

High-resolution structures with bound Mn^{2+} and Cd^{2+} map the metal import pathway in an Nramp transporter

Shamayeeta Ray¹, Samuel P Berry¹, Eric A Wilson², Casey H Zhang^{1†}, Mrinal Shekhar³, Abhishek Singharoy², Rachelle Gaudet^{1*}

¹Department of Molecular and Cellular Biology, Harvard University, Cambridge, United States; ²School of Molecular Sciences, Arizona State University, Tempe, United States; ³Broad Institute, Cambridge, United States

Abstract Transporters of the Nramp (Natural resistance-associated macrophage protein) family import divalent transition metal ions into cells of most organisms. By supporting metal homeostasis, Nramps prevent diseases and disorders related to metal insufficiency or overload. Previous studies revealed that Nramps take on a LeuT fold and identified the metal-binding site. We present high-resolution structures of *Deinococcus radiodurans* (Dra)Nramp in three stable conformations of the transport cycle revealing that global conformational changes are supported by distinct coordination geometries of its physiological substrate, Mn^{2+} , across conformations, and by conserved networks of polar residues lining the inner and outer gates. In addition, a high-resolution Cd^{2+} -bound structure highlights differences in how Cd^{2+} and Mn^{2+} are coordinated by DraNramp. Complementary metal binding studies using isothermal titration calorimetry with a series of mutated DraNramp proteins indicate that the thermodynamic landscape for binding and transporting physiological metals like Mn^{2+} is different and more robust to perturbation than for transporting the toxic Cd^{2+} metal. Overall, the affinity measurements and high-resolution structural information on metal substrate binding provide a foundation for understanding the substrate selectivity of essential metal ion transporters like Nramps.

***For correspondence:**
gaudet@mcb.harvard.edu

Present address: †Grossman School of Medicine, New York University, New York, United States

Competing interest: The authors declare that no competing interests exist.

Funding: See page 24

Preprinted: 09 September 2022

Received: 07 October 2022

Accepted: 06 April 2023

Published: 11 April 2023

Reviewing Editor: Merritt Maduke, Stanford University School of Medicine, United States

© Copyright Ray et al. This article is distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use and redistribution provided that the original author and source are credited.

Editor's evaluation

This manuscript provides fundamental new insight into protein conformational transitions underlying the transport mechanism of Nramps, an important and widespread transporter family that facilitates the uptake and movement of essential transition metals. Eight new crystallographic structures of the prokaryotic homolog DraNramp in a variety of ligand-bound and conformational states, along with companion molecular dynamics simulations and metal binding and transport assays, provide compelling evidence supporting most of the conclusions. These findings will be of broad interest to scientists studying transport mechanisms and ligand recognition.

Introduction

Transition metal ions like Mn^{2+} and Fe^{2+} are essential for various metabolic processes in all living cells and are usually required in low intracellular concentrations for optimal activity (Andrews, 2002; Bozzi and Gaudet, 2021). Excess or deficiency of transition metal ions leads to diseases (Bleackley and Macgillivray, 2011; Nies and Grass, 2009). For example, Fe^{2+} deficiency causes anemia and neurodegenerative diseases, whereas Fe^{2+} overload increases the risk of cancer by generating toxic reactive oxygen species (ROS) and mutations (Ekiz et al., 2005; Jung et al., 2015). Mn^{2+} overload in the

brain is linked to neurological disorders and deficiency causes metabolic defects and impairs growth (Budinger et al., 2021; Pittman, 2005). Other transition metals, like Cd^{2+} and Hg^{2+} , are toxic and their accumulation affects health by disrupting the physiological levels of essential metals or displacing them in enzyme active sites, thus inhibiting the proteins, or changing their activity (Andrews, 2002; Lin et al., 2009). Cells and organisms have evolved strategies to maintain metal ion homeostasis via highly regulated transport and storage processes (Bleackley and Macgillivray, 2011; Cellier and Gros, 2004; Nies and Grass, 2009).

Natural resistance-associated macrophage proteins (Nramps) are ubiquitous importers of Fe^{2+} and Mn^{2+} across cellular membranes into the cytosol (Bozzi and Gaudet, 2021; Cellier and Gros, 2004; Nevo and Nelson, 2006). In humans, Nramp1 extrudes essential metals from phagosomes of macrophages to aid in killing engulfed pathogens, and Nramp2 (DMT1) is expressed at low levels in the endosomes of all nucleated cells and imports Mn^{2+} and Fe^{2+} into the cytosol (Pujol-Giménez et al., 2017; Skamene et al., 1998; Vidal et al., 1993). Plant and fungal Nramps aid in Fe^{2+} and Mn^{2+} uptake and trafficking, and bacterial Nramps are involved in the acquisition of Mn^{2+} , an essential nutrient (Bozzi and Gaudet, 2021). In addition to the physiological substrates Fe^{2+} and Mn^{2+} , Nramps can also transport toxic metals like Cd^{2+} and Hg^{2+} but exclude the abundant alkaline earth metals like Mg^{2+} and Ca^{2+} (Bozzi and Gaudet, 2021).

Recent bacterial Nramp structures reveal a LeuT fold, three stable conformations (outward-open, occluded, and inward-open), and identify the metal-binding site residues, including conserved aspartate, asparagine, and methionine residues (Bozzi et al., 2016b; Bozzi et al., 2019b; Ehrnstorfer et al., 2014; Ehrnstorfer et al., 2017). The metal-binding methionine is essential to select against alkaline earth metals (Bozzi et al., 2016a). This finding is corroborated by the fact that a bacterial Nramp homolog which lacks a metal-binding methionine, NRMT (Nramp-related Mg^{2+} transporter), can transport Mg^{2+} (Ramanadane et al., 2022). However, little is known about whether the canonical Nramps can mechanistically distinguish between their physiological substrates (Fe^{2+} and Mn^{2+}) from non-essential ones like Cd^{2+} within their broad spectrum of transition metal substrates. Functional studies on *Deinococcus radiodurans* (Dra)Nramp revealed that Mn^{2+} and Cd^{2+} transport differ in their dependence on pH, proton flux, and membrane potential (Bozzi et al., 2019a; Bozzi et al., 2019b). However, we lack high-resolution structural information on binding of different metals to explain these differences.

We present high-resolution structures of DraNramp in three conformations in both Mn^{2+} -bound and metal-free states, providing the first molecular map of the entire Mn^{2+} transport cycle. The structures along with molecular simulations reveal that Nramps achieve alternate access during transport by adopting distinct Mn^{2+} -coordination spheres in different conformations. These different conformations are also supported by dynamic rearrangements of key polar-residue networks that gate the inner and outer vestibules. This Mn^{2+} transport cycle also informs on the transport of Fe^{2+} , the other common physiological Nramp substrate, because Fe^{2+} and Mn^{2+} have similar coordination preferences and chemical properties (Bozzi and Gaudet, 2021; Davidsson et al., 1989; Kawabata, 2019; Liu et al., 2021). Comparisons with an additional high-resolution structure of DraNramp bound to a non-physiological substrate, Cd^{2+} , and complementary binding and transport measurements and mutational analyses, suggest that Nramps can distinguish physiological from toxic substrates through thermodynamic differences in the conformational landscape of the transport cycle.

Results

DraNramp transports a mostly dehydrated Mn^{2+} ion

To visualize how the metal substrate is coordinated in Nramps, we determined crystal structures of DraNramp using lipid-mesophase based techniques (Supplementary file 1a). We obtained a structure of wildtype (WT) DraNramp in an occluded state bound to Mn^{2+} at 2.38 Å by soaking crystals with Mn^{2+} (WT• Mn^{2+} ; Table 1, Figure 1A–B). We resolved a comparable structure using co-crystallization with Mn^{2+} and the inward-locking mutation A47W (A47W• Mn^{2+} ; Supplementary file 1b; Cα RMSD=0.47 Å; all pairwise RMSD values listed in Supplementary file 1c; Bozzi et al., 2016b). The similarity of both structures, including a nearly identical Mn^{2+} -coordination sphere (Figure 1B, Figure 1—figure supplement 1A), suggests that the observations we make based on these two structures are robust. Both structures superimpose best with the published occluded metal-free G45R

Table 1. Data collection and refinement statistics for four new DraNramp structures.

Structure Conformation Bound metal ion substrate PDB ID	WT _{soak} Occluded none 8E5V	WT•Mn ²⁺ Occluded Mn ²⁺ 8E60	M230A•Mn ²⁺ Inward open Mn ²⁺ 8E6I	WT•Cd ²⁺ Inward open Cd ²⁺ 8E6M
Data Collection				
Beamline	GMCA 23IDB	GMCA 23IDB	GMCA 23IDB	NECAT 24IDC
Wavelength (Å)	1.033	1.033	1.033	0.984
Resolution range (Å)	41.23–2.36 (2.44–2.36)	41.28–2.38 (2.46–2.38)	45.32–2.52 (2.61–2.52)	45.54–2.48 (2.57–2.48)
Space group	P 2 21 21	P 2 21 21	P 2 21 21	P 2 21 21
Unit cell (a, b, c)	58.95, 71.04, 98.77	59.08, 71.10, 98.75	58.67, 71.35, 98.59	59.14, 71.37, 99.05
Unit cell (α, β, γ)	90, 90, 90	90, 90, 90	90, 90, 90	90, 90, 90
Number of crystals	1	1	3	1
Total reflections	58744 (5928)	57472 (5733)	146077 (14913)	76829 (7275)
Unique reflections	17477 (1718)	16468 (1646)	14548 (1427)	15351 (1507)
Redundancy	3.4 (3.4)	3.5 (3.5)	10.0 (10.4)	5.0 (4.8)
Completeness (%)	98.71 (98.85)	95.11 (96.92)	99.90 (99.79)	99.03 (99.47)
Mean I/σ (I)	8.89 (0.97)	8.92 (0.89)	8.47 (0.75)	9.90 (1.12)
Wilson B-factor	49.97	50.55	54.29	49.52
R _{merge}	0.106 (1.292)	0.109 (1.241)	0.269 (2.475)	0.158 (1.626)
R _{meas}	0.127 (1.511)	0.127 (1.447)	0.284 (2.603)	0.178 (1.831)
R _{pin}	0.067 (0.759)	0.063 (0.722)	0.090 (0.800)	0.078 (0.816)
CC1/2	0.99 (0.37)	0.99 (0.39)	0.98 (0.34)	0.99 (0.34)
Refinement				
Resolution range (Å)	41.23–2.36 (2.44–2.36)	41.28–2.38 (2.46–2.38)	45.32–2.52 (2.61–2.52)	45.54–2.48 (2.57–2.48)

Table 1 continued on next page

Table 1 continued

Structure Conformation Bound metal ion substrate PDB ID	WT _{peak} Occluded none 8E5V	WT•Mn ²⁺ Occluded Mn ²⁺ 8E60	M230A•Mn ²⁺ Inward open Mn ²⁺ 8E6I	WT•Cd ²⁺ Inward open Cd ²⁺ 8E6M
No. reflections	17441 (1714)	16438 (1636)	14547 (1425)	15291 (1507)
No. reflections in <i>R</i> _{free}	1743 (171)	1642 (164)	1454 (143)	1530 (151)
<i>R</i> _{work}	0.217 (0.340)	0.207 (0.316)	0.225 (0.313)	0.202 (0.319)
<i>R</i> _{free}	0.245 (0.350)	0.259 (0.358)	0.266 (0.349)	0.250 (0.354)
Number of atoms	3449	3385	3451	3321
Protein	2945	2933	2934	2905
Ligand	443	405	448	362
Water	61	47	69	54
Protein Residues	392	393	392	388
Ramachandran plot				
Favored (%)	98.46	98.47	98.21	98.96
Allowed (%)	1.54	1.53	1.79	1.04
Outliers (%)	0	0	0	0
Rotamer outliers (%)	0.33	1.00	1.01	1.01
Clashscore	8.25	8.97	7.15	5.57
RMS (bonds)	0.002	0.002	0.002	0.002
RMS (angles)	0.43	0.46	0.43	0.46
Average <i>B</i> -factor	65.12	64.98	66.61	64.82
Protein	63.36	63.23	65.04	62.68
Ligand	77.99	78.48	77.70	83.11
Water	56.34	58.24	61.33	57.45
No. of TLS groups	9	8	3	3

Values in parentheses are for highest-resolution shell. Data for M230A•Mn²⁺ merge reflections from three crystals. Data for the other structures were obtained from a single crystal. See **Supplementary file 1a** for details on soaking or co-crystallization procedures for bound metal ion substrates.

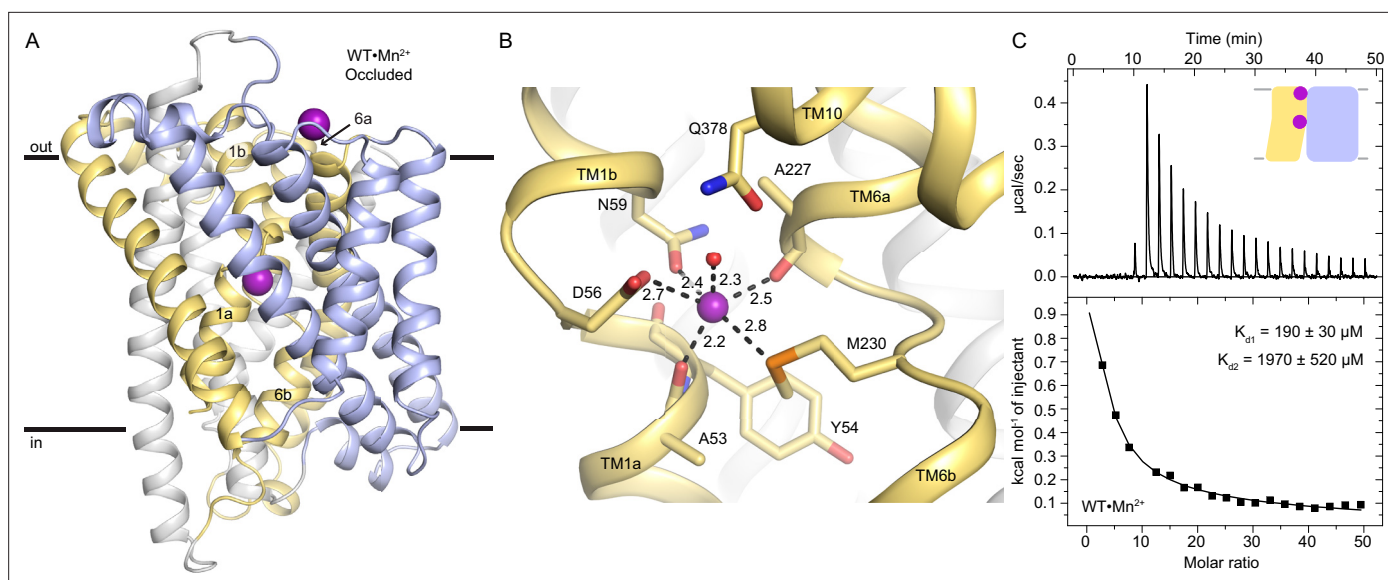


Figure 1. The occluded structure of DraNramp reveals a largely dehydrated Mn²⁺-coordination sphere. **(A)** Cartoon representation of WT•Mn²⁺ in an occluded state. Anomalous signal confirmed the presence of Mn²⁺ in both the orthosteric metal-binding site and an additional site at the mouth of the external vestibule (Figure 1—figure supplement 1A) which is less conserved across the Nramp family (Figure 1—figure supplement 3). TM1 and TM6 are labeled. **(B)** Detail of the orthosteric metal-binding site of WT•Mn²⁺ where D56, N59, M230, and the pseudo-symmetrically related carbonyls of A53 and A227 coordinate the Mn²⁺ ion (Figure 1—figure supplement 1B). A water molecule completes the six-ligand coordination sphere. Coordinating residues are shown as sticks, and coordinating distances are indicated in Å. **(C)** ITC measurement of the affinity of WT DraNramp for Mn²⁺. Top graph shows heat absorbed upon injection of Mn²⁺ solution to the protein solution. Bottom graph shows the fit of the integrated and corrected heat to a binding isotherm. The data show an endothermic mode of binding and fits best with a two-site sequential binding model. The figure shows one of three measurements and the average K_d values ± SEM (K_{d1}=190±30 μM, K_{d2}=1970±520 μM; see Appendix 1). Based on ITC experiments comparing Mn²⁺ binding to WT or DraNramp constructs with mutations at the external site (Figure 1—figure supplement 2A), we assigned K_{d1} to the orthosteric site. In all figures, unless otherwise noted, TMs 1, 5, 6, and 10 are pale yellow, TMs 2, 7, and 11 gray, TMs 3, 4, 8, and 9 light blue, and Mn²⁺ atoms are magenta spheres.

The online version of this article includes the following source data and figure supplement(s) for figure 1:

Source data 1. Multiple sequence alignment of 6172 Nramp homologs.

Source data 2. Maximum likelihood phylogenetic tree of Nramp homologs built with RAxML-NG.

Source data 3. Raw data of metal ion uptake into proteoliposomes measured at four ΔΨ values for each DraNramp construct.

Figure supplement 1. Structure of Mn²⁺ binding at the orthosteric and external sites of DraNramp.

Figure supplement 2. Affinity of Mn²⁺ for the orthosteric and external sites of DraNramp and transport activity of external-site variants.

Figure supplement 3. The external metal-binding site in DraNramp is somewhat conserved in clade A homologs, but poorly conserved across all Nramps.

Figure supplement 4. Representative time traces of metal ion uptake into proteoliposomes (n=2–3) measured at four ΔΨ values for each DraNramp construct.

structure (Bozzi et al., 2019b). Although the inner vestibule is occluded in both structures, WT•Mn²⁺ and A47W•Mn²⁺ differ in their TM1a position, with WT•Mn²⁺ nearly identical to G45R whereas the A47W•Mn²⁺ TM1a is displaced within the inner vestibule, likely to accommodate the bulky tryptophan sidechain. Therefore, we generally used the WT•Mn²⁺ structure for analysis of the occluded state. As in the metal-free G45R, the Mn²⁺-bound occluded structures have a completely sealed outer vestibule and a partially closed inner vestibule, with the Mn²⁺ occluded from bulk solvent (Figure 1A).

Anomalous difference Fourier maps confirmed presence of Mn²⁺ at the canonical, orthosteric Nramp metal-binding site between the unwound regions of TM1 and TM6 (Supplementary file 1d, Figure 1—figure supplement 1A). In WT•Mn²⁺, the Mn²⁺ is coordinated by conserved residues D56, N59, and M230, and backbone carbonyls of A53 and A227 (Figure 1A, Supplementary file 1e). A53 and A227 are pseudosymmetrically related in the inverted repeats of the LeuT fold of DraNramp (Figure 1—figure supplement 1B). The coordination sphere is completed by a water bridging Mn²⁺ with Q378, a residue previously proposed to directly coordinate Mn²⁺ in the occluded state (Bozzi

et al., 2019b). This yields a coordination number of 6, typical for Mn^{2+} , and a largely dehydrated metal-binding site with a distorted octahedral Mn^{2+} -coordination geometry (**Supplementary file 1f**), as often observed in other Mn^{2+} -protein complexes (*Barber-Zucker et al.*, 2017; *Couñago et al.*, 2014; *Dudev and Lim*, 2014).

At the mouth of the outer vestibule, an additional Mn^{2+} bridges D296 and D369 at the N termini of extracellular helix 2 (EH2) and TM10, respectively (**Figure 1—figure supplement 1A**). We denote this site as the ‘external site’ and the canonical substrate-binding site as the ‘orthosteric site’. Corroborating the structures, isothermal titration calorimetry (ITC) measurements reveal an endothermic mode of binding and are best fitted with a two-site model for WT ($K_{d1}=190\pm30\text{ }\mu\text{M}$, $K_{d2}=1970\pm520\text{ }\mu\text{M}$; **Figure 1C**) and A47W ($K_{d1}=125\pm5\text{ }\mu\text{M}$, $K_{d2}=2450\pm650\text{ }\mu\text{M}$; **Figure 1—figure supplement 2A**; all K_d values are in **Supplementary file 1g**; see Appendix 1 for a description of our ITC data analyses). To determine the affinity of the orthosteric site, we mutated the external-site aspartates. The D296A and D369A substitutions have little impact on Mn^{2+} transport (**Figure 1—figure supplement 2B**). The D296A and D369A variants each bind one Mn^{2+} with $K_d=370\pm30\text{ }\mu\text{M}$ and $420\pm30\text{ }\mu\text{M}$ respectively (**Figure 1—figure supplement 2A**), which is closest to K_{d1} of WT. Hence, the affinity for Mn^{2+} at the orthosteric site is higher than at the external site. A metal ion is present at the external site in all inward-open and occluded metal-bound DraNrap structures, but not outward-open structures, as opening the outer vestibule separates D296 and D369 and disrupts the site (**Figure 1—figure supplement 3A**). D296 and D369 are not conserved across Nramps, but they are more conserved within bacterial clade A, and there is a general abundance of acidic residues in the corresponding loop regions across all clades (**Figure 1—figure supplement 3B–C**). At present, our results provide little evidence of a biological role for this previously unidentified external site; perhaps the concentration of acidic residues at the mouth of the outer vestibule (**Figure 1—figure supplement 3D**) provides electrostatic attraction for metal cations.

