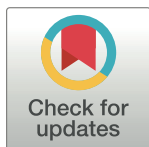


## RESEARCH ARTICLE

## Early-life exposure to Ivermectin alters long-term growth and disease susceptibility

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## Abstract

Ivermectin is a broad-spectrum antiparasitic medicine, which is often used as a treatment for parasites or as a prophylaxis. While studies have looked at the long-term effects of Ivermectin on helminths, studies have not considered the long-term impacts of this treatment on host health or disease susceptibility. Here, we tracked the effects of early life Ivermectin treatment in Cuban tree frogs (*Osteopilus septentrionalis*) on growth rates, mortality, metabolically expensive organ size, and susceptibility to *Batrachochytrium dendrobatidis* (Bd) infection. One year after exposure, there was no effect of Ivermectin exposure on frog mass ( $X^2_1 = 0.904$ ,  $p = 0.34$ ), but when tracked through the exponential growth phase (~2.5 years) the Ivermectin exposed individuals had lower growth rates and were ultimately smaller ( $X^2_1 = 7.78$ ,  $p = 0.005$ ;  $X^2_1 = 5.36$ ,  $p = 0.02$ , respectively). These results indicate that early life exposure is likely to have unintended impacts on organismal growth and potentially reproductive fitness. Additionally, we exposed frogs to Bd, a pathogenic fungus that has decimated amphibian populations globally, and found early life exposure to Ivermectin decreased disease susceptibility (disease load:  $X^2_1 = 17.57$ ,  $p = 0.0002$ ) and prevalence (control: 55%; Ivermectin: 22%) over 2 years after exposure. More research is needed to understand the underlying mechanism behind this phenomenon. Given that Ivermectin exposure altered disease susceptibility, proper controls should be implemented when utilizing this drug as an antiparasitic treatment in research studies.

## Introduction

Parasitic nematodes are cosmopolitan and infect an extreme variety of taxonomic groups (e.g. beetles [1,2], amphibians [3] and humans [4]). In humans alone parasitic soil helminths are estimated to infect more than 1.45 billion people globally [4], many of whom are at risk of increased morbidity or mortality. To this end, Ivermectin is often used as a long-term anthelmintic and is broadly effective against many different nematode infections (e.g. *Ascaris lumbricoides*- the most common human helminth infection, *Necator americanus*, *Ancylostoma duodenale* [4] and *Onchocerca vulvulus* [2]). In most cases, the health cost of taking Ivermectin is thought to be better than the alternative infection. Ivermectin is often only effective against the juvenile stages (microfilariae; [5]), which means early treatment and long-term

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administration of this drug in at-risk populations, (e.g. pre-pubescent children) is necessary to reduce the impact of infections [2].

Ivermectin is an incredibly important medication used in agriculture and pet trade, as well. For example, it is used to prevent *Dirofilaria immitis*, canine heartworm [6,7], to clear lung-worm infections in US cattle [8], and it is one of the most common anthelmintic medications used in sheep [9] and equines in the UK [10]. The adult nematodes can live for 5–10 years, and while Ivermectin does not kill the adults, it can be used reduce local transmission events by killing the microfilariae. Again, in both the agricultural system and with our own pets, we often utilize early life administration of Ivermectin to try to control nematode infections.

Amphibians, arguably the most at-risk taxonomic group in terms of declines and extinctions [11], are another group known to suffer from severe helminth infections [3]. Amphibians are declining globally and are often collected from the wild to be cared for by aquariums and zoos. In order to provide the highest chance of survival and to reduce the chance of parasitic transmission to the other captive individuals, facilities clear amphibians of their helminth infections with Ivermectin upon arrival [12]. Many of these species are threatened or endangered in the wild, and so the survival of these captive individuals is crucial.

