

The neuromodulatory role of the noradrenergic and cholinergic systems and their interplay in cognitive functions: a focused review

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Abstract

Noradrenergic and cholinergic modulation of functionally distinct regions of the brain has become one of the primary organizational principles behind understanding the contribution of each system to the diversity of neural computation in the central nervous system. Decades of work has shown that a diverse family of receptors stratified across different brain regions and circuit-specific afferent and efferent projections play a critical role in helping such widespread neuromodulatory systems obtain substantial heterogeneity in their role in neural information processing. This review briefly discusses the anatomical layout of both the noradrenergic and cholinergic systems, as well as the types and distributions of relevant receptors for each system. Previous work characterizing direct and indirect interaction between these two systems is discussed, especially in the context of high order cognitive functions such as attention, learning, and the decision making process. Though a substantial amount of work has been done to characterize the role of each neuromodulator, a cohesive understanding of the region-specific cooperation of these two systems is not yet fully realized. For the field to progress, new experiments will need to be conducted that capitalize on the modular subdivisions of the brain and systematically explore the role of norepinephrine and acetylcholine in each of these subunits and across the full range of receptors expressed in different cell types in these regions.

18 Introduction

19 The central nervous system performs an incredibly large number of continuous
20 computations, the result of which is to efficiently process the external world and execute a relevant
21 response. In the human brain, an estimated 10^{11} neurons make approximately 1000 average
22 connections to other neurons, forming up to 10^{14} distinct sites for information transmission. That
23 is likely an order of magnitude higher than the total number of cells in the entire human body¹.
24 Even this staggering number of physical connections understates the complexity of information
25 handling in the brain. Beyond simple neuron-to-neuron connections, multiple subtypes of glial
26 cells are also known to play a role in synaptic transmission^{2,3}. This ever-shifting structural
27 background, across which the flow of information proceeds throughout an individual's life, is then
28 capable of giving rise to a diverse array of orchestral melodies through the 100+ endogenous
29 substances that play a role in modulating synaptic transmission⁴. Some of these substances,
30 known as neurotransmitters, can act over varying physical distances through their interaction with
31 a much larger number of receptors.

32 Since the discovery of the first neurotransmitter, acetylcholine, in 1926 by Otto Loewi,
33 there has been an explosion in the identification and understanding of chemical
34 neurotransmission. Conceptually simplified, information transfer occurs in two modes: electrical
35 propagation within neurons or chemical propagation outside neurons. Neurotransmitters are the
36 chemicals that traverse the physical division between cells connecting the postsynaptic cell with
37 information from the presynaptic cell. This is primarily mediated through an array of specific
38 receptors on the postsynaptic cell. For any given neuron the combination of presynaptic inputs
39 will determine if a message is electrically transcribed and transmitted. If it is transmitted, an action
40 potential will travel down the length of a neuron, resulting in the release of extracellular
41 neurotransmitter onto the dendrites of postsynaptic cells. The substances released are usually
42 tightly regulated and reuptaken or degraded to limit the action of the substance on its target.

Generally, neurotransmitters may have excitatory, inhibitory, and neuromodulatory effects on neurons through the action of their receptors. Excitatory receptors, when activated by corresponding neurotransmitters, result in a membrane depolarization and the propagation of an action potential. Glutamate is the neurotransmitter that predominate mediates excitatory effects through its receptor, which is nearly ubiquitously expressed in all types of neurons and many types of glial cells⁵. Inhibitory receptors exert an opposing effect, with the binding of corresponding neurotransmitters resulting in a membrane hyperpolarization that limits the ability of a neuron to initiate an action potential. In the mature brain, γ -aminobutyric acid (GABA) is the primary neurotransmitter that exerts inhibitory effects on neurons through GABAergic receptors⁶. The interplay between these two competing systems has been studied in a variety of contexts⁷⁻⁹ and provides the foundation for how neurotransmission is thought to occur in the brain. The summation of excitatory and inhibitory inputs at every connection point in the brain determines the direction and pattern of information propagation in complex networks of neurons. The third type of neurotransmitters, known as neuromodulators, add an additional, but important, complexity to this paradigm by altering the balance of transmission on a micro-, meso-, or macroscale.

There are four primary neuromodulatory systems: acetylcholine, norepinephrine, dopamine, and serotonin. Each of these four molecules play an important function in altering basic synaptic transmission patterns. The groups of neurons responsible for delivery of these four neuromodulators are known as ascending neuromodulatory systems due to the fact each of these neurotransmitters originates in the brainstem, midbrain, or basal forebrain and projects to various brain structures. While early work characterized these ascending modulatory systems as highly collateralized and largely exerting their influence through global regulation of neural activity^{10,11}, more recent work has emphasized the subdivision of these systems into cortical-region-specific sub-systems that can differentially influence information processing^{12,13}. Subsequently there has been a shift away from viewing neuromodulatory systems as mere “state-setting” or “gating”

69 systems towards a framework in which there is also an understanding that a subset of highly
70 specific neuromodulatory projections to the forebrain, especially the cortex, are necessary for
71 specific cognitive functions¹⁴.

72 This review will focus specifically on the role of two of these neuromodulatory systems,
73 the noradrenergic and cholinergic systems, in modulating cognitive functions. Acetylcholine (ACh)
74 is an important neuromodulator long implicated in learning, memory, and synaptic plasticity^{15,16}.
75 More recent work, however, has also highlighted the role of ACh in attentional effort, orienting,
76 and detection of behaviorally significant stimuli^{17,18}. Norepinephrine (NE) has classically been
77 viewed as a major mediator of arousal that plays an important role in regulating the efficiency of
78 external sensory processing¹⁹⁻²⁵. Recent work expanding on this has revealed a varied and
79 complex role for the noradrenergic system in everything from memory formation²⁶, to executive
80 function and attention^{27,28}, to cognitive flexibility^{29,30}, to decision making³¹⁻³³. Dysfunction in either
81 of these two neuromodulatory systems or in the coordinated interaction between them is heavily
82 implicated in numerous neurodegenerative and neuropsychiatric diseases³⁴⁻⁴¹.

83 Understanding the complex and dynamic roles of neuromodulation on sophisticated
84 behavioral outcomes requires not only understanding the full range of impact of each
85 neuromodulator and its various anatomically distinct subdivisions, but also an understanding of
86 the interplay between multiple neuromodulatory systems. These interactions could occur at the
87 functional level seen in an organism's behavior, the anatomic level through connectomic
88 communication, the cellular level through intertwined signaling pathways, or even at the synaptic
89 level when important brain regions experience the release of multiple neurotransmitters. A more
90 complete understanding of the role that neuromodulation plays in brain functions will require a
91 better understanding of when and how two systems, such as the cholinergic and noradrenergic
92 systems, collectively work together. The implications will be critical for better understanding and
93 predicting normal brain function and for providing novel treatment strategies for aberrant brain
94 function.

Anatomical Overview

Cholinergic System

Sources

The cholinergic system is primarily comprised of groups of cells in the basal forebrain and midbrain that send diffuse but sparse projections to the rest of the brain⁴²⁻⁵². Only the striatum differs with a local supply of cholinergic neurons for local transmission⁵³. In primates, the cholinergic input to the cerebral cortex originates almost entirely from the nucleus basalis of Meynert (NBM), located in the basal forebrain⁵⁴. These neurons are large with extensive dendritic trees, and a single neuron can innervate multiple brain regions, though there is minimal overlap in the axonal fields^{50,55}. While cholinergic projections are widespread throughout the cortex, multiple studies have shown that there is a distinct pattern of segmented innervation depending on which nuclei in the basal forebrain the cholinergic neuron originates from^{17,51,56-58}.

A second major cholinergic source is found in the continuous array of multipolar neurons in the medial septum (MS)⁴⁶ and diagonal band of Broca (DB)^{59,60}. A whole-brain atlas of projections from these regions in the mouse has found that cholinergic neurons were distributed in an uneven pattern between different brain regions, with neuronal density varying more than 10-fold across various structures⁵⁰. Additionally, soma volume of cholinergic neurons varied by up to 8-fold across the examined regions⁵⁰. Taken together, this work has demonstrated the existence of region-specific subtypes of cholinergic neurons that perform varying functions, though unlike in the nucleus basalis, there does not seem to be a specific relationship between soma location in the nuclei and the location of projections⁵⁰.

Inputs

Cholinergic neurons in the nucleus basalis (NB) receives dopaminergic input from the ventral tegmental area and substantia nigra, serotonergic input from the raphe nuclei, and

121 noradrenergic input from the locus coeruleus^{47,61}. The main cholinergic afferents to the nucleus
122 basalis arrive from the midbrain pedunculo-pontine (PPT), the lateral dorsal tegmental (LDT)
123 nuclei, and the limbic cortex⁶²⁻⁶⁴. There is also GABAergic input in the form of symmetric
124 synapses, which could represent local inhibitory neurons or projections from other brain regions⁶⁵⁻
125 ⁶⁷. It has been shown in multiple studies that the cholinergic neurons originating in NB and
126 projecting to the cortex are regulated through GABAergic receptors^{47,61,65-67}. There is also
127 extensive bidirectional communication between the nucleus basalis and the prefrontal cortex, and
128 in the mouse the rostral-caudal distribution of cholinergic cells in the basal forebrain is associated
129 with projection to the superficial-deep layers of the ventral medial prefrontal cortex, respectively⁴⁹.

130 Both the PPT and LDT nuclei receive afferent projections from a widespread number of
131 common structures, most predominately the reticular formation in the brainstem, the midbrain
132 central gray region, and the lateral hypothalamus-zona incerta region^{68,69}. Retrograde tracing
133 studies have shown inputs arriving in the LDT nucleus from the midbrain reticular formation, the
134 periaqueductal gray, the medial preoptic nucleus, the anterior hypothalamic nucleus, the
135 perifornical and lateral hypothalamic areas, the premammillary nucleus, paraventricular
136 hypothalamic nucleus, zona incerta, and the lateral habenular nucleus^{46,70}. The PPT nucleus
137 receives afferent inputs from the wide range of regions, reviewed more in depth by Martinez-
138 Gonzalez in 2011⁷¹, but most notably including afferent connections from the cortex^{69,72,73}, various
139 locations in the basal ganglia^{69,74-81}, the locus coeruleus⁸², and the dorsal raphe⁸³.

140 The MS receives noradrenergic inputs from the locus coeruleus^{45,84} and serotonergic
141 inputs from the raphe nuclei⁸⁵, as well as additional inputs from several other brainstem nuclei⁸⁵.
142 Afferent projections to the DB have not been well characterized, partly due to its poorly defined
143 margins. Some studies have shown afferent connections arriving from the supermammillary
144 nucleus⁸⁶ and reciprocal connections with the CA2 subfield of the hippocampus⁵⁹.

145

Outputs

The nucleus basalis is an important source of ACh to the cerebral cortex^{43,51,52,56,57}, with efferent cholinergic projections that terminate on both pyramidal and GABAergic cells⁵⁵. The nucleus basalis also supplies several thalamic nuclei with ACh, including the intralaminar nuclei, medial dorsal nucleus, and reticular nucleus^{44,87}. Additional cells arise from the nucleus basalis that terminate throughout the amygdala, though most of these projections are GABAergic, with a minority being cholinergic^{88,89}.

The cholinergic nuclei in the midbrain, the PPT and LDT, have primary outputs that project to the nucleus accumbens, hypothalamus, raphe, and pontine and medullary reticular formations. They also project to the nucleus basalis, all thalamic nuclei, the amygdala, and the primary visual cortex^{48,90}. The PPT, specifically, has long been thought to act as an interface between the basal ganglia and motor systems, though more recent work highlights the role of the varied neuronal subtypes and projections to play a central role in updating behavioral states⁹¹. The LDT appears to play a significant role in activating the mesolimbic reward system⁹².

Noradrenergic System

Sources

Noradrenergic projection to the forebrain is exclusively provided by a single source: the locus coeruleus (LC), which is a small, bilateral nucleus located in the pons⁹³⁻⁹⁷. A complete review of the LC was provided by Poe et al in 2020⁹⁸, but a brief description will be provided here. Traditional investigations of the LC presumed it to be a broadly acting, primarily homogenous source of NE with broad implications^{93,94,99,100}, but more recent research has shown that the LC is composed of many distinct modules with highly specific functional roles throughout the brain⁹⁸. There are two major, complementary theories on how a diffusely projecting single source of norepinephrine can achieve such disparate functional results. The first is that the function of NE release relies on regional differences in postsynaptic receptor distribution and resulting

differences in spatiotemporal NE reuptake¹⁰¹⁻¹⁰³. The second is a corollary to the function of the noradrenergic system in the periphery, in which the sympathetic nervous system has discrete efferent limbs that are organ specific but capable of acting in a unified manner^{104,105}. In this theory, the LC provides localized neuromodulation to well-defined target regions and spiking is synchronized in highly specific subsets of LC neurons. For a more complete review see Totah et al, 2019¹⁰⁶.

Inputs

An important step in understanding the regional and modular functionality of the LC was achieved through an in-depth characterization of the afferent and efferent projections to and from the LC. The LC itself consists of a small, dense core, where cell bodies are found, and a peri-LC shell in which LC dendrites reside¹⁰⁷⁻¹⁰⁹. There are prominent afferent inputs to the LC core originating from the paragigantocellularis nucleus and the prepositus hypoglossi nuclei, both structures in the rostral medulla¹¹⁰. There are also additional inputs from the insular cortex, central nucleus of the amygdala, preoptic area, and the lateral and paraventricular hypothalamic areas^{108,111,112}. Cerebellar Purkinje cells and neurons from deep cerebellar nuclei also provide synaptic inputs onto the core of the LC¹⁰⁸.