Snapshots of the complete Mn^{2+} transport cycle by DraNrap

We also determined high-resolution DraNrap structures in metal-free occluded (WT) and Mn^{2+} -bound inward-open ($\text{M230A}\cdot\text{Mn}^{2+}$) states and re-refined a Mn^{2+} -bound outward-open conformation ($\text{G223W}\cdot\text{Mn}^{2+}$; **Table 1**, **Supplementary file 1a and b**). Along with the published structures of outward-open metal-free G223W and inward-open metal-free ‘Patch’ (which has a patch of mutations in intracellular loops) (*Bozzi et al.*, 2016b; *Bozzi et al.*, 2019b), these new structures allow us to map the entire Mn^{2+} transport cycle to three major conformations, each in Mn^{2+} -bound and metal-free states (**Figure 2A–B**). By ordering and comparing these six structures, we outline a molecular mechanism by which metal substrate binds, induces conformational change, and is released.

TMs 1, 5, 6, and 10 move the most as the Mn^{2+} -binding site accessibility switches from outward to inward across the conformations (**Figure 2B–C**, **Figure 2—figure supplement 1**), as also highlighted in previous studies (*Bozzi and Gaudet*, 2021; *Bozzi et al.*, 2019b; *Ehrnstorfer et al.*, 2017). As Mn^{2+} binds to the outward-open state, TM10 tilts toward TM1b, the upper half of TM5 toward TM7, and TM6a toward TM11 to seal the outer vestibule and yield an occluded state. The lower half of TM5 also moves away from TM1a, allowing it to swing upward to open the inner vestibule in the following transition. This swing of TM1a is the only noteworthy difference between the Mn^{2+} -bound occluded and inward-open states, allowing release of the Mn^{2+} into the inner vestibule. TM1a swings to a similar angle in the new inward-open $\text{M230A}\cdot\text{Mn}^{2+}$ as in the previous low-resolution inward-open metal-free structure (*Bozzi et al.*, 2016b), and structures of the homologous *Eggerthella lenta* Nrap-related magnesium transporter (EleNRMT), LeuT, and serotonin transporter (*Coleman et al.*, 2019; *Krishnamurthy and Gouaux*, 2012; *Ramanadane et al.*, 2022). Thus, most of the structural reorganization in Nramps occurs in the shift from outward open to occluded. The three metal-free DraNrap conformations are similar to their corresponding Mn^{2+} -bound structures, suggesting that once Mn^{2+} is released, the conformational transitions are reversed, including passing through an occluded metal-free intermediate, to reach the outward-open metal-free conformation ready to accept Mn^{2+} (**Figure 2A–B**, **Figure 2—figure supplements 1 and 2A**).

The substrate-binding TM1 and TM6 are well-resolved in our structures (**Figure 2—figure supplement 2B**) and pairwise superpositions reveal how their motions contribute to the conformational changes across the transport cycle (**Figure 2C**). TM6a tilts 22° and the unwound region of TM6 becomes more helical as it moves toward the Mn^{2+} to close the outer gate. The central unwound

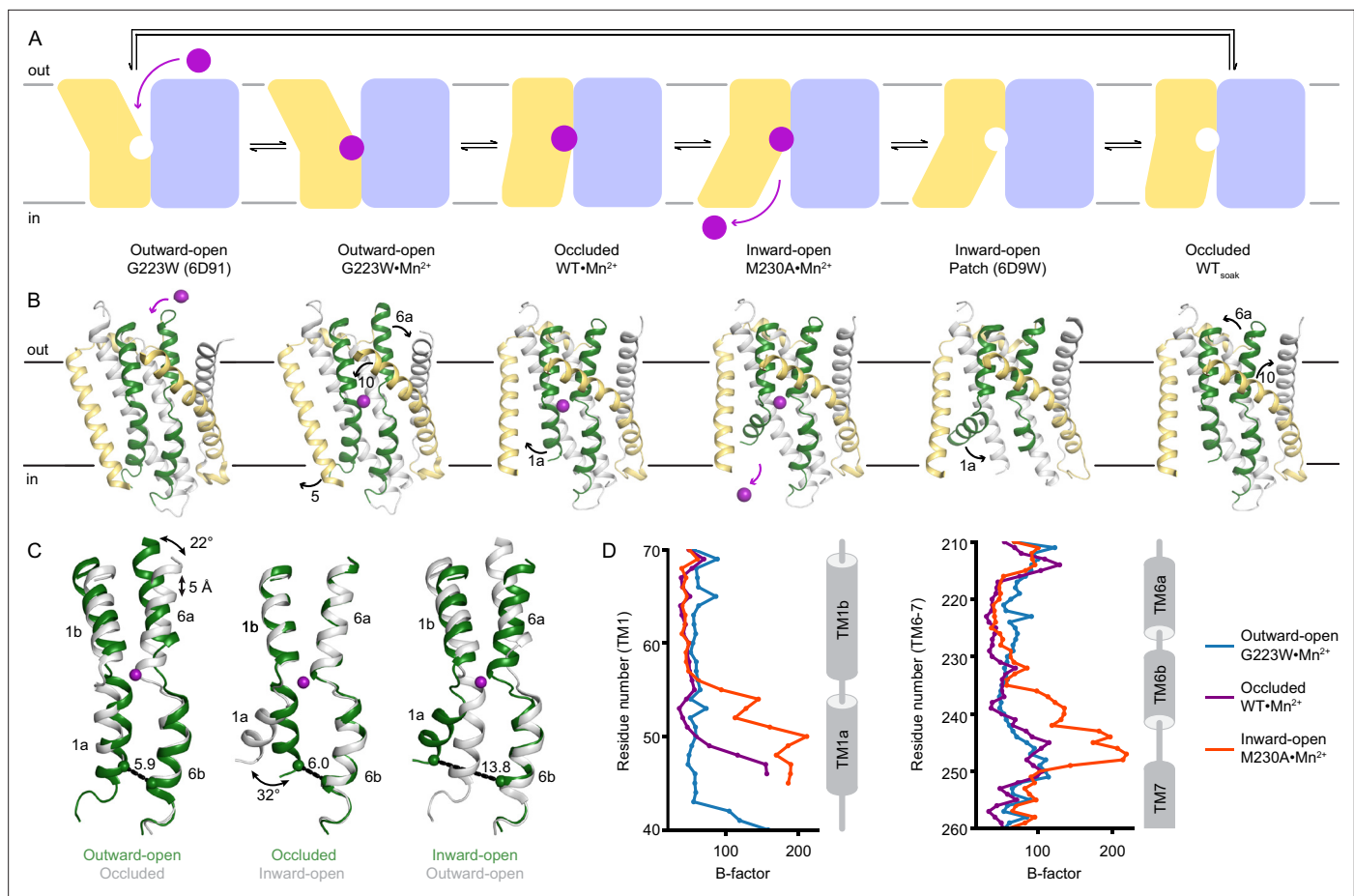


Figure 2. Structures in new conformations complete the Mn^{2+} transport cycle by DraNrap. **(A)** Schematic of the conformational states that DraNrap traverses to import Mn^{2+} . The mobile and stationary parts are pale yellow and light blue, respectively. **(B)** Corresponding structures of DraNrap, showing TMs 1 and 6 in green, TMs 5 and 10 in pale yellow, and TMs 2, 7, and 11 gray. Stationary TMs 3, 4, 8, and 9 are omitted to highlight the key motions in the mobile parts. Mn^{2+} ions are magenta. Black arrows indicate the key motions in TMs 1a and 6a detailed in panel **(C)**, and TMs 5 and 10 detailed in **Figure 2—figure supplement 1**. Full structures and the electron density for TMs 1 and 6 are illustrated in **Figure 2—figure supplement 2**. **(C)** Pairwise superpositions of whole Mn^{2+} -bound structures highlight the motions of TMs 1 and 6. Conformations are indicated at the bottom. The distance between residues 46 and 240 in TMs 1a and 6b, indicated for the green structures, increases from 5.9 Å to 13.8 Å from outward open to inward open. The large angular motions of TM1a and TM6b are also indicated. **(D)** Plots of B-factor by residue for the TM1 region (residues 40–70) and the TM6 region. The B-factors are highest for the inward-open state in which the interaction between TMs 1a and 6b (both in the inner leaflet) is broken.

The online version of this article includes the following figure supplement(s) for figure 2:

Figure supplement 1. Pairwise superpositions of Mn^{2+} -bound structures highlighting the motions of TMs 5 and 10.

Figure supplement 2. Structures of different conformational states of the DraNrap Mn^{2+} transport cycle.

regions of TM1 and TM6 are closest in the occluded state, resulting in an almost dehydrated Mn^{2+} -coordination sphere. Finally, the inner vestibule opens when TM1a tilts upward by 32°, increasing the distance between TM1a and TM6b by ~8 Å (**Figure 2C**). TM6b is largely static relative to the protein core, although in the inward-open structure it has high B-factors (**Figure 2D**), indicating that the interaction with TM1a stabilizes TM6b to close the inner vestibule.

Different conformations have distinct Mn^{2+} coordinations

Our Nrap structures provide snapshots of the complete Mn^{2+} -coordination sphere geometries in each conformation (**Figure 3**). Two new structures of DraNrap point-mutants reveal the coordination of Mn^{2+} in the inward-open state: M230A• Mn^{2+} and D296A• Mn^{2+} (C α RMSD of 0.42 Å). The Mn^{2+} is in the same location of the orthosteric site as in the occluded state, as confirmed by anomalous diffraction for D296A• Mn^{2+} (**Supplementary file 1d, Figure 3—figure supplement 1**). As in the occluded state, the Mn^{2+} binds D56, N59, and the A227 carbonyl, with a water replacing M230 in M230A• Mn^{2+}

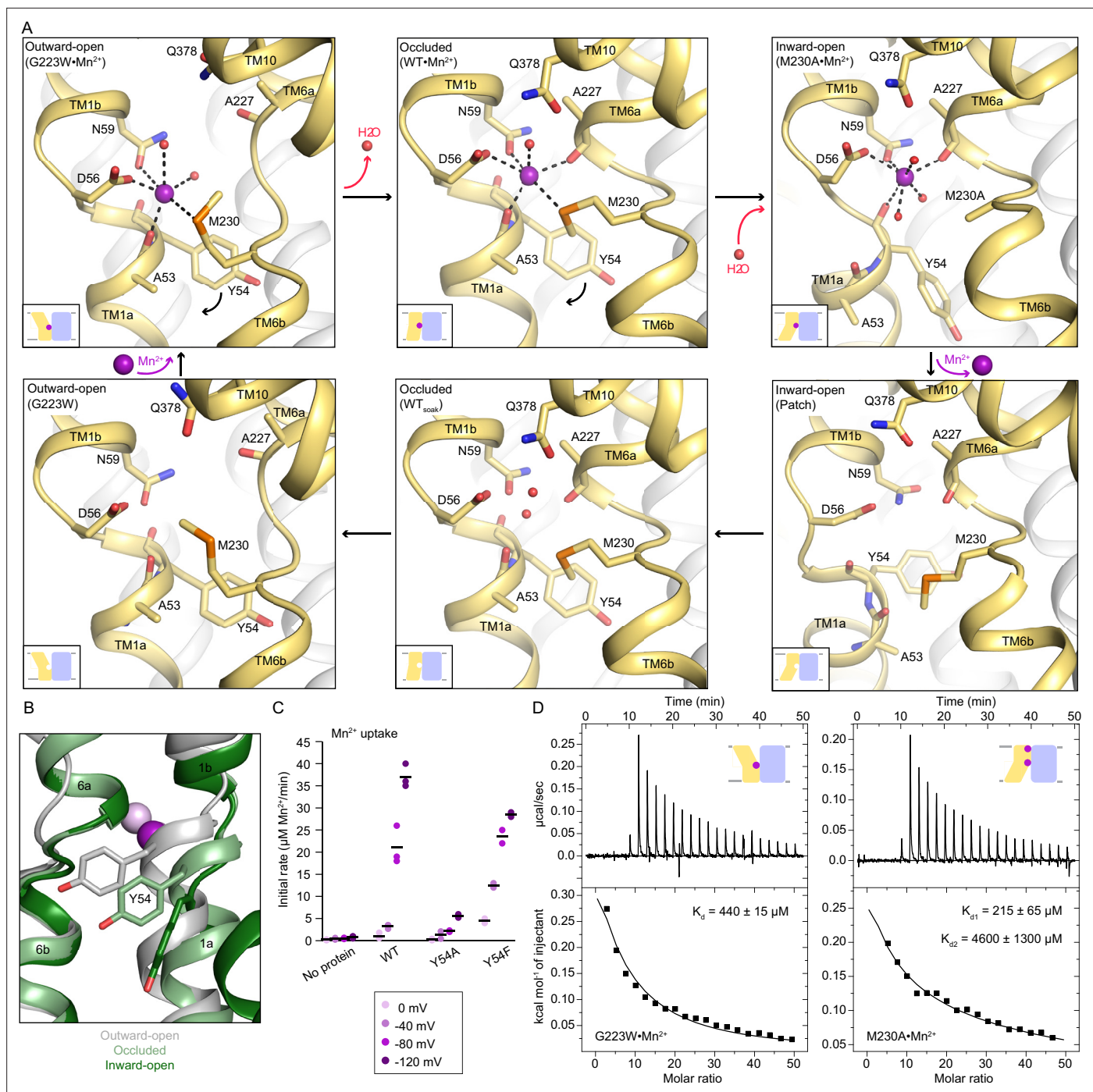


Figure 3. Coordination sphere changes across the Mn²⁺ transport cycle of DraNrpM. **(A)** Structures of the orthosteric metal-binding site in six conformations reveal the differences in coordination geometry and illustrate that the bound Mn²⁺ is more hydrated in the outward-open and inward-open states than the occluded state. In the occluded structure of metal-free WT DraNrpM a density we have assigned as water replaces Mn²⁺. Y54 in TM1a progressively moves to open the inner vestibule in the transition from outward to inward open, shown by black curved arrows. The view is rotated 180° along the vertical axis from **Figure 2C**. **(B)** TM1 and TM6 from a superposition of the three Mn²⁺-bound structures in panel A illustrate the swing of the Y54 sidechain as sticks. The view is rotated 180° along the vertical axis from **Figure 2C**. **(C)** Initial Mn²⁺ uptake rates for DraNrpM variants Y54A and Y54F at membrane potentials ranging from ΔΨ=0 to −120 mV (n=2–3; each data point is on the scatter plot and black bars are the mean values). The Mn²⁺ concentration was 750 μM, and the pH was 7 on both sides of the membrane. Y54A nearly abolishes transport whereas Y54F has near-wildtype initial transport rates. Corresponding time traces are plotted in **Figure 1—figure supplement 4**. **(D)** ITC measurements of G223W (left; one-site binding model with fixed n=1) and M230A (right; two-site sequential binding model) binding to Mn²⁺. One isotherm is shown of two measured, and the listed K_d values are the average ± SEM (see Appendix 1 for ITC analysis).

The online version of this article includes the following figure supplement(s) for figure 3:

Figure 3 continued on next page

Figure 3 continued

Figure supplement 1. Structure and Mn^{2+} -binding site architecture of D296A• Mn^{2+} .

Figure supplement 2. $2F_o - F_c$ maps (gray mesh; 1σ) of the Mn^{2+} -coordination sphere at the orthosteric site across different conformations of the Mn^{2+} transport cycle of DraNramp.

Figure supplement 3. Sequence logos highlighting that Y54 in TM1a is 80% conserved in all Nramps (3762 sequences), 100% conserved in bacterial clades A and C, but replaced by a phenylalanine in clade B.

(**Figure 3A**, **Supplementary file 1e**). However, with TM1a displaced, the A53 carbonyl no longer coordinates Mn^{2+} ; instead, the Y54 carbonyl approaches Mn^{2+} at a longer distance of 3.1 Å. Two more waters, one bound to Q378 and another from the inner vestibule, complete a seven-coordination sphere resembling a pentagonal bipyramidal geometry with substantial distortion (**Supplementary file 1f**). Seven coordination is infrequent but found in Mn^{2+} -coordinating proteins like MntR (**Chen and He, 2008; Glasfeld et al., 2003**). Our inward-open structures provide the first evidence that Y54 participates in the Mn^{2+} transport cycle.

The new occluded and inward-open Mn^{2+} -bound structures have a monodentate coordination of D56 with Mn^{2+} . For consistency, we reinterpreted the outward-open G223W• Mn^{2+} map (PDB ID: 6BU5) with a monodentate coordination of D56 with Mn^{2+} instead of previously modeled bidentate interaction (**Bozzi et al., 2019b**); the local and global model statistics are very similar to the original structure (**Supplementary file 1e**). Re-refined G223W• Mn^{2+} has six Mn^{2+} -coordinating ligands: D56, N59, M230, carbonyl of A53 and two waters (**Figure 3A**, **Supplementary file 1e**); the overall geometry resembles a distorted octahedron as in the occluded structure (**Supplementary file 1f**). The coordination spheres in all conformations of the transport cycle are well defined as confirmed by the $2F_o - F_c$ maps of the closeup snapshots of their Mn^{2+} -bound orthosteric site (**Figure 3—figure supplement 1C**, **Figure 5—figure supplement 2**).

Comparing the three Mn^{2+} -bound conformations (**Figure 3A**), the carbonyls of A53 in TM1a and A227 in TM6b alternately coordinate Mn^{2+} in the outward- and inward-open structures respectively, and both residues interact with Mn^{2+} in the occluded structure. The pseudosymmetrically related A53 and A227 may thus act as hinges altering the Mn^{2+} -coordination sphere as TM1 and TM6 move in turn to open the gates during Mn^{2+} transport. Furthermore, as DraNramp switches from outward- to inward-open, Y54 progressively swings downward, acting as a gate in concert with TM1a's upward swing to open the inner vestibule and allow metal release (**Figure 3A–B**). Our inward-open structures also suggest that the Y54 carbonyl may participate in Mn^{2+} release through interaction with the metal ion (**Figure 3A**). All 3796 Nramp sequences in our alignment have either a tyrosine or a phenylalanine at this position and Y54 is completely conserved among bacterial clades A (including DraNramp) and C, while the position is 40% and 100% phenylalanine among eukaryotes and bacterial clade B, respectively (**Figure 3—figure supplement 3**). To evaluate the significance of Y54 in Mn^{2+} transport, we purified and reconstituted into proteoliposomes the Y54A and Y54F variants. While Y54F has near-wildtype Mn^{2+} -transport activity, Y54A nearly eliminates Mn^{2+} transport (**Figure 3C**), indicating that an aromatic ring is essential for the gating motion required for transport.

We also measured the Mn^{2+} -binding affinity of the constructs that yielded inward- or outward- open structures, M230A and G223W, respectively. Mn^{2+} is present at the external site in the inward-open M230A• Mn^{2+} (**Figure 1—figure supplement 3A**), and consistently, the ITC data fit a two-site model ($K_{d1}=215\pm65\ \mu\text{M}$ assigned to the orthosteric site, $K_{d2}=4600\pm1300\ \mu\text{M}$ for the external site; **Figure 3D** and Appendix 1). The ITC data for G223W with Mn^{2+} fits only in a one-site model ($K_d=440\pm15\ \mu\text{M}$; **Figure 3D**), which we assign to the orthosteric site because the opening of the outer vestibule displaces TM10, disrupting the external site (**Figure 1—figure supplement 3A**).

Mn^{2+} binding does not significantly alter the three main DraNramp conformations

To compare the metal-bound states to analogous metal-free states of the transport cycle, we determined two metal-free occluded structures of wildtype DraNramp at a higher resolution than the previously reported G45R structure (**Bozzi et al., 2019b**), which we refer to as WT (2.38 Å; **Supplementary file 1b**) and WT_{soak} (2.36 Å; **Table 1**). The crystal used for WT_{soak} was mock-soaked (with no metal in the soaking solution). WT and WT_{soak} are nearly identical, confirming that the soaking process does

not influence the conformational state. We analyzed WT_{soak}, unless otherwise noted. In WT_{soak}, we observed density but no anomalous signal at the orthosteric site and modeled a water molecule at the position where Mn²⁺ sits in the occluded state (**Figure 3A**). WT_{soak} is nearly identical to WT•Mn²⁺, indicating that the occluded conformation is unchanged by the presence of metal ion substrate, and the metal-binding site is instead filled by ordered water molecules.

