

Characterizing canopy complexity of natural and restored intertidal oyster reefs (*Crassostrea virginica*) with a novel laser-scanning method

Running Head: Investigating surface complexity on oyster reefs

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Author Contributions

KMK, SCM, DJC conceived and designed the research; KMK, SCM collected the observations; DJC, SCM performed data analysis; DJC, KMK, JT, SCM wrote and edited the manuscript

Abstract

The structural complexity of oyster reef canopy plays a major role in promoting biodiversity, balancing the sediment budget, and modulating hydrodynamics in estuarine systems. While oyster canopy structure is both spatially and temporally heterogeneous, oyster canopies are generally characterized using simple first-order quantities, like oyster density, which may lack the ability to sufficiently parameterize reef roughness. In this study, a novel laser scan approach was used to map the surface of intact reference and restored reefs (restoration age: 1 – 4 years) during low tide, when the oyster canopy was fully exposed. Measurements were used to estimate hydrodynamically-relevant roughness characteristics over the entire reef surface ($>140 \text{ m}^2$; 0.50 m resolution), providing estimates of the canopy height (h_c), standard deviation (σ_c), rugosity index (R), and fractal dimension (D). Average canopy heights ranged from 3.6 – 4.9 cm, with canopy height standard deviations between 1.4 and 2.0 cm. Mean rugosity indices and fractal dimensions were relatively low on the youngest (1 year) restored reef ($R=1.21$; $D=2.67$), with substantial increases observed for more mature reef canopies (4 years: $R=1.51$; $D=2.71$). Structural complexity was consistently greater on reef margins than in reef interiors. Increases in complexity were linked to restoration age, with older reefs exhibiting more complex oyster canopies. The highest fractal dimension was observed on the intact reference reef, highlighting the importance of sustained reef growth for maintaining higher-order structural complexity. Results provide spatially explicit surface roughness characterizations for healthy, intertidal oyster reefs, with applications in both restoration science and natural and nature-based feature design.

Keywords: estuary, canopy, complexity, laser scan, oyster, roughness

Implications for Practice

- Typically measured oyster canopy characteristics, including live oyster density and shell length, are insufficient for accurately describing surface roughness on intact and restored reefs.
- The canopies of restored intertidal oyster reefs become more structurally complex over time, and proper restoration techniques can produce surface roughness characteristics that match or exceed those found on intact reference reefs.
- The spatially explicit canopy characteristics described in this study can be used to inform design of natural and nature-based features, especially those designed to provide ecosystem services controlled by surface complexity (e.g. habitat provision, wave attenuation, etc.).

Introduction

Oysters are considered “ecosystem engineers”, so-called for their capacity to alter the physical, biological, and chemical signatures of the environment they inhabit (e.g. Morris et al. 2019) in a process of niche construction (Post & Palkovacs 2009). Habitat alteration occurs through a combination of (1) filter-feeding and waste deposition, which enhances water quality (Dame et al. 1989; Filippini et al. 2022) and sequesters carbon and nutrients (Chambers et al. 2018), and (2) reef-building, which is linked to shoreline stabilization (Meyer et al. 1997; McClenachan et al. 2020), current and wave attenuation (Cannon et al. 2022a), and habitat provision (Coen & Luckenbach 2000). Living oysters build complex three-dimensional structures, with reefs composed of branching clusters made up of mature living oysters, dead oyster shells, and newly recruited oyster spat (Figure 1b,c). These structures form a rigid canopy, composed of hundred (or thousands) of individual oyster clusters on the reef surface. Oyster canopies attenuate flows (Kitsikoudis et al. 2020), provide refuge for prey species (Peterson et al. 2003), and subsequently enhance biodiversity on the reef (Loch et al. 2021), with research suggesting that ecosystem benefits are directly linked to the structural complexity of the canopy itself (e.g. Hill & Weissburg 2013; Humphries et al. 2011; Soniat et al. 2004).

Over the last century, exogenous pressures have precipitated large declines in the global oyster population (>85% loss: Beck et al. 2011), and oyster-mediated ecosystem services have been simultaneously lost in affected coastal habitats. In recent years, coastal management has targeted construction and restoration of oyster reef habitat to recover these lost services, with research focused on effective strategies for recruiting oysters to natural reef or engineered reef-mimic structures (Goelz et al. 2020; Nitsch et al. 2021; Walters et al. 2021). Although restoration efforts have successfully recruited live oysters, a global meta-analysis of faunal abundance and diversity concluded that even successfully restored historical reef systems carry a considerable recovery debt as compared to pre-disturbed conditions (Hemraj et al. 2022). This recovery debt includes ecosystem services that may permanently lost, even after living oysters are re-introduced to the reef. An even more challenging task is the creation of new self-sustaining reef using artificial structures, an activity that has gained traction as a nature-based coastal defense aimed at slowing erosion (Morris et al. 2020). In a reef creation scenario, site environmental conditions and larval abundance may be less optimal as compared to sites of known historical reef. Focusing the design of artificial reef-like structures toward enhancing the recruitment and retention of target species

may improve long-term success in transforming artificial structures into living reef. In particular, the role of structural complexity on the reef surface has not been investigated. Oyster canopies are expected to change in height and complexity over time, especially on recently restored or constructed reefs where the canopy is actively growing, and structure-related ecosystem services may develop over relatively long timescales (Cannon et al. 2022b; La Peyre et al. 2014). While understanding the temporal development of canopy structure is imperative for evaluating the success of oyster reef restoration efforts, parameterizing the structural complexity of natural intact reef canopies is an equally important baseline for the design of natural and nature-based reef mimic structures.

Although the structural complexity of many marine (coral: Leon et al. 2015; Zawada & Brock 2009; mussel: Commito & Rusignuolo 2000) and terrestrial (forest: Atkins et al. 2018; Franklin & Van Pelt 2004) canopies have been studied extensively over the last several decades, studies of intertidal oyster reefs are relatively rare (exceptions: Margiotta et al. 2016; Karp et al., 2018). When reported, oyster reef canopy characterizations are generally restricted to estimates of the mean canopy height, living oyster density, and shell length as averaged over relatively few randomly selected points on the reef surface (e.g. Garvis et al. 2015; Walters et al. 2021; etc.). These measurements require intensive manual sampling, but they do not provide the spatial resolution required to describe an entire reef canopy or to estimate higher-order structural complexity parameters (e.g. rugosity, fractal dimension). However, the structural characteristics of intertidal oyster reefs provide unique opportunities for roughness measurements. The crest elevation of healthy oyster reefs is biologically mediated, and canopies experience full inundation and full exposure during high and low tide, respectively. While direct manual measurements may be extremely labor intensive, especially on large reef flats, low-tide exposure allows for indirect methodologies, like terrestrial laser scanning, to provide novel, high-resolution canopy measurements over the entire reef surface.

