

1 **Reduction in animal abundance and oxygen availability during and after the**
2 **end-Triassic mass extinction**

3

4 **Abstract**

5 The end-Triassic biodiversity crisis was one of the most severe mass extinctions in the history of
6 animal life. However, the extent to which the loss of taxonomic diversity was coupled with a
7 reduction in organismal abundance remains to be quantified. Further, the temporal relationship
8 between organismal abundance and local marine redox conditions is lacking in carbonate
9 sections. To address these questions, we measured skeletal grain abundance in shallow-marine
10 limestones by point counting 293 thin sections from four stratigraphic sections across the
11 Triassic/Jurassic boundary in the Lombardy Basin and Apennine Platform of western Tethys.
12 Skeletal abundance decreased abruptly across the Triassic/Jurassic boundary in all stratigraphic
13 sections. The abundance of skeletal organisms remained low throughout the lower-middle
14 Hettangian strata and began to rebound during the late Hettangian and early Sinemurian. A two-
15 way ANOVA indicates that sample age ($p<0.01$, $\eta^2=0.30$) explains more of the variation in skeletal
16 abundance than the depositional environment or paleobathymetry ($p<0.01$, $\eta^2=0.15$). Measured
17 I/Ca ratios, a proxy for local shallow-marine redox conditions, show this same pattern with the
18 lowest I/Ca ratios occurring in the early Hettangian. The close correspondence between oceanic
19 water column oxygen levels and skeletal abundance indicates a connection between redox
20 conditions and benthic organismal abundance across the Triassic/Jurassic boundary. These
21 findings indicate that the end-Triassic mass extinction reduced not only the biodiversity but also
22 the carrying capacity for skeletal organisms in early Hettangian ecosystems, adding to evidence
23 that mass extinction of species generally leads to mass rarity among survivors.

24

25 **1 Introduction**

26 Predicting the responses of marine ecosystems to extreme environmental change is a task of
27 immense importance for the 21st century, as such predictions are needed to guide effective
28 conservation strategies. The major mass extinctions of the geological past provide our best
29 empirical constraints on such predictions (e.g., Harnik et al., 2012; Blois et al., 2013). These
30 biodiversity crises significantly altered the taxonomic composition and ecological structure of
31 marine ecosystems in response to environmental change (e.g., Droser et al., 2000; Bambach et al.,
32 2002; Wagner et al., 2006; Hull and Darroch, 2013).

33 Mass extinctions have traditionally been recognized by rapid reductions in taxonomic diversity at
34 regional to global scales (e.g., Raup and Sepkoski, 1982; Bambach, 2006). Taxonomic diversity,
35 however, only partially reflects the evolutionary and ecological effects of mass extinctions. Mass
36 extinction events also impact the biogeographic distributions of taxa (Finnegan et al., 2016; Penn
37 et al., 2018), the relative diversities of different ecological guilds (Schubert and Bottjer, 1995;
38 Roopnarine and Angielczyk, 2015; Dineen et al., 2019), body-size distributions within and among
39 species (Payne, 2005; Atkinson et al., 2019), and the abundances of biomimeticizing and
40 bioturbating taxa (Twitchett and Barras, 2004; Knaust, 2010; Cribb and Bottjer, 2020). The relative

41 taxonomic and ecological significances of mass extinction events often differ, meaning that
42 extinction and recovery must be understood through the analysis not only of taxonomic diversity
43 but also of other dimensions of biodiversity and ecosystem function (Droser et al., 2000; Greene
44 et al., 2011; Hull et al., 2011; McGhee et al., 2013). Some of the most taxonomically severe
45 extinction events, such as the Late Ordovician extinction, had comparatively little effect on the
46 structure of ecosystems (Brenchley et al., 2001; Christie et al., 2013). In other cases, the global
47 taxonomic losses do not consistently predict the composition of post-extinction ecosystems. For
48 instance, the end-Permian mass extinction preferentially reduced taxonomic diversity within
49 heavily calcified clades such as corals, brachiopods, and echinoderms relative to moderately
50 calcified clades such as bivalves and gastropods (Sepkoski et al., 1981; Knoll et al., 1996, 2007a),
51 but some earliest Triassic shallow-marine communities remained dominated by crinoids and
52 brachiopods rather than by bivalves and gastropods (Greene et al., 2011; Brosse et al., 2019).

53 Explaining and predicting differences between the responses of diversity and abundance across
54 intervals of rapid environmental change is one of the most important unsolved problems in the
55 study of mass extinction and the conservation of future biodiversity and ecosystem function.
56 Reduction in animal abundance, either overall or differentially across clades and functional groups,
57 can fundamentally alter the state of an ecosystem even in the absence of extinction. For example,
58 marine defaunation by commercial fishing has caused population blooms of species at lower
59 trophic levels, such as prawns and sea urchins, in some parts of the ocean (Scheffer et al., 2005;
60 Diaz and Rosenberg, 2008; McCauley et al., 2015). On land, the biomass of large mammals has
61 increased over the past several centuries even as their taxonomic diversity has declined due to the
62 enormous increases in the population sizes of humans as well as domestic cattle, pigs, and sheep
63 (Barnosky et al., 2012).

64 Despite the importance of overall biomass to trophic transfer efficiency and nutrient cycling, the
65 responses of overall biomass within ecosystems to past mass extinction events remains poorly
66 quantified. Only a few studies across the end-Silurian (Bowman et al., 2020), end-Permian (Payne
67 et al., 2006; Jacobsen et al., 2011), and end-Cretaceous (Hull et al., 2011; Sibert et al., 2014)
68 extinctions provide empirical constraint on changes in animal abundance across mass extinction
69 intervals. The key pattern emerging from this work is that changes in organismal abundance vary
70 considerably in both time and space across different extinction intervals. These few studies have
71 demonstrated that quantifying animal abundance across mass extinctions in a biogeochemical
72 framework can provide a robust picture of the ecological severity of mass extinctions (Hull, 2015).
73 From a functional standpoint, mass rarity may be as important or more important than the reduction
74 of biodiversity (Hull, 2015).

75 The end-Triassic mass extinction, ca. 201.5 Ma (Blackburn et al., 2013), was the most
76 taxonomically severe mass extinction experienced by a marine fauna that is taxonomically similar
77 to its modern counterpart (Sepkoski et al., 1981; Ritterbush et al., 2015a). The ecological severity
78 of the end-Triassic mass extinction trails only the end-Permian and end-Cretaceous mass
79 extinctions (McGhee Jr et al., 2004; McGhee et al., 2013; Bond and Grasby, 2017). It preferentially
80 impacted the diversity of reef-building taxa like scleractinian corals and sphinctozoan sponges
81 (Hautmann, 2004; Kiessling and Aberhan, 2007; Kiessling et al., 2007; Dunhill et al., 2018).
82 Geological and geochemical data indicate many parallels between the circumstances of the end-83
83 Triassic extinction and the “deadly trio” (Pörtner et al., 2004) of climate warming, ocean
84 deoxygenation, and ocean acidification that imperil extant marine animals. During the end-Triassic

85 mass extinction, volcanism in the Central Atlantic Magmatic Province (CAMP) caused the
86 massive injection of isotopically light carbon into the atmosphere (Ruhl and Kürschner, 2011;
87 Bachan et al., 2012; Fox et al., 2020). Sedimentary and geochemical proxy data indicate a resulting
88 drop in the ocean pH (Greene et al., 2011), rise in temperature of about 10 °C in the tropics (Pálfy
89 et al., 2007), and expansion of anoxic and euxinic bottom waters (van de Schootbrugge et al., 2007;
90 Richoz et al., 2012; Kasprak et al., 2015; Jost et al., 2017b; Atkinson and Wignall, 2019; Larina et
91 al., 2019; He et al., 2020).

92 Although the taxonomic severity and environmental circumstances of the end-Triassic extinction
93 are increasingly well constrained, their impact on the abundances of animals and other well-
94 fossilized taxa (e.g., foraminifera and calcareous algae) remain essentially unknown due to the
95 lack of relevant data. To fill this knowledge gap, here we quantify changes in organismal
96 abundance across the Triassic/Jurassic transition by generating limestone compositional data
97 across four stratigraphic sections spanning the Triassic/Jurassic boundary in the Tethys realm. By
98 standardizing skeletal content relative to rock volume in carbonate-rich depositional environments
99 and by investigating changes in skeletal content across depositional settings, we provide the first
100 estimate of overall changes in the biomass of skeletal animals, calcareous algae, and foraminifera.
101 Because physiological data and evidence from both the modern ocean (Deutsch et al., 2015) and
102 the end-Permian mass extinction (Penn et al., 2018b) indicate that the effects of oxygen availability
103 for aerobic organisms present an important control on extinction and biogeography, we also
104 reconstruct the variation in redox conditions for two of the sections through measurement of I/Ca
105 ratios.

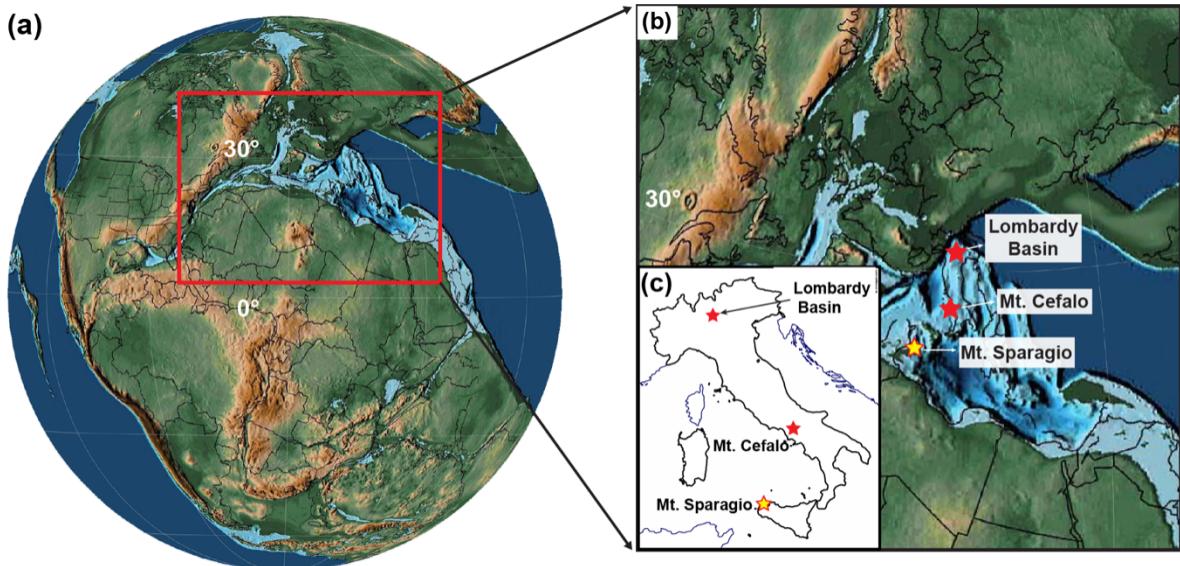
106

107 **2 Geological setting**

108 **2.1 Overview**

109 The shallow-marine carbonate sediments exposed in the southern Alps (Lombardy Basin) and
110 southern Apennines of Italy record sedimentation in two distinct paleogeographic domains of
111 western Tethys during the Mesozoic (Fig.1). These paleogeographic domains reflect the Triassic
112 rifting of Pangaea, which led to closure of the Paleotethys Ocean and expansion of the Neotethys
113 Ocean in the Late Triassic (Haas et al., 1995; Schettino and Turco, 2011). As a result of rifting, a
114 series of fault-controlled, intra-platform basins developed across the western Mediterranean.
115 Shallow-marine carbonate platforms developed during this time in the northern and southern parts
116 of western Tethys (Stampfli et al., 2001; Iannace and Zamparelli, 2002). The platforms exposed
117 in the northern, central, and southern Apennines of Italy developed in connection with the Ionian
118 Arm of Tethys, which opened in the Middle Triassic (Ciarapica and Passeri, 2005). Sedimentary
119 deposits of the Northern Calcareous Alps developed in connection with the Alpine Ocean (also
120 referred to as the Ligurian-Piedmont Ocean), which started opening in the Late Triassic to Early
121 Jurassic (Bertotti et al., 1993; Jadoul et al., 2005) (Fig.1).

122



123
124

125 *Figure 1: Paleogeography and study site locations.* Panel (a) shows the Early Jurassic
126 paleogeography modified using the GPLATES software. Panel (b) indicates the paleogeographic
127 locations of the Lombardy Basin (southern Alps), Mt. Cefalo (southern Apennines) and Mt.
128 Sparagio (Sicily) sections in the Tethys sea. Panel (c) shows the present-day locations of
129 aforementioned sections in Italy. The GPS coordinates for the sections studies in this study are:
130 Ital cementi: 45°46'33.83"N 9°31'9.91"E, Val Adrara: 45°43'29.33"N 9°57'32.29"E, Pozzo
131 Glaciale: 45°41'5.99"N 10°2'56.83"E, Mt. Cefalo: 41°15'32.05"N 13°32'10.50"E. The
132 approximate location of Mt. Sparagio section was obtained from He et al. (2022a). The
133 paleogeographic maps are from Scote se, (2016). The outline map of Italy was downloaded from
134 [www.d-maps.com](http://www.d-maps.com/carte.php?num_car=2322&lang=en) using this link: https://d-maps.com/carte.php?num_car=2322&lang=en

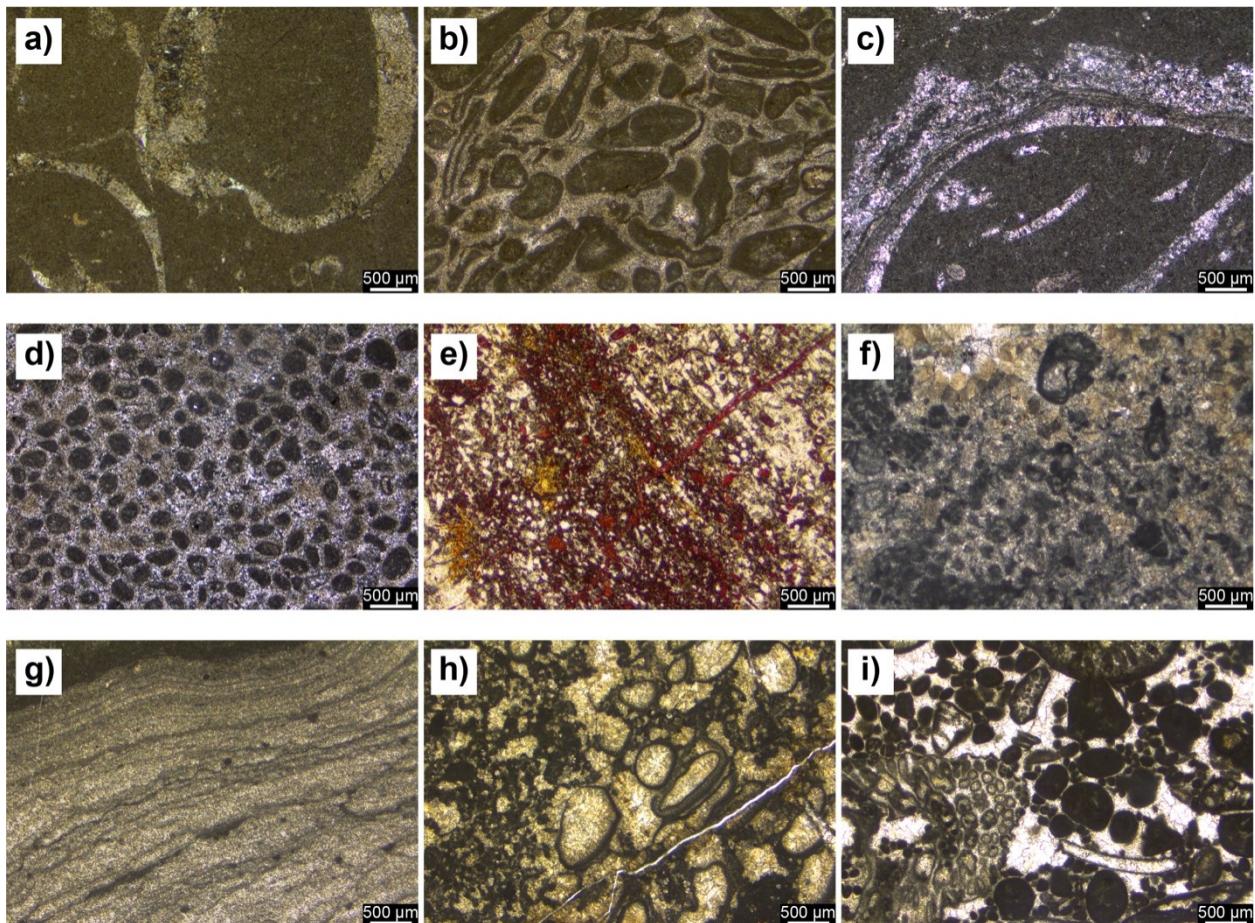
135

136 **2.2 Lombardy Basin**

137 The Lombardy Basin was in close proximity to the European continent during the Late Triassic
138 and Early Jurassic (Fig. 1) and was a site of sediment accumulation in an active rift basin (Jadoul
139 et al., 1992, 2004; Berra et al., 2010). Rifting led to development of horst and graben structures,
140 which became sites of carbonate sedimentation (Jadoul et al., 1992). During the Norian, shallow-
141 marine carbonates accumulated on structural highs and bedded turbiditic limestones accumulated
142 in deeper intra-platform troughs. Rifting slowed during the late Norian and a west-to-east-dipping
143 homoclinal ramp developed (Jadoul et al., 1992, 2004). The Rhaetian-Hettangian ramp carbonates
144 in the Lombardy Basin increase in thickness from west to east due to differential subsidence
145 associated with rifting. Basin-wide marine transgression during the late Hettangian led to
146 deposition of chert-rich sediments of the Sedrina and Moltrasio limestones. The platform
147 ultimately drowned early in the Sinemurian (McRoberts, 1994; Jadoul et al., 2004).

148 The Rhaetian-Hettangian sedimentary succession in the Lombardy Basin of the south-western
149 Alps consists of shallow-marine ramp carbonates (Galli et al., 2007). The Rhaetian succession of
150 the Lombardy Basin is represented by the Zu Limestone (Figs. 2(a, b, c), 3, 4). The Zu Limestone
151 is a fossiliferous packstone to framestone with a diverse open marine fauna consisting of bivalves,
152 gastropods, and large benthic foraminifera with subordinate corals, brachiopods, ostracods, and
153 echinoderms (Fig. 2). The Zu Limestone is abruptly overlain by the Malanotte Formation, which
154 has been assigned a Hettangian age based on pollen content and carbon isotope chemostratigraphy
155 (van de Schootbrugge et al., 2008). The lower Malanotte Formation is an unfossiliferous marl
156 (Figs. 3, 4). The marl is overlain by carbonate-rich mudstones and wackestones containing rare
157 mollusks (Fig. 2). The middle-upper Malanotte Formation is composed of oolitic sands and
158 peloidal packstone indicating a shallowing-upward trend (Galli et al., 2007; Jadoul and Galli,
159 2008). The Albenza Formation overlies the Malanotte Formation and is composed of oolitic
160 grainstone and oncoidal packstone indicating deposition in a shallow-marine environment (Figs.
161 2-5; Galli et al., 2007; Jadoul and Galli, 2008). The Albenza Formation carbonates also contain
162 some stratigraphically limited intervals exhibiting fabric destructive dolomitization. The Sedrina
163 Limestone, which overlies the Albenza Formation, is a cherty, sponge-rich limestone at the Val
164 Adrara locality. It is composed of thin- to medium-bedded mudstones and wackestones and lacks
165 visible bedforms, thus indicating deposition in a deep-water, distal ramp setting (McRoberts, 1994;
166 Bachan et al., 2012). At the Pozzo Glaciale section, the Sedrina Limestone is composed of oncotic
167 and oolitic grainstone to packstone, indicating deposition in a subtidal, open-marine environment
168 (McRoberts, 1994; Jadoul and Galli, 2008) (Figs. 3-5). The Pozzo Glaciale section has been
169 interpreted as the eastern margin of the Lombardian Platform (Fig. 5; McRoberts, 1994; Jadoul
170 and Galli, 2008). The Moltrasio Limestone, which overlies the entire Lombardy Basin, is
171 composed of chert-bearing mudstone and marks the final drowning of the Lombardian Platform
172 (Fig. 2; McRoberts, 1994; Jadoul and Galli, 2008).

173 In this study, we analyzed hand samples from the three measured stratigraphic sections within a
174 broad shallow-to-deep depositional gradient in the Lombardy Basin: Italcementi, Val Adrara, and
175 Pozzo Glaciale (Figs. 3-5). The sections can be correlated based on the formation boundaries,
176 supplemented by carbon isotope chemostratigraphy (Galli et al., 2005; Bachan et al., 2012).
177 Biostratigraphic age assignments are based on Gaetani (1970), Lakew (1990, 1994), McRoberts
178 (1994), and Muttoni et al. (2010).



179
180

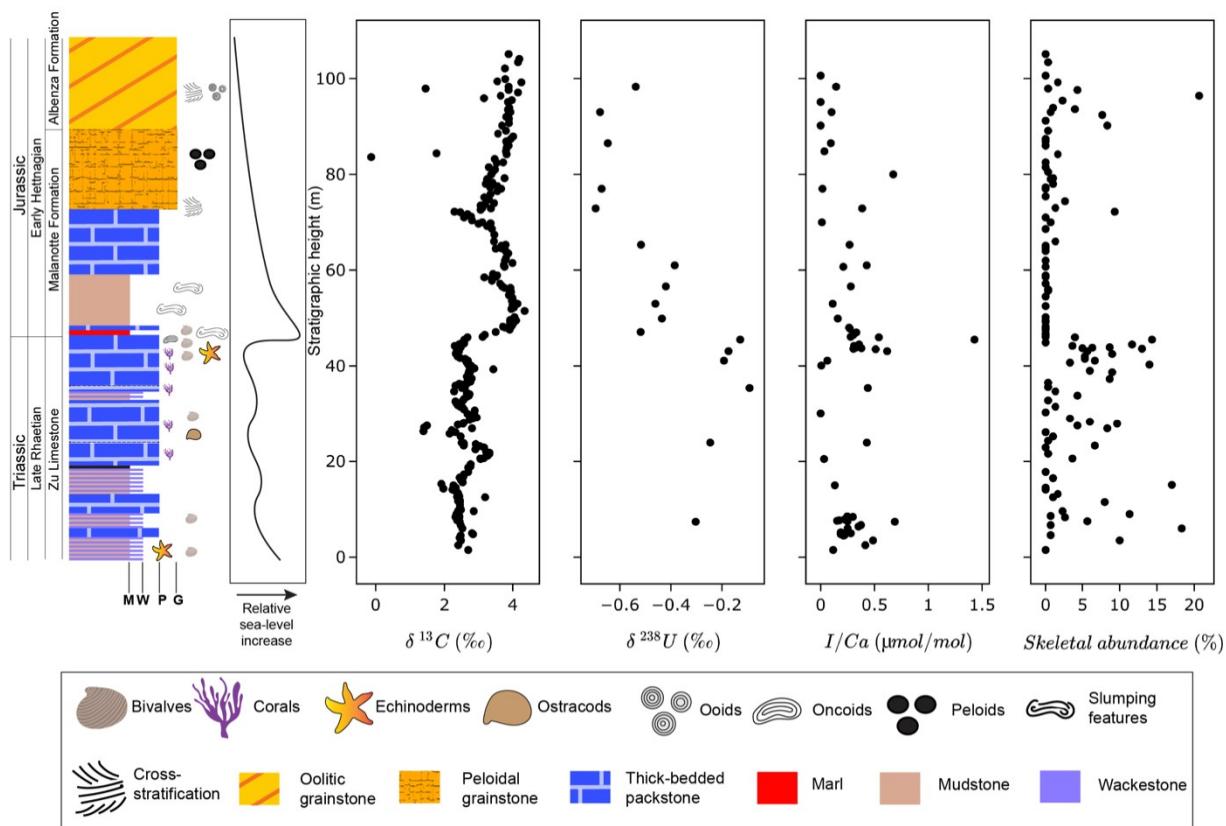
181 *Figure 2: Thin section photomicrographs illustrating microfacies from the Lombardy Basin and*
 182 *Apennine Platform. (a) Skeletal packstone from the Rhaetian Zu Limestone showing megalodontid*
 183 *bivalve fragments (Italcementi section), (b) Peloidal grainstone from the Rhaetian Zu Limestone*
 184 *(Italcementi section), (c) Bivalve-rich skeletal packstone from the Rhaetian Zu limestone (Val*
 185 *Adrara section), (d) Oolitic grainstone from lower Hettangian Albenza Formation (Val Adrara*
 186 *section), (e) Sponge spicules from early Sinemurian Moltrasio Limestone (Pozzo Glaciale section),*
 187 *(f) The Rhaetian association of dasyclad green algae (*Gryphoporella curvata*) and problematic*
 188 *algae (*Thaumatoporella parvovesiculifera*) from Mt. Cefalo section, (g) Laminar stromatolites*
 189 *from the early Hettangian (Mt. Cefalo section), (h) Problematic algae in a clotted microbial fabric*
 190 *(lower Hettangian, Mt. Cefalo section), (i) Open-marine, subtidal assemblage of calcareous green*
 191 *algae, bivalves, and peloids in upper Sinemurian-lower Pliensbachian Mt. Cefalo section.*

192

193 2.3 Apennine Platform

194 The Mt. Cefalo section is part of the vast, shallow-marine carbonate succession of the Apennine
 195 Platform (Fig. 1). It was deposited in the south-western Tethys close to the deep-water, Ionian arm
 196 of Tethys. The peritidal sedimentary successions from this shelf are exposed in the central and
 197 southern Apennines (Mancinelli et al., 2005; Di Stefano et al., 2015), western Sicily (Todaro et
 198 al., 2016, 2017), Greece (Romano et al., 2008), and Turkey (Coskun Tunaboylu et al., 2014).

