

Research



Cite this article: Barredo E, Raji JI, Ramon M, DeGennaro M, Theobald J. 2022 Carbon dioxide and blood-feeding shift visual cue tracking during navigation in *Aedes aegypti* mosquitoes. *Biol. Lett.* **18**: 20220270.
<https://doi.org/10.1098/rsbl.2022.0270>

Received: 13 June 2022
Accepted: 5 September 2022

Subject Areas:
behaviour, neuroscience

Keywords:
insect vision, optomotor responses, *Aedes aegypti*, insect flight

Authors for correspondence:

Matthew DeGennaro
e-mail: mdegenna@fiu.edu
Jamie Theobald
e-mail: theobald@fiu.edu

[†]These authors contributed equally.
[‡]These corresponding authors contributed equally.

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.6189616>.

Animal behaviour

Carbon dioxide and blood-feeding shift visual cue tracking during navigation in *Aedes aegypti* mosquitoes

Elina Barredo^{1,2,†}, Joshua I. Raji^{1,2,†}, Michael Ramon^{1,2}, Matthew DeGennaro^{1,2,‡} and Jamie Theobald^{1,‡}

¹Department of Biological Sciences, and ²Biomolecular Sciences Institute, Florida International University, Miami, FL 33199, USA

ID EB, 0000-0001-7515-3878; JIR, 0000-0002-6447-6986; MD, 0000-0003-3837-4430; JT, 0000-0001-7648-3772

Haematophagous mosquitoes need a blood meal to complete their reproductive cycle. To accomplish this, female mosquitoes seek vertebrate hosts, land on them and bite. As their eggs mature, they shift attention away from hosts and towards finding sites to lay eggs. We asked whether females were more tuned to visual cues when a host-related signal, carbon dioxide, was present, and further examined the effect of a blood meal, which shifts behaviour to ovipositing. Using a custom, tethered-flight arena that records wing stroke changes while displaying visual cues, we found the presence of carbon dioxide enhances visual attention towards discrete stimuli and improves contrast sensitivity for host-seeking *Aedes aegypti* mosquitoes. Conversely, intake of a blood meal reverses vertical bar tracking, a stimulus that non-fed females readily follow. This switch in behaviour suggests that having a blood meal modulates visual attention in mosquitoes, a phenomenon that has been described before in olfaction but not in visually driven behaviours.

1. Introduction

Anthropophilic mosquitoes are dangerous vectors of disease—taking a toll of over 700 000 human lives a year [1]. Mosquito control suffers from shortcomings due to insecticide resistance [2], inefficient repellents [3,4] and the lack of economical [5] and efficacious adult traps [6]. Adults emerging from aquatic stages cycle through distinct behavioural phases: nectar foraging, mating, host-seeking, resting and oviposition. To perform these tasks, they integrate visual, auditory and chemical stimuli, then make behavioural decisions according to the needs of each stage [7]. Newly emerged mosquitoes follow nectar cues for basic nutrient acquisition [8,9], which include plant semiochemicals and floral shapes and colours [9]. As they age and become reproductively mature [10,11], females increase their attraction towards human scent, relying on carbon dioxide (CO₂) emanations, body odour, heat and likely contact cues found on the skin [7,12]. Once gravid, they are drawn towards standing water, where they lay eggs [13].

To extract relevant information from complex visual environments, animals can shift visual attention [14]. This is crucial for airborne animals, who regulate flight in response to rapidly changing scenes [15]. They may follow, avoid or ignore features depending on external factors, such as heat or odour, internal factors, such as hunger, and reproductive phase [15–17]. Visual needs vary with light habitat and activity [18], but even within a species animals may attend to varying elements of their visual landscape [16,19]. Despite the seeming importance of visual attention in host-seeking mosquitoes, there is