We used previously reported inward-open 'Patch' and outward-open G223W metal-free structures for analysis of the Mn²⁺ transport cycle (**Figures 2B and 3A**; **Bozzi et al., 2016b**; **Bozzi et al., 2019b**). These structures, resolved at lower resolution than the ones described here, have no density at the orthosteric site. This is consistent with a more flexible organization of a metal-binding site open to bulk aqueous solvent. In contrast, metal-free WT has a more ordered orthosteric site, suggesting a stable occluded intermediate in the switch from inward- to outward-open.

Polar networks latch the gates to achieve alternating access

Vestibules providing access to the orthosteric site from the extracellular or intracellular side alternately open from the motions of DraNramp's TMs 1, 5, 6, and 10, which form the outer and inner gates during Mn²⁺ transport (**Bozzi et al., 2020**; **Bozzi et al., 2019b**). To pinpoint protein features that enable these motions, we used our Mn²⁺-bound structures to identify interaction networks with the following attributes: (i) they contain conserved polar residues from at least one of the four mobile helices; (ii) they line the gates; and (iii) they rearrange between the three resolved protein conformations (**Figure 4A**).

Two networks seal the outer gate. In the occluded and inward-open conformations, Q378 interacts with two waters, one coordinating the orthosteric Mn²⁺ and the other interacting with D56 and the carbonyl of T130. This network is disrupted in the outward-open conformation as Q378 and the rest of TM10 swing outward to open the outer vestibule (**Figure 4B**). In the second network, T228, N275, and N82 interact via a water in the occluded and inward-open states, but not in the outward-open state, where the extended unwound region of TM6a positions T228 farther from the orthosteric site and N275 and N82 (**Figure 4C**). This T228 network helps rearrange TM6a, closing the outer vestibule in the occluded state and generating a nearly dehydrated Mn²⁺-coordination sphere (**Figure 1B**). As the inner gate opens to release Mn²⁺, both networks persist, ensuring that the outer gate remains closed in the inward-open conformation.

All six residues in the Q378 and T228 networks are completely conserved across bacterial clade A and highly conserved across all Nramps (**Figure 4—figure supplement 1**). To investigate the robustness of these networks, we performed duplicates of molecular dynamics (MD) simulations starting in each of the three conformations and confirmed that within the first 250 ns of these simulations, the T228 and Q378 networks persist in simulations of the occluded and inward-open states and remain broken in simulations starting in the outward-open state (**Figure 4—figure supplement 2A–C**). This includes the coordinated waters, for example the water at the center of the Q378 network is present in more than 50% of the frames in occluded-state simulations (**Figure 4—figure supplement 2E**). In line with the X-ray snapshots and simulation outcomes and highlighting their key function in the conformational cycle of Nramps, mutations of any of the six residues across these networks reduces Mn²⁺ transport by DraNramp (**Bozzi et al., 2016a**; **Bozzi et al., 2019a**; **Bozzi et al., 2020**; **Bozzi et al., 2019b**).

The inner vestibule is gated by rearrangements of residues in two other polar networks, namely those of R244 and Q89 (**Figure 4D–E**). In the outward-open state, R244 forms an ion pair with E176 and interacts with the TM1a backbone, as does D263, keeping TM1a and TM5 close and the inner gate closed (**Figure 4D**; **Bozzi et al., 2020**). In the occluded state, the E176–R244 interaction breaks and TM5 moves away from TM6b, creating space for TM1a to swing up and open the inner vestibule in the inward-open state. Accordingly, the E176–R244 ion pair is stable in MD simulations of the outward-open state, while these residues are >10 Å apart in simulations of the occluded and inward-open states (**Figure 4—figure supplement 2D**). Supporting the importance of the R244 network, E176 is 100% and R244 is 85% conserved across all Nramps (**Figure 4—figure supplement 1**) and mutation of either residue reduces Mn²⁺ transport by DraNramp (**Bozzi et al., 2020**).

In the Q89 network, Q89 hydrogen-bonds with Y54 and H237 to seal the inner gate in the outward-open (**Bozzi et al., 2020**) and occluded states (**Figure 4E**). In the inward-open structure, the Q89–H237 hydrogen bond is broken and a rearranged Y54–Q89 hydrogen bond buttresses the opening of

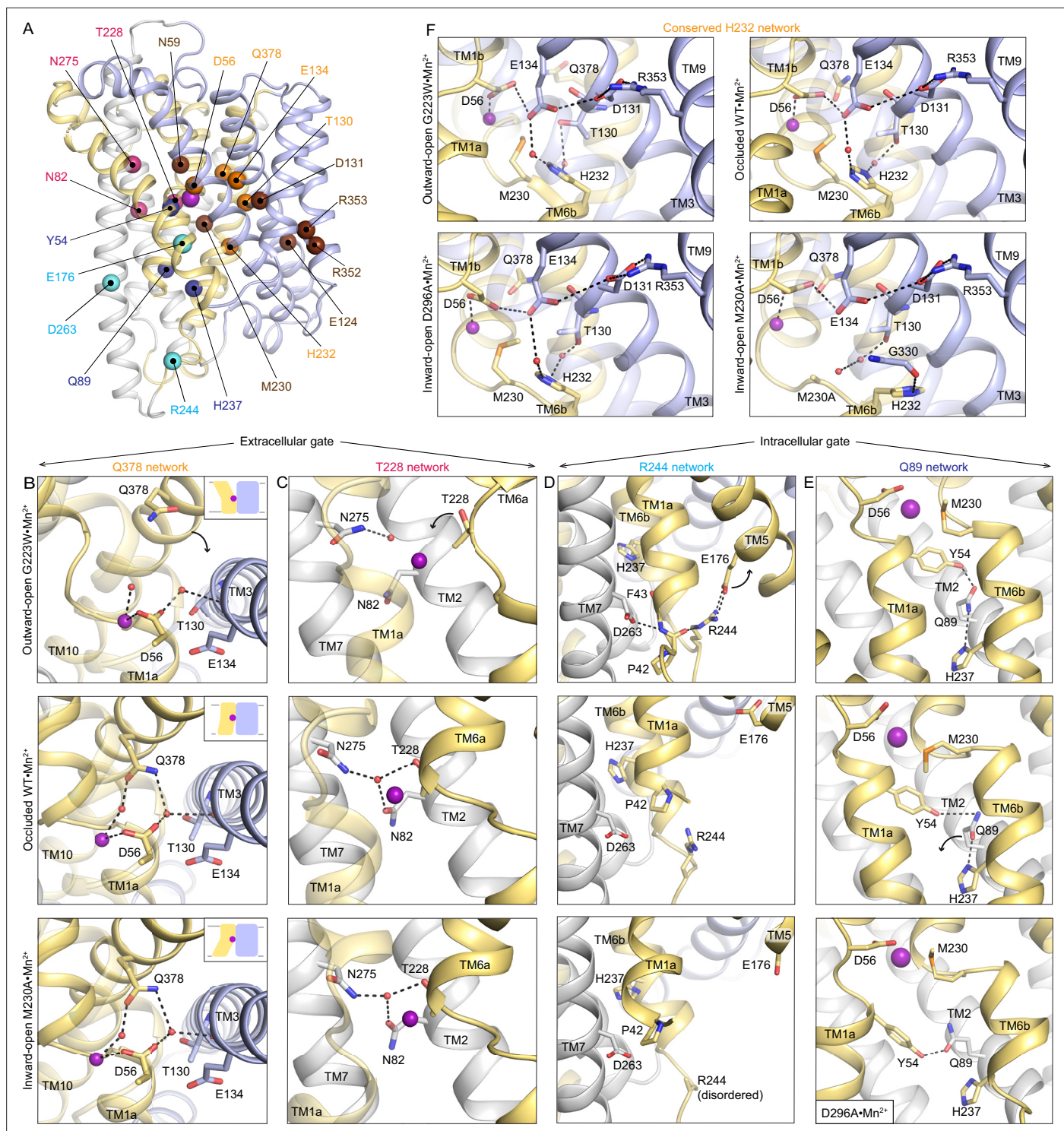


Figure 4. Networks of polar residues lining the outer and inner vestibules rearrange through the conformational transitions needed for Mn²⁺ transport. **(A)** The C α positions of residues in the Q378 (orange) and T228 (pink) networks lining the outer gate, R244 (cyan) and Q89 (blue) networks within the inner gate, and the H232 (orange) network coordinating with the proton pathway, are mapped on the occluded WT•Mn²⁺ structure. Metal-binding and proton pathway residues (Bozzi et al., 2019b) are represented as brown spheres. **(B)** The Q378 network forms as TM10 moves when DraNrap transitions from outward-open to occluded to close the outer vestibule. Water-mediated interactions form between Q378, D56, and T130. The other D56 carbonyl interacts directly with Mn²⁺. TM1 is transparent. **(C)** The T228 network forms with N275, N82, and T228 coordinating a water as TM6a moves to close the outer vestibule. TM1 is transparent. **(D)** In the R244 network, interactions between R244, E176, and D263 break as TM5 moves in the transition from outward-open to occluded state to initiate the opening of the inner vestibule. **(E)** In the Q89 network, Y54, Q89, and H237 rearrange from

Figure 4 continued on next page

Figure 4 continued

occluded to inward-open state as TM1a swings up to allow for metal release. (F) H232, which abuts the orthosteric Mn^{2+} -binding site, interacts with E134 and T130 through waters conserved in all conformations. In the $\text{M230A}\cdot\text{Mn}^{2+}$ structure, H232 flips and is replaced by a water, retaining the interaction with T130 but breaking the connection with E134, suggesting that M230 helps stably position H232. TM8 is transparent. In panels c-e, TMs 3, 4, 8, and 9 are omitted to better visualize the interactions. The illustrated structures are indicated on the figure.

The online version of this article includes the following figure supplement(s) for figure 4:

Figure supplement 1. Sequence logos showing conservation of polar residues important for opening and closing the gates during Mn^{2+} transport (highlighted in black boxes).

Figure supplement 2. Structural stability of gating residue networks of the Mn^{2+} transport cycle.

Figure supplement 3. The H232 rotamer is fixed in the outward-open state but dynamic in the occluded and inward-open states.

the inner vestibule (**Figure 4E**). As discussed above, Y54 is conserved and important for Mn^{2+} transport by *DraN*ramp (**Figure 3C** and **Figure 3—figure supplement 1C**). Similarly, Q89 and H237 are conserved (**Figure 4—figure supplement 1**), and mutations to either residue impair Mn^{2+} transport in a cell-based assay (Bozzi et al., 2020). Cysteine accessibility measurements showed that mutations to Q89 or H237 render the outer vestibule solvent-inaccessible (Bozzi et al., 2020), indicating that disrupting the Q89 network likely prevents closing of the inner gate.

H232 (TM6b) is conserved across all *N*ramps (**Figure 4—figure supplement 1**), highlighting its importance. H232 sits below the orthosteric site and forms a network conserved across all conformations, with water-mediated hydrogen bonds to E134 (involved in proton transfer to the salt-bridge residues in TMs 3 and 9) (Bozzi et al., 2019a; Bozzi et al., 2019b; Ehrnstorfer et al., 2017) and T130 in TM3 (**Figure 4F**). Waters occupy these two sites in MD simulations in all states, especially the water coordinated between H232 and T130 (**Figure 4—figure supplement 2F–G**). However, in $\text{M230A}\cdot\text{Mn}^{2+}$ H232 flips to interact with the G330 carbonyl (TM8; **Figure 4F**), suggesting that it may transiently move during the conformational cycle. Indeed, while the H232 sidechain rotamer is stable in MD simulations of the outward-open state, it explores other rotamers in simulations of the occluded and inward-open states (**Figure 4—figure supplement 3**).

Unlike Mn^{2+} binding, Cd^{2+} binding to *DraN*ramp is exothermic

*N*ramps transport divalent transition metals quite promiscuously, including both physiological (Fe^{2+} and Mn^{2+}) and non-physiological substrates (Cd^{2+} , Zn^{2+} , Co^{2+} , Ni^{2+} , Pb^{2+}), but select against alkaline earth metals (Mg^{2+} , Ca^{2+}) (I Bannon et al., 2002; Bozzi et al., 2016a; Bozzi and Gaudet, 2021; Bozzi et al., 2019b; Ehrnstorfer et al., 2017; Illing et al., 2012; Sacher et al., 2001). *DraN*ramp transports Cd^{2+} well, but without concomitant proton flux and with weaker voltage dependence (**Figure 5—figure supplement 1A**; Bozzi et al., 2019a; Bozzi and Gaudet, 2021; Bozzi et al., 2019b). To better understand the underlying mechanistic differences, we compared the binding, transport, and structures of *DraN*ramp with Mn^{2+} and Cd^{2+} .

In contrast to endothermic binding of Mn^{2+} , ITC measurements show exothermic binding of Cd^{2+} to WT *DraN*ramp (**Figure 5—figure supplement 1B**; see Appendix 1 for details of the ITC analyses). Like for Mn^{2+} , the Cd^{2+} isotherm fits best in a two-site model ($K_{d1}=55\pm15\text{ }\mu\text{M}$, $K_{d2}=220\pm20\text{ }\mu\text{M}$). Both D296A and D369A—containing mutations at the external metal-binding site—showed exothermic binding but fit best in a one-site model (**Figure 5A**). Based on their affinity ($K_d=120\pm1\text{ }\mu\text{M}$ for D296A and $K_d=70\pm10\text{ }\mu\text{M}$ for D369A), we conclude that the orthosteric site has higher affinity toward Cd^{2+} than the external site (**Figure 5A–B**). Both the orthosteric and external site shows higher affinity toward Cd^{2+} than Mn^{2+} . The affinities are comparable at the orthosteric site, in low micromolar range for both metals (K_d of $190\pm30\text{ }\mu\text{M}$ for Mn^{2+} vs. $55\pm15\text{ }\mu\text{M}$ for Cd^{2+}), whereas the external site has a much higher affinity toward Cd^{2+} (K_d of $1970\pm520\text{ }\mu\text{M}$ for Mn^{2+} vs. $220\pm20\text{ }\mu\text{M}$ for Cd^{2+}). Consistent with its inability to transport Mg^{2+} , a representative alkaline earth metal (Bozzi and Gaudet, 2021; Bozzi et al., 2019b), *DraN*ramp does not bind Mg^{2+} (**Figure 5A**). These are the first ITC measurements comparing the binding of different metals to an *N*ramp transporter and they show clear differences in the binding mode and affinity of different substrates toward *DraN*ramp (**Figure 5A**, **Figure 5—figure supplements 2 and 3**). In contrast to *DraN*ramp, previous ITC studies showed endothermic binding of Cd^{2+} to the *Staphylococcus capitis* *N*ramp homolog (*ScaDMT*) with $29\text{ }\mu\text{M}$ affinity (Ehrnstorfer et al., 2014). However, in the absence of ITC data with other metals, it is not

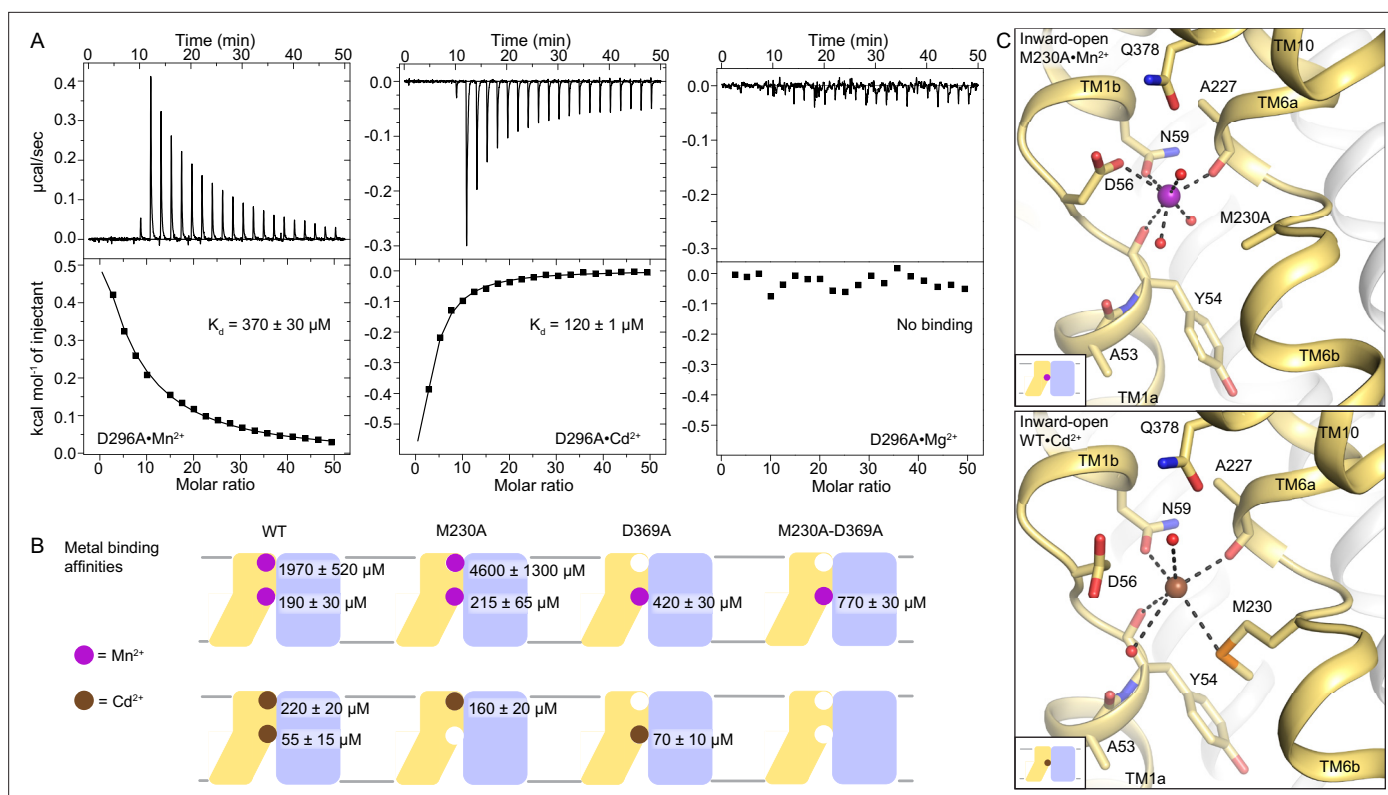


Figure 5. DraNrap binds differently to Mn²⁺ and Cd²⁺. **(A)** ITC measurements show that DraNrap binds Mn²⁺ in an endothermic mode, Cd²⁺ in an exothermic mode and does not bind Mg²⁺ (6 mM metal). ITC was performed with DraNrap in which the external site is mutated (D296A) to analyze binding at the orthosteric site. One isotherm is shown of 2 or 3 measured. These isotherms were fit using a one-site model with fixed n=1, and the listed K_d values are the average ± SEM (see Appendix 1 for ITC analysis). **(B)** Schematic showing the ITC-measured K_d values of various DraNrap constructs for Mn²⁺ (top) and Cd²⁺ (bottom). WT DraNrap binds Mn²⁺ and Cd²⁺ at the same two sites, with the external-site affinity ~10-fold higher and the orthosteric-site affinity ~threefold higher for Cd²⁺ than Mn²⁺. The M230A mutation eliminates binding of Cd²⁺ but not Mn²⁺ at the orthosteric site. The D369A mutation eliminates binding of either metal at the external site. A variant with mutations at both the orthosteric and external sites, M230A-D369A, does not bind Cd²⁺ but maintains orthosteric site binding for Mn²⁺. For all ITC data, the K_d values were computed while fixing the number of sites (*n*) to 1 or 2 and assigned to the external or orthosteric site based on knowledge of the crystal structures and mutational analysis (Appendix 1; **Supplementary file 1g**). ITC traces are shown in **Figure 5—figure supplement 1A**, **Figure 5—figure supplement 2** and **Figure 5—figure supplement 3**. **(C)** Comparison of the inward-open state bound to Mn²⁺ (M230A•Mn²⁺; top) and Cd²⁺ (WT•Cd²⁺; bottom) shows differences in coordination geometry at the orthosteric site. D56 is oriented differently and does not directly coordinate Cd²⁺. Cd²⁺ coordinates N59, M230, carbonyls of A227 and Y54, and two waters, for a total of six ligands compared to seven for Mn²⁺. Mn²⁺ and Cd²⁺ are magenta and brown spheres, respectively. Of note, we use M230A•Mn²⁺ here because soaking of WT crystals with Cd²⁺ yielded an inward-open WT•Cd²⁺ structure (**Figure 5—figure supplement 1C**) whereas soaking the same kind of crystals with Mn²⁺ yielded an occluded WT•Mn²⁺ structure (**Figure 1**).

