

Site-Differentiated $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ Complex Reproducing the Selective Assembly of Biological Heterobimetallic Mn/Fe Cofactors

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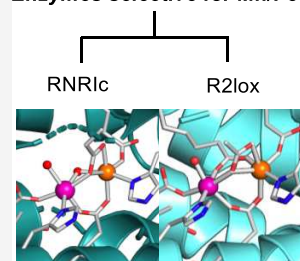
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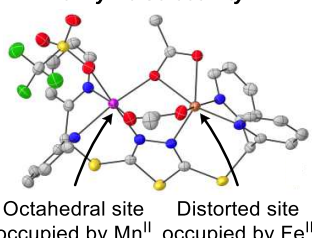
Supporting Information

ABSTRACT: Class Ic ribonucleotide reductases (RNRIc) and R2-like ligand-binding oxidases (R2lox) are known to contain heterobimetallic $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ cofactors. How these enzymes assemble $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ cofactors has been a long-standing puzzle due to the weaker binding affinity of Mn^{II} versus Fe^{II} . In addition, the heterobimetallic selectivity of RNRIc and R2lox has yet to be reproduced with coordination complexes, leading to the hypothesis that RNRIc and R2lox overcome the thermodynamic preference for coordination of Fe^{II} over Mn^{II} with their carefully constructed three-dimensional protein structures. Herein, we report the selective formation of a heterobimetallic $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ complex accomplished in the absence of a protein scaffold. Treatment of the ligand Py_4DMcT (L) with equimolar amounts of Fe^{II} and Mn^{II} along with two equivalents of acetate (OAc) affords $[\text{LMn}^{\text{II}}\text{Fe}^{\text{II}}(\text{OAc})_2(\text{OTf})]^+$ ($\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$) in 80% yield, while the diiron complex $[\text{LFe}^{\text{II}}\text{Fe}^{\text{II}}(\text{OAc})_2(\text{OTf})]^+$ ($\text{Fe}^{\text{II}}\text{Fe}^{\text{II}}$) is produced in only 8% yield. The formation of $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ is favored regardless of the order of addition of Fe^{II} and Mn^{II} sources. X-ray diffraction (XRD) of single crystals of $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ reveals an unsymmetrically coordinated carboxylate ligand—a primary coordination sphere feature shared by both RNRIc and R2lox that differentiates the two metal binding sites. Anomalous XRD studies confirm that $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ exhibits the same site selectivity as R2lox and RNRIc, with the Fe^{II} (d^6) center preferentially occupying the distorted octahedral site. We conclude that the successful assembly of $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ originates from (1) Fe-deficient conditions, (2) site differentiation, and (3) the inability of ligand L to house a dimanganese complex.

Enzymes selective for Mn/Fe



Synthetic model reproduces enzyme selectivity



INTRODUCTION

Heterobimetallic cofactors have been discovered in a number of proteins responsible for life-sustaining chemical transformations, including the heme a_3/Cu_B site in cytochrome *c* oxidase, $[\text{NiFe}]$ hydrogenase, Cu/Zn superoxide dismutase, and the Mn/Fe or Zn/Fe sites in purple acid phosphatases.¹ Mn/Fe cofactors have also been characterized in class Ic ribonucleotide reductases (RNRIc),^{2–9} R2-like ligand-binding oxidases (R2lox),^{10–25} and, most recently, in the chlamydia protein associated with death domains (CADD).²⁶ Since their discovery, heterobimetallic cofactors have piqued the interest of chemists and biologists alike. Understanding the mechanisms behind metal cofactor assembly can aid in answering important questions regarding protein evolution and metal selectivity in native and engineered proteins.^{27–29}

Of the heterobimetallic cofactors, the most intriguing are those that incorporate Mn^{II} and Fe^{II} —two metal ions of similar size and binding affinity—with apparent selectivity.³⁰ Despite sharing similar coordination environments to the diiron sites in canonical bacterial multicomponent monooxygenases (BMMs),^{31–34} both RNRIc and R2lox feature distinct, site-selective heterobimetallic Mn/Fe cofactors (Figure 1A).^{6,19,20}

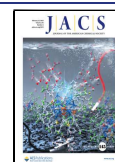
The mechanism of heterobimetallic $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ cofactor assembly *in vivo* has been the subject of intense scrutiny. Work by Högbom et al.,^{12,13,19–25} Shafaat et al.,^{14–17} Stubbe et

al.,^{3–5} and Bollinger et al.^{6–9} suggests that favorable $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ assembly is likely driven by a combination of metal availability and differential binding affinity. While *in vivo* studies have provided valuable insights into the selective formation of $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ cofactors, the complexity of the protein matrix has prevented the identification of specific structural features responsible for this process.

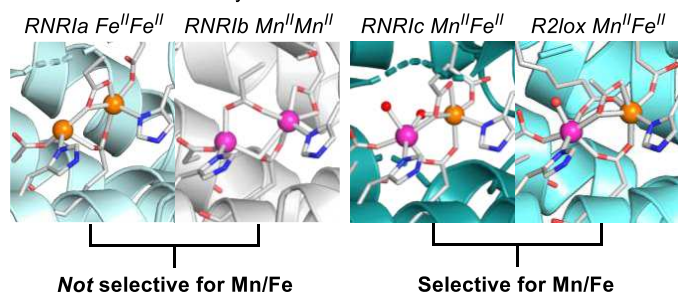
As studies on the assembly of RNRIc and R2lox continue, there have also been a number of attempts to replicate the primary coordination sphere, site differentiation, and selectivity of RNRIc and R2lox with coordination compounds.^{35–49} However, the synthesis of $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ species is often complicated by simultaneous formation and/or scrambling of the heterobimetallic complex to form homobimetallic $\text{Fe}^{\text{II}}\text{Fe}^{\text{II}}$ and $\text{Mn}^{\text{II}}\text{Mn}^{\text{II}}$ analogues (Figure 1B).^{38,39} To synthesize pure Mn/Fe complexes, pioneering work by Que et al. employed iron(III)/manganese(II) sources, along with a stepwise

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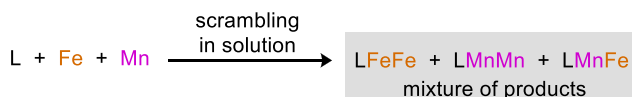


A. Nature can selectively form Mn/Fe active sites



B. Challenges to synthetically replicating Nature's selectivity

Contamination by homobimetallic analogues is common

C. This work: First selective self-assembly of an Mn^{II}Fe^{II} complex

Differentiation between Mn^{II} and Fe^{II} occurs via the unsymmetric bridging carboxylate

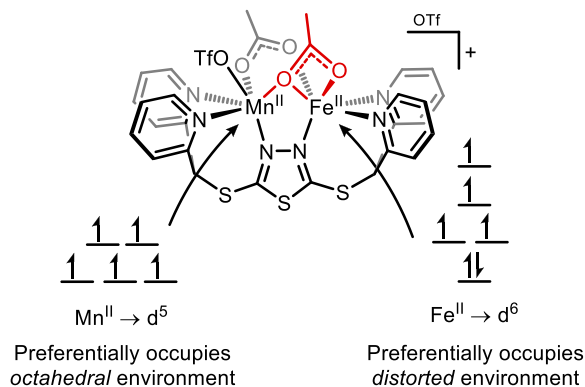


Figure 1. (A) Pymol rendered active sites of class Ia ribonucleotide reductase (PDB: 1PIY), Ib (PDB: 6TQW), Ic (PDB: 4M1I), and R2lox (PDB: 4HR4). (B) Scrambling of heterobimetallic complexes to form homobimetallic complexes. (C) Site-differentiated Mn^{II}Fe^{II} complex in this work.

synthetic route to differentiate the binding event of each metal (Figure 2A).^{35,44} A second synthetic strategy was developed by Wieghardt/Girerd et al., in which the heterobimetallic species was formed via self-assembly from two independently synthesized mononuclear complexes (Figure 2B).^{36,43} Following these early studies, Neves et al.,⁴⁵ Chaudhuri et al.,⁴⁶ Blondin/Latour et al.,⁴⁸ Borovik et al.,^{40,49} and Lu et al.³⁸ have utilized ligands with unsymmetric coordination environments to further bias the coordination of each metal to inequivalent sites (Figure 2C,D).