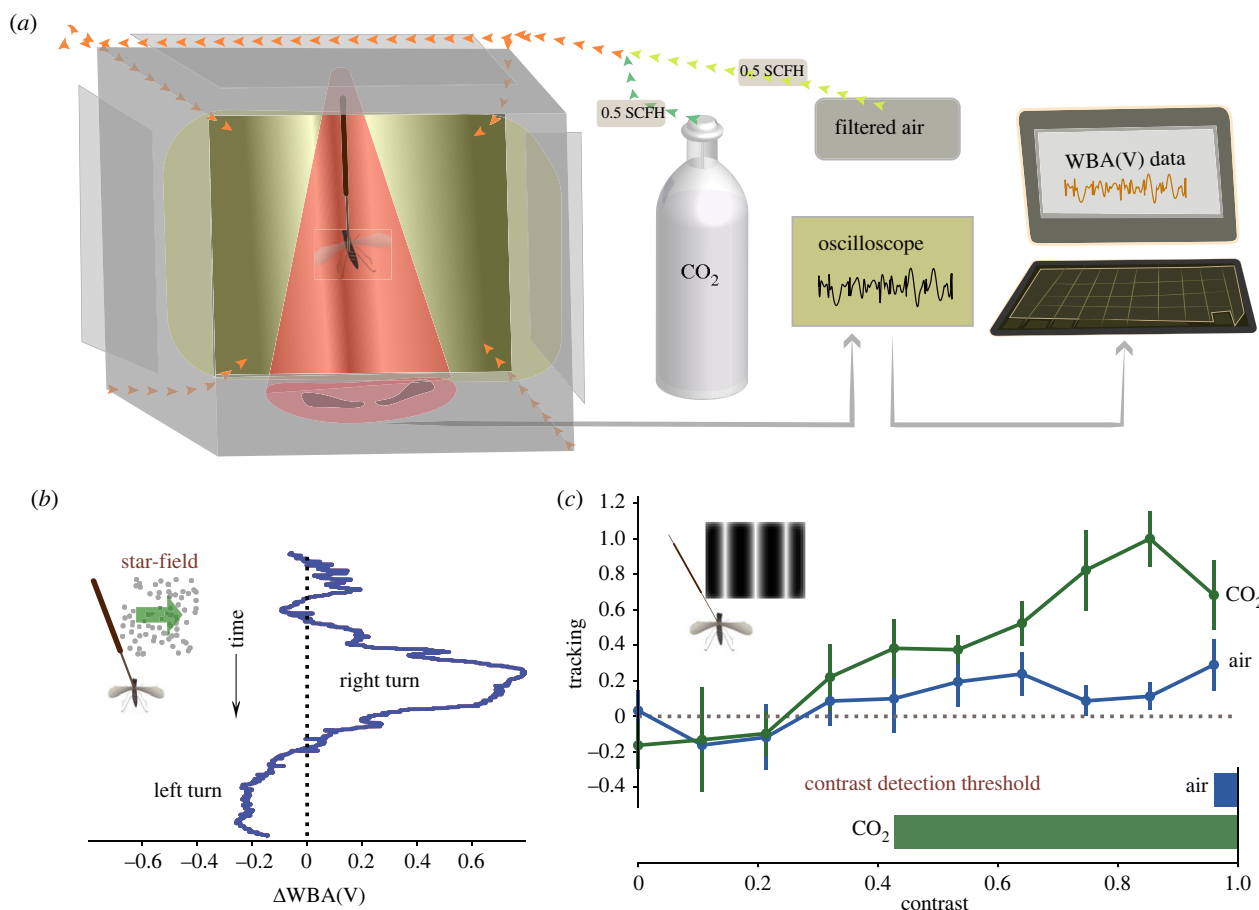


Figure 1. Flight arena system. (a) Flight arena (left) with a flying mosquito tethered under an infrared beam. CO₂ was delivered through each corner of the front side of the cube (yellow arrows). An oscilloscope (right) recorded insect WBA as voltage (V) and frequency in hertz (Hz) used for analysis. (b) Representative trace of a mosquito responding to visual simulations. ΔWBA is calculated by subtracting the right WBA from the left. Negative and positive ΔWBA values suggest left and right steering attempts, respectively. (c) *Ae. aegypti* average tracking responses \pm s.e.m. for each contrast with and without CO₂. The bottom bar graph summarizes the minimum contrast values for which mosquito tracking responses (blue = air, $n = 9$; green = CO₂, $n = 16$) deviated from 0 (p -values < 0.05).

limited knowledge about the relationship between their reproductive cycle stage and how they process information from their visual environments.

CO₂ is a general activator of mosquito host-seeking behaviour, but there is contradicting evidence as to whether it specifically triggers visual attention [20,21] or not [22] in host-seeking *Ae. aegypti*. To address this matter, we delivered CO₂ plumes to rigidly tethered flying mosquitoes and tested its effects on visual tracking. Next, we compared visual cue tracking between host-seeking and gravid female mosquitoes.

