

1    **Title:** Experimental and genetic analysis of selfing reveals no reinforcement in *Phlox*

2    *cuspidata*

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12    **Running header:** Selfing in *Phlox cuspidata*

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14    **Keywords:** reinforcement, self-fertilization, speciation, sympatry, reproductive isolation,

15    mating system, selfing

16 **Summary**

17 • Reinforcement is the process through which prezygotic reproductive barriers evolve in  
18 sympatry due to selection against hybridization between co-occurring, closely related  
19 species. The role of self-fertilization in reinforcement and reproductive isolation is  
20 uncertain in part because its efficiency as a barrier against heterospecific mating can  
21 depend on the timing of autonomous selfing.

22 • To investigate whether increased autonomous selfing has evolved as a mechanism for  
23 reinforcement, we compared *Phlox cuspidata* populations across their native Texas range  
24 using both estimates of genetic diversity and experimental manipulation with  
25 morphological measurements. Specifically, we investigated patterns of variation in floral  
26 traits and timing of selfing between individuals from allopatric populations of *P.*  
27 *cuspidata* and from populations sympatric with the closely related species, *P.*  
28 *drummondii*.

29 • We infer intermediate rates of selfing across field-collected individuals with no  
30 significant difference between allopatric and sympatric populations. Among greenhouse  
31 grown plants, we find no differences in timing of selfing or other floral traits including  
32 anther dehiscence timing, anther-stigma distances, autonomous selfing rate and self-seed  
33 count between allopatric and sympatric populations. However, our statistical analyses  
34 indicate that *P. cuspidata* individuals sympatric with *P. drummondii* seem to have  
35 generally larger flowers compared to allopatric individuals.

36 • Despite strong evidence of costly hybridization with *P. drummondii*, we find no evidence  
37 of trait divergence due to reinforcement in *P. cuspidata*. Although we document nearly  
38 complete autonomous self-seed set in the greenhouse, estimates of selfing rates from  
39 genetic data imply realized selfing is much lower in nature suggesting an opportunity for  
40 reinforcing selection to act on this trait.

41 **Introduction**

42 Determining how reproductive isolation (RI) evolves and is maintained between closely  
43 related species is key to understanding the speciation process (Coyne & Orr, 2004). RI is  
44 caused by the formation of multiple pre and post zygotic mechanisms. While these  
45 mechanisms vary among taxa, studies observing RI through time often note that prezygotic  
46 RI evolves with greater strength and faster in sympatry than in allopatry (Coyne & Orr, 1989;  
47 Noor, 1995). This pattern supports reinforcement as a broadly important process in species  
48 formation (Coyne & Orr, 1989; Tabah & Hiscock, 2003; Yukilevich, 2011). Reinforcement is  
49 the process through which prezygotic barriers to reproduction evolve in sympatry due to  
50 selection against costly hybridization between co-occurring, closely related species  
51 (Dobzhansky, 1940). Reinforcement often generates a pattern of reproductive character  
52 displacement (RCD), in which adaptive traits such as differences in reproductive timing or  
53 flower morphology evolve in sympatric populations but not allopatric populations (Levin,  
54 1985; Knowlton, 1993). This trait divergence minimizes deleterious reproductive interactions  
55 between species that have come into secondary contact and increases RI in the process  
56 (Levin, 1985; Lowry et al., 2008; Hopkins, 2013; Pfennig & Rice, 2014; Baack et al., 2015).  
57 Most research on prezygotic isolation in plants has focused on how patterns of pollinator  
58 visitation and associated floral trait evolution can drive RI (Schemske & Bradshaw, 1999;  
59 Cardona et al., 2020). However, mating system shifts and associated floral differences can  
60 also drive prezygotic barriers between sympatric species, yet these processes have not been as  
61 extensively studied (Fishman & Wyatt, 1999; Wendt et al., 2002; Grossenbacher & Whittall,  
62 2011; Castillo et al., 2016; Rausher, 2017).

63 Plants have evolved a wide range of mating system strategies that underly variation in  
64 outcrossing rates. For example, angiosperms have evolved genetic self-incompatibility (SI),

65 herkogamy, dichogamy, dioecy, or even sterile flowers (i.e., nonsexual display that can  
66 enhance pollinator visitation) (Yampolsky & Yampolsky, 1886; Webb & Lloyd, 1986;  
67 Barrett & Harder, 1996; Barrett, 2010; Takayama & Isogai, 2005). Driven by this variation in  
68 mating system strategy, species fall along a continuous distribution of realized selfing rates  
69 (Igic & Kohn, 2006). At the highly-selfing extreme of this spectrum, flowering plants often  
70 evolve correlated suites of floral trait variation. Specifically, the “selfing syndrome” is  
71 characterized by smaller seed size, shorter bud developmental stages, reduced flower size,  
72 and reduced pollen to ovule ratio compared to their outcrossing relatives (Stebbins, 1950;  
73 Wyatt, 1984; Steiner, 1988; Ritland & Ritland, 1989; Sicard & Lenhard, 2011; Renner,  
74 2014). These morphological characteristics consistently accompany transitions from  
75 outcrossing to selfing across angiosperm evolution.

76 The transition to autonomous selfing is one of the most frequent evolutionary  
77 transitions, with ~20% angiosperm species having evolved predominant selfing (autogamy)  
78 from outcrossing (Barrett, 2002). Frequent transitions towards selfing might seem surprising  
79 since selfers often incur the cost of inbreeding depression either through increased  
80 homozygosity of deleterious recessive alleles or through the loss of heterozygosity at loci  
81 exhibiting heterozygote advantage (Charlesworth, 1992; Possingham et al., 2001; Wright et  
82 al., 2013). If selfing occurs over enough generations, purging of deleterious recessive alleles  
83 by natural selection can effectively remove the harmful effects of inbreeding (Lande &  
84 Schemske, 1985; Charlesworth & Charlesworth, 1987). However, this purging can come at a  
85 cost of loss of genetic diversity and adaptive capacity. Despite this disadvantage, evolving  
86 autogamy provides reproductive assurance in situations where seed production is pollen  
87 limited (Lloyd & Schoen, 1992; Fishman & Wyatt, 1999; Busch & Delph, 2012; Toräng et  
88 al., 2017). Moreover, selfing plants transmit two copies of their genes to offspring compared

89 to the one copy transmitted by outcrossing plants, resulting in a two-fold transmission  
90 advantage (Fisher, 1941).

91 Shifts in mating system can also lead to increased reproductive isolation between  
92 species in secondary contact (Brys et al., 2014; Cardona et al., 2020). For example, increased  
93 selfing can decrease hybridization by reducing heterospecific pollen transfer. Several case  
94 studies have found that increased selfing can evolve to avoid hybridization with competing  
95 species, a pattern consistent with reinforcement of speciation (Brown & Wilson, 1956;  
96 Antonovics, 1968; Levin, 1972; Fishman & Wyatt, 1999). Furthermore, macro-evolutionary  
97 studies indicate divergence in mating system is greater in sympatric sister species compared  
98 to allopatric sister species, suggesting the general importance of autonomous selfing for  
99 reproductive isolation (Grossenbacher & Whittall, 2011).

100 The efficiency of selfing as a barrier against heterospecific mating largely depends on  
101 the timing of autonomous selfing (Brys et al., 2016). For increased selfing to decrease  
102 hybridization, self-pollination must either occur before outcross pollination or outcompete  
103 heterospecific pollen when concurrently deposited on the stigma. Evolving an earlier  
104 developmental timing of autonomous self-fertilization can give priority to self-pollen and  
105 thus effectively cause reproductive isolation. Out of the three modes of autonomous selfing  
106 (prior, competing or delayed), prior (or earlier) autonomous selfing significantly reduces the  
107 possibility of successful outcross pollination and thus hybridization (Brys et al., 2016). If  
108 reinforcement were to favor a trait to decrease hybridization by increasing self-fertilization,  
109 then we hypothesize that the timing of selfing will be favored to evolve earlier in sympatric  
110 populations relative to allopatric populations. This pattern would ensure selfing prior to  
111 heterospecific pollen deposition and hybrid formation (Elle et al., 2010; Brys et al., 2016;  
112 Yang et al., 2018). To our knowledge, there has been only one study investigating the timing  
113 of selfing as a contributor to RI which did find a pattern consistent with reinforcement such

114 that the selfing species exhibits earlier selfing in sympatric relative to allopatric populations  
115 (Randle et al., 2018).

116 The annual Texas wildflowers *Phlox drummondii* and *Phlox cuspidata* provide an  
117 ideal system to investigate the interaction between mating system evolution and reproductive  
118 isolation. These two species are partially sympatric and hybridize where they co-occur  
119 (Ferguson et al., 1999; Hopkins & Rausher, 2012; Roda & Hopkins, 2019). The species  
120 flower simultaneously and are visited by the same pollinating species (Burgin et al., 2023;  
121 Hopkins & Rausher, 2012; Ruane & Donohue, 2007). Hybrids between the two *Phlox* species  
122 have strong or complete sterility compared to the two parental species indicating that  
123 hybridization is costly (Suni et al., 2018). Previous research demonstrates that reinforcing  
124 selection favored flower color divergence in sympatric populations of *P. drummondii*, thus  
125 decreasing hybridization with closely related *P. cuspidata* (Hopkins & Rausher, 2012;  
126 Hopkins & Rausher, 2014). Unlike *P. drummondii*, it is unclear whether *P. cuspidata* has  
127 evolved any mechanisms to reduce hybridization in sympatry. *Phlox cuspidata* is  
128 hypothesized to be self-compatible, indicating that variation in autonomous selfing could  
129 reduce hybridization and respond to reinforcing selection (Levin, 1985; Roda et al., 2017). In  
130 this study, we aim to determine whether selfing rate, timing of anther dehiscence, and key  
131 flower traits associated with mating system variation differ between allopatric and sympatric  
132 individuals across several *P. cuspidata* populations.

