

**Enabling direct compression tablet formulation of celecoxib by simultaneously eliminating punch sticking, improving manufacturability, and enhancing dissolution through co-processing with a mesoporous carrier**

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1    **ABSTRACT**

2    The development of a high quality tablet of Celecoxib (CEL) is challenged by poor dissolution,  
3    poor flowability, and high punch sticking propensity of CEL. In this work, we demonstrate a  
4    particle engineering approach, by loading a solution of CEL in an organic solvent into a  
5    mesoporous carrier to form a coprocessed composite, to enable the development of tablet  
6    formulations up to 40% (w/w) of CEL loading with excellent flowability and tablettability,  
7    negligible punch sticking propensity, and a 3-fold increase in *in vitro* dissolution compared to  
8    a standard formulation of crystalline CEL. CEL is amorphous in the drug-carrier composite  
9    and remained physically stable after 6 months under accelerated stability conditions when the  
10   CEL loading in the composite was  $\leq$  20% (w/w). However, crystallization of CEL to different  
11   extents from the composites was observed under the same stability condition when CEL  
12   loading was 30-50% (w/w). The success with CEL encourages broader exploration of this  
13   particle engineering approach in enabling direct compression tablet formulations for other  
14   challenging active pharmaceutical ingredients.

15

16    **KEYWORDS:** Punch sticking, powder flow, dissolution rate, mesoporous carrier, direct  
17   compression.

18

19        **1. INTRODUCTION**

20        The tablet is the most commonly used pharmaceutical dosage form to treat or alleviate  
21        a disease condition in humans, because of better stability, higher patient-compliance, and lower  
22        manufacturing cost compared to other dosage forms (Arshad et al., 2021). Among the different  
23        processes for tablet manufacturing, direct compression (DC) is preferred if possible as the  
24        process is devoid of solvent usage and the most economical since it involves fewer steps than  
25        other processes. In fact, direct compression is an important route to continuous manufacturing  
26        of tablets meeting established quality standards (Lee et al., 2015). A successful DC process  
27        requires adequate tabletability and flowability of the formulations to ensure adequate  
28        mechanical strength and content uniformity of the finished dosage forms (C. C. Sun, 2010). In  
29        addition, formulation components should not stick to punches during compression, in order to  
30        meet aesthetic standards of the tablet appearance by avoiding dull appearance or pitted surface  
31        (Chattoraj et al., 2018). During large scale commercial manufacturing, such sticking problems  
32        must be eliminated in order to avoid stoppage of compression operations needed for cleaning  
33        and polishing of punches.

34        Celecoxib (CEL) is a non-steroidal anti-inflammatory drug (NSAID) frequently used  
35        in the treatment of osteoarthritis and rheumatoid arthritis. Tablet product development of CEL  
36        is challenging due to its poor dissolution performance (a weak acid with  $pK_a=11.1$  and intrinsic  
37        solubility of 3-7  $\mu\text{g}/\text{mL}$  in water) (Paulson et al., 2001) These problems have been addressed  
38        using several solubilizing strategies, such as solid dispersion by spray drying (Fouad et al.,  
39        2011), self-emulsifying drug delivery (Song et al., 2013), inclusion complexation (Sinha et al.,  
40        2005), **pharmaceutically acceptable solvates (Wang and Sun, 2021)**, spherical crystallization  
41        (Paradkar et al., 2002) and nanoparticles (Liu et al., 2010). Despite these efforts, tablet  
42        products of CEL are still not yet available. A successful commercial CEL tablet product  
43        requires effective resolution to these critical issues on manufacturability and slow dissolution.

44 Mesoporous materials have recently gained attention as a class of drug delivery carriers  
45 due to their high specific surface areas, tunable pore size to accommodate diverse active  
46 pharmaceutical ingredients (APIs), and excellent thermal stability (Bharti et al., 2015; Florek  
47 et al., 2017; Slowing et al., 2008). Additionally, amorphous APIs with acceptable physical  
48 stability can be achieved by confining API in the small pores of mesoporous carriers  
49 (Baumgartner and Planinšek, 2021; Zolotov et al., 2021), which could enhance API dissolution.  
50 Mesoporous carriers with different physico-chemical properties can be used to achieve  
51 flexibility in API loading without significantly impacting manufacturability or content  
52 uniformity (Sun et al., 2018). Here, we attempted to develop a DC tablet product of CEL by  
53 overcoming key manufacturability issues identified above, while simultaneously improving  
54 dissolution through using a CEL-carrier composite. We hypothesize that loading CEL inside  
55 a porous carrier would significantly reduce the probability of punch sticking by minimizing  
56 direct contact between CEL and punch surface during compression.

57

## 58 **2. MATERIALS AND METHODS**

### 59 **2.1. Materials**

60 Celecoxib (CEL, Form III,  $d_{50} = 11.2 \mu\text{m}$ ) (Aarti Labs Pvt. Ltd., Karnataka, India) was  
61 used as received. The same lot of CEL was also used in previous punch sticking studies (Paul  
62 et al., 2017d, 2017b, 2017c). Neusilin<sup>®</sup> (US2, Fuji Chemical Industries PVt. Ltd., Toyama,  
63 Japan) was employed as a mesoporous carrier. Microcrystalline cellulose (MCC; Avicel  
64 PH102, FMC Biopolymer, Philadelphia, PA) and lactose monohydrate (LM; Fastflo<sup>®</sup>,  
65 Foremost Farms, Clayton, WI) were used as tablet filler. Croscarmellose sodium (NaCMC;  
66 Ac-Di-Sol, /FMC Biopolymer, Philadelphia, PA) was used as a tablet disintegrant. Magnesium  
67 stearate (MgSt; HyQual<sup>TM</sup>, Mallinckrodt, St Louis, MO) was used as a lubricant. Dimethyl

68 formamide (DMF; Sigma Aldrich, Saint Louis, MO) was used as a solvent to prepare CEL  
69 solutions for loading into the carrier.

70 **2.2. Methods**

71

72 **2.2.1. Loading CEL in the carrier**

73 A constant ratio of Neusilin to solution 2:1 (w:v) was used so that solution fills most  
74 pores in Neusilin. An appropriate amount of CEL was first dissolved in DMF (a class II  
75 solvent) to form a solution with a desired concentration, which was then added dropwise into  
76 Neusilin while being mixed with a spatula. The concentration of CEL in DMF was varied to  
77 obtain 10-30% loading of CEL in Neusilin after drying. The moist powder was dried under  
78 house vacuum at 50°C overnight to remove DMF. Higher loadings of 40% and 50% CEL were  
79 achieved by repeated loading. The entrapment of CEL in Neusilin **was expected to be**  
80 essentially 100% since all of the solution went inside the carrier particles and CEL remained  
81 inside during drying. **This was confirmed by extracting CEL from the composite using both**  
82 **methanol and ethanol (data not shown).** The amount of residual DMF was not quantified in  
83 this work since it unlikely affects the assessed powder properties relevant to tablet  
84 manufacturing. However, residual solvents should be carefully monitored per the ICH  
85 guideline to make sure it does not exceed allowed safe levels in commercial tablets (ICH  
86 Guideline, 2021).

87 **2.2.2. Powder blending, tableting, and sticking assessment**

88 Five DC formulations prepared in this work comprised of two control formulations  
89 containing 20% as received crystalline CEL and three composite-based formulations (Table 1).  
90 CEL-Neusilin composites containing 10%, 30% and 50% CEL were used to prepare DC  
91 formulations containing 5%, 20% and 40% of overall CEL loading, respectively. These  
92 formulations contained 4% NaCMC as a disintegrant and 1% MgSt as a lubricant. All the

93 constituents were passed through a mesh #30 standard sieve before being blended in a mixer  
94 (Turbula, Glen Mills Inc., Clifton, NJ) for 3 min at 50 rpm. All blends were kept in ambient  
95 temperature and 33% relative humidity (RH), over a saturated MgCl<sub>2</sub> solution (O'Brien, 1948),  
96 for 2 days prior to compaction.