2. Methods

(a) Insect rearing and preparation

We reared Orlando strain *Ae. aegypti* and maintained them at 25–28°C, 75% relative humidity under a 14:10 light–dark cycle. Eggs were hatched in hatching broth –1 l of deionized, deoxygenated water and one pellet of Tetramin fish food (catalogue no. 16152, Tetra, Melle, Germany). We sorted first-instar larvae (approx. 230 per 2 l of water) and fed them two pellets of Tetramin. Controlled larval density and food ratio guaranteed even mosquito sizes. Adult mosquitoes were fed *ad libitum* on 10% sucrose until experimentation, 6–8 days post-eclosion.

Female subjects were cold anaesthetized for a maximum of 3 min, then glued by the dorsal thorax to a tungsten rod. They rested holding a piece of tissue paper on their legs, which stops spontaneous wing beating, for 10–30 min. We suspended tethered subjects at the centre of the flight arena and removed

the paper to initiate flight. Each insect was tested only once, or twice if the first trace had poor quality. We blood-fed female mosquitoes using an artificial feeder consisting of a glass tube warmed by 37°C running water. Stretched parafilm, scented by vigorous rubbing against the experimenter's arm, secured the blood (sheep blood, Fisher Scientific R54020) to the glass tube and allowed feeding through the membrane. Engorged females were separated into a cage 72 h before experiments. A custom feeder with cut transfer pipettes on the ceiling allowed capillary delivery of 10% sucrose and prevented females from laying eggs, ensuring they stayed gravid. Non-gravid females for these experiments were housed similarly.

As a reference model for our CO₂-flight arena system, we also ran the CO₂ experiments using the model insect, *Drosophila melanogaster* (electronic supplementary material, figure S1). Fruit fly subjects were female *D. melanogaster* from a laboratory colony, raised on a standard food medium, under a 12L:12D cycle at 21°C and collected 4–6 days post-eclosion as done previously [19].

(b) Tethered-flight arena and steering responses

Our custom-built flight arena can deliver simultaneous visual and olfactory stimulation to rigidly tethered insects. Back projection screen material covers five sides of a 200 mm cube, with the back open for access, and first-surface mirrors, affixed at 45° to the sides, allow a front projector to illuminate the five faces simultaneously [23]. This covers 10.47 steradians of the visual field to a subject in the centre and displays a perspective-corrected three-dimensional scene at 360 Hz (figure 1a).

Within the arena, an infrared beam above the insect casts wing shadows onto two photodiodes below (figure 1a), which

