

X-ray Tomographic Microscopy of Eocene Coprolites from Pipestone Springs Main Pocket, Southwest Montana

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10 **Contribution to the field statement [200 words]:** This contribution highlights the broadening
11 applications of high-powered imaging techniques such as x-ray tomographic microscopy (μ CT) in
12 the study of coprolites. Fossilized feces contain a plethora of information specific to the diet, feeding
13 habits, and the surrounding environment of past organisms. However, tapping into this
14 paleontological archive has typically involved destructive methods, requiring sub-sampling of the
15 fossil through mechanical or chemical disaggregation, or serial grinding to extract internal
16 constituents. Using μ CT it is possible to reveal the internal structures and features of these trace
17 fossils in a non-destructive manner. Herein, we used a sub-set of vertebrate carnivoran coprolites
18 from the Pipestone Springs Main Pocket, Southwest Montana to compare macro- through
19 microscopic features that can be observed via μ CT in contrast to mechanical disaggregation. Results
20 demonstrate that in such coprolites lacking significant secondary diagenetic alteration it is feasible to
21 virtually extract bone material from the phosphatic matrix. Moreover, internal porosity of the
22 coprolites shows irregular pores likely left by gaseous vacuoles in conjunction with a much smaller
23 contingent specific to the molds of hair follicles. Neither of these latter components have been noted
24 or measured previously in this coprolite assemblage, presumably because they have been overlooked
25 when employing destructive methods.

26 **Abstract**

27 The Eocene Pipestone Springs Main Pocket (Renova Formation, Jefferson County, Montana, USA)
28 is a locality renowned for its diverse Chadronian (late Eocene; ~38–33.9 million years ago)
29 mammalian fauna and abundant coprolites. Two distinct coprolite size classes were previously
30 identified in the trace fossil assemblage from which we selected representatives to investigate feeding
31 behaviors and dietary selection of the producers. A subset of the selected coprolites was analyzed
32 based on their compositional and taphonomic attributes using non-destructive x-ray tomographic
33 microscopy in combination with more traditional methods including thin-section petrography,
34 scanning electron microscopy, and energy dispersive spectroscopy. Among the features extracted in
35 the tomographic data were skeletal fragments, including those showing evidence of bone-crushing;
36 delicate hair molds; encrusted lithic fragments; and several irregular pores and cracks throughout the

37 coprolites. Segmentation and volumetric renders permit quantitative assessment of the relative
38 proportions of inclusions, revealing porosity as a primary volumetric element aside from the matrix
39 and bone inclusions. There was no significant difference in the total volume of bone extracted
40 between coprolite size class, though the smaller coprolites preserved a relatively higher volumetric
41 proportion of undigested skeletal material. This multi-visualization approach provides a means to
42 observe and evaluate differences in the coprolite gross morphology and inclusions across the two size
43 classes, thereby offering valuable insights into the broader paleoecology of the Pipestone Springs
44 Main Pocket coprolite producers and holding promise for comparable paleo-dietary studies of other
45 coprolite-rich deposits.

46 1 Introduction

47 Coprolites and affiliated trace fossils offer a rare glimpse into the feeding, digestive, and excretory
48 behaviors of their producers whilst simultaneously capturing unique paleoecological and
49 paleoenvironmental information (Hunt et al., 2012; Myrhvold, 2012). Examination of such fossils
50 typically requires sub-sampling of the specimen, often employing destructive sampling techniques
51 (e.g., disaggregation or dissolution) to either a portion of the specimen or its entirety (Bryant, 1970;
52 Fry, 1970; Shillito et al., 2020). Recent studies have utilized non-destructive three-dimensional (3D)
53 imaging techniques, such as x-ray tomographic microscopy (μ CT) and synchrotron
54 microtomography, to reveal macroscopic and microscopic inclusions that may otherwise be lost via
55 more traditional methods (Bravo-Cuevas et al., 2017; Qvarnström et al. 2017, 2019; Wang et al.,
56 2018; Romanuk et al., 2020; Abella et al., 2022). Challenges with x-ray-based methodologies
57 persist, however, often owing to the attributes of the sample. For instance, depending on the
58 preservation of the coprolite, some inclusions may be compositionally similar to the matrix material.
59 In such cases, overlapping intensity ranges will make discerning these features difficult via standard
60 segmentation or thresholding techniques. This lack of phase contrast can be exacerbated by
61 taphonomic processes caused during digestion wherein the bone margins become diffuse, or during
62 diagenesis when remobilization of soluble minerals causes secondary infilling of pores. Diagenesis
63 can also reduce or eliminate phase contrast through secondary remineralization of the entire
64 specimen. Nevertheless, for coprolites and inclusions that have not been substantially altered through
65 taphonomic or diagenetic processes, μ CT proves to be a powerful tool in extracting inclusions
66 virtually for qualitative analysis—although its applications for such research at present are in their
67 infancy (but see Huisman et al., 2014; Shillito et al., 2020). Where prior coprolites studies employing
68 μ CT have been predominantly qualitative, intent on the description of the types of inclusions, the
69 quantitative analysis of extracted components and associated potential to address testable hypotheses
70 is an area that requires further research.

71 Herein, we employ non-destructive μ CT in conjunction with targeted consumptive sampling to gain a
72 more holistic view of the internal composition and inclusions of a subset of coprolites from a well-
73 documented trace fossil assemblage. Specifically, we targeted coprolite material from the Pipestone
74 Springs Main Pocket (PSMP) assemblage (Renova Formation), Jefferson County, Montana. This site
75 is renowned for its diverse mammalian fauna and associated trace fossils (Lofgren et al., 2017), and
76 previous work has reported on the taxonomy of vertebrate inclusions and the likely identity of at least
77 one taxon of coprolite producer. Lofgren et al. (2017) examined coprolite surface morphology and
78 visible skeletal elements in 358 specimens, including 20 specimens that were mechanically prepared
79 by removing skeletal inclusions. It is worth noting that the regular shape, phosphatic composition,
80 low bone density, and highly digested, fragmented nature of the bones in these specimens supports
81 their interpretation as coprolites, rather than regurgitalites (Myrhvold, 2012; Gordon et al. 2020;
82 Serafini et al. 2022). Analysis of the measurable dimensions of this coprolite assemblage have

83 previously revealed two distinct size classes, including smaller forms ranging from 4–15 mm in
84 diameter and larger forms ranging from 16–29 mm in diameter (Lofgren et al., 2017). The larger
85 coprolites could not be attributed to any single genus with confidence, and are suggested to have
86 been produced by *Brachyrhynchocyon dodgei*. The producer of the smaller coprolites, with several
87 smaller carnivorous taxa as potential candidates was deduced to be *Hesperocyon gregarius* (Lofgren
88 et al., 2017). This prior work provides an important foundation from which we can both ascertain the
89 broader paleoecological framework for the coprolite assemblage and further build upon it by
90 assessing the taphonomic and diagenetic attributes of the coprolites.

91 Utilizing computed tomographic techniques provides a novel avenue of investigation to substantiate
92 previous findings on the gross morphology of the PSMP coprolites, as well as to (1) visualize 3D
93 specimen structure and identify the undigested skeletal and keratinous inclusions via virtual
94 segmentation; (2) quantify the relative proportions of bone inclusions and pores to feces matrix; and
95 (3) discern any difference between the two size classes with respect to the shape and volume of bone
96 inclusions. This work further serves to assess the potential applications (and limitations) of μ CT
97 techniques in the analysis of coprolites. Finally, corroborating previous findings with data herein, we
98 seek to further elucidate the feeding behavior of these ancient vertebrate producers across coprolite
99 size classes, while also detailing the taphonomic and diagenetic processes found within the PSMP
100 assemblage.

