Understanding Local Adaptation to Prepare Populations for Climate Change

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Adaptation within species to local environments is widespread in nature. Better understanding this local adaptation is critical to conserving biodiversity. However, conservation practices can rely on species' trait averages or can broadly assume homogeneity across the range to inform management. Recent methodological advances for studying local adaptation provide the opportunity to fine-tune efforts for managing and conserving species. The implementation of these advances will allow us to better identify populations at greatest risk of decline because of climate change, as well as highlighting possible strategies for improving the likelihood of population persistence amid climate change. In the present article, we review recent advances in the study of local adaptation and highlight ways these tools can be applied in conservation efforts. Cutting-edge tools are available to help better identify and characterize local adaptation. Indeed, increased incorporation of local adaptation in management decisions may help meet the imminent demands of managing species amid a rapidly changing world.

Keywords: adaptation, conservation practice, climate change, genomics, conservation

ocal adaptation (Williams 2018) is pervasive ■ across ecological systems and is a key evolutionary process that has generated much of the world's biodiversity. Local adaptation is the process by which populations have traits that confer higher survival and reproduction in the local environment than they would elsewhere because of the spatial match between adaptive genetic variation and environmental variation (Blanquart et al. 2013). Local adaptation is generally a result of divergent selection over one or more traits, which, if combined with geographical isolation, can lead to different evolutionary trajectories, including reproductive isolation and speciation (White and Butlin 2021). Local adaptation has been widely investigated across the tree of life (e.g., Kawecki and Ebert 2004, Blanquart et al. 2013), and we are rapidly increasing our understanding of the scale and pervasiveness of local adaptation (Sork 2017).

As our understanding of the processes that generate and maintain diversity has rapidly expanded, so has our interest in the consequences of local adaptation (Hereford 2009, Savolainen et al. 2013). One of the most important outcomes of local adaptation may be the maintenance of ecologically important genetic variation, which can be vital for species' persistence amid changing ecological conditions (Whitlock 2015). However, much of the spatial variability

in adaptive genetic diversity generated by local adaptation may be threatened by the multifaceted effects of global and anthropogenic change. In addition, adaptation to nonclimatic factors, such as other abiotic and biotic interactions (e.g., soil and water characteristics, diseases, food resources, predation, and competition), may be disrupted by climate change (Delph 2017). A shift from local adaptation to maladaptation (i.e., reduced fitness and survival of individuals, given a mismatch between genetically determined traits and the current environment) under fast-changing environments likely contributes to staggering losses of populations worldwide (Derry et al. 2019). Therefore, an improved understanding of local adaptation can help guide conservation actions to better conserve the world's biodiversity (e.g., figure 1). One caveat in this discussion of local adaptation and conservation, however, is that in extremely small populations, the larger forces of genetic drift and mutational load may prevent local adaptation from occurring (Willi et al. 2022). Therefore, managers in such situations may find more benefit from focusing on increasing overall genetic diversity and census size (Willi et al. 2022). Information on local adaptation, however, can provide valuable insights for improving the adaptive potential of populations that are not on the very brink of extinction, to maximize their likelihood



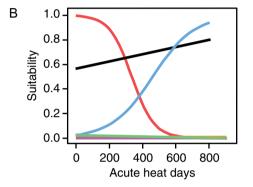


Figure 1. (a) The importance of incorporating local adaptation in conservation planning is highlighted by research on the American pika (Ochotona princeps). A juvenile pika from Darkhorse Creek Canyon, Beaverhead Mountains in the central-northern Rocky Mountain chain, along the Montana-Idaho border. Photograph: Peter Billman, Montana State University. (b) Different populations show different responses to the same range of values in the same climatic aspect. The plot shows the marginal response of each lineage to acute heat days, ignoring the effect of all other variables. Specifically, the increasing frequency of hotter days is beneficial to pikas in the northern Rocky Mountains (the blue line); it strongly decreases habitat suitability for pikas in the Sierra Nevada (the red line) and has no or nearly no effect on pikas in the three other genetic lineages (the green, orange, and purple lines). The composite model (the black line) masks this intraspecific diversity in how populations respond heterogeneously to climate. Output is from a LARS (least angle regression) model calibrated with an elastic net to obtain the optimal level of regularization. By design, the coefficients are biased toward 0 (which is from an unpublished analysis associated with Smith et al. 2019).

of long-term persistence (Bay et al. 2018, Flanagan et al. 2018). In addition, using local adaptation to inform translocations, reintroductions, captive breeding, and identification of conservation units and critical habitat may substantially increase the likelihood of successful outcomes (Flanagan et al. 2018). Despite this importance, relatively few studies

have incorporated local adaptation into predictions of how species' distributions and abundances will be affected by climate change (Smith et al. 2019) or have taken local adaptation into account when designing conservation and recovery plans (Peterson et al. 2019).

The time is ripe to evaluate local adaptation as a standard component of conservation planning and integrate it into conservation actions (Hellmann and Pfrender 2011). In the present article, our objectives are to provide a primer on local adaptation and its potential incorporation in conservation by highlighting the ways conservation biologists can merge classic and emerging tools to detect local adaptation, by exemplifying methods for incorporating local adaptation in conservation actions to better help populations withstand changing climate and other ecological stressors, and by describing the implications and applications of local adaptation for policy, management, and conservation and future directions for promoting the strategic incorporation of local adaptation into conservation decision-making (see box 1 for a glossary of terms used throughout). We acknowledge that numerous other ecological disturbances (e.g., land-use change, overharvest, pollution, disease) have all contributed to losses of biodiversity (Díaz et al. 2019). Given this, we primarily focus in the present article on climate change because of its iteratively increasing pace, near ubiquity, interaction with the abovementioned factors, and clear influence on fitness and population persistence.

Integrating classic and emerging tools to characterize and quantify local adaptation

Common garden experiments and reciprocal transplants have been the gold standard for identifying local adaptation and separating genetic contributions from phenotypic plasticity. However, these studies tend to be biased toward abundant, well studied, easy to translocate species, and parsing traits with complex genetic architecture requires using many study individuals (Ghalambor et al. 2018). Until recently, this limited the ability to incorporate local adaptation in conservation because species of conservation interest are generally understudied and rare, and many traits of conservation interest are likely controlled by a suite of genes, rather than by single genes of strong effect (Savolainen et al. 2013).

The conservation toolbox is now rapidly expanding as new technologies emerge for studying genomes (i.e., the entirety of an organism's genetic material) and as our understanding of the genomic basis for traits increases. We now have the tools to identify and incorporate local adaptation into conservation and management by integrating cutting-edge genomic data with traditional methods of studying local adaptation, such as common garden experiments (Hoban et al. 2016). We can generate high-resolution genomic data for almost any species (Whitlock 2015). Furthermore, many of these methods do not necessitate rearing or growing organisms in the lab, such as species distribution modeling and landscape genetics, making an understanding of the causes and consequences of local adaptation possible for

Box 1. Glossary.

Adaptive capacity: the ability of a species or its populations to cope with or accommodate a given change (e.g., climate change) by persisting in situ or shifting to more-suitable ranges or microhabitats

Adaptive potential: the ability of a species or population to adapt via evolutionary change

Assisted gene flow: intentional translocation of individuals to facilitate adaptation to anticipated local conditions

Common garden experiment: a method of assessing differential responses to environmental variables by placing two populations into the same environment. Variations of this include reciprocal transplants and provenance plots.