Although the projections of sensory afferents from the mesencephalic trigeminal sensory nucleus (Me5)^{113,114} and the nucleus of tractus solitarius (NTS)¹¹⁵ to the LC exert influences on cognitive functions¹¹⁶, an important regulatory component on the core noradrenergic neurons in the LC include the peri-LC afferent innervations. Noradrenergic LC neurons possess long dendrites that pass through the surrounding small nuclei-like regions around the LC, which receive separate inputs from a variety of brain regions, including the prefrontal and infralimbic cortex, the amygdala, and the dorsal raphe nucleus¹¹⁷. There are additionally cholinergic, serotonergic, and adrenergic inputs to the peri-LC area, representing potential points of indirect regulation from other neuromodulatory systems^{98,112}. The peri-LC zone also gives rise to a number of GABAergic inputs into the LC^{118,119}.

Outputs

The efferent projections from the LC are widespread but nonuniform to the neocortex in both rodents¹⁰² and primates^{120,121}. Collateral axons from the LC are distributed in a coordinated fashion to target circuits with a specific function^{98,108,122-126}. The efferent projections from the LC travel throughout the brain, providing NE input to the cortex, insula, hippocampus, thalamus, amygdala, and cerebellum. A full review of this system was provided by Schwarz and Luo in 2015²¹. Though the projections are widespread, the selective activation of specifically patterned noradrenergic neurons is poorly understood and likely involves a complex interplay between inputs into the LC and interacting systems²¹. Nevertheless, it has been shown that genetically distinct groups of noradrenergic neurons project to regionally and functionally specific circuits¹²⁷. Understanding the anatomically distinct efferent circuits underlying specific functional consequences is an ongoing area of research that will likely improve our understanding of the role of the LC in the context of localized function.

An important aspect of neuromodulation, the LC also directly projects to serotonergic, cholinergic, and dopaminergic nuclei, providing a centralized locus of control over, or feedback with, other neuromodulators^{63,128}.

Direct Communication between the Cholinergic and Noradrenergic Systems

Direct interactions between the cholinergic and noradrenergic systems are complex and likely highly dependent on regional context. Some example experiments have shed an early understanding of some of these direct actions. Post-synaptic NE release inhibits approximately 90% of rat brainstem cholinergic neurons through direct activation of inwardly rectifying K⁺ currents, most likely through the α_2 adrenoreceptor¹²⁹. In striatal cholinergic neurons, NE mediates depolarization, through β_1 adrenoreceptor activation¹³⁰. It is also interesting to note, that the LC has unidirectional input into the basal forebrain, where a mix of α_1 and β_1 adrenoceptors are found

on cholinergic neurons^{131,132}. In the same region however, GABAergic cells express α_2 , the activation of which suppresses neural activity. Overall, the net effect of LC modulation on the basal forebrain is enhancement of ACh release in the cortex^{21,133,134}, though more sophisticated studies should be performed to determine the binding preference and net effect with varying levels of NE input¹³⁵.

Conversely, ACh acting on noradrenergic neurons is mediated primarily through α_3 nicotinic receptors, leading to depolarization and NE release and activation of the hypothalamic-pituitary-adrenal axis¹³⁶. In the hippocampus, nicotinic receptors are also responsible for releasing NE from LC neuron terminals, likely through the NO/cGMP pathway^{136,137}. Beyond evidence for local modulation of noradrenergic neurons by ACh^{138,139}, cholinergic receptors, presumed to be muscarinic, on LC neurons can act to centrally modulate LC firing. ACh induces increased firing rates, though the cholinergic source is not well understood.

These direct forms of interactive modulation demonstrate that even in the simplest scenarios in which cholinergic or noradrenergic activity occurs, the other system is being engaged. Whether or not this results in a synergistic or antagonistic effect likely depends on the extent to which each system is activated, the distribution of receptors in targeted regions, and the relative concentrations of each.

Indirect Communication of Cholinergic and Noradrenergic Systems

Cholinergic and noradrenergic projections to the prefrontal cortex are important for a variety of cognitive and executive functions. As such, neuromodulatory connections to distinct areas in the cortex such as the anterior cingulate cortex, medial prefrontal cortex, and orbitofrontal cortex control important aspects of an animal's behavior¹⁴⁰⁻¹⁴³. An important study by Chandler et al in 2014, showed that, while both cholinergic and noradrenergic neurons projected to the cortex from their respective nuclei, their pattern of distribution varied across subregions in the prefrontal cortex¹⁴⁴. Cholinergic neurons appeared to occur throughout all regions in a relatively equal

distribution, while noradrenergic neurons projected to much more defined locations, which did not overlap with other monoaminergic projections¹⁴⁴.

Much of what is currently known about cholinergic and noradrenergic interaction has been researched in the context of various functional outcomes or neurological disease models. In a sheep model of chronic pain concentrations of ACh and NE in the cerebral spinal fluid were measured, and found to only be correlated to one another in those animals with pain¹⁴⁵. In Alzheimer's disease (AD), the close interplay between ACh and NE is being increasingly investigated under a theory that AD is a broad neuromodulatory disorder as opposed to a dysfunction of primarily the cholinergic system^{34,146}. Locomotor activity, a process often disrupted in a wide array of neurological disorders, has also been shown to be mediated through cholinergic interaction with the noradrenergic system¹⁴⁷. In attentional disorders, there is evidence indicating deficits in norepinephrine-mediated control of the cholinergic system in the parietal cortex¹⁴⁸.

An extremely important, though substantially under-researched mode by which indirect interactions between these neuromodulatory systems occur is in their differential effect in glial cells such as astrocytes. Specifically, it has been shown that each neuromodulator has a specific effect on astrocyte potassium clearance, thereby regulating the extracellular potassium concentration and influencing local synaptic transmission¹⁴⁹.

There are also examples in literature of the indirect engagement of the noradrenergic system that is likely mediated by α_7 nicotinic receptors on GABAergic neurons and a resulting disinhibitory effect¹⁵⁰. Supporting this is direct measurement of increased ACh and NE in the rat cortex after administration of a nicotinic agonist¹⁵¹. It is important to note here though, that a simple increase and decrease in neurotransmitter level is not enough to discern the role of that change in a functional capacity. A more thorough understanding of the location, receptors, and other systems involved is needed to unravel functional consequences.

Role of Acetylcholine in the Brain

Major Cholinergic Receptor Subtypes and Function

The cholinergic system exerts its action by binding to two distinct receptor classes: nicotinic and muscarinic. Nicotinic acetylcholine receptors (nAChRs) are composed of five subunits arranged around a central core, forming a transmembrane channel that conducts Na⁺, K⁺, and Ca²⁺ when bound to ACh, leading to a local membrane depolarization¹⁵². The assembled receptor is constructed using two primary families of α and β subunits¹⁵². The standard neuronal configuration includes combinations of $\alpha 2$ through $\alpha 6$ and $\beta 2$ through $\beta 4$ proteins^{153,154}. There are additional homomeric configurations composed of $\alpha 7$ through $\alpha 9$ subunits¹⁵⁵. The central nervous system appears to be predominately composed of $\alpha 4\beta 2$ or $\alpha 7$ nAChRs, of which the former has a higher affinity for ACh¹⁵⁶. In the adult rat brain, there is little anatomical overlap between the heteromeric and homomeric nAChR types, indicating two distinct modes of ACh activity within distinct anatomical regions¹⁵⁷. Overall, nAChRs are distributed widely, but sparsely, throughout the hippocampus and cortex at both pre- and postsynaptic locations¹⁵² and expressed on interneurons, pyramidal cells, and stellate cells¹⁵⁸⁻¹⁶¹. A series of previous experiments have shown that layers I, III, and V in the human cortex exhibit the highest binding of ACh. This was slightly different in the primary somatosensory cortex, where binding in layer III was highest, and in the primary motor cortex, where layers III and V were the highest¹⁶². The $\alpha 7$ receptors are expressed broadly across almost all glutaminergic and GABAergic neurons, though regional differences have been noted. The presence of a presynaptic nAChR almost universally results in an increased neurotransmitter release, across multiple types of neurotransmitters^{152,163}.

Muscarinic receptors (MAChR) are expressed throughout all layers of the cortex though layers II and V exhibit the highest concentration¹⁶⁴. There are a total of five known muscarinic receptors types, M₁ to M₅. In general, M₁ receptors are most abundant in the neocortex, hippocampus, and striatum¹⁶⁵; M₂ receptors are located throughout the entire brain¹⁶⁶; M₃ receptors only have a low level of expression throughout the brain¹⁶⁴; M₄ receptors are localized

in the striatum¹⁶⁷; and M₅ receptors are also widely distributed across the brain¹¹⁸. These receptors have a seven transmembrane region that is highly conserved in G-protein coupled receptors (GPCRs) and activate multiple intracellular signaling pathways, including phospholipase C (by M₁, M₃, M₅), inhibition of adenylyl cyclase (by M₂ and M₄), and regulation of several ion channels^{168,169}. MACHRs also activate mitogen-activated protein kinases (MAPKs), which regulates cell survival, differentiation, and synaptic plasticity¹⁷⁰⁻¹⁷².

Cholinergic Involvement in Learning and Decision Making

The cholinergic system plays an important role in higher cognitive functions, specifically in decision making and the learning process. Neuromodulation by acetylcholine is generally orchestrated through the differential activation of the nicotinic and muscarinic receptors. The nicotinic receptor, for example, has been shown to be easily desensitized and up-regulated through the presence of nicotine, inducing long term alterations in the decision-making process¹⁷³. Nicotine has also been implicated in increasing impulsivity and disinhibition in decision making¹⁷⁴. Studies have also shown how nAChRs seem to be important in adapting appropriate choices to a specific outcome¹⁷⁵. Specifically, the nAChR $\alpha 7$ receptor has been implicated in slowing learning rates in mice during knockout experiments⁴, as well as cognitive improvement during enhanced activation¹⁷⁶.

Both nAChRs and MACHRs have been identified to contribute to risk and uncertainty¹⁷⁷, with several studies showing that a complex interaction between these two receptors contributes to cholinergic interneuron patterned activity^{178,179}. This interaction seems to have a larger implication for network dynamics across different brain regions. Specifically, studies where MACHRs or nAChRs were activated or blocked demonstrated changes in neural synchrony across multiple EEG bands, invoking the emergence of theta-gamma coupling in the cortex and exhibiting a correlation to increased learning performance in item-context association behaviors^{180,181}.

Acetylcholine on a global level in the brain has been found to be relevant in almost every decision-making paradigm. Lesioning studies of the basal forebrain have shown disruptions in reversal learning in marmosets¹⁸², as well as being implicated in memory storage^{183,184}. The role of ACh in memory has been shown to act as a modulator of update speed and as a controller of metalearning¹⁸⁵. Here ACh modulates different neural systems throughout learning, regulating the appropriate amount of ACh in specific brain regions to appropriately and effectively learn and formulate memories¹⁸⁶.

Cholinergic Involvement in Attention

Cholinergic release primarily mediates attentional processing in the brain^{17,187}. In a 5-choice serial reaction time task (5-CSRTT), the nicotinic facilitation of attention was found to exist and the magnitude of this facilitation was dependent on the level of attentional engagement¹⁸⁸. In addition, by using the 5-CSRT, Robbins and colleagues demonstrated that nicotinic $\beta 2$ subunits in the prelimbic cortex are crucial for mice to successfully detect the cue¹⁸⁹. In an operant sustained attention task (SAT), both the detection of signals and the attentional performance were enhanced by the $\alpha 4\beta 2$ nAChR agonist-evoked ACh increases in the mPFC¹⁹⁰. In a knockout study, it was shown that task performance which relies on highly attentive control was impaired in a group of mice lacking the $\beta 2$ subunit in the mPFC, in comparison with their wild-type littermates¹⁹¹. Similarly, genetic deletion of this special subtype of nAChRs also results in compromised performance in an auditory discrimination paradigm¹⁹⁰, suggesting its critical role in selective auditory attention.

In addition, there is evidence implicating that the muscarinic system plays a role in directing attentional selection mechanisms¹⁹²⁻¹⁹⁴. Specifically, muscarinic receptors are believed to primarily modulate higher-level visual stimulus processing¹⁹⁴. Attending to the receptive field of certain V1 neurons evokes an increase in these neurons' firing rates and scopolamine, a muscarinic antagonist, reduces this attentional modulation¹⁹⁵. Interestingly, nicotinic antagonist

does not exert systematic effect¹⁹⁵. Yet another aspect which can also be associated with attentional control is adaptive behavioral control¹⁹⁶. Various genotypes of M₂ modulate the high-level inhibitory control processes that require the processing of prior information and suppression of irrelevant information¹⁹⁶.

Role of Norepinephrine in the Brain

Major Noradrenergic Receptor Subtypes

The noradrenergic system exerts influence over brain function through three receptor classes: α_1 , α_2 , and β receptors. Each of these receptors has control over specific processes of neurotransmission and sympathetic nervous system regulation. α_1 receptors are members of the adrenoceptor family, a subset of G-protein coupled receptors¹⁹⁷. They have been further classified into three distinct subtypes: α_{1A} , α_{1B} , and α_{1D} . Each subreceptor has demonstrated unique quantitative differences in effect¹⁹⁷. Several experiments have explored the different concentrations of these subtypes throughout the brain. Specifically, it has been shown that α_{1B} was more prominent in the thalamus, lateral amygdaloid nuclei, and cortical laminar areas, while α_{1A} was higher in the entorhinal cortex, amygdala, and general cerebral cortex areas¹⁹⁸. Furthermore, transgenic mouse experiments have allowed for specific receptors to be knocked out, uncovering that both α_{1A} and α_{1B} has similar expression throughout the central nervous system, just with different abundances¹⁹⁹. Around 55% of the brain was shown to express α_{1A} , 35% α_{1B} , and less than 10% α_{1D} ²⁰⁰⁻²⁰². The function of α_1 receptors is implicated in a variety of cognitive processes and synaptic efficacies. Beginning with synaptic involvement, α_1 receptors have been shown to increase the firing frequency of pyramidal and somatosensory neurons of the visual cortex through the protein kinase C signaling (PKC) pathway^{203,204}. They have also been implicated in the enhancement of glutamate and acetylcholine release as well as neuronal excitation via PKC pathways, calcium pathways, and excitatory synapses²⁰⁵⁻²⁰⁹. α_1 has also been shown to affect non-neuronal function as well, with modulation of synaptic transmission through

astrocytes and glial cells²¹⁰⁻²¹². In regards to cognitive functions, α_1 receptors have been shown to be implicated in memory, motor and motivational behavior, memory retention, and storage, but most of these are associated with general norepinephrine release in the brain²¹³.

