REVIEW





A review of the diet of the common vampire bat (*Desmodus rotundus*) in the context of anthropogenic change

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Abstract

The common vampire bat (*Desmodus rotundus*) maintains a diverse, sanguivorous diet, utilizing a broad range of prey taxa. As anthropogenic change alters the distribution of this species, shifts in predator–prey interactions are expected. Understanding prey richness and patterns of prey selection is, thus, increasingly informative from ecological, epidemiological, and economic perspectives. We reviewed *D. rotundus* diet and assessed the geographical, taxonomical, and behavioral features to find 63 vertebrate species within 21 orders and 45 families constitute prey, including suitable host species in regions of invasion outside *D. rotundus*' range. Rodentia contained the largest number of species utilized by *D. rotundus*, though cattle were the most commonly reported prey source, likely linked to the high availability of livestock and visibility of bite wounds compared to wildlife. Additionally, there was tendency to predate upon species with diurnal activity and social behavior, potentially facilitating convenient and nocturnal predation. Our review highlights the dietary heterogeneity of *D. rotundus* across its distribution. We define *D. rotundus* as a generalist predator, or parasite, depending on the ecological definition of its symbiont roles in an ecosystem (i.e., lethal vs. non-lethal blood consumption). In view of the eminent role of *D. rotundus* in rabies virus transmission and its range expansion, an understanding of its ecology would benefit public health, wildlife management, and agriculture.

Keywords Anthropogenic · Common vampire bat · Desmodus rotundus · Diet · Predator · Prey · Rabies

Introduction

It is widely recognized that human modification of natural landscapes continues to transform ecosystems at an immense rate (Barlow et al. 2016; Ellis et al. 2010; Grantham et al. 2020). Anthropogenic disturbances such as

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land-use change, exploitation of natural resources, climate anomalies, and agricultural expansion continue to alter biodiversity assemblages (Bengtsson et al. 2005; Bellard et al. 2012; Chaudhary and Mooers 2018; Hooper et al. 2012). In Latin America, this phenomenon remains disproportionately significant, given the abundant ecosystem services provided by this region (Nobre et al. 2016; Salazar et al. 2015). Deforestation remains the dominant land-use trend in this expanse, driven by a demand for cattle production and export-oriented monoculture agriculture (Dros 2004; Grau and Aide 2008). Moreover, climate projections anticipate substantial change in both precipitation and temperature in subtropical Latin America in the coming years, further altering ecosystem structure and function (Cabré et al. 2016; Nuñez et al. 2009).

Facing the constraints of the Anthropocene epoch and in response to increasingly unsuitable habitats, wildlife populations in Latin America are undergoing major geographic redistribution (Ancillotto et al. 2016; McCracken et al. 2018; Osland et al. 2021). Wildlife is significantly impacted by the diminished forests that occur secondary to land-use changes as well as the climate alterations that further relegate the



amount of wild landscapes that meet their ecological needs (Grantham et al. 2020; Willmott et al. 2022). Movement of non-native taxa into novel regions is, thus, increasingly common as species are driven to seek refuge elsewhere (Pecl et al. 2017; Willmott et al. 2022). Changes in the distribution and abundance of wildlife linked to land cover change have already been documented in Latin America for several taxa, including amphibians (Vieira et al. 2018), rodents (Campos-Krauer and Wisely 2011), birds (Hayes et al. 2018), artiodactyls such as peccaries (Altrichter and Boaglio 2004) and tapirs (Reyna-Hurtado et al. 2019), marsupials (Loyola et al. 2012), and numerous bat species (Barquez et al. 2013; LaVal 2004).

Historically, the common vampire bat, Desmodus rotundus, has been confined to tropical and subtropical regions of Latin America, spanning southward from Mexico to northern Argentina, central Chile, and southern Uruguay (Greenhall 1988; Zarza et al. 2017). This phyllostomid species can be found at sea level to over 3500 m of altitude (Greenhall et al. 1983; Greenhall 1988). Desmodus rotundus occupies diverse habitats ranging from rainforests to deserts, where it maintains an exclusively hematophagous diet and feeds on the blood of numerous vertebrate species (Greenhall 1972, 1988; McNab 1973). Rising temperatures, habitat fragmentation, and agricultural activities have altered the dispersal and driven this Neotropical sanguivore both northward and into higher altitudes (Botto Nuñez et al. 2020; Camargo et al. 2018; Rojas-Sereno et al. 2022). Desmodus rotundus has recently been documented close to the United States-Mexico border, a novel region for the species (Bodenchuck and Bergman 2020; Hayes and Piaggio 2018) and at higher elevations in Costa Rica than previously noted (LaVal 2004). Climate models have suggested potential range expansion of D. rotundus in Colombia, French Guyana, Suriname, Venezuela, Mexico, and at greater altitudes such as those maintained in the Andes Mountains of Chile (Lee et al. 2012; Mistry and Moreno-Valdez 2009; Zarza et al. 2017). In time, D. rotundus may enter the United States through the southern borders of Texas or Arizona (Hayes and Piaggio 2018; Mistry and Moreno-Valdez 2009).

As with any non-native taxa, the expansion of *D. rotundus* populations into novel regions holds potential for cascading repercussions (Andersen and Shwiff 2014; Borremans et al. 2019; Clavero and García-Berthou 2005; Paini et al. 2016). Through evolutionary time, wildlife has reliably adapted to utilize novel prey species upon migration, including livestock, igniting changes in the trophic structure and introducing pathogens into naïve populations and regions (El-Sabaawi 2018; Plowright et al. 2015). Known for their robust adaptive capacity, range expansion of *D. rotundus* is particularly important given that the species (i) feeds on a diverse repertoire of prey, (ii) has demonstrated considerable plasticity in adapting to novel landscapes and diet

sources, and (iii) maintains a strictly sanguivorous (i.e., whole blood) diet, providing opportunities for rapid transmission of pathogens in which the species acts as a reservoir for (Bobrowiec et al 2015; Greenhall 1988; Mayen 2003; Streicker and Allgeier 2016). These attributes suggest that distributional shifts of *D. rotundus* into new territories may result in establishment of the species as invasive, generating novel ecologic, epidemiologic, and economic human—wild-life conflicts (Acha and Alba 1988; Delpietro and Russo 1996; Ingala et al. 2019; Mayen 2003).

Importance of understanding prey richness

Ecologic

Changes in predator-prey dynamics and disruptions in trophic structure are an unavoidable and expected consequence of changes to biodiversity (Gámez et al. 2022; Guiden et al. 2019; Murphy et al. 2021). When wildlife migrates to novel regions or changes in abundance, interspecific interactions change correspondingly (Fleming and Bateman 2018; Guiden et al. 2019). For example, when the ranges of predators increasingly overlap and prey populations decline, predators often switch to generalist diets or utilize new taxa as sources of prey (Ballejo et al. 2018; Fleming and Bateman 2018; Murray et al. 2015; Rodewald et al. 2011; Wolfe et al. 2018). This may include livestock in landscapes subject to agricultural conversion, should natural prey become scarce or less accessible relative to domestic herbivores (Cardinale et al. 2006; Green et al. 2022; Michalski et al. 2006; Murphy et al. 2021; Patten et al. 2019). Novel landscape conditions may likewise promote phenological shifts such as changes in temporal patterns (Lebbin et al. 2007; Mandel and Bildstein 2007; Rich and Longcore 2013), behavioral adaptations related to sensory processes (Chan et al. 2010; Eisenbeis et al. 2009; Siemers and Schaub 2011; Simpson et al. 2016), and alterations in feeding (Grabrucker and Grabrucker 2010; Stafford-Bell et al. 2012; Valeix et al. 2012; Weir and Kacelnik 2006).

The ability of *D. rotundus* to readily adapt to new land-scapes and novel prey sources is documented in both wild and captive settings (Bolívar-Cimé et al. 2019; Greenhall 1988). Experimentally, *D. rotundus* has been offered and readily accepted a variety of non-natural diets including both live prey and extracted blood. The bats have attacked species such as armadillos and porcupines and demonstrated successful location of vulnerable areas for biting (Greenhall 1988). In the wild, studies have detailed *D. rotundus* feeding on invasive feral swine and other taxa which did not co-evolve as not part of their diet (Calfayan et al. 2019; Galetti et al. 2016; Grotta-Neto et al. 2021; Hernández-Pérez et al. 2019; Pereira et al. 2016). The prey consumed by *D.*



rotundus is suggested as largely dependent on surrounding fauna of the bats, that is, based on availability (Gonçalves et al. 2021). The success of this species in geographic regions with sparse wildlife or poor species richness has been achieved through the utilization of livestock on multiple accounts (Bobrowiec et al. 2015; Botto Nuñez et al. 2020; Delpietro et al. 1992; Voigt and Kelm 2006). The absence of native or abundance of introduced prey, or alternatively, the establishment of *D. rotundus* in alien regions may provide ample opportunity for dietary shifts and novel interspecific interactions (Gonçalves et al. 2021; Streicker and Allgeier 2016).

Epidemiologic

In general, bats are important pathogen reservoirs in emerging infectious disease (Plowright et al. 2015), as noted by outbreaks of Hendra virus (Halpin et al. 2000), Nipah virus (Yob et al. 2001), SARS (Wang et al. 2006), and other coronaviruses (Burki 2020; Hernández-Aguilar et al. 2021). Given their unique behavioral and ecological traits, as well as the plethora of pathogens harbored by the species, *D. rotundus* may be a key Chiropteran in heterospecific disease transmission (Johnson et al. 2014; Wray et al. 2016).

Desmodus rotundus is a known host for many viral pathogens, most notably as the primary rabies virus (RABV) reservoir in Latin America (Benavides et al. 2020a; Cárdenas-Canales et al. 2020; Johnson et al. 2014; Streicker et al. 2012). Several additional viral agents have been detected in the species, including adenoviruses, herpesviruses, vesiculoviruses, and coronaviruses, among others (Bergner et al. 2019; Wray et al. 2016). Recent metagenomic sequencing of D. rotundus fecal and saliva samples identified over 58 viral families, 17 of which are known to infect mammalian species (Bergner et al. 2021a). Alphacoronaviruses, a viral genus significant to mammal health, have been detected in D. rotundus subjects in Argentina (Arteaga et al. 2022), Brazil (Alves et al. 2021; Asano et al. 2016), and Peru (Bergner et al. 2020). A Betacoronavirus related to MERS-CoV was found in D. rotundus in Belize (Neely et al. 2020). The presence of deltaviruses, originally associated with hepatitis B co-infections in humans, has been confirmed in the saliva of D. rotundus as well. Phylogenetic analysis suggests that bat deltaviruses arose from other mammalian deltaviruses, and therefore hold potential for cross-species transmission (Bergner et al. 2021b). The viral agent responsible for foot and mouth disease, one of the most economically important diseases of livestock and greatest biosecurity threat to many nations, was successfully inoculated and recovered from D. rotundus in experimental settings (Lord et al. 1986).

Desmodus rotundus also hosts a variety of other pathogens, including *Leptospira* spp. (Ballados-González et al. 2018) and *Bartonella* spp. (Wray et al. 2016), both zoonotic

bacteria agents. Urination during feeding and flying has been noted as a possible means of leptospirosis transmission (Greenhall 1964). In addition, *D. rotundus* has recently also been implicated in fecal carriage of multidrug-resistant extended-spectrum beta-lactamase-producing *Escherichia coli* to livestock (Benavides et al. 2022). Numerous endoparasites have also been found on *D. rotundus*, such as *Toxoplasma gondii* (Zetun et al. 2009) and various trypanosomes including *Trypanosoma cruzi* (Quiroga et al. 2022). Ectoparasites include ixodid and argasid ticks (*Ixodes*, *Amblyomma*, and *Ornithodoros* sp.), trombiculid or spider mites (*Euschoengastia* spp.), mange mites (*Sarcoptes* sp.), Siphonapterid fleas, and numerous bat flies (Anciaux de Faveaux 1971; Greenhall et al. 1983; Rojas et al. 2008; Tamsitt and Fox 1970; Wenzel and Tipton 1966).

Distributional shifts of *D. rotundus* into novel regions may provide new opportunities for pathogen spillover when suitable hosts are available (Altizer et al. 2011; Borremans et al. 2019; Streicker et al. 2016). The nature of an obligate sanguivorous diet alone implies easy and robust transmission of saliva and blood-borne pathogens between prey and D. rotundus (Dunn 1932; Gupta 2005; Rocha and Dias 2020). Likewise, wide geographic ranges, prolonged exposure to prey during feeding, co-species roosting, and social behavior such as sharing blood meals and allogrooming have numerous implications for disease transmission (Rocha et al. 2020; Wray et al. 2016). Studies have hypothesized that emergence of vampire bat rabies in Uruguay resulted from a combination of land-use change and D. rotundus migration, leading to increased viral persistence and transmission to cattle (Nuñez et al. 2019). The potential for pathogen emergence is especially notable given the wide array of species D. rotundus has been documented to feed upon and their ability to utilize novel prey (Bobrowiec et al. 2015; Greenhall 1988; Herrera et al. 1998; Voigt and Kelm 2006). Furthermore, Desmodus rotundus bites can increase the susceptibility of prey species to secondary infections in the wound, even if pathogens were not directly transmitted by the bats (Johnson et al. 2014). Following a blood meal, prey may have residual hemorrhage for several hours, depending on the prey species and duration of D. rotundus feeding, leaving animals vulnerable to other pathogens and blood loss during this period of time (Greenhall et al. 1969).

Economic

In addition to infectious disease and ecologic concerns, *D. rotundus* causes massive economic loss to the livestock industry, primarily through the transmission and propagation of rabies. Although to a considerably less degree, *D. rotundus* may negatively impact livestock through means other than rabies, including production loss via decreased milk output, poor or diminished weight gain, anemia,

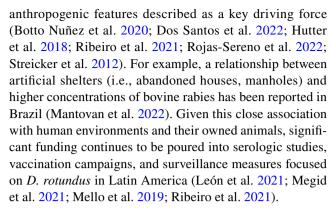


myiasis (fly strike), and damaged hides attributable to bite wounds following attack (Thompson et al. 1977; Acha and Alba 1988; Greenhall 1988; Mayen 2003). Nevertheless, this financial impact has been largely overlooked and is a rarely focused area of research compared to that of rabies, which occurs directly though the deaths of thousands of cattle and indirectly through the cost of rabies prevention and control measures (Acha and Alba 1988; Meltzer and Rupprecht 1998; Schmidt and Badger 1979).