97 **Table 1.** Compositions of different DC formulations

Formulation	CEL (%)	CEL-Neusilin composite (%)	Neusilin (%)	MCC (%)	Lactose (%)	NaCMC (%)	MgSt (%)
Control 1	20	-	30	45	-	4	1
Control 2	20	-	-	45	30	4	1
Drug-carrier 5F	-	50 <sup>a</sup>	-	45	-	4	1
Drug-carrier 20F	-	66 <sup>b</sup>	-	29	-	4	1
Drug-carrier 40F	-	80 <sup>c</sup>	-	15	-	4	1

98 CEL loading in composite: *a* = 10%, *b* = 30%, and *c* = 50%.

99

100 Tablets were prepared by compressing a powder with a 9.5 mm flat-faced punches over  
101 a pressure range of 25-300 MPa on a compaction simulator (Presster; Metropolitan Computing  
102 Corp., NJ) at a tableting speed corresponding to 25 ms dwell time (corresponding to 49,300  
103 tablets/h) by simulating Korsch XL100 press (10 stations).

104 Sticking assessment was conducted using an upper punch with a removable flat-faced  
105 tip (round, 12.7 mm diameter) at a compaction pressure of 150 MPa to compress a total of 50  
106 tablets for each formulation. The punch tip was removed and weighed after every 10 tablets to  
107 determine the amount of mass adhered on to the punch face. After each removal, the punch tip  
108 was weighed three times on a digital balance with precision of 0.01 mg and the average was  
109 reported. The cumulative amount of mass adhered after 50 compactions was used to quantify  
110 sticking propensity.

111 **2.2.3. Powder flowability**

112 The flow properties of different formulations were accessed in triplicate using a ring  
113 shear cell tester under ambient condition (23°C and 20% - 25% RH). A preshear stress of 3  
114 kPa was used with normal stress of 500, 1000, 1500, 2000 and 2500 Pa during shear testing to  
115 construct a yield locus. Unconfined yield strength ( $f_c$ ) and major principal stress ( $\sigma_n$ ) were  
116 obtained from each yield locus by drawing Mohr's circles. The flowability index,  $ff_c$ , was  
117 calculated using Eq. (1).

$$118 \quad ff_c = \frac{\sigma_n}{f_c} \quad (1)$$

119 **2.2.4. Powder true density determination**

120 As water can be adsorbed by MCC, NaCMC and Neusilin, true density of each powder  
121 blend,  $\rho_t$ , was determined by nonlinear regression of tablet density ( $\rho$ ) vs.  $P$  data according to  
122 Eq. 2 (Sun, 2004). This method was more suitable than helium pycnometry for determining  
123 true density of moisture-containing powders (Chang and Sun, 2017; C. (Calvin) Sun, 2005;  
124 Sun, 2008).

$$125 \quad P = \frac{1}{C} \left[ \left( 1 - \varepsilon_c \right) - \frac{\rho}{\rho_t} - \varepsilon_c \ln \left( \frac{1 - \frac{\rho}{\rho_t}}{\varepsilon_c} \right) \right] \quad (2)$$

126 Accurate  $\rho_t$  is critical for calculating accurate  $\varepsilon$  using Eq. 3 for reliable analyses of powder  
127 compression performance (Paul et al., 2017a; C. C. Sun, 2005).

$$128 \quad \varepsilon = 1 - \frac{\rho}{\rho_t} \quad (3)$$

129 **2.2.5. Tablet diametrical breaking test**

130 Tablets were broken on a texture analyzer (Texture Technologies Corp., Surrey, UK)  
131 at 0.01 mm/s. Using Eq. 4, tablet tensile strength ( $\sigma$ ) was calculated from the breaking force  
132 ( $F$ ), tablet diameter ( $D$ ), and thickness ( $h$ ) (Fell and Newton, 1970).

133 
$$\sigma = \frac{2F}{\pi \cdot D \cdot h} \quad (4)$$

134 **2.2.6. Compressibility analysis**

135 The tablet porosity ( $\varepsilon$ ) - compaction pressure ( $P$ ) data were analyzed by nonlinear  
136 regression using Eq. 5 (Kuentz and Leuenberger, 1999).

137 
$$P = \frac{1}{C} \left[ \varepsilon - \varepsilon_c - \varepsilon_c \ln \left( \frac{\varepsilon}{\varepsilon_c} \right) \right] \quad (5)$$

138 A total of 12-15 tablets were compressed over 20-300 MPa for each formulation. Two  
139 parameters,  $1/C$  and  $\varepsilon_c$ , related to plasticity of the material and the critical porosity were  
140 obtained from curve fitting (Paul and Sun, 2017; Sun, 2017).

141 **2.2.7. Compactibility analysis**

142 Compactibility profile ( $\sigma$  vs.  $\varepsilon$ ) of each formulation was analyzed by non-linear  
143 regression of data using Eq. 6 (Ryshkewitch, 1953).

144 
$$\sigma = \sigma_0 e^{-b \cdot \varepsilon} \quad (6)$$

145 Where  $\sigma_0$  is the maximum tensile strength of the tablet attained at zero porosity and  $b$  is an  
146 empirical constant that quantifies sensitivity of  $\sigma$  to changes in  $\varepsilon$ .  $\sigma_0$  can be used to quantify the  
147 apparent bonding strength.

148 **2.2.8. Expedited friability analysis**

149 A separate set of 12-15 tablets were compressed over 25-300 MPa pressure range and  
150 subjected to impact and attrition in a friabilator (Pharma Alliance Group Inc., Model F2, Santa  
151 Clarita, CA) for 4 min at 25 rpm. Each tablet was weighed before and after the test and the  
152 percent weight loss for each tablet was determined and plotted against pressure to determine  
153 the minimum pressure required for obtaining tablets with weight loss of less than 1.0%.

154 **2.2.9. Solid State properties of the composites**

155      **2.2.9.1. PXRD**

156              Samples of Neusilin loaded with different amounts of CEL were scanned over a  $2\theta$   
157          range of  $5^\circ$ – $35^\circ$  on a wide angle X-ray diffraction instrument (X’Pert Pro; PANalytical Inc.,  
158          West Borough, MA) using Cu  $K\alpha$  radiation (45 kV and 40 mA) at a step size of  $0.0167^\circ$  and a  
159          dwell time of 1.15 s. The percent crystallinity of CEL in composites was determined by PXRD  
160          from the calibration plot of total area of all peaks over the  $2\theta$  range of  $5$ – $35^\circ$  as a function of  
161          proportion of crystalline CEL (5–60%) in a physical mixture with Neusilin. All the  
162          diffractograms were baseline corrected before peak area determination.

163      **2.2.9.2. Thermal analyses**

164              Degradation temperature was determined using a thermogravimetry analyzer (Q50; TA  
165          Instruments) by heating each sample at  $10\text{ }^\circ\text{C}/\text{min}$  to  $350\text{ }^\circ\text{C}$ . The maximum temperature in  
166          subsequent DSC experiments was kept lower than the degradation temperature to avoid  
167          contamination to the DSC cell of a differential scanning calorimeter (DSC; Q2000; TA  
168          Instruments, New Castle, DE). Samples were heated to  $180\text{ }^\circ\text{C}$  at a heating rate of  $10\text{ }^\circ\text{C}/\text{min}$   
169          under  $50\text{ mL}/\text{min}$  nitrogen gas purge. An empty aluminum pan was used as reference in all  
170          cases.

171      **2.2.9.3. Karl Fischer titration (KFT)**

172              KFT was performed using a Metrohm 831 KF coulometer, equipped with a Metrohm  
173          703 Ti Stand mixer (Metrohm Inc., Riverview, FL, USA). Briefly, 50 mg of sample was  
174          directly added to the thermostatic titration vessel containing reagent solution. The amount of  
175          water in a sample is determined voltammetrically by applying Faraday’s law to calculate the  
176          amount of water reacted with iodine, which is generated from an iodide containing reagent  
177          under constant current. A start and stop drift of  $10\text{ }\mu\text{g}/\text{min}$  was used. Each sample was tested  
178          in triplicate.

179 **2.2.9.4. IR spectroscopy**

180 IR spectra of CEL, Neusilin, and 30% CEL loaded composite were recorded on a FTIR  
181 spectrophotometer (Nicolet iS50; Thermo Scientific, Waltham, MA) with a built-in diamond  
182 attenuated total reflection (ATR). Data was collected over the range of 400-4000  $\text{cm}^{-1}$  at a  
183 resolution of 4  $\text{cm}^{-1}$  and 32 scans was processed to obtain an average spectrum using OMNIC  
184 9.2 software.