α_2 receptors are also a type of G-protein coupled adrenoreceptor, classified into three subtypes: α_{2A} , α_{2B} , and α_{2C} . Specifically α_2 receptors have been implicated in orchestrating the presynaptic inhibition of norepinephrine in the central and peripheral nervous system²¹⁴⁻²¹⁶. This inhibition is critical for regulation of normal involuntary processes including physiological functions of the heart, vision, and gastrointestinal systems. Using pharmacological agents such as prazosin or oxymetazoline, α_{2A} and α_{2B} receptors have been shown to have significant control over sympathetic outflow and blood pressure²¹⁶. Several other studies have shown α_{2A} receptor agonists enhance both serotonin and norepinephrine release²¹⁶. Interestingly, the abundance of α_2 receptor subtypes is much more localized than α_1 . While literature here is limited, studies have shown that α_{2B} receptors are found almost exclusively in the thalamus; α_{2C} in the olfactory bulb, cerebral cortex, hippocampal formation, and dorsal root ganglia²¹⁷.

The final type of noradrenergic receptors, classified as β , are also a G-protein coupled receptor, divided into three subtypes: β_1 , β_2 , β_3 ²¹⁶. There have been studies linking β receptors to synaptic plasticity, with norepinephrine acting on β receptors to dictate synaptic strength in hippocampal neurons, as well as NE released from the locus coeruleus enhancing LTD-related memory processing²¹⁸.

Noradrenergic Involvement in Learning and Decision Making

The noradrenergic system has been implicated in a variety of decision-making paradigms as well as throughout the learning process. Studies using optogenetics, pharmacological agents, and lesioning have brought to light the effect norepinephrine has on cognition and higher-order thought processes. One theory regarding the role of NE in decision making involves the idea of network reset, acting as an “internal interrupt” signal^{219,220}. Here it is explained that the phasic

activation of locus coeruleus noradrenergic neurons, causes an increase of NE throughout the cortex, invoking cognitive shifts and potential reorganization of neural networks²²¹. This shifted brain state is hypothesized to be better equipped for rapid behavioral adaptation and enhanced decision making²²¹. Other theories point out how stimulus-induced firing patterns of the LC are closely attuned to behavioral performance, hypothesized from LC primate recordings in visual discrimination tasks²²². Similar phasic activation in primates has shown how the LC can respond to specific task-related decisions, modulating NE release and adapting future task-relevant decisions¹⁸⁶, as well as showcasing coordinated activity patterns in cortical networks derived from ascending NE projections²²³. Studies invoking NE release through an agonist have shown enhancement in sensory stimulation, allowing more rapid synaptic plasticity and faster behavioral responses²²⁴.

Several pharmacological experiments have investigated the specific role α_2 receptors play in the decision-making process. Studies using NE antagonists have shown α_{2A} receptor knockout leading to more risk-on behavior, with rats exhibiting greedier decisions²²⁵. α_{2A} agonists have been proven to enhance the efficiency of working memory and reduce impulsivity in primates²²⁶. This increased receptor uptake in the prefrontal cortex seems to be part of the shifted network brain state described earlier. The agonist guanfacine, another α_{2A} agent, was also shown to improve visual object discrimination performance during a reversal learning paradigm in primates²²⁷.

Noradrenergic Involvement in Attention

Noradrenergic modulation of attention has been studied for several decades²²⁸⁻²³⁰. Studies have established the theory that the LC-NE system regulates the efficacy of information processing during neuronal coding of detected cues^{20,231,232}. During behavioral tasks, selective attention enhances neuronal responsiveness to sensory cues^{233,234}. The firing rates of LC neurons is correlated with attentive behavior in an odd-ball task²³⁵, in which either high or low tonic firing rates corresponds to inattentive states, and medium firing rates associates with animals' best

performance. In a novel environment where more adaptive behaviors are required, changes in electrotonic coupling among LC neurons regulates the goal-directed exploration and preserves attentional selectivity²⁸. In addition, some studies have investigated the effects of NE agonists. It is shown that in a cued target detection task (CTD), the application of α_2 receptor agonists clonidine or guanfacine significantly impaired alerting behavior and the effect was dose-dependent²³⁶, while the effect was blocked by the α_2 antagonists idazoxan or yohimbine.

Most recent studies also show an association between the NE system and impulsivity control²³⁷⁻²³⁹. It was observed from the superior frontal theta band activity that the NE system dynamically gains and loses relevance to regulate inhibitory control under different responding modes²³⁹. This work has led to the use of the NE-specific reuptake inhibitor atomoxetine as a treatment of pediatric attention-deficit/hyperactivity disorder (ADHD)²³⁷. Furthermore, it is demonstrated that ADHD patients have a higher positron emission tomography (PET)-measured NET availability in comparison to healthy individuals, suggesting that there are underlying genetic and epigenetic mechanisms.

Functional Interplay between the Cholinergic and Noradrenergic Systems

ACh and NE in Attention

It has been widely acknowledged that both cholinergic and noradrenergic systems show graded and transient increases in their response to increased attention to environmental cues²⁴⁰⁻²⁴³. Noradrenergic axon activity starts ~1 s prior to the peak of pupil dilation while cholinergic axon activity lags ~0.5 s behind the peak, suggesting pupil-linked alertness, attention and mental effort are controlled differentially by the two neuromodulatory transmitter systems²⁴⁴. Studies have shown that basal forebrain (BF) and brainstem cholinergic systems interact differently with LC-NE system related to attention^{42,245}. It was demonstrated that sustained attentional performance necessarily requires the integrity of BF cholinergic projections but not their noradrenergic afferents^{246,247}. However, for thalamocortical information processing, ACh activation produces a

noisy broadband signal detection mode, while NE activation sets to a noise-free high-frequency signal detection mode, which seems to be more optimized for selective attention than brainstem cholinergic activation²⁴⁵.

Generally, cortical ACh-NE interaction plays a significant role in the modulation of attention²⁴⁸⁻²⁵⁰. Using fluorescent retrograde tracers in ACC, mPFC and OFC, it was uncovered that subsets of LC neurons might be responsible for modulating individual prefrontal subregions independently, yet subsets of NB neurons might produce universal influence in prefrontal subregions²⁴⁸, providing insights respecting prefrontal cortex's role of allocating attentional reserves. In a attentional set shifting task, McGaughy and colleagues pointed out that the specific impairments in animals' ability to shift attentional set were produced by noradrenergic instead of cholinergic deafferentation in prefrontal cortex²⁵⁰. Indeed, the cortical cholinergic system is very likely to be involved in aspects of established attentional performance while NE system is more competent in detecting shifts in the predictive relationship between action and reinforcement²⁴⁹.

Future studies are highly encouraged to explore the ACh-NE interaction in the contexts of their complementary roles regarding attention modulation. Systematic characterization and manipulation of the cholinergic and NE projections at biochemical, genetic, pharmacological and physiological levels would largely facilitate our understanding of the interaction between the two systems and inform the development of potential therapeutics for certain neurodegenerative and psychiatric diseases.

ACh and NE in Learning and Decision Making

The interplay between the noradrenergic and cholinergic systems in the decision making and learning processes is highly complex. Each neuromodulatory system plays a pivotal role in creating these complex cognitive brain states, integrating sensory information with positive and negative feedback loops through multiple brain regions. For example, the visual system of a macaque primate has integration and circuitry across all neuromodulators, inhibiting and exciting

different neurons, changing gene expression, and modulating synaptic circuitry²⁵¹. Due to the complexity of decision making, the unpredictability of environments, and the uncertainty of risk-taking during exploration, the relationship between neuromodulators is not yet well understood²⁵². Even with this challenge, there have been studies which attempted to look at isolated NE and ACh interaction in specific decision-making tasks. In isolated behaviors, specific neuromodulators can be seen regulating specific sub-tasks. In metalearning, NE can be seen contributing to the randomness of action selection, while ACh seems to solely dictate the speed of specific memory updates¹⁸⁵. Both neuromodulators also have been implicated in the information transmission during different behaviors^{19,253}. Theoretical modeling has been used to further understand the role both NE and ACh play in uncertainty, behaving both synergistically and antagonistically, enabling complex learning in challenging adaptive environments²⁵⁴. Understanding how both of these neuromodulators interact with one another in a variety of decision making and learning paradigms will be critical for developing translational treatments for neurological conditions that involve these neurotransmitters.

Future Directions

The functional consequences of overlapping and interacting neuromodulatory systems are as numerous as they are behaviorally important. An understanding has been slowly emerging over at least two decades that a diversity of functionally distinct circuits and heterogeneously distributed receptor subpopulations between neuromodulatory systems gives rise to many of the most interesting aspects of neural processing and adaptive behavioral outcomes. The differential influence of each neuromodulator on a specific circuit of interest is highly complex though and requires a very careful experimental framework in order to begin unraveling a more complete understanding of the influence of ACh and NE on specific behaviors. As has been demonstrated by the work covered in this review, overlapping, segmented receptive fields, non-uniform receptor distributions and the wide-range of actions of ACh and NE, in a spatially and temporally dependent

manner, make broad conclusions about each system difficult to generalize. It is critical for future work to use a modern arsenal of tools to dissect the role of these two systems in isolated circuits that have important behavioral relevance.

Given the highly region-dependent action of these neuromodulatory systems, future work may begin with the identification and isolation of a specific target. An example of such an isolation would be to examine the role of the regions comprising the mPFC in decision making^{175,255-258}. In order to understand the possible functional role of ACh and NE in modulating these regions, experiments should be devised to further characterize noradrenergic and cholinergic inputs to the mPFC. Initial experiments should also provide a basic characterization of the density and distribution of cholinergic and noradrenergic receptors in the target region using either traditional receptor expression profiling tools or the integration of new tools such as spatial single cell sequencing. Once the neuromodulatory input into the region of interest has been quantified and the range of receptors expressed summarized, an experimental framework can be established to systematically isolate one variable at a time.

Under this framework, a functionally important region of cortex can be isolated with respect to its neuromodulatory input and the relevant molecular targets identified. At this point systematic exploration of the release of NE and ACh in a representative behavioral task will provide the foundation for understanding the differential presence of each system in the specified brain region during normal behavior. More sophisticated single-unit electrophysiology or calcium imaging experiments that explore the change in network dynamics in response to neuromodulator release can be paired with selective cholinergic and noradrenergic activation or silencing to observe the response not only of the animal and the neuronal connectivity, but also in the response of the other neurotransmitter system. Systematic knock outs or implementation of inducible/repressible receptor expression in the specified region will also provide insights into how functional modules in the cortex are regulated by multiple neuromodulators.

The result of conducting many of these experiments in parallel will be an improved understanding of how broadly acting neuromodulator networks contribute to subdivided, and not necessarily unified, responses throughout the brain. Understanding the influence of ACh and NE on specific modules, and the afferent source of these neuromodulators, will provide improved targets for functional neural stimulation and greater degrees of freedom for higher bandwidth communication and dynamic network control through neural interfaces ²⁵⁹.

Conclusion

Cholinergic and noradrenergic modulation of functionally distinct regions of the brain has become one of the primary organizational principles behind understanding the contribution of each system to the diversity of neural computation in the central nervous system. Decades of work has shown that a diverse family of receptors, which stratify across different brain regions, and afferent and efferent projections that can be selectively activated, are critical in helping widespread neuromodulatory systems obtain substantial heterogeneity in the sophistication of their role in neural processes. The end result of such a complicated interplay between two diffuse modulatory systems is a dynamic and highly context dependent role for brain regions important in learning, memory, attention, and decision making. To further improve our understanding of these systems, it is essential to adopt approaches that are built on previous work to identify the interplay, at the receptor, circuit, and functional levels, between these systems in an isolated circuit that can be tied to a behaviorally functional outcome. The results will allow greater understanding and control over wide-ranging behaviors.