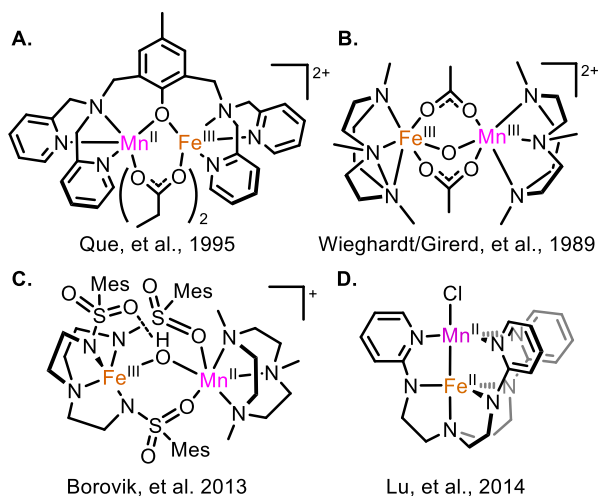


Figure 2. (A–D) Previously reported synthetic Mn/Fe complexes.

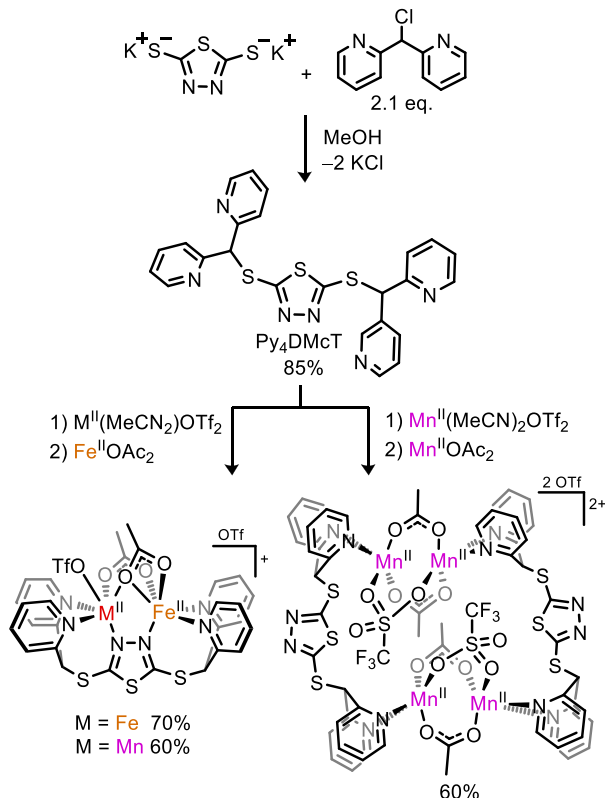
While previous synthetic studies have greatly increased our understanding of Mn/Fe complex formation, none have been able to shed light on the formation of MnFe cofactors under biologically relevant conditions, i.e., self-assembly in the presence of both Mn and Fe sources at a II,II state. Within the primary coordination spheres of RNR1c and R2lox, the only feature that differentiates the two metal binding sites is an unsymmetric μ -1,1 bridging carboxylate residue. The unsymmetric binding mode of the carboxylate leads to distinct geometries at each site: an octahedral site that preferentially

binds Mn^{II} and a distorted octahedral site that primarily binds Fe^{II} (Figure 1A).^{6,19,20}

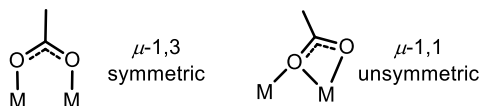
Herein, we report the first synthetic Mn^{II}Fe^{II} complex containing an unsymmetric μ -1,1 bridging acetate ligand. Despite the lack of a protein scaffold, the Mn^{II}Fe^{II} complex self-assembles in the presence of both Fe^{II} and Mn^{II} (Figure 1C). Regardless of the order of Fe^{II} and Mn^{II} addition to the ligand Py₄DMcT, the heterobimetallic complex remains the major product with >72% selectivity. Anomalous X-ray diffraction (XRD) studies confirm that the Mn^{II}Fe^{II} model complex has the same site selectivity as R2lox and RNR1c, where Mn^{II} occupies the octahedral site and Fe^{II} occupies the distorted site. The structural similarities of our model complex to R2lox and RNR1c, along with the same metal site selectivity, allow us to propose a mechanism for heterobimetallic Mn^{II}Fe^{II} formation that does not contradict the classical Irving–Williams series.

RESULTS AND DISCUSSION

Synthesis and Characterization. To design a ligand capable of supporting a Mn/Fe bimetallic center, we modified our previous Py₄DMB system^{50,51} by introducing a dimercaptan-1,3,4-thiadiazole linker. The new ligand L was synthesized in 85% yield (Py₄DMcT, L; Scheme 1). Treatment of L with one equivalent each of bis-acetonitrile iron(II) triflate (Fe^{II}(OTf)₂MeCN₂) and iron(II) acetate (Fe^{II}(OAc)₂) in a 1:1 mixture of chloroform/methanol, followed by recrystallization from dichloromethane/ether, affords orange crystals of [LFe^{II}₂(OAc)₂(OTf)](OTf) (Fe^{II}Fe^{II}) in 70% yield (Scheme 2, left). We found that the heterobimetallic analogue can be synthesized by replacing Fe^{II}(OTf)₂MeCN₂ with one equivalent of bis-acetonitrile manganese(II) triflate (Mn^{II}(OTf)₂MeCN₂) (Scheme 1, left). Recrystallization from dichloromethane/ether results in yellow crystals of [LMn^{II}Fe^{II}(OAc)₂(OTf)](OTf) (Mn^{II}Fe^{II}) in 60% yield (Figure 2B). Single-crystal XRD analysis reveals that both Fe^{II}Fe^{II} and Mn^{II}Fe^{II} contain two bridging acetate anions with symmetric and unsymmetric μ -1,3/ μ -1,1 binding modes, respectively (Scheme 2). Despite being isostructural, Fe^{II}Fe^{II} and Mn^{II}Fe^{II} crystallize with different unit cell parameters (Figure 3A,B; see the Supporting Information). The M–M separation in Mn^{II}Fe^{II} (3.4794(6) Å) is also slightly longer

Scheme 1. Synthesis of Py₄DMcT (L) and Metal Complexes

Scheme 2. Symmetric versus Unsymmetric Acetate Binding Modes



compared to the 3.46409(16) Å distance in $\text{Fe}^{II}\text{Fe}^{II}$, consistent with the larger atomic radius of Mn^{II} versus Fe^{II} .

While an unsymmetric μ -1,1 carboxylate ligand (Scheme 2) is a common structural feature in RNRs, R2lox, and BMMs,^{31–34} it is not as common in synthetic systems.^{52–57} Notably, a μ -1,1 carboxylate ligand has never been reported in a synthetic Mn/Fe complex until now. In our synthetic system, the unsymmetric acetate replicates the unsymmetric carboxylates in RNR1c (E227) and R2lox (E202), producing the same site-specific geometries as the enzymes. The unsymmetric μ -1,1 binding of acetate in $\text{Fe}^{II}\text{Fe}^{II}$ and $\text{Mn}^{II}\text{Fe}^{II}$ is likely promoted by the outward tilting of the metal z -axes following coordination of the metal centers to the N atoms of the thiadiazole ring. Similar outward tilting of the metal z -axes can be observed in RNR1c and R2lox (see the Supporting Information, Figure S36), although such a structural feature is not unique to these enzymes.^{31–34,58} In our complexes, the N atoms of the central ring form a template for a short M–M distance, making the six-membered metallacycle of μ -1,1/ μ -1,3 binding mode more favorable than the eight-membered metallacycle of μ -1,3/ μ -1,3 binding mode.

