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### Journal of Theoretical Biology

journal homepage: www.elsevier.com/locate/yjtbi





## Assessing the impact of insecticide-treated nets in the face of insecticide resistance on malaria control

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#### ARTICLE INFO

# Keywords: Mosquito resistance to insecticides Malaria control Asymptomatic infection Human behavior Infectious disease model

### ABSTRACT

The mosquito-borne disease, malaria, continues to impose a devastating health and economic burden worldwide. In malaria-endemic areas, insecticide-treated nets (ITNs) have been useful in curtailing the burden of the disease. However, mosquito resistance to insecticides, decay in ITN efficacy, net attrition, etc., undermine the effectiveness of ITNs in combatting malaria. In this study, mathematical models that account for asymptomatic infectious humans (through a partially immune class or a separate asymptomatic infectious class), insecticide resistance, and decay in ITN efficacy are proposed and analyzed. Analytical and numerical results of the models when ITN efficacy is constant show that there are parameter regimes for which a backward bifurcation occurs. Local and global sensitivity analyses are performed to identify parameters (some of which are potential targets for disease control) with the most significant influence on the control reproduction  $(R_c)$  and disease prevalence. These influential parameters include the maximum biting rate of resistant mosquitoes, ITN coverage, initial ITN efficacy against sensitive mosquitoes, the probability that an infectious mosquito (human) infects a susceptible human (mosquito), and the rate at which adult mosquitoes develop (lose) resistance to insecticides. Simulations of the models show that accounting for asymptomatic infectious humans through a separate class, or not accounting for the decay in ITN efficacy leads to an underestimation of disease burden. In particular, if the initial efficacy of ITNs against sensitive and resistance mosquitoes is 96%, the minimum ITN coverage required to reduce R<sub>c</sub> below one (and hence, contain malaria) is approximately 11% (27%) lower when ITN efficacy is averaged (constant) for a model with a separate asymptomatic class. For the model with a partially immune class and decaying ITN efficacy, reducing  $R_c$  below one is impossible even if the entire populace uses ITNs. The study shows that replacing ITNs before their prescribed lifespans, or designing ITNs with longer lifespans is important for malaria control. Furthermore, the study shows that piperonyl butoxide (PBO) ITNs (which inhibit or reverse insecticide resistance) outperform regular ITNs in malaria control. Hence, prospects for effectively controlling malaria are enhanced by widespread use of high quality ITNs (e.g. PBO ITNs), especially if the useful lifespans of the ITNs are long enough and the ITNs are replaced before the end of their useful lifespans.

### 1. Introduction

Malaria continues to wreak havoc on human health and economies around the world, especially in sub-Saharan Africa (SSA) (World Health Organization, 2021; Gallup and Sachs, 2001; Sachs and Malaney, 2002). In 2020, it was implicated for about 241 million clinical illness episodes (representing a 6.2% increase from 2019) and 627,000 deaths (representing a 12.4% increase from 2019) worldwide (World Health Organization, 2021). Disruptions in malaria control programs caused by the 2019 coronavirus (COVID-19) pandemic accounted for an estimated 68% of the additional malaria-related deaths reported in 2020 (World Health Organization, 2021). These staggering figures suggest that the global gains in the fight against malaria and the progress made

towards achieving the Global Technical Strategy (GTS) for malaria's goal of at least a 90% reduction in malaria incidence and mortality by 2030 are being reversed, especially in high-burden malaria countries.

Insecticide-treated nets (ITNs), i.e., long-lasting insecticidal nets (with an average lifespan of three years World Health Organization, 2011; World Health Organization et al., 2013) and natural nets that have been treated with insecticides have been instrumental in the fight against malaria (Lengeler, 2004a; Trape et al., 2011; Bhatt et al., 2015; Lindsay et al., 2021). In particular, ITNs have contributed in reducing the number of malaria cases by approximately 68% since 2000 (Bhatt et al., 2015). They shield those who sleep underneath

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them from mosquito-bites (by preventing mosquitoes from reaching them). Pyrethroids (i.e., the main insecticide that is commonly used on ITNs) can kill mosquitoes that come in contact with ITNs even at low doses. By killing adult mosquitoes (thereby limiting the available adult mosquito population), the protection received by individuals who sleep beneath ITNs within a community is extended to other members of the community, especially in communities with high ITN-use (Maxwell et al., 2002; Killeen and Smith, 2007). Also, high ITN-coverage can force mosquitoes that are unable to feed on humans to turn to other sources for blood meals (Takken and Verhulst, 2013; Iwashita et al., 2014; Stone and Gross, 2018). In 2020, 229 million ITNs were distributed to malaria-endemic nations, with approximately 91% going to SSA (World Health Organization, 2021). Furthermore, over 65% of SSA homes owned at least one ITN and approximately 43% of the population slept beneath an ITN in 2020 (World Health Organization, 2021). The protective efficacy of new ITNs is over 90%, but, this can fall below 70% over the effective lifespan of the ITNs and to about 50% for untreated nets or when all the insecticide in the net is lost (N'Guessan et al., 2007; Clarke et al., 2001). Although, ITNs have been embraced for malaria control and mitigation in many malaria endemic regions (owing to their cost-effectiveness and ease of use), their efficacy is threatened by several factors including decay in ITN efficacy, net attrition (Pulkki-Brännström et al., 2012; Briët et al., 2012; Diema et al., 2017), and mosquito resistance to insecticides used in ITNs (World Health Organization, 2013; Mawejje et al., 2013; Churcher et al., 2016; Ranson and Lissenden, 2016; Namias et al., 2021).

Resistance of mosquitoes to pyrethroid and other insecticides used in ITNs poses a significant problem to the long-term management and control of malaria (World Health Organization, 2013; Brogdon and McAllister, 1998.; Mawejje et al., 2013). According to the 2021 World Health Organization Report on Malaria, resistance to pyrethroids is widespread globally and nearly three-quarters of nations reporting resistance to pyrethroids experienced high intensity resistance in 2020 (World Health Organization, 2021). Resistance enables mosquitoes to evade or overcome vector control measures that rely on the use of insecticides. In particular, as the efficacy of ITNs decays, mosquitoes modify their behavior to evade the repelling and killing effect of pesticides used in the nets, thereby enhancing malaria transmission (Ngonghala et al., 2020). Decay in ITN efficacy, net attrition, mosquito resistance to insecticides, coupled with the decline in access to, and ITN-use in SSA since 2017 (World Health Organization, 2021) undermine the GTS and decade-long achievement in malaria control. This necessitates extensive research to have a better understanding of mosquito resistance to insecticides and to inform effective mosquito and insecticide resistance management strategies (Brooke, 2008; Namias et al., 2021). Synergists such as piperonyl-butoxide (PBO), which are combined with pyrethroids in the new pyrethroid-PBO nets are useful in managing insecticide resistance. Pyrethroid-PBO nets are designed to overcome pyrethroid-resistance and to improve the ability of ITNs to kill pyrethroid resistant mosquitoes. Hence, including PBO in ITNs improves their capability of killing resistant mosquitoes. Pyrethroid-PBO nets have been shown to lower malaria prevalence by reducing (increasing) mosquito blood feeding (mortality) in high pyrethroid resistant environments (Gleave et al., 2018; Martin et al., 2021). Exposure to synergists has been employed to boost the effectiveness of some insecticides (Bingham et al., 2011; Dadzie et al., 2017). Also, the use of pyrethroid-PBO nets has been able to restore insecticide susceptibility in mosquitoes in some countries (World Health Organization, 2021).

The impact of ITNs and the decay in ITN efficacy on the control of malaria have been assessed through mathematical models (Killeen et al., 2007; Chitnis et al., 2010; Okell et al., 2012b; Briët et al., 2012; Agusto et al., 2013; Ngonghala et al., 2014, 2016). Also, mathematical models to assess the effects of mosquito resistance to insecticides have been proposed and analyzed (Barbosa and Hastings, 2012; Birget and Koella, 2015; Barbosa et al., 2018; Mohammed-Awel et al., 2018,

2020; Ngonghala et al., 2020). Although there is overwhelming evidence of asymptomatic malaria transmission (i.e., transmission of the malaria disease by humans with malaria parasite burden, but who do not exhibit clinical symptoms of malaria) in different parts of the world (Bousema et al., 2004; Eke et al., 2006; Lindblade et al., 2013; Lopez-Perez et al., 2015; Sáenz et al., 2017; Tiono et al., 2014; Jiram et al., 2019; Chaumeau et al., 2019; Ramaswamy et al., 2020; Sumner et al., 2021), only few mathematical models have attempted to quantify the impact of asymptomatic infection on malaria spread and persistence through a separate asymptomatic human carrier class (e.g., Filipe et al. (2007) and Aguilar and Gutierrez (2020)). The existence of asymptomatic infectious humans complicates disease control efforts. Specifically, since asymptomatic infectious humans do not exhibit clinical symptoms of malaria and hence, cannot be detected through fever-based methods (Galatas et al., 2016; Ramaswamy et al., 2020), they serve as hidden reservoirs for malaria parasites that can be picked up my mosquitoes and conveyed to other humans. Hence, they are critical in the spread and persistence of malaria (Alves et al., 2002; Bousema et al., 2004; Eke et al., 2006; Lindblade et al., 2013; Dal-Bianco et al., 2007; Laishram et al., 2012). In particular, according to Lindblade et al. (2013), four to five times more asymptomatic than symptomatic malaria cases occur in malaria endemic regions. Also, Okell et al. (2012a), pointed out that subpatent cases (that are common in areas where malaria transmission is low) can account for up to half of all malaria transmissions. Thus, understanding the impact of asymptomatic infection on the dynamics of malaria in the face of decaying ITN efficacy and insecticide resistance is important for malaria control.

In this study, a mathematical model that accounts for asymptomatic infectious humans through a separate class, decay in ITN efficacy over time, and mosquito resistance to insecticides is proposed and analyzed rigorously. The model is used to assess the effects of (regular and PBO) ITNs, decay in ITN efficacy over time, disease transmission by asymptomatic infectious humans, and how mosquito resistance to insecticides affects malaria control efforts in endemic regions. Furthermore, the model is compared with one which fails to account for asymptomatic human transmission and one with partially immune individuals who transmit malaria, albeit a lower extent. To our knowledge, this is the first study that accounts for interactions between the decay in ITN efficacy over time and mosquito resistance to insecticides, as well as a separate asymptomatic infectious human class in a single framework.

### 2. The model with an infectious partially immune human class

In this section a susceptible–exposed–infectious–recovered – susceptible model for the dynamics of malaria that accounts for ITN-use as a control measure, mosquito resistance to insecticides, and a partially immune human class that contributes in disease transmission is proposed and studied. The model consists of an autonomous system of differential equations with no separate asymptomatic infectious class.

### 2.1. Model formulation

The human population is split into susceptible  $(S_h)$ , exposed  $(E_h)$ , infectious  $(I_h)$ , and partially immune  $(R_h)$  classes, so that the total human population  $(N_h)$  is given by  $N_h = S_h + E_h + I_h + R_h$ . Partially immune humans are recovered individuals, who are protected from severe infection (e.g., through natural immunity acquired from repeated exposure to malaria), but not fully protected from mild infection and hence contribute in spreading malaria. The mosquito population is split into sensitive susceptible  $(S_s)$ , resistant susceptible  $(S_r)$ , sensitive exposed  $(E_s)$ , resistant exposed  $(E_r)$ , sensitive infectious  $(I_s)$ , and resistant infectious  $(I_r)$  classes, so that the total mosquito population  $(N_v)$  is  $N_v = S_s + E_s + I_s + S_r + E_r + I_r$ . The flow-diagram of the model is presented

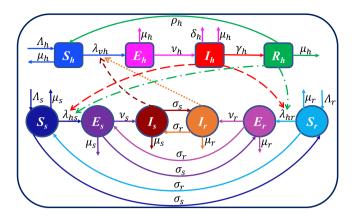


Fig. 1. Conceptual framework of the model depicting transitions of humans and mosquitoes between different epidemiological stages (solid lines) and interactions between humans and mosquitoes that result in infectious and partially immune humans infecting susceptible mosquitoes (long dashed and dash-dotted lines, respectively) and infectious sensitive and resistant mosquitoes infecting susceptible humans (short dashed and dotted lines, respectively). The human population is split into susceptible  $(S_h)$  exposed  $(E_h)$ , infectious  $(I_h)$ , and partially immune  $(R_h)$  compartments, while the mosquito population comprises susceptible sensitive and resistant  $(S_s$  and  $S_r)$ , exposed sensitive and resistant  $(E_s$  and  $E_r)$ , and infectious sensitive and resistant  $(I_s$  and  $I_r)$  compartments. The parameter  $A_s = (1-\theta)A_v$  and  $A_r = \theta A_v$ . Descriptions of the parameters are presented in the text and in Table 1.

in Fig. 1. Using the schematics in Fig. 1, the variable descriptions, and the parameter descriptions in Table 1 yields the model:

$$\dot{S}_{h} = \Lambda_{h} + \rho_{h} R_{h} - (\lambda_{vh} + \mu_{h}) S_{h}, 
\dot{E}_{h} = \lambda_{vh} S_{h} - (\mu_{h} + \nu_{h}) E_{h}, 
\dot{I}_{h} = \nu_{h} E_{h} - (\delta_{h} + \mu_{h} + \gamma_{h}) I_{h}, 
\dot{R}_{h} = \gamma_{h} I_{h} - (\mu_{h} + \rho_{h}) R_{h}, 
\dot{S}_{s} = (1 - \theta) \Lambda_{v} + \sigma_{r} S_{r} - (\lambda_{hs} + \sigma_{s} + \mu_{s}) S_{s}, 
\dot{E}_{s} = \lambda_{hs} S_{s} + \sigma_{r} E_{r} - (\sigma_{s} + \mu_{s} + \nu_{s}) E_{s}, 
\dot{I}_{s} = \nu_{s} E_{s} + \sigma_{r} I_{r} - (\sigma_{s} + \mu_{s}) I_{s}, 
\dot{S}_{r} = \theta \Lambda_{v} + \sigma_{s} S_{s} - (\lambda_{hr} + \sigma_{r} + \mu_{r}) S_{r}, 
\dot{E}_{r} = \lambda_{hr} S_{r} + \sigma_{s} E_{s} - (\sigma_{r} + \mu_{r} + \nu_{r}) E_{r}, 
\dot{I}_{r} = \nu_{r} E_{r} + \sigma_{s} I_{s} - (\sigma_{r} + \mu_{r}) I_{r},$$
(2.1)

where  $\Lambda_s=(1-\theta)\Lambda_v, \Lambda_r=\theta\Lambda_v$ , and the dynamics of the total human and mosquito populations are given by:

$$\dot{N}_h = \Lambda_h - \mu_h N_h - \delta_h I_h, \quad \dot{N}_v = \Lambda_v - \mu_s (S_s + E_s + I_s) - \mu_r (S_r + E_r + I_r).$$
 (2.2)

For notational convenience, we set

$$\begin{split} A_1 &= \mu_h + \nu_h, A_2 = \delta_h + \mu_h + \gamma_h, A_3 = \mu_h + \rho_h, B_s = \sigma_s + \mu_s, \\ \tilde{B}_s &= B_s + \nu_s, B_r = \sigma_r + \mu_r, \tilde{B}_r = B_r + \nu_r. \end{split} \tag{2.3}$$

The forces of infection ( $\lambda_{vh}$  and  $\lambda_{hj}$ ,  $j \in \{r,s\}$ ) are modeled as in Ngonghala et al. (2014, 2016, 2020), with the assumption that humans are bitten by sensitive and resistant mosquitoes at rates  $\beta_s$  and  $\beta_r$ , respectively, Wat'senga et al. (2018) and Fuseini et al. (2019). Thus,

$$\lambda_{vh} = \frac{p_{vh}(\beta_s I_s + \beta_r I_r)}{N_h} \quad \text{and} \quad \lambda_{hj} = \frac{p_{hv}\beta_j(I_h + \epsilon R_h)}{N_h}, \ j \in \{r, s\}, \ (2.4)$$

where  $\beta_j=\beta_{j1}-(\beta_{j1}-\beta_{j0})\varepsilon_{\beta_j}b_0$  is the biting rate of resistant and sensitive mosquitoes,  $0\leq b_0\leq 1$  is the ITN coverage (i.e., the fraction of persons who sleep under ITNs),  $0\leq \varepsilon_{\beta_j}\leq 1$  is the personal protective efficacy of ITNs against sensitive and resistant mosquitoes, i.e., the potential of ITNs to shield humans underneath them from sensitive mosquito bites (j=s) or resistant mosquito bites (j=r). The parameters  $p_{vh},p_{hv},\beta_{j1},\beta_{j0},j\in\{r,s\}$  are described in Table 1. It should be mentioned that the biting rate of mosquitoes is maximum

(i.e.,  $\beta_j=\beta_{j1}$ ) if  $b_0=0$  or  $\varepsilon_{\beta_j}=0$  and minimum (i.e.,  $\beta_j=\beta_{j0}$ ) if  $b_0=1$  and  $\varepsilon_{\beta}=1$ . As in Ngonghala et al. (2014, 2016), the mortality rates  $\mu_j, j\in\{r,s\}$  are given by  $\mu_j=\mu_{j0}+\mu_{j1}\varepsilon_{\mu_j}b_0$ , where  $0\leq \varepsilon_{\mu_j}\leq 1$  is the sensitive (j=s) and resistant (j=r) mosquito killing efficacy of ITNs (i.e., the potential of ITNs to kill sensitive or resistant mosquitoes that land on them) and  $\mu_{jk}, k\in\{0,1\}$  are as defined in Table 1. The mosquito death rate is minimum (i.e.,  $\mu_j=\mu_{j0}$ ) if  $b_0=0$  or  $\varepsilon_{\mu_j}=0$  and maximum (i.e.,  $\mu_j=\mu_{j0}+\mu_{j1}$ ) if  $b_0=1$  and  $\varepsilon_{\mu_j}=1$ . For simplicity, we set  $\varepsilon_{\beta_j}=\varepsilon_{\mu_j}=\varepsilon_{j0}$ , where  $\varepsilon_{j0}$  is constant in Sections 2 and 3.

### 2.2. Analytical results

In this section, we establish the well-posedness, positivity, and boundedness of solutions of the model system (2.1), compute the control reproduction number and hence the basic reproduction number, and establish the existence and stability of equilibrium points, as well as the existence of a sub-critical (backward) bifurcation.