101 2 Geological Setting

102 Coprolites are derived from the PSMP, which is considered part of the Climbing Arrow Member of
103 the Renova Formation in Jefferson County, Montana (Kuenzi and Fields, 1971; see Hanneman et al.,
104 2022, figures 1 and 2). Specimens documented herein were collected from the upper-most 15m-thick
105 silty mudstone unit of the Pistone Springs reference section. Outcropping strata are exposed on
106 opposing sides of an erosional gully though both isolated pockets are lithologically and faunally
107 indistinguishable, hence are often referred to as the same bed (Tabrum et al., 1996). Material herein
108 was collected from the RAM V200301 locality which is situated on the dip slope towards the
109 northern end of the Main Pocket exposures, in the same vicinity as MV 5811, MV 5902, and CM
110 3575 (Orr, 1958; see Hanneman et al., 2022, figures 3 and 4).

111 Based on prior biostratigraphic correlation and magnetostratigraphy (Prothero, 1984), the PSMP
112 assemblage has been estimated as middle to late Chadronian in age (35.0–35.5 Ma). The deposit is
113 exposed as a 50 cm–4 m thick layer of homogenous tuffaceous mudstone, lacking sedimentary
114 structures or obvious bioturbation (Lofgren et al., 2017). Coprolite and vertebrate remains are
115 randomly distributed (without size-sorting) throughout the deposit, both laterally and vertically,
116 precluding hydrodynamic sorting (Lofgren et al., 2017). Several interpretations concerning the
117 formation of these deposits have been proposed, with initial scenarios involving either repeated ash-
118 rich mud flows or inundated floodplain deposits (Kuenzi and Fields, 1971; Lofgren et al., 2017).
119 However, in the absence of clear debris-flow or fluvial sedimentary features, Hanneman et al. (2022)
120 interpret these deposits as having been formed by predominantly aeolian processes. Paleosols of
121 varying degrees of development and exhibiting extensive bioturbation are intercalated between
122 aeolian events represented by direct and reworked ashfall deposits, and loessites (Hanneman et al.,
123 2022).

124

125 3. Methods

126 The twelve coprolite specimens examined herein (Table 1) are housed in the collections of the
127 Raymond M. Alf Museum of Paleontology (RAM), Claremont, California. The external appearance
128 of the coprolites was examined using reflective light microscopy and photographed using a Magnify²
129 GIGAMacro Robotic Imaging System with Canon EOS Rebel T6i/T8i DSLR cameras and Nikon x1
130 objective to acquire gigapixel resolution photomosaics. Petrographic photomicrographs were
131 captured using a Nikon D3300 DSLR camera mounted to a Nikon Eclipse E200 polarizing
132 microscope. Background illumination was corrected using open-source imaging software FIJI
133 (Schindelin et al., 2012) and the method provided by Landini (2006).

134 Exterior physical features were described using the methods outlined in Jouy-Avantin et al.
135 (2003), including the identification of color, shape, hardness, texture, the presence or absence of
136 constrictions, and inclusions visible at the surface. Coprolite color was characterized based on the
137 Munsell Color Rock-Color Chart (Munsell Color, 2010). The overall shape and specific morphology
138 of the coprolite extremities were noted (e.g., sharp-ended, rounded, or broken), as were taphonomic
139 modifications including desiccation, abrasion, and surface markings/burrows (Supplementary Table
140 S1). Any inclusions visible on the surface were also noted (i.e., bones, adhering sediment). The
141 surface texture was examined to determine whether the coprolite displayed homogenous or
142 heterogeneous mixtures; compacted aggregates less than 1 mm, compacted aggregates greater than or
143 equal to 1 mm, homogenous mixtures with some aggregates, or many aggregates included in a
144 homogenous mixture.

145

146 3.1. Microscopic Analyses and Visualization

147

148 Coprolite samples were scanned using a Zeiss Xradia 510 Versa μ CT microscope at the X-ray
149 Microanalysis Laboratory (Mizzo μ X), University of Missouri, for non-destructive analyses. Optimal
150 scanning parameters for the coprolites varied, with source voltage ranging between 80–140 kV,
151 source power between 7–10 W, and exposure time between 1–5 second(s). All scans captured 1601
152 projections through 360° of rotation and used a 0.4X objective. Two types of Zeiss low-energy filters
153 were used based on sample transmittance values, with 12 scans using the LE5 filter, and two using
154 the LE2 filter. Voxel size ranged from 3.4618–30.12 μ m. The scanning parameters for each sample
155 are summarized in the Supplementary Table S2.

156 Visualization of the 3D data was achieved by importing serial tomogram stacks into
157 Dragonfly software v. 2020.2 Build 941–v. 2022.2 for Windows, Object Research Systems (ORS)
158 Inc, Montreal, Canada, 2018 (<http://www.theobjects.com/dragonfly>). Segmentation via labeled
159 voxels was performed using upper and lower Otsu thresholding of greyscale values, in combination
160 with other operations such as fill inner areas, Boolean calculations, and in certain cases manual
161 segmentation throughout the image stacks to extract internal constituents and features of the
162 coprolites (e.g., bones and pore spaces) from the matrix. Volume measurements and relative
163 volumetric proportions of pore space and bone inclusions were calculated for each coprolite
164 tomogram. Note for pores, the remaining porosity was measured, which excluded pores that were
165 secondarily infilled. For more delicate features such as the tubular voids inferred to be moldic
166 preservation of hair, a subsample of 300 slices was classified using the Trainable Weka Segmentation
167 Fiji plugin (Arganda-Carreras et al., 2017) to differentiate these features from other volumetric
168 elements, and subsequently imported into ORS Dragonfly for visualization and quantification in 3D.
169 Feret diameters (i.e., caliper diameter, defined as the distance between the two parallel planes

170 restricting the object perpendicular to that direction) for the bones and pores were measured within
171 Dragonfly. The minimum feret diameter for bones was set at 0.14 mm and the minimum feret
172 diameter for pores set at 0.196 mm. A multi-ROI (Region Of Interest) was extracted from the bone
173 segmentation into group-labeled voxels to identify individual bone components. Select components
174 were then extracted as meshes (.stl files) and smoothed for one iteration in Dragonfly before being
175 exported to Meshmixer [Autodesk Meshmixer 3.5, (RRID:SCR_015736)]. The 3D meshes were
176 rendered to remove islands and unrelated material and applied with a shader. Each bone was
177 examined individually for identification and to determine the general shape and signs of
178 fragmentation.

179 A single specimen (RAM 17540) was prepared and sectioned for examination via optical and
180 scanning electron microscopy (SEM). The coprolite was strengthened with PALEObond Penetrant
181 Stabilizer, embedded in epoxy, and cut diagonally along a section predetermined from observation of
182 the μ CT data. One of the two halves was polished using a Buehler EcoMet250, while the other was
183 left unpolished. Both halves were then analyzed using a Zeiss Sigma 500 VP SEM equipped with a
184 high-definition 5-segment backscattered electron detector at the Mizzou μ X lab and imaged using the
185 Bruker ATLAS workflow for large-area SEM mosaics. Elemental mapping was conducted on
186 specific regions of interest using dual Bruker XFlash energy dispersive X-ray spectrometers (EDS).
187 All SEM analyses were conducted at their optimal operating conditions of 20 keV beam accelerating
188 voltage, 40 nA beam current, 60 μ m aperture for imaging (120 μ m aperture for EDS elemental
189 mapping), chamber pressure at 20 Pa, and a working distance of 16.5 mm.

190

191 3.2. Statistical Analysis

192

193 Analyses were conducted and figures produced using software package *R* (R core Team, 2017;
194 Version 4.1.0) and associated R packages *boot*, *diptest*, *ggplot2*, *ggtreemap*, *ggpubr* and *mclust*
195 (Davison and Hinkley, 1997; Scrucca et al., 2016; Wickham, 2016; Arnold, 2021; Canty and Ripley,
196 2021; Maechler, 2021). Raw data is provided in Supplementary Tables S3–S6 along with *R* scripts
197 (Supplementary Scripts S1). Using the volumetric data in Supplementary Table S4, two bar graphs
198 were produced in Microsoft Excel in order to assess the respective contributions of matrix, bones,
199 and pores to total coprolite volume (mm^3) and the relative proportions of these components for each
200 sample.