In the 1960s, there were an estimated 90,000–100,000 rabies-related cattle deaths per year in Mexico (Acha 1967). Presently, there are over 400 deaths per 100,000 cattle head in Peru (Benavides et al. 2017) and 100,000-500,000 cattle deaths in Latin America annually (Mello et al. 2019). In the 1980s, livestock losses due to rabies, mostly ascribed to D. rotundus, were estimated to be around US\$50 million annually in Latin America (Acha and Szyfres 1986). In Brazil alone, this value is estimated at over US\$17 million (Horta et al. 2022). Currently, D. rotundus continues to be the principal RABV reservoir in the Latin American region (Constantine and Blehert 2009; Horta et al. 2022). Should D. rotundus spread to southern Texas in the United States, a livestock-dense portion of the country, the economic impact via rabies in cattle, swine, sheep, and goats is estimated to be between US\$7 million and US\$9.2 million annually (Anderson and Shwiff 2014).

Additionally, D. rotundus may transmit RABV to nonlivestock species, including both wildlife and companion animals in captive and free-range settings (Gonçalves et al. 2021; Schneider et al. 2009). Desmodus rotundus has historically been implicated in rabies cases found in free-ranging capybaras, deer, foxes, non-sanguivorous bat species, and other mammals known to play a role in both sylvatic and urban cycles of the disease (Delpietro et al. 2009; Gonçalves et al. 2021; Favoretto et al. 2002; Kobayashi et al. 2005). In captive settings, D. rotundus has been documented to prey on animals housed in zoos, farms, and residential sites (Benavides et al. 2020a, 2020b; Constantine 1979; Valderrama et al. 2006). A zoologic institution in Brazil recently reported a case of rabies encephalitis in a lowland tapir (Tapirus terrestris), with suspicions of D. rotundus involvement described (Pereira et al. 2022). Similarly, and also suspected to be incited by D. rotundus, a fatal infection was described for a captive white-tailed deer in Mexico (Franco-Molina et al. 2021). Both cases involved economic loss to the respective organizations, through the cost of employees' post-exposure prophylaxis, confirmatory diagnostics for the case itself, and preemptive testing, post-exposure vaccination, and quarantine of other animals on site (Franco-Molina et al. 2021; Pereira et al. 2022).

The prevalence and incidence of livestock, companion animal, and human rabies caused by *D. rotundus* continues to wax and wane throughout Latin America, with



In view of the ecologic, epidemiologic, and economic points described, a thorough understanding of the diet of D. rotundus is, therefore, critical in preparing for latitudinal and altitudinal expansion of the species' distribution (Bodenchuck and Bergman 2020; Hayes and Piaggio 2018; Mayen 2003; Schneider et al. 2009). The aim of this paper is, thus, to revise the diet of D. rotundus to identify major prey consumption trends in light of ongoing anthropogenic change. Here, we expand a previous contribution by Carter et al. (2021) on the taxonomic breadth of D. rotundus prey and account for species-level characteristics relevant to prey selection, provide an updated record of recognized prey taxa, and discuss these findings in the context of D. rotundus range expansion. Moreover, we discuss the relevance of D. rotundus prey presently occurring in the United States. Ultimately, this information may be used to anticipate pathogen emergence in new species, identify at-risk populations of domestic and wildlife taxa, and critically evaluate mitigation methods in specific localities.

Materials and methods

A literature search was conducted using Web of Science Core Collection (WoS), Scopus, and Science Direct databases to identify recorded prey species of D. rotundus. The search strategy used the following search terms in all fields: Desmodus or vampire bat* and prey or predation* or attack* or diet* or bite* or feeding* and was restricted to papers in English, Spanish, and Portuguese. Duplicates were removed, retrieving a total of 360 results. Inclusion criteria for study selection included a documented occurrence of D. rotundus feeding on an identifiable vertebrate species in a known geographical location. Although humans are a recognized prey species of D. rotundus, these reports were excluded from assessment, given the scope of the review and number of existing publications focusing on the public health components of D. rotundus. Prey detection methods utilized were extracted for each study included in the review. Taxonomic information of prey was standardized using the Integrated Taxonomic Information System (https://www.itis.gov/) for a



more accurate categorization of species. Information on the geographic, temporal, and social behavior was pulled from a variety of literature sources (Supplementary Table S3). Data were analyzed and visualized using JMP Pro 16.0.0 (SAS Institute Inc, Cary, NC) statistical software and QGIS (Bioalowieza Version 3.22.7; Open Source Geospatial Foundation, OR).

Literature trends

Desmodus rotundus diet has been studied over the last century with existing literature scattered across the scientific disciplines, from ecology and animal behavior to physiology and epidemiology (Arellano 1988; Freitas et al. 2003; Greenhall et al. 1971; Souza et al. 1997). Carter et al. (2021) summarized the diet of the three known vampire bat species, including D. rotundus, and revealed the broad array of species predated upon. From our literature review, we recovered a total of 67 publications containing 214 reports of D. rotundus feeding on prey during the period 1931–2020. There was an apparent rise in publications of D. rotundus between 1970 and 1979 and a later increase in the 2010's (Fig. 1A). The number of prey species documented each year has likewise continued to grow (Fig. 1B).

The liquid nature of D. rotundus' diet has limited the implementation of standard fecal analysis based on detritus in feces as a means of prey identification (Bobrowiec et al. 2015). Other attempts have been made to quantify target species, including using direct observation, camera traps, genomics, stable isotope analysis, and experimental studies (Greenhall et al. 1969; Greenhall 1988; Hernández-Pérez et al. 2019; Voigt and Kelm 2006). Methods for D. rotundus prey documentation from the articles reviewed included anecdotal evidence (57%), molecular and chemical techniques (49%), experiments performed in captivity (32%), bite wound visualization (27%), camera traps (24%), and live field observations of attack (22%). Anecdotal evidence includes interviews, authors' personal accounts, or any reports from community members, producers, and other individuals where D. rotundus is claimed to have bitten specific prey species within the publication, though attacks or wounds were not necessarily witnessed. Bite wound analysis comprises reports or studies that involve documented visualization of insults. Anecdotal reports and bite wound analysis were most common in literature dating before 1980. This lack of field-based methodology utilized in older research and the resultant uncertainty in data credibility and species identification is a major limitation when assessing the diet

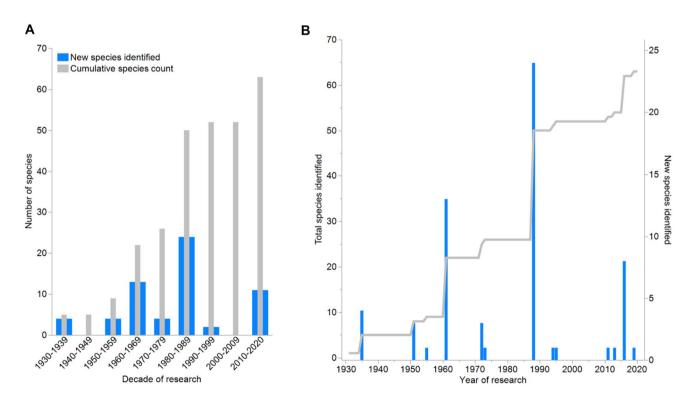


Fig. 1 Desmodus rotundus literature on prey species published between 1930 and 2020. **A** Decadal number of prey species. The number of new species discovered over the course of each decade (blue) and the cumulative number of *D. rotundus* prey species documented by the end of each decade (gray). No new species were

reported as *D. rotundus* prey between 1940–1949 and 2000–2009. Research throughout the last decade has resulted in documentation of 11 new prey species. **B** Cumulative number of prey species (gray) and new species reported annually (blue)



of *D. rotundus*. Today, ecological and genomic technology is increasingly utilized as a means to document predation. In the last decade, 35% of occurrences were documented via camera traps and 35% via molecular techniques. Contrarily, live field observations have become a less common methodology and were only used to document 2.8% of all occurrences between 2010 and 2020.

Prey diversity

Cumulatively, all existing literature supports the notion that *D. rotundus* is able to feed on a diverse collection of species, including those in the classes Mammalia, Reptilia, Aves, Amphibia, and Insecta (Bobrowiec et al. 2015; Greenhall 1988; Herrera et al. 1998; Voigt and Kelm 2006). A total of 63 prey species within 5 classes, 21 orders, and 45 families were recovered from the literature by 2020 and used to create a comprehensive dataset (Fig. 2, Supplementary Table S1). Note that these species include those documented under all conditions, including experiments performed in captivity. Insecta was documented as a prey class but for unspecified

taxa (Arata et al. 1967; Rouk and Glass 1970; Bohmann et al. 2018) and was, thus, excluded from our assessment.

Prey data were then examined in terms of taxonomic richness (i.e., number of species) (Fig. 2A) and number of publications (Fig. 2B). Most of the prey were mammals (75.4%) with 47 species documented, though several species from other taxa were identified, including 7 avian, 8 reptilian, and 1 amphibian species. Species were characterized as wildlife (i.e., free-ranging and undomesticated) or domestic (i.e., bred and raised for companionship or production/vocational purposes). Domestic species were further divided into those that function as livestock and those that act primarily as companion animals. Swine (*Sus scrofa*) were counted in both livestock and wildlife groups as there are reports of *D. rotundus* preying on both feral and domestic pigs.

The order Rodentia contained the largest number of D. rotundus prey species (n=12), followed by Artiodactyla (n=11) and Carnivora (n=9) (Fig. 2). Rodent species include those of the families Caviidae (n=2), Cricetidae (n=2), Sciuridae (n=2), Cuniculidae (n=1), Dasyproctidae (n=1), Echimyidae (n=1), Erethizontidae (n=1), Muridae

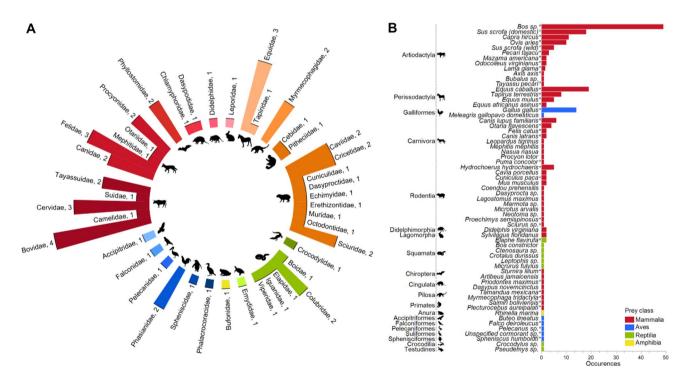


Fig. 2 Taxa of *Desmodus rotundus* prey. **A** Order (silhouettes), family (bar labels) and species (bar number) denoting the taxonomic range of *D. rotundus* prey. The 63 species known to be *D. rotundus* prey are grouped among the Artiodactyla (4 families, 11 species), Carnivora (5 families, 9 species), Chiroptera (1 family, 2 species), Cingulata (2 families, 2 species), Didelphimorphia (1 family, 1 species), Lagomorpha (1 family, 1 species), Perissodactyla (2 families, 4 species), Pilosa (1 family, 2 species), Primates (2 families, 2 species), Rodentia (9 families, 12 species), Crocodilia (1 family, 1 species), Squamata (5 families, 6 species), Testudines (1 family, 1 species), Anura (1 family, 6 species), Testudines (1 family, 1 species), Anura (1 family, 1 species)

ily, 1 species), Accipitriformes (1 family, 1 species), Falconiformes (1 family, 1 species), Galliformes (1 family, 1 species), Pelecaniformes (1 family, 1 species), Sphenisciformes (1 family, 1 species), and Suliformes (1 family, 1 species) orders. **B** Prey order (silhouettes on the left) and species (bars on the right) denoting the number of literature occurrences between 1931 and 2020 (y-axis) and grouped by class (Amphibia=yellow, Aves=blue, Mammalia=red, Reptilia=green). Species documented in non-captive settings are denoted by an asterisk. Humans are not included but are well-documented prey of *D. rotundus*



(n=1), and Octodontidae (n=1). Artiodactyl species include Bovidae (n=4), Cervidae (n=3), Camelidae (n=1), Suidae (n=1), and Tayassuidae (n=2). Bovidae contained the most Artiodactyl species (n=4). Families within Carnivora include Felidae (n=3), Canidae (n=2), Procyonidae (n=2), Mephitidae (n=1), and Otariidae (n=1). Other prey orders identified include Accipitriformes, Anura, Chiroptera, Cingulata, Crocodilia, Didelphimorphia, Falconiformes, Galliformes, Lagomorpha, Pelecaniformes, Perissodactyla, Pilosa, Primates, Sphenisciformes, Squamata, Suliformes, and Testudines.

Regarding methodology, 56% of all species were documented in a non-captive setting (i.e., via camera trap, field observation, chemical and molecular analysis, bite wound visualization). The remaining 33% were identified as *D. rotundus* prey in captive experiments and 10% via anecdotal evidence only. Anecdotally identified prey includes the turkey (*Meleagris gallopavo domesticus*), guanaco (*Lama glama*), Virginia opossum (*Didelphis virginiana*), agouti (*Dasyprocta* sp.), Argentine plains viscacha (*Lagostomus maximus*), and marmot (*Marmota* sp.). Species only identified under experimental conditions are described in the proceeding sections. The methods used to identify *D. rotundus* prey is a major limitation in the accuracy of these data, especially for wild settings, but could provide signal of prey use.

Wildlife as D. rotundus prey

Historically, *Desmodus rotundus* is described to prey on medium-to-large-bodied terrestrial mammals, particularly ungulates, although there are reports of the bat preying on small mammals, marine mammals, birds, and reptiles as well (Bobrowiec et al. 2015; Greenhall 1988; Voigt and Kelm 2006). Commonly discussed wildlife prey includes peccaries (Voigt and Kelm 2006), deer (Sánchez-Cordero et al. 2011), tapirs (Gnocchi et al. 2017), and capybaras (Gonçalves et al. 2021). Our review supported these claims, identifying a total of 52 wildlife species as *D. rotundus* prey. The South American tapir (*Tapirus terrestris*) (n=8), capybara (*Hydrochoerus hydrochaeris*) (n=5), feral pig (*Sus scrofa*) (n=5), and South American sea lion (*Otaria flavescens*) (n=4) were the most commonly reported wild mammals attacked by *D. rotundus*.