This study investigates the structural complexity of intertidal eastern oyster (*Crassostrea virginica*) canopies on intact reference and restored reefs in Mosquito Lagoon, Florida (USA). This work utilizes a space-for-time (SFT) approach to examine differences in reef characteristics related to restoration age, or time since restoration, with measurements collected 1-, 2- and 4-years post-restoration. We hypothesize that structural complexity will increase over time on restored

oyster reefs, with changes in canopy height, canopy height standard deviation, rugosity, and fractal dimension linked to restoration age. Furthermore, we hypothesize that oyster canopies may be more structurally complex than other rigid biological marine canopies (i.e. corals, mussels), which often lack the rough branching structures, length scale variability, and high element densities associated with intertidal oyster reefs. A novel method is proposed for characterizing canopy structure and roughness using a portable laser scanner, typically restricted to use in terrestrial settings. High-resolution laser scans were collected at low tide to help parameterize the structural complexity of oyster canopies on large reef flats (surface area: 144 – 440 m²) at relatively small spatial scales (0.25 m² computation grids). The results and analysis presented herein are among the first of their kind, providing valuable insight for coastal managers restoring natural reef habitats and engineers designing natural and nature-based coastal infrastructure that utilizes living oyster canopy.

Methods

Study area

Measurements were collected in Mosquito Lagoon, a biologically diverse subtropical estuary on the Atlantic coast of Florida (USA). Mosquito Lagoon is generally shallow (mean depth: 1 m) and microtidal, with seasonal water level fluctuations on the order of ± 50 cm. Tidal exchange with the Atlantic Ocean occurs through Ponce de Leon Inlet (tidal amplitude: ± 100 cm), which is positioned at the northernmost tip of the waterbody. Study sites were located approximately 15 km southeast of the inlet, where tidal amplitudes were 20 - 30 cm. The northern portion of the lagoon, including the study area, is characterized by a complex network of channels that flow through a maze of sandy shoals, mangroves, and salt marsh wetlands (Figure 1; Mehta & Brooks 1973). This study is focused on intertidal oyster (*Crassostrea virginica*) reefs, which are abundant in the northern reach of the lagoon where tidal amplitudes are high enough to promote periodic inundation (e.g. Garvis et al. 2015). Intertidal reef is a dominant landscape feature throughout this region, and oysters occupy the margins of mangrove wetlands and form extensive reefs on the shallow shoals between vegetated islands.

Laser scans were conducted in the summer of 2018 on four intertidal oyster reefs in northern Mosquito Lagoon (Figure 1; Table 1). This work uses a space-for-time (SFT) approach to investigate changes in the structural complexity of intertidal oyster reef canopies over time. Instead

of following the evolution of a single restored reef as it matures over years (or decades), multiple reefs of increasing restoration age are used to infer how oyster canopies may change over time. Measurements were collected on an intact reference-condition reef (Reference), as well as three restored reefs defined by their year of restoration: R-2014, R-2016, and R-2017. These restored reefs can also be defined using their restoration age, or time since restoration, where R-2017, R-2016, and R-2014, are approximately 1 year, 2 years, and 4 years old, respectively. As a consequence of the SFT study design, it is assumed that differences in observed reef canopy structure are largely linked restoration age, with only minor variability associated with reef identity. Further consideration of potential reef identity effects can be found in the discussion.

Identical restoration techniques were employed at each restored reef, as detailed in Garvis et al. (2015). Prior reef degradation was linked to recreational boating in the lagoon, with boat wakes acting to dislodge oyster clusters and pile disaggregated shells into mounds at the reef crest (Grizzle et al, 2002). Degraded reef crest elevations were initially lowered to match nearby intact reefs, and extruded polyethylene mesh (Vexar™) oyster mats (size: 0.5 x 0.5 m) were anchored to the reef surface using concrete weights. Each mat was constructed using 36 adult *C. virginica* shells distributed across the surface of the mat and attached vertically to mimic the structure of live reefs. These mats promoted natural oyster recruitment at the restored reefs, and live oyster densities at each site were similar to those observed on reference reefs within one year of restoration (Cannon et al. 2022a).

Field measurements

Surveys were conducted to estimate canopy characteristics on each study reef. Manual canopy characterizations are labor intensive and measuring the entire reef surface (surface area: 150 – 450 m²) is impractical. Instead, five 0.25m² sample quadrats were chosen on each reef to provide representative sample areas. Quadrat locations were selected haphazardly at low tide when the reef surface was exposed. Live oyster densities and shell lengths were estimated directly by counting and measuring all live *C. virginica* greater than 5mm in length in each quadrat. Individual quadrat measurements were used to compute reef-wide averages, medians, and standard errors. On each reef, a single quadrat was randomly selected and used to characterize the canopy height and canopy element density, defined as the total number of solid canopy elements, or oyster clusters, attached to the reef surface. Individual canopy elements were typically composed of both live and dead

oysters, as seen in Figure 2b. The mean canopy height and standard error were estimated by measuring the height of all canopy elements in the randomly selected quadrat, with heights measured from the reef surface to the highest point on the oyster cluster.

High-resolution canopy measurements were captured in detail using a FARO x330 terrestrial laser scanner. Although terrestrial laser scanners are more typically used for geospatial surveying in engineering operations, recent studies have demonstrated successful applications in complex biological structures, including forests (Wang et al. 2021) and blanket bogs (Chico et al. 2019). The FARO x330 is a phase-based laser scanner (pulse wavelength: 155 nm) with a maximum range of 330 m and a ranging error of ± 2 mm. The laser scanner was set to scan at maximum resolution, collecting 976 kpts/s at a step size of 0.009° , resulting in a point spacing of 1.5 mm at a distance of 10 m from the scanner. Laser scanning was completed at low tide during seasonal low water levels, ensuring that reef surfaces were maximally exposed. The laser scanner was deployed and repositioned multiple times on each reef to minimize the effects of shadows, which were created by individual canopy elements blocking the laser. Fixed spherical targets installed on tripods were used as reference points for aligning multiple scans, which were combined to produce a final point cloud for surface roughness analysis. Measurement positions were chosen such that the distance between the scanner and any individual target intended to remain in place for a subsequent scan position on a reef was never greater than approximately 10 m.