Genetic rescue: an increase in population fitness caused by gene flow inferred from some demographic vital rate or phenotypic trait, by more than can be attributed to the demographic contribution of immigrants

Inbreeding depression: reduced fitness of individuals with related parents due to lower survival or reproduction

Maladaptation: Suboptimal fitness, either in absolute terms or relative to another population

Outbreeding depression: reduced fitness (compared to the parental fitness) caused by crossing distantly related individuals

Phenotypic plasticity: in which a single genotype produces multiple phenotypes in response to environmental changes

Reciprocal transplant: a method of evaluating local adaptation by swapping individuals among locations and comparing their fitness in their home versus in foreign environments. This method requires local and nonlocal individuals be tested in each habitat of interest, whereas provenance plots do not.

most organisms of conservation importance (e.g., figure 2; Funk et al. 2019, Campbell-Staton et al. 2020). We can identify genotypes that are locally adapted, determine the environmental variables driving population differentiation, and pinpoint the genetic variation for specific traits underlying local adaptation, making it possible to then use that information to make predictions for future habitat suitability and guide conservation and restoration actions (Tiffin and Ross-Ibarra 2017). Increasing our understanding of the causes and consequences of local adaptation in species of conservation focus will allow conservation biologists to better understand which populations may be limited in their adaptive potential and how phenotypes and fitness will respond to climate heterogeneity. Ultimately, this information can be used for improved conservation planning. See supplemental table S1 for further exploration of these methods.

Using local adaptation to best equip populations for persistence under changing climates

Given the recent advances in our ability to identify local adaptation, it is now possible to incorporate local adaptation into species conservation and management to plan for future conditions under climate change (Capblancq et al. 2020). Managed gene flow and the incorporation of local adaptation in relocation and reintroduction planning each provide options for throwing a lifeline to populations that have become increasingly maladapted to their environment because of reduced genetic diversity or rapidly changing environments or to restore resilient populations in places where they have become locally extinct. Cryopreservation of gametes and gene editing are two additional technological advances that, if they are informed by an improved understanding of local adaptation, may provide options for saving species from the brink of extinction and allow a path for protecting species that are most vulnerable to the negative

effects of global change. Below, we discuss the benefits and concerns regarding these options and highlight how the understanding of local adaptation can inform these important conservation tools. We are not advocating that these tools should replace more classic conservation strategies, such as land and species protection and take prohibitions, which will continue to constitute pivotal approaches for biodiversity conservation. Rather, we aim to demonstrate that we also now have new tools to combat biodiversity loss.

Managed gene flow and translocations or reintroductions. Translocation, the human-assisted movement of individuals, has a long history of use in managing populations for conservation objectives (Griffith et al. 1989). Today, it can also be a potentially powerful management strategy for mitigating maladaptation due to rapidly changing climate conditions (Hoffmann et al. 2021). This can occur through the intentional translocation of individuals within a species' range to reintroduce populations that have gone locally extinct or to facilitate gene flow of locally adaptive alleles into existing populations. In addition, locally adapted individuals can be translocated to areas outside their current or historical range that are projected to provide suitable habitat under future climatic conditions on the basis of assessments of adaptive capacity (for a review of actions, see Thurman et al. 2022).

More specifically, translocation is an option for assisted gene flow to benefit maladapted populations and promote increased local adaptation through a number of nonmutually exclusive mechanisms (Aitken and Whitlock 2013). For example, immigrant genotypes can be sourced from an environment that matches projected future climate conditions, such that "preadapted" alleles are used for translocation or introduction into a maladapted recipient population (Catullo et al. 2019, Chen et al. 2022). These alleles may be locally

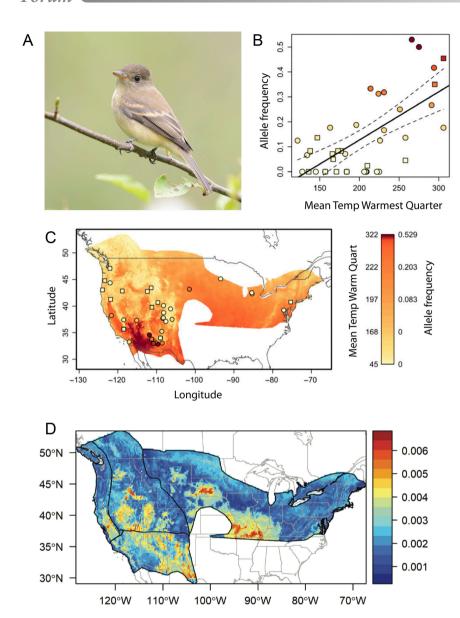


Figure 2. (a) Candidate single nucleotide polymorphisms (SNPs) appear tightly linked to temperature in the willow flycatcher; the southwestern willow flycatcher has been federally designated as endangered since 1995 (only an estimated 900-1100 pairs existed, as of 2002, USFWS 1995, 2013). Photograph: (a) V. J. Anderson, CC BY-SA 4.0 (https://creativecommons.org/licenses/by-sa/4.0), via Wikimedia Commons. (b) As mean temperature of the warmest quarter increases, so does the allele frequency of the temperature-associated SNP. The allele frequencies from the original genome scan data are indicated by squares, whereas allele frequencies based on the validation set are denoted by circles. The highly significant relationship between this SNP and 7 of the eight top-ranked climate variables in both the genome scan and validation results suggests a potential role for the SNP in climate adaptation across the region. (c) The geographically explicit representation of panel (b), showing the association between mean temperature of the warmest quarter (BIO10) and the SNP allele frequency across western North America; population allele frequencies are color-coded from high frequency (red) to low (yellow). (d) Map showing the genomic vulnerability (amount of mismatch between current and predicted future genotype-environment relationships) across the willow flycatcher breeding range; red, high genomic vulnerability; blue, low genomic vulnerability. Source: Reprinted from Ruegg and colleagues (2018).

adapted to these future conditions. This is most often discussed for long-lived and dispersal-limited species that are unlikely to otherwise keep pace with changes in local climate, such as trees (Browne et al. 2019) or corals (Bay and Palumbi 2014). A large body of literature also now supports the idea that gene flow into populations that have become small and isolated because of habitat fragmentation can facilitate genetic rescue-an increase in population growth due to the immigration of new genetic material (see box 2). Finally, over a longer time frame, increased genetic variation provided by assisted gene flow may be a critical source of raw genetic material that allows populations to adapt to novel selection pressures (Derry et al. 2019).

Although assisted gene flow and movement of individuals have many potential benefits, they also pose a number of possible risks. Outbreeding depression can occur if immigrants are too distantly related (particularly if they have differences in chromosomal structure or number) or if they introduce traits that are not well suited for the new environment (Frankham et al. 2011, Leroy et al. 2018). For example, reduced hybrid survival was observed when geographically separated salmon populations with distinct spawning phenotypes (even- versus odd-year spawn timing) were crossed (Gilk et al. 2004), highlighting the critical importance of a strong understanding of organismal natural history before carrying out assisted gene flow. Another concern is the loss of local genetic lineages through genetic swamping (Rhymer and Simberloff 1996, Gilk et al. 2004). This may occur if too many individuals with higher fitness than local individuals are introduced. It has also been shown that strong selection can maintain locally favored alleles, even in the face of high gene flow (Fitzpatrick et al. 2020). Although the benefits of assisted gene flow and genetic rescue within a species' range are increasingly recognized, movement of individuals to areas outside of the historical range of a species should be treated cautiously (Schwartz et al. 2012). Therefore, building on the lessons from empirical studies, demographic modeling and simulations

Box 2. Does genetic rescue constrain or facilitate local adaptation?