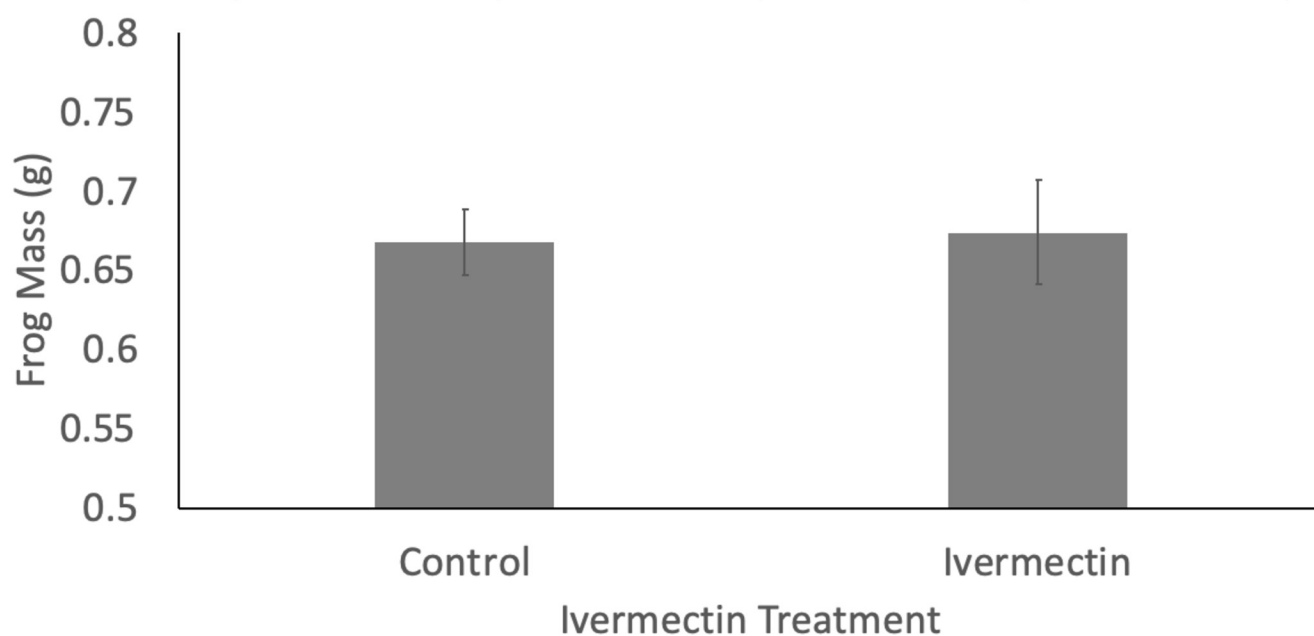
Ivermectin functions by altering the glutamate-gated chloride channel receptors [5] and causing nematode paralysis. It is important to note, however, that while these channels are found in the peripheral neuromuscular systems of invertebrates, they are also found in the central nervous system of vertebrates [13,14]. Consequently, this commonly used anthelmintic may be targeting invertebrates, while impacting the vertebrate host as well.

Here, we assessed the impacts of early life exposure of Ivermectin on long term growth, health and disease susceptibility in amphibians. Early life exposure to pesticides is known to have long-lasting and often unpredictable impacts on amphibians, including increased mortality and disease risk into adulthood [15]. Therefore, it seems plausible that Ivermectin exposure may similarly impact health. Amphibians are one of the groups often affected by Ivermectin exposure, and importantly, they share approximately 1700 genes with human disease associations [16]. Additionally, they are a useful model for exploring the impacts of pesticides like Ivermectin on humans because there is overlap between the two immune and endocrine systems [17]. Understanding the impacts of chemical exposure on amphibians themselves is important, and they are an underutilized model for studying the chemical exposure on human health.

## Results

There was no effect of Ivermectin treatment on mortality ( $X^2_1 = 0.904$ ,  $p = 0.34$ ; survival rates of 45% and 55% in control and Ivermectin-dosed groups, respectively) and there was no difference in frog mass at one-year post exposure (average mass  $\pm$  SEM: control:  $0.668 \pm 0.02$ ; Ivermectin:  $0.674 \pm 0.03$ ;  $X^2_1 = 0.02$ ,  $p = 0.877$ ; Fig 1). Individuals exposed to Ivermectin had reduced growth rate (2.5 years of growth:  $X^2_1 = 7.78$ ,  $p = 0.005$ ; Fig 2) and lower change in mass ( $X^2_1 = 5.36$ ,  $p = 0.02$ ) at the end of the experiment, relative to unexposed animals. There was no significant effect of Ivermectin treatment on metabolically expensive organ mass (liver:  $X^2_1 = 0.006$ ,  $p = 0.93$ , heart:  $X^2_1 = 0.730$ ,  $p = 0.39$ , or spleen:  $X^2_1 = 0.21$ ,  $p = 0.90$ ), and independent of treatment, the size of the frog was a significant correlate for liver and heart but not spleen (liver:  $X^2_1 = 5.33$ ,  $p = 0.02$ , heart:  $X^2_1 = 5.03$ ,  $p = 0.02$ , spleen:  $X^2_1 = 0.03$ ,  $p = 0.86$ ).

