

Surfactant proteins and innate immunity of otitis media

Innate /mmt.rility
2022 , Vol. 28 (7.S): 213- 223
© The Author(s) 2022
Art ide r euseguidelines:
sagepub.com/jour nal.s-permission s
DO I: 10 .1177/17534259221123309
jouma ls.sagepu b.co m/ hom e/in
(\$)SAGE

Osama Abdell-Razek ¹(E), Jason A udlin², Dennis S. Poe⁴
and Guirong Wang^{1,3}(E)

Abstract

Otitis media (OM) is the most common disease among young children and one of the most frequent reasons to visit the pediatrician. Development of OM requires nasopharyngeal colonization by a pathogen which must gain access to the tympanic cavity through the eustachian tube (ET) along with being able to overcome the defense mechanisms of the immune system and middle ear mucosa. OM can be caused by viral or bacterial infection. The three main bacterial pathogens are *Streptococcus pneumoniae*, *nontypeable Haemophilus influenzae* (NTHi), and *Moraxella catarrhalis*. Innate immunity is important in OM resolution as the disease occurs in very young children before the development of specific immunity. Elements of innate immunity include natural barriers and pattern recognition receptors such as Toll like receptors (TLRs), and Nod like receptors (NLRs). Surfactant proteins A (SP-A) and D (SP-D) act as pattern recognition receptors and are found in the lung and many other tissues including the ET and the middle ear where they probably function in host defense. Surfactant has a potential for use in the treatment of OM due to surface tension lowering function in the ET, and the possible immune functions of SP-D and SP-A in the middle ear and ET.

Keywords

Innate immunity, otitis media, otoscopy, surfactant proteins

Date received: 27 January 2022; revised: 15 July 2022; accepted: 15 August 2022

Introduction

Otitis media (OM) is the most common disease in young children and one of the most frequent reasons to visit the pediatrician.^{1,2} It is estimated that about 60%-80% of infants will have at least one episode of OM.³ Acute otitis media (AOM) is inflammation of the mucosa of the middle ear cleft which includes the mastoid process, tympanic cavity and, the Eustachian tube (ET).⁴ There is a significant global health burden of AOM; Thirty one million children of an estimated 709 million cases per year progress to develop chronic suppurative OM (OM with chronic ear discharge), and complications such as meningitis or brain abscess which result in about 21,000 deaths each year.⁵ The World Health Organization estimates that 51,000 deaths/year are attributable to AOM in children younger than 5 years old and that chronic otitis media is the major cause of hearing loss in developing countries.⁶ Studies show that before their third birthday, 80% of children will have suffered a minimum of one attack of OM and 40% will have at least six recurrent attacks by the age of 7 years. OM is also the primary indication for prescribing antibiotics among children.⁷ In the first two years of life, children spend a mean of 90 days on antibiotics for OM.⁸

Additionally, OM is the primary indication for ventilation tube insertion, which is the most commonly performed operation in children.⁷

Pathogenesis

There are three requirements for developing OM: 1) nasopharyngeal colonization by the pathogen;⁹ 2) ascending infection through the ET to the tympanic cavity; 3) pathogens must be able to overcome the defense mechanisms of the immune system and middle ear mucosa.^{10,11} A multitude of host and environmental factors significantly

¹Department of Surgery, SUNY Upstate Medical University, Syracuse, NY, USA

²Department of Otolaryngology, SUNY Upstate Medical University, Syracuse, NY, USA

³Department of Microbiology and Immunology, SUNY Upstate Medical University, Syracuse, NY, USA

⁴Harvard Medical School, Boston, MA USA

Corresponding author:

Guirong Wang, Department of Surgery, UH Room 8715, SUNY Upstate Medical University, 750 E Adams St, Syracuse, NY, 13210, USA.

Email: wangg@upstate.edu

influence the risk of developing OM. Host factors that can increase the risk of developing OM include, sex, young age, genetic susceptibility, adenoid hypertrophy, laryngopharyngeal reflux, race and ethnicity, and craniofacial malformations such as cleft palate, atopy, immunodeficiency, and viral upper respiratory tract infections (URTIs).¹² Family history of OM may be an especially important factor.¹³ Environmental factors that increase the risk of OM include: low socioeconomic status, exposure to tobacco smoke, having older siblings, day-care attendance and the use of a pacifier.¹⁴

Viral and bacterial pathogens induced OM

Acute otitis media can result from viral or bacterial infections. Viral infection causes around 20% of AOM. Respiratory syncytial virus (RSV), influenza viruses, adenoviruses, rhinoviruses, and enteroviruses, are the viruses that most commonly cause AOM.^{15, 16} A recent study by Heikkinen *et al.*, showed that among children younger than three years, the average RSV infection incidence rate was 275 per 1000 children per year and 58% of these children with RSV developed AOM.¹⁷ Bacterial infection is the predominant cause of acute and recurrent otitis media. It leads to hyperplasia of the tympanic epithelium, middle ear effusion and leukocytic infiltration of the tympanic cavity.^{18, 19} There are three main bacterial pathogens, *Streptococcus pneumoniae*, *nontypeable Haemophilus influenzae* (NTHi), and *Moraxella catarrhalis*.^{10, 20} Following the introduction of vaccines against *S. pneumoniae*, NTHi has become the most common pathogen in OM.^{19, 21} The risk for developing AOM after upper respiratory viral infection depends on the otopathogenic bacteria colonizing the nasopharynx; the risk is highest if the nasopharynx is colonized by all three pathogenic bacteria and lowest with no colonized bacteria.²² Viral infection negatively impacts the nasopharyngeal mucosa and the ET functions. Viral infection changes the nasopharyngeal mucosa by modifying host immune function, inducing cytokine activity and inflammatory mediators and upregulates host cell surface antigens that act as bacterial receptor sites and increases bacterial colonization and adherence. Viral infection impairs the ET through altering the properties of mucus and diminishing the normal mucociliary clearance by mucosal cells of the ER. Transient impairment of ET functions allows bacteria colonizing the nasopharynx to ascend into the middle ear and cause AOM.^{1, 23} Using the chinchilla otitis media model, studies show that inoculation with *S. pneumoniae* alone through the nasopharynx resulted in the development of OM in 20% of the animals, whereas adding adenovirus to the bacteria led to the development of OM in nearly 80% of them.²⁴ Initially, *M. Catarrhalis* adheres to mucosal surfaces through a dozen or more adhesins that it expresses. However, in AOM, adherence to the mucosal surface

alone is not sufficient to cause disease. A cofactor, such as a viral infection, is thought to be needed to precipitate migration to the ME through the Eustachian tube. *M. catarrhalis* is often isolated with *S. pneumoniae* and *H. influenzae* in respiratory tract cultures and may facilitate polymicrobial infection by sheltering these organisms from complement-mediated immune destruction, promoting biofilm formation, and releasing beta-lactamase into the local environment.^{25, 26} *M. catarrhalis* has shown the ability to overcome host innate immune response and to evade complement-mediated lysis.