199 The measured section is part of a thick Mesozoic carbonate succession deposited from the Late
 200 Triassic to Cretaceous in the northern, central, and southern Apennines of Italy (Fig. 6). The
 201 sediments from Mt. Cefalo were deposited in a shallow-marine, peritidal environment (Mancinelli
 202 et al., 2005, Bachan et al., 2012). The meter-scale cycles at Mt. Cefalo, the shallowest section of
 203 our four sections, are typically composed of allochem-rich packstone and grainstone capped by
 204 fenestral boundstones (Fig. 2). The cycles alternate between microfacies containing clotted and
 205 filamentous microbialites exhibiting fenestral fabrics indicating an intertidal to supratidal
 206 depositional environment and microfacies containing calcareous algae and benthic foraminifera
 207 and lacking fenestral fabrics, suggesting deposition in a shallow subtidal to restricted lagoon
 208 setting (Mancinelli et al., 2005, Bachan et al., 2012). The Rhaetian fossil assemblage is dominated
 209 by dasyclad green algae (*Gryphoporella curvata*), large benthic foraminifera (*Triasina hantkeni*),
 210 megalodontid bivalves, gastropods, echinoderms, ostracods, and problematic algae
 211 (*Thaumatoporella parvovesiculifera*) (Fig. 2). The Hettangian-lower Sinemurian (110-270 m)
 212 assemblage is distinguished by an absence of megalodontid bivalves, dasyclad algae, and large
 213 benthic foraminifera. The Hettangian-lower Sinemurian biota is dominated by problematic algae
 214 with minor mollusks. Dasyclad algae reappear and become more common in the upper
 215 Sinemurian-lower Pliensbachian strata (270-325 m) along with a rich assemblage of large benthic
 216 foraminifera (Mancinelli et al., 2005) (Fig. 2).



217
 218

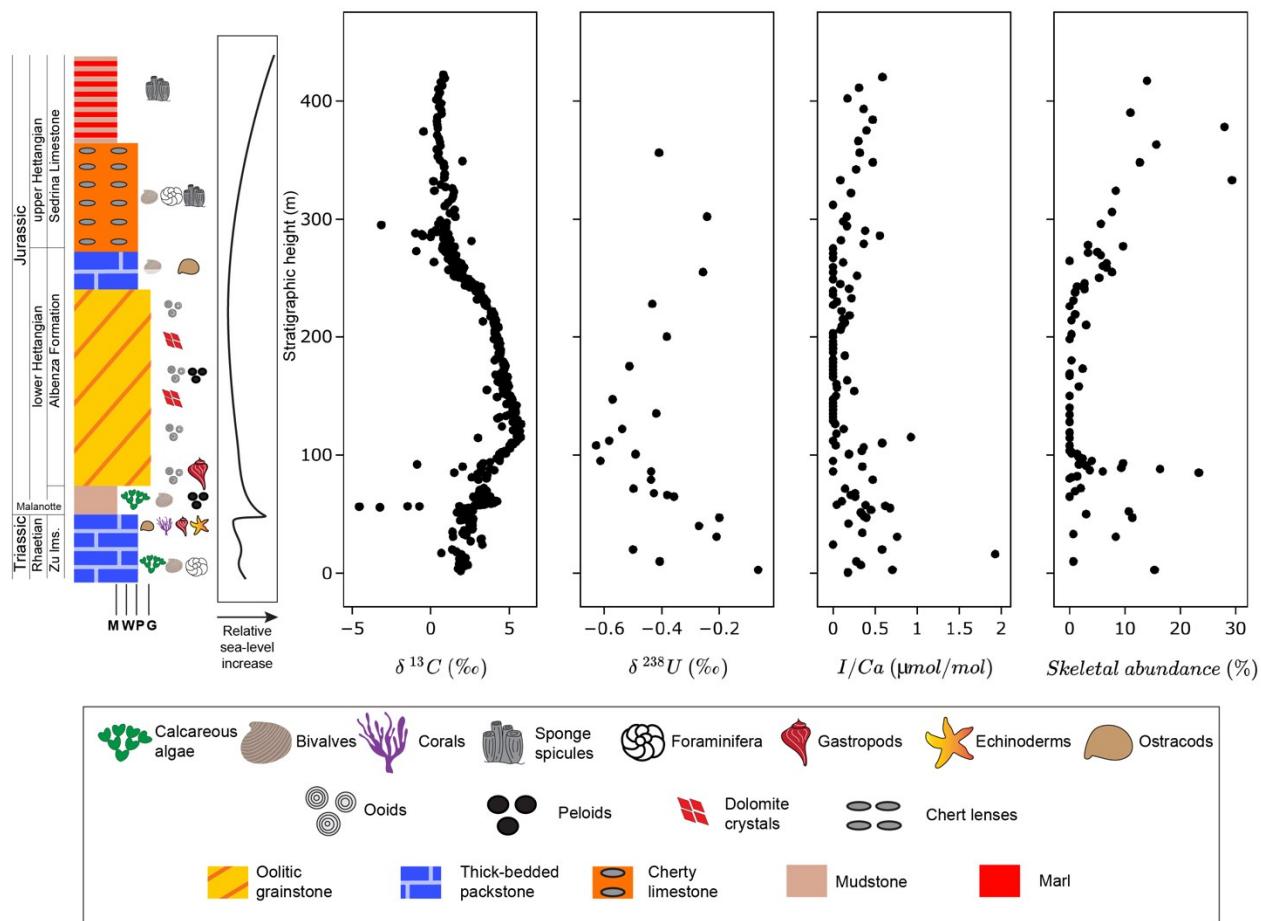
219 *Figure 3: The variations in carbonate carbon isotope ratios ($\delta^{13}\text{C}$; Bachan et al., 2012),*
 220 *uranium isotopes ratios ($\delta^{238}\text{U}$; Jost et al., 2017), I/Ca ratios and skeletal abundance at the*
 221 *Italcementi section in the Lombardy Basin (this study).*

223 **3 Materials and Methods**

224 The samples used in this study were collected from four measured stratigraphic sections, three
 225 from the Lombardy Basin (Figs. 3-5) in the southern Alps and one (Mt. Cefalo) from the Apennine
 226 Platform (Fig. 6) in the southern Apennines. The hand samples were used for the compositional
 227 analysis and I/Ca measurements. A total of 370 (Italcementi: 159, Val Adrara: 169, Pozzo Glaciale:
 228 42) samples were studied from the Lombardy Basin, and 67 samples were studied from the
 229 Apennine Platform section at Mt. Cefalo. The number of samples analyzed for both composition
 230 and I/Ca ratio amounted to 26 and 15 for the Italcementi Val Adrara sections respectively.

231 **3.1 Compositional analysis of marine limestones**

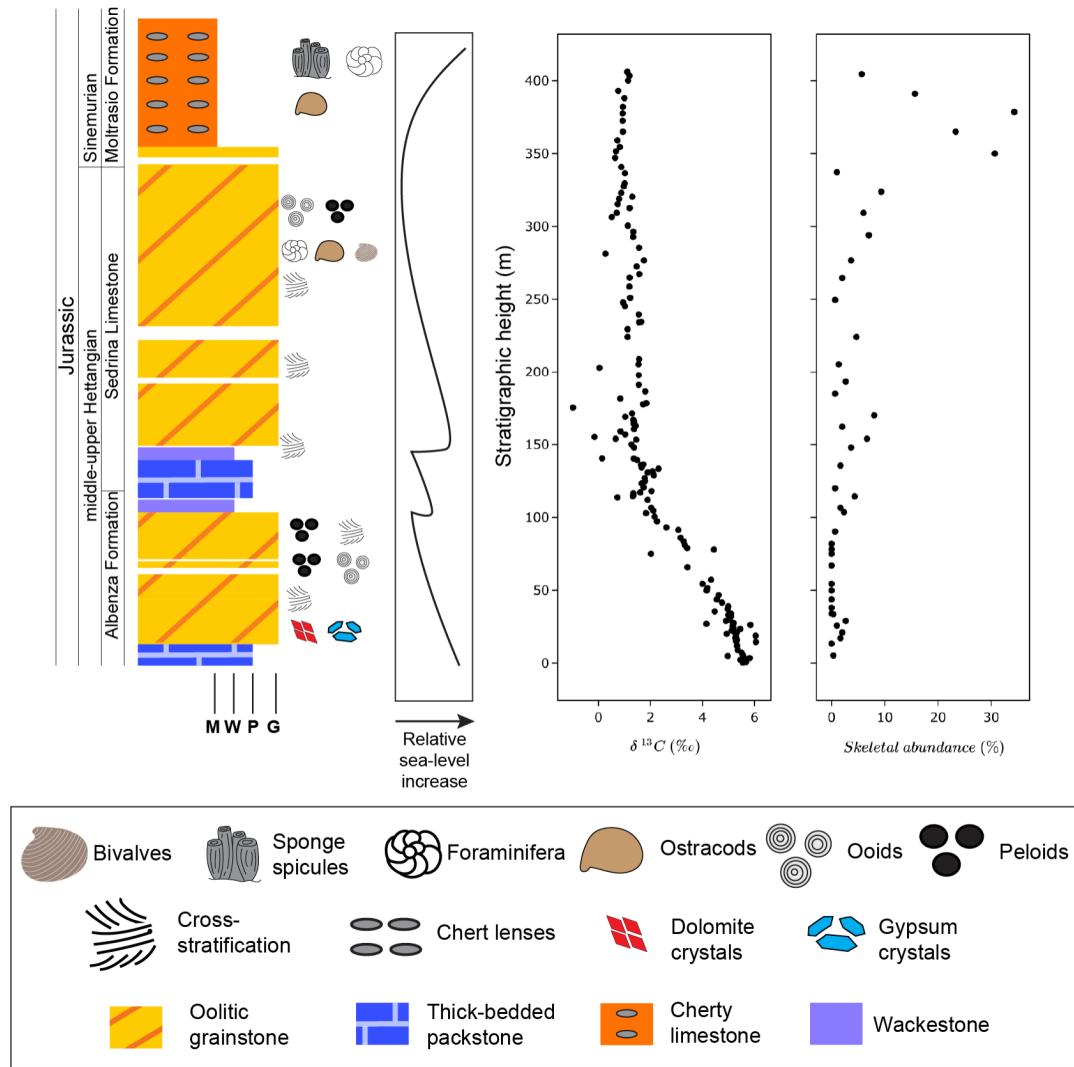
232 In this study, a total of 293 samples: 112, 72, 42 from Italcementi, Val Adrara and Pozzo Glaciale,
 233 respectively, and 67 from Mt. Cefalo were chosen for thin sectioning. The proportional content of



235 *Figure 4: The variations in carbonate carbon isotope ratios ($\delta^{13}\text{C}$; van de Schootbrugge et al.,
 236 2008; Bachan et al., 2012), uranium isotopes ratios ($\delta^{238}\text{U}$; Jost et al., 2017), I/Ca ratios and
 237 skeletal abundance at the Val Adrara section in the Lombardy Basin (this study). The age model
 238 is based on Bachan and Payne (2016).*

239 grains, matrix, cement, and void space was quantified via compositional analysis. Compositional
240 analysis was conducted by counting 300 points per thin section (size: 26mm X 46mm) across the
241 Rhaetian-Hettangian interval in the Lombardy Basin and Mt. Cefalo section from the Apennine
242 Platform (Table S1). Specifically, samples were point-counted using a mechanical stage following
243 the grain-solid method (Flügel, 2010). Skeletal grains were assigned to higher taxonomic groups
244 when identifiable, down to the Linnaean class level where possible. The categories used for
245 compositional analysis were: bivalves, gastropods, brachiopods, echinoderms, ostracods, corals,
246 bryozoans, sponges, foraminifera, red algae, green algae, problematica, unidentified biota,
247 filamentous microbial fabric, clotted microbial fabric, micrite, micrite clasts, peloid, radialxial
248 fibrous calcite cement, blocky calcite cement, microsparite, dolomite, and ooids. The microbial
249 components were point-counted by the presence of clotted and filamentous fabric in thin sections.
250 Microbial components were not included in the calculation of total skeletal abundance calculation,
251 which focused on enzymatically controlled carbonate skeletons.

252 The diversity of skeletal components in limestone samples was quantified by a comprehensive
253 survey of each thin section. The diversity of a thin section is represented by the number of distinct
254 taxonomic categories observed (Table S2). The total diversity of a thin section was quantified as
255 the number of distinct categories of enzymatically controlled skeletal grains present in the thin
256 section (see above for the categories used). For instance, if a thin section contained only calcareous
257 algae and bivalves, then its diversity would be noted as two. This approach ensured that a skeletal
258 grain that was not counted while point counting would still be accounted for in the total diversity.



259

260 *Figure 5: The variations in carbonate carbon isotope ratios ($\delta^{13}\text{C}$; van de Schootbrugge et al.,*
 261 *2008; Bachan et al., 2012) and skeletal abundance at the Pozzo Glaciale section in the eastern*
 262 *edge of Lombardy Basin (this study).*

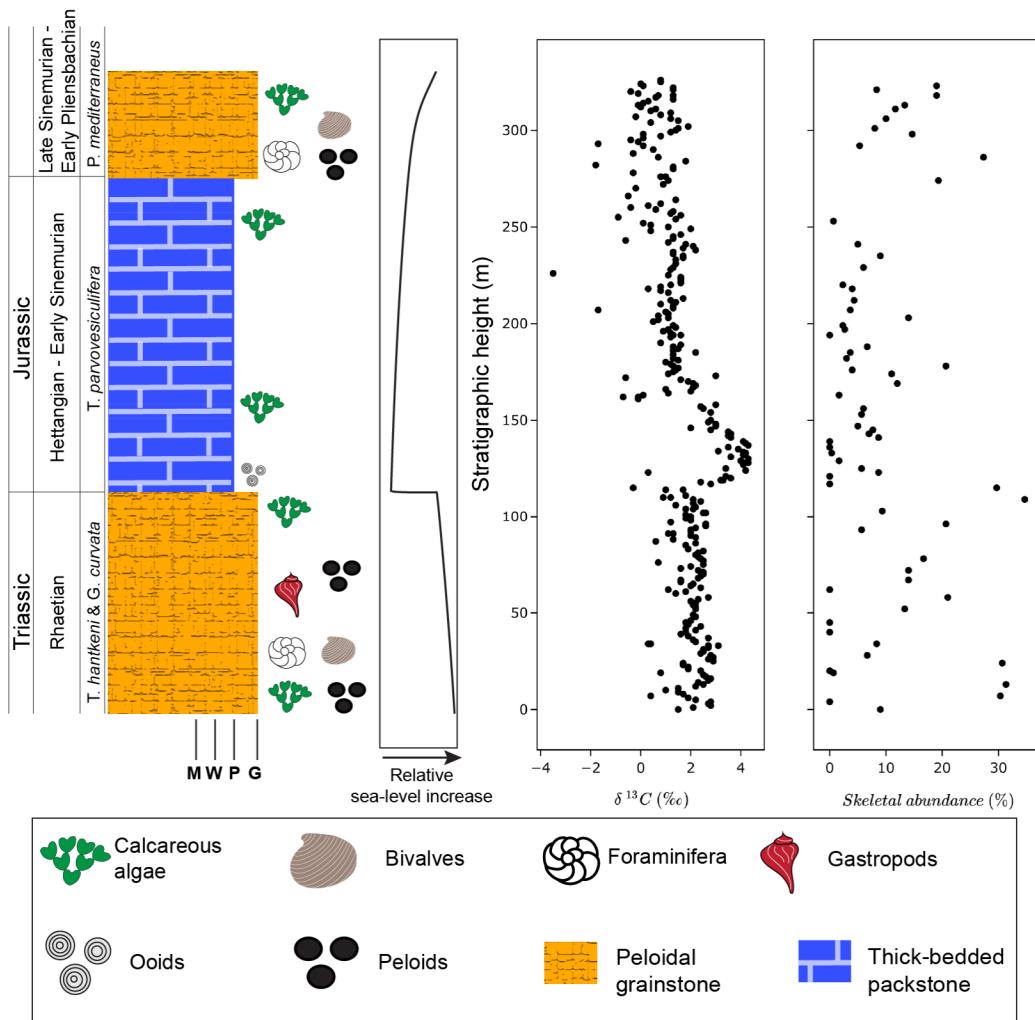
263

264 3.2 I/Ca as a redox proxy

265 The I/Ca ratio of marine limestone has been established as a paleo-proxy for shallow-marine redox
 266 conditions (Lu et al., 2010, 2018; Zhou et al., 2014, 2015; Hardisty et al., 2017). The residence
 267 time of iodine in seawater is ~ 300 ky, which is much greater than the seawater circulation time
 268 (~ 1 ky). Although the concentration of iodine in the ocean can be assumed to be globally uniform
 269 (Chance et al., 2014), its speciation is spatially variable. The dissolved inorganic iodine is
 270 dominantly present in its thermodynamically stable forms of iodate (IO_3^-) and iodide (I^-) ions in
 271 seawater (Wong and Brewer, 1977). The standard reduction potential of IO_3^-/I^- is very close to that
 272 of $\text{O}_2/\text{H}_2\text{O}$, thus making the speciation of iodine very sensitive to dissolved oxygen concentrations.
 273 In oxic conditions, most iodine occurs as iodate and is reduced to iodide in dysoxic to anoxic
 274 conditions.

275 In the modern ocean, the concentration of iodate qualitatively tracks the concentration of dissolved
276 oxygen in seawater (Truesdale et al., 2000; Lu et al., 2016). In the well-oxygenated upper ocean,
277 iodate is the dominant iodine species whereas in anoxic basins and in oxygen minimum zones
278 (OMZs), iodide is dominant (Wong and Brewer, 1977; Emerson et al., 1979). During formation of
279 carbonate sediment, only iodate is incorporated into the carbonate lattice (Lu et al., 2010).
280 Consequently, the concentration of iodine relative to calcium in limestone is sensitive to the
281 dissolved oxygen concentration in the seawater from which it was precipitated (Lu et al., 2010).

282 In total, 104 samples from the Val Adrara and Italcementi sections of the Lombardy Basin were
283 analyzed for I/Ca ratios following the methods of Lu et al. (2018) (Table S3). The I and Ca
284 concentrations were measured on a Bruker M90 quadrupole inductively-coupled-plasma mass
285 spectrometer (ICP-MS) housed at Syracuse University. For the analysis, 3-5 mg of powdered
286 limestone (extracted using 0.8 mm drill bit) samples were weighed out, thoroughly rinsed with de-
287 ionized water, and then dissolved in 3% HNO_3 acid. The solutions were diluted for analysis to
288 achieve \sim 50 ppm Ca. To stabilize iodine, 0.5% tertiary amine solution was added to each solution.
289 For analysis, 5 ppb of Sc and Cs were added as internal standards. The detection limit of I/Ca is
290 around 0.1 $\mu\text{mol/mol}$. The sensitivity of ^{127}I is tuned to about 80–100 kcps for a 1 ppb standard.
291 The precision of ^{127}I is typically better than 1%. The long-term precision is assessed with repeated
292 measurements of the reference material JCp-1 (Lu et al., 2010) and was $3.70 \pm 0.27 \mu\text{mol/mol}$ (1σ)
293 ($n=2,280$) (Lu et al., 2020). The I/Ca values of samples were corrected by adjusting the value of



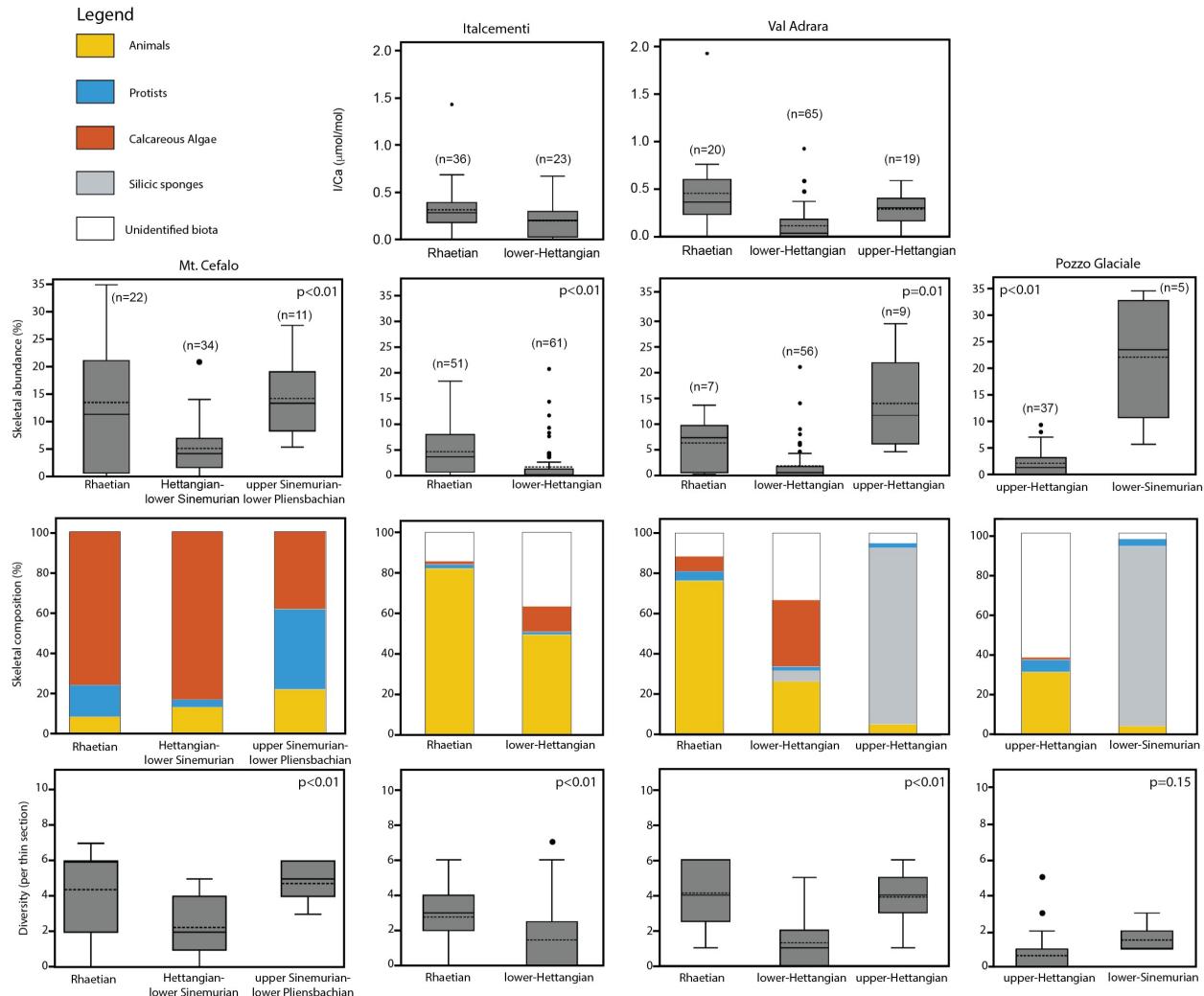
295
296 *Figure 6: The variations in carbonate carbon isotope ratios ($\delta^{13}\text{C}$; Bachan et al., 2012) and*
297 *skeletal abundance in the Mt. Cefalo section of the Apennine platform (this study).*

298

299 **4 Results**300 **4.1 Compositional analysis of marine limestones**

301 The Lombardy Basin and the Apennine Platform both record a drop in skeletal abundance across
302 the Triassic-Jurassic boundary (Figs. 3–6). All three stratigraphic sections indicate a statistically
303 significant change in the skeletal abundance across the Upper Triassic and Lower Jurassic time
304 intervals (binned into time intervals defined by the stratigraphic or biostratigraphic constraints for
305 each stratigraphic section as depicted in Figs. 3–6; one-way ANOVA results: Mt. Cefalo: $p<0.01$,
306 Italcementi: $p<0.01$, Val Adrara: $p=0.01$) (Fig. 7). Furthermore, a two-way ANOVA on all 293
307 samples across the four stratigraphic sections shows that the time interval ($p<0.01$, $\eta^2=0.30$)
308 explains more of the variation in skeletal content than which stratigraphic section the sample is

309 from ($p<0.01$, $\eta^2=0.15$). To test this finding further, we conducted another two-way ANOVA
 310 consisting of samples from the same ramp, but different water depths from the Ital cementi and Val
 311 Adrara sections. The results again showed similar results with more of the variation in skeletal
 312 abundance explained by time interval ($p<0.01$, $\eta^2=0.37$) than by stratigraphic section ($p<0.01$,
 313 $\eta^2=0.12$).