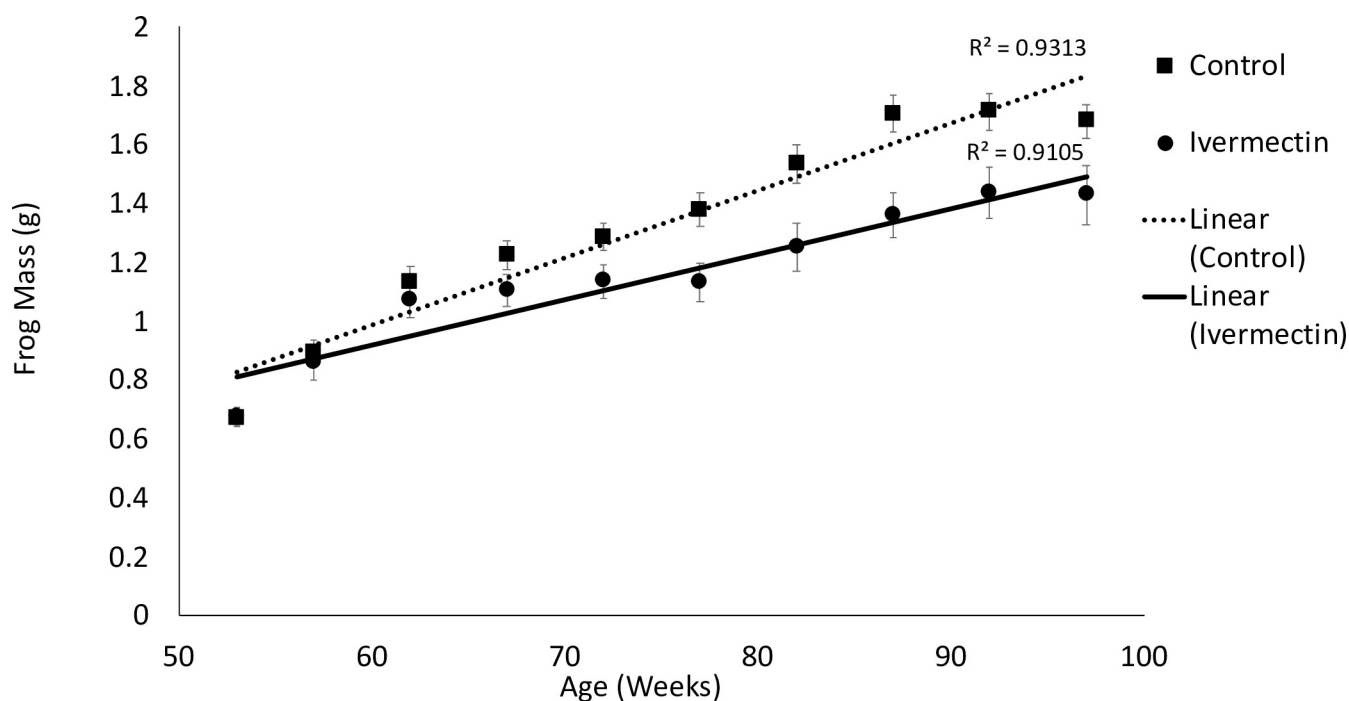
Frogs with early life Ivermectin exposure had decreased Bd prevalence (control: 55% and Ivermectin: 22%) and disease load ( $X^2_1 = 17.57$ ,  $p = 0.0002$ ; Fig 3). We also found spleen mass and Bd load correlated ( $X^2_1 = 6.60$ ,  $p = 0.01$ ), but that there was an interactive effect of Ivermectin exposure and spleen mass on Bd load ( $X^2_1 = 11.60$ ,  $p = 0.0007$ ; Fig 4). There was a



**Fig 1. The effects of early life exposure of Ivermectin on mass of Cuban tree frogs (*Osteopilus septentrionalis*) one year after metamorphosis.** No significant difference in mass was observed. Shown are means  $\pm$  SEM.

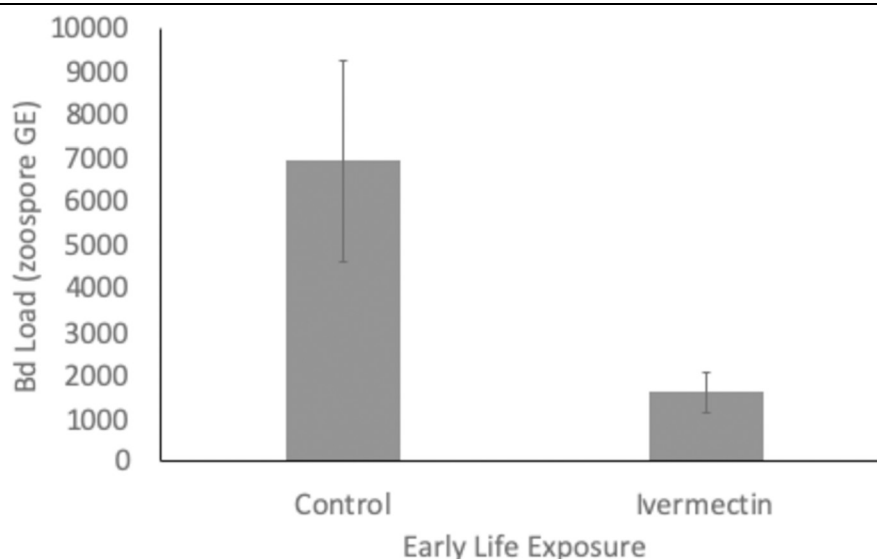
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negative correlation between spleen size and Bd load for individuals exposed to Ivermectin, while for the control individuals the correlation was positive (Fig 4).



**Fig 2. The effects of early life exposure of Ivermectin on the long-term growth of Cuban tree frogs (*Osteopilus septentrionalis*).** Individuals exposed to Ivermectin had slower growth rates and weighed less, relative to controls. Shown are means  $\pm$  SEM.

