

Orexin 1 Receptor Antagonism in the Basolateral Amygdala Shifts the Balance From Pro- to Antistress Signaling and Behavior

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ABSTRACT

BACKGROUND: Stress produces differential behavioral responses through select molecular modifications to specific neurocircuitry elements. The orexin (Orx) system targets key components of this neurocircuitry in the basolateral amygdala (BLA).

METHODS: We assessed the contribution of intra-BLA Orx₁ receptors (Orx₁Rs) in the expression of stress-induced phenotypes of mice. Using the Stress Alternatives Model, a social stress paradigm that produces two behavioral phenotypes, we characterized the role of intra-BLA Orx₁R using acute pharmacological inhibition (SB-674042) and genetic knockdown (AAV-U6-Orx₁R-shRNA) strategies.

RESULTS: In the BLA, we observed that Orx₁R (*Hcrtr1*) messenger RNA is predominantly expressed in CamKII α ⁺ glutamatergic neurons and rarely in GABAergic (gamma-aminobutyric acidergic) cells. While there is a slight overlap in *Hcrtr1* and Orx₂ receptor (*Hcrtr2*) messenger RNA expression in the BLA, we find that these receptors are most often expressed in separate cells. Antagonism of intra-BLA Orx₁R after phenotype formation shifted behavioral expression from stress-sensitive (Stay) to stress-resilient (Escape) responses, an effect that was mimicked by genetic knockdown. Acute inhibition of Orx₁R in the BLA also reduced contextual and cued fear freezing responses in Stay animals. This phenotype-specific behavioral change was accompanied by biased molecular transcription favoring *Hcrtr2* over *Hcrtr1* and *Mapk3* over *Plcb1* cell signaling cascades and enhanced *Bdnf* messenger RNA.

CONCLUSIONS: Functional reorganization of intra-BLA gene expression is produced by antagonism of Orx₁R, which promotes elevated *Hcrtr2*, greater *Mapk3*, and increased *Bdnf* expression. Together, these results provide evidence for a receptor-driven mechanism that balances pro- and antistress responses within the BLA.

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Stress-induced alterations in neurocircuitry result in divergent behavioral responses. Enhanced stress reactivity (prostress) in rodent models is similar to human affective dysfunction in mood disorders such as depression, fear- and anxiety-related disorders, or posttraumatic stress disorder (1). Current pharmacotherapies for affective disorders have had limited success, and a mechanistic understanding remains elusive.

Balance within key stress circuits may be disrupted during periods of intense or prolonged stress to shift signaling dynamics in pro- or antistress pathways (2–4). Stressful stimuli are interpreted, in part, through converging signals in the basolateral amygdala (BLA), where glutamatergic projection neurons are influenced by distinctive GABAergic (gamma-aminobutyric acidergic) interneurons, to direct behavioral responses (5). In addition, activity in the BLA is modified by hypothalamic orexinergic neurons, which are critical for panic (6,7) and motivation (8,9).

Orexin (Orx) A and Orx_B, neuromodulators derived from a single pre-propeptide, activate two G protein-coupled receptors: Orx₁ receptors (Orx₁Rs) bind Orx_A and Orx_B (half maximal effective concentration = 30 vs. 2500 nM), as do Orx₂Rs (half maximal effective concentration = 38 vs. 36 nM) (10,11). These receptors stimulate G_q proteins, which increase intracellular Ca²⁺ (11) by activating phospholipase C (PLC) pathways (12). The PLC β_1 isozyme variant is transcribed in the amygdala (13), and its dysfunction is linked to psychopathologies such as depression (14), bipolar disorder (15), addiction (16), and schizophrenia (17,18).

Stimulation of Orx₁R can also activate ERK (extracellular signal-regulated protein kinase). In the amygdala, recruitment of ERKs is important for consolidation, reconsolidation, and extinction of fear memories (19,20). While Orx₁Rs in the BLA are important in regulating fear (21,22), depression (23,24), and anxiety (25), it is unclear how shifts in molecular signaling

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cascades mediate such responses and initiate stress-induced phenotype development.

Using the Stress Alternatives Model (SAM), a behavioral paradigm that separates individuals into social stress-resilient (Escape; as validated by social interaction/preference test) and stress-vulnerable (Stay) (26) populations, we explored how Orx₁R activity in the BLA is involved in the formation of stress-related phenotypes. As a social interaction and avoidance paradigm in which smaller subjects encounter intense attacks from larger novel aggressors over a 4-day period, the SAM produces two separate subsets of animals exhibiting social avoidance or enhanced fear-conditioned responses (27,28). Unlike a traditional social defeat outcome, the SAM provides mice an opportunity to avoid social aggression by exiting the arena through escape tunnels only large enough for the smaller mouse. By the end of the second day of social interaction, test subjects commit to a phenotype: Escape or Stay. These stable phenotypes may be altered through pharmacological manipulations (Escape reduced by anxiogenic drugs, Stay reduced by anxiolytic drugs) administered on the third day of the SAM (28–30). Thus, the SAM is a useful tool for studying the development of stress-induced phenotypes while providing an opportunity to explore physiological and clinically relevant molecular mechanisms.

We investigated if inhibition of intra-BLA Orx₁Rs alters the formation of social stress-induced behavioral phenotypes. We predicted that pharmacological inhibition or genetic knock-down will shift behavioral patterns in vulnerable (Stay) populations toward resilience (Escape). Furthermore, we explored

if Orx₁R inhibition affects conditioned fear responses and alters expression of genes responsible for balancing signaling in pro- and antistress neurocircuitries. Together, these results allow us to propose a neurocircuit model that defines the role of intra-BLA Orx₁R signaling in the balance of pro- and antistress states.

METHODS AND MATERIALS

Social Stress and Choice Paradigm

Aggressive social interactions between larger novel CD1 and smaller male C57BL/6NHsd mice dyads in the SAM apparatus (Figure 1) involve four trials, lasting up to 5 minutes each, that allow test animals the opportunity to shorten stressful encounters by making use of size-restricted tunnels at the apical end of the oval open field interaction arena. A tone given during isolation in the SAM apparatus before social interaction permits comparisons between cued and contextual fear conditioning. The escape routes provide a choice, producing two stable phenotypes: active avoidance (Escape) and accepting confrontation (Stay), which may be modified by drug treatment on day 3. The treatment regimen allows for statistical comparisons between groups and within subjects by comparing responses to SAM interactions before and after treatment. All procedures were performed in accordance with the Guide for the Care and Use of Laboratory Animals (NIH Publications No. 80-23) and approved by the University of South Dakota Institutional Animal Care and Use Committee.

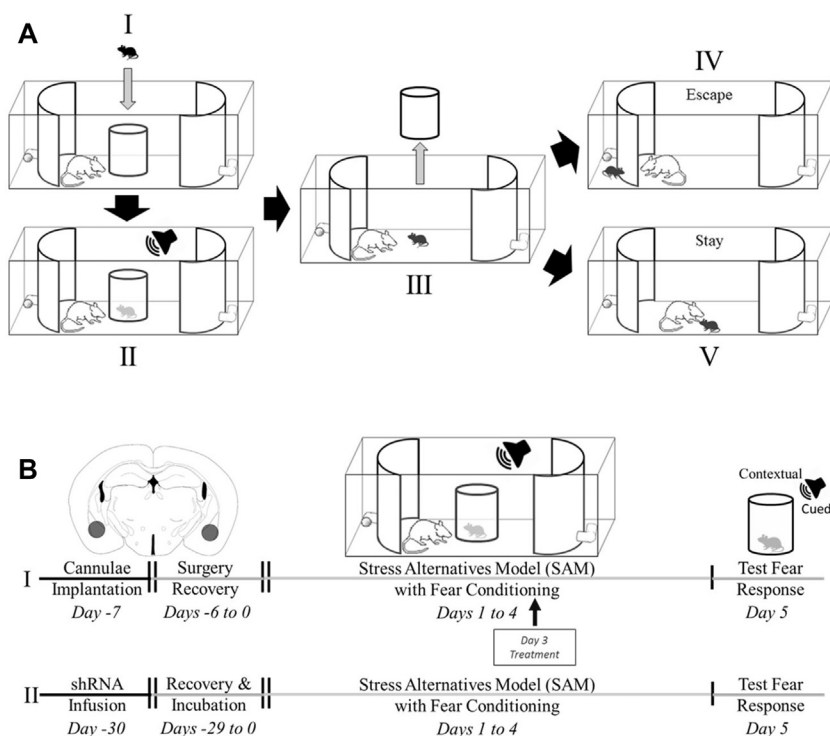


Figure 1. The SAM is used to assess the development of stress-induced phenotypes. **(A)** The SAM is a 4-day behavioral paradigm in which (I) a test mouse is placed into an opaque cylinder, (II) presented a tone, (III) exposed to social aggression, and commits to a phenotype: (IV) Escape or (V) Stay. **(B)** The behavioral timelines for (I) pharmacology and (II) genetic knockdown experiments (mice are the same age at testing) include surgeries targeting the basolateral amygdala, SAM exposure (days 1–4), and the testing of contextual and cued fear responses (day 5). SAM, Stress Alternatives Model; shRNA, short hairpin RNA.