Human-assisted gene flow resulting in genetic rescue has aided the recovery of several iconic species such as Florida panthers (Johnson et al. 2010), bighorn sheep (Hogg et al. 2006), and Australian marsupials (Weeks et al. 2017). However, despite increased evidence for its potential to reverse population declines in small and isolated populations, genetic rescue is not a widely used management strategy (Ralls et al. 2018). Genetic rescue is primarily thought to increase fitness through masking of deleterious alleles that become exposed in small and inbred populations. However, the genetic architecture underlying genetic rescue is rarely known. In addition to masking deleterious alleles, beneficial genetic variation introduced by gene flow could rescue populations through heterozygote advantage, adaptive evolution, or a combination of these processes (Scott et al. 2020).

One of the main concerns with gene-flow augmentation in management is the possibility that gene flow may constrain local adaptation (Edmands 2006). This concern warrants re-evaluation in the context of overwhelming evidence for inbreeding depression when populations become isolated and are rapidly reduced in size, increasing evidence that gene flow increases fitness under a wider set of conditions than what theory predicts (Whiteley et al. 2015), and the extent of maladaptation that many natural populations now face, given the pace of environmental change (Hoffmann and Sgrò 2011). In fact, gene flow may actually facilitate local adaptation, especially in cases in which populations lack the variation needed to respond to selection.

Recent empirical work supports the idea that moderate rates of gene flow, even from adaptively differentiated source populations, can maintain or even facilitate adaptation within recipient populations (figure 3). For example, experiments using Trinidadian guppies in the wild and in mesocosms showed increases in population growth without the loss of locally adapted traits (Fitzpatrick et al. 2016, Kronenberger et al. 2018). In fact, gene flow caused shifts in some traits in the predicted adaptive direction (Fitzpatrick et al. 2017). In addition, greenhouse crosses using the annual wildflower Clarkia pulchella found strongest benefits of gene flow during an anomalously warm year, highlighting the potential role of gene flow in aiding adaptation to warming climates (Bontrager and Angert 2019). As natural populations become fragmented and exposed to severe environmental stressors, gene flow may be an increasingly important source of variation necessary for persistence.

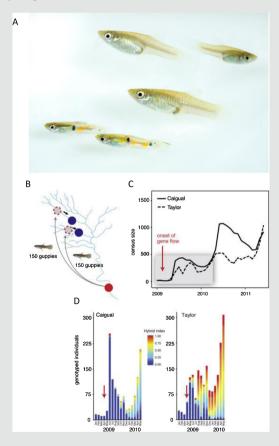


Figure 3. (a) Trinidad guppies. Photograph: David Herasimtschuk. (b) Populations have benefitted from gene flow from adaptively differentiated source populations. The gray arrows indicate initial translocations. The black arrows indicate the direction of gene flow. (c) Work by Fitzpatrick and colleagues (2020) showed increases in census size after gene flow in both populations. (d) Incorporation of new genotypes from translocation is shown by the increase in hybrid index in each population over time.

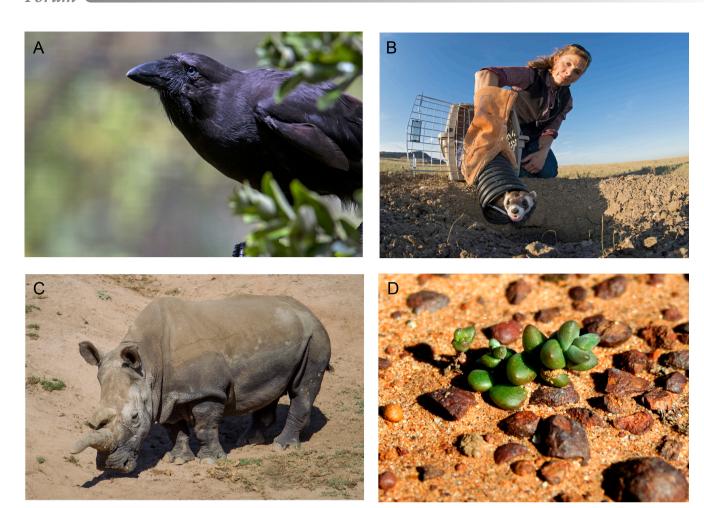


Figure 4. Organisms that have been cryopreserved in order to save genetic material and aid conservation efforts. (a) Semen from male Hawaiian' alalā is preserved by the Maui Bird Conservation Center to bank genetic diversity for the recovery effort. Photograph: San Diego Zoo Wildlife Alliance. (b) Black-footed ferret: World Wildlife Fund Northern Great Plains biologist and black-footed ferret expert Kristy Bly prepares to release a black-footed ferret at Snake Butte on the Fort Belknap Reservation, northern Montana. Photograph: Clay Bolt/WWF-US. (c) San Diego Zoo Global has cells from 12 individual northern white rhinos cryopreserved in the Frozen Zoo to help recover the species. Photograph: San Diego Zoo Wildlife Alliance. (d) Endangered Dudleya brevifolia seed is stored at the San Diego Zoo Wildlife Alliance Native Plant Seed Bank. Photograph: Matthew Luskin, wikimedia.

are increasingly used as powerful tools to both forecast the interaction of local adaptation with demographic, biotic, and abiotic factors (Landguth et al. 2020) and predict the spatial distribution of adaptive genetic variants and therefore determine appropriate current and future geographic areas for translocations (Razgour et al. 2019, Rochat et al. 2021). Simulations of assisted gene flow have shown that multiple translocations over several generations can lead to smaller reductions of fitness, especially if outbreeding depression is strong (Grummer et al. 2022). However, when traits are polygenic (i.e., controlled by multiple loci of small effect), the beneficial effects of assisted gene flow may only be tangible on a longer time scale than that necessary for most climate-related actions (Grummer et al. 2022). Given that the stage and evolutionary play are iteratively changing as was noted by G. Evelyn Hutchinson, conservation practitioners

must also ensure that viability of source populations is not compromised by assisted migration. These concerns deserve increased research to continue to increase the applicability of this important conservation tool (Fitzpatrick et al. 2020, Brodie et al. 2021).

Cryopreservation and seed banking. Another emerging tool that can be used to help support incorporation of local adaptation in conservation is seed banking and the cryopreservation (freezing) of gametes, embryos, or somatic cells (figure 4). These methods have been increasingly implemented in species for which *in situ* conservation methods have failed to maintain viable populations (Holt et al. 1996, Ryder and Onuma 2018). There are around 1300 seed banks worldwide, and ecologists are gaining an understanding of how best to design and sample populations to increase

the utility of seed banks for promoting local adaptation (Rajasekharan 2015, Hoban 2019). In addition, there have been rapid advances in cryopreservation techniques that allow maintenance of genetic material for animals and plants (e.g., tropical species) whose seed can't be stored (e.g., maintenance of tissue culture stocks; Ryder and Onuma 2018). Cryopreserved cells can theoretically be stored indefinitely and can serve as backups to preserve the genotypes of individuals. When combined with *in vitro* fertilization techniques, this approach can produce new offspring (Charlton et al. 2018, Ryder and Onuma 2018), which may then be used in genetic rescue and reintroduction efforts.

