



Single-Cell Transcriptomics of Epstein-Barr Virus and Human Herpesvirus 6 Coinfection

JJ L. Miranda^a

^aDepartment of Biology, Barnard College, Columbia University, New York, New York, USA

ABSTRACT Epstein-Barr virus (EBV) and human herpesvirus 6 (HHV-6) infections are widespread in human populations. Here, I describe single-cell RNA sequencing of two lymphoblastoid cell lines harboring both episomal EBV and inherited chromosomally integrated HHV-6. Rare instances of HHV-6 expression appear enriched with EBV reactivation.

Herpesviruses have ubiquitously spread among humans. Epstein-Barr virus (EBV), a gammaherpesvirus, infects ~90% of the population (1). The circular DNA genome establishes episomal latency (2) to facilitate lifelong persistence. The name “human herpesvirus 6” (HHV-6) collectively refers to two distinct viruses, human betaherpesvirus 6A and human betaherpesvirus 6B (3), which together infect ~70 to 80% of the population (4). HHV-6A and HHV-6B DNA integrates into telomeres (5), and this inherited chromosomally integrated HHV-6 (iciHHV-6) is vertically transmitted to all somatic cells in ~0.1 to 1% of people (6, 7). The widespread prevalence of these viruses predicts frequent coinfections at the cellular level.

Transformation of primary B cells by EBV *in vitro* generates lymphoblastoid cell lines (LCLs) (8) that have been extensively studied as tools for biology research. B cells from some donors contain iciHHV-6 (9). These resulting lines provide a practical opportunity to study the genomics of EBV/HHV6 coinfection.

The EBV-immortalized lines HG00362 and HG01277 (Coriell Institute for Medical Research, Camden, NJ) contain iciHHV-6B and iciHHV-6A, respectively (9). LCLs were cultured under standard conditions (10). Each line was prepared for single-cell RNA sequencing (scRNA-seq) using the Gel bead in EMulsion (GEM) droplet-based system (11) with a Chromium Next GEM Single Cell 3' GEM, Library & Gel Bead Kit v3.1 (10X Genomics, Pleasanton, CA). Sequencing of libraries from HG00362 and HG01277 cells yielded, respectively, 350 and 300 million paired-end reads of 101 nucleotides.

A pipeline optimized for simultaneously measuring viral and human transcripts (12, 13) was modified for the analysis of EBV and HHV-6. Cell Ranger (10X Genomics) was used to demultiplex raw base call files, trim template switching oligo and poly(A) sequences, align reads, and count unique molecular identifiers (UMIs). Custom references contained the genomes of human GRCh38, EBV B95-8 (GenBank accession number [NC_007605.1](#)), and either HHV-6A U1102 ([NC_001664.4](#)) or HHV-6B Z29 ([NC_000898.1](#)). Viral genomes were appended as extra chromosomes.

HHV-6 RNA is enriched with high levels of EBV gene expression (Fig. 1). The majority of cells in both LCLs display a range of EBV transcription across ~4 orders of magnitude. The upper threshold represents reactivating lytic EBV (14, 15). HHV-6 transcription, on the other hand, is rarely detectable in both LCLs. These scRNA-seq results contrast with those of bulk RNA-seq experiments (16), which could not detect any HHV-6 transcripts in LCLs. The HG00362 data set contains 5 cells with HHV-6B RNA. The HG01277 data set contains 3 cells with HHV-6A RNA. UMI counts fall in the 1 to 5 range, most 1 to 2. With only 1 exception, HHV-6 transcription is found coexpressed with high levels of EBV transcription. This is not related to sequencing depth, as HHV-6 expression is not overrepresented with high levels

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2023 Miranda. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

Address correspondence to jj@jlmirandalab.org.

The author declares no conflict of interest.