Structure and function of the middle ear and ET

The middle ear cleft is comprised of the tympanic cavity, the mastoid air cell system and Eustachian tube (ET). The middle ear cleft is an irregular shaped gas filled chamber that uses the ET to equalize middle ear pressure with the ambient pressure through the nasopharynx² (Figure 1). The middle ear mucosa is derived from two origins, neural crest which give origin to the dorsal region of the middle ear mucosa which is a non-ciliated epithelium and endoderm of the first pharyngeal pouch which give rise to the ventral part of the mucosa forming the ciliated epithelium near the ET orifice.²⁸ Embryologically, the ET develops from the first pharyngeal pouch and connects the nasopharynx to the tympanic cavity. The nose, palate, nasopharynx, ET, middle ear and mastoid air cells form a system of contiguous organs. The ET is not a tube in reality, but an organ consisting of a lumen with its mucosa, cartilage, surrounding soft tissue, and peritubal muscles.²⁹ The ET functions to aerate and protect the middle ear from excessive changes in the atmospheric pressure, also as a mucus drainage pathway, and to protect the middle ear from otopathogens and other inhaled noxious agents.³⁰ The ET is lined by respiratory epithelium, which is pseudostratified ciliated columnar epithelium with interspersed goblet cells, that produce both mucoid and serous mucus. The cilia propel mucus in combination with epithelial secretion of antimicrobial protein, through the ET, from the middle ear to the nasopharynx which helps to protect against bacterial colonization of the middle ear.¹⁴ The ET epithelium is the front-line defense against the passage and colonization of otopathogens from the nasopharynx.^{14, 31} ET dysfunction is the primary cause of middle ear infection and effusion.³² Poor or impaired function of the mucociliary system of the ET can result in bacterial entry into the tympanic cavity. OM is more common in infants which can be explained anatomically. The ET is shorter, wider and more horizontal in infants and young children (< 1 year of age) than in adults, which facilitates otopathogen transmission to the middle ear increasing the opportunity for antigen exposure or pathogen colonization while reducing the opportunity for mucosal immune responses thus increasing the risk of OM, compared to the adult morphology^{14, 33}

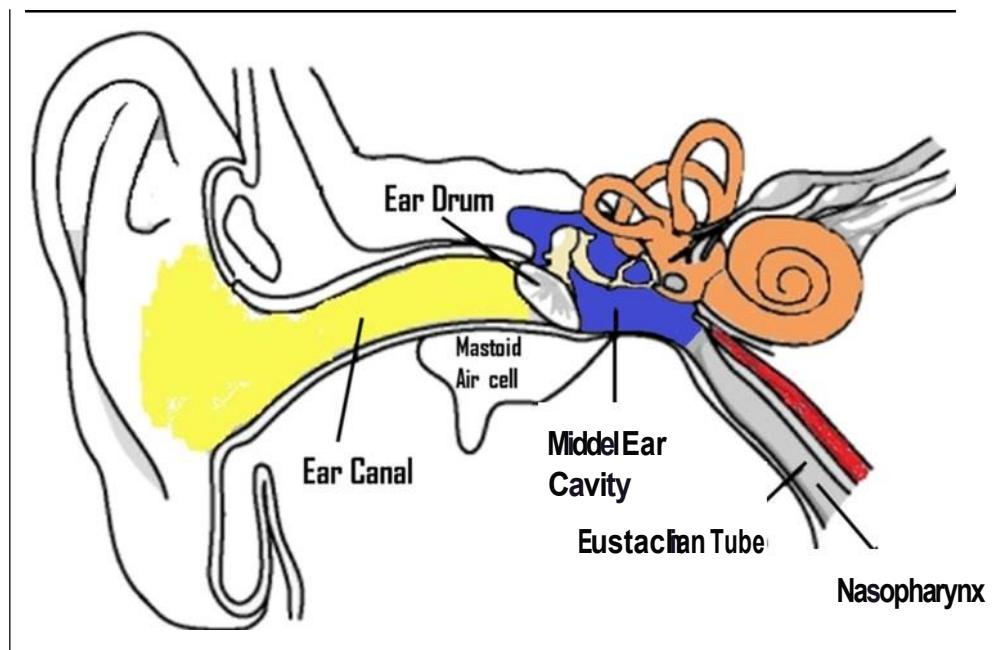


Figure I. Schematic representation of ear structure showing the external ear (Yellow), the middle ear (blue) and the inner ear (orange). The middle ear space connects to eustachian tube and the mastoid air cell system.

Innate immune response in OM

The innate immune response is a nonspecific, first line defense against pathogens that does not require prior sensitization (see Table 1). There are diverse players that contribute to the innate immunity (Figure 2) in the middle ear and ET. These innate immune elements can be classified as natural barriers, pathogen recognition receptors, antimicrobial proteins, and peptides, mucociliary system and various cellular components. Natural barriers function through mucous membranes and mucociliary system that employs a bilayer stream of mucus blanket with a thick mucus layer secreted by goblet cells and submucosal glands traveling on top of a thin serous layer secreted by the epithelial cells. Together these elements constitute the periciliary fluid. Both layers cover the middle ear and ET mucosa and are important for the proper functioning of the mucociliary system.³⁴

The innate immune system recognizes the presence of microbial infection by using pattern recognition receptors (PRRs) to identify pathogen-associated molecular patterns (PAMPs), which are the molecular signature of pathogens.³⁵ Toll-Like receptors (TLRs) play a key role in innate immunity. They are a class of PRRs recognizing molecules associated with microbial pathogens.^{36, 37} Recently, TLRs have emerged as key regulators of innate immune responses to infection in mammals. TLRs are found in the cytoplasm or on the cell membranes of the epithelial cells lining the middle ear, and also on mast cells and dendritic cells, which are abundant at middle ear mucosal surfaces. Contact with PAMPs leads to activation of the TLRs,

which activate a signaling cascades that regulate innate immune response to mobilize cytokines, chemokines and interferons which attract neutrophils and macrophages to clear the site of infection from the invading bacteria and can then stimulate both local and systemic adaptive immune responses (Figure 2).^{37, 38} There have been thirteen different members of the mammalian TLR family identified (10 in humans and 12 in mice).³⁹ Among the known 10 members of human TLRs, TLR2 is activated by pneumococcal cell wall components, such as lipoteichoic acid and lipoproteins and likely plays an important role in the pathogenesis of pneumococcal OM.^{38, 40} TLRs activate an intracellular signaling cascade through employing adapter proteins. Except for TLR3 all TLRs employ myeloid differentiation factor-88 (MyD88) as an adapter protein to induce nuclear factor-kappa B (NF-KB) or mitogen-activated protein (MAP) kinase-dependent proinflammatory gene expression to the production of proinflammatory cytokines such as tumor necrosis factor (TNF- α) and the interleukins (ILs).^{39, 40} Recovery from OM is impaired by the absence of key elements of TLR signaling, particularly TLR2, MyD88, and TNF- α .³⁷ In children with recurrent otitis media, the levels of IL-1, IL-6, and TNF- α in nasopharyngeal secretions were found to be lower than in healthy children.⁴¹ TLRs work by recognizing pathogen associated molecular patterns. The interactions between TLRs and these molecular patterns can activate intracellular signaling pathways, such as the NF- κ B pathway. The activation of NF- κ B pathway in turn upregulates the expression of proinflammatory genes involved in the production of cytokines