The overall structure of $\text{Mn}^{II}\text{Fe}^{II}$ more closely resembles the unsymmetric core of R2lox, as it lacks the solvent-derived O-atom bridge present in RNR1c. Further, the metal separation in $\text{Mn}^{II}\text{Fe}^{II}$ is more comparable to the M–M distance seen in R2lox (3.65 Å)¹⁹ than RNR1c (3.2 Å).²⁵ Given the structural

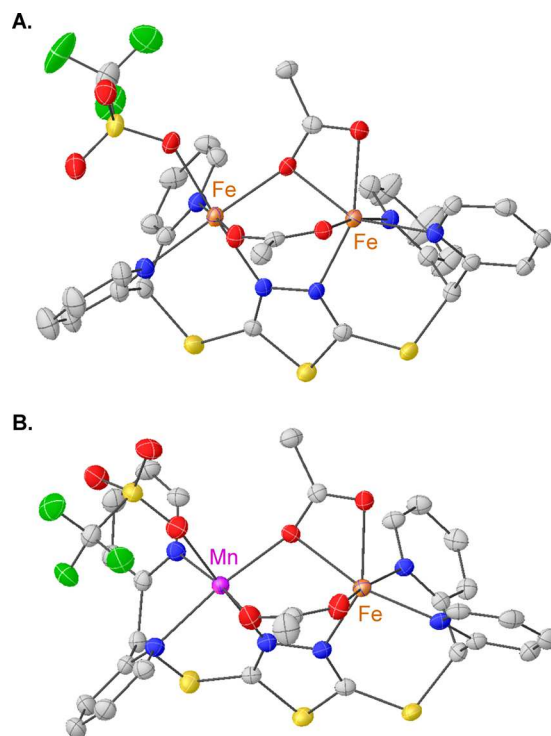


Figure 3. Solid-state structures of (A) $\text{Fe}^{II}\text{Fe}^{II}$ and (B) $\text{Mn}^{II}\text{Fe}^{II}$ with thermal ellipsoids shown at 50% level of probability. The outer sphere triflate counter anion and co-crystallized dichloromethane solvent molecules are omitted for clarity.

similarities between $\text{Mn}^{II}\text{Fe}^{II}$ and R2lox, we tentatively assigned the metal center at the distorted octahedral site as Fe^{II} . Such an assignment also results in a better agreement with the electron density map, where $R_1 = 4.30\%$ vs 4.46% with a reversed Fe/Mn assignment.

The heterobimetallic identity of $\text{Mn}^{II}\text{Fe}^{II}$ was further confirmed by spectroscopic analysis. The ^1H NMR spectrum of $\text{Mn}^{II}\text{Fe}^{II}$ reveals eight broad resonances from -30 to 140 ppm that are distinct from the paramagnetic peaks of $\text{Fe}^{II}\text{Fe}^{II}$ (Figure 4A), consistent with the unsymmetric nature of $\text{Mn}^{II}\text{Fe}^{II}$.³⁹ Mössbauer spectroscopy reveals that $\text{Fe}^{II}\text{Fe}^{II}$ is best fitted with two distinct iron sites, whereas $\text{Mn}^{II}\text{Fe}^{II}$ is fitted best with a single iron site (Figure 4B,C). The isomer shifts of $\text{Fe}^{II}\text{Fe}^{II}$ ($\delta_1 = 1.20 \text{ mm s}^{-1}$, $\delta_2 = 1.22 \text{ mm s}^{-1}$) and $\text{Mn}^{II}\text{Fe}^{II}$ ($\delta = 1.28 \text{ mm s}^{-1}$) are consistent with high-spin Fe^{II} centers and are similar to the reported value for the $\text{Mn}^{II}\text{Fe}^{II}$ cofactor of R2lox ($\delta = 1.28 \text{ mm s}^{-1}$).²⁰ This indicates that $\text{Mn}^{II}\text{Fe}^{II}$ accurately models not only the geometry of the heterobimetallic cofactor but also the spin state of the metal ions. X-band electron paramagnetic resonance spectroscopy (EPR) of $\text{Mn}^{II}\text{Fe}^{II}$ shows a broad $S = 1/2$ signal (Figure S7), similar to the EPR spectra of $\text{Mn}^{II}\text{Fe}^{II}$ complexes reported by Que et al. and Carboni et al.^{44,48} Inductively coupled plasma mass spectrometry (ICP-MS) analysis of $\text{Mn}^{II}\text{Fe}^{II}$ indicates the incorporation of Fe^{II} and Mn^{II} in approximately a 1:1 ratio (4.16% Fe^{II} to 4.74% Mn^{II}).

After the isolation and characterization of $\text{Fe}^{II}\text{Fe}^{II}$ and $\text{Mn}^{II}\text{Fe}^{II}$, we attempted to complete the series of bimetallic complexes by preparing the analogous dimanganese species. However, treatment of **L** with $\text{Mn}^{II}(\text{OAc})_2$ and $\text{Mn}(\text{OTf})_2\text{MeCN}_2$ resulted in the isolation of a colorless, NMR-silent tetramanganese paddlewheel complex, Mn_4^{II} (Scheme 2, right).^{39,59} Single-crystal XRD analysis shows two dimanganese

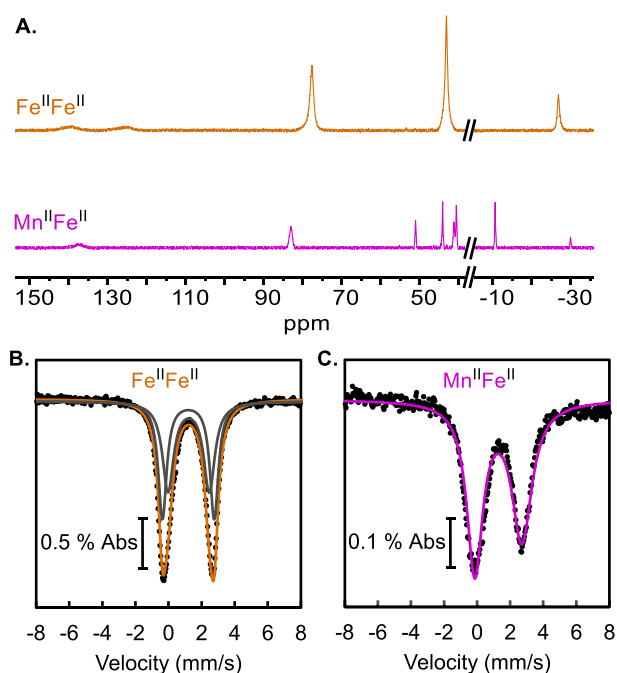


Figure 4. (A) ¹H NMR spectra of Fe^{II}Fe^{II} (orange) and Mn^{II}Fe^{II} (pink), 2.5 mM in CD₂Cl₂. Solid-state ⁵⁷Fe Mössbauer spectra of (B) Fe^{II}Fe^{II} and (C) Mn^{II}Fe^{II}.

paddlewheels sandwiched between two Py₄DMcT ligands, with all Mn^{II} ions symmetrically bridged by two acetate anions and one triflate anion, and a Mn–Mn distance of 3.860 Å (see the SI, Figure S35). The formation of the tetramanganese complex, rather than a dimanganese species, suggests that L is unable to accommodate two Mn^{II} centers in the same bimetallic environment, perhaps due to the larger size of Mn^{II} versus Fe^{II}. This observation also coincides with the lack of Mn^{II}Mn^{II} cofactor formation in R2lox.^{16,19,20,23,24}