133 **Materials and Methods**

134 *Study species*

135 *Phlox cuspidata* (Polemoniaceae) is an annual wildflower that occurs throughout  
136 Eastern Texas (Fig. 1). Flowers are five-parted with sexual organs (pistil and anthers) fully  
137 inserted into a fused corolla tube. The stigma is positioned beneath two rows of anthers

138 occurring at two heights in the corolla tube (Fig. 2). Flowers are light blue/purple, fragrant,  
139 and borne in clusters. Each flower produces a single, dehiscent fruit which contains up to  
140 three seeds. Based on its reduced floral size relative to other members of the *Phlox* genus, it  
141 is hypothesized that *P. cuspidata* is predominantly selfing (Levin, 1978; Levin, 1989). In a  
142 study using a single population and relying on a highly limited number of genetic markers,  
143 selfing rate in field conditions was inferred to be ~78% (Levin, 1978). To our knowledge,  
144 there have been no further quantifications of selfing rate in *P. cuspidata* in field or  
145 greenhouse conditions. *Phlox drummondii* has an active genetic self-incompatibility  
146 mechanism that prevents germination of self-pollen on the stigmatic surface (Roda &  
147 Hopkins, 2019). It is unknown whether *P. cuspidata* maintains any function of this genetic SI  
148 mechanism.

149 *P. cuspidata* grows in sympatry with a closely related species, *P. drummondii*, at the  
150 western edge of its range (Fig. 1). These *Phlox* species share pollinators and overlap  
151 temporally throughout their blooming season (Burgin et al., 2023; Hopkins & Rausher, 2012;  
152 Ruane & Donohue, 2007). As a result, hybridization can and does occur, resulting in the  
153 production of hybrids with significantly reduced fitness (Ferguson et al., 1999; Suni et al.,  
154 2018).

155 *Genetic estimates of selfing rate using field-collected individuals*

156 To infer rates of realized selfing in allopatric and sympatric populations, we grew  
157 seeds collected from natural populations and sequenced individuals to estimate genetic  
158 diversity. We collected seeds from naturally growing plants in 2019 and 2021 and grew one  
159 seed from 4-25 maternal plants from 12 populations (6 sympatric and 6 allopatric) for a total  
160 of 171 individuals. We incubated field-collected seeds in 500 ppm gibberellic acid in water  
161 for two days, vernalized planted seeds at 4 °C for ten days, and subsequently moved pots to

162 growth chambers with controlled temperatures of 23 °C during the day and 20 °C at night to  
163 initiate germination.

164 We extracted DNA from fresh leaf tissue using the EZNA Plant DNA kit buffers  
165 (Omega Bio-Tek) combined with a chloroform extraction and two cold 96% ethanol washes  
166 as in Goulet-Scott et al. (2021). We prepared DNA libraries with double-digest restriction  
167 site-associated DNA sequencing (ddRAD-seq) protocol (Peterson et al., 2012). We used a  
168 Pippin Prep (Sage Biosciences) to select for 300-500bp fragments and cleaned pooled  
169 libraries with a Monarch PCR and DNA Cleanup kit (New England Biolabs). Samples were  
170 paired end sequenced (2x150 bp) at the Bauer Core of Harvard University on one Illumina  
171 NovaSeq 6000 lane. Illumina reads are accessible in NCBI's Sequence Read Archive (SRA;  
172 ##).

173 Raw reads were demultiplexed and filtered as described in Goulet-Scott et al., 2021.  
174 Reads were aligned to a complete assembly of the *P. drummondii* genome (Wu et al. 2023,  
175 unpublished), using BWA-MEM, and individuals were genotyped using STACKS v2.64  
176 (Catchen et al., 2013; Li, 2013). Samples with a mean read depth under 10 were removed.  
177 Sites were filtered using BCFtools to include only biallelic SNPs with no more than 50%  
178 missingness across individuals (Danecek & Bonfield et al., 2021). Sites were further filtered  
179 using BCFtools to include only sites with a minor allele frequency greater than 5%. After  
180 these quality filtering steps, 70,118 sites remained. The final dataset included 86 sympatric  
181 and 68 allopatric individuals representing 6 sympatric and 6 allopatric populations (Table 1).  
182 Using the populations program in Stacks v2.64, we calculated the inbreeding coefficient  $F_{IS}$   
183 averaged across loci for each population (Wright, 1949; Hartl & Clark, 1997; Danecek et al.,  
184 2011; Catchen et al., 2013). We inferred selfing rate ( $s$ ) via its relationship to  $F_{IS}$  defined by  
185 the formula (see Hartl & Clark, 1997):

186 
$$F_{IS} = \frac{s}{2-s}$$

187 *Seed collection and plant growth for greenhouse experiments*

188 To quantify traits involved in autonomous self-pollination, seeds from multiple  
189 sympatric and allopatric populations of *P. cuspidata* were collected in 2019 and 2021. These  
190 populations represent some but not all of those included in the genetic analyses (Fig. 1; Table  
191 1). We generated maternal families by growing multiple seeds collected from the same plant  
192 in the field. Individuals within a maternal family range from full to half siblings sharing the  
193 same maternal parent. For a balanced design, we aimed to grow 20 maternal families from  
194 each of four sympatric and four allopatric populations. However, due to low germination  
195 rates, not all populations were included in subsequent analyses (Fig. 1; Table 1). In total, the  
196 experiment included 180 individuals representing 68 allopatric and 112 sympatric individuals  
197 (Table 1). Seeds were soaked in gibberellic acid and vernalized to induce germination as  
198 described above.

199 *Flower and plant size*

200 Germinated seeds were transplanted to 4.5-inch pots (using Pro-Mix HP Mycorrhizae  
201 potting media) and grown to flowering in a greenhouse with 16h of supplemented light and a  
202 temperature ranging from 23 °C to 26 °C. For each plant, we scanned six flowers (three face  
203 up and three on the side) and used ImageJ to calculate flower face diameter and flower length  
204 (Fig. A1). To estimate variation in plant height, the length between the second and third  
205 internode was recorded for each plant.

206 *Anther-dehiscence timing*

207 To determine timing of anther-dehiscence, four flowers were marked on one  
208 experimental plant per maternal family (n=79) at each of five bud developmental stages (e.g.,  
209 Stage 0 = pigmented bud a day before opening, Stage 1 = day 1 of fully opened flower, Stage

210 2 = day 2 of fully opened flower, Stage 3 = day 3 of fully opened flower, Stage 4 = day 4 of  
211 fully opened flower; Fig. 2). For each flower included in the experiment, the anthers and  
212 attached corolla were removed at the assigned stage, leaving behind the pistil. We visually  
213 assessed anthers on the day they were removed from the calyx and recorded whether pollen  
214 dehiscence had occurred as a binary trait. Plants were grown in the greenhouse where all  
215 pollinators are excluded thereby preventing pollen transfer between flowers.

216 *Timing of autonomous selfing*

217 To determine timing of autonomous selfing, we recorded total fruit set from the  
218 experimental flowers that had anthers removed at each developmental stage from Stage 0 to  
219 Stage 4 (n=79) as described above. For a fruit to set, autonomously deposited self-pollen  
220 must have reached the receptive stigma prior to the stage at which the anthers were removed  
221 (Fig. 2). If dehisced pollen has not yet reached the receptive stigma, no fruit will form even at  
222 developmental stages when anthers have dehisced.

223 *Anther-stigma distance*

224 To quantify herkogamy (anther-stigma distance), two additional buds/flowers were  
225 marked at both developmental Stage 0 and 1 on the 79 plants included in the autonomous  
226 selfing experiment. The heights of the stamens and the pistil were measured from the bottom  
227 of the ovary. We calculated the distance from the stigma to the anthers by subtracting the  
228 height of the shortest stamen (closest to the stigma) from the height of the pistil.

229 *Selfing rate and seed count*

230 To estimate the autonomous selfing rate in the greenhouse, we bagged 10-15  
231 unmanipulated flowers on one offspring per plant (n=79) and monitored for seed set. Selfing  
232 rate for each plant was calculated as the proportion of flowers that formed fruits. Seed count  
233 was estimated by counting the number of seeds developed from a known number of fruits.

234 *Statistical analyses*

235 We constructed a simple linear model that included zone (two categories: sympatry  
236 vs. allopatry) as a fixed factor to assess variation in  $F_{IS}$  and selfing rate inferred from genetic  
237 variation. Our greenhouse manipulative experiments aimed to test whether there were  
238 significant differences between sympatric and allopatric individuals of *P. cuspidata* in flower  
239 length and diameter, internode distance, timing of autonomous selfing, anther-stigma  
240 distance, selfing rate, and seed count. Specifically, we analyzed linear and generalized mixed  
241 models to test for differences in each of the seven traits independently. A linear mixed model  
242 was built for flower length, diameter and internode length separately with a normal  
243 distribution that included zone (two categories: allopatric and sympatric) as a fixed factor and  
244 random effects of maternal family to account for plants collected from the same mother and  
245 individual as multiple measurements were taken per plant.