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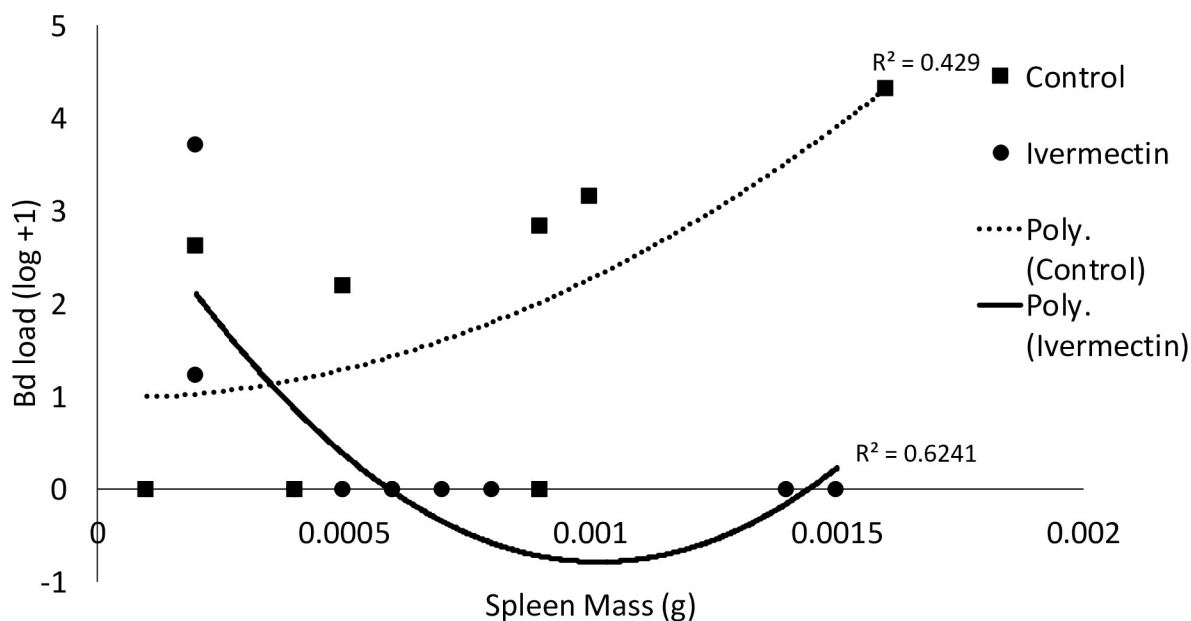


**Fig 3.** The effects of early life exposure of Ivermectin on the disease susceptibility in Cuban tree frogs (*Osteopilus septentrionalis*) to *Batrachochytrium dendrobatidis* (Bd; Bd load is reported in zoospore genome equivalents). Individuals exposed to Ivermectin were less susceptible to Bd infection compared to controls. Shown are means  $\pm$  SEM.

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## Discussion

Ivermectin is an extremely common antiparasitic medication used in humans and other vertebrates both prophylactically and in treatment regimes. Studies have assessed the mortality and short-term risk associated with administration of this drug, and generally the short-term side effects appear minimal especially when compared to the benefit of the drug [5,18,19]. Our results support this in that metamorphic amphibians exposed at an early developmental stage did not



**Fig 4.** There was an interaction between Ivermectin exposure and spleen size and their impact on *Batrachochytrium dendrobatidis* (Bd) load in Cuban tree frogs (*Osteopilus septentrionalis*).

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have altered growth up to one year after exposure (Fig 1). This is consistent with other studies that found short-term consequences were mild even with repeated or stronger than FDA recommended doses [18,19], however we do report long-term risks of early Ivermectin exposure.

Cuban tree frogs undergo sexual maturity at approximately 17 and 36 weeks for males and females, respectively (reviewed by [20]), but they continue to grow for several years. Here, we exposed adolescent Cuban tree frogs (~ 4 weeks post metamorphosis) to Ivermectin and tracked their growth through the critical growth period (approximately 2.5 years). Despite no evidence of growth differences 1 year after exposure, we found that individuals exposed to Ivermectin, just once as young developing metamorphs, had reduced growth late into their critical growth period (Fig 2). This puts the Ivermectin exposed individuals at a disadvantage long-term, as larger males and females are reproductively more fit [21]. Given that these individuals only experienced one dose, it is likely that our results underestimate the true impact of repeated Ivermectin exposure during early development.

Organisms operate with limited energetic resources, and during development these resources must be differentially allocated to growth, the development of crucial organs (e.g. heart and liver), and pathogen defense (e.g. immune response). We found that while there was a positive correlation between frog mass and organ (liver and heart) mass, which was expected, there was no effect of Ivermectin treatment on the mass of these metabolically expensive organs. Ivermectin exposure reduced whole body growth and growth rate when compared to control animals, but did not result in smaller hearts, livers, or spleens of these individuals. This indicates Ivermectin influences growth in organ-specific ways, as metabolically expensive and important organs likely received an increased allocation of resources relative to whole body growth following Ivermectin treatment. This disparate allocation is not surprising given that studies have shown that resources are shunted toward critical organs and tissues in vertebrates in tissue-specific ways and under resource-limiting conditions [22,23].