Experimental Overview

See the [Supplement](#) for further information. The primary treatments for these experiments are inhibition of BLA Orx₁R via the antagonist SB-674042 (0.3 nmol/0.3 μ L delivered bilaterally intra-BLA 1 hour prior to interaction on day 3), contrasted with Orx₁R stimulation (accomplished by Orx_A+Orx₂R antagonism), or short hairpin RNA (shRNA) knock-down (bilateral intra-BLA transfection beginning 30 days prior to SAM interaction). Considering the difference in timing of delivery, these treatments were done and analyzed separately with a priori hypotheses. All behavioral measures were performed during the dark cycle when the animals were active and included Escape (use of the apical tunnels), Stay (remaining in the SAM arena with the novel aggressor), time spent attentive to the escape hole, latency to escape (for Escape mice), fear-conditioned freezing (measured in response to the tone [conditioned stimulus (CS)] and context prior to the social interaction unconditioned stimulus [US] and as a conditioned response [CR] on day 5 in the absence of the US), and food intake. Thus, treatment groups included home-cage control animals and intra-BLA SB-674042 (or vehicle, Orx_A, Orx_A+MK-1064, MK-1064) injection of Escape and Stay mice. In addition, transgenic treatment groups included home-cage control animals, intra-BLA AAV-Orx₁R-shRNA injection, and intra-BLA AAV-scramble-shRNA injection. Brains and blood were collected for visual representations of gene expression (using RNAscope) of *Hcrt1*, *Hcrt2*, *Calb1*, *CamkII α* , *Gad1*, and *Pvalb* in the BLA and to measure plasma concentrations of the stress hormone corticosterone (by enzyme-linked immunosorbent assay). Gene expression (using reverse transcriptase quantitative polymerase chain reaction) of *Hcrt1*, *Hcrt2*, *Plcb1*, *Mapk1*, *Mapk3*, *Bdnf*, and *Gapdh* (housekeeping gene) was measured in the BLA tissue. All experimental designs and statistical analyses were based on a priori hypotheses, using two-way repeated-measures analysis of variance, two-way analysis of variance, one-way analysis of variance, regression analyses, and *t* test, followed (where appropriate) by post hoc analyses.

RESULTS

OrxR Expression in the BLA

The glutamatergic marker CamKII α identified the vast majority of BLA neurons (~80%) ([Figure S2](#)) and those expressing *Hcrt1* (31,32) (also in some calbindin-GABAergic neurons) ([Figure 2](#)). Few (<20%) BLA *Hcrt1*-possessing cells express *Gad1* (GABAergic marker) and co-express parvalbumin (~10%) ([Figure 2G–K](#)). Our results suggest that *Hcrt1* is expressed in 10% to 15% of BLA glutamatergic neurons and ~5% of GABA cells ([Figure 2K](#)). In BLA cells, messenger RNA (mRNA) for *Hcrt1* and *Hcrt2* largely does not overlap, and ~80% of *Hcrt1*⁺ cells do not co-express *Hcrt2* ([Figure 2L–O](#)). Specific BLA GABAergic neurons may predominantly localize Orx₂R ([Figure 2P](#)) (28).

Motivation for Active Avoidance (Escape)

In the SAM, animals evenly choose one of two stable (27–29,33) behavioral phenotypes, Escape (44.7%) or Stay (55.3%) ([Figure 1A](#) and [Figure S1B, C](#)) (26,27,29,33,34). Time

spent investigating escape routes predicts active avoidance and indicates motivation to escape (28). Time spent attentive to the hole was significantly greater in vehicle-treated Escape mice ([Figure 3A](#)), but intra-BLA infusion of the Orx₁R antagonist (Escape: [Figure 3B, C](#); Stay: [Figure 3B, D](#)) or AAV-U6-Orx₁R-shRNA ([Figure 3E](#)) increases attention to the escape route. Furthermore, receptor activation with Orx_A reduced time that Escape mice spent investigating the escape route ([Figure S3](#)).

Avoidance (Escape)

Upon intra-BLA injections of an Orx₁R antagonist on SAM day 3, a substantial number of Stay mice exhibited Escape behavior ([Figure 4A](#)), with a 30% shift that day and a significant increase the day after (day 4 = 70% increase). Intra-BLA activation of both Orx receptors with Orx_A or biased activation of Orx₁R (Orx_A+Orx₂R antagonist) blocked Escape behavior in a small, but not statistically significant, proportion of mice on days 3 and 4 ([Figure S4](#)), exhibiting deviation from stable phenotype behavioral patterns.

Because knockdown reduced Orx₁R expression prior to stressful interactions, we did not expect a dramatic change in behavior over the course of SAM trials, but AAV-U6-Orx₁R-shRNA yielded incrementally (although not significantly) more escape on the last 2 days ([Figure 4B](#)). By day 4, 72.7% of AAV-U6-Orx₁R-shRNA-treated mice displayed Escape compared with 54.5% of the scramble control mice.

Escape mice spent significantly less time in the SAM arena with the CD1 mouse on days 2 to 4 (26,27,29,33,35); therefore, escape latency was reduced ([Figure 4C](#)). Stay mice remained for the entire 5-minute period unless treated with Orx₁R antagonist, significantly reducing time spent with aggressive CD1 mice on day 4 ([Figure 4D, F](#)). Inhibition of Orx₁R did not influence escape latency in Escape animals ([Figure 4D](#)). Neither of the Orx₁R manipulations, antagonist or knockdown treatments, influenced arousal/locomotion ([Figure S5](#)) but did result in small but significant decreases in food intake and body weight ([Figure S6](#)).

Cued and Contextual Fear Conditioning

Cued fear responses significantly enhanced freezing in both Escape and Stay phenotypes, and Stay mice displayed heightened freezing behavior to context (opaque cylinder divider) as well ([Figure 5A, B](#)). Although inhibition of intra-BLA Orx₁R did not affect the fear freezing profile in Escape mice, antagonist-treated Stay mice exhibited significantly reduced contextual and cued fear responses ([Figure 5B](#) and [Figure S7; Table S1](#)). Similar to mice of the Escape phenotype, knockdown of BLA Orx₁R did not affect conditioned freezing behavior ([Figure S8](#)). Activation of intra-BLA Orx receptors with Orx_A did not change the fear freezing profile in Escape or Stay mice compared with vehicle ([Figure 5B](#) and [Figure S7; Table S1](#)). However, biased stimulation of Orx₁R in the BLA with a combination of Orx_A+Orx₂R antagonist eliminated the CR in Escape, but not Stay, mice ([Figure 5B](#) and [Figure S7; Table S1](#)). Furthermore, acute inhibition of Orx₂R in the BLA eliminated the cued freezing response in Escape mice and significantly reduced freezing during the post-tone period ([Figure 5B](#) and [Figure S7; Table S1](#)). Stay mice treated with an