Heterobimetallic Selectivity. With the spectroscopic characterization of Fe^{II}Fe^{II}, Mn^{II}Fe^{II}, and Mn^{II}₄, we wanted to determine whether L could reproduce RNR1c and R2lox's ability to selectively assemble a heterobimetallic Mn/Fe center. Importantly, Mn^{II}Fe^{II} is stable in CD₂Cl₂ at room temperature without noticeable metal scrambling for at least 12 h (Figure S8), allowing the use of in situ ¹H NMR to quantify Mn^{II}Fe^{II} formation. First, we investigated whether the order of addition of Fe^{II} and Mn^{II} sources to L influences the yield of Mn^{II}Fe^{II}. Ligand L was treated with Fe^{II} and Mn^{II} sources in different orders. Regardless of the order of addition, the resulting ¹H NMR spectra show that Mn^{II}Fe^{II} is produced in similar yields (74–77%; Table 1) with minimal (ca. 6%) Fe^{II}Fe^{II} contamination. As Mn^{II}Fe^{II} is the major product under all conditions, we posited that the formation of Mn^{II}Fe^{II} likely occurs selectively via the same assembly route, no matter the

order of metal addition. Indeed, treatment of L with a mixture of one equivalent each of Fe^{II} and Mn^{II} sources simultaneously affords Mn^{II}Fe^{II} in 80% yield with 8% Fe^{II}Fe^{II} contamination (Table 2). Our attempts to characterize a monometallic state

Table 2. ¹H NMR Yields of Mn^{II}Fe^{II} as a Function of the Ratio of Fe^{II}:Mn^{II}:L

Fe ^{II} :Mn ^{II} :L/Apo-protein	% Fe ^{II} Fe ^{II}	% Mn ^{II} Fe ^{II}	ref
1:1:1 L	8.0%	80.0%	this work
1.25:1.25:1 L	9.0%	75.0%	this work
1.50:1.50:1 L	20.0%	72.0%	this work
1.75:1.75:1 L	41.0%	49.0%	this work
2:2:1 L	61.0%	0%	this work
2:2:1 R2lox	61%	21%	ref 20

(L:M = 1:1) en route to Mn^{II}Fe^{II} were unsuccessful. Addition of one equivalent of M^{II} to L in the presence of acetate leads to the formation of the bimetallic complex as the major product (see the SI, Figures S34 and S35). When acetate is not present in the reaction mixture, the combination of M^{II} and L in a 1:1 ratio results in an intractable mixture of products.

We further investigated the influence of “metal availability” on heterobimetallic selectivity. The ligand L was subjected to a series of reactions with increasing amounts of equimolar Fe and Mn sources. In all reactions, Fe^{II}(OTf)₂MeCN₂ and Mn^{II}(OTf)₂MeCN₂ were premixed before being added to one equivalent of L. The reaction was allowed to stir for several minutes before acetate was added in the form of TBAOAc. The resulting solution was then analyzed by ¹H NMR spectroscopy (Table 2).

Strikingly, as the metal availability increased from one equivalent to two equivalents of Fe^{II}/Mn^{II} per L, the yield of Mn^{II}Fe^{II} decreases, and the yield of Fe^{II}Fe^{II} increases. When two equivalents of Fe^{II} and Mn^{II} ions are available, no Mn^{II}Fe^{II} formation is observed, implying that the formation of the heterobimetallic species is only favored under Fe^{II}-deficient conditions. With two equivalents of Fe^{II}/Mn^{II} per L, the binding of Fe^{II} outcompetes that of Mn^{II} completely, which is in agreement with the higher binding affinity of Fe^{II} over Mn^{II}, as predicted by the classical Irving–Williams series.³⁰ The availability of Mn^{II} vs Fe^{II} in biological environments has been invoked as a potential reason for why the heterobimetallic Mn/Fe cofactors form.^{3–5,11}

While there is not a complete reversal in selectivity of the RNR1c and R2lox cofactors under Fe-rich conditions, higher Fe^{II} concentrations have also been found to inhibit heterobimetallic cofactor assembly.²⁰ Thus, the increased yield of Fe^{II}Fe^{II} in our system under Fe-rich conditions is similar to that observed for R2lox and RNR1c.^{18–20} The stronger binding affinity of Fe^{II} versus Mn^{II} is further supported by a computational study.¹⁹ The relative stabilities of each M^{II}M^{II} complex (M = Fe or Mn) were determined by comparing the energy sum of the bimetallic complex with different combinations of solvated metal ions (Table 3).¹⁹ Broken-symmetry density functional theory (DFT) calculations at the B3LYP/def2-TZVP level of theory show that the formation of Fe^{II}Fe^{II} is more favorable than Mn^{II}Fe^{II} by 4.6 kcal/mol. The Fe^{II}Mn^{II} complex with a reversed metal occupancy and the theoretical Mn^{II}Mn^{II} species are less stable than Mn^{II}Fe^{II} by 2.5 and 5.6 kcal/mol, respectively. These results follow the same trend as an analogous study performed on the R2lox cofactor.¹⁹

Table 1. ¹H NMR Yields of Fe^{II}Fe^{II} and Mn^{II}Fe^{II} under Various Metalation Conditions

order of addition of metal salts	yield of Fe ^{II} Fe ^{II}	yield of Mn ^{II} Fe ^{II}
(1) Fe ^{II} (OTf) ₂ MeCN ₂ (2) Mn ^{II} (OAc) ₂	6.6%	77.0%
(1) Mn ^{II} (OTf) ₂ MeCN ₂ (2) Fe ^{II} (OAc) ₂	5.8%	75.4%
(1) Fe ^{II} (OAc) ₂ (2) Mn ^{II} (OTf) ₂ MeCN ₂	5.8%	73.8%
(1) Mn ^{II} (OAc) ₂ (2) Fe ^{II} (OTf) ₂ MeCN ₂	6.6%	77.0%

Table 3. DFT-Computed Relative Energies (B3LYP/def2-TZVP) for the Binding of Different Combinations of Fe^{II} and Mn^{II} to L

bimetallic complex	solvated ions	relative energy (kcal/mol)
Fe ^{II} Fe ^{II}	Mn ^{II} , Mn ^{II}	0.0
Mn ^{II} Fe ^{II} (Fe at distorted site)	Mn ^{II} , Fe ^{II}	+4.6
Fe ^{II} Mn ^{II} (Mn at distorted site)	Fe ^{II} , Mn ^{II}	+7.1
Mn ^{II} Mn ^{II}	Fe ^{II} , Fe ^{II}	+10.2

Anomalous XRD Study. In addition to the bioavailability of Fe^{II}, the inequivalent metal binding sites in R2lox (distorted versus octahedral site) have also been regarded as a potential influence on the selective formation of the heterobimetallic cofactor. As Mn^{II}Fe^{II} contains structurally analogous differentiated metal binding sites and exhibits similar selective assembly to R2lox and RNR1c, we wanted to further understand site selectivity in Mn^{II}Fe^{II} via single-crystal anomalous XRD.

Anomalous XRD allows for a quantitative determination of the amount of metal present at a particular crystallographic site.^{38,60,61} Unlike traditional XRD, metal centers with similar numbers of electrons can be differentiated by exploiting the differences in their K-edge absorption energies.⁶² At a synchrotron source, we collected several anomalous data sets at 50 eV above and below the Fe^{II} and Mn^{II} absorption edges. Additionally, a data set was collected at 30 keV, far from the absorption edges of any atoms. As the incident X-ray wavelength approaches the K-edge energy of a metal, the anomalous scattering terms (f' , f'') of the atomic scattering factor (f , where $f(\lambda) = f^0 + f' + if''$) become more pronounced (Figure 5A). Under these conditions, f' and f'' can be utilized to quantify the amount of metal present at each site (Figure 5B and Table 4).^{38,60,61} Gratifyingly, we observed primary occupation of Fe^{II} at the distorted site (82.0%) and primary occupation of Mn^{II} at the octahedral site (73.0%).