We found that individuals exposed to Ivermectin were less susceptible to infection with the pathogenic fungus Bd (Figs 3 and 4). Thus, Ivermectin treatment may actually provide a secondary protection against other infectious agents, consistent with the ongoing discovery of new therapeutic applications of the drug (reviewed in [5]) and despite the fact that Bd itself is immunosuppressive [24–26]. Interestingly, there was an interaction between spleen size and Ivermectin exposure in relation to Bd load. For individuals exposed to Ivermectin, there was a negative correlation between spleen size and Bd load and we saw the opposite for control individuals (Fig 4). Interestingly, Bd exposure alone was found to have no impact on spleen size [27,28], but frogs with an acquired immune response to Bd had increased splenocyte production [29]. While further research is needed to fully understand this dynamic, we hypothesize that the absence of an existing internal parasite load following Ivermectin treatment impacts immune system allocation in response to new infections. Potentially, this permits a larger proportion of the immune response to be directed against the new pathogen, Bd. Given that Bd infection is a major causative agent in the extreme extirpations and extinctions of amphibian species worldwide [11], reduction in disease load and prevalence is certainly an unexpected benefit. We caution against the use of this antiparasitic drug in treatment of Bd, as it has not been tested for this, and given that we did not eliminate the infection it is unlikely to be effective for that purpose. Future work should explore whether the secondary protection afforded by Ivermectin treatment extends to other species and other infectious agents. Consequently, administration of Ivermectin might have unintended positive clinical outcomes, despite evidence indicating resistance development in helminth populations [5].

Our results indicate just a single dose of Ivermectin during early-life can negatively impact growth rates over time, and therefore potentially reproductive fitness. It is important to note that if we had stopped tracking growth at one year, less than halfway through the frogs' critical

growth period, we would not have gotten a view of the long-term impacts of Ivermectin treatment. While we acknowledge the restraints of lab resources and time, our data indicate the time frame considered for short and long term studies should be evaluated in the context of life history stage and overall lifespan of the organism, as well as species-specific drug half-lives.

The clinical and pharmacological side effects of drug administration must be outweighed by the positive clinical outcomes. Ivermectin certainly meets that criterion given the reported significant reduction in microfilarial dermal loads after treatment and minimal side effects [reviewed by 5,30]. More recently, research has shown the potential efficacy of Ivermectin as therapeutic for SARS-CoV-2 *in vitro* [31], as a potential complimentary pharmaceutical treatment for combating malaria [32,33], and a recent meta-analysis provides promising evidence supporting the removal of the contraindications for children <15 kg [34]. However a growing literature suggests neonatal and early-childhood antibiotics impact child growth, BMI, and gut microbiome in age-at-dose and sex-specific ways (e.g., [35]), indicating early pharmaceutical interventions can have long-term implications on organism growth. While we are certainly not advocating that Ivermectin use be eliminated, we want to make sure that it is clear that our results indicate there may similarly be long-term impacts of Ivermectin exposure in early developmental stages. Additionally, given that Ivermectin exposure altered both growth and disease susceptibility in the long-term, the use of proper controls in scientific studies examining or utilizing Ivermectin is crucial [32,33]. Without these controls the impact of Ivermectin exposure may lead to erroneous experimental results.

## Methods

### Animal collection and maintenance

Eggs were collected from invasive, wild Cuban tree frogs (*Osteopilus septentrionalis*) by deploying plastic breeding refugia in local Tampa, FL, USA habitats (27.9506° N, 82.4572° W). No permits were required for egg collection as Cuban tree frogs are an invasive species in FL and all collections were done on private land with owner's permission. No additional collecting permits were required. Briefly, frogs deposited eggs into these refugia overnight, and the eggs were collected and brought to the lab during the morning hours. Eggs were maintained in artificial spring water (ASW; [36]) with weekly water changes and reared through metamorphosis. Animals were never exposed to soil and therefore should not have acquired any nematode infections. Tadpoles were fed organic spinach *ad libitum*. Metamorphic frogs (~ 4 weeks post metamorphosis) were divided evenly and randomly between control and Ivermectin-dosed groups (n = 20/treatment). They were all given a single dose orally of either ASW (control) or 1% Ivermectin (2 mg/kg body mass/treatment) at the same time. All animals were transferred to individual 1 L plastic containers with dampened paper towels and fed calcium dusted crickets *ad libitum*. Mortality was checked daily, container changes occurred weekly, and the frogs were weighed every week through their exponential growth phase (approximately 2.5 years), including during the final two week *Bd* infection period for both control and Ivermectin exposed groups. Throughout this process all amphibians were maintained at approximately 23°C.

### Bd culture preparation

We cultured *Batrachochytrium dendrobatidis* (*Bd*; strain JEL 419 isolated in Panamá) on 1% tryptone agar plates at 18°C for two weeks. We flooded these *Bd* positive (*Bd*+) plates with ASW to suspend the infectious zoospores, and this liquid was homogenized from all the *Bd* + plates. We quantified the zoospore concentration with a hemocytometer and the *Bd* + stock was diluted with ASW to  $1.3 \times 10^5$  zoospores/mL.