### 2.2.1. Well-posedness, positivity, and boundedness of solutions

For a full description of the model (2.1), initial conditions of the form  $X(0) = (S_h(0), E_h(0), I_h(0), R_h(0), S_s(0), E_s(0), I_s(0), S_r(0), E_r(0), I_r(0))$ , where  $N_h(0) = S_h(0) + E_h(0) + I_h(0) + R_h(0)$  and  $N_v(0) = S_s(0) + E_s(0) + I_s(0) + S_r(0) + E_r(0) + I_r(0)$  must be prescribed in an epidemiologically-feasible region. Since the state variables  $(S_h, E_h, I_h, R_h, S_s, E_s, I_s, S_r, E_r, \text{ and } I_r)$  represent human and mosquito populations, both the state variables and the corresponding initial conditions  $X(0) = (S_h(0), E_h(0), I_h(0), R_h(0), S_s(0), E_s(0), I_s(0), S_r(0), E_r(0), I_r(0))$  are non-negative. Hence, with given initial conditions of the form X(0) in the epidemiologically-feasible region:

$$\Omega = \{ (S_h, E_h, I_h, R_h, S_s, E_s, I_s, S_r, E_r, I_r) \in \mathbb{R}_+^{10} : 0 \le S_h + E_h + I_h + R_h \\ \le \Lambda_h / \mu_h, 0 \le S_s + E_s + I_s + S_r + E_r + I_r \le \Lambda_v / \mu_v \},$$

where  $\mu_v = min(\mu_s, \mu_r)$ , it can be verified (using the approach in Ngonghala et al. (2020) and methods in Jordan and Smith (2007) and Agarwal and O'Regan (2008)) that the model (2.1) is mathematically well-posed within the epidemiologically feasible region ( $\Omega$ ). That is, it can be verified that the model (2.1) has a unique solution that satisfies the initial conditions  $\forall t > 0$ , and that  $N_h(t) = 0$  ( $N_v(t) = 0$ )  $\forall t$  if  $N_h(0) = 0$  ( $N_v(0) = 0$ ), and  $N_h(t) > 0$  ( $N_v(t) > 0$ )  $\forall t$  if  $N_h(0) > 0$  ( $N_v(0) > 0$ ).

Since the dynamics of the total human population is described by the equation  $\dot{N}_h = \Lambda_h - \mu_h N_h - \delta_h I_h, \dot{N}_h \leq \Lambda_h - \mu_h N_h$ . Using an integrating factor or separating variables, we obtain:  $N_h(t) \leq \frac{\Lambda_h}{\mu_h} + \left(N_h(0) - \frac{\Lambda_h}{\mu_h}\right)e^{-\mu_h t}$ , where  $N_h(0)$  is the initial total human population. On the other hand, if  $\mu_v = \min(\mu_s, \mu_r)$ , then the population dynamics

of the total mosquito population described by the second equation of Eqs. (2.2) becomes:  $\dot{N}_v \leq \Lambda_v - \mu_v N_v$ . Therefore,  $N_v(t) \leq \frac{\Lambda_v}{\mu_v} + \left(N_v(0) - \frac{\Lambda_v}{\mu_v}\right)e^{-\mu_v t}$ , where  $N_v(0)$  is the initial total mosquito population. Clearly,  $N_h(t) \leq \Lambda_h/\mu_h$  and  $N_v(t) \leq \Lambda_v/\mu_v$  as  $t \to \infty$ . Hence, the epidemiologically feasible region  $\Omega = \{(S_h, E_h, I_h, R_h, S_s, E_s, I_s, S_r, E_r, I_r) \in \mathbb{R}^{10}_+ : 0 \leq S_h + E_h + I_h + R_h \leq \Lambda_h/\mu_h, 0 \leq S_s + E_s + I_s + S_r + E_r + I_r \leq \Lambda_v/\mu_v\}$  is positively invariant and attracting with respect to the model system (2.1). That is, any solution of Eqs. (2.1) with initial condition within  $\Omega$  remains trapped within  $\Omega$  for t > 0.

### 2.2.2. Disease-free equilibrium and the control reproduction number

The disease-free equilibrium  $(S_h^0, E_h^0, I_h^0, R_h^0, S_s^0, E_s^0, I_s^0, S_r^0, E_r^0, I_r^0)$ , of the model (2.1), obtained by setting the left-hand sides (LHS) and the disease related variables of Eqs. (2.1) to zero is

$$(S_h^0, E_h^0, I_h^0, R_h^0, S_s^0, E_s^0, I_s^0, S_r^0, E_r^0, I_r^0) = \left(\frac{A_h}{\mu_h}, 0, 0, 0, \frac{B_1}{B_3}, 0, 0, \frac{B_2}{B_3}, 0, 0\right), \tag{2.5}$$

where  $B_1 = \Lambda_s B_r + \Lambda_r \sigma_r$ ,  $B_2 = \Lambda_r B_s + \Lambda_s \sigma_s$ , and  $B_3 = B_s B_r - \sigma_r \sigma_s = \mu_r \mu_s + \mu_r \sigma_s + \mu_s \sigma_r$ . Local asymptotic stability of the disease-free equilibrium is established using the next generation matrix operator, which doubles as the process for deriving the reproduction number

Table 1
Table of parameter values and ranges used for the simulations of System (2.1). The dimension H, D, and M represent human, day, and mosquito, respectively. Dimensions are enclosed in parentheses at the end of parameter descriptions and excluded for dimensionless parameters.

Parameter	Description of parameter (dimension)	Value	Range	Reference		
$\Lambda_h$	Recruitment rate of humans (HD <sup>-1</sup> )	$5.1 \times 10^{-2}$	$[3.65, 9.13] \times 10^{-2}$	The world fact book (2022a)		
$\mu_h$	Human natural mortality rate $(D^{-1})$	$4.1 \times 10^{-5}$	$[3.3, 5.5] \times 10^{-5}$	The world fact book (2022b)		
$v_h$	Rate at which exposed humans	$7.1 \times 10^{-2}$	$[6.7, 20] \times 10^{-2}$	Mehlhorn (2001) and		
	become infectious $(D^{-1})$			Chitnis et al. (2008a)		
$\gamma_h$	Recovery rate of infectious humans $(D^{-1})$	$1.3 \times 10^{-2}$	$[1.4, 17] \times 10^{-3}$	Kakkilaya (2022) and Chitnis et al. (2008a)		
$\delta_h$	Malaria-induced death rate	$9.0 \times 10^{-5}$	$[0, 4.1] \times 10^{-4}$	Chitnis et al. (2008b)		
	for infectious humans $(D^{-1})$					
$ ho_h$	Rate at which partially immune	$8.3 \times 10^{-3}$	$[5.5, 1100] \times 10^{-5}$	Ngonghala et al. (2014) and Chitnis et al. (2008a)		
	humans loss immunity $(D^{-1})$					
$\theta$	Proportion reduction in sensitive mosquito	$1.0 \times 10^{-1}$	[0, 1]			
	recruitment due to resistance.					
$\Lambda_v$	Mosquito recruitment rate $(MD^{-1})$	83.3	$[0.333, 1.4] \times 10^3$	Ngonghala et al. (2020)		
$\nu_s$	Rate at which exposed sensitive	$7.7 \times 10^{-2}$	$[2.9, 33] \times 10^{-2}$	Macdonald et al. (1957) and Chitnis et al. (2008a)		
	mosquitoes become infectious $(D^{-1})$					
$\nu_r$	Rate at which exposed resistant	$7.7 \times 10^{-2}$	$[2.9, 33] \times 10^{-2}$	Macdonald et al. (1957) and Chitnis et al. (2008a)		
	mosquitoes become infectious $(D^{-1})$					
$\sigma_{_S}$	Mutation rate of sensitive mosquitoes $(D^{-1})$	$6.0 \times 10^{-1}$	[0, 1]	Teboh-Ewungkem et al. (2009)		
$\sigma_r$	Mutation rate of resistant mosquitoes $(D^{-1})$	$5.0 \times 10^{-1}$	[0, 1]	Wang and Zhao (2008a) and Chitnis et al. (2006)		
$\mu_{r0}$	Natural death rate of resistant mosquitoes $(D^{-1})$	$2.5 \times 10^{-2}$	$[1.2, 14] \times 10^{-2}$	Osoro et al. (2022), Davidson and Draper (1953)		
				and Giles and Warrel (1993)		
$\mu_{s0}$	Natural death rate of sensitive mosquitoes $(D^{-1})$	$3.3 \times 10^{-2}$	$[1.5, 14] \times 10^{-2}$	Osoro et al. (2022), Davidson and Draper (1953)		
				and Giles and Warrel (1993)		
$\mu_{r1}$	ITN death rate of resistant mosquitoes $(D^{-1})$	$2.5 \times 10^{-2}$	$[1.2, 14] \times 10^{-2}$	Osoro et al. (2022) and Lines et al. (1987)		
$\mu_{s1}$	ITN death rate of sensitive mosquitoes $(D^{-1})$	$3.3 \times 10^{-2}$	$[1.5, 14] \times 10^{-2}$	Osoro et al. (2022) and Lines et al. (1987)		
$p_{hv}$	Human to mosquito transmission probability	$4.8 \times 10^{-1}$	$[7.2, 64] \times 10^{-2}$	Chitnis et al. (2008b), Boyd (1949), Smalley and		
				Sinden (1977) and Nedelman (1984)		
$p_{vh}$	Mosquito to human transmission probability	$2.2 \times 10^{-2}$	$[1.0, 27] \times 10^{-2}$	Chitnis et al. (2008b) and Nedelman (1985)		
$\epsilon$	Immune human to mosquito transmission	0.1	[0, 1]	Chitnis et al. (2008b)		
	probability modification factor					
$\beta_{s1}$	Maximum sensitive mosquito biting rate $(D^{-1})$	$2.0 \times 10^{-1}$	[0, 2]	Molineaux et al. (1979), Chitnis et al. (2008b),		
				Gupta et al. (1994) and Bhatt et al. (2015)		
$\beta_{r1}$	Maximum resistant mosquito biting rate $(D^{-1})$	$6.0 \times 10^{-1}$	[0, 2]	Molineaux et al. (1979), Chitnis et al. (2008b),		
				Gupta et al. (1994) and Bhatt et al. (2015)		
$\beta_{s0}$	Minimum sensitive mosquito biting rate $(D^{-1})$	$1.0 \times 10^{-2}$	$[1.0, 100] \times 10^{-3}$	Mwangangi et al. (2013)		
$\beta_{r0}$	Minimum resistant mosquito biting rate $(D^{-1})$	$3.0 \times 10^{-2}$	$[1.0, 100] \times 10^{-3}$	Mwangangi et al. (2013)		
$b_0$	ITN coverage, i.e., the proportion of humans	$4.3 \times 10^{-1}$	$[1.0, 100] \times 10^{-2}$	World Health Organization (2021)		
	who are protected from mosquito-bites by ITNs	Varies	$[1.0, 100] \times 10^{-2}$	World Health Organization (2021)		
$\epsilon_{s0}$	Efficacy of ITNs against sensitive mosquitoes	Varies	$[1.0, 100] \times 10^{-2}$	N'Guessan et al. (2007)		
$\epsilon_{r0}$	Efficacy of ITNs against resistant mosquitoes	Varies	$[1.0, 100] \times 10^{-2}$			
p	Modification factor the efficacy $\epsilon_{r0}$	0.5	[0, 1]	N'Guessan et al. (2007)		
T	Useful life of ITNs	$3 \times 365$	$[0.5, 3] \times 365$	World Health Organization (2011) and		
				Pulkki-Brännström et al. (2012)		

of the model (2.1) (Diekmann et al., 1990; Driessche and Watmough, 2002). The matrices of new infections ( $\mathscr{F}$ ) and transitions ( $\mathscr{F}$ ), and the next generation matrix ( $\mathscr{F}\mathscr{V}^{-1}$ ) are given in Section S1.1 of the online supplementary information (SI). If we set  $B_4 = (\sigma_r + \mu_r + \nu_r)(\mu_s + \nu_s) + \sigma_s(\mu_r + \nu_r)$ ,  $B_5 = (B_r \nu_s + \tilde{B}_s \nu_r)\sigma_r$ ,  $B_6 = (B_s \nu_r + \tilde{B}_r \nu_s)\sigma_s$ ,  $B_7 = B_r \tilde{B}_r \nu_s + \nu_r \sigma_r \sigma_s$ ,  $B_8 = B_s \tilde{B}_s \nu_r + \nu_s \sigma_s \sigma_r$ ,  $B_0 = \beta_s^2 B_1 B_7 + \beta_r^2 B_2 B_8 + \beta_r \beta_s (B_1 B_6 + B_2 B_5)$ ,  $\beta_j = \beta_{j1} - (\beta_{j1} - \beta_{j0})\varepsilon_{j0}b_0$ , and  $\mu_j = \mu_{j0} + \mu_{j1}\varepsilon_{j0}b_0$ ,  $j \in \{r, s\}$ , then the control reproduction number of the model (2.1), i.e., the spectral radius of the next generation matrix ( $\mathscr{F}\mathscr{V}^{-1}$ ) is:

$$R_{c} = \sqrt{\frac{p_{hv}p_{vh}\mu_{h}\nu_{h}B_{0}(A_{3} + \epsilon\gamma_{h})}{\Lambda_{h}A_{1}A_{2}A_{3}B_{3}^{2}B_{4}}}.$$
(2.6)

The control reproduction number  $(R_c)$  quantifies the average number of new malaria cases created by an infectious individual in a population in which a portion of the humans is protected by ITNs. It becomes the basic reproduction number  $(R_0)$  in the absence of ITN-usage (i.e., when  $b_0=0$ ) and other malaria control and mitigation measures. In particular, if  $b_0=0$ , then  $\beta_j=\beta_{j1}, \mu_j=\mu_{j0}, B_0=\beta_{s1}^2B_1B_7+\beta_{r1}^2B_2B_8+\beta_{r1}\beta_{s1}(B_1B_6+B_2B_5)$ , and  $B_j=\sigma_j+\mu_{j0}, j\in\{r,s\}$ . The following result is derived from Theorem 2 of Driessche and Watmough (2002):

**Theorem 2.1.** The disease-free equilibrium of the model system given by Eqs. (2.1) is locally-asymptotically stable if the reproduction number  $(R_c)$  is less than one and unstable if the reproduction number is greater than one.

According to Theorem 2.1, a slight increase in malaria cases will not result in malaria establishing itself in a community provided  $R_c$  is kept below unity and there is no backward (sub-critical) bifurcation.

### 2.2.3. Endemic equilibria and backward bifurcation

Endemic equilibria of the model (2.1) are obtained by setting the LHS of the system to zero and solving for each of the variables in terms of equilibrium values  $\lambda_{vh}^*$  and  $\lambda_{hi}^*$ , of the forces of infection  $\lambda_{vh}$  and  $\lambda_{hj}$ :

$$S_{h}^{*} = \frac{\Lambda_{h}A_{1}A_{2}A_{3}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, E_{h}^{*} = \frac{\Lambda_{h}A_{2}A_{3}\lambda_{vh}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = \frac{\Lambda_{h}v_{h}A_{3}\lambda_{vh}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = \frac{\Lambda_{h}v_{h}A_{3}\lambda_{vh}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = S_{h}^{*} + E_{h}^{*} + I_{h}^{*} + R_{h}^{*} = \frac{\Lambda_{h}(A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = S_{h}^{*} + E_{h}^{*} + I_{h}^{*} + R_{h}^{*} = \frac{\Lambda_{h}(A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + E_{h}^{*} + I_{h}^{*} + R_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{4}\lambda_{vh}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + A_{h}^{*}}, I_{h}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*} = \frac{S_{h}^{*} + I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*}}, I_{h}^{*} = \frac{I_{h}^{*} + I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{h}^{*}}{\mu_{h}A_{1}A_{2}A_{3} + I_{h}^{*}}, I_{$$

$$\lambda_{hj}^{*} = \beta_{j} p_{hv} \left( \frac{I_{h}^{*} + \epsilon R_{h}^{*}}{N_{h}^{*}} \right) = \frac{\beta_{j} p_{hv} \nu_{h} (A_{3} + \epsilon \gamma_{h}) \lambda_{vh}^{*}}{A_{1} A_{2} A_{3} + A_{5} \lambda_{vh}^{*}}, j \in \{s, r\},$$
 (2.8)

$$C_{2} = \frac{A_{1}A_{2}A_{3} \left\{ v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s}v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) + 2A_{5} \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) \right] + 3A_{5}^{2}B_{3} \right\}}{A_{5} \left\{ v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s}v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) + A_{5} \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) \right] + A_{5}^{2}B_{3} \right\}} (1 - \mathcal{R}_{2})$$

$$C_{1} = \frac{\left( A_{1}A_{2}A_{3} \right)^{2} \left( v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) + 3A_{5}B_{3} \right)}{A_{5} \left\{ v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s}v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) + A_{5} \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) \right] + A_{5}^{2}B_{3} \right\}} (1 - \mathcal{R}_{1})$$

$$C_{0} = \frac{\left( A_{1}A_{2}A_{3} \right)^{3}B_{3}}{A_{5} \left\{ v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s}v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) + A_{5} \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) \right] + A_{5}^{2}B_{3} \right\}} (1 - \mathcal{R}_{c}^{2}),$$

$$\mathcal{R}_{2} = \frac{p_{vh}v_{h}p_{hv}} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s} \left( \beta_{s}B_{11} + \beta_{r}B_{12} \right) v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) + A_{5}^{2}B_{3} \right]}{A_{h}A_{1}A_{2}A_{3}B_{3}B_{4} \left\{ v_{h}p_{hv}} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s}v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) + A_{5}^{2} \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) \right] + 3A_{5}^{2}B_{3} \right\}},$$

$$\mathcal{R}_{1} = \frac{p_{vh}v_{h}p_{hv}} \left( A_{3} + \epsilon \gamma_{h} \right) \left[ \beta_{r}\beta_{s} \left( \beta_{s}B_{11} + \beta_{r}B_{12} \right) u_{h}v_{h}p_{hv}} \left( A_{3} + \epsilon \gamma_{h} \right) + \left( A_{5}\mu_{h} + A_{4} \right) B_{0} \right]}{A_{h}A_{1}A_{2}A_{3}B_{3}B_{4} \left[ v_{h}p_{hv} \left( A_{3} + \epsilon \gamma_{h} \right) \left( \beta_{s}B_{r} + \beta_{r}B_{s} \right) + 3A_{5}B_{3} \right]},$$

Box I.

$$\lambda_{vh}^{*} = p_{vh} \left( \frac{\beta_{s} I_{s}^{*} + \beta_{r} I_{r}^{*}}{N_{h}^{*}} \right) = p_{vh} \frac{1}{N_{h}^{*}} \left( \beta_{s} I_{s}^{*} + \beta_{r} I_{r}^{*} \right)$$

$$= p_{vh} \frac{\left( \mu_{h} A_{1} A_{2} A_{3} + A_{4} \lambda_{vh}^{*} \right)}{A_{h} \left( A_{1} A_{2} A_{3} + A_{5} \lambda_{vh}^{*} \right)} \times \left\{ \frac{\beta_{s} \left[ B_{2} B_{5} \lambda_{hr}^{*} + \left( B_{1} B_{7} + B_{11} \lambda_{hr}^{*} \right) \lambda_{hs}^{*} \right]}{B_{3} B_{4} \left[ B_{3} + B_{s} \lambda_{hr}^{*} + \left( B_{r} + \lambda_{hr}^{*} \right) \lambda_{hs}^{*} \right]} + \frac{\beta_{r} \left[ B_{1} B_{6} \lambda_{hs}^{*} + \left( B_{2} B_{8} + B_{12} \lambda_{hs}^{*} \right) \lambda_{hr}^{*} \right]}{B_{3} B_{4} \left[ B_{3} + B_{s} \lambda_{hr}^{*} + \left( B_{r} + \lambda_{hr}^{*} \right) \lambda_{hs}^{*} \right]} \right\}. \tag{2.9}$$

Substituting  $\lambda_{hj}^*, j \in \{r,s\}$  from Eq. (2.8) into Eq. (2.9) and collecting terms in powers of  $\lambda_{vh}^*$  results in

$$(\lambda_{vh}^{*3} + C_2 \lambda_{vh}^{*2} + C_1 \lambda_{vh}^{*} + C_0) \lambda_{vh}^{*} = 0,$$

where  $C_0, C_1$ , and  $C_2$  are given in Box I. This leads to  $\lambda_{vh}^* = 0$  (which corresponds to the disease-free equilibrium case), or the cubic equation

$$\lambda_{nh}^{*3} + C_2 \lambda_{nh}^{*2} + C_1 \lambda_{nh}^* + C_0 = 0, \tag{2.11}$$

with the coefficients  $(C_n, n=0,1,2)$  and the thresholds  $(\mathcal{R}_n, n=1,2)$  given by Eqs. (2.10) in Box I. It should be noted that  $\lambda_{vh}^*=0$  leads to an unstable disease-free equilibrium solution when  $R_c>1$ . The possibility of zero, one, or multiple endemic equilibria can be investigated by solving the cubic Eq. (2.11) or by studying the signs of its coefficients using Descartes Rule of Signs. It should be mentioned that the signs of  $C_0$  and  $C_n, n \in \{1,2\}$  are determined by the signs of  $1-R_c$  and  $1-\mathcal{R}_n$ , respectively. Specifically,  $C_n>(\leq)$  0 if  $\mathcal{R}_n<(\geq)$  1 and  $C_0>(\leq)$  0 if  $R_c<(\geq)$  1. Hence, the cubic Eq. (2.11) yields the following result:

**Theorem 2.2.** Let  $C_n$ ,  $n \in \{0,1,2\}$  be coefficients of the polynomial equation (2.11). Then the model (2.1) has

- (a) no endemic equilibrium if  $C_0 \ge 0, C_1 \ge 0$ , and  $C_2 \ge 0$ ;
- (b) a unique endemic equilibrium if  $C_0 < 0, C_1 \ge 0$ , and  $C_2 \ge 0$ , or  $C_0 \le 0, C_1 < 0$ , and  $C_2 \ge 0$ , or  $C_0 \le 0, C_1 \le 0$ , and  $C_2 \le 0$ ;

(c) zero or two possible endemic equilibria if  $C_0 \ge 0$ ,  $C_1 > 0$  and  $C_2 < 0$ , or  $C_0 > 0$ ,  $C_1 < 0$  and  $C_2 \ge 0$ , or  $C_0 > 0$ ,  $C_1 \le 0$  and  $C_2 < 0$ .