Seed banks and cryopreserved cells not only serve as genetic resource banks to conserve genetic diversity overall in an imperiled species but can also preserve locally adapted genotypes that are important for future adaptive potential (Holt et al. 1996). In black-footed ferrets, a species reliant on ex situ conservation and whose extremely low population sizes have led to strong inbreeding effects, the use of frozen spermatozoa and artificial-insemination techniques increased genetic diversity and lowered measures of inbreeding in released individuals (Howard et al. 2016). This approach can be combined with genomic analyses to identify and target genetic variants that are, or will be, important for promoting local adaptation. The northern white rhinoceros is extinct in the wild, but cells from multiple individuals have been cryopreserved. These cells have been tested not only for measures of genetic diversity but also for signatures of selection that may reflect locally adapted genotypes (Tunstall et al. 2018). These data can be used to prioritize cells that will lead to offspring with the greatest chance of success in the reintroduction effort (Howard et al. 2016, Tunstall et al. 2018). This approach was recently proven successful by cloning an individual black-footed ferret from cells that had been cryopreserved for over 30 years (USFWS 2021a). Promoting local adaptation amid ongoing global change via cryopreservation might specifically include ensuring that individuals from disjunct populations are well represented; including individuals from numerous edges of the bioclimatic envelope (or that have wide moisture and temperature tolerances), particularly from trailingedge populations; preserving populations that naturally experienced higher frequencies of extreme events; including individuals from transition zones; and including individuals from more populations of poorly dispersing species associated with past climates, such as cryosphere-associated species (Thurman et al. 2022). However, it will be important to also ensure any of these targets of cryopreservation also come from populations with high levels of overall genomic diversity to avoid the negative effects of bottlenecks.

Cryopreservation, however, requires technical knowledge from specialist veterinarians and regular maintenance at designated facilities. In addition, there is significant variation among species in the viability of frozen gametes for fertilization (Charlton et al. 2018, Ryder and Onuma 2018). Cryopreservation has also been shown to have negative

effects on the morphology of some species, including effects that can reduce the survivorship and reproductive output of individuals (Poo and Hinkson 2020). Nonetheless, seed banks and frozen zoos may be the last option for the future of some species, and research on how to improve seed banking and cryopreservation techniques to support future adaptation seems poised to provide invaluable insights.

Gene editing. Gene-editing technology has also emerged as a potentially viable tool for conservation biology in recent years, especially with the explosion of versatile CRISPR gene-editing techniques (Phelps et al. 2020). An example of gene-editing utility for conservation can be found in the 'i'iwi (Hawaiian honeycreeper). Simulation models of 'i'iwi reintroductions after devastating population loss driven by nonnative malaria suggest that the species has a much higher chance of success if genetically edited, malariaresistant individuals were to be introduced (Samuel et al. 2020). Although the 'i'iwi has not yet been gene edited, this study demonstrates the strong potential for this tool to assist conservation efforts if a gene for malaria resistance can be identified. This solution is especially important because future warming is projected to increase the rate of malaria-driven extinctions of other species of Hawaiian birds, which may require analogous conservation efforts (Paxton et al. 2016). Similar gene-editing schemes have been proposed for other conservation scenarios, such as introducing genes for temperature resistance in coral species, fungal-disease resistance in bats and amphibians, and plague resistance in black-footed ferrets (Piaggio et al. 2017). As gene-editing tools become more accessible and the ability to identify regions of the genome associated with adaptive traits becomes more precise, these technologies will become a more feasible part of the conservation toolkit (Supple and Shapiro 2018, Derry et al. 2019). For example, it is not outside the realm of possibility that, one day, we could identify adaptive genetic variation for thermal tolerance in an organism, using the techniques outlined in the table S1, and then introduce this variation into populations that will likely need it under future warming but are currently lacking it using gene-editing techniques (Thomas et al. 2013). This approach could have the added benefit of preserving the unique, local genetic diversity present in the population while introducing important alleles for future persistence. However, as with all the abovementioned conservation actions, it will be important and nontrivial for sufficient overall genetic variation to be produced and maintained in any gene-edited population to avoid inbreeding and promote long-term persistence and resilience. Local adaptation-oriented gene editing might specifically include seeking conservation of allelic diversity at multiple spatiotemporal scales, seeking maintenance of the evolutionary processes and pathways that maintain local adaptation, and balancing concerns about swamping and hybrid vigor with desire to facilitate population persistence in a warmer, drier, or more erratic future climate. In addition, any use of genome editing in a conservation context

would benefit from being implemented in an adaptive management framework, with sufficient data collection before, during, and after actions, to ensure learning and the possibility for future refinement of techniques.

Gene editing also has limitations and requires ethical considerations and community engagement before being implemented as a regular conservation tool (Kardos and Shafer 2018, Barnhill-Dilling and Delborne 2021). Geneediting technologies have progressed rapidly but still depend on the ability to accurately identify regions of the genome that are adaptive and loci that have large effects on the trait of interest (Phelps et al. 2020). This approach is unlikely to work well for traits that are determined by many loci of small effect. In addition, there is some public resistance to the use of gene-editing technologies for conservation purposes, and many people remain skeptical that the techniques are safe or ethical (Kohl et al. 2019). Therefore, it is important to engage with multiple stakeholders when considering genetically modifying organisms for conservation purposes (Kofler et al. 2018). Ethical analyses of other genomic technologies, such as cloning, have been conducted and could be adapted to evaluate gene editing as well (Sandler et al. 2021). In addition, we presume that specific policies around the use of gene editing for conservation will be developed iteratively over time (Burgiel et al. 2021). Although the practical and ethical considerations of gene editing should be carefully weighed, this technology may constitute the last chance for species when all other conservation efforts have failed. Conservation biologists may benefit from increasing engagement with these emerging technologies to reduce future biodiversity loss and plan for future climatic conditions (Piaggio et al. 2017).

Implications and applications for policy, management, and conservation

Local adaptation has profound implications for a wide array of conservation actions, as was described above. However, the full incorporation of local adaptation into the conservation toolbox will require both emphasizing the identification of local adaptation and having policies that promote the incorporation of local adaptation in conservation planning (Hällfors et al. 2016). This includes having flexibility in the legal and regulatory mechanisms for preserving biodiversity under future climate conditions while also providing clear directives. The European Union's Birds and Habitats Directive, for instance, provides regulatory agencies numerous options for planning and implementing actions to assist natural populations with adaptation to climate change. This flexibility, however, also has its limitations, because it does not require any specific actions or protections; consequently, planning for climate change is possible but voluntary (Verschuuren 2010).

