



Rare yet everywhere: phylogenetic position of the enigmatic deep-sea shrimp *Physetocaris microphthalmalma* Chace, 1940 (Decapoda, Caridea)

Pedro A. Peres^{A,*}  and Heather Bracken-Grissom^{A,B}

For full list of author affiliations and declarations see end of paper

***Correspondence to:**

Pedro A. Peres
Institute of Environment and Department of Biology, Florida International University (FIU), Miami, FL, USA
Email: pedro.peres27@gmail.com

Handling Editor:
Shane Ahyong

ABSTRACT

The mysterious deep-sea shrimp *Physetocaris microphthalmalma* Chace, 1940 remains a challenge for the understanding of caridean shrimp systematics. Upon first description in 1940, the unique morphology in combination with lack of material made the allocation of *P. microphthalmalma* to any family or superfamily difficult, therefore the monotypic superfamily *Physetocaridoidea* and family *Physetocarididae* were described. The rarity of the species, only documented a few times in scientific literature, in combination with a circumglobal distribution, makes the advancement of the systematics and biology of this shrimp challenging. Current literature places *Physetocaridoidea* as a superfamily with a sister relationship to *Pandaloidea* but this relationship has never been tested using molecular data. Recent expeditions to the northern Gulf of Mexico and north-eastern Pacific Ocean provided fresh material for inclusion in phylogenetic analyses. Here, we used a molecular systematics approach to investigate the phylogenetic placement of this species within the infraorder Caridea and test for cryptic diversity across oceanic basins. We sequenced five genes (*12S rRNA*, *16S rRNA*, *H3*, *NaK* and *PEPCK*) and built phylogenetic trees including specimens across *Pandaloidea* and other carideans ($n = 75$) using maximum-likelihood and Bayesian approaches. Our results strongly support the inclusion of *P. microphthalmalma* within the family *Pandalidae* and superfamily *Pandaloidea*, indicating that the superfamily *Physetocaridoidea* and family *Physetocarididae* are not valid. In addition, the inclusion of specimens from the Atlantic and Pacific Oceans does not support evidence of cryptic diversity, suggesting the global distribution of *P. microphthalmalma*. This is the first study to provide genetic data for this species, resulting in an updated classification for the infraorder Caridea and highlighting that deep-pelagic species can be rare yet still widely distributed.

Keywords: Caridea, deep-sea, Pandalidae, Pandaloidea, pelagic, phylogenetics, *Physetocaris*, shrimp, systematics.

Introduction

The deep sea (> 200 m) is the largest habitat on Earth yet is poorly known and explored (Ramirez-Llodra *et al.* 2011; Sinniger *et al.* 2016; Sutton *et al.* 2017). Below the epipelagic zone (> 200 m), light becomes dim, as animals transition from the photic zone (0–200 m) into the mesopelagic or twilight zone (200–1000 m) until light is completely lost and darkness dominates (bathypelagic or midnight zone, 1000–4000 m; abyssopelagic zone, 4000–6000 m; hadalpelagic zone, > 6000 m). Animals within the deep sea have adapted to a life of darkness, resulting in a suite of unique morphological adaptations often compared to science fiction monsters or space creatures. Within these waters, the deep-pelagic zone is no exception, containing many fascinating organisms only collected a few times or new to science (Webb *et al.* 2010).

Exploration of the deep sea and associated diversity is challenging due to financial and logistical constraints. However, in recent years, several deep-sea species have been described from meso- and bathypelagic environments (Pietsch and Sutton 2015; Varela and Bracken-Grissom 2021a; Judkins *et al.* 2022), and the use of genetic techniques

Received: 16 May 2023
Accepted: 12 July 2023
Published: 2 August 2023

Cite this:
Peres PA and Bracken-Grissom H (2023)
Invertebrate Systematics
37(8), 529–537. doi:10.1071/IS23024

© 2023 The Author(s) (or their employer(s)). Published by
CSIRO Publishing.

has significantly enhanced our understanding of evolutionary relationships and biodiversity (e.g. Tsang *et al.* 2009; Davis *et al.* 2014; Eilertsen and Malaquias 2015; Sinniger *et al.* 2016; Varela *et al.* 2021; Rodríguez-Flores *et al.* 2022). One of these mysterious taxa is the enigmatic *Physetocaris microphthalma* Chace, 1940 (Fig. 1), a small (max. carapace length = 16 mm) meso- and bathypelagic shrimp from the Infraorder Caridea. This species was originally described from Bermuda in 1940 and since discovery, *P. microphthalma* has only been collected eight other times according to scientific records. This species has a large distribution ranging from the south-western and north-eastern Pacific to the Eastern and North Atlantic oceans (Fig. 2), and a depth range of 200 m to over 2000 m within the pelagic environment. Diagnostic characters include two lateral carinae, a pleon without any dorsal carinae or spines, the second pereopod with a fixed finger curving subrectangularly around a short, broad movable finger, a greatly reduced maxilla and second maxilliped, and a telson deeply sulcate dorsally and broadly truncate at the tip (Chace 1940, 1992). Other distinguishing characteristics include an inflated rostrum, small eyes and overall body morphology

unlike any other caridean shrimp (pers. observation). This combination of characters was so unique that the original description states that,

It has been impossible to accommodate it in any known caridean family and even its relative position among the established families is uncertain. There is little doubt that it is one of the most specialized bathypelagic carideans known [Chace 1940, pp. 198–199].

The original description noted affinities of *P. microphthalma* to Processidae and Crangonidae, mentioned the lack of adult males and ovigerous females and speculated that the specimen was an undescribed larval form (Chace 1940). Owing to the rarity in nature and lack of material for study, the systematic position of *P. microphthalma* is under debate and remains a mystery.

Early classification grouped the families Phystocarididae, Pandalidae and Thalassocarididae into the superfamily Pandaloidea based on the chelae of first pair of pereiopods being microscopically small or absent and diagnosed



Fig. 1. *Physetocaris microphthalma* Chace, 1940 collected in the Gulf of Mexico (DP07) and used for phylogenetic analyses (HBG10751). Photo: Danté Fenolio|DEEPEND|RESTORE.