Of these 52 species, 27 were observed in non-captive settings, while the remaining 25 were documented under experimental conditions. Mammalian species observed as *D. rotundus* prey in field settings include the puma (*Puma concolor*), coyote (*Canis latrans*), chital deer (*Axis axis*), South American red brocket (*Mazama americana*), white-tailed deer (*Odocoileus virginianus*), wild pig (*Sus scrofa*), collared peccary (*Pecari tajacu*), white-lipped peccary (*Tayassu pecari*), South American sea lion (*Otaria flavescens*), little yellow-shouldered bat (*Sturnira lilium*), giant

armadillo (*Priodontes maximus*), South American tapir (*Tapirus terrestris*), giant anteater (*Myrmecophaga tridactyla*), Northern tamandua (*Tamandua mexicana*), madidi titi (*Plecturocebus aureipalati*), black-capped squirrel monkey (*Saimiri boliviensis*), lowland paca (*Cuniculus paca*), capybara (*Hydrochoerus hydrochaeris*), spiny rat (*Proechimys semispinosus*), squirrel (*Sciurus* sp.), and rat snake (*Elaphe flavirufa*).

Desmodus rotundus was reported to predate upon one amphibian species, a cane toad (Rhinella marina), which was only documented in a captive setting (Greenhall 1988). Similarly, the majority of literature on D. rotundus predating on eight reptilian species was done in captivity, with the exception of the rat snake (Elaphe flavirufa) (Greenhall 1988). Six avian species were identified as D. rotundus prey in wild settings, including the orange-breasted falcon (Falco deiroleucus), Humboldt penguin (Spheniscus humboldti), pelican (Pelicanus sp.), cormorant (Suliformes), and domestic chicken (Gallus gallus). Desmodus rotundus was also documented to feed on the red-shouldered hawk (Buteo lineatus) in captivity.

Livestock as D. rotundus prey

Despite the research efforts devoted to predation on native species, the general consensus in the literature is that D. rotundus selects livestock as prey over free-ranging wildlife when provided the opportunity (Voigt and Kelm 2006). There is inconsistency regarding which livestock species are most commonly targeted, with some reports suggesting a tendency to feed on cattle (Bohmann et al. 2018; Goodwin and Greenhall 1961), horses (Mialhe 2014; Turner 1975), or swine (Bobrowiec et al. 2015). Poultry is described as a possible second choice (Bobrowiec et al. 2015) and small ruminants (sheep and goats) as uncommon prey choices, although the reason for a plausible distaste is unknown (Turner 1975; Mialhe 2014). Our review revealed that cattle were the most commonly reported prey species across all publications (n=49). There were 26 reports documenting D. rotundus feeding on equine (Equus spp.), 18 on domestic swine (Sus scrofa), 14 on chickens, 11 on goats (Capra hircus), 10 on sheep (Ovis aries), 2 on llamas (Lama glama), and 1 on turkeys. Desmodus rotundus did predate upon livestock species more often than others in this review, with 121 total occurrences, compared to 80 for wildlife and 10 for companion animals.

Nonetheless, this tendency for *D. rotundus* to predate on livestock and the interpretation as a true predation pattern may be biased. There is a large effort devoted to studying these species with regard to the potential RABV transmission from *D. rotundus* (Mayen 2003). The likelihood of witnessing predation is also much greater for livestock than wildlife, given the relative exposure of humans to domestic

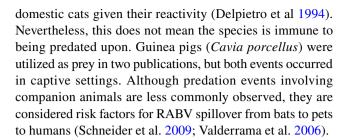


species (Mialhe 2014; Voigt and Kelm 2006). Livestock is often handled at a frequent cadence and bite wounds are readily evident as producers and large animals typically seek out any lesions or form of injury that may ultimately impact production or economics (Greenhall 1972). Likewise, it may be due to the limited research on *D. rotundus* feeding in wild settings and lack of information on livestock availability for many studies that cover wildlife predation (Bobrowiec et al. 2015). Additional research examining the influence of livestock availability on the utilization of wildlife prey is, therefore, needed to better understand predation patterns across the domestic and wild prey.

Although not unanimously agreed upon, convenience is the most commonly cited driving factor for prey selection (Delpietro et al. 1992; Gonçalves et al. 2021; Voigt and Kelm 2006). Anecdotal reports suggest that livestock is targeted due to its abundant and stable availability as compared with wildlife prey (Delpietro et al. 1992; Gonçalves et al. 2021; Voigt and Kelm 2006). These claims may, again, be biased considering that D. rotundus roosts are often located in close proximity to humans where these studies often take place (Ribeiro et al. 2021; Rojas-Sereno et al. 2022). Likewise, studies on prey availability versus richness are, thus, limited in pristine environments where wildlife are most abundant (Bobrowiec et al. 2015). Other studies hypothesize that prey richness is influenced by various facets of convenience including large prey size for larger volumes of blood (Delpietro et al. 1992; Johnson et al. 2014), proximity to roost site (Rocha et al. 2020), ease of attack (Mialhe 2014), lack of prey reactivity (Arellano and Greenhall 1971; Delpietro 1989; Delpietro et al. 1992), and herd size (Bobrowiec et al. 2015). Nutritional composition of blood has also been described as a factor (Voigt and Kelm 2006). For cattle specifically, there is a reported inclination to feed on calves over older cows and heifers, and likewise a breed difference in Holstein predation over various others (Arellano and Greenhall 1971). This is suspected to be due to prey temperament and lack of prey reactivity that would facilitate easier feeding (Arellano and Greenhall 1971).

Companion animals and other domestic species as *D. rotundus* prey

Desmodus rotundus also feeds on anthropogenic prey with reports describing attacks on dogs and humans, although these events are much less common in the literature (Schneider et al. 2001; Streicker and Allgeier 2016; Torres et al. 2005). We retrieved six reports of *D. rotundus* predation on domestic dogs (Canis familiaris). Nevertheless, the literature involving cats (Felis catus) describes the species as a livestock guardian of sorts (Delpietro et al 1994). The addition of barn cats was an effective means to deter *D. rotundus* from attacking livestock and it is proposed that the bats avoid



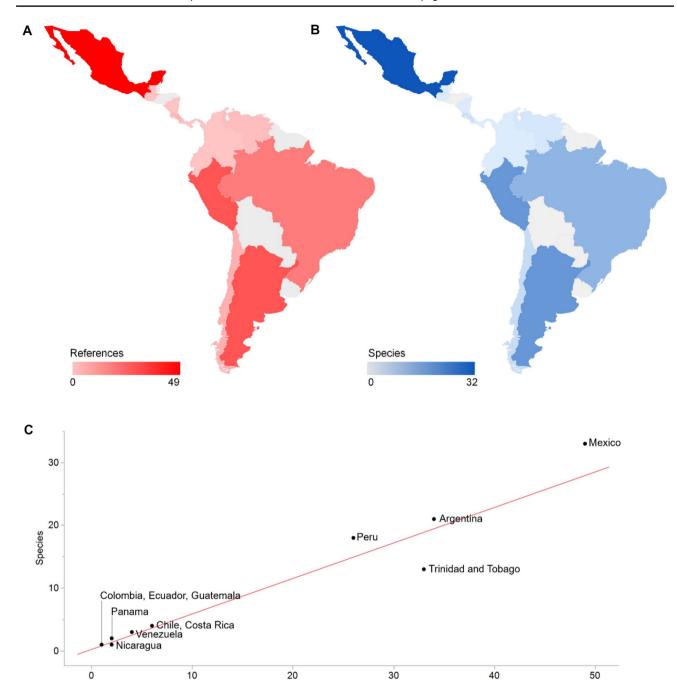
Geographic trends in D. rotundus predation

In terms of geography, there is a need for additional research on the prey of D. rotundus in certain countries, and a greater focus on less commonly targeted classes such as Aves, Reptilia, and Amphibia. Blood meal or fecal sample analysis for D. rotundus subjects obtained from regions with unaddressed research needs may provide a valuable tool for characterization of prey taxa. The number of documented species and relatedness to prey richness for each country versus true sampling bias due to sampling effort influencing the number of species detected must likewise be distinguished. For instance, across Latin America, no literature was identified for feeding occurrences in Belize, El Salvador, Honduras, Bolivia, Paraguay, Uruguay, Guyana, Suriname, or French Guiana (Fig. 3A). Feeding events were reported in Argentina, Brazil, Chile, Colombia, Costa Rica, Ecuador, Guatemala, Mexico, Nicaragua, Peru, Trinidad and Tobago, and Venezuela. Mexico was the only country reporting prey species in more than three classes, although research on mammalian species was evenly distributed across the countries. Nevertheless, amphibian and reptilian prey sources were only documented in Mexico, and avian species in Mexico, Guatemala, Peru, Chile, Trinidad and Tobago, Argentina, and Brazil. Despite distribution of many prey species throughout most of Latin America, number of prey species was higher in Mexico (n = 32), Peru (n = 19), and Argentina (n = 19). This does not necessarily correlate with the frequency of feeding events documented in each country (Fig. 3B). A total of 49 events of *D. rotundus* feeding in Mexico returned 32 different species, while 33 events in Brazil only returned 13 difference species. There was a significant positive relationship between the number of references and total species documented per country $(F_{1,11} = 113.61,$ p < 0.001, Fig. 3C), suggesting sampling bias where more research effort returns more prey species.

Behavioral trends in predation

Prey species reported had similar solitary (n = 32) and social behavior (n = 31) although most prey were diurnal (n = 29) over nocturnal (n = 19), crepuscular (n = 9), or cathemeral (n = 6) (Fig. 4A). The majority of diurnal prey were social species (33%) as opposed to solitary (13%).





Literature References

Fig. 3 Geographic distribution of *Desmodus rotundus* prey. A Number of references of *D. rotundus* prey per country denoting regions with none (gray), and low (light red) to high (dark red) number of publications. **B** Number of unique prey species documented in per country denoting areas with none (gray), and low (light blue) to high (dark blue) number of species detected. **C** Relationship between

the number of reports published (x-axis) and the number of species detected (y-axis) per country suggesting a significant, positive correlation suggesting that as the research effort increases, the number of new D. rotundus prey species may increase. Countries with the same number of references are separated by comma

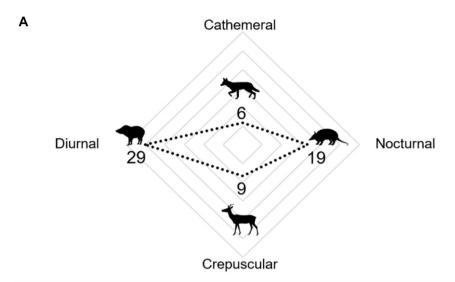
This was consistent at the prey class level (Supplementary Fig. S1), with mammalian and avian prey being primarily diurnal and social. Reptilian and amphibian prey were scattered across the temporal and behavioral patterns, but

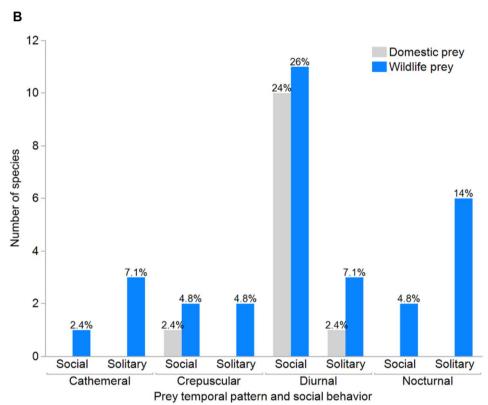
these species were primarily studied in captive settings, and therefore the findings are less extrapolatable.

When removing species solely studied in experimental settings, findings remain similar. The majority of prey



Fig. 4 Behavioral patterns of *Desmodus rotundus* prey. A Number of prey species (dashed line) grouped by temporal behavior as diurnal, nocturnal, crepuscular, cathemeral (vertices). B Number of prey species (y-axis) grouped by prey type (i.e., wildlife or domestic prey) (blue) and temporal patterns of activity with social behavior (x-axis). Only prey species documented in non-captive settings are represented within figure





species demonstrated diurnal activity patterns (59.5%), compared to nocturnal (19.0%), crepuscular (11.9%), and cathemeral (9.5%). Prey likewise demonstrated mainly social (n = 27) over solitary (n = 15) behavior. Combined, half of all prey documented in non-captive settings demonstrated diurnal temporal patterns and social behavior, with similar distributions between wildlife (n = 11) and domestic (n = 10) species for the dual conditions (Fig. 4B). These patterns of behavior suggest that D. rotundus are remarkably flexible in targeting prey with very different

habits but may increasingly predate upon diurnal species that congregate in groups. Nevertheless, the temporal and behavioral characteristics for each species were classified in a general sense (i.e., predominant patterns in taxa across settings) (Supplementary Table S3). That is, there is wide variation in the true patterns behavioral demonstrated by prey species, impacted by the environment and other conditions. For instance, cattle are also odd sleepers, with frequent periods of apparent alertness throughout the night,



which could affect their vulnerability to predation (Balsch 1955; Klefot et al. 2016; Ruckebusch 1972).

Contingency analysis was likewise used to investigate temporal patterns between wild versus domestic prey species. These findings include all species detected, including those in captive settings. Temporal patterns for each species were examined in a context outside of D. rotundus predation events, that is, for differences in the species themselves and not between those selected by D. rotundus over others. Results indicated a significant difference in temporal patterns, with 93.6% of domestic and only 31.6% of wildlife species demonstrating diurnal activity ($\chi^2_{(3)} = 111.3$, p < 0.001). Wildlife prey were primarily nocturnal (40.5%), followed by crepuscular (17.7%) and cathemeral (10.1%; Supplementary Fig. S2A). Upon removal of species documented under experimental conditions, a larger proportion of wildlife was found to be diurnal (44%), but still remained more widely distributed across temporal conditions than domestic species ($\chi^2_{(3)} = 10.2$, p = 0.02). The remaining wildlife species, prey documented in field conditions, were nocturnal (28.0%), followed by crepuscular (16.0%), and cathemeral (12.0%). Domestic species continued to demonstrate primarily diurnal activity (90.9%) and to a larger degree than wildlife taxa. These patterns may, in part, explain an increased tendency to feed on for livestock, given the predominantly diurnal behavior across species, and thus lack of reactivity during D. rotundus prime hunting hours.

Wildlife and domestic prey also demonstrated significant differences in social behavior ($\chi^2_{(1)} = 13.79$, p = 0.002) (Supplementary Fig. SB). Domestic species preyed upon by D. rotundus in all study conditions were primarily social (88.3%), compared to solitary wildlife species (61.7%). For prey species documented only under field conditions, 90.1% of domestic animals demonstrated social behavior. Nevertheless, wild prey were more evenly distributed across the two conditions, with 56% of species considered social and 44% considered solitary ($\chi^2_{(1)} = 4.8$, p = 0.03).

Suitable prey species in areas of invasion

Given the potential for northward range expansion, species distributed in the United States were examined within the dataset. Of the 63 total species identified, 36 are currently present in the United States, including 1 amphibian, 5 avian, 6 reptilian, and 24 mammalian species. Mammals in the United States that may serve as suitable prey for *D. rotundus* include 12 domestic and 23 wild species. Wild mammalian species include the white-tailed deer, wild pig, coyote, puma, Virginia opossum, marmot, squirrel, striped skunk (*Mephitis mephitis*), northern raccoon (*Procyon lotor*), nine-banded armadillo (*Dasypus novemcinctus*), eastern cottontail (*Sylvilagus floridanus*), packrat (*Neotoma* sp.), and house mouse (*Mus musculus*).