Received 23 April 2023

Accepted 6 June 2023

Published 20 June 2023

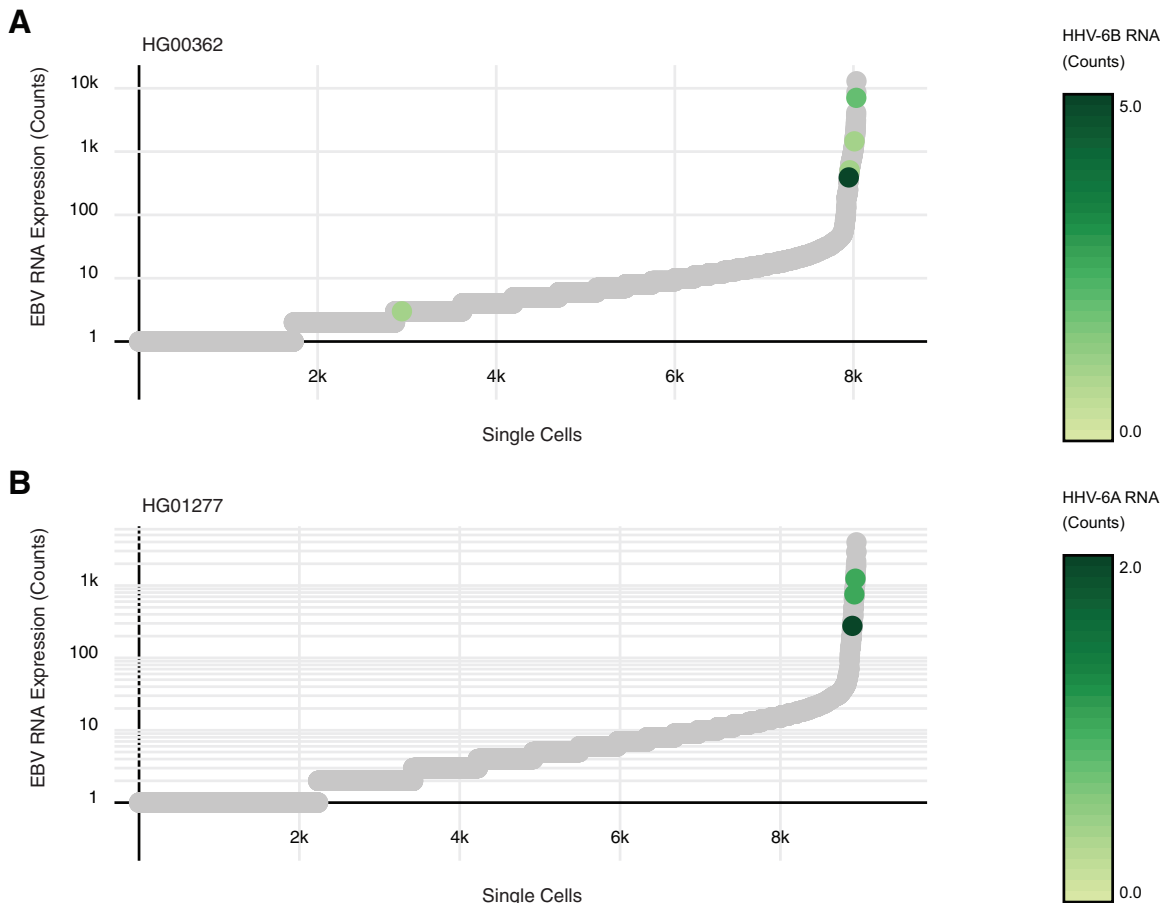


FIG 1 Coexpression of EBV and HHV-6 RNA in single cells. EBV and HHV-6 transcription in the HG00362 (A) and HG01277 (B) LCLs measured by scRNA-seq. Each individual dot on the x axis represents a single cell rank-ordered according to EBV RNA levels. Gene expression was measured by the number of UMI counts per cell. EBV transcription is denoted on a log scale on the y axis. HHV-6 transcription is depicted by green dots, colored as indicated on the linear-scale heatmap. The gray dots represent cells with no detectable HHV-6 transcription.

of human transcription. I call attention to this rare correlation between HHV-6 expression and EBV reactivation while simultaneously encouraging future scRNA-seq studies of EBV/HHV-6 coinfection.