185 **2.2.9.5. Raman spectroscopy**

186 Raman spectra of CEL, Neusilin, and 30% CEL loaded composite were recorded on a  
187 Raman microscope (Alpha300 R, WITec, Ulm, Germany). A point of interest in the powder  
188 was focused with a 100x magnification lens and an average of two spectra was obtained using  
189 a source laser (532 nm) at an integration time of either 1s (for CEL and carrier) or 10 s (for the  
190 composite).

191 **2.2.9.6. In vitro dissolution**

192 The *in vitro* dissolution of different formulations was evaluated using an artificial  
193 stomach and duodenum (ASD) apparatus. It consists of two jacketed beakers with temperature  
194 controlled at 37 °C by a water bath. This apparatus simulates stomach and duodenum fluid  
195 transfer by regulating the flow using a programmatically controlled peristaltic pump  
196 (Masterflex, L/S Easy-Load II, Cole-Parmer, Vernon Hills, IL).

197 To simulate human physiological conditions in the fast state, experiments were  
198 conducted with 0.01 N HCl (pH = 2) for the stomach and 0.1 M sodium phosphate buffer (pH  
199 = 6.8) for the duodenum. The initial volume of the stomach chamber was 250 mL, which was  
200 decreased to 50 mL by first-order emptying with a half-life of 15 min. The duodenum volume  
201 was maintained at 30 mL throughout the entire study, achieved by setting a vacuum line in the  
202 duodenum chamber at a calibrated height. In addition, the chambers were infused with fresh

203 gastric or duodenal secretion liquid at 2 mL/min to mimic *in vivo* secretion processes. Drug  
204 concentration was monitored by a fiber optic UV/Vis probe. Mixing was achieved by an  
205 overhead paddle stirrer in the stomach chamber and a magnetic stirrer in the duodenum  
206 chamber. Calibration of all pumps and spectrometers were performed before each run. CEL  
207 release from the CEL-Neusilin 20F and Control 2 formulations were determined. Dissolution  
208 media were degassed to avoid generation of bubbles that might affect the real-time  
209 concentration detection with a UV dip probe.

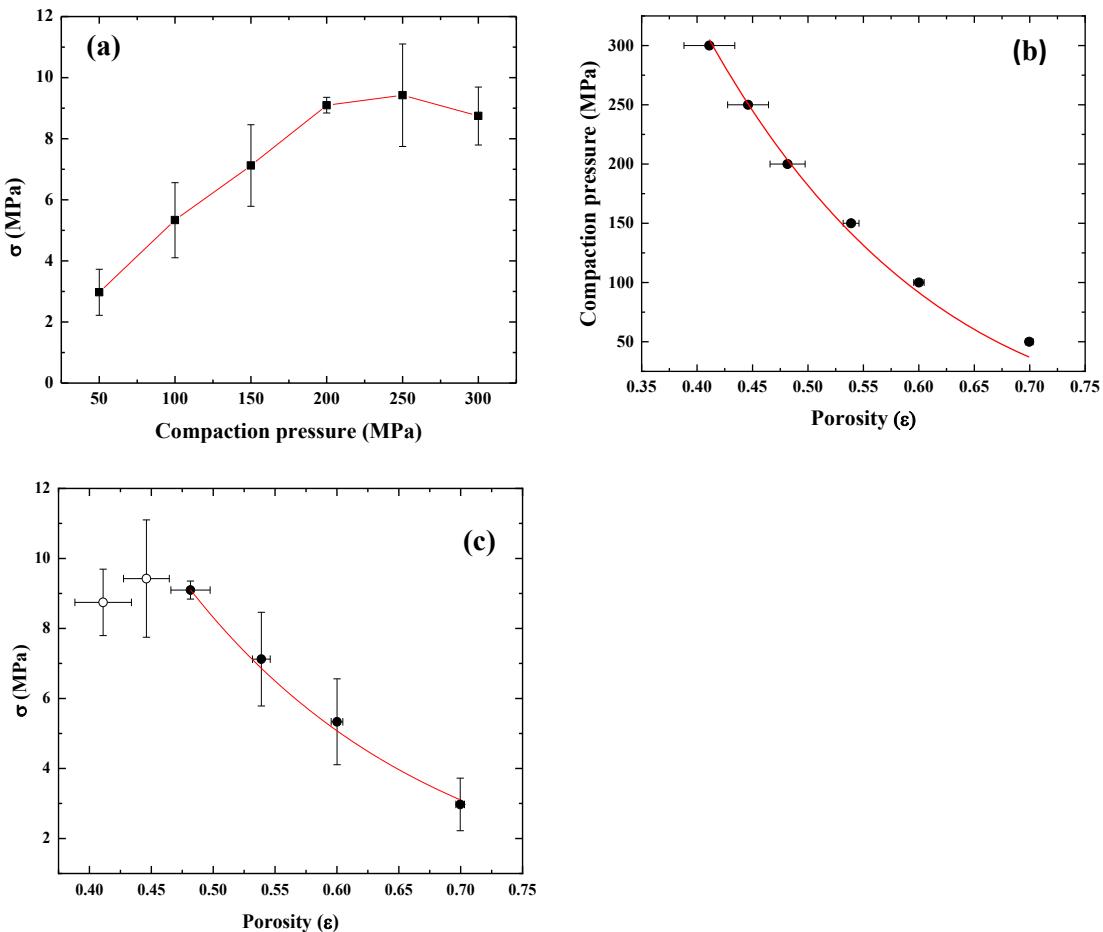
210 **2.2.9.7. Statistical analysis**

211 The statistical mean difference between two set of data of interest was determined by  
212 student's t-test method at a statistical significance level of  $p = 0.05$  using Origin Pro software  
213 (v17; Northampton, MA, USA).

214 **3. RESULTS AND DISCUSSION**

215 **3.1. Baseline characterization of the carrier**

216 Various mesoporous carriers from different manufacturers and different grades from  
217 the same manufacturer differ in solvent retention capacity (SRC) and compactibility. A greater  
218 SRC would allow a higher drug loading in the carrier while greater compactibility would favor  
219 the compression of tablets. We chose Neusilin US2, a magnesium aluminometasilicate, in this  
220 work because of its high SRC, neutral slurry pH (Sun et al., 2018), pharmaceutically acceptable  
221 safety profile for use in oral solid dosage form (Almotairy et al., 2023; Rowe et al., 2009), and  
222 excellent tabletability (Fig. 1a).



223

224 **Figure 1.** Compression properties of Neusilin US2 ( $n = 3$ ), a) Tableability, b) compressibility  
 225 and c) compactibility (two points at the lowest porosities were excluded from non-linear  
 226 regression because they are overcompressed).