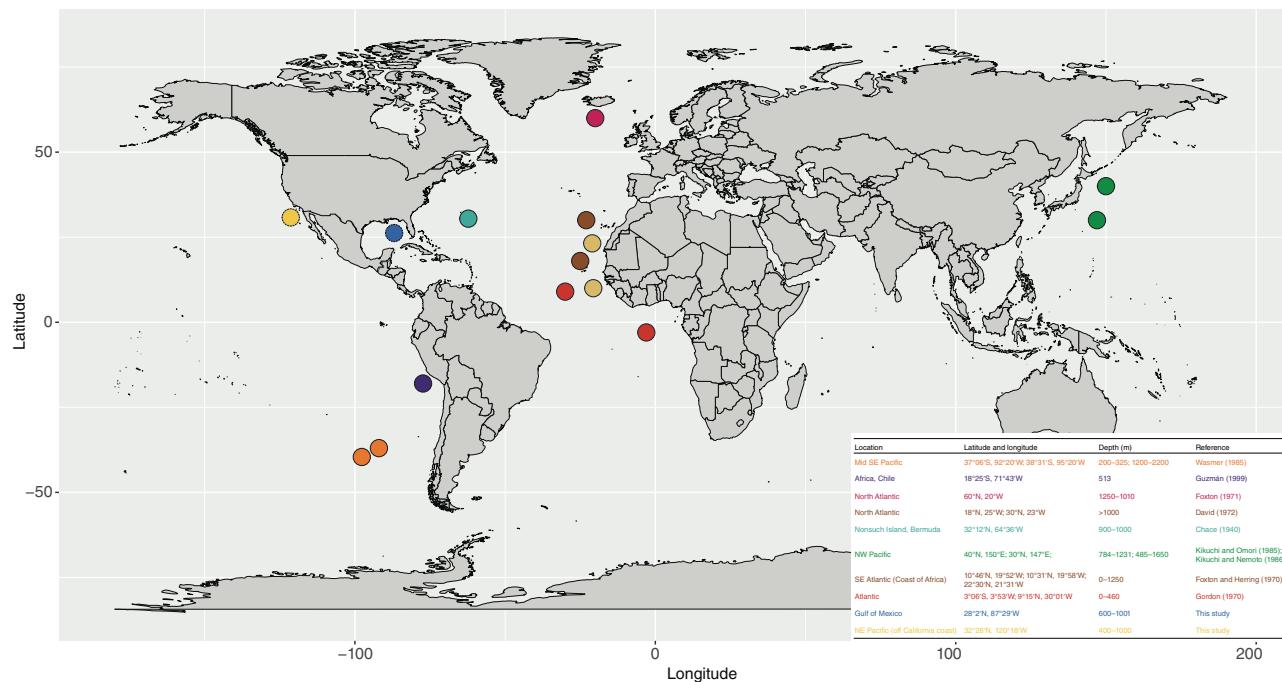


Fig. 2. Compilation of all known records of *Phisetocaris microphthalmalma* Chace, 1940. Full circles represent previous records, dashed circle represent the new records from this study (Gulf of Mexico and Northeast Pacific). Sources: Chace (1940); Foxton and Herring (1970); Gordon (1970); Foxton (1971); David (1972); Kikuchi and Omori (1985); Wasmer (1985); Kikuchi and Nemoto (1986); Guzmán (1999).

Physetocarididae by the inflated carapace and simple mandible with palp absent (Holthuis 1955). Later, the superfamily Physetocaridoidea, containing only Physetocarididae, was coined with no morphological description (Bowman and Abele 1982). The use of the superfamily rank Physetocaridoidea was probably accepted based on unique traits of the species and used in further publications even with a lack of morphological investigation and description that included characters that distinguished this from all other caridean groups (Chace 1992; Holthuis 1993). Since then, few studies have tried to solve this systematic and taxonomic conundrum. A phylogenetic hypothesis based on morphological data suggested the synonymisation of Physetocaridoidea with Pandaloidea because results indicated that *P. microphthalmalma* fell within the Pandaloidea (Christoffersen 1989). Although Christoffersen (1989) suggested synonymisation of the superfamily, the family Physetocarididae was kept within Pandaloidea. The same work also suggests that other genera should be included within Physetocarididae: *Stylopandalus*, *Chlorotocella*, *Chlorocurtis*, *Anachlorocurtis* and *Miropandalus* based on morphological data (Christoffersen 1989). However, in this analysis, Physetocarididae is represented by character reversal or loss of traits and that is likely the reason why this phylogenetic hypothesis was not totally accepted.

Advancements in molecular methods have helped to elucidate the phylogenetic position and classification of mysterious decapod species (Bracken *et al.* 2009, 2010; Bracken-Grissom *et al.* 2012; Wang *et al.* 2021). One group

in which molecular systematic methods were employed was the Pandaloidea, a superfamily that might include Physetocarididae or that is sister to Physetocaridoidea (Liao *et al.* 2019). These authors hypothesised that *P. microphthalmalma* could be part of the new family Chlorotocellidae, within Pandaloidea and sister to Pandalidae (Clade A in Komai *et al.* 2019; Liao *et al.* 2019). Once again, due to the rarity of *P. microphthalmalma*, this species was not included in the study or any other past molecular phylogenies.

Molecular methods are also useful in the detection of cryptic species complexes and the rarity of *P. microphthalmalma* coupled with the broad distribution (Fig. 2) could indicate undetected diversity. Although many deep-sea crustaceans are considered to represent a single panmictic population with distributions spanning oceanic basins (Bik *et al.* 2010; Havermans *et al.* 2013), DNA-based approaches have revealed surprising levels of genetic structure or cryptic speciation (Miyamoto *et al.* 2010; Baco *et al.* 2016; Varela *et al.* 2021). Studies that sample across global distributions are challenging due to financial and logistical constraints, therefore increased efforts are needed to investigate phylogenetic relationships and connectivity patterns in deep sea taxa, especially rare species.

Here, we provide the first molecular phylogenetic investigation of *Phisetocaris microphthalmalma* Chace, 1940 to resolve the phylogenetic placement of this enigmatic deep-sea species. As mentioned, this species is historically under debate but the most comprehensive checklist of all shrimp groups

follows the classification that considers the monotypic species *P. microphthalmalma* part of the family Phycetocarididae within the superfamily Phycetocardoidea (De Grave and Fransen 2011; Poore and Ahyong 2023; World Register of Marine Species, see <https://www.marinespecies.org>). We tested two alternative hypotheses: (1) the reciprocal monophyly of the superfamily Phycetocardoidea and Pandaloidea (*sensu* Liao *et al.* (2019) and Komai *et al.* (2019), [Pandalidae + 'Thalassocaridae'] + Chlorotocellidae) that would agree with the recent classification of the species (De Grave and Fransen 2011; Poore and Ahyong 2023); (2) *P. microphthalmalma* nested within Chlorotocellidae (clade A – Liao *et al.* 2019) and within Pandaloidea. Finally, considering that we had one individual from the Gulf of Mexico and one from the north-eastern Pacific, we tested the alternative hypotheses that (3) there is cryptic diversity within *P. microphthalmalma*; and (4) this is a circumglobally distributed species. Our work expands the records of *P. microphthalmalma* to the Gulf of Mexico and north-eastern Pacific, reveals the phylogenetic placement of this enigmatic shrimp and provides data that can support an updated classification for Infraorder Caridea.