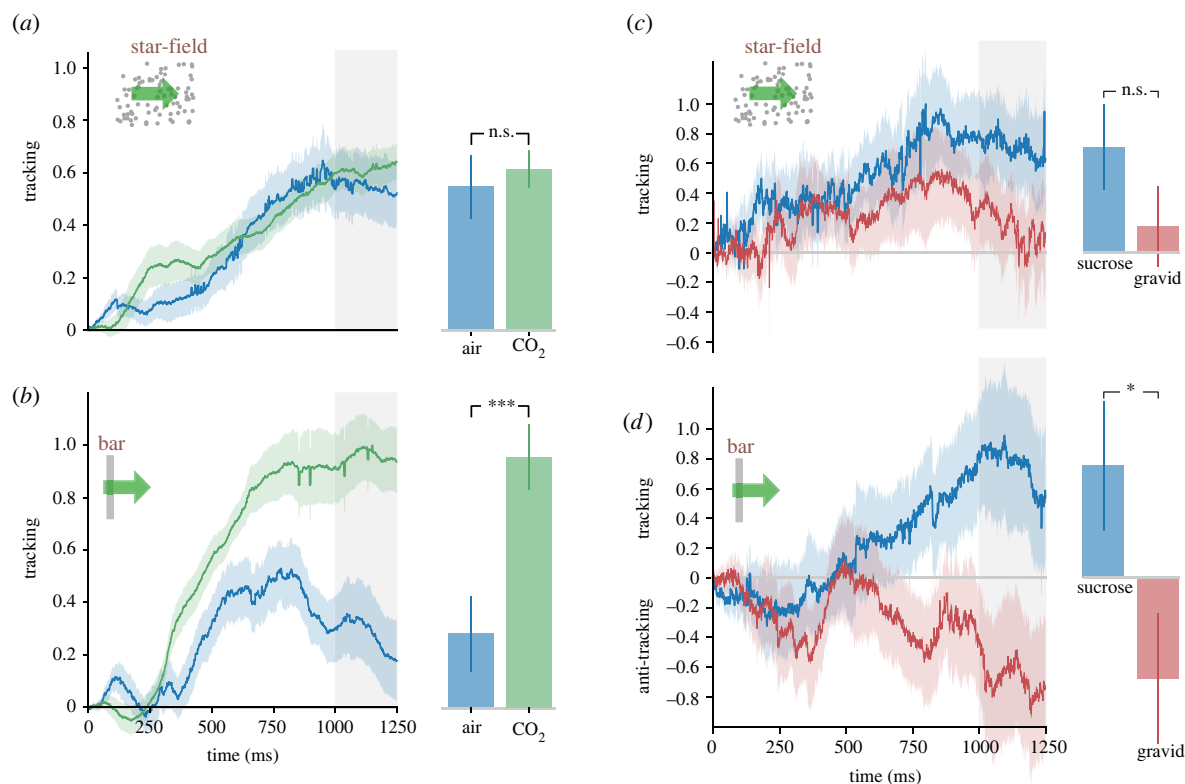


Figure 2. Visual attention is enhanced in the presence of CO_2 and depends on mosquito feeding status. Normalized responses of *Ae. aegypti* female mosquitoes ($n = 19$, green = CO_2 ; $n = 17$, blue = air) to a star-field (a) and a high-contrast bar (b) For the CO_2 trials, the concentration was kept within 2200–2800 ppm. Normalized responses of gravid, blood-fed ($n = 21$, red) and non-gravid, sucrose-fed ($n = 24$, blue) *Ae. aegypti* mosquitoes tracking a star-field (c) and bar (d) in the absence of CO_2 or air plumes. Positive ΔWBA values (tracking) are wing stroke deviations where mosquitoes attempt to turn towards the stimulus, while negative ΔWBA values represent turns in the opposite direction as the moving cue. In all line plots, the solid line represents the average response over time in milliseconds, and the shadow surrounding the line represents the s.e.m. for that treatment. Grey-shaded regions denote the time-frame compared for significance (1000–1250 ms) summarized in the bar plots \pm s.e.m. Asterisks denote a significant difference in ΔWBA between the two treatment groups (one-tailed t -test).

generates a voltage signal with every wing stroke. The relative amplitudes of right-wing and left-wing beats (figure 1b) indicate steering effort [24]. The average ΔWBA is calculated by subtracting the right WBA from the left for each experimental trial, divided by the total number of trials per group. Tracking values are deviations in the mean ΔWBA relative to the direction of the stimulus motion. For instance, an insect steering away from a stimulus receives a negative ΔWBA , which we call ‘anti-tracking’. For each experiment in every trial, we averaged ΔWBA between 1000 and 1250 ms (the shaded regions in figures below), which gave responses time to stabilize. One-tailed t -tests were conducted to compare the mean ΔWBA between groups of interest as we expected tracking to be enhanced for one of the treatments only.

(c) Visual and olfactory stimuli

For tracking stimuli, we used star-fields of small points (113 dots sr^{-1}) and frontal moving vertical bars ($11 \text{ by } 100^\circ$), both in open loop. For contrast sensitivity, we used frontal grating, ($0.04 \text{ cycles per degree at } 10 \text{ Hz}$) at 10 different contrasts, from 0 to 0.95 (Michelson contrast). Each mosquito saw each tracking stimulus one time for 1250 ms, in random order, in both left and right directions, interspersed with 3 s bouts of closed-loop bar fixation. Insects that failed to hold a stable bar between experiments, or failed to beat their wings throughout an experiment, were eliminated from further analysis.

Clear tubings affixed to each corner of the front screen provided CO_2 to the arena. An airflow meter (VFA-4-SSV Dwyer Instruments Inc., IN, USA) controlled the delivery rate. We found flight was stable when CO_2 and air were each set at 0.5 standard cubic feet per hour (SCFH), keeping the CO_2 at 2200–2800 ppm and a working value from previous studies [25]. We monitored CO_2 with a meter (catalogue no. CO2-100, Amprobe)

placed at the back of the arena, allowing CO_2 to be delivered continuously throughout the experiment. Control experiments were done with filtered air set at 1.0 SCFH to maintain the same airflow in all the assays.