246 A simple linear model was built for autonomous selfing rate in the greenhouse and  
247 seed count separately with zone (categorical) as a fixed factor and fruit number as an offset  
248 variable for seed count. To analyze variation in timing of selfing, a generalized linear model  
249 was built for fruit set with a binomial distribution that included zone as a categorical variable,  
250 stage as a numerical variable and their interaction, with individual as random effects as each  
251 plant included was observed for fruit set over the five developmental stages.

252 For anther-stigma distances, a linear model of the minimum distance between the  
253 stigma and anther was built with zone and stage as categorical fixed factors along with their  
254 interaction and maternal family and individual as random effect because the minimum  
255 distance was measured on two flowers per stage per plant. Because there were more than  
256 twice the number of sympatric than allopatric individuals (Table 1), an equal number of  
257 sympatric individuals were randomly sampled 1,000 times and p-values were compared for

258 all floral traits measured. All statistical analyses were performed in R (R Core Team, 2021)  
259 using package *lme4* (Bates et al., 2015).

260 **Results**

261 *Genetic estimates of selfing from field-collected individuals*

262 Genetic diversity measured from field collected individuals indicates some but not  
263 complete selfing in nature. After filtering for quality and coverage, we analyzed genetic  
264 diversity using 154 individuals (86 sympatric and 68 allopatric) from 12 populations (6  
265 sympatric and 6 allopatric). Estimates of  $F_{IS}$  were similar across all populations with no  
266 differences between sympatric and allopatric populations (allopatric = 0.3377, sympatric =  
267 0.3064) (Tables A1, A2). Inferred rates of selfing based on estimates of  $F_{IS}$  were intermediate  
268 and not significantly different between allopatric and sympatric populations (allopatric =  
269 49.94%, sympatric = 45.29%) (Fig. 3a; Tables A1, A2).

270 *Autonomous selfing in greenhouse-grown plants*

271 At developmental Stage 0 (the day before bud opening) no marked flowers had  
272 anthers dehisced. By developmental Stage 1 (day 1 of open flower), all marked flowers had  
273 dehisced anthers. Fruit set increased over time across all populations, with 0% fruit set at  
274 developmental Stage 0 to nearly 100% fruit set by developmental Stage 4 (day 4 of open  
275 flower) (Fig. 3b). We detected no variation in anther dehiscence timing, autonomous-self  
276 fruit set across developmental stages, autonomous selfing rate or seed count between  
277 sympatric and allopatric populations (Table 2; Fig. 2; Fig. 3c, d, Fig. A2). On average,  
278 sympatric and allopatric *P. cuspidata* individuals had a 92% and 93% selfing rate  
279 respectively with 2.7 seeds per fruit (out of a maximum of 3) (Fig. 3c, d; Fig. A2).

280 *Floral traits of greenhouse-grown plants*

281 Flower length and diameter were significantly greater for sympatric populations of *P.*  
282 *cuspidata* compared to allopatric populations (Table 2; Fig. 3e, f; Fig. A2). In contrast, there  
283 were no significant differences between sympatric and allopatric individuals in internode  
284 length or anther-stigma distance. Although these differences did not pass significance, mean  
285 internode distance is larger among sympatric plants indicating overall larger plants in  
286 sympatry (Table 2; Fig. 3g, h; Fig. A2).

287 **Discussion**

288 The cost of producing low fitness hybrids can generate reinforcing selection favoring  
289 traits that decrease hybridization between sympatric species. We tested the hypothesis that  
290 reinforcement has favored the evolution of increased and earlier selfing in *P. cuspidata*  
291 individuals sympatric with *P. drummondii*. These two species share a broad region of  
292 geographic overlap, are known to hybridize in nature, and produce highly sterile offspring  
293 making the conditions seemingly ideal for reinforcement. However, our results indicate that  
294 the selfing syndrome is not more exaggerated in sympatry than allopatry in *P. cuspidata*. In  
295 fact, plants from both sympatric and allopatric populations are efficient selfers with high  
296 autonomous selfing rates in the greenhouse. Population genetic estimates of selfing rate  
297 demonstrate intermediate selfing in natural conditions, with no difference between sympatric  
298 and allopatric populations, consistent with the lack of difference observed in greenhouse  
299 grown plants. Our analysis of *P. cuspidata* populations spanning its Texas range revealed no  
300 significant differences in timing of selfing or other floral traits associated with selfing  
301 syndrome including anther dehiscence timing and anther-stigma distances between the  
302 allopatric and sympatric populations growing with *P. drummondii*. Contrary to the hypothesis  
303 of reinforcement favoring increased selfing in sympatry, we find that sympatric individuals  
304 have larger floral length and diameter than allopatric individuals.

305 *Phlox cuspidata* displays a highly efficient selfing syndrome throughout its  
306 geographic range. It has evolved small tubular corollas and anthers dehisce as soon as the  
307 flower opens. Under greenhouse conditions, autonomous selfing results in nearly 100% fruit  
308 set within four days after a flower opens. Despite this high potential for selfing, patterns of  
309 genetic variation indicate that the selfing rate in nature is only around 45-50%. The large  
310 difference in selfing rate estimated in the greenhouse versus via genetic data may be driven  
311 by inbreeding depression if selfed offspring have lower fitness compared to outcrossed  
312 offspring in nature. While there may be some contribution of inbreeding depression to this  
313 apparent difference, it is clear from the population genetic data that natural populations of *P.*  
314 *cuspidata* reproduce through both selfing and outcrossing. This finding is consistent with the  
315 complex relationship between the potential for selfing and realized outcrossing rates  
316 described across angiosperms. Although strongly self-incompatible species tend to have high  
317 outcrossing rates, strongly self-compatible species span a range of outcrossing rates (Raduski  
318 et al., 2011). Our data suggest that *P. cuspidata* is strongly self-compatible with an  
319 intermediate outcrossing rate in nature (~45-50%). Previous field observations demonstrate  
320 pollinator visitation to *P. cuspidata* flowers, supporting our finding of intermediate  
321 outcrossing rates in natural populations (Hopkins & Rausher, 2012; Briggs et al., 2018).  
322 Because *P. cuspidata* achieves reproduction through both outcrossing and selfing, reinforcing  
323 selection may act on traits that vary the timing and efficiency of self-fertilization to decrease  
324 costly hybridization with *P. drummondii*.

325 Mating system variation is known to evolve to prevent costly hybridization in a  
326 variety of other systems across the angiosperm phylogeny. For example, case studies in  
327 *Arenaria uniflora* (Caryophyllaceae) (Fishman & Wyatt, 1999), *Collinsia rattanii*  
328 (Scrophulariaceae) (Randle et al., 2018) and *Clarkia xantiana* subsp. *parviflora* (Onagraceae)  
329 (Briscoe Runquist & Moeller, 2013) show evidence that reinforcement has favored the

330 evolution of increased selfing in sympatric populations. Furthermore, a metanalysis in  
331 *Mimulus* found that sympatric sister species are more likely to have one species with  
332 characteristic selfing-syndrome floral traits than allopatric sister species, which may be  
333 caused by selection to avoid hybridization in sympatry (Grossenbacher & Whittall, 2011).  
334 Finally, theoretical work supports increased selfing as a viable and, under some conditions,  
335 favorable mechanism for decreasing hybridization between sympatric species (Castillo et al.,  
336 2016; Rausher, 2017). Collectively, previous work demonstrates how a shift in selfing rate  
337 can serve as a reproductive isolating mechanism favored by reinforcement. Despite this  
338 strong empirical and theoretical basis provided by other systems, we find no evidence of  
339 increased selfing or selfing syndrome traits in sympatric *P. cuspidata* individuals. There is no  
340 obvious difference between *Phlox* and other systems previously investigated that might  
341 explain why we do not find evidence of reinforcement in *P. cuspidata*. We discuss several  
342 hypotheses to explain this finding here.

343 First, it may be that the net selection favoring increased selfing is not as strong as we  
344 originally predicted. Hybrids of *P. cuspidata* and *P. drummondii* are nearly completely  
345 sterile, yet if hybridization is exceedingly rare, the overall strength of reinforcing selection  
346 will be low. It has already been demonstrated that flower color variation in the sympatric  
347 species *P. drummondii* can decrease hybridization by 50% (Hopkins & Rausher, 2012). As a  
348 result, there may be very little current hybridization to generate selection favoring increased  
349 selfing in sympatric *P. cuspidata*. Theoretical analyses demonstrate that it is difficult for  
350 reinforcement to complete speciation (i.e., stop hybridization completely) because the  
351 strength of selection diminishes as reproductive isolation increases (Bank et al., 2011).

352 Additionally, it could be that realized selfing is higher than estimated by the  
353 population genetic measures presented here. If selfing in nature is as high as selfing in the  
354 greenhouse, selection would have little room to act to increase selfing rate in sympathy.

355 Selfing estimates based on population-level heterozygote deficiency ( $F_{IS}$ ) are imperfect  
356 because they rely on assumptions rarely met in nature. Previous studies demonstrate that  $F_{IS}$ -  
357 based calculations tend to overestimate the selfing rate relative to progeny arrays when the  
358 selfing rate is  $< 50\%$  (Jarne & Auld, 2006). Therefore, we expect the true selfing rate in  
359 natural *P. cuspidata* populations to be lower, or at least not much higher, than the estimates  
360 presented here. More precise estimates of selfing based on progeny arrays would provide  
361 additional clarity (Colicchio & Herman, 2020).

