traits are often considered separately from upper and lower thermal limits (Magozzi and Calosi, 2015). If diet and temperature together influence biological rates and thermal limits, ectotherms may be incentivized to make diet choices that improve their thermal responses. However, if fitness-enhancing traits are differentially affected by diet and temperature, there could be important performance consequences associated with an ectotherm's ultimate diet choices.

Opaleye (*Girella nigricans*) are temperate omnivorous fish that consume a greater proportion of algae in warmer water across their geographic range (Behrens and Lafferty, 2012), which makes them an ideal model for exploring diet effects on thermal acclimation responses. Here, we tested the hypothesis that when offered a simple choice omnivorous diet (*ad libitum* *Artemia* sp. and *Ulva* sp.) versus a carnivorous diet (*ad libitum* *Artemia* sp. only) at two ecologically relevant temperatures (12 and 20°C), opaleye would make diet choices that altered their thermal acclimation responses in trait-specific ways. As juveniles, opaleye live in the intertidal zone where they face many challenges, including escaping predators, maintaining growth rates and dealing with high daily thermal variation (Somero, 2010). Therefore, we adopted an integrative approach and assessed the opaleye's thermal acclimation responses at the whole-animal (growth, sprint speed, metabolism, critical thermal limits), organ (cardiac thermal limits) and cellular (glycolytic capacity, oxidative stress) levels to compare ecologically and physiologically relevant performance traits and thermal limits for the fish in their juvenile life stage and identify any trade-offs associated with the treatment diets. We hypothesized that biological rates would increase with temperature but have different thermal sensitivities depending on the diet treatment. Specifically, we predicted that there would be costs to consuming the omnivorous diet (e.g. higher digestive infrastructure costs resulting in higher maintenance metabolism or reduced growth from lower protein diet) that would be offset by increases in the performance of other traits (e.g.

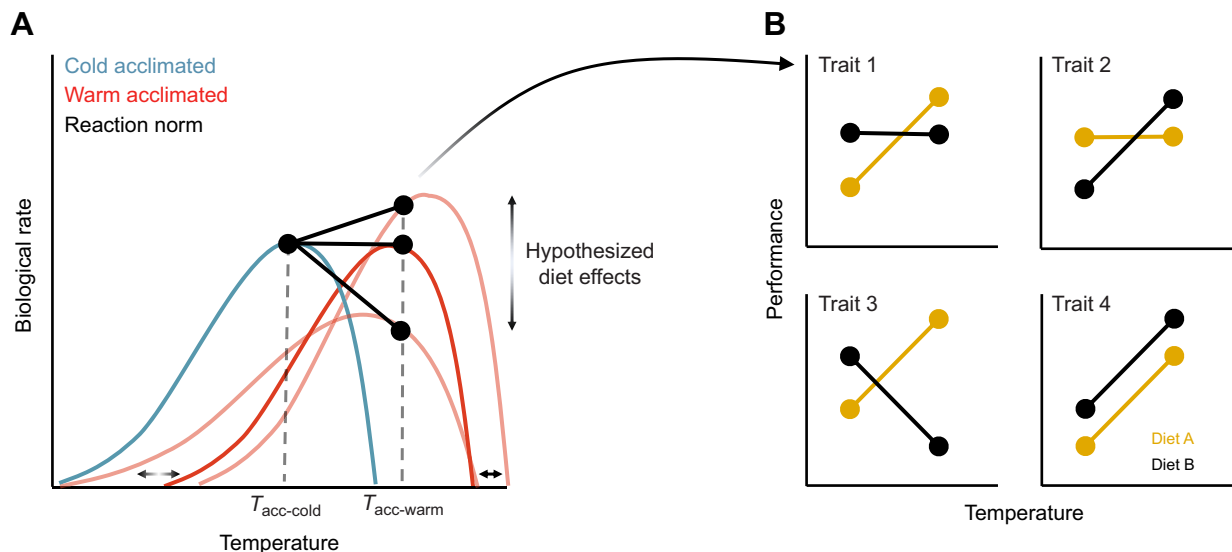


Fig. 1. Conceptual graphs illustrating how diet may affect thermal acclimation responses and how those effects could be trait specific. (A) Diet can affect thermal performance. The graph shows how acute thermal performance curves (TPC) shift towards the acclimation temperature (T_{acc}), where blue is the acute TPC after acclimation to cold and red is the acute TPC after acclimation to warm conditions. Note that both cold and warm acclimation may be affected by diet, but only potential effects of warm acclimation are displayed for simplicity. Diet may influence the shape (height and breadth) of those acute TPC (indicated by pink curves) or the location of the curve along the x-axis (i.e. temperature of peak performance). These effects could influence the slope of the line between reaction norms (performance at acclimation conditions, indicated by black lines). (B) Diet and temperature may interact and have trait-specific effects. The graphs are a series of hypothetical reaction norm plots for various traits. These traits can have different diet and temperature sensitivities, which could create performance trade-offs for ectotherms consuming different diets.

thermal limits, sprint performance, glycolytic capacity, oxidative stress).

MATERIALS AND METHODS

Fish collection

Juvenile opaleye, *Girella nigricans* (Ayres 1860), were collected in spring 2019 [experiment 1: respirometry, sprint, growth, critical thermal maxima (CT_{max}/CT_{min}); $N=144$, mean $\pm$ s.d. body mass (BM) 14.75 ± 3.53 g and total length (TL) 9.42 ± 0.76 cm] and in winter 2020 (experiment 2: Arrhenius breakpoint test; $N=126$, BM 19.5 ± 6.1 g and TL 10.5 ± 1.1 cm) by seine or hook and line from Santa Barbara Harbor, CA, USA (34.40829, -119.691389). Fish were transported in coolers ($>70\%$ air saturation) to the University of California, Santa Barbara and held in 95 l fiberglass flow-through seawater tanks (9–12 fish per tank). Prior to the start of acclimation, fish were held at ambient conditions (mean $\pm$ s.d. experiment 1: $13.9\pm 1.1^\circ\text{C}$ and experiment 2: $16.2\pm 0.6^\circ\text{C}$) and fed *ad libitum* omnivorous diets (*Ulva* sp. and *Artemia* sp.). All protocols were approved by the Institutional Animal Care and Use Committee at the University of California, Santa Barbara.

Acclimation and diet treatments

Fish were randomly assigned to one of two ecologically relevant temperatures (12 and 20°C , representative of the low and high seasonal temperatures experienced in Santa Barbara, CA, USA; Fig. 2) and fed one of two *ad libitum* diets (omnivorous: *Artemia* sp. and *Ulva* sp.; and carnivorous: *Artemia* sp.) in a factorial design with 3–4 replicate tanks per treatment. *Ulva* sp. (collected by hand from Goleta Beach, Goleta, CA, USA) and *Artemia* sp. (brineshrimpdirect.com) were replaced every morning. Diets were selected based on stomach content information from Barry and Ehret (1993), which demonstrated that both *Ulva* sp. and small crustaceans constitute a significant portion of wild opaleye diets, and Behrens and Lafferty (2007), which used *Ulva* sp. as a representative herbivorous diet for a lab study on opaleye. Preliminary data demonstrated that *Ulva* sp.-only diets were not nutritionally sufficient. Dietary proximate analysis is provided in Table S1.

Food consumption rates were assessed during preliminary trials. All tanks were fasted for 24 h, and then the fish were offered their

pre-weighed diet treatment for 1 h. The remaining *Artemia* sp. and *Ulva* sp. were removed and weighed. However, we were unable to obtain any measurable estimates of *Ulva* sp. consumption during these brief 1 h trials. At 20°C , *Artemia* sp. consumption was $\sim 23\%$ lower in the omnivorous treatment (7.39% body mass) than in the carnivorous treatment (9.58% body mass), suggesting that the fish in the omnivorous treatment were supplementing their diet with *Ulva* sp. This is consistent with our visual observations that the fish readily consumed *Ulva* sp. in the warm treatment (though they consistently ate more *Artemia* sp. compared with *Ulva* sp.). In contrast, at 12°C , *Artemia* sp. consumption was only $\sim 10\%$ lower in the omnivorous treatment (3.65% body mass) compared with the carnivorous (4.07% body mass) and we did not observe the fish consuming *Ulva* sp. at this temperature. This suggests that the fish either were not consuming *Ulva* sp. or were doing so in small amounts at the cold acclimation treatment. Further, opaleye ate less at 12°C compared to 20°C . Overall, *Artemia* sp. consumption was $\sim 55\%$ lower in the cold treatment compared with the warm.

Temperature and dissolved oxygen content were monitored 1–2 times daily by hand using a Digi-sense Traceable Single RTD thermometer (Cole Palmer, IL, USA) and an OxyGuard handy Polaris 2 dissolved oxygen meter (OxyGuard International A/S, Farum, Denmark). Oxygen was maintained at $>80\%$ air saturation throughout the study. The average temperature per treatment was 20.0 ± 0.4 and $12.2\pm 0.4^\circ\text{C}$ in the two experiments (mean $\pm$ s.d.; determined from in-tank Thermochron 4 K iButtons programmed to record every 20 min). Fish were acclimated to treatment conditions for 3 weeks prior to experimentation (14 h:10 h light:dark cycle). All individuals were fasted for 36–40 h prior to respirometry, thermal limit and sprint testing. All tests were performed at acclimation temperatures unless otherwise noted.

Intermittent flow respirometry

Respirometry was conducted using 12 respirometers in one of three sizes: 349, 579 and 711 ml. Water was flushed and recirculated through the chamber at a rate of 2.5 l min^{-1} (Eheim Universal 300 pumps, Eheim, Germany). Dissolved oxygen was measured continuously in each respirometer using a robust oxygen probe and Firesting optical oxygen meter (Pyroscience, Germany).

Fish were transferred with minimal air exposure (<10 s) to a cylindrical chase tank (20 l), where they were chased by hand for 5 min and then immediately placed in the respirometers to obtain an estimate of their maximum metabolic rate (MMR) (this chase protocol was the most effective at eliciting MMR in opaleye; data not shown). Chases occurred between 10:30 h and 14:30 h and were followed by ~ 20 h of automated measurement cycles (15 min total flush/recirculation cycle). Tanks were covered in shade cloth to minimize potential disturbance. Fish were held at either 20.1 ± 0.6 or $11.8\pm 0.4^\circ\text{C}$ (mean $\pm$ s.d.) across all tests. Background respiration was measured before and after each test for ≥ 3 full measurement cycles.

After 20 h in the respirometers, fish were removed and anesthetized in 80 mg l^{-1} MS-222 buffered with 80 mg l^{-1} NaHCO_3 (Sigma Aldrich Co., St Louis, MO, USA). Each fish was weighed (mass in g), measured for TL, and tagged with a unique color code using Visible Implant Elastomer Tags (Northwest Marine Fisheries Inc., Seattle, WA, USA).

