

Research paper

Modeling of cell distribution dynamics in cell-laden bioink with active circulation

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ABSTRACT

Three-dimensional bioprinting is recognized as the future of constructing bio-functional tissues and organs to satisfy the increasing demand of tissue regeneration and organ transplantation. During depositing the bioink consisting of biological materials and living cells, cells sediment due to dominant gravitational force, resulting in cell aggregation through cell-cell interaction at the bioink reservoir bottom. It significantly undermines the printing performance in terms of droplet formation and post-printing cell distribution, therefore being considered as a critical problem in 3D bioprinting. The previous study has demonstrated the feasibility of circulating bioink to mitigate sedimentation and associated cell aggregation with potentially superior effectiveness. This study focused on construction of sedimentation model to predict the dynamic performance of bioink reservoir in cell distribution considering the influence of cell aggregation in stationary bioink and in circulated bioink. The iterative and time-region-based sedimentation model has been provided, and the influence of cell aggregation has been quantified through least-squares optimization by Nelder-Mead simplex algorithm. The role of cell aggregation in sedimentation of cell-laden bioink during inkjet-based bioprinting has been investigated theoretically and experimentally. It has been discovered that (1) sedimentation model considering single cell underestimates the experimental quantifications of cell concentrations in stationary fluid and overestimates the pump capacity in circulated bioink, while sedimentation model considering cell aggregation generally agrees with both; (2) the formed cell aggregation expedites the cell sedimentation due to the larger size-induced higher sedimentation velocity in stationary bioink; (3) the formed cell aggregation reduces the pump capacity and mitigation effectiveness to cell aggregation in circulated bioink. Applying higher initial cell concentration from 1×10^6 to 3×10^6 to 5×10^6 cells/ml, lower polymer concentration from 1% to 0.5 to 0.25% (w/v) sodium alginate, and longer printing time from 30 to 60 to 90 minutes increase the chance of forming cell aggregates due to greater sedimentation-induced accumulation of cells and shorter distance-induced greater cell-cell interaction.

1. Introduction

Recently, great advancement has been made in three-dimensional (3D) bioprinting attributed to the scientific and technological progress in biology, manufacturing, and materials [1]. As a computer-aided manufacturing technology, 3D bioprinting precisely positions picolitre biologically relevant materials (with/without living cells) to fabricate functional biomimetic structures layer by layer in a predefined layout, particularly benefiting tissue engineering and regenerative medicine [2, 3]. Based on printing mechanism, it is generally classified into inkjet-based [4,5], extrusion-based [6,7], laser-based [8], and stereolithography-based [9,10] bioprinting. Inkjet-based bioprinting demonstrates its unique superiority in direct cell printing due to suitable

nozzle size and printing resolution for most living cells (cell size of 10–30 μm), high post-printing cell viability ($>90\%$), good precision on droplet size, freeform cell-laden biostructure fabrication, non-contact delivery, and easy scaleup [11–14]. Based upon droplet formation process, inkjet-based printing is categorized into two main types, namely continuous inkjet (CIJ) printing and drop-on-demand inkjet (DOD) printing. Driven by pneumatic pressure, CIJ printing discharges a continuous liquid jet, which breaks up into droplets due to Rayleigh instability [15]. Driven by thermal expansion [16] or piezoelectric actuation [17], DOD printing produces picolitre droplets of bioink when required. Piezoelectric DOD printing is selected in this study as it has been broadly used in 3D bioprinting [18,19]. The bioink, being used in 3D bioprinting, is a cell suspension containing solution of biological

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materials and living cells. Biological materials usually have attributes such as good biocompatibility and biodegradability, shear-thinning property, and other favorable rheological and mechanical properties [20]. Natural polymer (e.g., alginate [21], collagen [22] and fibrin [23]) and synthetic polymer (e.g., gelatin methacrylate [24] and polyethylene glycol [25]) are two commonly applied biological materials in 3D bioprinting to imitate natural extracellular matrix (ECM) [23]. Model cells have a variety of choices, such as fibroblasts [26], stem cells [27], cancer cells [28]. To date, various 3D artificial tissues have been fabricated by inkjet-based bioprinting, such as cardiac tissue [29], skin [30], cartilage [31] and vascular-like structure [32].

3D bio-printed functional tissues/organs require the printing process to be precise and reliable. However, as the technical hurdles to achieve precise and reliable inkjet bioprinting, cell sedimentation and the resultant cell aggregation have recently drawn extensive attentions [26] as they are widely reported to undermine the printing performance [4, 26]. As documented by Saunders et al. [18], the printing efficiency was largely reduced only after 20-minute inkjet printing due to nonuniformity of the bioink by cell sedimentation. Lee et al. [33] reported that the inkjet nozzle had a high risk of being clogged as aggregation accumulated over time, resulting in the hindrance of continuous jetting and other non-ideal jetting behaviors during printing. Chahafi et al. [28] also demonstrated that the actual number of cells per well (NCPW) was 1859, while the expected NCPW was 975 in post-printing of bioink containing 0.5×10^6 MCF-breast cancer cells/ml. Frequency of nozzle failure was reported to be 23% with average time to clogging of 8 min. Bioink with a certain critical cell concentration has spaces between cells, so the force of cell interaction is minimal. However, gravity causes cells to settle, leading to increased local cell concentration and decreased space between cells. At a critical point, such as $4 \mu\text{m}$ for erythrocytes [34], cell adhesion dominates, forming cell aggregates. Cell adhesion proteins mediate cell aggregation and are classified into families based on structure and function. The cadherin family forms adherens junctions between cells by homophilic interaction with adjacent cadherins (transmembrane proteins) in a calcium-dependent manner [35]. The integrin family mediates cell adhesion with extracellular matrix proteins and adjacent cells through heterodimeric transmembrane proteins [36].

There have been several studies focusing on quantification of cell sedimentation and cell aggregation during inkjet bioprinting. Xu et al. [37] conducted a force analysis on a single mouse fibroblast within the bioink containing 0.5–2% (w/v) sodium alginate (NaAlg) by estimating three major forces, namely gravitational force, buoyant force, and drag force. Cell sedimentation velocity was reported to range from 0 to $1.5 \mu\text{m/s}$ and to be inversely related to the polymer concentration. At 0.5% (w/v) NaAlg, the local cell concentration at the bottom of bioink reservoir was 4 times greater after 2-hour printing with prominent cell aggregation observed. Later, Xu et al. [5] quantified the local top, middle and bottom cell concentrations within bioink reservoir and the percentage of the cells forming cell aggregation due to cell sedimentation. It was reported that 60.9% of the cells at the bioink reservoir bottom formed aggregates at 0.5% (w/v) NaAlg and 2×10^6 fibroblasts/ml at 40-minute printing time. Pepper et al. [27] proposed a Stokes' Law-based mathematical model of cell concentration at the bottom of the bioink reservoir to predict cell output when applying the bioink containing D1 murine stem cells for inkjet printing. Output for cells linearly increased and then decreased owing to sedimentation with associated cell aggregation and cartridge surface. Xu et al. [4] also quantified the cell sedimentation-induced cell aggregation within the nozzle and the post-printing microspheres during and after inkjet printing of the bioink with 0.5% (w/v) NaAlg and 1.5×10^6 mouse fibroblasts/ml. After only 15-minute printing, 80% of the cells formed cell aggregates within the nozzle. The mean cell number per microsphere increased significantly from 0.38 to 1.05 with the maximum number of encapsulated cells being 10.

There also have been several studies focusing on mitigating cell sedimentation and cell aggregation during inkjet bioprinting. These

approaches fall into three main categories [26]: achieving neutral buoyancy, active stirring, and bioink circulation. For the first category, Xu et al. [37] reported increasing the polymer concentration of NaAlg from 0.5% to 1.5% (w/v) significantly retarded the mouse fibroblasts sedimentation velocity from $1.5 \mu\text{m/s}$ to $0.3 \mu\text{m/s}$ due to increased buoyant force provided by the bioink. Fewer cell aggregates were therefore observed. Chahafi et al. [28] formulated the bioink incorporating MCF breast cancer cells with the addition of Ficoll PM400 at 10–15% (w/v) concentration to achieve the nearly neutral buoyancy. When adding 10% (w/v) Ficoll PM400, the number of actual cell output was close to the expected one with little cell sedimentation. For the second category, Parsa et al. [38] applied a magnetic stirrer to agitate the bioink containing Hep G2 hepatoma cells during inkjet bioprinting. By intermittent stirring, the cell sediments at bioink reservoir bottom were dispersed locally but not re-floated evenly across the whole bioink. Furthermore, the cell viability decreased from 99% to 75% after 50-minute stirring. For the third category, Liu et al. [39] introduced active bioink circulation to assist inkjet printing of bioink incorporating 0.5% (w/v) NaAlg and 1×10^6 mouse fibroblasts/ml. It demonstrated the feasibility of bioink circulation in suppressing cell sedimentation and aggregation. The cell sedimentation was mitigated up to 95% and the percentage of individual cells was improved from 32.83% to 86.79% compared to the non-circulation control. Almost negligible effect on post-printing cell viability was reported when introducing active bioink circulation. The comparisons of these mitigation approaches are present in Table 1.

Despite of being validated, the mitigation approaches of the first and second categories hold limited universality in accommodation of multiple cells types and holistic mitigation, respectively. Our previously proposed bioink circulation overcomes these limitations and has been demonstrated feasible in suppressing cell sedimentation and aggregation with potentially high effectiveness [36]. This study focuses on construction of sedimentation model to predict the dynamic performance of bioink reservoir in cell distribution considering the influence of cell aggregation in stationary bioink and in circulated bioink. The role of cell aggregation in sedimentation of cell-laden bioink during inkjet-based bioprinting is investigated theoretically and experimentally. This study facilitates the prediction of bioink reservoir dynamic performance in cell distribution over printing and prediction of effectiveness of circulating bioink to mitigate sedimentation and associated cell aggregation, therefore promoting precise and reliable bioprinting.

Table 1
Comparison of cell aggregation mitigation approaches.