314
 315 *Figure 7: Box plots showing changes in I/Ca ratios, skeletal abundance, skeletal composition,*
 316 *diversity (per thin section) in Upper Triassic and Lower Jurassic limestones from the Lombardy*
 317 *Basin (Italcementi, Val Adrara, Pozzo Glaciale) and Apennine Platform (Mt. Cefalo) sections.*
 318 *“n” indicates the number of samples in each category. The figure broadly depicts significant*
 319 *changes in the skeletal abundance, biotic composition and diversity in marine ecosystems during*
 320 *the Triassic/Jurassic transition along with the redox chemistry as depicted by I/Ca ratios.*

321 The composition of skeletal material across Lombardy Basin and Apennine Platform stratigraphic
 322 sections differed significantly depending on the depositional environment and time interval (Fig.
 323 7). The main difference between the Lombardy Basin and Apennine Platform limestones is the
 324 dominance of non-poriferan animals as skeleton producers in the Lombardy Basin whereas
 325 calcareous algae and protists are the skeletal grains in the Apennine Platform (Fig. 7). The Rhaetian
 326 Zu Limestone at Italcementi and Val Adrara is primarily composed of non-poriferan animal

327 skeletons (~80%) with protists and calcareous algae forming a minor (<5%) component of skeletal
328 material. The proportion of non-poriferan animal skeletal material as a fraction of all skeletal
329 material decreased in the lower Hettangian strata in both Ital cementi and Val Adrara sections
330 whereas the proportion of calcareous algae increased. In the upper Hettangian Sedrina limestone,
331 the Val Adrara locality shows evidence of a rapid increase of water depth evident from the well-
332 bedded carbonate mudstone-wackestone facies rich in sponge spicules floating in a micritic matrix
333 (McRoberts, 1994; Bachan et al., 2012). The skeletal component of limestones at this location is
334 primarily hexactinellid sponges (88%) with minor contributions from non-poriferan calcifying
335 animals and protists. This composition contrasts with that of Pozzo Glaciale, where the identifiable
336 skeletal components are entirely composed of non-poriferan animals and protists. The deepening
337 trend continued across the entire Lombardy Basin during the late Sinemurian and led to deposition
338 of the Moltrasio Limestone, which primarily contains hexactinellid sponges (89%) (Fig. 4, 5, 7).

339 In the Apennine Platform, the peritidal facies at Mt. Cefalo are primarily composed of calcareous
340 algae (>75%) in the Rhaetian and Hettangian-early Sinemurian strata (Fig. 7). The composition of
341 skeletal material changed significantly in the upper Sinemurian and lower Pliensbachian strata
342 with a significant contribution from benthic foraminifera (~40%) and animals (~22%).

343 Similar to the skeletal abundance trends, the mean (and median) diversity per sample dropped
344 significantly across the Triassic-Jurassic boundary in both Lombardy Basin and Apennine
345 Platform sections (one-way ANOVA analysis, Mt. Cefalo: $p<0.01$, Ital cementi: $p<0.01$, Val
346 Adrara: $p<0.01$) (Fig. 7). The variations in taxonomic diversity across the four stratigraphic
347 sections are more strongly associated with changes in time interval (two-way ANOVA: $p<0.01$,
348 $\eta^2=0.19$) than with the identity of the stratigraphic section (two-way ANOVA: $p<0.01$, $\eta^2=0.09$).
349 Furthermore, the frequency with which biota from dominant taxonomic categories occurred in
350 samples also decreased sharply across the Triassic-Jurassic boundary (Fig. S1).

351 **4.2 I/Ca ratios of bulk carbonate samples**

352 Iodine/calcium background values in both sections are generally low (< 2 $\mu\text{mol/mol}$). The I/Ca
353 ratios decline between the Rhaetian and Hettangian at both Ital cementi and Val Adrara (Figs. 3, 4,
354 7). These declines are statistically significant or marginally significant in both the Val Adrara (one-
355 way ANOVA: $p<0.01$) and Ital cementi (one-way ANOVA: $p=0.05$) sections (Fig. 7). The
356 limestone samples from the Rhaetian (Zu Limestone) of the Lombardy Basin show I/Ca ratios in
357 the range of <0.01 to 0.70 $\mu\text{mol/mol}$ (Figs. 3, 4). In the Ital cementi section, the mean I/Ca ratio is
358 $0.30 \pm 0.25 \mu\text{mol/mol}$ (1σ , $n=36$) whereas the Val Adrara section shows a slightly higher mean
359 I/Ca ratio of $0.45 \pm 0.40 \mu\text{mol/mol}$ (1σ , $n=20$; Figs. 2, 3). Overall, the I/Ca ratios in the Rhaetian
360 Zu Limestone of Lombardy Basin decrease towards the Rhaetian-Hettangian boundary. The
361 decreasing trend continues into the lower Hettangian Malanotte and Albenza formations. At the
362 Ital cementi section, the mean I/Ca ratio is $0.20 \pm 0.16 \mu\text{mol/mol}$ (1σ , $n=23$) whereas in the Val
363 Adrara section, the mean I/Ca ratio is $0.10 \pm 0.17 \mu\text{mol/mol}$ (1σ , $n=65$) in the lower-Hettangian
364 strata. In the upper Hettangian Sedrina limestone exposed only at Val Adrara locality, the I/Ca
365 ratio shows a slight increasing trend towards the top of the section and reaches a mean ratio of 0.28
366 $\pm 0.16 \mu\text{mol/mol}$ (1σ , $n=19$) (Figs. 3, 4).

367

368 **5 Discussion**

369 **5.1 Ecological changes across the Triassic/Jurassic boundary in the Lombardy Basin**

370 The compositional changes in shallow-marine carbonates across the Triassic-Jurassic boundary in
371 the Lombardy Basin and Apennine Platform indicate a significant decrease in the abundance and
372 diversity of marine organisms in shallow-marine ecosystems across the extinction interval that
373 cannot be explained by local facies variation (Fig. 8). The decrease in skeletal abundance and
374 diversity occurs across a variety of depositional environments ranging from proximal to distal
375 ramp environments of the Lombardy Basin and to peritidal environments of the Apennine
376 Platform. In addition, variation in skeletal content among time intervals is larger than variation
377 among stratigraphic sections. Thus, the observed variations in the skeletal abundance are best
378 interpreted to reflect an external control acting across all sampled locations irrespective of the
379 depositional setting.

380 The decrease in skeletal content across the Triassic/Jurassic boundary cannot be explained by
381 related changes in the depositional environments represented across the two basins (Fig. 8). In the
382 Lombardy Basin, the Rhaetian Zu Limestone is dominantly composed of open-marine fauna
383 deposited in a shallow-marine environment in both the proximal ramp section at Ital cementi and
384 the more distal section at Val Adrara. The overlying Malanotte Formation is composed of marls
385 overlain by thin-bedded carbonate mudstones indicating a rise in relative sea level (Bachan et al.,
386 2012). This abrupt change in carbonate sedimentation has been interpreted as a transgressive event
387 that can be traced across the Lombardy Basin and into the Northern Calcareous Alps of western
388 Tethys (McRoberts et al., 1997). The subsequent shallowing indicated by deposition of cross-
389 bedded oolitic and peloidal limestones of the Albenza Formation did not, however, coincide with
390 an increase in skeletal grain abundance. Instead, the skeletal abundance decreased sharply in the
391 lower-middle Hettangian Malanotte Formation and stayed low throughout the lower-middle
392 Hettangian Albenza Formation. The low skeletal abundance at both Ital cementi (proximal ramp)
393 and Val Adrara (distal ramp) sections indicates that this pattern is not an artifact of local high-
394 energy environments but was a common occurrence across the entire ramp of Lombardy Basin
395 during that time.

396 This pattern of reduced skeletal abundance and a shift to a non-skeletal carbonate factory also
397 occurs at the Mt. Cefalo section and across the entire Apennine Platform (Barattolo and Romano,
398 2005; Mancinelli et al., 2005; Brandano et al., 2016; Todaro et al., 2017). The skeletal content at
399 Mt. Cefalo is consistent with observations of physical sedimentary structures that indicate
400 deposition in a peritidal environment across the Rhaetian/Hettangian transition (see Bachan et al.
401 (2012) for detailed stratigraphy). The Rhaetian intertidal facies are composed of stromatolitic
402 laminites whereas the subtidal facies are composed of bioturbated limestones rich in large benthic
403 foraminifera (*T. hantkeni*) and calcareous algae (*G. curvata*). The Triassic/Jurassic boundary is
404 marked by a subaerial exposure surface associated with a sea-level drop that can be traced across
405 the entire Apennine Platform (Berra, 2012; Cirilli et al., 2015; Todaro et al., 2017). The limestone
406 immediately above the exposure surface is oolitic grainstone containing very little skeletal
407 material, indicating the demise of the biological carbonate factory and a shift to a non-skeletal
408 mode of carbonate precipitation. The oolitic limestones are overlain by laminated fenestral
409 boundstones indicating a microbial mode of carbonate precipitation with few intervals rich in
410 calcareous algae but almost devoid of animals and protists. The compositional data from Mt.
411 Cefalo indicates that ecosystems in peritidal environments contained few skeletal animals
412 following the end-Triassic mass extinction and that carbonate precipitation was dominantly abiotic

413 and microbially induced. This shift from metazoan-dominated to microbial and abiotic modes of
414 carbonate precipitation in early-Hettangian is not unique to Mt. Cefalo: compositional data from
415 northern and central Apennines (Brandano et al., 2016) show similar shifts in limestone
416 composition across the Triassic/Jurassic boundary. This finding further supports the interpretation
417 that the facies shifts at Mt. Cefalo are indicative of a more widespread pattern in carbonate
418 deposition rather than a local shift in the depositional environment. The facies pattern in upper
419 Hettangian-lower Sinemurian at Mt. Cefalo records a shift in sedimentation in an open marine
420 peritidal environment with thicker subtidal intervals. However, the overall metazoan abundance
421 remained low throughout the Hettangian and early Sinemurian indicating that the changes in biotic
422 abundance transcended the shift in depositional environment (Fig. 6).

423 Like the abundance patterns, the diversity data is also indicative of a biotic crisis at the end of the
424 Triassic period in Lombardy Basin and Apennine platform. The overall diversity of taxa per
425 sample decreased sharply across the Triassic-Jurassic transition. The fossil occurrences of
426 dominant taxonomic categories when plotted as the percentage of samples in which they appear
427 indicates not only that the taxa became less abundant but also that the frequency with which they
428 occurred decreased substantially (Fig. S1).

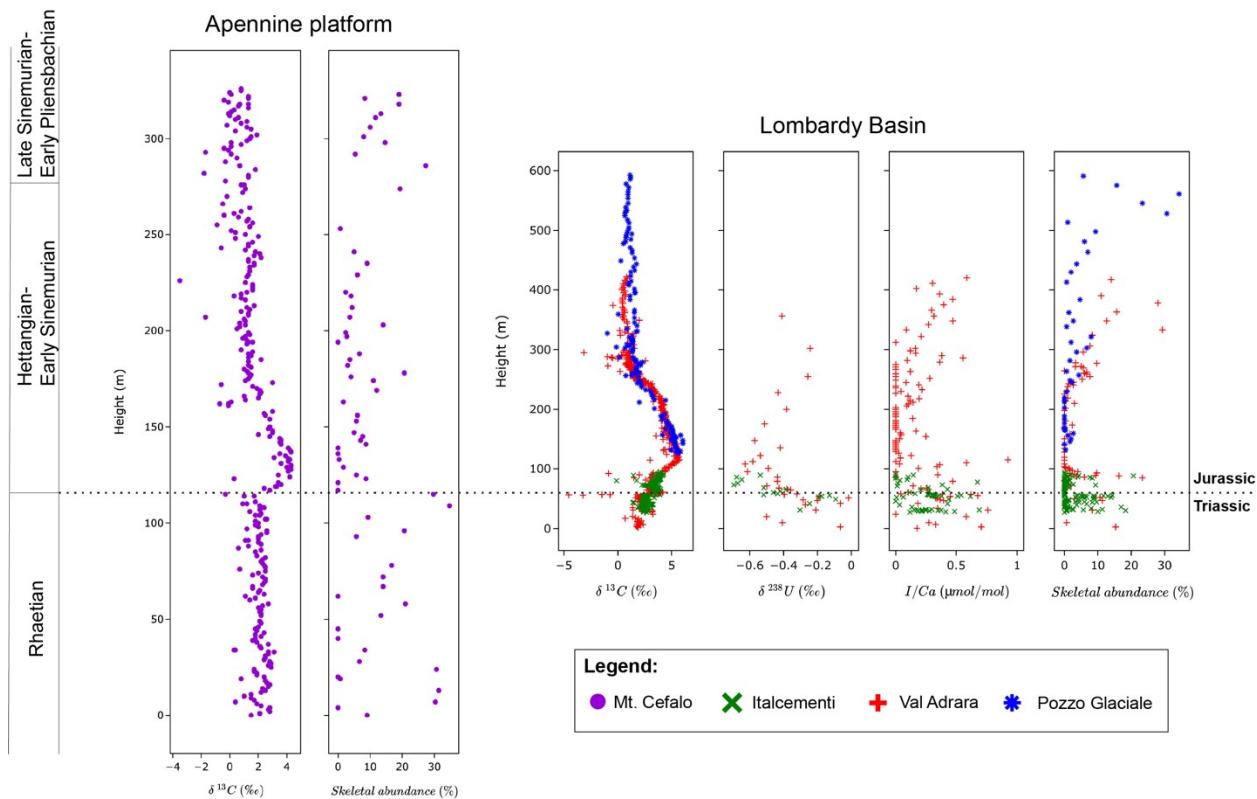
429 The close similarities in skeletal abundance and diversity patterns across the Triassic/Jurassic
430 boundary between the widely separated regions (~500 km) of the Lombardy Basin and the
431 Apennine Platform are also representative of a more general pattern across the western Tethys.
432 This pattern of reduced animal abundance across the Triassic/Jurassic boundary followed by
433 establishment of abiotic and microbial carbonate factories during the Early Jurassic occurred also
434 in the Northern Calcareous Alps in Austria (McRoberts et al., 1997), the Transdanubian Range in
435 Hungary (Palfy et al., 2021), the central and northern Apennines of Italy (Barattolo and Romano,
436 2005; Mancinelli et al., 2005; Brandano et al., 2016), western Sicily (Todaro et al., 2017, 2018),
437 Greece (Romano et al., 2008) and Turkey (Coskun Tunaboylu et al., 2014). The recovery of
438 skeletal abundance to pre-extinction levels in shallow-marine environments across western
439 Tethyan carbonate platforms occurred in upper Hettangian-Sinemurian limestones with an
440 increase in skeletal contribution from protists and mollusks. Overall, the Early Jurassic ecosystems
441 of western Tethys supported far less benthic biomass and diversity than their Late Triassic
442 counterparts.

443 The changes in shallow-marine ecosystems in the western Tethys are also representative of a global
444 biotic crisis that unfolded at the end of the Triassic. In the southern Tethys, the shallow-marine
445 platforms of the Arabian plate record a decrease in skeletal abundance across the Triassic/Jurassic
446 boundary and a shift to an abiotic carbonate factory dominated by ooids (Al-Suwaidi et al., 2016,
447 Ge et al., 2018). Evidence from eastern Panthalassa indicates a different extinction pattern than
448 Tethyan sections. During the Early Jurassic, calcifying organisms decreased in abundance and
449 were replaced by silicic demosponges in middle- to inner-ramp environments in Peru (Ritterbush
450 et al., 2014, 2015b; Corsetti et al., 2015) and Nevada (Ritterbush et al., 2015b, 2016). The post-
451 extinction facies in the Lombardy Basin (this study) and the Eiberg Basin of western Tethys
452 (Delecat and Reitner, 2005) show a similar increase in silicifying hexactinellid sponges in upper
453 Hettangian and lower Sinemurian strata. However, it is challenging to compare them with the
454 Panthalassic ramp sections because the intra-platform basins in the Northern Calcareous Alps
455 experienced a rise in relative sea level beginning in middle-late Hettangian that coincided with an
456 increase in the sponge abundance. Hence, it is unclear whether the increase in abundance of

457 sponges in the Lombardy and Eiberg Basins was a response to the end-Triassic biotic crisis or a
458 response to expansion of deeper marine benthic habitats associated with the rise in relative sea
459 level.

460 The decrease in the abundance of shell-forming organisms suggests that overall ecological activity
461 decreased sharply across the Triassic/Jurassic transition. The evidence provided above focuses
462 mainly on the composition of carbonate sediments deposited in shallow-marine environments. In
463 theory, the ecological and evolutionary dynamics of shell-forming organisms in carbonate
464 environments could differ from those of skeletal organisms in different environments or from those
465 of non-skeletal organisms. However, evidence of decreased metazoan activity across the
466 Triassic/Jurassic transition is also evident in ichnofacies patterns from both siliciclastic and
467 carbonate environments. The number of ichnotaxa, depth of bioturbation and size of burrows
468 decreased sharply during late Rhaetian-early Hettangian time in Panthalassic (New York Canyon),
469 Tethyan (Austria) and adjacent ocean basins (southern England) (Twitchett and Barras, 2004;
470 Barras and Twitchett, 2007). A sharp decrease in the bioturbation-associated porosity in limestones
471 also occurred in peritidal carbonates of western Tethys (Todaro et al., 2016). The presence of
472 wrinkle structures in shallow-marine sediments has been associated with reduced metazoan
473 activity in post-extinction environments (Pruss et al., 2004; Mata and Bottjer, 2009; Mariotti et al.,
474 2014). Similar wrinkle structures occur in siliciclastic subtidal environments during the Hettangian
475 (Häntzschel and Reineck, 1968; Peterffy et al., 2016).

476 The reduction in ecological functioning due to biogeochemical disturbance has occurred multiple
477 times during the Phanerozoic (Droser et al., 2002; Aberhan and Kiessling, 2015; Hull, 2015). The
478 mean total skeletal abundance in comparable peritidal environments was reduced by more than
479 90% across Permian-Triassic strata (from six stratigraphic sections in South China; Payne et al.,
480 2006) as compared to a ~60 % decrease across the Triassic/Jurassic strata (Fig. 9). Of the total
481 skeletal abundance, the calcareous algae show an especially interesting response to the end-
482 Permian and end-Triassic extinction events (Fig. 8). Macroscopic green algae were ecologically
483 important primary producers during the Paleozoic and early Mesozoic (Wray, 1977; Knoll et al.,
484 2007; Payne and van de Schootbrugge, 2007; Hull, 2017). Lower Triassic strata from shallow-
485 marine environments were nearly devoid of calcareous algae in the aftermath of the end-Permian
486 extinction (Bucur, 1999, Payne et al., 2006). Conversely, the calcareous algal abundance in the
487 Lower Jurassic strata decreased only slightly across the Triassic/Jurassic transition (Fig. 9). This
488 difference in algal abundance in the aftermaths of the two mass extinctions could indicate that the
489 end-Permian mass extinction was much more severe for calcifying primary producers than the
490 end-Triassic extinction (Bucur, 1999).



491
492 *Figure 8: A composite plot of variations in stable isotope proxies, I/Ca ratios and skeletal*
493 *abundance in Lombardy basin and Apennine platform. See figures 3-6 for data sources. The figure*
494 *indicates broad scale spatial and temporal changes in the abundance of skeleton-forming*
495 *organisms across the Triassic/Jurassic boundary in context of local (I/Ca) and global ($\delta^{238}\text{U}$)*
496 *redox proxies and carbon isotope ratios. The local and global redox proxies indicate a shift to*
497 *reducing redox conditions in the Lower Jurassic. The negative and positive carbon isotope*
498 *excursions in Lower Jurassic are global in extent and can be tracked across multiple basins. See*
499 *Bachan et al. (2012) for detailed description of carbon cycle changes across the Triassic/Jurassic*
500 *boundary.*

501

502 5.2 Marine redox conditions and environmental dynamics across the T/J boundary

503 The I/Ca ratios from the Lombardy Basin most likely record Triassic/Jurassic variations in marine
504 redox conditions. Typical carbonate diagenesis processes, such as alteration by meteoric fluids and
505 dolomitization, can decrease the concentrations of iodate in the carbonate crystal lattice (Hardisty
506 et al., 2017). The elemental ratios of Mg/Ca and Mn/Sr indicate low likelihood of substantial
507 diagenetic alteration in our samples. In the Lombardy Basin samples, there is weak correlation of
508 I/Ca with $\delta^{18}\text{O}$ (Pearson corr.(r), Italcementi: $r=0.27$, $p=0.04$, $n=54$; Val Adrara: $r=0.28$, $p=0.01$,
509 $n=81$), of I/Ca with Mg/Ca (Pearson corr.(r), Italcementi: $r=0.33$, $p=0.01$, $n=54$; Val Adrara: $r=$
510 0.03 , $p=0.75$, $n=81$) and of I/Ca with Mn/Sr (Pearson corr.(r), Italcementi: $r=0.11$, $p=0.69$, $n=13$;
511 Val Adrara: $r=0.12$, $p=0.43$, $n=42$) (Fig. S2). The Mn/Sr ratio, in particular, is indicative of the
512 degree of alteration through meteoric diagenesis (Brand and Veizer, 1980). In theory, the Mn/Sr
513 and I/Ca ratios should be inversely correlated in case of significant diagenesis in the meteoric zone
514 (Hardisty et al., 2017). However, the samples from the Lombardy Basin do not indicate significant

515 inverse correlation between the two ratios. Furthermore, He et al. (2022a) reported a sharp drop in
516 carbonate I/(Ca+Mg) ratios from a mean of 3.5 $\mu\text{mol/mol}$ during pre-extinction period to 1.4
517 $\mu\text{mol/mol}$ across the extinction horizon in another Italian section (Mt. Sparagio, Fig. 1),
518 independently supporting the temporal trend in local redox conditions observed in this study.

519 The lithology of the limestones also does not appear to play a major role in governing I/Ca ratios.
520 A one-way ANOVA ($p<0.05$, $\eta^2=0.16$) on the I/Ca ratios of different lithologies encountered in
521 our data indicates that although there are statistically significant differences in the means of
522 different lithologies, this factor overall explains only a minor percentage of variance in the I/Ca
523 ratios (Fig. S3). A two-way ANOVA of the Val Adrara section data, with age and lithology as the
524 independent variables, indicates that time period ($p=0.04$, $\eta^2=0.07$) is a significant predictor of
525 I/Ca ratios whereas the lithology is not ($p=0.62$, $\eta^2=0.02$). In the Ital cementi section, the lithology
526 ($p=0.07$, $\eta^2=0.45$) similarly explains more of the variation in I/Ca than does the time period
527 ($p=0.45$, $\eta^2=0.01$) but overall neither time period nor lithology is significantly associated with I/Ca
528 ratios in the two-way ANOVA. The small effect size and lack of statistically significant
529 associations could indicate that I/Ca ratios at Ital cementi section have a higher noise/signal ratio
530 than at Val Adrara. The source of noise could be diagenetic and/or lithologic factors. However,
531 the weight of the evidence points to temporal changes in primary I/Ca ratios across the
532 Triassic/Jurassic boundary.

533 The I/Ca values found in this study are relatively low (< 2 $\mu\text{mol/mol}$), whereas the Rhaetian
534 peritidal limestones from the southwestern Tethys (He et al., 2022a) have higher I/Ca values. These
535 observations can be explained in two ways: (1) the low I/Ca values in the Lombardy Basin are
536 largely primary signals indicating a low-oxygen water mass in close proximity to the sample
537 locations; or (2) post-depositional diagenetic alteration lowered the primary I/Ca of the rocks after
538 deposition. Support for a low-oxygen water mass (and a primary signal) comes from the argument
539 that the organic matter remineralization depth in the oceanic water column deepened during the
540 Mesozoic, which reduced the prevalence of shallow oxygen minimum zones compared to the
541 Paleozoic (Lu et. al, 2018). Although the exact timing of this transition is still unclear, a shallower
542 OMZ in the Triassic is consistent with current evidence, including previously published I/Ca data
543 from the Rhaetian (Lu et al., 2018). For instance, our mean I/Ca values during the Rhaetian (0.30
544 $\mu\text{mol/mol}$ and 0.45 $\mu\text{mol/mol}$ for Ital cementi and Val Adrara respectively) are close to values (0.22
545 $\mu\text{mol/mol}$) from the adjacent Lagonegro Basin (Lu et al., 2018). On the other hand, the Rhaetian
546 peritidal limestones from the southwestern Tethys exhibit high I/Ca ratios, around 3.5 $\mu\text{mol/mol}$
547 (He et al., 2022a). Therefore, it is possible that some low oxygen waters were close to the
548 Lombardy Basin sections throughout the study interval. Diagenetic loss of carbonate-associated
549 iodine during later diagenesis cannot be completely ruled out, but there is no evidence to suggest
550 that the stratigraphic trend in I/Ca is dominated by diagenesis based on relationships with $\delta^{18}\text{O}$,
551 Mg/Ca, and Mn/Sr (Fig. S2).

552 The I/Ca data (0.00-0.70 $\mu\text{mol/mol}$) from the Upper Triassic of the Lombardy Basin suggest a
553 water column where oxygen was present and relatively stable but not abundant (Fig. 8). The sub-
554 surface marine redox conditions at the end of the Triassic shifted toward anoxic/hypoxic conditions
555 based on a decrease in I/Ca (Fig. 8). Variation in I/Ca ratios indicating an expansion of local
556 dysoxic and anoxic waters across the Triassic/Jurassic boundary is consistent with other lines of
557 evidence for globally widespread anoxic and euxinic water column conditions in shallow marine
558 environments during the Early Jurassic (He et al., 2022a, 2022b). The uranium isotope ratios

560 ($\delta^{34}\text{U}$) from the Lombardy Basin record a negative excursion of 0.7 ‰, indicating a 30-fold
561 increase in anoxic seafloor area in a span of ~45 kyr at the Triassic/Jurassic boundary (Jost et al.,
562 2017a). Anoxia-driven isotopic excursions have also been interpreted from carbonate-associated
563 sulphate ($\delta^{34}\text{S}$) isotope ratios (He et al., 2020). The evidence of photic zone euxinia in shallow-
564 marine environments across the Triassic/Jurassic boundary has also been reported from changes
565 in nitrogen isotope ratios and fossil biomarker studies from Tethyan and Panthalassic sections
566 (Richoz et al., 2012; Jaraula et al., 2013; Kasprak et al., 2015; Fujisaki et al., 2020). The reducing
567 redox conditions remained prevalent in global oceans through the middle-late Hettangian with first
568 signs of shifts towards oxic environments occurring in the early Sinemurian (Richoz et al., 2012;
569 Ritterbush et al., 2015b; Thibodeau et al., 2016).