Data analysis for respirometry data

All oxygen consumption data were analyzed in R (version 3.5.1) using best practices as outlined in Rosewarne et al. (2016) and Chabot et al. (2016) (<http://www.R-project.org/>). The data were used to

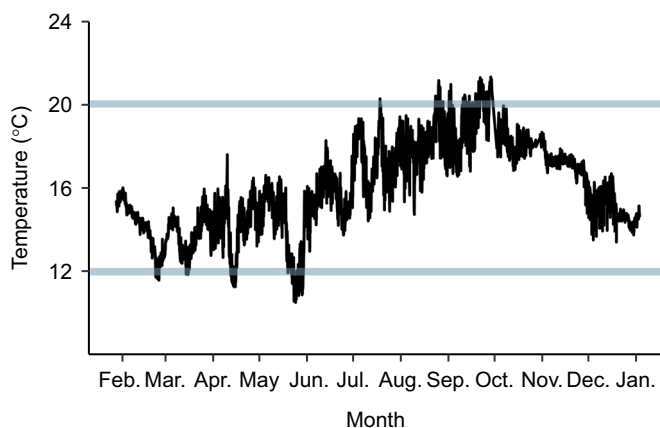


Fig. 2. Temperature data collected in 2019 at Stearns Wharf in Santa Barbara, CA, USA (located next to Santa Barbara Harbor, where fish were collected). The temperature sensor was mounted at 2 m depth and set to record every 4 min. Blue lines indicate treatment temperatures (12 and 20°C). Data source: Washburn, 2021 and Southern California Coastal Ocean Observing System (SCCOOS; https://www.sccoos.org/data/autoss/timeline/?main=single&station=stearns_wharf).

calculate four metabolic rate metrics which define an individual's aerobic energy budget. Standard metabolic rate (SMR) is the baseline metabolic rate needed to survive, and MMR is the maximum rate of energy expenditure. The difference between these two metabolic rates is the absolute aerobic scope (AAS=MMR–SMR), which is representative of the aerobic energy budget a fish has to perform all critical biological processes (Clark et al., 2013). Factorial aerobic scope (FAS=MMR/SMR) is another estimate of the aerobic energy budget and represents the scope for increasing metabolic rate proportional to SMR (Clark et al., 2013). Only fish for which >75% of measurements followed a linear decrease with $R^2 > 0.9$ were included in SMR analysis (i.e. >60 $\dot{M}_{O_2}$ measurements total per fish; $N=6-10$ per treatment). SMR was calculated as the lowest 15% quantile of all recorded measurement cycles (Chabot et al., 2016). MMR was calculated as the steepest 120 s slope during the first measurement period (Little et al., 2020b; $N=6-11$ per treatment). All presented metabolic rate measurements are body mass specific (i.e. presented as $\text{mg O}_2 \text{ kg}^{-1} \text{ min}^{-1}$). We tested for the need to account for body size scaling but given the small size range in the study (8–39 g), we did not find evidence of a scaling relationship when the data were plotted on log–log plots (data not shown). Given the lack of knowledge on a species-specific scaling exponent, we did not scale the metabolism to a common body size using an allometric scaling slope. Background respiration was assessed and, when applicable, a linear regression was fitted between the pre- and post-background measurements and subtracted from the corresponding $\dot{M}_{O_2}$ values (Rodgers et al., 2016; Rosewarne et al., 2016).

Sprint speed

We used a modified protocol based on Kraskura and Nelson (2018). The setup included a custom-built acrylic sprint chamber (128 cm×30 cm×30 cm, L×W×H; water height 10 cm; water volume 41.6 l) with a camera (Canon EOS Rebel T4i) positioned above the sprint chamber to ensure full view of the work area. Each sprint trial was recorded at 60 frames s^{-1} and saved for digital analysis. Fish were placed behind a gated and shaded area in a sprint chamber and habituated to the chamber for 10 min. The gate was removed and the fish were motivated to sprint by manual chasing. Each fish performed ≤ 7 trials with 5 min of recovery between trials. Only trial videos where the fish had undergone a sprint with (1) a straight path of >20 cm, (2) an unobstructed view of 50 cm of the chamber, (3) active and continued bursts, and (4) no back and forth swimming around the chamber were used in subsequent analysis. Temperature was monitored throughout all trials and remained within $\pm 1^\circ\text{C}$ of the test temperature. All videos used in analysis were visualized and validated by ≥ 2 researchers. Videos were tracked in ImageJ/MtrackJ and subsequently analyzed in R to determine the fastest 5 frames of sprinting (~ 10 cm) per trial; 2–3 trials were analyzed per fish and averaged to determine each fish's sprint performance (cm s^{-1}) (Figs S1 and S2; $N=8-10$ per treatment). All fish were given at least 3 weeks of recovery between respirometry or thermal limit experiments and the sprint test.

Thermal limits: $CT_{\max}$ and $CT_{\min}$ testing

We used a standardized critical thermal maxima and minima tests ($CT_{\max}$ and $CT_{\min}$, respectively) as described in Beitinger and Lutterschmidt (2011). $CT_{\max}$ and $CT_{\min}$ represent the most extreme temperatures a fish can survive at for a short period of time. Briefly, 3–4 fish per tank were transferred to the testing tank (78 cm×61 cm×18 cm, L×W×H; water height 12 cm, water volume 57 l) at their treatment temperature with minimal air exposure (<10 s; total $N=8-12$ per treatment). After a brief

habituation period (~ 5 min), the test tank was heated ($CT_{\max}$; 1500 W immersion heater, Process Technology, Willoughby, OH, USA) or cooled ($CT_{\min}$; 1 HP AquaEuro chiller, AquaEuroUSA, Los Angeles, CA, USA) at a rate of $\sim 0.3^\circ\text{C min}^{-1}$ until fish lost the ability to maintain their righting response. Temperature was recorded at the loss of equilibrium and the fish were immediately placed in a recovery tank at an intermediate temperature before being returned to their treatment tanks. All fish fully recovered following the critical thermal tests.

Growth

All fish were weighed and measured for length prior to acclimation and again after 8 weeks under treatment conditions (note that some had been randomly removed for tissue sample collection, see below) to estimate growth across tanks (i.e. average weight gain per week).

Dissections and frozen tissue assays

Fish from each treatment ($N=6$ per treatment) were euthanized by cerebral concussion followed by severing of the spinal cord on day 27–34 of acclimation after the aerobic scope trials were complete. Morphometrics (body mass and TL) were measured, and a white muscle and liver sample were flash frozen in liquid nitrogen and stored at -80°C for future analysis. Stomach and remaining gut contents were emptied and leftover fish remains were stored at -20°C for proximate analysis (see Supplementary Materials and Methods and Table S1).

Lactate dehydrogenase assays

Lactate dehydrogenase activity assays were performed as outlined in Little et al. (2020a) as a proxy for glycolytic capacity in white muscle, where high levels of lactate dehydrogenase activity indicate that the animal has a high capacity to support anaerobic ATP production to fuel burst swimming or hypoxia tolerance (Little et al., 2020a). Briefly, white muscle samples were homogenized in homogenization buffer (0.1% Triton, 50 mmol l^{-1} Hepes, 1 mmol l^{-1} EDTA, pH 7.4 buffer) before being run in triplicate (intra-assay coefficient of variation, CV <15%) on a SpectraMax iD3 Multi-Mode Microplate Reader (Molecular Devices) using a wavelength of 340 nm to measure the disappearance of NADH. The assay was repeated at 8, 12, 20, 26 and 32°C .

Lipid peroxidation assays

Thiobarbituric acid reactive substances were quantified in the liver to estimate lipid peroxidation as a proxy for oxidative stress using a commercially available fluorometric assay kit (Cayman Chemical). Here, higher levels of lipid peroxidation are indicative of greater oxidative damage to cellular components (Castro et al., 2012). Samples were homogenized in RIPA buffer and treated according to the manufacturer's instructions before being run in duplicate at an excitation wavelength of 544 nm and emission wavelength of 590 nm.

Thermal limits: Arrhenius breakpoint temperature test

Arrhenius breakpoint temperature (ABT) tests on the heart were conducted ($N=6-14$ per test and per treatment) as outlined in Casselman et al. (2012) and Gilbert et al. (2020). Briefly, fish were anesthetized in seawater containing 80 mg l^{-1} MS-222 buffered with 1 g l^{-1} NaHCO_3 before being placed ventral side up in an experimental sling in the test tank (10 l seawater containing buffered 65 mg l^{-1} MS-222). Water was circulated past the gills to irrigate them. Stainless Steel Needle Tip Electrodes (ADInstruments Inc., Colorado Springs, CO, USA) were inserted just under the skin to

detect an ECG signal, which was amplified using a Dual Bio Amp amplifier (ADInstruments Inc.) and filtered (filters: 60 Hz Notch filter; mains filter; low pass: 2 kHz; high pass: 10 Hz; range: 2 mV).

After a 30 min equilibration period at the acclimation temperature (Ferreira et al., 2014; Hansen et al., 2017), atropine sulfate was injected intraperitoneally (1.2 mg kg^{-1} in 0.9% NaCl) to block vagal tone followed by isoproterenol ($4 \mu\text{g kg}^{-1}$ in 0.9% NaCl) to maximally stimulate β -adrenoreceptors. Any fish that did not respond to the drug injections or for which experimental error occurred (e.g. water pump failure) were removed from the analysis and not considered further. These drug concentrations were tested prior to experimentation to ensure doubling the concentration did not further increase heart rate (f_H ; beats min^{-1}). Fifteen minutes after isoproterenol injection, water temperature was heated (warm ABT test) or cooled (cold ABT test) at 1°C every 6 min (Polystat recirculating heater/chiller; Cole-Palmer, Vernon Hills, IL, USA). At each 1°C interval, f_H and temperature were stabilized to record a value for f_H . This procedure was repeated until the onset of cardiac arrhythmia (T_{arr}), as indicated by a transition from rhythmic to arrhythmic beating or a missed QRS peak resulting in a precipitous drop in f_H (Casselman et al., 2012) or until the known average CT_{max} for the species (generally $0\text{--}5^\circ\text{C}$ higher than T_{arr} ; Chen et al., 2015; Muñoz et al., 2014; Safi et al., 2019), to ensure that curves could later be fitted to the data for comparisons of acute thermal performance curves across treatments. All fish were immediately euthanized at the end of the test.

Data analysis for ABT tests

All ECG analyses were performed in LabChart software (www.adinstruments.com). f_H was calculated for each temperature increment from 15 continuous seconds of measurements using automated ECG analysis software in LabChart (Gradil et al., 2016). The heart may set upper thermal tolerance temperatures in fish (Anttila et al., 2014; Muñoz et al., 2014). The warm ABT test measures three sublethal thermal limits on cardiac function (T_{AB} , T_{peak} , T_{arr}). Each of these limits indicates transition temperatures where the heart's capacity to transport oxygen, nutrients and immune cells becomes compromised (Anttila et al., 2014; Muñoz et al., 2014). The first thermal limit (T_{AB}) is highly correlated with the thermal optimum for aerobic scope in other teleosts (Anttila et al., 2013; Ferreira et al., 2014). T_{AB} was calculated by performing ABT tests on the rising phase of the thermal performance curve for f_H using *segmented* (v1.1-0; Muggeo, 2008) in R. The temperature corresponding to the breakpoint in f_H was defined as T_{AB} (warm ABT test) or $T_{\text{AB-cold}}$ (cold ABT test). Overall maximum heart rate ($f_{H,\text{max}}$) was defined as the highest f_H recorded during the 15 s measurement phases in the warm ABT test and minimum heart rate ($f_{H,\text{min}}$) was defined as the lowest f_H recorded during the 15 s measurement phases during the cold ABT test. Peak temperature (T_{peak}) was the temperature corresponding to $f_{H,\text{max}}$.

Statistical analysis

All data were statistically analyzed using R (version 3.5.1). All metrics were investigated for normality using Shapiro–Wilk tests and quantile–quantile plots, and for heteroscedasticity using Levene's test. Data that were not normally distributed were log-transformed before statistical analysis (only FAS). Data are displayed with untransformed values. All data were statistically analyzed (significance level $\alpha=0.05$) using a 2-way ANOVA (Car v3.0-2; Fox and Weisberg, 2011) with *post hoc* Tukey HSD, except for the thermal limits from the cold ABT, which were analyzed using a *t*-test. Differences between treatments were also assessed

using Cohen's *D*-tests (*effsize*; <https://CRAN.R-project.org/package=effsize>). Note that 12°C fish were not tested for the cold ABT because of complications surrounding COVID-19 forcing the early shutdown of the experiment. In all 2-way ANOVA tests, significance of interaction between diet and temperature was tested for and excluded when non-significant. Polynomial curves were fitted to f_H data and compared using Akaike information criterion (AIC), where the fit with the lowest AIC score was assigned the best fit model, but all models with $\Delta\text{AIC}<2$ were considered (Burnham and Anderson, 2002). Measures of thermal sensitivity for all biological rates were calculated for each diet treatment using Q_{10} values, where:

$$Q_{10} = \frac{R_2^{(10/(T_2-T_1))}}{R_1}, \quad (1)$$

R_1 is the treatment mean at 12°C , R_2 is the treatment mean at 20°C , T_1 is 12°C and T_2 is 20°C .