Mitigation approach	Mechanism	Pros/Cons
Ficoll PM400 [28]	Neutral buoyancy	Pros: No further modification of setup No shear stress introduced
Bovine Serum Albumin [40]		Careful formulation required
Polymer concentration [37]		Cons: Rheological modification of bioink Incompatibility with multiple kinds of cell No rheological modification of bioink
Magnetic-driven stir bar [38]	Active stirring	Pros: Compatibility with multiple kinds of cell
Axial flow impeller [41]		Further modification of setup
Cylindrical Nd magnet [41]		Cons: High shear stress Local agitation No rheological modification of bioink
Bioink circulation [39] (This study)	Circulation	Pros: Compatibility with multiple kinds of cell High cell viability Holistic circulation
		Cons: Further modification of setup

This paper is organized as: Section 2 presents the experimental materials, setup and conditions, indicators to characterize the mitigation effectiveness to cell sedimentation and aggregation, and sedimentation model with stationary and circulated bioink. Section 3 demonstrates the expedited sedimentation due to cell aggregation in stationary bioink. Section 4 presents the influence of cell aggregation on mitigation effectiveness to cell sedimentation and aggregation when using cell-laden bioink circulation. Section 5 discusses the effect of bioink volume, optimization, circulation outlet and cell viability. Section 6 draws the major conclusions and proposes the future research topic.

2. Materials and methods

2.1. Bioink preparation

This study applies bioink consisting of solution of biological materials and living cells. NaAlg was selected to imitate the ECM due to its favorable biocompatibility, biodegradability, hydrophilicity, and its economical-friendliness [42]. NIH 3T3 mouse fibroblast (ATCC, Rockville, MD) was selected for the model cell since it is extensively found in mammalian connective tissues [20,43]. Fibroblasts were cultured in an incubator at 37 °C with a humidity of 5% CO₂. Culturing fibroblasts required culture medium, which incorporates Dulbecco's Modified Eagles Medium (DMEM; Sigma-Aldrich, St. Louis, MO) supplemented with 10% Bovine Calf Serum (BCS; Hyclone, Manassas, VA) and 1% antibiotic/antimycotic solution (Corning, Manassas, VA). Changing culture medium every other day was required. Making 0–1% (w/v) NaAlg solution required NaAlg powder (Sigma-Aldrich) and incubation for five minutes. Cell suspension was then centrifuged in room temperature at speed of 1000 rpm for five minutes. The resulting cell pellet was dispersed in NaAlg solution, preparing final bioink with $1\text{--}5 \times 10^6$ cells/ml [44].

2.2. Experimental setup and conditions

A schematic of the inkjet-based bioprinting without and with bioink circulation is shown in Fig. 1, where six major parts are as follows: (1) a bioink reservoir (Microfab, Plano, TX) or a modified bioink reservoir associated with active circulation using an external peristaltic pump, (2) a pneumatic backpressure controller, (3) an inkjet dispenser with an orifice diameter of 120 µm (MicroFab, Plano, TX), (4) a waveform

generator to power the inkjet dispenser, (5) a highly resolved imaging module for the observation of droplet generation, and (6) a substrate holder to receive droplets to be crosslinked with solution of calcium chloride at 2% (w/v). The modified bioink reservoir associated with setup of circulation consists of a bioink reservoir in volume capacity of 1.5 ml, a silicone and bio-compatible tubing in length of 35 cm and inner diameter of 0.5 mm for connection of the bioink reservoir between the top end and bottom end, and a peristaltic pump to adjust flowrates. The 10-roller rotor-based peristaltic pump was utilized to replenish the top end with the bioink in the bottom end [39].

This study investigates the role of cell aggregation in affecting sedimentation of stationary and circulated cell-laden bioink within the bioink reservoir during inkjet-based bioprinting theoretically and experimentally. Table 2 summarizes the major operating conditions. 0.25%, 0.5% and 1% (w/v) NaAlg were selected for the polymer concentrations. 1×10^6 , 3×10^6 and 5×10^6 cells/ml were selected for the cell concentrations. 30, 60 and 90 min were selected for the printing time. The fixed bioink volume was 1 ml. The flowrate for circulating bioink was selected from 0 to 0.5 ml/min. The fixed excitation waveform parameters for processing were as follows: 20–40 V (excitation voltage), 3 µs (rise time), 25 µs (dwell time), 5 µs (fall time), 30 µs (echo time), and 3 µs (final rise time). Experiments were repeated for six times.

2.3. Indicators of mitigation effectiveness to cell sedimentation and cell aggregation

The active circulation was implemented into the bioink reservoir. At the printing time of 0, 30, 60, and 90 min, 10 µl bioink was sampled at the top and bottom of the bioink reservoir. A hemocytometer (Hausser Scientific, Horsham, PA) was utilized to measure the cell concentration. This study introduces pump capacity (C_p) as an indicator to show the mitigation effectiveness to cell sedimentation. It is formulated as:

Table 2

Major operational conditions.

Operating conditions	Unit	Value
Initial cell concentration	cells/ml	1×10^6 , 3×10^6 , 5×10^6
NaAlg concentration	w/v	0.25%, 0.5%, 1%
Printing time	minutes	30, 60, 90
Circulation flowrate	ml/min	0, 0.05, 0.1, 0.3, 0.5

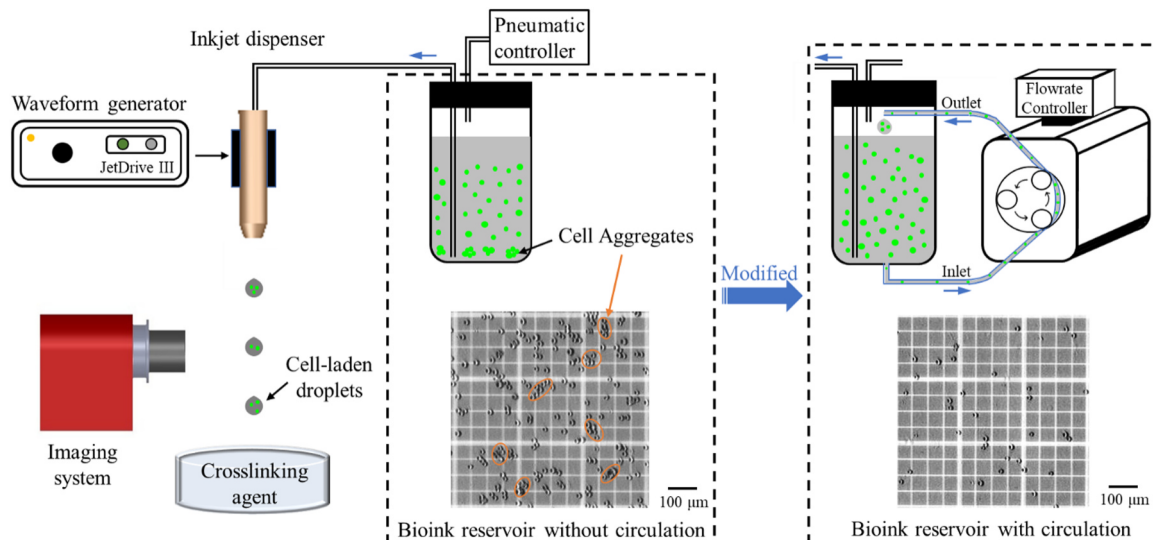


Fig. 1. A Schematic of inkjet-based bioprinting without and with setup of bioink circulation.

$$C_p = \frac{C_{top}}{C_{bot}} \quad (1)$$

where C_{top} denotes the top cell concentration of the bioink reservoir, and C_{bot} denotes the bottom cell concentration of the bioink reservoir. C_p ranges from 0 to 1. A higher C_p indicates a greater mitigation effectiveness on cell sedimentation. $C_p = 1$ indicates that pump could fully recover the cell sedimentation-resulting nonuniform cell distribution within the bioink reservoir. $C_p = 0$ indicates that pump could not recover any cell sedimentation-resulting nonuniform cell distribution.

The percentage of individual cells ($ic\%$), the percentage of small cell aggregates ($sa\%$) involving 2–4 cells, and the percentage of large cell aggregates ($la\%$) involving 5 or more cells are characterized, respectively, as: $ic\% = \frac{\sum_{c=1}^{ab} \text{Total number of cells}}{\sum_{c=1}^{ab} \text{Total number of cells}} \times 100\%$, $sa\% = \frac{\sum_{c=2}^4 \text{Total number of cells}}{\sum_{c=1}^{ab} \text{Total number of cells}} \times 100\%$, and $la\% = \frac{\sum_{c=5}^{ab} \text{Total number of cells}}{\sum_{c=1}^{ab} \text{Total number of cells}} \times 100\%$, where a : the number of cells within the individual cells or the cell aggregates, b : the corresponding

occurrence frequency of the individual cells or cell aggregates, and c : the greatest number of cells constituting the largest cell aggregates.

The mitigation effectiveness to cell aggregation ($E_a\%$) at bioink reservoir bottom is quantified in percentage, which is formulated as:

$$E_a\% = \frac{ic_{wc}\% - ic_{nc}\%}{ic_{control}\% - ic_{nc}\%} \quad (2)$$

where, $ic_{wc}\%$: the percentage of individual cells with active circulation at the printing time t , $ic_{nc}\%$: the percentage of individual cells with no active circulation at the same printing time t , and $ic_{control}\%$: the percentage of individual cells in the control case initially. $E_a\%$ ranges from 0% to 100%. A higher E_a indicates a greater mitigation effectiveness to cell aggregation. $E_a = 100\%$ indicates that pump could fully recover the cell sedimentation-resulting cell aggregation at bioink reservoir bottom. $E_a = 0\%$ indicates that pump could not recover any cell sedimentation-resulting cell aggregation.