362 Furthermore, the cost of hybridization favoring increased selfing could be partially  
363 countered by selection against selfing due to inbreeding depression and its associated costs. If  
364 selection towards maintaining genetic diversity with higher outcrossing rates is high, then net  
365 selection on the timing of selfing and selfing rate will be weak or counter to the expectations  
366 of reinforcement (Epinat & Lenormand, 2009; Pickup et al., 2019). This hypothesis is  
367 particularly intriguing for two reasons. First, we found that flowers from sympatric  
368 individuals were larger. Floral display size is an important signal for pollinators, and these  
369 findings may reflect selection to increase pollinator attraction. Consistent with increased  
370 pollinator attraction, we inferred that selfing rates in nature were somewhat, although not  
371 significantly, lower in sympatry than allopatry. Second, we found that autonomous selfing  
372 rate in the greenhouse is low on the first day a flower is open ( $\sim 10\%$ ) and remains low  
373 through the second day ( $\sim 25\%$ ). As a result, there is high potential for outcrossing during the  
374 first days of a flower's lifespan. These data are consistent with the delayed selfing hypothesis  
375 which suggests that selfing occurring after outcrossing opportunities have passed should be  
376 selected for under a wide range of ecological conditions (Goodwillie & Weber, 2018). If *P.*  
377 *cuspidata* experiences selection for delayed selfing as a mechanism to maintain opportunities  
378 for outcrossing, it may explain why we fail to find evidence for the evolution of earlier  
379 selfing in sympatric populations. Further investigations of pollinator visitation, timing of

380 selfing in natural populations, and the relative fitness of selfed versus outcrossed offspring  
381 could address these hypotheses.

382 Another possible explanation for the observed absence of reinforcing selection is a  
383 lack of genetic variation underlying selfing related traits available for selection to act upon.  
384 Across all individuals, from all populations, we found that all anthers dehisced on the first  
385 day of the open flower. Although we only monitored anther dehiscence once per day, we  
386 found no evidence that anthers could release pollen earlier than flower opening in this species  
387 or later than the morning of flower opening. Alternatively, there could be negative pleiotropic  
388 effects of adjusting anther dehiscence timing due to correlated developmental traits or  
389 fertility. If strong purifying selection results in highly coordinated timing of floral  
390 development, we would expect to find little to no variation in anther dehiscence timing  
391 consistent with our findings in *P. cuspidata*. Future work should focus on investigating the  
392 extent of adaptive divergence in other floral traits across populations of *P. cuspidata* to  
393 clarify whether anther dehiscence timing is a uniquely invariant trait.

394 In summary, we found no systematic difference between sympatric and allopatric  
395 individuals of *P. cuspidata* in the timing of selfing, anther dehiscence timing, anther-stigma  
396 distance, or self-seed set between the sympatric and allopatric populations. Despite the high  
397 potential for selfing in *P. cuspidata*, population genetic estimates of selfing rate indicate  
398 intermediate selfing rates in nature, with no difference in sympatric versus allopatric  
399 populations. Although reinforcement underlies mating system divergence in other systems,  
400 we find no evidence of reinforcing selection acting on the timing of selfing in *P. cuspidata*.  
401 Understanding why reinforcement occurs in some cases but not others can help clarify its  
402 overall importance to the speciation process and the dynamics under which reinforcement  
403 evolves.

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409 **Author Contribution**

410 BMS, GAB and RH designed the study. BMS and GAB collected the data. BMS and GAB  
411 analyzed the data. BMS, GAB and RH wrote the manuscript.

412 **Data Availability Statement**

413 The data and statistical code that support the findings of this study are available on Dryad.

414 **Conflict of Interest Statement**

415 The authors declare no conflict of interest.

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420 **References**

421 Antonovics, J. (1968). Evolution in closely adjacent plant populations V. Evolution of self-  
422 fertility. *Heredity*, 23(2), 219–238. <https://doi.org/10.1038/hdy.1968.30>

423 Baack, E., Melo, M. C., Rieseberg, L. H., & Ortiz-Barrientos, D. (2015). The origins of  
424 reproductive isolation in plants. *New Phytologist*, 207(4), 968–984.  
425 <https://doi.org/10.1111/NPH.13424>

426 Bank, C., Hermission, J., & Kirkpatrick, M. (2011). Can reinforcement complete speciation?  
427 *Evolution*, 66(1), 229–239. <https://doi.org/10.1111/j.1558-5646.2011.01423.x>

428 Barrett, S. C. H. (2002). The evolution of plant sexual diversity. *Nature Reviews. Genetics*,  
429 3(4), 274–284. <https://doi.org/10.1038/NRG776>

430 Barrett, S. C. H. (2010). Understanding plant reproductive diversity. *Philosophical  
431 Transactions of the Royal Society B: Biological Sciences*, 365(1537), 99.  
432 <https://doi.org/10.1098/RSTB.2009.0199>

433 Barrett, S. C. H., & Harder, L. D. (1996). Ecology and evolution of plant mating. *Trends in  
434 Ecology & Evolution*, 11(2), 73–79. [https://doi.org/10.1016/0169-5347\(96\)81046-9](https://doi.org/10.1016/0169-5347(96)81046-9)

435 Bates, D., Mächler, M., Zurich, E., Bolker, B. M., & Walker, S. C. (2015). Fitting linear  
436 mixed-effects models using lme4. *Journal of Statistical Software*, 67, 1–48.  
437 <https://doi.org/10.18637/jss.v067.i01>

438 Briggs, H. M., Graham, S., Switzer, C. M., & Hopkins, R. (2018). Variation in context-  
439 dependent foraging behavior across pollinators. *Ecology and Evolution*, 8(16), 7964–  
440 7973. <https://doi.org/10.1002/ece3.4303>

441 Briscoe Runquist, R. D., & Moeller, D. A. (2013). Floral and mating system divergence in  
442 secondary sympatry: Testing an alternative hypothesis to reinforcement in *Clarkia*.  
443 *Annals of Botany*, 113(2), 223–235. <https://doi.org/10.1093/aob/mct218>

444 Brown, W. L., & Wilson, E. O. (1956). Character displacement. *Systematic Zoology*, 5(2),  
445 49–64. <https://doi.org/10.2307/2411924>

446 Brys, R., van Cauwenberghe, J., & Jacquemyn, H. (2016). The importance of autonomous  
447 selfing in preventing hybridization in three closely related plant species. *Journal of  
448 Ecology*, 104(2), 601–610. <https://doi.org/10.1111/1365-2745.12524>

449 Brys, R., vanden Broeck, A., Mergeay, J., & Jacquemyn, H. (2014). The contribution of  
450 mating system variation to reproductive isolation in two closely related *Centaurea*  
451 species (Gentianaceae) with a generalized flower morphology. *Evolution*, 68(5), 1281–  
452 1293. <https://doi.org/10.1111/EVO.12345>

453 Burgin, G.A., Bronzo-Munich, O., Garner, A.G., Acevedo, I.A., Hopkins, R. (2023).  
454 Characterizing each step of pollination in *Phlox drummondii* reveals that a single  
455 butterfly species predominates in the pollinator assemblage. *American Journal of Botany*,  
456 <https://doi.org/10.1002/ajb2.16172>.

457 Busch, J. W., & Delph, L. F. (2012). The relative importance of reproductive assurance and  
458 automatic selection as hypotheses for the evolution of self-fertilization. *Annals of  
459 Botany*, 109(3), 553. <https://doi.org/10.1093/AOB/MCR219>

460 Cardona, J., Lara, C., & Ornelas, J. F. (2020). Pollinator divergence and pollination isolation  
461 between hybrids with different floral color and morphology in two sympatric *Penstemon*  
462 species. *Scientific Reports*, 10(1), 1–17. <https://doi.org/10.1038/s41598-020-64964-8>

463 Castillo, D. M., Gibson, A. K., & Moyle, L. C. (2016). Assortative mating and self-  
464 fertilization differ in their contributions to reinforcement, cascade speciation, and  
465 diversification. *Current Zoology*, 62(2), 169–181. <https://doi.org/10.1093/CZ/ZOW004>

466 Catchen, J., Hohenlohe, P. A., Bassham, S., Amores, A., & Cresko, W. A. (2013). Stacks: An  
467 analysis tool set for population genomics. *Molecular Ecology*, 22(11), 3124–3140.  
468 <https://doi.org/10.1111/mec.12354>

469 Charlesworth, B. (1992). Evolutionary rates in partially self-fertilizing species. *The American  
470 Naturalist*, 140(1), 126–148. <https://doi.org/10.1086/285406>

471 Charlesworth, D., & Charlesworth, B. (1987). Inbreeding depression and its evolutionary  
472 consequences. *Annu. Rev. Ecol. Syst.*, 237–268.  
473 <https://doi.org/10.1146/annurev.es.18.110187.001321>

474 Colicchio, J. M., & Herman, J. (2020). Empirical patterns of environmental variation favor  
475 adaptive transgenerational plasticity. *Ecology and Evolution*, 10(3), 1648–1665.  
476 <https://doi.org/10.1002/ece3.6022>

477 Coyne, J. A., & Orr, H. A. (1989). Patterns of speciation in *Drosophila*. *Evolution*, 43(2),  
478 362–381. <https://doi.org/10.1111/J.1558-5646.1989.TB04233.X>

479 Coyne, J. A., & Orr, H. A. (2004). *Speciation*. Sinauer Associates, Inc.

480 Danecek, P., Bonfield, J. K., Liddle, J., Marshall, J., Ohan, V., Pollard, M. O., Whitwham,  
481 A., Keane, T., McCarthy, S. A., Davies, R. M., & Li, H. (2021). Twelve years of  
482 SAMtools and BCFtools. *GigaScience*, 10(2).  
483 <https://doi.org/10.1093/gigascience/giab008>