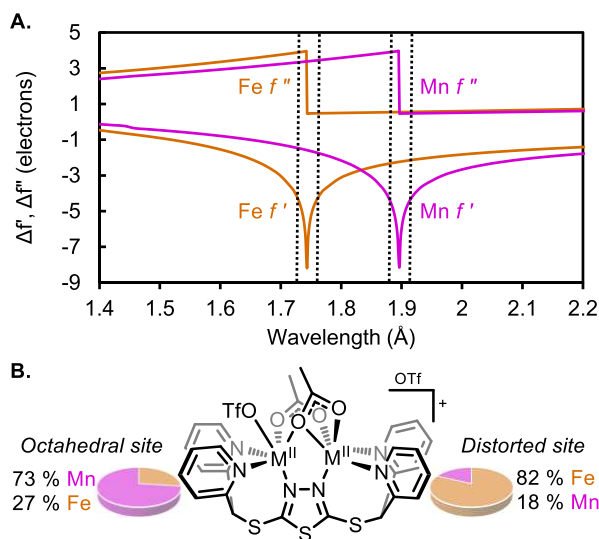


Figure 5. (A) Theoretical anomalous dispersion corrections with the real ($\Delta f'$) and imaginary ($\Delta f''$) scattering factors for Fe (orange) and Mn (pink) as functions of wavelength. Anomalous data were collected at the wavelengths highlighted with dashed lines. (B) Metal occupancies determined based on experimental f' values of four sets of anomalous XRD data.

Table 4. Results of Single-Crystal Anomalous XRD Studies for Mn^{II}Fe^{II}, R2lox, and RNR1c

	distorted site		octahedral site		ref
	% Fe	% Mn	% Fe	% Mn	
Mn ^{II} Fe ^{II} ^a	82%	18%	27%	73%	this work
RNR1c ^b	100%	0%	0%	100%	ref 6
R2lox ^c	80%	21%	48%	53%	ref 19
R2lox ^d	69%	31%	18%	82%	ref 19
R2lox ^e	92%	8%	20%	80%	ref 20

^aSingle crystals isolated from a solution with 1:1 Fe^{II} and Mn^{II} under an N₂ atmosphere. Average of two data sets. ^bProtein crystals isolated after sequential metal loading in the presence of O₂. ^cApo-R2lox crystals soaked for 1 h in excess, equal concentrations of Mn^{II}/Fe^{II} in the absence of O₂. Average of two data sets. ^dApo-R2lox crystals soaked for 1 h in excess, equal concentrations of Mn^{II}/Fe^{II} in the presence of O₂. Average of two data sets. ^eRecrystallization of R2lox after reconstitution in a 2:1 ratio of Mn^{II}/Fe^{II} in the presence of O₂. Average of two data sets.

Interestingly, the selectivity of the distorted and octahedral sites by Mn^{II}Fe^{II} is quite similar to RNR1c and R2lox (Table 4). Both R2lox and RNR1c display a preference for Fe^{II} coordination at the distorted site and Mn^{II} coordination at the octahedral site (Table 4).^{6,19,20} It was proposed that the facile activation of O₂ at the correctly assembled Mn^{II}Fe^{II} site enriches MnFe cofactors. Exposure to oxygen represents an irreversible step in the maturation process, in which the metals are “locked in place” as the less labile Mn^{III}Fe^{III} cofactors.¹⁶ Our study suggests that selective formation of the MnFe site can also be accomplished in high accuracy at the reduced state (II,II).

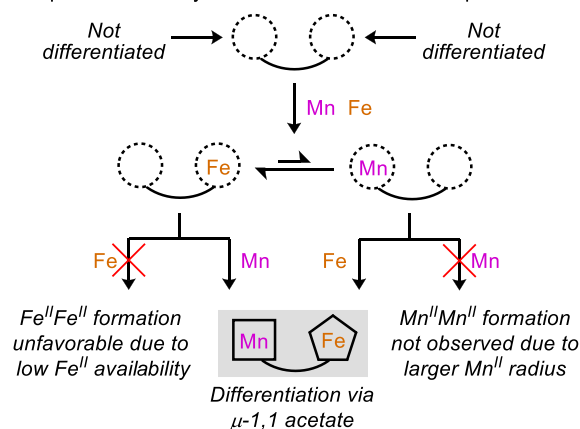
DISCUSSION

The simplicity of our synthetic system allows us to draw several conclusions regarding the mechanism of Mn^{II}Fe^{II} formation (Scheme 3A). We successfully determined that the binding of Fe^{II} is favored as both the first and the second metal, as the yield of Mn^{II}Fe^{II} is reduced to 0% when two equivalents of Fe^{II} are present. Under conditions where only one equivalent of each metal is present per L, the more favorable binding of Fe^{II} to L results in a Mn^{II}-rich environment, which promotes the loading of Mn^{II} as the second metal (Scheme 3A). Since the binding of Fe^{II} outcompetes that of Mn^{II} (in accordance with the Irving–Williams series), the formation of Mn^{II}Fe^{II} must result from the depletion of Fe^{II} in the solution. Fe^{II} depletion can be attributed to two potential reasons: (1) highly selective binding of Fe^{II} as the first metal and (2) the inability of L to coordinate two Mn^{II} ions. Even if the first binding step is not perfectly selective for Fe^{II}, the monometallic LMn^{II} cannot bind another Mn^{II} ion to form a bimetallic Mn^{II}Mn^{II} complex, preventing further consumption of Mn^{II} ions in the solution (Scheme 3A).

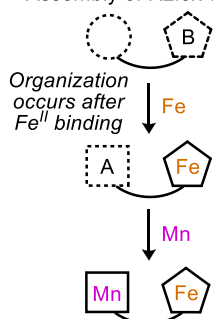
The proposed mechanism of assembly of Mn^{II}Fe^{II} more closely resembles that of R2lox, where the formation of the Mn/Fe cofactor occurs via initial coordination of Fe^{II}, followed by Mn^{II} (Scheme 3B). Rather than loading Mn^{II} and Fe^{II} in a stepwise fashion, apo-RNR1c first loads two Mn^{II} ions and then preferentially substitutes one Mn^{II} for Fe^{II} at the distorted site (Scheme 3C). A difference between our synthetic system and RNR1c and R2lox is that the natural proteins have either one (R2lox) or two (RNR1c) pre-organized metal binding sites, whereas the metal binding sites in our system are only

Scheme 3. Assembly of Synthetic Complex versus Enzyme Cofactors

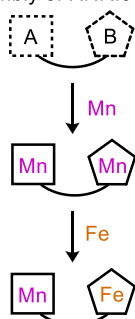
A. Proposed assembly of our heterobimetallic complex



B. Assembly of R2lox cofactor



C. Assembly of RNR1c cofactor



differentiated upon acetate binding. Nonetheless, the end result, following acetate coordination, is the same site-differentiated Mn/Fe species, with Mn^{II} occupying the octahedral site and Fe^{II} occupying the distorted site, suggesting that the unsymmetric carboxylate is an essential structural feature for correct site selectivity (Mn^{II} in octahedral site and Fe^{II} in distorted site). More importantly, however, metal availability (an iron deficiency) is the first key requirement that allows for Mn/Fe cofactor formation. The importance of metal availability is emphasized by the decreased yields of synthetic and enzymatic heterobimetallic species in the presence of excess Fe.

CONCLUSIONS

In summary, we have reported a series of synthetic model complexes, $\text{Fe}^{\text{II}}\text{Fe}^{\text{II}}$, $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$, and Mn^{II}_2 , which have shed light onto the mechanism of heterobimetallic assembly in R2lox and RNR1c. While **L** was not able to accommodate a dimanganese complex in the solid state, single-crystal X-ray analysis of $\text{Fe}^{\text{II}}\text{Fe}^{\text{II}}$ and $\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}$ displayed a bimetallic core containing an unsymmetrically bridged carboxylate ligand. The carboxylate differentiates the metal binding sites and results in a heterobimetallic complex that models the binding of Fe^{II} and Mn^{II} observed in RNR1c and R2lox, i.e., Fe^{II} coordination at the distorted site and Mn^{II} coordination at the octahedral site, which was confirmed via anomalous XRD.