Data availability. These data have been deposited in the NCBI Gene Expression Omnibus (17, 18) under accession number [GSE230239](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE230239), which links to NCBI Sequence Read Archive entries [SRX20032490](https://www.ncbi.nlm.nih.gov/sra/acc.cgi?acc=SRX20032490) and [SRX20032491](https://www.ncbi.nlm.nih.gov/sra/acc.cgi?acc=SRX20032491).

ACKNOWLEDGMENTS

I am grateful to the staff of the J.P. Sulzberger Columbia Genome Center Single Cell Analysis Core (Columbia University Irving Medical Center) for contributing to the scRNA-seq experiment. This material is based upon work supported by the National Science Foundation under award number IOS-2110223. This research was funded in part through the NIH/NCI Cancer Center support grant P30CA013696 and used the Genomics and High Throughput Screening Shared Resource.

REFERENCES

- Balfour HH, Sifakis F, Sliman JA, Knight JA, Schmeling DO, Thomas W. 2013. Age-specific prevalence of Epstein-Barr virus infection among individuals aged 6–19 years in the United States and factors affecting its acquisition. *J Infect Dis* 208:1286–1293. <https://doi.org/10.1093/infdis/jit321>.
- Chiu Y-F, Sugden B. 2016. Epstein-Barr Virus: the path from latent to productive infection. *Annu Rev Virol* 3:359–372. <https://doi.org/10.1146/annurev-virology-110615-042358>.
- Ablashi D, Agut H, Alvarez-Lafuente R, Clark DA, Dewhurst S, DiLuca D, Flamand L, Frenkel N, Gallo R, Gompels UA, Höllsberg P, Jacobson S, Luppi M, Lusso P, Malnati M, Medveczky P, Mori Y, Pellett PE, Pritchett JC, Yamanishi K, Yoshikawa T. 2014. Classification of HHV-6A and HHV-6B as distinct viruses. *Arch Virol* 159:863–870. <https://doi.org/10.1007/s00705-013-1902-5>.
- Okuno T, Takahashi K, Balachandra K, Shiraki K, Yamanishi K, Takahashi M, Baba K. 1989. Seroepidemiology of human herpesvirus 6 infection in normal children and adults. *J Clin Microbiol* 27:651–653. <https://doi.org/10.1128/jcm.27.4.651-653.1989>.
- Arbuckle JH, Medveczky MM, Luka J, Hadley SH, Luegmayr A, Ablashi D, Lund TC, Tolar J, De Meirleir K, Montoya JG, Komaroff AL, Ambros PF,