The white-tailed deer, wild pig, coyote, puma, and squirrel have been documented as *D. rotundus* prey in non-captive settings, while opossums and marmots have been reported anecdotally. The striped skunk, northern raccoon, ninebanded armadillo, eastern cottontail, and packrat have only been studied as *D. rotundus* prey in captive settings. Wildlife of other prey classes include the rat snake (*Elaphe flavirufa*), pelican (*Pelecanus* sp.), and cormorant of unspecified species, all of which have been described as prey in non-experimental settings.

Discussion

Desmodus rotundus as a generalist predator

This article provides a comprehensive overview of the diet of Desmodus rotundus and reinforces the broad range of prey taxa utilized by this sanguivorous bat species. In addition to their considerably sized dietary collection, D. rotundus maintains the ability to utilize a variety of novel prey in the absence of natural prey sources. This is especially true in cases where wildlife is scarce and alternative food sources, such as livestock, are abundant, as D. rotundus will readily change their dietary reserves. It is, therefore, reasonable to characterize D. rotundus as a generalist predator (Hassell and May 1986; Kang and Wedekin 2013; Schutt 2008) or parasite (Combes 2001; May and Anderson 1990) given the species' dietary array and plasticity. Differentiation between the two depends on the ecological definition of its symbiont roles in an ecosystem and the taxa of prey involved (Buck 2019; Hassell 1966; Raffel et al. 2008). Range expansion of D. rotundus in response to anthropogenic change, therefore, holds significant risks given the dietary breadth and behavior of this species. Identifying at-risk prey, therefore, becomes critical in both preventing and mitigating these issues in ethical and efficacious ways. A clear understanding of the species at risk of *D. rotundus* predation can be used to help key stakeholders anticipate the downstream effects and plan efforts such as vaccination protocols, public health outreach and education, and resource allocation.

Regarding livestock, swine, equine, and poultry are likely at risk, with numerous reports of the *D. rotundus* feeding on these production species. Nevertheless, cattle were the most commonly cited prey throughout the literature. This observation seems to be related to intensification of livestock rearing in areas inhabited by *D. rotundus*, combined with land-use changes that reduce available wildlife prey (Lanzagorta-Valencia et al. 2020; Mantovan et al. 2022; Rojas-Sereno et al. 2022). Likewise, it may be a true pattern of predation, explained by the convenience and availability of this food source (Delpietro et al. 1992; Gonçalves et al. 2021; Voigt and Kelm 2006). As *D. rotundus* expands its geographic



range to higher latitudes and elevations, it will become increasingly crucial that members of the agricultural industry, including producers, large animal veterinarians, and public health officials, continue to stay vigilant to prepare for the potentially increased risk of RABV transmission. This includes maintaining awareness of these distribution changes and the impact *D. rotundus* may have on livestock production systems. Surveillance is particularly important in areas where livestock are abundant (i.e., Texas rangeland), as *D. rotundus* may quickly take to using these agricultural species as a primary food source (Acha and Alba 1988; Anderson and Shwiff 2014).

Dietary shifts upon range expansion are also likely to include various wildlife, as noted by *D. rotundus*' plasticity and range of suitable natural prey. Rodentia has not previously been implicated as a high-risk group for *D. rotundus* predation, but our findings demonstrate the large number of rodent species utilized by *D. rotundus*. Predation on rodents may be due to the diversity of species within this order, but the actual drivers of this association are unknown. Even-toed ungulates and carnivores may also be at risk upon *D. rotundus* invasion, given the number of species currently preyed upon in these taxonomic groups.

In the United States specifically, several wildlife taxa may serve as suitable prey for D. rotundus. Many of those species, such as the white-tailed deer, are widely distributed across the country and comprise ample population sizes (Johns and Kilgo 2005). The large number of rodent species identified in the review may be especially notable in semiarid regions like Texas, where numerous rodents continue to thrive (Windberg 1998; Schmidly and Bradley 2016). Novel species like the nutria (*Myocastor coypus*) may be extremely suitable species for D. rotundus upon invasion, given their morphologic and anatomic similarities to known D. rotundus prey, although additional research is needed to confirm this hypothesis (Swank and Petrides 1954). Moreover, many of the suitable wildlife species in the United States already present significant disease burdens to both domestic and livestock health. For example, the striped skunk, an identified prey source of D. rotundus, is the main reservoir for sylvatic rabies in Texas (Oertli et al. 2009; Wohlers et al. 2018). Desmodus rotundus holds the potential for interspecies RABV transmission and perpetuation of rabies in current reservoirs like the striped skunk and also for introduction of novel virus variants into novel host taxa (Favoretto et al. 2002; Kobayashi et al. 2005). It is, therefore, reasonable to describe the possible impact of D. rotundus on United States wildlife as considerable, particularly in regard to the economics of rabies control. Between 1989 and 2004, the United States spent over \$34 million on oral rabies vaccines for coyotes, a D. rotundus prey species, and gray foxes in Texas alone (Sterner et al. 2009). The country continues to pour funding into efforts to control the sylvatic form of this disease (Slate et al. 2009; Fehlner-Gardiner 2018). Range expansion of *D. rotundus* is already estimated to be a costly event (Anderson and Shwiff 2014; Zarza et al. 2017), although this estimation neglects the costs of control in the reservoir (e.g., *D. rotundus* vaccination, population control, monitoring) and may, thus, be underestimated.

While the risk of RABV transmission and occurrence of clinical disease in companion animals is lower, given the robust vaccination protocols typically used in pets (Ma et al. 2018), D. rotundus may predate upon domestic dogs and cats. Alternatively, dogs and cats can predate D. rotundus (Delpietro et al. 1994). Both events have implications for public health considering that reports of pets positive to rabies derived from D. rotundus are common (Badilla et al. 2003; de Mattos et al. 1996; Ito et al. 2001; Horta et al. 2022; Schaefer et al. 2002). In Latin America, domestic dogs were once RABV reservoirs of significant public health importance, causing a large number of human rabies cases (Horta et al. 2022). The number of dog-mediated human cases eventually declined in certain regions through pet vaccination campaigns, public health efforts, and widely available post-exposure prophylaxis, though their present role is non-negligible (Horta et al. 2022). Range shifts of D. rotundus, driven by land-use changes, led this species to surpass domestic dogs as the primary RABV reservoir in the region (Horta et al. 2022). As a result, human rabies cases have risen in recent years in some regions of Latin America (Horta et al. 2022). As D. rotundus expands to new ranges and potentially becomes established in areas like the United States, the risks to humans and companion animals may rise similarly to those seen in Brazil.

Mitigating the effects of *Desmodus rotundus* invasion

Although D. rotundus range expansion seems to be imminent due to ongoing ecosystem change, actions may be taken to mitigate the potential consequences of rabies range expansion, including a sustained availability of wildlife prey through biodiversity and habitat management. Anthropogenic change is known to be closely amalgamated with conflict and disease spread between bats, domestic animals, and humans (Stoner-Duncan et al. 2014). The relationship between diminished natural environments, declines in wildlife populations, readily available domestic species, and distribution changes of D. rotundus is critical to consider in mitigation attempts. As previously described, D. rotundus prey choice is likely attributable to convenience (Delpietro et al. 1992; Goncalves et al. 2021; Voigt and Kelm 2006). In Latin America, land-use change not only increases the abundance of livestock, but in doing so, reduces the richness and distribution of wildlife taxa (Grau and Aide 2008; Michalski et al. 2006). Under these conditions, D. rotundus



readily predates upon human-supplied species such as livestock and companion animals (Acha and Alba 1988; Rojas-Sereno et al. 2022). As D. rotundus expands into new territories, it is reasonable to assume that domestic animal populations in invaded zones will follow similar patterns and utilize the domestic species readily available to them if wildlife is scarce or absent. An ecosystem-health approach to D. rotundus management, incorporating landscape, wildlife, and domestic species ecology, is, therefore, of great benefit to mitigation efforts. Conservations attempts in this context may serve as an indirect disease prevention tool and should be made not only in regard to wildlife richness, but native habitats and the larger ecological community as well. Ensuring abundant prey is available, via maintenance of suitable wildlife habitat, is especially critical in regions recently converted for agriculture use (Mayen 2003; Uhart and Milano 2002). This integrative approach to disease mitigation is an added benefit for bat population health, which must also be considered in discussions of ecological impartiality. The larger effects of bats on ecosystem health are well documented, and although less clearly defined for D. rotundus, conservation should be pursued when possible to avoid undue ecological damage associated with bat population declines (Kunz et al. 2011; Stoner-Duncan et al. 2014). In general, bats provide valuable ecosystem services including insect control, pollination, and seed dispersal (Kunz et al. 2011). Guano, produced by D. rotundus specifically, contains greater nitrogen content than that of various frugivorous and insectivorous bats (Emerson and Roark 2007; Hadas and Rosenberg 1992). Ultimately, the loss of bat species may lead to further ecologic impairment and agricultural loss (Kunz et al. 2011) and this may hold implications for *D. rotundus* itself.

A conservation-minded approach to disease control is not to undermine the public and veterinary health risks associated with D. rotundus. Nevertheless, some form of population control is considered to be required for *D. rotundus*, given their overabundance and potential for public health and economic harm in some sites (Gonzalez and Mitchell 1976; Johnson et al. 2014; León et al. 2021). Failing to control the species through selective measures may lead to a rise in the utilization of nonselective and unethical methods (Mayen 2003; Olival 2016). For example, D. rotundus have historically been subjected to and targeted by campaigns fueled by fear, with little evidence-based methodology behind them (Mayen 2003). Cementing roosts closed, destroying roosts with explosives, and poisoning D. rotundus have all been used to as forms of population control (Gonzalez and Mitchell 1976; Greenhall 1988). Many other traditional control methods have been attempted with often limited efficacy and longevity, including both chemical and physical means of *D. rotundus* depopulation (Arellano 1988; Gonçalves et al. 2002; Johnson et al. 2014; León et al. 2021;

Thompson et al. 1972). More recently, nonselective population reduction and control methods have been highlighted as ineffective (Streicker et al. 2012), counterproductive (Viana et al. 2023), inhumane (Olival 2016), and a threat to other bat species that may be critical in maintaining ecosystem services (Mayen 2003; O'Shea et al. 2016). Research suggests that for culling to be effective, it would need to occur at a scale that is essentially not feasible (Gonçalves et al. 2021; Streicker et al. 2012) or in bat populations with absence of RABV circulation (Viana et al 2023). Likewise, mass culling of D. rotundus may lead to increased disease spread via displacement of colonies in addition to the displacement already induced from habitat fragmentation itself (Streicker et al. 2012). This activity may contribute to the persistence of rabies via movement of infectious bats between colonies following frequent immunizing but non-lethal exposure (Blackwood et al. 2013). Similarly, the stress associated with culling activities may increase the likelihood of viral shedding (Olival 2016). Minimal disruption of roosts is now recommended in discussions of D. rotundus control (Delpietro et al. 2017; Rocha and Dias 2020; Streicker et al. 2012).

In Latin America, efforts to control D. rotundus have involved the administration of topical anticoagulants to cattle, ultimately causing death via hemorrhage to D. rotundus that feed on the bovids (Mayen 2003; Rocha and Dias 2020). Finely targeted control involves the direct application of 2\% warfarin in Vaseline on the mid-dorsum of D. rotundus to promote intoxication of the colony during grooming (Caraballo and Alejandro 1996). This solution is also applied along the walls of *D. rotundus* roosts, though this leads to indiscriminate killing of other bat species (Johnson et al. 2014). A more costly but selective variation involves the application of warfarin in paste formulation to bite wounds of cattle, targeting D. rotundus individuals that return to feed on cattle across multiple nights (Piccinini et al. 1998). Warfarin has also been injected directly into cattle to depopulate D. rotundus (Flores Crespo et al. 1979), but effects of this practice on the health of livestock and human consumers are unclear. Ultimately, no single population control strategy fits all scenarios and there is a need for continued and updated research on the topic. Sustainable and effective rabies control and prevention methods include diligent vaccination of livestock (Anderson et al. 2014) and oral vaccination of D. rotundus (Bakker et al. 2019). The latter is an ongoing development in terms of implementation (Delpietro et al. 2021; Sétien et al. 1998; Stoner-Duncan et al. 2014). It has more recently been under assessment in the context of D. rotundus social behavior, that is, utilizing social grooming as a means to introduce and spread vaccines throughout a colony (Delpietro et al. 2021; Sétien et al. 1998; Stoner-Duncan et al. 2014). The effects of rabies vaccination on D. rotundus and the prevalence of other bat-borne pathogens and bat demography are unknown. Reproductive drugs to sterilize bats

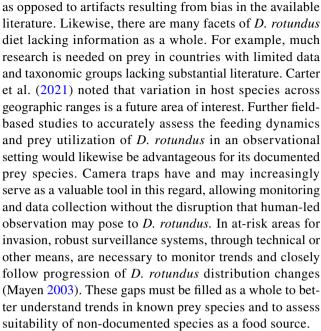


have also been discussed as a population control and rabies prevention tool for the species (Gonçalves et al. 2021; Serrano et al. 2007). Other efforts have focused on minimizing the incidence of bites and exposure of livestock to the bats (Stoner-Duncan et al. 2014). Modifications to agricultural practices that may deter *D. rotundus* establishment include netting, artificial light, and ultrasound acoustics (Arnett et al. 2013; Benavides et al. 2020a; Delpietro 1989; Gonçalves et al. 2021). Nevertheless, these interventions could also exacerbate the displacement of colonies to new areas, expanding the human—wildlife conflict. The use of repellant to selectively deter *D. rotundus* from preying on livestock has also been suggested (Mayen 2003).

Furthermore, and in regard to disease risk, much of the focus lies on D. rotundus as a disease reservoir and threat to domestic animal health. Given the diversity of wildlife species found in this review, it may be of value to examine the role of the different wildlife species in the transmission of RABV (Worsley-Tonks et al. 2020). This may guide decisions regarding resource allocation and target species for rabies prevention and control efforts. For example, rodents in particular may be an important prey species, but they do not play a relevant role in the epidemiology of rabies after contact with an infected D. rotundus (Winkler et al. 1972). Rodents, however, may serve as important reservoirs of other diseases that *D. rotundus* is known to host such as hantavirus (Sabino-Santos Jr et al. 2018) or leptospirosis (Cosson et al 2014). As previously mentioned, certain wildlife species (i.e., those that play a role in the propagation of rabies) in atrisk regions of invasion require additional research on their interactions with D. rotundus. The striped skunk, a primary RABV reservoir in Texas, was demonstrated as D. rotundus prey, but only in captive and experimental settings. Studies to clarify whether D. rotundus would target this species under field conditions are needed to better characterize the species-specific risks that might present following invasion distribution changes. This is particularly important given the role of skunks as reservoir of RABV in regions of potential D. rotundus invasion.