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References

- 1 Bianconi, E. *et al.* An estimation of the number of cells in the human body. *Ann Hum Biol* **40**, 463-471, doi:10.3109/03014460.2013.807878 (2013).
- 2 Newman, E. A. New roles for astrocytes: regulation of synaptic transmission. *Trends Neurosci* **26**, 536-542, doi:10.1016/S0166-2236(03)00237-6 (2003).
- 3 Marinelli, S., Basilico, B., Marrone, M. C. & Ragozzino, D. Microglia-neuron crosstalk: Signaling mechanism and control of synaptic transmission. *Semin Cell Dev Biol* **94**, 138-151, doi:10.1016/j.semcdb.2019.05.017 (2019).
- 4 Hyman, S. E. Neurotransmitters. *Curr Biol* **15**, R154-158, doi:10.1016/j.cub.2005.02.037 (2005).
- 5 Nedergaard, M., Takano, T. & Hansen, A. J. Beyond the role of glutamate as a neurotransmitter. *Nat Rev Neurosci* **3**, 748-755, doi:10.1038/nrn916 (2002).
- 6 Owens, D. F. & Kriegstein, A. R. Is there more to GABA than synaptic inhibition? *Nat Rev Neurosci* **3**, 715-727, doi:10.1038/nrn919 (2002).
- 7 Kurcyus, K. *et al.* Opposite Dynamics of GABA and Glutamate Levels in the Occipital Cortex during Visual Processing. *J Neurosci* **38**, 9967-9976, doi:10.1523/JNEUROSCI.1214-18.2018 (2018).
- 8 Carlsson, A. *et al.* Interactions Between Monoamines, Glutamate, and GABA in Schizophrenia: New Evidence. *Annual Review of Pharmacology and Toxicology* **41**, 237-260, doi:10.1146/annurev.pharmtox.41.1.237 (2001).
- 9 Marmigere, F., Rage, F. & Tapia-Arancibia, L. GABA-glutamate interaction in the control of BDNF expression in hypothalamic neurons. *Neurochem Int* **42**, 353-358, doi:10.1016/s0197-0186(02)00100-6 (2003).
- 10 Katz, P. & Edwards, D. *Beyond neurotransmission*. (Oxford University Press New York, 1999).
- 11 Agnati, L. F. *et al.* Volume transmission and wiring transmission from cellular to molecular networks: history and perspectives. *Acta Physiol (Oxf)* **187**, 329-344, doi:10.1111/j.1748-1716.2006.01579.x (2006).
- 12 Zaborszky, L. in *Progress in Brain Research* Vol. 136 359-372 (Elsevier, 2002).
- 13 Golmayo, L., Nunez, A. & Zaborszky, L. Electrophysiological evidence for the existence of a posterior cortical-prefrontal-basal forebrain circuitry in modulating sensory responses in visual and somatosensory rat cortical areas. *Neuroscience* **119**, 597-609, doi:10.1016/s0306-4522(03)00031-9 (2003).
- 14 Briand, L. A., Gritton, H., Howe, W. M., Young, D. A. & Sarter, M. Modulators in concert for cognition: modulator interactions in the prefrontal cortex. *Prog Neurobiol* **83**, 69-91, doi:10.1016/j.pneurobio.2007.06.007 (2007).
- 15 Blokland, A. Acetylcholine: a neurotransmitter for learning and memory? *Brain Research Reviews* **21**, 285-300 (1995).

- 16 Sarter, M. & Bruno, J. P. Cognitive functions of cortical acetylcholine: toward a unifying hypothesis. *Brain Res Brain Res Rev* **23**, 28-46, doi:10.1016/s0165-0173(96)00009-4 (1997).
- 17 Parikh, V., Kozak, R., Martinez, V. & Sarter, M. Prefrontal acetylcholine release controls cue detection on multiple timescales. *Neuron* **56**, 141-154, doi:10.1016/j.neuron.2007.08.025 (2007).
- 18 Goard, M. & Dan, Y. Basal forebrain activation enhances cortical coding of natural scenes. *Nat Neurosci* **12**, 1444-1449, doi:10.1038/nn.2402 (2009).
- 19 Rodenkirch, C. & Wang, Q. Rapid and transient enhancement of thalamic information transmission induced by vagus nerve stimulation. *J Neural Eng* **17**, 026027, doi:10.1088/1741-2552/ab6b84 (2020).
- 20 Rodenkirch, C., Liu, Y., Schriver, B. J. & Wang, Q. Locus coeruleus activation enhances thalamic feature selectivity via norepinephrine regulation of intrathalamic circuit dynamics. *Nat Neurosci* **22**, 120-133, doi:10.1038/s41593-018-0283-1 (2019).
- 21 Schwarz, L. A. & Luo, L. Organization of the locus coeruleus-norepinephrine system. *Curr Biol* **25**, R1051-R1056, doi:10.1016/j.cub.2015.09.039 (2015).
- 22 Devilbiss, D. M., Page, M. E. & Waterhouse, B. D. Locus Ceruleus Regulates Sensory Encoding by Neurons and Networks in Waking Animals. *The Journal of Neuroscience* **26**, 9860-9872, doi:10.1523/jneurosci.1776-06.2006 (2006).
- 23 Vazey, E. M., Moorman, D. E. & Aston-Jones, G. Phasic locus coeruleus activity regulates cortical encoding of salience information. *Proceedings of the National Academy of Sciences* **115**, E9439, doi:10.1073/pnas.1803716115 (2018).
- 24 Yang, H., Bari, B. A., Cohen, J. Y. & O'Connor, D. H. Locus coeruleus spiking differently correlates with S1 cortex activity and pupil diameter in a tactile detection task. *eLife* **10**, e64327, doi:10.7554/eLife.64327 (2021).
- 25 Liu, Y., Narasimhan, S., Schriver, B. J. & Wang, Q. Perceptual Behavior Depends Differently on Pupil-Linked Arousal and Heartbeat Dynamics-Linked Arousal in Rats Performing Tactile Discrimination Tasks. *Frontiers in systems neuroscience* **14**, 614248-614248, doi:10.3389/fnsys.2020.614248 (2021).
- 26 O'Dell, T. J., Connor, S. A., Guglietta, R. & Nguyen, P. V. beta-Adrenergic receptor signaling and modulation of long-term potentiation in the mammalian hippocampus. *Learn Mem* **22**, 461-471, doi:10.1101/lm.031088.113 (2015).
- 27 Spencer, R. C. & Berridge, C. W. Receptor and circuit mechanisms underlying differential procognitive actions of psychostimulants. *Neuropsychopharmacology* **44**, 1820-1827, doi:10.1038/s41386-019-0314-y (2019).
- 28 Usher, M., Cohen, J. D., Servan-Schreiber, D., Rajkowski, J. & Aston-Jones, G. The Role of Locus Coeruleus in the Regulation of Cognitive Performance. *Science* **283**, 549-554, doi:10.1126/science.283.5401.549 (1999).
- 29 Janitzky, K. *et al.* Optogenetic silencing of locus coeruleus activity in mice impairs cognitive flexibility in an attentional set-shifting task. *Frontiers in Behavioral Neuroscience* **9**, 286, doi:10.3389/fnbeh.2015.00286 (2015).
- 30 Aston-Jones, G., Rajkowski, J. & Cohen, J. in *Progress in Brain Research* Vol. Volume 126 (eds H.B.M. Uylings *et al.*) 165-182 (Elsevier, 2000).
- 31 Kalwani, R. M., Joshi, S. & Gold, J. I. Phasic Activation of Individual Neurons in the Locus Ceruleus/Subceruleus Complex of Monkeys Reflects Rewarded Decisions to Go But Not Stop. *The Journal of Neuroscience* **34**, 13656-13669, doi:10.1523/jneurosci.2566-14.2014 (2014).
- 32 Schriver, B., Bagdasarov, S. & Wang, Q. Pupil-linked arousal modulates behavior in rats performing a whisker deflection direction discrimination task. *Journal of Neurophysiology* **120**, 1655-1670, doi:10.1152/jn.00290.2018 (2018).
- 33 Schriver, B. J., Perkins, S. M., Sajda, P. & Wang, Q. Interplay between components of pupil-linked phasic arousal and its role in driving behavioral choice in Go/No-Go

perceptual decision-making. *Psychophysiology*, e13565, doi:10.1111/psyp.13565 (2020).

34 Slater, C. & Wang, Q. Alzheimer's disease: An evolving understanding of noradrenergic involvement and the promising future of electroceutical therapies. *Clinical and Translational Medicine* **11**, e397, doi:<https://doi.org/10.1002/ctm2.397> (2021).

35 Furey, M. L. The prominent role of stimulus processing: cholinergic function and dysfunction in cognition. *Curr Opin Neurol* **24**, 364-370, doi:10.1097/WCO.0b013e328348bda5 (2011).

36 Bohnen, N. I. & Albin, R. L. The cholinergic system and Parkinson disease. *Behavioural Brain Research* **221**, 564-573, doi:<https://doi.org/10.1016/j.bbr.2009.12.048> (2011).

37 Yamamoto, K.-i., Shinba, T. & Yoshii, M. Psychiatric symptoms of noradrenergic dysfunction: A pathophysiological view. *Psychiatry and Clinical Neurosciences* **68**, 1-20, doi:<https://doi.org/10.1111/pcn.12126> (2014).

38 Marien, M. R., Colpaert, F. C. & Rosenquist, A. C. Noradrenergic mechanisms in neurodegenerative diseases: a theory. *Brain Research Reviews* **45**, 38-78, doi:<https://doi.org/10.1016/j.brainresrev.2004.02.002> (2004).

39 Doppler, C. E. J. *et al.* Microsleep disturbances are associated with noradrenergic dysfunction in Parkinson's disease. *Sleep* **44**, doi:10.1093/sleep/zsab040 (2021).

40 Weinshenker, D. Long Road to Ruin: Noradrenergic Dysfunction in Neurodegenerative Disease. *Trends in Neurosciences* **41**, 211-223, doi:<https://doi.org/10.1016/j.tins.2018.01.010> (2018).

41 Gannon, M. *et al.* Noradrenergic dysfunction in Alzheimer's disease. *Frontiers in Neuroscience* **9** (2015).

42 Sarter, M. & Bruno, J. P. Cortical cholinergic inputs mediating arousal, attentional processing and dreaming: differential afferent regulation of the basal forebrain by telencephalic and brainstem afferents. *Neuroscience* **95**, 933-952, doi:[https://doi.org/10.1016/S0306-4522\(99\)00487-X](https://doi.org/10.1016/S0306-4522(99)00487-X) (1999).

43 Lehmann, J., Nagy, J. I., Atmadia, S. & Fibiger, H. C. The nucleus basalis magnocellularis: the origin of a cholinergic projection to the neocortex of the rat. *Neuroscience* **5**, 1161-1174, doi:10.1016/0306-4522(80)90195-5 (1980).

44 Levey, A. I., Hallanger, A. E. & Wainer, B. H. Cholinergic nucleus basalis neurons may influence the cortex via the thalamus. *Neuroscience Letters* **74**, 7-13, doi:[https://doi.org/10.1016/0304-3940\(87\)90042-5](https://doi.org/10.1016/0304-3940(87)90042-5) (1987).

45 Bergado, J. A., Frey, S., Lopez, J., Almaguer-Melian, W. & Frey, J. U. Cholinergic afferents to the locus coeruleus and noradrenergic afferents to the medial septum mediate LTP-reinforcement in the dentate gyrus by stimulation of the amygdala. *Neurobiol Learn Mem* **88**, 331-341, doi:10.1016/j.nlm.2007.05.003 (2007).

46 Satoh, K. & Fibiger, H. C. Cholinergic neurons of the laterodorsal tegmental nucleus: efferent and afferent connections. *J Comp Neurol* **253**, 277-302, doi:10.1002/cne.902530302 (1986).

47 Mesulam, M. M. Cholinergic circuitry of the human nucleus basalis and its fate in Alzheimer's disease. *J Comp Neurol* **521**, 4124-4144, doi:10.1002/cne.23415 (2013).

48 Higo, S., Matsuyama, T. & Kawamura, S. Direct projections from the pedunculo pontine and laterodorsal tegmental nuclei to area 17 of the visual cortex in the cat. *Neurosci Res* **26**, 109-118, doi:10.1016/s0168-0102(96)01079-6 (1996).

49 Bloem, B. *et al.* Topographic mapping between basal forebrain cholinergic neurons and the medial prefrontal cortex in mice. *J Neurosci* **34**, 16234-16246, doi:10.1523/JNEUROSCI.3011-14.2014 (2014).

50 Li, X. *et al.* Generation of a whole-brain atlas for the cholinergic system and mesoscopic projectome analysis of basal forebrain cholinergic neurons. *Proc Natl Acad Sci U S A* **115**, 415-420, doi:10.1073/pnas.1703601115 (2018).