201 **3.2.1. Testing for size classes of coprolites reported in Lofgren et al. (2017).** Size data of
202 coprolites reported in Lofgren et al.’s (2017) Figure 11 were extracted (in 1 mm bins) using
203 plotdigitizer.com (on March 8, 2023, Table S6). Gaussian finite mixture modelling of these data was
204 conducted using the *mclust* package in R.

205

206 **3.2.2. Testing for differences in size classes of our subset of Lofgren et al.’s coprolites.** We tested
207 for differences in nine variables by coprolite size class, including measures of length, width, mass,
208 proportion of pore volume, and proportion and size of bone inclusions. We first used the Shapiro test
209 for normality and the Dip test for unimodality (if the data were not normally distributed) to determine
210 if the Wilcoxon Rank-Sum tests were appropriate.

211 4. Results

212 4.1. Coprolite External Morphology

213

214 The coprolites were divided into two different size classes previously defined by Lofgren et al.,
215 (2017) based on their diameter, with larger coprolites ranging between 16–29 mm and smaller
216 coprolites between 4–15 mm. Within our subset we had seven larger class coprolites (Class I; RAM
217 17370, 17405, 17517, 17540, 17546, 17547, and RAM 18171) and five smaller class coprolites
218 (Class II; RAM 17557, 31209, 31211, 31212, and 31214). Size measurements (i.e., mass, length, and
219 width) are summarized in Table 1. Coprolites exhibited three different colors, including yellow gray
220 (5Y 7/2), grayish yellow (5Y 8/4), or yellowish gray (5Y 8/1), as defined in the Munsell Color Chart
221 (2010). There was no obvious color difference between the classes of coprolites, though all Class II
222 coprolites are yellowish gray, 5Y 8/1.

223 Most of the examined Class I coprolites (5/7) have a smooth, relatively homogenous surface
224 and tend to share a similar cylinder-like shape, circular in cross-section though occasionally flattened
225 on one side (i.e., RAM 17517) (Figure 1). Notable exceptions include RAM 17546 (Figure 1F) and
226 RAM 17370 (Figure 1G), which both display a rough, topographically complex surface, while
227 simultaneously showing signs of constrictions. The examined Class II coprolites also have
228 homogenous, smooth surfaces but are less uniform in shape. As noted by Lofgren et al. (2017), the
229 smaller forms exhibit blunt or tapered ends, and occasionally both. Though few, coprolites within
230 both size classes I and II show small black bone inclusions on their surface visible to the naked eye.
231 In-depth descriptions of each specimen can be found in the Supplementary Text S1.

232 [Insert Table 1 here]

233

234 4.2. Taphonomic Surface Features

235

236 **4.2.1 Desiccation Cracks.** Desiccation cracks occur prior to lithification of the coprolite and are
237 caused as a response to both climate and depositional environment (Northwood, 2005). Several
238 samples displayed desiccation cracks on the exterior coprolite surface (N=6, Table S1), varying in
239 size from large, conspicuous cracks with infilled sediment to small hair-line cracks on the surface of
240 the coprolite. An example of these features can be seen on RAM 17517 (Figure 2A), where the
241 cracks appear as small continuous lines that disrupt the specimen surface. RAM 18171 also displays
242 large, infilled cracks approximately midway along the sagittal length that range from 3–10 mm in
243 length (Figure 1A). Desiccation cracks were only noted in the larger Class I coprolites where both
244 distinct and finer cracks could be observed together on the same sample.

245 **4.2.2 Abrasion.** Abrasion relates to how smooth the surface has become in response to erosion
246 caused by water or wind-born particle scouring following excretion or transport and weathering post
247 lithification (Northwood, 2005). We employed the three categories designated by Northwood (2005)
248 to describe the degrees of abrasion including: A. showing no surface abrasions, B. showing little
249 surface abrasion, C. showing significant surface abrasion with a smooth surface. Surface abrasion
250 was common, with most samples (N=8) displaying evidence of significant abrasion and evenly across

251 the two size classes (Supplementary Table S1). Two specimens showed evidence of some abrasion
252 (RAM 17517, Figure 1B and RAM 17546, Figure 1F). Those showing no abrasion typically
253 preserved either topographically rough surfaces (RAM 17370, Figure 1G) or delicate features such as
254 radial parallel anal sphincter marks (RAM 31209, Figure 1L). There is no obvious difference in
255 abrasion pattern between coprolite size classes, as both exhibit the full range of abrasion profiles.

256 **4.2.3 Surface Marks.** Surface marks are more ambiguous in their origin and manifest as some sort of
257 imprint or trace left by either inanimate objects or a biological agent post defecation. Frequently,
258 these include traces from coprophagous invertebrates as surface scratches or burrows (Northwood,
259 2005; Eriksson et al., 2011). A few specimens show plausible evidence of invertebrate scratch marks,
260 displayed as short, radiating, unbranching traces with no unified orientation (Figure 2B). Whether
261 these markings are biogenic or abiogenic (i.e., abrasion) is difficult to discern with confidence, though
262 it might be expected that abrasion marks are less localized and more evenly distributed compared to
263 biogenic markings. Notably, the scale of these marks is also comparable to the surface expression of
264 exposed hair molds (i.e., RAM 17403 and 17557), which appear as fine linear impressions on the
265 coprolites' surface (Figure 2C).

266 **4.2.4 Breakage.** Breakage in coprolite samples is any sort of mark that shows that the sample has
267 broken after the fossilization process. Breakages are more likely to occur close to surface inclusions
268 within the coprolite, which tend to create weak points (Northwood, 2005). The extent of breakage
269 within the present dataset varies from small parts of the coprolite chipped off (Figure 1H, I, K) to
270 larger portions presumably missing (i.e., RAM 17546; Figure 1E). Some of the coprolite specimens
271 show partially smoothed fracture surfaces, such as RAM 18171 (Figure 1A), indicative of weathering
272 and exposure following the fracture, as compared to fresher, more angular fractures in other
273 specimens such as RAM 17547 (Figure 1E). Breakage was more prevalent in the Class I coprolites
274 than their smaller contemporaries.

275 **4.2.5 Decomposition and Distortion.** Decomposition of the feces prior to lithification is the main
276 cause of spherical to irregular cavities on the coprolite surfaces, which result from the accumulation
277 of decompositional gasses (Northwood, 2005). Such features were present in both Class I and Class
278 II coprolites and varied in size from sub-millimetric scales to spanning several millimeters. Class I
279 coprolites have larger surface pores, for example best preserved in RAM 17546 and 17370 (Figure
280 1F and G respectively). The large size of these cavities rules out their representing the remnants of
281 bacterial colonies (described in phosphatic coprolites by, for example, Hollocher et al., 2001). The
282 surface cavities were described based on their relative abundance (Table S1). One notable cavity was
283 a linear feature with closely spaced repeated circular depressions (Figure 2F). Select coprolites also
284 display a ventrally flattened surface associated with pre-lithification plasticity of the sample (e.g.,
285 RAM 17405 and 18171). This distortion provides some indication of the moisture content of the
286 original scat when excreted and is unique to the larger Class I coprolite samples in the subset
287 observed.

288

289 **4.3. Coprolite Internal Morphology and Inclusions**

290

291 **4.3.1 Matrix.** Petrographic analysis reveals a very-fine to medium-grained matrix of predominantly
292 amorphous phosphate interspersed with fine unidentifiable bone fragments (Figure 3). SEM-EDS
293 analysis of RAM 17546 confirms a calcium and phosphorous (carbonated hydroxyapatite) matrix

294 composition, with minor traces of silicon, aluminum, and iron (Figures 4 and 5). Evidence of
295 digestive corrosion surfaces and phosphate enrichment are visible along defined boundaries within
296 the matrix (Figure 3A), with some boundaries between different compositional textures notably more
297 diffuse (Figure 3B). There are also conspicuous contacts between the apatite matrix and adhered
298 sediment on the external surface of the coprolite (Figure 3E), characterized by detrital grains and
299 finer sediment containing aluminum and potassium (Figure 5B). Petrographic and μ CT analyses
300 additionally demonstrate evidence of desiccation cracks extending inwards from the surface of the
301 coprolite.