Materials and methods

Sample collection

Specimens of *Phycetocaris microphthalmalma* were collected during research expeditions in the Gulf of Mexico and Northeast Pacific (California, USA). The research cruises in the Gulf of Mexico were on the R/V *Point Sur* as part of the DEEPEND|RESTORE consortium (<http://www.deependconsortium.org>) and the Northeast Pacific sample was donated by Dr Anela Choy from Scripps Institution of Oceanography, University of California—San Diego (San Diego, CA, USA). The Gulf of Mexico sample was collected in May 2021 (DP07) and the Northeast Pacific sample in June 2021 with a multiple opening-closing net and environmental sensing system (MOC-10) rigged with six 3-mm mesh nets ranging from 0- to 1500-m depth, allowing for collected specimens to be assigned to a depth bin (0–200, 200–600, 600–1000, 1000–1200 and 1200–1500 m; the sixth net sampled from 0 to 1500 m). *Phycetocaris microphthalmalma* from the Gulf of Mexico was collected in the 600–1000-m bin and north-eastern Pacific in the 400–1000-m bin. Animals were preserved in 80% ethanol and later identified in the laboratory. Both specimens are deposited in the Florida International University Crustacean Collection (FICC; voucher numbers, HBG10571 and HBG11495).

DNA extraction, PCR amplification and sequencing

Total genomic DNA was extracted from pleon muscle tissue using DNeasy Blood and Tissue Kit (Qiagen). We selected two mitochondrial and three nuclear genes to perform the

phylogenetic analysis. The genes selected were the large ribosomal subunit (16S rRNA), small ribosomal subunit (12S rRNA), histone 3 (H3), sodium–potassium ATPase α -subunit (*NaK*) and phosphoenolpyruvate carboxykinase (*PEPCK*). These genes are well suited to investigate phylogenetic relationships in decapods (Timm and Bracken-Grissom 2015) and have been used to investigate the evolutionary relationships in Pandaloidea (Liao *et al.* 2019). Primers used were the same as indicated in Liao *et al.* (2019). PCR amplification contained 12.5 μ L of GoTaq, 8.5 μ L of water, 1 μ L of each primer (10 μ M), and 2 μ L of DNA per sample, representing 25 μ L of total PCR reaction. The thermal profile was: 3 min at 94°C for initial denaturation; 35 cycles of 30 s at 94°C, 30 s at the primer-specific temperature (45–64°C) and 1 min at 72°C; and ending in 5 min at 72°C for final extension. PCR products were purified and sequenced at TACGen (Richmond, CA, USA) using the BigDye Terminator Cycle Sequencing Kit (ver. 3.1, Applied Biosystems) and sequenced in a 3730xl DNA Analyzer (Applied Biosystems). *Phycetocaris microphthalmalma* sequences were quality-checked, trimmed and assembled using Geneious Prime software (ver. 2020.0.4, see <https://www.geneious.com/>). Protein coding genes were translated and examined for indels and stop codons to ensure that pseudogenes were not included (Song *et al.* 2008).

Phylogenetic analysis

We selected all Pandalidae species ($n = 62$) and outgroups (Alpheidae, Palaemonidae, Hippolytidae, Lysmatidae, Mergidae, Crangonidae and Glyphocrangonidae; $n = 11$) included in Liao *et al.* (2019), plus the two individuals of *P. microphthalmalma* (total $n = 75$). We retrieved all five genes for the species selected when these were available from GenBank (Supplementary Table S1). We aligned the sequences using MAFFT 7 (ver. 7.490, see <https://mafft.cbrc.jp/alignment/software/>; Katoh and Standley 2013) and gene alignments were concatenated using Geneious Prime software (ver. 2020.0.4).

The IQ-TREE 2 program (ver. 2.2.0.5, see <http://www.iqtree.org/>; Kalyaanamoorthy *et al.* 2017; Minh *et al.* 2020) was used to construct a phylogenetic hypothesis using a Maximum Likelihood (ML) approach. The best model of evolution and partitioning scheme was chosen based on the Bayesian Information Criterion (BIC) estimated by the ModelFinder implemented within the IQ-TREE (see <http://www.iqtree.org/ModelFinder/>). The support of branches was estimated through the ultrafast bootstrapping (UFboot, Hoang *et al.* 2018) and SH-like approximate likelihood ratio test (SH-aLRT, Guindon *et al.* 2010) methods with 1000 replications.

The MrBayes program (ver. 3.2.6, see <https://github.com/NBISweden/MrBayes/>; Ronquist *et al.* 2012) was used to construct a phylogenetic hypothesis using a Bayesian (BY) approach. Two independent runs were performed using the

partitioning scheme and substitution models selected by ModelFinder. When the selected model was not available in MrBayes, the next more complex model was selected following the MrBayes manual. Both runs contained four chains and the Markov Chain Monte Carlo (MCMC) algorithm ran for 10 000 000 generations, sampling every 1000 generations with a burn-in set to 25%. Convergence was assumed when the average standard deviation of split frequencies was below 0.01. Branch support was estimated through posterior probabilities (pp) computed on the 50% majority rule tree (consensus tree).

Phylogenetic analyses were run on the Florida International University High-Performance Computing Cluster (HPCC).

Genetic distance

Pairwise genetic distance analysis was performed to investigate intraspecific diversity within *P. microphthalmalma*.

We used the p-distance method to calculate genetic distance using the molecular marker 16S rRNA using MEGA11 (ver. 11.0.13, see <https://www.megasoftware.net>; Tamura *et al.* 2021) that was selected because this is one of the commonly used barcode genes in decapods (Varela *et al.* 2021).

Results

Our paper provides the first genetic sequences generated for *P. microphthalmalma*. In total, we sequenced five sequences per individual. Our analyses included 74 sequences of the marker 12S rRNA, 75 of 16S rRNA, 73 of H3, 73 of NaK and 71 of PEPCK (Supplementary Table S1). Topologies derived from the ML and BY analyses were similar (Fig. 3). Our main goal was to investigate the phylogenetic placement of *P. microphthalmalma*, therefore we will not discuss evolutionary relationships beyond our focal taxa