- 51 Woolf, N. J. Cholinergic systems in mammalian brain and spinal cord. *Prog Neurobiol* **37**, 475-524, doi:10.1016/0301-0082(91)90006-m (1991).
- 52 Rye, D. B., Wainer, B. H., Mesulam, M. M., Mufson, E. J. & Saper, C. B. Cortical projections arising from the basal forebrain: a study of cholinergic and noncholinergic components employing combined retrograde tracing and immunohistochemical localization of choline acetyltransferase. *Neuroscience* **13**, 627-643, doi:10.1016/0306-4522(84)90083-6 (1984).
- 53 Kaneko, S. *et al.* Synaptic integration mediated by striatal cholinergic interneurons in basal ganglia function. *Science* **289**, 633-637, doi:10.1126/science.289.5479.633 (2000).
- 54 Bickford, M. E., Gunluk, A. E., Van Horn, S. C. & Sherman, S. M. GABAergic projection from the basal forebrain to the visual sector of the thalamic reticular nucleus in the cat. *J Comp Neurol* **348**, 481-510, doi:10.1002/cne.903480402 (1994).
- 55 van der Zee, E. A. & Luiten, P. G. Muscarinic acetylcholine receptors in the hippocampus, neocortex and amygdala: a review of immunocytochemical localization in relation to learning and memory. *Prog Neurobiol* **58**, 409-471, doi:10.1016/s0301-0082(98)00092-6 (1999).
- 56 Kim, J.-H. *et al.* Selectivity of Neuromodulatory Projections from the Basal Forebrain and Locus Ceruleus to Primary Sensory Cortices. *The Journal of Neuroscience* **36**, 5314-5327, doi:10.1523/jneurosci.4333-15.2016 (2016).
- 57 Chavez, C. & Zaborszky, L. Basal Forebrain Cholinergic-Auditory Cortical Network: Primary Versus Nonprimary Auditory Cortical Areas. *Cereb Cortex* **27**, 2335-2347, doi:10.1093/cercor/bhw091 (2017).
- 58 Fournier, G. N., Semba, K. & Rasmusson, D. D. Modality- and region-specific acetylcholine release in the rat neocortex. *Neuroscience* **126**, 257-262, doi:<https://doi.org/10.1016/j.neuroscience.2004.04.002> (2004).
- 59 Liu, A. K. L. & Gentleman, S. M. in *Handbook of Clinical Neurology* Vol. 179 (eds Dick F. Swaab *et al.*) 175-187 (Elsevier, 2021).
- 60 Mechawar, N., Cozzari, C. & Descarries, L. Cholinergic innervation in adult rat cerebral cortex: A quantitative immunocytochemical description. *Journal of Comparative Neurology* **428**, 305-318, doi:[https://doi.org/10.1002/1096-9861\(20001211\)428:2<305::AID-CNE9>3.0.CO;2-Y](https://doi.org/10.1002/1096-9861(20001211)428:2<305::AID-CNE9>3.0.CO;2-Y) (2000).
- 61 Smiley, J. F. & Mesulam, M. M. Cholinergic neurons of the nucleus basalis of Meynert receive cholinergic, catecholaminergic and GABAergic synapses: an electron microscopic investigation in the monkey. *Neuroscience* **88**, 241-255, doi:10.1016/s0306-4522(98)00202-4 (1999).
- 62 Mesulam, M. M. The cholinergic innervation of the human cerebral cortex. *Prog Brain Res* **145**, 67-78, doi:10.1016/S0079-6123(03)45004-8 (2004).
- 63 Samuels, E. R. & Szabadi, E. Functional neuroanatomy of the noradrenergic locus coeruleus: its roles in the regulation of arousal and autonomic function part II: physiological and pharmacological manipulations and pathological alterations of locus coeruleus activity in humans. *Current neuropharmacology* **6**, 254-285, doi:10.2174/157015908785777193 (2008).
- 64 Jones, B. E. & Cuello, A. C. Afferents to the basal forebrain cholinergic cell area from pontomesencephalic--catecholamine, serotonin, and acetylcholine--neurons. *Neuroscience* **31**, 37-61, doi:10.1016/0306-4522(89)90029-8 (1989).
- 65 Khateb, A. *et al.* GABAergic input to cholinergic nucleus basalis neurons. *Neuroscience* **86**, 937-947, doi:10.1016/s0306-4522(98)00094-3 (1998).
- 66 Rodriguez-Garcia, G. & Miranda, M. I. Opposing Roles of Cholinergic and GABAergic Activity in the Insular Cortex and Nucleus Basalis Magnocellularis during Novel Recognition and Familiar Taste Memory Retrieval. *J Neurosci* **36**, 1879-1889, doi:10.1523/JNEUROSCI.2340-15.2016 (2016).

67 Casamenti, F., Deffenu, G., Abbamondi, A. L. & Pepeu, G. Changes in cortical acetylcholine output induced by modulation of the nucleus basalis. *Brain Res Bull* **16**, 689-695, doi:10.1016/0361-9230(86)90140-1 (1986).

68 Semba, K., Reiner, P. B., McGeer, E. G. & Fibiger, H. C. Brainstem afferents to the magnocellular basal forebrain studied by axonal transport, immunohistochemistry, and electrophysiology in the rat. *J Comp Neurol* **267**, 433-453, doi:10.1002/cne.902670311 (1988).

69 Semba, K. & Fibiger, H. C. Afferent connections of the laterodorsal and the pedunclopontine tegmental nuclei in the rat: a retro- and antero-grade transport and immunohistochemical study. *J Comp Neurol* **323**, 387-410, doi:10.1002/cne.903230307 (1992).

70 Brudzynski, S. M., Kadishevitz, L. & Fu, X. W. Mesolimbic component of the ascending cholinergic pathways: electrophysiological-pharmacological study. *J Neurophysiol* **79**, 1675-1686, doi:10.1152/jn.1998.79.4.1675 (1998).

71 Martinez-Gonzalez, C., Bolam, J. P. & Mena-Segovia, J. Topographical organization of the pedunclopontine nucleus. *Front Neuroanat* **5**, 22, doi:10.3389/fnana.2011.00022 (2011).

72 Matsumura, M. *et al.* Organization of somatic motor inputs from the frontal lobe to the pedunclopontine tegmental nucleus in the macaque monkey. *Neuroscience* **98**, 97-110, doi:10.1016/s0306-4522(00)00099-3 (2000).

73 Schofield, B. R. & Motts, S. D. Projections from auditory cortex to cholinergic cells in the midbrain tegmentum of guinea pigs. *Brain Res Bull* **80**, 163-170, doi:10.1016/j.brainresbull.2009.06.015 (2009).

74 Florio, T. *et al.* High-frequency stimulation of the subthalamic nucleus modulates the activity of pedunclopontine neurons through direct activation of excitatory fibres as well as through indirect activation of inhibitory pallidal fibres in the rat. *Eur J Neurosci* **25**, 1174-1186, doi:10.1111/j.1460-9568.2007.05360.x (2007).

75 Granata, A. R. & Kitai, S. T. Intracellular analysis of excitatory subthalamic inputs to the pedunclopontine neurons. *Brain Res* **488**, 57-72, doi:10.1016/0006-8993(89)90693-8 (1989).

76 Granata, A. R. & Kitai, S. T. Inhibitory substantia nigra inputs to the pedunclopontine neurons. *Exp Brain Res* **86**, 459-466, doi:10.1007/BF00230520 (1991).

77 Hammond, C., Rouzaire-Dubois, B., Feger, J., Jackson, A. & Crossman, A. R. Anatomical and electrophysiological studies on the reciprocal projections between the subthalamic nucleus and nucleus tegmenti pedunclopontinus in the rat. *Neuroscience* **9**, 41-52, doi:10.1016/0306-4522(83)90045-3 (1983).

78 Smith, Y., Bolam, J. P. & Von Krosigk, M. Topographical and Synaptic Organization of the GABA-Containing Pallidosubthalamic Projection in the Rat. *Eur J Neurosci* **2**, 500-511, doi:10.1111/j.1460-9568.1990.tb00441.x (1990).

79 Moriizumi, T. & Hattori, T. Separate neuronal populations of the rat globus pallidus projecting to the subthalamic nucleus, auditory cortex and pedunclopontine tegmental area. *Neuroscience* **46**, 701-710, doi:10.1016/0306-4522(92)90156-v (1992).

80 Haber, S. N., Lynd, E., Klein, C. & Groenewegen, H. J. Topographic organization of the ventral striatal efferent projections in the rhesus monkey: an anterograde tracing study. *J Comp Neurol* **293**, 282-298, doi:10.1002/cne.902930210 (1990).

81 Noda, T. & Oka, H. Nigral inputs to the pedunclopontine region: intracellular analysis. *Brain Res* **322**, 332-336, doi:10.1016/0006-8993(84)90128-8 (1984).

82 Jones, B. E. & Yang, T. Z. The efferent projections from the reticular formation and the locus coeruleus studied by anterograde and retrograde axonal transport in the rat. *J Comp Neurol* **242**, 56-92, doi:10.1002/cne.902420105 (1985).

83 Steininger, T. L., Rye, D. B. & Wainer, B. H. Afferent projections to the cholinergic pedunclopontine tegmental nucleus and adjacent midbrain extrapyramidal area in the

- albino rat. I. Retrograde tracing studies. *J Comp Neurol* **321**, 515-543, doi:10.1002/cne.903210403 (1992).
- 84 Segal, M. Brain stem afferents to the rat medial septum. *J Physiol* **261**, 617-631, doi:10.1113/jphysiol.1976.sp011577 (1976).
- 85 Vertes, R. P. Brainstem afferents to the basal forebrain in the rat. *Neuroscience* **24**, 907-935, doi:10.1016/0306-4522(88)90077-2 (1988).
- 86 Borhegyi, Z., Magloczky, Z., Acsady, L. & Freund, T. F. The supramammillary nucleus innervates cholinergic and GABAergic neurons in the medial septum-diagonal band of Broca complex. *Neuroscience* **82**, 1053-1065, doi:10.1016/s0306-4522(97)00301-1 (1998).
- 87 Steriade, M., Datta, S., Pare, D., Oakson, G. & Curro Dossi, R. C. Neuronal activities in brain-stem cholinergic nuclei related to tonic activation processes in thalamocortical systems. *J Neurosci* **10**, 2541-2559 (1990).
- 88 Steriade, M., Parent, A., Pare, D. & Smith, Y. Cholinergic and non-cholinergic neurons of cat basal forebrain project to reticular and mediodorsal thalamic nuclei. *Brain Res* **408**, 372-376, doi:10.1016/0006-8993(87)90408-2 (1987).
- 89 Mesulam, M. M., Mufson, E. J., Levey, A. I. & Wainer, B. H. Cholinergic innervation of cortex by the basal forebrain: cytochemistry and cortical connections of the septal area, diagonal band nuclei, nucleus basalis (substantia innominata), and hypothalamus in the rhesus monkey. *J Comp Neurol* **214**, 170-197, doi:10.1002/cne.902140206 (1983).
- 90 Steckler, T., Inglis, W., Winn, P. & Sahgal, A. The pedunculo pontine tegmental nucleus: a role in cognitive processes? *Brain Res Brain Res Rev* **19**, 298-318, doi:10.1016/0165-0173(94)90016-7 (1994).
- 91 Mena-Segovia, J. & Bolam, J. P. Rethinking the Pedunculo pontine Nucleus: From Cellular Organization to Function. *Neuron* **94**, 7-18, doi:10.1016/j.neuron.2017.02.027 (2017).
- 92 Kami, K., Tajima, F. & Senba, E. Activation of mesolimbic reward system via laterodorsal tegmental nucleus and hypothalamus in exercise-induced hypoalgesia. *Sci Rep* **8**, 11540, doi:10.1038/s41598-018-29915-4 (2018).
- 93 Grzanna, R. & Molliver, M. E. The locus coeruleus in the rat: an immunohistochemical delineation. *Neuroscience* **5**, 21-40, doi:10.1016/0306-4522(80)90068-8 (1980).
- 94 Swanson, L. W. & Hartman, B. K. The central adrenergic system. An immunofluorescence study of the location of cell bodies and their efferent connections in the rat utilizing dopamine-beta-hydroxylase as a marker. *J Comp Neurol* **163**, 467-505, doi:10.1002/cne.901630406 (1975).
- 95 Sara, S. J. & Bouret, S. Orienting and reorienting: the locus coeruleus mediates cognition through arousal. *Neuron* **76**, 130-141, doi:10.1016/j.neuron.2012.09.011 (2012).
- 96 Foote, S. L., Bloom, F. E. & Aston-Jones, G. Nucleus locus ceruleus: new evidence of anatomical and physiological specificity. *Physiological Reviews* **63**, 844-914, doi:10.1152/physrev.1983.63.3.844 (1983).
- 97 Dahlström, A. & Fuxe, K. *Demonstration of monoamines in the cell bodies of brain stem neurons*. (Verlag nicht ermittelbar, 1964).
- 98 Poe, G. R. *et al.* Locus coeruleus: a new look at the blue spot. *Nature Reviews Neuroscience* **21**, 644-659, doi:10.1038/s41583-020-0360-9 (2020).
- 99 Fallon, J. H., Koziell, D. A. & Moore, R. Y. Catecholamine innervation of the basal forebrain. II. Amygdala, suprarhinal cortex and entorhinal cortex. *J Comp Neurol* **180**, 509-532, doi:10.1002/cne.901800308 (1978).
- 100 Morrison, J. H., Molliver, M. E., Grzanna, R. & Coyle, J. T. Noradrenergic innervation patterns in three regions of medial cortex: an immunofluorescence characterization. *Brain Res Bull* **4**, 849-857, doi:10.1016/0361-9230(79)90022-4 (1979).