Data analysis

Laser scan data were initially imported and processed using FARO Scene Version 6. The key processing step is the registration (integration) of the multiple reef scans using the spherical targets that remained fixed for multiple scans. Data were filtered to remove erroneous stray points caused by dust and water vapor in the atmosphere. All measurements beyond the emergent edge of the reef, where the water surface intersected the reef canopy, were also removed. All point data were then exported to Matlab, where reef scans were subdivided into 0.25 m^2 computations grids ($50 \text{ cm} \times 50 \text{ cm}$) for further analysis. All computation grids with fewer than 2500 points (mean density: 1 point/ cm^2) were removed from analysis, as were all grids where the point bounding area (i.e. area of polygon bounding all points in grid cell) was less than 0.19 m^2 , or 75% of the total grid area, resulting in between 20 and 40% data reduction for each reef. Removed grid cells were largely (>90%) located along reef fringes, where fluctuating water levels led to inconsistent laser

measurements. This quality control helped reduce computational errors due to edge effects and low sampling densities.

In order to focus the analysis on oyster canopy structure, large-scale elevation changes related to the underlying bed morphology were removed from the point cloud (Figure 2). For each subsampled computation grid (50 cm x 50 cm), a plane was fit (least-squares) to the lowest 25% of points selected from each of 16 equally distributed cells (12.5 cm x 12.5 cm) within the larger subsample. The fit plane was then shifted to lowest measured elevation to represent the local sediment surface, or bed-plane (Figure 2a). This procedure allowed identification of the bed for a large range of point densities and bed slopes without relying on assumptions of spatial homogeneity in canopy heights, as would be required for simple random sampling. All computation grids with maximum bed slopes greater than 0.20 m/m were considered erroneous and removed from analysis.

All measured point elevations were converted to local canopy heights by subtraction from the bed-plane. Elevations were then averaged over 1 cm x 1 cm grid cells to normalize point densities across the measurement domain. This point density normalization was intended to reduce the effects of measurement resolution on roughness characterizations, as has been discussed in previous studies using manual measurement techniques (e.g. Knudby & LeDrew 2007). Although the choice of horizontal grid resolution (1cm) was somewhat arbitrary, it is consistent with the length of chain-links used in more high-resolution studies of coral canopy roughness (Knudby & LeDrew 2007). Canopy height estimates in grid cells without points were set equal to zero, and mean canopy heights (h_c) and canopy height standard deviations (σ_c) were estimated directly from non-zero grid cell measurements, consistent with typical field measurement techniques. Computation grids with average canopy heights greater than 15 cm were considered erroneous and removed from analysis (based on in-situ observations), resulting in less than 2% total data loss.

The rugosity (R), or tortuosity index, for each computational grid was estimated as the ratio between the surface area of the canopy (i.e. three-dimensional canopy-top) and the area of its orthogonal projection onto the bed plane (i.e. flat reef surface: 0.25 m²). Rugosity is a common metric used for parametrizing biological roughness, especially in rigid canopies like coral reefs (Shepard et al. 2001; Leon et al. 2015). In the current study, the canopy surface area was estimated using a three-dimensional surface linearly interpolated between each non-zero canopy element in the 0.5 x 0.5m grid (Figure 2b). The rugosity was defined as the total area of this irregular surface

divided by the area of the bed below the canopy (0.25 m^2). This methodology retained high spatial resolution while avoiding unrealistically sharp surface spikes associated with small interstitial areas in the canopy. The resulting surface is analogous to a sheet draped over the oyster canopy, and it is functionally similar to linear rugosity estimates, which rely on chain measurements with rigid links (length: 1 – 100 cm) which effectively filter out small-scale roughness elements (e.g. Knudby & LeDrew 2007).

The fractal dimension (D) of the oyster canopy was estimated from canopy height data using the variation method developed by Dubuc et al. (1989). The fractal dimension describes the complexity of an irregular surface, and it is strongly correlated with human visual perception of roughness (Pentland 1984). For three-dimensional surfaces, the fractal dimension ranges from 2 – 3, with higher values representing more complex surfaces. In this study, the fractal dimension was estimated following the methods described in Zhou & Lam (2005) and Zawada & Brock (2009). In short, a variable length window was defined for each computational grid (50 cm x 50 cm), with a side length $L=2\epsilon+1$ and $1 \text{ cm} \leq \epsilon \leq 12 \text{ cm}$. For each side length (3 – 25 cm), the window was positioned at all possible locations within the computational grid and the difference between the two most extreme within-window canopy height measurements (including zero-elements) was recorded and averaged to compute the mean variation $V(\epsilon)$. Finally, the best-fit slope (m) of the regression between $[V(\epsilon)]$ and $[\epsilon]$ was used to estimate the fractal dimension, defined as $D=3-m$. All computation grids with best-fit $R^2 < 0.85$ were removed from analysis, resulting in approximately 1% data loss.

Probability density functions (PDF) for all roughness variables (canopy height, canopy height standard deviation, rugosity, and fractal dimension) were estimated using a kernel density estimator, which provides a continuous, nonparametric estimate of the PDF without a priori assumptions about the distribution of the data (Bowman and Azzalini, 1997). PDFs were computed in Matlab using the *ksdensity* function with default bandwidth settings.

Results

Manually observed oyster characteristics varied across sample reefs, with live oyster densities and shell lengths that tended to increase with restoration age (Table 1). For restored reefs, live oyster densities (mean \pm SE) ranged from 208 ± 7 oysters/ m^2 (R-2016) to 475 ± 41 oysters/ m^2 (R-2014). Although average oyster densities were lower on R-2016 (208 oysters/ m^2) than on R-2017 (250

oysters/m²), shell lengths were over 30% larger on the older reef (50.7 ± 1.2 mm vs. 37.6 ± 0.9 mm), reflecting the growth and replacement of first-year oyster recruits. The intact reference reef was characterized by lower live oyster densities (184 ± 34 oysters/m²) and shorter shell lengths (40.5 ± 10 mm) than the oldest restored reef (R-2014). However, the number of rigid canopy elements (168 elements/m²), which includes both live oyster and non-living reef structure, was nearly twice as large on the reference reef than on restored reef surfaces (84 - 96 elements/m²).

Canopy heights (h_c) ranged from 3 – 10 cm, and reef-wide averages generally increased with restoration age (Table 1; Figure 3a-d; Figure 4a). Canopy heights at the youngest restored reef (R-2017) were smaller (mean \pm 95% CI here and after: 3.6 ± 0.1 cm) than those measured at all other study sites, with shorter canopies presumably linked to the younger (~1 year), smaller oysters inhabiting the reef surface and the lack of accumulation of new oysters over multiple recruitment years. Accordingly, the tallest oyster canopies were observed at R-2014 (h_c : 4.9 ± 0.1 cm), where mean canopy heights in individual computation grids often exceeded 7 cm. In general, observed canopy heights were largest at the reef margins and decreased towards the center of the reef (Figure 5a). This trend was especially evident at R-2016, where spatial heterogeneity created a bimodal canopy height distribution in the probability density plot (Figure 4a). Importantly, manual measurements of canopy heights on each reef (Figure 3a) generally fell within 0.5 cm of local laser-scan derived estimates (collected within 1m of quadrat), providing confidence in applied data-processing techniques.