## Bd exposure and tissue sampling

To evaluate the effects of Ivermectin exposure on Bd susceptibility, all surviving adult frogs in both control ( $n = 9$ ) and Ivermectin-exposed ( $n = 11$ ) groups were dosed with 1 mL of the Bd + stock during the final two weeks of the overall 2.5 year experimental period. Bd+ stock was squirted directly onto the dorsal surface of each frog. The extra liquid was allowed to collect on the damp paper towels in the 1 L plastic container to promote infection establishment. Following the singular Bd+ dose, the frogs experienced a two-week infection period, during which container changes occurred weekly, they were fed calcium dusted crickets *ad libitum*, and they were maintained at 18°C to promote Bd growth. Animals were euthanatized immediately at the end of this two-week infection period with an overdose of topical benzocaine and they were pithed post mortem. A ventral skin sample was collected from each frog (1 cm<sup>2</sup>) for quantitative PCR to determine Bd infection load (See [37] for the qPCR protocol used). In order to assess the potential effects of Ivermectin treatment on the partitioning of resources between whole animal growth and the growth of individual organs, we also sampled the metabolically-expensive heart, liver, and spleen from each euthanized individual. Organs were collected and rinsed in a 0.7% saline bath, and were weighed immediately. All animal collections and procedures were approved in writing by the University of Tampa Institutional Animal Care and Use Committee (IACUC# 2018–12).

## Frog health monitoring

All animals were monitored daily for health and mortality status. Any animal that appeared lethargic, was not alert upon inspection and did not appear to be healthy in any way were further examined. If an animal appeared unhealthy, they were placed on their back and if righting was not observed in 20 seconds (a conservative timeframe selected to reduce unnecessary discomfort) then the animal was considered moribund. At this point the animal would be euthanized with an overdose of topical benzocaine and they were pithed post mortem as an extra precaution. All animals that died during this experiment died quickly and naturally and did not experience an undue, prolonged unhealthy state; indeed, no animals were euthanized due to an unhealthy state. This was a long-term experiment lasting several years, and early life mortality is not unusual for amphibians. The research staff were trained directly by the chair of the University's IACUC committee.

## Statistical analyses

All statistical analyses were conducted in R statistical software [38]. We used a Cox-proportional hazards regression (package: survival, function: coxph) to determine if there was an effect of Ivermectin treatment on survival. We used a general linear model (package: stats; function: glm) to determine if there was an effect of treatment on: mass size at one year, growth rate, frog mass change, and organ size at the end of the experiment. We used a zero-inflated negative binomial Poisson regression to determine if treatment, spleen size or treatment\*spleen size had an effect on Bd load (package: pscl, function: zeroinfl).

## Supporting information

### S1 Checklist.

(DOCX)

### S1 Raw data. These are the raw data sets used for all statistical analyses and figures.

(XLSX)