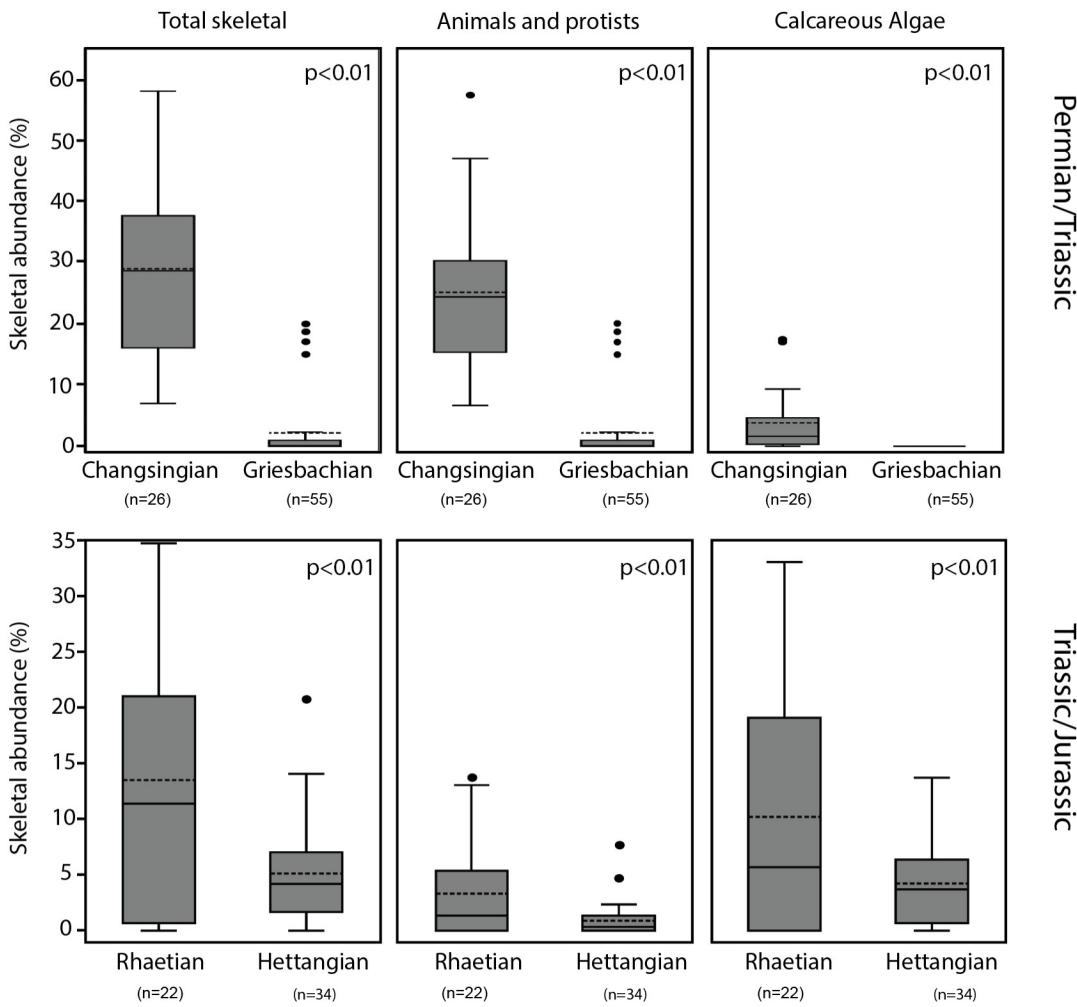
570 The fluctuating local redox conditions across the Triassic/Jurassic boundary and their close
571 correspondence with skeletal abundance indicate a connection between water-column anoxia and
572 the reduction in benthic faunal abundance across the extinction boundary. The fluctuating local
573 redox conditions well corresponded to changes in skeletal abundance, indicating water column
574 anoxia as a candidate controlling the benthic faunal abundance through the restriction of aerobic
575 habitat (Fig. 10). However, an overarching control, such as climate, which drove increased anoxia
576 in the Jurassic, could also have shaped variation in skeletal abundance. For instance, interaction
577 between temperature and oxygen availability explain the biogeography of many living species
578 through their influences on the ratio of oxygen supply to metabolic demand (Deutsch et al., 2015).
579 In this latter case, redox conditions may not have been directly or entirely responsible for the
580 changes in skeletal abundance but may have remained correlated with skeletal changes due to
581 shared controls (Fig. 10).

582

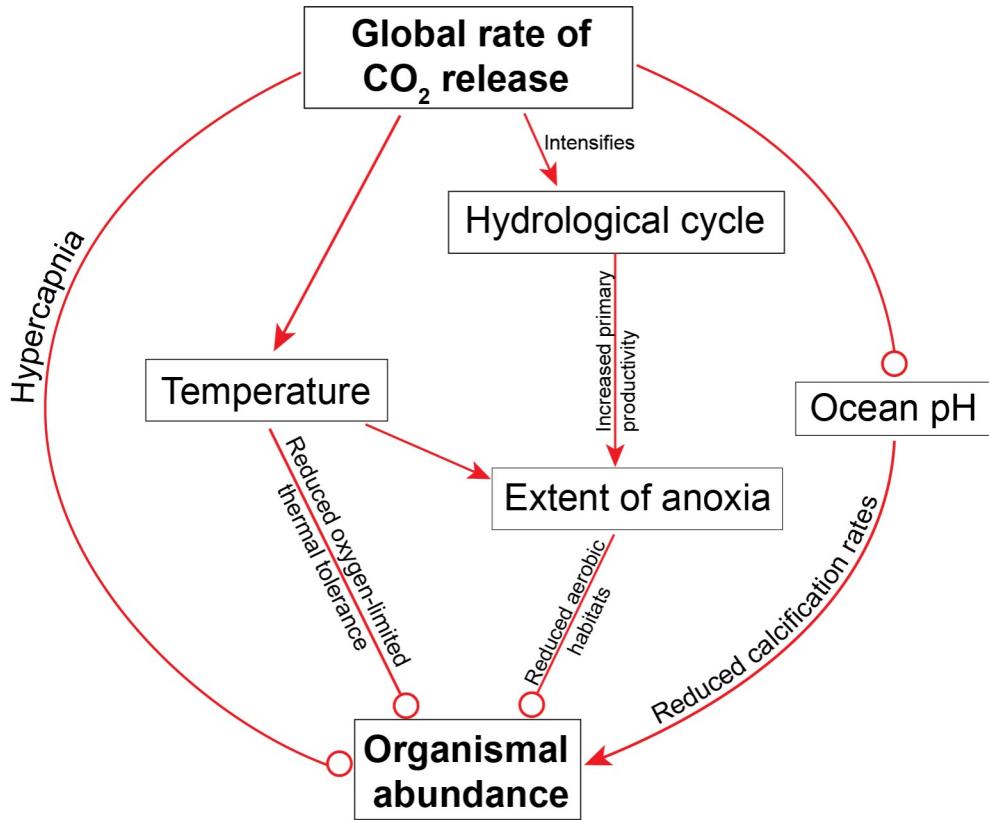
583 Geological evidence from the Triassic/Jurassic boundary interval supports the scenario involving
584 shared controls on abundance and redox conditions, for the study sites in the Lombardy Basin and
585 at Mt. Cefalo. Peritidal facies at Mt. Cefalo and shallow-marine facies including oolites in the
586 Lombardy Basin suggest that much of the deposition in the studied sections occurred above fair-
587 weather wave base and that the local water column would have been oxygenated by wind mixing
588 even if the I/Ca proxy recorded the mixing of more poorly oxygenated waters from deeper in the
589 basin due to the slower kinetics of iodide oxidation. Emplacement of the Central Atlantic
590 Magmatic Province (CAMP) at the Triassic/Jurassic boundary is a well-supported candidate for
591 the shared control on skeletal abundance and redox conditions. Carbon dioxide emissions from
592 CAMP led to an increase in atmospheric CO₂ (Bachan et al., 2012). Resultant warming (McElwain
593 et al., 1999; Ruhl and Kürschner, 2011) could have driven deoxygenation as evident from
594 observations in modern oceans (Levin, 2018). This warming would have increased the basal
595 metabolic demands of marine organisms and the elevated carbon dioxide concentrations and
596 reduced pH of marine waters would have added to physiological stresses (Pörtner et al., 2004;
597 Gazeau et al., 2013; Deutsch et al., 2015) (Fig. 10). The combined effects of multiple stressors,
598 especially warming and deoxygenation (Reddin et al., 2020), can potentially account for the lower
599 abundance of skeletal (and non-skeletal) animals in the aftermath of the mass extinction event.

600 Irrespective of the control, the geochemical proxy evidence indicates multiple episodes of carbon
601 dioxide release during the early Hettangian created deleterious environmental conditions by
602 sustaining high temperatures (McElwain et al., 1999; Huynh and Poulsen, 2005) and creating
603 unstable oceanic redox conditions in shallow-marine environments in Early Jurassic. The

604 prolonged hyperthermal and anoxic conditions could have reduced the carrying capacity of Early
 605 Jurassic marine ecosystems and delayed recovery until early Sinemurian time.



606
 607 *Figure 9: Box plots showing skeletal abundance variations in shallow marine environments across*
 608 *two major mass extinction boundaries: end-Permian mass extinction and end-Triassic mass*
 609 *extinction. In the top panel: the leftmost plot shows total skeletal abundance change across*
 610 *Changsingian (Upper Permian)- Griesbachian (Lower Triassic) boundary. The plots in the center*
 611 *and right show a breakdown of total skeletal abundance in animals and protists, and calcareous*
 612 *algae, respectively. The lower panel from left to right: total skeletal, animal & protists and*
 613 *calcareous algae abundance variations across the Rhaetian (Upper Triassic) - Hettangian (Lower*
 614 *Jurassic) boundary. The data for the end-Permian mass extinction was obtained from Payne et al.*
 615 *(2006). “n” indicates the number of thin sections in each category.*



616

617 *Figure 10: A systems analysis model showing the ocean biogeochemical changes associated with*
 618 *CAMP volcanism and its relationship with the Triassic/Jurassic biotic crisis. The parameters*
 619 *connected by arrows indicate a positive coupling between them and a negative coupling between*
 620 *those connected by open circles. For example, physiological effects due to increase in pCO₂ will*
 621 *lead to a decrease in skeletal abundance indicating an inverse relationship. Thus, this feedback is*
 622 *represented by an open circle.*

623

624 6 Conclusions

625 In this study, we measured skeletal abundance and I/Ca ratios in marine limestones in four Tethyan
 626 sections to reconstruct changes in faunal abundance and redox conditions across the
 627 Triassic/Jurassic boundary. We find evidence of a significant reduction in skeletal abundance
 628 across the end-Triassic extinction event that is reproducible among sections and cannot be
 629 explained by changes in depositional environment. I/Ca ratios indicate that the drop in skeletal
 630 abundance across the Triassic/Jurassic boundary was accompanied by an expansion of reducing
 631 conditions in shallow-marine environments in the regions of deposition. The eventual increase in
 632 skeletal abundance in the late Hettangian-Sinemurian is also associated with an increase in I/Ca.
 633 This correlation between skeletal abundance and I/Ca ratio indicates that the prevalence of dysoxic
 634 to anoxic environments during the early Hettangian was an important factor contributing to the
 635 decline in skeletal abundance. We hypothesize that physiological constraints posed by high
 636 oceanic temperatures and changing ocean pH resulting from carbon dioxide release during CAMP

637 volcanism acted in tandem with reduced oxygen availability to limit the abundance of skeletal
638 benthic organisms in Hettangian ecosystems.

639 Changes in abundance of benthic organisms, which are critical to understanding energy flow in
640 ecosystems, have remained among the most difficult to assess across mass extinctions. Studies like
641 this one provide a novel perspective for understanding the ecological severity of mass extinctions,
642 complementary to taxonomic diversity, counts of ecological modes of life, and food-web
643 reconstructions. They also provide a different perspective in directly assessing effects of ongoing,
644 human-induced mass rarity currently underway in modern oceans (Hull, 2015).

645
646

647 **References**

648 Aberhan, M., and Kiessling, W., 2015, Persistent ecological shifts in marine molluscan
649 assemblages across the end-Cretaceous mass extinction: *Proceedings of the National
650 Academy of Sciences*, v. 112, p. 7207–7212, doi:10.1073/pnas.1422248112.

651 Atkinson, J.W., and Wignall, P.B., 2019, How quick was marine recovery after the end-Triassic
652 mass extinction and what role did anoxia play? *Palaeogeography, Palaeoclimatology,
653 Palaeoecology*, v. 528, p. 99–119, doi:10.1016/j.palaeo.2019.05.011.

654 Atkinson, J.W., Wignall, P.B., Morton, J.D., and Aze, T., 2019, Body size changes in bivalves of
655 the family Limidae in the aftermath of the end-Triassic mass extinction: the Brobdingnag
656 effect: *Palaeontology*, v. 62, p. 561–582.

657 Bachan, A., and Payne, J.L., 2016, Modelling the impact of pulsed CAMP volcanism on pCO₂
658 and δ₁₃C across the Triassic-Jurassic transition: *Geological Magazine*, v. 153, p. 252–270,
659 doi:10.1017/S0016756815000126.

660 Bachan, A., Van De Shootbrugge, B., Fiebig, J., McRoberts, C.A., Ciarapica, G., and Payne,
661 J.L., 2012, Carbon cycle dynamics following the end-Triassic mass extinction: Constraints
662 from paired δ₁₃C_{carb} and δ₁₃C_{org} records: *Geochemistry, Geophysics, Geosystems*, v. 13,
663 p. 1–24, doi:10.1029/2012GC004150.

664 Bambach, R.K., 2006, Phanerozoic biodiversity mass extinctions: *Annu. Rev. Earth Planet. Sci.*,
665 v. 34, p. 127–155.

666 Bambach, R.K., Knoll, A.H., and Sepkoski, J.J., 2002, Anatomical and ecological constraints on
667 Phanerozoic animal diversity in the marine realm: *Proceedings of the National Academy of
668 Sciences*, v. 99, p. 6854–6859.

669 Barattolo, F., and Romano, R., 2005, Shallow carbonate platform bioevents during the Upper
670 Triassic-Lower Jurassic: An evolutive interpretation: *Bollettino della Societa Geologica
671 Italiana*, v. 124, p. 123–142.

672 Barnosky, A.D. et al., 2012, Approaching a state shift in Earth's biosphere: *Nature*, v. 486, p.
673 52–58.

674 Barras, C.G., and Twitchett, R.J., 2007, Response of the marine infauna to Triassic-Jurassic
675 environmental change: Ichnological data from southern England: *Palaeogeography,
676 Palaeoclimatology, Palaeoecology*, v. 244, p. 223–241, doi:10.1016/j.palaeo.2006.06.040.

677 Berra, F., 2012, Sea-level fall, carbonate production, rainy days: How do they relate? Insight
678 from Triassic carbonate platforms (Western Tethys, Southern Alps, Italy): *Geology*, v. 40,
679 p. 271–274, doi:10.1130/G32803.1.

680 Berra, F., Jadoul, F., and Anelli, A., 2010, Environmental control on the end of the Dolomia
681 Principale/Hauptdolomit depositional system in the central Alps: Coupling sea-level and
682 climate changes: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 290, p. 138–150,
683 doi:https://doi.org/10.1016/j.palaeo.2009.06.037.

684 Bertotti, G., Siletto, G.B., and Spalla, M.I., 1993, Deformation and metamorphism associated
685 with crustal rifting: The Permian to Liassic evolution of the Lake Lugano-Lake Como area
686 (Southern Alps): *Tectonophysics*, v. 226, p. 271–284, doi:[https://doi.org/10.1016/0040-6871\(93\)90122-Z](https://doi.org/10.1016/0040-6871(93)90122-Z).

688 Blackburn, T.J., Olsen, P.E., Bowring, S.A., McLean, N.M., Kent, D. V, Puffer, J., McHone, G.,
689 Rasbury, E.T., and Et-Touhami, M., 2013, Zircon U-Pb geochronology links the end-
690 Triassic extinction with the Central Atlantic Magmatic Province: *Science*, v. 340, p. 941–
691 945.

692 Blois, J.L., Zarnetske, P.L., Fitzpatrick, M.C., and Finnegan, S., 2013, Climate change and the
693 past, present, and future of biotic interactions: *Science*, v. 341, p. 499–504.

694 Bond, D.P.G., and Grasby, S.E., 2017, On the causes of mass extinctions: *Palaeogeography,
695 Palaeoclimatology, Palaeoecology*, v. 478, p. 3–29, doi:10.1016/j.palaeo.2016.11.005.

696 Bowman, C.N., Lindskog, A., Kozik, N.P., Richbourg, C.G., Owens, J.D., and Young, S.A.,
697 2020, Integrated sedimentary, biotic, and paleoredox dynamics from multiple localities in
698 southern Laurentia during the late Silurian (Ludfordian) extinction event: *Palaeogeography,
699 Palaeoclimatology, Palaeoecology*, v. 553, doi:10.1016/j.palaeo.2020.109799.

700 Brand, U., and Veizer, J., 1980, Chemical diagenesis of a multicomponent carbonate system; 1,
701 Trace elements: *Journal of Sedimentary Research*, v. 50, p. 1219–1236,
702 doi:10.1306/212F7BB7-2B24-11D7-8648000102C1865D.

703 Brandano, M., Corda, L., Tomassetti, L., and Tagliavento, M., 2016, Frequency analysis across
704 the drowning of a Lower Jurassic carbonate platform: The Calcare Massiccio Formation
705 (Apennines, Italy): *Marine and Petroleum Geology*, v. 78, p. 606–620,
706 doi:<https://doi.org/10.1016/j.marpetgeo.2016.09.013>.

707 Brenchley, P.J., Marshall, J.D., and Underwood, C.J., 2001, Do all mass extinctions represent an
708 ecological crisis? Evidence from the Late Ordovician: *Geological Journal*, v. 36, p. 329–
709 340.

710 Brosse, M., Bucher, H., Baud, A., Frisk, Å.M., Goudemand, N., Hagdorn, H., Nützel, A., Ware,
711 D., and Hautmann, M., 2019, New data from Oman indicate benthic high biomass
712 productivity coupled with low taxonomic diversity in the aftermath of the Permian–
713 Triassic Boundary mass extinction: *Lethaia*, v. 52, p. 165–187.

714 Bucur, I.I., 1999, Stratigraphic significance of some skeletal algae (Dasycladales, Caulerpales) of
715 the Phanerozoic: Depositional episodes and bioevents, p. 53–104.

716 Chance, R., Baker, A.R., Carpenter, L., and Jickells, T.D., 2014, The distribution of iodide at the
717 sea surface: *Environ. Sci.: Processes Impacts*, v. 16, p. 1841–1859,
718 doi:10.1039/C4EM00139G.

719 Christie, M., Holland, S.M., and Bush, A.M., 2013, Contrasting the ecological and taxonomic
720 consequences of extinction: *Paleobiology*, v. 39, p. 538–559.

721 Ciarapica, G., and Passeri, L., 2005, Late Triassic and Early Jurassic sedimentary evolution of

722 the Northern Apennines: An overview: *Bollettino della Societa Geologica Italiana*, v. 124,
723 p. 189–201.

724 Cirilli, S., Buratti, N., Gugliotti, L., and Frix, A., 2015, Palynostratigraphy and palynofacies of
725 the Upper Triassic Streppenosa Formation (SE Sicily, Italy) and inference on the main
726 controlling factors in the organic rich shale deposition: *Review of Palaeobotany and*
727 *Palynology*, v. 218, p. 67–79, doi:<https://doi.org/10.1016/j.revpalbo.2014.10.009>.

728 Corsetti, F.A. et al., 2015, Investigating the Paleoecological Consequences of Supercontinent
729 Breakup: Sponges Clean Up in the Early Jurassic: The Sedimentary Record, v. 13, p. 4–10,
730 doi:[10.2110/sedred.2015.2.4](https://doi.org/10.2110/sedred.2015.2.4).

731 Coskun Tunaboylu, B., Altiner, D., Isintek, I., and Demirci, D., 2014, Foraminiferal
732 biostratigraphy and sequence stratigraphy of peritidal carbonates at the Triassic-Jurassic
733 boundary (Karaburun Peninsula, Western Turkey): *Journal of Asian Earth Sciences*, v. 90,
734 p. 61–76, doi:[10.1016/j.jseaes.2014.04.015](https://doi.org/10.1016/j.jseaes.2014.04.015).

735 Cribb, A.T., and Bottjer, D.J., 2020, Complex marine bioturbation ecosystem engineering
736 behaviors persisted in the wake of the end-Permian mass extinction: *Scientific reports*, v.
737 10, p. 1–8.

738 Delecat, S., and Reitner, J., 2005, Sponge communities from the Lower Liassic of Adnet
739 (Northern Calcareous Alps, Austria): *Facies*, v. 51, p. 385–404, doi:[10.1007/s10347-005-0045-x](https://doi.org/10.1007/s10347-005-0045-x).

741 Deutsch, C., Ferrel, A., Seibel, B., Pörtner, H.-O., and Huey, R.B., 2015, Climate change
742 tightens a metabolic constraint on marine habitats: *Science*, v. 348, p. 1132–1135,
743 doi:[10.1126/science.aaa1605](https://doi.org/10.1126/science.aaa1605).

744 Di Stefano, P., Favara, R., Luzio, D., Renda, P., Cacciatore, M.S., Calò, M., Napoli, G., Parisi,
745 L., Todaro, S., and Zarcone, G., 2015, A regional-scale discontinuity in western Sicily
746 revealed by a multidisciplinary approach: A new piece for understanding the geodynamic
747 puzzle of the southern Mediterranean: *Tectonics*, v. 34, p. 2067–2085,
748 doi:[10.1002/2014TC003759](https://doi.org/10.1002/2014TC003759).

749 Diaz, R.J., and Rosenberg, R., 2008, Spreading dead zones and consequences for marine
750 ecosystems: *science*, v. 321, p. 926–929.

751 Dineen, A.A., Roopnarine, P.D., and Fraiser, M.L., 2019, Ecological continuity and
752 transformation after the Permo-Triassic mass extinction in northeastern Panthalassa:
753 *Biology Letters*, v. 15, doi:[10.1098/rsbl.2018.0902](https://doi.org/10.1098/rsbl.2018.0902).

754 Droser, M.L., Bottjer, D.J., Sheehan, P.M., and McGhee Jr, G.R., 2000, Decoupling of
755 taxonomic and ecologic severity of Phanerozoic marine mass extinctions: *Geology*, v. 28, p.
756 675–678.

757 Droser, M.L., Jensen, S., and Gehling, J.G., 2002, Trace fossils and substrates of the terminal
758 Proterozoic-Cambrian transition: Implications for the record of early bilaterians and
759 sediment mixing: *Proceedings of the National Academy of Sciences*, v. 99, p. 12572–

760 12576, doi:10.1073/pnas.202322499.

761 Dunhill, A.M., Foster, W.J., Azaele, S., Sciberras, J., and Twitchett, R.J., 2018, Modelling
762 determinants of extinction across two Mesozoic hyperthermal events: *Proceedings of the*
763 *Royal Society B: Biological Sciences*, v. 285, doi:10.1098/rspb.2018.0404.

764 Emerson, S., Cranston, R.E., and Liss, P.S., 1979, Redox species in a reducing fjord: equilibrium
765 and kinetic considerations: *Deep Sea Research Part A. Oceanographic Research Papers*, v.
766 26, p. 859–878, doi:[https://doi.org/10.1016/0198-0149\(79\)90101-8](https://doi.org/10.1016/0198-0149(79)90101-8).
767 Bond, D.P.G., and
768 Grasby, S.E., 2017, On the causes of mass extinctions: *Palaeogeography,*
769 *Palaeoclimatology, Palaeoecology*, v. 478, p. 3–29, doi:10.1016/j.palaeo.2016.11.005.

770 Bowman, C.N., Lindskog, A., Kozik, N.P., Richbourg, C.G., Owens, J.D., and Young, S.A.,
771 2020, Integrated sedimentary, biotic, and paleoredox dynamics from multiple localities in
772 southern Laurentia during the late Silurian (Ludfordian) extinction event: *Palaeogeography,*
773 *Palaeoclimatology, Palaeoecology*, v. 553, doi:10.1016/j.palaeo.2020.109799.

774 Brand, U., and Veizer, J., 1980, Chemical diagenesis of a multicomponent carbonate system; 1,
775 Trace elements: *Journal of Sedimentary Research*, v. 50, p. 1219–1236,
776 doi:10.1306/212F7BB7-2B24-11D7-8648000102C1865D.