Theorem 2.2(c) indicates that it might be possible for the model (2.1) to have two endemic equilibria when  $R_c < 1$ . This suggests that there is a parameter regime for which the model might exhibit a backward bifurcation (a phenomenon governed by the co-existence of a stable disease-free and a stable endemic equilibrium when  $R_c$  < 1) (Dushoff et al., 1998; Castillo-Chavez and Song, 2004; Gumel, 2012). The Center manifold theory can be used to prove the existence of a backward bifurcation rigorously (Castillo-Chavez and Song, 2004; Gumel, 2012; van den Driessche and Watmough, 2000). The existence of a backward bifurcation implies that although the condition  $R_c < 1$  is necessary, it is not sufficient for disease elimination (i.e., for reducing the number of malaria cases significantly to essentially very few or no case). Disease elimination in a backward bifurcation scenario is characterized by the initial population size and a second threshold denoted by  $R_c^{bb}$ , which is less than  $R_c$ . Specifically, in models with backward bifurcations, reducing the control reproduction number slightly below one might not be sufficient for disease elimination (as is the case for models with no backward bifurcations). For disease elimination to be possible in a model with a backward bifurcation, control measures must be applied and sustained until  $R_a$  falls below  $R^{bb}$  (i.e., within the region in which the disease-free equilibrium is globally asymptotically stable).

### 2.3. Numerical simulation results

The model (2.1) is simulated using the baseline parameter values in Table 1 together with different values of the disease-induced mortality rate  $(\delta_h)$  to assess the existence of a trans-critical (forward) and a sub-critical (backward) bifurcation. The results obtained, depicted in Fig. 2(a) show a forward bifurcation when the disease-induced mortality rate  $(\delta_h)$  is smaller than the natural mortality rate  $(\mu_h)$ , i.e., when

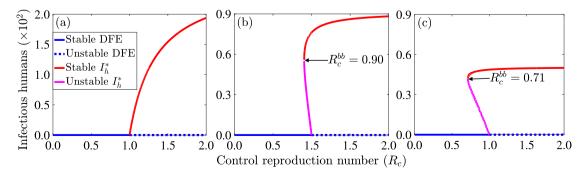


Fig. 2. Simulations of the model (2.1) illustrating: (a) a forward bifurcation when the disease-induced mortality rate  $\delta_h = 9.0 \times 10^{-5} < 4.1 \times 10^{-5} = \mu_h$ , and a backward bifurcation for (b)  $\delta_h = 4.5 \times 10^{-4} > 4.1 \times 10^{-5} = \mu_h$  and (c)  $\delta_h = 9.0 \times 10^{-4}$ . The other parameters used for the simulations are presented in Table 1.

 $\delta_h = 9.0 \times 10^{-5} < 4.1 \times 10^{-5} = \mu_h$ . In this case, there is a globally asymptotically stable disease-free equilibrium when the control reproduction number  $(R_c)$  is less than unity and a unique stable endemic equilibrium when  $R_c > 1$ . More simulation results of the model (2.1) for different ITN coverages ( $b_0$ ) and initial ITN efficacies against sensitive and resistant mosquitoes ( $\epsilon_{s0}$  and  $\epsilon_{r0}$ , respectively) that confirm the forward bifurcation scenario and highlight the importance of high usage of highly effective ITNs in the fight against malaria and the impact of other parameters on malaria dynamics are presented in Figs. S1-S2 of the SI. Fig. 2(b)-(c) depict backward bifurcation plots of the model (2.1) when the disease-induced mortality rate is smaller than the natural mortality rate. In this backward bifurcation case, the control reproduction number is less than one and the value of the backward bifurcation threshold quantity  $R_c^{bb}$  is 0.90 when  $\delta_h = 4.5 \times$  $10^{-4} > 4.1 \times 10^{-5} = \mu_h$  (Fig. 2(b)). In particular, there is a globally asymptotically stable disease-free equilibrium when  $R_c < R_c^{bb}$  and a locally stable disease-free equilibrium co-exists with a locally stable endemic equilibrium when  $R_c^{bb} \le R_c < 1$  (backward bifurcation region). Thus, intervention efforts must be maintained until  $R_c < R_c^{bb} = 0.90$ . This corresponds to an ITN coverage of  $\approx 88\%$ . If  $\delta_h = 9.0 \times 10^{-4}$ , this threshold reduces to about  $R_c^{bb} = 0.71$ , which corresponds to an ITN coverage that is greater than 100% (Fig. 2(c)). For this case, ITNs alone are not enough to contain the disease, even if everyone in the community uses them (see Fig. 2).

### 3. The model with a separate asymptomatic human carrier class

### 3.1. Model formulation

In this Section, the model formulated and analyzed in Section 2 will be modified to include a separate asymptomatic human infectious class denoted by  $I_{ah}$  and a partially immune class  $(R_h)$  that does not contribute to disease transmission. The asymptomatically infectious class consists of (infectious) humans with malaria parasite burden, who do not exhibit clinical symptoms of malaria at the end of the incubation period, and who have not recovered from infection yet. Here, a proportion (*m*) of exposed humans progress to the symptomatic infectious class  $(I_{sh})$ , while the remaining proportion (1 - m) progress to the asymptomatic infectious class. Symptomatic (asymptomatic) infectious humans develop immunity at rate  $\gamma_{sh}$  ( $\gamma_{ah}$ ). The total mosquito population remains the same, while the total human population is given by  $N_h = S_h + E_h + I_{sh} + I_{ah} + R_h$ . The force of infection  $\lambda_{hj}$ ,  $j \in \{r, s\}$ of the modified model is given by  $\lambda_{hj}=\beta_{j}p_{hv}(I_{sh}+\epsilon I_{ah})/N_{h},$  where  $\epsilon$  is a modification parameter representing the fact that asymptomatic infectious humans are less transmissible. The other functional forms are as defined in Section 2 and the other parameters are as defined in Table 1. Schematics of the modified model are presented in Fig. 3. The modified model is given by the system of equations:

$$\begin{split} \dot{S}_{h} &= \Lambda_{h} + \rho_{h} R_{h} - (\lambda_{vh} + \mu_{h}) S_{h}, \\ \dot{E}_{h} &= \lambda_{vh} S_{h} - (\mu_{h} + \nu_{h}) E_{h}, \\ \dot{I}_{sh} &= m \nu_{h} E_{h} - (\delta_{h} + \gamma_{sh} + \mu_{h}) I_{sh}, \\ \dot{I}_{ah} &= (1 - m) \nu_{h} E_{h} - (\gamma_{ah} + \mu_{h}) I_{ah}, \\ \dot{R}_{h} &= \gamma_{sh} I_{sh} + \gamma_{ah} I_{ah} - (\mu_{h} + \rho_{h}) R_{h}, \\ \dot{S}_{s} &= (1 - \theta) \Lambda_{v} + \sigma_{r} S_{r} - (\lambda_{hs} + \sigma_{s} + \mu_{s}) S_{s}, \\ \dot{E}_{s} &= \lambda_{hs} S_{s} + \sigma_{r} E_{r} - (\sigma_{s} + \mu_{s} + \nu_{s}) E_{s}, \\ \dot{I}_{s} &= v_{s} E_{s} + \sigma_{r} I_{r} - (\sigma_{s} + \mu_{s}) I_{s}, \\ \dot{S}_{r} &= \theta \Lambda_{v} + \sigma_{s} S_{s} - (\lambda_{hr} + \sigma_{r} + \mu_{r}) S_{r}, \\ \dot{E}_{r} &= \lambda_{hr} S_{r} + \sigma_{s} E_{s} - (\sigma_{r} + \mu_{r} + \nu_{r}) E_{r}, \end{split}$$

where  $A_s = \delta_h + \gamma_{sh} + \mu_h$ ,  $A_a = \gamma_{ah} + \mu_h$  and  $A_1$ ,  $A_3$ ,  $B_j$ , and  $\tilde{B}_j$ ,  $j \in \{r, s\}$  are as defined in Eq. (2.3).

 $\dot{I}_r = v_r E_r + \sigma_s I_s - (\sigma_r + \mu_r) I_r,$ 

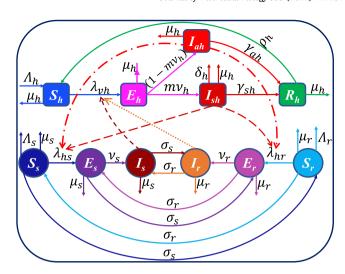


Fig. 3. Schematics of the model depicting transitions of humans and mosquitoes between epidemiological stages (solid lines) and interactions between humans and mosquitoes that result in asymptomatic and symptomatic infectious humans infecting susceptible mosquitoes (long dashed and dash-dotted lines, respectively) and infectious sensitive and resistant mosquitoes infecting susceptible humans (short dashed and dotted lines, respectively).

### 3.2. Analytical results

It can easily be verified as in Section 2 that the modified model system (3.1) is well-posed within the region

$$\begin{aligned} \Omega &= \{ (S_h, E_h, I_{sh}, I_{ah}, R_h, S_s, E_s, I_s, S_r, E_r, I_r) \in \mathbb{R}_+^{11} : 0 \le S_h + E_h + I_{sh} \\ &+ I_{ah} + R_h \le \Lambda_h / \mu_h, 0 \le S_s + E_s + I_s + S_r + E_r + I_r \le \Lambda_v / \mu_v \}, \end{aligned}$$

where  $\mu_v = min(\mu_s, \mu_r)$  and that this epidemiologically feasible region is positively invariant and attracting with respect to the model (3.1). That is, solutions of (3.1) originating from  $\Omega$  are trapped in  $\Omega$  for t > 0.

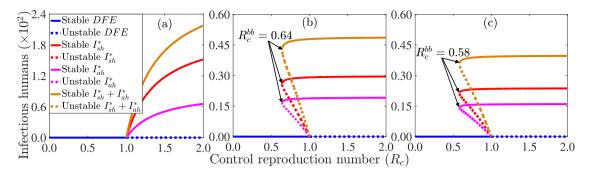
The disease-free equilibrium of the modified model (3.1) is the same as that of the model (2.1) (i.e., Eq. (2.5)), but the reproduction numbers are different. In particular, the reproduction number of Eqs. (3.1) is a function of the proportion of exposed humans who become symptomatically infectious at the end of the incubation period (m). Additionally, the reproduction number of the model (3.1) depends on the parameter groupings  $A_s = \delta_h + \gamma_{sh} + \mu_h$  and  $A_a = \gamma_{ah} + \mu_h$ , since the infectious human class of the model (2.1) is now split into symptomatic and asymptomatic infectious classes, with different recovery rates ( $\gamma_{sh}$  and  $\gamma_{ah}$ ). Following the next generation operator approach (see Section S2.1 of the SI for details), the reproduction number of the model (3.1) is

$$R_{c} = \sqrt{\frac{p_{hv}p_{vh}\mu_{h}\nu_{h}B_{0}\left[mA_{a} + (1 - m)\epsilon A_{s}\right]}{\Lambda_{h}A_{1}A_{a}A_{s}B_{3}^{2}B_{4}}}.$$
(3.2)

Theorem 3.1 on the stability of the disease-free equlibrium of Eqs. (3.1) follows directly from Theorem 2 in Driessche and Watmough (2002).

**Theorem 3.1.** The disease-free equilibrium of the model system given by Eqs. (3.1) is locally-asymptotically stable if  $R_c < 1$  and unstable if  $R_c > 1$ .

Following the approach in Section 2.2.3, endemic equilibrium values of each of the state variables can be expressed in terms of  $\lambda_{vh}^*$  and  $\lambda_{hj}^*, j \in \{r, s\}$ , i.e., the equilibrium values of the forces of infection  $\lambda_{vh}$  and  $\lambda_{hj}$ . Using the functional forms of the modified forces of infection leads to a cubic polynomial equation  $\lambda_{vh}^{*3} + C_2 \lambda_{vh}^{*2} + C_1 \lambda_{vh}^* + C_0 = 0$  (see Section S2.2 of the SI for the coefficients  $C_n, n \in \{0, 1, 2\}$ ) that can be used to determine the possible number of endemic equilibria of the modified model (3.1). This analysis leads to the following theorem:



**Fig. 4.** Simulations of the model (3.1) showing a: (a) forward bifurcation when the recovery rate of asymptomatic infectious humans ( $\gamma_{ah}$ ) is 1/80, and a backward bifurcation for (b)  $\gamma_{ah} = 1/120$ ,  $\delta_h = 9.0 \times 10^{-5}$  and (c)  $\gamma_{ah} = 1/120$ ,  $\delta_h = 6.3 \times 10^{-4}$ , where  $\delta_h$  is the malaria-induced mortality rate. The magenta and red curves represent the equilibrium populations of the asymptomatic infectious ( $I_{ah}$ ) and the symptomatic infectious ( $I_{sh}$ ) classes, respectively. The other parameters used for the simulations are presented in Table 1.

**Theorem 3.2.** Let  $C_n$ ,  $n \in \{0,1,2\}$  be coefficients of the polynomial Eq. (2.11). Then the model (3.1) has

- (a) no endemic equilibrium if  $C_0 \ge 0, C_1 \ge 0$ , and  $C_2 \ge 0$ ;
- (b) a unique endemic equilibrium if  $C_0 < 0, C_1 \ge 0$ , and  $C_2 \ge 0$ , or  $C_0 \le 0, C_1 < 0$ , and  $C_2 \ge 0$ , or  $C_0 \le 0, C_1 \le 0$ , and  $C_2 \le 0$ ;
- (c) zero or two possible endemic equilibria if  $C_0 \ge 0$ ,  $C_1 > 0$  and  $C_2 < 0$ , or  $C_0 > 0$ ,  $C_1 < 0$  and  $C_2 \ge 0$ , or  $C_0 > 0$ ,  $C_1 \le 0$  and  $C_2 < 0$ .

The case of a unique endemic equilibrium in Theorem 3.2(b) is illustrated in Fig. 4(a) and Fig. S3 of the SI, while the case of a backward bifurcation in Theorem 3.2(c) is illustrated in Fig. 4(b)–(c).

### 3.3. Numerical simulation results

In this section, simulations of the model (3.1) to assess the impact of malaria intervention measures such as ITNs, the impact of mosquito resistance to insecticides, the impact of asymptomatic transmission, etc., will be performed. Unless otherwise specified, simulations will be performed using the baseline parameter values in Table 1.

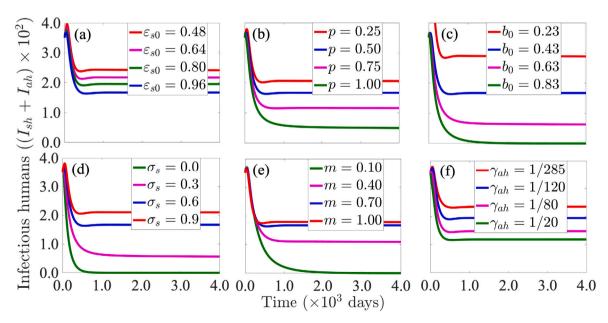
### 3.3.1. Forward versus backward bifurcations

The model (3.1) is simulated to assess the possibility of a forward and a backward bifurcation. The results show that the model exhibits a

forward bifurcation when the recovery rate of asymptomatic infectious humans ( $\gamma_{ah}$ ) is 1/80 (Fig. 4(a)). In this case,  $R_c > 1$ . The model exhibits a backward bifurcation when  $\gamma_{ah} = 1/120$ ,  $\delta_h = 9.0 \times 10^{-5}$  (Fig. 4(b)) and  $\gamma_{ah} = 1/120$ ,  $\delta_h = 4.5 \times 10^{-4}$  (Fig. 4(c)). In this backward bifurcation case,  $R_c < 1$  and the backward bifurcation threshold ( $R_c^{bb}$ ) is 0.64 for Fig. 4(b), which corresponds to an ITN coverage level of  $\approx 95\%$ , and 0.58 for Fig. 4(c). This corresponds to an ITN coverage level of > 100%. To contain the disease, control efforts must be sustained until  $R_c < R_c^{bb}$ . Achieving this using only ITNs is not possible in the case of Fig. 4(c).