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## Author Contributions

**Conceptualization:** Taegan A. McMahon, Jeffrey M. Grim.

**Data curation:** Taegan A. McMahon, Jeffrey M. Grim.

**Formal analysis:** Taegan A. McMahon.

**Funding acquisition:** Taegan A. McMahon, Jeffrey M. Grim.

**Investigation:** Taegan A. McMahon, Shannon Fernandez-Denmark, Jeffrey M. Grim.

**Methodology:** Taegan A. McMahon, Shannon Fernandez-Denmark, Jeffrey M. Grim.

**Project administration:** Taegan A. McMahon, Jeffrey M. Grim.

**Resources:** Taegan A. McMahon, Jeffrey M. Grim.

**Supervision:** Taegan A. McMahon.

**Writing – original draft:** Taegan A. McMahon, Shannon Fernandez-Denmark, Jeffrey M. Grim.

**Writing – review & editing:** Taegan A. McMahon, Shannon Fernandez-Denmark, Jeffrey M. Grim.

## References

1. Davis AK, Coogler B, Johnson I. The heartrate reaction to acute stress in horned passalus beetles (*Odontotaenius disjunctus*) is negatively affected by a naturally-occurring nematode parasite. *Insects*. 2017; 8(110). <https://doi.org/10.3390/insects8040110> PMID: 29057831
2. Moncayo AL, Vaca M, Amorim L, Rodriguez A, Erazo S, Oviedo G, et al. Impact of long-term treatment with Ivermectin on the prevalence and intensity of soil-transmitted helminth infections. *Plos Neglected Tropical Diseases*. 2008; 2(9). <https://doi.org/10.1371/journal.pntd.0000293> PMID: 18820741
3. Knutie SA, Wilkinson CL, Wu QC, Ortega CN, Rohr JR. Host resistance and tolerance of parasitic gut worms depend on resource availability. *Oecologia*. 2017; 183:1031–40. <https://doi.org/10.1007/s00442-017-3822-7> PMID: 28138818
4. Pullan RL, Smith JL, Jasrasaria R, Brooker SJ. Global numbers of infection and disease burden of soil transmitted helminth infections in 2010. *Parasites and Vectors*. 2014; 7(37). <https://doi.org/10.1186/1756-3305-7-37> PMID: 24447578
5. Laing R, Gillan V, Devaney E. Ivermectin—Old Drug, New Tricks? *Trends in Parasitology*. 2017; 33(6):463–72. <https://doi.org/10.1016/j.pt.2017.02.004> PMID: 28285851
6. Traversa D, Di Cesare A, Conboy G. Canine and feline cardiopulmonary parasitic nematodes in Europe: emerging and underestimated. *Parasites & vectors*. 2010; 3(1):62. <https://doi.org/10.1186/1756-3305-3-62> PMID: 20653938
7. Bowman DD, Atkins CE. Heartworm biology, treatment, and control. *Veterinary Clinics: Small Animal Practice*. 2009; 39(6):1127–58.
8. McArthur MJ, Reinemeyer CR. Herding the U.S. cattle industry toward a paradigm shift in parasite control. *Veterinary Parasitology*. 2014; 2014(204):34–43.
9. Burgess CGS, Bartley Y, Redman E, Skuce PJ, Nath M, Whitelaw F, et al. A survey of the trichostrongylid nematode species present on UK sheep farms and associated anthelmintic control practices. *Veterinary Parasitology*. 2012; 189:299–307. <https://doi.org/10.1016/j.vetpar.2012.04.009> PMID: 22560313
10. Relf VE, Morgan ER, Hodgkinson JE, Matthews JB. A questionnaire study on parasite control practices on UK breeding thoroughbred studs. *Equine Veterinary Journal*. 2012; 44:466–71.
11. Stuart SN, Chanson JS, Cox NA, Young BE, Rodrigues ASL, Fischman DL, et al. Status and trends of amphibian declines and extinctions worldwide. *Science*. 2004; 306(5702):1783–6. <https://doi.org/10.1126/science.1103538> PMID: 15486254