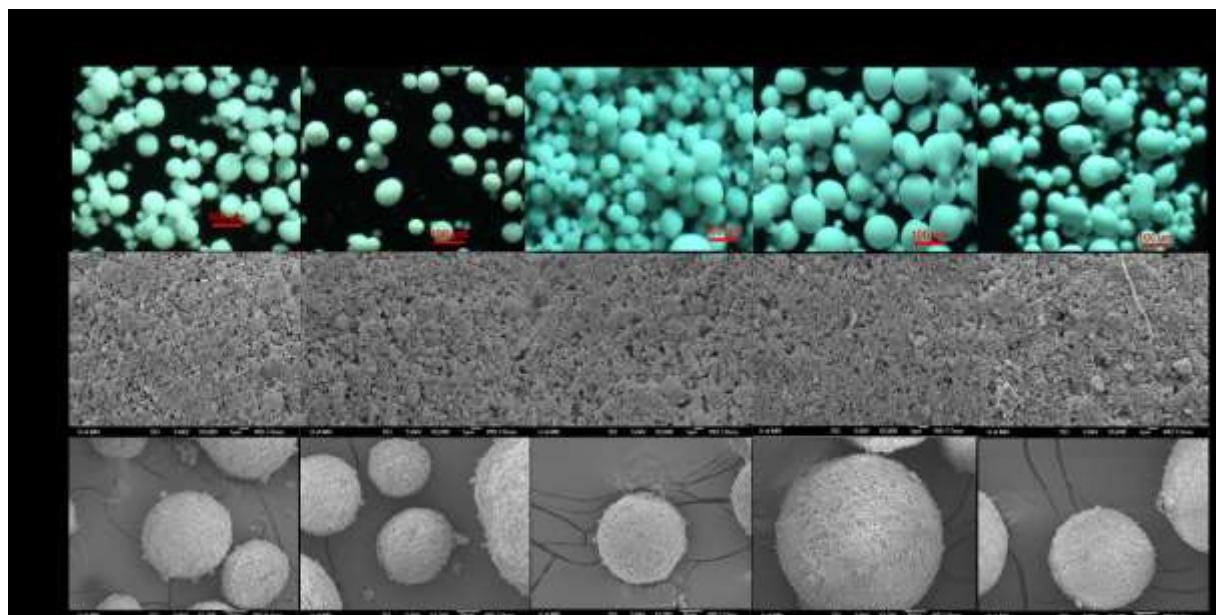
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228 Tablet porosity of Neusilin gradually decreases with increasing pressure, where a  
 229 porosity of 0.41 was attained at 300 MPa (Fig. 1b). The slow pore elimination of Neusilin  
 230 corresponds to a high  $1/C$  value ( $1,409 \pm 153$  MPa). By this measure, Neusilin is significantly  
 231 harder than lactose ( $1/C = 504 \pm 19$  MPa), significantly softer than anhydrous dicalcium  
 232 phosphate ( $1/C = 4203 \pm 77$  MPa), but close to a 60% DCPA and 40% mixture with MCC ( $1/C$   
 233 =  $1117 \pm 95$  MPa) (Paul and Sun, 2017; Vreeman and Sun, 2021). To address the expected  
 234 high ejection force during manufacturing of tablets of hard materials (Sun, 2015), 1% MgSt  
 235 was incorporated in all formulations in this work. Neusilin also showed high apparent bonding  
 236 strength ( $\sigma_0$ ) of ~90 MPa (Fig. 1c).

237

238 **3.2. Impact of CEL loading on particle size and morphology**

239 Neusilin particles are largely spherical, 50-100  $\mu\text{m}$  in diameter, with many open pores  
240 visible under high magnification (Fig. 2a). Loading of CEL up to 30% (w/w) did not cause  
241 obvious change in the size and shape of Neusilin (Fig. 2), indicating CEL was loaded inside  
242 the pores of Neusilin, instead of coating Neusilin particles. However, slight size enlargement  
243 was observed for composites of 40% and 50% CEL loadings prepared by the process of  
244 repeated drug loading-drying cycles (Fig. 2). No difference in surface texture was observed in  
245 the composites up to 30% loading, with pores clearly visible without extraneous particles.  
246 However, some fiber-like CEL particles were observed at 40% and 50% loadings (Fig. 2b,c).



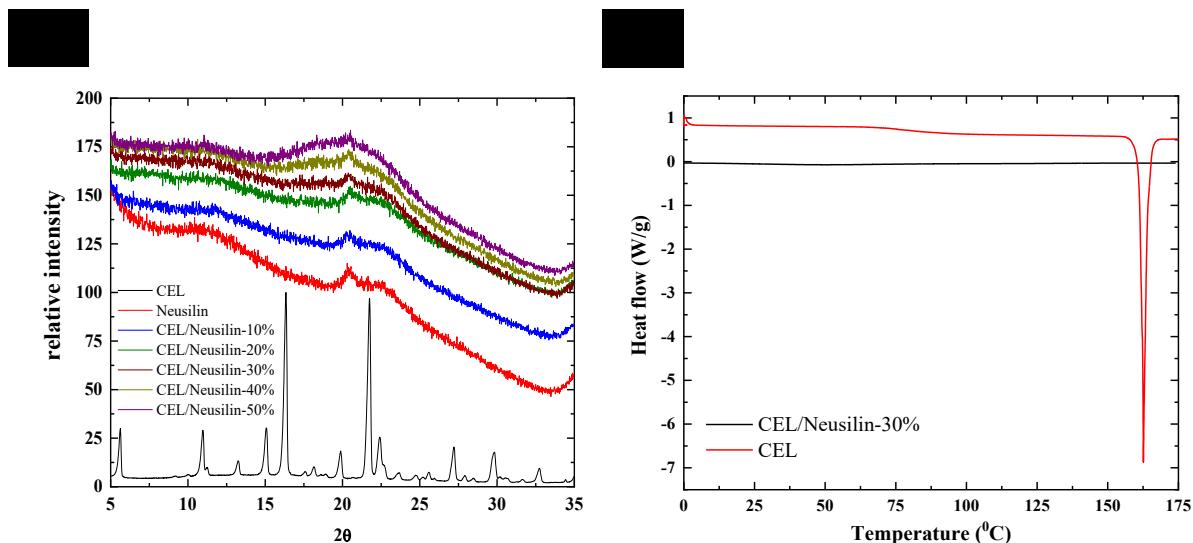
247

248 **Figure 2.** Morphology and surface textures of various CEL-Neusilin composites observed  
249 under (a) optical microscope (scale bar – 100  $\mu\text{m}$ ), and (b) SEM (scale bar = 1  $\mu\text{m}$ ), and (c)  
250 SEM (scale bar = 10  $\mu\text{m}$ ).

251

252 **3.3. Solid-state properties of the composite**

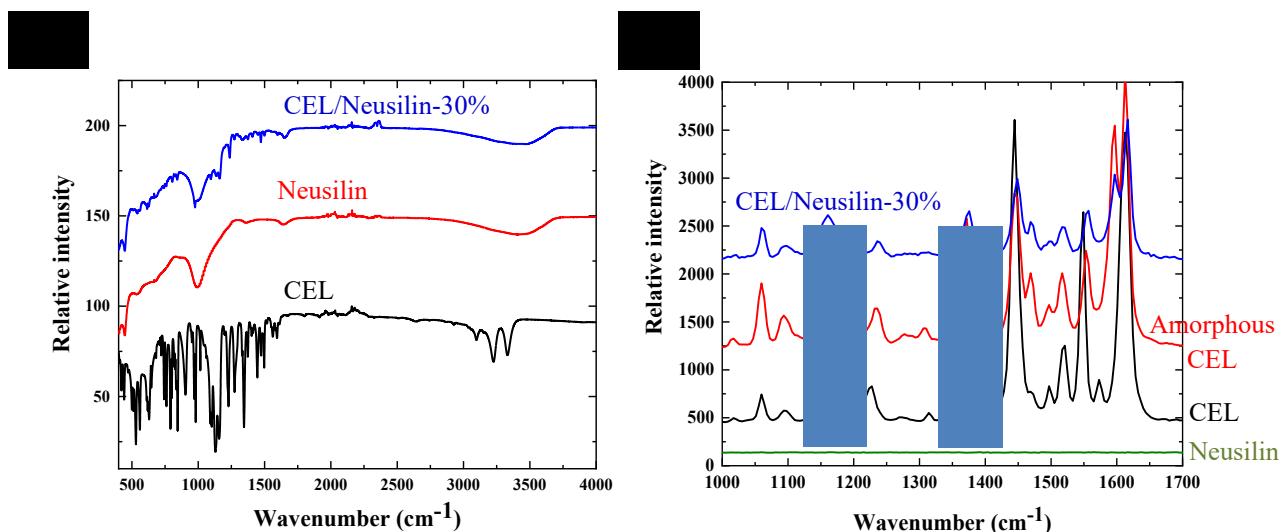
253 The absence of diffraction peaks in PXRD patterns suggests no crystalline CEL in  
 254 composites, containing up to 50% CEL (Fig. 3a). The DSC thermogram showed no melting  
 255 event for 30% CEL loaded composite. Melting of crystalline CEL (form III) at 160.8 °C was  
 256 consistent as previously reported (Wang and Sun, 2019). The absence of X-ray diffraction  
 257 peaks and melting events in the DSC thermograms of CEL-Neusilin composite also eliminate  
 258 the possibility of a crystalline DMF solvate of CEL (Bond and Sun, 2020; Chawla et al., 2003).