The online version of this article includes the following source data and figure supplement(s) for figure 5:

Source data 1. Source files (Origin files) of ITC experiments of magnesium binding to each DraNrap construct.

Figure supplement 1. Analysis of Cd²⁺ and Mn²⁺ binding to DraNrap reveal differences.

Figure supplement 2. ITC measurements comparing the affinity for Mn²⁺ and Cd²⁺ of DraNrap constructs with mutations at metal-binding sites.

Figure supplement 3. ITC measurements comparing Mn²⁺ and Cd²⁺ binding affinity of the conformation-locking mutants.

Figure supplement 4. Comparison of the gating networks in the inward-open Mn²⁺ and Cd²⁺-bound structures.

known whether ScaDMT also shows differences in the mode and affinity of binding to different metals like DraNrap.

A Cd²⁺-bound structure helps explain functional differences

To understand the differences in binding and transport of Mn²⁺ and Cd²⁺, we determined a 2.5 Å Cd²⁺-bound structure by soaking WT DraNrap crystals with 2 mM Cd²⁺ (**Table 1**). In agreement with the ITC data, WT•Cd²⁺ shows Cd²⁺ ions at both the external and orthosteric sites as confirmed by anomalous signal (**Figure 5—figure supplement 1C–D**, **Supplementary file 1d**). The higher affinity

for Cd^{2+} than Mn^{2+} at the external site suggests that local geometry favors non-physiological Cd^{2+} over the physiological substrate Mn^{2+} (**Figure 1—figure supplement 3A**). The low affinity of this site for Mn^{2+} , compared to Cd^{2+} , is consistent with the idea that this external site plays a role in general electrostatic attraction of substrate to the orthosteric site, rather than as a finely tuned binding site specific for Mn^{2+} .

Interestingly, $\text{WT}\cdot\text{Cd}^{2+}$ is inward open (**Figure 5—figure supplement 1C**), although both mocked-soaked and Mn^{2+} -soaked crystals under otherwise equivalent conditions yielded occluded structures (WT_{soak} and $\text{WT}\cdot\text{Mn}^{2+}$, respectively; **Figures 1 and 3**). The larger ionic radius of Cd^{2+} (~ 0.95 Å) compared to Mn^{2+} (~ 0.82 Å) (**Kumarevel et al., 2005; Vashishtha et al., 2016**) and different preferred coordination geometry may increase the stability of the inward-open state with Cd^{2+} bound. Compared with the inward-open $\text{M230A}\cdot\text{Mn}^{2+}$, D56 adopts a different rotamer in $\text{WT}\cdot\text{Cd}^{2+}$ and does not coordinate the Cd^{2+} bound at the orthosteric site (**Figure 5C**). The rest of the coordination sphere is similar and includes N59, M230, the A227 carbonyl, the Y54 carbonyl, a water that coordinates Q378 and another water from inner vestibule. Cd^{2+} has six coordinating ligands, and the distortion from ideal octahedral geometry is more pronounced compared to Mn^{2+} (**Supplementary file 1f**). The coordination distances are larger for Cd^{2+} than for Mn^{2+} (**Supplementary file 1e**), as seen in other proteins (**Begg et al., 2015; Yokoyama et al., 2012**) and consistent with its larger ionic radius and distinct charge distribution. Cd^{2+} is a soft metal, likely explaining why it retains coordination by the softer sulfur ligand of M230 (**Cammack and Hughes, 2008**) but not the hard oxygen of D56, although D56 can still provide favorable electrostatics.

Cd^{2+} binding is prone to perturbations and favors the inward-open state

ITC data analysis of different DraNrap variants highlights additional differences between Mn^{2+} and Cd^{2+} binding and how their binding affects the conformational preferences of DraNrap (**Figure 5B, Figure 5—figure supplements 2–3**). A general observation is that, at the orthosteric site, Mn^{2+} and Cd^{2+} binding are entropy- and enthalpy-driven, respectively, and entropy contributions ($-\Delta S$) are smaller for Cd^{2+} than Mn^{2+} (Appendix 1), suggesting that Cd^{2+} binding conformationally constrains the protein more, leads to less solvent release, or both (**Ferrante and Gorski, 2012; Olsson et al., 2008**).

ITC with the orthosteric-site mutants, M230A and D56A, shows metal-specific behavior. For M230A, Mn^{2+} binding fits with a two-site model and Cd^{2+} fits only with a one-site model (**Figure 5B and Figure 5—figure supplement 2**, Appendix 1). M230 is thus a crucial ligand for Cd^{2+} but not Mn^{2+} , which is further reflected in the transport behavior where M230A affects Cd^{2+} transport drastically but has negligible effect on Mn^{2+} (**Bozzi et al., 2016a; Bozzi et al., 2019a**). Data for D56A fit better with a two-site model with both metals, although D56A does not transport either metal (**Bozzi et al., 2019b**). This suggests that D56 is more important for catalyzing transport than for substrate binding. Data for variants with an additional mutation at the external site (D56A-D296A, D56A-D369A, M230A-D296A, and M230A-D369A) fit only with a one-site model for Mn^{2+} -binding isotherms, which we assigned as binding to the orthosteric site. However, all four double mutants showed complete loss of Cd^{2+} binding, which is expected for ones with M230A—which eliminates Cd^{2+} binding at the orthosteric site on its own, as we have also shown previously (**Bozzi et al., 2016a**)—but more surprising for double mutants with D56A. Thus, binding of the preferred physiological Mn^{2+} substrate at the orthosteric site is more robust to perturbations than binding of Cd^{2+} , a toxic metal.

The conformation-locking mutants—A47W, which is outward-closed (**Bozzi et al., 2016b**) and crystallized in an occluded state, and outward-open G223W (**Bozzi et al., 2019b**)—also show different behavior with Mn^{2+} and Cd^{2+} (**Figure 5—figure supplement 3**). ITC with Mn^{2+} agrees with the structural data, with two-site fit for A47W (**Figure 1—figure supplement 2A**), and one-site fit for A47W-D296A, A47W-D369A, and G223W (assigned to the orthosteric site). However, with Cd^{2+} , the ITC data for A47W fit best with a one-site model and we observed no binding with A47W-D296A, A47W-D369A, and G223W, indicating that there is no significant affinity for Cd^{2+} at the orthosteric site of A47W and G223W. This is consistent with our inability to obtain Cd^{2+} -bound structures in conformations other than inward-open, despite trying both co-crystallization and soaking with A47W and G223W. These data suggest that Cd^{2+} has highest affinity for the inward-open state and low affinity for other states, in

agreement with the fact that soaking of WT crystals (which yielded the metal-free occluded WT_{soak} structure) produced an inward-open Cd²⁺-bound structure.

Gating network differences in the Cd²⁺-bound structure are restricted to D56

To better understand the Cd²⁺ transport mechanism given that its binding at the orthosteric site seems less optimal and robust than Mn²⁺, we compared the gating networks described above for the inward-open Mn²⁺- and Cd²⁺-bound structures. The Q89, T228, and R244 networks are essentially identical in the inward-open state, regardless of whether Mn²⁺ or Cd²⁺ is bound (**Figure 5—figure supplement 4A–C**).

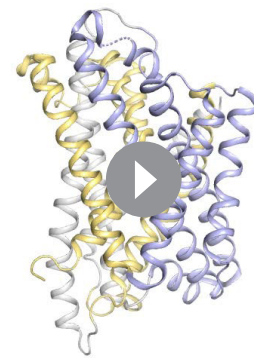
The other two networks, both of which involve D56, have some differences. In the Q378 network gating the outer vestibule, D56 does not coordinate Cd²⁺ but does interact with the conserved water that connects the metal ion with Q378 in all outward-closed structures (**Figure 5—figure supplement 4D**). This preserves a connection to both Cd²⁺ and Q378 to close the outer gate. Most of the H232 network is similar in the Mn²⁺- and Cd²⁺-bound structures, except for the orientation of D56 relative to E134 (**Figure 5—figure supplement 4E**).

Mutations of several residues in these polar networks reduced transport in proteoliposome-based assays (**Figure 5—figure supplement 4F**), corroborating results from cell-based assays (Bozzi et al., 2020). Y54A, H237A, and H232A mutations cause the largest decreases, Q89A causes a moderate decrease, and Y54F is similar to WT. Mn²⁺ and Cd²⁺ follow similar trends although with less voltage dependence for Cd²⁺ than Mn²⁺. These results support the idea that although Mn²⁺ and Cd²⁺ bind differently at the orthosteric site, the flexibility of the D56 sidechain enables the networks of polar residues that gate the outer and inner vestibules to engage and enable transport of both metals at similar rates.

Discussion

Our high-resolution structures in different conformations and analysis of substrate-binding affinities reveal a molecular map of the Mn²⁺ import pathway in DraNrpM (**Video 1**). Binding of the Mn²⁺ substrate at the orthosteric site takes on different coordination geometries through the three main conformational states in the transport cycle, all of which deviate substantially from ideal, consistent with the moderate binding affinities we measured. We identified several networks of polar interactions that gate both the outer and inner vestibules. Structures of DraNrpM bound to Mn²⁺ and Cd²⁺ and corresponding analyses of substrate binding demonstrate that the orthosteric site shows similar affinity for physiological (Mn²⁺) and toxic (Cd²⁺) substrates, but Cd²⁺ binding is less robust to various perturbations.

Our structures of one NrpM homolog, DraNrpM, in all conformations of the transport cycle provide an opportunity to update the overview of the conformational cycle, focusing on the polar interaction networks that gate the outer and inner vestibules (**Figure 6A**). The structures also provide the first molecular view of how local changes in the coordination spheres and the global conformational transitions coordinate to facilitate Mn²⁺ transport. Starting in the outward-open state, Mn²⁺ entry into the outer vestibule and binding at the orthosteric site triggers the transition to the occluded state



Video 1. Conformational rearrangement and distinct coordination geometry adopted by DraNrpM during Mn²⁺ import. The global conformational changes are highlighted first, with cartoon representations of each of the six structures in the Mn²⁺ transport cycle starting from the outward-open Mn²⁺-bound conformation rotating through the occluded Mn²⁺-bound, inward-open Mn²⁺-bound, and then the metal-free states transitioning back to outward-open Mn²⁺-bound form. The second set of scenes then focusses on the orthosteric site, cycling through the structures in the same order to illustrate the distinct coordination geometries of the bound Mn²⁺, and the corresponding positions of its interacting ligands (protein residues and water) in the metal-free states.

<https://elifesciences.org/articles/84006/figures#video1>

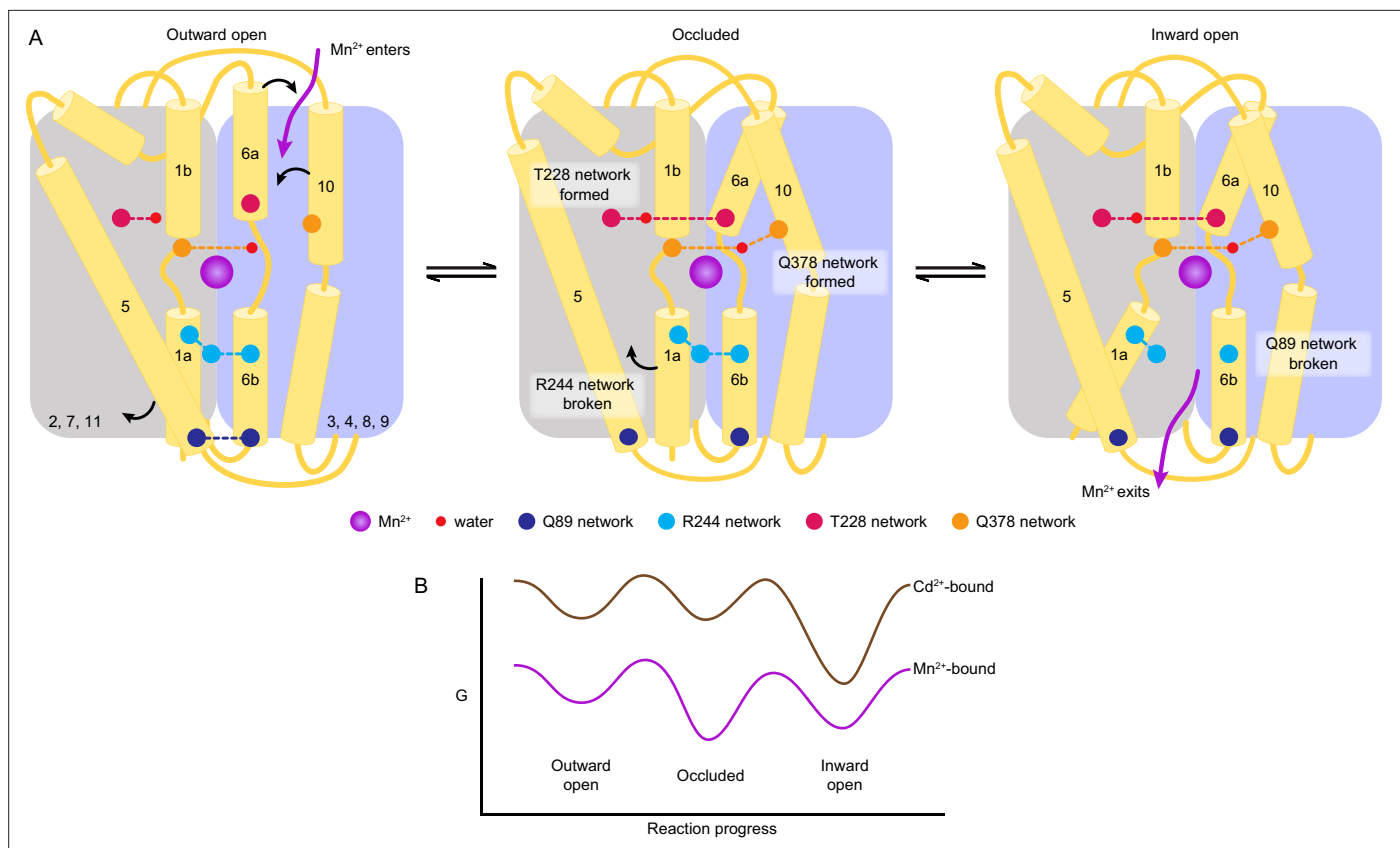


Figure 6. A structure-guided model of the conformational cycle and thermodynamic landscape of metal transport by Nramp. **(A)** After Mn²⁺ enters through the outer vestibule between TM6a and TM10 in the outward-open state, a bulk conformation change closes the outer gate. The occluded conformation arises through rearrangements of TM6a and TM10 facilitated by formation of the T228 and Q378 networks, respectively. The inner gate partially opens in the occluded state as the R244 network breaks and TM5 moves. To achieve the inward-open conformation, disruption of the Q89 network frees TM1a to swing up to fully open the inner vestibule for Mn²⁺ release into the cytosol. **(B)** Our data indicate that the most stable Mn²⁺-bound state is the occluded state, and the three main states are readily accessible to facilitate transport. In contrast, Cd²⁺ binding stabilizes the inward-open state.

The online version of this article includes the following figure supplement(s) for figure 6:

Figure supplement 1. Comparison of coordination spheres of different Mn²⁺-binding proteins: MntR, a Mn²⁺ regulator (PDB ID: 1ON1) (Glasfeld et al., 2003); PsaA, the solute-binding protein domain of Mn²⁺-transporting ATP-binding cassette transporter (PDB ID: 3ZTT) (Counago et al., 2014); and the DraNrap WT•Mn²⁺ occluded structure (this work).

by two major rearrangements: (i) closing of the outer gate as the T228 and Q378 networks form and support the reorientation of TM6a and TM10, respectively, and (ii) partial opening of the inner gate through motion of TM5 and breaking of the R244 network. TM6a and TM10 approach the orthosteric site and directly (A227 in TM6a) or indirectly (Q378 in TM10 through a conserved water) coordinate Mn²⁺ and restrict solvent access from the extracellular side in the occluded structure. A227 replaces a water of the outward-open Mn²⁺-coordination sphere, thus retaining a six-coordination geometry in both conformations. Mn²⁺ prefers octahedral (six) coordination (Chen and He, 2008; Dudev et al., 2006; Dudev and Lim, 2014) as in the outward-open and occluded structures, although we observed distortions (Supplementary file 1f).

The inward-open state is achieved when TM1a swings up, rupturing the Q89 network and opening the inner gate. The coordination sphere changes again as a water from the open inner vestibule replaces A53 from TM1a. The TM1a swing introduces Y54 as a long-range seventh ligand. The seven-coordination of Mn²⁺ in the inward-open structure is less favored and more distorted, which may facilitate Mn²⁺ release as a solvated ion in the cytosol. In contrast, the more favorable six-coordination in the outward-open and occluded structures could help energize the global conformational changes. Our metal-free structures indicate that after Mn²⁺ release to the cytosol, the protein resets to the

outward-open state through the same occluded state. The residues in the gating networks are highly conserved, suggesting that their role is conserved across the Nrap family.

Compared to other proteins that bind Mn^{2+} but are not metal transporters, like the Mn^{2+} regulator MntR (Glasfeld et al., 2003) and PsaA, the solute-binding protein (SBP) domain of an ATP-binding cassette transporter (Couñago et al., 2014), we observe longer Mn^{2+} -coordinating bond lengths for DraNrap (Figure 6—figure supplement 1). Typical manganese-oxygen bonding distances are 2.1–2.5 Å, although ‘weak interactions’ (2.6–3.2 Å) are occasionally part of a Mn^{2+} -coordination sphere (Harding, 2000; Harding, 2001). Metal-sulfur bonding distances are longer owing to the greater van der Waals radius of sulfur (Rulisek and Vondrásek, 1998). The non-ideal metal-ligand bonding distances and angles we observe in DraNrap may allow it to avoid getting trapped in an energy minimum and thus keep moving through the conformational transitions required to transport Mn^{2+} .

While the Mn^{2+} transport mechanism and associated conformational changes in DraNrap differ appreciably from other LeuT-fold transporters (Bozzi and Gaudet, 2021; Bozzi et al., 2019b), the presence of key residues defining the extracellular and intracellular gates is a common theme (Coleman et al., 2016; Coleman et al., 2019; Faham et al., 2008; Krishnamurthy and Gouaux, 2012; Krishnamurthy et al., 2009; Perez et al., 2012; Shimamura et al., 2010; Watanabe et al., 2010). Comparing the gating networks described for these transporters with DraNrap, the positions and nature of the gating networks are generally not conserved, but two common themes emerge. First, opening (or closing) a particular vestibule often involves a pair of changes, as in the DraNrap intracellular vestibule (Figure 6A). Such paired changes have been associated with ‘thin’ and ‘thick’ gates in Mhp1 (Simmons et al., 2014) and LeuT (Krishnamurthy and Gouaux, 2012), that is, gates based on sidechain and helix motions, respectively. Second, some gating residue positions are shared, but the networks are not, indicating that different families have evolved analogous networks to stabilize equivalent conformations (Coleman et al., 2016; Krishnamurthy and Gouaux, 2012).

The accumulated DraNrap structures also provide clues as to the thermodynamic landscape of the transport process (Figure 6B). Interestingly, both metal-free and Mn^{2+} -bound WT DraNrap crystallized in the occluded conformation, whereas the open conformations were achieved through conformation-locking (Bozzi et al., 2019b) or mutations of functionally important residues (Bozzi et al., 2016b). This occluded state more closely resembles an inward-open conformation (Bozzi et al., 2019b) and is an important intermediate in the Mn^{2+} transport cycle, with both local changes in coordination geometry and global changes in protein conformation in comparison to the outward-open state. Physiologically, we naïvely expect outward-open, rather than occluded, to be the preferred substrate-free conformation. However, the occluded or inward-open states of other LeuT-fold importers are also more stable, with changes in environmental conditions or the presence of substrate stabilizing specific states and lowering barriers to conformational transitions (Del Alamo et al., 2022). For example, SGLT1 and DraNrap require a negative membrane potential to transport substrates and this negative membrane potential stabilizes the outward-open state of SGLT1 (Bozzi et al., 2019a; Loo et al., 1998). Overall, our DraNrap structures suggest that the occluded state is most stable (at least in the absence of a membrane potential). The occluded and inward-open states may be energetically similar as relatively small perturbations yielded inward-open structures (M230A• Mn^{2+} and D296A• Mn^{2+}). Furthermore, our previous cysteine accessibility data indicate that the energy barriers between states are low enough for the protein to readily sample the outward- and inward-open states in cell membranes in the absence or presence of metal substrate (Bozzi et al., 2016b; Bozzi et al., 2019b).

