

# One-Step Regio- and Stereoselective Electrochemical Synthesis of Orexin Receptor Antagonist Oxidative Metabolites

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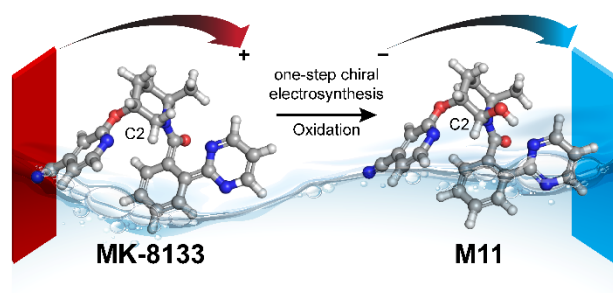
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## Abstract

Synthesis of drug metabolites, which often have complex structures, is an integral step in the evaluation of drug candidate metabolism, pharmacokinetic properties, and safety profiles. Frequently, such synthetic endeavors entail arduous, multiple-step *de novo* synthetic routes. Herein we present the one-step Shono-type electrochemical synthesis of milligrams of chiral  $\alpha$ -hydroxyl amide metabolites of two orexin receptor antagonists, MK-8133 and MK-6096, as revealed by a small-scale (pico- to nano-mole level) reaction screening using a lab-built online electrochemistry/mass spectrometry (EC/MS) platform. The electrochemical oxidation of MK-8133 and MK-6096 was conducted in aqueous media and found to produce the corresponding  $\alpha$ -piperidinols with exclusive regio- and stereoselectivity, as confirmed by high resolution NMR characterization of products. Based on DFT calculations, the exceptional regio- and stereoselectivity for this electrochemical oxidation are governed by more favorable energetics of the transition state leading to the preferred secondary carbon radical  $\alpha$  to the amide group and subsequent steric hindrance associated with the U-shaped conformation of the cation derived from the secondary  $\alpha$ -carbon radical, respectively.

**Keywords:** electrochemical synthesis; chiral synthesis; drug metabolites; mass spectrometry; NMR characterization; DFT.

## Introduction

Insomnia is a common disorder affecting 10–15% of the population and is often associated with other physical and psychological problems.<sup>1</sup> Following the discovery of orexin, the novel therapies of dual OX1R and OX2R orexin receptor antagonists (DORAs) and selective orexin receptor antagonists (SORAs) have advanced into late stage clinical development. Examples of such therapies are MK-8133 and MK-6096 (see Scheme 1).<sup>2-7</sup> These drugs incorporate a piperidine core in their structures. Piperidine moieties are an integral feature of many alkaloid structures and drug development studies have led to the preparation of many highly efficient drugs containing substituted piperidine and piperazine structural moieties.<sup>2</sup> During a preclinical safety study, a major circulating metabolite M11 from MK-8133 (Scheme 1) was detected, isolated from pooled rat plasma, and characterized by mass spectrometry and NMR spectroscopy. Metabolic studies are an integral component of drug candidate pharmacokinetic (PK) and safety profiling in the drug development process.<sup>3-7</sup> To further understand biological activity of the M11 metabolite, in addition to the potential for interaction with other drugs, milligram quantities of the metabolite were needed. Historically, the required milligram quantities of metabolites for testing was fulfilled using classical organic synthetic procedures, which can be a demanding and lengthy process, necessitating a significant and continuing resource commitment.<sup>8,9</sup> Due to the structural complexity of the MK-8133, coupled with enantiomeric purity requirements, it was estimated that at a minimum, a four-step linear

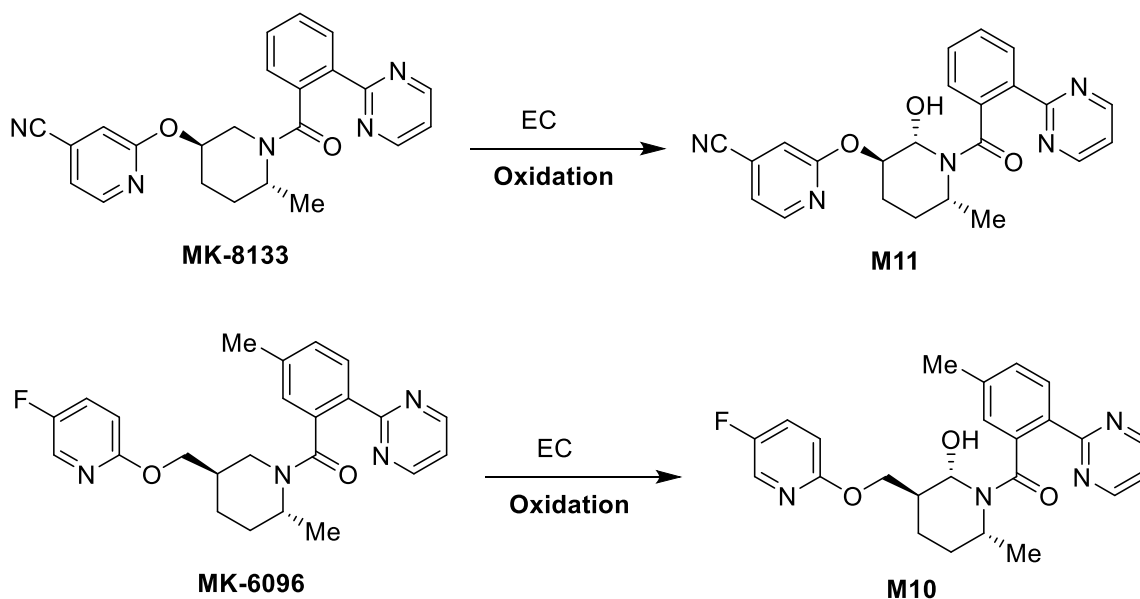
synthesis (e.g., the synthesis of metabolite of suvorexant required a four-step linear synthesis<sup>10</sup>) would be needed to provide the requisite quantities of the M11 metabolite.

Electrochemical synthetic procedures afford a powerful and convenient method for the oxidative modification of complex organic molecules and have garnered considerable interest.<sup>11-24</sup> Much milder oxidants and reductants can be employed in electrochemical redox reactions, with protons from water serving as the oxidant in many cases, and toxic transition metal catalysts can often be avoided entirely. Electrochemical reactions produce little to no waste and comply with the tenets of “green chemistry.”<sup>25</sup> In addition, electrochemical redox reactions occur at ambient temperature and pressure, avoiding the inherent sensitivity issues associated with many synthetic organic transformations.<sup>25</sup> In concert, these attributes make electrochemical synthesis highly attractive and numerous remarkable transformations *via* electrochemical reactions have been reported.<sup>26-35</sup> For instance, Shono oxidations<sup>36-38</sup> are among the most widely employed electrochemical methods for the functionalization of C-H bonds adjacent to nitrogen atoms. In Shono oxidation, amides or carbamates in alcohol solvents are electrochemically oxidized to give the corresponding *N*- or *O*-acetals. However, such oxidations simultaneously affording both regio- and stereoselective electrochemical synthesis have not been reported. We reasoned that M11 could be prepared from its parent molecule MK-8133 via direct Shono-type electro-oxidation, considering that MK-8133 has an amide core structure.

The combination of electrochemistry (EC) with mass spectrometry (MS), EC/MS, can be versatile, for mimicking drug metabolism, cleaving proteins/peptides, reducing protein disulfide bonds, facilitating protein/peptide quantitation, and determining the location of unsaturated double bonds in lipids.<sup>39-44</sup> The method can also be used to observe

elusive reaction intermediates<sup>45-54</sup> and for electrosynthetic reaction screening.<sup>55-57</sup> Significant progress has been made recently in the application of EC/MS to mimic drug metabolism, especially in the case of oxidative metabolites whose formation is catalyzed by cytochrome P450.<sup>58-61</sup> For example, reactions such as aliphatic or aromatic hydroxylation, *N*-dealkylation or epoxidation can be mimicked using an online EC/MS platform.<sup>62-68</sup>

In this study, we describe a clean, one-step Shono-type electrochemical synthesis of the  $\alpha$ -piperidinol amide metabolites of MK-8133 and MK-6096 (Scheme 1), which otherwise would involve a multiple step *de novo* synthesis. Optimal conditions for the desired metabolite formation were determined via screening using online EC/MS using minimal amounts of substrates (pico- to nano-mole level). Once established, the optimal conditions were employed with a large electrochemical cell to prepare milligram quantities of the hydroxylated metabolites. Strikingly, the oxidation was observed to be exclusively regio- and stereoselective as determined by high resolution NMR. Although the electrolytic yield varied from a moderate 40-50% for small scale electrolysis in thin-layer electrochemical flow cell to as low as 14-17% for larger scale electrolysis in the SynthesisCell<sup>TM</sup>; nevertheless, milligram quantities of desired metabolites (M11 and M10, respectively, Scheme 1) were successfully obtained in this one-step electrochemical procedure. Following the characterization of the metabolite structures and the confirmation of the stereochemical orientation of electrochemically introduced 2-hydroxyl substituent, computational work was next undertaken to better understand the unique regio- and stereoselectivity observed for the oxidation of MK-8133 and MK-6096.

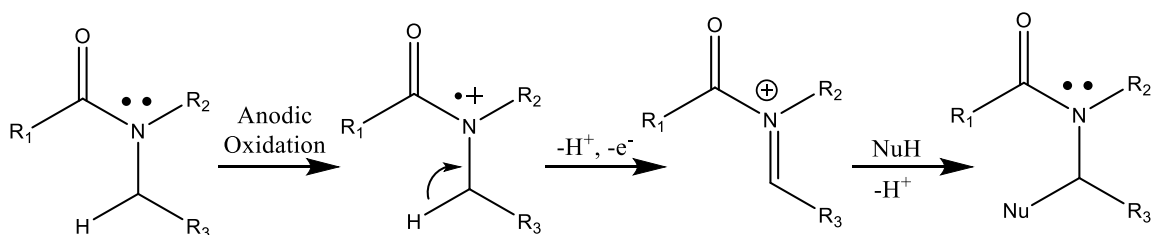


**Scheme 1.** One-step electrochemical oxidation of MK-8133 and MK-6096 to produce hydroxylated metabolites, M11 and M10, respectively.