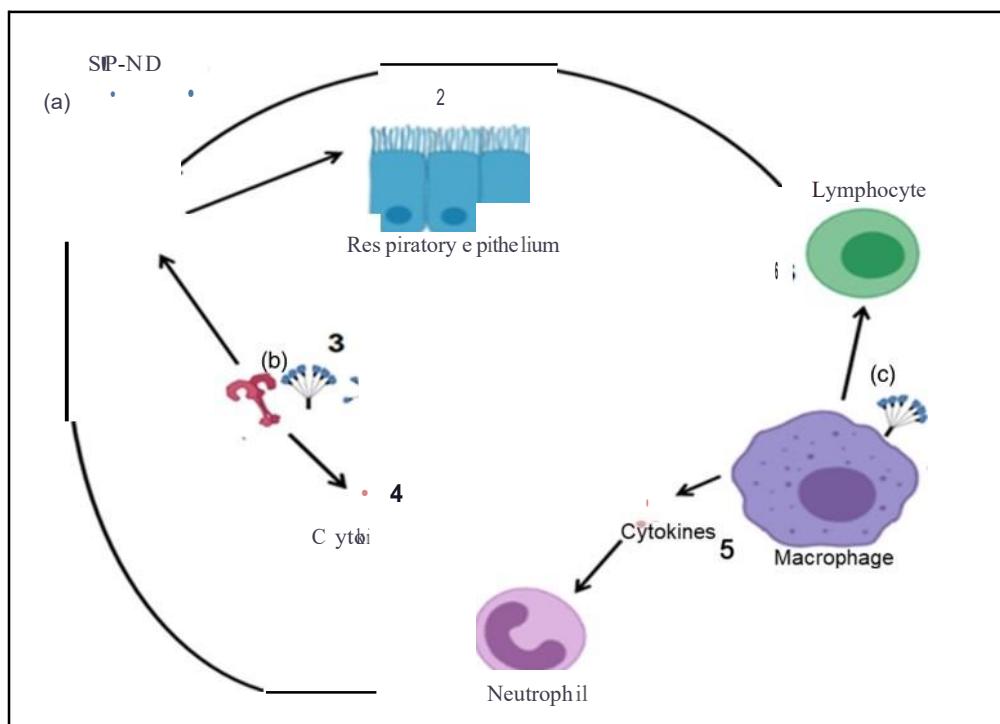


Figure 2. Schematic representation of the major players in the innate immune system. (1) pathogens attack the host; (2) Physical and chemical barriers as epithelium with the mucociliary system; (3) Pathogen recognition receptors as TLRs; (4) activation of cytokines which attract innate immune cells; (5) to attack the pathogens. Macrophage help to (6) activate adaptive immunity. SP-A and SP-D can attack pathogens directly (a) act as PRR (b) or enhance phagocytosis (c).

and chemokines and activation of the adaptive immune system. TLR2, a member of TLR family, reported to regulate the pathogenesis of NTHi induced AOM as it recognizes pathogenic patterns of NTHi and regulates the inflammatory responses of the host.⁴² Lipoprotein P6 of the NTHi uses TLR2 signaling to activate NF- κ B in human epithelial cells. Similarly, TLR4 mediates lipopolysaccharide (LPS) and lipooligosaccharide (LOS) responsiveness and recognizes gram-negative bacteria via the LPS/LOS moiety present on the surfaces of these pathogens. Although the structure of LOS and LPS is different, lipid A, which is a common component to both, is thought to be responsible for TLR4 signaling.⁴³ LPS-TLR4 signaling and neutrophil reduced nicotinamide adenine dinucleotide phosphate oxidase activate NF- κ B signaling and upregulate TLR2 expression in endothelial cells, and this increased TLR2 expression via NF- κ B signaling results in increased intercellular adhesion molecule 1 expression and enhanced neutrophil migration. TLR4 signaling appears to induce TLR2 expression, and TLR2 activation is critical for bacterial clearance and rapid resolution of OM.⁴⁴ A recent study demonstrated that the TLR4 gene locus, regulating the innate immune response, influences the genetic predisposition to childhood OM in a subpopulation of patients.⁴⁵ TLR4 Polymorphisms have been found to associate with the risk of recurrent OM, but the data are

partly conflicting.⁴⁶ Additionally, TLR9 is localized to endosomes and when exposed to bacterial DNA it recognizes a common motif present in bacterial but not mammalian DNA, unmethylated cytidine-phosphate-guanosine (CpG). This results in signaling molecules recruitment, that eventually result in the production of pro-inflammatory cytokines and other target genes expression. TLR9 facilitates the development of the inflammasome, which might contribute in the development of effective adaptive immune response.³¹ Leichtle et. al. recently showed that deletion of TLR9 significantly prolonged the inflammatory response induced by NTHi in the ME and delayed bacterial clearance.³¹

Other PRRs including cytoplasmic (NOD)-like receptors (NLRs), and C-type lectin receptors (CLRs) are found to be involved in the innate immunity of OM.⁷ The NOD-like receptors (NLRs) are cytoplasmic proteins that regulate inflammatory and apoptotic responses. The NOD proteins NOD1 (which is encoded by the caspase-recruitment domain 4 gene, *CARD4*) recognizes a molecule called meso-diaminopimelic acid, a peptidoglycan constituent only of Gram negative bacteria. NOD2 (which is encoded by *CARD15*) recognizes intracellular muramyl dipeptide (**MDP**), a peptidoglycan constituent of both Gram positive and Gram negative bacteria.^{47,48}

Defensins are cationic proteins released by the middle ear epithelium and function as antimicrobial agent with a wide range of activity against viruses, bacteria, fungi and protozoa. Defensins can also upregulate cytokines to attract neutrophils, mast cells, and T-lymphocytes.⁴⁹ Lysozyme is an antimicrobial molecule secreted by epithelial cells in the airway and act by degrading the peptidoglycan in the bacterial cell wall.⁵⁰ Special inflammatory and immune-response-relevant cells such as mucosal dendritic cells (DCs) and mast cells have been found to play a role in the pathogenesis of OM.