3. Results

(a) CO_2 greatly enhances mosquito contrast sensitivity

We assessed *Ae. aegypti* responses to visual stimuli in the presence and absence of CO_2 . Without CO_2 , mosquitoes tracked only weakly at all contrasts less than 0.95. When CO_2 was added, their contrast sensitivity increased dramatically. This difference was significant for all contrast gratings of value 0.43 or greater (air $n = 9$, CO_2 $n = 16$, figure 1c). To confirm our assay was robust, we tested *Drosophila* as well and found that CO_2 only enhanced fly contrast sensitivity at lower concentrations, in line with previous studies [26] (electronic supplementary material, figure S1A).

(b) CO_2 affects mosquito bar, but not star-field visual tracking

During star-field optical flow, CO_2 elicited little effect on mosquito tracking (air $n = 17$, CO_2 $n = 19$, $p = 0.32$, figure 2a). However, mosquitoes tracked a bar much more strongly with added CO_2 (air $n = 17$, CO_2 $n = 19$, $p < 0.001$, figure 2b). When CO_2 was delivered to *Drosophila* at the same rate as we used for mosquitoes (2200–2800 ppm), it substantially reduced fruit fly tracking during star-field optical flow as

well as bar tracking, nearly disrupting *Drosophila* flight completely (air $n=11$, CO₂ $n=15$, $p<0.001$, electronic supplementary material, figure S1B,C).

(c) The blood-fed group of mosquitoes reversed rigidly tethered bar tracking

Finally, to assess the dramatic state change that occurs in the days following a blood meal, we examined the tracking of sucrose-fed female mosquitoes, with identically aged females that had ingested a blood meal 72 h before. For star-field flow, the blood treatment resulted in a visibly reduced tracking, but that was not statistically significant at our sample size (sucrose $n=24$, blood $n=21$, $p=0.098$, figure 2c). For moving bars, however, sucrose and blood-fed mosquitoes displayed opposite responses, with the sucrose group tracking and the blood group anti-tracking—moving opposite to the bar motion, an effect we had not seen in other experiments, (sucrose $n=24$, blood $n=21$, $p=0.015$, figure 2d).

4. Discussion

Drosophila melanogaster has been studied under restricted flight conditions for decades [27], and we took advantage of this assay to explore the effects of CO₂ in mosquito visual attention. Here we used contrast as a general metric of stimulus strength required for steering, star-fields as a measure for course correction and bars as a measure of target fixation, a cue that may resemble a host. Our findings align with others showing CO₂ guides mosquitoes by enhancing object salience. When vision is impaired, as in *Ae. aegypti* *op1*, *op2* double mutants, mosquitoes lose tracking behaviour towards black spots with CO₂ plumes [28]. CO₂ is a food cue for vinegar flies as well [29] and reduced the contrast at which they followed a grating (electronic supplementary material, figure S1A). Nonetheless, the valence of fruit fly responses to CO₂ varies depending on context (such as feeding state or CO₂ concentration) [30], modulating the neural processing pathways that determine attractive and aversive behaviours [31].

The CO₂ signals emitted by a vertebrate's breath reach mosquitoes at intermittent and variable doses, which triggers upwind flight to search for potential hosts [20,29,32]. Because tethered mosquitoes are flying when tested, we cannot credit CO₂ as a general flight activator. But by enhancing bar tracking and not star-field tracking, it appears to depend on context, similar to how it enhances *Ae. aegypti* attraction for specific colours and not others [33]. We also explored if CO₂ affected contrast sensitivity, the ability to discriminate an object from its background, a critical visual property, especially for crepuscular and nocturnal insects [34]. The eye anatomy of *Ae. aegypti* suggests that they have relatively poor resolution but may adequately detect contrast changes typical of sunset hours [35]. Since they also prefer to land on low-reflectance and high-contrast objects, and are

specifically attracted to visual patterns with high vertical contrast [36], we speculate that CO₂ alerts them to find landing targets. And although another study found no CO₂ enhancement of tracking in tethered flight [22], that study used a free-yaw tether, in which an insect turning might rapidly affect the odour concentration the insect experiences, while the rigid tether assay here is open loop, keeping concentration constant throughout the experiment.