Future directions

Nonrandom bias exists in study location, prey detection methods, number of samples per site, and type of *D. rotundus* prey (i.e., wildlife versus livestock), among others. These biases make interpretation of the proposed trends problematic and increase uncertainty in any extrapolations of the patterns to other areas of potential invasion. It is, therefore, difficult to infer patterns of prey selection and potential driving factors behind it, particularly without baseline data to control for species richness or density within each study location. The trends identified in this review cannot be confirmed as true biological variation



Lastly, continuing outreach on the distributional ecology of D. rotundus is needed given their impending range expansion, the broad taxonomic range of suitable prey, and the magnitude of the impact that invasion of this species holds. Engagement of key stakeholders, including wildlife health professionals, researchers, veterinarians, public health officials, and producers is critical. This is particularly vital in areas of potential D. rotundus range expansion, such as the United States-Mexico border region and the Andes region in South America. Given the dietary adaptations of and its plasticity to use disparate prey, D. rotundus functions as an ideal biological model to investigate the effects of global change on predator-prey dynamics at the human-wildlife interface. The ecology of D. rotundus in the context of ongoing anthropogenic change is deserving of continued research attention, with significant implications for the health of humans, livestock, and wildlife alike.

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Author contributions NB and LEE devised the project, developed the proof outline, and analyzed relevant data. NB performed data collection and wrote the first draft of the manuscript in consultation with LEE. All authors approved the final version of the article.



Data availability The authors confirm that the data supporting the findings of this study are available within the article [and/or] its supplementary materials.

Declarations

Conflicts of interest On behalf of all authors, the corresponding author states that there is no conflict of interest.

References

- Acha PN (1967) Epidemiology of paralytic bovine rabies and bat rabies. Bull off Int Epizoot 67:343–382
- Acha PN, Alba AM (1988) Economic losses due to *Desmodus rotundus*. In: Greenhall AM, Schmidt U (eds) Natural history of vampire bats. CRC Press, Boca Raton, pp 208–214
- Acha P, Szyfres B (1986) Zoonosis y enfermedades transmisibles comunes al hombre y a los animals. Organización Panamericana de la Salud Publicación Científica, Washington
- Altizer S, Bartel R, Han BA (2011) Animal migration and infectious disease risk. Science 331:296–302. https://doi.org/10.1126/science.1194694
- Altrichter M, Boaglio GI (2004) Distribution and relative abundance of peccaries in the Argentine Chaco: associations with human factors. Biol Conserv 116:217–225. https://doi.org/10.1016/s0006-3207(03)00192-7
- Alves RS, do Canto Olegário J, Weber MN, da Silva MS, Canova R, Sauthier JT, Baumbach LF, Witt AA, Varela APM, Mayer FQ, da Fontoura Budaszewski R, Canal CW (2021) Detection of coronavirus in vampire bats (*Desmodus rotundus*) in southern Brazil. Transbound Emerg Dis 69:2384–2389. https://doi.org/10.1111/tbed.14150
- Anciaux de Faveaux M (1971) Catalogue des acariens parasites et commensaux des chiroptères. Institut Royal des Sciences Naturelles de Belgique, Bruxelles
- Ancillotto L, Santini L, Ranc N, Maiorano L, Russo D (2016) Extraordinary range expansion in a common bat: the potential roles of climate change and urbanisation. Sci Nat 3:103–108. https://doi.org/10.1007/s00114-016-1334-7
- Anderson A, Shwiff S (2014) Economic impact of the potential spread of vampire bats into south Texas. Proc Vertebr Pest Conf 26:305–309. https://doi.org/10.5070/V426110474
- Arata AA, Vaughn JB, Thomas ME (1967) Food habits of certain Colombian bats. J Mamm 48:653–655. https://doi.org/10.2307/1377595
- Arellano SC (1988) Biology, ecology, and control of the vampire bat. Rev Infect Dis 10:615–619. https://doi.org/10.1093/clinids/10. supplement_4.s615
- Arellano SC, Greenhall AM (1971) Preferencia de la predación del vampiro en relación a la edad y raza del ganado ya la época del año. Técnica Pecuaria 17:20–23
- Arnett EB, Hein CD, Schirmacher MR, Huso MMP, Szewczak JM (2013) Evaluating the effectiveness of an ultrasonic acoustic deterrent for reducing bat fatalities at wind turbines. PLoS ONE 8:e65794. https://doi.org/10.1371/journal.pone.0065794
- Arteaga FL, Miragaya M, Molina N, Mondino MA, Bracamonte JC, Capitelli GM, Mundo S, Bratanich AC (2022) Circulation of coronavirus in bats from northern and central Argentina: preliminary study. Int J Infect Dis 116:S76–S77. https://doi.org/10.1016/j.ijid.2021.12.181
- Asano KM, Hora AS, Scheffer KC, Fahl WO, Iamamoto K, Mori E, Brandão PE (2016) Alphacoronavirus in urban Molossidae and

- Phyllostomidae bats, Brazil. Virol J 13:110. https://doi.org/10. 1186/s12985-016-0569-4
- Badilla X, Pérez-Herra V, Quirós L, Morice A, Jiménez E, Sáenz E, Salazar F, Fernández R, Orciari L, Yager P, Whitfield S, Rupprecht CE (2003) Human rabies: a reemerging disease in Costa Rica? Emerg Infect Dis 9:721–723. https://doi.org/10.3201/eid0906.020632
- Bakker KM, Rocke TE, Osorio JE, Abbott RC, Tello C, Carrera JE, Valderrama W, Shiva C, Falcon N, Streicker DG (2019) Fluorescent biomarkers demonstrate prospects for spreadable vaccines to control disease transmission in wild bats. Nat Ecol Evol 3:1697–1704. https://doi.org/10.1038/s41559-019-1032-x
- Ballados-González GG, Sánchez-Montes S, Romero-Salas D, Colunga Salas P, Gutiérrez-Molina R, León-Paniagua L, Becker I, Méndez-Ojeda ML, Barrientos-Salcedo C, Serna-Lagunes R, Cruz-Romero A (2018) Detection of pathogenic *Leptospira* species associated with phyllostomid bats (Mammalia: Chiroptera) from Veracruz, Mexico. Transbound Emerg Dis 65:773–781. https://doi.org/10.1111/tbed.12802
- Ballejo F, Lambertucci SA, Trejo A, Santis LJMD (2018) Trophic niche overlap among scavengers in Patagonia supports the condor-vulture competition hypothesis. Bird Conserv Int 28:390–402. https://doi.org/10.1017/s0959270917000211
- Barlow J, Lennox GD, Ferreira J, Berenguer E, Lees AC, Nally RM, Thomson JR, de Ferraz SFB, Louzada J, Oliveira VHF, Parry L, de Castro R, Solar R, Vieira ICG, Aragão LEOC, Begotti RA, Braga RF, Cardoso TM, de Oliveira RC, Souza CM Jr, Moura NG, Nunes SS, Siqueira JV, Pardini R, Silveira JM, Vaz-de-Mello FZ, Veiga RCS, Venturieri A, Gardner TA (2016) Anthropogenic disturbance in tropical forests can double biodiversity loss from deforestation. Nature 535:144–147. https://doi.org/10.1038/nature18326
- Barquez RM, Carbajal MN, Failla M, Díaz MM (2013) New distributional records for bats of the Argentine Patagonia and the southernmost known record for a molossid bat in the world. Mammalia 77:119–126. https://doi.org/10.1515/mammalia-2012-0053
- Bellard C, Bertelsmeier C, Leadley P, Thuiller W, Courchamp F (2012) Impacts of climate change on the future of biodiversity. Ecol Lett 15:365–377. https://doi.org/10.1111/j.1461-0248.2011.01736.x
- Benavides JA, Paniagua ER, Hampson K, Valderrama W, Streicker DG (2017) Quantifying the burden of vampire bat rabies in Peruvian livestock. PLoS Negl Trop Dis 11:e0006105. https://doi.org/10.1371/journal.pntd.0006105
- Benavides JA, Valderrama W, Recuenco S, Uieda W, Suzán G, Avila-Flores R, Velasco-Villa A, Almeida M, de Andrade FAG, Molina-Flores B, Vigilato MAN, Pompei JCA, Tizzani P, Carrera JE, Ibanez D, Streicker DG (2020a) Defining new pathways to manage the ongoing emergence of bat rabies in Latin America. Viruses 12:1002. https://doi.org/10.3390/v12091002
- Benavides JA, Velasco-Villa A, Godino LC, Satheshkumar PS, Nino R, Rojas-Paniagua E, Shiva C, Falcon N, Streicker DG (2020b) Abortive vampire bat rabies infections in Peruvian peridomestic livestock. PLoS Negl Trop Dis 14:e0008194. https://doi.org/10.1371/journal.pntd.0008194
- Benavides JA, Godreuil S, Opazo-Capurro A, Mahamat OO, Falcon N, Oravcova K, Streicker DG, Shiva C (2022) Long-term maintenance of multidrug-resistant *Escherichia coli* carried by vampire bats and shared with livestock in Peru. Sci Total Environ 810:152045. https://doi.org/10.1016/j.scitotenv.2021.152045
- Bengtsson J, Ahnström J, Weibull A-C (2005) The effects of organic agriculture on biodiversity and abundance: a meta-analysis. J Appl Ecol 42:261–269. https://doi.org/10.1111/j.1365-2664. 2005.01005.x
- Bergner LM, Orton RJ, da Silva FA, Shaw AE, Becker DJ, Tello C, Biek R, Streicker DG (2019) Using noninvasive metagenomics



- to characterize viral communities from wildlife. Mol Ecol Resour 19:128–143. https://doi.org/10.1111/1755-0998.12946
- Bergner LM, Orton RJ, Streicker DG (2020) Complete genome sequence of an alphacoronavirus from common vampire bats in Peru. Microbiol Resour Announc 9:e00742-e820. https://doi.org/10.1128/mra.00742-20
- Bergner LM, Mollentze N, Orton RJ, Tello C, Broos A, Biek R (2021a) Characterizing and evaluating the zoonotic potential of novel viruses discovered in vampire bats. Viruses 13:252. https://doi.org/10.3390/v13020252
- Bergner LM, Orton RJ, Broos A, Tello C, Becker DJ, Carrera JE (2021b) Diversification of mammalian deltaviruses by host shifting. Proc Natl Acad Sci U S A 3:e2019907118. https://doi.org/10.1073/pnas.2019907118
- Blackwood JC, Streicker DG, Altizer S, Rohani P (2013) Resolving the roles of immunity, pathogenesis, and immigration for rabies persistence in vampire bats. Proc Natl Acad Sci U S A 110:20837. https://doi.org/10.1073/pnas.1308817110
- Bobrowiec PED, Lemes MR, Gribel R (2015) Prey preference of the common vampire bat (*Desmodus rotundus*, Chiroptera) using molecular analysis. J Mamm 96:54–63. https://doi.org/10.1093/jmamma/gyu002
- Bodenchuk MJ, Bergman DL (2020) Vampire bats: preparing for range expansion into the U.S. Proc Vertebr Pest Conf 29:1–2
- Bohmann K, Gopalakrishnan S, Nielsen M, dos Nielsen LSB, Jones G, Streicker DG, Gilbert MTP MTP (2018) Using DNA metabarcoding for simultaneous inference of common vampire bat diet and population structure. Mol Ecol Resour 18:1050–1063. https://doi.org/10.1111/1755-0998.12891
- Bolívar-Cimé B, Flores-Peredo R, García-Ortíz SA, Murrieta-Galindo R, Laborde J (2019) Influence of landscape structure on the abundance of *Desmodus rotundus* (Geoffroy 1810) in northeastern Yucatan, Mexico. Ecosist Recur Agropecu 6:263–271. https://doi.org/10.19136/era.a6n17.1968
- Borremans B, Faust C, Manlove KR, Sokolow SH, Lloyd-Smith JO (2019) Cross-species pathogen spillover across ecosystem boundaries:mechanisms and theory. Philos Trans R Soc B Biol Sci 374:20180344. https://doi.org/10.1098/rstb.2018.0344
- Botto Nuñez G, Becker DJ, Lawrence RL, Plowright RK (2020) Synergistic effects of grassland fragmentation and temperature on bovine rabies emergence. EcoHealth 17:203–216. https:// doi.org/10.1007/s10393-020-01486-9
- Buck JC (2019) Indirect effects explain the role of parasites in ecosystems. Trends Parasitol 35:835–847. https://doi.org/10.1016/j.pt.2019.07.007
- Burki T (2020) The origin of SARS-CoV-2. Lancet Infect Dis 20:1018–1019. https://doi.org/10.1016/S1473-3099(20) 30641-1
- Cabré MF, Solman S, Núñez M (2016) Regional climate change scenarios over southern South America for future climate (2080–2099) using the MM5 Model. Mean, interannual variability and uncertainties. Atmósfera 29:35–60. https://doi.org/10.20937/atm. 2016.29.01.04
- Calfayan L, Bonnot G, Gómez Villafañe I (2019) Case reports of common vampire bats *Desmodus rotundus* (Chiroptera:Phyllostomid ae:Desmodontinae) attacking wild exotic mammals in Argentina. Notas Sobre Mamíferos Sudam 1:1–6. https://doi.org/10.31687/saremnms.19.0.05
- Camargo A, Olivera R, Santiago M, González JC (2018) Genetic relatedness of *Desmodus rotundus* from northern Uruguay with populations from the remainder of its distribution range. Bol Soc Zool Urug 1:14–18. https://doi.org/10.26462/27.1.5
- Campos-Krauer JM, Wisely SM (2011) Deforestation and cattle ranching drive rapid range expansion of capybara in the Gran Chaco ecosystem. Glob Chang Biol 17:206–218. https://doi.org/10.1111/j.1365-2486.2010.02193.x

