# Non-paraxial multiple scattering model for intensity diffraction tomography

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**Abstract:** We propose a novel intensity diffraction tomography reconstruction algorithm based on the split-step non-paraxial model for recovering the 3D refractive index distribution of multiple-scattering biological samples. © 2022 The Author(s)

### 1. Introduction

3D quantitative phase imaging (QPI) are attractive for characterizing thick biological samples by providing refractive index (RI) information of the samples. Recently, intensity diffraction tomography (IDT) has been developed [1–4]. As a phase-less technique, IDT can be easily implemented on a standard microscope using a programmable LED array. Our group has developed two strategies to push the acquisition speed enabling visualizing dynamic biological samples. The multiplexed IDT (mIDT) [4] using multiple LEDs to illuminate the sample simultaneously with a widely used LED matrix as shown in Fig.1(a). The annular IDT (aIDT) [3] using an LED ring matching the objective's numerical aperture (NA) as shown in Fig.1(b).

IDT reconstruction algorithms relying on the single scattering models [2–4] are limited to weak scattering samples, and algorithms based on multiple-scattering beam propagation method (BPM) [1] have degraded accuracy for high-resolution imaging using high-NA optics. The split-step non-paraxial (SSNP) method was recently

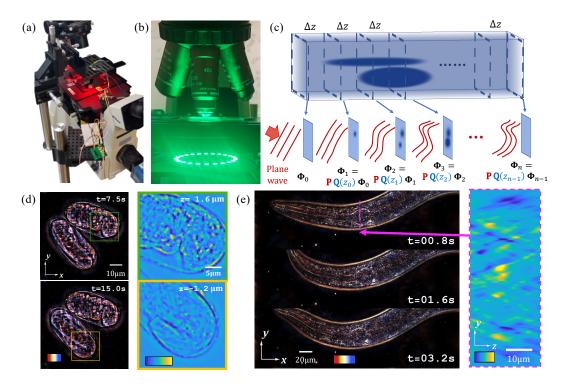


Fig. 1. (a) Our IDT/mIDT setup uses an LED matrix. (b) Our annular IDT setup uses an LED ring. (c) The SSNP model involves successive propagation to compute the scattered field. (d) Live *C. elegans* embryos reconstruction result from mIDT. (e) Live *C. elegans* worm reconstruction result from aIDT.

proposed as an alternative multiple-scattering model for ODT [5] to overcome the limitations in the first Born and BPM methods. We extended the SSNP method to IDT measurements [6] and demonstrate high-quality, large field-of-view (FOV) reconstruction results of live *C. elegans* embryos from mIDT and live *C. elegans* worm from aIDT.

# 2. Procedure of SSNP-based IDT model

#### 2.1. Forward SSNP IDT model

The SSNP model discretizes a 3D sample into a series of axial (z) slices and calculates the internal field slice-byslice (xy), as illustrated in Fig. 1(c). The scattering process of SSNP-based IDT model  $S\{\cdot\}$  with single LED can be written as:

$$S\{\mathbf{\Phi}_{xy}(z_0)\} = \left|\mathbf{F}\mathscr{P}_{NA}\mathbf{P}_{\Delta z_f}\mathbf{P}\mathbf{Q}(z_{n-1})\dots\mathbf{P}\mathbf{Q}(z_1)\mathbf{P}\mathbf{Q}(z_0)\mathbf{\Phi}_{xy}(z_0)\right|^2,\tag{1}$$

where  $\Phi_{xy}(z_0)$  denotes the illumination field, **P** operator denotes propagation in homogeneous media,  $\mathbf{Q}(z)$  operator denotes scattering of the slice at axial position z,  $\mathbf{P}_{\Delta z_f}$  denotes propagation operator to the focal plane;  $\mathscr{P}_{NA}$  denotes low-pass filtering by the pupil function, **F** operator denotes extracting the forward-propagating field from the field vector.

For aIDT and mIDT, the estimated intensities are the combination of the scattered result calculated using SSNP algorithm from corresponding illuminations:

$$I_{seq}^{l} = S\{\mathbf{\Phi}_{xy}^{l}(z_{0})\}, \quad I_{mul}^{l} = \sum_{m=1}^{M} S\{\mathbf{\Phi}_{xy}^{l,m}(z_{0})\}$$
 (2)

where  $I_{seq}^l$  and  $I_{mul}^l$  denotes the lth intensity for aIDT and mIDT respectively, and  $\Phi_{xy}^{l,m}(z_0)$  specifies the illumination field from the mth LED in the lth pattern.

### 2.2. Inverse problem of SSNP-based IDT model

We formulate the aIDT and mIDT reconstruction as the following optimization problem:

$$\hat{n}_r = \underset{n_r \in \Theta}{\operatorname{argmin}} \{ \sum_{l=1}^{L} \|I_{SSNP}^l - I_{meas}^l\|_2^2 + \tau R_{TV}(n_r) \}$$
(3)

where  $I_{SSNP}^l$  denotes the *l*th intensity estimation from SSNP-based aIDT and mIDT model;  $I_{meas}^l$  denotes the *l*th intensity measurement;  $\tau$  is the regularization parameter, and  $R_{TV}$  is the total variation regularizer which suppresses the noise and artifacts. We perform reconstruction by iteratively refining the 3D RI estimation  $n_r(x, y, z)$ .

# 3. Results

We first apply our algorithm to a thick multi-scattering sample of live C. elegans embryos. In Fig. 1(d), the colorbars shows depth ranging from  $-9.7 \,\mu m$  to  $9.7 \,\mu m$  and RI ranging from 1.327 to 1.336. From the color-coded view (left), how the worms are folded can be clearly observed; from the single-depth cross section (right), the morphological details of the cells' outline, the buccal cavity, and the tail of the worm are reconstructed.

We also apply our algorithm to a live C. elegans worm sample from aIDT. In Fig. 1(d), the colorbars shows depth ranging from  $-9.6 \,\mu m$  to  $9.6 \,\mu m$  and RI ranging from 1.32 to 1.34. Figure 1(e) shows three consecutive volumes reconstructed by our SSNP algorithm displayed as color-coded depth projections, and a zoomed-in YZ cross-sectional views around the terminal pharyngeal bulb region.

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