777 Deutsch, C., Ferrel, A., Seibel, B., Pörtner, H.-O., and Huey, R.B., 2015, Climate change
778 tightens a metabolic constraint on marine habitats: *Science*, v. 348, p. 1132–1135,
779 doi:10.1126/science.aaa1605.

780 Finnegan, S., Rasmussen, C.M., and Harper, D.A.T., 2016, Biogeographic and bathymetric
781 determinants of brachiopod extinction and survival during the Late Ordovician mass
782 extinction: *Proceedings of the Royal Society B: Biological Sciences*, v. 283, p. 20160007.

783 Gazeau, F., Parker, L.M., Comeau, S., Gattuso, J.-P., O'Connor, W.A., Martin, S., Pörtner, H.-
784 O., and Ross, P.M., 2013, Impacts of ocean acidification on marine shelled molluscs:
785 *Marine Biology*, v. 160, p. 2207–2245, doi:10.1007/s00227-013-2219-3.

786 He, T. et al., 2022a, Shallow ocean oxygen decline during the end-Triassic mass extinction:
787 *Global and Planetary Change*, v. 210, p. 103770,
788 doi:<https://doi.org/10.1016/j.gloplacha.2022.103770>.

789 He, T., Wignall, P.B., Newton, R.J., Atkinson, J.W., Keeling, J.F.J., Xiong, Y., and Poulton,
790 S.W., 2022b, Extensive marine anoxia in the European epicontinental sea during the end-
791 Triassic mass extinction: *Global and Planetary Change*, v. 210, p. 103771,
792 doi:<https://doi.org/10.1016/j.gloplacha.2022.103771>.

793 Huynh, T.T., and Poulsen, C.J., 2005, Rising atmospheric CO₂ as a possible trigger for the end-
794 Triassic mass extinction: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 217, p.
795 223–242, doi:<https://doi.org/10.1016/j.palaeo.2004.12.004>.

796 Levin, L.A., 2018, Manifestation, Drivers, and Emergence of Open Ocean Deoxygenation:
797 *Annual Review of Marine Science*, v. 10, p. 229–260, doi:10.1146/annurev-marine-121916-
798 063359.

798 Lu, Z., Lu, W., Rickaby, R.E.M., and Thomas, E., 2020, Earth History of Oxygen and the
799 iprOxy: Cambridge University Press, Elements in Geochemical Tracers in Earth System
800 Science, doi:10.1017/9781108688604.

801 McElwain, J.C., Beerling, D.J., and Woodward, F.I., 1999, Fossil Plants and Global Warming at
802 the Triassic-Jurassic Boundary: *Science*, v. 285, p. 1386–1390,
803 doi:10.1126/science.285.5432.1386.

804 Pörtner, H.O., Langenbuch, M., and Reipschläger, A., 2004, Biological impact of elevated ocean
805 CO₂ concentrations: lessons from animal physiology and earth history: *Journal of
806 Oceanography*, v. 60, p. 705–718.

807 Flügel, E., 2010, Quantitative Microfacies Analysis, *in* Microfacies of Carbonate Rocks:
808 Analysis, Interpretation and Application, Berlin, Heidelberg, Springer Berlin Heidelberg, p.
809 243–266, doi:10.1007/978-3-642-03796-2_6.

810 Fox, C.P., Cui, X., Whiteside, J.H., Olsen, P.E., Summons, R.E., and Grice, K., 2020, Molecular
811 and isotopic evidence reveals the end-Triassic carbon isotope excursion is not from massive
812 exogenous light carbon: *Proceedings of the National Academy of Sciences of the United
813 States of America*, v. 117, p. 30171–30178, doi:10.1073/pnas.1917661117.

814 Fujisaki, W., Fukami, Y., Matsui, Y., Sato, T., Sawaki, Y., and Suzuki, K., 2020, Redox
815 conditions and nitrogen cycling during the Triassic-Jurassic transition: A new perspective
816 from the mid-Panthalassa: *Earth-Science Reviews*, v. 204,
817 doi:10.1016/j.earscirev.2020.103173.

818 Gaetani, M., 1970, Faune Hettangiane della parte orientale della provincia di Bergamo: *Riv.
819 Italiana Paleontologia Stratigrafia*, v. 76, p. 355–442.

820 Galli, M.T., Jadoul, F., Bernasconi, S.M., Cirilli, S., and Weissert, H., 2007, Stratigraphy and
821 palaeoenvironmental analysis of the Triassic-Jurassic transition in the western Southern
822 Alps (Northern Italy): *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 244, p. 52–
823 70, doi:10.1016/j.palaeo.2006.06.023.

824 Galli, M.T., Jadoul, F., Bernasconi, S.M., and Weissert, H., 2005, Anomalies in global carbon
825 cycling and extinction at the Triassic/Jurassic boundary: evidence from a marine C-isotope
826 record: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 216, p. 203–214,
827 doi:<https://doi.org/10.1016/j.palaeo.2004.11.009>.

828 Gazeau, F., Parker, L.M., Comeau, S., Gattuso, J.-P., O'Connor, W.A., Martin, S., Pörtner, H.-
829 O., and Ross, P.M., 2013, Impacts of ocean acidification on marine shelled molluscs:
830 *Marine Biology*, v. 160, p. 2207–2245, doi:10.1007/s00227-013-2219-3.

831 Greene, S.E., Bottjer, D.J., Hagdorn, H., and Zonneveld, J.-P., 2011, The Mesozoic return of
832 Paleozoic faunal constituents: A decoupling of taxonomic and ecological dominance during
833 the recovery from the end-Permian mass extinction: *Palaeogeography, Palaeoclimatology,
834 Palaeoecology*, v. 308, p. 224–232.

835 Haas, J., Kovács, S., Krystyn, L., and Lein, R., 1995, Significance of Late Permian-Triassic

836 facies zones in terrane reconstructions in the Alpine-North Pannonian domain:
837 *Tectonophysics*, v. 242, p. 19–40, doi:[https://doi.org/10.1016/0040-1951\(94\)00157-5](https://doi.org/10.1016/0040-1951(94)00157-5).

838 Häntzschel, W., and Reineck, H.E., 1968, Fazies-Untersuchungen im Hettangium von Helmstedt
839 (Niedersachsen): Mitteilungen aus dem Geologischen Staatsinstitut in Hamburg, v. 37, p. 5–
840 39.

841 Hardisty, D.S. et al., 2017, Perspectives on Proterozoic surface ocean redox from iodine contents
842 in ancient and recent carbonate: *Earth and Planetary Science Letters*, v. 463, p. 159–170,
843 doi:<https://doi.org/10.1016/j.epsl.2017.01.032>.

844 Harnik, P.G., Simpson, C., and Payne, J.L., 2012, Long-term differences in extinction risk
845 among the seven forms of rarity: *Proceedings of the Royal Society B: Biological Sciences*,
846 v. 279, p. 4969–4976.

847 Hautmann, M., 2004, Effect of end-Triassic CO₂ maximum on carbonate sedimentation and
848 marine mass extinction: *Facies*, v. 50, p. 257–261.

849 He, T. et al., 2020, An enormous sulfur isotope excursion indicates marine anoxia during the
850 end-Triassic mass extinction: *Science Advances*, v. 6, p. eabb6704,
851 doi:[10.1126/sciadv.abb6704](https://doi.org/10.1126/sciadv.abb6704).

852 He, T. et al., 2022a, Shallow ocean oxygen decline during the end-Triassic mass extinction:
853 *Global and Planetary Change*, v. 210, p. 103770,
854 doi:<https://doi.org/10.1016/j.gloplacha.2022.103770>.

855 He, T., Wignall, P.B., Newton, R.J., Atkinson, J.W., Keeling, J.F.J., Xiong, Y., and Poulton,
856 S.W., 2022b, Extensive marine anoxia in the European epicontinental sea during the end-
857 Triassic mass extinction: *Global and Planetary Change*, v. 210, p. 103771,
858 doi:<https://doi.org/10.1016/j.gloplacha.2022.103771>.

859 Hull, P.M., 2017, Emergence of modern marine ecosystems: *Current Biology*, v. 27, p. R466–
860 R469, doi:<https://doi.org/10.1016/j.cub.2017.04.041>.

861 Hull, P., 2015, Life in the aftermath of mass extinctions: *Current Biology*, v. 25, p. R941–R952,
862 doi:[10.1016/j.cub.2015.08.053](https://doi.org/10.1016/j.cub.2015.08.053).

863 Hull, P.M., and Darroch, S.A.F., 2013, Mass extinctions and the structure and function of
864 ecosystems: *The Paleontological Society Papers*, v. 19, p. 115–156.

865 Hull, P.M., Norris, R.D., Bralower, T.J., and Schueth, J.D., 2011, A role for chance in marine
866 recovery from the end-Cretaceous extinction: *Nature Geoscience*, v. 4, p. 856–860.

867 Huynh, T.T., and Poulsen, C.J., 2005, Rising atmospheric CO₂ as a possible trigger for the end-
868 Triassic mass extinction: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 217, p.
869 223–242, doi:<https://doi.org/10.1016/j.palaeo.2004.12.004>.

870 Iannace, A., and Zamparelli, V., 2002, Upper Triassic platform margin biofacies and the
871 paleogeography of Southern Apennines: *Palaeogeography, Palaeoclimatology,*
872 *Palaeoecology*, v. 179, p. 1–18, doi:[https://doi.org/10.1016/S0031-0182\(01\)00362-5](https://doi.org/10.1016/S0031-0182(01)00362-5).

873 Jacobsen, N.D., Twitchett, R.J., and Krystyn, L., 2011, Palaeoecological methods for assessing
874 marine ecosystem recovery following the Late Permian mass extinction event:
875 *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 308, p. 200–212,
876 doi:10.1016/j.palaeo.2010.04.024.

877 Jadoul, F., Berra, F., and Frisia, S., 1992, Stratigraphic and paleogeographic evolution of a
878 carbonate platform in an extensional tectonic regime: the example of the Dolomia
879 Principale in Lombardy (Italy): *Rivista Italiana di Paleontologia e Stratigrafia*, v. 98, p. 29–
880 44, doi:10.13130/2039-4942/8939.

881 Jadoul, F., and Galli, M.T., 2008, The hettangian shallow water carbonates after the
882 triassic/jurassic biocalcification crisis: The albenza formation in the Western Southern
883 ALPS: *Rivista Italiana di Paleontologia e Stratigrafia*, v. 114, p. 453–470,
884 doi:10.13130/2039-4942/5911.

885 Jadoul, F., Galli, M.T., Berra, F., Cirilli, S., Ronchi, P., and Paganoni, A.M., 2004, The Late
886 Triassic-Early Jurassic of the Lombardy Basin: stratigraphy, palaeogeography and
887 palaeontology. Field guide book excursion P86.

888 Jadoul, F., Galli, M.T., Calabrese, L., and Gnaccolini, M., 2005, Stratigraphy of Rhaetian to
889 lower Sinemurian carbonate platforms in western Lombardy (southern alps, Italy):
890 paleogeographic implications: v. 111, p. 285–303.

891 Jaraula, C.M.B., Grice, K., Twitchett, R.J., Böttcher, M.E., LeMetayer, P., Dastidar, A.G., and
892 Opazo, L.F., 2013, Elevated pCO₂ leading to Late Triassic extinction, persistent photic
893 zone euxinia, and rising sea levels: *Geology*, v. 41, p. 955–958, doi:10.1130/G34183.1.

894 Jost, A.B., Bachan, A., van de Schootbrugge, B., Brown, S.T., DePaolo, D.J., and Payne, J.L.,
895 2017a, Additive effects of acidification and mineralogy on calcium isotopes in
896 Triassic/Jurassic boundary limestones: *Geochemistry, Geophysics, Geosystems*, v. 18, p.
897 113–124, doi:10.1002/2016GC006724.

898 Jost, A.B., Bachan, A., van de Schootbrugge, B., Lau, K. V., Weaver, K.L., Maher, K., and
899 Payne, J.L., 2017b, Uranium isotope evidence for an expansion of marine anoxia during the
900 end-Triassic extinction: *Geochemistry, Geophysics, Geosystems*, v. 18, p. 3093–3108,
901 doi:10.1002/2017GC006941.

902 Kasprak, A.H., Sepúlveda, J., Price-Waldman, R., Williford, K.H., Schoepfer, S.D., Haggart,
903 J.W., Ward, P.D., Summons, R.E., and Whiteside, J.H., 2015, Episodic photic zone euxinia
904 in the northeastern Panthalassic Ocean during the end-Triassic extinction: *Geology*, v. 43, p.
905 307–310, doi:10.1130/G36371.1.

906 Kiessling, W., and Aberhan, M., 2007, Environmental determinants of marine benthic
907 biodiversity dynamics through Triassic–Jurassic time: *Paleobiology*, v. 33, p. 414–434.

908 Kiessling, W., Aberhan, M., Brenneis, B., and Wagner, P.J., 2007, Extinction trajectories of
909 benthic organisms across the Triassic–Jurassic boundary: *Palaeogeography,*
910 *Palaeoclimatology, Palaeoecology*, v. 244, p. 201–222.

911 Knaust, D., 2010, The end-Permian mass extinction and its aftermath on an equatorial carbonate
912 platform: insights from ichnology: *Terra Nova*, v. 22, p. 195–202.

913 Knoll, A.H., Bambach, R.K., Canfield, D.E., and Grotzinger, J.P., 1996, Comparative Earth
914 history and Late Permian mass extinction: *Science*, v. 273, p. 452–457.

915 Knoll, A.H., Bambach, R.K., Payne, J.L., Pruss, S., and Fischer, W.W., 2007a, Paleophysiology
916 and end-Permian mass extinction: *Earth and Planetary Science Letters*, v. 256, p. 295–313,
917 doi:10.1016/j.epsl.2007.02.018.

918 Knoll, A.H., Summons, R.E., Waldbauer, J.R., and Zumberge, J.E., 2007b, CHAPTER 8 - The
919 Geological Succession of Primary Producers in the Oceans, in Falkowski, P.G. and Knoll,
920 A.H. eds., *Evolution of Primary Producers in the Sea*, Burlington, Academic Press, p. 133–
921 163, doi:<https://doi.org/10.1016/B978-012370518-1/50009-6>.

922 Lakew, T., 1994, Diagenesis of a Rhaetian Patch Reef (Lombardian Basin, southern Alps):
923 *Rivista Italiana di Paleontologia e Stratigrafia*, v. 100.

924 Lakew, T., 1990, Microfacies and cyclic sedimentation of the Upper Triassic (Rhaetian) Calcare
925 di zu (Southern Alps): *Facies*, v. 22, p. 187–231, doi:10.1007/BF02536952.

926 Larina, E., Bottjer, D.J., Corsetti, F.A., Zonneveld, J.P., Celestian, A.J., and Bailey, J. V., 2019,
927 Uppermost Triassic phosphorites from Williston Lake, Canada: link to fluctuating euxinic-928
anoxic conditions in northeastern Panthalassa before the end-Triassic mass extinction:
929 *Scientific Reports*, v. 9, doi:10.1038/s41598-019-55162-2.

930 Levin, L.A., 2018, Manifestation, Drivers, and Emergence of Open Ocean Deoxygenation:
931 *Annual Review of Marine Science*, v. 10, p. 229–260, doi:10.1146/annurev-marine-121916-
932 063359.

933 Lu, W. et al., 2018, Late inception of a resiliently oxygenated upper ocean: *Science*, v. 361, p.
934 174–177, doi:10.1126/science.aar5372.

935 Lu, Z., Hoogakker, B.A.A., Hillenbrand, C.-D., Zhou, X., Thomas, E., Gutschess, K.M., Lu, W.,
936 Jones, L., and Rickaby, R.E.M., 2016, Oxygen depletion recorded in upper waters of the
937 glacial Southern Ocean: *Nature Communications*, v. 7, p. 11146,
938 doi:10.1038/ncomms11146.

939 Lu, Z., Jenkyns, H.C., and Rickaby, R.E.M., 2010, Iodine to calcium ratios in marine carbonate
940 as a paleo-redox proxy during oceanic anoxic events: *Geology*, v. 38, p. 1107–1110,
941 doi:10.1130/G31145.1.

942 Lu, Z., Lu, W., Rickaby, R.E.M., and Thomas, E., 2020, *Earth History of Oxygen and the
943 iprOxy: Cambridge University Press, Elements in Geochemical Tracers in Earth System
944 Science*, doi:10.1017/9781108688604.

945 Mancinelli, A., Chiocchini, M., Chiocchini, R.A., and Romano, A., 2005, Biostratigraphy of
946 Upper Triassic-Lower Jurassic carbonate platform sediments of the central-southern
947 Apennines (Italy): *Rivista Italiana di Paleontologia e Stratigrafia*, v. 111, p. 271–283,
948 doi:10.13130/2039-4942/6314.

949 Mariotti, G., Pruss, S.B., Perron, J.T., and Bosak, T., 2014, Microbial shaping of sedimentary
950 wrinkle structures: *Nature Geoscience*, v. 7, p. 736–740, doi:10.1038/ngeo2229.

951 Mata, S.A., and Bottjer, D.J., 2009, The paleoenvironmental distribution of Phanerozoic wrinkle
952 structures: *Earth-Science Reviews*, v. 96, p. 181–195,
953 doi:<https://doi.org/10.1016/j.earscirev.2009.06.001>.

954 McCauley, D.J., Pinsky, M.L., Palumbi, S.R., Estes, J.A., Joyce, F.H., and Warner, R.R., 2015,
955 Marine defaunation: animal loss in the global ocean: *Science*, v. 347.

956 McElwain, J.C., Beerling, D.J., and Woodward, F.I., 1999, Fossil Plants and Global Warming at
957 the Triassic-Jurassic Boundary: *Science*, v. 285, p. 1386–1390,
958 doi:10.1126/science.285.5432.1386.

959 McGhee, G.R., Clapham, M.E., Sheehan, P.M., Bottjer, D.J., and Droser, M.L., 2013, A new
960 ecological-severity ranking of major Phanerozoic biodiversity crises: *Palaeogeography,
961 Palaeoclimatology, Palaeoecology*, v. 370, p. 260–270, doi:10.1016/j.palaeo.2012.12.019.

962 McGhee Jr, G.R., Sheehan, P.M., Bottjer, D.J., and Droser, M.L., 2004, Ecological ranking of
963 Phanerozoic biodiversity crises: ecological and taxonomic severities are decoupled:
964 *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 211, p. 289–297.

965 McRoberts, C.A., 1994, The Triassic-Jurassic ecostratigraphic transition in the Lombardian Alps,
966 Italy: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 110, p. 145–166,
967 doi:10.1016/0031-0182(94)90114-7.

968 McRoberts, C.A., Furrer, H., and Jones, D.S., 1997, Palaeoenvironmental interpretation of a
969 Triassic-Jurassic boundary section from Western Austria based on palaeoecological and
970 geochemical data: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 136, p. 79–95,
971 doi:[https://doi.org/10.1016/S0031-0182\(97\)00074-6](https://doi.org/10.1016/S0031-0182(97)00074-6).

972 Muttoni, G., Kent, D. V, Jadoul, F., Olsen, P.E., Rigo, M., Galli, M.T., and Nicora, A., 2010,
973 Rhaetian magneto-biostratigraphy from the Southern Alps (Italy): Constraints on Triassic
974 chronology: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 285, p. 1–16,
975 doi:<https://doi.org/10.1016/j.palaeo.2009.10.014>.

976 Pálfy, J. et al., 2007, Triassic–Jurassic boundary events inferred from integrated stratigraphy of
977 the Csővár section, Hungary: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 244,
978 p. 11–33, doi:<https://doi.org/10.1016/j.palaeo.2006.06.021>.

979 Payne, J.L., 2005, Evolutionary dynamics of gastropod size across the end-Permian extinction
980 and through the Triassic recovery interval: *Paleobiology*, v. 31, p. 269–290,
981 doi:10.1666/0094-8373(2005)031[0269:EDOGSA]2.0.CO;2.

982 Payne, J.L., Lehrmann, D.J., Wei, J., and Knoll, A.H., 2006, The pattern and timing of biotic
983 recovery from the end-Permian extinction on the Great Bank of Guizhou, Guizhou
984 province, China: *Palaios*, v. 21, p. 63–85, doi:10.2110/palo.2005.p05-12p.

985 Payne, J.L., and Van De Schootbrugge, B., 2007, CHAPTER 9 - Life in Triassic Oceans: Links
986 Between Planktonic and Benthic Recovery and Radiation, *in* Falkowski, P.G. and Knoll,

987 A.H. eds., *Evolution of Primary Producers in the Sea*, Burlington, Academic Press, p. 165–
988 189, doi:<https://doi.org/10.1016/B978-012370518-1/50010-2>.

989 Penn, J.L., Deutsch, C., Payne, J.L., and Sperling, E.A., 2018b, Temperature-dependent hypoxia
990 explains biogeography and severity of end-Permian marine mass extinction: *Science*, v.
991 362, doi:[10.1126/science.aat1327](https://doi.org/10.1126/science.aat1327).

992 Peterffy, O., Calner, M., and Vajda, V., 2016, Early Jurassic microbial mats—A potential
993 response to reduced biotic activity in the aftermath of the end-Triassic mass extinction
994 event: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 464, p. 76–85,
995 doi:<https://doi.org/10.1016/j.palaeo.2015.12.024>.

996 Pörtner, H.O., Langenbuch, M., and Reipschläger, A., 2004, Biological impact of elevated ocean
997 CO₂ concentrations: lessons from animal physiology and earth history: *Journal of*
998 *Oceanography*, v. 60, p. 705–718.

999 Pruss, S., Fraiser, M., and Bottjer, D.J., 2004, Proliferation of Early Triassic wrinkle structures:
1000 Implications for environmental stress following the end-Permian mass extinction: *Geology*,
1001 v. 32, p. 461–464, doi:[10.1130/G20354.1](https://doi.org/10.1130/G20354.1).

1002 Raup, D.M., and Sepkoski, J.J., 1982, Mass extinctions in the marine fossil record: *Science*, v.
1003 215, p. 1501–1503.

1004 Reddin, C.J., Nätscher, P.S., Kocsis, Á.T., Pörtner, H.O., and Kiessling, W., 2020, Marine clade
1005 sensitivities to climate change conform across timescales: *Nature Climate Change*, v. 10, p.
1006 249–253, doi:[10.1038/s41558-020-0690-7](https://doi.org/10.1038/s41558-020-0690-7).

1007 Richoz, S. et al., 2012, Hydrogen sulphide poisoning of shallow seas following the end-Triassic
1008 extinction: *Nature Geoscience*, v. 5, p. 662–667, doi:[10.1038/ngeo1539](https://doi.org/10.1038/ngeo1539).

1009 Ritterbush, K.A., Bottjer, D.J., Corsetti, F.A., and Rosas, S., 2014, New evidence on the role of
1010 siliceous sponges in ecology and sedimentary facies development in eastern Panthalassa
1011 following the Triassic–Jurassic mass extinction: *PALAIOS*, v. 29, p. 652–668,
1012 doi:[10.2110/palo.2013.121](https://doi.org/10.2110/palo.2013.121).

1013 Ritterbush, K.A., Ibarra, Y., and Tackett, L.S., 2016, Post-extinction biofacies of the first
1014 carbonate ramp of the early Jurassic (Sinemurian) in NE Panthalassa (New York canyon,
1015 Nevada, USA): *PALAIOS*, v. 31, p. 141–160, doi:[10.2110/palo.2015.021](https://doi.org/10.2110/palo.2015.021).

1016 Ritterbush, K.A., Rosas, S., Corsetti, F.A., Bottjer, D.J., and West, A.J., 2015a, Andean sponges
1017 reveal long-term benthic ecosystem shifts following the end-Triassic mass extinction:
1018 *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 420, p. 193–209,
1019 doi:[10.1016/j.palaeo.2014.12.002](https://doi.org/10.1016/j.palaeo.2014.12.002).

1020 Romano, R., Masett, D., Carras, N., Barattolo, F., and Roghi, G., 2008, The triassic/jurassic
1021 boundary in a peritidal carbonate platform of the pelagonian domain: The mount messapion
1022 section (Chalkida, Greece): *Rivista Italiana di Paleontologia e Stratigrafia*, v. 114, p. 431–
1023 452, doi:[10.13130/2039-4942/5910](https://doi.org/10.13130/2039-4942/5910).

1024 Roopnarine, P.D., and Angielczyk, K.D., 2015, Community stability and selective extinction

1025 during the Permian-Triassic mass extinction: *Science*, v. 350, p. 90–93.

1026 Ruhl, M., and Kürschner, W.M., 2011, Multiple phases of carbon cycle disturbance from large
1027 igneous province formation at the Triassic-Jurassic transition: *Geology*, v. 39, p. 431–434,
1028 doi:10.1130/G31680.1.

1029 Scheffer, M., Carpenter, S., and de Young, B., 2005, Cascading effects of overfishing marine
1030 systems: *Trends in ecology \& evolution*, v. 20, p. 579–581.

1031 Schettino, A., and Turco, E., 2011, Tectonic history of the western Tethys since the Late
1032 Triassic: *GSA Bulletin*, v. 123, p. 89–105, doi:10.1130/B30064.1.

1033 Van De Schootbrugge, B., Payne, J.L., Tomasovich, A., Pross, J., Fiebig, J., Benbrahim, M.,
1034 Föllmi, K.B., and Quan, T.M., 2008, Carbon cycle perturbation and stabilization in the
1035 wake of the Triassic-Jurassic boundary mass-extinction event: *Geochemistry, Geophysics,
1036 Geosystems*, v. 9, doi:10.1029/2007GC001914.