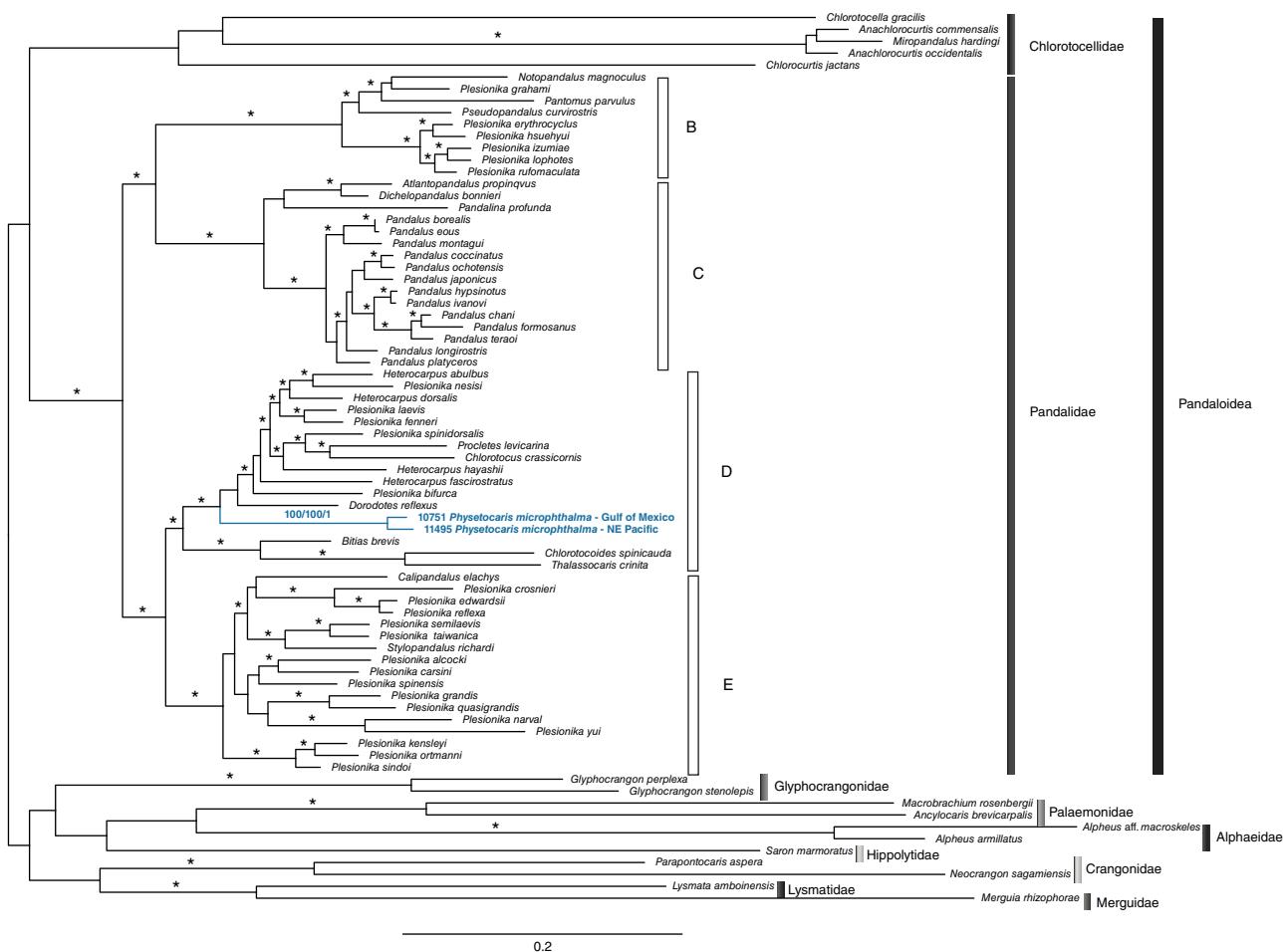


Fig. 3. Phylogenetic hypothesis for the placement of *Phyetocaris microphthalmalma* Chace, 1940 within the infraorder Caridea, with a focus on Pandaloidea. The phylogram is based on the genes 12S rRNA, 16S rRNA, H3, NaK and PEPCK, and represents a combination of the Maximum-likelihood (ML) and Bayesian (BY) trees. Branch support values are indicated only for the *P. microphthalmalma* clade (SH-aLRT/UFboots/pp). For all other branches, support is represented by an asterisk (*) when all metrics are significant (SH-aLRT > 80/UFboots > 95/pp > 1). Branch support is not indicated if any of the metrics are below the threshold indicated. Clades B, C, D and E follow Liao *et al.* (2019) notation. Clade A (*sensu* Liao *et al.* 2019) is represented as Chlorotocellidae.

(but see Liao *et al.* (2019) for a discussion of evolutionary relationships across genera). Our phylogenetic hypotheses show *P. microphthalmalma* nested within Pandalidae, refuting hypothesis 1 (monophyly of the superfamily Phycetocaridoidea and Pandaloidea). Our phylogenetic hypothesis also refutes hypothesis 2 (*P. microphthalmalma* nested within Chlorotocellidae and within Pandaloidea). *Phycetocaris microphthalmalma* examined from the Gulf of Mexico and from the Northeast Pacific were recovered in the same clade showing short branch lengths (SH-aLRT, 100; UFboot, 100; pp, 1) and low genetic distance (16S rRNA, 0.173%) indicating that both represent the same species. Both specimens formed a clade with *Dorodotes reflexus*, *Heterocarpus dorsalis*, *Plesionika bifurca*, *Plesionika fenneri*, *Plesionika laevis*, *Plesionika nesisi*, *Heterocarpus abulbus*, *Heterocarpus fascirostratus*, *Heterocarpus hayashii*, *Plesionika spinidorsalis*, *Chlorotocus crassicornis* and *Proctetes levicarina* (SH-aLRT, 100; UFboot, 100; pp, 1). This clade represents clade D recovered by Liao *et al.* (2019).

We provide evidence against our prediction 3 (there is cryptic diversity within *P. microphthalmalma*) and confirm our prediction 4 (this is a circumglobally distributed species). Therefore, our study also provides two new records for *P. microphthalmalma*, extending the range of the species to the Gulf of Mexico and north-eastern Pacific (Fig. 2).

Discussion

Based on phylogenetic analyses, our results show that the rare and enigmatic deep-sea shrimp *Phycetocaris microphthalmalma* is nested within the caridean family Pandalidae. This outcome challenges the current taxonomic classification of this species and indicates that the superfamily Phycetocaridoidea and family Phycetocarididae should not be accepted. Therefore, the genus *Phycetocaris* should be transferred to the superfamily Pandaloidea and family Pandalidae, being *P. microphthalmalma* part of a monotypic genus. The inclusion of material from the Gulf of Mexico and north-eastern Pacific also indicates that *P. microphthalmalma* represents a single species with a very broad geographic distribution; however, the inclusion of material from across the entire distribution would confirm these findings.