RESULTS

Metabolic rates

Metabolism was influenced by diet and temperature, but each metabolic rate responded differently (Figs 3 and 4). SMR was 28% higher in the 20°C omnivorous diet treatment compared with the carnivorous diet treatment, resulting in a significant interaction between diet and temperature (Fig. 3A, Table 1). This was further supported by a large effect size between the 20°C treatments (Cohen's *D*-test). In contrast, MMR significantly increased with acclimation temperature, but did not differ across diet treatments (Fig. 3B,C, Table 1). There was a marginal, but not significant diet and temperature effect on AAS (Fig. 3C, Table 1). This was likely driven by the high individual variability in MMR, as the effect of diet on AAS had a *P*-value of 0.087 and medium effect sizes between diet treatments at 20°C . In contrast, FAS in the 20°C omnivorous diet treatment was significantly lower than that in the carnivorous diet treatment (diet: d.f.=1, $F=5.796$, $P=0.023$), which was largely driven by a 44% higher FAS (with a large effect size) in the carnivorous versus omnivorous diet (Fig. 3D, Table 1).

Other biological rates and traits

Growth rate, sprint speed, lipid peroxidation and lactate dehydrogenase activity were inconsistently affected by diet and temperature (Fig. 4). Growth was significantly higher at 20°C but did not differ across diets (Fig. 4, Table 1; Fig. S2). Unexpectedly, the Q_{10} value for growth was higher than that for all other rates (28.05) and not close to any of the Q_{10} values for metabolic rate (range 1.28–4.12; Fig. 4, Table 1; Fig. S2). Proximate analyses for whole tissue did not differ between treatment groups (Table S1).

In contrast with growth, maximum sprint speed did not differ in response to thermal acclimation ($Q_{10}=1.04$) or diet treatment (Fig. 4, Table 1; Figs S1 and S2). Similarly, lipid peroxidation in liver tissue did not differ in response to temperature and showed a marginal but insignificant effect of diet (Fig. 4; Fig. S2; temperature: d.f.=1, $F=0.318$, $P=0.579$; diet: d.f.=1, $F=3.260$, $P=0.085$). Lactate dehydrogenase activity in white muscle was moderately affected by temperature acclimation ($Q_{10}=2.10$), being higher at 20°C compared with 12°C , but did not differ across diets (Fig. 4; Fig. S3). Lactate dehydrogenase activity also increased with acute temperature exposure (Fig. S3). Overall, Q_{10} values for the reaction norms differed dramatically across biological rates, with sprint speed having a Q_{10} of 1.04 (insensitive to temperature), while growth rate had a Q_{10} of 28.05 (highly sensitive to temperature) (Fig. 4).

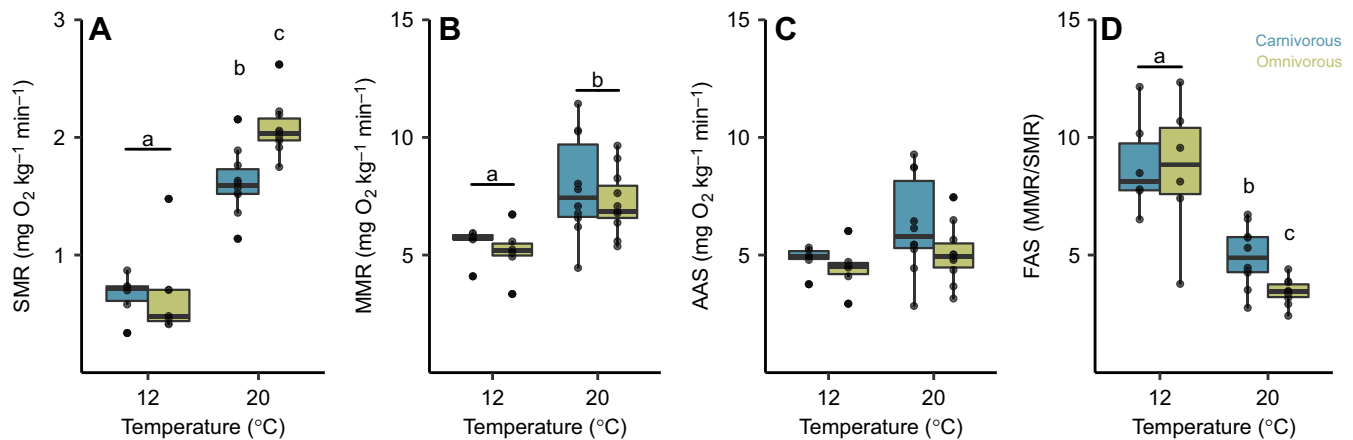


Fig. 3. Oxygen uptake rate ($\text{mg O}_2 \text{ kg}^{-1} \text{ min}^{-1}$) after an exhaustive chase protocol in opaleye acclimated to 12 or 20°C and fed either a carnivorous or omnivorous diet. (A) Standard metabolic rate (SMR), (B) maximum metabolic rate (MMR), (C) absolute aerobic scope (AAS; $\text{MMR} - \text{SMR}$) and (D) factorial aerobic scope (FAS; MMR/SMR). Blue, carnivorous diet (*Artemia* sp.); green, omnivorous diet (*Artemia* sp. and *Ulva* sp.). Lowercase letters indicate significant differences ($P < 0.05$) between treatment groups where applicable (see Table S1 for 2-way ANOVA outputs). Boxplots represent interquartile ranges (boxes and whiskers), median values (solid lines) and outliers (>1.5 beyond interquartile range) plotted as data points outside the whiskers.

Thermal tolerance

All thermal limits increased with acclimation to 20°C (Fig. 5, Table 2). Upper thermal limits (CT_{max} , T_{AB} , T_{peak} , T_{arr}) increased by 2.6–5.3°C and CT_{min} increased by 2.1–2.3°C with warm acclimation (Fig. 5, Table 2). Surprisingly, $f_{\text{H,max}}$ was significantly lower in the omnivorous treatments, which was driven by a 10% lower $f_{\text{H,max}}$ in the 20°C omnivorous treatment relative to the carnivorous treatment (Fig. 5, Table 2). As expected, f_{H} followed the

shape of an acute thermal performance curve (TPC), where it increased with an acute temperature increase until T_{peak} , at which point f_{H} began declining with temperature until the onset of cardiac arrhythmia (T_{arr} ; Fig. 5A). Thermal acclimation to 20°C shifted the acute TPC for f_{H} to the right of the TPC at 12°C (Fig. 5A). There was evidence to support an effect of diet on model selection for f_{H} in the warm ABT test, where the best fit model by AIC was a third-order polynomial curve that incorporated an interaction of acclimation

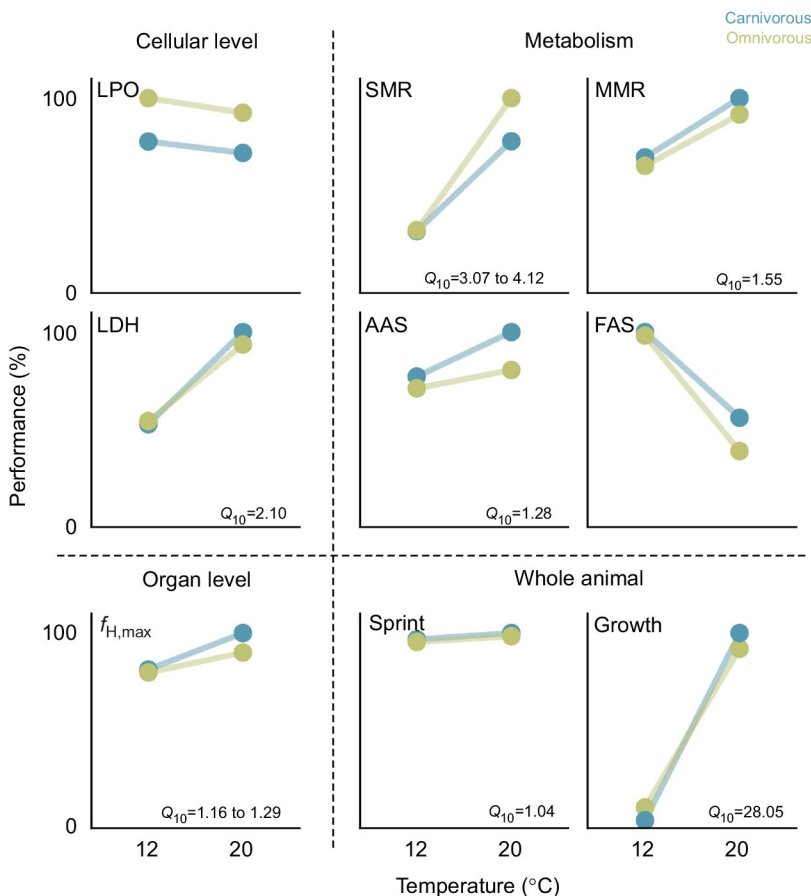


Fig. 4. Reaction norms plotted across all measured biological rates and traits. Average data from each treatment, scaled to the maximum average treatment value (i.e. the maximum treatment value is equal to 100%). All values are from fish tested at their acclimation temperature. Graphs are arranged by level of biological organization (cellular, organ, whole animal) and labelled by trait: SMR, MMR, AAS ($\text{MMR} - \text{SMR}$), FAS (MMR/SMR), maximum overall heart rate ($f_{\text{H,max}}$), maximum sprint speed, growth rate, lipid peroxidation (LPO) and lactate dehydrogenase activity (LDH). Q_{10} values are listed for all biological rate measurements (i.e. everything except LPO and FAS), as a range when the diet treatments were statistically different and as individual values when the diet treatments were not statistically different. Lines and circles indicate reaction norms and colors indicate diet treatment (blue, carnivorous diet; green, omnivorous diet).

Table 1. Summary statistics for biological rates

		Results						Statistical parameters						
	Temp.	Carnivorous		Omnivorous			Diet			Temperature			Diet×Temperature	
Biological rate	(°C)	<i>n</i>	Mean±s.e.m.	<i>n</i>	Mean±s.e.m.	d.f.	<i>F</i>	<i>P</i>	d.f.	<i>F</i>	<i>P</i>	d.f.	<i>F</i>	<i>P</i>
SMR (mg O ₂ kg ⁻¹ min ⁻¹)	12	6	0.66±0.07	7	0.67±0.14	1	8.577	0.007	1	147.093	<0.001	1	5.282	0.029
	20	10	1.62±0.09	10	2.08±0.07									
MMR (mg O ₂ kg ⁻¹ min ⁻¹)	12	6	5.49±0.28	6	5.15±0.45	1	1.061	0.311	1	16.542	<0.001			
	20	10	7.89±0.69	11	7.23±0.40									
AAS (mg O ₂ kg ⁻¹ min ⁻¹)	12	6	4.84±0.23	6	4.46±0.41	1	3.131	0.087	1	3.656	0.066			
	20	10	6.27±0.66	10	5.05±0.40									
FAS (MMR/SMR)	12	6	8.81±0.83	6	8.65±1.21	1	5.796	0.023	1	52.242	<0.001			
	20	10	4.94±0.41	10	3.43±0.17									
Growth (g week ⁻¹)	12	3	0.03±0.19	3	0.08±0.01	1	0.003	0.958	1	23.498	<0.001			
	20	3	0.75±0.15	3	0.69±0.16									
Sprint speed (cm s ⁻¹)	12	10	112.36±4.88	9	110.85±5.26	1	0.099	0.756	1	0.456	0.504			
	20	8	116.10±5.46	8	114.26±6.01									

Means±s.e.m. for each test group and ANOVA results are presented. SMR, standard metabolic rate; MMR, maximum metabolic rate; AAS, absolute aerobic scope; FAS, factorial aerobic scope; Temp., temperature, d.f., degrees of freedom.

temperature, acute test temperature and diet (Fig. 5A; Table S2). These models demonstrated that the acute TPC for f_H was lower across temperatures in the 20°C choice treatment compared with 20°C carnivorous treatment, which was consistent with the observed differences in $f_{H,max}$ (Fig. 5A). Altogether, these results demonstrate that for the 20°C acclimated fish, the omnivorous diet reduced overall cardiac function across temperatures but did not affect the upper thermal tolerance limits on the heart.