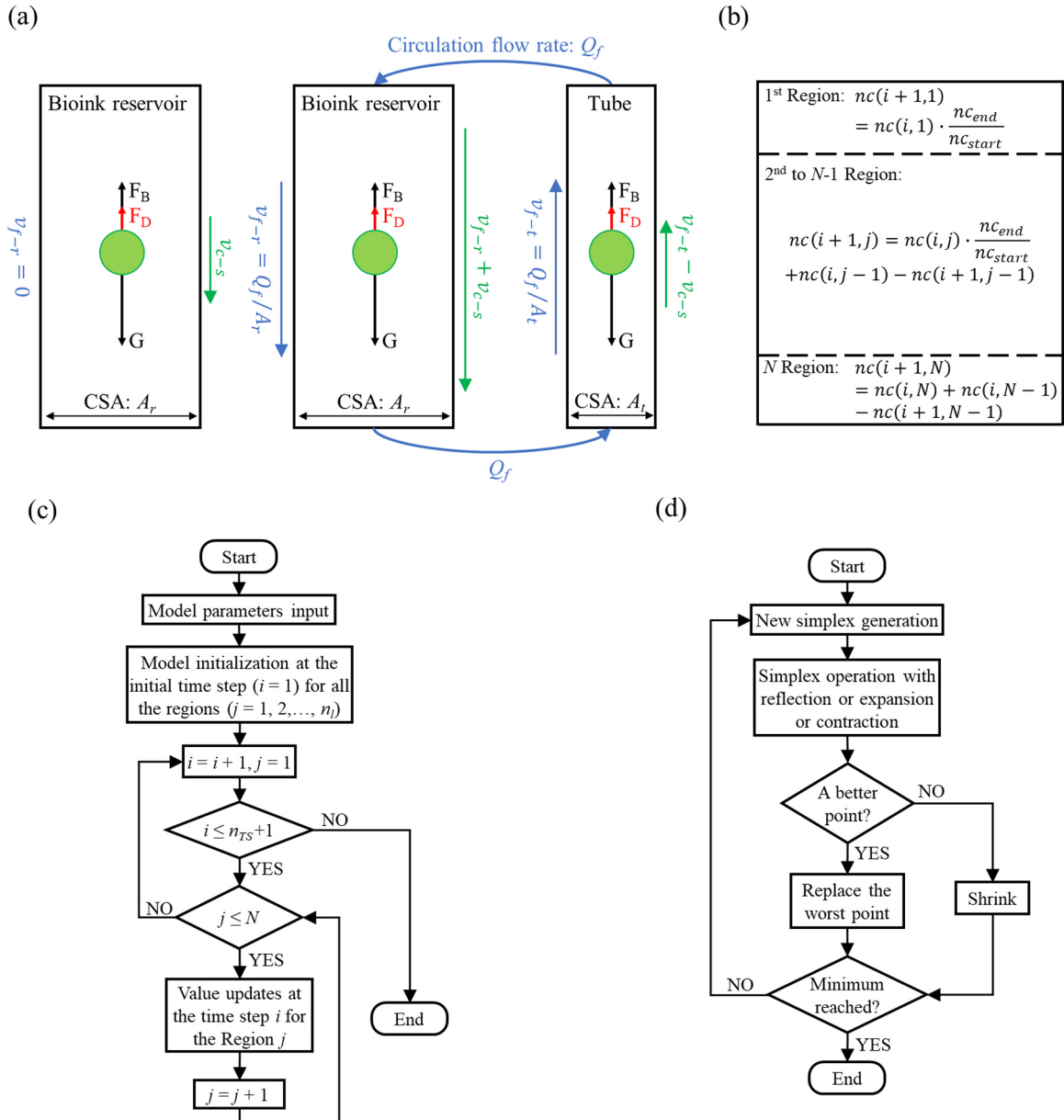


Fig. 2. Cell sedimentation model (a) force and velocity analysis at stationary and circulated bioink (CSA denotes the cross-section area); (b) divided region-based governing equations; (c) flowchart for time-region iteration algorithm; (d) flowchart for Nelder-Mead simplex algorithm.

2.4. Statistical Analysis

Data is represented as mean $\pm$ standard deviation. One-way and two-way analysis of variance (ANOVA) test are performed to examine the significance of the operational conditions on cell distribution and mitigation effectiveness with $P < 0.05$ (*) being statistically significant.

2.5. Sedimentation model of cell-laden bioink

Three major forces—cell gravitational force ($G = \rho_c V_c g$, where ρ_c : cell density, g : gravitational acceleration, and V_c : volume of the cell), buoyant force ($F_B = \rho_f V_c g$, where ρ_f : fluid density), and drag force (F_D) play important roles in cell sedimentation phenomenon [45]. In stationary bioink, suspended cells sediment to the bioink reservoir bottom since $G > F_B$ at initial. Then F_D , defined as the resistance force due to the relative motion of cells against the surrounding fluid, is introduced and estimated by $F_D = \frac{1}{2} \rho_f v_c^2 C_d A$, where v_c : velocity of cells in motion relative to the surrounding fluid, C_d : drag coefficient, and A : cross-sectional area of the cells. In incompressible fluid, C_d is related to the Reynolds number Re , defined as $Re = \frac{\rho_f v_c D}{\mu}$, where D : cell diameter, and μ : fluid dynamic viscosity [46]. v_c is considered to influence the drag force as the ρ_f , A , D , and μ are assumed constant during the sedimentation process. Force equilibrium is then reached where $G = F_B + F_D$. In circulated bioink, cell sedimentation mainly occurs in the bioink reservoir due to the comparability between fluid velocity in bioink reservoir (v_r) and v_c [39]. Same force equilibrium is reached where $G = F_B + F_D$ shown in Fig. 2(a). Bioink circulation mitigates the cell sedimentation through increasing the space between neighboring cells. The cell adhesion through cell-cell interaction can be significantly reduced and cell aggregation is suppressed. Sedimentation model with stationary and circulated bioink is based upon individual cells without aggregation. $v_{f,r}$ is expressed as:

$$v_{f,r} = \begin{cases} 0, & \text{for stationary fluid } Q_f = 0, \\ \frac{Q_f}{A_r}, & \text{for circulated fluid } Q_f > 0, \end{cases} \quad (3)$$

where Q_f : the applied circulation flowrate, and A_r : the area of bioink reservoir in cross-section.

The cell velocity within bioink reservoir is calculated by:

$$v_{c,r} = v_{f,r} + v_{c,s} \quad (4)$$

where $v_{c,r}$: cell velocity in the bioink reservoir, $v_{c,s}$: cell sedimentation velocity at stationary fluid. Assuming minimal time and space, the number of cells at time period end over that at the start can be calculated by:

$$\frac{nc_{end}}{nc_{start}} = \begin{cases} 1, & \text{for } v_{f,r} = 0, \\ \frac{C \bullet A_r \bullet v_{f,r} \bullet t}{C \bullet A_r \bullet v_{f,r} \bullet t + C \bullet A_r \bullet v_{c,s} \bullet t} = \frac{v_{f,r}}{v_{f,r} + v_{c,s}}, & \text{for } v_{f,r} > 0, \end{cases} \quad (5)$$

where nc_{end} and nc_{start} : the respective number of cells at the end of the time step and that at the start, C : local concentration of cells in this space, and t : time step. The bioink bulk height is H , which is divided into N regions. Each region is H/N in height. t is calculated as $t = \frac{H/N}{v_{c,s}}$. Sedimentation model requires iterative computation of the number of cells of i timesteps for each j regional division. Fig. 2(b) shows governing equations of the number of cells for each divided region in the timestep iteration. Calculation follows the conservation of number of cells within the bioink reservoir. The 0.15 ml bioink at the top and the bottom is considered for quantification of local cell concentration. The top cell concentrations and the bottom cell concentration are calculated, respectively, using:

$$C_{top} = \frac{\sum_{j=1}^{5N} nc(n_{TS}, j)}{0.15 \bullet A_r \bullet H}, \quad (6)$$

$$C_{bot} = \frac{\sum_{j=0.85N}^N nc(n_{TS}, j)}{0.15 \bullet A_r \bullet H}, \quad (7)$$

where n_{TS} : time step number, $nc(n_{TS}, j)$: the number of cells for the region j and the time step n_{TS} . The algorithm flowchart is shown in Fig. 2(c). The major parameters for sedimentation model are tabulated in Table 3.

2.6. Model considering cell aggregation

Cell aggregation formed during cell sedimentation is dependent on four experimental conditions: initial cell concentration (C_i), polymer concentration-based cell sedimentation velocity in stationary fluid ($v_{c,s}$), printing time (t), and the system circulation flowrate (Q_f). Cell aggregation effect on local cell concentration (C_a) is characterized by the power-law relationship:

$$C_a = a C_i^b \bullet v_{c,s}^c \bullet t^d \bullet Q_f^e, \quad (8)$$

where a , b , c , d , and e are all positive. Noticeably, Q_f is reversely proportional to the C_a , since the large Q_f suppresses the cell aggregation. Therefore, the local top cell concentration and bottom cell concentration in the bioink reservoir become, respectively, as:

$$C'_{top} = C_{top} - C_{a,top}, \quad (9)$$

$$C'_{bot} = C_{bot} + C_{a,bot}. \quad (10)$$

In this study, the least-squares optimization by the Nelder-Mead simplex algorithm (NMSA) is leveraged [47]. Experimental dataset includes m points of (x_k, y_k) , where $k = 1, 2, \dots, m$. The residuals r_k of the dependent variable y_k from the observation and the independent variables x_k of experimental conditions in n dimensions is expressed as:

$$r_k = y_k - f(x_k), \quad (11)$$

where f : the predictive function in n dimensions shown in Eq. 8 (power-law relationship in four dimensions). y_k herein denotes the difference of actual observation between the experiment and the single cell-based sedimentation model. The sum of the squared residuals S is expressed as:

$$S = \sum_{k=1}^m (r_k)^2. \quad (12)$$

The NMSA is a numerical direct search method to find the minimum/maximum of a multidimensional function [47]. In this study, the sum of the squared residuals S is the objective function $u(v)$ to be minimized by the NMSA:

$$\min u(v), \quad (13)$$

where the algorithm utilizes the geometric simplex with $n + 1$ vertices for n dimensional vectors v . The algorithm sequences the points of vertices $\{v_i\}_{i=1}^{n+1}$ in the simplices from the lowest to the highest at each

Table 3

Major parameters for sedimentation model.