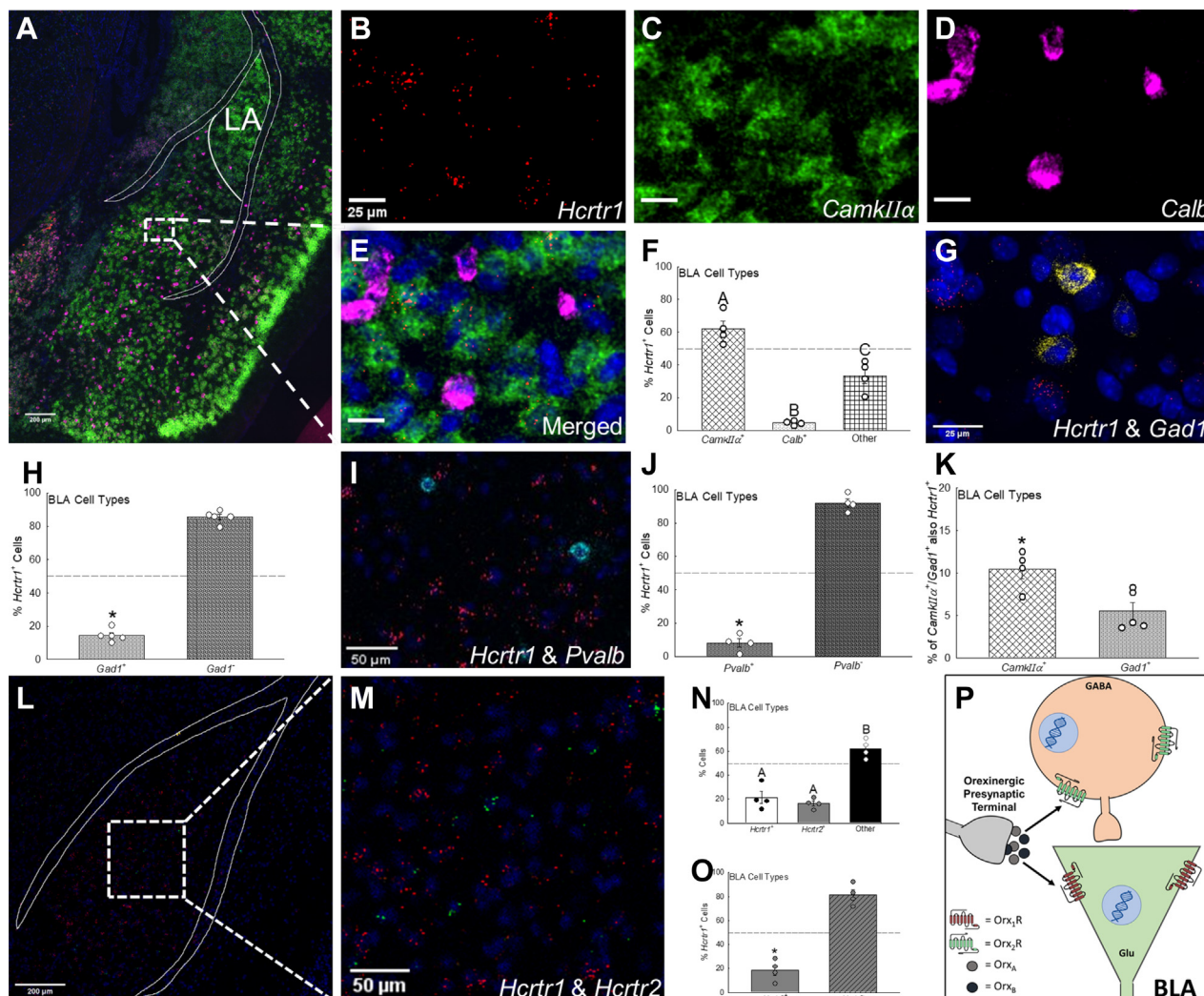


Figure 2. In the untreated BLA, Orx₁Rs are expressed predominantly in glutamatergic neurons and are rarely co-expressed with Orx₂Rs. (A) Imaged sections containing BLA cells stained with probes targeting mRNA of (B) *Hcrt1* (red), (C) *CamkIIα* (green), and (D) *Calb* (magenta) revealed when (E) merged (with DAPI) that (F) Orx₁R⁺ cells mostly co-express the glutamatergic cell marker *CamkIIα* ($n = 4$, $F_{2,9} = 54.4$, $p < .001$; *CamkIIα*⁺ vs. *Calb*⁺: $t_6 = 10.4$, $p < .001$; *CamkIIα*⁺ vs. other: $t_6 = 5.2$, $p < .001$; bars are statistically different from one another as illustrated with unique letters, e.g., A is significantly different from B and C; $p < .001$). (G) Expression of *Hcrt1* (red) and GAD₆₇ (*Gad1*) mRNA (yellow) infrequently overlap with (H) most *Hcrt1*⁺ cells being absent of the GABAergic marker ($n = 5$, $t_8 = 29.5$, $p < .001$). (I) While a subset of BLA GABAergic neurons produce the calcium-binding protein parvalbumin (*Pvalb*⁺), (J) *Hcrt1*⁺ (red) cells are mostly absent of *Pvalb* expression (light blue) with (K) <10% being both *Hcrt1*⁺ and *Pvalb*⁺ ($n = 4$, $t_6 = 23.1$, $p < .001$). (L) Furthermore, more BLA glutamatergic (*CamkIIα*⁺) neurons (compared with GABAergic → *GAD1*⁺) also express *Hcrt1* ($n = 9$, $t_7 = 3.2$, $p < .015$). (M) Images of BLA cells with fluorescent markers labeling (N) *Hcrt1* mRNA (red) and *Hcrt2* mRNA (green) demonstrate that (O) most BLA cells express neither *Hcrt1* nor *Hcrt2* ($n = 4$, $F_{2,9} = 42.1$, $p < .001$; *Hcrt1*⁺ vs. other, $t_6 = 7.5$, $p < .001$; *Hcrt2*⁺ vs. other, $t_6 = 8.4$, $p < .001$; bars are statistically different from one another as illustrated with unique letters, e.g., A is significantly different from B). (P) Most *Hcrt1*⁺ cells in the BLA do not express *Hcrt2* ($n = 4$, $t_6 = 10.1$, $p < .001$), as depicted in (P) showing Orx₁R on glutamatergic neurons. BLA, basolateral amygdala; GABA, gamma-aminobutyric acid; Glu, glutamatergic neuron; LA, lateral amygdala; mRNA, messenger RNA; Orx₁R, orexin 1 receptor; Orx₂R, orexin 2 receptor.

Orx₂R antagonist displayed no statistical differences in the levels of contextual and cued freezing (Figure 5B and Figure S7; Table S1).

Corticosterone Concentrations

Social stress in SAM interactions increases corticosterone concentrations in both Escape and Stay animals (27,28,33),

although Stay mice have higher levels of corticosterone than Escape mice. Inhibition of BLA Orx₁R decreased Stay corticosterone concentrations compared with vehicle-treated Stay animals and did not differ significantly from nonstressed mice (Figure 5C). Treatments with Orx_A or the combination of Orx_A and an Orx₂R antagonist did not change corticosterone levels relative to vehicle-treated control mice; however, the differences between Escape and Stay were eliminated and levels