Surfactant proteins and innate immunity in OM

Surfactant proteins (SPs) represent a small percentage of surfactant composition but they are essential for surfactant homeostasis.⁵¹ Four surfactant proteins, named as SP-A, SP-B, SP-C, and SP-D, have been discovered and characterized in lung biology. SP-B and SP-C are hydrophobic proteins, and play a key role in lowering surface tension,⁵² as well as surfactant metabolism and recycling in the lung.⁵¹ SP-A and SP-D belongs to the family of C-type lectins, or collectins, they are hydrophilic proteins and mainly contribute to the innate immunity.⁵³⁻⁵⁵ At initial studies, all surfactant proteins are synthesized and secreted by alveolar type II cells and SP-A and SP-D were also found to be secreted by sub-mucosal and Clara cells in the lung. SPs are present at the luminal surface of pulmonary epithelial cells, and are secreted into alveolar spaces.⁵⁶⁻⁵⁸ Several recent studies have demonstrated that SP-A and SP-D are expressed in multiple tissues other than the lung, including the tongue and oral epithelium; the digestive, urinary and reproductive tracts; synovial and pericardial fluid; the spleen, thymus, pancreas, kidney, the middle ear and Eustachian tube.⁵⁹ Although SP-B and SP-C were thought to be expressed solely in the pulmonary epithelium,⁶⁰⁻⁶² and various phospholipids were found also expressed in the eustachian tube. Therefore, all main components of pulmonary surfactant are produced in this tissue by fully competent cells having a complete surfactant machinery.⁶³

The innate immune response is very important for susceptibility to OM in young age before acquiring specific immunity.³ Collectins protect the host through the identification of PAMPs on pathogens and allergens and respond by activation of multiple processes of innate immunity, including phagocytosis, cytokine secretion, and complement activation.⁵³ SP-A and SP-D play an important role in host defense through opsonization and complement activation (Figure 3).⁶⁴ In a recent study by our group we found that SP-A mediated NTHi aggregation and killing along with enhanced bacterial phagocytosis by macrophages in vitro and modulated inflammation of the ME in otitis media in vivo.⁵⁹ In another study (results not published

yet) we found that mice lacking SP-D gene had prolonged inflammation and slow resolution of the ME inflammation in response to NTHi induced otitis media.

Collectins are structurally and functionally related to the first component of the classical complement pathway Clq, with the exception that collectins are exclusively present in the extracellular matrix.⁶⁵⁻⁶⁷ Collectins are formed of collagen like regions associated with non-collagen domains. The carbohydrate recognition domain (CRD) is the part that recognizes carbohydrate epitopes from multiple microorganisms. CRD is attached to the neck and collagen domain which is attached to N-terminal domain.⁶⁸ CRD of collectins can differentiate between self and non-self carbohydrates. For example, all collectins have affinity to mannose, SP-A has affinity to L-fructose and N-acetylmannosamine, and SP-D to glucose, mannose and inositol. On the other hand, they poorly recognize galactose and sialic acid which make clusters of oligosaccharides in most vertebral animals. Direct agglutination or neutralization of microorganisms can occur when they bind to collectins. This agglutination provides a first line of defense, which can be augmented by killing and clearance mechanisms mediated by phagocytic cells that carry receptors for SP-A and SP-D.¹ Collectins also help in opsonization and the presentation of bound microbes directly to phagocytes, or activate complement cascade via the lectin pathway. The lectin domains recognize and bind to carbohydrate structures found on a variety of microorganisms such as viruses, bacteria, yeast and fungi. The collagenous regions are ligands for the collectin receptor on phagocytes and also mediate Clq-independent activation of the classical complement pathway.⁶⁹⁻⁷⁰ SP-A enhances production of secretory leukoprotease inhibitor (SLPI). SLPI is a 12 kDa protein that is a constitutively expressed, up-regulatable inhibitor of serine proteases. It is found in airway surface epithelial cells, Clara cells, and associated with elastin fibers of the lung interstitium and monocytes, alveolar macrophages and neutrophils. SLPI targets neutrophil elastase, cathepsin G, chymotrypsin and trypsin. It has antimicrobial properties against bacteria and it regulates the production of TNF- α by inhibiting the LPS-induced NF- κ B activation and inhibiting I κ B- α degradation.⁷¹

Because SP-A works as a pattern recognition receptor, it functions as one of the first lines of defense before the development of specific antimicrobial antibodies. SP-A binds to and increases the phagocytosis of *S. pneumoniae* and *H. influenzae*, the most common otopathogens.^{3, 61} SP-A opsonizes gram negative bacteria and modifies LPS for macrophage binding. It also modulates proinflammatory cytokines IL-1 β , IL-6, and TNF- α which play an important role in fighting infections. SP-A and SP-D are expressed in the normal human and porcine eustachian tube.⁵⁴ SP-A is the most abundant surfactant associated protein in the body and also in the ET, where it is synthesized most profusely at the mucosal

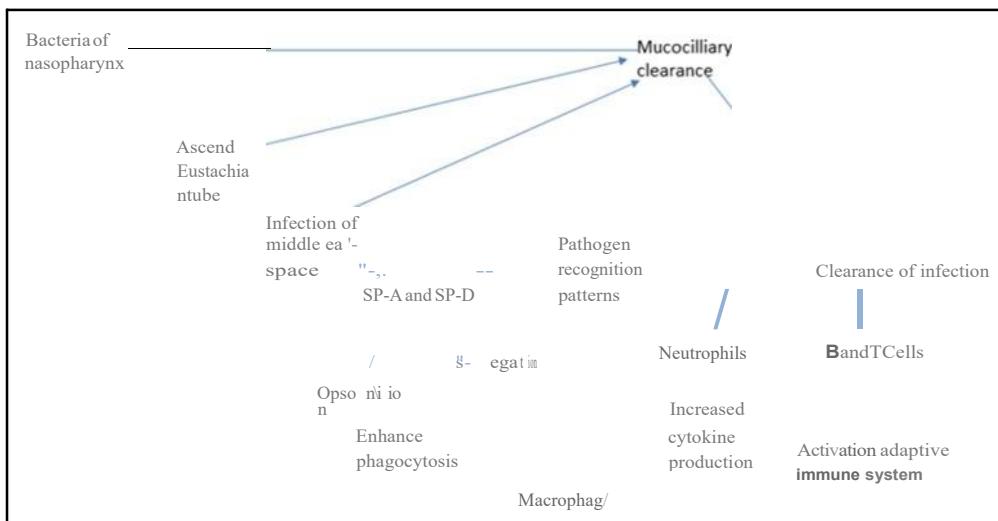


Figure 3. Schematic representation of the pathogenesis of **AOM** and the roles of **SP-A** and **SP-D** in the innate immune response of **OM**.

Table I. Innate immune components and their acting mechanisms.