302 **4.3.2 Pores.** All coprolites examined reveal pores as a major structural element, with voids ranging in
303 volume between 0.001–1389.2 mm³, and feret diameters of 0.196–50.562 mm. When distinguished
304 from other moldic inclusions (e.g., hair molds, discussed below), no universal pore shapes could be
305 identified, with morphologies ranging from long and thin to large and irregular in shape (Figures 3C–
306 D, 6, and 7). Light microscopy and SEM analysis of RAM 17540 reveal that the pores are frequently
307 lined with crusts of botryoidal silica (Figure 3C–D, 5C–D). Interconnected porosity is observed but
308 limited and few pores are in contact with the external surface; instead, most pores are isolated within
309 the matrix (Figure 6). By volume, pores compose the second largest constituent of the coprolite on
310 average after the matrix, comprising between 3.35–14.32% across both coprolite size classes
311 (Supplementary Table S4). The number of pores per specimen varied markedly, with their
312 distribution throughout the sample relatively even except when influenced by the position of bone
313 inclusions within the coprolite matrix (Figure 6J and K).

314 **4.3.3 Hair Inclusions.** Notable features detected using μ CT are micron-scale tubules that represent
315 the moldic remains of fossil hair within the coprolite matrix. Unlike the irregular pore spaces or other
316 taphonomic features, these structures have a distinct and consistent morphology that does not display
317 random branching attributed to desiccation cracks, burrows by coprophagous organisms or traces of
318 fungal hyphae (Chin, 2007). Fossilized hair was observable via tomography in 10 of the 12
319 coprolites. In thin section, hair molds appear as elongated, parallel, straight-edged tubules with
320 circular cross-sections (Figure 3F) and are typically infilled with silica (Figure 5D). Measured cross-
321 sections reveal molds range in width from 0.015mm–0.092mm (n=60, mean=0.040). Hair molds are
322 also discernible in tomographic slice data; for example, the targeted high-resolution scan (an ~1 cm
323 cylindrical ROI) of specimen RAM 17540 revealed small elongate, tube-like structures, each only a
324 few millimeters in length (Figure 7). While hair molds were distributed relatively evenly throughout
325 the scanned volume (Figure 7B–D), their distribution throughout the entire coprolite was more
326 variable.

327 **4.3.4 Bone Inclusions.** Bone inclusions visible on the exterior of the coprolites were often dark in
328 color relative to the surrounding matrix. The sectioned surface and corresponding thin section of
329 RAM 17540 shows that bone inclusions varied in their quality of preservation; the better-preserved
330 fragments displayed darker inclusions with clear external margins (Figures 3A, 4A) while poorly
331 preserved, heavily corroded, buff-colored inclusions displayed ill-defined margins (Figure 5E).
332 Compositionally, the bones are similar to the matrix, differing by their increased concentrations of
333 sulfur and decreased concentrations of silicon and iron as compared to the matrix (Figures 5A, 5E).
334 Virtually extracted material from μ CT data shows a considerable variation in the size and degree of
335 fragmentation of bone inclusion, with a maximum bone feret diameter of 16.56 mm for the Class I
336 coprolites and 25.73 mm for Class II. Except for a few coprolites, principally those preserving parts
337 of long bones (inc. RAM18171, Figure 8A; RAM 17557, Figure 8H; RAM31212, Figure 8J), there is
338 limited evidence of preferred bone orientation due to the overly fragmented nature of the inclusions.

339 More typically, smaller bone fragments are visible ‘floating’ around the larger bones within the
340 coprolite matrix.

341 A total of 437 bone inclusions (excluding volumes of $<0.001 \text{ mm}^3$) were virtually extracted
342 from the 12 coprolites examined. Due to their predominantly fragmentary nature (most $<1 \text{ mm}^3$), the
343 majority could not be identified to any single taxon, nor categorized anatomically. However, some
344 bones preserved sufficient morphological detail outside of general shape to allow attribution to a
345 particular bone type, i.e., long and short, cancellous, flat and irregular bones. Full descriptions of the
346 most complete extracted bone material figured in Figures 9 and 10 are available in the Supplementary
347 Text S2.

348

349 **4.4. Statistical Analyses**

350

351 The relationships between coprolite volumetric measurements and relative proportion of inclusions
352 (i.e., matrix, pores, bones) are compared for individual coprolite samples (Figures 11A and B,
353 respectively). The proportion of pores within the matrix was consistently greater when compared to
354 that of bone inclusions (Figure 11B). The lowest percentage of pores in any sample was in RAM
355 17370, with 3.89%, and the highest percentage was 14.32% in RAM 31211. Bone inclusions, on the
356 other hand range from 0.09% in RAM 17546 to 11.64% in RAM 31211.

357 All 12 coprolites (Class I, $n=7$; Class II, $n=5$) were included in statistical analyses to test for
358 differences between the two classes of coprolites. The distribution of coprolite widths based on
359 Lofgren et al.’s (2017) data is best modelled as two Gaussian distributions with unequal variance
360 values. Component 1, the small size class, has a median width of 10.6 mm and comprises 79% of the
361 data. Component 2, the large size class, has a median width of 18.4 mm and comprises 21% of the
362 data. We interpret the coprolite specimens analyzed in this study to cleanly fall into one of the two
363 size categories ($n_{\text{small}} = 5$, $n_{\text{large}} = 7$) (Figure S1).

364 Coprolite length, width, mass, proportion of pore volume, and proportion of bone volume
365 values for coprolites cannot be shown to not be normally distributed (Table S7). The Shapiro tests
366 results for bone volume, maximum feret diameter, mean feret diameter, and minimum feret diameter
367 suggest that these values are not normally distributed, however, the Dip Test for Unimodality results
368 cannot refute unimodality (Table S7), therefore the Wilcoxon test is appropriate for these
369 comparisons.

370 The Wilcoxon Rank-Sum test revealed significant differences in length ($p = 0.003$), width ($p = 0.003$), and mass ($p = 0.003$) between Class I and Class II coprolites (Figure 11C). We bootstrapped
371 95% confidence intervals for the median proportion of pore volume between Class I (0.054 mm^3) and
372 Class II (0.082 mm^3) coprolites and found no significant difference. Conversely, bootstrapping for
373 the median proportion of skeletal material between Class I and Class II suggests that there is a
374 significant difference in the median.

376 Differences in median bone size between the two coprolite classes were tested with a Wilcoxon
377 Rank-Sum test, comparing bone volume, maximum feret diameter, mean feret diameter, and minimum
378 feret diameter (Figure 12A–D; Supplementary Table S7). In total 437 bone values were used in the
379 analysis (Class I, $n = 172$; Class II, $n = 265$). There was no significant difference ($p = 0.176$) between
380 the median bone volumes of Class I (0.1 mm^3) and Class II (0.09 mm^3). The median value of the

381 maximum feret diameters of bones for Class I (1.58 mm) was significantly larger ($p = 0.002$) than
382 Class II (1.250 mm). Similarly, there was a significant difference ($p < 0.001$) between the median value
383 of mean feret diameters of bones of Class I (1.015 mm) and Class II (0.870 mm). The same result can
384 be seen with the median value of minimum feret diameters of bones, with Class I (0.495 mm)
385 significantly larger ($p=0.004$) than Class II (0.460 mm).