1037 van de Schootbrugge, B., Tremolada, F., Rosenthal, Y., Bailey, T.R., Feist-Burkhardt, S.,
1038 Brinkhuis, H., Pross, J., Kent, D. V., and Falkowski, P.G., 2007, End-Triassic calcification
1039 crisis and blooms of organic-walled “disaster species”: *Palaeogeography,
1040 Palaeoclimatology, Palaeoecology*, v. 244, p. 126–141, doi:10.1016/j.palaeo.2006.06.026.

1041 Schubert, J.K., and Bottjer, D.J., 1995, Aftermath of the Permian-Triassic mass extinction event:
1042 Paleoecology of Lower Triassic carbonates in the western USA: *Palaeogeography,
1043 Palaeoclimatology, Palaeoecology*, v. 116, p. 1–39.

1044 Scotese, C.R., 2016, Tutorial: PALEOMAP PaleoAtlas for GPlates and the PaleoData Plotter
1045 Program, PALEOMAP Project:, doi:10.13140/RG2.2.34367.00166.

1046 Sepkoski, J.J., Bambach, R.K., Raup, D.M., and Valentine, J.W., 1981, Phanerozoic marine
1047 diversity and the fossil record: *Nature*, v. 293, p. 435–437.

1048 Sibert, E.C., Hull, P.M., and Norris, R.D., 2014, Resilience of Pacific pelagic fish across the
1049 Cretaceous/Palaeogene mass extinction: *Nature Geoscience*, v. 7, p. 667–670,
1050 doi:10.1038/ngeo2227.

1051 Stampfli, G.M., Borel, G.D., Cavazza, W., Mosar, J., and Ziegler, P.A., 2001, Palaeotectonic and
1052 palaeogeographic evolution of the western Tethys and PeriTethyan domain (IGCP Project
1053 369): *Episodes*, v. 24, p. 222–227, doi:10.18814/epiugs/2001/v24i4/001.

1054 Thibodeau, A.M., Ritterbush, K., Yager, J.A., West, A.J., Ibarra, Y., Bottjer, D.J., Berelson,
1055 W.M., Bergquist, B.A., and Corsetti, F.A., 2016, Mercury anomalies and the timing of
1056 biotic recovery following the end-Triassic mass extinction: *Nature Communications*, v. 7, p.
1057 11147, doi:10.1038/ncomms11147.

1058 Todaro, S., Hollis, C., and Di Stefano, P., 2016, Spongy-like porosity in peritidal carbonates: An
1059 interaction of cyclic sea-level oscillations, fresh water supply and sediment texture:
1060 *Sedimentary Geology*, v. 333, p. 70–83, doi:10.1016/j.sedgeo.2015.12.005.

1061 Todaro, S., Rigo, M., Randazzo, V., and Di Stefano, P., 2018, The end-Triassic mass extinction:

1062 A new correlation between extinction events and $\delta^{13}\text{C}$ fluctuations from a Triassic-Jurassic
1063 peritidal succession in western Sicily: *Sedimentary Geology*, v. 368, p. 105–113,
1064 doi:<https://doi.org/10.1016/j.sedgeo.2018.03.008>.

1065 Todaro, S., Di Stefano, P., Zarcone, G., and Randazzo, V., 2017, Facies stacking and extinctions
1066 across the Triassic–Jurassic boundary in a peritidal succession from western Sicily: *Facies*,
1067 v. 63, p. 1–21, doi:10.1007/s10347-017-0500-5.

1068 Truesdale, V.W., Bale, A.J., and Woodward, E.M.S., 2000, The meridional distribution of
1069 dissolved iodine in near-surface waters of the Atlantic Ocean: *Progress in Oceanography*, v.
1070 45, p. 387–400, doi:[https://doi.org/10.1016/S0079-6611\(00\)00009-4](https://doi.org/10.1016/S0079-6611(00)00009-4).

1071 Twitchett, R.J., and Barras, C.G., 2004, Trace fossils in the aftermath of mass extinction events:
1072 Geological Society, London, Special Publications, v. 228, p. 397–418.

1073 Wagner, P.J., Kosnik, M.A., and Lidgard, S., 2006, Abundance distributions imply elevated
1074 complexity of post-Paleozoic marine ecosystems: *Science*, v. 314, p. 1289–1292.

1075 Wong, G.T.F., and Brewer, P.G., 1977, The marine chemistry of iodine in anoxic basins:
1076 *Geochimica et Cosmochimica Acta*, v. 41, p. 151–159, doi:[https://doi.org/10.1016/0016-7037\(77\)90195-8](https://doi.org/10.1016/0016-7037(77)90195-8).

1078 Wray, J.J., 1977, Late Paleozoic Calcareous Red Algae, in Flügel, E. ed., *Fossil Algae*, Berlin,
1079 Heidelberg, Springer Berlin Heidelberg, p. 167–176.

1080 Zhou, X., Jenkyns, H.C., Owens, J.D., Junium, C.K., Zheng, X.Y., Sageman, B.B., Hardisty,
1081 D.S., Lyons, T.W., Ridgwell, A., and Lu, Z., 2015, Upper ocean oxygenation dynamics
1082 from I/Ca ratios during the Cenomanian-Turonian OAE 2: *Paleoceanography*, v. 30, p. 510–
1083 526, doi:10.1002/2014PA002741.

1084 Zhou, X., Thomas, E., Rickaby, R.E.M., Winguth, A.M.E., and Lu, Z., 2014, I/Ca evidence for
1085 upper ocean deoxygenation during the PETM: *Paleoceanography*, v. 29, p. 964–975,
1086 doi:10.1002/2014PA002702.

Supporting information for “Reduction in animal abundance and oxygen availability during and after the end-Triassic mass extinction”

Contents of this file

1. Figure S1
2. Figure S2
3. Figure S3

Additional supporting information (files uploaded separately)

1. Table S1: Point count values used in the study
2. Table S2: Diversity count values used in the study
3. Table S3: I/Ca ratios from Ital cementi and Val Adrara sections

Figure S1: Plots of diversity changes in major taxa across the Triassic/Jurassic boundary in Lombardy Basin (sections Val Adrara and Ital cementi combined) and Apennine platform (Mt. Cefalo section).

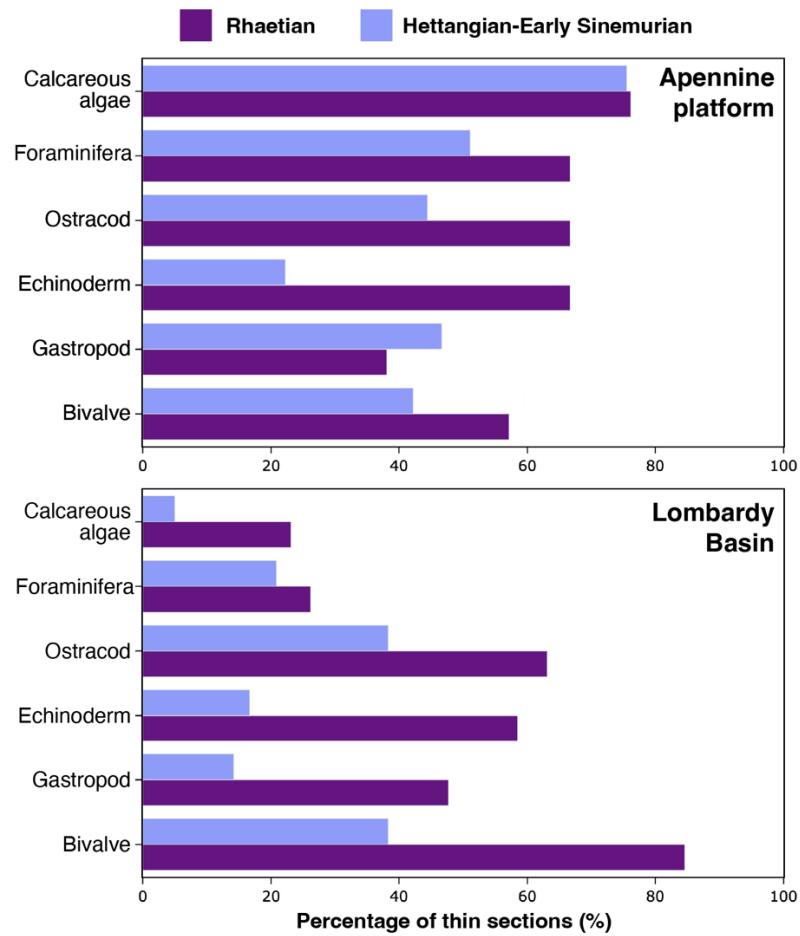
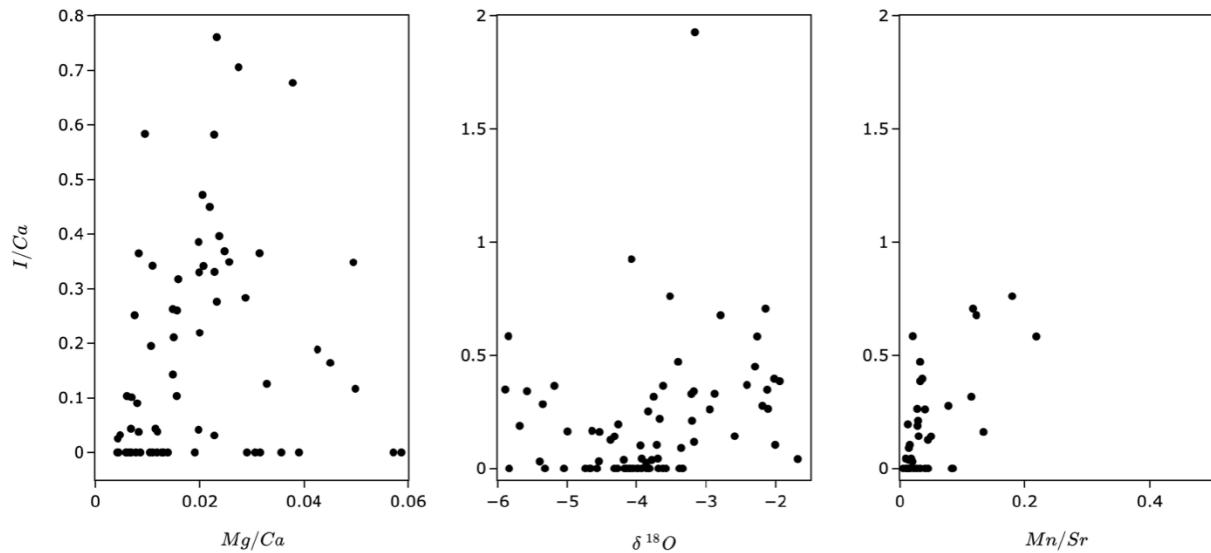


Figure S2: Plots of I/Ca vs $\delta^{18}O$, I/Ca vs Mg/Ca and I/Ca vs Mn/Sr in samples from Italcementi and Val Adrara sections.

Italcementi



Val Adrara

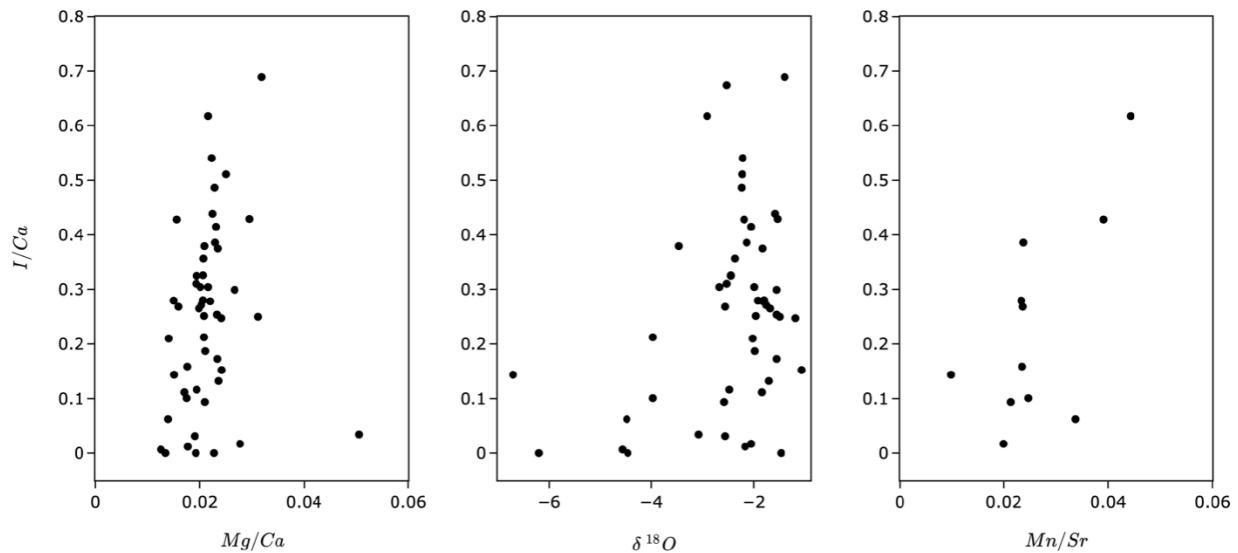
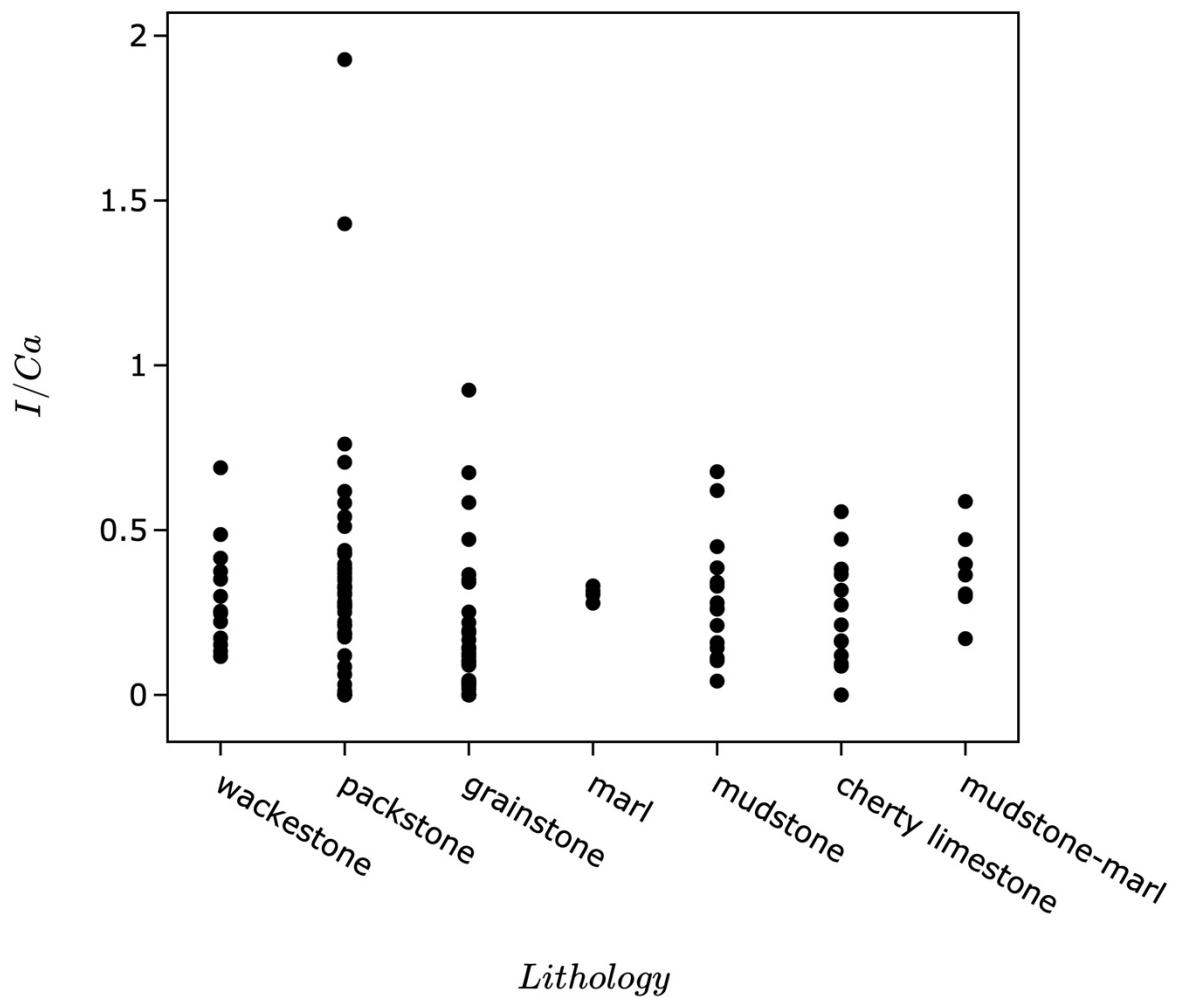


Figure S3: I/Ca ratios associated with every lithology in the Val Adrara and Italcementi sections.



height	organic matter	microsparite	bivalve	flame	microbial	clot	micrite	sponges	peloid	oooids	bunter	studnid	biota	foraminifer	green algae	bivalves	echinoderms	problematic	costracods	gastropods	soft	blocky	cl.	total	skeletal
2.8	0	0	0	0	239	0	3	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	300	15.3
9.8	0	0	0	0	298	0	0	0	0	0	0	0	0	6	12	0	4	0	0	0	0	0	0	300	0.7
30.8	0	0	0	0	275	0	0	0	0	3	0	0	0	0	1	0	0	0	0	0	0	0	300	8.3	
33	0	0	0	0	298	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	300	0.7
46.9	0	0	0	0	258	0	0	0	0	5	5	2	7	5	0	8	2	8	2	8	300	11.3			
49.9	0	0	0	0	286	0	0	0	0	0	0	0	0	6	0	0	0	3	0	5	300	3.0			
52.2	0	9	0	0	236	0	12	0	0	3	2	0	16	6	3	2	0	11	0	11	300	10.7			
64.7	0	0	0	0	300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0	
69.3	0	0	0	0	297	0	0	0	0	2	0	0	0	1	0	0	0	0	0	0	0	0	300	1.0	
71.8	0	4	0	0	288	0	0	0	0	2	0	0	0	3	0	1	0	0	0	0	0	2	300	2.0	
80	0	190	0	0	3	0	95	12	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0	
81	0	34	0	0	15	0	8	233	0	0	0	0	0	1	0	0	0	0	0	0	9	300	0.3		
82	0	13	0	0	64	0	73	122	0	1	0	2	1	0	0	0	0	0	0	0	24	300	1.3		
85	0	59	0	0	0	0	74	40	0	7	0	33	28	0	0	2	0	0	0	0	57	300	23.3		
86	0	31	0	0	3	0	48	173	0	0	0	16	2	0	0	0	0	0	0	0	27	300	6.0		
87	0	25	0	0	27	0	24	180	0	2	0	3	5	0	0	0	1	33	300	3.7					
88	0	34	0	0	16	0	63	65	0	7	0	35	5	0	0	0	2	73	300	16.3					
89	0	40	0	0	10	0	103	89	0	1	1	20	3	0	1	0	2	30	300	9.3					
91	0	7	0	0	0	0	53	152	0	0	0	8	1	0	0	0	0	79	300	3.0					
92	0	39	0	0	0	0	28	168	0	0	0	5	0	0	0	0	0	60	300	1.7					
93	0	27	0	0	120	0	76	28	0	5	0	6	1	3	14	0	0	0	20	300	9.7				
95	0	123	0	0	51	0	79	31	0	2	0	9	0	1	0	0	0	4	300	4.0					
97	0	18	0	0	98	0	174	2	1	1	0	2	0	3	1	0	0	0	0	0	300	2.3			
98	0	124	0	0	62	0	107	0	0	2	0	2	0	0	1	0	0	2	300	1.7					
100.6	0	24	0	0	202	0	41	4	0	2	0	1	0	0	1	0	0	25	300	1.3					
101.4	0	0	0	0	279	0	1	0	0	0	0	0	1	0	0	0	0	19	300	0.3					
102.5	0	0	0	0	279	0	0	0	0	0	0	0	0	1	0	0	0	0	20	300	0.3				
103.5	0	0	0	0	292	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	8	300	0.0		
108	0	0	0	0	300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0		
114.4	10	0	0	0	7	282	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	300	0.0		
119	0	0	0	0	300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0		
128	0	0	0	5	293	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	300	0.0		
134	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0						
140	0	0	0	0	300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0		
150	0	0	0	0	300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0		
158	0	0	0	0	292	0	0	0	0	0	1	0	0	3	0	0	1	0	3	0	0	300	1.7		
167	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	299	300	0.0					
168	0	1	0	0	299	0	0	0	0	0	0	0	0	0	0	0	0	1	301	0.0					
169	0	0	0	0	8	0	0	0	0	0	0	0	0	0	0	0	0	292	300	0.0					
173	0	0	0	0	293	0	0	0	0	5	0	0	2	0	0	0	0	0	0	0	0	300	2.3		
180	0	92	0	0	151	0	56	0	0	0	0	0	0	0	0	0	0	1	0	0	0	300	0.3		
198	0	60	0	0	232	0	8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0		
202	0	128	0	0	159	0	11	0	0	1	0	0	0	0	0	0	0	0	0	0	1	300	0.3		
210	0	1	0	0	264	0	0	0	0	6	0	0	0	1	0	2	0	26	300	3.0					
214	0	4	0	0	294	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	300	0.3		
219	0	27	0	0	1	0	269	0	0	0	0	0	0	0	0	0	3	0	0	0	0	300	1.0		
226	0	50	0	0	96	0	154	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0	
231	1	1	0	0	292	0	4	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	300	0.7	
238	0	22	0	0	275	0	0	0	0	0	0	0	0	0	0	0	3	0	0	0	0	0	300	1.0	
240.75	0	147	0	0	7	0	80	45	0	2	5	0	0	0	0	0	1	0	13	300	2.7				
242.9	0	6	0	0	283	0	6	0	0	0	2	0	0	0	0	0	2	0	1	0	0	300	1.3		
245.6	0	0	0	0	291	0	0	0	0	6	0	0	1	0	0	1	0	0	0	0	0	299	2.7		
250.3	0	1	0	0	274	0	0	0	1	3	2	0	2	0	0	7	2	7	2	7	299	5.3			
255	0	6	0	0	268	0	0	0	0	23	0	0	0	0	0	0	0	0	3	0	0	300	7.7		
259	0	0	0	0	278	0	0	0	0	6	0	0	2	0	0	12	0	0	2	0	0	300	6.7		
260.4	0	2	0	0	280	4	0	0	0	13	0	0	0	0	0	0	1	0	0	0	0	300	6.0		
262.5	0	1	0	0	277	0	2	0	0	18	0	0	1	0	0	0	1	0	0	0	0	0	300	6.7	
264.5	0	0	0	0	285	0	15	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	300	0.0	
269.5	0	1	0	0	279	0	0	0	0	13	0	0	2	0	0	2	0	3	0	0	0	0	300	5.7	
271.5	0	4	3	0	134	0	148	0	1	2	0	0	8	0	0	0	0	0	0	0	0	0	300	3.3	
272	0	21	0	0	201	0	63	0	0	13	0	0	1	0	0	1	0	0	1	0	0	0	300	5.0	
277	0	6	0	0	261	18	2	0	0	10	0	0	1	0	0	0	0	0	0	0	2	0	300	9.7	
278	0	13	0	0	255	4	22	0	0	5	0	0	0	0	0	0	1	0	0	0	0	0	300	3.3	
296	0	5	0	0	265	11	2	0	0	3	0	0	0	2	0	0	1	0	11	0	0	0	300	5.7	
306	0	28	0	0	185	16	63	0	0	3	0	0	4	0	0	0	0	0	1	0	0	0	300	7.7	
324	1	15	0	0	175	3	0	31	0	8	5	0	2	0	0	0	5	2	52	299	8.3				
333	0	2	0	0	207	87	1	0	0	0	0	0	0	0	0	0	1	0	0	2	0	0	300	29.3	
348	0	12	0	0	213	33	0	0	0	3	1	0	1	0	0	0	0	0	0	0	0	0	300	12.7	
363	0	6	0	0	247	45	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	300	15.7	
378	0	13	0	0	203	81	0	0	0	0	3														

height	bivalve	gastropod	ostracod	foram	greenalga	echinoderm	peloid	oid	sponge
1.5	1	0	1	0	0	0	0	0	0
3.5	1	1	1	1	0	0	0	0	0
4.6	1	1	1	1	0	0	0	1	0
6	1	1	1	1	0	0	0	1	0
6.7	1	0	1	0	0	0	0	0	0
7.5	0	0	0	0	0	0	0	0	0
8.31	1	0	1	0	0	0	0	0	0
8.6	1	0	1	0	0	0	0	0	0
9	1	1	1	0	0	1	0	0	0
9.6	1	0	1	0	0	0	0	0	0
11.5	1	0	0	0	0	1	0	0	0
12.5	1	0	1	1	0	0	0	0	0
13.2	1	0	0	0	0	0	0	0	0
14.1	0	0	1	1	0	0	0	0	0
14.5	1	0	1	0	0	0	1	0	0
15.3	1	1	1	0	1	1	0	0	0
16.5	1	0	0	0	0	0	0	0	0
17.8	1	0	1	0	0	0	0	0	0
20.6	1	0	1	0	0	1	0	0	0
21.6	0	0	0	1	0	0	0	0	0
22.9	1	0	0	0	0	0	0	0	0
23.3	1	1	1	0	0	1	0	0	0
24.3	1	0	1	1	0	1	0	0	0
25.2	0	1	0	0	0	1	0	0	0
26.1	1	0	1	1	0	0	0	0	0
26.9	0	0	1	0	0	0	0	0	0
27.5	1	0	1	0	0	0	0	0	0
27.9	1	1	0	0	1	1	0	0	0
28.3	1	1	1	0	0	1	0	1	0
29	1	1	1	0	1	1	0	0	0
30.3	1	0	0	0	0	0	1	0	0
31.5	1	0	0	0	0	0	0	0	0
32.8	0	0	0	0	0	1	0	0	0
33.8	1	1	0	1	0	1	0	0	0
34.7	1	1	0	0	0	0	0	0	0
35.6	0	0	0	0	0	0	0	0	0
36.5	1	0	0	0	0	0	1	0	0
37.3	1	1	1	0	0	1	0	0	0
38.7	1	1	0	0	0	1	1	0	0
39	1	1	1	0	0	1	1	0	0
40.3	1	1	0	0	0	1	1	0	0
40.7	1	1	1	0	0	1	1	0	0
41.1	1	1	0	0	0	1	1	0	0