Parameter	Unit	Value
Diameter of bioink reservoir	mm	8
Bioink height, H	mm	19.9
Circulation flowrate, Q_f	ml/min	0, 0.05, 0.1, 0.3, 0.5
Relative cell sedimentation velocity, $v_{c,s}$	$\mu\text{m/s}$	0.65, 1.45, 1.97
Number of regions, N		10,000
Printing time	s	1800, 3600, 5400
Time step, t	s	1.37

iteration:

$$u(v_1) \leq u(v_2) \leq \dots \leq u(v_{n+1}). \quad (14)$$

The worst vertex v_{n+1} is replaced with another better vertex to update the simplex. The algorithm contains four operations, namely reflection (α), expansion (β), contraction (γ) and shrink (δ) to spot the next better vertex with the defaulted scalar parameters being $\alpha = 1$, $\beta = 2$, $\gamma = 1/2$, and $\delta = 1/2$. The centroid of the n better vertices is calculated by:

$$\bar{v} = \frac{1}{n} \sum_{i=1}^n v_i. \quad (15)$$

The algorithm flowchart is shown in Fig. 3(d). For one iteration of

NMSA:

1. Sequence. Evaluate and order the objective function u at $n+1$ vertices in Eq. 14.
2. Reflection. Calculate the reflective point $v_r = v + \alpha(v - v_{n+1})$ and evaluate $u(v_r)$ with $u(v_p)$. If $u(v_1) \leq u(v_r) < u(v_{n+1})$, update v_{n+1} with v_r . If $u(v_r) < u(v_1)$, proceed to Step 3. If $u(v_r) \leq u(v_p) < u(v_{n+1})$, proceed to Step 4. If $u(v_r) \geq u(v_{n+1})$, proceed to Step 5.
3. Expansion. Calculate the expanded point $v_e = v + \beta(v_r - v)$. Evaluate $u(v_e)$ with $u(v_r)$. If $u(v_e) < u(v_r)$, update v_{n+1} with v_e . Otherwise, update v_{n+1} with v_r .
4. Contraction Outside. Calculate the contraction point at outside $v_{co} = v + \gamma(v_r - v)$. Evaluate $u(v_{co})$ with $u(v_p)$. If $u(v_{co}) \leq u(v_p)$, update v_{n+1} with v_{co} . Otherwise, proceed to Step 6.

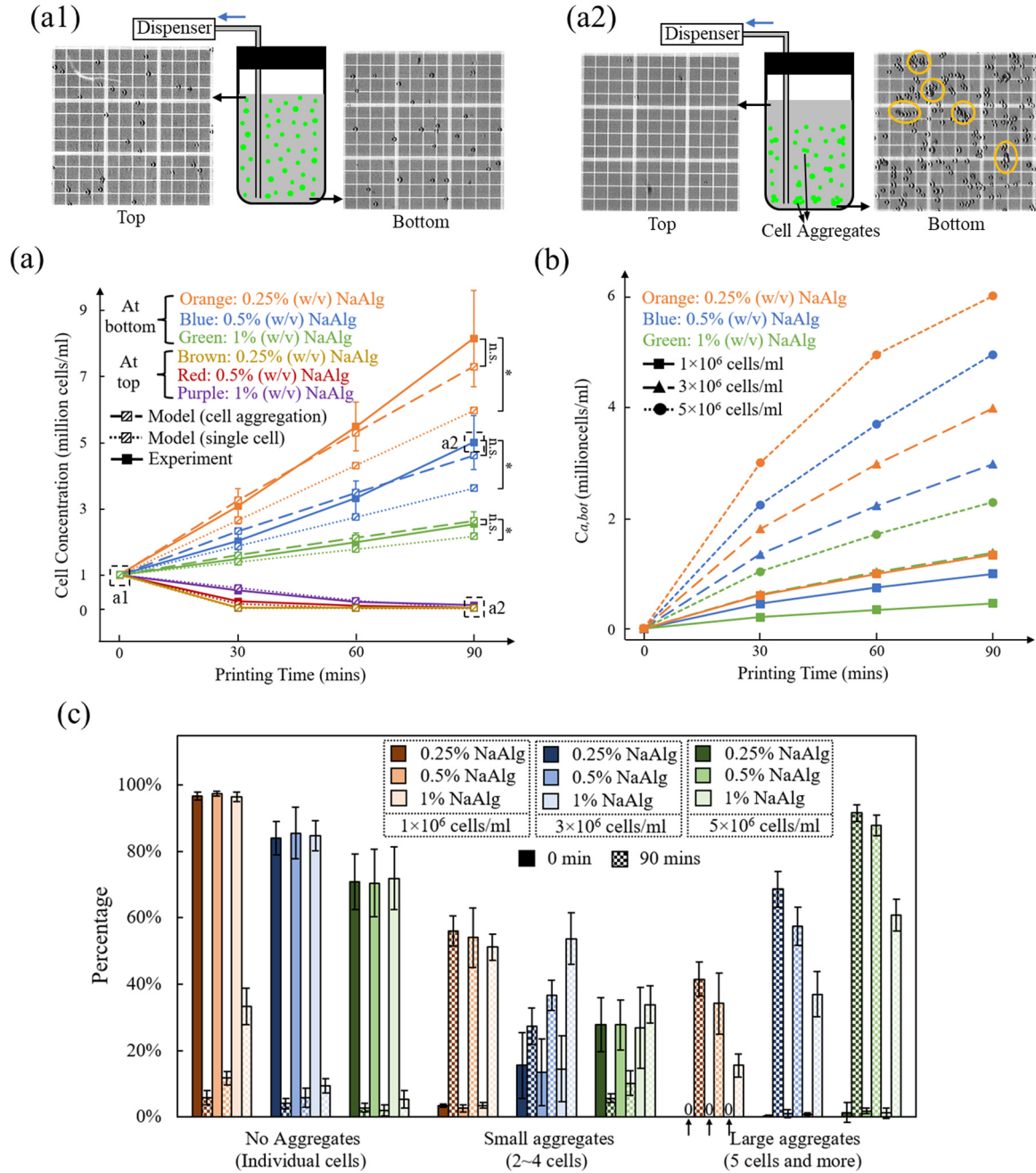


Fig. 3. Cell sedimentation and cell aggregation at stationary fluid. (a) cell sedimentation at 1×10^6 cells/ml over 90-min printing; (b) $C_{a,bot}$ at different cell concentration, different polymer concentration, and different printing time; (c) individual cell and cell aggregation in percentage; (a1) schematic of bioink reservoir at 0 min; (a2) schematic of bioink reservoir at 90 min.

5. Contraction finside . Compute the contraction point at $\text{finside } v_{ci} = v_{ci}^* - v_{ci}$. Evaluate $u(v_{ci})$ with $u(v_{n+1})$. If $u(v_{ci}) < u(v_{n+1})$, update v_{n+1} with v_{ci} . Otherwise, proceed to Step 6.
6. Shrink. For $2 \leq l \leq n+1$, redefine the $v_l = v_l + \delta(v_l - v_1)$.

3. Influence of cell aggregation on sedimentation of stationary bioink

The role of cell aggregation in affecting cell sedimentation process within stationary bioink during inkjet-based bioprinting is studied. Specifically, the experimental quantifications to local cell concentrations, that predicted by sedimentation model considering single cell assumption, and that predicted by sedimentation model considering cell aggregation are studied and compared. Top and bottom cell concentration and bottom cell aggregation at bioink reservoir are quantified to demonstrate the homogeneity of bioink.

Fig. 3(a) shows the local top cell concentration and bottom cell concentration within the bioink reservoir at initial 1×10^6 cells/ml over 90-minute printing. Two respective schematics of bioink reservoir at 0 and 90-minute are shown in Fig. 3(a1) and Fig. 3(a2). With printing time increasing from 0 to 30–60 to 90 min, the observed top cell concentration decreases from 1.00 to 0.54–0.20 to 0.04×10^6 cells/ml, from 1.00 to 0.20–0.03 to 0.01×10^6 cells/ml, and from 1.01 to 0.02–0.01 to 0.01×10^6 cells/ml at 1%, 0.5%, and 0.25% (w/v) NaAlg, respectively. In contrast, those predicted by sedimentation model considering single cell assumption are from 1.00 to 0.60–0.22 to 0.00×10^6 cells/ml, from 1.00 to 0.14–0.00 to 0.00×10^6 cells/ml, and from 1.00 to 0.00–0.00 to 0.00×10^6 cells/ml, respectively. Experimental quantification agrees well with the sedimentation model that top cell concentration decreases faster at lower polymer concentration. This is because cell sedimentation velocity is higher at lower polymer concentration [37]. With printing time increasing from 0 to 30–60 to 90 min, the observed bottom cell concentration increases from 1.00 to 1.49–2.00 to 2.54×10^6 cells/ml, from 1.01 to 2.03–3.33 to 5.01×10^6 cells/ml, and from 1.02 to 3.08–5.49 to 8.15×10^6 cells/ml at 1%, 0.5%, and 0.25% (w/v) NaAlg, respectively. In contrast, those predicted by sedimentation model considering single cell assumption are from 1.00 to 1.39–1.78 to 2.18×10^6 cells/ml, from 1.00 to 1.87–2.75 to 3.62×10^6 cells/ml, and from 1.00 to 2.66–4.32 to 5.98×10^6 cells/ml, respectively. Those predicted by sedimentation model considering cell aggregation are from 1.00 to 1.59–2.12 to 2.63×10^6 cells/ml, from 1.01 to 2.32–3.49 to 4.62×10^6 cells/ml, and from 1.01 to 3.26–5.31 to 7.31×10^6 cells/ml, respectively. Experimental quantification fails to agree with the sedimentation model considering single cell but agrees with the sedimentation model considering cell aggregation. This highlights the greater cell aggregation formed during cell sedimentation process at lower polymer concentration would expedite the sedimentation according to Stokes' Law since aggregation generally has larger equivalent radius. Higher cell sedimentation velocity could also increase the uncertainty of cell motion and the chance of forming cell aggregation.