Our model study provides an alternative explanation as to why R2lox and RNR1c appear to be violating the fundamental thermodynamic rules governing coordination chemistry. The favorable binding of Fe^{II} over Mn^{II} along with a site-differentiating μ -1,1-carboxylate was sufficient for correct

heterobimetallic assembly. Despite the overall results of the reaction appearing to be contrary to the binding affinities of Fe^{II} versus Mn^{II} , the Irving–Williams series was not being circumvented. The μ -1,1-carboxylate is a feature shared by both R2lox and RNR1c in the reduced state, as is the observation of favorable diiron cofactor formation under increased concentrations of iron. Therefore, metal availability and site differentiation are two of the most critical factors to correct cofactor assembly in RNR1c and R2lox. This hypothesis is also consistent with the observation that the intracellular availability of metal ions plays a role in metal selection.^{4,5,19} Beyond correct cofactor formation, we believe that site differentiation likely has implications toward oxidative reactivity, as observed in R2lox.¹⁶ These intriguing questions will be the subject of our future studies.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/jacs.2c11930>.

Experimental details, including characterization data, spectra, computational procedures, and results, and crystallographic data for $[\text{Fe}^{\text{II}}_2\text{Py}_4\text{DMcT}(\text{OAc})_2(\text{OTf})][\text{OTf}]$ (CSD: 2218376), $[\text{Mn}^{\text{II}}\text{Fe}^{\text{II}}\text{Py}_4\text{DMcT}(\text{OAc})_2(\text{OTf})][\text{OTf}]$ (CSD: 2218377), and $[\text{Mn}^{\text{II}}_4(\text{Py}_4\text{DMcT})_2(\text{OAc})_4(\text{OTf})_2][\text{OTf}]_2$ (CSD: 2218378) (PDF).

Accession Codes

CCDC 2218376–2218378 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Mitić, N.; Smith, S. J.; Neves, A.; Guddat, L. W.; Gahan, L. R.; Schenk, G. The Catalytic Mechanisms of Binuclear Metallohydrolases. *Chem. Rev.* **2006**, *106*, 3338–3363.
- (2) Nordlund, P.; Reichard, P. Ribonucleotide Reductases. *Annu. Rev. Biochem.* **2006**, *75*, 681–706.
- (3) Cotruvo, J. A., Jr.; Stubbe, J. Class I Ribonucleotide Reductases: Metallocofactor Assembly and Repair In Vitro and In Vivo. *Annu. Rev. Biochem.* **2011**, *80*, 733–767.
- (4) Cotruvo, J. A., Jr.; Stubbe, J. Metallation and Mismetallation of Iron and Manganese Proteins in Vitro and in Vivo: The Class I Ribonucleotide Reductases as a Case Study. *Metallomics* **2012**, *4*, 1020–1036.
- (5) Huang, M.; Parker, M. J.; Stubbe, J. Choosing the Right Metal: Case Studies of Class I Ribonucleotide Reductases. *J. Biol. Chem.* **2014**, *289*, 28104–28111.
- (6) Dassama, L. M. K.; Krebs, C.; Bollinger, J. M., Jr.; Rosenzweig, A. C.; Boal, A. K. Structural Basis for Assembly of the Mn^{IV}/Fe^{III} Cofactor in the Class Ic Ribonucleotide Reductase from Chlamydia Trachomatis. *Biochemistry* **2013**, *52*, 6424–6436.
- (7) Jiang, W.; Yun, D.; Saleh, L.; Barr, E. W.; Xing, G.; Hoffart, L. M.; Maslak, M. A.; Krebs, C.; Bollinger, J. M. A Manganese(IV)/Iron(III) Cofactor in Chlamydia Trachomatis Ribonucleotide Reductase. *Science* **2007**, *316*, 1188–1191.
- (8) Jiang, W.; Hoffart, L. M.; Krebs, C.; Bollinger, J. M. A Manganese(IV)/Iron(IV) Intermediate in Assembly of the Manganese(IV)/Iron(III) Cofactor of Chlamydia Trachomatis Ribonucleotide Reductase[†]. *Biochemistry* **2007**, *46*, 8709–8716.
- (9) Jiang, W.; Yun, D.; Saleh, L.; Bollinger, J. M., Jr.; Krebs, C. Formation and Function of the Manganese(IV)/Iron(III) Cofactor in Chlamydia Trachomatis Ribonucleotide Reductase. *Biochemistry* **2008**, *47*, 13736–13744.
- (10) Högbom, M. The Manganese/Iron-Carboxylate Proteins: What Is What, Where Are They, and What Can the Sequences Tell Us? *J. Biol. Inorg. Chem.* **2010**, *15*, 339–349.
- (11) Högbom, M. Metal Use in Ribonucleotide Reductase R2, Di-Iron, Di-Manganese and Heterodinuclear—an Intricate Bioinorganic Workaround to Use Different Metals for the Same Reaction. *Metallomics* **2011**, *3*, 110–120.
- (12) Griese, J. J.; Kositzki, R.; Haumann, M.; Högbom, M. Assembly of a Heterodinuclear Mn/Fe Cofactor Is Coupled to Tyrosine–Valine Ether Cross-Link Formation in the R2-like Ligand-Binding Oxidase. *J. Biol. Inorg. Chem.* **2019**, *24*, 211–221.
- (13) Diamanti, R.; Srinivas, V.; Johansson, A. I.; Nordström, A.; Griese, J. J.; Lebrette, H.; Högbom, M. Comparative Structural Analysis Provides New Insights into the Function of R2-like Ligand-Binding Oxidase. *FEBS Lett.* **2022**, *596*, 1600–1610.
- (14) Miller, E. K.; Trivelas, N. E.; Maugeri, P. T.; Blaesi, E. J.; Shafaat, H. S. Time-Resolved Investigations of Heterobimetallic Cofactor Assembly in R2lox Reveal Distinct Mn/Fe Intermediates. *Biochemistry* **2017**, *56*, 3369–3379.
- (15) Maugeri, P. T.; Griese, J. J.; Branca, R. M.; Miller, E. K.; Smith, Z. R.; Eirich, J.; Högbom, M.; Shafaat, H. S. Driving Protein Conformational Changes with Light: Photoinduced Structural Rearrangement in a Heterobimetallic Oxidase. *J. Am. Chem. Soc.* **2018**, *140*, 1471–1480.
- (16) Kiseropoulos, E. C.; Griese, J. J.; Smith, Z. R.; Branca, R. M. M.; Schneider, C. R.; Högbom, M.; Shafaat, H. S. Key Structural Motifs Balance Metal Binding and Oxidative Reactivity in a Heterobimetallic Mn/Fe Protein. *J. Am. Chem. Soc.* **2020**, *142*, 5338–5354.
- (17) Kiseropoulos, E. C.; Gan, Y. J.; Greer, S. M.; Hazel, J. M.; Shafaat, H. S. Pulsed Multifrequency Electron Paramagnetic Resonance Spectroscopy Reveals Key Branch Points for One- vs Two-Electron Reactivity in Mn/Fe Proteins. *J. Am. Chem. Soc.* **2022**, *144*, 11991–12006.
- (18) Griese, J. J.; Srinivas, V.; Högbom, M. Assembly of Nonheme Mn/Fe Active Sites in Heterodinuclear Metalloproteins. *J. Biol. Inorg. Chem.* **2014**, *19*, 759–774.
- (19) Griese, J. J.; Roos, K.; Cox, N.; Shafaat, H. S.; Branca, R. M. M.; Lehtiö, J.; Gräslund, A.; Lubitz, W.; Siegbahn, P. E. M.; Högbom, M. Direct Observation of Structurally Encoded Metal Discrimination and Ether Bond Formation in a Heterodinuclear Metalloprotein. *Proc. Natl. Acad. Sci. U. S. A.* **2013**, *110*, 17189–17194.
- (20) Kutin, Y.; Srinivas, V.; Fritz, M.; Kositzki, R.; Shafaat, H. S.; Birrell, J.; Bill, E.; Haumann, M.; Lubitz, W.; Högbom, M.; Griese, J. J.; Cox, N. Divergent Assembly Mechanisms of the Manganese/Iron Cofactors in R2lox and R2c Proteins. *J. Inorg. Biochem.* **2016**, *162*, 164–177.
- (21) Högbom, M.; Stenmark, P.; Voevodskaya, N.; McClarty, G.; Gräslund, A.; Nordlund, P. The Radical Site in Chlamydial Ribonucleotide Reductase Defines a New R2 Subclass. *Science* **2004**, *305*, 245–248.
- (22) Andersson, C. S.; Högbom, M. A Mycobacterium Tuberculosis Ligand-Binding Mn/Fe Protein Reveals a New Cofactor in a Remodeled R2-Protein Scaffold. *Proc. Natl. Acad. Sci. U. S. A.* **2009**, *106*, 5633–5638.
- (23) Shafaat, H. S.; Griese, J. J.; Pantazis, D. A.; Roos, K.; Andersson, C. S.; Popović-Bijelić, A.; Gräslund, A.; Siegbahn, P. E. M.; Neese, F.; Lubitz, W.; Högbom, M.; Cox, N. Electronic Structural Flexibility of Heterobimetallic Mn/Fe Cofactors: R2lox and R2c Proteins. *J. Am. Chem. Soc.* **2014**, *136*, 13399–13409.
- (24) Griese, J. J.; Kositzki, R.; Schrapers, P.; Branca, R. M. M.; Nordström, X. A.; Lehtiö, J.; Haumann, M.; Högbom, X. M. Structural Basis for Oxygen Activation at a Heterodinuclear Manganese/Iron Cofactor. *J. Biol. Chem.* **2015**, *290*, 25254–25272.
- (25) Kutin, Y.; Kositzki, R.; Branca, R. M. M.; Srinivas, V.; Lundin, D.; Haumann, M.; Högbom, M.; Cox, N.; Griese, J. J. Chemical Flexibility of Heterobimetallic Mn/Fe Cofactors: R2lox and R2c Proteins. *J. Biol. Chem.* **2019**, *294*, 18372–18386.
- (26) Manley, O. M.; Phan, H. N.; Stewart, A. K.; Mosley, D. A.; Xue, S.; Cha, L.; Bai, H.; Lightfoot, V. C.; Rucker, P. A.; Collins, L.; Williams, T. I.; Chang, W. C.; Guo, Y.; Makris, T. M. Self-Sacrificial Tyrosine Cleavage by an Fe/Mn Oxygenase for the Biosynthesis of Para-Aminobenzoate in Chlamydia Trachomatis. *Proc. Natl. Acad. Sci. U. S. A.* **2022**, *119*, 1–7.
- (27) Waldron, K. J.; Rutherford, J. C.; Ford, D.; Robinson, N. J. Metalloproteins and Metal Sensing. *Nature* **2009**, *460*, 823–830.
- (28) Smethurst, D. G. J. J.; Shcherbik, N. Interchangeable Utilization of Metals: New Perspectives on the Impacts of Metal Ions Employed in Ancient and Extant Biomolecules. *J. Biol. Chem.* **2021**, *297*, 1–23.
- (29) Choi, T. S.; Tezcan, F. A. Overcoming Universal Restrictions on Metal Selectivity by Protein Design. *Nature* **2022**, *603*, 522–527.
- (30) Irving, H.; Williams, R. J. P. The Stability of Transition-Metal Complexes. *J. Chem. Soc.* **1953**, 3192–3210.