The results of the cold ABT test showed similar evidence of an effect of diet on f_H and $f_{H,min}$. Here, diet did not have a significant effect on any cold thermal limits (CT_{min} , $T_{AB-cold}$, $T_{arr-cold}$), but $f_{H,min}$ was lower (16%) in the omnivorous treatment compared with the carnivorous treatment at 20°C. There was also evidence to support an effect of diet on model selection, where the best fit model by AIC was a fourth order polynomial curve that incorporated an effect of acute test temperature and diet (Fig. 5C; Table S2).

DISCUSSION

Using an omnivorous fish, opaleye, as a model species, we found evidence that diet can influence thermal performance in ectotherms and does so in trait-specific ways. This has critical implications for our understanding of species responses to temperature change. Specifically, we examined three concepts outlined in Fig. 1: (1) whether diet and temperature acclimation can affect thermal performance and limits, (2) whether ecologically important traits vary in their diet and temperature sensitivity (integrating multiple levels of biological organization) and (3) whether trait-specific variation in diet and temperature responses creates performance trade-offs for opaleye fed an omnivorous diet compared with a carnivorous diet.

Thermal limits increased with temperature but did not differ across diet treatments

We assessed thermal limits using a standard and commonly used critical thermal (CT) test, as well as an ABT test, which measures thermal limits of the heart (Casselman et al., 2012). The heart may be a primary regulator of functional thermal tolerance in fishes. It is responsible for oxygen, immune cell, metabolite and waste transport around the body and is thought to be the first organ system to fail at extreme temperatures (Christen et al., 2018; Eliason and Anttila, 2017). The heart starts showing declines in performance (at T_{AB}) ~10–20°C lower than CT_{max} and fails (T_{arr}) at temperatures ~0–5°C lower than CT_{max} in fishes (Chen et al., 2015; Muñoz et al., 2014; Safi et al., 2019). As expected, all thermal limits increased with

temperature acclimation and CT_{max} was 2.7–3.8°C higher than T_{arr} . Opaleye showed a highly plastic acclimation response across all thermal limits, consistent with other temperate marine ectotherms (Vinagre et al., 2016). However, their thermal limits did not differ between diets.

While we did not observe a diet difference here, diet quality and quantity can alter thermal limits in ectotherms (fishes: Hoar and Cottle, 1952; Craig et al., 1995; Abdel-Ghany et al., 2019; Gomez Isaza et al., 2019; Lee et al., 2016; Turko et al., 2020; Woiwode and Adelman, 1992). Most previous studies used formulated diets varying in lipid composition; thus, dietary lipid composition may be a primary factor affecting thermal limits. This is critical to consider in the context of aquaculture, where animal feeds should be designed to ensure farmed animals have adequate thermal performance and resistance to suboptimal temperatures. To our knowledge, this is the first study to test the effect of quasi-natural diets on the thermal acclimation of critical and cardiac thermal limits in ectotherms. Research with natural diets is ecologically relevant for ectotherms that consume broad diets and especially for fish such as opaleye that seem to change their diet in response to temperature (Behrens and Lafferty, 2007, 2012; Emde et al., 2016; González-Bergonzoni et al., 2016; Guinan et al., 2015; Vejříková et al., 2016). Climate change is altering the nutritional landscape for many aquatic ectotherms (Birmie-Gauvin et al., 2017; Huey and Kingsolver, 2019). Managers and biologists should consider the effects of this on ectotherm thermal limits. Thus, future research should explore whether other natural diets can influence thermal limits in ectotherms, as this has critical implications for how they may respond to global climate change.

Thermal acclimation responses did not scale with temperature and diet equally across biological rates

The biological rates assessed here did not scale with diet and temperature equally. Thermal sensitivity differed dramatically across biological rates, where sprint speed did not differ with diet and temperature acclimation ($Q_{10}=1.04$), lactate dehydrogenase activity was moderately affected by temperature ($Q_{10}=2.10$), but insensitive to diet, and growth was highly temperature sensitive ($Q_{10}=28.05$), but also not sensitive to the diet treatments used in this study. These results are consistent with large scale meta-analyses, which have revealed that Q_{10} values differ across biological rates (Dell et al., 2011; Seebacher et al., 2015). Diet sensitivity was also inconsistent across rates, where SMR and $f_{H,max}$ were sensitive to diet,

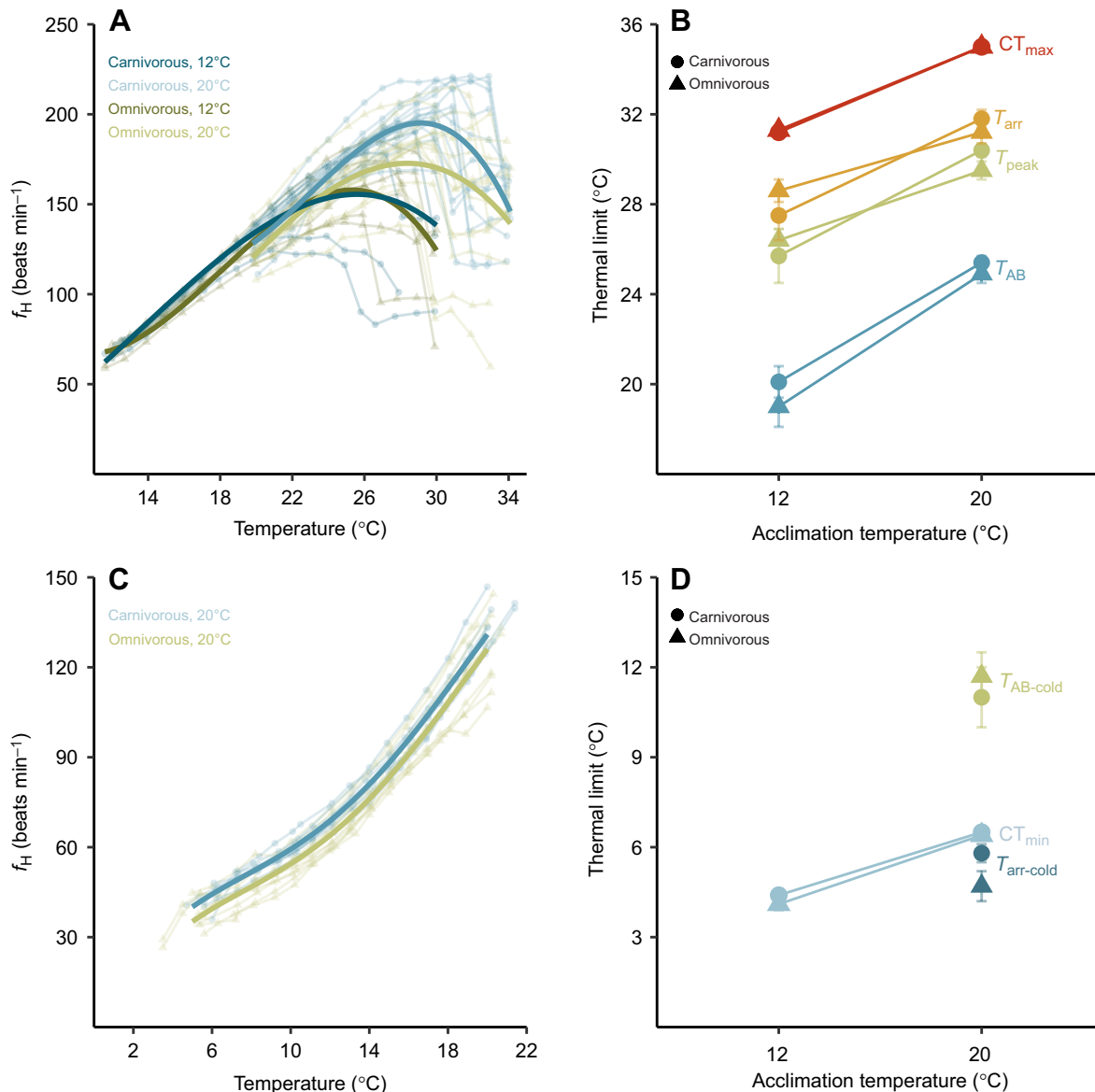


Fig. 5. Cardiac thermal performance. (A) Individual- and treatment-level responses of heart rate (f_H) of opaleye during acute warming from 12°C (dark blue, carnivorous diet, $N=6$; and dark green, omnivorous diet, $N=7$) or 20°C (light blue, carnivorous diet, $N=13$; and light green, omnivorous diet, $N=14$). Curves are third-order polynomials that account for the interaction diet $\times$ acclimation temperature $\times$ acute temperature. This was determined to be the best fit model by AIC (Table S2). (B) Upper thermal limits across diet and temperature treatments in opaleye. Circles and triangles indicate mean ($\pm$ s.e.m.) values for the carnivorous and omnivorous diet treatments, respectively. (C) Individual- and treatment-level responses in f_H from opaleye during acute cooling from 20°C (light blue, carnivorous diet, $N=9$; and light green, omnivorous diet, $N=11$). Curves are fourth-order polynomials that account for a fixed effect of diet and acute temperature. This was determined to be the best fit model by AIC (Table S2). (D) Lower thermal limits across diet and temperature treatments in opaleye. Circles indicate mean ($\pm$ s.e.m.) values for the carnivorous diet treatment and triangles indicate mean ($\pm$ s.e.m.) values for the omnivorous diet treatments. Note that f_H limits are missing from the 12°C treatments because of issues surrounding COVID-19. T_{arr} , temperature at the onset of cardiac arrhythmia; T_{AB} , breakpoint temperature of the heart; T_{peak} , temperature corresponding to maximum heart rate; CT_{max} , critical thermal maximum; CT_{min} , critical thermal minimum.

but all other rates were not. This was surprising for rates that are known to be impacted by diet, such as growth rate. Diet has been shown to have interactive effects with temperature on growth in other ectotherms (Sengalese sole: Guerreiro et al., 2012; rohu: Mishra and Samantaray, 2004; yellowtail kingfish: Ilham and Fotedar, 2016; crustaceans: Malzahn et al., 2016; Persson et al., 2011; Ruiz et al., 2020; Starke et al., 2020; insects: Kingsolver et al., 2006; Lee and Roh, 2010). However, *Ulva* sp. supplementation has had mixed effects on growth in aquaculture fish, with some studies showing modest amounts of *Ulva* sp. reducing growth, while others finding positive or no effect of *Ulva* sp. supplementation on growth (Wan

et al., 2019). Future work should explore how broader diet differences affect growth in relation to other important biological rates.