Fig. 3(b) shows the $C_{a,bot}$ at different cell concentration and different polymer concentration over 90-minute printing. At initial 1% (w/v) NaAlg, with printing time increasing from 0 to 30–60 to 90 min, $C_{a,bot}$ increase from 0.00 to 0.21–0.34 to 0.46×10^6 cells/ml, from 0.00 to 0.62–1.02 to 1.37×10^6 cells/ml, and from 0.00 to 1.03–1.70 to 2.29×10^6 cells/ml at initial 1×10^6 , 3×10^6 , 5×10^6 cells/ml, respectively. $C_{a,bot}$ increases as initial cell concentration increases. This is because higher initial cell concentration reduces the space between neighboring cells, which increases the cell-cell interaction, expediting sedimentation by forming greater cell aggregation at initial and during sedimentation. Similar greater increments of $C_{a,bot}$ are observed if the polymer concentration is decreased to 0.5% and to 0.25% (w/v) NaAlg. Fig. 3(c) shows the individual cell and cell aggregation in percentage at different cell concentration, different polymer concentration at initial and at 90-minute printing time. At initial 1×10^6 cells/ml, with printing

time increasing from 0 to 90 min, individual cells percentage decreases from 96.68% to 5.75%, from 97.40% to 11.72%, and from 96.50% to 33.27%, small aggregates percentage increases from 3.32% to 56.07%, from 2.60% to 54.10%, and from 3.5% to 51.20%, large aggregates percentage increases from 0% to 41.52%, from 0% to 34.18%, and from 0% to 15.53% at initial 0.25%, 0.5%, and 1% (w/v) NaAlg, respectively. Most individual cells become cell aggregation as the printing time increases and greater cell aggregation is formed as polymer concentration increases. This echoes the aforementioned discussion that bottom cell concentration becomes greater due to the expedited sedimentation by formed cell aggregation and accumulation of cell sedimentation over time at lower polymer concentration. At initial 0.25% (w/v) NaAlg, if initial cell concentration is increased from 1×10^6 to 3×10^6 to 5×10^6 cells/ml, individual cells percentage decreases from 96.68% to 84.10–70.88% and increases from 5.75% to 4.00–2.80%, small aggregates percentage increases from 3.32% to 15.50–27.77% and decreases from 56.07% to 27.38–5.60%, large aggregates percentage increases from 0% to 0.40–1.35% and increases from 41.52% to 68.62–91.60%, respectively, at 0-min and 90-min printing time. Cell aggregation becomes greater as initial cell concentration increases. This echoes the aforementioned discussion that there is greater cell aggregation formed at initial and during sedimentation since higher cell concentration reduces the space between neighboring cells. Similar greater cell aggregation is observed if the polymer concentration is increased to 0.5% and to 1% (w/v) NaAlg.

4. Influence of cell aggregation on mitigation effectiveness of bioink circulation

The role of cell aggregation in affecting cell sedimentation process and mitigation effectiveness of bioink circulation during inkjet-based bioprinting is studied. Mitigation effectiveness to cell sedimentation is characterized by C_p and the mitigation effectiveness to cell aggregation is characterized by E_a . Specifically, in theory and experiment, Section 4.1 investigates the effect of cell concentration on mitigation effectiveness; Section 4.2 investigates the effect of polymer concentration on mitigation effectiveness; and Section 4.3 investigates the effect of printing time on mitigation effectiveness. 0.5% (w/v) NaAlg, 3×10^6 cells/ml initial cell concentration and 60 min printing time were considered as the basis.

4.1. Cell concentration

By varying initial cell concentration from 1×10^6 to 3×10^6 to 5×10^6 cells/ml while fixing other operating conditions to 0.5% (w/v) NaAlg and 60 min printing time, the effect of cell concentration on mitigation effectiveness is investigated. Fig. 4(a) demonstrates the mitigation effectiveness to cell sedimentation in C_p with different cell concentration under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. Two respective schematics of bioink reservoir at initial 1×10^6 and 5×10^6 cells/ml are shown in Fig. 4(a1) and Fig. 4(a2). At initial 1×10^6 cells/ml, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, the observed C_p increases from 0.02 to 0.63–0.77 to 0.87–0.90. C_p is increased significantly by 0.88 when applying flowrate from 0 to 0.5 ml/min with greater improvement by 0.75 from 0 to 0.1 ml/min compared to 0.13 from 0.1 to 0.5 ml/min. This reflects the superior effectiveness of circulating bioink to mitigate cell sedimentation but large flowrate has slow increments [39]. The C_p predicted by sedimentation model considering single cell assumption increases from 0 to 0.73–0.86 to 0.95–0.97. Single cell-based sedimentation model fails to agree with the experimental quantification with overestimation. In contrast, the C_p predicted by sedimentation model considering cell aggregation increases from 0 to 0.59–0.72 to 0.84–0.87. Experimental quantification agrees well with sedimentation model considering cell aggregation. When the cell concentration is further increased to 3×10^6 cells/ml and to 5×10^6 cells/ml, the observed C_p

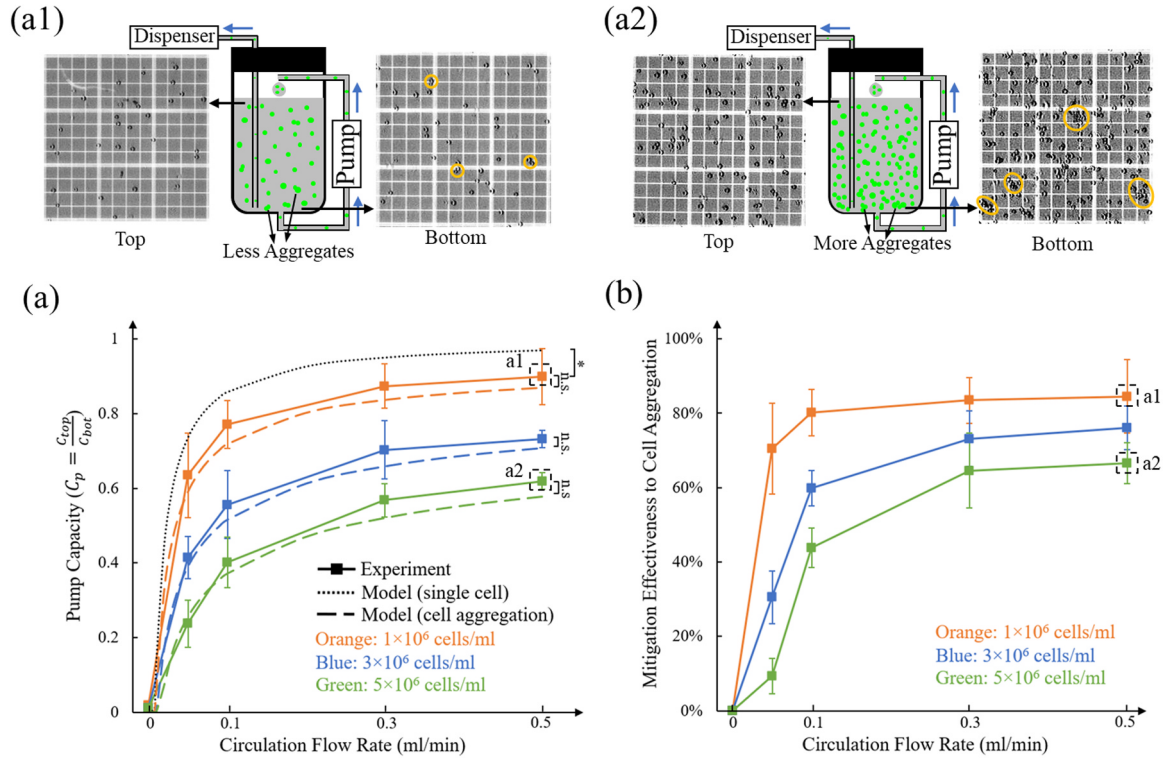


Fig. 4. Mitigation effectiveness to (a) cell sedimentation and (b) cell aggregation with different cell concentrations; (a1) schematic of bioink reservoir at 1×10^6 cells/ml; (a2) schematic of bioink reservoir at 5×10^6 cells/ml.

increases from 0.11 to 0.41–0.56 to 0.70–0.73 and increases from 0.12 to 0.24–0.40 to 0.56–0.62, respectively. In contrast, the C_p predicted by sedimentation model considering cell aggregation increases from 0 to 0.39–0.52 to 0.66–0.71 and increases from 0 to 0.26–0.37 to 0.52–0.58, respectively. Experimental quantification agrees well with sedimentation model considering cell aggregation. The mitigation effectiveness to cell sedimentation is generally reduced as cell concentration increases. This is because the space between neighboring cells becomes less with a higher cell concentration, which could result in stronger cell adhesion and greater cell aggregation due to cell-cell interaction. Pump could only recover the expedited sedimentation-resulting nonuniform cell distribution within the bioink reservoir to a less degree because greater cell aggregation has higher sedimentation velocity.

Fig. 4(b) demonstrates the mitigation effectiveness to cell aggregation with different cell concentration under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. At initial 1×10^6 cells/ml, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, E_a increases from 0% to 70.49–80.15% to 83.45–84.54%. E_a is increased significantly by 84.54% when applying flowrate from 0 to 0.5 ml/min with greater improvement by 80.15% from 0 to 0.1 ml/min compared to 4.39% from 0.1 to 0.5 ml/min. This reflects the superior effectiveness of circulating bioink to mitigate cell aggregation but large flowrate has slow increments. In contrast, when the cell concentration is increased to 3×10^6 cells/ml and to 5×10^6 cells/ml, E_a increases from 0% to 30.55–59.83% to 73.16–76.11% and increases from 0% to 9.35–43.86% to 64.60–66.58%, respectively. This indicates the cell concentration has a significant effect on the mitigation effectiveness to cell aggregation, which is generally reduced as cell concentration increases. This is because the active circulation is less capable to mitigate the larger cell aggregates formed at the higher cell concentration. Circulation flowrate needs to be increased to secure the commensurate effectiveness due to the higher sedimentation velocity of aggregation as discussed above. In summary, sedimentation model considering cell aggregation is capable to predict C_p with good accuracy in cell concentration effect. Higher cell concentration would result in

greater cell aggregation, reducing both of the mitigation effectiveness to cell sedimentation and cell aggregation.