- Medveczky PG. 2010. The latent human herpesvirus-6A genome specifically integrates in telomeres of human chromosomes in vivo and in vitro. *Proc Natl Acad Sci U S A* 107:5563–5568. <https://doi.org/10.1073/pnas.0913586107>.
6. Leong HN, Tuke PW, Tedder RS, Khanom AB, Eglin RP, Atkinson CE, Ward KN, Griffiths PD, Clark DA. 2007. The prevalence of chromosomally integrated human herpesvirus 6 genomes in the blood of UK blood donors. *J Med Virol* 79:45–51. <https://doi.org/10.1002/jmv.20760>.
 7. Tanaka-Taya K, Sashihara J, Kurahashi H, Amo K, Miyagawa H, Kondo K, Okada S, Yamanishi K. 2004. Human herpesvirus 6 (HHV-6) is transmitted from parent to child in an integrated form and characterization of cases with chromosomally integrated HHV-6 DNA. *J Med Virol* 73:465–473. <https://doi.org/10.1002/jmv.20113>.
 8. Bird AG, McLachlan SM, Britton S. 1981. Cyclosporin A promotes spontaneous outgrowth in vitro of Epstein-Barr virus-induced B-cell lines. *Nature* 289:300–301. <https://doi.org/10.1038/289300a0>.
 9. Telford M, Navarro A, Santpere G. 2018. Whole genome diversity of inherited chromosomally integrated HHV-6 derived from healthy individuals of diverse geographic origin. *Sci Rep* 8:3472. <https://doi.org/10.1038/s41598-018-21645-x>.
 10. Phan AT, Fernandez SG, Somberg JJ, Keck KM, Miranda JL. 2016. Epstein-Barr virus latency type and spontaneous reactivation predict lytic induction levels. *Biochem Biophys Res Commun* 474:71–75. <https://doi.org/10.1016/j.bbrc.2016.04.070>.
 11. Zheng GXY, Terry JM, Belgrader P, Ryvkin P, Bent ZW, Wilson R, Ziraldo SB, Wheeler TD, McDermott GP, Zhu J, Gregory MT, Shuga J, Montesclaros L, Underwood JG, Masquelier DA, Nishimura SY, Schnall-Levin M, Wyatt PW, Hindson CM, Bharadwaj R, Wong A, Ness KD, Beppu LW, Deeg HJ, McFarland C, Loeb KR, Valente WJ, Ericson NG, Stevens EA, Radich JP, Mikkelsen TS, Hindson BJ, Bielas JH. 2017. Massively parallel digital transcriptional profiling of single cells. *Nat Commun* 8:14049. <https://doi.org/10.1038/ncomms14049>.
 12. Rondeau NC, Finlayson MO, Miranda JL. 2020. Widespread traces of lytic Kaposi sarcoma-associated herpesvirus in primary effusion lymphoma at single-cell resolution. *Microbiol Resour Announc* 9:e00851–20. <https://doi.org/10.1128/MRA.00851-20>.
 13. Rondeau NC, Miranda JL. 2021. Rheostat coordination of latent Kaposi sarcoma-associated herpesvirus RNA expression in single cells. *J Virol* 95:e0003221. <https://doi.org/10.1128/JVI.00032-21>.
 14. SoRelle ED, Dai J, Bonglack EN, Heckenberg EM, Zhou JY, Giamberardino SN, Bailey JA, Gregory SG, Chan C, Luftig MA. 2021. Single-cell RNA-seq reveals transcriptomic heterogeneity mediated by host-pathogen dynamics in lymphoblastoid cell lines. *Elife* 10:e62586. <https://doi.org/10.7554/eLife.62586>.
 15. Bristol JA, Brand J, Ohashi M, Eichelberg MR, Casco A, Nelson SE, Hayes M, Romero-Masters JC, Baiu DC, Gumperz JE, Johannsen EC, Dinh HQ, Kenney SC. 2022. Reduced IRF4 expression promotes lytic phenotype in type 2 EBV-infected B cells. *PLoS Pathog* 18:e1010453. <https://doi.org/10.1371/journal.ppat.1010453>.
 16. Hill JA, Ikoma M, Zerr DM, Basom RS, Peddu V, Huang M-L, Hall Sedlak R, Jerome KR, Boeckh M, Barcy S. 2019. RNA sequencing of the in vivo human herpesvirus 6B transcriptome to identify targets for clinical assays distinguishing between latent and active infections. *J Virol* 93:e01419–18. <https://doi.org/10.1128/JVI.01419-18>.
 17. Edgar R, Domrachev M, Lash AE. 2002. Gene Expression Omnibus: NCBI gene expression and hybridization array data repository. *Nucleic Acids Res* 30:207–210. <https://doi.org/10.1093/nar/30.1.207>.
 18. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, Marshall KA, Phillippy KH, Sherman PM, Holko M, Yefanov A, Lee H, Zhang N, Robertson CL, Serova N, Davis S, Soboleva A. 2013. NCBI GEO: archive for functional genomics data sets—update. *Nucleic Acids Res* 41:D991–D995. <https://doi.org/10.1093/nar/gks1193>.