In contrast to WT DraNrap, ScaDMT was crystallized in an inward-open state, although its TM1a was deleted from the protein construct (Ehrnstorfer et al., 2014). This deletion would prevent inner vestibule closure, and thus likely affects its energetically preferred conformation. In the case of the *Eremococcus coleocola* Nrap homolog (EcoDMT), both substrate-free and inhibitor-bound conformations are outward open (Ehrnstorfer et al., 2017; Manatschal et al., 2019), suggesting that the outward-open state is its most stable state. Of note, EcoDMT was crystallized in detergent, whereas the DraNrap structures were obtained in a monoolein lipid bilayer environment. Further studies will be needed to determine to what extent the thermodynamic landscape we begin to outline here for DraNrap is conserved in other Nrap homologs.

Previous metal selectivity studies indicate that Nrap imports different substrates with distinct mechanisms (Bozzi et al., 2016a; Bozzi et al., 2019a; Bozzi and Gaudet, 2021; Ehrnstorfer et al.,

2014; Nevo and Nelson, 2006). These differences correlate with chemical properties (Irving and Williams, 1953) and preferred coordination chemistry (Ma et al., 2009; Singh et al., 2020) of these transition metals as observed in other transition metal binding proteins like the Psa permease (Begg et al., 2015; Couñago et al., 2014), and cation diffusion facilitators (CDFs) (Barber-Zucker et al., 2017). That WT•Cd²⁺ retains the M230 sulfur as a coordinating ligand but excludes the D56 carboxylate can be rationalized by the fact that Cd²⁺ is a softer metal than Mn²⁺. This difference in coordination likely alters the pKa of nearby residues and water molecules to perturb the proton pathway such that DraNrap co-transporters protons with Mn²⁺ but not Cd²⁺ (Bozzi et al., 2019a; Bozzi et al., 2019b), although our structures do not yet fully elucidate this mechanistic difference. Moreover, owing to its larger radius, Cd²⁺ tends to form weaker complexes than Mn²⁺ with comparable coordination numbers in complexes dominated by harder ligands (Rulisek and Vondrásek, 1998; Singh et al., 2020). The architecture of the orthosteric site in DraNrap appears to facilitate six- or seven-coordinated metal complexes, and thus most Cd²⁺-bound DraNrap conformations will be less stable than the Mn²⁺-bound states. Our ITC data also indicate that binding of the toxic Cd²⁺ to DraNrap is less robust to perturbations compared to its physiological substrate Mn²⁺. Furthermore, Cd²⁺ only binds well to the inward-open state, and its binding is exothermic rather than endothermic for Mn²⁺ (Figure 5A). The crystallographic data similarly suggest that inward-open is the most stable Cd²⁺-bound state (Figure 6B), based on the following observations: (i) soaking crystals of occluded WT yielded inward-occluded WT•Cd²⁺, (ii) co-crystallization efforts yielded no other Cd²⁺-bound structures, and (iii) soaking Cd²⁺ into crystals of outward-locked G223W yielded very poor diffraction and no structures.

Overall, our data show that the orthosteric metal-binding site of DraNrap, conserved across all Nramps, is best suited to the physiological substrate Mn²⁺ (and likely the similar ion Fe²⁺). The distinct interactions of Nramps with Mn²⁺ and Cd²⁺ could be leveraged for the design of therapies for metal toxicity and prevention strategies for toxic metal accumulation in crops. These results also lay a foundation for future studies of how metal ion transporters like Nramps evolve their substrate selectivity, for example in response to different environmental conditions. Finally, the first complete set of structures with the same homolog, both in substrate-free and substrate-bound states, suggest a substrate-specific thermodynamic landscape of the transport cycle and provide a framework for future experiments and simulations to fully define this landscape, and for comparisons to other LeuT-fold transporters.

Materials and methods

Key resources table

Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Gene (<i>Deinococcus radiodurans</i>)	DraNrap	Genomic DNA	Uniprot: Q9RTP8	
Strain, strain background (<i>Escherichia coli</i>)	C41(DE3)	Lucigen	60442-1	Chemically competent cells
Strain, strain background (<i>Escherichia coli</i>)	DH5α	Invitrogen	18265-017	Chemically competent cells
Recombinant DNA reagent	pET21a	Novagen	69740-3	Vector backbone for cloning DraNrap
Chemical compound, drug	n-Dodecyl-β-D-Maltopyranoside	Anatrace	D310S	1% for solubilizing membrane, 0.03% used in wash buffer
Chemical compound, drug	n-Decyl-β-D-Maltopyranoside	Anatrace	D322S	0.1% used in exchange buffer
Chemical compound, drug	Lauryl maltose neopentyl glycol	Anatrace	NG310	0.01% in elution buffer, 0.003% used in SEC buffer
Chemical compound, drug	Monoolein	Anatrace	LCP18	1:1.5 (protein: monoolein) for LCP crystallization
Chemical compound, drug	Fura-2 Pentapotassium Salt, cell impermeant	Life Technologies	F-1200	5 mM mixed with in proteoliposome
Chemical compound, drug	1-Palmitoyl-2-oleoyl-sn-glycero-3-phospho-(1'-rac-glycerol) (POPG)	Avanti Polar Lipids	850457 C	lipid mixture (3 POPE: 1 POPG) and protein in 400:1 ratio for proteoliposome preparation
Chemical compound, drug	1-Palmitoyl-2-oleoyl-sn-glycero-3-phosphoethanolamine (POPE)	Avanti Polar Lipids	840757 C	lipid mixture (3 POPE: 1 POPG) and protein in 400:1 ratio for proteoliposome preparation
Chemical compound, drug	Valinomycin	Sigma-Aldrich	V0627	Creates membrane potential by transporting K ⁺ in proteoliposome assay

Continued on next page

Continued

Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Commercial assay, kit	Ni Sepharose High Performance Resin	Cytiva	95055–838	IMAC resin
Commercial assay, kit	Superdex 200 10/300 GL	Cytiva	89497–272	SEC column
Commercial assay, kit	PD-10 Desalting Column	Cytiva	95017–001	Buffer exchange
Software, algorithm	XDS	PMID:20124692	RRID:SCR_015652	Data Porcessing
Software, algorithm	PHASER	PMID:19461840	RRID:SCR_014219	Model Building
Software, algorithm	PHENIX	PMID:22505256	RRID:SCR_014224	Refinement
Software, algorithm	coot	PMID:20383002	RRID:SCR_014222	Refinement and model building
Software, algorithm	PyMOL	Schrödinger	RRID:SCR_000305	Figure making
Software, algorithm	HMMER	hmmer.org	RRID:SCR_005305	Collected sequences with jackhmmer and aligned with hmmalign
Software, algorithm	MUSCLE	PMID:15318951	RRID:SCR_011812	Aligning sequences
Software, algorithm	RAXML-NG	PMID:31070718	RRID:SCR_022066	Tree building
Software, algorithm	PropKa	PMID:26596171		Identifying protonation states
Software, algorithm	CHARMM-GUI	PMID:25130509		Building MD system
Software, algorithm	NAMD	PMID:20675161	RRID:SCR_014894	Running MD simulations
Software, algorithm	mdtraj	PMID:26488642		Analyzing MD data
Software, algorithm	VMD	PMID:8744570	RRID:SCR_001820	Analyzing MD data

Cloning and protein expression vectors

The DraNrapm WT and mutant constructs were cloned into pET21a-N8H (Bozzi et al., 2016a). Primer sequences for the mutations are listed in **Supplementary file 1h**. All constructs used for crystallization had a truncation of 31 residues at the N-terminus (ΔN31, which does not impair metal transport), except for A47W which was full-length and transport deficient (Bozzi et al., 2016b). For proteoliposome-based transport assays, the full-length versions of each construct were used. For ITC, full-length DraNrapm constructs were cloned into pET21-NStrep (Bozzi et al., 2016a) to avoid background signal from metals (Mn²⁺, Cd²⁺) binding to the His-tag. All mutations were introduced by site-directed mutagenesis using the Quikchange mutagenesis protocol (Stratagene) and confirmed by Sanger DNA sequencing.

Protein expression

Protein expression was performed as previously described (Bozzi et al., 2016a). Briefly, transformed *Escherichia coli* C41(DE3) (Lucigen) were induced with 0.1 mM isopropyl-β-D-thiogalactopyranoside and cultured at 18 °C for 16 hr. Cell pellets from 10 L of culture were harvested and flash frozen in liquid nitrogen.

Protein purification for crystallography

Cells were thawed and resuspended in 50 mL load buffer (20 mM sodium phosphate, pH 7.5, 55 mM imidazole pH 7.5, 500 mM NaCl, 10% (v/v) glycerol) supplemented with 1 mM PMSF, 1 mM benzamidine, 0.3 mg/mL DNase I and 0.3 mg/mL lysozyme and lysed by sonication on ice (six cycles of 45 s with a Branson Sonifier 450 under duty cycle of 65% and output 10). Lysates were cleared by centrifuging for 20 min at 20,000 rpm (Beckman JA-20) and membranes pelleted from the supernatant by ultracentrifugation at 45,000 rpm (Beckman type 45Ti) for 70 min. Membranes were homogenized in 70 mL load buffer using a glass Potter-Elvehjem grinder, solubilized for 1 hr in 1% (w/v) n-dodecyl-β-D-maltopyranoside (DDM), then ultracentrifuged at 35,000 (Beckman type 45Ti) for 35 min to remove insoluble debris. Pre-equilibrated Ni-Sepharose beads (2 mL; GE Healthcare) were incubated with the supernatant for 90 min at 4 °C, then washed with 20 column volumes (CV) of each of the following buffers sequentially (i) load buffer containing 0.03% DDM, (ii) load buffer containing 0.5% lauryl maltose neopentyl glycol (LMNG), and (iii) load buffer containing 0.1% LMNG. Protein was eluted in 20 mM sodium phosphate, pH 7.5, 450 mM imidazole pH 7.5, 500 mM NaCl, 10%

(v/v) glycerol, 0.01% LMNG, concentrated to <0.5 mL in a 50 kDa molecular weight cutoff (MWCO) centrifugal concentrator (EMD Millipore), and purified by size exclusion chromatography (SEC) using a Superdex S200 10/300 (GE Healthcare) pre-equilibrated with SEC buffer (10 mM HEPES pH 7.5, 150 mM NaCl, 0.003% LMNG). Peak protein fractions enriched in DraNrap were combined, concentrated to ~25–40 mg/mL using a 50 kDa MWCO centrifugal concentrator, aliquoted and flash frozen in liquid nitrogen and stored at –80 °C. Purifications of each protein construct were performed at least twice and resulted in similar data.

Purification of DraNrap for ITC

To purify protein for ITC, harvested cells expressing strep-tagged DraNrap from 10 L of culture were resuspended in 50 mL of buffer W (100 mM Tris, pH 8.0, 150 mM NaCl), and membranes were isolated, homogenized and solubilized in 1% DDM as above. The supernatant was incubated with Strep-Tactin Superflow resin (3 mL; IBA) pre-equilibrated with buffer W+0.03% DDM and washed with the following buffers sequentially: (i) 1 CV buffer W+0.03% DDM, (ii) 2 CV buffer W+0.5% LMNG and (iii) 2 CV buffer W+0.1% LMNG. Protein was eluted with 3 CV buffer W+0.01% LMNG+2.5 mM desthiobiotin. The eluted protein was concentrated up to 2.5 mL using a 50 kDa MWCO centrifugal concentrator and buffer-exchanged into 150 mM NaCl, 10 mM HEPES, pH 7.5, and 0.003% LMNG using disposable PD-10 desalting columns (GE healthcare). Protein was concentrated to ~2.5 mg/mL a 50 kDa MWCO centrifugal concentrator and flash frozen in liquid nitrogen and stored at –80 °C. Purifications of each protein construct were performed at least twice and resulted in similar data.

DraNrap crystallization

Crystallization of all constructs was performed using lipidic cubic phase (LCP). Protein was mixed with monoolein in 1:1.5 volume ratio using the syringe reconstitution method (Bozzi et al., 2019b). The protein bolus (60 nL) and 720 nL precipitant were dispensed onto custom-made 96 well glass sandwich plates using an NT8 drop-setting robot (Formulatrix). Metal-free crystals (WT) and metal supplemented (5 mM MnCl_2) co-crystals (A47W• Mn^{2+} , M230A• Mn^{2+} , D296A• Mn^{2+}) were grown in different precipitant conditions (Supplementary file 1a), harvested within 7–10 days (after reaching their optimal size of 30–40 μm rods) using mesh loops (MiTeGen) and flash-frozen in liquid nitrogen prior to data collection. Some structures (WT• Cd^{2+} , WT• Mn^{2+} , WT_{soak}) were obtained by soaking WT DraNrap crystals grown in metal-free precipitant (Supplementary file 1a) for 7–10 days: The glass covering the wells was broken without disturbing the bolus and 2 μL soak solution (Supplementary file 1a) were added before resealing with a fresh siliconized glass coverslip, incubating overnight (16–18 hr), then harvesting and flash freezing for data collection.

X-ray diffraction data collection and processing

Diffraction data for structure determination and refinement were collected at beamlines 24-ID-C or 23-ID-B of the Advanced Photon Source at wavelengths of 0.984 Å or 1.033 Å, respectively. We used anomalous signals to confirm the presence of metals (Mn^{2+} or Cd^{2+}) in the binding site. For D296A• Mn^{2+} and A47W• Mn^{2+} , the same data we used for structure refinement and collected at 1.033 Å, 0.984 Å, respectively, provided strong anomalous signal in the metal-binding sites. For WT• Cd^{2+} , we were able to collect data at 1.904 Å (near the low-energy boundary for the beamline), to maximize the anomalous signal. Locations of the crystals in the mesh loops were identified by grid scanning with a 20 μm beam at 10% transmission followed by data collection with a 10 μm beam at 15% transmission. Data were indexed in XDS (Kabsch, 2010) and scaled in CCP4 AIMLESS (Version 7.0) (Evans and Murshudov, 2013; Winn et al., 2011). For datasets collected from several crystals, data from each crystal were independently indexed and integrated, then combined during scaling using CCP4 AIMLESS (Evans and Murshudov, 2013; Winn et al., 2011) to obtain complete datasets. The resolution cut-off of each structure was defined based on CC1/2 values of 0.3 and above (Karplus and Diederichs, 2012; Karplus and Diederichs, 2015). Initial phases for all structures were determined by molecular replacement in PHENIX (Version 1.17.1–3660) (Liebschner et al., 2019) using an occluded structure of DraNrap (PDB ID 6C3I chain A) (Bozzi et al., 2019b) as search model. Data statistics are listed in Table 1 and Supplementary file 1b and d.

Model building, refinement, and analysis

Models were built in COOT (Version 7.0) (*Emsley et al., 2010*) and refined in PHENIX (*Liebschner et al., 2019*), with macrocycles including reciprocal space, TLS groups, and individual B-factor refinement, and optimization of the X-ray/stereochemistry and X-ray/ADP weights. For WT•Cd²⁺, ‘anomalous group refinement’ was used to improve the fit to density of the Cd²⁺ ions, with Cd²⁺ as an ‘anomalous group’ with the reference *f'* and *f''* values suggested by phenix.form.factor (−0.462 and 2.132, respectively). Ligand restraints for monoolein and spermidine were generated in Phenix.elbow with automatic geometry optimization. All structures contain one protein molecule in the asymmetric unit. The final structures span from residues 45–48 to residues 433–436, except that residues 240–249 and 240–247 were not modeled in D296•Mn²⁺ and WT•Cd²⁺, respectively, because of lack of interpretable electron density map. Model refinement statistics are listed in **Table 1** and **Supplementary file 1b and d**. Pairwise RMSD for all structures are listed in **Supplementary file 1c**. Anomalous difference Fourier maps for Mn²⁺ (D296A•Mn²⁺ and A47W•Mn²⁺) and Cd²⁺ (WT•Cd²⁺) were generated in phenix.maps using a high-resolution cutoff of 3.5–4.5 Å. Polder maps omitting the metal ions were generated in phenix.polder to appropriately define the coordination sphere for Mn²⁺ and Cd²⁺ in all structures. All software were provided by SGrid (*Morin et al., 2013*).

Metal binding measurements using ITC

ITC experiments were performed using MicroCal iTC200 (GE Healthcare) to determine the affinity and thermodynamic parameters of binding of divalent metals (Mn²⁺, Cd²⁺ and Mg²⁺) to DraNrap (WT and its mutants) (*Leavitt and Freire, 2001; Wiseman et al., 1989*). All protein and metal solutions were prepared in ITC buffer (150 mM NaCl, 10 mM HEPES, pH 7.5, and 0.003% LMNG). The sample cell containing 25 μM protein was titrated with 20 2 μL injections of 6 mM metal, with an interval of 120 s between each successive 5 s injection, with a 750 rpm stirring rate at 25 °C. To nullify the heat of dilution, the data from titration of a metal solution into ITC buffer (‘buffer blank’ runs) were subtracted from the metal-protein titration curves prior to model fitting. Data were fitted and analyzed as detailed in Appendix 1. Briefly, as per best practice when *c* values (association constant × molar protein concentration) are below 1 (*Piccolo et al., 2009; Tellinghuisen, 2008; Turnbull and Daranas, 2003*), all the data reported for each construct are fitted fixing the number of sites (one-site binding model with fixed *n*=1 or sequential binding model with fixed *n*=2). The binding stoichiometry was selected based the model fits and knowledge from the crystal structures and mutational analysis. Data were fitted with Origin 7 software. The mean *K_d* values ± SEM from 2 to 3 repeats (from independent protein purifications) for each sample are reported in **Supplementary file 1g**.

Proteoliposome-based in vitro transport assays

Protein purification, liposome preparation, and metal transport assays were performed as described (*Bozzi et al., 2016a; Bozzi et al., 2019a*). Purifications of each protein construct were performed at least twice and resulted in similar data. For each construct, the presented data originates from two to three batches of reconstituted liposomes, each from an independent protein purification, represented as scatter plots of each data point and the corresponding mean.

Sequence alignments

We used 92 Nrap sequences from Pfam (*El-Gebali et al., 2019*) to build a seed alignment using MUSCLE (*Edgar, 2004*). We collected 15,451 sequences from Uniprot (*Bateman et al., 2021*) using HMMER (*Potter et al., 2018*). We used HMMER’s hmalign, with a hidden Markov model profile from the seed alignment as an input, to align all 15,451 sequences. We applied filters to retain sequences 400–600 residues in length and sequences with under 90% pairwise sequence identity, respectively. The final alignment contains 6712 sequences and is well aligned at biologically relevant residues (**Figure 1—source data 1**). A maximum-likelihood phylogenetic tree was generated via RAXML-NG *Kozlov et al., 2019* using the LG substitution model (*Le and Gascuel, 2008*), with the likeliest final tree selected from 10 parallel optimization trials (**Figure 1—source data 2**). The canonical Nrap clade in this tree was identified based on conservation of the ‘DPGN’ and ‘MPH’ motifs in trans-membrane helices 1 and 6, respectively, and contained 3796 sequences. The Nrap-related magnesium transporters were used to root the canonical Nrap phylogeny. Sequence analysis was done

with Biopython (Cock et al., 2009) and sequence logos were generated with logomaker (Tareen and Kinney, 2020) using a 'chemistry' color scheme inspired by WebLogo (Crooks et al., 2004).

Molecular dynamics simulation

Molecular dynamics (MD) simulations were initialized from three high-resolution structures of DraN-ramp: the outward-open G223W•Mn²⁺ structure (6BU5) with Mn²⁺ removed and W223 mutated back to the native glycine residue in silico, the inward-open WT•Cd²⁺ structure with Cd²⁺ removed, and the inward-occluded WT structure. Crystallographic waters were retained, and protonation states of key titratable residues were selected with PROPKA (Olsson et al., 2011; Søndergaard et al., 2011) assuming a pH of 5.0 for residues exposed to external solvent and a pH of 7.0 for residues exposed to cytosol, a condition under which DraN-ramp exhibits high activity. All structures were oriented in the membrane with the PPM web server and membrane systems were prepared with CHARMM-GUI (Jo et al., 2008; Lee et al., 2016). A POPC membrane of surface area 99×99 Å was constructed in the XY plane around the protein (Wu et al., 2014), the system was solvated in a 100×100×100 Å³ rectangular box using TIP3 waters and electronically neutralized using potassium and chlorine ions at an overall concentration of 150 mM. The overall system size was approximately 103,000 atoms.

All-atom simulations were run using GPU-accelerated NAMD (Phillips et al., 2008) and the CHARMM36m forcefield (Huang et al., 2017). Prior to simulation, the energy of each system was minimized for 10,000 steps using a conjugate gradient and line search algorithm native to NAMD. To improve simulation stability, the system was initially equilibrated using an NVT-ensemble with harmonic restraints placed on protein and lipid heavy atoms. The harmonic restraints were then incrementally relaxed over a period of 675 ps according to established CHARMM-GUI protocols (Lee et al., 2016). The system was then simulated at a constant pressure, utilizing the Langevin piston method to maintain 1 atm at 303.15 K, from anywhere between 617 and 1176 ns depending on the starting conformation. Simulations were performed using periodic boundary conditions and a time step of 3.0 fs with all bonds to hydrogens being constrained. Large integration timesteps were enabled by employing hydrogen mass repartitioning (Hopkins et al., 2015). Long-range electrostatic interactions were calculated using the particle mesh Ewald (PME) method with nonbonded interactions being cut off at 12 Å. Each simulation was performed in duplicate resulting in approximately 2 μs of total sampling for each system. Simulations are summarized in **Supplementary file 1i**.