- 101 Berridge, C. W. & Waterhouse, B. D. The locus coeruleus-noradrenergic system: modulation of behavioral state and state-dependent cognitive processes. *Brain Res Brain Res Rev* **42**, 33-84, doi:10.1016/s0165-0173(03)00143-7 (2003).
- 102 Agster, K. L., Mejias-Aponte, C. A., Clark, B. D. & Waterhouse, B. D. Evidence for a regional specificity in the density and distribution of noradrenergic varicosities in rat cortex. *J Comp Neurol* **521**, 2195-2207, doi:10.1002/cne.23270 (2013).
- 103 Giustino, T. F. & Maren, S. Noradrenergic Modulation of Fear Conditioning and Extinction. *Front Behav Neurosci* **12**, 43, doi:10.3389/fnbeh.2018.00043 (2018).
- 104 Jänig, W. *Integrative Action of the Autonomic Nervous System: Neurobiology of Homeostasis*. (Cambridge University Press, 2006).
- 105 Farmer, D. G. S. *et al.* On the presence and functional significance of sympathetic premotor neurons with collateralized spinal axons in the rat. *J Physiol* **597**, 3407-3423, doi:10.1113/JP277661 (2019).
- 106 Totah, N. K. B., Logothetis, N. K. & Eschenko, O. Noradrenergic ensemble-based modulation of cognition over multiple timescales. *Brain Res* **1709**, 50-66, doi:10.1016/j.brainres.2018.12.031 (2019).
- 107 Shipley, M. T., Fu, L., Ennis, M., Liu, W.-L. & Aston-Jones, G. Dendrites of locus coeruleus neurons extend preferentially into two pericoerulear zones. *Journal of Comparative Neurology* **365**, 56-68, doi:[https://doi.org/10.1002/\(SICI\)1096-9861\(19960129\)365:1<56::AID-CNE5>3.0.CO;2-I](https://doi.org/10.1002/(SICI)1096-9861(19960129)365:1<56::AID-CNE5>3.0.CO;2-I) (1996).
- 108 Schwarz, L. A. *et al.* Viral-genetic tracing of the input-output organization of a central noradrenaline circuit. *Nature* **524**, 88-92, doi:10.1038/nature14600 (2015).
- 109 Totah, N. K., Neves, R. M., Panzeri, S., Logothetis, N. K. & Eschenko, O. The Locus Coeruleus Is a Complex and Differentiated Neuromodulatory System. *Neuron* **99**, 1055-1068.e1056, doi:<https://doi.org/10.1016/j.neuron.2018.07.037> (2018).
- 110 Aston-Jones, G., Ennis, M., Pieribone, V. A., Nickell, W. T. & Shipley, M. T. The brain nucleus locus coeruleus: restricted afferent control of a broad efferent network. *Science* **234**, 734-737 (1986).
- 111 Cedarbaum, J. M. & Aghajanian, G. K. Afferent projections to the rat locus coeruleus as determined by a retrograde tracing technique. *Journal of Comparative Neurology* **178**, 1-15, doi:<https://doi.org/10.1002/cne.901780102> (1978).
- 112 Aston-Jones, G. *et al.* in *Progress in Brain Research* Vol. 88 (eds C. D. Barnes & O. Pompeiano) 47-75 (Elsevier, 1991).
- 113 De Ciccio, V. *et al.* Trigeminal, Visceral and Vestibular Inputs May Improve Cognitive Functions by Acting through the Locus Coeruleus and the Ascending Reticular Activating System: A New Hypothesis. *Frontiers in neuroanatomy* **11**, doi:10.3389/fnana.2017.00130 (2018).
- 114 Tramonti Fantozzi, M. P. *et al.* Trigeminal input, pupil size and cognitive performance: From oral to brain matter. *Brain Research* **1751**, 147194, doi:<https://doi.org/10.1016/j.brainres.2020.147194> (2021).
- 115 Rodenkirch, C., Carmel, J. & Wang, Q. Rapid Effects of Vagus Nerve Stimulation on Sensory Processing Through Activation of Neuromodulatory Systems. *Frontiers in Neuroscience* **in press**, doi:10.3389/fnins.2022.922424 (2022).
- 116 Tramonti Fantozzi, M. P. *et al.* Chewing and Cognitive Improvement: The Side Matters. *Frontiers in Systems Neuroscience* **15** (2021).
- 117 Luppi, P. H., Aston-Jones, G., Akaoka, H., Chouvet, G. & Jouvet, M. Afferent projections to the rat locus coeruleus demonstrated by retrograde and anterograde tracing with cholera-toxin B subunit and Phaseolus vulgaris leucoagglutinin. *Neuroscience* **65**, 119-160 (1995).
- 118 Abrams, P. *et al.* Muscarinic receptors: their distribution and function in body systems, and the implications for treating overactive bladder. *Br J Pharmacol* **148**, 565-578, doi:10.1038/sj.bjp.0706780 (2006).

- 119 Breton-Provencher, V. & Sur, M. Active control of arousal by a locus coeruleus GABAergic circuit. *Nature Neuroscience* **22**, 218-228, doi:10.1038/s41593-018-0305-z (2019).
- 120 Lewis, D. A. & Morrison, J. H. Noradrenergic innervation of monkey prefrontal cortex: a dopamine-beta-hydroxylase immunohistochemical study. *J Comp Neurol* **282**, 317-330, doi:10.1002/cne.902820302 (1989).
- 121 Morrison, J. H. & Foote, S. L. Noradrenergic and serotonergic innervation of cortical, thalamic, and tectal visual structures in Old and New World monkeys. *J Comp Neurol* **243**, 117-138, doi:10.1002/cne.902430110 (1986).
- 122 Kebschull, J. M. *et al.* High-Throughput Mapping of Single-Neuron Projections by Sequencing of Barcoded RNA. *Neuron* **91**, 975-987, doi:10.1016/j.neuron.2016.07.036 (2016).
- 123 Hirschberg, S., Li, Y., Randall, A., Kremer, E. J. & Pickering, A. E. Functional dichotomy in spinal- vs prefrontal-projecting locus coeruleus modules splits descending noradrenergic analgesia from ascending aversion and anxiety in rats. *Elife* **6**, doi:10.7554/eLife.29808 (2017).
- 124 Chandler, D. J., Gao, W. J. & Waterhouse, B. D. Heterogeneous organization of the locus coeruleus projections to prefrontal and motor cortices. *Proc Natl Acad Sci U S A* **111**, 6816-6821, doi:10.1073/pnas.1320827111 (2014).
- 125 Zerbi, V. *et al.* Rapid Reconfiguration of the Functional Connectome after Chemogenetic Locus Coeruleus Activation. *Neuron* **103**, 702-718 e705, doi:10.1016/j.neuron.2019.05.034 (2019).
- 126 Liu, Y., Rodenkirch, C., Moskowitz, N., Schriver, B. & Wang, Q. Dynamic Lateralization of Pupil Dilation Evoked by Locus Coeruleus Activation Results from Sympathetic, Not Parasympathetic, Contributions. *Cell reports* **20**, 3099–3112, doi:<https://doi.org/10.1016/j.celrep.2017.08.094> (2017).
- 127 Robertson, S. D., Plummer, N. W., de Marchena, J. & Jensen, P. Developmental origins of central norepinephrine neuron diversity. *Nat Neurosci* **16**, 1016-1023, doi:10.1038/nn.3458 (2013).
- 128 Jones, B. E. in *Progress in Brain Research* Vol. 88 (eds C. D. Barnes & O. Pompeiano) 15-30 (Elsevier, 1991).
- 129 Williams, J. A. & Reiner, P. B. Noradrenaline hyperpolarizes identified rat mesopontine cholinergic neurons in vitro. *J Neurosci* **13**, 3878-3883 (1993).
- 130 Pisani, A. *et al.* Activation of β 1-Adrenoceptors Excites Striatal Cholinergic Interneurons through a cAMP-Dependent, Protein Kinase-Independent Pathway. *The Journal of Neuroscience* **23**, 5272-5282, doi:10.1523/jneurosci.23-12-05272.2003 (2003).
- 131 Manns, I. D., Lee, M. G., Modirrousta, M., Hou, Y. P. & Jones, B. E. Alpha 2 adrenergic receptors on GABAergic, putative sleep-promoting basal forebrain neurons. *European Journal of Neuroscience* **18**, 723-727, doi:<https://doi.org/10.1046/j.1460-9568.2003.02788.x> (2003).
- 132 Szabadi, E. Functional neuroanatomy of the central noradrenergic system. *Journal of Psychopharmacology* **27**, 659-693, doi:10.1177/0269881113490326 (2013).
- 133 Carter, M. E. *et al.* Tuning arousal with optogenetic modulation of locus coeruleus neurons. *Nature Neuroscience* **13**, 1526-1533, doi:<https://doi.org/10.1038/nn.2682> (2010).
- 134 Xu, M. *et al.* Basal forebrain circuit for sleep-wake control. *Nature Neuroscience* **18**, 1641-1647, doi:10.1038/nn.4143 (2015).
- 135 Rho, H.-J., Kim, J.-H. & Lee, S.-H. Function of Selective Neuromodulatory Projections in the Mammalian Cerebral Cortex: Comparison Between Cholinergic and Noradrenergic Systems. *Frontiers in Neural Circuits* **12**, doi:10.3389/fncir.2018.00047 (2018).

- 136 Lena, C. *et al.* Diversity and distribution of nicotinic acetylcholine receptors in the locus
ceruleus neurons. *Proc Natl Acad Sci U S A* **96**, 12126-12131,
doi:10.1073/pnas.96.21.12126 (1999).
- 137 Woo, R. S. *et al.* Mechanism of nicotine-evoked release of 3H-noradrenaline in human
cerebral cortex slices. *Br J Pharmacol* **137**, 1063-1070, doi:10.1038/sj.bjp.0704975
(2002).
- 138 El-Etri, M. M., Ennis, M., Griff, E. R. & Shipley, M. T. Evidence for cholinergic regulation
of basal norepinephrine release in the rat olfactory bulb. *Neuroscience* **93**, 611-617,
doi:[https://doi.org/10.1016/S0306-4522\(99\)00169-4](https://doi.org/10.1016/S0306-4522(99)00169-4) (1999).
- 139 Lomax, P., Foster, R. S. & Kirkpatrick, W. E. Cholinergic and adrenergic interactions in
the thermoregulatory centers of the rat. *Brain Research* **15**, 431-438,
doi:[https://doi.org/10.1016/0006-8993\(69\)90165-6](https://doi.org/10.1016/0006-8993(69)90165-6) (1969).
- 140 Brown, V. J. & Bowman, E. M. Rodent models of prefrontal cortical function. *Trends*
Neurosci **25**, 340-343, doi:10.1016/s0166-2236(02)02164-1 (2002).
- 141 Dalley, J. W., Cardinal, R. N. & Robbins, T. W. Prefrontal executive and cognitive
functions in rodents: neural and neurochemical substrates. *Neurosci Biobehav Rev* **28**,
771-784, doi:10.1016/j.neubiorev.2004.09.006 (2004).
- 142 Furuyashiki, T. & Gallagher, M. Neural encoding in the orbitofrontal cortex related to
goal-directed behavior. *Ann N Y Acad Sci* **1121**, 193-215, doi:10.1196/annals.1401.037
(2007).
- 143 Robbins, T. W. Chemical neuromodulation of frontal-executive functions in humans and
other animals. *Exp Brain Res* **133**, 130-138, doi:10.1007/s002210000407 (2000).
- 144 Chandler, D. J., Lamperski, C. S. & Waterhouse, B. D. Identification and distribution of
projections from monoaminergic and cholinergic nuclei to functionally differentiated
subregions of prefrontal cortex. *Brain Res* **1522**, 38-58,
doi:10.1016/j.brainres.2013.04.057 (2013).
- 145 Eisenach, J. C., Detweiler, D. J., Tong, C., D'Angelo, R. & Hood, D. D. Cerebrospinal
Fluid Norepinephrine and Acetylcholine Concentrations During Acute Pain. *Anesthesia &*
Analgesia **82** (1996).
- 146 Decker, M. W. & McGaugh, J. L. The role of interactions between the cholinergic system
and other neuromodulatory systems in learning and memory. *Synapse* **7**, 151-168,
doi:<https://doi.org/10.1002/syn.890070209> (1991).
- 147 Mason, S. T. & Fibiger, H. C. Interaction between noradrenergic and cholinergic systems
in the rat brain: Behavioural function in locomotor activity. *Neuroscience* **4**, 517-525,
doi:[https://doi.org/10.1016/0306-4522\(79\)90128-3](https://doi.org/10.1016/0306-4522(79)90128-3) (1979).
- 148 Beane, M. & Marrocco, R. T. Norepinephrine and acetylcholine mediation of the
components of reflexive attention: implications for attention deficit disorders. *Progress in*
Neurobiology **74**, 167-181, doi:<https://doi.org/10.1016/j.pneurobio.2004.09.001> (2004).
- 149 Wotton, C. A., Cross, C. D. & Bekar, L. K. Serotonin, norepinephrine, and acetylcholine
differentially affect astrocytic potassium clearance to modulate somatosensory signaling
in male mice. *Journal of Neuroscience Research* **98**, 964-977,
doi:<https://doi.org/10.1002/jnr.24597> (2020).
- 150 Barik, J. & Wonnacott, S. Indirect Modulation by $\alpha 7$ Nicotinic Acetylcholine Receptors of
Noradrenaline Release in Rat Hippocampal Slices: Interaction with Glutamate and
GABA Systems and Effect of Nicotine Withdrawal. *Molecular Pharmacology* **69**, 618-
628, doi:10.1124/mol.105.018184 (2006).
- 151 Summers, K. L. & Giacobini, E. Effects of local and repeated systemic administration of
(-)nicotine on extracellular levels of acetylcholine, norepinephrine, dopamine, and
serotonin in rat cortex. *Neurochemical Research* **20**, 753-759, doi:10.1007/BF01705545
(1995).
- 152 Dani, J. A. Overview of nicotinic receptors and their roles in the central nervous system.
Biol Psychiatry **49**, 166-174, doi:10.1016/s0006-3223(00)01011-8 (2001).