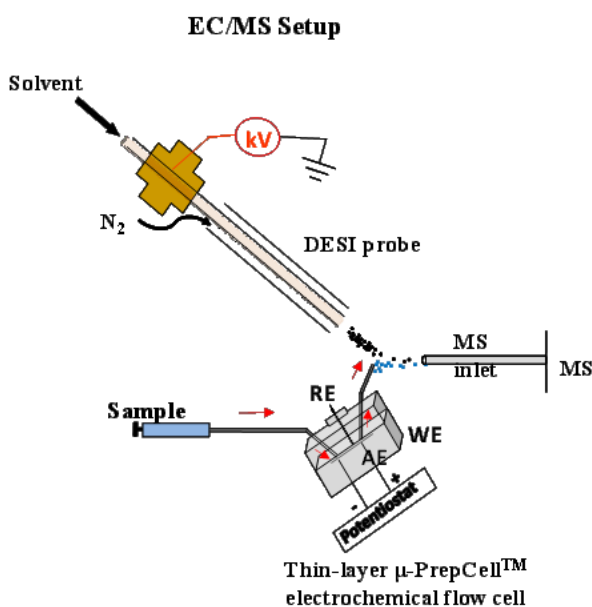
## Results and Discussion

### *Direct electrochemical oxidation of MK-8133*

Electrochemistry has been increasingly employed in recent years to mimic oxidative metabolism typically encountered during phase I clinical trials of drug candidates, especially those oxidations catalyzed by cytochrome P450.<sup>69-72</sup> Anodic oxidation of amides can generate *N*-acyliminium ions, which can subsequently undergo elimination, or conversely, these ions can be “trapped” by nucleophiles, functionalizing the  $\alpha$ -position of the amide.<sup>73-75</sup> Analogously, if the anodic oxidation is conducted in aqueous media, it is possible to form the  $\alpha$ -hydroxyl amide product ( $N_u=OH^+$ , Scheme 2). In this study, we envisioned that electrochemistry might be a highly attractive alternative to synthesize the desired  $\alpha$ -hydroxyl metabolites like M11 and M10.



**Scheme 2.** Schematic representation of the anodic electrochemical oxidation of an amide where NuH = nucleophile; e.g., water in this study.

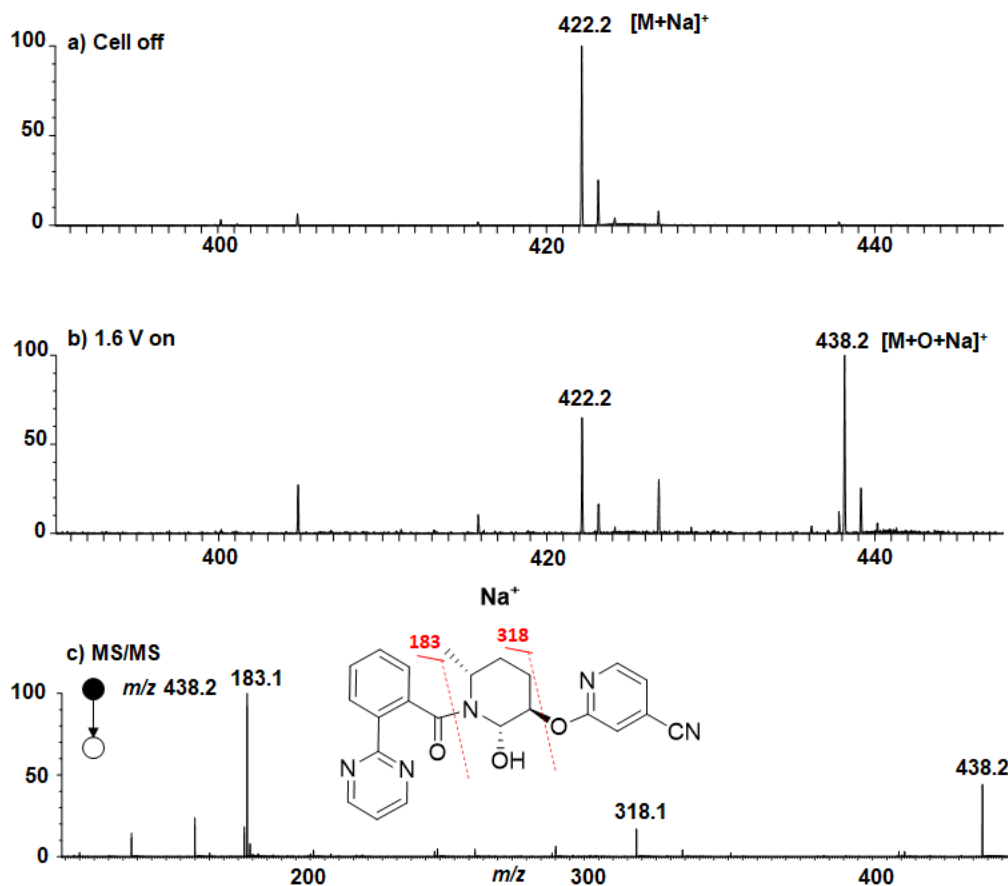


**Scheme 3.** EC/MS apparatus for online electrosynthesis monitoring/screening, and the optimization of the electrosynthesis parameters such as the initial concentration, solvents, applied potential, electrolysis time, pH and electrode materials.

We first tested our hypothesis with the anodic oxidation of MK-8133 using an online EC/MS setup (Scheme 3). EC/MS, particularly with ESI or DESI as the ionization interface, is a very powerful tool to quickly screen different electrochemical experimental parameters (e.g., applied potential, sample flow rate and pH variation), since the electrochemical oxidation or reduction reaction is monitored online with MS.<sup>76-78</sup>

Oxidation of MK-8133 at a low concentration (100  $\mu$ M) in water containing 10 mM  $\text{NH}_4\text{OAc}$  was first conducted using the EC/MS apparatus (Scheme 3) (Data is shown in Figure S1; Supporting Information). With no potential applied to the cell, only protonated MK-8133  $[(\text{M}+\text{H})^+]$  was detected at  $m/z$  400 (Figure S1-a, Supporting Information). When a positive potential of 1.6 V was applied to the cell, a small peak corresponding to the oxidation product  $[\text{M}+\text{O}]^+$  was detected at  $m/z$  416 (Figure S1-b, Supporting Information). 1.6 V was the optimized potential. This value is close to the reported amide oxidation potentials varying from 1.47 to 2.21 V.<sup>79</sup> However, when 20 mM  $\text{Na}_2\text{HPO}_4/\text{NaH}_2\text{PO}_4$  (pH=7.0) in  $\text{H}_2\text{O}$  was used, the oxidation yield was enhanced, likely attributed to the strong electrolytic character of the phosphate buffer, as shown in Figure 1. With no potential applied to the cell, only the sodium adduct ion of MK-8133 was detected at  $m/z$  422 (Figure 1a). When a positive potential of 1.6 V was applied to the cell, the intensity of the  $m/z$  422 ion decreased and another ion was observed at  $m/z$  438 (Figure 1b), corresponding to the sodium adduct ion of M11. Upon collision induced dissociation (CID), the  $m/z$  438 ion primarily yielded fragment ions at  $m/z$  318 and 183, consistent with oxidation of piperidine fragment, as was anticipated (Figure 1c). The in-spectrum oxidation yield was estimated to be >50%, as the relative intensity of  $m/z$  438 was higher than the remaining  $m/z$  422. Note that the test of electrochemical oxidation of MK-8133 using our online EC/MS setup in this study was not only fast and took just few minutes, it also only consumed a minimal amount of sample (pico- to nano-moles of substrate).





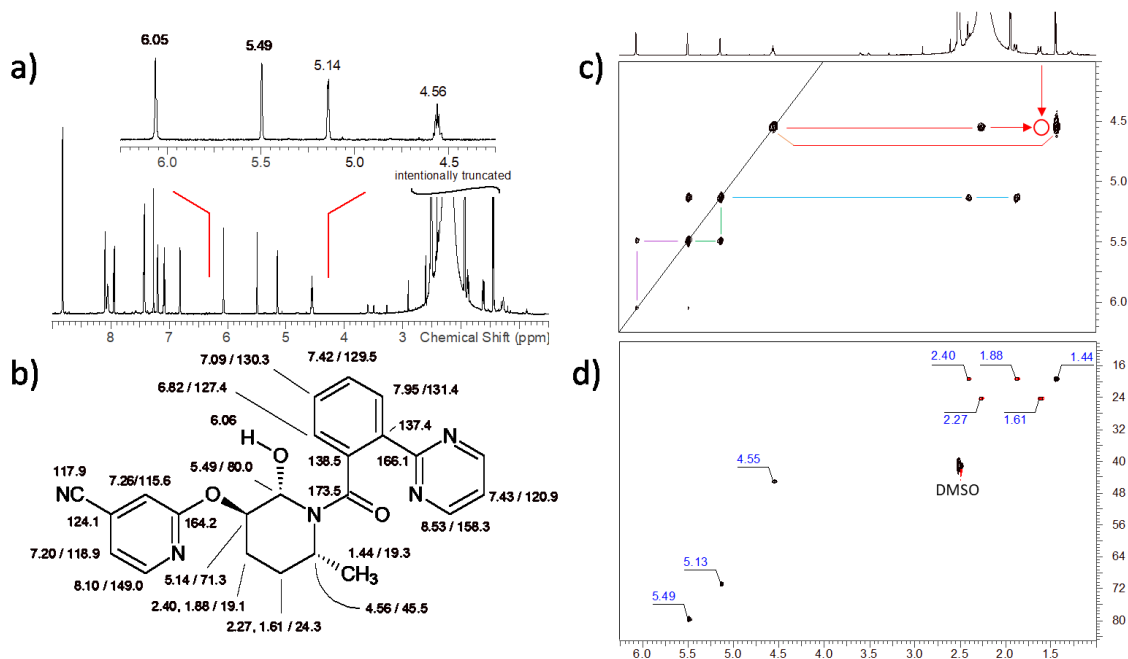
**Figure 1.** DESI-MS spectra acquired when a solution of 100  $\mu\text{M}$  MK-8133 in water containing 20 mM  $\text{Na}_2\text{HPO}_4/\text{NaH}_2\text{PO}_4$  (pH=7.0) was flowed through the thin-layer electrochemical cell a) without and b) with an applied 1.6 V; c) CID MS/MS spectrum of  $m/z$  438.