484 Dobzhansky, T. (1940). Speciation as a stage in evolutionary divergence. *The American*  
485 *Naturalist*, 74(753), 312–321. <https://doi.org/10.1086/280899>

486 Elle, E., Gillespie, S., Guindre-Parker, S., & Parachnowitsch, A. L. (2010). Variation in the  
487 timing of autonomous selfing among populations that differ in flower size, time to  
488 reproductive maturity, and climate. *American Journal of Botany*, 97(11), 1894–1902.  
489 <https://doi.org/10.3732/AJB.1000223>

490 Epinat, G., & Lenormand, T. (2009). The evolution of assortative mating and selfing with in-  
491 and outbreeding depression. *Evolution*, 63(8), 2047–2060.  
492 <https://doi.org/10.1111/j.1558-5646.2009.00700.x>

493 Ferguson, C. J., Levin, D. A., & Jansen, R. K. (1999). Natural hybridization between an  
494 outcrossing and a selfing *Phlox* (Polemoniaceae): The maternal species of F1 hybrids.  
495 *Plant Systematics and Evolution*, 218(1–2), 153–158. <https://doi:10.1007/bf01087043>

496 Fisher, R. A. (1941). Average excess and average effect of a gene substitution. *Annals of*  
497 *Eugenics*, 11(1), 53–63. <https://doi.org/10.1111/J.1469-1809.1941.TB02272.X>

498 Fishman, L., & Wyatt, R. (1999). Pollinator-mediated competition, reproductive character  
499 displacement, and the evolution of selfing in *Arenaria uniflora* (Caryophyllaceae).  
500 *Evolution*, 53(6), 1723. <https://doi.org/10.2307/2640435>

501 Goodwillie, C., & Weber, J. J. (2018). The best of both worlds? A review of delayed selfing  
502 in Flowering plants. *American Journal of Botany*, 105(4), 641–655.  
503 <https://doi.org/10.1002/ajb2.1045>

504 Goulet-Scott, B. E., Garner, A. G., & Hopkins, R. (2021). Genomic analyses overturn two  
505 long-standing homoploid hybrid speciation hypotheses. *Evolution*, 75(7), 1688–1710.

506 <https://doi.org/10.1111/evo.14279>

507 Grossenbacher, D. L., & Whittall, J. B. (2011). Increased floral divergence in sympatric  
508 monkeyflowers. *Evolution*, 65(9), 2712–2718. [https://doi.org/10.1111/J.1558-5646.2011.01306.X](https://doi.org/10.1111/J.1558-<br/>509 5646.2011.01306.X)

510 Hartl, D. L., and A. G. Clark. (1997). *Principles of population genetics*. Sinauer Associates,  
511 Sunderland, MA.

512 Hopkins, R. (2013). Reinforcement in plants. *The New Phytologist*, 197(4), 1095–1103.  
513 <https://doi.org/10.1111/NPH.12119>

514 Hopkins, R., & Rausher, M. D. (2012). Pollinator-mediated selection on flower color allele  
515 drives reinforcement. *Science*, 335, 1090–1092. <https://doi.org/10.1126/science.1215198>

516 Hopkins, R., & Rausher, M. D. (2014). The cost of reinforcement: selection on flower color  
517 in allopatric populations of *Phlox drummondii*. *The American Naturalist*, 183(5), 693–  
518 710. <https://doi.org/10.1086/675495>

519 Igic, B., & Kohn, J. R. (2006). The distribution of plant mating systems: Study bias against  
520 obligately outcrossing species. *Evolution*, 60(5), 1098. <https://doi.org/10.1554/05-383.1>

521 Jarne, P., & Auld, J. R. (2006). Animals mix it up too: The distribution of self-fertilization  
522 among hermaphroditic animals. *Evolution*, 60(9), 1816–1824.  
523 <https://doi.org/10.1111/j.0014-3820.2006.tb00525.x>

524 Knowlton, N. (1993). Sibling species in the sea. *Annu. Rev. Ecol. Syst*, 24, 189–216.  
525 [www.annualreviews.org](http://www.annualreviews.org)

526 Lande, R., & Schemske, D. W. (1985). The evolution if self-fertilization and inbreeding  
527 depression in plants. I. Genetic models. *Evolution*, 39(1), 24–40.  
528 <https://doi.org/10.1111/J.1558-5646.1985.TB04077.X>

529 Levin, D. A. (1972). The adaptedness of corolla-color variants in experimental and natural  
530 populations of *Phlox drummondii*. *The American Naturalist*, 106(947), 57–70.  
531 <https://doi.org/10.1086/282751>

532 Levin, D. A. (1978). Genetic variation in annual Phlox: Self-compatible versus self-  
533 incompatible species. *Evolution*, 32(2), 245. <https://doi:10.2307/2407593>

534 Levin, D. A. (1985). Reproductive character displacement in Phlox. *Evolution*, 39(6), 1275.  
535 <https://doi.org/10.2307/2408784>

536 Levin, D. A. (1989). Inbreeding depression in partially self-fertilizing phlox. *Evolution*,  
537 43(7), 1417. <https://doi:10.2307/2409457>

538 Li, Heng. (2013). Aligning sequence reads, clone sequences and assembly contigs with BWA-  
539 MEM. arXiv. <https://doi.org/10.48550/arXiv.1303.3997>

540 Lloyd, D. G., & Schoen, D. J. (1992). Self- and cross-fertilization in Plants. I. Functional  
541 Dimensions. *International Journal of Plant Sciences*, 153(3), 358–369.  
542 <https://doi.org/10.1086/297040>

543 Lowry, D. B., Modliszewski, J. L., Wright, K. M., Wu, C. A., & Willis, J. H. (2008). The  
544 strength and genetic basis of reproductive isolating barriers in flowering plants.  
545 *Philosophical Transactions of the Royal Society B: Biological Sciences*, 363(1506),  
546 3009–3021. <https://doi.org/10.1098/RSTB.2008.0064>

547 Noor, M. A. (1995). Speciation driven by natural selection in drosophila. *Nature*, 375(6533),  
548 674–675. <https://doi.org/10.1038/375674a0>

549 Peterson, B. K., Weber, J. N., Kay, E. H., Fisher, H. S., & Hoekstra, H. E. (2012). Double  
550 digest radseq: An inexpensive method for de novo SNP discovery and genotyping in  
551 model and non-model species. *PLoS ONE*, 7(5).  
552 <https://doi.org/10.1371/journal.pone.0037135>

553 Pfennig, K. S., & Rice, A. M. (2014). Reinforcement generates reproductive isolation  
554 between neighbouring conspecific populations of spadefoot toads. *Proceedings of the  
555 Royal Society B: Biological Sciences*, 281(1789).  
556 <https://doi.org/10.1098/RSPB.2014.0949>

557 Pickup, M., Brandvain, Y., Fraïsse, C., Yakimowski, S., Barton, N. H., Dixit, T., Lexer, C.,  
558 Cereghetti, E., & Field, D. L. (2019). Mating system variation in hybrid zones:  
559 Facilitation, barriers and asymmetries to gene flow. *New Phytologist*, 224(3), 1035–  
560 1047. <https://doi.org/10.1111/nph.16180>

561 Possingham, H. P., Lindenmayer, D. B., & McCarthy, M. A. (2001). Population viability  
562 analysis. *Encyclopedia of Biodiversity*, 831–843. [https://doi.org/10.1016/B0-12-226865-2/00356-4](https://doi.org/10.1016/B0-12-226865-<br/>563 2/00356-4)

564 R Core Team. 2021. R: A language and environment for statistical computing. R Foundation  
565 for Statistical Computing, Vienna, Austria.

566 Raduski, A. R., Haney, E. B., & Igić, B. (2011). The expression of self-incompatibility in  
567 angiosperms is bimodal. *Evolution*, 66(4), 1275–1283. [https://doi.org/10.1111/j.1558-5646.2011.01505.x](https://doi.org/10.1111/j.1558-<br/>568 5646.2011.01505.x)

569 Randle, A. M., Spigler, R. B., & Kalisz, S. (2018). Shifts to earlier selfing in sympatry may  
570 reduce costs of pollinator sharing. *Evolution*, 72(8), 1587–1599.  
571 <https://doi.org/10.1111/EVO.13522>

572 Rausher, M. D. (2017). Selfing, local mate competition, and reinforcement. *The American*  
573 *Naturalist*, 189(2), 87–104. <https://doi.org/10.1086/690009>

574 Renner, S. S. (2014). The relative and absolute frequencies of angiosperm sexual systems:  
575 dioecy, monoecy, gynodioecy, and an updated online database. *American Journal of*  
576 *Botany*, 101(10), 1588–1596. <https://doi.org/10.3732/AJB.1400196>

577 Ritland, C., & Ritland, K. (1989). Variation of sex allocation among eight taxa of the  
578 *Mimulus guttatus* species complex (Scrophulariaceae). *American Journal of Botany*,  
579 76(12), 1731–1739. <https://doi.org/10.2307/2444472>

580 Roda, F., & Hopkins, R. (2019). Correlated evolution of self and interspecific incompatibility  
581 across the range of a Texas wildflower. *New Phytologist*, 221(1), 553–564.  
582 <https://doi.org/10.1111/NPH.15340>

583 Roda, F., Mendes, F. K., Hahn, M. W., & Hopkins, R. (2017). Genomic evidence of gene  
584 flow during reinforcement in Texas Phlox. *Molecular Ecology*, 26(8), 2317–2330.  
585 <https://doi.org/10.1111/MEC.14041>

586 Ruane, L. G., & Donohue, K. (2007). Environmental effects on pollen-pistil compatibility  
587 between *Phlox cuspidata* and *P. drummondii* (Polemoniaceae): implications for  
588 hybridization dynamics. *American Journal of Botany*, 94(2), 219–227.  
589 <https://doi.org/10.3732/AJB.94.2.219>

590 Schemske, D. W., & Bradshaw, H. D. (1999). Pollinator preference and the evolution of  
591 floral traits in monkeyflowers (*Mimulus*). *Proceedings of the National Academy of*

592 *Sciences of the United States of America*, 96(21), 11910–11915.