386

387 **5. Discussion**

388

389 **5.1. Taphonomic Features and Internal Structures**

390

391 The PSMP coprolites are well-preserved, excluding extensive surface abrasion, and possess minor
392 diagenetic alteration or secondary mineralization. The limited evidence of adverse taphonomic
393 processes within the subsample conforms to deposition in relatively dry and stable
394 paleoenvironmental conditions. In addition to their in-tact or complete nature, surface features of the
395 coprolites indicate that specimens may have been exposed at the surface for a prolonged period pre-
396 burial due to high degrees of abrasion, likely from wind erosion, and the presence of desiccation
397 cracks. Distortion exhibited by flattened ventral surfaces of the coprolites also suggests these rested
398 relatively undisturbed soon after defecation. Desiccation cracks are relatively sparse and are most
399 conspicuous on the larger size Class I specimens. Post-burial diagenesis is similarly limited with
400 minimal evidence of compaction and only minor influence from percolating fluids. Thin botryoidal
401 silica crusts on the surface of internal pores indicate incursion of silica-rich fluids through the porous
402 matrix. Silica is likely derived from adjacent ashfall lapilli tuffs and the coarse fraction (including
403 volcanic glass, quartz, plagioclase, potassium feldspar, and granitic fragments) of the silty mudstone
404 which characterizes the Pipestone Springs strata (Hanneman et al., 2022). Silica infill observed in
405 thin section and SEM data is notably thin, coating pores to a thickness of between ~8–354 μm . The
406 lack of homogeneous silica infill or secondary remobilization of phosphate suggests that between
407 phases of burial and exposure, conditions were predominantly devoid of moisture.

408 The porous nature of the PSMP coprolites is a key morphological attribute that has been
409 overlooked in prior studies. Pores compose the largest percentage of internal structural components
410 after the matrix (Figures 11A and B). Moreover, this represents a minimum estimate of the true
411 porosity, as several pores have been secondarily infilled with silica. As seen in Figure 6, the irregular
412 pores (excluding hair molds) dramatically vary in size from 0.196–50.562 mm and are distributed
413 relatively evenly throughout the matrix when not occupied by skeletal remains. Though pores are
414 known to frequently occur within the matrix of coprolites (Horwitz and Goldberg, 1989, Herbig,
415 1993), their origins remain uncertain, owing to several different processes associated with their
416 formation. One explanation for their formation is decay of some degradable materials within the
417 feces, including smaller bone fragments, soft tissue (muscle, tendons, ligaments, etc.), or insect parts
418 that might not have been digested, and subsequently decayed to form these pores. Given the
419 relatively even distribution of pores throughout the matrix, this seems unlikely. An alternative
420 explanation is that the pores were already present at defecation (including on the surface) caused by
421 trapped gases within the feces—a product of bacterial respiration in the intestine (Herbig, 1993;
422 Magondu, 2021). Gases produced during digestion are typical and varied (Levitt and Bond, 1970);
423 however, reporting the preservation of these gaseous vesicles and their relative abundance within a

424 coprolite is often neglected. As excess gas can relate to bacterial overgrowth within the gut, which
425 may have deleterious consequences for the individual or indicate an intestinal disease (Suarez and
426 Levitt, 2000; Pimentel et al., 2006; Kalantar-Zadeh et al., 2019), the prevalence of such gaseous
427 vesicles may be useful in understanding the gut biome and physiological attributes of coprolite
428 producers. A third explanation for the pores relates to desiccation; as the sample loses moisture,
429 smaller pores (possibly those formed by gases) are enlarged in conjunction with the reduction in
430 overall coprolite volume. As few actualistic studies exist investigating the complete taphonomic
431 processes (from defecation to burial) of modern feces in natural environments, much of this remains
432 speculative (Northwood, 2005; though see Brachaniec et al., 2022).

433 Another constituent of the porous volumes within the coprolites are the moldic remains of
434 keratinous material, specifically hair. Whilst targeted higher resolution scanning was only performed
435 on one specimen (RAM 17540; Figure 7), other coprolites from this sample subset exhibit similar
436 structures. These small tube-like pores generally fall within the shaft shape and size range of hair,
437 with distinct circular cross-sections. Because these pores are small, isolated, and many terminate
438 without contacting the surface of the coprolite, these tubules can readily be distinguished from
439 desiccation cracks, insect burrows or fungal hyphae. Hair molds can also be observed on the surface
440 of select PSMP coprolites (see RAM 17540, Figure 2C). However, not all small pores exhibit this
441 tube-like form and hence hairs cannot account for all the small topographic depressions observed.
442 Due to keratinous material possessing highly resistant molecules that few extant taxa are able to
443 digest (Leprince et al., 1980), hair is relatively resistant to digestive and early lithification processes
444 (Taru and Backwell, 2013). Though the hair eventually decayed, it was preserved long enough to
445 form molds of the shafts within the interior of the coprolite. This pathway conforms with previous
446 studies where hair tubules have been found as casts and impressions both on the surface and interior
447 of coprolites, often preserving exceptional details of the cuticular surface (Crooper et al., 1997; Taru
448 and Backwell, 2013; Bajdek et al., 2016). Similar to the irregular pores, these molds display a
449 siliceous coating caused by secondary infilling during diagenesis.

450

451 **5.2. Insights into Feeding Behavior**

452

453 Qualitative and quantitative analyses of the two coprolite size classes observed reveal notable
454 differences with respect to the morphology of the inclusions, which may inform about feeding habits
455 and physiological differences of their respective producers. Results of our analyses reveal a
456 discernable statistical difference in the relative proportions of bone inclusions to total coprolite
457 volume between Classes I and II. Notably, the proportion of bone volume was greater in the smaller
458 Class II coprolites by comparison to the larger Class I coprolites (Figure 12A). However, there was
459 no significant difference in the extracted bone volumes between the two classes indicating that,
460 despite the relative size of the smaller Class II coprolites, the bone constituent was comparable to that
461 observed in larger Class I coprolites.

462 In the case of Class I coprolites, bones tended to be larger overall with respect to their feret
463 diameters (Figure 12B–D). Bone inclusions within the Class I coprolites also showed more
464 degradation and evidence of intense fragmentation from mastication, such that few bones could be
465 reliably identified. This implies that the Class I producers were capable of consuming larger prey
466 compared to the Class II producers. Bones extracted from the Class I coprolites, including Bone 1 of
467 RAM 17547 (Figure 9K) and Bone 1 of RAM 17517 (Figure 9L), lacked an identifiable shape but

468 revealed internal features such as cancellous tissue associated with larger bones (e.g., pelvic bones,
469 vertebrae, etc.). The ambiguous shapes, incomplete condition of the bones, and preponderance of
470 homogeneous phosphatic matrix contained within Class I coprolites, aligns with specific feeding
471 habits and digestive processes. Principally, the evidence presented herein supports a durophagous
472 carnivorous producer with a bone crushing habit, which is comparable to previous findings in
473 relation to late Miocene carnivoran coprolites from California (Wang et al., 2018). The producers of
474 these latter coprolites are inferred to have been borophagine canids that filled a unique ecological
475 niche in North America, comparable to extant hyenas, until their disappearance approximately 2
476 million years ago (Lofgren et al 2017; for further discussion see section 2.4.2 below). Like the
477 California specimens, the PSMP coprolites also exhibit a powdered homogenous matrix of bone
478 residues (Figure 3A, B) indicative of a producer with a highly acidic gastrointestinal system (Wang et
479 al., 2018). Examples of corrosion are visible in Bone 1 RAM 17547 (Figure 9K) where internal thin
480 trabecular material is revealed. Bone dissolution could also account for the diffuse boundaries
481 exhibited by several bone inclusions during the segmentation process.