Components	Innate immune system elements	Action
Natural Barriers:	Mucous membranes Mucociliary system	Forms an effective barrier to invading microorganisms Trap microorganisms in the mucous layer and transported by the ciliary action towards the nose/pharynx
Cells:	Neutrophils Macrophages Dendritic Cells	Direct attack of pathogens, release of cytokines to recruit more cells to the site of infection and act as antigen presenting cells to activate adaptive immune cells
Pathogen recognition receptors	Toll-Like receptors (TLRs) (NOD)-like receptors (NLRs) C-type lectin receptors (CLRs)	Identify pathogen-associated molecular patterns (PAMPs) activate a signaling cascades that regulate innate immune response to mobilize cytokines, chemokines and interferons which attract neutrophils and macrophages stimulate both local and systemic adaptive immune responses
Antimicrobial proteins and peptides	<i>Defensins</i> Antimicrobial agent with a wide range of activity against viruses, bacteria, fungi and protozoa. <i>Lysozyme</i> Antimicrobial molecule secreted by epithelial cells in the airway	Upregulate cytokines to attract neutrophils, mast cells, and T-lymphocytes Act by degrading the peptidoglycan in the bacterial cell wall
Surfactant collectins	Surfactant proteins A and D	Induce pathogen aggregation and enhance pathogen phagocytosis by macrophages and neutrophils; Regulate innate inflammation and resolution of tissue injury.

fold of the tubal floor, where ciliated cells predominate. The role played by surfactant in the pathobiology of OM is complex and entails both protein and lipid fractions.⁶³ Phosphatidylcholine (PC) is the same primary phospholipid which is found in lung as well as ET surfactant, but the difference is in the secondary phospholipid content. As in the lung, phosphatidylcholine (PC) is the main component of ear surfactants, however, phosphatidylethanolamine was higher and phosphatidylglycerol was undetectable in the middle ear.^{63,72}

Regarding PC to sphingomyelin ratio, ET lavage had a 2:1 ratio versus a 67:1 ratio found in pulmonary lavage. Variations in phospholipid content may be related to middle ear pathology. Svane-Knudsen *et al.* examined the concentrations of PC in lung and ET aspirates of normal subjects versus patients with secretory OM. They found that PC lung concentrations are the same in both groups, but they found a significant decrease in PC concentration in EF in patients with OM. Surfactant phospholipids did not significantly reduce surface tension in the air/ME

mucosa interface, but they have a cleansing function, facilitating the opening of the Ef and mucociliary transport, thus reducing $\text{mg}^b \text{ acten}^b \text{ al}^b \text{ mvas1}^b \text{ on}^{63} \text{ n}^{63}$

The ET surfactant appears to be an anti-adherent agent, rather than a surface tension lowering agent. As such, it may be perfectly suited to the structure and function of the eustachian tube.⁶⁰ Although the roles of SP-A and SP-D in the defense of the tubotympanum remains to be proven, it is likely that a deficiency in these host defense molecules may contribute to the pathogenesis of OM.⁷³

Adaptive immunity of OM

Immune system of mucosal surfaces can be divided into inductive site and effector site. In inductive sites, macrophages and dendritic cells process antigens and present it to naive B or T lymphocytes, whereas the transformation of B cells to antibody producing plasma cells takes place in effector sites. Inductive sites are organized in mucosal-associated lymphoid tissue (MALT), which is known more specifically in the nasal area as nasal associated lymphoid tissue (NALT). NALT consists of the adenoids and palatine tonsils which form Waldeyer's ring in humans.^{74, 75} Recently, respiratory microfold (M) cells were identified in the murine airways.⁷⁶ Respiratory M cells were capable of sampling inhaled bacterial antigens to initiate an Ag-specific immune response. It is widely accepted that NALT cells are key players in the uptake of nasally delivered antigens for the subsequent induction of antigen-specific IgA immune responses.^{77, 78} If these cells are present in the middle ear, they may result in the induction of a specific antibody response within the middle ear, particularly in patients who have chronic OM where germinal centers can be identified in the middle ear and Eustachian tube epithelium.^{79, 80} The main antibody that protects the mucosa of the middle ear is the secretory IgA and its presence highlights that the ME mucosa is an effector site of the mucosal immune system.⁷⁹

Genetic susceptibility of OM

Due to the high prevalence of OM, and the consequences of recurrence and chronicity of the disease, it is very important to understand its genetic background and susceptibility. Genetic differences could result in differences in anatomy of structures such as Ef or the nasopharynx or differences in immunological factors such as cytokines and mucins.¹³ Genetic studies of OM are based on twin studies, genome-wide linkage studies and genome-wide association studies (GWAS). In a monozygotic twins and triplets study, heritability was estimated to be 0.73 in both males and females ($p < 0.001$).^{13, 81} Genome-wide linkage studies (GWLS) identify regions of the genome that harbor disease susceptibility loci by typing microsatellite markers or single nucleotide polymorphisms (SNPs) spaced across the genome in sets of affected relatives.⁸¹ An evaluation of 588 patients undergoing tympanostomy tube insertion for chronic or

recurrent OM with DNA analysis, three important chromosomal regions were identified as important influencers, 10q, 19q, and 3p.¹³ Casselbrant *et al.* used the Genetic Relationships across Implicated Loci tool to identify possible candidate genes within their linkage regions. This identified a cluster of chemokine genes on 17q12 and several surfactant protein genes near 10q22.3. Possible candidate genes at these sites are surfactant protein gene SFP-A2 in the 10q22.3 region.⁸² The human SP-A locus in chromosome 10q22-q23 consists of 2 very similar genes, SP-A1 and SP-A2. Both gene products are required for fully functional SP-A protein. Several alleles that differ by single amino acid have been identified for each gene. The frequency of specific surfactant protein-A haplotypes and genotypes differs between children with recurrent otitis media compared with a control population.⁶¹ Using a candidate gene approach, Ramet *et al.* reported an over-representation of the 6A⁴-1A⁵ haplotype in children with recurrent otitis media and in children diagnosed with their first episode of AOM before 6 months; there was also an underrepresentation of the 6A²-1A⁰ haplotype in the latter subgroup.^{61, 83} A study by Wiertsema *et al.* on Polymorphisms of Mannose-binding Lectin (MBL) in relation to AOM found that MBL variant type was associated with an increased number of otitis media at 12 to 24 months of age.⁸⁴

Potential use of surfactant and surfactant proteins in the treatment of otitis media

The intranasal instillation or local nebulization of natural surfactants has a strong background and preclinical evidence showing positive effects on OM.⁶³ It is conceivable to suggest the administration of exogenous surfactant will improve the Ef function by reducing the surface tension, which helps in opening the Eustachian tube. Many studies have investigated a possible role for pulmonary surfactant in the treatment of OM with effusion (OME). Some of these studies have shown that using surfactant reduced the resolution time for OME.⁸⁵⁻⁸⁷ A recent study also showed that surfactant treatment for OME restored Ef function by reducing the opening pressure and increasing Ef compliance through the reduction of the mucosa-air surface tension.⁸⁸ The study also showed improvement of hearing threshold as well as improvement in the Ef mucosa and ciliary morphology compared with saline treated guinea pigs.⁸⁸ Another study showed that the administration of intranasal surfactant in OM chinchilla model lead to decrease in the severity and duration of infection.³²