593 <https://doi.org/10.1073/pnas.96.21.11910>

594 Sicard, A., & Lenhard, M. (2011). The selfing syndrome: a model for studying the genetic  
595 and evolutionary basis of morphological adaptation in plants. *Annals of Botany*, 107(9),  
596 1433–1443. <https://doi.org/10.1093/AOB/MCR023>

597 Stebbins, G. L. (1950). *Variation and evolution in plants*. Columbia University Press.  
598 <https://doi.org/10.7312/STEB94536/HTML>

599 Steiner, K. E. (1988). Dioecism and its correlates in the cape flora of South Africa. *American  
600 Journal of Botany*, 75(11), 1742–1754. [https://doi.org/10.1002/J.1537-2197.1988.TB11250.X](https://doi.org/10.1002/J.1537-<br/>601 2197.1988.TB11250.X)

602 Suni, S. S., & Hopkins, R. (2018). The relationship between postmating reproductive  
603 isolation and reinforcement in *Phlox*. *Evolution*, 72, 1387–1398.  
604 <https://doi.org/10.1111/evo.13507>

605 Tabah, D. A., & Hiscock, S. J. (2003). Flowering and reproduction | Sporophytic self–  
606 incompatibility. *Encyclopedia of Applied Plant Sciences*, 342–349.  
607 <https://doi.org/10.1016/B0-12-227050-9/00020-X>

608 Takayama, S., & Isogai, A. (2005). Self-incompatibility in plants. *Annual Review of Plant  
609 Biology*, 56, 467–489. <https://doi.org/10.1146/annurev.arplant.56.032604.144249>

610 Toräng, P., Vikström, L., Wunder, J., Wötzl, S., Coupland, G., & Ågren, J. (2017).  
611 Evolution of the selfing syndrome: Anther orientation and herkogamy together determine  
612 reproductive assurance in a self-compatible plant. *Evolution*, 71(9), 2206–2218.  
613 <https://doi.org/10.1111/EVO.13308>

614 Webb, C. J., & Lloyd, D. G. (1986). The avoidance of interference between the presentation  
615 of pollen and stigmas in angiosperms II. Herkogamy. *New Zealand Journal of Botany*,  
616 24(1), 163–178. <https://doi.org/10.1080/0028825X.1986.10409726>

617 Wendt, T., Canela, M. B. F., Klein, D. E., & Rios, R. I. (2002). Selfing facilitates  
618 reproductive isolation among three sympatric species of Pitcairnia (Bromeliaceae). *Plant  
619 Systematics and Evolution*, 232(3), 201–212. <https://doi.org/10.1007/S006060200043>

620 Wright, S. (1949). The genetical structure of populations. *Annals of Eugenics*, 15(4), 323–  
621 354. <https://doi.org/10.1111/j.1469-1809.1949.tb02451.x>

622 Wright, S. I., Kalisz, S., & Slotte, T. (2013). Evolutionary consequences of self-fertilization  
623 in plants. *Proceedings of the Royal Society B: Biological Sciences*, 280(1760).  
624 <https://doi.org/10.1098/RSPB.2013.0133>

625 Wu, F., D. Kohst, S. Chaturvedi, P. McKenzie, A. Garner, T. Sackton, R. Hopkins. (2023).  
626 Unpublished Phlox Genome.

627 Wyatt, R. (1984). The evolution of self-pollination in granite outcrop species of Arenaria  
628 (Caryophyllaceae). I. Morphological Correlates. *Evolution*, 38(4), 804.  
629 <https://doi.org/10.2307/2408392>

630 Yampolsky, C., & Yampolsky, H. (1886). *Distribution of sex forms in the phanerogamic  
631 flora*.

632 Yang, J. Q., Fan, Y. L., Jiang, X. F., Li, Q. J., & Zhu, X. F. (2018). Correlation between the  
633 timing of autonomous selfing and floral traits: a comparative study from three selfing  
634 Gentianopsis species (Gentianaceae). *Scientific Reports*, 8(1), 1–7.  
635 <https://doi.org/10.1038/s41598-018-21930-9>

636 Yukilevich, R. (2011). Asymmetrical patterns of speciation uniquely support reinforcement  
637 in *Drosophila*. *Evolution*, 66(5), 1430–1446. [https://doi.org/10.1111/J.1558-  
638 5646.2011.01534.X](https://doi.org/10.1111/J.1558-5646.2011.01534.X)

639

640 **Figure captions**

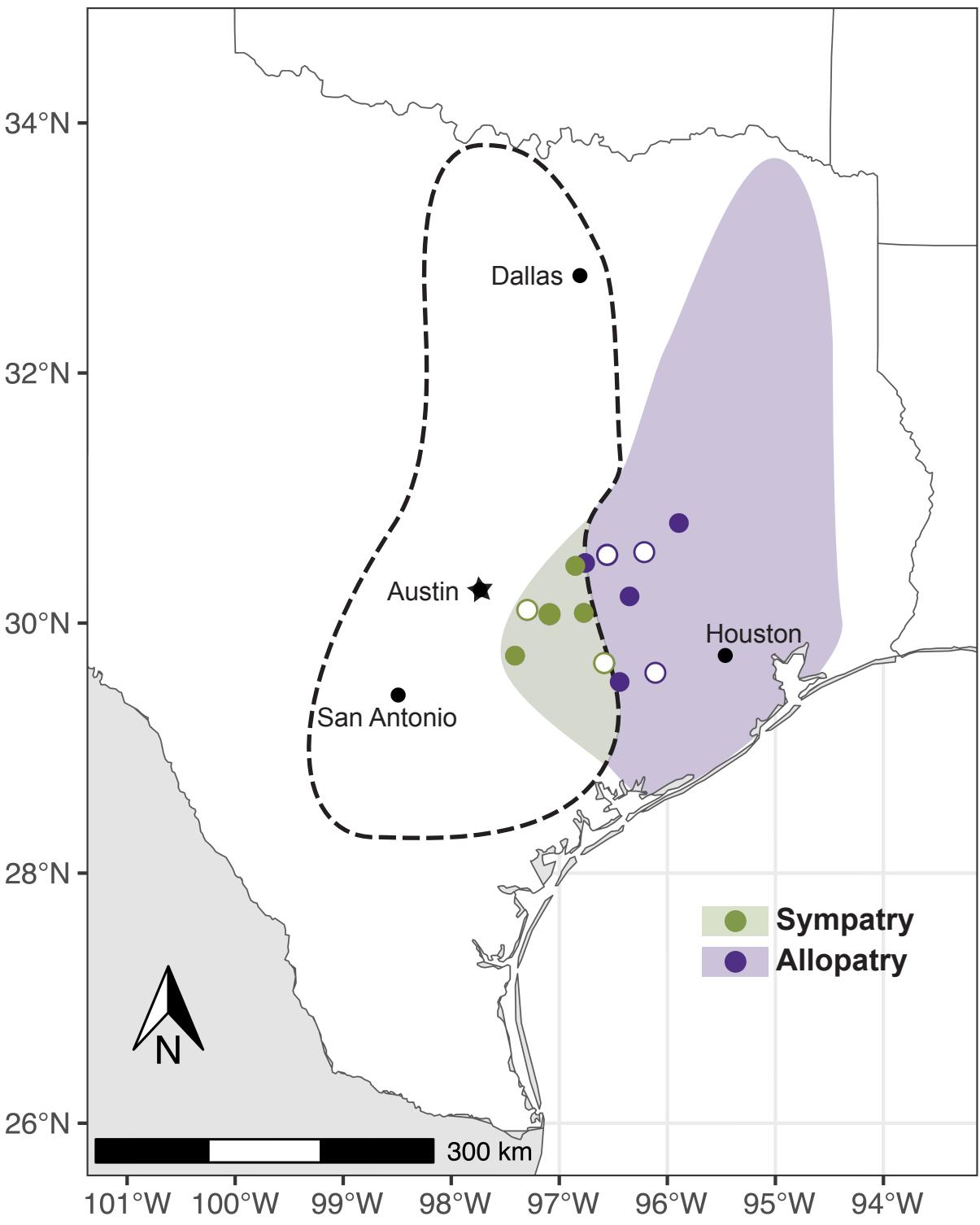
641 Figure 1: The range of *Phlox cuspidata* extends throughout Eastern Texas. The range of *P.*  
642 *drummondii* is indicated with a dashed black line. The allopatric portion of *P. cuspidata*'s  
643 range is shaded purple while the sympatric range is shaded green. All marked points  
644 represent locations of sampled populations used in this experiment (allopatric populations are  
645 shown in purple while sympatric populations are shown in green) with solid circles represent  
646 those included in the greenhouse experiment.