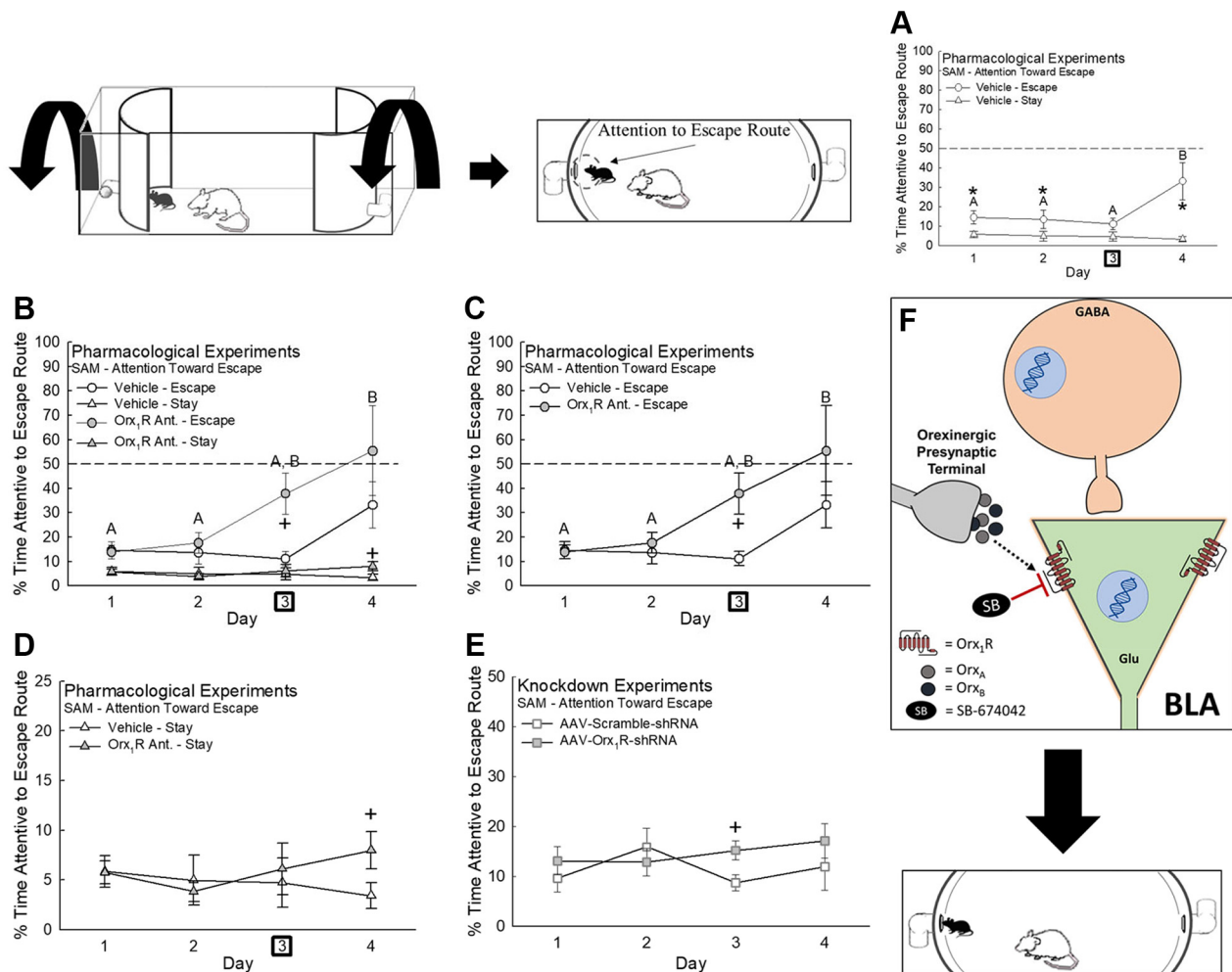


Figure 3. Motivation toward Escape behavior is effected through inhibition of intra-BLA Orx₁Rs. **(A)** Escape mice, as compared with those expressing the Stay phenotype, spend a greater percentage of time investigating the SAM escape routes ($n = 19$; phenotype effect: $F_{1,51} = 16.4$, $p < .001$; Escape vs. Stay: day 1, $t_{17} = 2.6$, $^*p \leq .018$; day 2, $t_{17} = 2.5$, $^*p \leq .017$; day 4, $t_{17} = 4.2$, $^*p < .001$). **(B)** While Escape mice, in general, explore the escape routes more often, **(C)** inhibition of intra-BLA Orx₁R promotes even more attention toward the escape tunnels ($n = 34$; treatment effect: $F_{1,30} = 7.7$, $p \leq .019$; day 3 vehicle escape vs. Orx₁R Ant. Escape, $t_{10} = 2.5$, $^*p \leq .018$). **(D)** Antagonism of intra-BLA Orx₁R only slightly stimulates escape route exploration in Stay mice (day 4 vehicle \times Orx₁R Ant., $t_{20} = 2.1$, $^+p \leq .05$). **(E)** Knockdown of intra-BLA Orx₁R temporarily and minimally increases attention toward escape on day 3 of the SAM ($n = 22$; day 3 scramble vs. AAV-Orx₁R-shRNA, $t_{20} = 2.4$, $^+p \leq .024$). **(F)** Illustration demonstrating that inhibition of intra-BLA Orx₁Rs predominantly on glutamatergic neurons promotes attention toward the escape route in the SAM arena. In pharmacological experiments, drug treatment is administered on day 3 as designated by the bold square. Note that the data plotted in panels **(A)**, **(C)**, and **(D)** are the same as those graphed in panels **(B)**; we have separated out these individual comparisons for the sake of clarity. Ant., antagonist; BLA, basolateral amygdala; GABA, gamma-aminobutyric acid neuron; Glu, glutamatergic neuron; Orx₁R, orexin 1 receptor; Orx_A, orexin A; Orx_B, orexin B; SAM, Stress Alternatives Model.

were elevated compared with Orx₁R antagonist-treated mice (Figure 5C). Inhibition of BLA Orx₁R not only reduces social fear responses but also reverses social stress responsiveness.

Antagonism of Intra-BLA OrxR Recruits Alternative Signaling

Although *Hcrt1* expression was unaltered following vehicle treatment, Orx₁R antagonism reduced intra-BLA *Hcrt1* in Escape mice compared with nonstressed cage control mice (Figure 6A) and simultaneously elevated *Hcrt2* expression in

Stay mice compared with Escape and vehicle-treated Stay mice (Figure 6B; Table S2). In vehicle control animals, *Hcrt2* expression was higher in Escape mice than both Stay and Orx₁R antagonist-treated Escape mice (Figure 6B; Table S2). A reduction in *Hcrt1* gene expression after Orx₂R antagonism was observed, but only in Stay animals relative to vehicle (Figure 6A; Table S2). Expression of *Hcrt2* in the BLA was reduced in both Escape and Stay phenotypes after blocking Orx₂R, contrasting with Orx₁R antagonism, which enhanced *Hcrt2* mRNA levels in Stay mice (Figure 6B; Table S2).

Transcription of BLA PLC β_1 (*Plcb1*) mRNA (13) is important for Orx₁R signaling (36). We predicted that Orx₁R antagonist