41.51	1	0	1	0	1	1	1	0	0
42	0	0	0	0	0	0	0	0	0
42.5	1	1	1	0	1	1	1	0	0
43.1	1	1	1	0	0	1	0	0	0
43.6	1	1	1	0	1	1	1	0	0
43.7	1	0	0	1	0	1	1	0	0
43.8	1	1	1	0	1	1	1	0	0
43.9	1	1	1	1	1	1	1	0	0
44.2	1	1	1	0	1	1	0	0	1
44.5	1	1	1	1	1	1	1	0	1
44.9	0	0	0	0	0	0	0	0	0
45.5	1	1	0	1	1	1	1	0	1
46	1	0	1	0	0	1	0	0	1
46.5	0	0	0	0	0	0	0	0	0
47.6	0	0	0	0	0	0	0	0	0
47.8	0	0	0	0	0	0	0	0	0
48.2	0	0	0	0	0	0	0	0	0
49.2	0	0	0	0	0	0	0	0	0
49.5	0	0	0	0	0	0	0	0	0
50.2	0	0	0	0	0	0	0	0	0
52.5	0	0	0	0	0	0	0	0	0
54.5	0	0	0	0	0	0	0	0	0
55.7	0	0	0	0	0	0	0	0	0
56	0	0	0	0	0	0	0	0	0
57.2	0	0	0	0	0	0	0	0	0
58.6	0	0	0	0	0	0	0	0	0
58.9	0	0	0	0	0	0	0	0	0
60.7	0	0	0	0	0	0	0	0	0
62	0	0	0	0	0	0	0	0	0
64	0	0	0	0	0	0	0	0	0
64.6	0	0	0	0	0	0	0	0	0
65.1	0	0	0	0	0	0	0	0	0
66	1	0	1	0	0	0	0	0	0
68.6	1	0	1	0	0	0	0	0	0
70	1	0	1	0	0	0	0	0	0
71	0	0	0	0	0	0	0	0	0
72.2	1	0	1	0	0	0	1	0	0
73	0	1	1	1	0	0	1	0	0
74.4	1	0	1	0	0	0	1	0	0
75.4	0	0	0	0	0	0	0	0	0
77	0	0	0	0	0	0	0	0	0
77.3	0	0	1	1	0	0	1	0	0
78	1	1	1	0	0	1	1	0	0
79	1	1	1	1	0	0	1	0	0

|tacementi

height	bivalve	gastropod	brachiopod	echinoderm	ostracod	coral	bryozoan	sponge	foram
0	1	1	0	0	1	0	0	0	1
7	1	0	0	0	0	0	0	0	0
13	1	0	0	1	0	0	0	0	1
19	0	0	0	0	0	0	0	0	0
20	0	0	0	0	0	0	0	0	0
24	1	1	0	1	1	0	0	0	1
28	0	0	0	1	1	0	0	0	0
34	1	1	0	1	1	0	0	0	1
40	0	0	0	0	0	0	0	0	0
45	0	0	0	0	0	0	0	0	0
52	0	1	0	1	1	0	0	0	1
58	1	0	0	1	1	0	0	0	1
62	0	0	0	0	0	0	0	0	0
67	1	1	0	1	1	0	0	0	1
72	0	0	0	1	1	0	0	0	1
78	1	0	1	1	1	0	0	0	1
93	1	1	0	1	1	0	0	0	1
96	1	1	0	1	1	0	0	0	1
103	1	0	0	1	1	0	0	0	1
109	1	0	0	1	1	0	0	0	1
115	0	1	0	1	1	0	0	0	1
117	0	0	0	0	0	0	0	0	0
121	0	0	0	0	0	0	0	0	0
123	0	0	0	0	0	0	0	0	1
125	1	1	0	0	0	0	0	0	0
129	1	1	0	1	0	0	0	0	0
133	0	0	0	0	1	0	0	0	0
136	0	0	0	0	0	0	0	0	0
139	0	0	0	0	0	0	0	0	0
141	0	1	0	0	1	0	0	0	0
143	0	0	0	0	1	0	0	0	1
145	0	0	0	0	0	0	0	0	1
147	0	1	0	0	1	0	0	0	1
153	0	0	0	0	1	0	0	0	1
156	0	0	0	0	0	0	0	0	1
163	1	0	0	0	0	0	0	0	1
169	0	0	0	0	0	0	0	0	0
174	0	0	0	0	0	0	0	0	0
176	0	1	0	0	0	0	0	0	0
178	1	1	0	0	0	0	0	0	0
182	0	0	0	0	0	0	0	0	0
185	0	0	0	0	0	0	0	0	1
188	1	1	0	0	1	0	0	0	1

194	0	0	0	0	0	0	0	0	0
197	1	1	0	0	0	0	0	0	1
199	1	1	0	1	0	0	0	0	0
203	1	1	0	0	0	0	0	0	0
207	0	1	0	0	1	0	0	0	0
212	0	0	0	0	0	0	0	0	0
218	0	0	0	0	0	0	0	0	0
220	0	0	0	0	0	0	0	0	0
229	1	1	0	0	1	0	0	0	1
235	1	0	0	0	1	0	0	0	1
241	0	0	0	1	1	0	0	0	1
253	0	0	0	0	0	0	0	0	0
274	1	1	0	1	1	0	0	0	1
286	1	1	0	1	1	0	0	0	1
292	0	1	0	1	1	0	0	0	1
298	1	1	0	1	1	0	0	0	1
301	1	1	0	0	1	0	0	0	1
306	1	0	0	0	1	0	0	0	1
311	1	1	0	0	0	0	0	0	0
313	1	1	0	1	1	0	0	0	1
318	0	1	0	1	1	0	0	0	1
321	1	1	0	1	1	0	0	0	1
323	1	0	0	0	1	0	0	0	1

0	0	0
1	0	0
1	0	1
1	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	0
1	0	0
1	0	0
0	0	0
1	0	0
0	0	0
0	0	0
1	0	0
0	0	1
0	0	1
0	0	0
0	0	1

198	0	0	0	0	0	0	0	0	0
202	0	0	0	0	0	0	0	0	0
210	0	0	0	0	0	0	0	0	0
214	0	0	0	0	0	0	0	0	0
219	0	0	0	0	0	0	0	0	0
226	0	0	0	0	0	0	0	0	1
231	0	0	1	0	0	0	0	0	0
238	0	0	1	0	0	0	0	0	0
240.75	1	0	0	0	0	0	0	1	1
242.9	0	0	1	0	0	0	0	0	0
245.6	1	0	1	0	0	0	1	0	0
250.3	1	1	1	0	0	0	1	0	0
255	1	1	1	1	0	0	1	1	0
259	1	1	1	1	0	0	1	0	0
260.4	1	1	1	0	0	1	0	1	0
262.5	1	1	1	0	0	0	1	0	0
264.5	0	0	1	0	0	0	0	1	0
269.5	1	0	1	0	0	0	1	0	0
271.5	1	0	1	0	0	0	0	0	0
272	1	1	1	0	0	0	1	0	0
277	0	0	1	0	0	1	0	0	0
278	0	0	1	1	0	1	0	0	0
296	1	1	1	1	0	1	0	1	0
306	1	1	1	0	0	1	1	0	0
324	1	1	1	1	0	1	1	1	0
333	1	0	1	0	0	1	0	0	0
348	1	0	1	1	0	1	0	0	0
363	0	0	1	1	0	1	0	0	0
378	0	0	0	0	0	1	0	0	0
390	1	0	1	1	0	1	1	0	0
417	0	0	1	0	0	1	1	1	0

|Val Adrara

height	bivalve	gastropod	ostracod	foram	greenalga	echinoderm	peloid	oid	sponge
1.5	1	0	1	0	0	0	0	0	0
3.5	1	1	1	1	0	0	0	0	0
4.6	1	1	1	1	0	0	0	1	0
6	1	1	1	1	0	0	0	1	0
6.7	1	0	1	0	0	0	0	0	0
7.5	0	0	0	0	0	0	0	0	0
8.31	1	0	1	0	0	0	0	0	0
8.6	1	0	1	0	0	0	0	0	0
9	1	1	1	0	0	1	0	0	0
9.6	1	0	1	0	0	0	0	0	0
11.5	1	0	0	0	0	1	0	0	0
12.5	1	0	1	1	0	0	0	0	0
13.2	1	0	0	0	0	0	0	0	0
14.1	0	0	1	1	0	0	0	0	0
14.5	1	0	1	0	0	0	1	0	0
15.3	1	1	1	0	1	1	0	0	0
16.5	1	0	0	0	0	0	0	0	0
17.8	1	0	1	0	0	0	0	0	0
20.6	1	0	1	0	0	1	0	0	0
21.6	0	0	0	1	0	0	0	0	0
22.9	1	0	0	0	0	0	0	0	0
23.3	1	1	1	0	0	1	0	0	0
24.3	1	0	1	1	0	1	0	0	0
25.2	0	1	0	0	0	1	0	0	0
26.1	1	0	1	1	0	0	0	0	0
26.9	0	0	1	0	0	0	0	0	0
27.5	1	0	1	0	0	0	0	0	0
27.9	1	1	0	0	1	1	0	0	0
28.3	1	1	1	0	0	1	0	1	0
29	1	1	1	0	1	1	0	0	0
30.3	1	0	0	0	0	0	1	0	0
31.5	1	0	0	0	0	0	0	0	0
32.8	0	0	0	0	0	1	0	0	0
33.8	1	1	0	1	0	1	0	0	0
34.7	1	1	0	0	0	0	0	0	0
35.6	0	0	0	0	0	0	0	0	0
36.5	1	0	0	0	0	0	1	0	0
37.3	1	1	1	0	0	1	0	0	0
38.7	1	1	0	0	0	1	1	0	0
39	1	1	1	0	0	1	1	0	0
40.3	1	1	0	0	0	1	1	0	0
40.7	1	1	1	0	0	1	1	0	0
41.1	1	1	0	0	0	1	1	0	0

41.51	1	0	1	0	1	1	1	0	0
42	0	0	0	0	0	0	0	0	0
42.5	1	1	1	0	1	1	1	0	0
43.1	1	1	1	0	0	1	0	0	0
43.6	1	1	1	0	1	1	1	0	0
43.7	1	0	0	1	0	1	1	0	0
43.8	1	1	1	0	1	1	1	0	0
43.9	1	1	1	1	1	1	1	0	0
44.2	1	1	1	0	1	1	0	0	1
44.5	1	1	1	1	1	1	1	0	1
44.9	0	0	0	0	0	0	0	0	0
45.5	1	1	0	1	1	1	1	0	1
46	1	0	1	0	0	1	0	0	1
46.5	0	0	0	0	0	0	0	0	0
47.6	0	0	0	0	0	0	0	0	0
47.8	0	0	0	0	0	0	0	0	0
48.2	0	0	0	0	0	0	0	0	0
49.2	0	0	0	0	0	0	0	0	0
49.5	0	0	0	0	0	0	0	0	0
50.2	0	0	0	0	0	0	0	0	0
52.5	0	0	0	0	0	0	0	0	0
54.5	0	0	0	0	0	0	0	0	0
55.7	0	0	0	0	0	0	0	0	0
56	0	0	0	0	0	0	0	0	0
57.2	0	0	0	0	0	0	0	0	0
58.6	0	0	0	0	0	0	0	0	0
58.9	0	0	0	0	0	0	0	0	0
60.7	0	0	0	0	0	0	0	0	0
62	0	0	0	0	0	0	0	0	0
64	0	0	0	0	0	0	0	0	0
64.6	0	0	0	0	0	0	0	0	0
65.1	0	0	0	0	0	0	0	0	0
66	1	0	1	0	0	0	0	0	0
68.6	1	0	1	0	0	0	0	0	0
70	1	0	1	0	0	0	0	0	0
71	0	0	0	0	0	0	0	0	0
72.2	1	0	1	0	0	0	1	0	0
73	0	1	1	1	0	0	1	0	0
74.4	1	0	1	0	0	0	1	0	0
75.4	0	0	0	0	0	0	0	0	0
77	0	0	0	0	0	0	0	0	0
77.3	0	0	1	1	0	0	1	0	0
78	1	1	1	0	0	1	1	0	0
79	1	1	1	1	0	0	1	0	0

height	bivalve	gastropod	brachiopod	echinoderm	ostracod	coral	bryozoan	sponge	foram
0	1	1	0	0	1	0	0	0	1
7	1	0	0	0	0	0	0	0	0
13	1	0	0	1	0	0	0	0	1
19	0	0	0	0	0	0	0	0	0
20	0	0	0	0	0	0	0	0	0
24	1	1	0	1	1	0	0	0	1
28	0	0	0	1	1	0	0	0	0
34	1	1	0	1	1	0	0	0	1
40	0	0	0	0	0	0	0	0	0
45	0	0	0	0	0	0	0	0	0
52	0	1	0	1	1	0	0	0	1
58	1	0	0	1	1	0	0	0	1
62	0	0	0	0	0	0	0	0	0
67	1	1	0	1	1	0	0	0	1
72	0	0	0	1	1	0	0	0	1
78	1	0	1	1	1	0	0	0	1
93	1	1	0	1	1	0	0	0	1
96	1	1	0	1	1	0	0	0	1
103	1	0	0	1	1	0	0	0	1
109	1	0	0	1	1	0	0	0	1
115	0	1	0	1	1	0	0	0	1
117	0	0	0	0	0	0	0	0	0
121	0	0	0	0	0	0	0	0	0
123	0	0	0	0	0	0	0	0	1
125	1	1	0	0	0	0	0	0	0
129	1	1	0	1	0	0	0	0	0
133	0	0	0	0	1	0	0	0	0
136	0	0	0	0	0	0	0	0	0
139	0	0	0	0	0	0	0	0	0
141	0	1	0	0	1	0	0	0	0
143	0	0	0	0	1	0	0	0	1
145	0	0	0	0	0	0	0	0	1
147	0	1	0	0	1	0	0	0	1
153	0	0	0	0	1	0	0	0	1
156	0	0	0	0	0	0	0	0	1
163	1	0	0	0	0	0	0	0	1
169	0	0	0	0	0	0	0	0	0
174	0	0	0	0	0	0	0	0	0
176	0	1	0	0	0	0	0	0	0
178	1	1	0	0	0	0	0	0	0
182	0	0	0	0	0	0	0	0	0
185	0	0	0	0	0	0	0	0	1
188	1	1	0	0	1	0	0	0	1

194	0	0	0	0	0	0	0	0	0
197	1	1	0	0	0	0	0	0	1
199	1	1	0	1	0	0	0	0	0
203	1	1	0	0	0	0	0	0	0
207	0	1	0	0	1	0	0	0	0
212	0	0	0	0	0	0	0	0	0
218	0	0	0	0	0	0	0	0	0
220	0	0	0	0	0	0	0	0	0
229	1	1	0	0	1	0	0	0	1
235	1	0	0	0	1	0	0	0	1
241	0	0	0	1	1	0	0	0	1
253	0	0	0	0	0	0	0	0	0
274	1	1	0	1	1	0	0	0	1
286	1	1	0	1	1	0	0	0	1
292	0	1	0	1	1	0	0	0	1
298	1	1	0	1	1	0	0	0	1
301	1	1	0	0	1	0	0	0	1
306	1	0	0	0	1	0	0	0	1
311	1	1	0	0	0	0	0	0	0
313	1	1	0	1	1	0	0	0	1
318	0	1	0	1	1	0	0	0	1
321	1	1	0	1	1	0	0	0	1
323	1	0	0	0	1	0	0	0	1

0	0	0
1	0	0
1	0	1
1	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	0
1	0	0
1	0	0
0	0	0
1	0	0
0	0	0
0	0	0
1	0	0
0	0	1
0	0	1
0	0	0
0	0	1

198	0	0	0	0	0	0	0	0	0
202	0	0	0	0	0	0	0	0	0
210	0	0	0	0	0	0	0	0	0
214	0	0	0	0	0	0	0	0	0
219	0	0	0	0	0	0	0	0	0
226	0	0	0	0	0	0	0	0	1
231	0	0	1	0	0	0	0	0	0
238	0	0	1	0	0	0	0	0	0
240.75	1	0	0	0	0	0	0	1	1
242.9	0	0	1	0	0	0	0	0	0
245.6	1	0	1	0	0	0	1	0	0
250.3	1	1	1	0	0	0	1	0	0
255	1	1	1	1	0	0	1	1	0
259	1	1	1	1	0	0	1	0	0
260.4	1	1	1	0	0	1	0	1	0
262.5	1	1	1	0	0	0	1	0	0
264.5	0	0	1	0	0	0	0	1	0
269.5	1	0	1	0	0	0	1	0	0
271.5	1	0	1	0	0	0	0	0	0
272	1	1	1	0	0	0	1	0	0
277	0	0	1	0	0	1	0	0	0
278	0	0	1	1	0	1	0	0	0
296	1	1	1	1	0	1	0	1	0
306	1	1	1	0	0	1	1	0	0
324	1	1	1	1	0	1	1	1	0
333	1	0	1	0	0	1	0	0	0
348	1	0	1	1	0	1	0	0	0
363	0	0	1	1	0	1	0	0	0
378	0	0	0	0	0	1	0	0	0
390	1	0	1	1	0	1	1	0	0
417	0	0	1	0	0	1	1	1	0

|Val Adrara

height	bivalve	gastropod	ostracod	foram	greenalga	echinoderm	peloid	oid	sponge
1.5	1	0	1	0	0	0	0	0	0
3.5	1	1	1	1	0	0	0	0	0
4.6	1	1	1	1	0	0	0	1	0
6	1	1	1	1	0	0	0	1	0
6.7	1	0	1	0	0	0	0	0	0
7.5	0	0	0	0	0	0	0	0	0
8.31	1	0	1	0	0	0	0	0	0
8.6	1	0	1	0	0	0	0	0	0
9	1	1	1	0	0	1	0	0	0
9.6	1	0	1	0	0	0	0	0	0
11.5	1	0	0	0	0	1	0	0	0
12.5	1	0	1	1	0	0	0	0	0
13.2	1	0	0	0	0	0	0	0	0
14.1	0	0	1	1	0	0	0	0	0
14.5	1	0	1	0	0	0	1	0	0
15.3	1	1	1	0	1	1	0	0	0
16.5	1	0	0	0	0	0	0	0	0
17.8	1	0	1	0	0	0	0	0	0
20.6	1	0	1	0	0	1	0	0	0
21.6	0	0	0	1	0	0	0	0	0
22.9	1	0	0	0	0	0	0	0	0
23.3	1	1	1	0	0	1	0	0	0
24.3	1	0	1	1	0	1	0	0	0
25.2	0	1	0	0	0	1	0	0	0
26.1	1	0	1	1	0	0	0	0	0
26.9	0	0	1	0	0	0	0	0	0
27.5	1	0	1	0	0	0	0	0	0
27.9	1	1	0	0	1	1	0	0	0
28.3	1	1	1	0	0	1	0	1	0
29	1	1	1	0	1	1	0	0	0
30.3	1	0	0	0	0	0	1	0	0
31.5	1	0	0	0	0	0	0	0	0
32.8	0	0	0	0	0	1	0	0	0
33.8	1	1	0	1	0	1	0	0	0
34.7	1	1	0	0	0	0	0	0	0
35.6	0	0	0	0	0	0	0	0	0
36.5	1	0	0	0	0	0	1	0	0
37.3	1	1	1	0	0	1	0	0	0
38.7	1	1	0	0	0	1	1	0	0
39	1	1	1	0	0	1	1	0	0
40.3	1	1	0	0	0	1	1	0	0
40.7	1	1	1	0	0	1	1	0	0
41.1	1	1	0	0	0	1	1	0	0

41.51	1	0	1	0	1	1	1	0	0
42	0	0	0	0	0	0	0	0	0
42.5	1	1	1	0	1	1	1	0	0
43.1	1	1	1	0	0	1	0	0	0
43.6	1	1	1	0	1	1	1	0	0
43.7	1	0	0	1	0	1	1	0	0
43.8	1	1	1	0	1	1	1	0	0
43.9	1	1	1	1	1	1	1	0	0
44.2	1	1	1	0	1	1	0	0	1
44.5	1	1	1	1	1	1	1	0	1
44.9	0	0	0	0	0	0	0	0	0
45.5	1	1	0	1	1	1	1	0	1
46	1	0	1	0	0	1	0	0	1
46.5	0	0	0	0	0	0	0	0	0
47.6	0	0	0	0	0	0	0	0	0
47.8	0	0	0	0	0	0	0	0	0
48.2	0	0	0	0	0	0	0	0	0
49.2	0	0	0	0	0	0	0	0	0
49.5	0	0	0	0	0	0	0	0	0
50.2	0	0	0	0	0	0	0	0	0
52.5	0	0	0	0	0	0	0	0	0
54.5	0	0	0	0	0	0	0	0	0
55.7	0	0	0	0	0	0	0	0	0
56	0	0	0	0	0	0	0	0	0
57.2	0	0	0	0	0	0	0	0	0
58.6	0	0	0	0	0	0	0	0	0
58.9	0	0	0	0	0	0	0	0	0
60.7	0	0	0	0	0	0	0	0	0
62	0	0	0	0	0	0	0	0	0
64	0	0	0	0	0	0	0	0	0
64.6	0	0	0	0	0	0	0	0	0
65.1	0	0	0	0	0	0	0	0	0
66	1	0	1	0	0	0	0	0	0
68.6	1	0	1	0	0	0	0	0	0
70	1	0	1	0	0	0	0	0	0
71	0	0	0	0	0	0	0	0	0
72.2	1	0	1	0	0	0	1	0	0
73	0	1	1	1	0	0	1	0	0
74.4	1	0	1	0	0	0	1	0	0
75.4	0	0	0	0	0	0	0	0	0
77	0	0	0	0	0	0	0	0	0
77.3	0	0	1	1	0	0	1	0	0
78	1	1	1	0	0	1	1	0	0
79	1	1	1	1	0	0	1	0	0

height	bivalve	gastropod	brachiopod	echinoderm	ostracod	coral	bryozoan	sponge	foram
0	1	1	0	0	1	0	0	0	1
7	1	0	0	0	0	0	0	0	0
13	1	0	0	1	0	0	0	0	1
19	0	0	0	0	0	0	0	0	0
20	0	0	0	0	0	0	0	0	0
24	1	1	0	1	1	0	0	0	1
28	0	0	0	1	1	0	0	0	0
34	1	1	0	1	1	0	0	0	1
40	0	0	0	0	0	0	0	0	0
45	0	0	0	0	0	0	0	0	0
52	0	1	0	1	1	0	0	0	1
58	1	0	0	1	1	0	0	0	1
62	0	0	0	0	0	0	0	0	0
67	1	1	0	1	1	0	0	0	1
72	0	0	0	1	1	0	0	0	1
78	1	0	1	1	1	0	0	0	1
93	1	1	0	1	1	0	0	0	1
96	1	1	0	1	1	0	0	0	1
103	1	0	0	1	1	0	0	0	1
109	1	0	0	1	1	0	0	0	1
115	0	1	0	1	1	0	0	0	1
117	0	0	0	0	0	0	0	0	0
121	0	0	0	0	0	0	0	0	0
123	0	0	0	0	0	0	0	0	1
125	1	1	0	0	0	0	0	0	0
129	1	1	0	1	0	0	0	0	0
133	0	0	0	0	1	0	0	0	0
136	0	0	0	0	0	0	0	0	0
139	0	0	0	0	0	0	0	0	0
141	0	1	0	0	1	0	0	0	0
143	0	0	0	0	1	0	0	0	1
145	0	0	0	0	0	0	0	0	1
147	0	1	0	0	1	0	0	0	1
153	0	0	0	0	1	0	0	0	1
156	0	0	0	0	0	0	0	0	1
163	1	0	0	0	0	0	0	0	1
169	0	0	0	0	0	0	0	0	0
174	0	0	0	0	0	0	0	0	0
176	0	1	0	0	0	0	0	0	0
178	1	1	0	0	0	0	0	0	0
182	0	0	0	0	0	0	0	0	0
185	0	0	0	0	0	0	0	0	1
188	1	1	0	0	1	0	0	0	1