- 153 Colquhoun, L. M. & Patrick, J. W. Pharmacology of neuronal nicotinic acetylcholine
receptor subtypes. *Adv Pharmacol* **39**, 191-220, doi:10.1016/s1054-3589(08)60072-1
(1997).
- 154 Le Novere, N. & Changeux, J. P. Molecular evolution of the nicotinic acetylcholine
receptor: an example of multigene family in excitable cells. *J Mol Evol* **40**, 155-172,
doi:10.1007/BF00167110 (1995).
- 155 McGehee, D. S. & Role, L. W. Physiological diversity of nicotinic acetylcholine receptors
expressed by vertebrate neurons. *Annu Rev Physiol* **57**, 521-546,
doi:10.1146/annurev.ph.57.030195.002513 (1995).
- 156 Charpentier, E., Barneoud, P., Moser, P., Besnard, F. & Sgard, F. Nicotinic acetylcholine
subunit mRNA expression in dopaminergic neurons of the rat substantia nigra and
ventral tegmental area. *Neuroreport* **9**, 3097-3101, doi:10.1097/00001756-199809140-
00033 (1998).
- 157 Tribollet, E., Bertrand, D., Marguerat, A. & Raggenbass, M. Comparative distribution of
nicotinic receptor subtypes during development, adulthood and aging: an
autoradiographic study in the rat brain. *Neuroscience* **124**, 405-420,
doi:10.1016/j.neuroscience.2003.09.028 (2004).
- 158 Alkondon, M., Pereira, E. F., Eisenberg, H. M. & Albuquerque, E. X. Nicotinic receptor
activation in human cerebral cortical interneurons: a mechanism for inhibition and
disinhibition of neuronal networks. *J Neurosci* **20**, 66-75 (2000).
- 159 Frazier, C. J., Buhler, A. V., Weiner, J. L. & Dunwiddie, T. V. Synaptic potentials
mediated via alpha-bungarotoxin-sensitive nicotinic acetylcholine receptors in rat
hippocampal interneurons. *J Neurosci* **18**, 8228-8235 (1998).
- 160 Porter, J. T. *et al.* Selective excitation of subtypes of neocortical interneurons by nicotinic
receptors. *J Neurosci* **19**, 5228-5235 (1999).
- 161 Levy, R. B. & Aoki, C. Alpha7 nicotinic acetylcholine receptors occur at postsynaptic
densities of AMPA receptor-positive and -negative excitatory synapses in rat sensory
cortex. *J Neurosci* **22**, 5001-5015 (2002).
- 162 Sihver, W., Gillberg, P. G. & Nordberg, A. Laminar distribution of nicotinic receptor
subtypes in human cerebral cortex as determined by [3H](-)nicotine, [3H]cytisine and
[3H]epibatidine in vitro autoradiography. *Neuroscience* **85**, 1121-1133,
doi:10.1016/s0306-4522(97)00652-0 (1998).
- 163 Albuquerque, E. X. *et al.* Properties of neuronal nicotinic acetylcholine receptors:
pharmacological characterization and modulation of synaptic function. *J Pharmacol Exp
Ther* **280**, 1117-1136 (1997).
- 164 Smythies, J. *The Neuromodulators*. Vol. 64 (2005).
- 165 Adem, A., Jolkkonen, M., Bogdanovic, N., Islam, A. & Karlsson, E. Localization of M1
muscarinic receptors in rat brain using selective muscarinic toxin-1. *Brain Res Bull* **44**,
597-601, doi:10.1016/s0361-9230(97)00281-5 (1997).
- 166 Mrzljak, L., Levey, A. I., Belcher, S. & Goldman-Rakic, P. S. Localization of the m2
muscarinic acetylcholine receptor protein and mRNA in cortical neurons of the normal
and cholinergically deafferented rhesus monkey. *J Comp Neurol* **390**, 112-132 (1998).
- 167 Wei, J., Walton, E. A., Milici, A. & Buccafusco, J. J. m1-m5 muscarinic receptor
distribution in rat CNS by RT-PCR and HPLC. *J Neurochem* **63**, 815-821,
doi:10.1046/j.1471-4159.1994.63030815.x (1994).
- 168 Marino, M. J., Rouse, S. T., Levey, A. I., Potter, L. T. & Conn, P. J. Activation of the
genetically defined m1 muscarinic receptor potentiates N-methyl-D-aspartate (NMDA)
receptor currents in hippocampal pyramidal cells. *Proc Natl Acad Sci U S A* **95**, 11465-
11470, doi:10.1073/pnas.95.19.11465 (1998).
- 169 Fisahn, A. *et al.* Muscarinic induction of hippocampal gamma oscillations requires
coupling of the M1 receptor to two mixed cation currents. *Neuron* **33**, 615-624,
doi:10.1016/s0896-6273(02)00587-1 (2002).

- 170 Hamilton, S. E. & Nathanson, N. M. The M1 receptor is required for muscarinic activation
of mitogen-activated protein (MAP) kinase in murine cerebral cortical neurons. *J Biol
Chem* **276**, 15850-15853, doi:10.1074/jbc.M011563200 (2001).
- 171 Berkeley, J. L. *et al.* M1 muscarinic acetylcholine receptors activate extracellular signal-
regulated kinase in CA1 pyramidal neurons in mouse hippocampal slices. *Mol Cell
Neurosci* **18**, 512-524, doi:10.1006/mcne.2001.1042 (2001).
- 172 Berkeley, J. L. & Levey, A. I. Muscarinic activation of mitogen-activated protein kinase in
PC12 cells. *J Neurochem* **75**, 487-493, doi:10.1046/j.1471-4159.2000.0750487.x (2000).
- 173 Naude, J., Dongelmans, M. & Faure, P. Nicotinic alteration of decision-making.
Neuropharmacology **96**, 244-254, doi:10.1016/j.neuropharm.2014.11.021 (2015).
- 174 Kolokotroni, K. Z., Rodgers, R. J. & Harrison, A. A. Acute nicotine increases both
impulsive choice and behavioural disinhibition in rats. *Psychopharmacology (Berl)* **217**,
455-473, doi:10.1007/s00213-011-2296-2 (2011).
- 175 Pittaras, E. C. *et al.* Neuronal Nicotinic Receptors Are Crucial for Tuning of E/I Balance
in Prelimbic Cortex and for Decision-Making Processes. *Front Psychiatry* **7**, 171,
doi:10.3389/fpsy.2016.00171 (2016).
- 176 Thomsen, M. S., Hansen, H. H., Timmerman, D. B. & Mikkelsen, J. D. Cognitive
improvement by activation of alpha7 nicotinic acetylcholine receptors: from animal
models to human pathophysiology. *Curr Pharm Des* **16**, 323-343,
doi:10.2174/138161210790170094 (2010).
- 177 Mendez, I. A., Gilbert, R. J., Bizon, J. L. & Setlow, B. Effects of acute administration of
nicotinic and muscarinic cholinergic agonists and antagonists on performance in different
cost-benefit decision making tasks in rats. *Psychopharmacology (Berl)* **224**, 489-499,
doi:10.1007/s00213-012-2777-y (2012).
- 178 Goldberg, J. A., Ding, J. B. & Surmeier, D. J. Muscarinic modulation of striatal function
and circuitry. *Handb Exp Pharmacol*, 223-241, doi:10.1007/978-3-642-23274-9_10
(2012).
- 179 Silveira, M. M., Malcolm, E., Shoaib, M. & Winstanley, C. A. Scopolamine and
amphetamine produce similar decision-making deficits on a rat gambling task via
independent pathways. *Behav Brain Res* **281**, 86-95, doi:10.1016/j.bbr.2014.12.029
(2015).
- 180 Howe, W. M. *et al.* Acetylcholine Release in Prefrontal Cortex Promotes Gamma
Oscillations and Theta-Gamma Coupling during Cue Detection. *J Neurosci* **37**, 3215-
3230, doi:10.1523/jneurosci.2737-16.2017 (2017).
- 181 Tort, A. B., Komorowski, R. W., Manns, J. R., Kopell, N. J. & Eichenbaum, H. Theta-
gamma coupling increases during the learning of item-context associations. *Proc Natl
Acad Sci U S A* **106**, 20942-20947, doi:10.1073/pnas.0911331106 (2009).
- 182 Ridley, R. M., Baker, H. F., Drewett, B. & Johnson, J. A. Effects of ibotenic acid lesions
of the basal forebrain on serial reversal learning in marmosets. *Psychopharmacology
(Berl)* **86**, 438-443, doi:10.1007/BF00427905 (1985).
- 183 Groman, S. M. The Neurobiology of Impulsive Decision-Making and Reinforcement
Learning in Nonhuman Animals. *Curr Top Behav Neurosci* **47**, 23-52,
doi:10.1007/7854_2020_127 (2020).
- 184 Hasselmo, M. E. & Bower, J. M. Acetylcholine and memory. *Trends Neurosci* **16**, 218-
222, doi:10.1016/0166-2236(93)90159-j (1993).
- 185 Doya, K. Metalearning and neuromodulation. *Neural Netw* **15**, 495-506,
doi:10.1016/s0893-6080(02)00044-8 (2002).
- 186 Gold, P. E. Acetylcholine modulation of neural systems involved in learning and memory.
Neurobiol Learn Mem **80**, 194-210, doi:10.1016/j.nlm.2003.07.003 (2003).
- 187 Passetti, F., Dalley, J. W., O'Connell, M. T., Everitt, B. J. & Robbins, T. W. Increased
acetylcholine release in the rat medial prefrontal cortex during performance of a visual

- attentional task. *Eur J Neurosci* **12**, 3051-3058, doi:10.1046/j.1460-9568.2000.00183.x (2000).
- 188 Mirza, N. R. & Stolerman, I. P. The role of nicotinic and muscarinic acetylcholine
189 receptors in attention. *Psychopharmacology (Berl)* **148**, 243-250,
doi:10.1007/s002130050048 (2000).
- 189 Robbins, T. W. The 5-choice serial reaction time task: behavioural pharmacology and
190 functional neurochemistry. *Psychopharmacology (Berl)* **163**, 362-380,
doi:10.1007/s00213-002-1154-7 (2002).
- 190 Howe, W. M. *et al.* Enhancement of attentional performance by selective stimulation of
191 alpha4beta2(*) nAChRs: underlying cholinergic mechanisms.
192 *Neuropsychopharmacology* **35**, 1391-1401, doi:10.1038/npp.2010.9 (2010).
- 191 Guillem, K. *et al.* Nicotinic acetylcholine receptor beta2 subunits in the medial prefrontal
192 cortex control attention. *Science* **333**, 888-891, doi:10.1126/science.1207079 (2011).
- 192 Furey, M. L., Pietrini, P., Haxby, J. V. & Drevets, W. C. Selective effects of cholinergic
193 modulation on task performance during selective attention. *Neuropsychopharmacology*
194 **33**, 913-923, doi:10.1038/sj.npp.1301461 (2008).
- 193 Erskine, F. F. *et al.* Evidence for synergistic modulation of early information processing
194 by nicotinic and muscarinic receptors in humans. *Hum Psychopharmacol* **19**, 503-509,
doi:10.1002/hup.613 (2004).
- 194 Mentis, M. J. *et al.* Muscarinic versus nicotinic modulation of a visual task. a pet study
195 using drug probes. *Neuropsychopharmacology* **25**, 555-564, doi:10.1016/S0893-
196 133X(01)00264-0 (2001).
- 195 Herrero, J. L. *et al.* Acetylcholine contributes through muscarinic receptors to attentional
196 modulation in V1. *Nature* **454**, 1110-1114, doi:10.1038/nature07141 (2008).
- 196 Zink, N., Bensmann, W., Arning, L., Stock, A. K. & Beste, C. CHRM2 Genotype Affects
197 Inhibitory Control Mechanisms During Cognitive Flexibility. *Mol Neurobiol* **56**, 6134-6141,
doi:10.1007/s12035-019-1521-6 (2019).
- 197 Ahlquist, R. P. A study of the adrenotropic receptors. *Am J Physiol* **153**, 586-600,
doi:10.1152/ajplegacy.1948.153.3.586 (1948).
- 198 Blendy, J. A., Grimm, L. J., Perry, D. C., West-Johnsrud, L. & Kellar, K. J.
199 Electroconvulsive shock differentially increases binding to alpha-1 adrenergic receptor
200 subtypes in discrete regions of rat brain. *J Neurosci* **10**, 2580-2586,
doi:10.1523/jneurosci.10-08-02580.1990 (1990).
- 199 Papay, R. *et al.* Localization of the mouse alpha1A-adrenergic receptor (AR) in the brain:
200 alpha1AAR is expressed in neurons, GABAergic interneurons, and NG2 oligodendrocyte
progenitors. *J Comp Neurol* **497**, 209-222, doi:10.1002/cne.20992 (2006).
- 200 Tanoue, A. *et al.* The alpha1D-adrenergic receptor directly regulates arterial blood pressure
201 via vasoconstriction. *The Journal of Clinical Investigation* **109**, 765-775,
doi:10.1172/JCI14001 (2002).
- 201 Cavalli, A. *et al.* Decreased blood pressure response in mice deficient of the
202 α1b-adrenergic receptor. *Proceedings of the National
Academy of Sciences* **94**, 11589-11594, doi:10.1073/pnas.94.21.11589 (1997).
- 202 Rokosh, D. G. & Simpson, P. C. Knockout of the α1A/C-adrenergic receptor
203 subtype: The α1A/C is expressed in resistance arteries and is required to
maintain arterial blood pressure. *Proceedings of the National Academy of Sciences* **99**,
204 9474-9479, doi:10.1073/pnas.132552699 (2002).
- 203 Kobayashi, M. *et al.* Presynaptic and postsynaptic modulation of glutamatergic synaptic
transmission by activation of alpha1- and beta-adrenoceptors in layer V pyramidal neurons of
rat cerebral cortex. *Synapse* **63**, 269-281, doi:https://doi.org/10.1002/syn.20604 (2009).
- 204 Mouradian, R. D., Sessler, F. M. & Waterhouse, B. D. Noradrenergic potentiation of
excitatory transmitter action in cerebrocortical slices: evidence for mediation by an alpha1