259  
 260 **Figure 3.** (a) X-ray diffractograms of CEL, Neusilin US2, and CEL-Neusilin composites (10-  
 261 50% loading), (b) DSC thermograms of crystalline CEL and CEL-Neusilin composite at 30%  
 262 loading.

263 The amount of DMF in the composite was probed by combining two methods, i.e. water  
 264 content determination by KFT and weight loss by TGA. The as-received Neusilin had ~16%  
 265 water content, which was slightly reduced to ~14% after vacuum drying (Fig. S2a). The CEL-  
 266 Neusilin 30% composite had ~8% water content by KFT. The TGA data (Fig. S2b) indicates  
 267 the corresponding weight loss of ~8% up to 220 °C, which is well above the boiling point of  
 268 DMF (153 °C). These findings suggest that the amount of DMF solvent in the composite was  
 269 negligible.

270 The IR spectra showed several signature peaks of the crystalline CEL and Neusilin (Fig.  
 271 4a). In CEL, the asymmetric and symmetric stretching frequencies of N-H were observed at  
 272 3332 and 3326  $\text{cm}^{-1}$  and aromatic stretching of C-H at 3097  $\text{cm}^{-1}$ . The S=O (1331 and 1062  
 273  $\text{cm}^{-1}$ ) and C-F stretching (1403 and 1374  $\text{cm}^{-1}$ ) and N-H bending (1596 and 1562  $\text{cm}^{-1}$ ) were  
 274 also observed in the CEL fingerprint region. Neusilin exhibited broad O-H signature peak at  
 275 3415  $\text{cm}^{-1}$ , corresponding to the presence of several silanol groups. In addition, a characteristic  
 276 peak of Al-O-Si group was observed at 993  $\text{cm}^{-1}$ . In contrast, the spectrum of CEL-Neusilin  
 277 composite appears to be that of Neusilin overlaid with weak signals of CEL in the fingerprint  
 278 region. This could be attributed to only 30% presence of CEL in the composite and limited  
 279 penetration depth of the IR light rays into the composite particles, since CEL remains inside  
 280 Neusilin particles.



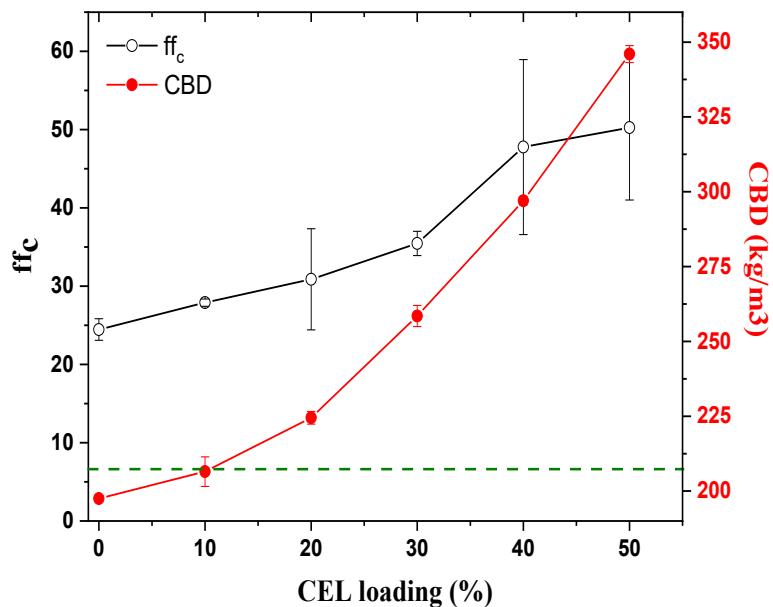
281  
 282 **Figure 4.** (a) IR and (b) Raman spectra for crystalline CEL, Neusilin, and CEL-Neusilin  
 283 composite. Raman spectrum of amorphous CEL is shown for comparison, where regions of  
 284 spectroscopic difference are shaded.

285  
 286 Raman spectra of various powders were also collected to further gain insights into the  
 287 nature of CEL in the composites (Fig. 4b). Neusilin did not show any Raman signal. The

288 symmetric S=O stretching of crystalline CEL was observed at 1160-1200  $\text{cm}^{-1}$  where  
289 broadened peaks with low intensity were observed for CEL-Neusilin composite, matching with  
290 that of amorphous CEL obtained by cryomilling. This is consistent with the amorphous nature  
291 of CEL in the composite, suggested by PXRD and DSC. Similar observation was noted for C-  
292 F stretching at 1230  $\text{cm}^{-1}$  and N-H bending vibrations at 1560  $\text{cm}^{-1}$  (Andrews et al., 2010). The  
293 blue shift of N-H bending for both the composite and amorphous CEL to a higher frequency  
294 suggests N-H groups are involved in stronger interactions than those in the crystalline CEL.  
295 CEL in the composite and the amorphous form both exhibited a doublet around 1620  $\text{cm}^{-1}$   
296 (Andrews et al., 2010), which could be ascribed to combined vibration of C-C with amino  
297 stretching (Tammer, 2004). Overall, the spectroscopic data support that CEL inside Neusilin  
298 particles is in amorphous state.

299 **3.4. Flowability of composite powders**

300 The flowability indices,  $ff_c$ , of all the composites were significantly higher than that of  
301 Avicel PH102 (Fig. 5), implying excellent flowability (Sun CC, 2010). The  $ff_c$  values increased  
302 with increasing CEL loading, which is in part due to increased particle density with increasing  
303 CEL loading in Neusilin. This is confirmed by the increase in consolidated bulk density (CBD)  
304 with increasing CEL loading (Fig. 5).



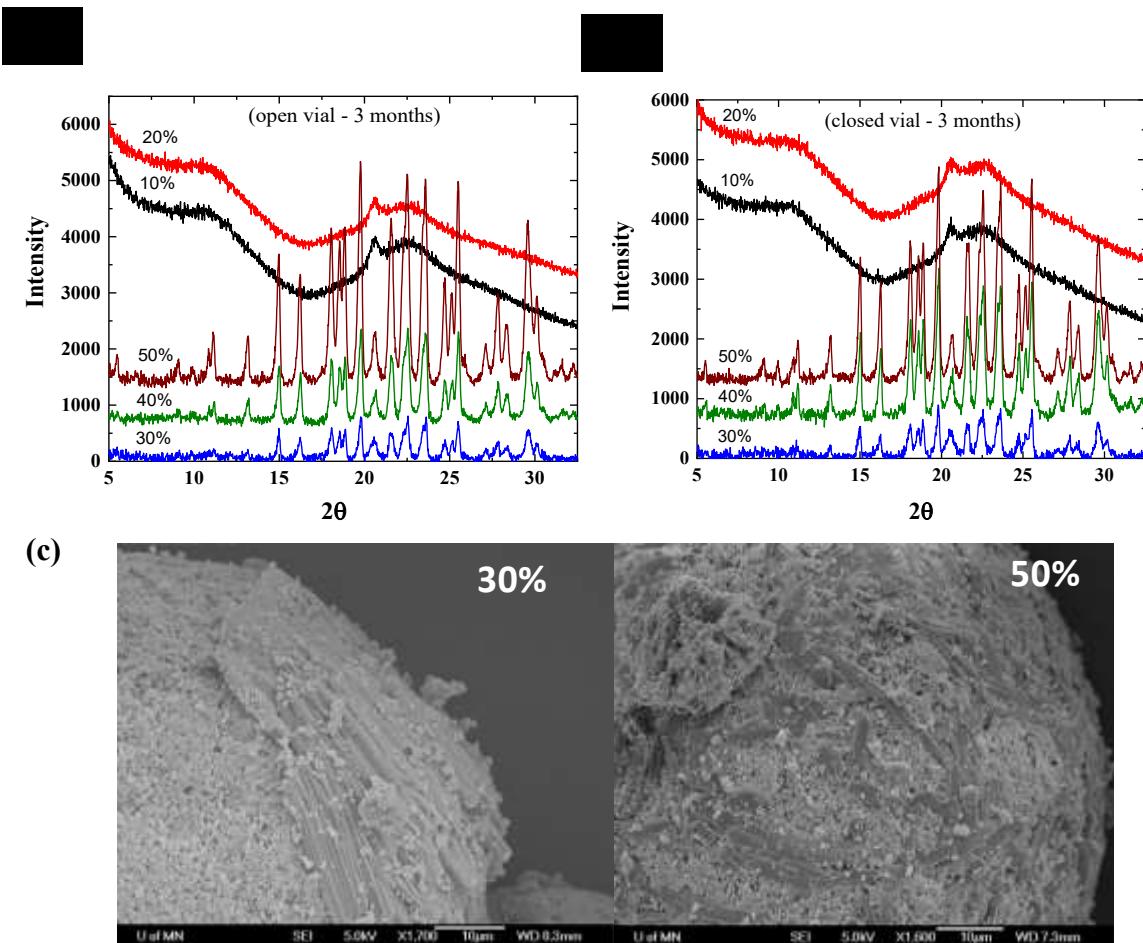
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306 **Figure 5.** Flowability index and consolidated bulk density (CBD) of various Neusilin and  
 307 CEL-Neusilin composites ( $n = 3$ ). The horizontal dashed line indicates the  $ff_c$  of Avicel  
 308 PH102.