It is also possible to isolate the electrochemically generated metabolite, M11. In an initial experiment, a higher concentration of MK-8133 (500  $\mu\text{M}$ ) in acetonitrile: aqueous 20 mM sodium phosphate buffer (10:90; v/v) was continuously infused to the thin-layer  $\mu\text{-PrepCell}^{\text{TM}}$  (a flow cell, applied potential: 1.8 V) at a flow rate of 30  $\mu\text{L}/\text{min}$ . The oxidation solution collected was recycled through the electrochemical cell for a second time to further enhance the oxidation conversion yield, which was found to approximately 40% by the LC/UV measurement (based on the ratio of the product UV absorption peak area at 254 nm vs. the total of UV absorption peak areas of the remaining reactant and the product at

254 nm, Figure S2-a, Supporting Information). A subsequent scale-up reaction for producing M11 by anodic oxidation of MK-8133 using the SynthesisCell™, a batch reactor cell with a large boron doped diamond (BDD) working electrode (3cm x 3cm) was conducted, employing the optimized conditions obtained from the online EC/MS screening. In the first attempt, 80 mL of 500  $\mu$ M MK-8133 (16 mg starting material) was prepared in acetonitrile: aqueous sodium phosphate buffer (10:90; v/v) and added into the SynthesisCell™ (a batch reactor). A 21% conversion (3.4 mg product was obtained) was achieved within 3 h. Next, we attempted the oxidation of MK-8133 at high concentration to improve the product quantity. An 80 mL solution of 1.25 mM MK-8133 solution (40 mg starting material) in DMSO: aqueous 20 mM sodium phosphate buffer (40:60; v/v) was added to the SynthesisCell™. The reaction was monitored by LC/MS, 11% conversion was observed (4.4 mg M11 produced) within 12 h, the highest conversion was achieved at approximately 17% yield (6.9 mg M11 produced) with reaction duration of about 72 h (Figure S2-b, Supporting Information). Although the conversion yield in the SynthesisCell™ was low (17% from a 72 h electrolysis), there was little byproduct formed and the remaining 80% was recoverable MK-8133. It is plausible that the working electrode of SynthesisCell™ needed to be cleaned following a long duration electrolytic reaction. Further improvement of the electrosynthetic yield may be possible by using a suitable porous electrode for electrolysis since the majority of parent MK-8133 remained. Remarkably, as shown below, the isolated product analyzed by solution NMR was shown to be identical to that isolated from the *in vivo* matrix with the correct regio- and stereochemical configuration.

***NMR characterization of the electrochemically prepared M11 metabolite***

Definitive characterization of the oxidative metabolite, M11, of MK-8133 was accomplished using high resolution NMR techniques. Based on the multiplicity-edited HSQC spectrum of the metabolite, it was immediately apparent that only two, rather than the three methylene resonances of the parent molecule remained in the structure of the metabolite, indicating that, as expected, one of the methylenes had undergone oxidation in the electrochemical reaction. The two remaining methylene groups 2.27, 1.61/24.3 and 2.40, 1.88/19.1 ppm ( $^1\text{H}/^{13}\text{C}$ ) were consistent with the proton and carbon chemical shifts of the 5- and 4-positions, respectively, of the piperidine ring of the parent. The resonances for the 6-methine (4.56/45.5 ppm) and the methyl (1.44, *d*,  $J_{\text{HH}} = 6.9$  Hz/19.3 ppm) were also comparable to those of the parent, strongly suggesting that oxidation had occurred at the 2-methylene (Figure 2). A new methine resonance was observed in the multiplicity-edited HSQC spectrum at 5.49/80.0 ppm, which is consistent with hydroxylation at the 2-position of the piperidine ring; the carbon chemical shift is consistent for an aminol carbon resonance. Homonuclear proton-proton correlations in the COSY spectrum afforded further confirmation of hydroxylation at the 2-position of the piperidine ring (Figure 2).

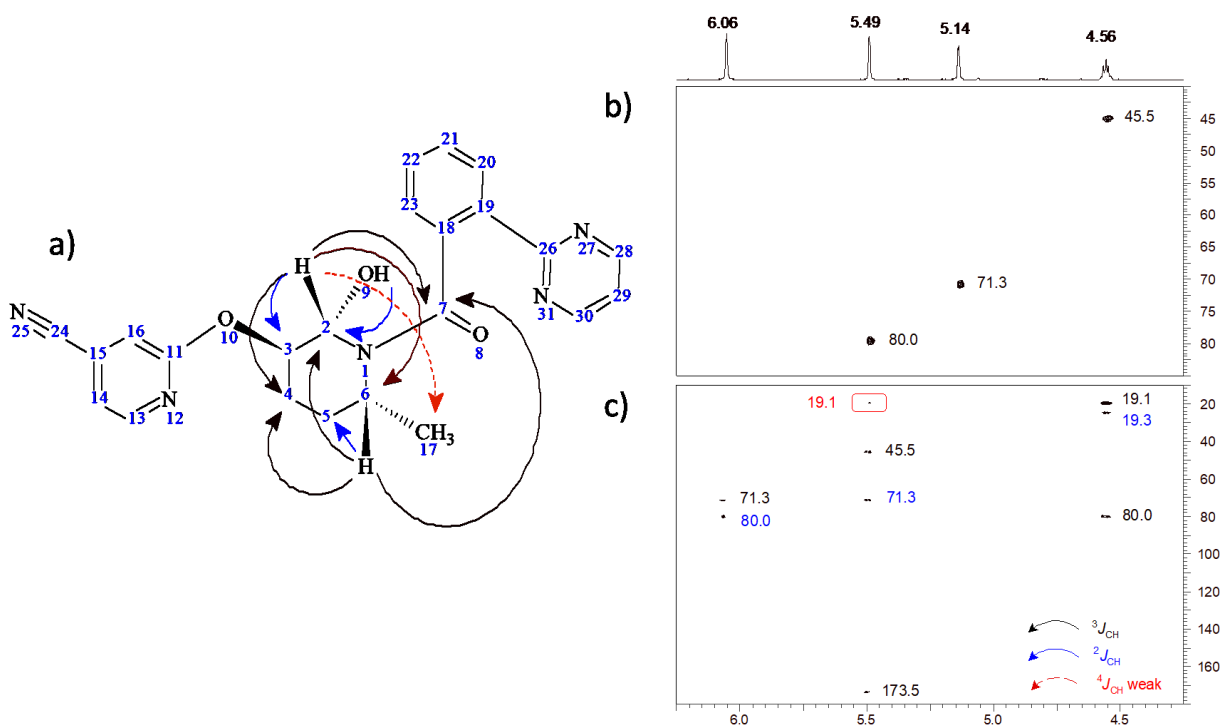


**Figure 2.** a) 600 MHz  $^1\text{H}$  NMR spectrum of the M-11 metabolite of MK-8133 in  $d_6$ -DMSO. b)  $^1\text{H}$  and  $^{13}\text{C}$  NMR resonance assignments for the M-11 metabolite made from the ensemble of 2D NMR data acquired for the sample. c) Pertinent region of the COSY spectrum showing correlations for the aliphatic resonances of the piperidine moiety in the structure. The proton resonating at 4.56 ppm affords correlations to the vicinal proton resonating at 2.27 ppm and to the methyl group resonating at 1.44 ppm. The open circle corresponds to a very weak correlation to the vicinal proton resonating at 1.61 ppm (see also the multiplicity-edited HSQC data shown in panel d) that is below the threshold used to plot the COSY data. d) Correlations for the aliphatic resonances of the M-11 metabolite observed in the multiplicity-edited HSQC spectrum.