Phylogenetic placement of the rare deep-sea shrimp *Phycetocaris microphthalmalma*

Some of the earliest morphological studies suggested *P. microphthalmalma* to be nested within Pandaloidea (Christoffersen 1989). The most comprehensive molecular tree of pandalids (Liao *et al.* 2019) used eight molecular markers and examined the relationship between Pandalidae and Thalassocarididae, the two families in Pandaloidea but these lacked samples of *P. microphthalmalma* (Liao *et al.* 2019). The authors suggested that *P. microphthalmalma* would fall within the newly designated

Clade A (*Chlorotocella*, *Chlorocurtis*, *Anachlorocurtis* and *Miropandalus*) (Liao *et al.* 2019). Later, Clade A was established as the family Chlorotocellidae in a follow-up study (Komai *et al.* 2019). The inclusion of *P. microphthalmalma* in Chlorotocellidae (Clade A) was based on the argument that this same clade had been recovered in a previous morphological phylogeny (Christoffersen 1989). *Phycetocaris microphthalmalma* is unlikely to be part of Chlorotocellidae (Clade A). In the morphological study (Christoffersen 1989), the clade recovered as Phycetocarididae (= *Stylopandalus* (synonymised to *Plesionika*) + Chlorotocellidae genera + *Phycetocaris*) is supported by synapomorphies represented by characters reversion or acquisition of a lost trait casting doubts if this is a feasible grouping. The character that supports Phycetocarididae (*sensu* Christoffersen 1989) is the absence of palp, an apomorphic character of *Phycetocaris* but not a synapomorphy of Chlorotocellidae (Christoffersen 1989; Komai *et al.* 2019), casting doubt on the inclusion of *Phycetocaris* within Chlorotocellidae. When comparing the morphological traits defining Phycetocarididae (*sensu* Christoffersen 1989) and Clade A (Liao *et al.* 2019), we find no shared characters. Most characters recovered by the ancestral state reconstruction of Clade A (Liao *et al.* 2019) are not characteristic of *P. microphthalmalma* because of this enigmatic shrimp's unique features. Another argument against *P. microphthalmalma* being part of Chlorotocellidae is that this family is represented by mostly shallow-water species while *P. microphthalmalma* is found within the deep-pelagic environment. In short, multiple lines of evidence question the position of *P. microphthalmalma* within Chlorotocellidae.

Our study was the first to include *P. microphthalmalma* into a molecular phylogeny of Pandaloidea and both specimens are strongly supported within a clade including the genera *Dorodotes*, *Chlorotocus*, *Proctetes*, some *Heterocarpus* and some *Plesionika*. This grouping is supported from a morphological perspective because *P. microphthalmalma* contains a long postrostral carina found to be a synapomorphy uniting this group (Liao *et al.* 2019). The fact that all genera within this clade represent deep-sea pelagic species provides further evidence of affinity. Although we have strong molecular evidence to support *Phycetocaris* + *Dorodotes*, *Chlorotocus* + *Proctetes* + some *Heterocarpus*, and + some *Plesionika* many genera within Pandalidae are not recovered as monophyletic, indicating that the whole family needs further investigation (Liao *et al.* 2019) and substantial revision. Pandalid systematics are highly convoluted due to high morphological disparity and biodiversity. Recently, the superfamily Pandaloidea was rearranged with the synonymisation of the family Thalassocaridae to Pandalidae (Liao *et al.* 2019) and the creation of the new family Chlorotocellidae (Komai *et al.* 2019). Similarly, our results suggest the synonymisation of Phycetocarididae and Phycetocaridoidea with Pandalidae and Pandaloidea, respectively. Based on recent changes at the family-level (Komai *et al.* 2019; Liao *et al.* 2019; this study), we should

expect new arrangements as more genus-level studies are performed.

Rare yet everywhere, *Physetocaris microphthalmalma* is circumglobally distributed

After a literature search, we compiled all records for *P. microphthalmalma* and found this species to be recorded only eight times (Fig. 2) prior to this study. Records include the type locality in the north-western Atlantic (Bermudas) and offshore locations in the North Atlantic, south-eastern Atlantic, north-western Pacific and south-eastern Pacific. Our work contributes with two more new records: the Gulf of Mexico and the north-eastern Pacific (off-shore California). Many cosmopolitan deep-sea species, such as chaetognaths and shrimps, represent cryptic species only discovered after molecular investigations (Miyamoto *et al.* 2010; Varela *et al.* 2021). However, similar investigation across nematodes and amphipods reveals true cosmopolitan distributions (Bik *et al.* 2010; Havermans *et al.* 2013). In the case of *P. microphthalmalma*, cryptic diversity is a feasible hypothesis considering that no molecular or morphological analyses have been performed that include individuals from many locations. The lack of records might indicate that natural populations of *P. microphthalmalma* have small population sizes. Genetic drift strongly affects small populations, causing a more rapid accumulation of genetic differences among populations that would also support cryptic diversification within this species (Charlesworth 2009). Surprisingly, when looking at our phylogenetic results, the individuals from the Gulf of Mexico and the north-eastern Pacific fall within the same clade, representing the same species and no cryptic diversification is supported. This indicates that *P. microphthalmalma* occurs in the North and South Atlantic, North and South Pacific and can potentially be found across the globe.

We propose a few explanations for this 'rare yet everywhere' pattern. The first is that *P. microphthalmalma* is a small deep-sea shrimp that can be carried by oceanic currents across different ocean basins, reaching further locations. Sampling data show *P. microphthalmalma* being collected at different depths, indicating that this species participates in diel vertical migration (DVM). Here, DVM is defined as a predator avoidance behaviour characterised by the vertical movement of pelagic organisms going from deep to shallow waters (dusk) and back (dawn) over a 24-h cycle (Brierley 2014; Bandara *et al.* 2021). Vertical migration has been shown to potentially influence dispersal and connectivity using simulated particles (Gary *et al.* 2020) and empirical data provide evidence that strong vertical migrators (deep-sea shrimps *Systellaspis debilis* and *Acanthephyra purpurea*) are more connected to adjacent ocean basins than weak vertical migrators (*Robustosergia robusta*) (Timm *et al.* 2020). Previous observations of a living specimen document strong buoyancy that could facilitate transportation across oceanic currents (Foxton and Herring 1970).

The transportation could also occur during the larval phases (Shanks 2009). Little is known about deep-sea pelagic larvae, especially the total number of larval phases of a determined species and the morphological variations along the multiple stages (Varela and Bracken-Grissom 2021b). The only description of a gravid *P. microphthalmalma* female indicated three large eggs attached to the female's pleopods (Foxton and Herring 1970). The authors could not confirm whether the female lost some of the embryos or if this is a low fecundity species. The embryos were kept in an aquarium until hatching (44 and 46 days) and reached the zoea II stage 12 days after hatching but no further stages were recorded (Foxton and Herring 1970). Another deep-sea shrimp, *Systellaspis debilis*, is notably known as a cosmopolitan and produces large eggs and only four zoea phases (Bartolotti and Dos Santos 2019) that might be similar to *P. microphthalmalma*. Only a few larval stages are sufficient to guarantee that populations across the globe are connected (Cowen and Sponaugle 2009; Weersing and Toonen 2009; Baco *et al.* 2016; Gary *et al.* 2020). Another idea is that *P. microphthalmalma* may be overlooked during midwater sampling expeditions. *Physetocaris microphthalmalma* is believably a common midwater species but due to the lack of taxonomic expertise or studies on midwater organisms, this species is infrequently identified and reported. Equally believable, gear type (i.e. mesh size) can influence sampling success, indicating that the species is present but the gear is not adequate for the collection of the species (Kaartvedt *et al.* 2012). Lastly, a combination of all these explanations (buoyancy of adults, the presence of larval phases, DVM behaviour, lack of taxonomic expertise and gear type) may play a role in explaining our results indicating that *P. microphthalmalma* is circumglobally distributed across oceanic basins.