The multi-modal nature of host-seeking has generated extensive research into olfaction and CO₂ sensing [12,25,37,38], but after a satisfactory blood meal, females become refractory to both host volatiles and CO₂ [39]. Such changes in behaviour are in tune with transcriptional regulation of genes in blood-fed *Ae. aegypti* [40] and *Anopheles gambiae* [41]. In addition to olfactory genes, visual genes such as opsins are transcriptionally downregulated following blood-feeding in *An. gambiae* [42]. In behavioural assays, blood-fed *Ae. aegypti* lose interest in their hosts [13,40,43]. The bar avoidance in our blood-fed group could be a defensive strategy to avoid hosts when the priority is ovipositing, which could be further explored in free-flight assays [44]. Testing whether mosquitoes recover their bar tracking ability after laying eggs could provide additional evidence that visual attention depends on the female reproductive stage. This is the first study, to our knowledge, that presents a shift in visual attention for gravid females, and we expect the roles of CO₂ sensing and blood-fed status in the modulation of visual attention to be largely conserved across haematophagous mosquitoes.

Data accessibility. Data are available from the Dryad Digital Repository <https://doi.org/10.5061/dryad.z8w9ghxfv> [45].

The data are provided in the electronic supplementary material [46].

Authors' contributions. E.B.: conceptualization, formal analysis, investigation, visualization, writing—original draft and writing—review and editing; J.I.R.: conceptualization, investigation and writing—review and editing; M.R.: investigation and writing—review and editing; M.D.: conceptualization, funding acquisition, project administration, resources, supervision, validation, visualization and writing—review and editing; J.T.: conceptualization, data curation, formal analysis, funding acquisition, methodology, project administration, resources, software, supervision, validation, visualization and writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. This research was supported by grants from the National Science Foundation: IOS-1750833 to J.T. and the Centers for Disease Control and Prevention (CDC) Southeastern Center of Excellence in Vector Borne Diseases (U01CK000510) to M.D. The content is solely the responsibility of the authors and does not represent the official views of the CDC.

Acknowledgements. We thank Yash Sondhi and André Luis Costa-da-Silva for their helpful insights and comments on the manuscript. We also thank all the members of the DeGennaro and Theobald laboratories at FIU. This is contribution no. 1485 from the Institute of Environment at Florida International University.

References

1. World Health Organization. 2020 *WHO Fact sheets*. <https://www.who.int/news-room/fact-sheets>.
2. Liu N. 2015 Insecticide resistance in mosquitoes: impact, mechanisms, and research directions. *Annu. Rev. Entomol.* **60**, 537–559. (doi:10.1146/annurev-ento-010814-020828)