482 Class II coprolites overall have a statistically larger proportion of bones compared to total
483 coprolite volume than Class I, though there is notable variation within the sampled dataset (Figure
484 11D). Such variation is also evident in the condition of extracted inclusions. Most identifiable bones
485 in the PSMP material are associated with specimens in Class II, though this is slightly skewed as
486 most of these bones were also recovered from a single specimen, RAM 31212 (Figure 8J). This
487 coprolite has the second largest amount of bone inclusions of those examined here (Supplementary
488 Table S4, 11.28%), while RAM 31211 has the highest overall proportion (Supplementary Table S4,
489 11.64%). Three of the Class II coprolites (RAM 31211, 31212, and 17557) are more densely packed
490 with larger bone inclusions suggesting that the producers were capable of consuming the bones
491 whole, with limited mastication of prey items compared to the producers of the Class I coprolites. In
492 this case, producers of these Class II coprolites were able to extract nutrients with minimal bone-
493 crushing required to consume the prey item. Most likely, this is attributed to the smaller size of the
494 prey item itself for which durophagous mastication was not necessary (Pokines and Tersigni-Tarrant,
495 2012). On the other hand, the Class II coprolites also preserve smaller bones compared to Class I
496 coprolites, as seen by the significantly smaller median values in feret diameters in Class II compared
497 to Class I coprolites (Figure 12B–D). These smaller bones show evidence of intense fragmentation
498 (RAM 31209; Figure 8I and RAM 31214; Figure 8L) which lends support to the capacity for bone-
499 crushing when required for larger prey items, or once the higher-return nutrient-rich organs had been
500 eaten (Pokines and Tersigni-Tarrant, 2012). More direct evidence for a bone-crushing habit is
501 displayed in Bone 1 of RAM 31209 (Figure 10A) which features a clear indent on the flat surface of
502 the bone where the bone has been partially crushed, as well as tooth marks in Bone 6 of RAM 17517
503 (Figure 9E). There is also some tentative evidence to suggest gnawing or bone cracking in Bone 12
504 of RAM 31212 (Figure 9D) exhibited by the somewhat rounded edges on the proximal base plate. It
505 is worth noting that the degree to which stomach acids have caused shrinkage or subsequent breakage
506 of the ingested bone material in the Class II coprolites is difficult to discern and could potentially
507 further account for some reduction in bone fragment size (Fernández-Jalvo et al., 2014). However,
508 the clear boundaries visible between the matrix and the bone material via the CT tomogram slices
509 might attest to the relative resistance of these bone fragments to acid dissolution.

510 In a previous study employing comparable μ CT methods, evidence of an osteophagous diet
511 was inferred from two carnivoran coprolites of discrete sizes from the late Miocene, Spain (Abella et
512 al., 2022). Evidence of osteophagy was based on abundant skeletal inclusions including fragments
513 that appeared to belong to larger bones while others display depressions resembling partial tooth
514 marks. Similar to the Class I PSMP coprolites, the larger specimen of the Spanish coprolites

515 (specimen BAT-3'9.178) preserves irregular bone fragments not identifiable to a specific anatomical
516 bone but does show evidence of digestive corrosion (Abella et al., 2022). The smaller coprolite
517 (BAT-3'10.153), which is comparable in length to the PSMP Class I coprolites exhibits more
518 complete skeletal elements and has a greater proportion of bone inclusions relative to the larger
519 coprolite at the same locality (Abella et al., 2022, figure 4 and 6). Several medium-sized carnivores
520 were suggested as the producer of the smaller coprolite though the most probable was *Protictitherium*
521 *crassum*, a member of Hyaenidae.

522 These findings conform with the borophagine canids interpretation of Lofgren et al. (2017),
523 to the extent that (1) some of the predators consumed whole bones; (2) there was a degree of bone
524 fracturing that occurred, especially in the smaller coprolites; and (3) evidence of digestive corrosion
525 was present.

526

527 **5.3.Identity of the Producer**

528

529 In previous work, Lofgren et al. (2017) hypothesize that the relationship between the diameter of a
530 producer's feces and its body mass, in conjunction with the relative abundance of the respective
531 carnivore species in the PSMP could be applied to determine the likelihood of the producer itself.
532 Several different mammalian predators have been recovered from the PSMP site, belonging to eight
533 different species (Lofgren et al., 2017) including *Hyaenodon microdon* (Mellet, 1977), *Hyaenodon*
534 *crucians* (Leidy, 1853), *Hesperocyon gregarius* (Cope, 1873), *Mustelavus priscus* (Clark, 1936, in
535 Scott and Jepsen, 1936), *Brachyrhynchocyon dodgei* (Scott, 1898), *Parictis montanus* (Clark and
536 Guensburg, 1972), *Daphoenictis tedfordi* (Hunt, 1974), and *Palaeogale sectoria* (Gervais, 1848,
537 1852). After estimating the overall body mass of the predators and their mean prey mass (see Table
538 2), Lofgren et al. (2017) deduced that the range in diameters of larger coprolites may have
539 represented multiple species, concluding that *B. dodgei* was most likely due to the abundance of
540 dentigerous elements in the deposit (77% of the larger species). Similarly, the smaller coprolites
541 could have been produced by *Palaeogale sectoria*, *Parictis montanus*, *Hyaenodon crucians*,
542 *Hyaenodon microdon*, and *Hesperocyon gregarius* (Lofgren et al., 2017). Although the body mass
543 estimates are congruent with *Hy. crucians* as a likely producer, the bone alteration observed in the
544 smaller coprolites is more consistent with extant canids and hence *Hes. gregarius* was inferred as the
545 main producer of the smaller coprolites (Lofgren et al., 2017).

546 [Insert Table 2 here]

547 While the coprolite interior volumes and bone inclusions observed herein did not reveal any
548 unambiguous details as to which taxon might have excreted the coprolites, they did show that at least
549 the Class I producers had aggressive gastrointestinal environments that could digest bone material
550 and cartilage. This is presumably comparable to the bone-crushing habits of modern spotted hyenas
551 (*Crocuta crocuta*), which are also known to consume (and occasionally regurgitate) indigestible
552 materials such as hair (Kruuk, 1972; Di Silvestre et al., 2000). However, as noted by Wang et al.,
553 (2018) in their assessment of the California carnivoran coprolites, there is a paucity of literature
554 investigating gastric pH across broader Carnivora, and hence it is highly speculative to make further
555 inferences as to the identity of the producer based on broad assumptions of a bone-dissolving
556 gastrointestinal system alone.

557

558 **5.4.Evaluation of µCT for Analysis of PSMP Coprolites**

559

560 The value of using µCT as a non-destructive method in extracting information on the internal
561 inclusions of coprolite samples has been well-demonstrated (Qvarnström et al., 2017, 2019).
562 However, specific advantages, limitations, and implicit biases encountered herein are worthy of
563 discussion. We note that while X-ray microscopy can certainly add value to the study of coprolites,
564 results depend on the resolution of the CT scan and the nature of the specimen. In some cases, using
565 µCT alone may not be sufficient to fully describe the internal constituents of bromalites for a
566 particular investigation, and it is recommended to use it in conjunction with other techniques for a
567 more comprehensive analysis. The difficulty of identifying bone inclusions within the PSMP
568 coprolites using µCT to a level beyond generic bone type was not a limitation of µCT itself, but
569 rather a consequence of how much the bones were masticated and digested. Similar results were
570 documented by Abella et al., (2022), wherein they solely used µCT, but found many of the bones
571 were not identifiable to a certain anatomical bone or taxon, and instead the authors relied on
572 describing the general shape and features. Studies comparing the results of mechanical extraction and
573 µCT analysis have encountered similar difficulties. For example, investigations of neolithic midden
574 deposits from Swifterbant, Netherlands, used paired µCT and physical extraction methods to examine
575 phytoliths and bone fragments from finely layered and coprolite-hosting deposits (Huisman et al.,
576 2014). Results showed that many bone features were visible using both methods, though comparably
577 more recognizable bone elements were identified in the sieved material (n=32) compared to µCT
578 (n=7 with confidence, 5 unable to be attributed to species level) (Huisman et al., 2014). The benefit
579 of using µCT in the case of the Swifterbant material, was the ability to provide microstratigraphic
580 context as well as recognize articulated remains and material too small to appear in the sieved
581 fraction (Huisman et al., 2014). A noteworthy finding comparable to observations made in the
582 material herein was that identification of the skeletal components was not dependent on how the
583 bones were extracted, but rather on how completely the bones were preserved (Huisman et al., 2014).
584 Bones with few distinguishable features as a consequence of mastication and digestive processes are
585 going to appear almost identical whether observed virtually or as physically extracted material. In the
586 case of the present study, surface renders of select bones were blurred by ill-defined edges and
587 significant degradation due to the digestive processes which reduced the phase contrast between bone
588 inclusions and the surrounding phosphatic matrix. This further emphasizes that potential limitations
589 on µCT are associated with the degree of preservation of the inclusions and not solely the imaging
590 technique.