- Caraballo H, Alejandro J (1996) Outbreak of vampire bat biting in a Venezuelan village. Rev Saude Publ 30:483–484
- Cárdenas-Canales EM, Gigante CM, Greenberg LA, Velasco-Villa A, Ellison JA, Satheshkumar PS, Medina-Magües LG, Griesser R, Falendysz E, Amezcua I, Osorio JE, Rocke TE (2020) Clinical presentation and serological responses to natural outbreaks of rabies in a captive colony of common vampire bats. Trop Med Infect Dis. https://doi.org/10.3390/tropicalmed5010034
- Cardinale BJ, Srivastava DS, Emmett Duffy J, Wright JP, Downing AL, Sankaran M, Jouseau C (2006) Effects of biodiversity on the functioning of trophic groups and ecosystems. Nature 443:989–992. https://doi.org/10.1038/nature05202
- Carter G, Brown B, Razik I, Ripperger S (2021) Penguins, falcons, and mountain lions: the extraordinary host diversity of vampire bats. In: Lim BK, Fenton MB, Brigham RM, Mistry S, Kurta A, Gillam EH, Russell A, Ortega J (eds) 50 years of bat research: foundations and new frontiers. Springer Cham, New York, pp 151–170
- Chan AAY-H, Giraldo-Perez P, Smith S, Blumstein DT (2010) Anthropogenic noise affects risk assessment and attention: the distracted prey hypothesis. Biol Lett 6:458–461. https://doi.org/10.1098/rsbl.2009.1081
- Chaudhary A, Mooers AO (2018) Terrestrial vertebrate biodiversity loss under future global land use change scenarios. Sustainability 10:2764. https://doi.org/10.3390/su10082764
- Clavero M, García-Berthou E (2005) Invasive species are a leading cause of animal extinctions. Trends Ecol Evol 20:110. https://doi.org/10.1016/j.tree.2005.01.003
- Combes C (2001) Parasitism: the ecology and evolution of intimate interactions. University of Chicago Press, London
- Constantine DG (1979) Bat rabies and bat management. Bull Soc Vector Ecol 4:1–9
- Constantine DG, Blehert DS (2009) Bat rabies and other *Lyssavirus* infections. US Geological Survey Circular 1329. https://doi.org/10.3133/cir1329
- Cosson JF, Picardeau M, Mielcarek M, Tatard C, Chaval Y, Suputtamongkol Y, Buchy P, Jittapalapong S, Herbreteau V, Morand S (2014) Epidemiology of *Leptospira* transmitted by rodents in Southeast Asia. PLoS Negl Trop Dis 8:e2902. https://doi.org/10.1371/journal.pntd.0002902
- de Mattos CA, de Mattos CC, Smith JS, Miller ET, Papo S, Utrera A, Osburn BI (1996) Genetic characterization of rabies field isolates from Venezuela. J Clin Microbiol 34:1553–1558. https://doi.org/ 10.1128/jcm.34.6.1553-1558.1996
- Delpietro HA (1989) Case reports on defensive behaviour in equine and bovine subjects in response to vocalization of the common vampire bat (*Desmodus rotundus*). Appl Anim Behav Sci 22:377–380. https://doi.org/10.1016/0168-1591(89)90032-4
- Delpietro HA, Russo RG (1996) Ecological and epidemiologic aspects of the attacks by vampire bats and paralytic rabies in Argentina and analysis of the proposals carried out for their control. Rev Sci Tech 15:971–984
- Delpietro HA, Marchevsky N, Simonetti E (1992) Relative population densities and predation of the common vampire bat (*Desmodus rotundus*) in natural and cattle-raising areas in north-east Argentina. Prev Vet Med 14:13–20. https://doi.org/10.1016/0167-5877(92)90080-v
- Delpietro H, Konolsaisen F, Marchevsky N, Russo G (1994) Domestic cat predation on vampire bats (*Desmodus rotundus*) while foraging on goats, pigs, cows and human beings. Appl Anim Behav Sci 39:141–150. https://doi.org/10.1016/0168-1591(94)90134-1
- Delpietro HA, Lord RD, Russo RG, Gury-Dhomen F (2009) Observations of sylvatic rabies in northern Argentina during outbreaks of paralytic cattle rabies transmitted by vampire bats (*Desmodus rotundus*). J Wildl Dis 45:1169–1173. https://doi.org/10.7589/0090-3558-45.4.1169



- Delpietro HA, Russo RG, Carter GG, Lord RD, Delpietro GL (2017) Reproductive seasonality, sex ratio and philopatry in Argentina's common vampire bats. R Soc Open Sci 4:160959. https://doi.org/ 10.1098/rsos.160959
- Delpietro HA, Russo RG, Rupprecht CE, Delpietro GL (2021) Towards development of an anti-vampire bat vaccine for rabies management: inoculation of vampire bat saliva induces immune-mediated resistance. Viruses 13:515. https://doi.org/10.3390/v1303 0515
- Dos Santos AJF, Ferreira JM, Baptista F, Alexandrino B, da Silva MAG, Gomes JEC, Júnior JPV, Tavares RM, de Sousa AK (2022) Statistical analysis between 2006 and 2019 and forecast of rabies in cattle from 2020 to 2022 in Tocantins State (Brazil), by using the R Studio software. Epi Inf. https://doi.org/10.1017/s0950268822000553
- Dros JM (2004) Managing the soy boom: two scenarios of soy production. AIDEnvironment, Amsterdam
- Dunn LH (1932) Experiments in the transmission of *Trypanonoma hippicum* (Darling) with the vampire bat, *Desmodus rotundus murinus* (Wagner), as a vector in Panama. J Prev Med 6:415–424
- Eisenbeis G, Hänel A, McDonnell M, Hahs A, Breuste J (2009) Light pollution and the impact of artificial night lighting on insects. In: McDonnell MJ, Hahs AK, Breuste JH (eds) Ecology of cities and towns: a comparative Approach. Cambridge University Press, New York, pp 243–263
- Ellis EC, Klein Goldewijk K, Siebert S, Lightman D, Ramankutty N (2010) Anthropogenic transformation of the biomes, 1700 to 2000. Glob Ecol Biogeogr 19:589–606. https://doi.org/10.1111/j. 1466-8238.2010.00540.x
- El-Sabaawi R (2018) Trophic structure in a rapidly urbanizing planet. Funct Ecol 32:1718–1728. https://doi.org/10.1111/1365-2435. 13114
- Emerson JK, Roark AM (2007) Composition of guano produced by frugivorous, sanguivorous, and insectivorous bats. Acta Chiropt 9:261–267. https://doi.org/10.3161/1733-5329(2007)9[261: COGPBF]2.0.CO;2
- Favoretto SR, Carrieri ML, Cunha EMS, Aguiar EAC, Silva LHQ, Sodré MM, Souza MCAM, Kotait I (2002) Antigenic typing of Brazilian rabies virus samples isolated from animals and humans, 1989–2000. Rev Inst Med Trop Sao Paulo 44:91–95. https://doi. org/10.1590/s0036-46652002000200007
- Fehlner-Gardiner C (2018) Rabies control in North America—past, present and future. Rev off Int Epizoot 37:421–437. https://doi.org/10.20506/rst.37.2.2812
- Fleming PA, Bateman PW (2018) Novel predation opportunities in anthropogenic landscapes. Anim Behav 138:145–155. https://doi.org/10.1016/j.anbehav.2018.02.011
- Flores Crespo R, Said Fernández S, De Anda LD, Ibarra Velarde F, Anaya RM (1979) Nueva técnica para el combate de los vampiros: Warfarina por vía intramuscular al ganado bovino. Bol Oficina Sanit Panam 87:4
- Franco-Molina MA, Santana-Krímskaya SE, Cortés-García B, Sánchez-Aldana-Pérez JA, García-Jiménez O, Kawas J (2021) Fatal case of rabies in a captive white-tailed deer: a case report from Chiapas, Mexico. Trop Med Inf Dis 6:135. https://doi.org/10.3390/tropicalmed6030135
- Freitas MB, Welker AF, Millan SF, Pinheiro EC (2003) Metabolic responses induced by fasting in the common vampire bat *Desmodus rotundus*. J Comp Phys B 173:703–707. https://doi.org/10.1007/s00360-003-0383-3
- Galetti M, Pedrosa F, Keuroghlian A, Sazima I (2016) Liquid lunch-vampire bats feed on invasive feral pigs and other ungulates. Front Ecol Environ 14:505–506. https://doi.org/10.1002/fee.1431
- Gámez S, Potts A, Mills KL, Allen AA, Holman A, Randon PM, Linson O, Harris NC (2022) Downtown diet:a global metaanalysis of increased urbanization on the diets of vertebrate

- predators. Proc R Soc B 289:20212487. https://doi.org/10.1098/rspb.2021.2487
- Gnocchi AP, Srbek-Araujo AC, Gnocchi AP, Srbek-Araujo AC (2017) Common vampire bat (*Desmodus rotundus*) feeding on lowland tapir (*Tapirus terrestris*) in an Atlantic forest remnant in southeastern Brazil. Biota Neotrop 17:e20170326. https://doi.org/10.1590/1676-0611-bn-2017-0326
- Gonçalves MAS, Sá-Neto RJ, Brazil TK (2002) Outbreak of aggressions and transmission of rabies in human beings by vampire bats in northeastern Brazil. Rev Soc Bras Med Trop 35:453–460. https://doi.org/10.1590/s0037-86822002000500006
- Gonçalves F, Galetti M, Streicker DG (2021) Management of vampire bats and rabies: a precaution for rewilding projects in the neotropics. Perspect Ecol Conserv 19:37–42. https://doi.org/10.1016/j.pecon.2020.12.005
- Gonzalez R, Mitchell GC (1976) Vampire bat control programs in Latin America. Proc Vertebr Pest Conf 7:7
- Goodwin GG, Greenhall AM (1961) A review of the bats of Trinidad and Tobago:descriptions, rabies infection, and ecology. Bull Am Mus 122:3. https://doi.org/10.2307/1377397
- Grabrucker S, Grabrucker AM (2010) Rare feeding behavior of great-tailed grackles (*Quiscalus mexicanus*) in the extreme habitat of Death Valley. Open Ornithol J 3:1. https://doi.org/ 10.2174/1874453201003010101
- Grantham HS, Duncan A, Evans TD, Jones KR, Beyer HL, Schuster R, Walston J, Ray JC, Robinson JG, Callow M, Clements T, Costa HM, DeGemmis A, Elsen PR, Ervin J, Franco P, Goldman E, Goetz S, Hansen A, Hofsvang E, Jantz P, Jupiter S, Kang A, Langhammer P, Laurance WF, Lieberman S, Linkie M, Malhi Y, Maxwell S, Mendez M, Mittermeier R, Murray NJ, Possingham H, Radachowsky J, Saatchi S, Samper C, Silverman J, Shapiro A, Strassburg B, Stevens T, Stokes E, Taylor R, Tear T, Tizard R, Venter O, Visconti P, Wang S, Watson JEM (2020) Anthropogenic modification of forests means only 40% of remaining forests have high ecosystem integrity. Nat Commun 11:5978. https://doi.org/10.1038/s41467-020-19493-3
- Grau H, Aide M (2008) Globalization and land-use transitions in Latin America. Ecol Soc 13:2. https://doi.org/10.5751/es-02559-130216
- Green AM, Barnick KA, Pendergast ME, Şekercioğlu ÇH (2022) Species differences in temporal response to urbanization alters predator-prey and human overlap in northern Utah. Glob Ecol Conserv 36:e02127. https://doi.org/10.1016/j.gecco.2022.e02127
- Greenhall AM (1964) Bats: their public health importance and control with special reference to Trinidad. Proc Vertebr Pest Conf. https://escholarship.org/uc/item/6r04h653. Accessed 22 June 23
- Greenhall AM (1972) The biting and feeding habits of the vampire bat, *Desmodus rotundus*. J Zool 168:451–461. https://doi.org/10.1111/j.1469-7998.1972.tb01361.x
- Greenhall AM (1988) Natural history of vampire bats. CRC Press, Boca Raton
- Greenhall AM, Schmidt U, Lopez Forment CM (1969) Field observations on the mode of attack of the vampire bat (*Desmodus rotundus*) in Mexico. An Inst Biol UNAM Zool 40:245–252. https://doi.org/10.2307/2989817
- Greenhall AM, Schmidt U, Lopez-Forment W (1971) Attacking behavior of the vampire bat, *Desmodus rotundus*, under field conditions in Mexico. Biotropica 3:136–141. https://doi.org/10.2307/2989817
- Greenhall AM, Joermann G, Schmidt U, Seidel MR (1983) Desmodus rotundus. Mamm Species 1983:2021–2026. https://doi.org/10.1093/mspecies/202.1a
- Grotta-Neto F, Peres PHF, Piovezan U, Passos FC, Duarte JMB (2021) Hunting practices of feral pigs (Sus scrofa) and predation by vampire bats (Desmodus rotundus) as a potential route of rabies