Canopy height standard deviations (σ_c) followed trends observed in the mean canopy height (Table 1; Figure 3e-h), and larger standard deviations were observed on older reefs with taller canopies and higher densities of mature oyster. Reef-wide standard deviation averages ranged from 1.4 ± 0.1 cm at R-2017 to 2.0 ± 0.1 cm at R-2014. Following initial recruitment (R-2017), standard deviations increased on the reef margins (R-2016), widening the range of the distribution (Figure 4b; Figure 5b). As the reef continued to mature, additional oysters were recruited to the structure and distributions shifted to higher means (R-2014: 2.0 ± 0.1 cm). Canopy height standard deviations on the oldest restored reef (R-2014) were higher than those observed on the reference reef (Reference: 1.7 ± 0.1 cm), where estimates at the reef margins (1.9 ± 0.1 cm) were nearly 20% greater than those observed in the reef interior (1.6 ± 0.1 cm; Figure 5b).

Estimates of the rugosity index (R) ranged from 1.0 – 3.0, with an average rugosity of $R = 1.37 \pm 0.01$ measured across all study reefs (Table 1; Figure 3j-m; Figure 4c). As with other roughness parameters, reef surface rugosity was linked to restoration age, increasing from 1.28 ± 0.08 at 1-year post-restoration (R-2017) to 1.56 ± 0.01 at 4-years post-restoration (R-2014). Mean rugosity estimates at R-2014 were higher than those measured at any other sample location, including the intact reference reef, where $R=1.31 \pm 0.01$. Rugosity indices were spatially heterogeneous on the reference reef, with reef margins that were significantly rougher than the reef interior (Figure 5c) ($R=1.39 \pm 0.02$ vs. $R=1.28 \pm 0.01$).

Fractal dimension (D) estimates were consistent across study reefs, with reef-wide means between 2.67 and 2.74 (Table 1; Figure 3n-q, Figure 4d). The spatial distribution of fractal dimension (Figure 3n-q; Figure 5d) consistently highlighted complex reef margins, with less complexity in the reef interiors. The lowest fractal dimensions were observed in the interiors of the youngest study reefs (R-2017, R-2016), where fractal dimensions fell below 2.5 for ~5% of all computation cells. Probability density distributions (Figure 4d) narrowed with time since restoration, with a wider range of D observed on younger reefs. The fractal dimension was highest on the reference reef ($D=2.74 \pm 0.01$), despite live oyster densities and canopy heights that were generally lower than those observed at R-2014.

Discussion

Roughness characteristics of intertidal oyster reef

Reef-scale roughness characteristics computed from the laser scan point clouds were similar to those described based on manual measurements of other natural marine canopies, including coral reefs (e.g. Knudky & LeDrew 2007; Leon et al. 2015; Miller et al. 2021), mussel beds (e.g. Commito & Rusignuolo 2000; Shynn Lim et al. 2020), and oyster reefs (e.g. Margiotta et al. 2016; Karp et al. 2018). Although direct comparisons are complicated by variable measurement techniques, which can have significant effects on inferred surface complexity (e.g. Yanovski et al. 2017; Knudly & LeDrew 2007), it is still useful to place the results of this study in the broader context of marine canopy literature. For example, rugosity indices on study reefs (range: 1.0 – 2.2) agreed well with those reported by Karp et al. (2018), Margiotta et al. (2015), and Rodney & Paynter (2006), who measured linear rugosity indices between 1.2 and 3.0 on restored and relic oyster reefs. On the other hand, the intertidal oyster reefs investigated in the current study were

markedly different than corals reefs, with prior studies reporting higher rugosity indices (Knudsky & LeDrew 2007: 1 - 2.25; Burns et al. 2015: 1.5; Yanovski et al. 2017: 2.22; Carlot et al. 2020: 2 - 3.75) and lower fractal dimensions (Zawada et al. 2010: 2 - 2.5; Leon et al. 2015: 2.2 – 2.6; Miller et al. 2021: 2 – 2.5) for corals in comparison to oysters (\bar{R} =1.2 – 1.5; \bar{D} = 2.67 – 2.74). Differences in roughness characteristics between corals and oysters may be due, at least in part, to differences in reef growth and development. Intertidal oyster reefs develop through gregarious settlement of oyster larvae and individual oyster growth, creating complex, intertwining canopy structures composed of multiple age and size classes. Conversely, coral reefs predominantly develop through clonal colony growth, with many species producing large, relatively sparse canopy elements. These larger canopy elements are likely responsible for the higher rugosity indices reported for coral reefs, since the metric is defined based on *relative* changes in the canopy height compared to surrounding canopy elements. Higher fractal dimensions in oyster reefs may reflect the more random development process (i.e. larval attachment and growth), which produces characteristically irregular shells, sharp edges, and branching arms throughout the canopy. Additional studies focused on parallel measurements of oyster and coral reefs using identical methodologies may provide additional insight for differences in structural complexity between both marine canopies.

Temporal evolution of structural complexity on restored oyster reefs

In the current study, differences in canopy structure were well correlated with time since restoration, and older, more mature reefs were characterized by more complex surface structures. The observations presented here suggest that continued oyster recruitment and growth may increase surface complexity over several years post-restoration. All restored reefs started with no living oysters, and spat recruitment during the first season led to nearly immediate increases in live oyster density, canopy height, and canopy complexity (i.e. σ_c , R, D), as seen in R-2017. Following initial recruitment, living oysters continued to grow (i.e. increasing shell lengths) and the canopy developed distinct spatial patterns, with taller, more complex oyster clusters at the leading edge of the reef driving bimodal distributions in canopy height and canopy height standard deviation. The consistent patterns in spatial heterogeneity suggest that oyster spat may preferentially settle on the reef margins, at least during the early phases of reef development when growth is accelerated (i.e. Ridge et al. 2015). However, as similar patterns were also evident in the reference reef, spatial distributions may be linked to other ecological processes, including increased food availability (i.e.

nutrient concentration drawdown caused by filter feeding) and/or differential predation pressures. Following the first year of reef development, additional oyster growth and recruitment created increasingly complex canopy structures across the entire reef surface. The distributions of roughness characteristics narrowed and shifted to higher means, mirroring those observed on the intact reference reef within four years post-restoration. Ecosystem services moderated by canopy complexity, including habitat provision and energy (i.e. wave and current) attenuation, are expected to evolve simultaneously, as described in Cannon et al. (2022a,b). It is important to note that variability in surface complexity may also be linked, at least partially, to reef identity, with individual reef characteristics, including location, size, and sediment biogeochemistry, potentially playing a role in reef-to-reef canopy structure variability. While the study design (i.e. space-for-time) makes it difficult to distinguish between variability linked to restoration age and reef identity, the shared geological history and close proximity of the sample sites (i.e. within 3.5km) suggest that restoration age likely plays a larger role in modulating canopy evolution than variability in environmental factors.