RMSD, residue distance, and dihedral analyses were performed using mdtraj version 1.9.8 (McGibbon et al., 2015). For distance analysis, the minimum interatomic distances were identified between the specified residues across each frame. For water analysis, simulations were centered and wrapped in VMD and a Tcl script was used to produce water density maps. Contour maps of these densities were then visualized in PyMOL. Water occupancies of sites coordinated by specific residues were also calculated in an alignment-agnostic manner by determining for each frame in each simulation whether a water was present within 2.5 Å of both specified residues. Distinct rotamers were identified from dihedrals using spectral clustering as implemented in scikit-learn version 1.0.

Data availability

Atomic coordinates and structure factors for the crystal structures reported in this work have been deposited to the Protein Data Bank under accession numbers 8E5S (WT), 8E5V (WT_{soak}), 8E60 (WT•Mn²⁺), 8E6H (A47W•Mn²⁺), 8E6I (M230A•Mn²⁺), 8E6L (D296A•Mn²⁺), 8E6M (WT•Cd²⁺), and 8E6N (re-refined G223W•Mn²⁺). Corresponding X-ray diffraction images have been deposited to the SBGrid Data Bank under the respective accession numbers 962 (doi:10.15785/SBGRID/962), 963 (doi:10.15785/SBGRID/963), 964 (doi:10.15785/SBGRID/964), 966 (doi:10.15785/SBGRID/966), 967 (doi:10.15785/SBGRID/967), 968 (doi:10.15785/SBGRID/968), 969 (doi:10.15785/SBGRID/969), and previously deposited 564 (doi:10.15785/SBGRID/564). The multiple sequence alignment and phylogenetic tree have been provided as **Figure 1—source data 1** and **Figure 1—source data 2**, respectively. All liposome-based transport data are provided in **Figure 1—source data 3**. Code for analysis of molecular dynamics data, as well as the raw data plotted in **Figure 4—figure supplement 2** and **Figure 4—figure supplement 3**, can be found at <https://github.com/samberry19/nramp-md> (MIT license). Raw molecular dynamics trajectory files are available on Dryad (<https://doi.org/10.5061/dryad.tx95x6b2b>). Source files (origin files) of all ITC experiments are provided in **Appendix 1—table 1—source data 1**.

(Mn²⁺ isotherms), **Appendix 1—table 2—source data 1** (Cd²⁺ isotherms) and **Figure 5—source data 1** (Mg²⁺ isotherms).

Acknowledgements

We thank Arghya Deb for discussions of metal coordination chemistry, and previous and current members of the Gaudet lab for discussions and assistance, particularly Gerardo Zavala and Edward Lee for contributions to preliminary molecular dynamics analyses, and José Velilla for help with crystal soaking and fishing prior to data collection. This work was funded by NIGMS grant R01GM120996 (RG), a National Science Foundation (NSF) CAREER award MCB-1942763 (AS), AstraZeneca and ASU-Mayo Foundation (EW), and the NSF-Simons Center for Mathematical and Statistical Analysis of Biology at Harvard (award number 1764269) and the Harvard Quantitative Biology Initiative (SB). Diffraction data reported in this study were collected at NE-CAT beamline 24IDC and GM/CA beamline 23IDB in the Advanced Photon Source. NE-CAT is funded by NIGMS grant P30 GM124165 and GM/CA is funded by the National Cancer Institute (ACB-12002) and the National Institute of General Medical Sciences (AGM-12006, P30GM138396). The Eiger 16 M detector at GM/CA-XSD is funded by NIH grant S10 OD012289. The Advanced Photon Source is a U.S. Department of Energy Facility operated by Argonne National Laboratory under Contract No. DE-AC02-06CH11357. The molecular simulations used the Extreme Science and Engineering Discovery Environment (XSEDE) supported by NSF (ACI-1548562), and Oak Ridge Leadership Computing Facility, supported by the Office of Science, Department of Energy (DE-AC05-00OR22725).

Additional information

Funding

Funder	Grant reference number	Author
National Institute of General Medical Sciences	R01GM120996	Rachelle Gaudet
National Science Foundation	MCB-1942763	Abhishek Singharoy
National Science Foundation	1764269	Samuel P Berry

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

Author contributions

Shamayeeta Ray, Conceptualization, Data curation, Validation, Investigation, Visualization, Methodology, Writing - original draft, Writing - review and editing; Samuel P Berry, Data curation, Formal analysis, Investigation, Visualization, Writing - original draft, Writing - review and editing; Eric A Wilson, Data curation, Formal analysis, Investigation, Methodology, Writing - review and editing; Casey H Zhang, Data curation, Formal analysis, Validation, Methodology, Writing - review and editing; Mrinal Shekhar, Data curation, Investigation, Methodology, Writing - review and editing; Abhishek Singharoy, Resources, Supervision, Funding acquisition, Methodology, Project administration, Writing - review and editing; Rachelle Gaudet, Conceptualization, Resources, Supervision, Funding acquisition, Validation, Investigation, Visualization, Project administration, Writing - review and editing

Author ORCIDs

Shamayeeta Ray  <http://orcid.org/0000-0001-7906-0572>

Rachelle Gaudet  <http://orcid.org/0000-0002-9177-054X>

Decision letter and Author response

Decision letter <https://doi.org/10.7554/eLife.84006.sa1>

Author response <https://doi.org/10.7554/eLife.84006.sa2>

Additional files

Supplementary files

- Supplementary file 1. Additional tables with methodological details and results from data analyses. (a) Construct, precipitant, and soaking solutions used for each structure. (b) Data collection and refinement statistics for the four supporting new DraNramp structures. (c) Cα RMSD in Å for all DraNramp structure pairs (number of aligned residues in parentheses). (d) Data collection statistics for anomalous maps. (e) Distances to metal (Å) for coordinating atoms at the orthosteric site. (f) Coordination number and geometry of metal ions in the orthosteric site. (g) Binding affinity of metals to various DraNramp constructs. (h) Primers for Mutagenesis (5' to 3' sequence). (i) Summary of molecular dynamics simulations.
- MDAR checklist

Data availability

Atomic coordinates and structure factors for the crystal structures reported in this work have been deposited to the Protein Data Bank under accession numbers 8E5S (WT), 8E5V (WTsoak), 8E60 (WT•Mn²⁺), 8E6H (A47W•Mn²⁺), 8E6I (M230A•Mn²⁺), 8E6L (D296A•Mn²⁺), 8E6M (WT•Cd²⁺), and 8E6N (re-refined G223W•Mn²⁺). Corresponding X-ray diffraction images have been deposited to the SBGrid Data Bank under the respective accession numbers 962 (doi:10.15785/SBGRID/962), 963 (doi:10.15785/SBGRID/963), 964 (doi:10.15785/SBGRID/964), 966 (doi:10.15785/SBGRID/966), 967 (doi:10.15785/SBGRID/967), 968 (doi:10.15785/SBGRID/968), 969 (doi:10.15785/SBGRID/969), and previously deposited 564 (doi:10.15785/SBGRID/564). The multiple sequence alignment and phylogenetic tree have been provided as Figure 1-source data 1 and Figure 1-source data 2, respectively. All liposome-based transport data are provided in Figure 1-source data 3. Code for analysis of molecular dynamics data, as well as the raw data plotted in Figure 4-figure supplement 2 and Figure 4-figure supplement 3, can be found at <https://github.com/samberry19/nramp-md> (MIT license). Raw molecular dynamics trajectory files are available on Dryad (<https://doi.org/10.5061/dryad.tx95x6b2b>). Source files (origin files) of all ITC experiments are provided in Appendix 1-table 1-source data 1 (Mn²⁺ isotherms), Appendix 1-table 2-source data 1 (Cd²⁺ isotherms) and Figure 5-source data 1 (Mg²⁺ isotherms).

The following datasets were generated:

Author(s)	Year	Dataset title	Dataset URL	Database and Identifier
Wilson EA, Berry SP, Shekhar M, Gaudet R, Singharoy A	2022	Molecular dynamics simulations in: High-resolution structures with bound Mn2+ and Cd2+ map the metal import pathway in an Nramp transporter	https://dx.doi.org/10.5061/dryad.tx95x6b2b	Dryad Digital Repository, 10.5061/dryad.tx95x6b2b
Ray S, Gaudet R	2022	X-ray structure of the Deinococcus radiodurans Nramp/MntH divalent transition metal transporter WT in an occluded state	https://www.rcsb.org/structure/8E5S	RCSB Protein Data Bank, 8E5S
Ray S, Gaudet R	2022	X-ray structure of the Deinococcus radiodurans Nramp/MntH divalent transition metal transporter WTsoak in an occluded state	https://www.rcsb.org/structure/8E5V	RCSB Protein Data Bank, 8E5V
Ray S, Gaudet R	2022	X-ray structure of the Deinococcus radiodurans Nramp/MntH divalent transition metal transporter WT in an occluded, manganese-bound state	https://www.rcsb.org/structure/8E60	RCSB Protein Data Bank, 8E60

Continued on next page

Continued

Author(s)	Year	Dataset title	Dataset URL	Database and Identifier
Ray S, Gaudet R	2022	X-ray structure of the <i>Deinococcus radiodurans</i> Nramp/MntH divalent transition metal transporter A47W mutant in an occluded, manganese-bound state	https://www.rcsb.org/structure/8E6H	RCSB Protein Data Bank, 8E6H
Ray S, Gaudet R	2022	X-ray structure of the <i>Deinococcus radiodurans</i> Nramp/MntH divalent transition metal transporter M230A mutant in an inward-open, manganese-bound state	https://www.rcsb.org/structure/8E6I	RCSB Protein Data Bank, 8E6I
Ray S, Gaudet R	2022	X-ray structure of the <i>Deinococcus radiodurans</i> Nramp/MntH divalent transition metal transporter D296A mutant in an inward-open, manganese-bound state	https://www.rcsb.org/structure/8E6L	RCSB Protein Data Bank, 8E6L
Ray S, Gaudet R	2022	X-ray structure of the <i>Deinococcus radiodurans</i> Nramp/MntH divalent transition metal transporter WT in an inward-open, cadmium-bound state	https://www.rcsb.org/structure/8E6M	RCSB Protein Data Bank, 8E6M
Ray S, Gaudet R	2022	X-ray structure of the <i>Deinococcus radiodurans</i> Nramp/MntH divalent transition metal transporter G223W mutant in an outward-open, manganese-bound state	https://www.rcsb.org/structure/8E6N	RCSB Protein Data Bank, 8E6N
Ray S, Gaudet R	2022	X-Ray Diffraction data from WT Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E5S structure	https://data.sbgrid.org/dataset/962	SBGrid Data Bank, 10.15785/SBGRID/962
Ray S, Gaudet R	2022	X-Ray Diffraction data from WTsoak Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E5V structure	https://data.sbgrid.org/dataset/963	SBGrid Data Bank, 10.15785/SBGRID/963
Ray S, Gaudet R	2022	X-Ray Diffraction data from Manganese-bound WT Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E60 structure	https://data.sbgrid.org/dataset/964	SBGrid Data Bank, 10.15785/SBGRID/964
Ray S, Gaudet R	2022	X-Ray Diffraction data from Mn-bound A47W mutant Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E6H structure	https://data.sbgrid.org/dataset/966	SBGrid Data Bank, 10.15785/SBGRID/966

Continued on next page

Continued

Author(s)	Year	Dataset title	Dataset URL	Database and Identifier
Ray S, Gaudet R	2022	X-Ray Diffraction data from Mn-bound M230A mutant Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E6I structure	https://data.sbgrid.org/dataset/967	SBGrid Data Bank, 10.15785/SBGRID/967
Ray S, Gaudet R	2022	X-Ray Diffraction data from Mn-bound D296A mutant Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E6L structure	https://data.sbgrid.org/dataset/968	SBGrid Data Bank, 10.15785/SBGRID/968
Ray S, Gaudet R	2022	X-Ray Diffraction data from Cadmium-bound WT Nramp/MntH divalent transition metal transporter from <i>Deinococcus radiodurans</i> , source of 8E6M structure	https://data.sbgrid.org/dataset/969	SBGrid Data Bank, 10.15785/SBGRID/969

The following previously published dataset was used:

Author(s)	Year	Dataset title	Dataset URL	Database and Identifier
Bozzi AT, Nicoludis JM, Gaudet R	2019	X-Ray Diffraction data from <i>Deinococcus radiodurans</i> Nramp/MntH divalent transition metal transporter in the outward-open, mangan, source of 6BU5 structure	https://data.sbgrid.org/dataset/564/	SBGrid Data Bank, 10.15785/SBGRID/564

References

- Andrews NC.** 2002. Metal transporters and disease. *Current Opinion in Chemical Biology* **6**:181–186. DOI: [https://doi.org/10.1016/s1367-5931\(02\)00307-1](https://doi.org/10.1016/s1367-5931(02)00307-1), PMID: 12039002
- Barber-Zucker S, Shaanan B, Zarivach R.** 2017. Transition metal binding selectivity in proteins and its correlation with the phylogenomic classification of the cation diffusion facilitator protein family. *Scientific Reports* **7**:16381. DOI: <https://doi.org/10.1038/s41598-017-16777-5>, PMID: 29180655
- Bateman A, Martin MJ, Orchard S, Magrane M, Agivetova R, Ahmad S, Alpi E, Bowler-Barnett EH, Britto R, Bursteinas B, Bye-A-Jee H, Coetzee R, Cukura A, Da Silva A, Denny P, Dogan T, Ebenezer T, Fan J, Castro LG, Garmiri P, et al.** 2021. UniProt: the universal protein knowledgebase in 2021. *Nucleic Acids Research* **49**:D480–D489. DOI: <https://doi.org/10.1093/nar/gkaa1100>, PMID: 33237286
- Begg SL, Eijkelkamp BA, Luo Z, Couñago RM, Morey JR, Maher MJ, Ong C-LY, McEwan AG, Kobe B, O'Mara ML, Paton JC, McDevitt CA.** 2015. Dysregulation of transition metal ion homeostasis is the molecular basis for cadmium toxicity in *Streptococcus pneumoniae*. *Nature Communications* **6**:6418. DOI: <https://doi.org/10.1038/ncomms7418>, PMID: 25731976
- Bleackley MR, Macgillivray RTA.** 2011. Transition metal homeostasis: from yeast to human disease. *Biomaterials* **24**:785–809. DOI: <https://doi.org/10.1007/s10534-011-9451-4>, PMID: 21479832
- Bozzi AT, Bane LB, Weihofen WA, McCabe AL, Singharoy A, Chipot CJ, Schulten K, Gaudet R.** 2016a. Conserved methionine dictates substrate preference in nramp-family divalent metal transporters. *PNAS* **113**:10310–10315. DOI: <https://doi.org/10.1073/pnas.1607734113>, PMID: 27573840
- Bozzi AT, Bane LB, Weihofen WA, Singharoy A, Guillen ER, Ploegh HL, Schulten K, Gaudet R.** 2016b. Crystal structure and conformational change mechanism of a bacterial nramp-family divalent metal transporter. *Structure* **24**:2102–2114. DOI: <https://doi.org/10.1016/j.str.2016.09.017>, PMID: 27839948
- Bozzi AT, Bane LB, Zimanyi CM, Gaudet R.** 2019a. Unique structural features in an nramp metal transporter impart substrate-specific proton cotransport and a kinetic bias to favor import. *The Journal of General Physiology* **151**:1413–1429. DOI: <https://doi.org/10.1085/jgp.201912428>, PMID: 31619456
- Bozzi AT, Zimanyi CM, Nicoludis JM, Lee BK, Zhang CH, Gaudet R.** 2019b. Structures in multiple conformations reveal distinct transition metal and proton pathways in an Nramp transporter. *eLife* **8**:e41124. DOI: <https://doi.org/10.7554/eLife.41124>, PMID: 30714568

- Bozzi AT**, McCabe AL, Barnett BC, Gaudet R. 2020. Transmembrane helix 6b links proton and metal release pathways and drives conformational change in an nramp-family transition metal transporter. *The Journal of Biological Chemistry* **295**:1212–1224. DOI: <https://doi.org/10.1074/jbc.RA119.011336>, PMID: 31882536
- Bozzi AT**, Gaudet R. 2021. Molecular mechanism of nramp-family transition metal transport. *Journal of Molecular Biology* **433**:166991. DOI: <https://doi.org/10.1016/j.jmb.2021.166991>, PMID: 33865868
- Budinger D**, Barral S, Soo AKS, Kurian MA. 2021. The role of manganese dysregulation in neurological disease: emerging evidence. *The Lancet. Neurology* **20**:956–968. DOI: [https://doi.org/10.1016/S1474-4422\(21\)00238-6](https://doi.org/10.1016/S1474-4422(21)00238-6), PMID: 34687639
- Cammack R**, Hughes M. 2008. Considerations for the Specification of Enzyme Assays Involving Metal Ions. Proceedings of the 3rd Beilstein ESCEC Symposium “Experimental Standard Conditions Of Enzyme Characterization.” Rüdesheim, Germany.
- Cellier M**, Gros P. 2004. *The NRAMP Family* New York, NY: Springer.
- Chen PR**, He C. 2008. Selective recognition of metal ions by metalloregulatory proteins. *Current Opinion in Chemical Biology* **12**:214–221. DOI: <https://doi.org/10.1016/j.cbpa.2007.12.010>, PMID: 18258210
- Cock PJA**, Antao T, Chang JT, Chapman BA, Cox CJ, Dalke A, Friedberg I, Hamelryck T, Kauff F, Wilczynski B, de Hoon MJL. 2009. Biopython: freely available python tools for computational molecular biology and bioinformatics. *Bioinformatics* **25**:1422–1423. DOI: <https://doi.org/10.1093/bioinformatics/btp163>, PMID: 19304878
- Coleman JA**, Green EM, Gouaux E. 2016. X-Ray structures and mechanism of the human serotonin transporter. *Nature* **532**:334–339. DOI: <https://doi.org/10.1038/nature17629>, PMID: 27049939
- Coleman JA**, Yang D, Zhao Z, Wen PC, Yoshioka C, Tajkhorshid E, Gouaux E. 2019. Serotonin transporter-ibogaine complexes illuminate mechanisms of inhibition and transport. *Nature* **569**:141–145. DOI: <https://doi.org/10.1038/s41586-019-1135-1>, PMID: 31019304
- Couñago RM**, Ween MP, Begg SL, Bajaj M, Zuegg J, O’Mara ML, Cooper MA, McEwan AG, Paton JC, Kobe B, McDevitt CA. 2014. Imperfect coordination chemistry facilitates metal ion release in the PSA permease. *Nature Chemical Biology* **10**:35–41. DOI: <https://doi.org/10.1038/nchembio.1382>, PMID: 24212134
- Crooks GE**, Hon G, Chandonia JM, Brenner SE. 2004. Weblogo: a sequence logo generator. *Genome Research* **14**:1188–1190. DOI: <https://doi.org/10.1101/gr.849004>, PMID: 15173120
- Davidsson L**, Lönnnerdal B, Sandström B, Kunz C, Keen CL. 1989. Identification of transferrin as the major plasma carrier protein for manganese introduced orally or intravenously or after in vitro addition in the rat. *The Journal of Nutrition* **119**:1461–1464. DOI: <https://doi.org/10.1093/jn/119.10.1461>, PMID: 2585137
- Del Alamo D**, Meiler J, Mchaourab HS. 2022. Principles of alternating access in leut-fold transporters: commonalities and divergences. *Journal of Molecular Biology* **434**:167746. DOI: <https://doi.org/10.1016/j.jmb.2022.167746>, PMID: 35843285
- Dudev M**, Wang J, Dudev T, Lim C. 2006. Factors governing the metal coordination number in metal complexes from cambridge structural database analyses. *The Journal of Physical Chemistry. B* **110**:1889–1895. DOI: <https://doi.org/10.1021/jp054975n>, PMID: 16471760
- Dudev T**, Lim C. 2014. Competition among metal ions for protein binding sites: determinants of metal ion selectivity in proteins. *Chemical Reviews* **114**:538–556. DOI: <https://doi.org/10.1021/cr4004665>, PMID: 24040963
- Edgar RC**. 2004. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* **5**:113. DOI: <https://doi.org/10.1186/1471-2105-5-113>, PMID: 15318951
- Ehrnstorfer IA**, Geertsma ER, Pardon E, Steyaert J, Dutzler R. 2014. Crystal structure of a SLC11 (NRAMP) transporter reveals the basis for transition-metal ion transport. *Nature Structural & Molecular Biology* **21**:990–996. DOI: <https://doi.org/10.1038/nsmb.2904>, PMID: 25326704
- Ehrnstorfer IA**, Manatschal C, Arnold FM, Laederach J, Dutzler R. 2017. Structural and mechanistic basis of proton-coupled metal ion transport in the SLC11/NRAMP family. *Nature Communications* **8**:14033. DOI: <https://doi.org/10.1038/ncomms14033>, PMID: 28059071
- Ekiz C**, Agaoglu L, Karakas Z, Gurel N, Yalcin I. 2005. The effect of iron deficiency anemia on the function of the immune system. *The Hematology Journal* **5**:579–583. DOI: <https://doi.org/10.1038/sj.thj.6200574>, PMID: 15692603
- El-Gebali S**, Mistry J, Bateman A, Eddy SR, Luciani A, Potter SC, Qureshi M, Richardson LJ, Salazar GA, Smart A, Sonnhammer ELL, Hirsh L, Paladin L, Piovesan D, Tosatto SCE, Finn RD. 2019. The pfam protein families database in 2019. *Nucleic Acids Research* **47**:D427–D432. DOI: <https://doi.org/10.1093/nar/gky995>, PMID: 30357350
- Emsley P**, Lohkamp B, Scott WG, Cowtan K. 2010. Features and development of coot. *Acta Crystallographica. Section D, Biological Crystallography* **66**:486–501. DOI: <https://doi.org/10.1107/S0907444910007493>, PMID: 20383002
- Evans PR**, Murshudov GN. 2013. How good are my data and what is the resolution? *Acta Crystallographica. Section D, Biological Crystallography* **69**:1204–1214. DOI: <https://doi.org/10.1107/S0907444913000061>, PMID: 23793146
- Faham S**, Watanabe A, Besserer GM, Cascio D, Specht A, Hirayama BA, Wright EM, Abramson J. 2008. The crystal structure of a sodium galactose transporter reveals mechanistic insights into na⁺/sugar symport. *Science* **321**:810–814. DOI: <https://doi.org/10.1126/science.1160406>, PMID: 18599740
- Ferrante A**, Gorski J. 2012. Enthalpy-Entropy compensation and cooperativity as thermodynamic epiphenomena of structural flexibility in ligand-receptor interactions. *Journal of Molecular Biology* **417**:454–467. DOI: <https://doi.org/10.1016/j.jmb.2012.01.057>, PMID: 22342886