4.2. Polymer concentration

By varying polymer concentration from 0.25% to 0.5–1% (w/v) NaAlg while fixing other operational conditions to initial 3×10^6 cells/ml and 60 min printing time, the effect of polymer concentration on mitigation effectiveness is investigated. Fig. 5(a) demonstrates the mitigation effectiveness to cell sedimentation in C_p with different polymer concentration under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. Two respective schematics of bioink reservoir at 1% and 0.25% (w/v) NaAlg are shown in Fig. 5(a1) and Fig. 5(a2). At 0.25% (w/v) NaAlg, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, the observed C_p increases from 0.00 to 0.21–0.39 to 0.56–0.61. C_p is increased significantly by 0.61 when applying flowrate from 0 to 0.5 ml/min with greater improvement by 0.38 from 0 to 0.1 ml/min compared to 0.23 from 0.1 to 0.5 ml/min. This reflects the superior effectiveness of circulating bioink to mitigate cell sedimentation but large flowrate has slow increments. The C_p predicted by sedimentation model considering single cell assumption increases from 0 to 0.60–0.75 to 0.91–0.95. Single cell-based sedimentation model fails to agree with the experimental quantification with overestimation. In contrast, the C_p predicted by sedimentation model considering cell aggregation increases from 0 to 0.24–0.40 to 0.58–0.64. Experimental quantification agrees well with sedimentation model considering cell aggregation. When the NaAlg concentration is further increased to 0.5% (w/v) and to 1% (w/v), the observed C_p increases from 0.01 to 0.41–0.56 to 0.70–0.73 and increases from 0.10 to 0.60–0.72 to 0.78–0.79, respectively. They agree well with the sedimentation model considering cell aggregation being 0–0.39 to 0.52–0.66 to 0.71 and being 0.10–0.52 to 0.72–0.80 to 0.82, respectively. The mitigation effectiveness to cell sedimentation is generally increased as polymer concentration increases. This is because increasing polymer concentration from 0.25% to 0.5–1% (w/v) NaAlg reduces the

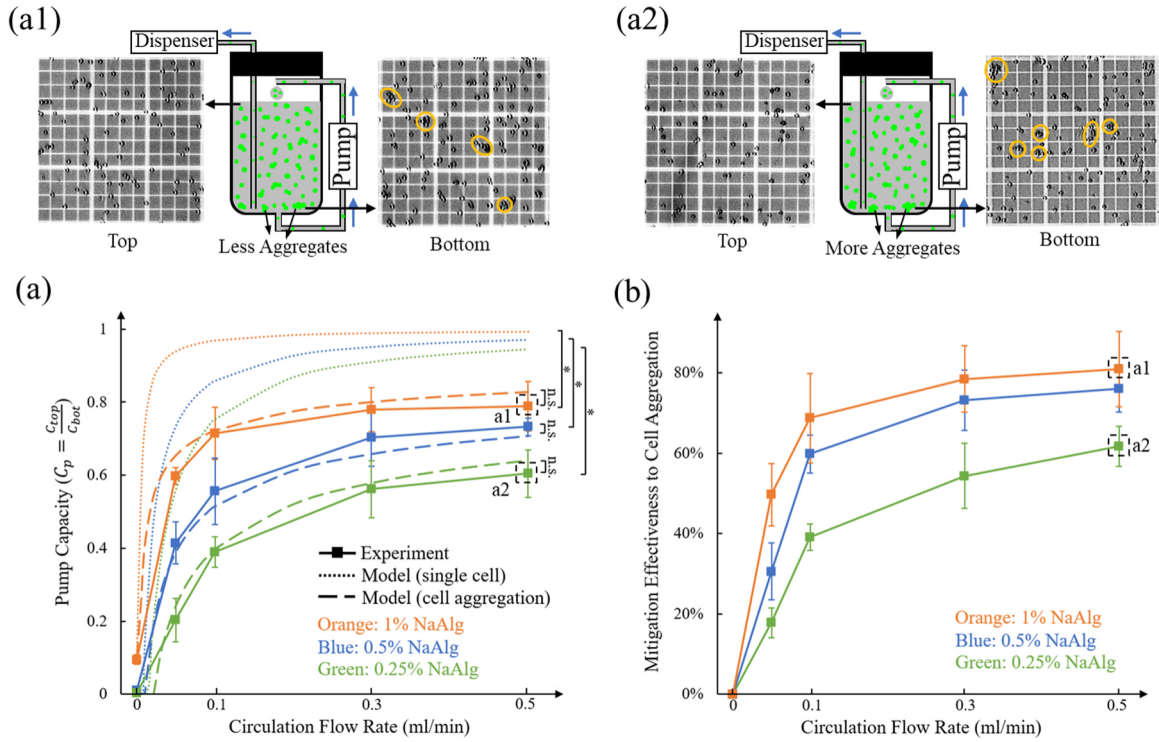


Fig. 5. Mitigation effectiveness to (a) cell sedimentation and (b) cell aggregation with different polymer concentrations; (a1) schematic of bioink reservoir at 1% (w/v) NaAlg; (a2) schematic of bioink reservoir at 0.25% (w/v) NaAlg.

cell sedimentation velocity as previously reported [37]. This retards the expedited sedimentation process to the bottom of the bioink reservoir and reduces the formation chance of cell aggregation due to larger distance-induced less cell-cell interaction. Pump could recover the cell sedimentation-resulted nonuniform cell distribution within the bioink reservoir to a greater degree because of the less cell aggregation.

Fig. 5(b) demonstrates the mitigation effectiveness to cell aggregation in E_a % with different polymer concentration under the active bioink circulation flowrate ranging from 0.05 to 0.5 ml/min. At 0.25% (w/v) NaAlg, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, E_a % increases from 0% to 17.79–39.11% to 54.34–61.70%. E_a % is increased significantly by 61.70% when applying flowrate from 0 to 0.5 ml/min with greater improvement by 39.11% from 0 to 0.1 ml/min compared to 22.59% from 0.1 to 0.5 ml/min. This reflects the superior effectiveness of circulating bioink to mitigate cell aggregation but large flowrate has slow increments. In contrast, when the NaAlg concentration is increased to 0.5% and to 1% (w/v), E_a % increases from 0% to 30.55–59.83% to 73.16–76.11% and increases from 0% to 49.68–68.76% to 78.47–81.02%, respectively. This indicates the polymer concentration has a significant effect on the mitigation effectiveness to cell aggregation, which is generally increased as polymer concentration increases. This is because the active circulation is more capable to mitigate the smaller cell aggregates formed at the higher polymer concentration. Less circulation flowrate could secure the commensurate effectiveness due to the slower sedimentation velocity of less aggregation as discussed above. In summary, sedimentation model considering cell aggregation is capable to predict C_p with good accuracy in polymer concentration effect. Higher polymer concentration would result in less cell sedimentation and cell aggregation, improving both of the mitigation effectiveness to cell sedimentation and cell aggregation.

4.3. Printing time

By varying the printing time from 30 to 60–90 min while fixing other experimental conditions to initial 3×10^6 cells/ml, and 0.5% (w/v)

NaAlg concentration, the effect of printing time on mitigation effectiveness is investigated. Fig. 6(a) demonstrates the mitigation effectiveness to cell sedimentation in C_p with different printing time under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. Two respective schematics of bioink reservoir at 30 and 90-minute are shown in Fig. 6(a1) and Fig. 6(a2). At 30-minute printing time, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, the observed C_p increases from 0.05 to 0.49–0.67 to 0.77–0.79. C_p is increased significantly by 0.74 when applying flowrate from 0 to 0.5 ml/min with greater improvement by 0.62 from 0 to 0.1 ml/min compared to 0.12 from 0.1 to 0.5 ml/min. This reflects the superior effectiveness of circulating bioink to mitigate cell sedimentation but large flowrate has slow increments. The C_p predicted by sedimentation model considering single cell assumption increases from 0.07 to 0.86–0.93 to 0.97–0.98. Single cell-based sedimentation model fails to agree with the experimental quantification with overestimation. In contrast, the C_p predicted by sedimentation model considering cell aggregation increases from 0.05 to 0.53–0.63 to 0.74–0.78. Experimental quantification agrees well with sedimentation model considering cell aggregation. When the printing time is further increased to 60 min and to 90 min, the observed C_p increases from 0.01 to 0.41–0.56 to 0.70–0.73 and increases from 0.00 to 0.27–0.45 to 0.62–0.66, respectively. They agree well with the sedimentation model considering cell aggregation being 0.00–0.39 to 0.52–0.66 to 0.71 and being 0.00–0.29 to 0.43–0.60 to 0.66, respectively. The mitigation effectiveness to cell sedimentation is generally decreased as the printing time increases. This is because increasing printing time would increase the accumulation of expedited sedimentation and increase the cell concentration at bioink reservoir bottom, resulting in greater cell aggregation due to shorter distance-induced greater cell-cell interaction. It becomes more difficult for active circulation to recover the highly nonuniform cell distribution within the bioink reservoir back to the initially uniform cell distribution.