Conclusions

Historically, *P. microphthalmalma* has been accepted within the family Physetocarididae and superfamily Physetocaridoidea, sister to Pandaloidea (Pandalidae + Thalassocaridae). However, the phylogenetic placement of *P. microphthalmalma* has been under debate for many decades due to the rarity in collection records and lack of molecular-grade material. Our study is the first to perform a molecular phylogeny with inclusion of this species and provides strong evidence that this is nested within the family Pandalidae, indicating that the superfamily Physetocaridoidea and family Physetocarididae should not be accepted. The deep sea is a fascinating and mysterious realm, teeming with hidden biodiversity waiting to be discovered. We propose increased collaboration across midwater researchers in an effort to conduct comprehensive investigations across oceanic basins. We can better understand this unique ecosystem and uncover new and exciting discoveries through collaborative efforts. Our 'rare yet everywhere'

shrimp is the perfect example of how much more we still have left to discover in the deep-sea and pelagic environments.

Supplementary material

Supplementary material is available [online](#).

References

Baco AR, Etter RJ, Ribeiro PA, von der Heyden S, Beerli P, Kinlan BP (2016) A synthesis of genetic connectivity in deep-sea fauna and implications for marine reserve design. *Molecular Ecology* **25**(14), 3276–3298. doi:[10.1111/mec.13689](https://doi.org/10.1111/mec.13689)

Bandara K, Varpe Ø, Wijewardene L, Tverberg V, Eiane K (2021) Two hundred years of zooplankton vertical migration research. *Biological Reviews* **96**(4), 1547–1589. doi:[10.1111/brv.12715](https://doi.org/10.1111/brv.12715)

Bartolotti C, Dos Santos A (2019) The secret life of deep-sea shrimps: ecological and evolutionary clues from the larval description of *Systellaspis debilis* (Caridea: Oplophoridae). *PeerJ* **7**, e7334. doi:[10.7717/peerj.7334](https://doi.org/10.7717/peerj.7334)

Bik HM, Thomas WK, Lunt DH, Lambson PJD (2010) Low endemism, continued deep-shallow interchanges, and evidence for cosmopolitan distributions in free-living marine nematodes (order Enoplogida). *BMC Evolutionary Biology* **10**(1), 389. doi:[10.1186/1471-2148-10-389](https://doi.org/10.1186/1471-2148-10-389)

Bowman TE, Abele LG (1982) Classification of the recent Crustacea. In 'The Biology of Crustacea, Decapoda. Vol. 1'. (Ed. LG Abele) pp. 1–27. (Academic Press: New York, NY, USA)

Bracken HD, De Grave S, Felder DL, Martin JW (2009) Phylogeny of the infraorder Caridea based on mitochondrial and nuclear genes (Crustacea: Decapoda). *Decapod Crustacean Phylogenetics* **18**, 274–298.

Bracken HD, De Grave S, Toon A, Felder DL, Crandall KA (2010) Phylogenetic position, systematic status, and divergence time of the Procarididea (Crustacea: Decapoda). *Zoologica Scripta* **39**(2), 198–212. doi:[10.1111/j.1463-6409.2009.00410.x](https://doi.org/10.1111/j.1463-6409.2009.00410.x)

Bracken-Grissom HD, Felder DL, Vollmer NL, Martin JW, Crandall KA (2012) Phylogenetics links monster larva to deep-sea shrimp. *Ecology and Evolution* **2**(10), 2367–2373. doi:[10.1002/ece3.347](https://doi.org/10.1002/ece3.347)

Brierley AS (2014) Diel vertical migration. *Current Biology* **24**(22), R1074–R1076. doi:[10.1016/j.cub.2014.08.054](https://doi.org/10.1016/j.cub.2014.08.054)

Chace FA (1940) Plankton of the Bermuda Oceanographic Expeditions. IX. The Bathypelagic Caridean Crustacea. *Zoologica* **25**(11), 117–209. doi:[10.5962/p.184703](https://doi.org/10.5962/p.184703)

Chace Jr FA (1992) On the classification of the Caridea (Decapoda). *Crustaceana* **63**(1), 70–80. doi:[10.1163/156854092X00299](https://doi.org/10.1163/156854092X00299)

Charlesworth B (2009) Effective population size and patterns of molecular evolution and variation. *Nature Reviews Genetics* **10**(3), 195–205. doi:[10.1038/nrg2526](https://doi.org/10.1038/nrg2526)

Christoffersen ML (1989) Phylogeny and classification of the Pandaloidea (Crustacea, Caridea). *Cladistics* **5**(3), 259–274. doi:[10.1111/j.1096-0031.tb00489.x](https://doi.org/10.1111/j.1096-0031.tb00489.x)

Cowen RK, Sponaugle S (2009) Larval dispersal and marine population connectivity. *Annual Review of Marine Science* **1**(1), 443–466. doi:[10.1146/annurev.marine.010908.163757](https://doi.org/10.1146/annurev.marine.010908.163757)

David PM (1972) RRS Discovery Cruise 45, February–April 1971, Plankton investigations at 11°N 20°W, 18°N 25°W and 30°N 26°W. Cruise Report Number 50 (Issued July 1972), National Institute of Oceanography.

Davis MP, Holcroft NI, Wiley EO, Sparks JS, Leo Smith W (2014) Species-specific bioluminescence facilitates speciation in the deep sea. *Marine Biology* **161**(5), 1139–1148. doi:[10.1007/s00227-014-2406-x](https://doi.org/10.1007/s00227-014-2406-x)

De Grave S, Fransen CHJM (2011) Carideorum catalogus: the recent species of the dendrobranchiate, stenopodidean, procarididean and caridean shrimps (Crustacea: Decapoda). *Zoologische Mededelingen* **85**(9), 195–589.