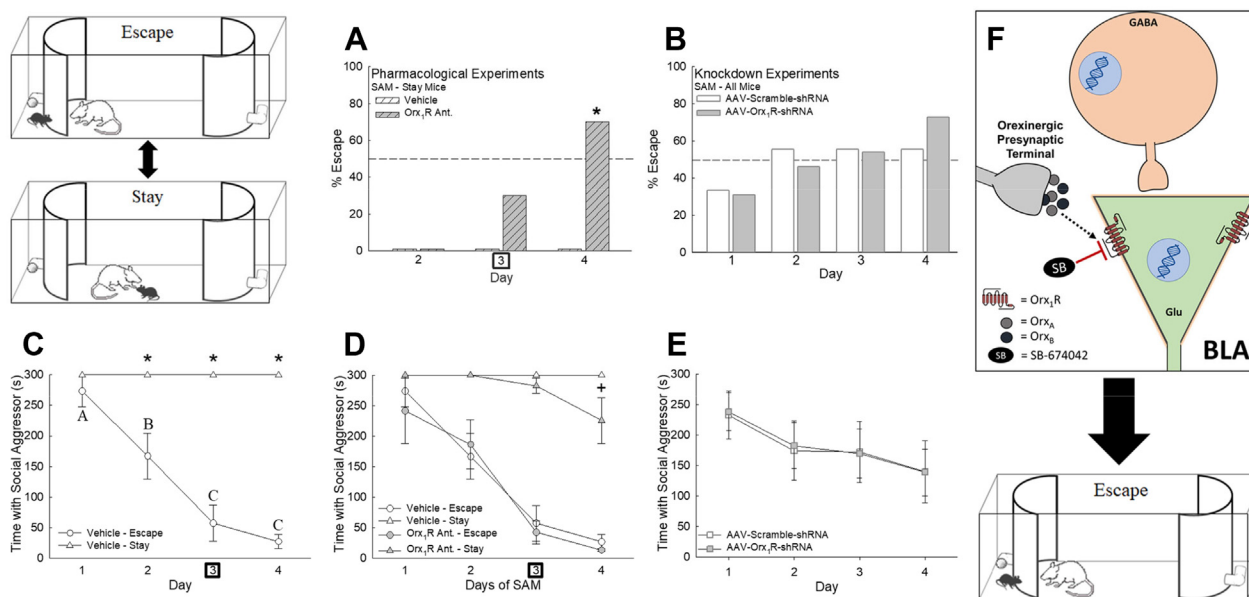


Figure 4. Intra-BLA Orx₁Rs mediate stress-related behavioral phenotype development. **(A)** Infusion of an Orx₁R Ant. (SB-674042) into the BLA promotes escape behavior in Stay mice ($n = 22$; day 4, χ^2 : $F_{1,45} = 9.3$, $p < .001$). **(B)** Knockdown of Orx₁R (AAV-Orx₁R-shRNA) upsets normal day 2 phenotype commitment behavior (as observed with AAV-Scramble-shRNA controls), inducing more escape behavior on days 3 and 4 ($n = 22$). **(C)** Escape animals learn to efficiently use the escape route to avoid social aggression (escape latency = time with social aggressor) over the course of 4 days while Stay mice remain with the aggressor ($n = 19$; phenotype effect: $F_{1,45} = 175.3$, $p < .001$; time effect: $F_{3,45} = 26.1$, $p < .001$; interaction effect: $F_{3,45} = 26.1$, $p < .001$; Escape vs. Stay: day 2, $t_{17} = 5.8$, $p < .001$; day 3, $t_{17} = 10.6$, $p < .001$; day 4, $t_{17} = 11.9$, $p < .001$; within-Escape phenotype comparison, $F_{3,18} = 17.8$, $p < .001$, day 1 vs. day 3, $t_6 = 5.7$, $p < .001$; day 1 vs. day 4, $t_6 = 6.5$, $p < .001$; day 2 vs. day 3, $t_6 = 2.9$, $p \leq .009$; day 2 vs. day 4, $t_6 = 3.7$, $p \leq .002$; $p < .05$ for days marked with unique lettering, e.g., A is different from B and C). **(D)** Antagonizing intra-BLA Orx₁Rs promotes aggressor avoidance in Stay mice ($n = 34$; time effect: $F_{3,54} = 2.9$, $p \leq .043$; interaction effect: $F_{3,54} = 2.9$, $p \leq .043$; day 4 vehicle Stay vs. Orx₁R Ant. Stay, $t_{20} = 3.4$, $p < .001$) but has no effect on those animals exhibiting the Escape phenotype. **(E)** Knockdown of intra-BLA Orx₁R does not affect the overall latency of aggressor avoidance ($n = 22$). Overall, **(F)** inhibition of Orx₁R in the BLA appears to prompt escape behavior. In pharmacological experiments, drug treatment is administered on day 3 as designated by the bold square. Ant., antagonist; BLA, basolateral amygdala; GABA, gamma-aminobutyric acid; Glu, glutamatergic neuron; Orx₁R, orexin 1 receptor.

might limit *Pcb1* expression levels (Figure 6C). Escape mice in both vehicle and Orx₁R antagonist groups expressed lower amounts of *Pcb1* than Stay and cage control animals (Figure 6C). Furthermore, greater *Pcb1* followed intra-BLA Orx₂R inhibition compared with vehicle-treated Escape mice (Figure 6C; Table S2).

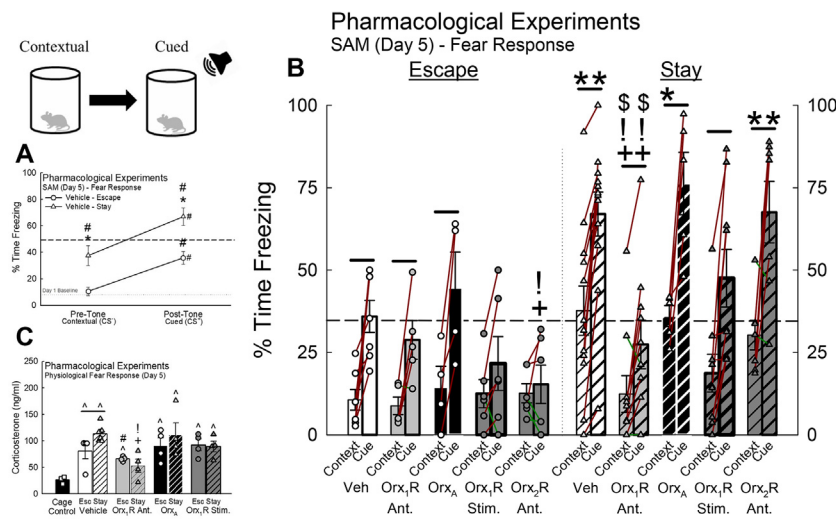
Alternative molecular pathways recruited during G_q activation are driven by ERK genes (*Mapk1* and *Mapk3*). In Stay mice, Orx₁R antagonism resulted in a significant increase in *Mapk3* expression (*Mapk1* mRNA was unaffected) (Figure S9) compared with similarly treated Escape, vehicle-treated Stay, and nonstressed cage control mice (Figure 6D; Table S2). Inhibition of intra-BLA Orx₂R did not alter *Mapk3* gene expression (Figure 6D; Table S2).

The transcription of BDNF (*Bdnf* [brain-derived neurotrophic factor]) is tied to neuroplasticity (37,38) and behavioral changes such as extinction of fear memories (39), so we predicted that an increase in *Bdnf* might be associated with intra-BLA Orx₁R inhibition (Figure 6E). As hypothesized, intra-BLA Orx₁R antagonism resulted in elevated *Bdnf* in Stay mice compared with Escape mice and vehicle-treated Stay mice (Figure 6E and Table S2). Finally, Orx₂R antagonist treatment enhanced *Bdnf* expression in Escape mice while diminishing transcription in Stay animals, an effect that is phenotypically opposite to that observed after Orx₁R inhibition (Figure 6E;

Table S2). Because Stay mice treated with an Orx₁R antagonist experienced shifts from stress-vulnerable to stress-resilient behavioral responses, the alterations in gene expression reported here (Figure 6F, G) may be implicit in this behavioral plasticity.

Molecular Restructuring Is Related to Fear Responsiveness

Expression levels of *Hcrt2*, but not *Hcrt1*, in both vehicle-treated and Orx₁R antagonist-treated mice are negatively correlated with cued freezing (Figure 7A, B). Relative expression levels of *Pcb1* were positively correlated with cued freezing behavior in vehicle-treated mice (Figure 7C); however, this relationship is not observed after intra-BLA Orx₁R inhibition (Figure 7D). Contextual freezing behavior was associated with *Mapk3* expression in only vehicle-treated mice (Figure S10). By contrast, intra-BLA antagonism of Orx₁R cued freezing behavior was negatively correlated to *Mapk3* expression (Figure 7F), but not in vehicle-treated mice (Figure 7E). The lack of gene expression correlations with cued fear freezing when phenotypes were assessed independently (Figure S11) indicates that behavioral and transcriptional relationships exist within collective operational adaptations that link behavioral change to molecular modification. No



parisons. (C) Mice exposed to social stress produce elevated levels of stress hormone ($n = 39$, $F_{2,12} = 24.3$, $p < .001$; cage control vs. vehicle Escape, $t_5 = 3.1$, $p \leq .028$; cage control vs. vehicle Stay, $t_9 = 9.9$, $p < .001$); however, Stay animals have the highest concentration (vehicle Escape vs. Stay, $t_{10} = 2.6$, $p \leq .025$). Inhibition of intra-BLA Orx₁R reduces corticosterone levels in Stay mice (vehicle Stay vs. Orx₁R Ant. Stay, $t_{10} = 5.1$, $p < .001$; Orx₁R Ant. Stay vs. Orx_A Stay, $t_6 = 3.3$, $p \leq .002$). Ant., antagonist; BLA, basolateral amygdala; CS, conditioned stimulus; Esc, escape; Orx₁R, orexin 1 receptor; Orx₂R, orexin 2 receptor; Orx_A, orexin A; SAM, Stress Alternatives Model; Stim., stimulation; Veh, vehicle.

relationships between gene expression and conditioned fear freezing were observed for any of the tested cell signaling markers after Orx₂R antagonism (not Orx₁R antagonism) except for *Bdnf*, in which a significant negative correlation was revealed (Figure S12E). Together, these results suggest a functional connection between Orx₁R antagonist-induced shifts in gene expression and fear-related behaviors.

Potential Molecular Mechanism Behind Intra-BLA Orx₁R Antagonism

To help generate a theoretical mechanism to explain the physiological basis surrounding the observed behavioral and phenotypic shifts resulting from intra-BLA inhibition of Orx₁R, we explored transcriptional relationships in systems that exhibited similar regression patterns (Figure 8). With antagonism of Orx₁R, there is a strongly positive relationship between *Hcrtr2* and *Mapk3* expression (Figure 8A). This association does not exist after vehicle or Orx₂R antagonist treatment (Figure S13). While there are no observed relationships between *Bdnf* and *Hcrtr2* expression levels (Figure 8B and Figure S13), *Bdnf* expression is positively correlated to *Mapk3* expression in animals treated with an Orx₁R antagonist (Figure 8C). Notably, no relationships exist between *Hcrtr1* expression and the other genes of interest (Figure S13). These data allowed us to predict a working model to explain how BLA Orx₁R may function to establish behavioral patterns consistent with stress-induced phenotype development (Figure 9).