309

310 **3.5. Physical stability of composites**

311 The physical stability of composites with different CEL loadings was studied under  
 312 accelerated stability conditions at 40 °C/75% RH per the ICH guideline. Both open and closed  
 313 vials were used to allow the separation of the influence of heat and moisture on physical  
 314 stability of composites. After 3 months of exposure to the stressed stability conditions in both  
 315 open and closed vials, 10% and 20% CEL loaded composites did not show any crystalline  
 316 peaks in their X-ray diffractograms, indicating excellent physical stability (Fig. 6a,b).  
 317 However, crystalline CEL peaks appeared for composites containing 30% or more CEL in both  
 318 open and closed vials, where peak intensity increased with increasing CEL loading (Fig. 6a,b).



319

320 **Figure 6.** Different CEL-Neusilin after 3 months of exposure to stressed stability conditions  
 321 (a) PXRD of samples under open conditions; (b) PXRD of samples under closed conditions  
 322 and (c) SEM of samples under open conditions.

323

324 Using the calibration curve constructed with a set of physical mixtures of crystalline  
 325 CEL and Neusilin in different proportions (Fig. S1a), the percent crystallinity of samples after  
 326 storage under different stability conditions was estimated. Under open condition, the percent  
 327 crystallized CEL at the end of a 3 months period, was essentially the same as that after 6 months  
 328 (Table 2). Thus, crystallization of CEL had mostly completed after 3 months under this  
 329 condition.

330 **Table 2.** Percent crystallinity of CEL-Neusilin composites under different stressed stability  
 331 conditions.

CEL in composite (%)	% CEL crystallized			
	3 months		6 months	
	closed vial	open vial	closed vial	open vial
10	0	0	0	0
20	0	0	0	0
30	2.3 ± 0.02	2.6 ± 0.4	2.4 ± 0.01	2.6 ± 0.01
40	5.3 ± 0.8	7.9 ± 0.7	6.5 ± 0.3	7.6 ± 0.4
50	12.4 ± 1.8	14.8 ± 0.7	12.8 ± 1.3	13.5 ± 0.5

332

333 SEM revealed the appearance of elongated features on the surface of the carrier in the  
 334 composites containing 30% and 50% CEL (Fig. 6c), which encompassed a greater area for 50%  
 335 CEL composite. In combination with the X-ray data, these new features are attributed to  
 336 crystalline CEL formed during stability storage. The crystallized CEL content increased with  
 337 increasing CEL loading in the composites (Table 2). The extent of crystallization was only  
 338 slightly lower in closed vials than that in open vials, implying that 75% RH only minimally  
 339 impacted crystallization of amorphous CEL after a prolonged period. It is possible that  
 340 crystallization at earlier time points could be faster under the open conditions. However, this  
 341 requires a separate stability study to establish. It is useful to point out that the PXRD data  
 342 suggested incomplete crystallization of amorphous CEL in 30%, 40%, and 50% CEL-loaded  
 343 composites. For example, in 30% CEL-loaded composite after storage for 6 months, 2.4%  
 344 (under closed conditions) and 2.6% (under open conditions) crystalline CEL was detected i.e.  
 345 27.6% and 27.4% of CEL remained amorphous. For the 40% CEL-loaded composite, 33.5%  
 346 (closed vial) and 32.4% (open vial) CEL remained amorphous. Similarly, for the 50% CEL-  
 347 loaded composite, 37.2% (closed vial) and 36.5% (open vial) CEL remained amorphous.  
 348 These results are consistent with the observation that no crystalline CEL was detected in  
 349 composites containing 10% and 20% CEL because they affirm that up to 27% amorphous CEL  
 350 in composites remain physically stable even under stressed stability conditions.

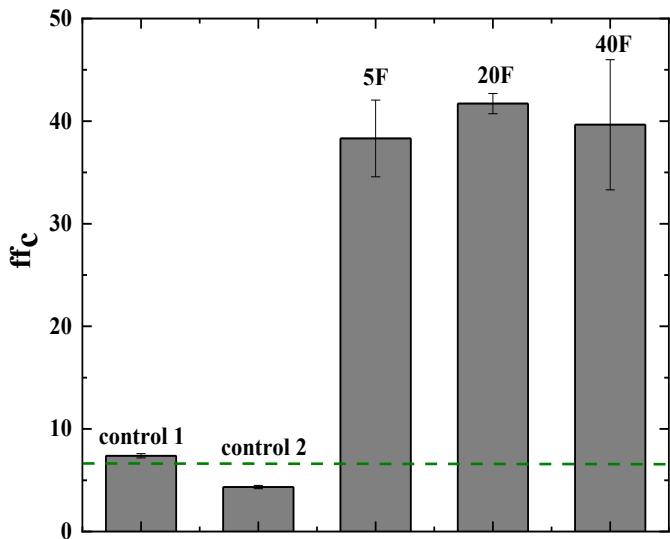
351 **3.6. Evaluation of the suitability of composites for DC formulations**

352 Characterization results of the CEL-Neusilin composites suggest good physical  
353 stability and excellent powder properties, which make them suitable for developing a DC tablet  
354 formulation. To this end, the five DC formulations in Table 1 were prepared and systematically  
355 evaluated based on tablet manufacturability and key performance of tablets.

356

357 **3.6.1. Flowability of formulations**

358 For both control formulations, 20% CEL loading drastically reduces the flowability,  
359 despite only DC grade excipients with good flowability were used. The flowability of the  
360 control 1 formulation was comparable to that of Avicel PH102 (Fig. 7), indicating its marginal  
361 flowability for a high speed tableting process (Sun CC, 2010). When Neusilin was replaced  
362 by LM in the control 2 formulation, the flowability was significantly lower than that of Avicel  
363 PH102, indicating inadequate flowability to sustain a high speed tablet manufacturing process.  
364 In contrast, all three CEL-Neusilin composite based formulations had similar and excellent  
365 flow, as shown by very high  $ff_c$  values (Fig. 7). The flowability of these formulations are  
366 expected to be excellent for a high speed tablet manufacturing process. The insensitivity of  
367 flowability to CEL loading in composite is consistent with the fact that CEL remained in the  
368 pores of the carrier particles. Thus, its impact on the size and morphology of composite  
369 particles is small (Fig. 2).



370  
371 **Figure 7.** Flowability index of control and composite based CEL formulations (n = 3). The  
372 horizontal dashed line indicates the *ffc* of Avicel PH102.