Genomic advances are reshaping how we identify conservation units. There is ongoing vigorous debate among scientists and practitioners regarding how to weigh patterns shown in a small number of genes versus genome-wide patterns in conservation decisions and policy (Kardos et al. 2021, Teixeira and Huber 2021, Waples et al. 2022). For example, genomic research has recently identified one genomic region (GREB1L to ROCK1) as being tightly linked with migration timing, an important life-history trait driven by local adaptation, in Pacific salmon (Prince et al. 2017, Waples and Lindley 2018). Researchers have demonstrated dramatic reductions in diversity at this locus after anthropogenic habitat modification, potentially reducing fitness and population sustainability (Thompson et al. 2019). Other components of fitness in salmonids, however, including growth rate and age at maturity, appear to be less homogeneous in their genetic basis (Waters et al. 2018). Consequently, it can be challenging to determine how much weight should be given to single genomic regions versus genome-wide patterns when designing conservation actions, particularly given that overall genomic diversity will provide the building blocks for future adaptation (DeWoody et al. 2021, García-Dorado and Caballero 2021).

Assisted gene flow and assisted colonization, as was discussed above, are potentially powerful methods for helping dispersal-limited species to cope with climate change and increase overall genetic diversity (Williams et al. 2021). However, for assisted gene flow and assisted colonization to become effective policy, clear protocols and benchmarks will be needed to guide when and how translocations can occur (Schwartz et al. 2012). Translocations triggered by clear population declines, a high likelihood of future declines, or the loss of ecosystem function are consistent with the preventive principle (addressing known ongoing declines in threatened species) and precautionary principle (addressing anticipated or hypothetical future declines) outlined by the Rio Declaration on Environment and Development (Sansilvestri et al. 2015).

In addition to directly assisting movement, another critical component of conservation planning relevant to local adaptation is reserve design that helps assist movement through human-dominated habitats, assuming that organisms (and their associated traits) can disperse sufficiently quickly (Kostyack et al. 2011). Reserves can be designed to both promote movement between important habitats and to conserve the highest number of adaptively differentiated populations. Reserve design and conservation planning can combine methods for identifying and mapping out local adaptation (outlined above) with spatial analyses of climate refugia (see Michalak et al. 2020, Stralberg et al. 2020, Saunders et al. 2023) to prioritize areas for protection. For example, Saunders and colleauges (2023) conduct a reserveselection prioritization to highlight priority areas for reserve design that both complement the current protected area network and would protect climate refugia for the highest number of taxonomic groups. One could expand on this analysis further to ensure that adaptively differentiated populations within species are represented across the reserve network. Such combinations would improve our ability to conserve

adaptive variation that will be important for species persistence now and into the future.

Under national laws aimed at species conservation, such as the US Endangered Species Act, addressing extant conventional threats has arguably often garnered higher priority than preemptive management in the face of climate change (Delach et al. 2019). Nonetheless, the US Fish and Wildlife Service is evaluating (as of November 2021) the use of techniques such as translocations, reintroductions, genetic supplementation of in situ populations and the use of captive breeding to create ex situ insurance populations and head starting offspring for numerous species whose range has become markedly more fragmented than the historical range and for which local adaptation was used to help delineate conservation or recovery units (e.g., the rusty-patched bumble bee, USFWS 2021b; the eastern massasauga rattlesnake, USFWS 2021c; the Dakota skipper, USFWS 2019). Such actions seek to achieve persistence of locally adapted populations across diverse selective regimes, higher genetic diversity, avoidance of inbreeding depression, and long-term persistence amid increasingly stochastic environmental conditions. Analogously, given declining population sizes and declining heterozygosity and after considering risks of genetic swamping and loss of local adaptation, translocations of numerous mountain pygmy possum (an endangered Australian marsupial) individuals from a genetically diverged population achieved genetic rescue and demographic recovery of one of the three remaining extant populations of the species (Weeks et al. 2017).

The IUCN Guidelines for Reintroductions and Other Conservation Translocations (IUCN 2013) can be used for developing national and more localized guidance for using these tools amid climate change. The guidelines acknowledge the importance of information on adaptations to local ecological conditions, but many other factors also warrant attention when considering human-assisted movement of individuals for conservation. Similarly, Weeks and colleagues (2011) provided pragmatic decision trees to evaluate possible translocations (see figure 2, supplemental table S2), suggesting that local adaptation would constitute one of seven suites of factors governing the likelihood of both resilience and persistence of translocated populations. Determining whether and how translocated individuals will be moved may require modification of existing laws or policies. For example, the Endangered Species Act provisions governing the movement of endangered or threatened plants are currently much less restrictive than those for animals and may warrant new or revised guidelines or policy to be able to apply more broadly to animals (Shirey and Lamberti 2010). Aitken and Whitlock (2013) argued in their incisive review that to robustly weigh the risks of translocations (i.e., outbreeding depression and disrupting local adaptation to nonclimatic factors) against their ability to mitigate maladaptation due to climate change, both pattern of gene flow and extent of local adaptation need to be known.

Looking forward, conservation actions necessitating and enabling the movement of individuals and genes because of climate change-induced shifts in suitable habitat will at times require collaboration among multiple governmental and nongovernmental organizations and, in many, cases across national boundaries. Current transboundary agreements for climate change often fail to accommodate this reality (Trouwborst 2012). A logical early step for transboundary collaborations would involve synthesizing data gathered by scientists and managers across species' ranges to identify patterns of local adaptation. This approach can help in assessing potential benefits and risks of assisted migration or colonization across borders to reduce extinction risk, approaching reserve design at a continental scale, identifying alleles associated with higher fitness under different climatic conditions, and determining populations for which seed banking or cryopreservation ought to be given high priority to preserve options for future conservation efforts. Organizations such as zoos, botanical gardens, and biobanks involved in cryopreservation or captive propagation of threatened species constitute key partners in coordinating efforts and providing genetic material or individuals for translocations. In addition, the resources required to prepare populations for climate-change threats will be much greater than the resources available to meet the need for all taxa threatened by climate change. Consequently, conservation practitioners will necessarily have to prioritize actions and employ cost-benefit analyses to inform when to apply these tools. Collaborations across the researcher-practitioner-policy spectrum may be beneficial to help managers identify the best combination of strategies, tools, and resources for adaptively managing species with local adaptation in mind.

Conclusions

We now have the technical capabilities to meaningfully integrate local adaptation into conservation planning. Although taking a local adaptation-focused conservation approach may necessitate greater initial investment in funds and human resources, we believe taking this approach will often have large returns on investment in terms of higher mean fitness, long-term sustainability, and greater effectiveness in buffering against the negative effects of climate change. Thorough and nuanced understanding of local adaptation is increasingly pivotal to inform the climate-adaptation conservation and management actions described in the present article, such as assisted migration, seed sourcing for restoration, active management of fragmented metapopulations and low-dispersal species, identification of conservation units, reserve design, and more-accurate predictions of climate-mediated range shifts. Indeed, local adaptation tools and approaches may provide the best last-ditch option for some species, such as those with long generation times, or limited and infrequent gene flow. We acknowledge, however, that local adaptation-relevant data are lacking for many species, including climate-sensitive species and despite massive

advances in tools, data, and analytical approaches, numerous research frontiers remain. The climatic factors and mechanisms involved, critical life stages, and population dynamics most strongly affecting persistence can be unknown, for parts or all of a species' range. In such cases, prioritizing the collection of such data could reap large rewards, and it is our hope that the information provided in supplemental table S1 can help in that data generation process. Ultimately, land and wildlife managers and other conservation practitioners must weave local adaptation information into an amalgamation of climate-change vulnerability involving not only adaptive capacity but also sensitivity, exposure, and anthropogenic disturbances. Indeed, a recent conceptual framework suggested that a primary pathway for reducing a species' adaptive capacity (including its evolutionary potential to create local adaptation) from its fundamental to realized level was mediated by extrinsic factors such as pollution, habitat fragmentation, and invasive species (see figure 1 of Beever et al. 2015). However, by thoroughly considering local adaptation, conservation practitioners will be better equipped to incorporate the myriad factors involved and prepare populations for long-term persistence under a changing climate.