- in the Brazilian Pantanal. Austral Ecol 46:324–328. https://doi.org/10.1111/aec.12971
- Guiden PW, Bartel SL, Byer NW, Shipley AA, Orrock JL (2019) Predator–prey interactions in the Anthropocene: reconciling multiple aspects of novelty. Trends Ecol Evol 34:616–627. https://doi.org/10.1016/j.tree.2019.02.017
- Gupta R (2005) Recent outbreak of rabies infections in Brazil transmitted by vampire bats. Euro Surveill 10(E051110):3. https://doi.org/10.2807/esw.10.45.02831-en
- Hadas A, Rosenberg R (1992) Guano as a nitrogen source for fertigation in organic farming. Fert Res 31:209–214. https://doi.org/10.1007/BF01063294
- Halpin K, Young PL, Field HE, Mackenzie JSY (2000) Isolation of Hendra virus from pteropid bats:a natural reservoir of Hendra virus. J Gen Virol 81:1927–1932. https://doi.org/10.1099/ 0022-1317-81-8-1927
- Hassell MP (1966) Evaluation of parasite or predator responses. J Anim Ecol. https://doi.org/10.2307/2690
- Hassell MP, May RM (1986) Generalist and specialist natural enemies in insect predator-prey interactions. J Anim Ecol 55:923–940. https://doi.org/10.2307/4425
- Hayes M, Piaggio A (2018) Assessing the potential impacts of a changing climate on the distribution of a rabies virus vector. PLoS ONE 13:e0192887. https://doi.org/10.1371/journal.pone.0192887
- Hayes FE, Lecourt P, del Castillo H (2018) Rapid southward and upward range expansion of a tropical songbird, the thrush-like wren (*Campylorhynchus turdinus*), in South America:a consequence of habitat or climate change? Rev Bras Ornitol 26:57–64. https://doi.org/10.1007/bf03544416
- Hernández-Aguilar I, Lorenzo C, Santos-Moreno A, Naranjo EJ, Navarrete-Gutiérrez D (2021) Coronaviruses in bats: a review for the Americas. Viruses 13:1226. https://doi.org/10.3390/v13071226
- Hernández-Pérez EL, Castillo-Vela G, García-Marmolejo G, Sanvicente López M, Reyna-Hurtado R (2019) Wild pig (*Sus scrofa*) as prey of the common vampire bat (*Desmodus rotundus*). Therya 10:195–199. https://doi.org/10.12933/therya-19-685
- Herrera LG, Fleming TH, Sternberg LS (1998) Trophic relationships in a neotropical bat community: a preliminary study using carbon and nitrogen isotopic signatures. Trop Ecol 39:23–29
- Hooper DU, Adair EC, Cardinale BJ, Byrnes JEK, Hungate BA, Matulich KL, Gonzalez A, Duffy JE, Gamfeldt L, O'Connor MI (2012) A global synthesis reveals biodiversity loss as a major driver of ecosystem change. Nature 486:105–108. https://doi.org/ 10.1038/nature11118
- Horta MA, Ledesma LA, Moura WC, Lemos ERS (2022) From dogs to bats:concerns regarding vampire bat-borne rabies in Brazil. Negl Trop Dis 16:e0010160. https://doi.org/10.1371/journal. pntd.0010160
- Hutter SE, Käsbohrer A, González SLF, León B, Brugger K, Baldi M, Mario Romero L, Gao Y, Chaves LF (2018) Assessing changing weather and the El Niño Southern Oscillation impacts on cattle rabies outbreaks and mortality in Costa Rica (1985–2016). BMC Vet Res 14:1–14. https://doi.org/10.1186/s12917-018-1588-8
- Ingala MR, Becker DJ, Holm JB, Kristiansen K, Simmons NB (2019) Habitat fragmentation is associated with dietary shifts and microbiota variability in common vampire bats. Ecol Evol 9:6508–6523. https://doi.org/10.1002/ece3.5228
- Ito M, Arai YT, Itou T, Sakai T, Ito FH, Takasaki T, Kurane I (2001) Genetic characterization and geographic distribution of rabies virus isolates in Brazil: identification of two reservoirs, dogs and vampire bats. Virology 284:214–222. https://doi.org/10. 1006/viro.2000.0916
- Johns PE, Kilgo JC (2005) White-tailed deer. In: Kilgo JC, Blake JI (eds) Ecology and management of a forested landscape. Island Press, Washington, DC, pp 380–389

- Johnson N, Aréchiga-Ceballos N, Aguilar-Setien A (2014) Vampire bat rabies: ecology, epidemiology and control. Viruses 6:1911–1928. https://doi.org/10.3390/v6051911
- Kang Y, Wedekin L (2013) Dynamics of an intraguild predation model with generalist or specialist predator. J Math Biol 67:1227–1259. https://doi.org/10.1007/s00285-012-0584-z
- Klefot JM, Murphy JL, Donohue KD, O'Hara BF, Lhamon ME, Bewley JM (2016) Development of a noninvasive system for monitoring dairy cattle sleep. J Dairy Sci 99:8477–8485. https://doi.org/10.3168/jds.2015-10695
- Kobayashi Y, Sato G, Shoji Y, Sato T, Itou T, Cunha EMS, Samara SI, Carvalho AAB, Nociti DP, Ito FH, Sakai T (2005) Molecular epidemiological analysis of bat rabies viruses in Brazil. J Vet Med Sci 67:647–652. https://doi.org/10.1292/jvms.67.647
- Kunz TH, Braun de Torrez E, Bauer D, Lobova T, Fleming TH (2011) Ecosystem services provided by bats. Ann NY Acad Sci 1223:1–38. https://doi.org/10.1111/j.1749-6632.2011.06004.x
- Lanzagorta-Valencia K, Fernández-Méndez J, Medellín R, Rodas-Martínez A, Avila-Flores R (2020) Landscape and cattle management attributes associated with the incidence of *Desmodus rotundus* attacks on cattle. Ecosist Recurs Agropec 7:1–10. https://doi.org/10.19136/era.a7n1.2164
- LaVal RK (2004) Impact of global warming and locally changing climate on tropical cloud forest bats. J Mamm 85:237–244. https://doi.org/10.1644/bwg-016
- Lebbin DJ, Harvey MG, Lenz TC, Andersen MJ, Ellis JM (2007) Nocturnal migrants foraging at night by artificial light. Wilson J Ornithol 119:506–508. https://doi.org/10.1676/06-139.1
- Lee DN, Papeş M, Van den Bussche RA (2012) Present and potential future distribution of common vampire bats in the Americas and the associated risk to cattle. PLoS One 7:e42466. https://doi.org/10.1371/journal.pone.0042466
- León B, González SF, Solís LM, Ramírez-Cardoce M, Moreira-Soto A, Cordero-Solórzano JM, Hutter SE, González-Barrientos R, Rupprecht CE (2021) Rabies in Costa Rica—next steps towards controlling bat-borne rabies after its elimination in dogs. Yale J Biol Med 94:311. https://doi.org/10.1089/vbz.2015.1906
- Lord RD, de Chaurell AO, Elsy CA (1986) Experimental infection of vampire bats with foot and mouth disease virus. J Wildl Dis 22:413–414. https://doi.org/10.7589/0090-3558-22.3.413
- Loyola RD, Lemes P, Faleiro FV, Trindade-Filho J, Machado RB (2012) Severe loss of suitable climatic conditions for marsupial species in Brazil: challenges and opportunities for conservation. PLoS ONE 7:e46257
- Ma X, Monroe BP, Cleaton JM, Orciari LA, Yager P, Li Y, Kirby JD, Blanton JD, Petersen BW, Wallace RM (2018) Rabies surveillance in the United States during 2016. J Am Vet Med Assoc 252:945–957. https://doi.org/10.2460/javma.252.8.945
- Mandel JT, Bildstein KL (2007) Turkey vultures use anthropogenic thermals to extend their daily activity period. Wilson J Ornithol 119:102–105. https://doi.org/10.1676/05-154.1
- Mantovan KB, Menozzi BD, Paiz LM, Sevá AP, Brandão PE, Langoni H (2022) Geographic distribution of common vampire bat *Desmodus rotundus* (Chiroptera: Phyllostomidae) shelters: implications for the spread of rabies virus to cattle in Southeastern Brazil. Pathogens 118:942. https://doi.org/10.3390/pathogens1 1080942
- May RM, Anderson RM (1990) Parasite—host coevolution. Parasitology 100:S89–S101. https://doi.org/10.1038/npg.els.0001765
- Mayen F (2003) Hematophagous bats in Brazil, their role in rabies transmission, impact on public health, livestock industry and alternatives to an indiscriminate reduction of bat population. J Vet Med 50:469–472. https://doi.org/10.1046/j.1439-0450.2003. 00713.x
- McCracken GF, Bernard RF, Gamba-Rios M, Wolfe R, Krauel JJ, Jones DN, Russell AL, Brown VA (2018) Rapid range expansion of



- the Brazilian free-tailed bat in the southeastern United States, 2008–2016. J Mamm 99:312–320. https://doi.org/10.1093/jmammal/gyx188
- McNab BK (1973) Energetics and the distribution of vampires. J Mammal 54:131–144. https://doi.org/10.2307/1378876
- Megid J, Benavides Tala JA, Belaz Silva LD, Castro Castro FF, Ribeiro BLD, Appolinário CM, Katz ISS, Scheffer KC, Silva SR, Rosa AR (2021) Serological surveillance of rabies in free-range and captive common vampire bats *Desmodus rotundus*. Front Vet Sci 8:967. https://doi.org/10.3389/fyets.2021.681423
- Mello AKM, Brumatti RC, Neves DA, Alcântara LO, Araújo FS, Gaspar AO, Lemos RA (2019) Bovine rabies: economic loss and its mitigation through antirabies vaccination. Pesq Vet Bras 39:179–185. https://doi.org/10.1590/1678-5150-PVB-6201
- Meltzer MI, Rupprecht CE (1998) A review of the economics of the prevention and control of rabies. Part 1: global impact and rabies in humans. Pharmacoeconomics 14:365–383. https://doi.org/10. 2165/00019053-199814040-00004
- Mialhe PJ (2014) Preferential prey selection by *Desmodus rotundus* (E. Geoffroy, 1810, Chiroptera, Phyllostomidae) feeding on domestic herbivores in the municipality of São Pedro. Braz J Biol 74:579–584. https://doi.org/10.1590/bjb.2014.0086
- Michalski F, Boulhosa RLP, Faria A, Peres CA (2006) Human–wildlife conflicts in a fragmented Amazonian forest landscape: determinants of large felid depredation on livestock. Anim Conserv 9:179–188. https://doi.org/10.1111/j.1469-1795.2006.00025.x
- Mistry S, Moreno-Valdez A (2009) COS 85–2: climate change, vampire bats, and rabies: modeling range shifts on the US-Mexico border. In: Conf Proc 94th Ecol Soc Amer
- Murphy A, Diefenbach DR, Ternent M, Lovallo M, Miller D (2021) Threading the needle: How humans influence predator–prey spatiotemporal interactions in a multiple-predator system. J Anim Ecol 90:2377–2390. https://doi.org/10.1111/1365-2656.13548
- Murray M, Cembrowski A, Latham ADM, Lukasik VM, Pruss S, St Clair CC (2015) Greater consumption of protein-poor anthropogenic food by urban relative to rural coyotes increases diet breadth and potential for human–wildlife conflict. Ecography 38:1235–1242. https://doi.org/10.1111/ecog.01128
- Neely BA, Janech MG, Fenton MB, Simmons NB, Bland AM, Becker DJ (2020) Surveying the vampire bat (*Desmodus rotundus*) serum proteome:a resource for identifying immunological proteins and detecting pathogens. J Proteome Res 20:2547–2559. https://doi.org/10.1021/acs.jproteome.0c00995.s007
- Nobre CA, Sampaio G, Borma LS, Castilla-Rubio JC, Silva JS, Cardoso M (2016) Land-use and climate change risks in the Amazon and the need of a novel sustainable development paradigm. Proc Natl Acad Sci USA 113:10759–10768. https://doi.org/10.1073/pnas.1605516113
- Nuñez MN, Solman SA, Cabré MF (2009) Regional climate change experiments over southern South America: climate change scenarios in the late twenty-first century. Clim Dyn 32:1081–1095. https://doi.org/10.1007/s00382-008-0449-8
- Nuñez GB, Becker DJ, Plowright RK (2019) The emergence of vampire bat rabies in Uruguay within a historical context. Epidemiol Infect 147:e180. https://doi.org/10.1017/s0950268819000682
- O'Shea TJ, Cryan PM, Hayman DTS, Plowright RK, Streicker DG (2016) Multiple mortality events in bats: a global review. Mamm Rev 46:175–190. https://doi.org/10.1111/mam.12064
- Oertli EH, Wilson PJ, Hunt PR, Sidwa TJ, Rohde RE (2009) Epidemiology of rabies in skunks in Texas. J Am Vet Med Assoc 234:616–620
- Olival KJ (2016) To cull, or not to cull, bat is the question. EcoHealth 13:6–8. https://doi.org/10.1007/s10393-015-1075-7
- Osland MJ, Stevens PW, Lamont MM, Brusca RC, Hart KM, Waddle JH, Langtimm CA, Williams CM, Keim BD, Terando AJ (2021) Tropicalization of temperate ecosystems in North America: The

- northward range expansion of tropical organisms in response to warming winter temperatures. Glob Change Biol 27:3009–3034. https://doi.org/10.1111/gcb.15563
- Paini DR, Sheppard AW, Cook DC, De Barro PJ, Worner SP, Thomas MB (2016) Global threat to agriculture from invasive species. Proc Natl Acad Sci U S A 113:7575–7579. https://doi.org/10.1073/pnas.1602205113
- Patten MA, Burger JC, Mitrovich M (2019) The intersection of human disturbance and diel activity, with potential consequences on trophic interactions. PLoS ONE 14:e0226418. https://doi.org/ 10.1371/journal.pone.0226418
- Pecl GT, Araújo MB, Bell JD, Blanchard J, Bonebrake TC, Chen I-C, Clark TD, Colwell RK, Danielsen F, Evengård B, Falconi L, Ferrier S, Frusher S, Garcia RA, Griffis RB, Hobday AJ, Janion-Scheepers C, Jarzyna MA, Jennings S, Lenoir J, Linnetved HI, Martin VY, McCormack PC, McDonald J, Mitchell NJ, Mustonen T, Pandolfi JM, Pettorelli N, Popova E, Robinson SA, Scheffers BR, Shaw JD, Sorte CJB, Strugnell JM, Sunday JM, Tuanmu M-N, Vergés A, Villanueva C, Wernberg T, Wapstra E, Williams SE (2017) Biodiversity redistribution under climate change: impacts on ecosystems and human well-being. Science 355:eaai9214. https://doi.org/10.1126/science.aai9214
- Pereira A, Batista C, Bender D, Reis N, Bazílio S (2016) Report on Desmodus rotundus (Chiroptera, Phyllostomidae) feeding on Sus scrofa (Artiodactyla, Suidae) blood. Bol Soc Bras Mastozool 77:151–153
- Pereira FM, de Oliveira AR, Soares-Neto LL, Munhoz L, Fernandes LS, Dos Santos DO, de Carvalho TP, Langoni H, Santos RL (2022) Rabies in a captive lowland tapir (*Tapirus terrestris*). J Comp Path 198:29–32. https://doi.org/10.1016/j.jcpa.2022.07. 008
- Piccinini RS, Peracchi AL, Souza JC, Tannure AM, Raimundo SD, Albuquerque ST, Furtado LL (1998) Métodos de tratamento tópico de bovinos com Warfarina Técnica em pasta a 2% para controle de morcegos hematófagos *Desmodus rotundus*. Rev Bras Med Vet 20:69–73
- Plowright RK, Eby P, Hudson PJ, Smith IL, Westcott D, Bryden WL, Middleton D, Reid PA, McFarlane RA, Martin G, Tabor GM, Skerratt LF, Anderson DL, Crameri G, Quammen D, Jordan D, Freeman P, Wang L-F, Epstein JH, Marsh GA, Kung NY, McCallum H (2015) Ecological dynamics of emerging bat virus spillover. Proc R Soc B 282:20142124. https://doi.org/10.1098/rspb.2014.2124
- Quiroga N, Campos-Soto R, Yañez-Meza A, Rodríguez-San Pedro A, Allendes JL, Bacigalupo A, Botto-Mahan C, Correa JP (2022) Trypanosoma cruzi DNA in Desmodus rotundus (common vampire bat) and Histiotus montanus (small big-eared brown bat) from Chile. Acta Trop 225:106206. https://doi.org/10.1016/j. actatropica.2021.106206
- Raffel TR, Martin LB, Rohr JR (2008) Parasites as predators:unifying natural enemy ecology. Trends Ecol Evol 23:610–618. https://doi.org/10.1016/j.tree.2008.06.015
- Reyna-Hurtado R, Sima-Pantí D, Andrade M, Padilla A, Retana-Guaiscón Ó, Sánchez-Pinzón K, Martínez W, Meyer N, Moreira-Ramírez JF, Carrillo-Reyna N, Rivero-Hernández CM, Serrano Mac-Gregor I, Calme S, Arias Domínguez N (2019) Tapir population patterns under the disappearance of free-standing water. Therya 10:353–358. https://doi.org/10.12933/therya-19-902
- Ribeiro J, Vieira RGV, Martins CM, Ferreira F, Araujo JP, Ullmann LS, Dos Santos AP, Biondo AW (2021) Spatial distribution of bat shelters and livestock rabies in Southern Brazil. Vector Borne Zoonotic Dis 21:785–795. https://doi.org/10.1089/vbz.2020.2730
- Rich C, Longcore T (2013) Ecological consequences of artificial night lighting. Island Press, Washington, DC
- Rocha F, Dias RA (2020) The common vampire bat *Desmodus* rotundus (Chiroptera:Phyllostomidae) and the transmission of