Fig. 6(b) demonstrates the mitigation effectiveness to cell aggregation in E_a % with different printing time under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. At 30 min, with

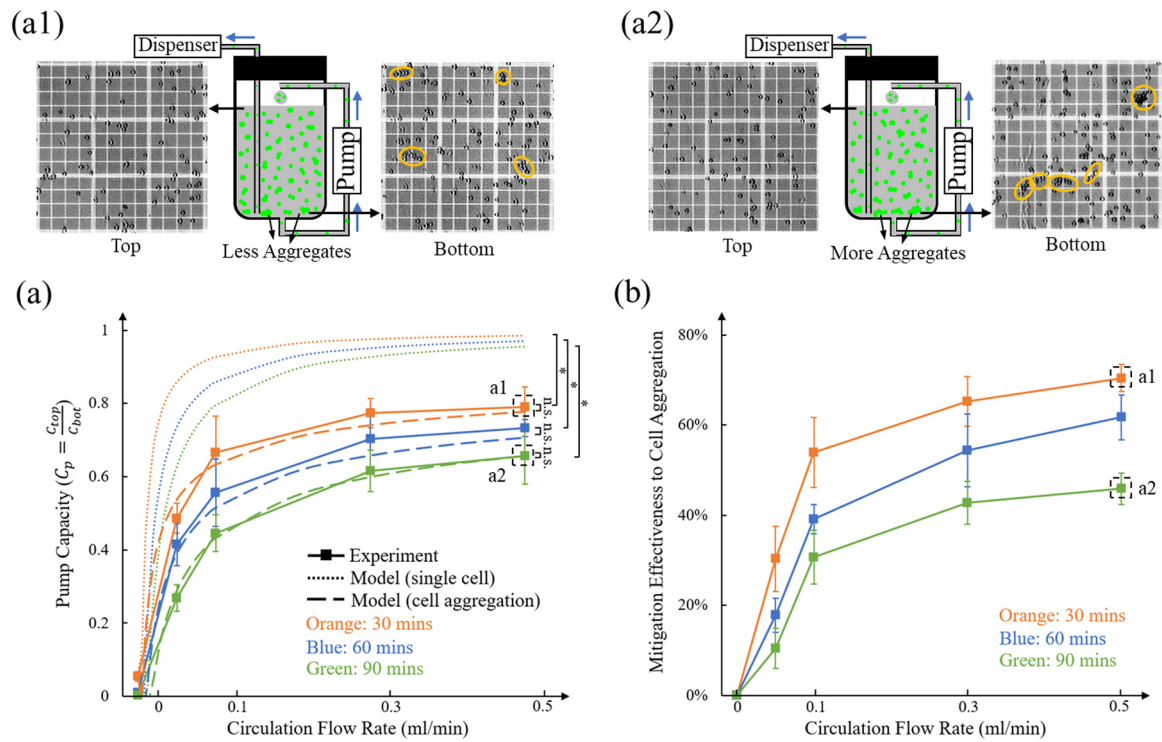


Fig. 6. Mitigation effectiveness to (a) cell sedimentation and (b) cell aggregation with different printing time; (a1) schematic of bioink reservoir at 30-minute printing time; (a2) schematic of bioink reservoir at 90-minute printing time.

circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, $E_a\%$ increases from 0% to 30.30–53.85% to 65.16–70.40%. $E_a\%$ is increased significantly by 70.40% when applying flowrate from 0 to 0.5 ml/min with more significant improvement by 53.85% from 0 to 0.1 ml/min compared to 16.55% from 0.1 to 0.5 ml/min. This reflects

the superior effectiveness of circulating bioink to mitigate cell aggregation but large flowrate has slow increments. In contrast, when the printing time is increased to 60 min and to 90 min, $E_a\%$ increases from 0% to 17.79–39.11% to 54.34–61.70% and increases from 0% to 10.39–30.62% to 42.70–45.83%, respectively. This indicates the

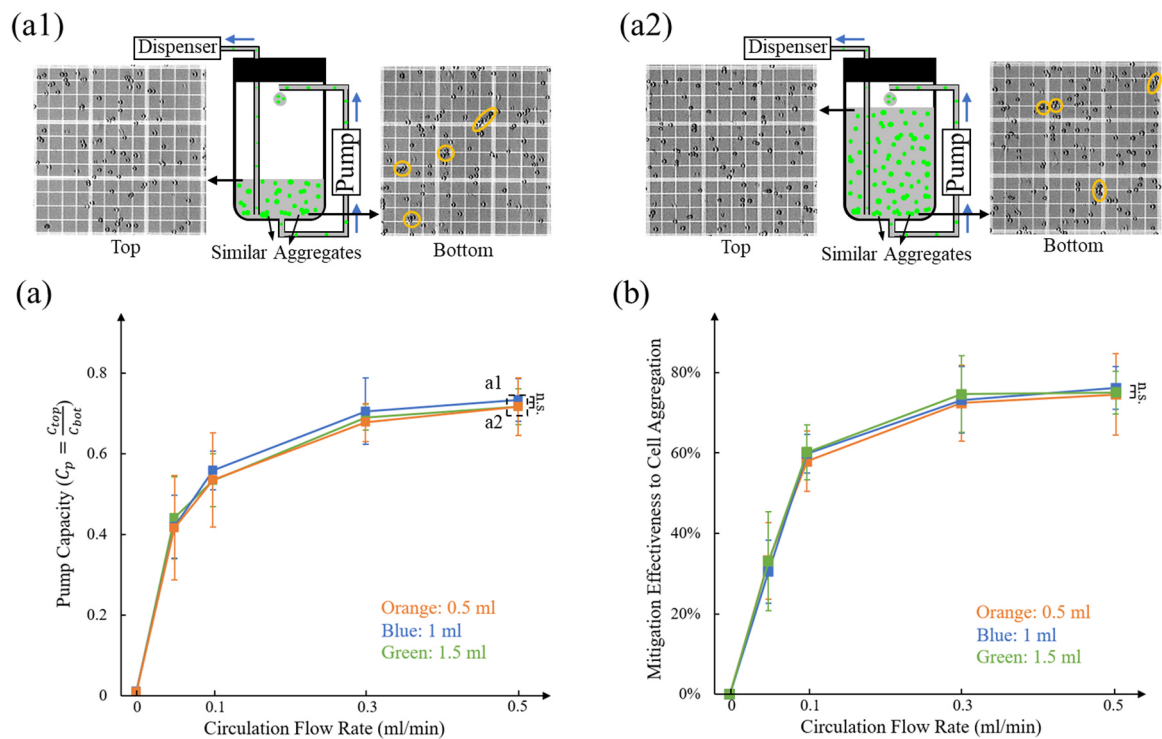


Fig. 7. Mitigation effectiveness to (a) cell sedimentation and (b) cell aggregation with different bioink volume; (a1) schematic of bioink reservoir at 0.5 ml bioink volume; (a2) schematic of bioink reservoir at 1.5 ml bioink volume.

printing time has a significant effect on the mitigation effectiveness to cell aggregation, which is generally decreased as printing time increases. This is because the active circulation is less capable to mitigate the greater sedimentation-induced larger cell aggregates at the longer printing time as discussed above. In summary, sedimentation model considering cell aggregation is capable to predict C_p with good accuracy in printing time effect. Longer printing time would result in greater cell sedimentation and cell aggregation, reducing both of the mitigation effectiveness to cell sedimentation and cell aggregation.

5. Discussions

5.1. Bioink volume within the bioink reservoir

By varying initial bioink volume from 0.5 to 1–1.5 ml while fixing other operational conditions at initial 3×10^6 cells/ml, 0.5% (w/v) NaAlg and 60 min printing time, the potential effect of bioink volume on mitigation effectiveness is investigated. Fig. 7(a) demonstrates the mitigation effectiveness to cell sedimentation in C_p with different bioink volume under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. Two respective schematics of bioink reservoir at 0.5 ml and 1.5 ml are shown in Fig. 7(a1) and Fig. 7(a2). At 0.5 ml, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, the observed C_p increases from 0.01 to 0.42–0.54 to 0.68–0.72 with bioink volume of 0.5 ml. In contrast, when bioink volume is further increased to 1 ml and to 1.5 ml, the observed C_p increases from 0.01 to 0.42–0.56 to 0.70–0.73 and increases from 0.01 to 0.44–0.53 to 0.69–0.72, respectively. This indicates different bioink volume barely affects the mitigation effectiveness to cell sedimentation. This is because increasing bioink volume while applying the geometrically identical bioink reservoir only increases the height of the bioink column. It does not alter the moving fluid velocity within the bioink reservoir and the cell sedimentation velocity relative to the moving fluid, therefore not altering the space between neighboring cells and cell-cell interaction in general. Accumulation of cell sedimentation and the cell concentration at the bioink reservoir top and bottom remains similar. Fig. 6(b) demonstrates the mitigation effectiveness to cell aggregation in $E\%$ with different bioink volume under the active bioink circulation flowrate ranging from 0 to 0.5 ml/min. At 0.5 ml, with circulation flowrate increasing from 0 to 0.05–0.1 to 0.3–0.5 ml/min, $E\%$ increases from 0% to 33.20–57.92% to 72.43–74.57%. In contrast, when the bioink volume is increased to 1 ml and to 1.5 ml, $E\%$ increases from 0% to 30.55–59.83% to 73.16–76.11% and increases from 0% to 33.07–60.20% to 74.61–75.01%, respectively. This indicates different bioink volume barely affects the mitigation effectiveness to cell aggregation. The main reason is that the resultant cell aggregation due to the accumulation of cell sedimentation remains similar as discussed above. It is noted that the model in this paper doesn't consider the decrease of the bioink volume in the reservoir due to printing. The bioink consumption flow rate during the printing depends on the printing conditions, such as droplet size and frequency. Generally speaking, the bioink circulation effectiveness is expected to be slightly increased considering bioink consumption. Because the cell aggregates at the reservoir bottom can be partially extracted and consumed, promoting the effectiveness.

5.2. Optimizations

This study uses root-mean-square error (RMSE) and coefficient of determination (R^2) to demonstrate the model accuracy. RMSE measures the standard deviation of the residuals and R^2 demonstrates the variations in the dependent variables predicted from the independent variables. RMSE and R^2 are calculated, respectively, by:

$$RMSE = \sqrt{\frac{RSS}{N_{sample}}} \quad (16)$$

$$R^2 = 1 - \frac{RSS}{TSS} \quad (17)$$

where RSS: the sum of the squared deviations of predictions from the actual observations, TSS: the sum of the squared deviations of actual observations from the mean, and N_{sample} : the number of samples. R^2 ranges from 0 to 1. $R^2 = 1$ indicates a full explanation of all the variability of data around its mean. $R^2 = 0$ indicates the model fails to explain the variability of data around its mean at all. For predicting C_p at the bottom cell concentration in the stationary bioink shown in Fig. 8(a), $R^2 = 0.8357$ indicates a general good agreement between actual observation and prediction model except for a slight underestimate at the high observation values. This might be due to the greater cell aggregation-induced larger cell-cell interaction. For predicting C_p at the top cell concentration and that at the bottom cell concentration in the circulated bioink shown in Fig. 8(b) and Fig. 8(b), $R^2 = 0.9323$ and $R^2 = 0.9426$, respectively, indicating a good prediction accuracy. The fitting parameters are tabulated in Table 4.