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Supplemental material

Supplemental data are available at BIOSCI online.

References cited

- Aitken SN, Whitlock MC. 2013. Assisted gene flow to facilitate local adaptation to climate change. Annual Review of Ecology, Evolution, and Systematics 44: 367–388.
- Barnhill-Dilling SK, Delborne JA. 2021, Whose intentions? What consequences? Interrogating "Intended consequences" for conservation with environmental biotechnology. Conservation Science and Practice 3: e406
- Bay RA, Harrigan RJ, Underwood VLe, Gibbs HL, Smith TB, Ruegg K. 2018. Genomic signals of selection predict climate-driven population declines in a migratory bird. Science 359: 83–86.
- Bay RA, Palumbi SR. 2014. Multilocus adaptation associated with heat resistance in reef-building corals. Current Biology 24: 2952–2956.
- Beever EA, et al. 2015. Improving conservation outcomes with a new paradigm for understanding species' fundamental and realized adaptive capacity. Conservation Letters 9: 131–137.
- Blanquart F, Kaltz O, Nuismer SL, Gandon S. 2013. A practical guide to measuring local adaptation. Ecology Letters 16:1195–1205.
- Bontrager M, Angert AL. 2019. Gene flow improves fitness at a range edge under climate change. Evolution Letters 3: 55–68.
- Brodie JF, Lieberman S, Moehrenschlager A, Redford KH, Rodr\u00e4Guez JP, Schwartz M, Seddon PJ, Watson JEM. 2021. Global policy for assisted colonization of species. Science 372: 456–458.
- Browne L, Wright JW, Fitz-Gibbon S, Gugger PF, Sork VL. 2019. Adaptational lag to temperature in valley oak can be mitigated by genome-informed

- assisted gene flow. Proceedings of the National Academy of Sciences 116: 25179–25185.
- Burgiel SW, Baumgartner B, Brister E, Fisher J, Gordon DR, Novak B, Palmer MJ, Seddon PJ, Weber M. 2021. Exploring the intersections of governance, constituencies, and risk in genetic interventions. Conservation Science and Practice 3: e380.
- Campbell-Staton SC, Winchell KM, Rochette NC, Fredette J, Maayan I, Schweizer RM, Catchen J. 2020. Parallel selection on thermal physiology facilitates repeated adaptation of city lizards to urban heat islands. Nature Ecology and Evolution 4: 652–658.
- Capblancq T, Fitzpatrick MC, Bay RA, Exposito-Alonso M, Keller SR. 2020. Genomic prediction of (mal)adaptation across current and future climatic landscapes. Annual Review of Ecology, Evolution, and Systematics 51: 245–269.
- Catullo RA, Llewelyn J, Phillips BL, Moritz CC. 2019. The potential for rapid evolution under anthropogenic climate change. Current Biology 29: R996–R1007.
- Charlton SJ, Nielsen MB, Pedersen CR, Thomsen L, Kristjansen MP, Sørensen TB, Pertoldi C, Strand J. 2018. Strong heterogeneity in advances in cryopreservation techniques in the mammalian orders. Zoological Science 35: 1–22.
- Chen Z, Grossfurthner L, Loxterman JL, Masingale J, Richardson BA, Seaborn T, Smith B, Waits LP, Narum SR. 2022. Applying genomics in assisted migration under climate change: Framework, empirical applications, and case studies. Evolutionary Applications 15: 3–21.
- Delach A, Caldas A, Edson KM, Krehbiel R, Murray S, Theoharides KA, Vorhees LJ, Malcom JW, Salvo MN, Miller JRB. 2019. Agency plans are inadequate to conserve US endangered species under climate change. Nature Climate Change 9: 999–1004.
- Delph LF. 2017. The study of local adaptation: A thriving field of research. Journal of Heredity 109: 1–2.
- Derry AM, et al. 2019. Conservation through the lens of (mal)adaptation: Concepts and meta-analysis. Evolutionary Applications 12: 1287–1304.
- Dewoody JA, Harder AM, Mathur S, Willoughby JR. 2021. The longstanding significance of genetic diversity in conservation. Molecular Ecology 30: 4147–4154.
- Díaz S, et al. 2019. Pervasive human-driven decline of life on Earth points to the need for transformative change. Science 366: aax3100.
- Edmands S. 2006. Between a rock and a hard place: Evaluating the relative risks of inbreeding and outbreeding for conservation and management. Molecular Ecology 16: 463–475.
- Fitzpatrick SW, et al. 2016. Gene flow from an adaptively divergent source causes rescue through genetic and demographic factors in two wild populations of Trinidadian guppies. Evolutionary Applications 9: 879–891.
- Fitzpatrick SW, Handelsman CA, Torres-Dowdall J, Ruell EW, Broder ED, Kronenberger JA, Reznick DN, Ghalambor CK, Angeloni LM, Funk WC. 2017. Gene flow constrains and facilitates genetically based divergence in quantitative traits. Copeia 105: 462–474.
- Fitzpatrick SW, Bradburd GS, Kremer CT, Salerno PE, Angeloni LM, Funk WC. 2020. Genomic and fitness consequences of genetic rescue in wild populations. Current Biology 30: 517–522.
- Flanagan SP, Forester BR, Latch EK, Aitken SN, Hoban S. 2018. Guidelines for planning genomic assessment and monitoring of locally adaptive variation to inform species conservation. Evolutionary Applications 11: 1035–1052.
- Frankham R, Ballou JD, Eldridge MDB, Lacy RC, Ralls K, Dudash MR, Fenster CB. 2011. Predicting the probability of outbreeding depression. Conservation Biology 25: 465–475.
- Funk WC, Forester BR, Converse SJ, Darst C, Morey S. 2019. Improving conservation policy with genomics: A guide to integrating adaptive potential into U.S. Endangered Species Act decisions for conservation practitioners and geneticists. Conservation Genetics 20: 115–134.
- García-Dorado A, Caballero A. 2021. Neutral genetic diversity as a useful tool for conservation biology. Conservation Genetics 22: 541–545.