591 The primary advantage of using µCT in this study was the ability to reveal volumes, including
592 pores and moldic remains of fossil hair, that would not otherwise be attainable via traditional
593 disaggregation methods. These structures were readily visible in tomographic slices, in both standard
594 and targeted higher resolution scans, and could be extracted to display their morphology as well as
595 their distribution in 3D space (Figure 7). Because hair tubules represent external molds, mechanical
596 or chemical extraction of inclusions would impede the detection of such features at all. Even thin
597 sectioning only provides a very shallow 3D view of these fossil hair follicles and may preclude
598 unambiguous identification of the elongate molds from other potential explanations (i.e., burrows,
599 desiccation cracks). Hair imprints in phosphate aggregates from a Palaeolithic cave site in Western
600 Slovenia have similarly been revealed by µCT methods (Turk et al., 2015). Though the PSMP hair
601 molds could not be attributed to any specific taxon, their prevalence throughout the coprolite may
602 provide insight into the higher taxonomic affiliation to prey items (i.e., mammalian), or alternatively
603 could allude to the affinity of the consumer (e.g., if they ingested their own hair while grooming).

604 Based on analysis of modern vertebrate carnivore scat, the digestibility of hair varies between
605 predator and prey item, however, under most circumstances, it is significantly less digestible
606 compared to bone (Ackerman et al., 1984; Baker et al., 1993; Gamberg and Atkinson, 2016). The
607 presence of hair in feces also can persist for several days after initial consumption depending on the
608 rate of passage through the intestine (Helm, 1984; Kelly and Garton, 1997; Pires et al., 2011).
609 Consequently, we hypothesize that coprolites have a higher likelihood of preserving hair remains
610 when compared to body fossils, and introduce taphonomic bias toward bromalites as a source of
611 information on hair structures and morphology. This supports previous studies that demonstrate the
612 potential of bromalites to facilitate exceptional preservation of soft tissues (Chin et al., 2003;
613 Qvarnstrom et al., 2017; Gordon et al., 2020).

614 Internal porosity and associated characteristics of the pores were overlooked in previous
615 examination of this material (Lofgren et al., 2017) and are easily removed via consumptive sampling
616 methods. Though it is difficult to ascertain whether such original features were considered or
617 overlooked in other coprolite studies (Wang et al., 2018, Romaniuk et al., 2020; Abella et al., 2022),
618 this raises concern for a potential reporting bias in the visualization of segmented volume renders in
619 isolation to tomographic slice data. Ideally, pairing cross-sectional tomographic data directly with
620 volumetric reconstructions (e.g., Turk et al. 2015, figs. 4, 5; Shillito et al., 2020, fig. 3) promotes
621 transparency in documenting coprolite internal composition. Moreover, tomographic slices should be
622 made available upon publication (whether via the journal or repository institution) to facilitate
623 reproducibility of the analysis. Porosity of coprolites is likely an important primary and taphonomic
624 feature. The presence of pores may have broader implications for gut function and health, hence
625 understanding pore morphology, abundance, and distribution is relevant to revealing both
626 palaeobiological data of the producer and the diagenetic history of the overall coprolite specimen.

627

628 **6. Conclusion**

629

630 X-ray tomographic microscopy of twelve coprolites examined herein provides key insights into the
631 diet, feeding behavior, and physiology of the producers from the Miocene Pipestone Spring Main
632 Pocket, Montana. The application of μ CT was a viable alternative to consumptive sampling and was
633 able to differentiate internal inclusions and volumes, including bones and pores, from the overall
634 matrix. Across the two coprolite size classes, several different bone types of varying quality are
635 preserved. Although μ CT methods were able to extract the skeletal inclusions in detail, the condition
636 of the bones as a result of the overall digestive process precluded taxonomic identification and hence
637 limited descriptions to general bone type. A primary benefit to employing μ CT methods is the ability
638 to extract details of internal structures that would be unattainable via consumptive sampling methods.
639 Foremost is the presence of pores and tubular hair molds within the coprolite matrix, which we
640 describe in coprolites of this locality for the first time.

641 The size, shape, and abundance of bone inclusions in conjunction with hair impressions
642 demonstrate these predators consumed vertebrate prey. Depending on the size of the prey, the
643 respective producers of the coprolite size classes may have adapted different feeding habits to
644 maximize the return on nutrients. The larger coprolites contained comparatively fewer complete
645 bones but were highly fragmented suggesting bone-crushing habits. Alternatively, the variability in
646 the smaller coprolites containing both complete bone morphologies in greater proportions as well as
647 fragmented bone implies both whole consumption and bone-crushing behavior.

648 Overall, application of µCT methodologies will allow for further standardization of coprolite
649 studies. With the growth of virtual paleontology, an opportunity has presented itself for the expansion
650 of coprolite research and the integration of novel qualitative and quantitative analyses. Where
651 coprolite samples were once damaged or destroyed via traditional methods, µCT provides a valuable
652 means to preemptively survey specimens to target regions of interest that maximize data return whilst
653 minimizing consumptive sampling for thin sectioning, SEM and elemental analyses. Within the
654 sample subset described here, certain coprolites possess minimal inclusions and consequently would
655 offer a limited view of the internal heterogeneity present within the sample population. Moreover, it
656 is desirable that prior to destructive sampling, a virtual record of the coprolite is retained for
657 reference. To this end, µCT may be instrumental in the long-term preservation and curation of
658 coprolite samples, and simultaneously promote sharing 3D datasets via online repositories.
659 Ultimately, this facilitates opportunities to share qualitative and quantitative µCT datasets via digital
660 repositories. Such methodologies enable greater standardization of methods and allow for more
661 comparative analyses to expand the field of coprolite research.

662

663 **7. Conflict of Interest**

664 The authors declare that the research was conducted in the absence of any commercial or financial
665 relationships that could be construed as a potential conflict of interest.

666 **8. Author Contributions**

667 SMJ: Conceptualization, Methodology, Supervision, Writing – Review and Editing, Investigation,
668 Resources, Visualization, Project administration, Data Curation, Funding acquisition. JLW: Writing
669 – Original Draft, Software, Investigation, Visualization. JWH: Formal analysis, Visualization,
670 Editing. TS: Visualization, Funding acquisition. JDS: Visualization, Resources, Funding acquisition.
671 All authors contributed toward edits to the final submitted manuscript.

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848 **12. Supplementary Material**

849 The Supplementary Material for this article can be found online at: XXXXXXXXXXXX.

850 **12 Data Availability Statement**

851 The datasets generated for this study can be found in the online 3D repository Morphosource
 852 <https://www.morphosource.org/>

853

854 **TABLE 1.** Coprolite dimensions and weight arranged by size class and mass (g).

Class	Sample Name (RAM)	Length (mm)	Width (mm)	Mass (g)
Class I	17517	50.69	24.11	33.22
Class I	17540	45.94	23.19	27.75
Class I	17405	44.81	22.15	22.00
Class I	17547	28.89	28.90	19.19
Class I	17546	42.50	20.55	17.65

Class I	17370	39.97	21.56	15.97
Class I	18171	64.36	24.03	39.14
Class II	17557	26.14	11.05	3.61
Class II	31214	24.03	10.71	3.27
Class II	31312	26.30	10.63	2.57
Class II	31211	22.65	9.91	2.08
Class II	31209	14.38	11.26	1.68

855

856 **TABLE 2.** Summary of the main carnivorous species found in the PSMP. Data derived from
 857 LaGarry (2004)^a; Lofgren et al. (2017)^b; Christison et al. (2022)^c .