3. Mappin F, DeGennaro M. 2022 Multimodal mechanisms of repellency in arthropods. In *Advances in arthropod repellents*, pp. 113–130. Amsterdam, The Netherlands: Elsevier.
4. DeGennaro M. 2015 The mysterious multi-modal repellency of DEET. *Fly* **9**, 45–51. (doi:10.1080/19336934.2015.1079360)
5. Hou J *et al.* 2021 Field evaluation of two mosquito traps in Zhejiang Province, China. *Sci. Rep.* **11**, 294. (doi:10.1038/s41598-020-80618-1)
6. Brown HE, Paladini M, Cook RA, Kline D, Barnard D, Fish D. 2008 Effectiveness of mosquito traps in measuring species abundance and composition. *J. Med. Entomol.* **45**, 517–521. (doi:10.1603/0022-2585(2008)45[517:EOMTIM]2.0.CO;2)
7. Raji JI, DeGennaro M. 2017 Genetic analysis of mosquito detection of humans. *Curr. Opin. Insect Sci.* **20**, 34–38. (doi:10.1016/j.cois.2017.03.003)
8. Foster WA. 1995 Mosquito sugar feeding and reproductive energetics. *Annu. Rev. Entomol.* **40**, 443–474. (doi:10.1146/annurev.en.40.010195.002303)
9. Barredo E, DeGennaro M. 2020 Not just from blood: mosquito nutrient acquisition from nectar sources. *Trends Parasitol.* **36**, 473–484. (doi:10.1016/j.pt.2020.02.003)
10. Foster WA, Takken W. 2004 Nectar-related vs. human-related volatiles: behavioural response and choice by female and male *Anopheles gambiae* (Diptera: Culicidae) between emergence and first feeding. *Bull. Entomol. Res.* **94**, 145–157. (doi:10.1079/BER2003288)
11. Tallon AK, Hill SR, Ignell R. 2019 Sex and age modulate antennal chemosensory-related genes linked to the onset of host seeking in the yellow-fever mosquito, *Aedes aegypti*. *Sci. Rep.* **9**, 43. (doi:10.1038/s41598-018-36550-6)
12. McMeniman CJ, Corfas RA, Matthews BJ, Ritchie SA, Vosshall LB. 2014 Multimodal integration of carbon dioxide and other sensory cues drives mosquito attraction to humans. *Cell* **156**, 1060–1071. (doi:10.1016/j.cell.2013.12.044)
13. Klowden MJ. 1990 The endogenous regulation of mosquito reproductive behavior. *Experientia* **46**, 660–670. (doi:10.1007/BF01939928)
14. Carrasco M. 2011 Visual attention: the past 25 years. *Vision Res.* **51**, 1484–1525. (doi:10.1016/j.visres.2011.04.012)
15. Tammero LF, Dickinson MH. 2002 Collision-avoidance and landing responses are mediated by separate pathways in the fruit fly, *Drosophila melanogaster*. *J. Exp. Biol.* **205**, 2785–2798. (doi:10.1242/jeb.205.18.2785)
16. Nityananda V. 2016 Attention-like processes in insects. *Proc. R. Soc. B* **283**, 20161986. (doi:10.1098/rspb.2016.1986)
17. Maimon G, Straw AD, Dickinson MH. 2008 A simple vision-based algorithm for decision making in flying *Drosophila*. *Curr. Biol.* **18**, 464–470. (doi:10.1016/j.cub.2008.02.054)
18. Land MF, Nilsson DE. 2012 *Animal eyes*. Oxford, UK: Oxford University Press.
19. Palermo N, Theobald J. 2019 Fruit flies increase attention to their frontal visual field during fast forward optic flow. *Biol. Lett.* **15**, 20180767. (doi:10.1098/rsbl.2018.0767)
20. van Breugel F, Riffell J, Fairhall A, Dickinson MH. 2015 Mosquitoes use vision to associate odor plumes with thermal targets. *Curr. Biol.* **25**, 2123–2129. (doi:10.1016/j.cub.2015.06.046)
21. Vinauger C, Van Breugel F, Locke LT, Tobin KKS, Dickinson MH, Fairhall AL, Akbari OS, Riffell JA. 2019 Visual–olfactory integration in the human disease vector mosquito *Aedes aegypti*. *Curr. Biol.* **29**, 2509–2516.e5. (doi:10.1016/j.cub.2019.06.043)
22. Liu MZ, Vosshall LB. 2019 General visual and contingent thermal cues interact to elicit attraction in female *Aedes aegypti* mosquitoes. *Curr. Biol.* **29**, 2250–2257.e4. (doi:10.1016/j.cub.2019.06.001)
23. Cabrera S, Theobald JC. 2013 Flying fruit flies correct for visual sideslip depending on relative speed of forward optic flow. *Front. Behav. Neurosci.* **7**, 76. (doi:10.3389/fnbeh.2013.00076)
24. Frye MA, Dickinson MH. 2004 Motor output reflects the linear superposition of visual and olfactory inputs in *Drosophila*. *J. Exp. Biol.* **207**, 123–131. (doi:10.1242/jeb.00725)
25. Raji JI, Melo N, Castillo JS, Gonzalez S, Saldana V, Stensmyr MC, DeGennaro M. 2019 *Aedes aegypti* mosquitoes detect acidic volatiles found in human odor using the IR8a pathway. *Curr. Biol.* **29**, 1253–1262.e7. (doi:10.1016/j.cub.2019.02.045)