The effects of temperature on sprint speed in comparison to lactate dehydrogenase activity were similarly unexpected. Sprinting in fish is primarily driven by glycolytic fast twitch white muscle (McDonald et al., 1998; Kraskura and Nelson, 2018). Lactate dehydrogenase is a critical enzyme in lactic acid fermentation during glycolysis. Thus, we expected that sprint performance would have a comparable thermal sensitivity to lactate dehydrogenase activity. Lactate dehydrogenase activity increased with temperature acclimation, suggesting the opaleye had a greater anaerobic capacity

Table 2. Summary statistics for all thermal limits

Thermal limit	Temp. (°C)	Results				Statistical parameters					
		Carnivorous		Omnivorous		Diet			Temperature		
		<i>n</i>	Mean±s.e.m.	<i>n</i>	Mean±s.e.m.	d.f.	<i>F</i> (or <i>t</i> *)	<i>P</i>	d.f.	<i>F</i>	<i>P</i>
CT _{min}	12	9	4.4±0.2	10	4.1±0.2	1	1.984	0.168	1	226.206	<0.001
	20	12	6.5±0.1	8	6.4±0.1						
T _{AB}	12	6	20.1±0.7	7	19.0±0.9	1	2.127	0.153	1	110.743	<0.001
	20	13	25.4±0.3	14	24.9±0.4						
T _{peak}	12	6	25.7±1.2	7	26.4±0.5	1	0.612	0.439	1	46.990	<0.001
	20	13	30.4±0.3	14	29.5±0.4						
T _{arr}	12	6	27.5±1.1	7	28.6±0.5	1	0.021	0.886	1	33.766	<0.001
	20	13	31.8±0.4	14	31.2±0.5						
CT _{max}	12	10	31.2±0.2	10	31.3±0.2	1	0.547	0.465	1	500.078	<0.001
	20	8	35.0±0.2	10	35.0±0.1						
f _{H,max}	12	6	166.6±12.5	7	163.5±6.2	1	5.243	0.028	1	17.669	<0.001
	20	13	204.8±4.4	14	184.4±5.4						
T _{arr-cold}	12	NA	NA	NA	NA	18	1.790*	0.090			
	20	9	5.8±0.3	11	4.7±0.5						
T _{AB-cold}	12	NA	NA	NA	NA	14	−0.493*	0.630			
	20	8	11.0±1.0	8	11.7±0.8						
f _{H,min}	12	NA	NA	NA	NA	18	2.407*	0.027			
	20	9	44.5±1.9	11	37.4±2.2						

Means±s.e.m. for each test group and ANOVA results are presented. *Note that *t*-test results are reported on the results of the cold ABT test, as this test was only run on 20°C acclimation fish because of complications surrounding COVID-19. CT_{min}, critical thermal minimum; T_{AB}, breakpoint temperature of the heart; T_{peak}, temperature corresponding to maximum heart rate; T_{arr}, temperature at the onset of cardiac arrhythmia; CT_{max}, critical thermal maximum; f_{H,max}, maximum heart rate across entire warm ABT test; f_{H,min}, minimum heart rate during cold ABT test.

at 20°C (McDonald et al., 1998). However, this did not translate to increases in sprint performance. Other enzymes in glycolysis could be rate-limiting steps (e.g. phosphofructokinase; McDonald et al., 1998) and more highly correlated with sprint speed. Sprint speed has been examined in one other diet×temperature study in fishes, which measured macronutrient selection and temperature effects on damselfish (Rowe et al., 2018). Macronutrient selection did not change with temperature, but sprint speed was highest in the colder temperature treatment (Rowe et al., 2018). Fish use two different swimming modes: anaerobically powered burst swimming and sustained aerobically powered swimming. These swimming modes may be differentially affected by diet and temperature. The effect of diet and temperature on aerobically powered swimming and maximum swim speeds (i.e. maximum swimming speed, U_{max} , and critical swimming speed, U_{crit}) have been assessed in grey mullet (Vagner et al., 2015, 2019), with no observed effect of diet, but an effect of temperature on U_{max} and no effect of diet or temperature on U_{crit} . Given that two metabolic traits, SMR and FAS, were affected by diet and temperature in this study, future work should explore how aerobically powered swimming is impacted by different levels of omnivory.

The diverse diet and temperature responses that we observed across biological rates resulted in no consistent pattern in the thermal sensitivity of any metabolic rates or levels of biological organization. The small number of papers that have measured the interactive effects of diet and temperature on metabolic rates in other ectotherms have found mixed results (i.e. insects: Alton et al., 2020; Schmitz and Rosenblatt, 2017; fish: Pérez-Casanova et al., 2010; Vagner et al., 2015). For example, Vagner et al. (2015) found an interactive effect of dietary fatty acid composition and temperature on the AAS of juvenile grey mullet. In contrast, Pérez-Casanova et al. (2010) did not find any effects of macronutrient ratios on metabolism in juvenile Atlantic cod and haddock. Our results were especially surprising for SMR, as it is often assumed that maintenance metabolism is coupled to all other biological rates and should have the same thermal sensitivity (Gillooly et al., 2001;

Brown et al., 2004). Aerobic scope is also considered a master physiological factor that determines the capacity for all other aerobically powered functions (Fry, 1947; Brett, 1971; Claireaux and Lefrançois, 2007; Pörtner, 2010), although this idea has been heavily debated (Clark et al., 2013; Schulte, 2015). Here, AAS did not differ in response to diet or temperature (although FAS did) and there was also no observed pattern between AAS, SMR and other biological rates. These results indicate that caution should be taken when using AAS and SMR as indicators of overall performance or proxies for other biological rates, especially when predicting the effects of multiple factors, such as diet and temperature.

Performance costs and benefits for diet choice at different acclimation temperatures

Contrary to our hypothesis, we did not find evidence of a performance trade-off to the omnivorous diet used in this study. Instead, opaleye that consumed the omnivorous diet displayed several higher costs: lower f_H across a thermal gradient, higher SMR and reduced FAS when acclimated to 20°C. Given that the thermal limits of the heart did not change with diet, it was remarkable that diet downshifted the thermal performance curve for $f_{H,max}$. A reduction in $f_{H,max}$ indicates a reduced capacity to transport oxygen, metabolites, immune cells and waste around the body. There are several potential mechanisms that could have driven this reduction in $f_{H,max}$ across acute temperatures. For example, differences in the lipid composition of the diet can impact membrane remodeling, which can affect cardiac function in fish (Chatelier et al., 2006; McKenzie, 2001). Further, although *Ulva* sp. contains no known herbivore deterrents, our results suggest some sort of anti-nutrient effect of *Ulva* sp. supplementation, which could have caused the observed reductions in cardiac thermal performance.

The higher SMR observed in the 20°C omnivorous treatment was expected as omnivorous and herbivorous animals generally have higher digestive infrastructure costs than carnivores (e.g. broader digestive enzymes, higher gut surface area; Caruso and Sheridan, 2011; Clements et al., 2009; Horn, 1989). Consistent with the higher

SMR at 20°C, the opaleye in the omnivorous treatment had a reduced FAS (3.43) that was less than half that at 12°C (8.65). In contrast, the carnivorous diet maintained a 44% higher FAS at 20°C (4.94) compared with the omnivorous diet. FAS is indicative of the amount of scope available to perform critical biological functions that scale proportional to SMR (Farrell, 2016). Digestion generally requires at least a doubling of SMR in many fishes (i.e. $FAS > 2$; Chabot et al., 2016; Farrell, 2016; McCue, 2006). This suggests that the opaleye in the omnivorous treatment at 20°C may have been on the threshold of having limited scope for activities beyond digestion. As climate change increases the seasonal extreme temperatures and the frequency of marine heatwaves in the rocky intertidal zone (IPCC, 2019), the opaleye's FAS will decrease further, which could limit other important biological functions, such as digestion, and exacerbate energetic tradeoffs associated with different diets. Thus, another avenue for future research will be to untangle diet × temperature effects on digestive costs (i.e. specific dynamic action) in relation to AAS and digestion efficiency (Jutfelt et al., 2021).

Given that opaleye and other omnivorous fishes eat more plants in warmer water in the wild, we expected there to be a performance advantage to the omnivorous diet. We measured a suite of traits to test for any performance benefits that may offset the costs of the omnivorous treatment. Specifically, we explored how diet and temperature affected (1) thermal tolerance, (2) maximum sprint speed, (3) glycolytic capacity and (4) oxidative stress. We predicted that because opaleye eat more plants in warmer water (Behrens and Lafferty, 2012), the omnivorous diet would result in higher thermal limits than the carnivorous diet at 20°C. However, we did not find evidence of any benefits to the omnivorous diet. We also expected that the higher lipid and protein content of the carnivorous treatment would raise oxidative stress relative to the omnivorous diet. Analysis of lipid peroxidation in liver tissue revealed a marginal trend of higher oxidative stress in the omnivorous diet treatment. However, antioxidant capacity could still have been higher in the omnivorous treatment. For example, Coggins et al. (2017) examined the effect of dietary glutathione supplementation on thermal limits, antioxidant capacity and lipid peroxidation in *Daphnia* sp. As the glutathione concentration increased, so did total antioxidant capacity, but glutathione supplementation did not alter lipid peroxidation or thermal limits (Coggins et al., 2017). In contrast, Castro et al. (2012) examined the effect of macronutrient ratios (45% and 55% protein) and temperature acclimation (12 and 18°C) on multiple antioxidant enzymes and lipid peroxidation in juvenile Senegalese sole. Lipid peroxidation differed across temperature treatments and was highest in the 55% protein diet (Castro et al., 2012). The limited amount of research on these interactions and inconclusiveness across studies indicate that more research is needed to elucidate the role of dietary antioxidants and macronutrient ratios in regulating oxidative stress across temperatures and taxa.

While it was not within the scope of this study, other important performance traits may have differed between the diet treatments. These include, but are not limited to, differences in microbiome diversity and function, visual acuity, cognitive ability, digestion efficiency and digestive costs relative to aerobic scope, immune function, aerobic swimming performance and cardiac stroke volume (Glencross and Rutherford, 2010; Koven et al., 2018; Vagner et al., 2014, 2019). It remains unclear whether opaleye and other omnivorous ectotherms consume different proportions of plant to animal to regulate their thermal responses. Here, the opaleye's omnivorous diet did not maximize their performance compared with a more specialized carnivorous diet. However, many other factors govern diet choice in the wild, including life history (Zhang et al.,

2020), competition (Pfenning, 1990), predation (Schmitz et al., 2016), food availability (MacArthur and Pianka, 1966) and habitat structure (Behrens and Lafferty, 2007). Therefore, the ecological benefits of consuming an omnivorous diet may outweigh the physiological costs.

Many ectotherms change their diets with temperature; either directly because their diet preference changes in response to temperature (Boersma et al., 2016b; Carreira et al., 2016; Devries and Appel, 2014; Lee et al., 2015; Lemoine et al., 2014; Rho and Lee, 2017; Schmitz et al., 2016; Vejříková et al., 2016) or indirectly because food availability or the nutritional content of a diet item changes with temperature (Alton et al., 2020; Boersma et al., 2016b; Cross et al., 2015; Ho et al., 2010). In either scenario, generalist ectotherms that have the capacity to adjust their diet may be at an advantage compared with those with more specialized inflexible diets. However, the ultimate diet choices that ectotherms make may not always be 'better' or 'worse' because diet and temperature can interact and have trait-specific effects. Not all diet choices are necessarily adaptive. Irrespective of the reasons why an ectotherm eats what it eats, our work here demonstrates that diet choices have consequences. These consequences have far-reaching implications, including whether diet choice can facilitate geographic range expansion, or colonization of warmer or cooler habitats; and further, whether specialist diets constrain thermal niches or whether diet can facilitate differences in acclimation rates or performance under fluctuating temperatures. Overall, diet should be treated as an interacting factor that has the capacity to modify the thermal responses of ectotherms.

Concluding remarks

Thermal acclimation is a key mechanism that ectotherms employ to maintain performance across a range of temperatures. Acclimation requires energy and nutritional building blocks that ectotherms obtain from their diet. Here, we explored whether different diets mediated distinct thermal acclimation responses in an omnivorous fish, opaleye. We found clear evidence that diet influences thermal acclimation responses. However, there was no consistent pattern in how different biological rates responded to the temperature and diet treatments, with Q_{10} values ranging from 1 to 28. When confronted with a seasonal warm temperature (20°C), the opaleye in the omnivorous diet treatment had inferior performance (higher SMR, lower FAS and lower cardiac performance) relative to the opaleye fed a carnivorous diet treatment. Global climate change is already changing the average and extreme temperatures that marine ectotherms experience as well as their nutritional landscape (IPCC, 2019; Bimie-Gauvin et al., 2017). These environmental changes are likely to interact and alter many ectotherms' thermal performance in the wild. Incorporating multiple interacting factors into our understanding of species' responses to global climate change is the next step in ensuring that researchers capture the resilience of different species and populations (Jackson et al., 2021). Accordingly, diet is essential to consider when predicting ectotherm performance in variable environments and in response to global climate change.