Current treatment modalities for otitis media

Watchful waiting (WW), and withholding antibiotic therapy in mild to moderate AOM episodes is advocated by most guidelines. Antibiotic treatment is advised to be reserved for more

severe cases.^{14, 89} Children should be carefully monitored by caregivers with clear instructions to return to the doctor if symptoms persist or the child's condition worsened.⁹⁰ Usually, by the end of the first day after diagnosis, 61% of children with AOM have decreased symptoms, increasing to 80% after 2 to 3 days with or without antibiotics treatment, and approximately 75% have resolution of their symptoms by the end of the first week.⁹¹ Assessment and management of pain is an essential part of AOM treatment and for this, anti-inflammatory analgesics may be warranted. Topical and oral antibiotics are not recommended in all cases, immediate antibiotic treatment of AOM is recommended in very young children <6 months, children with craniofacial anomalies or who are immunocompromised, as well as those with severe illness due to AOM.⁹² Suppurative complications tend to occur whether initial antibiotics are provided (0.24%) or withheld (0.12%).⁹¹ Topical and systemic decongestants, antihistamines and corticosteroids are either ineffective or have shown conflicting results in AOM symptoms resolution. Antihistamine use should be avoided during AOM, since the drug may prolong the duration of middle ear effusion (MEE) and are therefore not recommended.⁹³ Myringotomy, a small incision of the ear drum that permits drainage of MEE, and can help in identifying the pathogens causing AOM, but is not an effective treatment for AOM. In case of recurrent AOM despite the proper medical treatment, surgical treatment with ventilation tube insertion is recommended.⁹⁴

Future directions

Vaccination efforts constitute some of the important future directions in the prevention of OM and include targeting viruses associated with OM²³ and targeting pneumococci through the development of new vaccines using pneumococcal proteins, targeting *M. catarrhalis*,⁹⁵ and development of noninvasive methods to immunize against.^{96, 97} Some of the future directions in surgery include The Acclarent, an automated ventilation tube insertion system which is a safe and effective device for placement of tympanostomy tubes in patients with chronic OME and recurrent AOM.⁹⁸ There is also a novel biodegradable drug-eluting ofloxacin-loaded ventilation tube with sustained drug release technology that has been tested to potentially treat chronic OME patients.⁹⁴ Balloon dilation of the ET is another treatment modality that improves the management of OME patients. This technology hopefully will be applied in the future for the treatment of children with chronic ET dysfunction.⁹⁹

Author contributions

O.A. reviewed publications and drafted the manuscript. J.A. edited and made one figure. D.S. edited the manuscript. G.W. reviewed publications and wrote the manuscript.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was supported in part by NIH R01HL136706, NSF research award (1722630), and one Award from Clark Endowment for Pediatric Research Fund (G.W.).

ORCID iDs

Osama Abdel-Razek  <https://orcid.org/0000-0002-8450-4067>
Guirong Wang  <https://orcid.org/0000-0002-3288-8572>

References

1. Ryan AF, et al. Mouse models of induced otitis media. *Brain Res* 2006; 1091: 3- 8.
2. Kostic M, et al. Clinical and audiological findings in children with acute otitis media. *Acta Otolaryngol* 2015; 135: 645-650.
3. Pettigrew MM, et al. Association of surfactant protein A polymorphisms with otitis media in infants at risk for asthma. *BMC Med Genet* 2006; 7: 68.
4. Qureishi A, Lee Y, Belfield K, et al. Update on otitis media - prevention and treatment. *Infect Drug Resist* 2014; 7: 15-24.
5. Monasta L, et al. Burden of disease caused by otitis media: systematic review and global estimates. *PLoS one* 2012; 7: e36226.
6. Pichichero ME. Immunologic dysfunction contributes to the otitis prone condition. *J Infect* 2020; 80: 614- 622.
7. Mittal R, et al. Role of innate immunity in the pathogenesis of otitis media. *International Journal of Infectious Diseases : IJID : Official Publication of the International Society for Infectious Diseases* 2014; 29: 259--267.
8. Paradise JL, et al. Otitis media in 2253 Pittsburgh-area infants: prevalence and risk factors during the first two years of life. *Pediatrics* 1997; 99: 318-333.
9. Ruohola A, Laine MK and Tahtinen PA. Effect of antimicrobial treatment on the resolution of middle-ear effusion after acute otitis media. *J Pediatric Infect Dis Soc* 2018; 7: 64-70.
10. Murphy TF, et al. Nontypeable *haemophilus influenzae* as a pathogen in children. *Pediatr Infect Dis J* 2009; 28: 43-48.
11. Lim DJ, et al. Cell biology of tubotympanum in relation to pathogenesis of otitis media - a review. *Vaccine* 2000; 19: S17-S25.
12. Toivonen L, et al. Burden of recurrent respiratory tract infections in children: a prospective cohort study. *Pediatr Infect Dis J* 2016; 35: e362- e369.
13. Post JC. Genetics of otitis media. *Adv Oto-Rhino-Laryngol* 2011; 70: 135-140.
14. Schilder AG, et al. Otitis media. Nature reviews. *Disease Primers* 2016; 2: 16063.
15. Leibovitz E, et al. Epidemiologic and microbiologic characteristics of culture-positive spontaneous otorrhea in children with acute otitis media. *Pediatr Infect Dis J* 2009; 28: 381-384.

16. Co ker TR, et al. Diagnosis, microbial epidemiology, and antibiotic treatment of acute otitis media in children: a systematic review. *JAMA* 2010; 304:2161-2169.

17. Heikkinen T, Ojala E and Waris M. Clinical and socio-economic burden of respiratory syncytial virus infection in children. *J Infect Dis* 2017; 215: 17-23.

18. Leibovitz E. The challenge of recalcitrant acute otitis media: pathogens, resistance, and treatment strategy. *Pediatr Infect Dis J* 2007; 26: S8-S11.

19. Leibovitz E, Jacobs MR and Dagan R. Haemophilus influenzae: a significant pathogen in acute otitis media. *Pediatr Infect Dis J* 2004; 23: 1142-1152.

20. Ruohola A, et al. Microbiology of acute otitis media in children with tympanostomy tubes: prevalences of bacteria and viruses. *Clinical Infectious Diseases : An Official Publication of the Infectious Diseases Society of America* 2006; 43: 1417- 1422.

21. Fletcher MA and Fritzell B. Pneumococcal conjugate vaccines and otitis media: an appraisal of the clinical trials. *Int J Otolaryngol* 2012; 2012: 312935.

22. Revai K, Mamidi D and Chonmaitree T. Association of nasopharyngeal bacterial colonization during upper respiratory tract infection and the development of acute otitis media. *Clinical Infectious Diseases : An Official Publication of the Infectious Diseases Society of America* 2008;46:e34-e37.

23. Marom T, Nokso-Koivisto J and Chonmaitree T. Viral-bacterial interactions in acute otitis media. *Curr Allergy Asthma Rep* 2012; 12: 551-558.

24. Giebink GS, Ripley ML and Wright PF. Eustachian tube histopathology during experimental influenza A virus infection in the chinchilla. *Ann Otol Rhinol Laryngol* 1987; %: 199-206.

25. Pettigrew MM, et al. Viral-bacterial interactions and risk of acute otitis media complicating upper respiratory tract infection. *J Clin Microbiol* 2011; 49: 3750-3755.

26. DeMuri GP, Gem JE, Eickhoff JC, et al. Dynamics of bacterial colonization with streptococcus pneumoniae, haemophilus influenzae, and moraxella catarrhalis during symptomatic and asymptomatic viral upper respiratory tract infection. *Clin Infect Dis* 2018; 66: 1045- 1053.