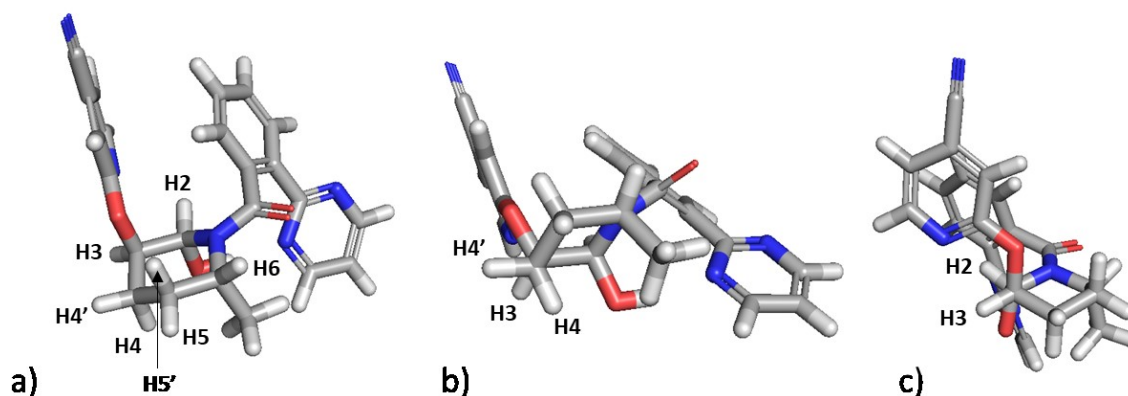
Long-range heteronuclear correlations were observed from the H2 methine proton (5.49 ppm) to carbons resonating at 173.5 (C7), 71.3 (C3), 45.5 (C6) and 19.1 (C4) ppm in an 8 Hz optimized HMBC experiment (Figure 3). The correlation to the C7 carbonyl was quite weak, which is to be expected based on the rotation about the bond between C7 and piperidine nitrogen. In similar fashion, correlations were observed from the H6 (4.56 ppm) proton to the C7 carbonyl, as well as carbons resonating at 80.0 (C2), 24.3 (C5) and 19.1 (C4) ppm (see numbered structure in Figure 3). In concert, these correlations

unequivocally establish the site of electrochemical oxidation as the 2-position of the piperidine ring.

Analysis of a resolution enhanced  $^1\text{H}$  NMR spectrum of the M11 metabolite revealed the H2 methine proton as a doublet  $J_{\text{HH}} = 2.5$  Hz; the H3 methine proton was observed as an apparent quartet due to the three equivalent homonuclear couplings of  $J_{\text{HH}} = 2.6$  Hz. These homonuclear coupling constants are consistent with the gauche relationship between the H2 and the H4' and H4'' protons with the H3 resonance, which can be seen looking along the C2-C3 bond axis of the energy-minimized structure of the M11 metabolite shown in Figure 4c. Further confirmation of the orientation of the 2-hydroxyl substituent was afforded by an observed NOE correlation between the 2-hydroxyl proton and the 6-methyl group in a NOESY spectrum (see Figure S10, Supporting Information).



**Figure 3.** a) Structure of the M-11 metabolite showing long-range  $^1\text{H}$ - $^{13}\text{C}$  correlations observed in the 8 Hz optimized HMBC spectrum. Correlation pathways are color-coded on the structure, with the most prevalent  $^3J_{\text{CH}}$  correlations denoted by black arrows;  $^2J_{\text{CH}}$  correlations are denoted by blue arrows; the single, weak  $^4J_{\text{CH}}$  correlation observed is denoted by the dashed red arrow. b) Aliphatic moiety correlations in the multiplicity-edited HSQC spectrum of the M11 metabolite. c) Correlations observed in the 8 Hz optimized HMBC spectrum. Chemical shift labels are color-coded as a function of the long-range heteronuclear correlation pathway involved. The very weak  $^4J_{\text{CH}}$  correlation from the proton resonating at 5.49 ppm to the methyl doublet is enclosed in the red box. It is interesting to note that no HMBC correlations were observed from the H3 proton resonating at 5.14 ppm to any of the aliphatic carbon resonances in the HMBC spectrum.



**Figure 4.** a) Energy-minimized structure of the M11 metabolite of MK-8133. The orientation of the H3 resonance relative to the H2 and H4' and H4'' resonances lead to dihedral angles that are consistent, based on the Karplus relationship, with the observed three small coupling constants exhibited by the H3 resonance. b) View along the C-4-C3 bond axis showing the *gauche* relationship of the H3 resonance to both H4 protons. c) View along the C2-C3 bond axis of the piperidine ring showing the *gauche* relationship of the H2 and H3 protons (see computational details below).

### ***Electrochemical oxidation of MK-6096 to produce M10***

Following the success in obtaining desired M11 metabolite from electrochemical oxidation of MK-8133, electrochemical oxidation of another parent compound, MK-6096, was investigated to determine whether the selective oxidation is broadly applicable and capable of producing the desired metabolite M10 (structure is shown in Scheme 1). As shown in Figure S3-a (Supporting Information), with no potential applied to the thin-layer  $\mu$ -PrepCell<sup>TM</sup> flow cell, the protonated and sodiated MK-6096 ions were observed at  $m/z$

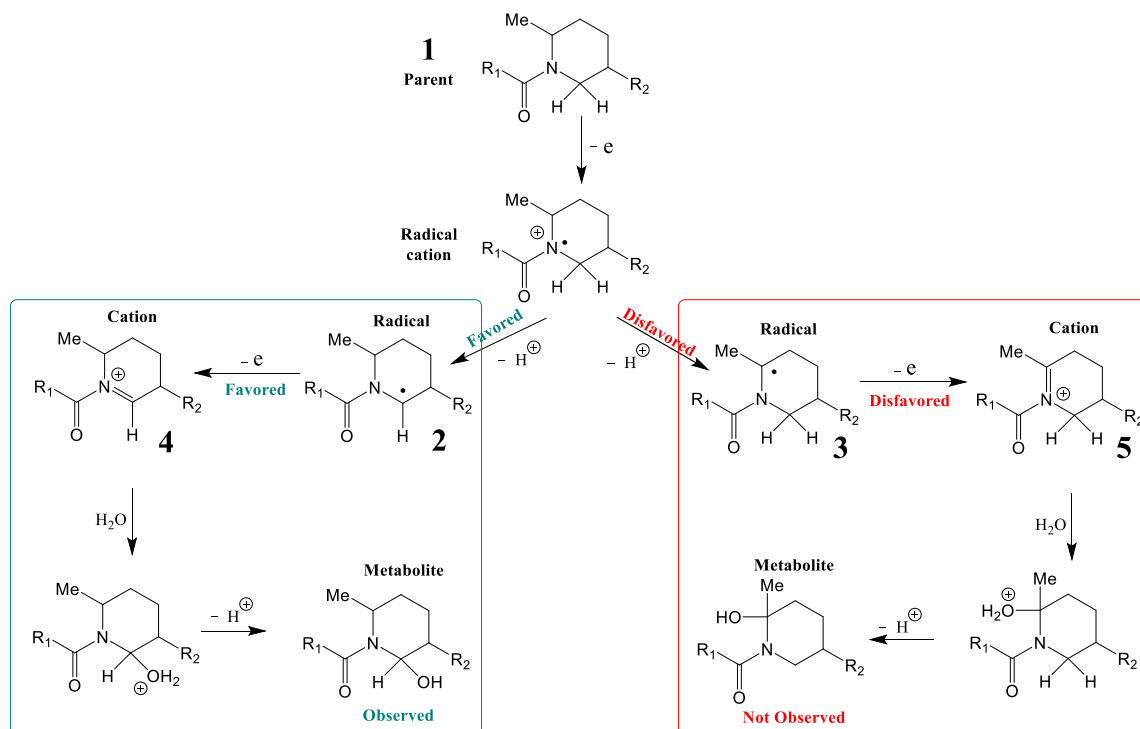
421 and 443, respectively, when 100  $\mu$ M MK-6096 in acetonitrile/water (50:50; v/v) containing 10 mM ammonium formate and 1% formic acid (pH 2.2) was passed through the cell at a flow rate of 10  $\mu$ L/min. When 1.8 V was applied to the BDD working electrode, two new ions were detected at  $m/z$  437 and 459 (Figure S3-b, Supporting Information), corresponding to the protonated and sodiated oxidation products of MK-6096, respectively. Based on the relative intensities of the remaining MK-6096 peaks and the product peak, the yield for electrosynthesis of the oxidative metabolite M10 was estimated to be  $\approx$ 40%. MS/MS of the  $m/z$  459 ion predominantly afforded fragment ions at  $m/z$  143, 169, 197 and 346, consistent with the sodium adduct of M10 structure (Figure S3-c, Supporting Information).

Based on the success of small-scale electrochemistry, the reaction was scaled up in an effort to produce usable quantities; mg quantities were obtained by using electrolysis in the SynthesisCell<sup>TM</sup>. For the mg scale electrosynthesis experiment, 0.5 mg/mL solution of MK-6096 dissolved in acetonitrile/water (50:50; v/v) containing 10 mM ammonium formate and 1% formic acid (pH 2.2) was prepared and 80 mL of this solution was added to the SynthesisCell<sup>TM</sup>. Conversion to the desired hydroxylated metabolite, M10, from MK-6096 was monitored by LC/MS. As shown in Figure S4 (Supporting Information), optimal conversion at about 14% occurred when the duration of the reaction was approximately 3.5 h. The desired hydroxylated metabolite was isolated (5.6 mg) and characterized by NMR analysis, which established the structure of the metabolite as M10. Detailed NMR analysis is shown in Figures 11S-16S and Table S3 (Supporting Information). Characterization of the structure of the isolated product of the

electrochemical reaction confirmed that selective oxidation occurs at the C2 position of the MK-6096 piperidine ring.