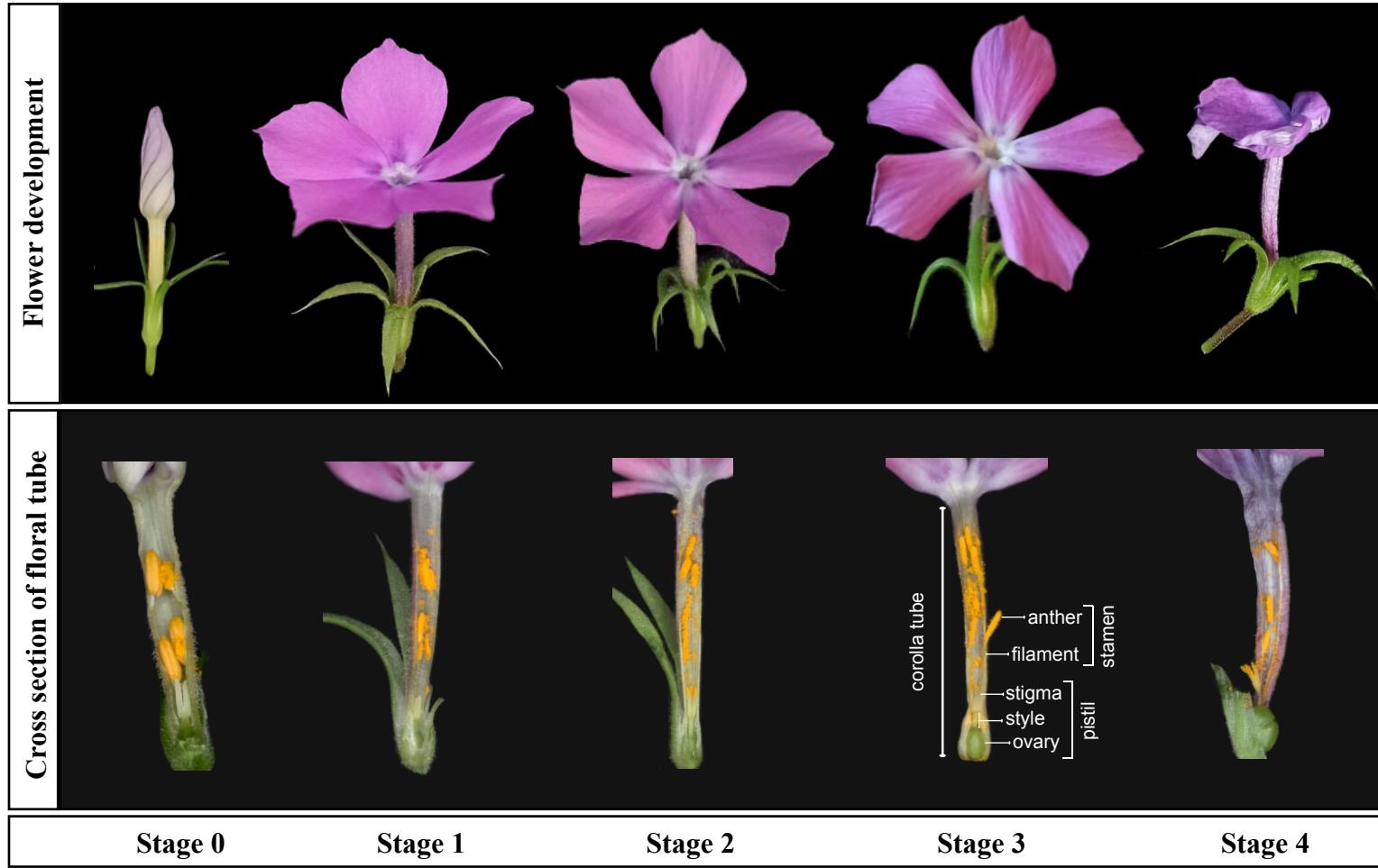
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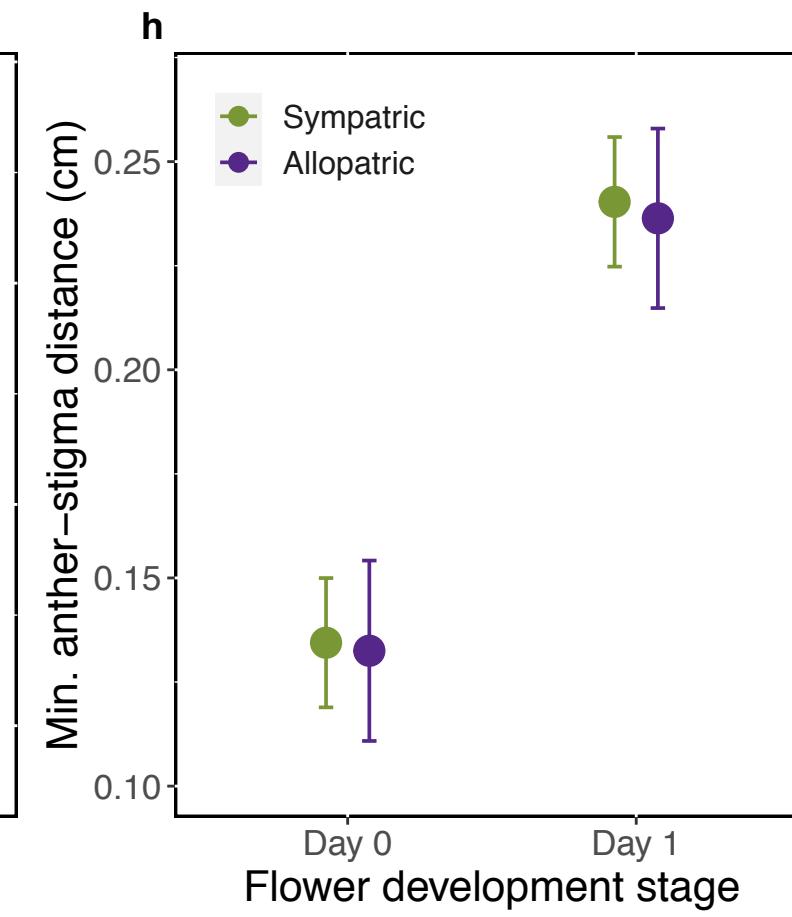
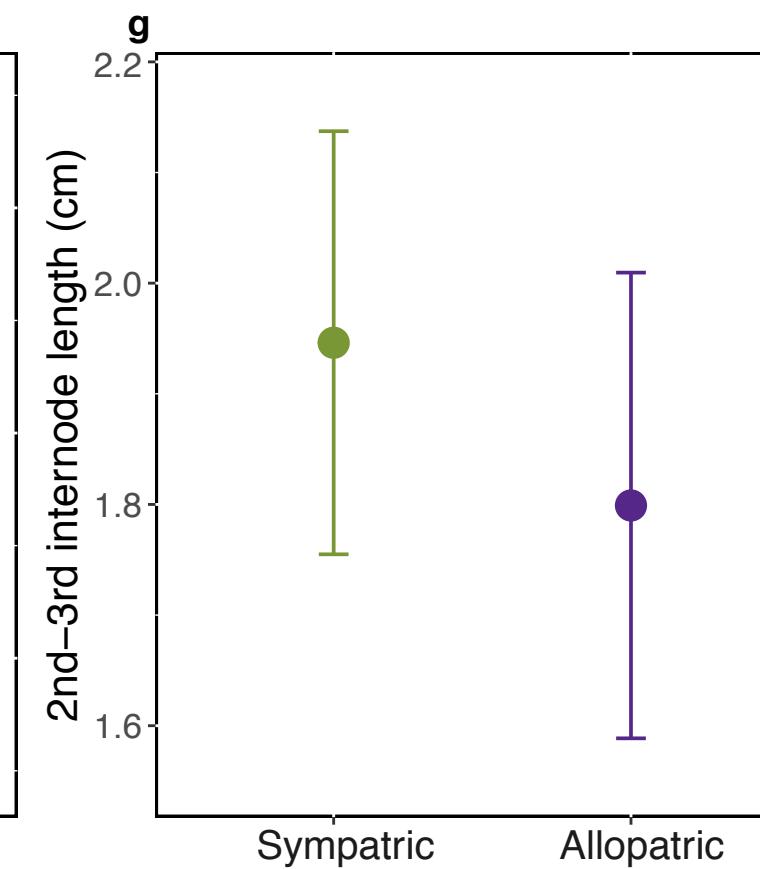