### 3.3.2. Assessing the impact of ITNs, symptomatic transmission, and insecticide resistance on malaria dynamics

The model (3.1) is simulated to assess the impact of ITN efficacy against sensitive mosquitoes ( $\epsilon_{s0}$ ), the modification factor (p) of ITN efficacy against resistant mosquitoes ( $\epsilon_{r0}$ ), ITN coverage ( $b_0$ ), the rate at which adult mosquitoes develop resistance to insecticides ( $\sigma_s$ ), the proportion of exposed humans who become symptomatically infectious at the end of the incubation period (m), and the recovery rate of asymptomatic infectious humans ( $r_{ah}$ ) on malaria burden, i.e., the total number of infectious humans ( $r_{sh}+r_{ah}$ ). All comparisons will be against the baseline scenario depicted by blue curves in the time series plots. The generated simulation results displayed in Fig. 5, show that reducing the efficacy of ITNs against sensitive mosquitoes from the baseline



**Fig. 5.** Simulations of the model (3.1) showing the impact of (a) ITN efficacy against sensitive mosquitoes ( $\epsilon_{s0}$ ), (b) the modification factor (p) of ITN efficacy against resistant mosquitoes ( $\epsilon_{r0}$ ), (c) ITN coverage ( $b_0$ ), (d) the rate at which adult mosquitoes develop resistance to insecticides ( $\sigma_s$ ), (e) the proportion of exposed humans who become symptomatically infectious at the end of the incubation period (m), and (f) the recovery rate of asymptomatic infectious humans ( $\gamma_{ah}$ ) on the total infectious human population ( $I_{sh} + I_{ah}$ ). The other parameters used for the simulations are given in Table 1.

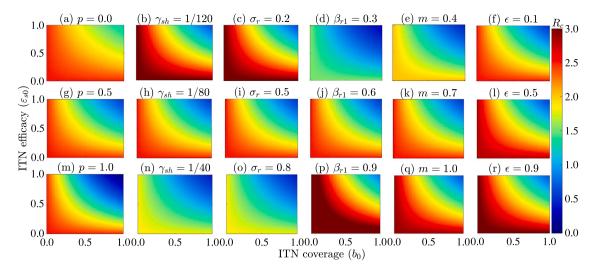


Fig. 6. Heatmaps of the control reproduction number ( $R_c$ ) of the model (3.1) as a function of ITN coverage ( $b_0$ ) and the efficacy of ITNs against sensitive mosquitoes ( $\epsilon_{s0}$ ) for different values of the modification factor for the efficacy of ITNs against resistant mosquitoes, p ((a), (g), and (m)), the recovery rate of symptomatic infectious humans,  $\gamma_{sh}$  ((b), (h), and (n)), the rate at which adult mosquitoes develop resistance to insecticides,  $\sigma_s$  ((c), (i), and (o)), the maximum biting rate of resistant mosquitoes,  $p_{r1}$  ((d), (j), and (p)), the proportion of exposed humans who become symptomatic at the end of the incubation period, m ((e), (k), and (q)), and the asymptomatic infectious human-to-mosquito transmission probability adjustment parameter,  $\epsilon$  ((f), (l), and (r)). Other parameter values of the model are as presented in Tables 1.

value of  $\varepsilon_{s0}$  = 0.96 (depicted by the blue curve) by 16% (i.e., to  $\varepsilon_{s0}$  = 0.80), will lead to an  $\approx 17\%$  increase in malaria burden (green curve in Fig. 5(a)). Further reductions in the efficacy will result in higher disease burdens. In particular, if the baseline value of  $\varepsilon_{s0}$  is halved (i.e., if  $\varepsilon_{s0} = 0.48$ ), the burden of malaria will increase by  $\approx 44\%$  (red curve in Fig. 5(a)). On the other hand, if the modification factor of the efficacy against resistant mosquitoes is halved (i.e., reduced from its baseline value of p = 0.5 to p = 0.25), an  $\approx 23\%$  increase in disease burden is observed (red curve in Fig. 5(b)), while if the baseline value is doubled (i.e., if p = 1.0), an  $\approx 70\%$  reduction in disease burden is observed (green curve in Fig. 5(b)). If the ITN coverage is 20% lower than the baseline value of  $b_0 = 0.43$  (i.e., if  $b_0 = 0.23$ ), the burden of malaria witnesses a 75% rise (red curve in Fig. 5(c)), while if the ITN coverage increases by 20% from the baseline value (i.e., if  $b_0 = 0.63$ ), the burden of malaria drops by 62% (magenta curve in Fig. 5(c)). A further increase in ITN coverage by 20%, i.e., increasing the ITN coverage to  $b_0 = 0.83$  from the baseline value, can result in malaria elimination (green curve in Fig. 5(c)). Also, if the rate at which adult mosquitoes develop resistance to insecticides increases from the baseline value of  $\sigma_s = 0.6$  per day to  $\sigma_s = 0.9$  per day, the burden of malaria increases by 25% (magenta curve in Fig. 5(d)), while if this rate reduces from the baseline value to  $\sigma_s = 0.3$  per day, the burden of malaria reduces by 66% (green curve in Fig. 5(d)). The diseases can be contained if no mosquito develops resistance to insecticides, i.e., if  $\sigma_s = 0.0$  per day. The simulation results also indicate that a 30% increase in the proportion of exposed humans who exhibit malaria symptoms at the end of the incubation period from the baseline value of m = 0.7 (i.e., m = 1.0), will trigger a 7% increase in the burden of malaria (red curve in Fig. 5(e)). A 30% reduction from the baseline value (i.e., m = 0.4) will lead to a 35% reduction in the burden of malaria (magenta curve in Fig. 5(e)), while reducing the proportion of exposed humans who develop malaria symptoms at the end of the incubation period from m = 0.7 to m = 0.1 can result in malaria elimination (green curve in Fig. 5(e)). Furthermore, the simulations show that if the recovery rate of asymptomatic infectious humans is reduced from its baseline value of  $\gamma_{ah} = 1/120$  to  $\gamma_{ah} = 1/285$  per day, the burden of malaria increases by 41% (magenta curve in Fig. 5(f)), while if this rate is increased from the baseline value to  $\gamma_{ah} = 1/20 \ per$ day, the burden of malaria reduces by 29% (green curve in Fig. 5(f)).

Fig. 6 shows heatmaps of the control reproduction number ( $R_c$ ) of the model (3.1) as a function of ITN coverage ( $b_0$ ) and the efficacy of ITNs against sensitive mosquitoes ( $\varepsilon_{s0}$ ) for various values of the modification factor for the efficacy of ITNs against resistant mosquitoes,

p (Fig. 6(a), (g), and (m)), the recovery rate of symptomatic infectious humans,  $\gamma_{sh}$  (Fig. 6(b), (h), and (n)), the rate at which adult mosquitoes develop resistance to insecticides,  $\sigma_s$  (Fig. 6(c), (i), and (o)), the maximum biting rate of resistant mosquitoes,  $\beta_{r1}$  ((d), (j), and (p)), the proportion of exposed humans who become symptomatic at the end of the incubation period, m (Fig. 6(e), (k), and (q)), and the asymptomatic infectious human-to-mosquito transmission probability adjustment parameter,  $\epsilon$  (Fig. 6(f), (l), and (r)). Fig. 6(a), (g), and (m) show that if resistance to insecticides lowers the efficacy of ITNs significantly then higher ITN coverage is required for reducing the control reproduction number significantly. In particular, if the efficacy of ITNs against resistant mosquitoes is zero (i.e., if p = 0), reducing the control reproduction number below one will be unattainable even if everybody in the community uses ITNs whose efficacy against sensitive mosquitoes is 100% (Fig. 6(a)). If p = 0.5 and  $\varepsilon_{s0} = 0.64$  ( $\varepsilon_{s0} = 0.96$ ), then 100% (71%) ITN coverage is required to reduce  $R_c$  below one (Fig. 6(g)), while if p = 1 (i.e.,  $\varepsilon_{r0} = \varepsilon_{s0}$ ), then 86% ITN coverage is required to reduce  $R_c$  below one if  $\varepsilon_{s0} = 0.48$  (Fig. 6(m)). Furthermore, if p=1 and  $\varepsilon_{s0}=0.64$  ( $\varepsilon_{s0}=0.96$ ), then 63% (43%) of the populace is required to sleep under ITNs to reduce  $R_c$  below one (Fig. 6(m)).

Next, Fig. 6(d), (j), and (p) show that lower ITN coverage is required in communities with low mosquito concentration and biting rates. Specifically, if the maximum biting rate of resistant mosquitoes is  $\beta_{r1}$  = 0.3, and the efficacy of ITNs against sensitive mosquitoes is only 48% (i.e.,  $\varepsilon_{s0}$  = 0.48), then 61% of the population is required to sleep under ITNs to reduce  $R_c$  below one (Fig. 6(d)). But if  $\beta_{r1} = 0.3$  and the efficacy of ITNs against sensitive mosquitoes is 64% (96%), then 45% (31%) ITN coverage is required to reduce  $R_c$  below one (Fig. 6(d)). If  $\beta_{r1} = 0.6$  and  $\varepsilon_{s0} = 0.48$ , reducing  $R_c$  below one will be impossible even if everybody sleeps under an ITN (Fig. 6(j)). However, if  $\beta_{r1} = 0.6$ and the efficacy of ITNs is  $\varepsilon_{s0} = 0.64$  ( $\varepsilon_{s0} = 0.96$ ), then 98% (65%) ITN coverage is required to reduce  $R_c$  below one (Fig. 6(j)). If individuals in a community with high mosquito densities and/or biting rates settle for ITNs of low efficacy to sensitive mosquitoes, it might be impossible to reduce  $R_c$  below one (hence, the disease will continue to spread in the population). However, if the populace settle for the baseline case with ITNs of efficacy 96% against sensitive mosquitoes, then at least 90% of the population is required to use ITNs in order to reduce  $R_c$  below unity if  $\beta_{r1} = 0.9$  (Fig. 6(p)). More outcomes from Fig. 6 are summarized in Table 2, while heatmaps of disease prevalence as a function of ITN coverage and efficacy, and heatmaps of  $R_c$  and malaria prevalence as functions of the rate at which adult mosquitoes develop resistance to insecticides ( $\sigma_s$ ) and the rate at which resistant mosquitoes lose their resistance ( $\sigma_r$ ) are presented in Figs. S4–S6 of the SI.

**Table 2** Proportion of humans required to use ITNs to reduce the control reproduction number ( $R_c$ ) below one for various values of the recovery rate of symptomatic infectious humans ( $\gamma_{sh}$ ), various rates at which adult mosquitoes develop resistance to insecticides ( $\sigma_s$ ), various proportions of exposed humans who become symptomatic at the end of the incubation period (m), and for various values of the asymptomatic infectious human-to-mosquito transmission probability adjustment factor ( $\epsilon$ ).

ITN	Minimum ITN coverage $(b_0)$									
efficacy	$\gamma_{sh} = 1/120$	$\gamma_{sh} = 1/40$	$\sigma_r = 0.2$	$\sigma_r = 0.8$	m = 0.4	m = 1.0	$\epsilon = 0.5$	$\epsilon = 0.9$		
$\epsilon_{s0} = 0.48$	>100%	86%	>100%	88%	100%	>100%	>100%	>100%		
$\varepsilon_{s0} = 0.64$	>100%	65%	>100%	65%	76%	>100%	>100%	>100%		
$\varepsilon_{s0} = 0.96$	80%	43%	80%	43%	51%	75%	65%	80%		

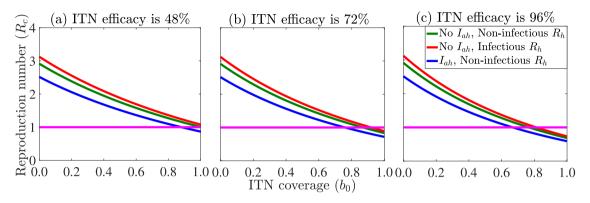


Fig. 7. Profile of the control reproduction number  $(R_c)$  as a function of ITN coverage  $(b_0)$  for different ITN efficacies  $(\varepsilon_{s0})$  and for the case in which there is no asymptomatic infectious class, i.e., m=1 and recovered individuals do not transmit the disease (green curve), there is an asymptomatic infectious class, i.e., 0 < m < 1 and recovered individuals do not transmit the disease (blue curve), and there is no asymptomatic infectious class, i.e., m=1 and recovered individuals can transmit the disease (red curve). (a)  $\varepsilon_{s0} = 0.48$ , (b)  $\varepsilon_{s0} = 0.72$ , and (c)  $\varepsilon_{s0} = 0.96$ . The other parameters used for the simulation are presented in Table 1.

### 3.3.3. The impact of incorporating a separate asymptomatic infectious class

Additional simulations of the model (3.1) were performed to assess the impact of incorporating a separate asymptomatic infectious human class. The results obtained (Fig. 7), show that accounting for asymptomatic transmission through a separate asymptomatic infectious human class leads to a model with a smaller control reproduction number. In particular, if the efficacy of ITNs ( $\varepsilon_{s0}$ ) is 48% and a separate asymptomatic infectious human class is incorporated, while partially immune humans do not contribute to disease transmission (i.e., the Model (3.1)), then 89% ITN coverage is required to contain the disease (blue curve in Fig. 7(a)). However, if there is no asymptomatic infectious human class and partially immune humans do not contribute to disease transmission, or if there is no separate asymptomatic infectious human class and partially immune humans contribute to disease transmission (i.e., the Model (2.1)), containing the disease is not possible even if the entire population uses ITNs (green and red curves in Fig. 7(a)). On the other hand, if the efficacy of ITNs against sensitive mosquitoes is 96% and a separate asymptomatic infectious human class is incorporated, then only 67% ITN coverage is required to contain the disease (blue curve in Fig. 7(c)). A higher coverage level is required if asymptomatic infectious humans are not accounted for. In particular, if there is no asymptomatic infectious human class and partially immune humans do not contribute to disease transmission, 77% ITN coverage is required to contain the disease (green curve in Fig. 7(c)), while if there is no asymptomatic infectious human class and partially immune humans contribute to disease transmission, 81% ITN coverage is required to contain the disease (red curve in Fig. 7(c)).

### 4. The models (2.1) and (3.1) with decaying ITN efficacy

There is overwhelming evidence that the efficacy of ITNs decays over time (N'Guessan et al., 2007; Briët et al., 2012; Obala et al., 2015). In this section, the models (2.1) and (3.1) are extended to account for decaying ITN efficacy and periodic ITN replacement, i.e.,  $\varepsilon_l$ ,  $l \in \{\beta_j, \mu_j\}$ ,  $j \in \{r, s\}$  is now a decreasing function of time over the useful life of an ITN. As in Ngonghala et al. (2014, 2016), we model the (waning of) ITN-efficacy in preventing mosquitoes from reaching

humans sleeping beneath them  $(\epsilon_{\beta_j})$  and in killing mosquitoes that land on them  $(\epsilon_{\mu_i})$  through the functional forms:

$$\varepsilon_{\beta}(t) = \frac{2^{n} + 1}{2^{n+1}} \left( \frac{2^{n} - 1}{2^{n} + 1} + \frac{1}{1 + \left( \frac{t \mod T}{T/2} \right)^{n}} \right) \varepsilon_{j0},$$

$$\varepsilon_{\mu}(t) = \frac{2^{n} + 1}{2^{n}} \left( -\frac{1}{2^{n} + 1} + \frac{1}{1 + \left( \frac{t \mod T}{T/2} \right)^{n}} \right) \varepsilon_{j0},$$
(4.1)

where  $0 \le \varepsilon_{i0} \le 1$  is the initial efficacy of ITNs, T is the useful lifespan of ITNs, and n > 1 is a shape constant. It should be mentioned that these functional forms take into account the fact that upon complete waning of insecticides from ITNs, the ITNs provide about half their original protective and killing efficacies (Clarke et al., 2001) and that ITNs are replaced with new ones (a process that restores the original efficacies of the ITNs) after the effective useful lifespan of the ITNs. As in Ngonghala et al. (2014, 2016),  $(S_r^*(t), S_r^*(t))$  is the unique globally attractive Tperiodic positive solution of the sensitive and resistant susceptible mosquito components of the disease-free system of the non-autonomous versions of the models (2.1) and (3.1), with  $\varepsilon_l$ ,  $l \in \{\beta, \mu\}$  given by Eq. (4.1) (Ngonghala et al., 2014; Nakata and Kuniya, 2010). The disease-free equilibria of the non-autonomous versions of the models (2.1) and (3.1) are  $(S_h^*, E_h^*, I_h^*, R_h^*, S_s^*(t), E_s^*(t), I_s^*(t), S_r^*(t), E_r^*(t), I_r^*(t)) = \left(\frac{\Lambda_h}{\mu_h}, 0, 0, 0, S_s^*(t), 0, 0, S_r^*(t), 0, 0\right)$  and  $(S_h^*, E_h^*, I_{sh}^*, I_{ah}^*, R_h^*, S_s^*(t), E_s^*(t), I_s^*(t), S_r^*(t), I_r^*(t)) = \left(\frac{\Lambda_h}{\mu_h}, 0, 0, 0, 0, S_s^*(t), 0, 0, S_r^*(t), 0, 0\right),$ respectively. See Ngonghala et al. (2014) for a sample proof of the wellposedness of the non-autonomous versions of the models (2.1) and (3.1) (i.e., the models (2.1) and (3.1) with  $\varepsilon_l$  given by Eq. (4.1)).

### 4.1. Results

Unless otherwise specified, the parameter values in Table 1 will be used to simulate the non-autonomous versions of the models (2.1) and (3.1) with  $\epsilon_l$ ,  $l \in \{\beta, \mu\}$  given by Eq. (4.1) to assess the impact of decay

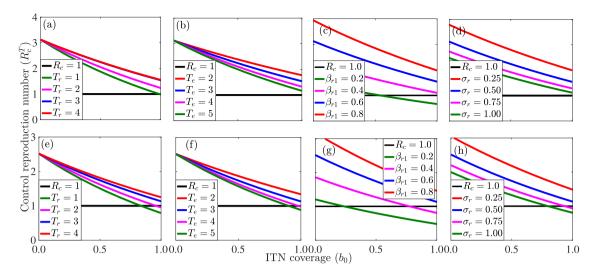


Fig. 8. Profiles of the control reproduction numbers ( $R_c^T$ ) of the models (2.1) ((a)–(d)) and (3.1) ((e)–(h)) with decaying ITN efficacy and periodic ITN replacement as functions of ITN coverage ( $b_0$ ) for various values of the ITN replacement times,  $T_r$  ((a) and (e)), useful lifespan of ITNs,  $T_e$  ((b) and (f)), the maximum biting rate of mosquitoes,  $\beta_{r1}$  ((c) and (g)), and the rate at which mosquitoes lose resistance to insecticides,  $\sigma_e$  ((d) and (h)). The other parameter values are given in Table 1. See Fig. S11 of the SI for more results.

in ITN efficacy, mosquito resistance to insecticides, the useful lifespan of ITNs  $(T=T_e)$ , and the effective replacement time of ITNs  $(T=T_r)$  on the burden of malaria. The minimum ITN coverage required to reduce the control reproduction number below unity (i.e., the possibility of malaria elimination) under different scenarios will be identified. The numerical technique presented in Wang and Zhao (2008b) and Ngonghala et al. (2014) will be used to compute the reproduction number  $(R_c^T)$  of the time-dependent periodic systems as functions of the ITN coverage  $(b_0)$ . Since the results obtained by comparing the models (2.1) and (3.1) for the case in which ITN efficacy decays over time are similar to those for the case in which ITN efficacy is constant, we focus mostly on the non-autonomous version of the model (3.1) (with decaying ITN efficacy) and present related results for the non-autonomous version of the model (2.1) in the SI (Section S3). The non-autonomous version of the model (3.1) will be referred to as the extended model.