### *Elucidation of electrochemical oxidation regioselectivity and stereoselectivity*

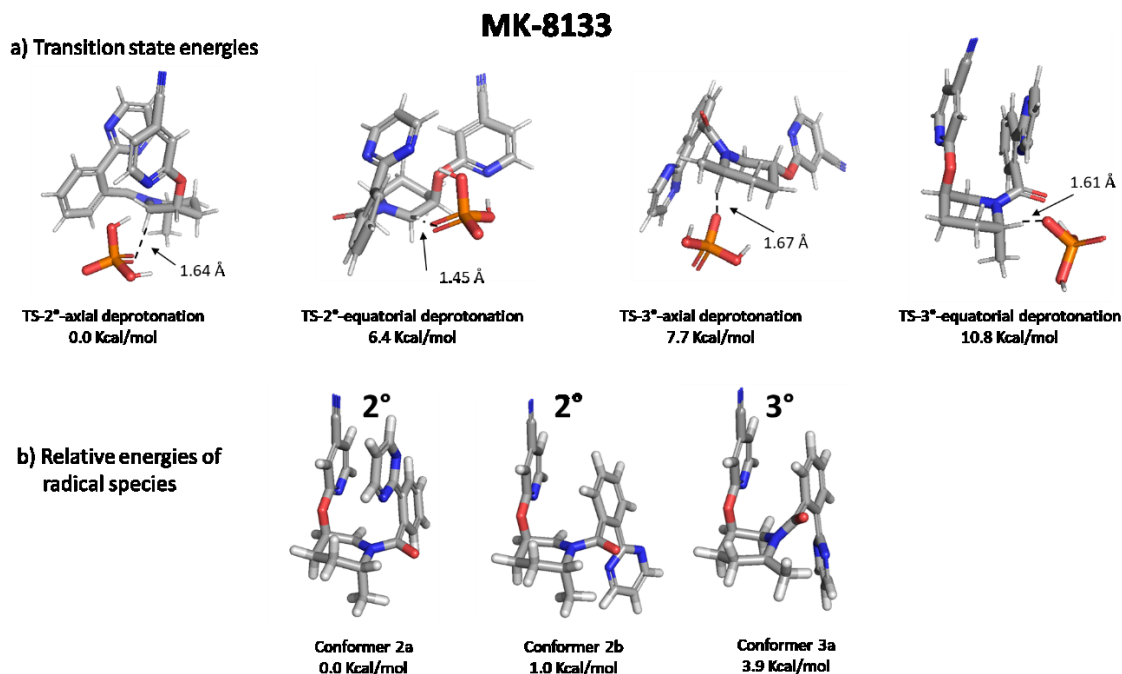
Scheme 4 shows the proposed mechanism for the electrochemical hydroxylation of MK-8133, which is similar to the anodic oxidation of an amide (Scheme 2). Hydroxylation on the tertiary carbon alpha to the nitrogen atom should be the predominant product since tertiary carbon radicals are normally more stable than secondary carbon radicals (**3** in Scheme 4). However, our results indicate that the electrochemical oxidation of MK-8133 and MK-6096 yielded primarily hydroxylation products on the secondary carbon, alpha to the nitrogen atom.



**Scheme 4.** The proposed mechanism for the electrochemical hydroxylation of MK-8133 and MK-6096.



A desire to rationalize this unexpected regioselectivity prompted further investigation using density functional theory (DFT) calculations. Conformational space was exhaustively sampled using three conformer generators (a combination of rules-based and distance geometry-based methods) followed by molecular mechanics minimization using MMFF94, a workflow that has been previously published.<sup>80</sup> Initial mechanistic rationale was explored based on comparing energies of the penultimate iminium cations, however, the relative energies of the cations predicted the formation of **5** and not **4** (Scheme 4). The relative stability of the iminium cations has, however, been used to rationalize the experimentally observed regioselectivity for a related series of compounds.<sup>81</sup> Libendi, *et al.* proposed that the regioselectivity for a cyano or ester-substituted piperidine could be attributed to the relative stability of the LUMO energies of the cations (Figure S17 and additional discussion in the SI), however, as will be shown here, we propose that the regioselectivity for this class of reactions was instead governed by kinetic control of the deprotonation step.



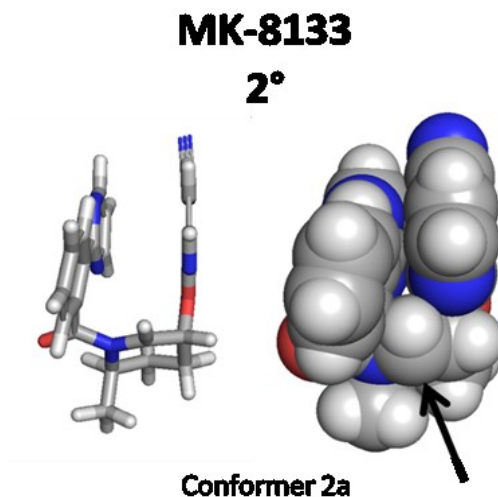
**Figure 5.** (a) Transition states for deprotonation, by a dihydrogen phosphate anion, of the radical cation leading to either the secondary or tertiary radicals. (b) Low energy conformations of the secondary and tertiary radicals for MK-8133. Carbon = grey, oxygen = red, nitrogen = blue, hydrogen = white, phosphorous = orange.

Transition states for the deprotonation step forming radical **2** from the radical cation of **1** were identified for MK-8133 (Figure 5). While there are several possibilities for what base may be deprotonating the intermediate radicals, we have selected to model a dihydrogen phosphate anion as the probable base present in the reaction solution. In the presence of an implicit solvent model, the preferred (quasiharmonic corrected Gibbs energy at 298 K) kinetic pathway is favored by 7.7 kcal/mol in support of formation of the observed product (see additional details in SI including Table S4). The lowest energy transition state for the observed secondary radical pathway derives from deprotonation of the axial proton from the lowest energy conformation of the MK-8133 starting material which adopts a chair conformation of the central piperidine core. Deprotonation of the axial proton allows for optimal overlap of the resultant p-orbitals of the carbon being

deprotonated and the piperidine nitrogen, driving stabilization of the observed pathway. The higher energy of the TS for the analogous axial deprotonation at the tertiary  $\alpha$ -carbon may be explained by steric strain from the equatorial methyl group with the carbonyl group bonded to N. The same steric and electronic factors are proposed to account for the selectivity observed with MK-6096.

Figure 5b depicts the three energetically significant conformers of the secondary and tertiary radicals of MK-8133. Conformer **2a** (conformers of secondary radical are denoted “2<sup>o</sup>” while conformers of the tertiary radical are denoted “3<sup>o</sup>” in Figure 5) dominates the free energy surface and exhibits an intramolecular stacking interaction between the pyridine and biaryl systems as in the preferred transition state. Rotation about the amide-biaryl dihedral angle leads to conformer **2b** of MK-8133 which is 1.0 Kcal/mol higher in energy. The tertiary radical is sterically disfavored owing to the methyl substituent at the radical center wanting to sit in the same space as the amide carbonyl which is not possible (this same steric argument explains why Libendi, *et al.*, observed different regioselectivity for the less sterically bulky cyano moiety compared to the ester functionality<sup>81</sup>). Annotated as conformer **3a**, the conformation of the tertiary radical is 3.9 Kcal/mol higher in energy than the lowest energy secondary radical conformer **2a**. Additional steric repulsion eliminates the amide-biaryl rotamer equivalent to conformer **2b** from consideration for the tertiary radical. In Figure S18, a series of calculations was performed on model systems containing either a truncated amide substituted piperidine or a modification of this core where the carbonyl was replaced with a methylene (shifts sp<sup>2</sup> amide geometry to one that allows the piperidine nitrogen to be more pyramidal). Calculations indicate that the methylene species allows for a more stable tertiary radical

compared to the secondary; when the carbonyl is in place, the secondary radical is preferred for reasons specified above.



**Figure 6.** Global minima conformation of the MK-8133 iminium cation **4** (derived from the secondary (2°) radical) shown in both a stick and space-filling representation. The arrow indicates favorable ‘bottom’ side approach in the axial position for the nucleophile whereas nucleophilic approach from the ‘top’ side is sterically blocked. Carbon = grey, oxygen = red, nitrogen = blue, hydrogen = white.

Following a second one-electron oxidation, **2** is converted to an iminium cation **4** (Scheme 4), which undergoes nucleophilic addition in the presence of water to give the 2-hydroxypiperidine product. It can be seen from Figure 6, that the global minima of the MK-8133 iminium cation generated from the radical precursor adopts a U-shaped conformation. Steric hindrance caused by the 2-phenylpyrimidine ring and cyanopyridine directs the nucleophilic attack to the bottom (*re* face) of *N*-acyl iminium ion in the axial position of the piperidine chair conformation. This is consistent with complete stereoselectivity observed during the electrochemical oxidation of MK-8133. Based on relative kinetic barrier heights and low energy conformations, we propose that the high regio- and stereoselectivity observed during electrochemical oxidation is driven by initial oxidation of the piperidine nitrogen of MK-8133, which then follows a reaction cascade driven

largely by the steric properties of the piperidine exocyclic methyl group with a significant preference for deprotonation along the pathway towards the secondary radical derived product.

Verification of the proposed factors governing selectivity observed with MK-8133 was conducted through electrochemical oxidation of a second orexin receptor antagonist, MK-6096, which possesses a similar molecular scaffold. As mentioned above, the product M10 from electrolytic oxidation of MK-6096 was isolated and characterized by NMR, which established the metabolite to be M10 with complete regio- and stereoselectivity similar to that of M11. Similar *in vacuo* radical stability calculations were carried out for MK-6096. Figure S19 shows the relative stabilities of transient radicals with the 2°  $\alpha$ -carbon radicals having lower conformational energies compared to the 3°  $\alpha$ -carbon radicals. Though the energy difference (1.0 Kcal/mol) is smaller compared to those calculated for MK-8133 (3.9 Kcal/mol), the MK-6096 carbon radical should predominantly form on the secondary  $\alpha$ -carbon given the transition states identified for MK-8133 deprotonation. A larger number of conformers are energetically accessible for MK-6096, likely due to the methylene-spaced ether linkage, which affords more flexibility to the pyridine ring. This flexibility provides additional space for the biaryl to be positioned closing the gap on the energetic preference of the secondary radical. Experimentally, high regioselectivity was again observed. It can be seen in Figure S20 that MK-6096 possesses the same sterically blocked U-shaped conformation for the iminium cation generated from the secondary radical. For MK-6096, owing to similar conformer energies, two conformations must be considered, but as seen in Figure S20, it is evident that both conformations are more accessible for bottom side (as depicted) attack in the axial position

of the chair conformation of the central piperidine. Transition states for the deprotonation step of MK-6096 were not undertaken as it is expected that the energetic preferences will mirror that of MK-8133.