DISCUSSION

Antagonism of Orx₁R in the BLA can reverse or diminish expression of stress-related behavior. Our results suggest that BLA Orx₁R play a central role in stress responsiveness (40,41) and related behavioral, physiological, and molecular outcomes that are important components of affective disorders (42,43),

such as anxiety (7), depression, and posttraumatic stress disorder. Acute inhibition of intra-BLA Orx₁R promotes Escape over Stay responses and limits freezing during fear conditioning in a phenotype-dependent way. Furthermore, inhibition of Orx₁R alters gene expression associated with critical signaling cascades. Following intra-BLA Orx₁R antagonism, transcription for receptors and intracellular signaling becomes biased toward Orx₂R (*Hcrtr2*) over Orx₁R (*Hcrtr1*) and ERK1 (*Mapk3*) over PLC β 1 (*Plcb1*) pathways. Even when BLA Orx₁R are inhibited, native Orx_A and Orx_B will bind Orx₂R. The relationship of these behavioral and molecular changes to enhanced expression of *Hcrtr2* mRNA, potentially in BLA neurons that do not contain Orx₁R (Figure 2L–O), suggests receptor-mediated mechanisms that balance pro- and anti-stress responses in BLA microcircuits.

Aggressive social interactions in SAM produced two behavioral phenotypes that represent risk assessment and choice: Escape and Stay. These phenotypes, similar to those exposed to social defeat paradigms (44,45), exhibit resilience (tightly linked to Escape) and susceptibility (highly correlated with Stay) in the social interaction/preference test (28). However, unlike traditional social defeat, SAM-separated phenotypes are expressed early in the behavioral paradigm, providing insight into the development and progression of stress-induced behavior and pathophysiology. Anxiolytic drugs (such as CRF₁ receptor antagonist antalarmin and the Orx₂R agonist [Ala¹¹, d-Leu¹⁵]-Orx_B) promote escape, while anxiogenic drugs (such as the α_2 antagonist yohimbine and the Orx₂R antagonist MK-1064) delay and/or block escape behavior (28,29). Surprisingly, neither the Orx₁R antagonist (Figure 4D) nor knockdown (Figure 4E) influenced escape latency, although it is reduced by anxiolytic factors such as exercise, neuropeptide S, and antalarmin and increased by anxiogenic factors such as yohimbine (29). We posit that enhanced escape on day 4, following BLA Orx₁R inhibition (on

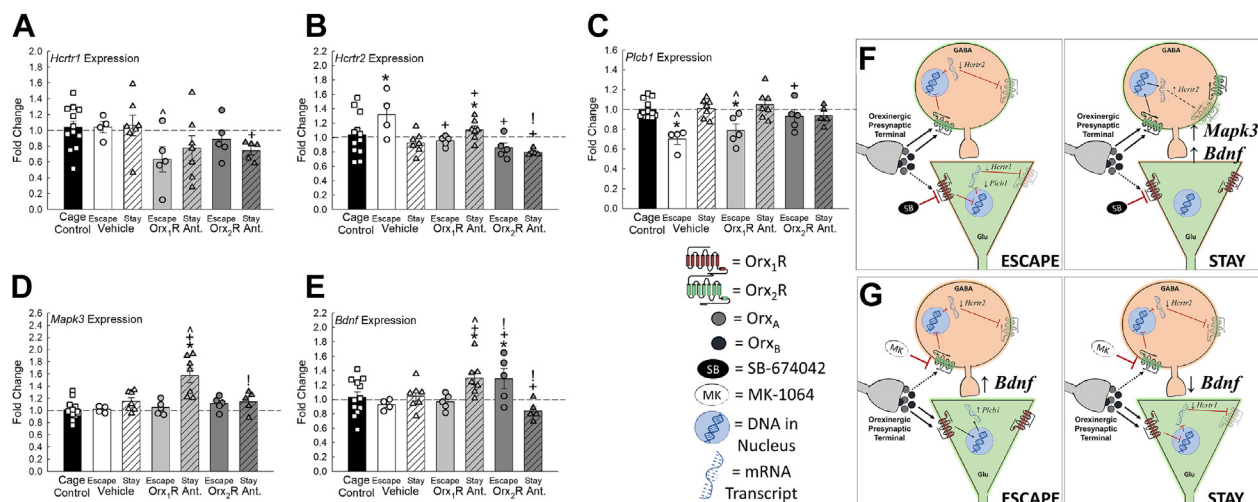


Figure 6. Transcriptional changes (relative to home-cage naïve controls) in BLA after Orx₁R or Orx₂R antagonism shifts signaling profile. **(A)** Antagonism of Orx₁R in the BLA reduces *Hcrtr1* expression ($n = 45$; treatment effect: $F_{2,27} = 3.5$, $p \leq .043$), but only significantly so in animals expressing the Escape phenotype (cage control vs. Orx₁R Ant. Escape, $t_{11} = 2.2$, $\hat{p} \leq .050$), whereas infusion of an Orx₂R Ant. in the BLA reduces *Hcrtr1* expression in Stay mice compared with vehicle animals of the same phenotype ($t_{10} = 2.2$, $\hat{p} \leq .044$). **(B)** While Escape mice (treatment effect: $F_{2,27} = 9.8$, $p < .001$; interaction effect: $F_{2,27} = 8.6$, $p < .001$) treated with vehicle express higher *Hcrtr2* levels than Stay mice ($t_9 = 3.0$; $\hat{p} \leq .016$) and Orx₁R or Orx₂R Ant.-treated Escape animals (vehicle vs. Orx₁R Ant., $t_7 = 2.6$, $\hat{p} \leq .035$; vehicle vs. Orx₂R Ant.: $t_7 = 4.5$, $\hat{p} < .001$; Orx₁R Ant. vs. Orx₂R Ant.: $t_8 = 3.5$, $\hat{p} < .001$), Orx₁R antagonism results in elevated levels (Escape vs. Stay, $t_{10} = 2.2$, $\hat{p} \leq .05$; vehicle vs. Orx₁R Ant., $t_{12} = 2.4$, $\hat{p} \leq .034$) while Orx₂R inhibition leads to a reduction (vehicle vs. Orx₂R Ant.: $t_{10} = 3.5$, $\hat{p} \leq .002$; Orx₁R Ant. vs. Orx₂R Ant.: $t_{10} = 4.7$, $\hat{p} < .001$) of *Hcrtr2* in Stay mice. **(C)** A reduction of *Plcb1* (phenotype effect: $F_{1,27} = 19.1$, $p < .001$; interaction effect: $F_{2,27} = 4.3$, $p \leq .023$) that is found in Escape mice under control conditions (cage control vs. vehicle Escape, $t_{10} = 5.1$, $\hat{p} < .001$; Escape vs. Stay, $t_9 = 5.0$, $\hat{p} < .001$) and Orx₁R antagonism (Escape vs. Stay, $t_{10} = 3.1$, $\hat{p} \leq .012$; cage control vs. Orx₁R Ant., $t_{11} = 3.3$, $\hat{p} \leq .007$) was eliminated with intra-BLA Orx₂R antagonism (vehicle vs. Orx₂R Ant.: $t_7 = 2.8$, $\hat{p} \leq .017$). **(D)** While Stay mice treated with an Orx₂R Ant. express higher levels of *Mapk3* (phenotype effect: $F_{1,27} = 11.3$, $p \leq .002$; treatment effect: $F_{2,27} = 4.3$, $p \leq .023$; interaction effect: $F_{2,27} = 5.1$, $p \leq .013$) in the BLA compared with vehicle control animals ($t_{12} = 3.1$, $\hat{p} < .001$), administration of an Orx₂R Ant. does not induce the same transcriptional response (Orx₁R Ant. vs. Orx₂R Ant.: $t_{10} = 2.7$, $\hat{p} \leq .022$). **(E)** Expression of *Bdnf* in the BLA after treatment (interaction effect: $F_{2,27} = 10.6$, $p < .001$) with an Orx₂R Ant. was enhanced in Escape mice (Orx₂R Ant. Escape vs. Stay, $t_8 = 2.2$, $\hat{p} \leq .019$; vehicle vs. Orx₂R Ant.: $t_7 = 2.7$, $\hat{p} \leq .013$; Orx₁R Ant. vs. Orx₂R Ant.: $t_8 = 2.5$, $\hat{p} \leq .017$) and reduced in Stay animals (vehicle vs. Orx₂R Ant.: $t_{10} = 2.2$, $\hat{p} \leq .05$; Orx₁R Ant. vs. Orx₂R Ant.: $t_{10} = 3.9$, $\hat{p} < .001$); a phenotypically opposite effect was observed after Orx₁R antagonism (Escape vs. Stay, $t_{10} = 2.8$, $\hat{p} \leq .018$; Orx₁R Ant. Stay vs. vehicle Stay, $t_{12} = 2.2$, $\hat{p} \leq .049$). Transcriptional changes after **(F)** intra-BLA Orx₁R antagonism and **(G)** Orx₂R inhibition were differentially regulated in a phenotype-dependent fashion. Ant., antagonist; BLA, basolateral amygdala; GABA, gamma-aminobutyric acid; Glu, glutamatergic neurons; mRNA, messenger RNA; Orx₁R, orexin 1 receptor; Orx₂R, orexin 2 receptor; Orx_A, orexin A; Orx_B, orexin B.

day 3 drug treatment), is a reflection of the shift toward anti-stress signaling indicated by downregulation in prostress signaling (*Hcrtr1*) and upregulation of antistress systems (*Hcrtr2*, *Mapk3*, *Bdnf*). Thus, BLA dual Orx₁R/Orx₂R inhibition may not promote behavioral change. These stress-induced effects are paired with important learning and motivational components during SAM interactions (27,29,33,35) and in human affective disorders (46).