### 4.1.1. The critical ITN coverage level for containing malaria

We identify the minimum proportion of the human population required to use ITNs to reduce the control reproduction number  $(R_a^T)$ of the non-autonomous versions of the models (2.1) and (3.1) with decaying ITN efficacy and periodic ITN replacement below one under different scenarios. These scenarios involve ITN policies that lead to various effective ITN replacement times  $(T_r)$ , useful lifespans of ITNs  $(T_e)$ , maximum biting rates of resistant mosquitoes  $(\beta_{r1})$ , and various rates of loss of resistance by adult mosquitoes ( $\sigma_r$ ). Fig. 8 depicts plots of  $R_c^T$  as functions of ITN coverage  $(b_0)$  under various scenarios (involving different values of  $T_r$ ,  $T_e$ ,  $\beta_{r1}$ , and  $\sigma_r$ ). All other parameters are fixed at their baseline values given in Table 1. The baseline case obtained using the baseline parameter values in Table 1 show that for the non-autonomous version of the model (2.1) ((3.1)) with decaying ITN efficacy and periodic ITN replacement, reducing  $R^T$  below one only through the use of ITNs is impossible even if the ITN coverage level is 100% (blue curves in Fig. 8). If an ITN policy that relies on replacing ITNs (of prescribed useful lifespan three years) every year is implemented, then it might be possible to reduce  $R_c^T$  below one using Model (2.1) ((3.1)), if 99% (83%) of the population use ITNs for personal protection (green curves in Fig. 8(a) and (e)). However, if ITNs (of prescribed useful lifespans of three years) are replaced every four years, then reducing  $R_a^T$  below one will never be possible even if everyone in the community uses ITNs for personal protection (red curves in Fig. 8(a) and (d)). On the other hand, if an ITN policy that requires the use of ITNs of shorter useful lifespans is implemented, then more humans will be required to use ITNs. In particular, if the useful

lifespan of ITNs is two years (i.e., one year shorter than the baseline useful lifespan prescribed by WHO), it will not be possible to reduce  $R_c^T$  below one even if everybody uses ITNs for personal protection (red curves in Fig. 8(b) and (f)). But, if the prescribed lifespan of ITNs is increased from the baseline value of three years to five years, then 90% ITN coverage is required to reduce  $R_c^T$  below one for the model (3.1), while reducing  $R_c^T$  below one is impossible for the model (2.1) (green curves in Fig. 8(b) and (f)). In communities with lower mosquito densities or biting rates, lower ITN coverage is required to reduce  $R_c^T$  below one. In particular, if the maximum biting rate of resistant mosquitoes is  $\beta_{r1} = 0.2$ , then using Model (2.1) ((3.1)), 54% (23%) ITN coverage is required to reduce  $R_c^T$  below unity (green curves in Fig. 8(c) and (g)). Furthermore, if the rate at which adult mosquitoes develop resistance to insecticides is reduced (possibly through the use of synergists that inhibit the development of resistance), lower ITN coverage is required to reduce  $R_c^T$  below one. Specifically, if the rate at which resistant mosquitoes lose resistance to insecticides is  $\sigma_r = 0.25 \, per$ day reducing  $R_c^T$  below one will be impossible (red curve in Fig. 8(c)), while if  $\sigma_r = 1.0$  per day, 80% ITN coverage is required for Model (3.1), while reducing  $R_c^T$  below one is impossible for Model (2.1) (green curves in Fig. 8(d) and (h)). Results for the initial ITN efficacies ( $\varepsilon_{s0}$  and  $\varepsilon_{r0}$ ) and the proportion of exposed humans who are symptomatic at the end of the incubation period (m) are presented in Section 5.

### 5. Outcomes of the autonomous versus non-autonomous model

In this section, the autonomous and non-autonomous versions of the models (2.1) and (3.1) (i.e., the models (2.1) and (3.1) with constant ITN efficacy and with decaying ITN efficacy and periodic ITN replacement) are analyzed and compared under various scenarios. Since results for the autonomous and non-autonomous versions of the model (2.1) are similar to those of the autonomous and non-autonomous versions of the model (3.1), we present them in the SI (with the exception of Fig. 9, which compares the long-term dynamics of all the models).

### 5.1. Long-term dynamics of the autonomous and non-autonomous models

Fig. 9 depicts the long-term dynamics of the autonomous and non-autonomous versions of the models (2.1) and (3.1) (i.e., the models (2.1) and (3.1) with constant ITN efficacy and with decaying ITN efficacy and periodic ITN replacement) under different ITN efficacies and ITN coverages. For ITNs with high efficacies and high coverage (e.g., if  $\varepsilon_{s0}=0.96$  and  $b_0=0.90$ ), the autonomous models (2.1)

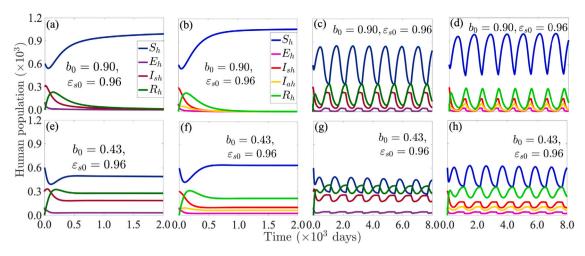


Fig. 9. Long-term dynamics of the model (2.1) with constant ITN efficacy ((a) and (e)), the model (3.1) with constant ITN efficacy ((b) and (f)), the model (2.1) with decaying ITN efficacy ((c) and (g)), and the model (3.1) with decaying ITN efficacy ((d) and (h)) for different values of the initial ITN efficacy against sensitive mosquitoes ( $\epsilon_{s0}$ ) and ITN coverages ( $b_0$ ). The other parameter values used for the simulation are presented in Table 1.

and (3.1) relax at the disease-free equilibrium (Fig. 9(a) and (b), respectively), while the non-autonomous models (2.1) and (3.1) exhibit oscillatory dynamics with non-zero infectious populations (Fig. 9(c) and (d), respectively). Thus, although the autonomous systems show that disease elimination is possible for this level of ITN coverage and ITN efficacy (Fig. 9(a)-(b)), the non-autonomous models show that disease elimination is not possible (Fig. 9(c)-(d)). But, for high ITN efficacy and low ITN coverage (e.g., if  $\varepsilon_{s0} = 0.96$  and  $b_0 = 0.43$ ), the autonomous models (2.1) and (3.1) relax on an their respective endemic equilibria (Fig. 9(e) and (f), respectively), while the nonautonomous models (2.1) and (3.1) exhibit oscillatory dynamics with period equivalent to the replacement time of ITNs (Fig. 9(g) and (h), respectively). These oscillations are due to the periodic replacement of ITNs after the useful lifespan. Although the autonomous systems show that disease elimination might be possible (Fig. 9(a)-(b)), the non-autonomous models show that disease elimination might not be possible in the same parameter regime due to the added realism from ITN-use implementation (Fig. 9(c)-(d)). This is consistent with results from previous studies (e.g., Ngufor et al. (2011), West et al. (2014) and Protopopoff et al. (2015)) showing that ITN-use or indoor residual spraying as a single intervention is not sufficient for containing malaria; however, combining the two interventions offers better performance in controlling the disease. Studies in Ngufor et al. (2011), Djènontin et al. (2010) and Djènontin et al. (2009) also show that combining the two measures improve insecticide-resistance management (since different insecticides with different modes of action are used at the same location and time). Furthermore, this result is consistent with results from Killeen and Smith (2007), Govella et al. (2010), Bugoro et al. (2011) and Govella and Ferguson (2012) indicating that the possibility of out-door feeding or resting by some mosquitoes can reduce the effectiveness of ITNs in malaria control significantly thereby rendering elimination challenging.

In both the autonomous and non-autonomous models, the disease burden (i.e., the total infectious human population) for the model (2.1) is greater than the disease burden for the model (3.1). Hence, the model (3.1) underestimates disease burden.

### 5.2. Assessing the impact of parameters on disease burden through a global uncertainty and sensitivity analysis

The malaria models (2.1) (from Section 2) and (3.1) from Section 3 with constant ITN efficacy together with the non-autonomous versions with decaying ITN efficacy and periodic ITN replacement times described in Section 4 have large numbers of parameters, the estimation of which involves uncertainty and variability. Consequently, it is useful

to examine the effect of these uncertainties and variability on the outputs of the model. Specifically, it is critical to identify the model parameters that have the greatest effect on disease dynamics. Here, we consider the model (2.1) with (a) constant ITN efficacy, (b) waning ITN efficacy and periodic ITN replacement time, and the model (3.1) with (c) constant ITN efficacy, and (d) waning ITN efficacy and periodic ITN replacement time. Malaria prevalence in humans is used as the response function. Similar results are obtained for the models (2.1) and (3.1) with constant ITN efficacy when the control reproduction number is used as the response function (Figs. S7 (a) and S12 (a) of the SI). As in Ngonghala et al. (2014, 2016, 2020), global uncertainty and sensitivity analysis will be performed on the response functions of the models utilizing the Latin Hypercube Sampling (LHS) and Partial Rank Correlation Coefficients (PRCCs) approach (Blower and Dowlatabadi, 1994; Marino et al., 2008). The uncertainty and sensitivity analysis results obtained for the four models are presented in Fig. 10. The analysis shows that for each of the four model scenarios, the maximum biting rate of resistant mosquitoes  $(\beta_{r1})$  and the ITN coverage  $(b_0)$  have the greatest impact on malaria prevalence (Fig. 10(a)-(d)) and the other response function (Figs. S7 and S12 of the SI). The next set of highly influential parameters are the probability that an infectious mosquito (human) infects a susceptible human,  $p_{vh}$  (mosquito,  $p_{hv}$ ), the ITN efficacy for sensitive mosquitoes ( $\varepsilon_{s0}$ ), and the rate at which adult mosquitoes lose (develop) resistance to insecticides,  $\sigma_r$  ( $\sigma_s$ ). The strong negative correlation between disease prevalence and the ITN coverage  $b_0$  and/or the efficacy of ITNs against sensitive mosquitoes indicates that high usage of high quality ITNs is important in mitigating the health burden of malaria. This is consistent with studies in Lindsay et al. (2021) calling for an increase in ITN coverage and improvements in the lifetime of active chemicals used in ITNs, as well as the physical integrity of ITNs in order to enhance the impact of ITNs in combating malaria. Also, studies in Clarke et al. (2001), Lengeler (2004b,a), Pryce et al. (2018) and Yang et al. (2018) have shown that ITNs contribute significantly in preventing the spread of malaria. Furthermore, ITNs have been shown to perform better in preventing the spread of malaria than untreated nets (Clarke et al., 2001; Pryce et al., 2018), while PBO ITNs have been shown to contribute more in reducing malaria prevalence in regions with high pyrethroid resistance by killing resistant mosquitoes and reducing the rate at which mosquitoes acquire blood meals (Gleave et al., 2018; Martin et al., 2021) than traditional ITNs. There is a positive correlation between the parameters  $\beta_{r1}$ ,  $p_{vh}$ ,  $p_{hv}$ ,  $\sigma_{\rm s}$  and malaria prevalence. Other significantly impactful parameters that correlate negatively with the response function are  $\sigma_r$ , the natural mortality rates of sensitive and resistant mosquitoes ( $\mu_{s0}$  and  $\mu_{r0}$ ), the recovery rate of symptomatically infectious humans  $(\gamma_{sh})$ , and the

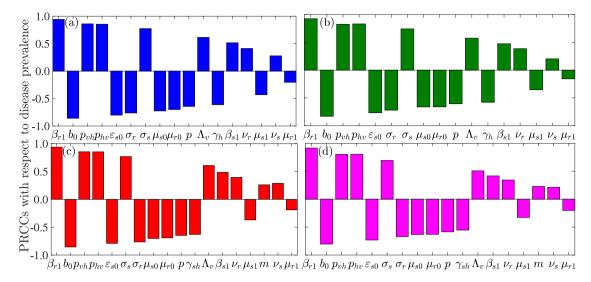


Fig. 10. Partial rank correlation coefficients (PRCCs) showing the contributions of parameters of the model (2.1) with (a) constant ITN efficacy, (b) waning ITN efficacy and periodic ITN replacement, and the model (3.1) with (c) constant ITN efficacy, (d) waning ITN efficacy and periodic ITN replacement to variability in human malaria prevalence  $((E_h + I_{sh} + I_{ah})/N_h)$ . A positive (negative) PRCC depicts a positive (negative) correlation between malaria prevalence in humans and the corresponding parameter, while the magnitude of the PRCC depicts the strength of the correlation with  $\pm 1$  depicting the strongest correlation and zero, depicting no correlation. Parameters with very low significance are not shown. The baseline parameters used for the simulation are presented in Table 1.

modification factor of the efficacy of ITNs for resistant mosquitoes (p), while other significantly impactful parameters that correlate positively with the response function are the mosquito recruitment rate  $(\Lambda_v)$  and the maximum biting rate of sensitive mosquitoes  $(\beta_{s1})$ . Parameters such as the proportion of mosquitoes that develop resistance at birth  $(\theta)$ , the useful lifespan of ITNs (T), and the minimum biting rates of sensitive and resistant mosquitoes  $(\beta_{s0}$  and  $\beta_{r0}$ , respectively,) are among the least influential parameters (and are not shown in Fig. 10). This confirms the minimal malaria risk in places with low mosquito concentrations or where mosquitoes are prevented from biting humans a lot.

### 5.3. Assessing the impact of ITN efficacy

In this Section, we compute the control reproduction number of the model (3.1) with constant ITN efficacy ( $R_c$ ), the time-averaged reproduction number of the non-autonomous version of the model (3.1) ( $\bar{R}_c$ ), and the actual reproduction number of the non-autonomous model (3.1) ( $R_c^T$ ) as functions of ITN coverage ( $b_0$ ) for various values of the initial ITN efficacies against sensitive and resistant mosquitoes ( $\epsilon_{s0}$  and  $\epsilon_{r0}$ , respectively). The numerical scheme described in Wang and Zhao (2008b) and Ngonghala et al. (2014) is used to compute the reproduction number ( $R_c^T$ ). The results obtained and depicted in Fig. 11 show that the model with constant ITN efficacy underestimates

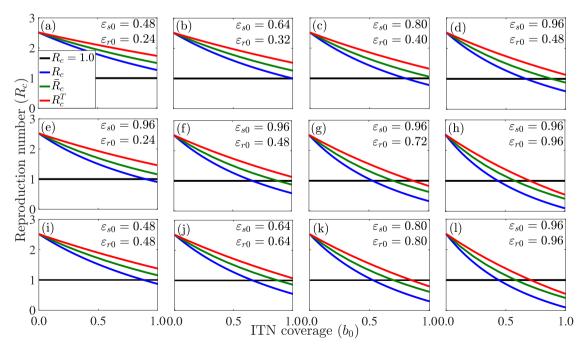


Fig. 11. Profiles of the control reproduction number of the model (3.1) with constant ITN efficacy denoted by  $R_c$  (blue curve), time-averaged ITN efficacy denoted by  $\bar{R}_c$  (green curves), and periodic ITN efficacy denoted by  $R_c^T$  (red curves) as functions of ITN coverage  $(b_0)$  for various initial ITN efficacies against sensitive and resistant mosquitoes  $(\epsilon_{s0})$  and  $\epsilon_{r0}$ , respectively). (a)–(d): The efficacy  $(\epsilon_{s0})$  is varied, while p=0.5. (e)–(h): The efficacy  $(\epsilon_{r0})$  is varied, while  $\epsilon_{r0}$  is fixed. (i)–(l): Both  $\epsilon_{s0}$  and  $\epsilon_{r0}$  are varied with  $\epsilon_{s0}=\epsilon_{r0}$ . The black horizontal line representing a control reproduction number of one is denoted by  $R_c^1$ . Other parameter values used are presented in Table 1.

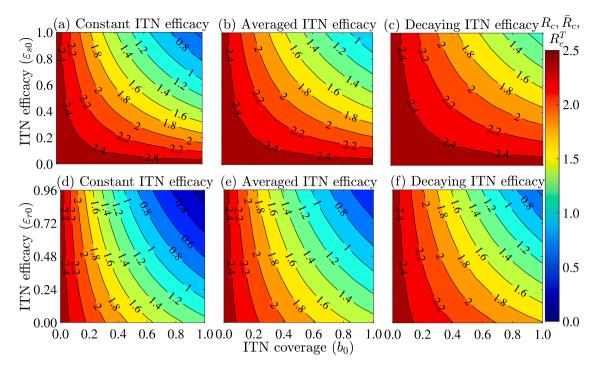


Fig. 12. Contour plots of the control reproduction number of the model (3.1) with constant ITN efficacy ( $R_c$ ), time-averaged ITN efficacy ( $\bar{R}_c$ ), and periodic ITN efficacy ( $R_c^T$ ) as functions of ITN coverage ( $b_0$ ) and initial ITN efficacies against (a)–(d): sensitive mosquitoes ( $\epsilon_{s0}$ ) and (e)–(h): resistant mosquitoes ( $\epsilon_{r0}$ ). Values of the other parameters used for generating the contour plots are given in Table 1

the control reproduction number (and hence the burden of the disease) and that reducing the control reproduction number below zero might be impossible if the efficacy of ITNs against sensitive or resistant mosquitoes is very low. In particular, if  $\varepsilon_{s0}=0.48$  or  $\varepsilon_{s0}=0.64$  and p=0.5 (which corresponds to  $\varepsilon_{r0}=0.24$  or  $\varepsilon_{r0}=0.32$ ), reducing any of the control reproduction numbers  $(R_c,\bar{R}_c,\text{ or }R_c^T)$  will never be possible even if everybody sleeps under an ITN (Fig. 11(a)-(b)). For populations that prioritize ITNs of higher efficacies, lower ITN coverage levels are required to reduce  $R_c$ ,  $\bar{R}_c$ , or  $R_c^T$  below unity. In particular for ITNs that are 80% efficacious against sensitive mosquitoes (i.e., if  $\varepsilon_{s0} = 0.80$ ) and p = 0.5 (or  $\varepsilon_{r0} = 0.40$ ), at least 79% of the population is required to use ITNs in order to reduce the reproduction number of the model (3.1) with constant ITN-efficacy below one (blue curve in Fig. 11(c)). However, for the model with periodic ITN efficacy, reducing any of the control reproduction numbers  $\bar{R}_c$ , or  $R_c^T$  below unity under this 80% ITN efficacy scenario is impossible, even if everybody uses ITNs (green and red curves in Fig. 11(c)). Furthermore, if the efficacy of ITNs against sensitive mosquitoes is 96% and p = 0.5 (or  $\varepsilon_{r0} = 0.48$ ), at least 66% of the population is required to use ITNs in order to reduce the reproduction number  $(R_c)$  below one for the model with constant ITN efficacy (blue curve in Fig. 11(d)), 87% ITN coverage is required for the model with a time-averaged ITN efficacy (green curve in Fig. 11(d)), and reducing  $R_c^T$  below one is unattainable even if everyone in the community uses highly efficacious ITNs (red curve in Fig. 11 (d)).