- the rabies virus to livestock: a contact network approach and recommendations for surveillance and control. Prev Vet Med 174:104809. https://doi.org/10.1016/j.prevetmed.2019.104809
- Rocha F, Ulloa-Stanojlovic FM, Rabaquim VCV, Fadil P, Pompei JC, Brandão PE, Dias RA (2020) Relations between topography, feeding sites, and foraging behavior of the vampire bat, *Desmodus rotundus*. J Mamm 101:164–171. https://doi.org/10.1093/jmammal/gyz177
- Rodewald AD, Kearns LJ, Shustack DP (2011) Anthropogenic resource subsidies decouple predator-prey relationships. Ecol Appl 21:936–943. https://doi.org/10.1890/10-0863.1
- Rojas A, Jiménez A, Vargas M, Zumbado M, Herrero MV (2008) Ectoparásitos del vampiro común (*Desmodus rotundus*) en Costa Rica: tasas de parasitismo y tendencias biogeográficas. Mastozool Neotrop 15:181–187
- Rojas-Sereno ZE, Streicker DG, Medina-Rodríguez AT, Benavides JA (2022) Drivers of spatial expansions of vampire bat rabies in Colombia. Viruses 14:2318. https://doi.org/10.3390/v1411 2318
- Rouk CS, Glass BP (1970) Comparative gastric histology of five north and central American bats. J Mamm 51:455–490. https://doi.org/10.2307/1378387
- Ruckebusch Y (1972) The relevance of drowsiness in the circadian cycle of farm animals. Anim Behav 20:637–643. https://doi.org/10.1016/S0003-3472(72)80136-2
- Sabino-Santos G Jr, Maia FG, Martins RB, Gagliardi TB, Souza WM, Muylaert RL, Luna LK, Melo DM, Cardoso RD, Barbosa ND, Pontelli MC (2018) Natural infection of neotropical bats with hantavirus in Brazil. Sci Rep 8:9018. https://doi.org/10.1038/s41598-018-27442-w
- Salazar A, Baldi G, Hirota M, Syktus J, McAlpine C (2015) Land use and land cover change impacts on the regional climate of non-Amazonian South America: a review. Glob Planet Change 128:103–119. https://doi.org/10.1016/j.gloplacha.2015.02.009
- Sánchez-Cordero V, Botello F, Magaña-Cota G, Iglesias J (2011) Vampire bats, *Desmodus rotundus*, feeding on white-tailed deer, *Odocoileus virginianus*. Mammalia 75:91–92. https:// doi.org/10.1515/mamm.2010.065
- Schaefer R, Caldas E, Schmidt E, King A, Roehe P (2002) First case of cat rabies in southern Brazil for 11 years. Vet Rec 150:216–217. https://doi.org/10.1136/vr.150.7.216
- Schmidly DJ, Bradley RD (2016) Order rodentia—rodents. In: Schmidly DJ, Bradley RD (eds) The mammals of Texas, 7th edn. The University of Texas Press, Austin, pp 436–630
- Schmidt KM, Badger DD (1979) Some social and economic aspects in controlling vampire bats. Proc Okla Acad Sci 59:112–114
- Schneider MC, Aron J, Santos-Burgoa C, Uieda W, Ruiz-Velazco S (2001) Common vampire bat attacks on humans in a village of the Amazon region of Brazil. Cad Saude Publ 17:1531–1536. https://doi.org/10.1590/s0102-311x2001000600025
- Schneider MC, Romijn PC, Uieda W, Tamayo H, da Silva DF, Belotto A, da Silva JB, Leanes LF (2009) Rabies transmitted by vampire bats to humans: an emerging zoonotic disease in Latin America? Rev Panam Salud Publ 25:260–269. https://doi.org/10.1590/s1020-49892009000300010
- Schutt B (2008) Dark banquet: blood and the curious lives of bloodfeeding creatures. Three Rivers Press, New York
- Serrano H, Pérez-Rivero J-J, Aguilar-Setién A, de-Paz O, Villa-Godoy A (2007) Vampire bat reproductive control by a naturally occurring phytooestrogen. Reprod Fertil Dev 19:470–472. https://doi.org/10.1071/rd06100
- Sétien AA, Brochier B, Tordo N, De Paz O, Desmettre P, Péharpré D, Pastoret P-P (1998) Experimental rabies infection and oral vaccination in vampire bats (*Desmodus rotundus*). Vaccine 16:1122–1126. https://doi.org/10.1016/s0264-410x(98) 80108-4

- Siemers BM, Schaub A (2011) Hunting at the highway: traffic noise reduces foraging efficiency in acoustic predators. Proc R Soc B 278:1646–1652. https://doi.org/10.1098/rspb.2010.2262
- Simpson SD, Radford AN, Nedelec SL, Ferrari MCO, Chivers DP, McCormick MI, Meekan MG (2016) Anthropogenic noise increases fish mortality by predation. Nat Commun 7:10544. https://doi.org/10.1038/ncomms10544
- Slate D, Algeo TP, Nelson KM, Chipman RB, Donovan D, Blanton JD, Niezgoda M, Rupprecht CE (2009) Oral rabies vaccination in North America: opportunities, complexities, and challenges. Negl Trop Dis 3:e549. https://doi.org/10.1371/journal.pntd. 0000549
- Souza M, Bernardi F, Ito FH (1997) Epidemiology of rabies: biological and serological aspects of rabies in vampire bats *Desmodus rotundus* (E. Geoffroy) captured in Vale do Paraíba, southeastern of Brazil. Arq Inst Biol 64:91–101
- Stafford-Bell R, Scarr M, Scarpaci C (2012) Behavioural responses of the Australian fur seal (Arctocephalus pusillus doriferus) to vessel traffic and presence of swimmers in Port Phillip Bay, Victoria, Australia. Aquat Mamm 3:8. https://doi.org/10.1578/am. 38.3.2012.241
- Sterner RT, Meltzer MI, Shwiff SA, Slate D (2009) Tactics and economics of wildlife oral rabies vaccination, Canada and the United States. Emerg Inf Dis 15:1176–1184
- Stoner-Duncan B, Streicker DG, Tedeschi CM (2014) Vampire bats and rabies: toward an ecological solution to a public health problem. Negl Trop Dis 8:e2867. https://doi.org/10.1371/journal.pntd. 0002867
- Streicker DG, Allgeier JE (2016) Foraging choices of vampire bats in diverse landscapes: potential implications for land-use change and disease transmission. J Appl Ecol 53:1280–1288. https://doi.org/10.1111/1365-2664.12690
- Streicker DG, Recuenco S, Valderrama W, Gomez Benavides J, Vargas I, Pacheco V, Condori Condori RE, Montgomery J, Rupprecht CE, Rohani P (2012) Ecological and anthropogenic drivers of rabies exposure in vampire bats: implications for transmission and control. Proc R Soc B 279:3384–3392. https://doi.org/10.1098/rspb.2012.0538
- Streicker DG, Winternitz JC, Satterfield DA, Condori-Condori RE, Broos A, Tello C, Recuenco S, Velasco-Villa A, Altizer S, Valderrama W (2016) Host–pathogen evolutionary signatures reveal dynamics and future invasions of vampire bat rabies. Proc Natl Acad Sci U S A 113:10926–10931. https://doi.org/10.1073/pnas. 1606587113
- Swank WG, Petrides GA (1954) Establishment and food habits of the nutria in Texas. Ecology 35:172–176
- Tamsitt JR, Fox I (1970) Records of bat ectoparasites from the Caribbean region (Siphonaptera, Acarina, Diptera). Can J Zool 48:1093–1097. https://doi.org/10.1139/z70-193
- Thompson RD, Mitchell GC, Burns RJ (1972) Vampire bat control by systemic treatment of livestock with an anticoagulant. Science 177:806–808. https://doi.org/10.1126/science.177.4051.806
- Thompson RD, Elias DJ, Mitchell GC (1977) Effects of vampire bat control on bovine milk production. J Wildl Manag 41:736–739. https://doi.org/10.2307/3799998
- Torres FD, Valença C, de Andrade Filho GV (2005) First record of *Desmodus rotundus* in urban area from the city of Olinda, Pernambuco, Northeastern Brazil: a case report. Rev Inst Med Trop Sao Paulo 47:107–108. https://doi.org/10.1590/s0036-46652 005000200010
- Turner DC (1975) The vampire bat. The Johns Hopkins University Press. Baltimore
- Uhart M, Milano F (2002) Multiple species production systems: reversing underdevelopment and nonsustainability in Latin America. Ann NY Acad Sci 969:20–23. https://doi.org/10.1111/j.1749-6632.2002.tb04345.x



- Valderrama J, García I, Figueroa G, Rico E, Sanabria J, Rocha N, Parra E, Saad C, Páez A (2006) Outbreaks of human rabies transmitted by vampire bats in Alto Baudó and Bajo Baudó municipalities, department of Chocó, Colombia, 2004–2005. Biomedica 26:387–396
- Valeix M, Hemson G, Loveridge AJ, Mills G, Macdonald DW (2012) Behavioural adjustments of a large carnivore to access secondary prey in a human-dominated landscape. J Appl Ecol 49:73–81. https://doi.org/10.1111/j.1365-2664.2011.02099.x
- Viana M, Benavides JA, Broos A, Ibañez Loayza D, Niño R, Bone J, da Silva FA, Orton R, Valderrama Bazan W, Matthiopoulos J, Streicker DG (2023) Effects of culling vampire bats on the spatial spread and spillover of rabies virus. Sci Adv 9:eadd7437. https:// doi.org/10.1126/sciadv.add7437
- Vieira KS, Guedes Montenegro PF, Santana GG, da Silvia Vieira WL (2018) Effect of climate change on distribution of species of common horned frogs in South America. PLoS ONE 13:e0202813. https://doi.org/10.1371/journal.pone.0202813
- Voigt CC, Kelm DH (2006) Host preference of the common vampire bat (*Desmodus rotundus*; Chiroptera) assessed by stable isotopes. J Mamm 87:1–6. https://doi.org/10.1644/05-mamm-f-276r1.1
- Wang L-F, Shi Z, Zhang S, Field H, Daszak P, Eaton BT (2006) Review of bats and SARS. Emerg Infect Dis 12:1834–1840. https://doi. org/10.3201/eid1212.060401
- Weir AAS, Kacelnik A (2006) A New Caledonian crow (Corvus moneduloides) creatively re-designs tools by bending or unbending aluminum strips. Anim Cogn 9:317. https://doi.org/10.1007/s10071-006-0052-5
- Wenzel RL, Tipton VJ (1966) Ectoparasites of panama. Field Museum of Natural History, Chicago
- Willmott NJ, Wong BBM, Lowe EC, McNamara KB, Jones TM (2022) Wildlife exploitation of anthropogenic change:interactions and consequences. Q Rev Biol 97:15–35. https://doi.org/10.1086/ 718748
- Windberg LA (1998) Population trends and habitat associations of rodents in southern Texas. Am Midl Nat 140:153–160. https:// doi.org/10.1674/0003-0031(1998)140[0153:PTAHAO]2.0.CO;2
- Winkler WG, Schneider NJ, Jennings WL (1972) Experimental rabies infection in wild rodents. J Wildl Dis 8:99–103. https://doi.org/ 10.7589/0090-3558-8.1.99
- Wohlers A, Lankau EW, Oertli EH, Maki J (2018) Challenges to controlling rabies in skunk populations using oral rabies vaccination:

- a review. Zoonoses Public Health 65:373–385. https://doi.org/10.1111/zph.12471
- Wolfe AK, Bateman PW, Fleming PA (2018) Does urbanization influence the diet of a large snake? Curr Zool 64:311–318. https://doi.org/10.1093/cz/zox039
- Worsley-Tonks KEL, Escobar LE, Biek R, Castaneda-Guzman M, Craft ME, Streicker DG, White LA, Fountain-Jones NM (2020) Using host traits to predict reservoir host species of rabies virus. Negl Trop Dis 14:e0008940. https://doi.org/10.1371/journal.pntd. 0008940
- Wray AK, Olival KJ, Morán D, Lopez MR, Alvarez D, Navarrete-Macias I, Liang E, Simmons NB, Lipkin WI, Daszak P, Anthony SJ (2016) Viral diversity, prey preference, and *Bartonella* prevalence in *Desmodus rotundus* in Guatemala. EcoHealth 13:761–774. https://doi.org/10.1007/s10393-016-1183-z
- Yob JM, Field H, Rashdi AM, Morrissy C, van der Heide B, Rota P, bin Adzhar A, White J, Daniels P, Jamaluddin A (2001) Nipah virus infection in bats (order Chiroptera) in peninsular Malaysia. Emerg Infect Dis 7:439. https://doi.org/10.3201/eid0703.017312
- Zarza H, Martínez-Meyer E, Suzán G, Ceballos G, Zarza H, Martínez-Meyer E, Suzán G, Ceballos G (2017) Geographic distribution of Desmodus rotundus in Mexico under current and future climate change scenarios: implications for bovine paralytic rabies infection. Vet Mex 4:10–25. https://doi.org/10.21753/ymoa.4.3.390
- Zetun CB, Hoffmann JL, Silva RC, Souza LC, Langoni H (2009) *Leptospira* spp. and *Toxoplasma gondii* antibodies in vampire bats (*Desmodus rotundus*) in Botucatu region SP, Brazil. J Venom Anim Toxins Incl Trop Dis 15:546–552. https://doi.org/10.1590/s1678-91992009000300014

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