- Ghalambor CK, Hoke KL, Ruell EW, Fischer EK, Reznick DN, Hughes KA. 2018. Erratum: Non-adaptive plasticity potentiates rapid adaptive evolution of gene expression in nature. Nature 555: 688.
- Gilk SE, Wang IA, Hoover CL, Smoker WW, Taylor SG, Gray AK, Gharrett AJ. 2004. Outbreeding depression in hybrids between spatially separated pink salmon, Oncorhynchusgorbuscha, populations: Marine survival, homing ability, and variability in family size. Genetics of Subpolar Fish and Invertebrates 69: 287-297.
- Griffith B, Scott JM, Carpenter JW, Reed C. 1989. Translocation as a species conservation tool: Status and strategy. Science 245: 477-480.
- Grummer JA, Booker TR, Matthey-Doret R, Nietlisbach P, Thomaz AT, Whitlock MC. 2022. The immediate costs and long-term benefits of assisted gene flow in large populations. Conservation Biology 36: e13911.
- Hällfors MH, Liao J, Dzurisin J, Grundel R, Hyvã¤Rinen M, Towle K, Wu GC, Hellmann JJ. 2016. Addressing potential local adaptation in species distribution models: Implications for conservation under climate change. Ecological Applications 26: 1154-1169.
- Hellmann JJ, Pfrender ME. 2011. Future human intervention in ecosystems and the critical role for evolutionary biology. Conservation Biology 25: 1143-1147.
- Hereford J. 2009. A quantitative survey of local adaptation and fitness tradeoffs. American Naturalist 173: 579-588.
- Hoban S. 2019. New guidance for ex situ gene conservation: Sampling realistic population systems and accounting for collection attrition. Biological Conservation 235: 199-208.
- Hoban S, Kelley JL, Lotterhos KE, Antolin MF, Bradburd G, Lowry DB, Poss ML, Reed LK, Storfer A, Whitlock MC. 2016. Finding the genomic basis of local adaptation: Pitfalls, practical solutions, and future directions. American Naturalist 188: 379-397.
- Hoffmann AA, Miller AD, Weeks AR, 2021. Genetic mixing for population management: From genetic rescue to provenancing. Evolutionary Applications 14: 634-652.
- Hoffmann AA, Sgrò CM. 2011. Climate change and evolutionary adaptation. Nature 470: 479-485.
- Hogg JT, Forbes SH, Steele BM, Luikart G. 2006. Genetic rescue of an insular population of large mammals. Proceedings of the Royal Society B 273: 1491-1499.
- Holt WV, Bennett PM, Volobouev V, Watwon PF. 1996. Genetic resource banks in wildlife conservation. Journal of Zoology 238: 531-544.
- Howard JG, Lynch C, Santymire RM, Marinari PE, Wildt DE. 2016. Recovery of gene diversity using long-term cryopreserved spermatozoa and artificial insemination in the endangered black-footed ferret. Animal Conservation 19: 102-111.
- [IUCN] International Union for Conservation of Nature. 2013. Guidelines for Reintroductions and Other Conservation Translocations, ver. 1.0. IUCN Species Survival Commission.
- Johnson WE, et al. 2010. Genetic restoration of the Florida panther. Science 329: 1641-1645.
- Kardos M, Armstrong EE, Fitzpatrick SW, Hauser S, Hedrick PW, Miller JM, Tallmon DA, Funk WC. 2021. The crucial role of genome-wide genetic variation in conservation. Proceedings of the National Academy of Sciences 118: e2104642118 https://doi.org/10.1073/pnas.2104642118.
- Kardos M, Shafer ABA. 2018. The peril of gene-targeted conservation. Trends in Ecology and Evolution 33: 827-839.
- Kawecki TJ, Ebert D. 2004. Conceptual issues in local adaptation. Ecology Letters 7: 1225-1241.
- Kofler N, et al. 2018. Editing nature: Local roots of global governance. Science 362: 527-529.
- Kohl PA, Brossard D, Scheufele DA, Xenos MA. 2019. Public views about editing genes in wildlife for conservation. Conservation Biology 33: 1286-1295.
- Kostyack J, Lawler JJ, Goble DD, Olden JD, Scott JM. 2011. Beyond reserves and corridors: Policy solutions to facilitate the movement of plants and animals in a changing climate. BioScience 61: 713-719.
- Kronenberger JA, Gerberich JC, Fitzpatrick SW, Broder ED, Angeloni LM, Funk WC. 2018. An experimental test of alternative population augmentation scenarios. Conservation Biology 32: 838-848.

- Landguth EL, Forester BR, Eckert AJ, Shirk AJ, Menon M, Whipple A, Day CC, Cushman SA. 2020. Modelling multilocus selection in an individual-based, spatially explicit landscape genetics framework. Molecular Ecology Resources 20: 605-615.
- Leroy G, Carroll EL, Bruford MW, Dewoody JA, Strand A, Waits L, Wang J. 2018. Next-generation metrics for monitoring genetic erosion within populations of conservation concern. Evolutionary Applications 11: 1066-1083.
- Michalak JL, Stralberg D, Cartwright JM, Lawler JJ. 2020. Combining physical and species-based approaches improves refugia identification. Frontiers in Ecology and the Environment 18: 254-260.
- Paxton EH, Camp RJ, Gorresen PM, Crampton LH, Leonard DL, Vanderwerf EA. 2016. Collapsing avian community on a Hawaiian island. Science Advances 2: e1600029.
- Peterson ML, Doak DF, Morris WF. 2019. Incorporating local adaptation into forecasts of species' distribution and abundance under climate change. Global Change Biology 25: 775-793.
- Phelps MP, Seeb LW, Seeb JE. 2020. Transforming ecology and conservation biology through genome editing. Conservation Biology 34: 54-65.
- Piaggio AJ, et al. 2017. Is it time for synthetic biodiversity conservation? Trends in Ecology and Evolution 32: 97-107.
- Poo S, Hinkson KM. 2020. Amphibian conservation using assisted reproductive technologies: Cryopreserved sperm affects offspring morphology, but not behavior, in a toad. Global Ecology and Conservation 21:
- Prince DJ, O'Rourke SM, Thompson TQ, Ali OA, Lyman HS, Saglam IK, Hotaling TJ, Spidle AP, Miller MR. 2017. The evolutionary basis of premature migration in Pacific salmon highlights the utility of genomics for informing conservation. Science Advances 3: e1603198.
- Rajasekharan PE. 2015. Gene banking for ex situ conservation of plant genetic resources. Plant Biology and Biotechnology 445-459.
- Ralls K, Ballou JD, Dudash MR, Eldridge MDB, Fenster CB, Lacy RC, Sunnucks P, Frankham R. 2018. Call for a paradigm shift in the genetic management of fragmented populations. Conservation Letters 11: e12412.
- Razgour O, Forester B, Taggart JB, Bekaert M, Juste J, Ibáñez C, Puechmaille SJ, Novella-Fernandez R, Alberdi A, Manel S. 2019. Considering adaptive genetic variation in climate change vulnerability assessment reduces species range loss projections. Proceedings of the National Academy of Sciences 116: 10418-10423.
- Rhymer JM, Simberloff D. 1996. Extinction by hybridization and introgression. Annual Review of Ecology and Systematics 27: 83-109.
- Rochat E, Selmoni O, Joost S. 2021. Spatial areas of genotype probability: Predicting the spatial distribution of adaptive genetic variants under future climatic conditions. Diversity and Distributions 27: 1076-1090.
- Ruegg K, Bay RA, Anderson EC, Saracco JF, Harrigan RJ, Whitfield M, Paxton EH, Smith TB. 2018. Ecological genomics predicts climate vulnerability in an endangered southwestern songbird. Ecology Letters 21: 1085-1096.
- Ryder OA, Onuma M. 2018. Viable cell culture banking for biodiversity characterization and conservation. Annual Review of Animal Biosciences 6: 83-98.
- Samuel MD, Liao W, Atkinson CT, Lapointe DA. 2020. Facilitated adaptation for conservation: Can gene editing save Hawaii's endangered birds from climate driven avian malaria? Biological Conservation 241:
- Sandler RL, Moses L, Wisely SM. 2021. An ethical analysis of cloning for genetic rescue: Case study of the black-footed ferret. Biological Conservation 257: 109118.
- Sansilvestri R, Frascaria-Lacoste N, Fernández-Manjarrés JF. 2015. Reconstructing a deconstructed concept: Policy tools for implementing assisted migration for species and ecosystem management. Environmental Science and Policy 51: 192-201.
- Savolainen O, Lascoux M, Merilä J. 2013. Ecological genomics of local adaptation. Nature Reviews Genetics 14: 807-820.