Next, the impact of the efficacy of ITNs against resistant mosquitoes  $(\varepsilon_{r0})$  on the three control reproduction numbers  $(R_c, \bar{R}_c, \text{ and } R_c^T)$ , when the efficacy of ITNs against sensitive mosquitoes  $(\varepsilon_{s0})$  is held at its baseline value of 96% is assessed. The results obtained and presented in Fig. 11(e)–(h) show that for a low efficacy of  $\varepsilon_{r0}=0.24$ , 90% ITN coverage is required to reduce  $R_c$  below one, while reducing  $\bar{R}_c$  or  $R_c^T$  below one will never be possible (green and red curves in Fig. 11(e)). The case for which  $\varepsilon_{r0}=0.24$  (i.e., Fig. 11(f)) has already been discussed under Fig. 11(d) above. For a moderately high efficacy of  $\varepsilon_{r0}=0.72$ , reducing  $R_c$  below one requires a 53% ITN coverage (blue curve in Fig. 11(g)), reducing  $\bar{R}_c$  below one requires a 70% ITN coverage (green curve in Fig. 11(g)), while reducing  $R_c^T$  below one requires a 87% ITN coverage (red curve in Fig. 11(g)). The high efficacy case (i.e., the case when  $\varepsilon_{r0}=0.96$ ) in Fig. 11(h) is discussed below.

To assess the impact of pyrethroid-PBO nets on malaria control, the control reproduction numbers  $(R_c, \bar{R}_c, \text{ and } R_c^T)$  are computed as functions of ITN coverage  $(b_0)$  for various values of ITN efficacies when the modification factor (p) of the ITN efficacy against resistance mosquitoes is one (i.e., when  $\varepsilon_{s0} = \varepsilon_{r0}$ ). The results obtained (depicted in Fig. 11(i)–(l)), show that if  $\epsilon_{s0}=\epsilon_{r0}=0.48$ , at least 89% ITN coverage is required to reduce  $R_c$  below one (blue curve in Fig. 11(i)), while reducing  $\bar{R}_c$  or  $R_c^T$  below one will never be possible even if the entire populace uses ITNs (green and red curves in Fig. 11(i)). On the other hand, if  $\varepsilon_{s0} = \varepsilon_{r0} = 0.80$ , at least 53% (70%) ITN coverage is required to reduce  $R_c$  ( $\bar{R}_c$ ) below one, while reducing  $R_c^T$ below one requires 85% of the community to use ITNs (red curve in Fig. 11(i)). Even lower ITN coverages are required for higher efficacies. Specifically, if  $\varepsilon_{s0} = \varepsilon_{r0} = 0.96$ , reducing  $R_c$  below one requires only a 44% ITN coverage (blue in Fig. 11(h) and (l)), while reducing  $\bar{R}_c$ below one requires a 60% ITN coverage (blue in Fig. 11(h) and (l)). For this same case in which  $\varepsilon_{s0}=\varepsilon_{r0}=0.96,~R_c^T$  can be brought below unity, with a 71% ITN coverage (red curve in Fig. 11(h) and (l)). Similar results for the model (2.1) are presented in Figs. S8-S9 of the SI.

In summary, the results depicted in Fig. 11 show a significant reduction of about 46% in the proportion of the population required to use ITNs in order to reduce  $R_c$  below one when ITNs of efficacy  $\varepsilon_{s0}=\varepsilon_{r0}=0.96$  compared to ITNs of efficacy  $\varepsilon_{s0}=0.96$  and  $\varepsilon_{r0}=0.24$ are used (comparing the blue curves in Fig. 11(e) and (h)) and that although reducing  $\bar{R}_c$  ( $R_c^T$ ) below one is possible with 60% (71%) ITN coverage, when  $\varepsilon_{s0} = \varepsilon_{r0} = 0.96$ , reducing  $\bar{R}_c$  ( $R_c^T$ ) below one is impossible when  $\varepsilon_{s0}=0.96$  and  $\varepsilon_{r0}=0.24$  (comparing the green curves in Fig. 11(e) and (h) and the red curves in Fig. 11(e) and (h)). Under the very high (96%) ITN efficacy against sensitive and resistant mosquito scenario, the autonomous model with constant ITN efficacy underestimates the reproduction number of the non-autonomous version by about 27% (comparing the intersection points of the blue and red curves with the black curve in Fig. 11(h) and (l)), while the model with averaged ITN efficacy underestimates the reproduction number of the non-autonomous model by about 11% (comparing the intersection points of the blue and green curves with the black curve in Fig. 11(h) and (1)). The scenario with  $\varepsilon_{s0} = \varepsilon_{r0}$  can be associated with control measures involving the use of pyrethroid-PBO nets. Detailed profiles for various combinations of ITN coverages and ITN efficacies required to reduce the reproduction numbers  $R_c$ ,  $\bar{R}_c$ , and  $R_c^T$  below unity are depicted by the contour plots in Fig. 12. Results for various values of the useful lifespan of ITNs, the replacement time of ITNs, the proportion of exposed humans who become symptomatic after the incubation period, and the rate at which adult mosquitoes develop resistance to insecticides are depicted in Figs. S11, S13 and S14 of the SI.

### 5.4. Assessing the impact of waning (decaying) ITN efficacy

In this section, the model (3.1) with constant, time averaged, and waning ITN efficacy are simulated to assess the impact of widespread use of ITNs of different efficacies on the control reproduction number  $(R_c)$  of the autonomous version of the model (3.1) with constant ITN efficacy and  $(\bar{R}_c \text{ and } R_c^T)$  of the non-autonomous version of the model (3.1) with waning ITN efficacy as functions of ITN coverage  $(b_0)$  for different values of initial ITN efficacies ( $\varepsilon_{i0}$ , j = r, s), the rate at which adult mosquitoes lose resistance to insecticides ( $\sigma_r$ ), and the proportion of exposed humans who become symptomatically infectious at the end of the incubation period (m). The results obtained and depicted in Fig. 13, indicate that for low efficacious ITNs, e.g., if  $\varepsilon_{s0} = 0.48$ and p = 0.5, reducing the control reproduction number below unity and hence containing the disease is impossible even if everybody in the community uses ITNs (red curves in Fig. 13(a), (e), and (i)). The proportion of the population required to use ITNs in order to reduce the control reproduction number below one reduces in communities that prioritize ITNs of higher efficacies. Specifically, for ITNs of moderately high efficacy (e.g., if  $\varepsilon_{s0}=0.80$  and p=0.5), at least 79% of the population is required to use ITNs in order to reduce  $R_c$  below unity (green curve in Fig. 13(a)), while reducing  $\bar{R}_c$  or  $R_c^T$  below one is impossible (green curves in Fig. 13(e) and (i)). It should be mentioned that for  $\varepsilon_{s0} = 0.80$  or  $\varepsilon_{s0} = 0.96$ , reducing either  $\bar{R}_c$  or  $R_c^T$  below one is not possible for the model (2.1) (Fig. S8 of the SI). Furthermore, for ITNs of very high efficacy (e.g., if  $\varepsilon_{s0} = 0.96$  and p = 0.5), a 66% ITN coverage is required to reduce  $R_c$  below one for the model (3.1) with constant ITN efficacy (blue curve in Fig. 13(a)), an 87% ITN coverage is

required to reduce  $\bar{R}_a$  below one for the model (3.1) with time-averaged ITN efficacy (blue curve in Fig. 13(e)), while reducing  $R^T$  below one is still impossible for the model (3.1) with waning ITN efficacy (blue curve in Fig. 13(i)). On the other hand, for ITNs with very low efficacy against resistant mosquitoes (e.g., if  $\varepsilon_{r0} = 0.24$  and  $\varepsilon_{s0} = 0.96$ ), 90% of the population is required to use ITNs to reduce  $R_c$  below one (blue curve in Fig. 13(b)), while reducing  $\bar{R}_c$  or  $R_c^T$  below one is unattainable (blue curves in Fig. 13(f) and (j)). However, for ITNs of very high efficacy against resistant mosquitoes (e.g., if  $\varepsilon_{r0} = \varepsilon_{s0} = 0.96$ ), only 44% of the population is required to use ITNs in order to reduce  $R_c$ below one (blue curve in Fig. 13(b)), 60% of the population is required to use ITNs in order to reduce  $\bar{R}_c$  below one (blue curve in Fig. 13(f)), while to reduce  $R^T$  below one requires up to 71% of the population to use ITNs (blue curve in Fig. 13(j)). For the model (2.1) and ITNs of efficacy 96% against resistant mosquitoes, at least 53% ITN coverage is necessary to bring R<sub>c</sub> below one, 70% ITN coverage is required to bring  $\bar{R}_c$  below one, and 83% ITN coverage is required to bring  $R_c^T$  below one (Fig. S8 of the SI). In summary, the model (3.1) with constant ITN efficacy underestimates  $R_c$  compared to the model (2.1) with constant ITN efficacy, while the model (3.1) with waning ITN efficacy and periodic ITN replacement time underestimates  $\bar{R}_c$  and  $R_c^T$ compared to the model (2.1) with waning ITN efficacy and periodic ITN replacement time. Additionally, the model (3.1) with constant ITN efficacy underestimates the control reproduction number compared to the model (3.1) with waning ITN efficacy and periodic ITN replacement time (i.e.,  $R_c < \bar{R}_c < R_c^T$  always).

To assess the impact of losing resistance to insecticides (or restoring susceptibility of resistant mosquitoes to insecticides), the control reproduction numbers ( $R_c$ ,  $\bar{R}_c$ , and  $R_c^T$ ) of the model (3.1) are studied as functions of ITN coverage ( $b_0$ ) for different values of the rate at which adult mosquitoes lose resistance to insecticides ( $\sigma_r$ ). The results depicted in Fig. 13(c), (g), and (k) show that when more adult mosquitoes lose resistance to insecticides, lower ITN coverage is required for containing the malaria disease. In particular, if the rate at which adult mosquitoes lose resistance to insecticides is as low as  $\sigma_r = 0.25$  per day, then 87% of the population is required to use ITNs to reduce  $R_c$ 

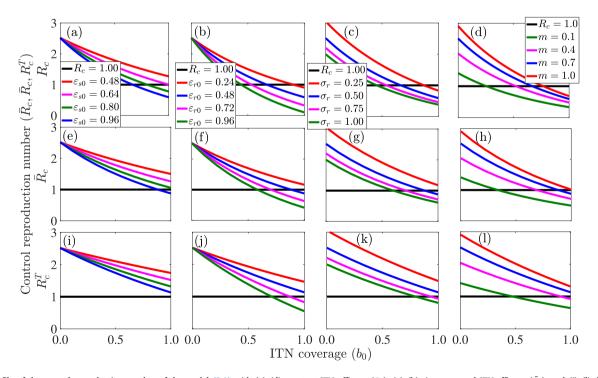


Fig. 13. Profile of the control reproduction number of the model (3.1) with (a)–(d) constant ITN efficacy ( $R_c$ ), (e)–(h) time-averaged ITN-efficacy ( $\bar{R}_c$ ), and (i)–(l) decaying ITN efficacy and periodic ITN replacement time ( $R_c^T$ ) as functions of ITN coverage ( $b_0$ ) for different values of (a), (e), and (i): ITN efficacy ( $\epsilon_{s0}$ ), (b), (f), and (j): ITN efficacy ( $\epsilon_{s0}$ ), (c), (g), and (k): rates at which adult mosquitoes lose resistance to insecticides ( $\sigma_r$ ), and (d), (h), and (l): the proportion of exposed humans who are symptomatic at the end of the incubation period (m). The values of the other parameters used for the simulations are presented in Table 1.

below one (red curve in Fig. 13(c)), while reducing  $\bar{R}_c$  or  $R_c^T$  below one is impossible even if everybody sleeps under an ITN (red curves in Fig. 13(g) and (k)). The proportion of humans required to sleep under ITNs reduces with increases in the rate at which adult mosquitoes lose resistance to insecticides. Specifically, if  $\sigma_r = 0.75$  per day, then at least 53% (70%) of the population must use ITN protection to reduce  $R_c$  ( $\bar{R}_c$ ) below one (magenta curves in Fig. 13(c) and Fig. 13(g), respectively), while 92% of the population must adopt ITN protection to reduce  $R_c^T$  below one (magenta curve in Fig. 13(k)). The reduction in the required ITN coverage is more significant if the rate at which resistant mosquitoes lose resistance is higher. Results for the model (2.1) with constant and decaying ITN efficacy are presented in Fig. S10 of the SI.

The impact of incorporating a separate asymptomatic infectious human class on malaria dynamics is highlighted in Fig. 13(d), (h), and (l). This result reveals that disease burden is higher in models in which asymptomatic infectious humans are not accounted for through a separate asymptomatic infectious human class. In particular, if the efficacy of ITNs against sensitive mosquitoes is 96% and m=1.0 (i.e., if every exposed human becomes symptomatic), then at least 76% of the population is required to use ITNs in order to reduce  $R_c$  below one (red curve in Fig. 13(d)), while reducing  $\bar{R}_c$  or  $R_c^T$  below unity is impossible (red curves in Fig. 13(h) and (l)). But if only 10% of exposed humans become symptomatic at the end of the incubation period (i.e., if m=0.1), then only about 25% (34%) of the human population is required to use ITNs to reduce  $R_c$  ( $\bar{R}_c$ ) below one (green curves in Fig. 13(d) and (h), respectively), and 48% ITN coverage is required to reduce  $R_c^T$  below one (green curve in Fig. 13(l)).

#### 6. Discussion and conclusion

Insecticide-treated nets are the most inexpensive and widely used control measures against malaria in many malaria-endemic countries (Lengeler, 2004a; Phillips-Howard et al., 2003; Trape et al., 2011; Bhatt et al., 2015; Lindsay et al., 2021). Since even untreated bed-nets still protect humans from mosquito bites, introducing insecticides in bednets, as is the case with ITNs, provides an additional layer of protection (by repelling mosquitoes from humans or killing mosquitoes that get into contact with the ITNs). Insecticide-treated nets have contributed significantly in decreasing malaria morbidity and mortality among children and pregnant women in Africa (D'Alessandro et al., 1995; Nevill et al., 1996; Habluetzel et al., 1997; Ter Kuile et al., 2003; Trape et al., 2011). Unfortunately, these, and other benefits of using ITNs in the fight against malaria are threatened by factors such as net attrition, natural and human-induced decay in ITN efficacy (Minakawa et al., 2008; Briët et al., 2012; McLean et al., 2014; Bush et al., 2017; Diema et al., 2017; Short et al., 2018; Sherrard-Smith et al., 2019; Solomon et al., 2019)), and resistance exhibited by mosquitoes to insecticides (e.g. pyrethroids) used in ITNs (World Health Organization, 2013; Brogdon and McAllister, 1998.; Mawejje et al., 2013; Churcher et al., 2016; Ranson and Lissenden, 2016; Namias et al., 2021).

In this study, mathematical models for the dynamics of malaria that account for the decay in ITN efficacy over time and mosquito resistance to insecticides are formulated and analyzed both analytically and numerically. One version of the models accounts for partially immune humans who contribute in disease transmission, while the other factors in an asymptomatic infectious human class, with a partially immune class that does not contribute to disease transmission. Both versions are considered for the case in which ITN efficacy is constant and the case in which ITN efficacy decays over time. Analytical results of versions of both models in which the efficacy of ITNs is constant show that both models exhibit a forward bifurcation when the control reproduction number  $(R_c)$  is greater than one and that a backward bifurcation is possible for some parameter regimes for which  $R_c < 1$ . These results are confirmed using numerical simulations. The existence of a forward bifurcation depicts a situation in which the disease can invade or establish itself within a community, while the existence of a backward bifurcation in a disease model renders control or elimination difficult since the model's reproduction number must be lowered substantially below unity (i.e., below another threshold,  $R_a^{bb}$ ) for the unique diseasefree equilibrium to be globally asymptotically stable. Thus, disease control measures must be implemented and sustained until  $R_c < R_c^{bb}$ . Furthermore, the study suggests the possibility of disease elimination in the autonomous models due to the assumption of constant ITN efficacy over time. However, this is not the case with the non-autonomous models, due to the additional realism of decaying ITN efficacy built in. Specifically, due to replacement of ITNs after their useful life, the dynamics of the non-autonomous models are oscillatory with period corresponding to the replacement time of ITNs. This finding is in line with results from other studies that have shown that ITNs as the only control and mitigation measure against malaria are insufficient for containing the disease (Ngufor et al., 2011; West et al., 2014; Protopopoff et al., 2015; Killeen and Smith, 2007; Govella et al., 2010; Bugoro et al., 2011; Govella and Ferguson, 2012).

A global uncertainty and sensitivity analysis of each of the models considered in this study suggests that uncertainty or variability in the maximum biting rate of resistant mosquitoes  $(\beta_{ri})$ , the ITN coverage  $(b_0)$ , the probability of an infectious mosquito (human) infecting a susceptible human (mosquito),  $p_{vh}$  ( $p_{hv}$ ), the initial ITN efficacy ( $\epsilon_{s0}$ ), and the rate at which resistant (sensitive) mosquitoes lose (develop) resistance to insecticides,  $\sigma_r$  ( $\sigma_s$ ) will introduce the greatest uncertainty and variability in the control reproduction number for the models with constant ITN efficacy and in the cumulative disease prevalence for both the model with constant and decaying ITN-efficacy. In particular, increases in  $\beta_{r1}, p_{vh}, p_{hv}$ , and  $\sigma_s$  will lead to significant increases in the control reproduction number and disease prevalence, while increases in  $b_0, \varepsilon_{s0}$ , and  $\sigma_r$  will lead to significant reductions in the control reproduction number and disease prevalence. Other parameters that have a significant impact on the control reproduction number and malaria prevalence are the natural mortality rates of sensitive and resistant mosquitoes ( $\mu_{s0}$  and  $\mu_{r0}$ , respectively), the modification factor for the efficacy of ITNs against resistant mosquitoes, the recovery rate of symptomatically infectious humans ( $\gamma_{sh}$ ), the mosquito recruitment rate  $(\Lambda_v)$ , and the maximum biting rate of sensitive mosquitoes  $(\beta_{s1})$ . Some of these parameters can be associated with malaria control measures. Specifically, mosquito-biting rates can be lowered by increasing the use of personal protective measures such as ITNs or making home changes to limit exposure to mosquitoes. The high impact of  $b_0$ ,  $\varepsilon_{s0}$ , and p implies that replacing ITNs regularly or when the efficacy has waned to a certain level, and ensuring that as many humans as possible sleep under ITNs is important for controlling malaria. Bio-control measures involving the use of transmission-blocking Wolbachia bacteria can reduce the likelihood of mosquito-to-human transmission (Kambris et al., 2010; Hughes et al., 2011; Walker and Moreira, 2011). Furthermore, the mosquito reproduction rate  $(\Lambda_v)$  can be lowered by removing vector breeding grounds near human dwellings and through the use of larvicides, while the adult mosquito death rate can be increased through indoor residual spraying and the use of ITNs and adulticides. The recovery rate  $(\gamma_{sh})$  can be boosted via anti-malaria drug therapy (e.g., artemether-lumefantrine, artesunate-amodiaquine, artesunate-pyronaridine, and dihydroartemisinin-piperaquine against Plasmodium falciparum in the WHO African region World Health Organization, 2021). Also, since the rate at which adult mosquitoes develop resistance  $(\sigma_r)$  or lose resistance  $(\sigma_r)$  to insecticides has a significant impact on the control reproduction number and malaria prevalence albeit a lesser extent compared to  $\beta_{r1}$ ,  $b_0$ , and  $\epsilon_{s0}$ , designing new methods for managing insecticide resistance is critical for malaria control. For example, resistance to insecticides can be reduced through the use of new chemicals to which mosquitoes are not resistant or chemicals that can suppress or reverse insecticide resistance in mosquitoes.