12. Clayton LA, Nelson J, Payton ME, McDonnell JJ, Rossi BH, King-Rahn K, et al. Clinical signs, management and outcome of presumptive Ivermectin overdose in a group of Dendrobatid frogs. *Journal of Herpetological Medicine and Surgery*. 2012; 22(1):5–11.
13. Aller MI, Janusonis S, Fite KV, Fernánde z-Lo'pez A. Distribution of the GABAA receptor complex beta 2/3 subunits in the brain of the frog *Rana pipiens*. *Neuroscience Letters*. 1997; 225:65–8. [https://doi.org/10.1016/s0304-3940\(97\)00191-2](https://doi.org/10.1016/s0304-3940(97)00191-2) PMID: 9143019
14. Ōmura S, Crump A. Ivermectin: panacea for resource-poor communities? *Trends in Parasitology*. 2014; 30(9):445–55. <https://doi.org/10.1016/j.pt.2014.07.005> PMID: 25130507
15. Rohr JR, Halstead NT, Johnson SA, Raffel TR, McMahon TA, Martin LB, editors. Pesticide exposure during development increases mortality to infections in adulthood. *Integrative and Comparative Biology*; 2011 Mar.
16. Hellsten U, Harland RM, Gilchrist MJ, Hendrix D, Jurka J, Kapitonov V, et al. The Genome of the Western Clawed Frog *Xenopus tropicalis*. *Science*. 2010; 328(5978):633–6. <https://doi.org/10.1126/science.1183670> PMID: 20431018
17. Hayes TB. Welcome to the revolution: Integrative biology and assessing the impact of endocrine disruptors on environmental and public health. *Integrative and Comparative Biology*. 2005; 45(2):321–9. <https://doi.org/10.1093/icb/45.2.321> PMID: 21676775
18. Pacqué M, Munoz B, Greene BM, Taylor HR. Community-based treatment of onchocerciasis with ivermectin: safety, efficacy, and acceptability of yearly treatment. *Journal of infectious diseases*. 1991; 163(2):381–5.
19. Guzzo CA, Furtek CI, Porras AG, Chen C, Tipping R, Clineschmidt CM, et al. Safety, tolerability, and pharmacokinetics of escalating high doses of ivermectin in healthy adult subjects. *The Journal of Clinical Pharmacology*. 2002; 42(10):1122–33. <https://doi.org/10.1177/009127002401382731> PMID: 12362927
20. Rouse Holzwart K, Campbell T, Smith A. A Risk Assessment of the Cuban Treefrog (*Osteopilus septentrionalis*) Invasion in Florida 2011.
21. Howard RD, Kluge AG. Proximate mechanisms of sexual selection in wood frogs. *Evolution*. 1985; 39(2):260–77. <https://doi.org/10.1111/j.1558-5646.1985.tb05665.x> PMID: 28564216
22. Brigelius-Flohé R. Tissue-specific functions of individual glutathione peroxidases. *Free Radical Biology and Medicine*. 1999; 27(9–10):951–65. [https://doi.org/10.1016/s0891-5849\(99\)00173-2](https://doi.org/10.1016/s0891-5849(99)00173-2) PMID: 10569628
23. Grim JM, Hyndman KA, Kriska T, Girotti AW, Crockett EL. Relationship between oxidizable fatty acid content and level of antioxidant glutathione peroxidases in marine fish. *Journal of Experimental Biology*. 2011; 214(22):3751–9. <https://doi.org/10.1242/jeb.058214> PMID: 22031739
24. Rosenblum EB, Poorten TJ, Settles M, Murdoch GK, Robert J, Maddox N, et al. Genome-wide transcriptional response of *Silurana* (*Xenopus*) *tropicalis* to infection with the deadly chytrid fungus. *PloS one*. 2009; 4(8):e6494. <https://doi.org/10.1371/journal.pone.0006494> PMID: 19701481
25. Ribas L, Li M-S, Doddington BJ, Robert J, Seidel JA, Kroll JS, et al. Expression profiling the temperature-dependent amphibian response to infection by *Batrachochytrium dendrobatidis*. *PloS one*. 2009; 4(12):e8408. <https://doi.org/10.1371/journal.pone.0008408> PMID: 20027316
26. Fites JS, Ramsey JP, Holden WM, Collier SP, Sutherland DM, Reinert LK, et al. The invasive chytrid fungus of amphibians paralyzes lymphocyte responses. *Science*. 2013; 342(6156):366–9. <https://doi.org/10.1126/science.1243316> PMID: 24136969
27. Fites JS, Reinert LK, Chappell TM, Rollins-Smith LA. Inhibition of local immune responses by the frog-killing fungus *Batrachochytrium dendrobatidis*. *Infect Immun*. 2014; 82(11):4698–706. <https://doi.org/10.1128/IAI.02231-14> PMID: 25156734
28. Brannelly LA, Webb RJ, Skerratt LF, Berger L. Effects of chytridiomycosis on hematopoietic tissue in the spleen, kidney and bone marrow in three diverse amphibian species. *Pathog Dis*. 2016; 74(7). <https://doi.org/10.1093/femspd/ftw069> PMID: 27422094
29. McMahon TA, Sears BF, Venesky MD, Bessler SM, Brown JM, Deutsch K, et al. Amphibians acquire resistance to live and dead fungus overcoming fungal immunosuppression. *Nature*. 2014; 511(7508):224–7. <https://doi.org/10.1038/nature13491> PMID: 25008531
30. Crump A. Ivermectin: enigmatic multifaceted 'wonder' drug continues to surprise and exceed expectations. *The Journal of antibiotics*. 2017; 70(5):495. <https://doi.org/10.1038/ja.2017.11> PMID: 28196978
31. Caly L, Druce JD, Catton MG, Jans DA, Wagstaff KM. The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 in vitro. *Antiviral Research*. 2020; 178:104787. <https://doi.org/10.1016/j.antiviral.2020.104787> PMID: 32251768
32. Slater HC, Foy BD, Kobylinski K, Chaccour C, Watson OJ, Hellewell J, et al. Ivermectin as a novel complementary malaria control tool to reduce incidence and prevalence: a modelling study. *The Lancet Infectious Diseases*. 2020. [https://doi.org/10.1016/S1473-3099\(19\)30633-4](https://doi.org/10.1016/S1473-3099(19)30633-4) PMID: 31948767

33. Roadmappers I. A Roadmap for the Development of Ivermectin as a Complementary Malaria Vector Control Tool. *The American journal of tropical medicine and hygiene*. 2020; 102(2s):3–24. <https://doi.org/10.4269/ajtmh.19-0620> PMID: 31971144
34. Jittamala P, Monteiro W, Smit MR, Pedrique B, Specht S, Chaccour CJ, et al. A systematic review and an individual patient data meta-analysis of ivermectin use in children weighing less than fifteen kilograms: Is it time to reconsider the current contraindication? *PLOS Neglected Tropical Diseases*. 2021; 15(3):e0009144. <https://doi.org/10.1371/journal.pntd.0009144> PMID: 33730099
35. Uzan-Yulzari A, Turta O, Belogolovski A, Ziv O, Kunz C, Perschbacher S, et al. Neonatal antibiotic exposure impairs child growth during the first six years of life by perturbing intestinal microbial colonization. *Nature Communications*. 2021; 12(1):443. <https://doi.org/10.1038/s41467-020-20495-4> PMID: 33500411
36. Cohen LM, Neimark HL, Everland LK. *Schistosoma mansoni*: Response to cercariae to a thermal gradient. *Journal of Parasitology*. 1980; 66(2):362–4. PMID: 7391881
37. Hyatt AD, Boyle DG, Olsen V, Boyle DB, Berger L, Obendorf D, et al. Diagnostic assays and sampling protocols for the detection of *Batrachochytrium dendrobatidis*. *Diseases of Aquatic Organisms*. 2007; 73(3):175–92. <https://doi.org/10.3354/dao073175> PMID: 17330737
38. Development Core Team R. R: A language and environment for statistical computing R Foundation for Statistical Computing. 2013;Version 3.1.3.