- (31) Rosenzweig, A. C.; Frederick, C. A.; Lippard, S. J.; Nordlund, P. Crystal Structure of a Bacterial Non-Haem Iron Hydroxylase That Catalyses the Biological Oxidation of Methane. *Nature* **1993**, *366*, 537–543.
- (32) Rosenzweig, A. C.; Lippard, S. J. Determining the Structure of a Hydroxylase Enzyme That Catalyzes the Conversion of Methane to Methanol in Methanotrophic Bacteria. *Acc. Chem. Res.* **1994**, *27*, 229–236.
- (33) Rosenzweig, A. C.; Nordlund, P.; Takahara, P. M.; Frederick, C. A.; Lippard, S. J. Geometry of the Soluble Methane Monooxygenase Catalytic Diiron Center in Two Oxidation States. *Chem. Biol.* **1995**, *2*, 409–418.
- (34) Whittington, D. A.; Lippard, S. J. Crystal Structures of the Soluble Methane Monooxygenase Hydroxylase from *Methylococcus Capsulatus* (Bath) Demonstrating Geometrical Variability at the Dinuclear Iron Active Site. *J. Am. Chem. Soc.* **2001**, *123*, 827–838.
- (35) Borovik, A. S.; Que, L.; Papaefthymiou, V.; Muenck, E.; Taylor, L. F.; Anderson, O. P. Heterobimetallic Complexes with $(\mu$ -Phenoxo)Bis(μ -Carboxylato) Cores. *J. Am. Chem. Soc.* **1988**, *110*, 1986.
- (36) Bossek, U.; Weyhermüller, T.; Wieghardt, K.; Bonvoisin, J.; Gierd, J. J. Synthesis, e.s.r. Spectrum and Magnetic Properties of a Heterobinuclear Complex Containing the $\{\text{Fe}^{\text{III}}(\mu\text{-O})(\mu\text{-MeCO}_2)_2\text{Mn}^{\text{III}}\}^{2+}$ Core. *J. Chem. Soc., Chem. Commun.* **1989**, 633–636.
- (37) Das, B.; Daver, H.; Singh, A.; Singh, R.; Haukka, M.; Demeshko, S.; Meyer, F.; Lisensky, G.; Jarenmark, M.; Himio, F.; Nordlander, E. A Heterobimetallic $\text{Fe}^{\text{III}}\text{Mn}^{\text{II}}$ Complex of an Unsymmetrical Dinucleating Ligand: A Structural and Functional Model Complex for the Active Site of Purple Acid Phosphatase of Sweet Potato. *Eur. J. Inorg. Chem.* **2014**, *2014*, 2204–2212.
- (38) Tereniak, S. J.; Carlson, R. K.; Clouston, L. J.; Young, V. G.; Bill, E.; Maurice, R.; Chen, Y.-S.; Kim, H. J.; Gagliardi, L.; Lu, C. C. Role of the Metal in the Bonding and Properties of Bimetallic Complexes Involving Manganese, Iron, and Cobalt. *J. Am. Chem. Soc.* **2014**, *136*, 1842–1855.
- (39) Kerber, W. D.; Goheen, J. T.; Perez, K. A.; Siegler, M. A. Enhanced Stability of the $\text{Fe}^{\text{II}}/\text{Mn}^{\text{II}}$ State in a Synthetic Model of Heterobimetallic Cofactor Assembly. *Inorg. Chem.* **2016**, *55*, 848–857.
- (40) Sano, Y.; Lau, N.; Weitz, A. C.; Ziller, J. W.; Hendrich, M. P.; Borovik, A. S. Models for Unsymmetrical Active Sites in Metalloproteins: Structural, Redox, and Magnetic Properties of Bimetallic Complexes with $\text{M}^{\text{II}}(\mu\text{-OH})\text{-Fe}^{\text{III}}$ Cores. *Inorg. Chem.* **2017**, *56*, 14118–14128.
- (41) Crossland, P. M.; Guo, Y.; Que, L., Jr. Spontaneous Formation of an Fe/Mn Diamond Core: Models for the Fe/Mn Sites in Class 1c Ribonucleotide Reductases. *Inorg. Chem.* **2021**, *60*, 8710–8721.
- (42) Buchanan, R. M.; Mashuta, M. S.; Richardson, J. F.; Webb, R. J.; Oberhausen, K. J.; Nanny, M. A.; Hendrickson, D. N. Synthesis, Structure, and Properties of a Novel Heterobimetallic Iron^{III}Manganese^{II} Complex Containing a Septadentate Polyimidazole Ligand. *Inorg. Chem.* **1990**, *29*, 1299–1301.
- (43) Hotzelmann, R.; Wieghardt, K.; Floerke, U.; Haupt, H. J.; Weatherburn, D. C.; Bonvoisin, J.; Blondin, G.; Gierd, J. J. Spin Exchange Coupling in Asymmetric Heterodinuclear Complexes Containing the μ -Oxo-Bis(μ -Acetato)Dimetal Core. *J. Am. Chem. Soc.* **1992**, *114*, 1681–1696.
- (44) Holman, T. R.; Wang, Z.; Hendrich, M. P.; Que, L. Structural and Spectroscopic Properties of Antiferromagnetically Coupled $\text{Fe}^{\text{III}}\text{Mn}^{\text{II}}$ and $\text{Fe}^{\text{II}}\text{Mn}^{\text{II}}$ Complexes. *Inorg. Chem.* **1995**, *34*, 134–139.
- (45) Karsten, P.; Neves, A.; Bortoluzzi, A. J.; Lanznaster, M.; Drago, V. Synthesis, Structure, Properties, and Phosphatase-Like Activity of the First Heterodinuclear $\text{Fe}^{\text{III}}\text{Mn}^{\text{II}}$ Complex with the Unsymmetric Ligand H_2BPBPMP as a Model for the PAP in Sweet Potato. *Inorg. Chem.* **2002**, *41*, 4624–4626.