## Conclusions

In conclusion, we have demonstrated the one-step electrochemical synthesis of milligram quantities of  $\alpha$ -hydroxyl amide metabolites of MK-8133 and MK-6096 with exclusive regio- and stereoselectivity in aqueous media, the structures and stereochemical features confirmed by NMR spectroscopy. Based on density functional theory (DFT) calculations, the reaction regio- and stereoselectivity is governed by the relative stability of the transition states for deprotonation of the transient radical cations and steric hindrance directing axial addition experienced by the subsequent iminium species, respectively. Finally, it is shown that online EC/MS is a powerful tool in quickly identifying and optimizing the reaction conditions for desired products. Adapting this tool in the process can shorten the time cycle for electrochemical synthesis greatly.

## Experimental Section

### *Electrochemical Synthesis*

Small-scale reaction screening was undertaken using a lab-built EC/MS apparatus (Scheme 3) which consisted of a thin-layer electrochemical flow cell coupled with a Waters Xevo QToF mass spectrometer. Liquid sample desorption electrospray ionization (DESI) was used the EC/MS interface, as described previously in detail.<sup>82-84</sup> The thin-layer  $\mu$ -PrepCell<sup>TM</sup> (ANTEC BV, Netherlands) equipped with a glassy carbon working electrode (30x12 mm<sup>2</sup>) was used for oxidizing MK-8133; a BDD working electrode (30x12 mm<sup>2</sup>)

was used to oxidize MK-6096. A HyREF<sup>TM</sup> electrode was used as a reference electrode and titanium was used as the counter electrode. The electrochemical reaction product flowed out of the cell via a short piece of fused silica connection capillary (i.d., 0.1 mm, length 4.0 cm) and underwent interactions with the charged microdroplets from DESI solvent spray for ionization. The capillary outlet was placed about 1 mm downstream from the DESI spray probe tip and kept in-line with the sprayer tip and the mass spectrometer's inlet. The spray solvent for DESI was methanol/water (1:1 by volume) containing 1% acetic acid and a high voltage of 5 kV was applied to the spray probe. The flow rates for both the DESI probe solvent and the sample solutions passing through the electrochemical cell for electrolysis were 10  $\mu$ L/min. The EC/MS setup allowed direct online monitoring of the electrosynthetic product or offline collection of the product from the  $\mu$ -PrepCell<sup>TM</sup>.

Large-scale electrochemical synthesis was undertaken using an apparatus consisting of a ROXY potentiostat (ANTEC BV, Netherlands) with an extended current range (up to 20 mA), controlled by Antec Dialogue software, and a bulk SynthesisCell<sup>TM</sup> (80 mL volume, ANTEC BV, Netherlands). The SynthesisCell<sup>TM</sup> was equipped with a flat smooth BDD working electrode (30x30 mm<sup>2</sup>), a HyREF<sup>TM</sup> reference electrode and an auxiliary electrode without a frit. The reaction solution consisted of 80 mL of 0.5 mg/mL of MK-8133 dissolved in DMSO/20 mM sodium phosphate buffer (40:60; v/v) or 80 mL of 1.2 mM of MK-6096 dissolved in acetonitrile/water (50:50; v/v) containing 10 mM ammonium formate and 1% formic acid. A square-wave pulse potential was applied to the SynthesisCell<sup>TM</sup> for oxidizing both MK-8133 and MK-6096. Under the optimized conditions, the potentials were +2.6 V (E<sub>1</sub>) and +2.2 V (E<sub>2</sub>) for oxidizing MK-8133 and the potentials were +1.8 V (E<sub>1</sub>) and +1.4 V (E<sub>2</sub>) for oxidizing MK-6096. Time intervals were

1,990 ms ( $t_1$ ) and 1,010 ms ( $t_2$ ). The progress of the synthesis was checked every 10 min by taking an aliquot of 1  $\mu$ L from the SynthesisCell<sup>TM</sup> solution. The sample was diluted by a factor of 1000 prior to liquid chromatography/mass spectrometry (LC/MS) analysis (see details in Supporting Information). Both M11 obtained from MK-8133 electrolysis (6.9 mg, 17% yield) and M10 obtained from MK-6090 electrolysis (5.6 mg, 14% yield) were white powder solid.

### ***NMR Characterization***

High resolution nuclear magnetic resonance (NMR) data were collected on either Bruker Avance III HD 700 MHz or Bruker Avance III HD 600 MHz NMR spectrometers (Bruker BioSpin Corporation, Billerica, MA); both spectrometers were equipped with a 1.7 mm HCN TCI MicroCryoProbe<sup>TM</sup>. Approximately 50  $\mu$ g of the purified hydroxylated metabolite of MK-6096 was dissolved in 35  $\mu$ L of CD<sub>3</sub>CN and used for solution NMR experiments utilizing standard pulse sequences (proton, COSY, ME-HSQC, 8 Hz optimized HMBC, and ROESY with mixing times of 400 msec and 500 msec). Comparable NMR experiments were performed on a 50  $\mu$ g of the purified hydroxylated M11 metabolite of MK-8133 dissolved in 35  $\mu$ L of  $d_6$ -DMSO.

### ***Computational chemistry***

Conformational searches were performed according to a computational workflow that combined several conformer generators with energetic evaluation using a series of force field and low level density functional methods.<sup>80</sup> After identification of an initial conformer ensemble, more accurate Boltzmann distributions based on free energies (298K) were calculated using M06-2X/6-31G\*\* for conformer distributions with diffuse functions



added when calculating transition states for deprotonation. (see additional computational details in SI including quasiharmonic corrections to the transition state free energies).<sup>85</sup> Stationary points were confirmed by vibrational frequency analysis. Transition states were confirmed with IRC (intrinsic reaction coordinate) following and confirmed minimizations to reactants and products. All calculations were run using Gaussian g09.d01 or g16.a03.<sup>86</sup> Implicit solvation was modeled using SMD.<sup>87</sup> For ground state species, analysis was focused on conformers contributing >2% to the overall ensemble for each species.

### **Conflicts of interest**

There are no conflicts to declare.

### **Supporting Information**

Additional experimental procedure, MS, MS/MS, NMR and computational data are included. The data underlying this study are available in the published article and its online supplementary material.

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### **References**

1. Benca, R. M., Diagnosis and Treatment of Chronic Insomnia: A Review. *Psych. Serv.* **2005**, 56 (3), 332-343.

2. Compennolle, F.; Saleh, M. A.; Van den Branden, S.; Toppet, S.; Hoornaert, G., Regioselective oxidation of piperidine-3 derivatives: a synthetic route to 2,5-substituted piperidines. *J. Org. Chem.* **1991**, *56* (7), 2386-2390.
3. Pelkonen, O.; Turpeinen, M.; Uusitalo, J.; Rautio, A.; Raunio, H., Prediction of drug metabolism and interactions on the basis of in vitro investigations. *Basic Clin. Pharmacol. Toxicol.* **2005**, *96* (3), 167-175.
4. Plant, N., Strategies for using in vitro screens in drug metabolism. *Drug Discov. Today* **2004**, *9* (7), 328-336.
5. Brandon, E. F. A.; Raap, C. D.; Meijerman, I.; Beijnen, J. H.; Schellens, J. H. M., An update on in vitro test methods in human hepatic drug biotransformation research: pros and cons. *Toxicol. Appl. Pharmacol.* **2003**, *189* (3), 233-246.
6. Baillie, T. A., Metabolism and Toxicity of Drugs. Two Decades of Progress in Industrial Drug Metabolism. *Chem. Res. Toxicol.* **2008**, *21* (1), 129-137.
7. Obach, S. R.; Dalvie, D. K.; Walker, G. S. In *Identification of drug metabolites*, John Wiley & Sons, Inc.: 2012; pp 121-175.
8. Stepan, A. F.; Mascitti, V.; Beaumont, K.; Kalgutkar, A. S., Metabolism-guided drug design. *MedChemComm* **2013**, *4* (4), 631-652.
9. Cusack, K. P.; Koolman, H. F.; Lange, U. E. W.; Peltier, H. M.; Piel, I.; Vasudevan, A., Emerging technologies for metabolite generation and structural diversification. *Bioorg. Med. Chem. Lett.* **2013**, *23* (20), 5471-5483.
10. Sherry, B. D.; Yin, J.; Fleitz, F. J., Enantioselective Synthesis of a DualOrexin Receptor Antagonist Ian K. Mangion. *Org. Lett.* **2012**, *14*, 3458-3461.