5.3. Circulation outlet and cell viability

This section discusses circulation outlet and demonstrates cell viability assessment when applying bioink circulation setup to the inkjet bioprinting. Circulation tube outlet is situated right above the bioink level. Two problems will be raised if submerging the tube outlet into bioink shown in Fig. 9(a):

1. If the tube outlet is close to the bioink liquid level, it is difficult to continuously flow the outlet end of the tube to match the bioink liquid level during printing. During the printing process, the bioink is continuously consumed and the bioink liquid level in the reservoir decreases.
2. If the tube outlet is below the bioink liquid level too much, it could not achieve a holistic/entire circulation within the bioink reservoir. In this case, only partial bioink will be circulated, which may significantly reduce the circulation effectiveness.

Additionally, Xu et al. [37] observed no significant alteration on the single cell sedimentation velocity v_{cs} during the cell sedimentation process. It was also reported that the measured v_{cs} was around 1.45 $\mu\text{m/s}$ and the estimated time for the whole cell sedimentation process was around 5.7 h within the 3 cm height (h_{bioink}) bioink reservoir containing 0.5% (w/v) NaAlg and fibroblasts. This study defines the time to finish the whole cell sedimentation process as complete cell sedimentation time (t_{ccs}). The t_{ccs} under stationary fluid is dependent upon the v_{cs} and the h_{bioink} . The t_{ccs} under active circulation (t_{ccs-c}) is dependent upon fluid velocity in bioink reservoir v_{f-r} , v_{cs} , and h_{bioink} . Increasing circulation flowrate increases the t_{ccs-c} by increasing the v_{f-r} . Increasing polymer concentration increases the t_{ccs-c} by decreasing the v_{cs} . Increasing the bioink volume within bioink reservoir increases the t_{ccs-c} by increasing h_{bioink} . Different operating conditions result in different t_{ccs-c} , which might affect the mitigation effectiveness, especially in the cases considering expedited sedimentation by influence of cell aggregation with flow circulation flowrates, flow polymer concentrations, flow bioink height and long printing time.

A fluorescence assay (cefepime acetoxymethyl ester and ethidium homodimer III, Biotium, Fremont, CA) was used to determine the viability of cells. The printing conditions are 1% (w/v) NaAlg and flow rate of 0.5 ml/min representing the potentially highest level of shear stress induced by bioink circulation under the current experimental design. At the reservoir bottom, the cell viability is around 87.7% after 90 min circulation compared to standby control of 92.1%. The results of the cell viability show that the majority of cells remain alive during this process, representing the feasibility of the proposed active circulation approach in 3D bioprinting applications.

6. Conclusions and future work

In this study, we constructed a sedimentation model to predict the

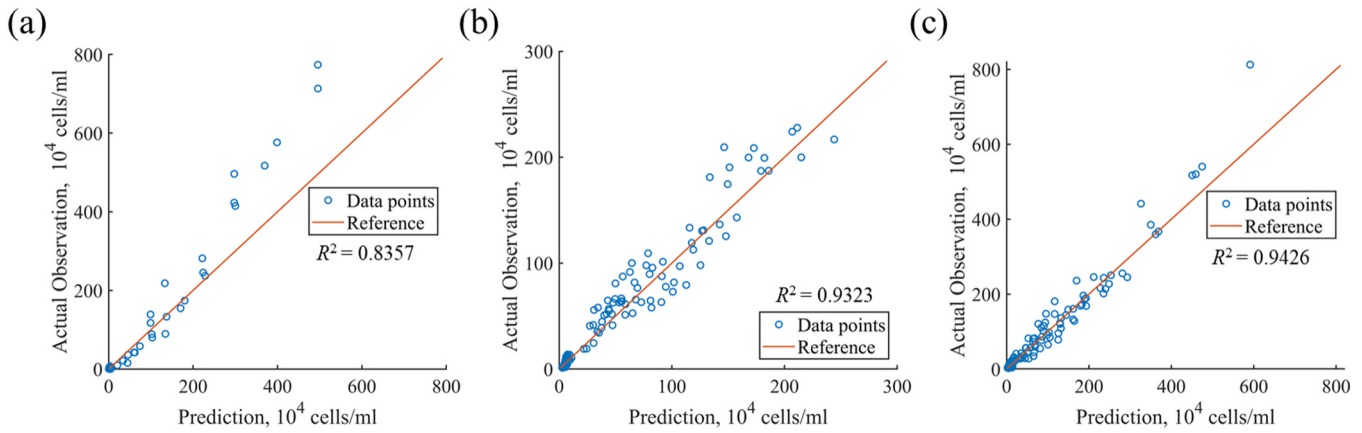


Fig. 8. Actual observation vs model prediction. (a) $C_{a,bot}$ at stationary biofink; (b) $C_{a,top}$ at cfirculated biofink; (c) $C_{a,bot}$ at cfirculated biofink. Reference represents perfect prediction.

Table 4

Major optimization parameters.

	<i>a</i>	<i>b</i>	<i>c</i>	<i>d</i>	<i>e</i>	RMSE	R^2
$C_{a,bot}$ (stationary biofink)	0.0268	0.9994	0.9647	0.7237	-	87.0352	0.8357
$C_{a,top}$ (cfirculated biofink)	0.9323	1.9279	0.5441	0.3143	0.2081	16.3673	0.9323
$C_{a,bot}$ (cfirculated biofink)	3.39E-05	2.0119	0.8314	0.5413	0.3915	32.5192	0.9426

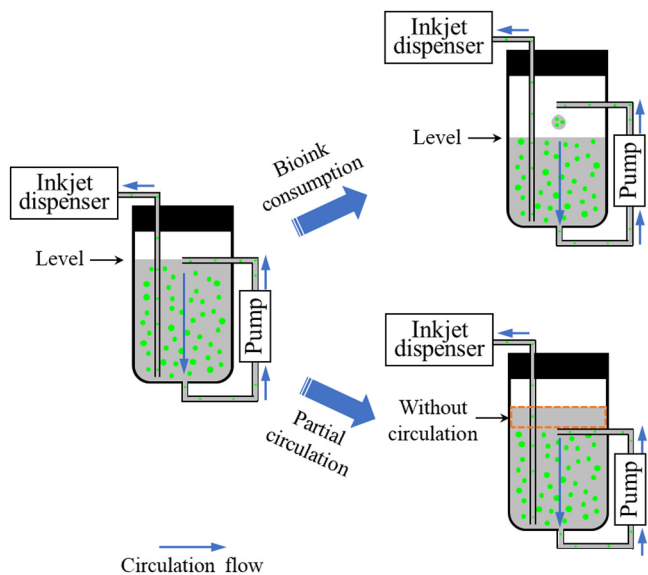


Fig. 9. Two problems due to the submerging of the tube outlet into biofink: biofink fluid level changes due to biofink consumption and partial biofink cfirculation.

performance of biofink reservoir in cell distribution considering the influence of cell aggregation in stationary biofink and in cell sedimentation and aggregation mitigation approach—active biofink cfirculation. The iterative and time-region-based sedimentation model was provided, and the influence of cell aggregation was quantified through least-squares optimization by Nelder-Mead simplex algorithm. The role of cell aggregation in sedimentation of cell-laden biofink during inkjet-based bioprinting was investigated theoretically and experimentally. This study facilitates the prediction of biofink reservoir dynamic performance in cell distribution over printing and prediction of effectiveness of cfirculating biofink to mitigate sedimentation and associated cell aggregation, therefore promoting precise and reliable bioprinting. Major

conclusions are: (1) Sedimentation model considering single cell underestimates the experimental quantifications of cell concentrations in stationary fluid and overestimates the pump capacity in cfirculated biofink, while sedimentation model considering cell aggregation generally agrees with both. Influence of cell aggregation during sedimentation needs to be considered herein; (2) In stationary biofink, the formed cell aggregation expedites the cell sedimentation due to the larger size-induced higher sedimentation velocity, increasing the chance of further forming cell aggregation; (3) Applying higher initial cell concentration from 1×10^6 to 3×10^6 to 5×10^6 cells/ml increases the chance of forming cell aggregates during printing due to less space-induced greater cell-cell interaction which expedites sedimentation process, therefore reducing the mitigation effectiveness to cell sedimentation and cell aggregation; (4) Applying higher polymer concentration from 1% to 0.5% to 0.25% (w/v) NaAlg retards the expedited sedimentation process by reducing cell sedimentation velocity relative to the fluid around and lessens the chance of forming cell aggregates due to greater space-induced less cell-cell interaction, therefore improving the mitigation effectiveness to cell sedimentation and cell aggregation; (5) Applying longer printing time from 30 to 60 to 90 minutes increases the chance of forming cell aggregates due to more expedited sedimentation-induced accumulation of cells and the less space-induced greater cell-cell interaction, therefore reducing the mitigation effectiveness to cell sedimentation and cell aggregation. Future work may involve modeling of cell distribution dynamics within microspheres considering biofink consumption, and development of complete working diagram of the biofink cfirculation using hybrid predictive modeling.

CRediT authorship contribution statement

Jiachen Liu: Writing – review & editing, Writing – original draft, Methodology, Investigation, Conceptualization. **Heqi Xu:** Writing – review & editing. **Md Shahriar:** Writing – review & editing. **Changxue Xu:** Writing – review & editing, Supervision, Resources, Project administration, Methodology, Funding acquisition, Conceptualization.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability

Data will be made available on request.

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