- receptor-linked second messenger pathway. *Brain Research* **546**, 83-95, doi:[https://doi.org/10.1016/0006-8993\(91\)91162-T](https://doi.org/10.1016/0006-8993(91)91162-T) (1991).
- 205 Gordon, G. R. J. & Bains, J. S. Priming of Excitatory Synapses by α_1 -Adrenoceptor-Mediated Inhibition of Group III Metabotropic Glutamate Receptors. *The Journal of Neuroscience* **23**, 6223-6231, doi:10.1523/jneurosci.23-15-06223.2003 (2003).
- 206 Luo, F., Tang, H., Li, B.-m. & Li, S.-h. Activation of α_1 -adrenoceptors enhances excitatory synaptic transmission via a pre- and postsynaptic protein kinase C-dependent mechanism in the medial prefrontal cortex of rats. *European Journal of Neuroscience* **39**, 1281-1293, doi:<https://doi.org/10.1111/ejn.12495> (2014).
- 207 Velásquez-Martínez, M. C., Vázquez-Torres, R. & Jiménez-Rivera, C. A. Activation of α_1 -adrenoceptors enhances glutamate release onto ventral tegmental area dopamine cells. *Neuroscience* **216**, 18-30, doi:<https://doi.org/10.1016/j.neuroscience.2012.03.056> (2012).
- 208 Chen, Q., Li, D.-P. & Pan, H.-L. Presynaptic α_1 -Adrenergic Receptors Differentially Regulate Synaptic Glutamate and GABA Release to Hypothalamic Presympathetic Neurons. *Journal of Pharmacology and Experimental Therapeutics* **316**, 733-742, doi:10.1124/jpet.105.094797 (2006).
- 209 Marek, G. J. & Aghajanian, G. K. 5-HT_{2A} receptor or α_1 -adrenoceptor activation induces excitatory postsynaptic currents in layer V pyramidal cells of the medial prefrontal cortex. *European Journal of Pharmacology* **367**, 197-206, doi:[https://doi.org/10.1016/S0014-2999\(98\)00945-5](https://doi.org/10.1016/S0014-2999(98)00945-5) (1999).
- 210 Shao, Y. & Sutín, J. Expression of adrenergic receptors in individual astrocytes and motor neurons isolated from the adult rat brain. *Glia* **6**, 108-117, doi:<https://doi.org/10.1002/glia.440060205> (1992).
- 211 Bekar, L. K., He, W. & Nedergaard, M. Locus Coeruleus α -Adrenergic-Mediated Activation of Cortical Astrocytes In Vivo. *Cerebral Cortex* **18**, 2789-2795, doi:10.1093/cercor/bhn040 (2008).
- 212 Kulik, A., Haentzsch, A., Lückermann, M., Reichelt, W. & Ballanyi, K. Neuron–Glial Signaling via α_1 -Adrenoceptor-Mediated Ca^{2+} Release in Bergmann Glial Cells *In Situ*. *The Journal of Neuroscience* **19**, 8401-8408, doi:10.1523/jneurosci.19-19-08401.1999 (1999).
- 213 Perez, D. M. α_1 -Adrenergic Receptors in Neurotransmission, Synaptic Plasticity, and Cognition. *Front Pharmacol* **11**, 581098, doi:10.3389/fphar.2020.581098 (2020).
- 214 Trendelenburg, A. U., Limberger, N. & Rump, L. C. α_2 -Adrenergic receptors of the α_2c subtype mediate inhibition of norepinephrine release in human kidney cortex. *Molecular Pharmacology* **45**, 1168-1176 (1994).
- 215 Giovannitti, J. A., Jr., Thoms, S. M. & Crawford, J. J. α_2 -Adrenergic receptor agonists: a review of current clinical applications. *Anesth Prog* **62**, 31-39, doi:10.2344/0003-3006-62.1.31 (2015).
- 216 Bylund, D. B. in *xPharm: The Comprehensive Pharmacology Reference* (eds S. J. Enna & David B. Bylund) 1-9 (Elsevier, 2007).
- 217 Philipp, M., Brede, M. & Hein, L. Physiological significance of α_2 -adrenergic receptor subtype diversity: one receptor is not enough. *Am J Physiol Regul Integr Comp Physiol* **283**, R287-295, doi:10.1152/ajpregu.00123.2002 (2002).
- 218 Hager, H., Hansen, N. & Manahan-Vaughan, D. β -Adrenergic Control of Hippocampal Function: Subserving the Choreography of Synaptic Information Storage and Memory. *Cerebral Cortex* **26**, 1349-1364, doi:10.1093/cercor/bhv330 (2016).
- 219 Dayan, P. & Yu, A. J. Phasic norepinephrine: a neural interrupt signal for unexpected events. *Network* **17**, 335-350, doi:10.1080/09548980601004024 (2006).

- 220 Ego-Stengel, V., Bringuier, V. & Shulz, D. E. Noradrenergic modulation of functional selectivity in the cat visual cortex: an in vivo extracellular and intracellular study. *Neuroscience* **111**, 275-289, doi:10.1016/s0306-4522(02)00011-8 (2002).
- 221 Bouret, S. & Sara, S. J. Network reset: a simplified overarching theory of locus coeruleus noradrenaline function. *Trends Neurosci* **28**, 574-582, doi:10.1016/j.tins.2005.09.002 (2005).
- 222 Usher, M., Cohen, J. D., Servan-Schreiber, D., Rajkowski, J. & Aston-Jones, G. The role of locus coeruleus in the regulation of cognitive performance. *Science* **283**, 549-554, doi:10.1126/science.283.5401.549 (1999).
- 223 Joshi, S. & Gold, J. I. Context-dependent relationships between locus coeruleus firing patterns and coordinated neural activity in the anterior cingulate cortex. *eLife* **11**, e63490, doi:10.7554/eLife.63490 (2022).
- 224 Gu, Q. Neuromodulatory transmitter systems in the cortex and their role in cortical plasticity. *Neuroscience* **111**, 815-835, doi:10.1016/s0306-4522(02)00026-x (2002).
- 225 Montes, D. R., Stopper, C. M. & Floresco, S. B. Noradrenergic modulation of risk/reward decision making. *Psychopharmacology (Berl)* **232**, 2681-2696, doi:10.1007/s00213-015-3904-3 (2015).
- 226 Kim, S., Bobeica, I., Gamo, N. J., Arnsten, A. F. & Lee, D. Effects of alpha-2A adrenergic receptor agonist on time and risk preference in primates. *Psychopharmacology (Berl)* **219**, 363-375, doi:10.1007/s00213-011-2520-0 (2012).
- 227 Steere, J. C. & Arnsten, A. F. The alpha-2A noradrenergic receptor agonist guanfacine improves visual object discrimination reversal performance in aged rhesus monkeys. *Behav Neurosci* **111**, 883-891, doi:10.1037//0735-7044.111.5.883 (1997).
- 228 Aston-Jones, G., Chiang, C. & Alexinsky, T. Discharge of noradrenergic locus coeruleus neurons in behaving rats and monkeys suggests a role in vigilance. *Prog Brain Res* **88**, 501-520 (1991).
- 229 Foote, S. L., Berridge, C. W., Adams, L. M. & Pineda, J. A. Electrophysiological evidence for the involvement of the locus coeruleus in alerting, orienting, and attending. *Prog Brain Res* **88**, 521-532, doi:10.1016/s0079-6123(08)63831-5 (1991).
- 230 Aston-Jones, G. & Bloom, F. E. Activity of norepinephrine-containing locus coeruleus neurons in behaving rats anticipates fluctuations in the sleep-waking cycle. *J Neurosci* **1**, 876-886 (1981).
- 231 Livingstone, M. S. & Hubel, D. H. Effects of sleep and arousal on the processing of visual information in the cat. *Nature* **291**, 554-561, doi:10.1038/291554a0 (1981).
- 232 McCormick, D. A., Pape, H. C. & Williamson, A. in *Progress in Brain Research* Vol. 88 (eds C. D. Barnes & O. Pompeiano) 293-305 (Elsevier, 1991).
- 233 Mountcastle, V. B., Andersen, R. A. & Motter, B. C. The influence of attentive fixation upon the excitability of the light-sensitive neurons of the posterior parietal cortex. *The Journal of Neuroscience* **1**, 1218, doi:10.1523/JNEUROSCI.01-11-01218.1981 (1981).
- 234 Hyvarinen, J., Poranen, A. & Jokinen, Y. Influence of attentive behavior on neuronal responses to vibration in primary somatosensory cortex of the monkey. *J Neurophysiol* **43**, 870-882, doi:10.1152/jn.1980.43.4.870 (1980).
- 235 Aston-Jones, G., Chiang, C. & Alexinsky, T. in *Progress in Brain Research* Vol. 88 (eds C. D. Barnes & O. Pompeiano) 501-520 (Elsevier, 1991).
- 236 Coull, J. T., Nobre, A. C. & Frith, C. D. The Noradrenergic $\alpha 2$ Agonist Clonidine Modulates Behavioural and Neuroanatomical Correlates of Human Attentional Orienting and Alerting. *Cerebral Cortex* **11**, 73-84, doi:10.1093/cercor/11.1.73 (2001).
- 237 Spencer, T. *et al.* An open-label, dose-ranging study of atomoxetine in children with attention deficit hyperactivity disorder. *J Child Adolesc Psychopharmacol* **11**, 251-265, doi:10.1089/10445460152595577 (2001).

- 238 Sigurdardottir, H. L. *et al.* Association of norepinephrine transporter methylation with in
vivo NET expression and hyperactivity-impulsivity symptoms in ADHD measured with
PET. *Mol Psychiatry* **26**, 1009-1018, doi:10.1038/s41380-019-0461-x (2021).
- 239 Dippel, G., Mückschel, M., Ziemssen, T. & Beste, C. Demands on response inhibition
processes determine modulations of theta band activity in superior frontal areas and
correlations with pupillometry – Implications for the norepinephrine system during
inhibitory control. *NeuroImage* **157**, 575-585,
doi:<https://doi.org/10.1016/j.neuroimage.2017.06.037> (2017).
- 240 Eggermann, E., Kremer, Y., Crochet, S. & Petersen, Carl C. H. Cholinergic Signals in
Mouse Barrel Cortex during Active Whisker Sensing. *Cell Reports* **9**, 1654-1660,
doi:<https://doi.org/10.1016/j.celrep.2014.11.005> (2014).
- 241 Aston-Jones, G. & Cohen, J. D. An integrative theory of locus coeruleus-norepinephrine
function: adaptive gain and optimal performance. *Annu Rev Neurosci* **28**, 403-450,
doi:10.1146/annurev.neuro.28.061604.135709 (2005).
- 242 in *Brain Control of Wakefulness and Sleep* 35-54 (Springer US, 2005).
- 243 in *Brain Control of Wakefulness and Sleep* 55-138 (Springer US, 2005).
- 244 Reimer, J. *et al.* Pupil fluctuations track rapid changes in adrenergic and cholinergic
activity in cortex. *Nat Commun* **7**, 13289, doi:10.1038/ncomms13289 (2016).
- 245 Hirata, A., Aguilar, J. & Castro-Alamancos, M. A. Noradrenergic activation amplifies
bottom-up and top-down signal-to-noise ratios in sensory thalamus. *J Neurosci* **26**,
4426-4436, doi:10.1523/jneurosci.5298-05.2006 (2006).
- 246 Koger, S. M. & Mair, R. G. Depletion of cortical norepinephrine in rats by 6-
hydroxydopamine does not impair performance of a delayed-nonmatching-to-sample
task. *Behavioral Neuroscience* **106**, 718-721, doi:10.1037/0735-7044.106.4.718 (1992).
- 247 McGaughy, J., Sandstrom, M., Ruland, S., Bruno, J. P. & Sarter, M. Lack of effects of
lesions of the dorsal noradrenergic bundle on behavioral vigilance. *Behav Neurosci* **111**,
646-652, doi:10.1037//0735-7044.111.3.646 (1997).
- 248 Chandler, D. & Waterhouse, B. D. Evidence for broad versus segregated projections
from cholinergic and noradrenergic nuclei to functionally and anatomically discrete
subregions of prefrontal cortex. *Front Behav Neurosci* **6**, 20,
doi:10.3389/fnbeh.2012.00020 (2012).
- 249 Dalley, J. W. *et al.* Distinct Changes in Cortical Acetylcholine and Noradrenaline Efflux
during Contingent and Noncontingent Performance of a Visual Attentional Task. *The
Journal of Neuroscience* **21**, 4908, doi:10.1523/JNEUROSCI.21-13-04908.2001 (2001).
- 250 McGaughy, J., Ross, R. S. & Eichenbaum, H. Noradrenergic, but not cholinergic,
deafferentation of prefrontal cortex impairs attentional set-shifting. *Neuroscience* **153**,
63-71, doi:<https://doi.org/10.1016/j.neuroscience.2008.01.064> (2008).
- 251 Disney, A. A. Neuromodulatory Control of Early Visual Processing in Macaque. *Annu
Rev Vis Sci* **7**, 181-199, doi:10.1146/annurev-vision-100119-125739 (2021).
- 252 Doya, K. Modulators of decision making. *Nat Neurosci* **11**, 410-416, doi:10.1038/nn2077
(2008).
- 253 Sethuramanujam, S. *et al.* Rapid multi-directed cholinergic transmission in the central
nervous system. *Nat Commun* **12**, 1374, doi:10.1038/s41467-021-21680-9 (2021).
- 254 Yu, A. J. & Dayan, P. Uncertainty, neuromodulation, and attention. *Neuron* **46**, 681-692,
doi:<https://doi.org/10.1016/j.neuron.2005.04.026> (2005).
- 255 Deppe, M., Schwindt, W., Kugel, H., Plaßmann, H. & Kenning, P. Nonlinear Responses
Within the Medial Prefrontal Cortex Reveal When Specific Implicit Information Influences
Economic Decision Making. *Journal of Neuroimaging* **15**, 171-182,
doi:<https://doi.org/10.1111/j.1552-6569.2005.tb00303.x> (2005).
- 256 Euston, David R., Gruber, Aaron J. & McNaughton, Bruce L. The Role of Medial
Prefrontal Cortex in Memory and Decision Making. *Neuron* **76**, 1057-1070,
doi:<https://doi.org/10.1016/j.neuron.2012.12.002> (2012).

1319 257 Orsini, C. A. *et al.* Contributions of medial prefrontal cortex to decision making involving
1320 risk of punishment. *Neuropharmacology* **139**, 205-216,
1321 doi:<https://doi.org/10.1016/j.neuropharm.2018.07.018> (2018).
1322 258 Sul, J. H., Kim, H., Huh, N., Lee, D. & Jung, M. W. Distinct Roles of Rodent Orbitofrontal
1323 and Medial Prefrontal Cortex in Decision Making. *Neuron* **66**, 449-460,
1324 doi:<https://doi.org/10.1016/j.neuron.2010.03.033> (2010).
1325 259 Rodenkirch, C., Schriver, B. & Wang, Q. in *Neural Engineering* (eds L. Zhang & D.
1326 Kaplan) (Springer, 2016).
1327