373

374 **3.6.2. Tableting performance of formulations**

375 The tableting performance of the five formulations followed the ascending order of control 2 <  
376 control 1 < CEL-Neusilin-40F < CEL-Neusilin -20F  $\approx$  CEL-Neusilin-5F (Fig. 8a). The  
377 apparent bonding strength of the formulations, assessed by  $\sigma_0$ , also followed a similar order,  
378 control 2 < control 1 < CEL-Neusilin-40F < CEL-Neusilin-5F < CEL-Neusilin-20F (Fig. S3c  
379 and Table 3). The significantly better tableting performance of control 1 formulation than control 2  
380 formulation (Fig. 8a) is attributed to the excellent tableting performance of Neusilin than lactose. Among  
381 the three composite based formulations, the tableting performance of 5F and 20F formulation was similar  
382 in the entire pressure range (Fig. 8a). Tablet tensile strength of 40F formulation is similar to  
383 5F and 20F formulations when compaction pressure is < 100 MPa, but significantly lower when  
384 pressure is > 150 MPa (Fig. 8a). It is useful to note that tablet porosity of the 40F formulation  
385 is always lower than that of 5F formulation, which means higher bonding area of the 40F  
386 formulation than the 5F formulation. Hence, the lower tableting performance of the 40F formulation at

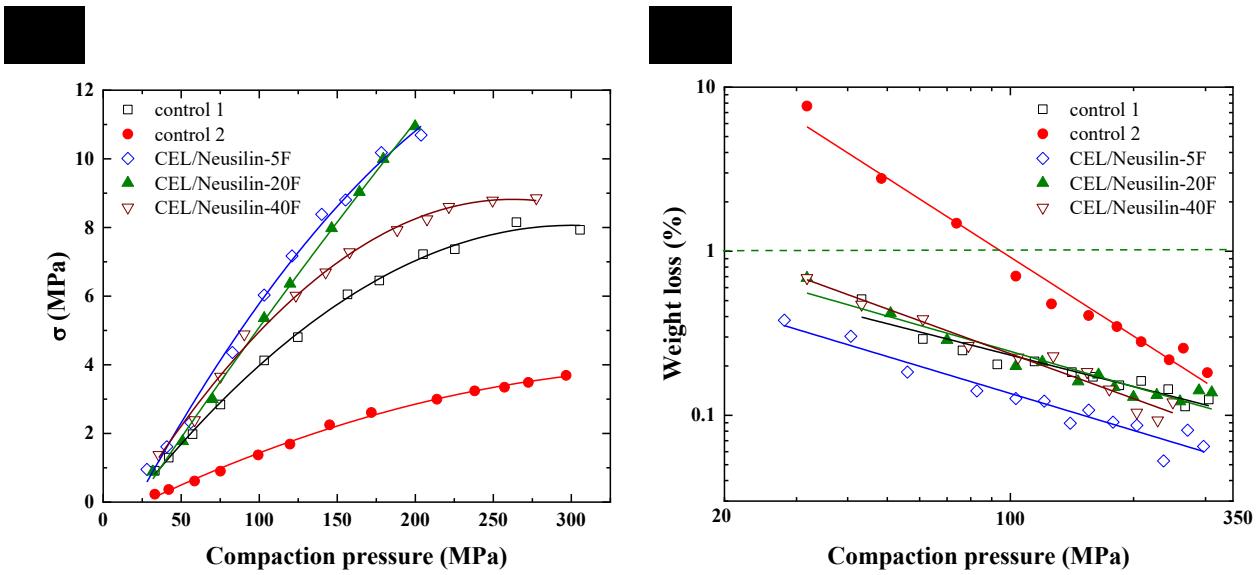
387 high pressures is attributed to its lower apparent bonding strength (Table 2), according to the  
388 bonding area-bonding strength interplay model (Osei-Yeboah et al., 2016; Paul et al., 2020).

389 **Table 3.** Compressibility and compactibility analysis of different formulations of CEL.  
390 Standard errors of fitting are shown in parentheses.

Formulations	KL fitting			Ryshkewitch fitting	
	1/C (MPa)	$\epsilon_c$	$R^2$	$\sigma_0$ (MPa)	$R^2$
Control 1	292.0 (14.6)	0.82 (0.02)	0.997	11.7 (0.6)	0.97
Control 2	298.8 (16.2)	0.84 (0.02)	0.996	4.1 (0.06)	0.995
CEL-Neusilin-5F	199.9 (4.9)	0.88 (0.01)	0.999	38.5 (4.3)	0.968
CEL-Neusilin-20F	138.0 (13.9)	0.7 (0.04)	0.992	66.3 (10.7)	0.975
CEL-Neusilin-40F	379.4 (8.1)	0.89 (0.008)	0.999	20 (2.0)	0.961

391  
392 As expected, friability is lower when tableability is higher, owing to their stronger  
393 resistance to particle dislodging during impact (Fig. 8b). Only control 2 formulation showed  
394 more than 1.0% friability below 100 MPa pressure, while all other formulations could produce  
395 tablets that pass the USP friability criterion (< 1.0%) even when compressed at pressures as  
396 low as 25 MPa. The ability to make sufficiently strong tablets at relatively low pressures is  
397 beneficial for APIs that are sensitive to mechanical stress, such as solid form change (Fabbiani  
398 and Pulham, 2006) and loss of biological activities of therapeutic microbial or fragile proteins  
399 (Klukkert et al., 2015).

400



401

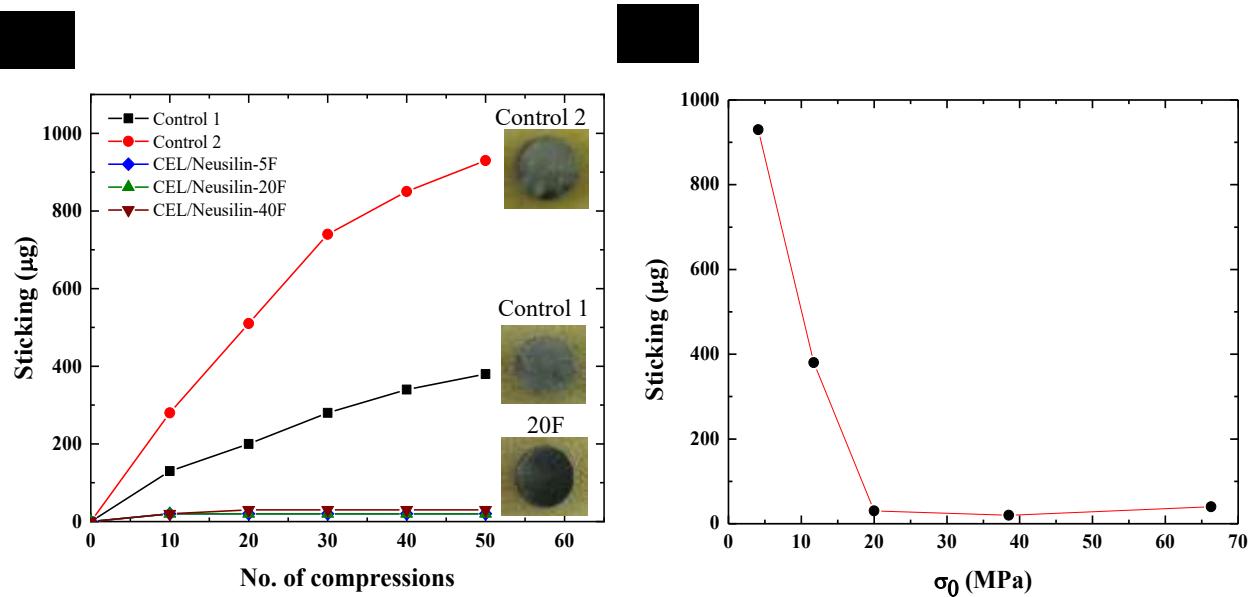
402 **Figure 8.** Tableting (a) and friability (b) profiles of different formulations. Each point  
403 represents result from a single tablet.

404

405 The consolidation behaviors of the five formulations differed as indicated by the  
406 parameter  $1/C$  (Table 3 and Fig. S3), which quantifies plasticity of a powder (Kuentz and  
407 Leuenberger, 1999; Paul and Sun, 2017). By this measure, replacing LM with Neusilin did not  
408 affect the deformability of Control 1 and 2 formulations because of their similar  $1/C$  values  
409 (Table 3). The plasticity of the three CEL-Neusilin composite based formulations follows the  
410 order of 20F > 5F > 40F (Table 3). With a lower proportion of plastic MCC in the formulation,  
411 the plasticity of CEL-Neusilin-20F formulation was surprisingly higher than 5F formulation.  
412 A possible explanation is that a composite containing more CEL may be more plastic so that it  
413 compensates the impact by the lower amount of MCC. However, the  $1/C$  of CEL Neusilin-  
414 40F (containing 80% of composite of 50% CEL loading) was higher than those of 5F and 20F  
415 formulations. Thus, the overall plasticity of these formulations is a complex of interplay  
416 between the impact of CEL loading on plasticity of the composite and weight fraction of the  
417 composite in formulation. A dedicated study would be required to fully understand the  
418 underlying mechanisms.