Untreated nets (i.e., nets with no insecticides) serve as physical shields between humans and mosquitoes, while ITNs constitute both physical and chemical shields for humans sleeping beneath them from

mosquitoes. Hence, both untreated nets and ITNs have a positive impact on reducing the transmission of malaria. However, ITNs contribute more significantly in preventing the spread of malaria (Clarke et al., 2001; Lengeler, 2004b,a; Pryce et al., 2018; Yang et al., 2018). In particular, the efficacy of untreated nets is about half the original efficacy of ITNs (Clarke et al., 2001). The effectiveness of ITNs in combating malaria is threatened by mosquito resistance to insecticides used in ITNs such as pyrethroids. Since mosquito resistance to insecticides reduces the efficacy of ITNs and hence constitute a serious challenge to the sustained management of many mosquito-borne infections (Machani et al., 2020), boosting the effectiveness of ITNs with synergists such as PBOs that improve the ability of ITNs to kill resistant mosquitoes, as well as restrict their human biting capability is critical (Bingham et al., 2011; Dadzie et al., 2017). It has been reported in some countries that the use of PBO can completely restore mosquito susceptibility to insecticides (World Health Organization, 2021). This study assesses the impact of traditional ITNs and PBO ITNs on malaria prevalence and control through two different ITN efficacies - the efficacy of ITNs against sensitive mosquitoes and the efficacy of ITNs against resistant mosquitoes. These efficacies are different for traditional ITNs and both sensitive and resistant mosquitoes retain the same efficacy for PBO ITNs. Results of this study show that PBO ITNs can be more effective than traditional ITNs in malaria control, especially in areas with high mosquito resistance to insecticides and especially when the efficacies of the PBO ITNs are high. This is consistent with results in Staedke et al. (2020), Gleave et al. (2018) and Martin et al. (2021).

The study highlights the importance of factoring in asymptomatic infectious humans and the decay in ITN efficacy over time with periodic ITN replacement. Specifically, the results show that the control reproduction number (and hence disease burden) is underestimated in a model in which asymptomatic infectious humans are accounted for through a separate asymptomatic infectious compartment, compared to a model with no separate asymptomatic infectious human class in which asymptomatic infectious humans are accounted for through a partially immune class. This result is consistent with that in Aguilar and Gutierrez (2020). Furthermore, the study indicates that the model (2.1) with constant ITN efficacy underestimates the control reproduction number (and hence disease burden) compared to the model (2.1) with decaying ITN efficacy, while the model (3.1) with constant ITN efficacy underestimates the control reproduction number compared to the model (3.1) with decaying ITN efficacy. In particular, the ITN coverage level required to reduce the control reproduction number below one is lowest for the versions of the models with constant ITN efficacy followed by the versions of the models with time-averaged ITN efficacy, and then the versions of the models with decaying ITN efficacy. Depending on the initial ITN efficacy, it might or might not be possible to reduce the control reproduction number below one. For example, for ITNs with high initial efficacy against susceptible and resistant mosquitoes of 96%, it is possible to reduce the control reproduction number of the model (3.1) with decaying ITN efficacy below one through a 71% ITN coverage, whereas for the model (2.1), 83% of the population is required to use ITNs to reduce the control reproduction number below one. The study also shows that eliminating malaria is unattainable if ITN efficacy or use is low and ITNs are not supplemented with other control measures. This is in line with results in Lengeler et al. (1998) and Ngonghala et al. (2020).

Additionally, the study shows that ITNs that can retain their efficacy and integrity longer perform better in malaria control than ITNs with shorter lifespans and that replacing ITNs before their useful lifespans is better for malaria control than replacing them after the lifespan. This outcome is consistent with results in Ngonghala et al. (2014) and a study in Gnanguenon et al. (2014) that recommended a serviceable lifespan of two years for ITNs. The integrity of ITNs deteriorates over time due to several factors including net attrition, frequent washing, especially for light colored ITNs (Gnanguenon et al., 2014; Mutuku et al.,

2013; Githinji et al., 2010), etc. Hence, to maximize the protective and killing efficacy of ITNs, they should be replaced in a timely manner.

The models in this study are based on some assumptions. These include the fact that all ITNs are replaced at the same time. Accounting for variable ITN replacement times might lead to slightly different outcomes. The study also fails to account for the impact of human decision to use ITNs properly or improperly, which can lead to useful public health information. This aspect is under study and will be reported in a separate paper. Nonetheless, the study shows that introducing a separate class (from the partially immune class) to account for asymptomatic infectious humans, or failing to account for the decay in ITN efficacy, underestimates the control reproduction number and hence the burden of the disease and the effort required to contain the disease. Furthermore, the study shows that it is critically important for malaria control to replace ITNs before their prescribed useful lifespans, design ITNs with longer lifespans, and to use PBO ITNs (which are more effective in inhibiting or reversing insecticide resistance and in preventing malaria transmission than regular ITNs). Consequently, by using highly effective ITNs widely, chances of effectively controlling malaria are improved, particularly if the ITNs have long useful lives and are replaced before their useful lifespans expire.

### CRediT authorship contribution statement

**Calistus N. Ngonghala:** Conceptualization, Formal analysis, Data curation, Investigation, Methodology, Visualization, Writing – original draft, Writing – review & editing.

### **Declaration of competing interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

### Data availability

No data was used for the research described in the article.

### Acknowledgments

CNN acknowledges the support of the Simons Foundation, United States of America (Award #627346) and the National Science Foundation, United States of America (Grant Number: DMS #2151870).

### Appendix A. Supplementary data

Supplementary material related to this article can be found online at https://doi.org/10.1016/j.jtbi.2022.111281.

### References

Agarwal, R.P., O'Regan, D., 2008. Existence and Uniqueness of Solutions of Systems. Springer.

Aguilar, J.B., Gutierrez, J.B., 2020. An epidemiological model of malaria accounting for asymptomatic carriers. Bull. Math. Biol. 82 (3), 1–55.

Agusto, F., Valle, S., Blayneh, K., Ngonghala, C., Goncalves, M., Li, N., Zhao, R., Gong, H., 2013. The impact of bed-net use on malaria prevalence.. J. Theoret. Biol. 320, 58-65.

Alves, F.P., Durlacher, R.R., Menezes, M.J., Krieger, H., Silva, L., Camargo, E.P., 2002. High prevalence of asymptomatic Plasmodium vivax and Plasmodium falciparum infections in native Amazonian populations. Amer. J. Trop. Med. Hygiene 66 (6), 641–648.

Barbosa, S., Hastings, I.M., 2012. The importance of modelling the spread of insecticide resistance in a heterogeneous environment: the example of adding synergists to bed nets. Malar. J. 11 (1), 258.

Barbosa, S., Kay, K., Chitnis, N., Hastings, I.M., 2018. Modelling the impact of insecticide-based control interventions on the evolution of insecticide resistance and disease transmission. Parasites Vectors 11 (1), 482.

- Bhatt, S., Weiss, D., Cameron, E., Bisanzio, D., Mappin, B., Dalrymple, U., Battle, K., Moyes, C., Henry, A., Eckhoff, P., et al., 2015. The effect of malaria control on Plasmodium falciparum in Africa between 2000 and 2015. Nature 526 (7572), 207–211.
- Bingham, G., Strode, C., Tran, L., Khoa, P.T., Jamet, H.P., 2011. Can piperonyl butoxide enhance the efficacy of pyrethroids against pyrethroid-resistant Aedes aegypti? Trop. Med. Int. Health 16 (4), 492–500.
- Birget, P.L., Koella, J.C., 2015. A genetic model of the effects of insecticide-treated bed nets on the evolution of insecticide-resistance. Evol. Med. Public Health 2015 (1), 205–215.
- Blower, S.M., Dowlatabadi, H., 1994. Sensitivity and uncertainty analysis of complex models of disease transmission: an HIV model, as an example. Int. Stat. Rev. Revue Int. Stat. 62 (2), 229–243.
- Bousema, J.T., Gouagna, L.C., Drakeley, C.J., Meutstege, A.M., Okech, B.A., Akim, I.N., Beier, J.C., Githure, J.I., Sauerwein, R.W., 2004. Plasmodium falciparum gametocyte carriage in asymptomatic children in western Kenya. Malar. J. 3 (1), 1–6
- Boyd, M.F., 1949. Epidemiology: factors related to the definitive host. Malariology 1, 608-697.
- Briët, O.J., Hardy, D., Smith, T.A., 2012. Importance of factors determining the effective lifetime of a mass, long-lasting, insecticidal net distribution: a sensitivity analysis. Malar. J. 11 (1), 20.
- Brogdon, W., McAllister, J., 1998.. Simplification of adult mosquito bioassays through use of time-mortality determinations in glass bottles.. J. Am. Mosquito Control Assoc. 14, 159–164.
- Brooke, B., 2008. Can a single mutation produce an entire insecticide resistance phenotype? Trans. R. Soc. Trop. Med. Hygiene 102, 524–525..
- Bugoro, H., Cooper, R.D., Butafa, C., Iro'ofa, C., Mackenzie, D.O., Chen, C.-C., Russell, T.L., 2011. Bionomics of the malaria vector anopheles farauti in Temotu Province, Solomon Islands: issues for malaria elimination. Malar. J. 10 (1), 1–11.
- Bush, E.R., Short, R.E., Milner-Gulland, E.J., Lennox, K., Samoilys, M., Hill, N., 2017.
  Mosquito net use in an artisanal East African fishery. Conserv. Lett. 10 (4), 451–459.
- Castillo-Chavez, C., Song, B., 2004. Dynamical models of tuberculosis and their applications. Math. Biosci. Eng. 3, 361–404.
- Chaumeau, V., Kajeechiwa, L., Fustec, B., Landier, J., Naw Nyo, S., Nay Hsel, S., Phatharakokordbun, P., Kittiphanakun, P., Nosten, S., Thwin, M.M., et al., 2019. Contribution of asymptomatic Plasmodium infections to the transmission of malaria in Kayin State, Myanmar. J. Infect. Dis. 219 (9), 1499–1509.
- Chitnis, N., Cushing, J.M., Hyman, J., 2006. Bifurcation analysis of a mathematical model for malaria transmission. SIAM J. Appl. Math. 67 (1), 24–45.
- Chitnis, N., Hyman, J.M., Cushing, J.M., 2008a. Determining important parameters in the spread of Malaria through the sensitivity analysis of a mathematical model. Bull. Math. Biol. 70, 1272–1296.
- Chitnis, N., Hyman, J.M., Cushing, J.M., 2008b. Determining important parameters in the spread of malaria through the sensitivity analysis of a mathematical model. Bull. Math. Biol. 70 (5), 1272.
- Chitnis, N., Schapira, A., Smith, T., Steketee, R., 2010. Comparing the effectiveness of malaria vector-control interventions through a mathematical model. Am. J. Trop. Med. Hygiene 83 (2), 230–240.
- Churcher, T.S., Lissenden, N., Griffin, J.T., Worrall, E., Ranson, H., 2016. The impact of pyrethroid resistance on the efficacy and effectiveness of bednets for malaria control in Africa. Elife 5, e16090.
- Clarke, S.E., Bøgh, C., Brown, R.C., Pinder, M., Walraven, G.E., Lindsay, S.W., 2001. Do untreated bednets protect against malaria? Trans. Royal Soc. Trop. Med. Hygiene 95 (5), 457–462.
- Dadzie, S.K., Chabi, J., Asafu-Adjaye, A., Owusu-Akrofi, O., Baffoe-Wilmot, A., Malm, K., Bart-Plange, C., Coleman, S., Appawu, M.A., Boakye, D.A., 2017. Evaluation of piperonyl butoxide in enhancing the efficacy of pyrethroid insecticides against resistant *Anopheles gambiae sl* in Ghana. Malar. J. 16 (1), 1–11.
- Dal-Bianco, M.P., Koster, K.B., Kombila, U.D., Kun, J.F., Grobusch, M.P., Ngoma, G.M., Matsiegui, P.B., Supan, C., Salazar, C.L.O., Missinou, M.A., et al., 2007. High prevalence of asymptomatic Plasmodium falciparum infection in Gabonese adults. Am. J. Trop. Med. Hyg. 77 (5), 939–942.
- D'Alessandro, U., Olaleye, B., Langerock, P., Aikins, M., Thomson, M., Cham, M., Greenwood, B., McGuire, W., Bennett, S., Cham, B., 1995. Mortality and morbidity from malaria in Gambian children after introduction of an impregnated bednet programme. Lancet 345 (8948), 479–483.
- Davidson, G., Draper, C., 1953. Field studies of some of the basic factors concerned in the transmission of malaria. Trans. R. Soc. Trop. Med. Hygiene 47 (6), 522–535.
- Diekmann, O., Heesterbeek, J., Metz, J.J., 1990. On the definition and the computation of the basic reproduction ratio  $R_0$  in models for infectious diseases in heterogeneous populations. J. Math. Biol. 28 (4), 365–382.
- Diema, K.K., Dodam, K.K., Aarah-Bapuah, M., Asibi, A.J., 2017. Barriers to sustained use of the insecticide treated bed net in the upper east region of ghana. Int. J. Community Med. Public Health 4 (2), 500–505.
- Djènontin, A., Chabi, J., Baldet, T., Irish, S., Pennetier, C., Hougard, J.-M., Corbel, V., Akogbéto, M., Chandre, F., 2009. Managing insecticide resistance in malaria vectors by combining carbamate-treated plastic wall sheeting and pyrethroid-treated bed nets. Malar. J. 8 (1), 1–9.

- Djènontin, A., Chandre, F., Dabiré, K.R., Chabi, J., N'guessan, R., Baldet, T., Akog-béto, M., Corbel, V., 2010. Indoor use of plastic sheeting impregnated with carbamate combined with long-lasting insecticidal mosquito nets for the control of pyrethroid-resistant malaria vectors. Am. J. Trop. Med. Hygiene 83 (2), 266.
- Driessche, Watmough, 2002. Reproduction numbers and sub-threshold endemic equilibria for compartmental models of disease transmission. Math. Biosci. 180, 29–48.
- Dushoff, J., Huang, W., Castillo-Chavez, C., 1998. Backwards bifurcations and catastrophe in simple models of fatal diseases.. J. Math. Biol. 36, 227–248.
- Eke, R., Chigbu, L., Nwachukwu, W., 2006. High prevalence of asymptomatic Plasmodium infection in a suburb of Aba Town, Nigeria. Annals African Med. 5 (1), 42-45.
- Filipe, J.A.N., Riley, E.M., Drakeley, C.J., Sutherland, C.J., Ghani, A.C., 2007. Determination of the processes driving the acquisition of immunity to malaria using a mathematical transmission model. PLoS Comput. Biol. 3 (12), e255.
- Fuseini, G., Nguema, R.N., Phiri, W.P., Donfack, O.T., Cortes, C., Von Fricken, M.E., Meyers, J.I., Kleinschmidt, I., Garcia, G.A., Maas, C., et al., 2019. Increased biting rate of insecticide-resistant culex mosquitoes and community adherence to IRS for malaria control in urban malabo, Bioko Island, Equatorial Guinea. J. Med. Entomol. 56 (4), 1071–1077.
- Galatas, B., Bassat, Q., Mayor, A., 2016. Malaria parasites in the asymptomatic: looking for the hay in the haystack. Trends Parasitol. 32 (4), 296–308.
- Gallup, J.L., Sachs, J.D., 2001. The economic burden of malaria. Am. J. Trop. Med. Hygiene 64 (1\_suppl), 85–96.
- Giles, H., Warrel, D., 1993. Bruce Chwatts' Essential of Malariology, third ed. Edward Arnold, London, pp. 124–163.
- Githinji, S., Herbst, S., Kistemann, T., Noor, A.M., 2010. Mosquito nets in a rural area of Western Kenya: ownership, use and quality. Malar. J. 9 (1), 1-6.
- Gleave, K., Lissenden, N., Richardson, M., Choi, L., Ranson, H., 2018. Piperonyl butoxide (PBO) combined with pyrethroids in insecticide-treated nets to prevent malaria in Africa. Cochrane Database Syst. Rev. (11).
- Gnanguenon, V., Azondekon, R., Oke-Agbo, F., Beach, R., Akogbeto, M., 2014. Durability assessment results suggest a serviceable life of two, rather than three, years for the current long-lasting insecticidal (mosquito) net (LLIN) intervention in Benin. BMC Infect. Dis. 14 (1), 1–10.
- Govella, N.J., Ferguson, H., 2012. Why use of interventions targeting outdoor biting mosquitoes will be necessary to achieve malaria elimination. Front. Physiol. 3, 199.
- Govella, N.J., Okumu, F.O., Killeen, G.F., 2010. Insecticide-treated nets can reduce malaria transmission by mosquitoes which feed outdoors. Am. J. Trop. Med. Hygiene 82 (3), 415–419.
- Gumel, A., 2012. Causes of backward bifurcations in some epidemiological models. J. Math. Anal. Appl. 395 (1), 355–365.
- Gupta, S., Swinton, J., Anderson, R.M., 1994. Theoretical studies of the effects of heterogeneity in the parasite population on the transmission dynamics of malaria. Proc. R. Soc. B 256 (1347), 231–238.
- Habluetzel, A., Diallo, D., Esposito, F., Lamizana, L., Pagnonl, F., Lengeler, C., Traore, C., Cousens, S., 1997. Do insecticide-treated curtains reduce all-cause child mortality in Burkina Faso? Trop. Med. Int. Health 2 (9), 855–862.
- Hughes, G.L., Koga, R., Xue, P., Fukatsu, T., Rasgon, J.L., 2011. Wolbachia infections are virulent and inhibit the human malaria parasite Plasmodium falciparum in Anopheles gambiae. PLoS Pathogens 7 (5), e1002043.
- Iwashita, H., Dida, G.O., Sonye, G.O., Sunahara, T., Futami, K., Njenga, S.M., Chaves, L.F., Minakawa, N., 2014. Push by a net, pull by a cow: can zooprophylaxis enhance the impact of insecticide treated bed nets on malaria control? Parasites Vectors 7 (1), 1–15.
- Jiram, A.I., Ooi, C.H., Rubio, J.M., Hisam, S., Karnan, G., Sukor, N.M., Artic, M.M., Ismail, N.P., Alias, N.W., 2019. Evidence of asymptomatic submicroscopic malaria in low transmission areas in Belaga district, Kapit division, Sarawak, Malaysia. Malar. J. 18 (1), 1–12.
- Jordan, D., Smith, P., 2007. Nonlinear Ordinary Differential Equations: An Introduction for Scientists and Engineers, third ed. OUP Oxford.
- Kakkilaya, B., 2022. Immunity. (Accessed on January 4, 2022),
- Kambris, Z., Blagborough, A.M., Pinto, S.B., Blagrove, M.S., Godfray, H.C.J., Sinden, R.E., Sinkins, S.P., 2010. Wolbachia stimulates immune gene expression and inhibits Plasmodium development in Anopheles gambiae. PLoS Pathogens 6 (10), e1001143.
- Killeen, G.F., Smith, T.A., 2007. Exploring the contributions of bed nets, cattle, insecticides and excitorepellency to malaria control: a deterministic model of mosquito host-seeking behaviour and mortality. Trans. Royal Soc. Trop. Med. Hygiene 101 (9), 867–880.
- Killeen, G.F., Smith, T.A., Ferguson, H.M., Mshinda, H., Abdulla, S., Lengeler, C., Kachur, S.P., 2007. Preventing childhood malaria in africa by protecting adults from mosquitoes with insecticide-treated nets. PLoS Med. 4 (7).
- Laishram, D.D., Sutton, P.L., Nanda, N., Sharma, V.L., Sobti, R.C., Carlton, J.M., Joshi, H., 2012. The complexities of malaria disease manifestations with a focus on asymptomatic malaria. Malar. J. 11 (1), 1–15.
- Lengeler, C., 2004a. Insecticide-treated bed nets and curtains for preventing malaria. Cochrane Database Systematic Rev. (2).
- Lengeler, C., 2004b. Insecticide-treated bed nets and curtains for malaria control. Cochane Lib. 3.