The oldest restored reef investigated in this study (R-2014) matched or exceeded many surface roughness characteristics observed on the reference intact reef, suggesting that intertidal oyster reef restoration can be an effective method of recreating natural coastal infrastructure. The mean canopy height (4.9 ± 0.1 vs. 4.1 ± 0.1 cm), canopy height standard deviation (2.0 ± 0.1 vs. 1.7 ± 0.1 cm), and rugosity index (1.56 ± 0.01 vs. 1.31 ± 0.01) were all greater at the restored reef than at the reference reef. These differences in canopy characteristics mirror observed differences in (manually measured) live oyster density and shell length, suggesting that the restored reef also supported a larger, more abundant oyster population per unit area. However, the slightly elevated mean fractal dimension of the reference reef (2.74 ± 0.01 vs. 2.71 ± 0.01) indicates the prevalence of additional sources of three-dimensional complexity that are not captured by the canopy height and rugosity indices. Persistent historic colonization at the reference site has created a more fundamentally diverse canopy of live oysters and nonliving oyster shell structures, with complex branching surface structures built over centuries (or millenia) of oyster life cycles (recruitment, growth, decay) since the development of the barrier island ecosystem (~7000 years ago; Brooks 1972). For restored reefs, reaching a similar level of complexity, as measured by the fractal dimension, would likely take decades. The history of the reference reef also reflects human pressures. While the restored reefs are posted and protected from harvesting, fishermen can freely

harvest live oysters on the reference reef, where they generally target large, mature oysters over younger, smaller oysters, effectively reducing the live oyster density and shifting the distribution towards younger recruits. It is also possible that harvesting may have a direct impact on the structural complexity of the reef surface, with shifts in the size class distributions producing more (or less) complex oyster canopies over time. Although beyond the scope of the current work, additional research could help determine “optimal” harvest regulations for meeting and maintaining restored oyster reef complexity goals.

Applications to natural and nature-based feature design

Conventional infrastructure approaches alone are unlikely to effectively manage the hazards of flooding and erosion that threaten many coastal communities (Morris et al. 2020; Spalding et al. 2014). Engineered structures often disrupt sediment transport and accretion processes in coastal systems, exacerbating rather than ameliorating the effects of sea level transgression and more frequent strong storms (Temmerman et al. 2013). Adaptation plans that integrate natural features, such as oyster reefs, into multi-layered adaptation approaches are an attractive alternative to stakeholders given both their low cost and extensive provision of inter-related ecosystem services. Beyond incorporating living materials into designs, nature-based infrastructures depend on these organisms for performance-related processes, such as wave attenuation, sediment and carbon retention or self-repair. Investments to restore or create oyster reef, for example, may be selected to influence local hydrodynamic patterns, filter water and sequester carbon/nutrients, or create habitat complexity (Grabowski et al. 2012), with the expectation that the reef crest will self-adjust in response to disturbance and environmental changes. Each of these desired processes are related to specific attributes of the organism community. The ultimate hydrodynamic influence of a reef will be determined from the structural form of the reef and oyster canopy relative to the flow environment (e.g., Cannon et al. 2022b). The reef and canopy form in turn are shaped by a combination of material properties and ecological processes, including habitat tolerances and life history traits of oysters, as well as selective pressures of competition and predation.

The restored reefs featured in this study were notably successful from the perspective of recruiting oyster larvae, retaining spat, and promoting survival as evidenced by the healthy growth in oyster density, canopy height and complexity over time since restoration and the multi-generational size classes represented by varied shell lengths. The structural complexity of the oyster reef canopy

observed in this study relates directly to alteration of the near-reef flow field and to larval settlement and survival. For example, Cannon et al. (2022b) found that even as mean velocities within oyster canopy remained low, turbulent dissipation within oyster canopy increased many times over during a single year of oyster recruitment to a newly-restored reef. Drag coefficients estimated within complex reference-condition oyster canopy were two times greater than those observed on the surface of a degraded reef with no live canopy (Kitsikoudis et al. 2020) and an order of magnitude greater than those reported over bare sand and fine sediments (Styles 2015; Whitman & Reidenbach 2012). Notably, drag coefficients reported within oyster canopy also exceed those observed in shear layers that develop above the top of the canopy (Styles 2015), where Reynolds stresses peak and velocity profiles become logarithmic (Whitman & Reidenbach 2012).

It is likely that the differing hydrodynamic conditions observed within and above the oyster canopy work in tandem to attract and promote settlement of oyster larvae and then to retain and allow growth of spat. For example, larval *C. virginica* have been observed in lab studies to expend energy to propel themselves downward (e.g. dive) with greater frequency and force as turbulent dissipation rates increase (Fuchs et al. 2013; Fuchs et al. 2017). Turbulence created by the canopy may also signal larvae to swim to the bed and enhance the probability of settlement on established canopy structures (Fuchs & Reidenbach 2013). Additionally, high retention of spat settled within the relatively small spaces between shell or tile substrates (Whitman & Reidenbach 2012; Nestlerode et al. 2007; Lavan 2019) highlight the importance of interstitial niches of particular size for retention and survival. Whether the canopy niche size provides protection from predation or other physical habitat functions is unknown. Artificial reef created from materials that lack the complex 3-dimensional structure of natural reef canopy may fail to either recruit or retain larvae, and restoration design could include efforts to reproduce structural characteristics of natural, healthy reefs, as described in this work, to boost early colonization.

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Tables

Table 1: Canopy characteristics as measured on restored (R-2017, R-2016, R-2014) and reference reefs. Measurements include reef area (m^2), live oyster density (oysters/ m^2), live oyster shell length (cm), canopy height (cm), canopy standard deviation (cm), rugosity index, and fractal dimension. Mean and [median] canopy characteristics ($\pm 95\%$ CI) were estimated over all grid cells (0.25 m^2) in a given reef area, while average ($\pm \text{SE}$) live oyster densities and shell lengths were estimated from manual oyster counts on each reef. Measurements in curly brackets were manually measured using a single quadrat on each reef.