- Glasfeld A, Guedon E, Helmann JD, Brennan RG. 2003. Structure of the manganese-bound manganese transport regulator of *Bacillus subtilis*. *Nature Structural Biology* **10**:652–657. DOI: <https://doi.org/10.1038/nsb951>, PMID: 12847518
- Harding MM. 2000. The geometry of metal-ligand interactions relevant to proteins. II. angles at the metal atom, additional weak metal-donor interactions. *Acta Crystallographica. Section D, Biological Crystallography* **56**:857–867. DOI: <https://doi.org/10.1107/s0907444900005849>, PMID: 10930832
- Harding MM. 2001. Geometry of metal-ligand interactions in proteins. *Acta Crystallographica. Section D, Biological Crystallography* **57**:401–411. DOI: <https://doi.org/10.1107/s0907444900019168>, PMID: 11223517
- Hopkins CW, Le Grand S, Walker RC, Roitberg AE. 2015. Long-time-step molecular dynamics through hydrogen mass repartitioning. *Journal of Chemical Theory and Computation* **11**:1864–1874. DOI: <https://doi.org/10.1021/ct5010406>, PMID: 26574392
- Huang J, Rauscher S, Nawrocki G, Ran T, Feig M, de Groot BL, Grubmüller H, MacKerell AD. 2017. CHARMM36m: an improved force field for folded and intrinsically disordered proteins. *Nature Methods* **14**:71–73. DOI: <https://doi.org/10.1038/nmeth.4067>, PMID: 27819658
- Bannon D, Portnoy ME, Olivi L, Lees PSJ, Culotta VC, Bressler JP. 2002. Uptake of lead and iron by divalent metal transporter 1 in yeast and mammalian cells. *Biochemical and Biophysical Research Communications* **295**:978–984. DOI: [https://doi.org/10.1016/s0006-291x\(02\)00756-8](https://doi.org/10.1016/s0006-291x(02)00756-8), PMID: 12127992
- Illing AC, Shawki A, Cunningham CL, Mackenzie B. 2012. Substrate profile and metal-ion selectivity of human divalent metal-ion transporter-1. *The Journal of Biological Chemistry* **287**:30485–30496. DOI: <https://doi.org/10.1074/jbc.M112.364208>, PMID: 22736759
- Irving H, Williams RJP. 1953. 637. the stability of transition-metal complexes. *Journal of the Chemical Society* **3192**. DOI: <https://doi.org/10.1039/jr9530003192>
- Jo S, Kim T, Iyer VG, Im W. 2008. CHARMM-GUI: a web-based graphical user interface for CHARMM. *Journal of Computational Chemistry* **29**:1859–1865. DOI: <https://doi.org/10.1002/jcc.20945>, PMID: 18351591
- Jung M, Mertens C, Brüne B. 2015. Macrophage iron homeostasis and polarization in the context of cancer. *Immunobiology* **220**:295–304. DOI: <https://doi.org/10.1016/j.imbio.2014.09.011>, PMID: 25260218
- Jurrus E, Engel D, Star K, Monson K, Brandi J, Felberg LE, Brookes DH, Wilson L, Chen J, Liles K, Chun M, Li P, Gohara DW, Dolinsky T, Konecny R, Koes DR, Nielsen JE, Head-Gordon T, Geng W, Krasny R, et al. 2018. Improvements to the APBS biomolecular solvation software suite. *Protein Science* **27**:112–128. DOI: <https://doi.org/10.1002/pro.3280>, PMID: 28836357
- Kabsch W. 2010. XDS. *Acta Crystallographica. Section D, Biological Crystallography* **66**:125–132. DOI: <https://doi.org/10.1107/S09074449090047337>, PMID: 20124692
- Karplus PA, Diederichs K. 2012. Linking crystallographic model and data quality. *Science* **336**:1030–1033. DOI: <https://doi.org/10.1126/science.1218231>, PMID: 22628654
- Karplus PA, Diederichs K. 2015. Assessing and maximizing data quality in macromolecular crystallography. *Current Opinion in Structural Biology* **34**:60–68. DOI: <https://doi.org/10.1016/j.sbi.2015.07.003>, PMID: 26209821
- Kawabata H. 2019. Transferrin and transferrin receptors update. *Free Radical Biology & Medicine* **133**:46–54. DOI: <https://doi.org/10.1016/j.freeradbiomed.2018.06.037>, PMID: 29969719
- Kozlov AM, Darriba D, Flouri T, Morel B, Stamatakis A, Wren J. 2019. RAXML-NG: a fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. *Bioinformatics* **35**:4453–4455. DOI: <https://doi.org/10.1093/bioinformatics/btz305>, PMID: 31070718
- Krishnamurthy H, Piscitelli CL, Gouaux E. 2009. Unlocking the molecular secrets of sodium-coupled transporters. *Nature* **459**:347–355. DOI: <https://doi.org/10.1038/nature08143>, PMID: 19458710
- Krishnamurthy H, Gouaux E. 2012. X-ray structures of leut in substrate-free outward-open and apo inward-open states. *Nature* **481**:469–474. DOI: <https://doi.org/10.1038/nature10737>, PMID: 22230955
- Kumarevel T, Mizuno H, Kumar PKR. 2005. Characterization of the metal ion binding site in the anti-terminator protein, hupP, of *Bacillus subtilis*. *Nucleic Acids Research* **33**:5494–5502. DOI: <https://doi.org/10.1093/nar/gki868>, PMID: 16192572
- Le SQ, Gascuel O. 2008. An improved general amino acid replacement matrix. *Molecular Biology and Evolution* **25**:1307–1320. DOI: <https://doi.org/10.1093/molbev/msn067>, PMID: 18367465
- Leavitt S, Freire E. 2001. Direct measurement of protein binding energetics by isothermal titration calorimetry. *Current Opinion in Structural Biology* **11**:560–566. DOI: [https://doi.org/10.1016/s0959-440x\(00\)00248-7](https://doi.org/10.1016/s0959-440x(00)00248-7), PMID: 11785756
- Lee J, Cheng X, Swails JM, Yeom MS, Eastman PK, Lemkul JA, Wei S, Buckner J, Jeong JC, Qi Y, Jo S, Pande VS, Case DA, Brooks CL, MacKerell AD, Klauda JB, Im W. 2016. CHARMM-gui input generator for namd, gromacs, amber, openmm, and CHARMM/openmm simulations using the charmm36 additive force field. *Journal of Chemical Theory and Computation* **12**:405–413. DOI: <https://doi.org/10.1021/acs.jctc.5b00935>, PMID: 26631602
- Liebschner D, Afonine PV, Baker ML, Bunkóczi G, Chen VB, Croll TI, Hintze B, Hung LW, Jain S, McCoy AJ, Moriarty NW, Oeffner RD, Poon BK, Prisant MG, Read RJ, Richardson JS, Richardson DC, Sammito MD, Sobolev OV, Stockwell DH, et al. 2019. Macromolecular structure determination using x-rays, neutrons and electrons: recent developments in phenix. *Acta Crystallographica. Section D, Structural Biology* **75**:861–877. DOI: <https://doi.org/10.1107/S2059798319011471>, PMID: 31588918
- Lin Z, Fernández-Robledo JA, Cellier MF, Vasta GR. 2009. Metals and membrane metal transporters in biological systems: the role (S) of Nramp in host-parasite interactions. *J Argent Chem Soc* **97**:210–225.

- Liu Q, Barker S, Knutson MD. 2021. Iron and manganese transport in mammalian systems. *Biochimica et Biophysica Acta. Molecular Cell Research* **1868**:118890. DOI: <https://doi.org/10.1016/j.bbamcr.2020.118890>, PMID: 33091506
- Loo DD, Hirayama BA, Gallardo EM, Lam JT, Turk E, Wright EM. 1998. Conformational changes couple Na⁺ and glucose transport. *PNAS* **95**:7789–7794. DOI: <https://doi.org/10.1073/pnas.95.13.7789>, PMID: 9636229
- Ma Z, Jacobsen FE, Giedroc DP. 2009. Metal transporters and metal sensors: how coordination chemistry controls bacterial metal homeostasis. *Chemical Reviews* **109**:4644. DOI: <https://doi.org/10.1021/cr900077w>
- Manatschal C, Pujol-Giménez J, Poirier M, Reymond J-L, Hediger MA, Dutzler R. 2019. Mechanistic basis of the inhibition of SLC11/NRAMP-mediated metal ion transport by bis-isothiourea substituted compounds. *eLife* **8**:e51913. DOI: <https://doi.org/10.7554/eLife.51913>, PMID: 31804182
- McGibbon RT, Beauchamp KA, Harrigan MP, Klein C, Swails JM, Hernández CX, Schwantes CR, Wang L-P, Lane TJ, Pande VS. 2015. MDTraj: A modern open library for the analysis of molecular dynamics trajectories. *Biophysical Journal* **109**:1528–1532. DOI: <https://doi.org/10.1016/j.bpj.2015.08.015>, PMID: 26488642
- Morin A, Eisenbraun B, Key J, Sanschagrin PC, Timony MA, Ottaviano M, Sliz P. 2013. Collaboration gets the most out of software. *eLife* **2**:e01456. DOI: <https://doi.org/10.7554/eLife.01456>, PMID: 24040512
- Nevo Y, Nelson N. 2006. The NRAMP family of metal-ion transporters. *Biochimica et Biophysica Acta* **1763**:609–620. DOI: <https://doi.org/10.1016/j.bbamcr.2006.05.007>, PMID: 16908340
- Nies DH, Grass G. 2009. Transition metal homeostasis. *EcoSal Plus* **3**. DOI: <https://doi.org/10.1128/ecosalplus.5.4.4.3>, PMID: 26443772
- Olsson TSG, Williams MA, Pitt WR, Ladbury JE. 2008. The thermodynamics of protein-ligand interaction and solvation: insights for ligand design. *Journal of Molecular Biology* **384**:1002–1017. DOI: <https://doi.org/10.1016/j.jmb.2008.09.073>, PMID: 18930735
- Olsson MHM, Søndergaard CR, Rostkowski M, Jensen JH. 2011. PROPKA3: consistent treatment of internal and surface residues in empirical pKa predictions. *Journal of Chemical Theory and Computation* **7**:525–537. DOI: <https://doi.org/10.1021/ct100578z>, PMID: 26596171
- Perez C, Koshy C, Yildiz O, Ziegler C. 2012. Alternating-Access mechanism in conformationally asymmetric trimers of the betaine transporter BetP. *Nature* **490**:126–130. DOI: <https://doi.org/10.1038/nature11403>, PMID: 22940865
- Phillips JC, Stone JE, Schulten K. 2008. Adapting a message-driven parallel application to GPU-accelerated clusters. SC - International Conference for High Performance Computing, Networking, Storage and Analysis. . DOI: <https://doi.org/10.1109/SC.2008.5214716>
- Piccolo A, Malvezzi M, Houtman JCD, Accardi A. 2009. Basis of substrate binding and conservation of selectivity in the CLC family of channels and transporters. *Nature Structural & Molecular Biology* **16**:1294–1301. DOI: <https://doi.org/10.1038/nsmb.1704>, PMID: 19898476
- Pittman JK. 2005. Managing the manganese: molecular mechanisms of manganese transport and homeostasis. *The New Phytologist* **167**:733–742. DOI: <https://doi.org/10.1111/j.1469-8137.2005.01453.x>, PMID: 16101910
- Potter SC, Luciani A, Eddy SR, Park Y, Lopez R, Finn RD. 2018. HMMER web server: 2018 update. *Nucleic Acids Research* **46**:W200–W204. DOI: <https://doi.org/10.1093/nar/gky448>, PMID: 29905871
- Pujol-Giménez J, Hediger MA, Gyimesi G. 2017. A novel proton transfer mechanism in the SLC11 family of divalent metal ion transporters. *Scientific Reports* **7**:6194. DOI: <https://doi.org/10.1038/s41598-017-06446-y>, PMID: 28754960
- Ramanadane K, Straub MS, Dutzler R, Manatschal C. 2022. Structural and functional properties of a magnesium transporter of the SLC11/NRAMP family. *eLife* **11**:e74589. DOI: <https://doi.org/10.7554/eLife.74589>, PMID: 35001872
- Rulísek L, Vondrášek J. 1998. Coordination geometries of selected transition metal ions (Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, and Hg²⁺) in metalloproteins. *Journal of Inorganic Biochemistry* **71**:115–127. DOI: [https://doi.org/10.1016/s0162-0134\(98\)10042-9](https://doi.org/10.1016/s0162-0134(98)10042-9), PMID: 9833317
- Sacher A, Cohen A, Nelson N. 2001. Properties of the mammalian and yeast metal-ion transporters DCT1 and Smf1p expressed in *Xenopus laevis* oocytes. *The Journal of Experimental Biology* **204**:1053–1061. DOI: <https://doi.org/10.1242/jeb.204.6.1053>, PMID: 11222124
- Shimamura T, Weyand S, Beckstein O, Rutherford NG, Hadden JM, Sharples D, Sansom MSP, Iwata S, Henderson PJF, Cameron AD. 2010. Molecular basis of alternating access membrane transport by the sodium-hydantoin transporter mhp1. *Science* **328**:470–473. DOI: <https://doi.org/10.1126/science.1186303>, PMID: 20413494
- Simmons KJ, Jackson SM, Brueckner F, Patching SG, Beckstein O, Ivanova E, Geng T, Weyand S, Drew D, Lanigan J, Sharples DJ, Sansom MSP, Iwata S, Fishwick CWG, Johnson AP, Cameron AD, Henderson PJF. 2014. Molecular mechanism of ligand recognition by membrane transport protein, mhp1. *The EMBO Journal* **33**:1831–1844. DOI: <https://doi.org/10.1525/embj.201387557>, PMID: 24952894
- Singh J, Nanda Srivastva A, Singh N, Singh A. 2020. Stability and applications of coordination compounds. *Stability and Applications of Coordination* InTechOpen. DOI: <https://doi.org/10.5772/intechopen.83186>
- Skamene E, Schurr E, Gros P. 1998. Infection genomics: nramp1 as a major determinant of natural resistance to intracellular infections. *Annual Review of Medicine* **49**:275–287. DOI: <https://doi.org/10.1146/annurev.med.49.1.275>, PMID: 9509263
- Søndergaard CR, Olsson MHM, Rostkowski M, Jensen JH. 2011. Improved treatment of ligands and coupling effects in empirical calculation and rationalization of pKa values. *Journal of Chemical Theory and Computation* **7**:2284–2295. DOI: <https://doi.org/10.1021/ct200133y>, PMID: 26606496

- Tareen A**, Kinney JB. 2020. Logomaker: beautiful sequence logos in python. *Bioinformatics* **36**:2272–2274. DOI: <https://doi.org/10.1093/bioinformatics/btz921>, PMID: 31821414
- Tellinghuisen J**. 2008. Isothermal titration calorimetry at very low c. *Analytical Biochemistry* **373**:395–397. DOI: <https://doi.org/10.1016/j.ab.2007.08.039>, PMID: 17920027
- Turnbull WB**, Daranas AH. 2003. On the value of C: can low affinity systems be studied by isothermal titration calorimetry? *Journal of the American Chemical Society* **125**:14859–14866. DOI: <https://doi.org/10.1021/ja036166s>, PMID: 14640663
- Vashishtha AK**, Wang J, Konigsberg WH. 2016. Different divalent cations alter the kinetics and fidelity of DNA polymerases. *The Journal of Biological Chemistry* **291**:20869–20875. DOI: <https://doi.org/10.1074/jbc.R116.742494>, PMID: 27462081
- Vidal SM**, Malo D, Vogan K, Skamene E, Gros P. 1993. Natural resistance to infection with intracellular parasites: isolation of a candidate for bcg. *Cell* **73**:469–485. DOI: [https://doi.org/10.1016/0092-8674\(93\)90135-d](https://doi.org/10.1016/0092-8674(93)90135-d), PMID: 8490962
- Watanabe A**, Choe S, Chaptal V, Rosenberg JM, Wright EM, Grabe M, Abramson J. 2010. The mechanism of sodium and substrate release from the binding pocket of vsgrt. *Nature* **468**:988–991. DOI: <https://doi.org/10.1038/nature09580>, PMID: 21131949
- Winn MD**, Ballard CC, Cowtan KD, Dodson EJ, Emsley P, Evans PR, Keegan RM, Krissinel EB, Leslie AGW, McCoy A, McNicholas SJ, Murshudov GN, Pannu NS, Potterton EA, Powell HR, Read RJ, Vagin A, Wilson KS. 2011. Overview of the CCP4 suite and current developments. *Acta Crystallographica. Section D, Biological Crystallography* **67**:235–242. DOI: <https://doi.org/10.1107/S0907444910045749>, PMID: 21460441
- Wiseman T**, Williston S, Brandts JF, Lin LN. 1989. Rapid measurement of binding constants and heats of binding using a new titration calorimeter. *Analytical Biochemistry* **179**:131–137. DOI: [https://doi.org/10.1016/0003-2697\(89\)90213-3](https://doi.org/10.1016/0003-2697(89)90213-3), PMID: 2757186
- Wu EL**, Cheng X, Jo S, Rui H, Song KC, Dávila-Contreras EM, Qi Y, Lee J, Monje-Galvan V, Venable RM, Klauda JB, Im W. 2014. CHARMM-GUI membrane builder toward realistic biological membrane simulations. *Journal of Computational Chemistry* **35**:1997–2004. DOI: <https://doi.org/10.1002/jcc.23702>
- Yokoyama H**, Tsuruta O, Akao N, Fujii S. 2012. Crystal structure of *Helicobacter pylori* neutrophil-activating protein with a di-nuclear ferroxidase center in a zinc or cadmium-bound form. *Biochemical and Biophysical Research Communications* **422**:745–750. DOI: <https://doi.org/10.1016/j.bbrc.2012.05.073>, PMID: 22618234

Appendix 1

ITC analysis

In this appendix, we describe our choices of models to fit our ITC isotherms. Based on the isotherm shapes and preliminary fits, the c values (association constant $\times$ molar protein concentration) for all isotherms are below 1. Curve-fitting is most robust for $5 < c < 500$, a range which generally allows to fit binding site stoichiometry (n) in addition to the dissociation constant, K_d , (or association constant, K_a) and the enthalpy of association (ΔH) (Turnbull and Daranas, 2003). The combination of low metal ion binding affinity and the achievable protein amounts and concentration for DraNrp makes it impractical to achieve a higher c value. However, previous studies have demonstrated that data with $c < 1$ can still provide useful information on K_d values (Piccolo et al., 2009; Tellinghuisen, 2008; Turnbull and Daranas, 2003). Hence, as recommended in those studies, we fit the isotherms with either a one-site model with a fixed $n=1$, or a two-site sequential binding model (fixed $n=2$; Appendix 1—tables 1 and 2 and Appendix 1—figure 1). The assumptions inherent to fixing n are that we have accurately determined the concentrations of metal ion ligand and protein, but fixing n has little impact on the resulting K_d value (Tellinghuisen, 2008). We then used the quality of fit and information from our crystal structures, mutational data to select the most appropriate fit to interpret each ITC isotherm dataset. Below we justify our choice of model for pairs of protein construct and metal ion ligand:

Appendix 1—table 1. ITC analysis of all Mn²⁺ binding isotherms, with chosen model and values shaded.