11. Schäfer, H. J., Comparison of Chemical and Electrochemical Methods in Organic Synthesis. In *Encyclopedia of Electrochemistry*, Wiley-VCH Verlag GmbH & Co. KGaA: 2007.
12. Wyatt, P. B., Organic Electrochemistry (4th Edition), Edited by H. Lund and O. Hammerich, Marcel Decker, 2001, 1393. *Electrochim. Acta* **2002**, 47, 1163.
13. Horn, E. J.; Rosen, B. R.; Chen, Y.; Tang, J.; Chen, K.; Eastgate, M. D.; Baran, P. S., Scalable and sustainable electrochemical allylic C-H oxidation. *Nature* **2016**, 533 (7601), 77-81.
14. Badalyan, A.; Stahl, S. S., Cooperative electrocatalytic alcohol oxidation with electron-proton-transfer mediators. *Nature* **2016**, 535 (7612), 406-10.
15. Schulz, L.; Enders, M.; Elsler, B.; Schollmeyer, D.; Dyballa, K. M.; Franke, R.; Waldvogel, S. R., Reagent- and Metal-Free Anodic C-C Cross-Coupling of Aniline Derivatives. *Angew. Chem. Int. Ed. Engl.* **2017**, 56 (17), 4877-4881.
16. Yan, M.; Kawamata, Y.; Baran, P. S., Synthetic Organic Electrochemical Methods Since 2000: On the Verge of a Renaissance. *Chem. Rev.* **2017**, 117 (21), 13230-13319.
17. Okada, Y.; Chiba, K., Redox-Tag Processes: Intramolecular Electron Transfer and Its Broad Relationship to Redox Reactions in General. *Chem. Rev.* **2018**, 118 (9), 4592-4630.
18. Moeller, K. D., Using Physical Organic Chemistry To Shape the Course of Electrochemical Reactions. *Chem. Rev.* **2018**, 118 (9), 4817-4833.
19. Francke, R.; Little, R. D., Redox catalysis in organic electrosynthesis: basic principles and recent developments. *Chem. Soc. Rev.* **2014**, 43 (8), 2492-2521.

20. Fu, N.; Sauer, G. S.; Lin, S., A general, electrocatalytic approach to the synthesis of vicinal diamines. *Nature Protocols* **2018**, *13* (8), 1725-1743.
21. Mahanty, K.; Maiti, D.; De Sarkar, S., Regioselective C–H Sulfonylation of 2H-Indazoles by Electrosynthesis. *J. Org. Chem.* **2020**, *85* (5), 3699-3708.
22. Yuan, Y.; Lei, A., Is electrosynthesis always green and advantageous compared to traditional methods? *Nature Commun.* **2020**, *11* (1), 1-3.
23. Schotten, C.; Nicholls, T. P.; Bourne, R. A.; Kapur, N.; Nguyen, B. N.; Willans, C. E., Making electrochemistry easily accessible to the synthetic chemist. *Green Chem.* **2020**, *22* (11), 3358-3375.
24. Leech, M. C.; Lam, K., A practical guide to electrosynthesis. *Nature Rev. Chem.* **2022**, *6* (4), 275-286.
25. Elsler, B.; Schollmeyer, D.; Dyballa, K. M.; Franke, R.; Waldvogel, S. R., Metal- and reagent-free highly selective anodic cross-coupling reaction of phenols. *Angew. Chem. Int. Ed.* **2014**, *53*, 5210-5213.
26. Schaefer, H. J., CC bonds at anodes and cathodes. *Angew. Chem.* **1981**, *93*, 978-1000.
27. Tang, F.; Moeller, K. D., Intramolecular Anodic Olefin Coupling Reactions: The Effect of Polarization on Carbon-Carbon Bond Formation. *J. Am. Chem. Soc.* **2007**, *129*, 12414-12415.
28. Xu, H.-C.; Moeller, K. D., Intramolecular Anodic Olefin Coupling Reactions: Use of the Reaction Rate To Control Substrate/Product Selectivity. *Angew. Chem., Int. Ed.* **2010**, *49*, 8004-8007.

29. Morofuji, T.; Shimizu, A.; Yoshida, J.-i., Metal- and Chemical-Oxidant-Free C-H/C-H Cross-Coupling of Aromatic Compounds: The Use of Radical-Cation Pools. *Angew. Chem., Int. Ed.* **2012**, *51*, 7259-7262.
30. Leow, W. R.; Lum, Y.; Ozden, A.; Wang, Y.; Nam, D.-H.; Chen, B.; Wicks, J.; Zhuang, T.-T.; Li, F.; Sinton, D.; Sargent, E. H., Chloride-mediated selective electrosynthesis of ethylene and propylene oxides at high current density. *Science* **2020**, *368*, 1228–1233.
31. Hu, X.; Nie, L.; Zhang, G.; Lei, A., Electrochemical Oxidative [4+2] Annulation for the  $\pi$ -Extension of Unfunctionalized Hetero-biaryl Compounds. *Angew. Chem. Int. Ed.* **2020**, *59*, 15238-15243.
32. Sattler, L. E.; Otten, C. J.; Hilt, G., Alternating Current Electrolysis for the Electrocatalytic Synthesis of Mixed Disulfide via Sulfur-Sulfur Bond Metathesis towards Dynamic Disulfide Libraries. *Chem. - Eur. J.* **2020**, *26* (14), 3129-3136.
33. Zhang, W.; Carpenter, K. L.; Lin, S., Electrochemistry Broadens the Scope of Flavin Photocatalysis: Photoelectrocatalytic Oxidation of Unactivated Alcohols. *Angew. Chem. Int. Ed. Eng.* **2020**, *59*, 409-417.
34. Zhu, X.; Zhou, X.; Jing, Y.; Li, Y., Electrochemical synthesis of urea on MBenes. *Nature Commun.* **2021**, *12* (1), 1-9.
35. Xu, C.; Ma, B.; Gao, Z.; Dong, X.; Zhao, C.; Liu, H., Electrochemical DNA synthesis and sequencing on a single electrode with scalability for integrated data storage. *Science Adv.* **2021**, *7* (46), eabk0100.

36. Shono, T.; Matsumura, Y.; Tsubata, K., Electroorganic chemistry. 46. A new carbon-carbon bond forming reaction at the .alpha.-position of amines utilizing anodic oxidation as a key step. *J. Am. Chem. Soc.* **1981**, *103*, 1172.
37. Wang, F.; Rafiee, M.; Stahl, S. S., Electrochemical Functional-Group-Tolerant Shono-type Oxidation of Cyclic Carbamates Enabled by Aminoxyl Mediators. *Angew. Chem. Int. Ed.* **2018**, *57*, 6686–6690.
38. Jones, A. M.; Banks, C. E., The Shono-type electroorganic oxidation of unfunctionalised amides. Carbon–carbon bond formation via electrogenerated N-acyliminium ions *Beilstein J. Org. Chem.* **2014**, *10*, 3056–3072.
39. Diehl, G.; Karst, U., On-line electrochemistry – MS and related techniques. *Anal. Bioanal. Chem.* **2002**, *373*, 390-398.
40. Permentier, H. P.; Bruins, A. P.; Bischoff, R., Electrochemistry-mass spectrometry in drug metabolism and protein research. *Mini Rev. Med. Chem.* **2008**, *8*, 46-56.
41. Li, J.; Dewald, H. D.; Chen, H., Online Coupling of Electrochemical Reactions with Liquid Sample Desorption Electrospray Ionization-Mass Spectrometry. *Anal. Chem.* **2009**, *81*, 9716-9722.
42. Zhao, P.; Wang, Q.; Kaur, M.; Kim, Y.-I.; Dewald, H. D.; Mozziconacci, O.; Liu, Y.; Chen, H., Absolute Quantitation of Proteins by Coulometric Mass Spectrometry. *Anal. Chem.* **2020**, *92*, 7877-7883.
43. Chintalapudi, K.; Badu-Tawiah, A. K., An integrated electrocatalytic nESI-MS platform for quantification of fatty acid isomers directly from untreated biofluids. *Chem. Sci.*, **2020**, *11*, 9891-9897

44. Tang, S.; Fan, L.; Cheng, H.; Yan, X., Incorporating electro-epoxidation into electrospray ionization mass spectrometry for simultaneous analysis of negatively and positively charged unsaturated glycerophospholipids. *J. Am. Soc. Mass Spectrom.* **2020**, *32* (9), 2288-2295.
45. Hu, J.; Zhang, N.; Zhang, P.-K.; Chen, Y.; Xia, X.-H.; Chen, H.-Y.; Xu, J.-J., Coupling a Wireless Bipolar Ultramicroelectrode with Nano-electrospray Ionization Mass Spectrometry: Insights into the Ultrafast Initial Step of Electrochemical Reactions. *Angew. Chem. Int. Ed. Engl.* **2020**, *59*, 18244-18248.
46. Brown, T. A.; Chen, H.; Zare, R. N., Identification of Fleeting Electrochemical Reaction Intermediates Using Desorption Electrospray Ionization Mass Spectrometry. *J. Am. Chem. Soc.* **2015**, *137*, 7274-7277.
47. Brown, T. A.; Chen, H.; Zare, R. N., Detection of the Short-Lived Radical Cation Intermediate in the Electrooxidation of N,N-Dimethylaniline by Mass Spectrometry. *Angew. Chem. Int. Ed. Engl.* **2015**, *54*, 11183-11185.
48. Qiu, R.; Zhang, X.; Luo, H.; Shao, Y., Mass spectrometric snapshots for electrochemical reactions. *Chem. Sci.* **2016**, *7*, 6684-6688.
49. Yu, J.; Zhou, Y.; Hua, X.; Liu, S.; Zhu, Z.; Yu, X.-Y., Capturing the transient species at the electrode–electrolyte interface by in situ dynamic molecular imaging. *Chem. Commun.* **2016**, *52*, 10952-10955.
50. Wang, Z.; Zhang, Y.; Liu, B.; Wu, K.; Thevuthasan, S.; Baer, D. R.; Zhu, Z.; Yu, X.-Y.; Wang, F., In Situ Mass Spectrometric Monitoring of the Dynamic Electrochemical Process at the Electrode–Electrolyte Interface: a SIMS Approach. *Anal. Chem.* **2017**, *89*, 960-965.