Reef Name	Reef Area m^2	Live Oyster Density oysters/ m^2	Live Oyster Shell Length Mm	Canopy Height h_c ; cm	Canopy Standard Deviation σ_c ; cm	Rugosity Index R; unitless	Fractal Dimension D; unitless
R-2017	144	250 ± 13 {84 el/ m^2 }	37.6 ± 0.9	3.6 ± 0.1 [3.5 \pm 0.1]	1.4 ± 0.1 [1.3 \pm 0.1]	1.28 ± 0.03 [1.22 \pm 0.02]	2.67 ± 0.01 [2.70 \pm 0.01]
R-2016	326	208 ± 7 {96 el/ m^2 }	50.7 ± 1.2	4.0 ± 0.1 [4.2 \pm 0.1]	1.5 ± 0.1 [1.4 \pm 0.1]	1.32 ± 0.02 [1.27 \pm 0.02]	2.69 ± 0.01 [2.70 \pm 0.01]
R-2014	282	475 ± 41 {88 el/ m^2 }	58.8 ± 1.4	4.9 ± 0.1 [4.9 \pm 0.1]	2.0 ± 0.1 [2.0 \pm 0.1]	1.56 ± 0.01 [1.54 \pm 0.01]	2.71 ± 0.01 [2.72 \pm 0.01]
Reference	446	184 ± 34 {168 el/ m^2 }	40.5 ± 1.0	4.1 ± 0.1 [4.0 \pm 0.1]	1.7 ± 0.1 [1.6 \pm 0.1]	1.31 ± 0.01 [1.28 \pm 0.01]	2.74 ± 0.01 [2.74 \pm 0.01]

Figure Captions

Figure 1: (a) Map of study area with oyster reef locations displayed as colored squares (Reference: green; R-2014: black; R-2016: magenta; R-2017: blue). Inset diagram shows the study area location within Mosquito Lagoon, Florida, USA. Inset pictures highlight (b) typical canopy element, or oyster cluster, composed of dead oyster shells and living oyster (c) intertidal oyster reef and in-situ laser scan set-up, and (d) an example of raw point-cloud data collected on the reef surface.

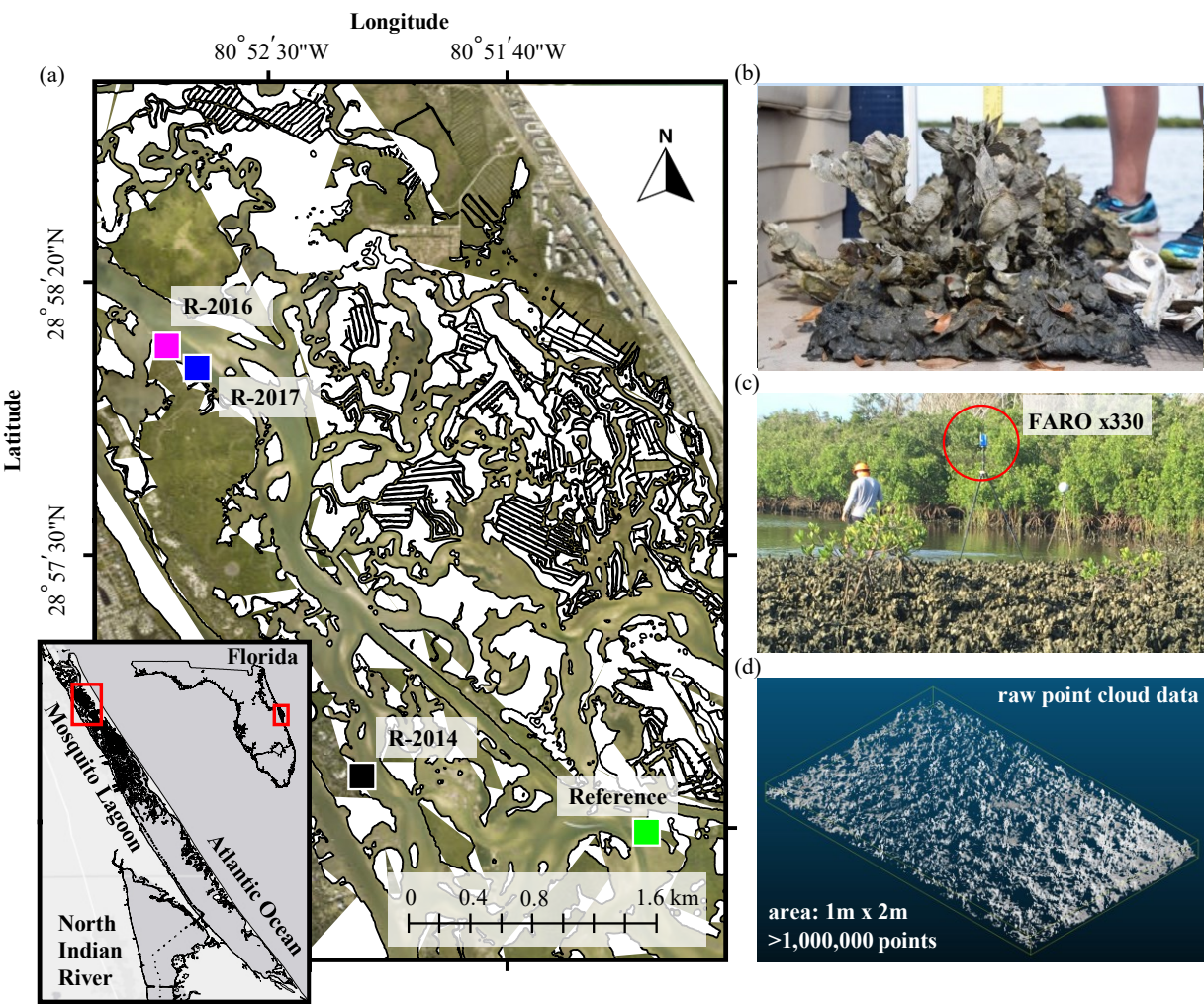
Figure 2: An example of processing steps for laser-scan analysis. (a) After sub-sampling raw elevation data (sample area: 50 cm x 50 cm), a plane (red plane) was fit to the lowest 25% of points (red points) selected from 16 equally-distributed grid cells (12.5 cm x 12.5 cm) and shifted to the lowest elevation measured in the sub-sample (blue plane). (b) All points were converted to elevation above plane (Z) and averaged over 1 cm x 1 cm grid cells, with the resulting canopy heights used to fit a 3D canopy surface. The elevation of grid cells without sample points was set equal to zero.