419 **3.6.3. Punch sticking propensity of formulations**

420 CEL exhibits a high punch sticking propensity, where severe punch sticking was  
421 observed in DC formulations at  $\leq 20\%$  CEL loading (Paul and Sun, 2018). Hence, punch  
422 sticking is a key manufacturing problem that must be addressed in order to develop a DC tablet  
423 formulation of CEL. The high sticking propensity of CEL was confirmed in this work using  
424 control formulations 1 and 2. The lower sticking propensity of control 1 is consistent with its  
425 higher tabletability (Fig. 8a). For the same API, a formulation having a stronger bonding among  
426 particles in tablet tends to exhibit lower punch sticking (Paul and Sun, 2018). However, punch  
427 sticking is still severe even for Control 1 formulation. In contrast, formulations of CEL-  
428 Neusilin composite exhibited no sticking to punch (clean punch tip after 50 tablets), even for  
429 the formulation containing 40% CEL (Fig. 9a). When all five formulations are considered, the  
430 severity of punch sticking followed a nonlinear negative dependence with  $\sigma_0$  (Fig. 9b).  
431 Although the stronger bonding strength of the three composite based formulations does favor  
432 lower punch sticking propensity of CEL, a more important reason is the fact that CEL residing  
433 inside the pores of Neusilin does not come in contact with punch tip, unless extensive fracture  
434 of the composite particle occurred. Even in that case, the probability of CEL directly interacts  
435 with punch tip is still low. The assessment results clearly show that co-processing CEL with  
436 Neusilin is effective in mitigating, if not eliminating, punch sticking of CEL. The strategy  
437 should be universally applicable for reducing punch sticking problem of other APIs.



438

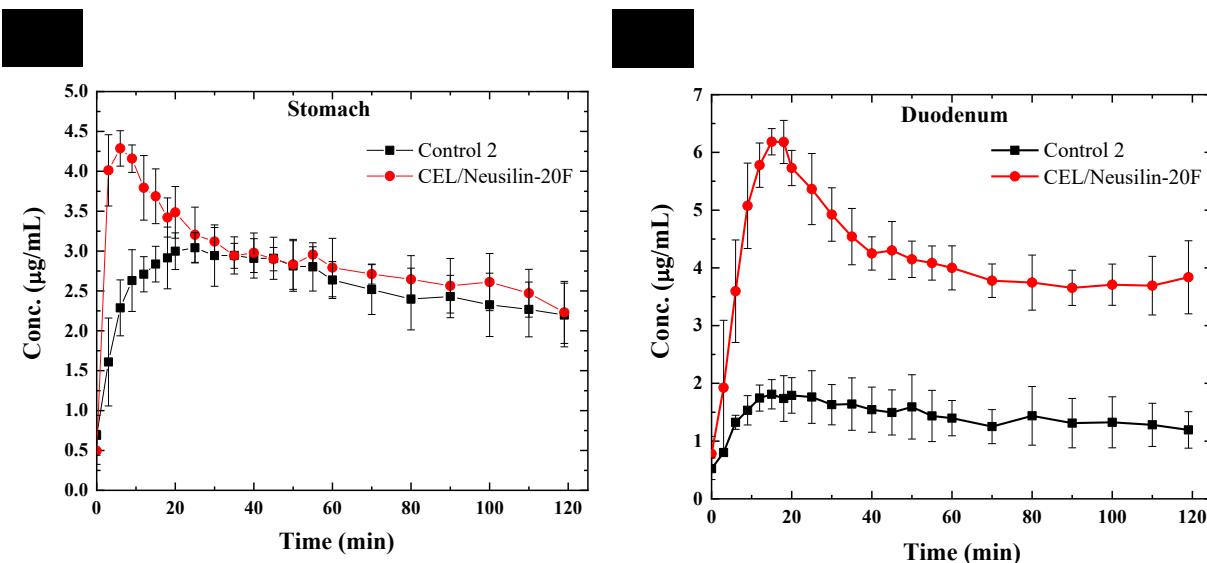
439 **Figure 9.** Dependence of sticking on (a) number of compressions (each point represents result  
440 from a single tablet.) and (b) bonding strength of five different DC formulations of CEL.

441

#### 442 3.6.4. Dissolution performance of formulations

443 The dissolution performance of a representative composite based CEL tablet, CEL-  
444 Neusilin-20F, was evaluated using an artificial stomach and duodenum apparatus (ASD). The  
445 ASD more closely mimics the physiological conditions in human guts than the USP dissolution  
446 apparatus because it simulates the pH in stomach and duodenum and transfer both liquid and  
447 particles from stomach to duodenum. The drug concentration – time profile in the duodenum  
448 chamber was shown to be proportional to bioavailability of BCS class II APIs (Carino et al.,  
449 2010, 2006). Thus, it is a reliable *in vitro* dissolution method for rank ordering *in vivo*  
450 bioavailability of different formulations of the same BCS II API. At 20% loading, the  
451 dissolution profile of the CEL-Neusilin-20F composite formulation in the stomach chamber is  
452 higher than that of the control 2 formulation (Fig. 10a). The maximum CEL concentration at  
453 ~10 min is 4.3 μg/mL, which is 40% higher than the peak concentration at ~ 23 min for the  
454 control 2 formulation. The higher dissolution rate of the CEL-Neusilin formulation is  
455 attributed to the amorphous nature of CEL (Fig. 3). After 10 min, the CEL concentration

456 decreased quickly to become approximately the same as that from the control formulation at  
457 30 min, indicating crystallization of CEL from the supersaturated solution.



458  
459 **Figure 10.** Dissolution profiles of tablets ( $n = 3$ ) containing a total of 20% CEL (either CEL-  
460 Neusilin or as-received) in (a) stomach chamber and (b) duodenum chamber.

461  
462 The concentration-time profile of CEL in the duodenum chamber showed a marked  
463 difference between the two formulations, where the composite based formulation showed  
464 significantly higher concentration profile and the area under the time-concentration profiles  
465 (AUC) (Fig. 10b). It reached a peak concentration of  $6.5 \mu\text{g/mL}$  at  $\sim 20$  min, which is more  
466 than 3 times that of the control 2 formulation ( $2 \mu\text{g/mL}$ ). The AUC of the CEL-Neusilin  
467 composite based formulation (20F) is also approximately 3 times that of the control 2  
468 formulation. Even without further formulation optimization, the composite based formulation  
469 already exhibits much improved dissolution performance than the control formulation. If  
470 desired, a higher CEL concentration of the composite based formulation could be achieved  
471 through the general strategy of incorporating a sufficient amount of an effective precipitation

472 inhibitor in the formulation (Bi et al., 2011; Budiman et al., 2022; Guo and Sun, 2022; Ozaki  
473 et al., 2013; Yamashita and Sun, 2019).

474

#### 475 **4. Conclusion**

476 This study shows the high potential of particle engineering to enable direct compression  
477 formulation of a challenging API, CEL, by forming composites with a mesoporous carrier. Up  
478 to 50% CEL could be loaded into Neusilin US2 by repeated loading, which remained  
479 amorphous after loading. Amorphous CEL undergoes partial crystallization under stressed  
480 conditions when the CEL loading was  $\geq 30\%$  (w/w), but remained physically stable at  $\leq 20\%$   
481 CEL loading even under stressed stability conditions. When the composite was used in a  
482 formulation, flowability, tablettability, and punch sticking performance were all excellent,  
483 indicating a high possibility to developing a DC formulation amenable for high speed tablet  
484 manufacturing. By the measure of AUC in the duodenum, the bioavailability of the composite  
485 based tablet formulation (20% drug loading) is more than 3 times that of the formulation  
486 containing the same amount of crystalline CEL. Further property enhancement of the  
487 composite is possible, if needed. Thus, this approach may find broad applications in  
488 developing robust DC tablet formulations with excellent manufacturability and dissolution  
489 performance.

#### 490 **Acknowledgement**

491 We thank Kunlin Wang for providing the Raman data of amorphous celecoxib.

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