26. Wasserman S, Salomon A, Frye MA. 2013 *Drosophila* tracks carbon dioxide in flight. *Curr. Biol.* **23**, 301–306. (doi:10.1016/j.cub.2012.12.038)
27. Götz KG. 1968 Flight control in *Drosophila* by visual perception of motion. *Kybernetik* **4**, 199–208. (doi:10.1007/BF00272517)
28. Zhan Y, Alonso San Alberto D, Rusch C, Riffell JA, & Montell C. 2021 Elimination of vision-guided target attraction in *Aedes aegypti* using CRISPR. *Curr. Biol.* **31**, 4180–4187.e6. (doi:10.1016/j.cub.2021.07.003)
29. Guerenstein PG, Hildebrand JG. 2008 Roles and effects of environmental carbon dioxide in insect life. *Annu. Rev. Entomol.* **53**, 161–178. (doi:10.1146/annurev.ento.53.103106.093402)
30. van Breugel F, Huda A, Dickinson MH. 2018 Distinct activity-gated pathways mediate attraction and aversion to CO₂ in *Drosophila*. *Nature* **564**, 420–424. (doi:10.1038/s41586-018-0732-8)
31. Cheng KY, Colbath RA, Frye MA. 2019 Olfactory and neuromodulatory signals reverse visual object avoidance to approach in *Drosophila*. *Curr. Biol.* **29**, 2058–2065.e2. (doi:10.1016/j.cub.2019.05.010)
32. Dekker T, Geier M, Cardé RT. 2005 Carbon dioxide instantly sensitizes female yellow fever mosquitoes to human skin odours. *J. Exp. Biol.* **208**, 2963–2972. (doi:10.1242/jeb.01736)
33. Alonso San Alberto D, Alberto D, Rusch C, Zhan Y, Straw AD, Montell C, Riffell JA. 2022 The olfactory gating of visual preferences to human skin and visible spectra in mosquitoes. *Nat. Commun.* **13**, 555. (doi:10.1038/s41467-022-28195-x)
34. Warrant E, Dacke M. 2011 Vision and visual navigation in nocturnal insects. *Annu. Rev. Entomol.* **56**, 239–254. (doi:10.1146/annurev-ento-120709-144852)
35. Muir LE, Thorne MJ, Kay BH. 1992 *Aedes aegypti* (Diptera: Culicidae) vision: spectral sensitivity and other perceptual parameters of the female eye. *J. Med. Entomol.* **29**, 278–281. (doi:10.1093/jmedent/29.2.278)
36. Muir LE, Kay BH, Thorne MJ. 1992 *Aedes aegypti* (Diptera: Culicidae) vision: response to stimuli from the optical environment. *J. Med. Entomol.* **29**, 445–450. (doi:10.1093/jmedent/29.3.445)
37. Cardé RT. 2015 Multi-cue integration: how female mosquitoes locate a human host. *Curr. Biol.* **25**, R793–R795. (doi:10.1016/j.cub.2015.07.057)
38. DeGennaro M, McBride CS, Seeholzer L, Nakagawa T, Dennis EJ, Goldman C, Jasinskiene N, James AA, Vosshall LB. 2013 Orco mutant mosquitoes lose strong preference for humans and are not repelled by volatile DEET. *Nature* **498**, 487–491. (doi:10.1038/nature12206)
39. Klowden MJ. 1981 Initiation and termination of host-seeking inhibition in *Aedes aegypti* during oöcyte maturation. *J. Insect. Physiol.* **27**, 799–803. (doi:10.1016/0022-1910(81)90071-8)
40. Matthews BJ, McBride CS, DeGennaro M, Despo O, Vosshall LB. 2016 The neurotranscriptome of the *Aedes aegypti* mosquito. *BMC Genomics* **17**, 32. (doi:10.1186/s12864-015-2239-0)
41. Rinker DC, Pitts RJ, Zhou X, Suh E, Rokas A, Zwiebel LJ. 2013 Blood meal-induced changes to antennal transcriptome profiles reveal shifts in odor sensitivities in *Anopheles gambiae*. *Proc. Natl Acad. Sci. USA* **110**, 8260–8265. (doi:10.1073/pnas.1302562110)
42. Ribeiro JMC. 2003 A catalogue of *Anopheles gambiae* transcripts significantly more or less expressed following a blood meal. *Insect. Biochem. Mol. Biol.* **33**, 865–882. (doi:10.1016/S0965-1748(03)00080-8)
43. Liesch J, Bellani LL, Vosshall LB. 2013 Functional and genetic characterization of neuropeptide Y-like receptors in *Aedes aegypti*. *PLoS Negl. Trop. Dis.* **7**, e2486. (doi:10.1371/journal.pntd.0002486)
44. Cribellier A, Straw AD, Spitzner J, Pieters RPM, Van Leeuwen JL, Muijres FT. 2022 Diurnal and nocturnal mosquitoes escape looming threats using distinct flight strategies. *Curr. Biol.* **32**, 1232–1246.e5. (doi:10.1016/j.cub.2022.01.036)
45. Barredo E, Raji JI, Ramon M, DeGennaro M, Theobald J. 2022 Data from: Carbon dioxide and blood-feeding shift visual cue tracking during navigation in *Aedes aegypti* mosquitoes. Dryad Digital Repository. (doi:10.5061/dryad.z8w9ghxfv)
46. Barredo E, Raji JI, Ramon M, DeGennaro M, Theobald J. 2022 Carbon dioxide and blood-feeding shift visual cue tracking during navigation in *Aedes aegypti* mosquitoes. Figshare. (doi:10.6084/m9.figshare.c.6189616)