Figure 3: Spatial maps for reef structural complexity parameters as estimated from high-resolution surface scans for restored (R-2017, R-2016, R-2014) and reference reefs. Parameters include the mean canopy height (a-d), the standard deviation of the canopy height (e-h), the rugosity index (j-m), and the fractal dimension (n-q). Reefs are drawn to scale, and horizontal and vertical tick marks represent 10m intervals. Red boxes in (a) represent approximate locations for manual canopy height measurement quadrats. Manually measured (*m*) and laser-scan (*ls*) derived canopy heights are included for reference, with *ls* indicating the range of canopy heights estimated within 1m of approximate quadrat locations.

Figure 4: Probability density plots comparing structural complexity parameters for restored (R-2017: blue, R-2016: magenta, R-2014: black) and reference (green) reefs. Parameters include the mean canopy height (a), the standard deviation of the canopy height (b), the rugosity index (c), and the fractal dimension (d). Vertical lines are used to show the mean of each distribution (values given in Table 1).

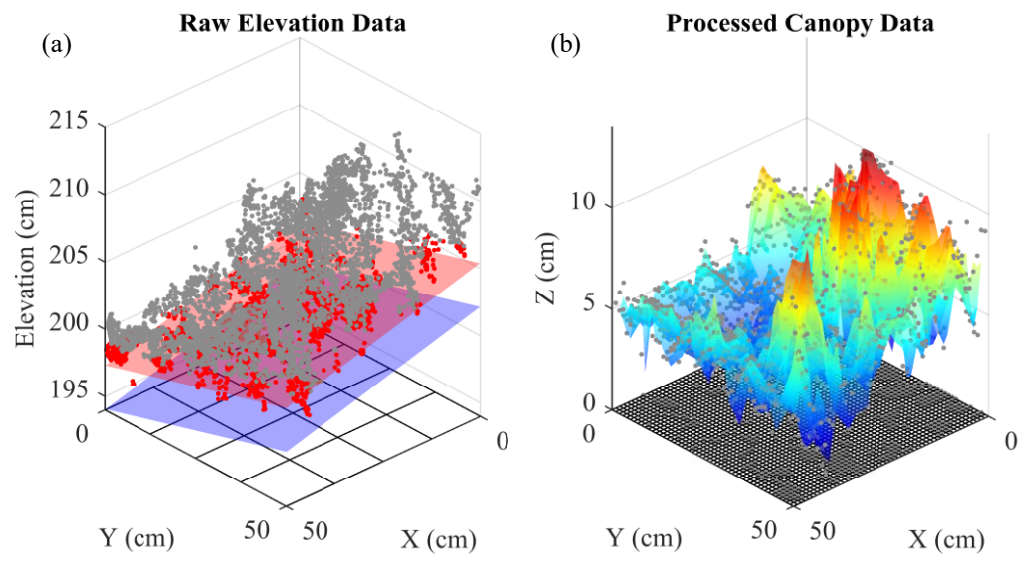
Figure 5: Distributions of observed canopy heights (a), canopy height standard deviations (b), rugosity indices (c), and fractal dimensions (d) within 2 m of the edge (red) and in the center (blue) of each reef (R-2017, R-2016, R-2014, Reference). Notches in boxplots represent 95% confidence intervals on sample medians.