- Saunders S, Grand J, Bateman B, Meek M, Wilsey C, Forstenhaeusler N, Graham E, Warren R, Price J. 2023. Integrating climate change refugia in 30 by 30 conservation planning in North America. Frontiers in Ecology and the Environment. In press.
- Schwartz MW, et al. 2012. Managed relocation: Integrating the scientific, regulatory, and ethical challenges. BioScience 62: 732-743.
- Scott PA, Allison LJ, Field KJ, Averill-Murray RC, Shaffer HB. 2020. Individual heterozygosity predicts translocation success in threatened desert tortoises. Science 370: 1086-1089.
- Shirey PD, Lamberti GA. 2010. Assisted colonization under the U.S. Endangered Species Act. Conservation Letters 3: 45-52.
- Smith AB, et al. 2019. Alternatives to genetic affinity as a context for within-species response to climate. Nature Climate Change 9:
- Sork VL. 2017. Genomic studies of local adaptation in natural plant populations. Journal of Heredity 109: 3-15.
- Stralberg D, Carroll C, Nielsen SE. 2020. Toward a climate-informed North American protected area network: Incorporating climate-change refugia and corridors in conservation planning. Conservation Letters 13: e12712.
- Supple MA, Shapiro B. 2018. Conservation of biodiversity in the genomics era. Genome Biology 19: 131.
- Teixeira J&OC, Huber CD. 2021. The inflated significance of neutral genetic diversity in conservation genetics. Proceedings of the National Academy of Sciences 118: e2015096118.
- Thomas MA, Roemer GW, Donlan CJ, Dickson BG, Matocq M, Malaney J. 2013. Gene tweaking for conservation. Nature 501: 485–486.
- Thompson TQ, et al. 2019. Anthropogenic habitat alteration leads to rapid loss of adaptive variation and restoration potential in wild salmon populations. Proceedings of the National Academy of Sciences 116:
- Thurman LL, Gross JE, Mengelt C, Beever EA, Thompson LM, Schuurman GW, Hoving CL, Olden JD. 2022. Applying assessments of adaptive capacity to inform natural-resource management in a changing climate. Conservation Biology 36: e13838.
- Tiffin P, Ross-Ibarra J. 2017. Advances and limits of using population genetics to understand local adaptation. Trends in Ecology and Evolution 29: 673-680.
- Trouwborst A. 2012. Transboundary wildlife conservation in a changing climate: Adaptation of the Bonn Convention on migratory species and its daughter instruments to climate change. Diversity 4: 258-300.
- Tunstall T, Kock R, Vahala J, Diekhans M, Fiddes I, Armstrong J, Paten B, Ryder OA, Steiner CC. 2018. Evaluating recovery potential of the northern white rhinoceros from cryopreserved somatic cells. Genome Research 28: 780-788.
- [USFWS] US Fish and Wildlife Service. 1995. Final Rule Determining Endangered Status for the Southwestern Willow Flycatcher. USFWS. Federal Register no. 60 FR 10694.
- [USFWS] US Fish and Wildlife Service. 2013. Designation of Critical Habitat for Southwestern Willow Flycatcher. USFWS. Federal Register no. 78 FR 343.
- [USFWS] US Fish and Wildlife Service. 2019. Recovery Plan for the Dakota Skipper (Hesperia dacotae). USFWS.
- [USFWS] US Fish and Wildlife Service. 2021a. Innovative Genetic Research Boosts Black-Footed Ferret Conservation: Efforts by USFWS and Partners. USFWS. www.fws.gov/mountain-prairie/ pressrel/2021/02182021-USFWS-and-Partners-Innovative-Genetic-Cloning-Research-Black-footed-Ferret-Conservation.php.
- [USFWS] US Fish and Wildlife Service. 2021b. Recovery Plan for the Rusty Patched Bumble Bee (Bombus affinis). USFWS.
- [USWFS] US Fish and Wildlife Service. 2021c. Recovery Plan for the Eastern Massasauga Rattlesnake (Sistrurus catenatus). USFWS.

- Verschuuren J. 2010. Climate change: Rethinking restoration in the European Union's birds and habitats directives. Ecological Restoration 28: 431-439.
- Waples RS, et al. 2022. Implications of large-effect loci for conservation: A review and case study with Pacific salmon. Journal of Heredity 113:
- Waples RS, Lindley ST. 2018. Genomics and conservation units: The genetic basis of adult migration timing in Pacific salmonids. Evolutionary Applications 11: 1518-1526.
- Waters CD, Hard JJ, Brieuc MSO, Fast DE, Warheit KI, Knudsen CM, Bosch WJ, Naish KA. 2018. Genome-wide association analyses of fitness traits in captive-reared Chinook salmon: Applications in evaluating conservation strategies. Evolutionary Applications 11: 853-868.
- Weeks AR, et al. 2011. Assessing the benefits and risks of translocations in changing environments: A genetic perspective. Evolutionary Applications 4: 709-725.
- Weeks AR, Heinze D, Perrin L, Stoklosa J, Hoffmann AA, Van Rooven A, Kelly T, Mansergh I. 2017. Genetic rescue increases fitness and aids rapid recovery of an endangered marsupial population. Nature Communications 8: 1071.
- White NJ, Butlin RK. 2021. Multidimensional divergent selection, local adaptation, and speciation. Evolution 75: 2167-2178.
- Whiteley AR, Fitzpatrick SW, Funk WC, Tallmon DA. 2015. Genetic rescue to the rescue. Trends in Ecology and Evolution 30: 42-49.
- Whitlock MC. 2015. Modern approaches to local adaptation. American Naturalist 186 suppl. 1: S1-S4.
- Williams GC. 2018. Adaptation and Natural Selection: A Critique of Some Current Evolutionary Thought. Princeton University Press.
- Williams JW, Ordonez A, Svenning JC. 2021. A unifying framework for studying and managing climate-driven rates of ecological change. Nature Ecology and Evolution 5: 17-26.
- Willi Y, Kristensen TN, Sgrò CM, Weeks AR, Ørsted M, Hoffmann AA. 2022. Conservation genetics as a management tool: The five bestsupported paradigms to assist the management of threatened species. Proceedings of the National Academy of Sciences 119: e2105076119.

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