Acknowledgements

We thank Mason Tittle, Vincent Han Lee, Samantha Csik, Bella Giglio and Terra Dressler for assistance in the lab and with fishing; Bartholomew Difiore, Joseph Curtis, Sevan Esaian, Nicholas Lee, Hope Hardison, Osborne Hardison, Brendan Shanny, Dr Alexander Lill, Terra Dressler, Dr Alexander Little, Bella Giglio, Anhadh Jassal, Claire Anderson, Jennay Argiris, Garret Parsons, Tyler Parsons and Alecia Dezzani for help fishing; David Davis and the entire marine operations staff at the University of California, Santa Barbara; Dr Adrian Stier and Dr Elizabeth Wilbanks for lending equipment; and Dr Christopher Jerde, Dr Gretchen Hofmann, Dr Kevin Lafferty, Dr Alexander Little and Dr Elizabeth Wilbanks for advice and feedback.

Competing interests

The authors declare no competing or financial interests.

Author contributions

Conceptualization: E.A.H., E.J.E.; Methodology: E.A.H., K.K., J.V.W., E.J.E.; Formal analysis: E.A.H., K.K., T.N.; Investigation: E.A.H., K.K., J.V.W., T.N., E.J.E.; Resources: E.J.E.; Data curation: E.A.H.; Writing - original draft: E.A.H., E.J.E.; Writing - review & editing: E.A.H., K.K., J.V.W., T.N., E.J.E.; Visualization: E.A.H.; Supervision: E.J.E.; Project administration: E.J.E.; Funding acquisition: E.J.E.

Funding

This work was supported by a Hellman Family Faculty Fellowship and the University of California, Santa Barbara. Additional funding for E.A.H. and J.V.W. was provided by National Science Foundation Graduate Research Fellowships. Additional support for T.N. was provided by the University of California LEADS Program. Open access funding provided by University of California, Santa Barbara. Deposited in PMC for immediate release. We thank the Santa Barbara Coastal Long Term Ecological Research Project under the National Science Foundation Cooperative Agreement #OCE-1831937.

Data availability

Data are available in Dryad (Hardison et al., 2021): D98P66.

References

- Abdel-Ghany, H. M., El-Sayed, A.-F. M., Ezzat, A. A., Essa, M. A. and Helal, A. M. (2019). Dietary lipid sources affect cold tolerance of Nile tilapia (*Oreochromis niloticus*). *J. Therm. Biol.* **79**, 50–55. doi:10.1016/j.jtherbio.2018.11.009
- Altton, L. A., Kutz, T. C., Bywater, C. L., Beaman, J. E., Arnold, P. A., Mirth, C. K., Sgrò, C. M. and White, C. R. (2020). Developmental nutrition modulates metabolic responses to projected climate change. *Funct. Ecol.* **34**, 2488–2502. doi:10.1111/1365-2435.13663
- Anttila, K., Casselman, M. T., Schulte, P. M. and Farrell, A. P. (2013). Optimum temperature in juvenile salmonids: connecting subcellular indicators to tissue function and whole-organism thermal optimum. *Physiol. Biochem. Zool.* **86**, 245–256. doi:10.1086/669265
- Anttila, K., Couturier, C. S., Øverli, Ø., Johnsen, A., Marthinsen, G., Nilsson, G. E. and Farrell, A. P. (2014). Atlantic salmon show capability for cardiac acclimation to warm temperatures. *Nat. Commun.* **5**, 6–11. doi:10.1038/ncomms5252
- Arnold, K. E., Ramsay, S. L., Henderson, L. and Larcombe, S. D. (2010). Seasonal variation in diet quality: antioxidants, invertebrates and blue tits *Cyanistes caeruleus*. *Biol. J. Linn. Soc.* **99**, 708–717. doi:10.1111/j.1095-8312.2010.01377.x
- Barry, J. P. and Ehret, M. J. (1993). Diet, food preference, and algal availability for fishes and crabs on intertidal reef communities in southern California. *Env. Biol. Fish.* **37**, 75–95. doi:10.1007/BF00000715
- Behrens, M. D. and Lafferty, K. D. (2007). Temperature and diet effects on omnivorous fish performance: implications for the latitudinal diversity gradient in herbivorous fishes. *Can. J. Fish. Aquat. Sci.* **64**, 867–873. doi:10.1139/f07-063
- Behrens, M. D. and Lafferty, K. D. (2012). Geographic Variation in the Diet of Opaleye (*Girella nigricans*) with Respect to Temperature and Habitat. *PLoS ONE*, **7**, e45901. doi:10.1371/journal.pone.0045901
- Beitinger, T. and Lutterschmidt, W. (2011). Temperature Measures of thermal tolerance. In *Encyclopedia of Fish Physiology: From Genome to Environment* (ed. A. P. Farrell), pp. 1695–1702. Elsevier Ltd.
- Bernhardt, J. R. and Leslie, H. M. (2013). Resilience to climate change in coastal marine ecosystems. *Ann. Rev. Mar. Sci.* **5**, 371–392. doi:10.1146/annurev-marine-121211-172411
- Birnie-Gauvin, K., Peiman, K. S., Raubenheimer, D. and Cooke, S. J. (2017). Nutritional physiology and ecology of wildlife in a changing world. *Conserv. Physiol.* **5**, cox030. doi:10.1093/conphys/cox030
- Boersma, M., Mathew, K. A., Niehoff, B., Schoo, K. L., Franco-Santos, R. M., Meunier, C. L. (2016a). Temperature-driven changes in the diet preference of omnivorous copepods: no more meat when it's hot? A response to Winder et al. *Ecol. Lett.* **19**, 1386–1388. doi:10.1111/ele.12666
- Boersma, M., Mathew, K. A., Niehoff, B., Schoo, K. L., Franco-Santos, R. M. and Meunier, C. L. (2016b). Temperature driven changes in the diet preference of omnivorous copepods: No more meat when it's hot? *Ecol. Lett.* **19**, 45–53. doi:10.1111/ele.12541
- Brett, J. R. (1971). Energetic responses of salmon to temperature. A study of some thermal relations in the physiology and freshwater ecology of sockeye salmon (*Oncorhynchus nerka*). *Amer. Zool.* **11**, 99–113. doi:10.1093/icb/11.1.99
- Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. and West, G. B. (2004). Toward a Metabolic Theory of Ecology. *Ecology* **85**, 1771–1789. doi:10.1890/03-9000
- Burnham, K. P. and Anderson, D. R. (2002). *Model Selection and Multi-model Inference: A Practical Information-theoretic approach*. New York: Springer.
- Carreira, B. M., Segurado, P., Orizaola, G., Gonçalves, N., Pinto, V., Laurila, A. and Rebelo, R. (2016). Warm vegetarians? Heat waves and diet shifts in tadpoles. *Ecology* **97**, 2964–2974. doi:10.1002/ecy.1541
- Caruso, M. A. and Sheridan, M. A. (2011). Gut anatomy and morphology | Pancreas. In *Encyclopedia of Fish Physiology*, Vol. 2. (ed. A. P. Farrell), pp. 1276–1283. London: Academic Press.
- Casselman, M. T., Anttila, K. and Farrell, A. P. (2012). Using maximum heart rate as a rapid screening tool to determine optimum temperature for aerobic scope in Pacific salmon *Oncorhynchus* spp. *J. Fish Biol.* **80**, 358–377. doi:10.1111/j.1095-8649.2011.03182.x
- Castro, C., Pérez-Jiménez, A., Guerreiro, I., Peres, H., Castro-Cunha, M. and Oliva-Teles, A. (2012). Effects of temperature and dietary protein level on hepatic oxidative status of Senegalese sole juveniles (*Solea senegalensis*). *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **163**, 372–378. doi:10.1016/j.cbpa.2012.07.003
- Chabot, D., Koenker, R. and Farrell, A. P. (2016). The measurement of specific dynamic action in fishes. *Fish Biol.* **88**, 152–172. doi:10.1111/jfb.12836
- Chatelier, A., McKenzie, D. J., Prinnet, A., Galois, R., Robin, J., Zambonino, J. and Claireaux, G. (2006). Associations between tissue fatty acid composition and physiological traits of performance and metabolism in the seabass (*Dicentrarchus labrax*). *J. Exp. Biol.* **209**, 3429–3439. doi:10.1242/jeb.02347
- Chen, Z., Devlin, R. H. and Farrell, A. P. (2015). Upper thermal tolerance of wild-type, domesticated and growth hormone-transgenic coho salmon *Oncorhynchus kisutch*. *J. Fish Biol.* **87**, 763–773. doi:10.1111/jfb.12736
- Christen, F., Desrosiers, V., Dupont-Cyr, B. A., Vandenberg, G. W., Le François, N. R., Tardif, J.-C., Dufresne, F., Lamarre, S. G. and Blier, P. U. (2018). Thermal tolerance and thermal sensitivity of heart mitochondria: Mitochondrial integrity and ROS production. *Free Radic. Biol. Med.* **116**, 11–18. doi:10.1016/j.freeradbiomed.2017.12.037
- Chung, D. J. and Schulte, P. M. (2020). Mitochondria and the thermal limits of ectotherms. *J. Exp. Biol.* **223**, jeb227801. doi:10.1242/jeb.227801
- Claireaux, G. and Lefrançois, C. (2007). Linking environmental variability and fish performance: integration through the concept of scope for activity. *Philos. Trans. R. Soc. B Biol. Sci.* **362**, 2031–2041. doi:10.1098/rstb.2007.2099
- Clark, T. D., Sandblom, E. and Jutfelt, F. (2013). Aerobic scope measurements of fishes in an era of climate change: respirometry, relevance and recommendations. *J. Exp. Biol.* **216**, 2771–2782. doi:10.1242/jeb.084251
- Clements, K. D., Raubenheimer, D. and Choat, J. H. (2009). Nutritional ecology of marine herbivorous fishes: ten years on. *Funct. Ecol.* **23**, 79–92. doi:10.1111/j.1365-2435.2008.01524.x
- Coggins, B. L., Collins, J. W., Holbrook, K. J. and Yampolsky, L. Y. (2017). Antioxidant capacity, lipid peroxidation, and lipid composition changes during long-term and short-term thermal acclimation in *Daphnia*. *J. Comp. Physiol. B Biochem. Syst. Environ. Physiol.* **187**, 1091–1106. doi:10.1007/s00360-017-1090-9
- Craig, S. R., Neill, W. H. and Gatlin, D. M. (1995). Effects of dietary lipid and environmental salinity on growth, body composition, and cold tolerance of juvenile red drum (*Sciaenops ocellatus*). *Fish Physiol. Biochem.* **14**, 49–61. doi:10.1007/BF00004290
- Cross, W. F., Hood, J. M., Benstead, J. P., Hurns, A. D. and Nelson, D. (2015). Interactions between temperature and nutrients across levels of ecological organization. *Glob. Chang. Biol.* **21**, 1025–1040. doi:10.1111/gcb.12809
- Daufresne, M., Lengfellner, K. and Sommer, U. (2009). Global warming benefits the small in aquatic ecosystems. *Proc. Natl. Acad. Sci. USA* **106**, 12788–12793. doi:10.1073/pnas.0902080106
- Dell, A. I., Pawar, S. and Savage, V. M. (2011). Systematic variation in the temperature dependence of physiological and ecological traits. *Proc. Natl. Acad. Sci. USA* **108**, 10591–10596. doi:10.1073/pnas.1015178108
- Devries, Z. C. and Appel, A. G. (2014). Effects of temperature on nutrient self-selection in the silverfish *lepisma saccharina*. *Physiol. Entomol.* **39**, 217–221. doi:10.1111/phen.12064
- Ekström, A., Hellgren, K., Gräns, A., Pichaud, N. and Sandblom, E. (2016). Dynamic changes in scope for heart rate and cardiac autonomic control during warm acclimation in rainbow trout. *J. Exp. Biol.* **219**, 1106–1109.
- Eliason, E. J. and Anttila, K. (2017). 4 - Temperature and the Cardiovascular System. In *The Cardiovascular System* (ed. A. K. Gamperl, T. E. Gillis, A. P. Farrell and C. J. Brauner), pp. 235–297. Academic Press.
- Emde, S., Kochmann, J., Kuhn, T., Dörge, D. D., Plath, M., Miesen, F. W. and Klimpel, S. (2016). Cooling water of power plant creates "hot spots" for tropical fishes and parasites. *Parasitol. Res.* **115**, 85–98. doi:10.1007/s00436-015-4724-4
- Farrell, A. P. (2016). Pragmatic perspective on aerobic scope: peaking, plummeting, pejus and apportioning. *J. Fish Biol.* **88**, 322–343. doi:10.1111/jfb.12789
- Ferreira, E. O., Anttila, K. and Farrell, A. P. (2014). Thermal optima and tolerance in the eurythermic goldfish (*Carassius auratus*): relationships between whole-animal aerobic capacity and maximum heart rate. *Physiol. Biochem. Zool.* **87**, 599–611. doi:10.1086/677317
- Floeter, S. R., Behrens, M. D., Ferreira, C. E. L., Paddack, M. J. and Horn, M. H. (2005). Geographical gradients of marine herbivorous fishes: patterns and processes. *Mar. Biol.* **147**, 1435–1447. doi:10.1007/s00227-005-0027-0
- Fox, J. and Weisberg, S. (2011). *An R companion to applied regression*, 2nd edn. Thousand Oaks, CA: Sage.