Protein construct	Two-site sequential binding model (fixed n=2)								One-site fit with fixed n=1			
	K _{d1} (μM)	ΔH ₁ (kcal/mol)	-TΔS ₁ (kcal/mol)	ΔG ₁ (kcal/mol)	K _{d2} (μM)	ΔH ₂ (kcal/mol)	-TΔS ₂ (kcal/mol)	ΔG ₂ (kcal/mol)	K _d (μM)	ΔH (kcal/mol)	-TΔS (kcal/mol)	ΔG (kcal/mol)
WT	220	3.2	-8.1	-5	961	5.4	-9.2	-3.8	650	8.8	-13.1	-4.3
	125	5.7	-11.1	-5.4	2700	17.1	-20.5	-4.6	380	12.6	-16.9	-4.3
	220	8	-12.8	-5	2250	13.7	-17.3	-3.4	500	15.4	-19.6	-4.2
	190±30	5.6±1.3	-10.6±1.4	-5.1±0.1	1970±520	12.1±3.3	-15.7±3.1	-4.0±0.3	510±80	12.3±1.9	-16.5±1.8	-4.2±0.03
A47W	130	6.3	-11.3	-5	3100	20.6	-23.8	-3.2	380	13.6	-18.1	-4.5
	120	6.2	-11.3	-5.1	1800	11.4	-14.9	-3.5	300	11.9	-16.6	-4.7
	125±5	6.2±0.5	-11.3±0.0	-5.0±0.1	2450±650	16±4.5	-19.3±4.4	-3.3±0.1	340±40	12.7±0.8	-17.3±0.7	-4.6±0.1
D56A	305	2.4	-7.1	-4.7	4800	20.4	-23.5	-3.1	2200	16.3	-19.6	-3.3
	140	1.8	-6.8	-5	2700	22.6	-25.9	-3.3	2500	22.6	-26.2	-3.6
	230±80	2.1±0.3	-6.9±0.1	-4.8±0.1	3800±1,100	21.5±1.1	-24.7±0.1	-3.2±0.1	2350±150	19.4±3	-22.9±3.3	-3.4±0.1
M230A	280	3	-7.7	-4.7	3300	14.9	-18.1	-3.2	1500	13.3	-16.9	-3.6
	150	2.9	-8	-5.1	5800	29.9	-33.1	-3.2	970	12.5	-16.3	-3.8
	215±65	2.9±0.05	-7.8±0.1	-4.9±0.2	4600±1,300	22.4±7.5	-25.6±7.5	-3.2±0	1240±270	12.9±0.4	-16.6±0.3	-3.7±0.1
G223W	Did not fit								460	5.8	-10.1	-4.3
									430	3	-7.4	-4.4
									440±15	4.4±1.4	-8.7±1.3	-4.3±0.1
									430	8.9	-13.4	-4.5
D296A	Did not fit								320	9.5	-14.3	-4.8
									370	10.1	-14.6	-4.5
									370±30	9.5±0.3	-14.1±0.3	-4.6±0.1
									390	9.5	-14	-4.5
D369A	Did not fit								480	8.9	-13.4	-4.5
									380	8.3	-12.8	-4.5
									420±30	8.9±0.3	-13.4±0.3	-4.5±0

Appendix 1—table 1 Continued on next page

Appendix 1—table 1 Continued

Protein construct	Two-site sequential binding model (fixed n=2)								One-site fit with fixed n=1			
	K _{d1} (μM)	ΔH ₁ (kcal/mol)	-TΔS ₁ (kcal/mol)	ΔG ₁ (kcal/mol)	K _{d2} (μM)	ΔH ₂ (kcal/mol)	-TΔS ₂ (kcal/mol)	ΔG ₂ (kcal/mol)	K _d (μM)	ΔH (kcal/mol)	-TΔS (kcal/mol)	ΔG (kcal/mol)
A47W-D296A	Did not fit								300	6.1	-10.7	-4.6
									210	13.2	-17.5	-4.3
									255±45	9.6±3.5	-14.1±3.4	-4.4±0.1
A47W-D369A	Did not fit								210	7.5	-12.5	-5
									400	6.5	-11.1	-4.6
									300±95	7±0.5	-11.8±0.7	-4.8±0.1
D56A-D296A	Did not fit								170	5.3	-10.4	-5.1
									330	4.8	-9.5	-4.7
									250±80	5.0±0.2	-9.9±0.4	-4.9±0.2
D56A-D369A	Did not fit								280	4.9	-9.5	-4.6
									145	4	-8.9	-4.9
									210±70	4.4±0.4	-9.2±0.3	-4.7±0.1
M230A-D296A	Did not fit								320	7.1	-11.6	-4.5
									140	3.7	-8.9	-5.2
									230±90	5.4±1.7	-10.2±1.3	-4.8±0.3
M230A-D369A	Did not fit								750	11.5	-15.4	-3.9
									790	7.9	-11.9	-4
									770±20	9.7±1.8	-13.6±1.7	-3.9±0.05

The online version of this article includes the following source data for appendix 1—table 1:

Appendix 1—table 1—source data 1. Source files (Origin files) of ITC experiments of manganese binding to each DraNrmamp construct.

Appendix 1—table 2. ITC analysis of all Cd²⁺-binding isotherms, with chosen model and values shaded.

Protein construct	Two-site sequential binding model (fixed n=2)								One-site fit with fixed n=1			
	K _{d1} (μM)	ΔH ₁ (kcal/mol)	-TΔS ₁ (kcal mol ⁻¹)	K _{d1} (μM)	ΔH ₁ (kcal/mol)	ΔH ₂ (kcal mol ⁻¹)	K _{d1} (μM)	ΔH ₁ (kcal/mol)	K _d (μM)	K _{d1} (μM)	ΔH ₁ (kcal/mol)	ΔG (kcal mol ⁻¹)
WT	85	-4.5	-0.9	-5.5	260	-2.6	-2	-4.6	105	-2.3	-2.9	-5.2
	50	-4.7	-0.9	-5.6	220	-4.1	-0.6	-4.7	105	-9	3.5	-5.5
	30	-2.9	-2.9	-5.8	180	-4.7	-0.3	-5	115	-7.6	2.1	-5.5
	55±15	-4.0±0.5	-1.6±0.6	-5.6±0.1	220±20	-3.8±0.6	-1.0±0.5	-4.8±0.1	110±3	6±2	-2.8±1.9	-5.4±0.1
	220	-4.4	-0.2	-4.6	130	0.6	-5.6	-5	165	-4.8	-0.2	-5
A47W	Did not fit								140	-3.3	-1.7	-5
									150±10	-4.0±0.7	-0.9±0.5	-5.0±0.0
	100	-6.1	0.6	-5.5	3000	-11.2	7.7	-3.5	190	-9.2	4.1	-5.1
D56A	55	-4.5	-1.2	-5.7	1500	-6.2	2.3	-3.9	130	-6.9	1.5	-5.4
	80±20	-5.3±0.8	-0.6±0.9	-5.6±0.1	2250±750	-8.7±2.5	5.0±0.5	-3.7±0.1	160±30	-8.0±1	-2.8±1.3	-5.2±0.1
									142	-6.1	0.9	-5.2
M230A	Did not fit								182	-3.1	-1.7	-4.8
									160±20	-4.6±1.5	-0.4±0.3	-5.0±0.2
G223W	No Binding								122	-3.5	-1.8	-6.8
									120	-3.7	-1.5	-5.2
									120±1	-3.6±0.1	-1.6±0.1	-6.0±0.8
D296A	Did not fit											

Appendix 1—table 2 Continued on next page

Appendix 1—table 2 Continued

Protein construct	Two-site sequential binding model (fixed n=2)								One-site fit with fixed n=1			
	K _{d1} (μM)	ΔH ₁ (kcal/mol)	-TΔS ₁ (kcal mol ⁻¹)	K _{d1} (μM)	ΔH ₁ (kcal/mol)	ΔH ₂ (kcal mol ⁻¹)	K _{d1} (μM)	ΔH ₁ (kcal/mol)	K _d (μM)	K _{d1} (μM)	ΔH ₁ (kcal/mol)	ΔG (kcal mol ⁻¹)
D369A	100	-1.6	-3.5	-5.1	5350	2.8*	-5.9	-3.1	80	-1.4	-4.1	-5.5
	90	-2.5	-2.6	-5.7	4850	5*	-8	-3	60	-2.1	-3.8	-5.9
	95±5	-5.3±0.8	-3.1±0.4	-5.4±0.1	5100±250	3.9±1.0*	-6.9±1.0	-3.0±0.1	70±10	-1.7±0.3	-3.9±0.1	-5.7±0.1
A47W-D296A	No Binding											
A47W-D369A	No Binding											
D56A-D296A	No Binding											
D56A-D369A	No Binding											
M230A-D296A	No Binding											
M230A-D369A	No Binding											

*Positive values of ΔH corresponding to an exothermic mode of Cd²⁺ binding to protein indicates inappropriate fit

The online version of this article includes the following source data for appendix 1—table 2:

Appendix 1—table 2—source data 1. Source files (Origin files) of ITC experiments of cadmium binding to each DraNrap construct.

WT binding to Mn²⁺

The WT•Mn²⁺ crystal structure revealed two Mn²⁺ ions bound at distinct sites, and the overall fit of the ITC data is better for the two-site sequential binding model than the one-site model (**Appendix 1—figure 1A**), with the two K_d values differing by ~20 fold. Based on ITC of the constructs with mutations of the external site (D296A and D236A; see below), we assigned the higher affinity K_{d1} to the orthosteric site (190±30 μM) and the lower affinity K_{d2} to the external site (1970±520 μM).

A47W binding to Mn²⁺

The A47W•Mn²⁺ crystal structure revealed two Mn²⁺ ions bound to the same sites as with WT•Mn²⁺, and the overall fit of the ITC data is better for the two-site sequential binding model than for the one-site model (**Appendix 1—figure 1B**). The two K_d values and the ITC data for the constructs with external-site mutations (A47W-D296A and A47W-D236A, both of which can only be fitted with a one-site model) are consistent with the K_d assignments for WT, with the orthosteric site having higher affinity for Mn²⁺ (K_{d1}=125±5 μM) than the external site (K_{d2}=2450±650 μM).

M230A binding to Mn²⁺

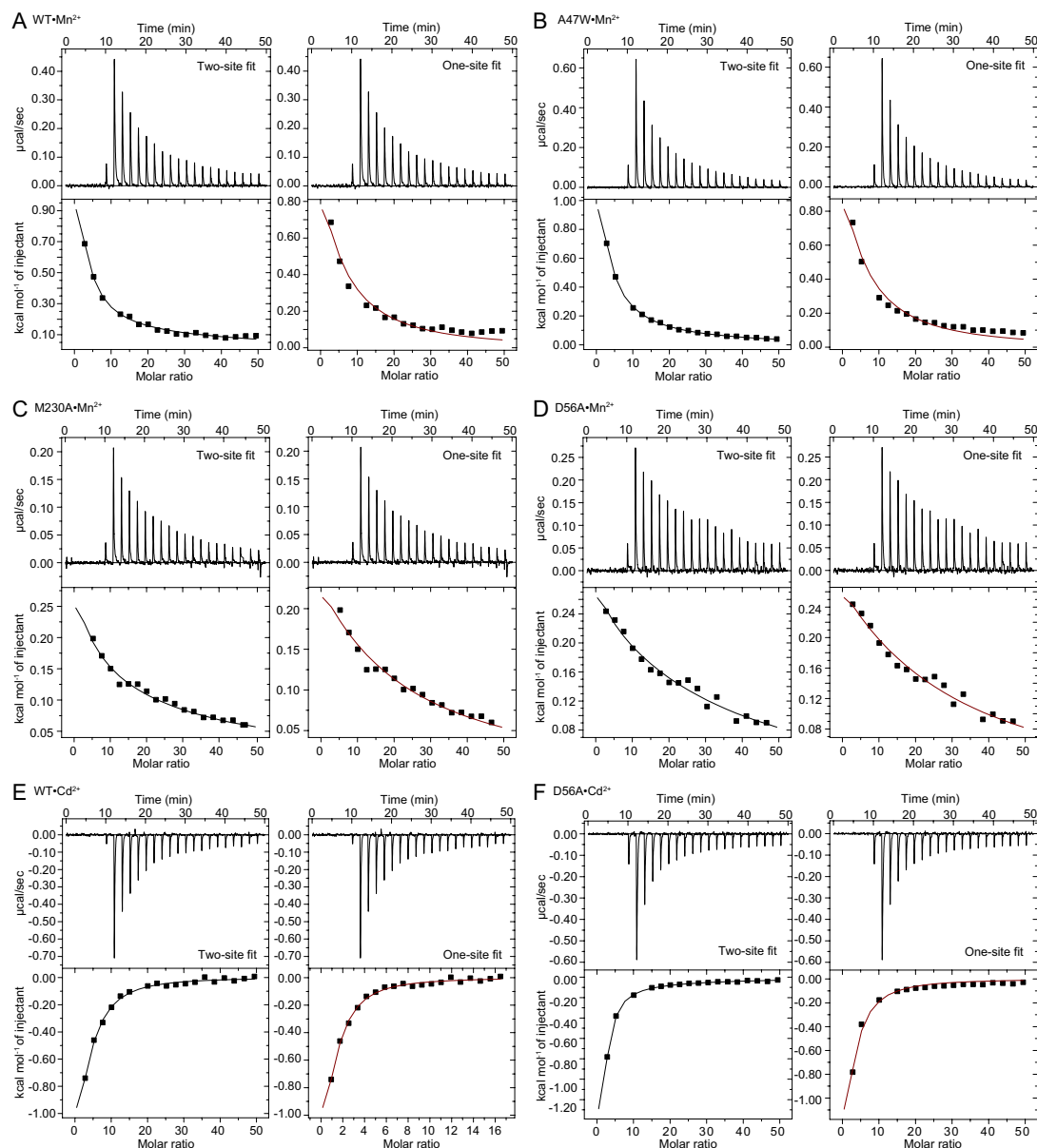
The M230A•Mn²⁺ crystal structure again revealed two bound Mn²⁺, and the overall fit of the ITC data is again better for the two-site sequential binding model than the one-site model (**Appendix 1—figure 1C**). The two K_d values and the ITC data for the constructs with external-site mutations (M230A-D296A and M230A-D236A, both of which only fit with a one-site model) are consistent with the K_d assignments for WT, with the orthosteric site having higher affinity for Mn²⁺ (K_{d1}=215±65 μM) than the external site (K_{d2}=4600±1300 μM).

D56A binding to Mn²⁺

The ITC data can be fitted using either the two-site sequential model or the one-site model (**Appendix 1—figure 1D**). In contrast, the data for constructs with external-site mutations (D56A-D296A and D56A-D369A) only fit well using a one-site model. Hence, we selected the two-site sequential binding model to fit D56A binding to Mn²⁺, as it agrees best with the overall analysis, with the orthosteric site having higher affinity for Mn²⁺ (K_{d1}=230±80 μM) than the external site (K_{d2}=3800±1100 μM).

WT binding to Cd²⁺

The WT•Cd²⁺ crystal structure reveals two Cd²⁺ ions bound at the same two distinct sites where Mn²⁺ bound. Based on this observation we chose a two-site model rather than a one-site model to fit the ITC data, although the fits look similar (**Appendix 1—figure 1E**). In contrast to Mn²⁺ binding,



Appendix 1—figure 1. Comparison of one-site and two-site fit to ITC binding isotherms for each case where both types of fits gave results. We show fits to a two-site sequential binding model (fixed $n=2$; left) or a one-site model with a fixed $n=1$ (right) for each case where both types of fits gave a result (**Appendix 1—tables 1 and 2**). For constructs•ligand pairs WT•Mn²⁺ (A), A47W•Mn²⁺ (B), M230A•Mn²⁺ (C), and D56A•Cd²⁺ (F), the two-site fit is better than the one-site fit, which corroborates findings from the crystal structures. For pairs WT•Cd²⁺ (E) and D56A•Mn²⁺ (D), the fits are similar, and we used additional biochemical and structural information to select the most appropriate fit.

the two K_d values obtained using a two-site sequential binding model are more similar to each other (~4 fold difference). Based on ITC of the constructs with external-site mutations (D296A and D236A; see below), we assigned the higher affinity K_d ($K_{d1}=55\pm15\ \mu\text{M}$) to the orthosteric site, and the lower affinity one to the external site ($K_{d2}=220\pm20\ \mu\text{M}$).

A47W binding to Cd²⁺

One-site model-based fits of the ITC data consistently yield better thermodynamic binding parameters; the two-site sequential binding model either does not fit or only fits with a positive ΔH which is inappropriate considering the observed exothermic binding. Furthermore, ITC

measurements on the corresponding constructs with external-site mutations (A47W-D296A and A47W-D369A) showed no binding. Therefore, we chose the one-site model to fit the ITC data of A47W binding to Cd^{2+} and we assigned the resulting K_d value ($150 \pm 10 \mu\text{M}$) to the external site. We conclude that Cd^{2+} does not bind to the A47W orthosteric site with measurable affinity.

M230A binding to Cd^{2+}

The ITC data could only be fitted with the one-site model. The ITC measurements on the corresponding constructs with external-site mutations (M230A-D296A and M230A-D369A) showed no binding. We thus assigned the K_d value obtained using the one-site model ($160 \pm 20 \mu\text{M}$) to the external site, and we conclude that Cd^{2+} does not bind to the M230A orthosteric site with measurable affinity.

D56A binding to Cd^{2+}

The ITC data fit better when using a two-site sequential binding model than a one-site model (**Appendix 1—figure 1F**). The $\text{WT} \cdot \text{Cd}^{2+}$ structure shows no direct interaction of D56 with Cd^{2+} , suggesting that the D56A mutation may not completely impair Cd^{2+} binding at the orthosteric site. We thus chose to use the two-site sequential binding model and assign higher affinity ($K_{d1} = 80 \pm 20 \mu\text{M}$) to the external site, and lower affinity ($K_{d2} = 2250 \pm 750 \mu\text{M}$) to the orthosteric site.

G223W binding to Mn^{2+}

Consistent with the $\text{G223W} \cdot \text{Mn}^{2+}$ crystal structure in which Mn^{2+} is only bound at the orthosteric site, the ITC data could only be fitted with the one-site model. We assigned the resulting K_d value ($440 \pm 15 \mu\text{M}$) to the orthosteric site.

G223W binding to Cd^{2+}

The ITC data show no binding, which is consistent with our inability to obtain a crystal structure of G223W bound to Cd^{2+} . This indicates that the outward-locked G223W does not bind Cd^{2+} with measurable affinity at the orthosteric site.

D296A binding to Mn^{2+}

Consistent with the $\text{D296A} \cdot \text{Mn}^{2+}$ crystal structure in which Mn^{2+} is bound at the orthosteric site but not at the external site, the ITC data could only be fitted with the one-site model. We assigned the resulting K_d value ($370 \pm 30 \mu\text{M}$) to the orthosteric site.

D296A binding to Cd^{2+}

The ITC data could only be fitted with the one-site model. We assigned the resulting K_d value ($120 \pm 10 \mu\text{M}$) to the orthosteric site, because the D296A mutation removes an aspartate ligand at the external site.

D369A binding to Mn^{2+}

The ITC data could only be fitted with the one-site model. We assigned the resulting K_d value ($420 \pm 30 \mu\text{M}$) to the orthosteric site, because the D369A mutation removes an aspartate ligand at the external site.

D369A binding to Cd^{2+}

Fits of the ITC data using the one-site model consistently yielded better thermodynamic binding parameters; the two-site sequential binding model either resulted in no fit or in a fit with positive ΔH which is inappropriate considering the observed exothermic binding. We thus chose to fit the data with the one-site model, and we assigned the resulting K_d value ($70 \pm 10 \mu\text{M}$) to the orthosteric site.