858

Taxon	Family	Body Mass (kg)	Mean prey mass (kg)
<i>Hyaenodon microdon</i>	<i>Hyaenodontidae</i>	1.6 ^b / 27.2 ^c	49.1 ^b
<i>Hyaenodon crucians</i>	<i>Hyaenodontidae</i>	~17 ^a / 3.3 ^b	NA
<i>Hesperocyon gregarius</i>	<i>Canidae</i>	3 ^a / ~2.9 ^b / 3.5 ^c	6.4 ^b
<i>Mustelavus priscus</i>	<i>Mustelidae</i>	1.9 ^b	NA
<i>Brachyrhynchocyon dodgei</i>	<i>Amphicyonidae</i>	13.9 ^c	25.1 ^b
<i>Daphoenictis tedfordi</i>	<i>Amphicyonidae</i>	NA	NA
<i>Parictis montanus</i>	<i>Ursidae</i>	2.5 ^b	NA
<i>Palaeogale sectoria</i>	<i>Palaeogalidae</i>	1.7 ^b	NA

859

860 **FIGURE 1.** Coprolites arranged by mass (g) into Class I (A–G) and Class II (H–L). **A.** RAM 18171.
 861 **B.** RAM 17517. **C.** RAM 17540. **D.** RAM 17405. **E.** RAM 17547. **F.** RAM 17546. **G.** RAM 17370.
 862 **H.** RAM 17557. **I.** RAM 31214 **J.** RAM 31212 **K.** RAM 31211 **L.** RAM 31209.

863 **FIGURE 2.** Taphonomic and morphological features observed on the surface of coprolite samples. **A.**
 864 Examples of small desiccation cracks (RAM 17517). **B.** Shows the surface scratches that have no
 865 consistent length or pattern to them (RAM 17517). **C.** Hair molds (RAM 17540). **D.** Large surface pore
 866 and likely mold (RAM 17546). Red boxes in overview images represent corresponding zoomed-in
 867 images. Scale bars represent 1 mm; insets all 10 mm.

868 **FIGURE 3.** Photomicrographs and backscattered electron images of interior composition of
 869 specimen RAM 17546. **A.** Phosphate enriched inclusion embedded within matrix of coprolite. **B.**
 870 Contact between two fabrics of matrix in the interior of the coprolite. **C.** Backscattered electron
 871 image of 3D pore space showing botryoidal silica crust. **D.** Pore spaces lined with silica crust and
 872 bone inclusions. **E.** Contact (dashed line) between coprolite matrix and adhered layer of sediment. **F.**
 873 Transverse (white arrows) and longitudinal (black arrows) cross sections of hair molds. B= bone;
 874 P=pore; C=crust; M=matrix; S=sediment; R=resin. Scale bars represent 250 μ m.

875 **FIGURE 4.** Internal and compositional characteristics of RAM 17540 **A.** Plane view of the
 876 unpolished sectioned surface under plain light. **B.** Backscatter electron image of the same surface
 877 figured in A. **C.** Back-scatter electron image of polished surface of sectioned sample. Insets shown in
 878 Figure 5. Scale bars represent 5 mm.

879 **FIGURE 5.** EDS elemental maps of areas in specimen RAM 17546 from Figure 4. **A.** Well-
880 preserved bone inclusion and surrounding matrix. **B.** Exterior edge of the coprolite showing sediment
881 adhered to the outer surface. **C.** Etched bone inclusions with diffuse edges. **D.** Matrix and hair molds.
882 **E.** Etched bone inclusions with diffuse edges. Scale bars represent 1 mm.

883 **FIGURE 6.** Internal view of pores of Class I Coprolites (A–G) and Class II (H–J). Pores are colored
884 brown while the external surface of the coprolite is represented by light blue. **A.** RAM 18171. **B.**
885 RAM 17517. **C.** RAM 17540. **D.** RAM 17405. **E.** RAM 17547. **F.** RAM 17546. **G.** RAM 17370. **H.**
886 RAM 17557. **I.** RAM 31214 **J.** RAM 31212 **K.** RAM 31211 **L.** RAM 31209. Scale bars represent 10
887 mm.

888 **FIGURE 7.** Hair molds in RAM 17540. **A.** Site of targeted ROI for high resolution scan. **B.** Volume
889 render of targeted region showing a subset of the hair molds within the volume. **C–D.** Extracted molds
890 using multi-ROI to visualize individual isolated (C) and connected (D) components. Scale bars
891 represent 10 mm.

892 **FIGURE 8.** Internal view of coprolite bone inclusions divided into size Class I (A–G) and Class II
893 (H–L). Bones of interest labeled. Scale bars 10mm.

894 **FIGURE 9.** Extracted bones with identifiable features. **A.** Bone 1 of RAM 31212 seen from four
895 different angles. Shows the condyle of a long bone, or capitulum (e.g., femoral head) . **B.** Bone 2 of
896 RAM 17517 seen from three angles showing the zygomatic arch. **C.** Bone 5 of RAM 31212 seen from
897 two angles, showing acetabulum (socket) of a hipbone. **D.** Bone 12 of RAM 31212 seen from three
898 different angles, showing potential elements of a pelvic girdle (ischium, or base plate and iliac blade)
899 related to C. **E.** Bone 6 of RAM 17517 seen from three angles. Unlikely to be mammal material;
900 unbroken edge and the overall geometry suggest this could be a squamate opisthotic. **F.** Bone 10 of
901 RAM 31212 seen from two views. Bone identified as a phalange. **G.** Bone 11 RAM 31212 of RAM
902 31212 seen from two views. Bone identified as a phalange. **H–I.** Bone 4 of RAM 17557 and Bone 3
903 of RAM 17517 each from two views. Potentially ribs; both are flat, hollow short bones with triangular
904 cross-section. **J.** Bone 5 of RAM 17517 seen in two views. Rib, based on triangular cross-section. **K–**
905 **L.** Bone 1 of RAM 17547 and Bone 1 of 17517 show the cancellous structures of the bones contained
906 within the interior. Scale bars represent 1 mm.

907 **FIGURE 10.** Extracted bones based on type. **A–H.** Flat bones. **A.** Plane and oblique view of Bone 1
908 in RAM 31209. **B.** Plane views of opposite sides to Bone 5 in RAM 31211. **C.** Plane and side views of
909 Bone 1 of RAM 17405; possible ilium. **D.** Plane and side views of Bone 1 in RAM 18171; possible
910 cranial element. **E–F.** Oblique and plane views of opposite sides to Bone 1 and 3 in RAM 31214. **G.**
911 Opposing flattened sides of Bone 2 in RAM 18171; resembles fibula. **H.** Opposing flattened sides in
912 Bone 1 of RAM 17370. **I–K.** Irregular bones. **I.** Patella in anterior, lateral, and posterior views of Bone
913 6 of RAM 31212. **J.** Plane view of Bone 2 of RAM 17405. **K.** Plane and side views of Bone 3 of RAM
914 17557; possible lower jaw joint. Scale bars represent 1 mm.

915 **FIGURE 11.** Comparison of coprolite shape in addition to volume and proportion of internal
916 components across classes. **A.** Volumetric analysis of all coprolites, comparing total volume of matrix,
917 bones and pores. **B.** Volumetric analysis of all coprolites, comparing total percentage of matrix, bones
918 and pores. **C.** A x-y plot of coprolite samples length vs. width. The size of the dots indicates the mass
919 of the coprolite specimen. **D.** The proportion of pore and bone material per coprolite specimen between
920 the two classes. Median pore proportion in Class I is 0.054 (95% CI: 0.015, 0.155); and for Class II is
921 0.082 (95% CI: 0.007, 0.225). Median skeletal proportion in Class I is 0.005 (95% CI: 0.004, 0.024);
922 and for Class II is 0.055 (95% CI: 0.044, 0.172).

923 **FIGURE 12.** Comparison of bone volume and feret diameters across coprolite classes. **A.** Median
924 difference between Class I and Class II of bone volumes (Class I=0.1m³ and Class II=0.09mm³). **B.**

925 Median difference between Class I and Class II maximum feret diameters of bones for coprolites (Class
926 I= 1.575 mm, Class II= 1.25 mm, $p=0.001863$). **C.** Median difference between Class I and Class II
927 mean feret diameters of bones for coprolites (Class I= 1.015 mm, Class II= 0.87 mm, $p=0.0006583$).
928 **D.** Median difference between Class I and Class II minimum feret diameters of bones for coprolites
929 (Class I= 0.495 mm, Class II= 0.460 mm, $p=0.004129$).