In addition to species-specific anxious behavior and learning, social stress promotes behavioral inhibition, depressed motivation, and depressed behavioral drive in some individuals (47), plus a lower rate of adaptive behavior (48). Behavioral depression reveals two distinctive phenotypes related to stress responsiveness in humans and other animals (45,49,50). In SAM social interactions, Stay animals do less exploration of the escape route (Figure 3A) and show indecisiveness relative to escape (35). Measuring motivation in the SAM is derived from a simple choice process, Escape or Stay (26,27). Antagonism and knockdown of Orx₁Rs increases interest in the escape route for both Stay and Escape mice (Figure 3C, D). Thus, BLA Orx₁Rs regulate stress-induced motivational behaviors, greatest in Escape mice but marking

a dramatic behavioral reversal in Stay mice that typically avoid the escape route (Figure 3B, C). Attention to the escape route happens prior to escape and is thus the first evidence of phenotypic differentiation in the SAM (28,35). Latency to escape and escape behavior also are influenced by motivation, although as previously demonstrated, these behaviors are strongly affected by stress and fearfulness associated with familiarity of the SAM or social interaction (27–29,33,35). Our results, similar to those of others, suggest that Orx activity plays a fundamental role in motivation (8,51) and, in this case, specifically in the BLA for behaviors associated with stress-related motivation and choice.

Understanding the development of choice and motivation in the SAM is enhanced by pairing aversive aggression (US) with a nonthreatening stimulus (tone CS) prior to interaction, promoting potent cued and contextual CR similar to standard fear conditioning approaches that use foot shock as a US (52). While the CRs elicited are similar, e.g., freezing (53), the ethological and ecological relevance of the US to the subject are not. By associating naturally aversive US with a benign stimulus (54), the SAM allows views into development of fear learning as it relates to the etiology of stress-provoked

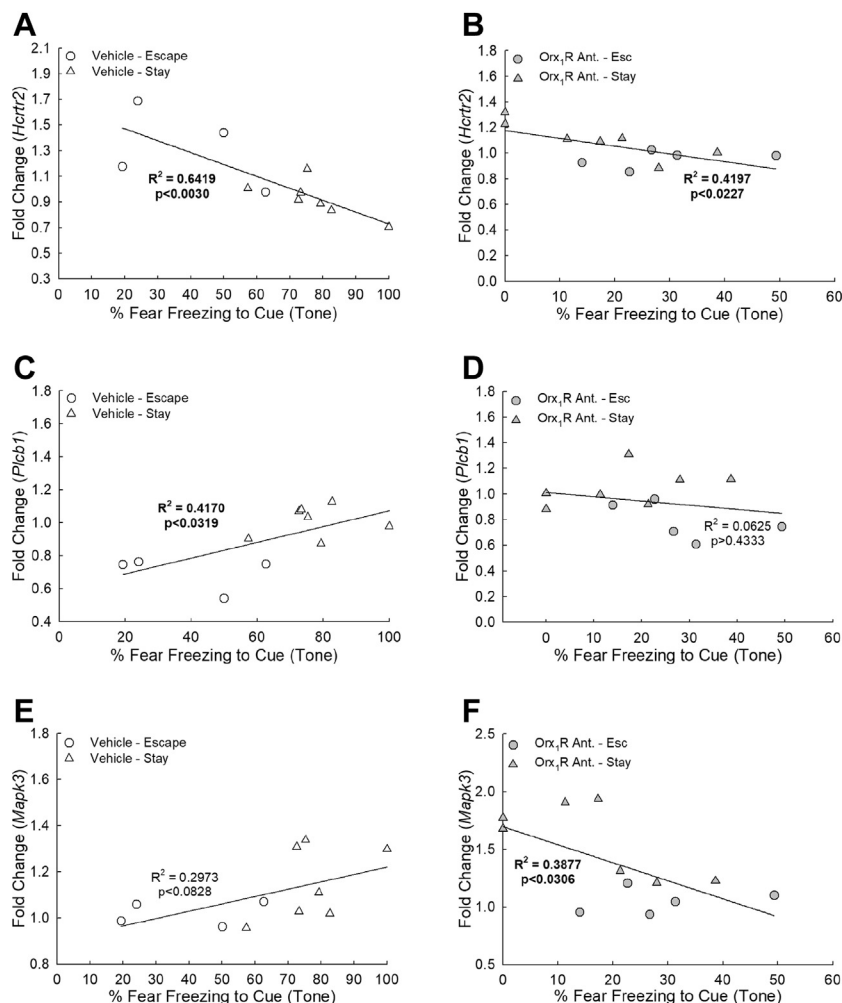


Figure 7. Conditioned fear freezing response is related to gene expression changes (fold change relative to home-cage naïve control mice) resulting from intra-basolateral amygdala Orx₁R antagonism. In both **(A)** vehicle-treated ($n = 11$, $F_{1,9} = 16.1$, $R^2 = 0.6419$, $p \leq .003$) and **(B)** Orx₁R Ant.-treated ($n = 12$, $F_{1,10} = 7.2$, $R^2 = 0.4197$, $p \leq .023$) animals, a negative correlation exists between *Hcrt2* expression and cued fear freezing. **(C)** With vehicle treatment, relative *Plcb1* expression is positively associated with cued fear freezing ($F_{1,9} = 6.4$, $R^2 = 0.417$, $p \leq .0319$). **(D)** This relationship is not observed in mice that were administered an Orx₁R Ant. ($F_{1,10} = 0.7$, $R^2 = 0.0625$, $p \geq .4333$). **(E)** While there is not a significant association between *Mapk3* expression and cued fear freezing after vehicle treatment ($F_{1,9} = 3.8$, $R^2 = 0.2973$, $p \geq .0828$), **(F)** a significant negative correlation is observed after Orx₁R antagonism ($F_{1,10} = 6.3$, $R^2 = 0.3877$, $p \leq .0306$). Ant., antagonist; Esc, escape; Orx₁R, orexin 1 receptor.

neurocircuitry changes and demonstrates a connection between stress-induced fear expression and phenotype (Figure 5). While early work suggested that only Stay mice exhibited cued fear learning (27,33), it is now clear that both Stay and Escape mice respond to auditory cues with enhanced freezing compared with pretone freezing, and Stay mice also show contextual (prior to the cue) fear conditioning (Figure 5A).

Fear responses are mediated through Orx₁R activity in the amygdala and in the locus coeruleus, which connects to the amygdala (22,55–57). Our results similarly demonstrate that Orx₁R, but not Orx₂R, inhibition diminishes both contextual and cued conditioned fear freezing in Stay animals (Figure 5B; Table S1). While antagonizing Orx₁R reduces fear- and panic-induced freezing (7,56,58), Orx₂R antagonism appears to eliminate fear learning in Escape mice, suggesting a phenotype-dependent effect (Figure 5B; Table S1). Although Orx₂R antagonism in the BLA reduced cued freezing only in Escape mice, we have previously demonstrated a potential anxiogenic effect of blocking receptor function (25,28). This response may be dependent on brain region because Orx₂R activity in the nucleus accumbens shell and prelimbic

prefrontal cortex may enhance anxious behavior (59,60). Furthermore, Orx₂R antagonism has demonstrated antidepressive capabilities in a clinical setting (61).