51. Khanipour, P.; Löffler, M.; Reichert, A. M.; Haase, F. T.; Mayrhofer, K. J. J.; Katsounaros, I., Cover Picture: Electrochemical Real-Time Mass Spectrometry (EC-RTMS): Monitoring Electrochemical Reaction Products in Real Time (Angew. Chem. Int. Ed. 22/2019). *Angew. Chem. Int. Ed.* **2019**, *58*, 7145-7145.
52. Hasa, B.; Jouny, M.; Ko, B. H.; Xu, B.; Jiao, F., Flow Electrolyzer Mass Spectrometry with a Gas-Diffusion Electrode Design. *Angew. Chem. Int. Ed.* **2021**, *60* (6), 3277-3282.
53. Liu, J.; Yu, K.; Zhang, H.; He, J.; Jiang, J.; Luo, H., Mass spectrometric detection of fleeting neutral intermediates generated in electrochemical reactions. *Chem. Sci.* **2021**, *12* (27), 9494-9499.
54. Hu, J.; Wang, T.; Zhang, W. J.; Hao, H.; Yu, Q.; Gao, H.; Zhang, N.; Chen, Y.; Xia, X. H.; Chen, H. Y., Dissecting the Flash Chemistry of Electrogenenerated Reactive Intermediates by Microdroplet Fusion Mass Spectrometry. *Angew. Chem. Int. Ed.* **2021**, *133* (34), 18642-18646.
55. Wan, Q.; Chen, S.; Badu-Tawiah, A. K., An integrated mass spectrometry platform enables picomole-scale real-time electrosynthetic reaction screening and discovery. *Chem. Sci.*, **2018**, *9*, 5724-5729.
56. Wang, Q.; Wang, Q.; Zhang, Y.; Mohamed, Y. M.; Pacheco, C.; Zheng, N.; Zare, R. N.; Chen, H., Electrocatalytic redox neutral [3 + 2] annulation of N-cyclopropylanilines and alkenes. *Chem. Sci.* **2020**, *12* (3), 969-975.
57. Cheng, H.; Yang, T.; Edwards, M.; Tang, S.; Xu, S.; Yan, X., Picomole-Scale Transition Metal Electrocatalysis Screening Platform for Discovery of Mild C–C



- Coupling and C–H Arylation through in Situ Anodically Generated Cationic Pd. *J. Am. Chem. Soc.* **2022**, 144, 1306–1312.
58. Jurva, U.; Wikstrom, H. V.; Weidolf, L.; Bruins, A. P., Comparison between electrochemistry/mass spectrometry and cytochrome P450 catalyzed oxidation reactions. *Rapid Commun. Mass Spectrom.* **2003**, 17 (8), 800-810.
59. Karst, U., Electrochemistry/mass spectrometry (EC/MS)--a new tool to study drug metabolism and reaction mechanisms. *Angew. Chem. Int. Ed.* **2004**, 43 (19), 2476-8.
60. Johansson Mali'n, T.; Weidolf, L.; Castagnoli Jr, N.; Jurva, U., P450-catalyzed vs. electrochemical oxidation of haloperidol studied by ultra-performance liquid chromatography/electrospray ionization mass spectrometry. *Rapid Commun. Mass Spectrom.* **2010**, 24 (9), 1231-1240.
61. Nouri-Nigjeh, E.; Bischoff, R.; Bruins, A. P.; Permentier, H. P., Electrochemistry in the mimicry of oxidative drug metabolism by cytochrome P450s. *Curr. Drug Metab.* **2011**, 12 (4), 359-371.
62. Lohmann, W.; Baumann, A.; Karst, U., Electrochemistry and LC-MS for metabolite generation and identification: Tools, technologies, and trends. *LCGC North Am.* **2010**, 28 (6), 470-478.
63. Stalder, R.; Roth, G. P., Preparative Microfluidic Electrosynthesis of Drug Metabolites. *ACS Med. Chem. Lett.* **2013**, 4 (11), 1119-1123.
64. Mautjana, N. A.; Estes, J.; Eyler, J. R.; Brajter-Toth, A., Antioxidant Pathways and One-Electron oxidation of Dopamine and Cysteine in Electrospray and On-line

- Electrochemistry Electrospray Ionization Mass Spectrometry. *Electroanalysis* **2008**, *20*, 1959-1967.
65. Permentier, H. P.; Bruins, A. P.; Bischoff, R., Electrochemistry-mass Spectrometry in drug metabolism and protein research. *Mini-Rev. Med. Chem.* **2008**, *8*, 46-56.
66. Gun, J.; Bharathi, S.; Gutkin, V.; Rizkov, D.; Voloshenko, A.; Shelkov, R.; Sladkevich, S.; Kyi, N.; Rona, M.; Wolanov, Y.; Rizkov, D.; Koch, M.; Mizrahi, S.; Pridkhochenko, P. V.; Modestov, A.; Lev, O., Highlights in Coupled Electrochemical Flow Cell-Mass Spectrometry, EC/MS. *Israel J. Chem.* **2010**, *50*, 360-373.
67. Jurva, U.; Wikstrom, H. V.; Bruins, A. P., In vitro mimicry of metabolic oxidation reactions by electrochemistry/mass spectrometry. *Rapid Commun. Mass Spectrom.* **2000**, *14*, 529-533.
68. Karst, U., Electrochemistry/mass spectrometry (EC/MS)-A new tool to study drug metabolism and reaction mechanisms. *Angew. Chem. Int. Ed.* **2004**, *43*, 2476-2478.
69. Bussy, U.; Boujtita, M., Advances in the Electrochemical Simulation of Oxidation Reactions Mediated by Cytochrome P450. *Chem. Res. Toxicol.* **2014**, *27*, 1652-1668.
70. Nowak, P.; Woźniakiewicz, M.; Kościelniak, P., Simulation of drug metabolism. *TrAC* **2014**, *59*, 42-49.
71. Yuan, T.; Permentier, H.; Bischoff, R., Surface-modified electrodes in the mimicry of oxidative drug metabolism. *TrAC* **2015**, *70*, 50-57.
72. Pedersen, A.; Ambach, L.; König, S.; Weinmann, W., Electrochemical simulation of Phase I metabolism for 21 drugs using four different working electrodes in an automated screening setup with MS detection. *Bioanalysis* **2014**, *6*, 2607-21.

73. Shono, T., Synthesis of alkaloidal compounds using an electrochemical reaction as a key step. *Top. Curr. Chem.* **1988**, *148* (Electrochemistry 3), 131-51.
74. Siu, T.; Li, W.; Yudin, A. K., Parallel Electrosynthesis of  $\alpha$ -Alkoxy carbamates,  $\alpha$ -Alkoxy amides, and  $\alpha$ -Alkoxy sulfonamides Using the Spatially Addressable Electrolysis Platform (SAEP). *J. Combin. Chem.* **2000**, *2* (5), 545-549.
75. Yudin, A. K.; Siu, T., Combinatorial electrochemistry. *Curr. Opin. Chem. Biol.* **2001**, *5* (3), 269-272.
76. Zheng, Q.; Chen, H., Development and Applications of Liquid Sample Desorption Electrospray Ionization Mass Spectrometry. *Ann. Rev. Anal. Chem.* **2016**, *9* (1), 411-448.
77. Liu, P.; Zheng, Q.; Dewald, H. D.; Zhou, R.; Chen, H., The study of electrochemistry with ambient mass spectrometry. *TrAC* **2015**, *70*, 20-30.
78. Zhou, F.; Berkel, G. J. V., Electrochemistry Combined Online with Electrospray Mass Spectrometry. *Anal. Chem.* **1995**, *67* (97), 3643-3649.
79. Golub, T.; Becker, J. Y., Electrochemical oxidation of amides of type Ph 2 CHCONHAr. *Org. Biomol. Chem.* **2012**, *10* (19), 3906-3912.
80. Sherer, E. C.; Lee, C. H.; Shpungin, J.; Cuff, J. F.; Da, C.; Ball, R.; Bach, R.; Crespo, A.; Gong, X.; Welch, C. J., Systematic Approach to Conformational Sampling for Assigning Absolute Configuration Using Vibrational Circular Dichroism. *J. Med. Chem.* **2014**, *57* (2), 477-494.
81. Libendi, S. S.; Demizu, Y.; Matsumura, Y.; Onomura, O., High regioselectivity in electrochemical  $\alpha$ -methoxylation of N-protected cyclic amines. *Tetrahedron* **2008**, *64* (18), 3935-3942.

82. Lu, M.; Liu, Y.; Helmy, R.; Martin, G. E.; Dewald, H. D.; Chen, H., Online Investigation of Aqueous-Phase Electrochemical Reactions by Desorption Electrospray Ionization Mass Spectrometry. *J. Am. Soc. Mass Spectrom.* **2015**, *26*, 1676-1685.
83. Cai, Y.; Zheng, Q.; Chen, H.; Liu, Y.; Helmy, R.; Loo, J. A., Integration of electrochemistry with ultra-performance liquid chromatography/mass spectrometry. *Eur. J. Mass. Spectrom.* **2015**, *21*, 341-51.
84. Zheng, Q.; Zhang, H.; Tong, L.; Wu, S.; Chen, H., Cross-Linking Electrochemical Mass Spectrometry for Probing Protein Three-Dimensional Structures. *Anal. Chem.* **2014**, *86*, 8983-8991.
85. Zhao, Y.; Truhlar, D. G., The M06 suite of density functionals for main group thermochemistry, thermochemical kinetics, noncovalent interactions, excited states, and transition elements: two new functionals and systematic testing of four M06-class functionals and 12 other functionals. *Theor. Chem. Acc.* **2008**, *120*, 215-241.
86. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery Jr., J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M. J.; Heyd, J.; Brothers, E. N.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A. P.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, N. J.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.;

- Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. *Gaussian 09*, Gaussian, Inc.: Wallingford, CT, USA, **2009**.
87. Marenich, A. V.; Cramer, C. J.; Truhlar, D. G., Universal Solvation Model Based on Solute Electron Density and on a Continuum Model of the Solvent Defined by the Bulk Dielectric Constant and Atomic Surface Tensions. *J. Chem. Phys. B* **2009**, *113* (18), 6378-6396.