- Fry, F. E. J. (1947). *Effects of the Environment on Animal Activity*, Vol. 68, pp. 1–52. Toronto: University of Toronto Press.
- Gilbert, M. J. H., Harris, L. N., Malley, B. K., Schimnowski, A., Moore, J.-S. and Farrell, A. P. (2020). The thermal limits of cardiorespiratory performance in anadromous Arctic char (*Salvelinus alpinus*): a field-based investigation using a remote mobile laboratory. *Conserv. Physiol.* **8**, coaa036. doi:10.1093/conphys/coaa036
- Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M. and Charnov, E. L. (2001). Effects of size and temperature on metabolic rate. *Science* **293**, 2248–2251. doi:10.1126/science.1061967
- Glanville, E. J. and Seebacher, F. (2006). Compensation for environmental change by complementary shifts of thermal sensitivity and thermoregulatory behaviour in an ectotherm. *J. Exp. Biol.* **209**, 4869–4877. doi:10.1242/jeb.02585
- Glencross, B. and Rutherford, N. (2010). Dietary strategies to improve the growth and feed utilization of barramundi, *Lates calcarifer* under high water temperature conditions. *Aquac. Nutr.* **16**, 343–350. doi:10.1111/j.1365-2095.2009.00670.x
- Gomez Isaza, D. F., Cramp, R. L., Smullen, R., Glencross, B. D. and Franklin, C. E. (2019). Coping with climatic extremes: Dietary fat content decreased the thermal resilience of barramundi (*Lates calcarifer*). *Comp. Biochem. Physiol.* **230**, 64–70. doi:10.1016/j.cbpa.2019.01.004
- González-Bergonzoni, I., Meerhoff, M., Davidson, T. A., Teixeira-de Mello, F., Baattrup-Pedersen, A. and Jeppesen, E. (2012). Meta-analysis shows a consistent and strong latitudinal pattern in fish omnivory across ecosystems. *Ecosystems* **15**, 492–503. doi:10.1007/s10021-012-9524-4
- González-Bergonzoni, I., Jeppesen, E., Vidal, N., Teixeira-de Mello, F., Goyenola, G., López-Rodríguez, A. and Meerhoff, M. (2016). Potential drivers of seasonal shifts in fish omnivory in a subtropical stream. *Hydrobiologia* **768**, 183–196. doi:10.1007/s10750-015-2546-0
- Gradiš, K. J., Garner, S. R., Wilson, C. C., Farrell, A. P. and Neff, B. D. (2016). Relationship between cardiac performance and environment across populations of Atlantic salmon (*Salmo salar*): a common garden experiment implicates local adaptation. *Evol. Ecol.* **30**, 877–886. doi:10.1007/s10682-016-9847-2
- Guerreiro, I., Peres, H., Castro-Cunha, M. and Oliva-Teles, A. (2012). Effect of temperature and dietary protein/lipid ratio on growth performance and nutrient utilization of juvenile Senegalese sole (*Solea senegalensis*). *Aquac. Nutr.* **18**, 98–106. doi:10.1111/j.1365-2095.2011.00884.x
- Guinan, M. E., Kapuscinski, K. L. and Teece, M. A. (2015). Seasonal diet shifts and trophic position of an invasive cyprinid, the rudd *Scardinius erythrophthalmus* (Linnaeus, 1758), in the upper Niagara river. *Aquat. Invasions* **10**, 217–225. doi:10.3391/ai.2015.10.2.10
- Hansen, A. K., Byriel, D. B., Jensen, M. R., Steffensen, J. F. and Svendsen, M. B. S. (2017). Optimum temperature of a northern population of Arctic charr (*Salvelinus alpinus*) using heart rate Arrhenius breakpoint analysis. *Pol. Biol.* **40**, 1063–1070. doi:10.1007/s00300-016-2033-8
- Hardison, E., Kraskura, K., Van Wert, J., Nguyen, T. and Eliason, E. J. (2021). Diet mediates thermal performance traits: implications for marine ectotherms. *Dryad Dataset*. doi:10.25349/D98P66
- Healy, T. M. and Schulte, P. M. (2012). Thermal acclimation is not necessary to maintain a wide thermal breadth of aerobic scope in the common killifish (*Fundulus heteroclitus*). *Physiol. Biochem. Zool.* **85**, 107–119. doi:10.1086/664584
- Ho, C. K., Pennings, S. C. and Carefoot, T. H. (2010). Is diet quality an overlooked mechanism for Bergmann's rule? *Am. Nat.* **175**, 269–276. doi:10.1086/649583
- Hoar, W. S. and Cottle, M. K. (1952). Dietary fat and temperature tolerance of goldfish. *Can. J. Zool.* **30**, 41–48. doi:10.1139/z52-003
- Hofmann, G. E. and Todgham, A. E. (2010). Living in the now: physiological mechanisms to tolerate a rapidly changing environment. *Annu. Rev. Physiol.* **72**, 127–145. doi:10.1146/annurev-physiol-021909-135900
- Horn, M. H. (1989). Biology of marine herbivorous fishes. *Ocean. Mar. Biol. Annu. Rev.* **27**, 167–272.
- Huey, R. B. and Kingsolver, J. G. (2019). Climate warming, resource availability, and the metabolic meltdown of ectotherms. *Am. Nat.* **194**, E140–E150. doi:10.1086/705679
- Huey, R. B., Kearney, M. R., Krockenberger, A., Holtum, J. A. M., Jess, M. and Williams, S. E. (2012). Predicting organismal vulnerability to climate warming: Roles of behaviour, physiology and adaptation. *Philos. Trans. R. Soc. B Biol. Sci.* **367**, 1665–1679. doi:10.1098/rstb.2012.0005
- Ihham, I. and Fotedar, R. (2016). Growth, antioxidant capacity and muscle histochemistry of yellowtail kingfish (*Seriola lalandi* Valenciennes 1883): Selenium and temperature interaction. *Anim. Feed Sci. Technol.* **217**, 76–86. doi:10.1016/j.anifeedsci.2016.04.009
- IPCC (2019). *IPCC Special Report on the Ocean and Cryosphere in a Changing Climate* (ed. H.-O. Pörtner et al.).
- Jackson, M. C., Pawar, S. and Woodward, G. (2021). The temporal dynamics of multiple stressor effects: from individuals to ecosystems. *Trends Ecol. Evol.* **36**, 402–410. doi:10.1016/j.tree.2021.01.005
- Jang, T., Rho, M. S., Koh, S.-H. and Lee, K. P. (2015). Host-plant quality alters herbivore responses to temperature: A case study using the generalist *Hyphantria cunea*. *Entomol. Exp. Appl.* **154**, 120–130. doi:10.1111/eea.12261
- Jobling, M. (2016). Fish nutrition research: past, present and future. *Aquac. Int.* **24**, 767–786. doi:10.1007/s10499-014-9875-2
- Johnson, J. S., Clements, K. D. and Raubenheimer, D. (2017). The nutritional basis of seasonal selective feeding by a marine herbivorous fish. *Mar. Biol.* **164**, 201. doi:10.1007/s00227-017-3223-9
- Jutfelt, F., Norin, T., Åsheim, E. R., Rowsey, L. E., Andreassen, A. H., Morgan, R., Clark, T. D. and Speers-Roesch, B. (2021). 'Aerobic scope protection' reduces ectotherm growth under warming. *Funct. Ecol.* **35**, 1397–1407. doi:10.1111/1365-2435.13811
- Kaiser, M. J. and Hughes, R. N. (1993). Factors affecting the behavioural mechanisms of diet selection in fishes. *Mar. Behav. Physiol.* **23**, 105–118. doi:10.1080/10236249309378860
- Kellermann, V., Chown, S. L., Schou, M. F., Aitkenhead, I., Janion-Scheepers, C., Clemson, A., Scott, M. T. and Sgrò, C. M. (2019). Comparing thermal performance curves across traits: how consistent are they? *J. Exp. Biol.* **222**, jeb193433. doi:10.1242/jeb.193433
- Kingsolver, J. G., Shlichta, J. G., Ragland, G. J. and Massie, K. R. (2006). Thermal reaction norms for caterpillar growth depend on diet. *Evol. Ecol. Res.* **8**, 703–715.
- Koven, W., Nixon, O., Allon, G., Gaon, A., El Sadin, S., Falcon, J., Besseau, L., Escande, M., Vassallo Agius, R., Gordin, H. et al. (2018). The effect of dietary DHA and taurine on rotifer capture success, growth, survival and vision in the larvae of Atlantic bluefin tuna (*Thunnus thynnus*). *Aquaculture* **482**, 137–145. doi:10.1016/j.aquaculture.2017.09.039
- Kraskura, K. and Nelson, J. A. (2018). Hypoxia and sprint swimming performance of juvenile striped bass, *Morone saxatilis*. *Physiol. Biochem. Zool.* **91**, 682–690. doi:10.1086/694933
- Lee, K. P. and Roh, C. (2010). Temperature-by-nutrient interactions affecting growth rate in an insect ectotherm. *Entomol. Exp. Appl.* **136**, 151–163. doi:10.1111/j.1570-7458.2010.01018.x
- Lee, K. P., Jang, T., Ravzanaadii, N. and Rho, M. S. (2015). Macronutrient balance modulates the temperature-size rule in an ectotherm. *Amer. Nat.* **186**, 212–222. doi:10.1086/682072
- Lee, S., Hung, S. S. O., Fangue, N. A., Haller, L., Verhille, C. E., Zhao, J. and Todgham, A. E. (2016). Effects of feed restriction on the upper temperature tolerance and heat shock response in juvenile green and white sturgeon. *Comp. Biochem. Physiol. A* **198**, 87–95. doi:10.1016/j.cbpa.2016.04.016
- Lemoine, N. P., Giery, S. T. and Burkepile, D. E. (2014). Differing nutritional constraints of consumers across ecosystems. *Oecologia* **174**, 1367–1376. doi:10.1007/s00442-013-2860-z
- Little, A. G., Hardison, E., Kraskura, K., Dressler, T., Prystay, T. S., Hendriks, B., Pruitt, J. N., Farrell, A. P., Cooke, S. J., Patterson, D. A. et al. (2020a). Reduced lactate dehydrogenase activity in the heart and suppressed sex hormone levels are associated with female-biased mortality during thermal stress in Pacific salmon. *J. Exp. Biol.* **223**, jeb214841. doi:10.1242/jeb.214841
- Little, A. G., Dressler, T., Kraskura, K., Hardison, E., Hendriks, B., Prystay, T., Farrell, A. P., Cooke, S. J., Patterson, D. A., Hinch, S. G. et al. (2020b). Maxed out: optimizing accuracy, precision and power for field measures of maximum metabolic rate in fishes. *Physiol. Biochem. Zool.* **93**, 243–254. doi:10.1086/708673
- MacArthur, R. H. and Pianka, E. R. (1966). On optimal use of a patchy environment. *Amer. Nat.* **100**, 603–609. doi:10.1086/282454
- Magozzi, S. and Calosi, P. (2015). Integrating metabolic performance, thermal tolerance, and plasticity enables for more accurate predictions on species vulnerability to acute and chronic effects of global warming. *Glob. Chang. Biol.* **21**, 181–194. doi:10.1111/gcb.12695
- Malzahn, A. M., Doerfler, D. and Boersma, M. (2016). Junk food gets healthier when it's warm. *Limnol. Oceanogr.* **61**, 1677–1685. doi:10.1002/lno.10330
- McCue, M. D. (2006). Specific dynamic action: a century of investigation. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **144**, 381–394. doi:10.1016/j.cbpa.2006.03.011
- McDonald, D. G., McFarlane, W. J. and Milligan, C. L. (1998). Anaerobic capacity and swim performance of juvenile salmonids. *Can. J. Fish. Aquat. Sci.* **55**, 1198–1207. doi:10.1139/f98-002
- McKenzie, D. J. (2001). Effects of dietary fatty acids on the respiratory and cardiovascular physiology of fish. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **128**, 605–619. doi:10.1016/S1095-6433(00)00338-X
- Mishra, K. and Samantary, K. (2004). Interacting effects of dietary lipid level and temperature on growth, body composition and fatty acid profile of rohu, *Labeo rohita* (Hamilton). *Aquac. Nutr.* **10**, 359–369. doi:10.1111/j.1365-2095.2004.00311.x
- Mugge, V. M. R. (2008). Segmented: an R package to fit regression models with broken-line relationships. *R News* **8**, 20–25.
- Muñoz, N. J., Anttila, K., Chen, Z., Heath, J. W., Farrell, A. P. and Neff, B. D. (2014). Indirect genetic effects underlie oxygen-limited thermal tolerance within a coastal population of chinook salmon. *Proc. R. Soc. B Biol. Sci.* **281**, 20141082. doi:10.1098/rspb.2014.1082
- Pérez-Casanova, J. C., Lall, S. P. and Gamperl, A. K. (2010). Effects of dietary protein and lipid level, and water temperature, on the post-feeding oxygen