Stimulation of intra-BLA Orx₁Rs and Orx₂Rs using Orx_A in Stay mice produces no reduction in contextual or cued fear conditioning (Figure 5B; Table S1), suggesting that the inhibition of both types of learned fear responses result specifically from Orx₁R inhibition in Stay mice. To clarify the roles of Orx₁R and Orx₂R, we administered Orx_A while concurrently inhibiting Orx₂R (MK1064), leaving Orx₁R stimulated, and again there was no statistically significant reduction in either type of fear conditioning response (Figure 5B; Table S1). Knockdown of Orx₁R did not affect the fear freezing profile (Figure S8). Because knockdown occurred before the introduction of social stress, activity levels of Orx₁R after SAM exposure allowed for fear learning (higher freezing after CS), but did not diminish freezing as observed with acute antagonism after stress and phenotype development (Figure 5B).

Molecular gene expression during SAM fear conditioning and phenotype development indicated potential shifts in receptor-linked intracellular signaling cascades (Figure 6).

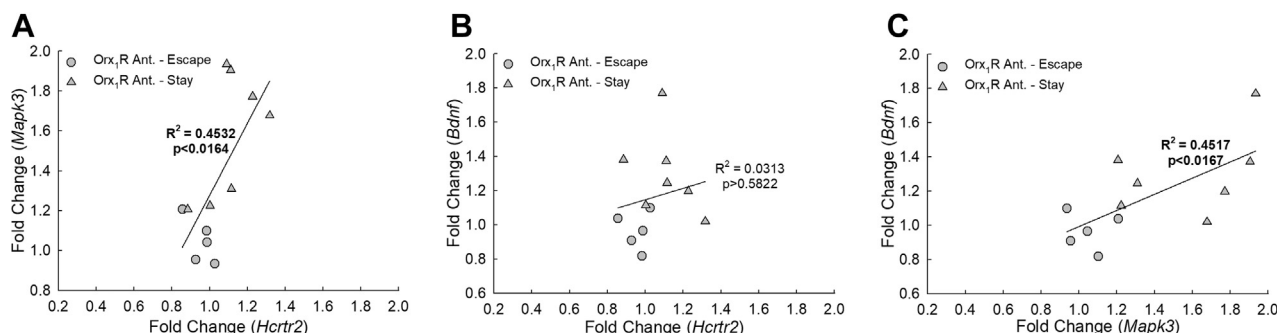


Figure 8. The basolateral amygdala transcriptional changes (relative to home-cage naïve control animals) that result from Orx₁R antagonism form relationships that hint at molecular timelines and signaling dynamics. **(A)** While relative gene expression of *Mapk3* is positively correlated to the transcriptional changes of *Hcrt2* ($n = 12$, $F_{1,10} = 8.3$, $R^2 = 0.4532$, $p \leq .0164$), **(B)** there is no association between *Bdnf* and *Hcrt2* ($F_{1,10} = 0.3$, $R^2 = 0.0313$, $p \geq .5822$). However, **(C)** a positive relationship emerges when comparing *Bdnf* expression to that of *Mapk3* ($F_{1,10} = 8.2$, $R^2 = 0.4517$, $p \leq .0167$). Ant., antagonist; Orx₁R, orexin 1 receptor.

Acute inhibition of intra-BLA Orx₁R, by means of a feedforward rather than feedback mechanism, lowered *Hcrt1* expression in Escape mice while enhancing *Hcrt2* mRNA in Stay animals (Figure 6A, B). Antagonism of Orx₂R in the BLA did the opposite, reducing *Hcrt1* mRNA only in Stay mice, and in a similar feedforward way, decreasing *Hcrt2* expression in both phenotypes (Figure 6A, B). Mice exhibiting escape and reduced fear freezing, expressed lower *Plcb1* compared with the Stay phenotype, an effect unaltered by SB-674042 treatment but reversed by Orx₂R antagonism (Figure 6C). However, intra-BLA Orx₁R antagonism increased *Mapk3* and *Bdnf* expression in Stay animals only, with Orx₂R inhibition having no effect on expression of *Mapk3* and enhancing *Bdnf*, but only in Escape mice, while reducing *Bdnf* in Stay mice (Figure 6D–G; Table S2). These results suggest that social stress disrupts gene expression and potentially alters BLA signaling pathways depending on an individual's stress state. Therefore, pharmacological interventions (such as acute Orx₁R antagonism) may functionally amend behavior through signaling adaptations that are phenotype dependent.

Fear conditioning responses appear to be related to specific transcriptional reorganization taking place during/after intra-BLA Orx₁R inhibition (Figure 7). In treated animals,

negative regressions exist between cued fear freezing behavior and *Hcrt2* as well as *Mapk3* (62) transcriptional changes (Figure 7B, F). Without treatment (vehicle), cued freezing was positively linked to *Plcb1* gene expression (Figure 7C), an effect not observed with Orx₁R antagonism (Figure 7D). These associations provide evidence for potential mechanistic remodeling (Figure 9) in the BLA during periods of stress that is tied to phenotype formation and involves Orx receptor activity. This balancing act between Orx₁R and Orx₂R creates an influence over BLA microcircuits, which further defines downstream signaling dynamics, in a way that can modify stress-induced behavior (2). Because changes in *Hcrt2* expression after intra-BLA Orx₁R inhibition are positively associated with *Mapk3* but not *Bdnf* transcription levels (Figure 8A, B), it appears that the adjusted bias of Orx₂R over Orx₁R activity favors ERK₁ signaling (Figure 9). Amplification of ERK₁, in turn, may lead to enhanced *Bdnf* expression (Figure 8C) and plastic changes within BLA microcircuits (Figure 9) (62,63). These findings highlight a role of intra-BLA Orx₁Rs in establishing prostress behavioral states but expose a receptor-driven balance that takes part in the fluid, not static, appearance of phenotype-specific behavior.

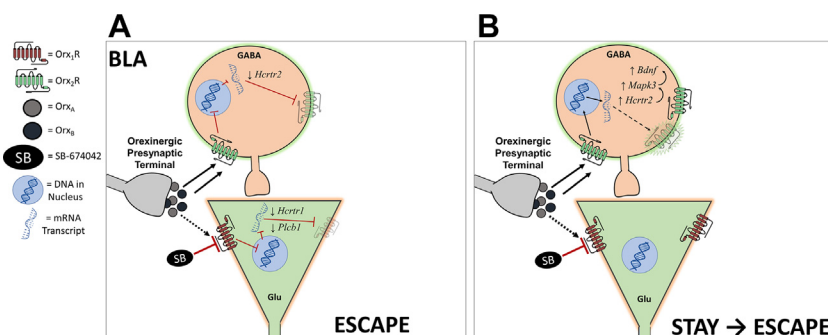


Figure 9. Predicted circuit demonstrates the influence of intra-BLA Orx₁R antagonism, during endogenous stimulation through Orx_A and Orx_B release, on microcircuit dynamics in a phenotype-dependent fashion. **(A)** Escape mice treated with an Orx₁R Ant. (SB-674042) undergo molecular shifts, including a feedforward reduction of *Hcrt1* and reduced *Plcb1* transcription, leading to diminished orexin activity on glutamatergic neurons in the BLA. Escape mice also have a feedforward decrease in *Hcrt2* expression, potentially via (undiagrammed) negative circuit feedback, even while Orx₂Rs are stimulated. **(B)** While Orx_B and Orx_A maintain stimulation of some GABAergic neurons through Orx₂R,

antagonism of some pyramidal neurons via intra-BLA Orx₁R inhibition differentially modifies molecular mechanisms in Stay mice through enhancement of Orx₂R (*Hcrt2*), extracellular signal-regulated kinase 1 (*Mapk3*), and *Bdnf* transcription and increased orexin activity in Orx₂R-containing neurons (likely GABAergic cells). BLA, basolateral amygdala; GABA, gamma-aminobutyric acid; Glu, glutamatergic neurons; mRNA, messenger RNA; Orx₁R, orexin 1 receptor; Orx₂R, orexin 2 receptor; Orx_A, orexin A; Orx_B, orexin B.

Conclusions

Modulation of BLA stress-regulatory pathways via Orx₁Rs found predominantly on glutamatergic pyramidal neurons modifies gene expression and behavior. Modulation of prostress BLA microcircuits via Orx₁R inhibition reduces stress-induced behavior. In the process, Orx₁R BLA inhibition modifies gene expression of *Hcrtr2*, which impedes prostress responses. Concurrently, transcription levels for downstream molecular signaling systems associated with Orx receptor signaling are also tilted toward increased ERK₁ (*Mapk3*) rather than PLC β ₁ (*Plcb1*) signaling pathways, potentially altering behavior.

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