Eilertsen MH, Malaquias MAE (2015) Speciation in the dark: Diversification and biogeography of the deep-sea gastropod genus *Scaphander* in the Atlantic Ocean. *Journal of Biogeography* **42**(5), 843–855. doi:[10.1111/jbi.12471](https://doi.org/10.1111/jbi.12471)

Foxton P (1971) RRS Discovery Cruise 39, April–June 1971, Plankton investigations at 60°N 20°W, 53°N 20°. Cruise Report 40 (Issued October 1971), National Institute of Oceanography.

Foxton P, Herring PJ (1970) Recent records of *Physetocaris microphthalmus* Chace with notes on the male and description of the early larvae (Decapoda, Caridea). *Crustaceana* **18**(1), 93–104. doi:[10.1163/156854070X00103](https://doi.org/10.1163/156854070X00103)

Gary SF, Fox AD, Biastoch A, Roberts JM, Cunningham SA (2020) Larval behaviour, dispersal and population connectivity in the deep sea. *Scientific Reports* **10**(1), 10675. doi:[10.1038/s41598-020-67503-7](https://doi.org/10.1038/s41598-020-67503-7)

Gordon I (1970) Two early "discovery" records of *Physetocaris* Chace (Decapoda, Caridea). *Crustaceana* **18**(1), 105–107. doi:[10.1163/156854070X00112](https://doi.org/10.1163/156854070X00112)

Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Systematic Biology* **59**(3), 307–321. doi:[10.1093/sysbio/syq010](https://doi.org/10.1093/sysbio/syq010)

Guzmán G (1999) Primer registro de *Physetocaris microphthalmus* Chace, 1940 para el Pacífico Sur oriental (18°25'S–71°43'W) (Crustacea: Decapoda: Physetocaridae). [First record of *Physetocaris microphthalmus* Chace, 1940 for the eastern South Pacific (18°25'S–71°43'W) (Crustacea: Decapoda: Physetocaridae).] *Museo Nacional de Historia Natural, Santiago* **337**, 3–5. [In Spanish]

Havermans C, Sonet G, d'Udekem d'Acoz C, Nagy ZT, Martin P, Brix S, Riehl T, Agrawal S, Held C (2013) Genetic and morphological divergences in the cosmopolitan deep-sea amphipod *Eurythenes gryllus* reveal a diverse abyss and a bipolar species. *PLoS One* **8**(9), e74218. doi:[10.1371/journal.pone.0074218](https://doi.org/10.1371/journal.pone.0074218)

Hoang DT, Chernomor O, Von Haeseler A, Minh BQ, Vinh LS (2018) UFBoot2: improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution* **35**(2), 518–522. doi:[10.1093/molbev/msx281](https://doi.org/10.1093/molbev/msx281)

Holthuis LB (1955) The recent genera of the caridean and stenopodidean shrimps (Class Crustacea, Order Decapoda, Supersection Natantia) with keys for their determination. *Zoologische Verhandelingen* **26**, 1–157.

Holthuis LB (1993) 'The Recent Genera of the Caridean and Stenopodidean Shrimps (Crustacea, Decapoda) with an Appendix on the Order Amphionidacea.' (Nationale Natuurhistorisch Museum: Leiden, Netherlands)

Judkins H, Rose-Mann L, Lindgren A, Taite M, Bush S, Vecchione M (2022) A newly discovered *Helicocranchia* species (Cephalopoda: Cranchiidae: Taoniinae) in the northern Gulf of Mexico. *Bulletin of Marine Science* **98**(3), 419–430. doi:[10.5343/bms.2021.0048](https://doi.org/10.5343/bms.2021.0048)

Kaartvedt S, Staby A, Aksnes DL (2012) Efficient trawl avoidance by mesopelagic fishes causes large underestimation of their biomass. *Marine Ecology Progress Series* **456**, 1–6. doi:[10.3354/meps09785](https://doi.org/10.3354/meps09785)

Kalyaanamoorthy S, Minh BQ, Wong TKF, Von Haeseler A, Jermiin LS (2017) ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature Methods* **14**(6), 587–589. doi:[10.1038/nmeth.4285](https://doi.org/10.1038/nmeth.4285)

Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* **30**(4), 772–780. doi:[10.1093/molbev/mst010](https://doi.org/10.1093/molbev/mst010)

Kikuchi T, Nemoto T (1986) List of pelagic shrimps (Crustacea, Decapoda) from the western North Pacific. *Bulletin of Biogeographical Society of Japan* **41**, 51–59.

Kikuchi T, Omori M (1985) Vertical distribution and migration of oceanic shrimps at two locations off the Pacific coast of Japan. *Deep-Sea Research – A. Oceanographic Research Papers* **32**(7), 837–851. doi:[10.1016/0198-0149\(85\)90119-0](https://doi.org/10.1016/0198-0149(85)90119-0)

Komai T, Chan TY, De Grave S (2019) Establishment of a new shrimp family Chlorotocellidae for four genera previously assigned to Pandalidae (Decapoda, Caridea, Pandaloidea). *Zoosystematics and Evolution* **95**, 391–402. doi:[10.3897/zse.95.35999](https://doi.org/10.3897/zse.95.35999)

Liao Y, Ma KY, De Grave S, Komai T, Chan TY, Chu KH (2019) Systematic analysis of the caridean shrimp superfamily Pandaloidea (Crustacea: Decapoda) based on molecular and morphological evidence. *Molecular Phylogenetics and Evolution* **134**, 200–210. doi:[10.1016/j.ympev.2019.02.006](https://doi.org/10.1016/j.ympev.2019.02.006)

Minh B Q, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, Von Haeseler A, Lanfear R (2020) IQ-TREE 2: new models and efficient methods for phylogenetic inference in the genomic era. *Molecular Biology and Evolution* **37**(5), 1530–1534. doi:[10.1093/molbev/msaa015](https://doi.org/10.1093/molbev/msaa015)

Miyamoto H, Machida RJ, Nishida S (2010) Genetic diversity and cryptic speciation of the deep sea chaetognath *Caecosagitta macrocephala* (Fowler, 1904). *Deep-Sea Research – II. Topical Studies in Oceanography* 57(24–26), 2211–2219. doi:10.1016/j.dsr2.2010.09.023

Pietsch TW, Sutton TT (2015) A new species of the ceratioid anglerfish genus *Lasiognathus regan* (Lophiiformes: Oneirodidae) from the northern Gulf of Mexico. *Copeia* 103(2), 429–432. doi:10.1643/CI-14-181

Poore GC, Ahyong ST (2023) 'Marine Decapod Crustacea: a Guide to Families and Genera of the World.' (CSIRO Publishing)

Ramirez-Llodra E, Tyler PA, Baker MC, Bergstad OA, Clark MR, Escobar E, Levin LA, Menot L, Rowden AA, Smith CR, Van Dover CL (2011) Man and the last great wilderness: human impact on the deep sea. *PLoS One* 6(8), e22588. doi:10.1371/journal.pone.0022588