194	0	0	0	0	0	0	0	0	0
197	1	1	0	0	0	0	0	0	1
199	1	1	0	1	0	0	0	0	0
203	1	1	0	0	0	0	0	0	0
207	0	1	0	0	1	0	0	0	0
212	0	0	0	0	0	0	0	0	0
218	0	0	0	0	0	0	0	0	0
220	0	0	0	0	0	0	0	0	0
229	1	1	0	0	1	0	0	0	1
235	1	0	0	0	1	0	0	0	1
241	0	0	0	1	1	0	0	0	1
253	0	0	0	0	0	0	0	0	0
274	1	1	0	1	1	0	0	0	1
286	1	1	0	1	1	0	0	0	1
292	0	1	0	1	1	0	0	0	1
298	1	1	0	1	1	0	0	0	1
301	1	1	0	0	1	0	0	0	1
306	1	0	0	0	1	0	0	0	1
311	1	1	0	0	0	0	0	0	0
313	1	1	0	1	1	0	0	0	1
318	0	1	0	1	1	0	0	0	1
321	1	1	0	1	1	0	0	0	1
323	1	0	0	0	1	0	0	0	1

0	0	0
1	0	0
1	0	1
1	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	0
1	0	0
1	0	0
0	0	0
1	0	0
0	0	0
0	0	0
1	0	0
0	0	1
0	0	1
0	0	0
0	0	1

198	0	0	0	0	0	0	0	0	0
202	0	0	0	0	0	0	0	0	0
210	0	0	0	0	0	0	0	0	0
214	0	0	0	0	0	0	0	0	0
219	0	0	0	0	0	0	0	0	0
226	0	0	0	0	0	0	0	0	1
231	0	0	1	0	0	0	0	0	0
238	0	0	1	0	0	0	0	0	0
240.75	1	0	0	0	0	0	0	1	1
242.9	0	0	1	0	0	0	0	0	0
245.6	1	0	1	0	0	0	1	0	0
250.3	1	1	1	0	0	0	1	0	0
255	1	1	1	1	0	0	1	1	0
259	1	1	1	1	0	0	1	0	0
260.4	1	1	1	0	0	1	0	1	0
262.5	1	1	1	0	0	0	1	0	0
264.5	0	0	1	0	0	0	0	1	0
269.5	1	0	1	0	0	0	1	0	0
271.5	1	0	1	0	0	0	0	0	0
272	1	1	1	0	0	0	1	0	0
277	0	0	1	0	0	1	0	0	0
278	0	0	1	1	0	1	0	0	0
296	1	1	1	1	0	1	0	1	0
306	1	1	1	0	0	1	1	0	0
324	1	1	1	1	0	1	1	1	0
333	1	0	1	0	0	1	0	0	0
348	1	0	1	1	0	1	0	0	0
363	0	0	1	1	0	1	0	0	0
378	0	0	0	0	0	1	0	0	0
390	1	0	1	1	0	1	1	0	0
417	0	0	1	0	0	1	1	1	0

|Val Adrara

height	bivalve	gastropod	ostracod	foram	greenalga	echinoderm	peloid	oid	sponge
1.5	1	0	1	0	0	0	0	0	0
3.5	1	1	1	1	0	0	0	0	0
4.6	1	1	1	1	0	0	0	1	0
6	1	1	1	1	0	0	0	1	0
6.7	1	0	1	0	0	0	0	0	0
7.5	0	0	0	0	0	0	0	0	0
8.31	1	0	1	0	0	0	0	0	0
8.6	1	0	1	0	0	0	0	0	0
9	1	1	1	0	0	1	0	0	0
9.6	1	0	1	0	0	0	0	0	0
11.5	1	0	0	0	0	1	0	0	0
12.5	1	0	1	1	0	0	0	0	0
13.2	1	0	0	0	0	0	0	0	0
14.1	0	0	1	1	0	0	0	0	0
14.5	1	0	1	0	0	0	1	0	0
15.3	1	1	1	0	1	1	0	0	0
16.5	1	0	0	0	0	0	0	0	0
17.8	1	0	1	0	0	0	0	0	0
20.6	1	0	1	0	0	1	0	0	0
21.6	0	0	0	1	0	0	0	0	0
22.9	1	0	0	0	0	0	0	0	0
23.3	1	1	1	0	0	1	0	0	0
24.3	1	0	1	1	0	1	0	0	0
25.2	0	1	0	0	0	1	0	0	0
26.1	1	0	1	1	0	0	0	0	0
26.9	0	0	1	0	0	0	0	0	0
27.5	1	0	1	0	0	0	0	0	0
27.9	1	1	0	0	1	1	0	0	0
28.3	1	1	1	0	0	1	0	1	0
29	1	1	1	0	1	1	0	0	0
30.3	1	0	0	0	0	0	1	0	0
31.5	1	0	0	0	0	0	0	0	0
32.8	0	0	0	0	0	1	0	0	0
33.8	1	1	0	1	0	1	0	0	0
34.7	1	1	0	0	0	0	0	0	0
35.6	0	0	0	0	0	0	0	0	0
36.5	1	0	0	0	0	0	1	0	0
37.3	1	1	1	0	0	1	0	0	0
38.7	1	1	0	0	0	1	1	0	0
39	1	1	1	0	0	1	1	0	0
40.3	1	1	0	0	0	1	1	0	0
40.7	1	1	1	0	0	1	1	0	0
41.1	1	1	0	0	0	1	1	0	0

41.51	1	0	1	0	1	1	1	0	0
42	0	0	0	0	0	0	0	0	0
42.5	1	1	1	0	1	1	1	0	0
43.1	1	1	1	0	0	1	0	0	0
43.6	1	1	1	0	1	1	1	0	0
43.7	1	0	0	1	0	1	1	0	0
43.8	1	1	1	0	1	1	1	0	0
43.9	1	1	1	1	1	1	1	0	0
44.2	1	1	1	0	1	1	0	0	1
44.5	1	1	1	1	1	1	1	0	1
44.9	0	0	0	0	0	0	0	0	0
45.5	1	1	0	1	1	1	1	0	1
46	1	0	1	0	0	1	0	0	1
46.5	0	0	0	0	0	0	0	0	0
47.6	0	0	0	0	0	0	0	0	0
47.8	0	0	0	0	0	0	0	0	0
48.2	0	0	0	0	0	0	0	0	0
49.2	0	0	0	0	0	0	0	0	0
49.5	0	0	0	0	0	0	0	0	0
50.2	0	0	0	0	0	0	0	0	0
52.5	0	0	0	0	0	0	0	0	0
54.5	0	0	0	0	0	0	0	0	0
55.7	0	0	0	0	0	0	0	0	0
56	0	0	0	0	0	0	0	0	0
57.2	0	0	0	0	0	0	0	0	0
58.6	0	0	0	0	0	0	0	0	0
58.9	0	0	0	0	0	0	0	0	0
60.7	0	0	0	0	0	0	0	0	0
62	0	0	0	0	0	0	0	0	0
64	0	0	0	0	0	0	0	0	0
64.6	0	0	0	0	0	0	0	0	0
65.1	0	0	0	0	0	0	0	0	0
66	1	0	1	0	0	0	0	0	0
68.6	1	0	1	0	0	0	0	0	0
70	1	0	1	0	0	0	0	0	0
71	0	0	0	0	0	0	0	0	0
72.2	1	0	1	0	0	0	1	0	0
73	0	1	1	1	0	0	1	0	0
74.4	1	0	1	0	0	0	1	0	0
75.4	0	0	0	0	0	0	0	0	0
77	0	0	0	0	0	0	0	0	0
77.3	0	0	1	1	0	0	1	0	0
78	1	1	1	0	0	1	1	0	0
79	1	1	1	1	0	0	1	0	0

height	bivalve	gastropod	brachiopod	echinoderm	ostracod	coral	bryozoan	sponge	foram
0	1	1	0	0	1	0	0	0	1
7	1	0	0	0	0	0	0	0	0
13	1	0	0	1	0	0	0	0	1
19	0	0	0	0	0	0	0	0	0
20	0	0	0	0	0	0	0	0	0
24	1	1	0	1	1	0	0	0	1
28	0	0	0	1	1	0	0	0	0
34	1	1	0	1	1	0	0	0	1
40	0	0	0	0	0	0	0	0	0
45	0	0	0	0	0	0	0	0	0
52	0	1	0	1	1	0	0	0	1
58	1	0	0	1	1	0	0	0	1
62	0	0	0	0	0	0	0	0	0
67	1	1	0	1	1	0	0	0	1
72	0	0	0	1	1	0	0	0	1
78	1	0	1	1	1	0	0	0	1
93	1	1	0	1	1	0	0	0	1
96	1	1	0	1	1	0	0	0	1
103	1	0	0	1	1	0	0	0	1
109	1	0	0	1	1	0	0	0	1
115	0	1	0	1	1	0	0	0	1
117	0	0	0	0	0	0	0	0	0
121	0	0	0	0	0	0	0	0	0
123	0	0	0	0	0	0	0	0	1
125	1	1	0	0	0	0	0	0	0
129	1	1	0	1	0	0	0	0	0
133	0	0	0	0	1	0	0	0	0
136	0	0	0	0	0	0	0	0	0
139	0	0	0	0	0	0	0	0	0
141	0	1	0	0	1	0	0	0	0
143	0	0	0	0	1	0	0	0	1
145	0	0	0	0	0	0	0	0	1
147	0	1	0	0	1	0	0	0	1
153	0	0	0	0	1	0	0	0	1
156	0	0	0	0	0	0	0	0	1
163	1	0	0	0	0	0	0	0	1
169	0	0	0	0	0	0	0	0	0
174	0	0	0	0	0	0	0	0	0
176	0	1	0	0	0	0	0	0	0
178	1	1	0	0	0	0	0	0	0
182	0	0	0	0	0	0	0	0	0
185	0	0	0	0	0	0	0	0	1
188	1	1	0	0	1	0	0	0	1

194	0	0	0	0	0	0	0	0	0
197	1	1	0	0	0	0	0	0	1
199	1	1	0	1	0	0	0	0	0
203	1	1	0	0	0	0	0	0	0
207	0	1	0	0	1	0	0	0	0
212	0	0	0	0	0	0	0	0	0
218	0	0	0	0	0	0	0	0	0
220	0	0	0	0	0	0	0	0	0
229	1	1	0	0	1	0	0	0	1
235	1	0	0	0	1	0	0	0	1
241	0	0	0	1	1	0	0	0	1
253	0	0	0	0	0	0	0	0	0
274	1	1	0	1	1	0	0	0	1
286	1	1	0	1	1	0	0	0	1
292	0	1	0	1	1	0	0	0	1
298	1	1	0	1	1	0	0	0	1
301	1	1	0	0	1	0	0	0	1
306	1	0	0	0	1	0	0	0	1
311	1	1	0	0	0	0	0	0	0
313	1	1	0	1	1	0	0	0	1
318	0	1	0	1	1	0	0	0	1
321	1	1	0	1	1	0	0	0	1
323	1	0	0	0	1	0	0	0	1

0	0	0
1	0	0
1	0	1
1	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	1
0	0	0
1	0	0
1	0	0
0	0	0
1	0	0
0	0	0
0	0	0
1	0	0
0	0	1
0	0	1
0	0	0
0	0	1

198	0	0	0	0	0	0	0	0	0
202	0	0	0	0	0	0	0	0	0
210	0	0	0	0	0	0	0	0	0
214	0	0	0	0	0	0	0	0	0
219	0	0	0	0	0	0	0	0	0
226	0	0	0	0	0	0	0	0	1
231	0	0	1	0	0	0	0	0	0
238	0	0	1	0	0	0	0	0	0
240.75	1	0	0	0	0	0	0	1	1
242.9	0	0	1	0	0	0	0	0	0
245.6	1	0	1	0	0	0	1	0	0
250.3	1	1	1	0	0	0	1	0	0
255	1	1	1	1	0	0	1	1	0
259	1	1	1	1	0	0	1	0	0
260.4	1	1	1	0	0	1	0	1	0
262.5	1	1	1	0	0	0	1	0	0
264.5	0	0	1	0	0	0	0	1	0
269.5	1	0	1	0	0	0	1	0	0
271.5	1	0	1	0	0	0	0	0	0
272	1	1	1	0	0	0	1	0	0
277	0	0	1	0	0	1	0	0	0
278	0	0	1	1	0	1	0	0	0
296	1	1	1	1	0	1	0	1	0
306	1	1	1	0	0	1	1	0	0
324	1	1	1	1	0	1	1	1	0
333	1	0	1	0	0	1	0	0	0
348	1	0	1	1	0	1	0	0	0
363	0	0	1	1	0	1	0	0	0
378	0	0	0	0	0	1	0	0	0
390	1	0	1	1	0	1	1	0	0
417	0	0	1	0	0	1	1	1	0

|Val Adrara

height	I/Ca ratios	stage	Italcementi
1.5	0.11638436	Rhaetian	
2.5	0.4147545	Rhaetian	
3.5	0.48662269	Rhaetian	
4.5	0.21234732	Rhaetian	
4.6	0.22124713	Rhaetian	
4.8	0.18699916	Rhaetian	
5	0.28009251	Rhaetian	
5.2	0.18705932	Rhaetian	
6	0.2511927	Rhaetian	
6.4	0.35096098	Rhaetian	
6.7	0.37493768	Rhaetian	
7.4	0.68923119	Rhaetian	
7.5	0.24974417	Rhaetian	
7.6	0.15222062	Rhaetian	
7.7	0.17262048	Rhaetian	
7.9	0.22184572	Rhaetian	
8.3	0.25365962	Rhaetian	
8.4	0.29917112	Rhaetian	
8.5	0.24714651	Rhaetian	
15	0.13219031	Rhaetian	
20.5	0.03130065	Rhaetian	
23.9	0.42906931	Rhaetian	
30.1	0	Rhaetian	
35.4	0.43851522	Rhaetian	
40.1	0.00652526	Rhaetian	
41.1	0.06208281	Rhaetian	
43.1	0.61769158	Rhaetian	
43.5	0.51095725	Rhaetian	
43.6	0.3043029	Rhaetian	
43.7	0.37933989	Rhaetian	
43.8	0.32461183	Rhaetian	
43.9	0.32562269	Rhaetian	
44.2	0.31032334	Rhaetian	
44.5	0.35676672	Rhaetian	
45.5	1.4293723	Rhaetian	
46	0.54052791	Rhaetian	
46.1	0.278198	Hettangian	
46.5	0.30434453	Hettangian	
46.8	0.3156856	Hettangian	
47	0.33077032	Hettangian	
47.8	0.27103742	Hettangian	
48	0.26484055	Hettangian	
49.9	0.1585904	Hettangian	
53	0.11168259	Hettangian	
56.6	0.27941121	Hettangian	
60.7	0.21001301	Hettangian	

61	0.42809386	Hettangian
65.3	0.26825504	Hettangian
70	0.01185157	Hettangian
72.9	0.38574635	Hettangian
77	0.01708235	Hettangian
80	0.67437623	Hettangian
84.8	0.03429122	Hettangian
86.5	0.09389566	Hettangian
90.2	0	Hettangian
93	0.10102217	Hettangian
95.1	0	Hettangian
98.3	0.14356164	Hettangian
100.6	0	Hettangian

height	I/Ca ratios	stage
0.5	0.17553944	Rhaetian
2.8	0.70584274	Rhaetian
6.8	0.33071266	Rhaetian
9.8	0.27584429	Rhaetian
16	1.92757485	Rhaetian
20	0.58230292	Rhaetian
24	0	Rhaetian
30.8	0.76108244	Rhaetian
34	0.34825307	Rhaetian
41.9	0.18360507	Rhaetian
46.9	0.39625049	Rhaetian
48	0.36897473	Rhaetian
50.3	0.34138592	Rhaetian
51.3	0.32985263	Rhaetian
53.5	0.44993818	Rhaetian
55	0.67721403	Rhaetian
57	0.61993325	Rhaetian
57.6	0.38564853	Rhaetian
57.9	0.04191127	Rhaetian
60.7	0.10373532	Rhaetian
64.7	0.26250336	lower-Hettangian
66.2	0.21074568	lower-Hettangian
67.7	0.26016131	lower-Hettangian
71.4	0.14259066	lower-Hettangian
79	0.47159107	lower-Hettangian
86	0	lower-Hettangian
90	0.34901169	lower-Hettangian
95	0	lower-Hettangian
100.6	0.18866823	lower-Hettangian
103.5	0.34182784	lower-Hettangian
106.5	0.36518116	lower-Hettangian
108	0.03116112	lower-Hettangian
110	0.58361463	lower-Hettangian
112	0	lower-Hettangian
115	0.92470904	lower-Hettangian
118	0.03855055	lower-Hettangian
122	0.12578035	lower-Hettangian
126	0.02515991	lower-Hettangian
129	0	lower-Hettangian
132	0	lower-Hettangian
135	0	lower-Hettangian
138	0	lower-Hettangian
141	0	lower-Hettangian
144	0	lower-Hettangian
147	0	lower-Hettangian
150	0.03213083	lower-Hettangian

154 0.25153159 lower-Hettangian
157 0.04376268 lower-Hettangian
160 0.03796047 lower-Hettangian
163 0.16692414 lower-Hettangian
166 0 lower-Hettangian
169 0 lower-Hettangian
172 0 lower-Hettangian
175 0 lower-Hettangian
178 0 lower-Hettangian
181 0 lower-Hettangian
184 0.13907046 lower-Hettangian
186.8 0 lower-Hettangian
190 0 lower-Hettangian
193 0 lower-Hettangian
196 0 lower-Hettangian
200 0 lower-Hettangian
203 0 lower-Hettangian
206 0.09016142 lower-Hettangian
209 0.10369134 lower-Hettangian
212 0.1417996 lower-Hettangian
215 0.11684494 lower-Hettangian
218 0.19509616 lower-Hettangian
222 0.1011641 lower-Hettangian
227 0 lower-Hettangian
230 0.04381678 lower-Hettangian
233 0.21926847 lower-Hettangian
236 0 lower-Hettangian
239 0 lower-Hettangian
241.03 0.19148491 lower-Hettangian
245 0.08528757 lower-Hettangian
249.1 0 lower-Hettangian
252 0.28336275 lower-Hettangian
255 0 lower-Hettangian
259.5 0 lower-Hettangian
263.2 0.11941185 lower-Hettangian
267.1 0 lower-Hettangian
271 0 lower-Hettangian
275 0 lower-Hettangian
279 0.36533796 lower-Hettangian
282 0.09414858 upper-Hettangian
286 0.5558097 upper-Hettangian
290 0.38184862 upper-Hettangian
294 0.1641992 upper-Hettangian
298 0.11979282 upper-Hettangian
302 0.16174032 upper-Hettangian
312 0 upper-Hettangian
322 0.21270885 upper-Hettangian

333 0.08711916 upper-Hettangian
342 0.27298681 upper-Hettangian
348 0.47240743 upper-Hettangian
356 0.31754261 upper-Hettangian
366 0.2982428 upper-Hettangian
375 0.39690318 upper-Hettangian
384 0.47134035 upper-Hettangian
393 0.36377763 upper-Hettangian
402 0.17061947 upper-Hettangian
411 0.30656529 upper-Hettangian
420 0.58691452 upper-Hettangian

height	I/Ca ratios	stage	Italcementi
1.5	0.11638436	Rhaetian	
2.5	0.4147545	Rhaetian	
3.5	0.48662269	Rhaetian	
4.5	0.21234732	Rhaetian	
4.6	0.22124713	Rhaetian	
4.8	0.18699916	Rhaetian	
5	0.28009251	Rhaetian	
5.2	0.18705932	Rhaetian	
6	0.2511927	Rhaetian	
6.4	0.35096098	Rhaetian	
6.7	0.37493768	Rhaetian	
7.4	0.68923119	Rhaetian	
7.5	0.24974417	Rhaetian	
7.6	0.15222062	Rhaetian	
7.7	0.17262048	Rhaetian	
7.9	0.22184572	Rhaetian	
8.3	0.25365962	Rhaetian	
8.4	0.29917112	Rhaetian	
8.5	0.24714651	Rhaetian	
15	0.13219031	Rhaetian	
20.5	0.03130065	Rhaetian	
23.9	0.42906931	Rhaetian	
30.1	0	Rhaetian	
35.4	0.43851522	Rhaetian	
40.1	0.00652526	Rhaetian	
41.1	0.06208281	Rhaetian	
43.1	0.61769158	Rhaetian	
43.5	0.51095725	Rhaetian	
43.6	0.3043029	Rhaetian	
43.7	0.37933989	Rhaetian	
43.8	0.32461183	Rhaetian	
43.9	0.32562269	Rhaetian	
44.2	0.31032334	Rhaetian	
44.5	0.35676672	Rhaetian	
45.5	1.4293723	Rhaetian	
46	0.54052791	Rhaetian	
46.1	0.278198	Hettangian	
46.5	0.30434453	Hettangian	
46.8	0.3156856	Hettangian	
47	0.33077032	Hettangian	
47.8	0.27103742	Hettangian	
48	0.26484055	Hettangian	
49.9	0.1585904	Hettangian	
53	0.11168259	Hettangian	
56.6	0.27941121	Hettangian	
60.7	0.21001301	Hettangian	

61	0.42809386	Hettangian
65.3	0.26825504	Hettangian
70	0.01185157	Hettangian
72.9	0.38574635	Hettangian
77	0.01708235	Hettangian
80	0.67437623	Hettangian
84.8	0.03429122	Hettangian
86.5	0.09389566	Hettangian
90.2	0	Hettangian
93	0.10102217	Hettangian
95.1	0	Hettangian
98.3	0.14356164	Hettangian
100.6	0	Hettangian

height	I/Ca ratios	stage
0.5	0.17553944	Rhaetian
2.8	0.70584274	Rhaetian
6.8	0.33071266	Rhaetian
9.8	0.27584429	Rhaetian
16	1.92757485	Rhaetian
20	0.58230292	Rhaetian
24	0	Rhaetian
30.8	0.76108244	Rhaetian
34	0.34825307	Rhaetian
41.9	0.18360507	Rhaetian
46.9	0.39625049	Rhaetian
48	0.36897473	Rhaetian
50.3	0.34138592	Rhaetian
51.3	0.32985263	Rhaetian
53.5	0.44993818	Rhaetian
55	0.67721403	Rhaetian
57	0.61993325	Rhaetian
57.6	0.38564853	Rhaetian
57.9	0.04191127	Rhaetian
60.7	0.10373532	Rhaetian
64.7	0.26250336	lower-Hettangian
66.2	0.21074568	lower-Hettangian
67.7	0.26016131	lower-Hettangian
71.4	0.14259066	lower-Hettangian
79	0.47159107	lower-Hettangian
86	0	lower-Hettangian
90	0.34901169	lower-Hettangian
95	0	lower-Hettangian
100.6	0.18866823	lower-Hettangian
103.5	0.34182784	lower-Hettangian
106.5	0.36518116	lower-Hettangian
108	0.03116112	lower-Hettangian
110	0.58361463	lower-Hettangian
112	0	lower-Hettangian
115	0.92470904	lower-Hettangian
118	0.03855055	lower-Hettangian
122	0.12578035	lower-Hettangian
126	0.02515991	lower-Hettangian
129	0	lower-Hettangian
132	0	lower-Hettangian
135	0	lower-Hettangian
138	0	lower-Hettangian
141	0	lower-Hettangian
144	0	lower-Hettangian
147	0	lower-Hettangian
150	0.03213083	lower-Hettangian

154 0.25153159 lower-Hettangian
157 0.04376268 lower-Hettangian
160 0.03796047 lower-Hettangian
163 0.16692414 lower-Hettangian
166 0 lower-Hettangian
169 0 lower-Hettangian
172 0 lower-Hettangian
175 0 lower-Hettangian
178 0 lower-Hettangian
181 0 lower-Hettangian
184 0.13907046 lower-Hettangian
186.8 0 lower-Hettangian
190 0 lower-Hettangian
193 0 lower-Hettangian
196 0 lower-Hettangian
200 0 lower-Hettangian
203 0 lower-Hettangian
206 0.09016142 lower-Hettangian
209 0.10369134 lower-Hettangian
212 0.1417996 lower-Hettangian
215 0.11684494 lower-Hettangian
218 0.19509616 lower-Hettangian
222 0.1011641 lower-Hettangian
227 0 lower-Hettangian
230 0.04381678 lower-Hettangian
233 0.21926847 lower-Hettangian
236 0 lower-Hettangian
239 0 lower-Hettangian
241.03 0.19148491 lower-Hettangian
245 0.08528757 lower-Hettangian
249.1 0 lower-Hettangian
252 0.28336275 lower-Hettangian
255 0 lower-Hettangian
259.5 0 lower-Hettangian
263.2 0.11941185 lower-Hettangian
267.1 0 lower-Hettangian
271 0 lower-Hettangian
275 0 lower-Hettangian
279 0.36533796 lower-Hettangian
282 0.09414858 upper-Hettangian
286 0.5558097 upper-Hettangian
290 0.38184862 upper-Hettangian
294 0.1641992 upper-Hettangian
298 0.11979282 upper-Hettangian
302 0.16174032 upper-Hettangian
312 0 upper-Hettangian
322 0.21270885 upper-Hettangian

333 0.08711916 upper-Hettangian
342 0.27298681 upper-Hettangian
348 0.47240743 upper-Hettangian
356 0.31754261 upper-Hettangian
366 0.2982428 upper-Hettangian
375 0.39690318 upper-Hettangian
384 0.47134035 upper-Hettangian
393 0.36377763 upper-Hettangian
402 0.17061947 upper-Hettangian
411 0.30656529 upper-Hettangian
420 0.58691452 upper-Hettangian