676 **Figures**



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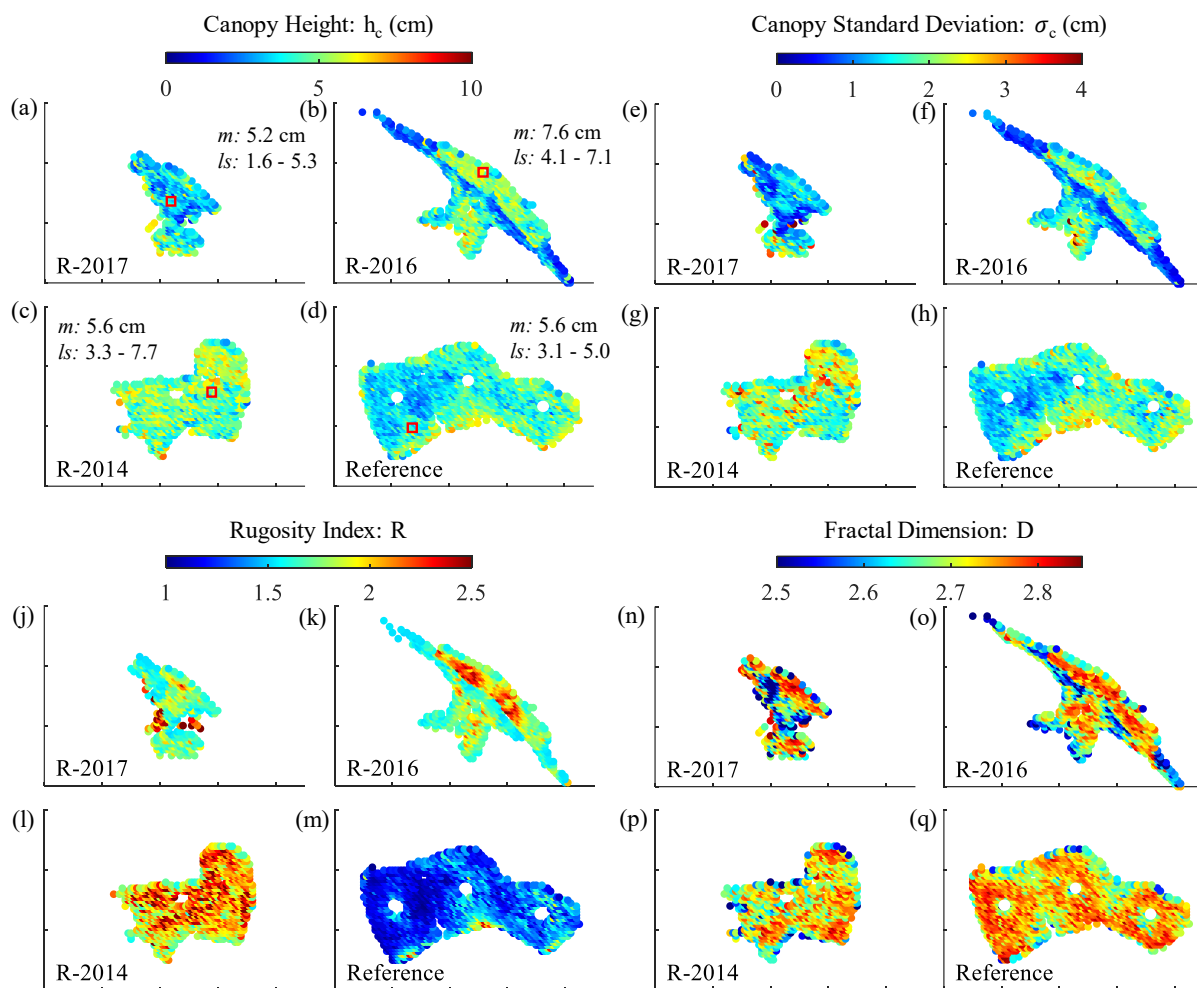
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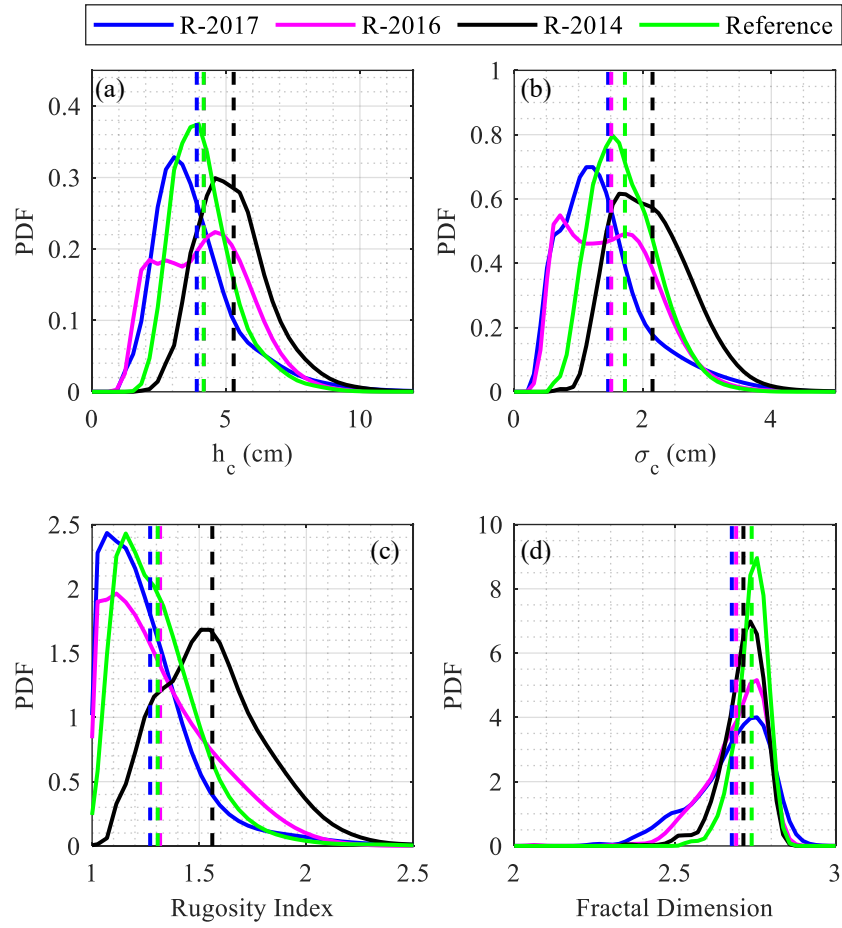


Figure 4.

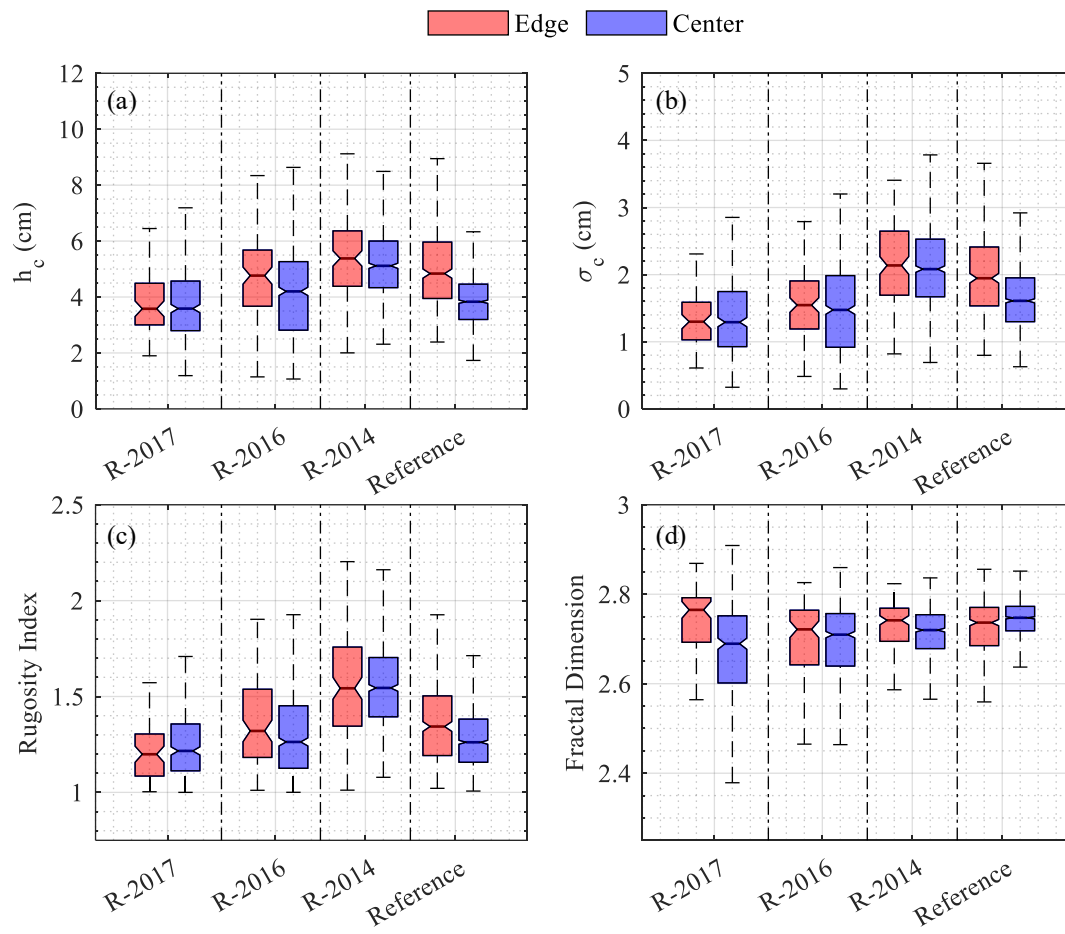


Figure 5.