27. Cinamon U, et al. The middle ear cleft status in a "natural" cohort with eustachian tube dysfunction. *Otology & neurotology : official publication of the American Otological Society. American Neurotology Society [and] European Academy of Otology and Neurotology* 2017; 38: I133-1139.

28. Luo W, et al. Cilia distribution and polarity in the epithelial lining of the mouse middle ear cavity. *Sci Rep* 2017;7: 45870.

29. Bluestone CD. Pathogenesis of otitis media: role of eustachian tube. *Pediatr Infect Dis J* 1996; 15: 281-291.

30. Paananen R, Sormunen R, Glumoff V, et al. Surfactant proteins A and D in eustachian tube epithelium. *American Journal of Physiology. Lung Cellular and Molecular Physiology* 2001; 281:L660-L667.

31. Leichtle A, et al. The role of DNA sensing and innate immune receptor TLR9 in otitis media. *Innate Immun* 2012; 18: 3-13.

32. Chandrasekhar SS and Mautone AJ. Otitis media: treatment with intranasal aerosolized surfactant. *Laryngoscope* 2004; 114: 472-485.

33. Bluestone CD and Doyle WJ. Anatomy and physiology of eustachian tube and middle ear related to otitis media. *J Allergy Clin Immunol* 1988; 81: 997-1003.

34. Kang SH, et al. Expression of water channel proteins (aquaporins) in the rat eustachian tube and middle ear mucosa. *Acta Otolaryngol* 2001; 127: 687-692.

35. Sasai Mand Yamamoto M. Pathogen recognition receptors: ligands and signaling pathways by toll-like receptors. *Int Rev Immunol* 2013; 32: 116- 133.

36. Medzhitov R. Toll-like receptors and innate immunity. *Nature reviews. Immunology* 2001; 1 : 135- 145.

37. Leichtle A, Lai Y, Wollenberg B, et al. Innate signaling in otitis media: pathogenesis and recovery. *Curr Allergy Asthma Rep* 2011; 11: 78-84.

38. Huang Y, et al. TLR2 Promotes macrophage recruitment and streptococcus pneumoniae clearance during mouse otitis media. *Pediatr Res* 2016; 80: 886-893.

39. Beutler B. Inferences, questions and possibilities in toll-like receptor signalling. *Nature* 2004; 430: 257- 263.

40. Akira S and Takeda K Toll-like receptor signalling. *Nature reviews. Immunology* 2004; 4: 499- 511.

41. Emonts M, et al. Genetic polymorphisms in irnmunoresponse genes TNFA, IL6, IL10, and TLR4 are associated with recurrent acute otitis media. *Pediatrics* 2007; 120: 814- 823.

42. Shuto T, et al. Glucocorticoids synergistically enhance non-typeable haemophilus influenzae-induced toll-like receptor 2 expression via a negative cross-talk with p38 MAP kinase. *J Biol Chem* 2002; 277: 17263- 17270.

43. Hirano T, Kodama S, Fujita K, et al. Role of toll-like receptor 4 in innate immune responses in a mouse model of acute otitis media. *FEMS Immunol Med Microbiol* 2007; 49: 75- 83.

44. Leichtle A, et al. TLR4-mediated Induction of TLR2 signalling is critical in the pathogenesis and resolution of otitis media. *Innate Immun* 2009; 15: 205-215.

45. Hafren L, et al. Predisposition to childhood otitis media and genetic polymorphisms within the toll-like receptor 4 (TLR4) locus. *PLoS One* 2015; 10: e0132551.

46. Toivonen L, et al. Polymorphisms of mannose-binding lectin and toll-like receptors 2, 3, 4, 7 and 8 and the risk of respiratory infections and acute otitis media in children. *Pediatr Infect Dis J* 2017; 36: e14-e122.

47. Strober W, Murray PJ, Kitani A, et al. Signalling pathways and molecular interactions of NOD1 and NOD2. *Nat Rev Immunol* 2006; 6: 9-20.

48. Kirn SH, et al. Age-dependent changes in pattern recognition receptor and cytokine mRNA expression in children with otitis media with effusion. *Int J Pediatr Otorhinolaryngol* 2015; 79: 229-234.

49. Yang D, et al. Defensin participation in innate and adaptive immunity. *Curr Pharm Des* 2001; 13: 3131-3139.

50. Parker D and Prince A. Innate immunity in the respiratory epithelium. *Am J Respir Cell Mol Biol* 2011; 45: 189- 201.

51. Somaschini M, Presi S, Ferrari M, et al. Surfactant proteins gene variants in premature newborn infants with severe respiratory distress syndrome. *Journal of Perinatology : Official Journal of the California Perinatal Association* 2018; 38: 337-344.

52. Weaver TE and Conkright JJ. Function of surfactant proteins B and C. *Annu Rev Physiol* 2001; 63: 555-578.

53. Gupta G and Surolia A. Collectins: sentinels of innate immunity. *BioEssays: News and Reviews in Molecular, Cellular and Developmental Biology* 2007; 29: 452-464.

54. Li L, et al. Expression of surfactant protein-A during LPS-induced otitis media with effusion in mice. *Otolaryngology--head and Neck Surgery : Official Journal of American Academy of Otolaryngology-Head and Neck Surgery* 2015; 153: 433-439.

55. Zhang L, et al. Surfactant proteins-A and -D attenuate LPS-induced apoptosis in primary intestinal epithelial cells (IECs). *Shock* 2018; 49: 90-98.

56. LeVine AM, et al. Distinct effects of surfactant protein A or D deficiency during bacterial infection on the lung. *J Immunol* 2000; 165: 3934-3940.

57. Oberley RE and Snyder JM. Recombinant human SP-A1 and SP-A2 proteins have different carbohydrate-binding characteristics. *American Journal of Physiology. Lung Cellular and Molecular Physiology* 2003; 284: L871-L881.

58. Vieira F, Kung JW and Bhatti F. Structure, genetics and function of the pulmonary associated surfactant proteins A and D: the extra-pulmonary role of these C type lectins. *Annals of Anatomy= Anatomischer Anzeiger : Official Organ of the Anatomische Gesellschaft* 2017; 211: 184-201.

59. Abdel-Razek O, Ni L, Yang F, et al. Innate immunity of surfactant protein A in experimental otitis media. *Immunol Lett* 2019; 25: 391-400.

60. McGuire JF. Surfactant in the middle ear and eustachian tube: a review. *Int J Pediatr Otorhinolaryngol* 2002; 66: 1-15.

61. Ramet M, Lofgren J, Alho OP, et al. Surfactant protein-A gene locus associated with recurrent otitis media. *J Pediatr* 2001; 138: 266-268.

62. Bourbon JR and Chailley-Heu B. Surfactant proteins in the digestive tract, mesentery, and other organs: evolutionary significance. *Comp Biochem Physiol A Mol Integr Physiol* 2001; 129: 151-161.