- Lengeler, C., Armstrong-Schellenberg, J., D'Alessandro, U., Binka, F., Cattani, J., 1998.
  Relative versus absolute risk of dying reduction after using insecticide-treated nets for malaria control in africa.. Trop. Med. Int. Health 3, 286–290.
- Lindblade, K.A., Steinhardt, L., Samuels, A., Kachur, S.P., Slutsker, L., 2013. The silent threat: asymptomatic parasitemia and malaria transmission. Expert Rev. Anti-Infect. Therapy 11 (6), 623–639.
- Lindsay, S.W., Thomas, M.B., Kleinschmidt, I., 2021. Threats to the effectiveness of insecticide-treated bednets for malaria control: thinking beyond insecticide resistance. Lancet Global Health 9 (9), e1325–e1331.
- Lines, J., Myamba, J., Curtis, C., 1987. Experimental hut trials of permethrinimpregnated mosquito nets and eave curtains against malaria vectors in Tanzania. Med. Vet. Entomol. 1 (1), 37–51.
- Lopez-Perez, M., Alvarez, A., Gutierrez, J.B., Moreno, A., Herrera, S., Arévalo-Herrera, M., 2015. Malaria-related anemia in patients from unstable transmission areas in Colombia. Am. J. Trop. Med. Hygiene 92 (2), 294.
- Macdonald, G., et al., 1957. The epidemiology and control of malaria. Epidemiol. Control Malaria.
- Machani, M.G., Ochomo, E., Zhong, D., Zhou, G., Wang, X., Githeko, A.K., Yan, G., Afrane, Y.A., 2020. Phenotypic, genotypic and biochemical changes during pyrethroid resistance selection in Anopheles gambiae mosquitoes. Sci. Rep. 10 (1), 1–8.
- Marino, S., Hogue, I.B., Ray, C.J., Kirschner, D.E., 2008. A methodology for performing global uncertainty and sensitivity analysis in systems biology. J. Theoret. Biol. 254 (1), 178–196.
- Martin, J.L., Mosha, F.W., Lukole, E., Rowland, M., Todd, J., Charlwood, J.D., Mosha, J.F., Protopopoff, N., 2021. Personal protection with PBO-pyrethroid synergist-treated nets after 2 years of household use against pyrethroid-resistant Anopheles in Tanzania. Parasit. Vectors 14 (1), 1–8.
- Mawejje, H., Wilding, C., Rippon, E., Hughes, A., Weetman, D., Donnelly, M., 2013. Insecticide resistance monitoring of eld-collected *Anopheles gambiae s.l.* populations from Jinja, Eastern Uganda, identifies high levels of pyrethroid resistance. Med. Vet. Entomol. 27, 276–283.
- Maxwell, C.A., Msuya, E., Sudi, M., Njunwa, K., Carneiro, I., Curtis, C., 2002. Effect of community-wide use of insecticide-treated nets for 3–4 years on malarial morbidity in Tanzania. Trop. Med. Int. Health 7 (12), 1003–1008.
- McLean, K.A., Byanaku, A., Kubikonse, A., Tshowe, V., Katensi, S., Lehman, A.G., 2014.
  Fishing with bed nets on Lake Tanganyika: a randomized survey. Malar. J. 13 (1), 1–5.
- Mehlhorn, H., 2001. Encyclopedic reference of parasitology: diseases, treatment, therapy. Encyclopedia Parasitol..
- Minakawa, N., Dida, G.O., Sonye, G.O., Futami, K., Kaneko, S., 2008. Unforeseen misuses of bed nets in fishing villages along Lake Victoria. Malar. J. 7 (1), 1-6.
- Mohammed-Awel, J., Agusto, F., Mickens, R.E., Gumel, A.B., 2018. Mathematical assessment of the role of vector insecticide resistance and feeding/resting behavior on malaria transmission dynamics: Optimal control analysis. Infect. Dis. Modell. 3, 301–321.
- Mohammed-Awel, J., Iboi, E.A., Gumel, A.B., 2020. Insecticide resistance and malaria control: A genetics-epidemiology modeling approach. Math. Biosci. 325, 108368.
- Molineaux, L., Gramiccia, G., World Health Organization, et al., 1980. The Garki Project: Research on the Epidemiology and Control of Malaria in the Sudan Savanna of West Africa. World Health Organization.
- Molineaux, L., Shidrawi, G., Clarke, J., Boulzaguet, J., Ashkar, T., 1979. Assessment of insecticidal impact on the malaria mosquito's vectorial capacity, from data on the man-biting rate and age-composition. Bull. World Health Organ. 57 (2), 265.
- Mutuku, F.M., Khambira, M., Bisanzio, D., Mungai, P., Mwanzo, I., Muchiri, E.M., King, C.H., Kitron, U., 2013. Physical condition and maintenance of mosquito bed nets in Kwale County, coastal Kenya. Malar. J. 12 (1), 1–14.
- Mwangangi, J., Muturi, E., Muriu, S., Nzovu, J., Midega, J., Mbogo, C., 2013. The role of Anopheles arabiensis and Anopheles coustani in indoor and outdoor malaria transmission in Taveta District, Kenya. Parasites Vectors 6, 114.
- Nakata, Y., Kuniya, T., 2010. Global dynamics of a class of SEIRS epidemic models in a periodic environment. J. Math. Anal. Appl. 363 (1), 230–237.
- Namias, A., Jobe, N.B., Paaijmans, K.P., Huijben, S., 2021. The need for practical insecticide-resistance guidelines to effectively inform mosquito-borne disease control programs. Elife 10, e65655.
- Nedelman, J., 1984. Inoculation and recovery rates in the malaria model of Dietz, Molineaux, and Thomas. Math. Biosci. 69 (2), 209–233.
- Nedelman, J., 1985. Introductory review: some new thoughts about some old malaria models. Math. Biosci. 73 (2), 159–182.
- Nevill, C., Some, E., Mung'Ala, V., Muterni, W., New, L., Marsh, K., Lengeler, C., Snow, R., 1996. Insecticide-treated bednets reduce mortality and severe morbidity from malaria among children on the Kenyan coast. Trop. Med. Int. Health 1 (2), 139–146.
- Ngonghala, C.N., Del Valle, S.Y., Zhao, R., Mohammed-Awel, J., 2014. Quantifying the impact of decay in bed-net efficacy on malaria transmission. J. Theoret. Biol. 363, 247–261.
- Ngonghala, C.N., Mohammed-Awel, J., Zhao, R., Prosper, O., 2016. Interplay between insecticide-treated bed-nets and mosquito demography: implications for malaria control. J. Theoret. Biol. 397, 179–192.

- Ngonghala, C.N., Wairimu, J., Adamski, J., Desai, H., 2020. Impact of adaptive mosquito behavior and insecticide-treated nets on malaria prevalence. J. Biol. Systems 28 (02), 515–542.
- N'Guessan, R., Corbel, V., Akogbéto, M., Rowland, M., 2007. Reduced efficacy of insecticide-treated nets and indoor residual spraying for malaria control in pyrethroid resistance area, benin. Emerg. Infect. Diseases 13 (2), 199.
- Ngufor, C., N'Guessan, R., Boko, P., Odjo, A., Vigninou, E., Asidi, A., Akogbeto, M., Rowland, M., 2011. Combining indoor residual spraying with chlorfenapyr and long-lasting insecticidal bed nets for improved control of pyrethroid-resistant Anopheles gambiae: an experimental hut trial in Benin. Malar. J. 10 (1), 1–7.
- Obala, A.A., Mangeni, J.N., Platt, A., Aswa, D., Abel, L., Namae, J., Prudhomme O'Meara, W., 2015. What is threatening the effectiveness of insecticide-treated bednets? A case-control study of environmental, behavioral, and physical factors associated with prevention failure. PLoS One 10 (7), e0132778.
- Okell, L.C., Bousema, T., Griffin, J.T., Ouédraogo, A.L., Ghani, A.C., Drakeley, C.J., 2012a. Factors determining the occurrence of submicroscopic malaria infections and their relevance for control. Nature Commun. 3 (1), 1–9.
- Okell, L.C., Paintain, L.S., Webster, J., Hanson, K., Lines, J., 2012b. From intervention to impact: modelling the potential mortality impact achievable by different long-lasting, insecticide-treated net delivery strategies. Malar. J. 11 (1), 327.
- Osoro, J.K., Machani, M.G., Ochomo, E., Wanjala, C., Omukunda, E., Githeko, A.K., Yan, G., Afrane, Y.A., 2022. Insecticide resistant *Anopheles gambiae* have enhanced longevity but reduced reproductive fitness and a longer first gonotrophic cycle. Sci. Rep. 12 (1), 1–7.
- Phillips-Howard, P.A., Nahlen, B.L., Kolczak, M.S., Hightower, A.W., Ter Kuile, F.O., Alaii, J.A., Gimnig, J.E., Arudo, J., Vulule, J.M., Odhacha, A., et al., 2003. Efficacy of permethrin-treated bed nets in the prevention of mortality in young children in an area of high perennial malaria transmission in western Kenya. Amer. J. Trop. Med. Hygiene 68 (4\_suppl), 23–29.
- Protopopoff, N., Wright, A., West, P.A., Tigererwa, R., Mosha, F.W., Kisinza, W., Kleinschmidt, I., Rowland, M., 2015. Combination of insecticide treated nets and indoor residual spraying in northern Tanzania provides additional reduction in vector population density and malaria transmission rates compared to insecticide treated nets alone: a randomised control trial. PLoS One 10 (11), e0142671.
- Pryce, J., Richardson, M., Lengeler, C., 2018. Insecticide-treated nets for preventing malaria. Cochrane Database Syst. Rev. (11).
- Pulkki-Brännström, A.-M., Wolff, C., Brännström, N., Skordis-Worrall, J., 2012. Cost and cost effectiveness of long-lasting insecticide-treated bed nets-a model-based analysis. Cost Effect. Resource Allocat. 10 (1), 1–13.
- Ramaswamy, A., Mahabala, C., Basavaiah, S.H., Jain, A., Chouhan, R.R.S., 2020. Asymptomatic malaria carriers and their characterization in hotpops of malaria at Mangalore. Trop. Parasitol. 10 (1), 24.
- Ranson, H., Lissenden, N., 2016. Insecticide resistance in african anopheles mosquitoes: a worsening situation that needs urgent action to maintain malaria control. Trends Parasitol. 32 (3), 187–196.
- Sachs, J., Malaney, P., 2002. The economic and social burden of malaria. Nature 415 (6872), 680.
- Sáenz, F.E., Arévalo-Cortés, A., Valenzuela, G., Vallejo, A.F., Castellanos, A., Poveda-Loayza, A.C., Gutierrez, J.B., Alvarez, A., Yan, Y.H., Benavides, Y., et al., 2017.
  Malaria epidemiology in low-endemicity areas of the northern coast of Ecuador: high prevalence of asymptomatic infections. Malar. J. 16 (1), 1–10.
- Sherrard-Smith, E., Skarp, J.E., Beale, A.D., Fornadel, C., Norris, L.C., Moore, S.J., Mihreteab, S., Charlwood, J.D., Bhatt, S., Winskill, P., et al., 2019. Mosquito feeding behavior and how it influences residual malaria transmission across Africa. Proc. Natl. Acad. Sci. 116 (30), 15086–15095.
- Short, R., Gurung, R., Rowcliffe, M., Hill, N., Milner-Gulland, E., 2018. The use of mosquito nets in fisheries: A global perspective. PLoS One 13 (1), e0191519.
- Smalley, M., Sinden, R., 1977. *Plasmodium falciparum* gametocytes: their longevity and infectivity. Parasitology 74 (1), 1–8.
- Solomon, T., Loha, E., Deressa, W., Gari, T., Overgaard, H.J., Lindtjørn, B., 2019. Low use of long-lasting insecticidal nets for malaria prevention in south-central Ethiopia: A community-based cohort study. PLoS One 14 (1), e0210578.
- Staedke, S.G., Gonahasa, S., Dorsey, G., Kamya, M.R., Maiteki-Sebuguzi, C., Lynd, A., Katureebe, A., Kyohere, M., Mutungi, P., Kigozi, S.P., et al., 2020. Effect of long-lasting insecticidal nets with and without piperonyl butoxide on malaria indicators in Uganda (LLINEUP): a pragmatic, cluster-randomised trial embedded in a national LLIN distribution campaign. Lancet 395 (10232), 1292–1303.
- Stone, C., Gross, K., 2018. Evolution of host preference in anthropophilic mosquitoes. Malar. J. 17 (1), 1–11.
- Sumner, K.M., Mangeni, J.N., Obala, A.A., Freedman, E., Abel, L., Meshnick, S.R., Edwards, J.K., Pence, B.W., Prudhomme-O'Meara, W., Taylor, S.M., 2021. Impact of asymptomatic Plasmodium falciparum infection on the risk of subsequent symptomatic malaria in a longitudinal cohort in Kenya. Elife 10, e68812.
- Takken, W., Verhulst, N.O., 2013. Host preferences of blood-feeding mosquitoes. Annual Rev. Entomol. 58, 433–453.
- Teboh-Ewungkem, M., 2009. Malaria control: the role of local communities as seen through a mathematical model in a changing population-Cameroon. pp. 103–140.
- Teboh-Ewungkem, M., Tchuenche, J., Mukandavire, Z., 2009. Malaria control: the role of local communities as seen through a mathematical model in a changing population-Cameroon. In: in Disease Epidemiology, A. (Ed.), Adv. Dis. Epidemiol. 101–138, Nova Science Publishers; New York.

- Ter Kuile, F.O., Terlouw, D.J., Phillips-Howard, P.A., Hawley, W.A., Friedman, J.F., Kariuki, S.K., Shi, Y.P., Kolczak, M.S., Lal, A.A., Vulule, J.M., et al., 2003. Reduction of malaria during pregnancy by permethrin-treated bed nets in an area of intense perennial malaria transmission in western Kenya. Amer. J. Trop. Med. Hygiene 68 (4\_suppl), 50–60.
- The world fact book, 2022a. Birth Rate: Country Comparison. Central Intelligence Agency, Accessed on August 13, 2022. URL: https://www.cia.gov/the-world-factbook/field/birth-rate/country-comparison.
- The world fact book, 2022b. Life Expectancy at Birth. Central Intelligence Agency, Accessed on August 13, 2022. URL: https://www.cia.gov/the-world-factbook/field/life-expectancy-at-birth/.
- Tiono, A.B., Ouédraogo, A., Diarra, A., Coulibaly, S., Soulama, I., Konaté, A.T., Barry, A., Mukhopadhyay, A., Sirima, S.B., Hamed, K., 2014. Lessons learned from the use of HRP-2 based rapid diagnostic test in community-wide screening and treatment of asymptomatic carriers of Plasmodium falciparum in Burkina Faso. Malar. J. 13 (1), 1–9.
- Trape, J.-F., Tall, A., Diagne, N., Ndiath, O., Ly, A.B., Faye, J., Dieye-Ba, F., Roucher, C., Bouganali, C., Badiane, A., et al., 2011. Malaria morbidity and pyrethroid resistance after the introduction of insecticide-treated bednets and artemisinin-based combination therapies: a longitudinal study. Lancet Infect. Dis. 11 (12), 925–932.
- van den Driessche, P., Watmough, J., 2000. A simple SIS epidemic model with a backward bifurcation. J. Math. Biol. 40 (6), 525–540.
- Walker, T., Moreira, L.A., 2011. Can Wolbachia be used to control malaria? Memórias Inst. Oswaldo Cruz 106, 212–217.
- Wang, W., Zhao, X.-Q., 2008a. Threshold dynamics for compartmental epidemic models in periodic environments. J. Dynam. Differential Equations 20, 699–717.

- Wang, W., Zhao, X.-Q., 2008b. Threshold dynamics for compartmental epidemic models in periodic environments. J. Dynam. Differential Equations 20 (3), 699–717.
- Wat'senga, F., Manzambi, E.Z., Lunkula, A., Mulumbu, R., Mampangulu, T., Lobo, N., Hendershot, A., Fornadel, C., Jacob, D., Niang, M., et al., 2018. Nationwide insecticide resistance status and biting behaviour of malaria vector species in the Democratic Republic of Congo. Malar. J. 17 (1), 129.
- West, P.A., Protopopoff, N., Wright, A., Kivaju, Z., Tigererwa, R., Mosha, F.W., Kisinza, W., Rowland, M., Kleinschmidt, I., 2014. Indoor residual spraying in combination with insecticide-treated nets compared to insecticide-treated nets alone for protection against malaria: a cluster randomised trial in Tanzania. PLoS Med. 11 (4), e1001630.
- World Bank Data, 2020. Life Expectancy at Birth, Total (Years) Sub-Saharan Africa. World Bank, (Accessed on January 3, 2020).
- World Health Organization, 2011. Guidelines for Monitoring the Durability of Long-Lasting Insecticidal Mosquito Nets under Operational Conditions, WHO/HTM/NTD/WHOPES/2011.5. World Health Organization.
- World Health Organization, 2013. World Malaria Report 2013 Geneva. World Health Organization Malaria Report 2013. WHO Genva.
- World Health Organization, 2021. World Malaria Report 2021. World Health Organization, Geneva, Assessed on December 16, 2021. URL: https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2021.
- World Health Organization, et al., 2013. Guidelines for Laboratory and Field-Testing of Long-Lasting Insecticidal Nets. Technical Report, World Health Organization.
- Yang, G.-g., Kim, D., Pham, A., Paul, C.J., 2018. A meta-regression analysis of the effectiveness of mosquito nets for malaria control: the value of long-lasting insecticide nets. Int. J. Environ. Res. Public Health 15 (3), 546.