- consumption of Atlantic cod and haddock. *Aquac. Res.* **41**, 198–209. doi:10.1111/j.1365-2109.2009.02318.x
- Persson, J., Wojewodziec, M. W., Hessen, D. O. and Andersen, T. (2011). Increased risk of phosphorus limitation at higher temperatures for *Daphnia magna*. *Oecologia* **165**, 123–129. doi:10.1007/s00442-010-1756-4
- Pfenning, D. W. (1990). The adaptive significance of an environmentally-cued developmental switch in an anuran tadpole. *Oecologia* **85**, 101–107. doi:10.1007/BF00317349
- Pörtner, H. O. (2001). Climate change and temperature-dependent biogeography: oxygen limitation of thermal tolerance in animals. *Naturwissenschaften* **88**, 137–146. doi:10.1007/s001140100216
- Pörtner, H.-O. (2010). Oxygen- and capacity-limitation of thermal tolerance: a matrix for integrating climate-related stressor effects in marine ecosystems. *J. Exp. Biol.* **213**, 881–893. doi:10.1242/jeb.037523
- Prejs, A. (1984). Herbivory by temperate freshwater fishes and its consequences. *Environ. Biol. Fishes* **10**, 281–296. doi:10.1007/BF00001481
- Raubenheimer, D., Zemke-White, W. L., Phillips, R. J. and Clements, K. D. (2005). Algal macronutrients and food selection by the omnivorous marine fish *Girella trichoptera*. *Ecology* **86**, 2601–2610. doi:10.1890/04-1472
- Rho, M. S. and Lee, K. P. (2017). Temperature-driven plasticity in nutrient use and preference in an ectotherm. *Oecologia* **185**, 401–413. doi:10.1007/s00442-017-3959-4
- Rodgers, G. G., Tenzing, P. and Clark, T. D. (2016). Experimental methods in aquatic respirometry: the importance of mixing devices and accounting. *J. Fish. Biol.* **88**, 65–80. doi:10.1111/jfb.12848
- Rosewarne, P. J., Wilson, J. M. and Svendsen, J. C. (2016). Measuring maximum and standard metabolic rates using intermittent-flow respirometry: a student laboratory investigation of aerobic metabolic scope and environmental hypoxia in aquatic breathers. *J. Fish. Biol.* **88**, 265–283. doi:10.1111/jfb.12795
- Rowe, C. E., Figueira, W., Raubenheimer, D., Solon-Biet, S. M. and Capuska, G. E. M. (2018). Effects of temperature on macronutrient selection, metabolic and swimming performance of the Indo-Pacific Damselfish (*Abudefduf vaigiensis*). *Mar. Biol.* **165**, 178. doi:10.1007/s00227-018-3435-7
- Rubio, V. C., Sánchez-Vázquez, F. J. and Madrid, J. A. (2003). Macronutrient selection through postingestive signals in sea bass fed on gelatine capsules. *Physiol. Behav.* **78**, 795–803. doi:10.1016/S0031-9384(03)00082-9
- Rubio, V. C., Boluda Navarro, D., Madrid, J. A. and Sánchez-Vázquez, F. J. (2009). Macronutrient self-selection in *Solea senegalensis* fed macronutrient diets and challenged with dietary protein dilutions. *Aquaculture* **291**, 95–100. doi:10.1016/j.aquaculture.2009.02.040
- Ruiz, T., Koussoroplis, A.-M., Danger, M., Aguer, J.-P., Morel-Desrosiers, N. and Bec, A. (2020). U-shaped response Unifies views on temperature dependency of stoichiometric requirements. *Ecol. Lett.* **23**, 860–869. doi:10.1111/ele.13493
- Safi, H., Zhang, Y., Schulte, P. M. and Farrell, A. P. (2019). The effect of acute warming and thermal acclimation on maximum heart rate of the common killifish *Fundulus heteroclitus*. *J. Fish Biol.* **95**, 1441–1446. doi:10.1111/jfb.14159
- Sánchez-Vázquez, F. J., Yamamoto, T., Akiyama, T., Madrid, J. A. and Tabata, M. (1998). Selection of macronutrients by goldfish operating self-feeders. *Physiol. Behav.* **65**, 211–218. doi:10.1016/S0031-9384(98)00047-X
- Schmitz, O. J. and Rosenblatt, A. E. (2017). The temperature dependence of predation stress and prey nutritional stoichiometry. *Front. Ecol. Evol.* **5**, 1–8. doi:10.3389/fevo.2017.00073
- Schmitz, O. J., Rosenblatt, A. E. and Smylie, M. (2016). Temperature dependence of predation stress and the nutritional ecology of a generalist herbivore. *Ecology* **97**, 3119–3130. doi:10.1002/ecy.1524
- Schulte, P. M. (2015). The effects of temperature on aerobic metabolism: towards a mechanistic understanding of the responses of ectotherms to a changing environment. *J. Exp. Biol.* **218**, 1856–1866. doi:10.1242/jeb.118851
- Seebacher, F., White, C. R. and Franklin, C. E. (2015). Physiological plasticity increases resilience of ectothermic animals to climate change. *Nat. Clim. Chang.* **5**, 61–66. doi:10.1038/nclimate2457
- Sinclair, B. J., Marshall, K. E., Sewell, M. A., Levesque, D. L., Willett, C. S., Slotsbo, S., Dong, Y., Harley, C. D. G., Marshall, D. J., Helmuth, B. S. et al. (2016). Can we predict ectotherm responses to climate change using thermal performance curves and body temperatures? *Ecol. Lett.* **19**, 1372–1385. doi:10.1111/ele.12686
- Somero, G. N. (2010). The physiology of climate change: how potentials for acclimatization and genetic adaptation will determine “winners” and “losers”. *J. Exp. Biol.* **213**, 912–920. doi:10.1242/jeb.037473
- Somero, G. N. (2011). Comparative physiology: a “crystal ball” for predicting consequences of global change. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **301**, 1–14. doi:10.1152/ajpregu.00719.2010
- Starke, C. W. E., Jones, C. L. C., Burr, W. S. and Frost, P. C. (2020). Interactive effects of water temperature and stoichiometric food quality on *Daphnia pulex*. *Freshw. Biol.* **66**, 1–10. doi:10.1111/fwb.13633
- Stillman, J. H. (2003). Acclimation capacity underlies susceptibility to climate change. *Science* **301**, 65. doi:10.1126/science.1083073
- Turko, A. J., Nolan, C. B., Balshine, S., Scott, G. R. and Pitcher, T. E. (2020). Thermal tolerance depends on season, age and body condition in imperilled redeye dace *Clinostomus elongatus*. *Conserv. Physiol.* **8**, coaa062. doi:10.1093/conphys/coaa062
- Vagner, M., Zambonino-Infante, J.-L., Mazurais, D., Imbert-Auvray, N., Ouillon, N., Dubillot, E., Le Delliou, H., Akbar, D. and Lefrançois, C. (2014). Reduced n-3 highly unsaturated fatty acids dietary content expected with global change reduces the metabolic capacity of the golden grey mullet. *Mar. Biol.* **161**, 2547–2562. doi:10.1007/s00227-014-2526-3
- Vagner, M., Lacoue-Labarthe, T., Infante, J.-L. Z., Mazurais, D., Dubillot, E., Le Delliou, H., Quazuguel, P. and Lefrançois, C. (2015). Depletion of essential fatty acids in the food source affects aerobic capacities of the golden grey mullet *Liza aurata* in a warming seawater context. *PLoS ONE* **10**, e0126489. doi:10.1371/journal.pone.0126489
- Vagner, M., Pante, E., Viricel, A., Lacoue-Labarthe, T., Zambonino-Infante, J.-L., Quazuguel, P., Dubillot, E., Huet, V., Le Delliou, H., Lefrançois, C. et al. (2019). Ocean warming combined with lower omega-3 nutritional availability impairs the cardio-respiratory function of a marine fish. *J. Exp. Biol.* **222**, jeb187179. doi:10.1242/jeb.187179
- Vejříková, I., Vejřík, L., Syvāranta, J., Kiljunen, M., Čech, M., Blabolil, P., Vašek, M., Sajdlová, Z., Chung, S. H. T., Šmejkal, M. et al. (2016). Distribution of herbivorous fish is frozen by low temperature. *Sci. Rep.* **6**, 39600. doi:10.1038/srep39600
- Vinagre, C., Leal, I., Mendonça, V., Madeira, D., Narciso, L., Diniz, M. S. and Flores, A. A. V. (2016). Vulnerability to climate warming and acclimation capacity of tropical and temperate coastal organisms. *Ecol. Indic.* **62**, 317–327. doi:10.1016/j.ecolind.2015.11.010
- Wan, A. H. L., Davies, S. J., Soler-Vila, A., Fitzgerald, R. and Johnson, M. P. (2019). Macroalgae as a sustainable aquafeed ingredient. *Rev. Aquac.* **11**, 458–492. doi:10.1111/raq.12241
- Woiwode, J. G. and Adelman, I. R. (1992). Effects of starvation, oscillating temperatures, and photoperiod on the critical thermal maximum of hybrid striped×white bass. *J. Therm. Biol.* **17**, 271–275. doi:10.1016/0306-4565(92)90066-O
- Zhang, P., van Leeuwen, C. H. A., Bogers, D., Poelma, M., Xu, J. and Bakker, E. S. (2020). Ectothermic omnivores increase herbivory in response to rising temperature. *Oikos* **129**, 1028–1039. doi:10.1111/oik.07082