63. Foligno S, et al. Extrapulmonary surfactant therapy: review of available data and research/development issues. *J Clin Pharmacol* 2020; 60: 1561-1572.

64. Madhukaran SP, Kishore U, Jamil K, et al. Decidual expression and localization of human surfactant protein SP-A and SP-D, and complement protein Clq. *Mol Immunol* 2015; 66: 197-207.

65. Konishi M, et al. Alloioococcus otitidis is a ligand for collectins and toll-like receptor 2, and its phagocytosis is enhanced by collectins. *Eur J Immunol* 2006; 36: 1527-1536.

66. Eggleton P and Reid KB. Lung surfactant proteins involved in innate immunity. *Curr Opin Immunol* 1999; 11: 28-33.

67. Malhotra R, Lu J, Holmskov U, et al. Collectins, collectin receptors and the lectin pathway of complement activation. *Clin Exp Immunol* 1994; 97: 4-9.

68. Petersen SV, Thiel S and Jensenius JC. The mannose-binding lectin pathway of complement activation: biology and disease association. *Mol Immunol* 2001; 38: 133-149.

69. Jakel A, Qaseem AS, Kishore U, et al. Ligands and receptors of lung surfactant proteins SP-A and SP-D. *Front Biosci* 2013; 18: 1129-1140.

70. Holmskov U, Malhotra R, Sim RB, et al. Collectins: collagenous C-type lectins of the innate immune defense system. *Immunol Today* 1994; 15: 67-74.

71. Ramadas RA, Wu L and LeVine AM. Surfactant protein A enhances production of secretory leukoprotease inhibitor and protects it from cleavage by matrix metalloproteinases. *J Immunol* 2009; 182: 1560-1567.

72. Paananen R, Postle AD, Clark G, et al. Eustachian tube surfactant is different from alveolar surfactant: determination of phospholipid composition of porcine eustachian tube lavage fluid. *J Lipid Res* 2002; 43: 99-106.

73. Park K and Lim DJ. Luminal development of the eustachian tube and middle ear: murine model. *Yonsei Med J* 1992; 33: 159-167.

74. Neutra MR and Kozlowski PA. Mucosal vaccines: the promise and the challenge. *Nat Rev Immunol* 2006; 6: 148-158.

75. Sabirov A and Metzger DW. Intranasal vaccination of infant mice induces protective immunity in the absence of nasal-associated lymphoid tissue. *Vaccine* 2008; 26: 1566-1576.

76. MacArthur CJ, Pillers DA, Pang J, et al. Altered expression of middle and inner ear cytokines in mouse otitis media. *Laryngoscope* 2011; 121: 365-371.

77. Kiyono H and Fukuyama S. NALT- versus Peyer's-patch-mediated mucosal immunity. *Nat Rev Immunol* 2004; 4: 699-710.

78. Kim DY, et al. The airway antigen sampling system: respiratory M cells as an alternative gateway for inhaled antigens. *J Immunol* 2011; 186: 4253-4262.

79. Massa HM, Lim DJ, Kurono Y, et al. Chapter 101 - Middle Ear and Eustachian Tube Mucosa! Immunology A2 - Mestecky, Jiri. In: Strober W, Russell MW, Kelsall BL, Cheroutre H and Lambrecht BN (eds) *Mucosal Immunology (Fourth Edition)*. 1923-1942. Boston: Academic Press, 2015, pp. 1934-1935.

80. Matsune S, Takahashi H and Sando I. Mucosa-associated lymphoid tissue in middle ear and eustachian tube in children. *Int J Pediatr Otorhinolaryngol* 1996; 34: 229-236.

81. Rye MS, Blackwell JM and Jamieson SE. Genetic susceptibility to otitis media in childhood. *Laryngoscope* 2012; 122: 665-675.

82. Casselbrant ML, et al. Otitis media: a genome-wide linkage scan with evidence of susceptibility loci within the 17q12 and 10q22.3 regions. *BMC Med Genet* 2009; 10: 85.

83. Casselbrant ML and Mandel EM. Genetic susceptibility to otitis media. *Curr Opin Allergy Clin Immunol* 2005; 5: 1-4.

84. Wiertsema SP, et al. Functional polymorphisms in the mannose-binding lectin 2 gene: effect on MBL levels and otitis media. *J Allergy Clin Immunol* 2006; 117: 1344-1350.

85. Koten M, et al. Nebulized surfactant as a treatment choice for otitis media with effusion: an experimental study in the rabbit. *J Laryngol Otol* 2001; 115: 363-368.

86. Venkatayam N, Troublefield YL, Connelly PE, et al. Intranasal surfactant aerosol therapy for otitis media with effusion. *Laryngoscope* 2000; 110: 1857-1860.

87. Venkatayam N, Connelly PE, Mautone AJ, et al. Dosage regimens of intranasal aerosolized surfactant on otitis media with effusion in an animal model. *Otolaryngology--head and Neck Surgery : Official Journal of American Academy of Otolaryngology-Head and Neck Surgery* 2001; 124: 388-393.

88. Zhu ZH, Shan YJ, Han Y, et al. Pathological study of otitis media with effusion after treatment with intranasal pulmonary surfactant. *Laryngoscope* 2013; 123: 3148-3155.

89. Rothman S, et al. Appropriate and Inappropriate Treatment of Acute Otitis Media in the Pediatric Emergency Department. *Pediatr Infect Dis J* 2017; 37(6): 520-525.

90. Lieberthal AS, et al. The diagnosis and management of acute otitis media. *Pediatrics* 2013; 131: e964-e999.
91. Rosenfeld RM and Kay D. Natural history of untreated otitis media. *Laryngoscope* 2003; 113: 1645-1657.
92. Hellstrom S, et al Ventilation tube treatment: a systematic review of the literature. *Otolaryngology--head and Neck Surgery : Official Journal of American Academy of Otolaryngology-Head and Neck Surgery* 2011; 145: 383- 395.
93. Chonmaitree T, et al A randomized, placebo-controlled trial of the effect of antihistamine or corticosteroid treatment in acute otitis media. *J Pediatr* 2003; 143: 377-385.
94. Gan CW, et al. Development of a novel biodegradable drug-eluting ventilation tube for chronic otitis media with effusion. *Laryngoscope* 2013; 123: 1770--1777.
95. Brockson ME, et al. Respiratory syncytial virus promotes *moraxella ca tarrhalis*-induced ascending experimental otitis media. *PloS one* 2012; 7: e40088.
96. Preciado D (ed.). *Otitis Media: State of the Art Concepts and Treatment*. xx: Springer International Publishing, 2015.
97. De Magistris MT. Mucosa! delivery of vaccine antigens and its advantages in pediatrics. *Adv Drug Delivery Rev* 2006; 58: 52--o7.
98. Syms CA GA, Zeiders JW and Faw KD. Automated tube deployment: success demonstrated in OR. *Otolaryngol Head Neck Surg* 2011; 145: 51.
99. Poe D, et al. Balloon dilation of the eustachian tube for dilatory dysfunction: A randomized co ntrolled trial *Laryngoscope* 2017; 128: 1200--1206.