Rodríguez-Flores PC, Macpherson E, Schnabel KE, Ahyong ST, Corbari L, Machordom A (2022) Depth as a driver of evolution and diversification of ancient squat lobsters (Decapoda, Galatheoidea, Phyllodiorhynchus). *Molecular Phylogenetics and Evolution* 171, 107467. doi:10.1016/j.ympev.2022.107467

Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61(3), 539–542. doi:10.1093/sysbio/sys029

Shanks AL (2009) Pelagic larval duration and dispersal distance revisited. *The Biological Bulletin* 216(3), 373–385. doi:10.1086/BBLv216n3p373

Sinniger F, Pawłowski J, Harii S, Gooday AJ, Yamamoto H, Chevaldonné P, Cedhagen T, Carvalho G, Creer S (2016) Worldwide analysis of sedimentary DNA reveals major gaps in taxonomic knowledge of deep-sea benthos. *Frontiers in Marine Science* 3, 92. doi:10.3389/fmars.2016.00092

Song H, Buhay JE, Whiting MF, Crandall KA (2008) Many species in one: DNA barcoding overestimates the number of species when nuclear mitochondrial pseudogenes are coamplified. *Proceedings of the National Academy of Sciences* 105(36), 13486–13491. doi:10.1073/pnas.0803076105

Sutton TT, Clark MR, Dunn DC, Halpin PN, Rogers AD, Guinotte J, Bograd SJ, Angel MV, Perez JAA, Wishner K, Haedrich RL, Lindsay DJ, Drazen JC, Vereshchaka A, Piątkowski U, Morato T, Blachowiak-Samołyk K, Robison BH, Gjerde KM, Pierrot-Bults A, Bernal P, Reygondeau G, Heino M (2017) A global biogeographic classification of the mesopelagic zone. *Deep-Sea Research – I. Oceanographic Research Papers* 126, 85–102. doi:10.1016/j.dsr.2017.05.006

Tamura K, Stecher G, Kumar S (2021) MEGA11: molecular evolutionary genetics analysis version 11. *Molecular Biology and Evolution* 38(7), 3022–3027. doi:10.1093/molbev/msab120

Timm L, Bracken-Grissom HD (2015) The forest for the trees: evaluating molecular phylogenies with an emphasis on higher-level Decapoda. *Journal of Crustacean Biology* 35(5), 577–592. doi:10.1163/1937240X-00002371

Timm LE, Isma LM, Johnston MW, Bracken-Grissom HD (2020) Comparative population genomics and biophysical modeling of shrimp migration in the Gulf of Mexico reveals current-mediated connectivity. *Frontiers in Marine Science* 7, 19. doi:10.3389/fmars.2020.00019

Tsang LM, Chan TY, Cheung MK, Chu KH (2009) Molecular evidence for the southern hemisphere origin and deep-sea diversification of spiny lobsters (Crustacea: Decapoda: Palinuridae). *Molecular Phylogenetics and Evolution* 51(2), 304–311. doi:10.1016/j.ympev.2009.01.015

Varela C, Bracken-Grissom HD (2021a) Primer registro del género *Oediceroides* (Amphipoda: Amphelochidea: Oedicerotidae) del Golfo de México, con la descripción de una especie nueva. [First record of the genus *Oediceroides* (Amphipoda: Amphelochidea: Oedicerotidae) for the Gulf of Mexico, with the description of a new species.] *Novitates Caribaea* 18, 18–27. [In Spanish] doi:10.33800/nc.vi18.261

Varela C, Bracken-Grissom H (2021b) A mysterious world revealed: larval-adult matching of deep-sea shrimps from the Gulf of Mexico. *Diversity* 13(10), 457. doi:10.3390/d13100457

Varela C, Golightly C, Timm LE, Wilkins B, Frank T, Fenolio D, Collins SB, Bracken-Grissom HD (2021) DNA barcoding enhances large-scale biodiversity initiatives for deep-pelagic crustaceans within the Gulf of Mexico and adjacent waters. *The Journal of Crustacean Biology* 41(1), ruab005. doi:10.1093/jcbiol/ruab005

Wang Y, Ma KY, Tsang LM, Wakabayashi K, Chan TY, De Grave S, Chu KH (2021) Confirming the systematic position of two enigmatic shrimps, Amphionidae and Procarididae (Crustacea: Decapoda). *Zoologica Scripta* 50(6), 812–823. doi:10.1111/zsc.12509

Wasmer RA (1985) New record for *Physetocaris microphthalmus* Chace (Decapoda, Caridea, Physetocarididae) from the South Pacific. *Crustaceana* 49(3), 315–318. doi:10.1163/156854085X00648

Webb TJ, Vanden Berghe E, O'Dor R (2010) Biodiversity's big wet secret: the global distribution of marine biological records reveals chronic under-exploration of the deep pelagic ocean. *PLoS One* 5(8), e10223. doi:10.1371/journal.pone.0010223

Weersing K, Toonen RJ (2009) Population genetics, larval dispersal, and connectivity in marine systems. *Marine Ecology Progress Series* 393, 1–12. doi:10.3354/meps08287

Data availability. Sequences generated for *Physetocaris microphthalmus* are deposited in GenBank (accession numbers: OR345167–OR345172, OR359961, OR359962, Supplementary Table S1). Alignments used to build trees are available as supplemental materials. Cruise data are publicly available through the Gulf of Mexico Research Initiative Information and Data Cooperative (GRIIDC) at <https://data.gulfresearchinitiative.org> (10.7266/N7XP7385, 10.7266/N7902234, 10.7266/n7-ac8e-0240, 10.7266/N70POX3T).

Conflicts of interest. The authors declare that they have no conflicts of interest.

Declaration of funding. This research was supported by the DEEPEND|RESTORE consortium provided in part by the Gulf of Mexico Research Initiative and in part by the National Oceanic and Atmospheric Administration's RESTORE Science Program under award NA19NOS4510193 to Nova Southeastern University and Florida International University. The Pacific Ocean specimen was collected with funding awarded to Anela Choy, from the UC Ship Funds Program (University of California—San Diego, Scripps Institution of Oceanography) and National Science Foundation (NSF) Award OCE-1829812.

Acknowledgements. We thank all researchers involved in the DEEPEND|RESTORE consortium for support in sample collections. We thank Dr Anela Choy and her team for collecting the specimen from north-eastern Pacific. This is contribution #1597 from the Coastlines and Oceans Division of the Institute of Environment at Florida International University.

Author affiliations

^AInstitute of Environment and Department of Biology, Florida International University (FIU), Miami, FL, USA.

^BDepartment of Invertebrate Zoology, National Museum of Natural History, Smithsonian Institution, 10th and Constitution Avenue NW, Washington, DC, USA.