- (46) Ross, S.; Weyhermüller, T.; Bill, E.; Bothe, E.; Flörke, U.; Wieghardt, K.; Chaudhuri, P. Asymmetric Heterodinuclear $\text{Fe}^{\text{III}}\text{M}^{\text{II}}$ ($\text{M} = \text{Zn}, \text{Cu}, \text{Ni}, \text{Fe}, \text{Mn}$), $\text{Co}^{\text{III}}\text{Fe}^{\text{II}}$ and $\text{Fe}^{\text{II}}\text{Co}^{\text{III}}$ Species: Synthesis, Structure, Redox Behavior, and Magnetism. *Eur. J. Inorg. Chem.* **2004**, *5*, 984–997.
- (47) Jarenmark, M.; Haukka, M.; Demeshko, S.; Tuczak, F.; Zuppiroli, L.; Meyer, F.; Nordlander, E. Synthesis, Characterization, and Reactivity Studies of Heterodinuclear Complexes Modeling Active Sites in Purple Acid Phosphatases. *Inorg. Chem.* **2011**, *50*, 3866–3887.
- (48) Carboni, M.; Clémancey, M.; Molton, F.; Pécaut, J.; Lebrun, C.; Dubois, L.; Blondin, G.; Latour, J.-M. Biologically Relevant Heterodinuclear Iron–Manganese Complexes. *Inorg. Chem.* **2012**, *51*, 10447–10460.
- (49) Sano, Y.; Weitz, A. C.; Ziller, J. W.; Hendrich, M. P.; Borovik, A. S. Unsymmetrical Bimetallic Complexes with $\text{M}^{\text{II}}(\mu\text{-OH})\text{-M}^{\text{III}}$ Cores ($\text{M}^{\text{II}}\text{M}^{\text{III}} = \text{Fe}^{\text{II}}\text{Fe}^{\text{III}}, \text{Mn}^{\text{II}}\text{Fe}^{\text{III}}, \text{Mn}^{\text{II}}\text{Mn}^{\text{III}}$): Structural, Magnetic, and Redox Properties. *Inorg. Chem.* **2013**, *52*, 10229–10231.
- (50) Tao, W.; Bower, J. K.; Moore, C. E.; Zhang, S. Dicopper M-oxo, M-nitrosyl Complex from the Activation of NO or Nitrite at a Dicopper Center. *J. Am. Chem. Soc.* **2019**, *141*, 10159–10164.
- (51) Tuttle, M. R.; Walter, C.; Brackman, E.; Moore, C. E.; Espe, M.; Rasik, C.; Adams, P.; Zhang, S. Redox-Active Zinc Thiolates for Low-Cost Rechargeable Aqueous Zn-Ion Batteries. *Chem. Sci.* **2021**, *12*, 15253–15262.
- (52) LeCloux, D. D.; Barrios, A. M.; Mizoguchi, T. J.; Lippard, S. J. Modeling the Diiron Centers of Non-Heme Iron Enzymes. Preparation of Sterically Hindered Diiron(II) Tetracarboxylate Complexes and Their Reactions with Dioxide. *J. Am. Chem. Soc.* **1998**, *120*, 9001–9014.
- (53) He, C.; Lippard, S. J. Synthesis and Electrochemical Studies of Diiron Complexes of 1,8-Naphthyridine-Based Dinucleating Ligands to Model Features of the Active Sites of Non-Heme Diiron Enzymes. *Inorg. Chem.* **2001**, *40*, 1414–1420.
- (54) Do, L. H.; Lippard, S. J. Toward Functional Carboxylate-Bridged Diiron Protein Mimics: Achieving Structural Stability and Conformational Flexibility Using a Macrocyclic Ligand Framework. *J. Am. Chem. Soc.* **2011**, *133*, 10568–10581.
- (55) Burger, B.; Dechert, S.; Große, C.; Demeshko, S.; Meyer, F. Visualising the Carboxylate Shift at a Bioinspired Diiron(II) Site in the Solid State. *Chem. Commun.* **2011**, *47*, 10428–10430.
- (56) Minier, M. A.; Lippard, S. J. ^{19}F NMR Study of Ligand Dynamics in Carboxylate-Bridged Diiron(II) Complexes Supported by a Macrocyclic Ligand. *Dalton Trans.* **2015**, *44*, 18111–18121.
- (57) Wang, F.; Becker, S.; Minier, M. A.; Loas, A.; Jackson, M. N.; Lippard, S. J. Tuning the Diiron Core Geometry in Carboxylate-Bridged Macrocyclic Model Complexes Affects Their Redox Properties and Supports Oxidation Chemistry. *Inorg. Chem.* **2017**, *56*, 11050–11058.
- (58) Barynin, V. V.; Whittaker, M. M.; Antonyuk, S. V.; Lamzin, V. S.; Harrison, P. M.; Artymiuk, P. J.; Whittaker, J. W. Crystal Structure of Manganese Catalase from *Lactobacillus Plantarum*. *Structure* **2001**, *9*, 725–738.
- (59) Bertini, I.; Turano, P.; Vila, A. J. Nuclear Magnetic Resonance of Paramagnetic Metalloproteins. *Chem. Rev.* **2002**, *93*, 2833–2932.
- (60) Freedman, D. E.; Han, T. H.; Prodi, A.; Müller, P.; Huang, Q. Z.; Chen, Y. S.; Webb, S. M.; Lee, Y. S.; McQueen, T. M.; Nocera, D. G. Site Specific X-Ray Anomalous Dispersion of the Geometrically Frustrated Kagomé Magnet, Herbertsmithite, $\text{ZnCu}_3(\text{OH})_6\text{Cl}_2$. *J. Am. Chem. Soc.* **2010**, *132*, 16185–16190.
- (61) Powers, T. M.; Gu, N. X.; Fout, A. R.; Baldwin, A. M.; Hernández Sánchez, R.; Alfonso, D. M.; Chen, Y. S.; Zheng, S. L.; Betley, T. A. Synthesis of Open-Shell, Bimetallic Mn/Fe Trinuclear Clusters. *J. Am. Chem. Soc.* **2013**, *135*, 14448–14458.
- (62) Garino, C.; Borfecchia, E.; Gobetto, R.; van Bokhoven, J. A.; Lamberti, C. Determination of the Electronic and Structural Configuration of Coordination Compounds by Synchrotron-Radiation Techniques. *Coord. Chem. Rev.* **2014**, *277–278*, 130–186.