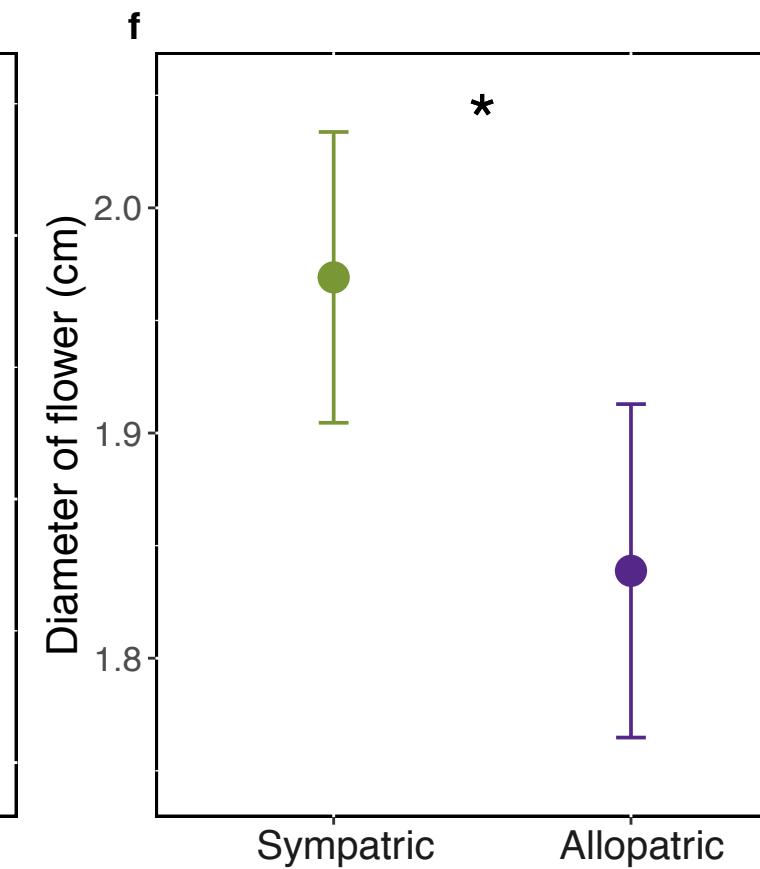
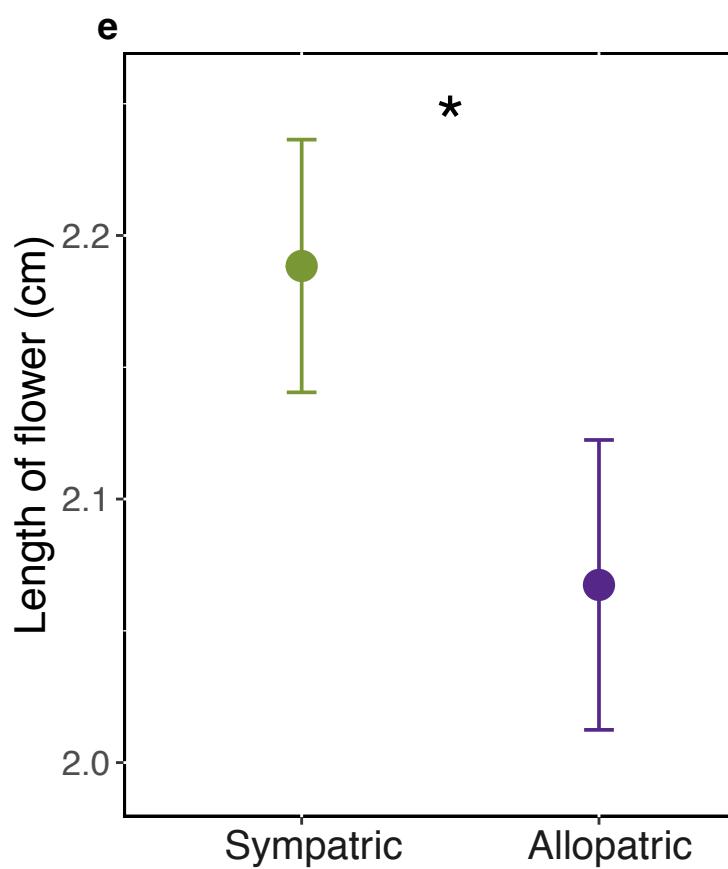
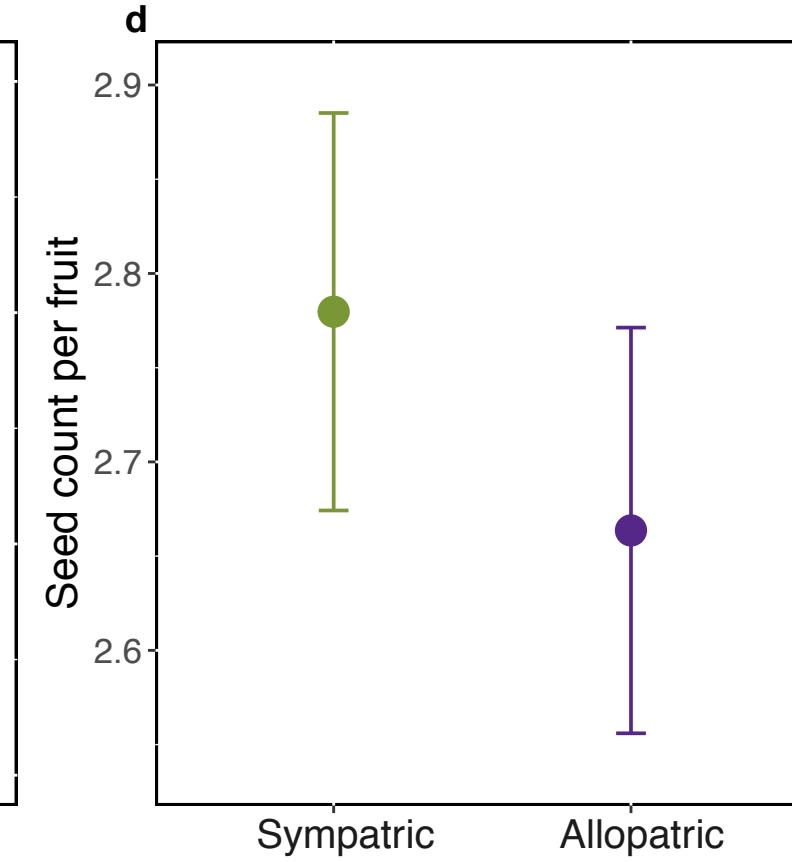
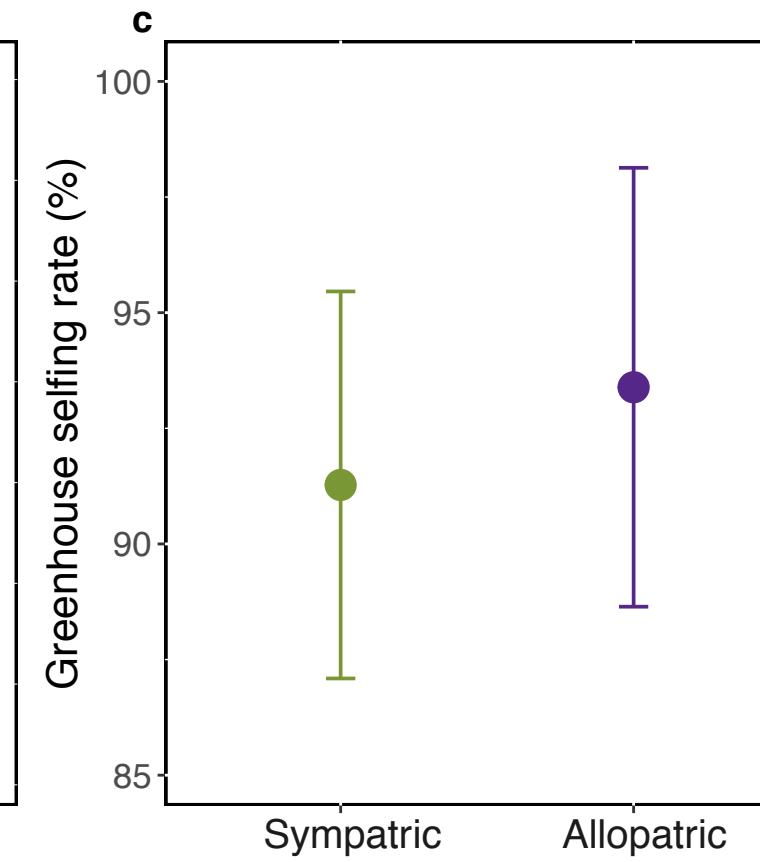
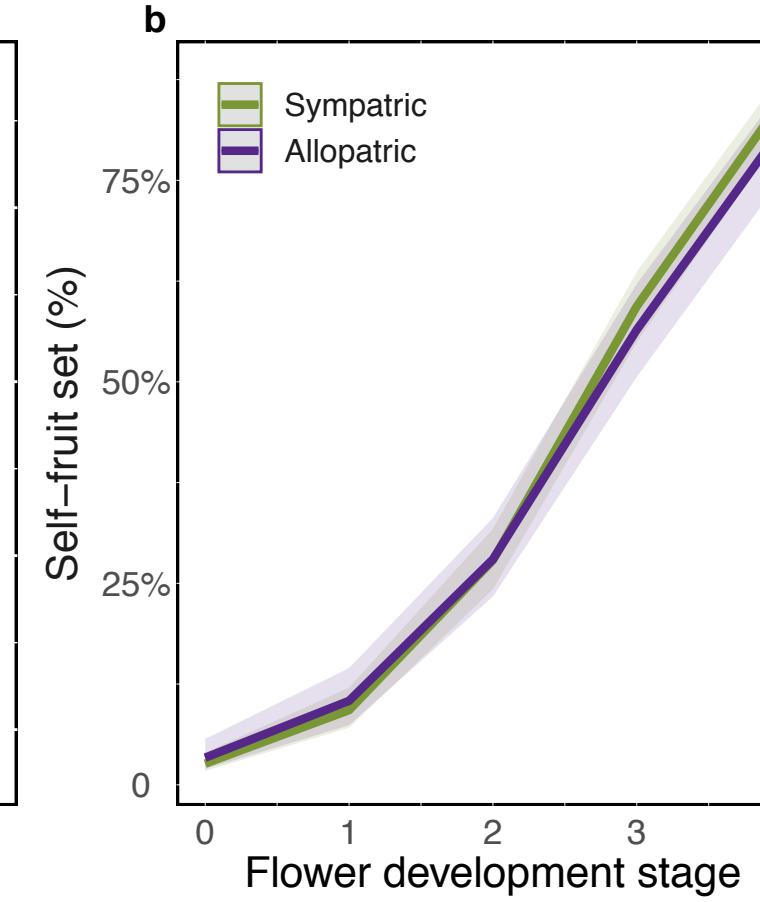
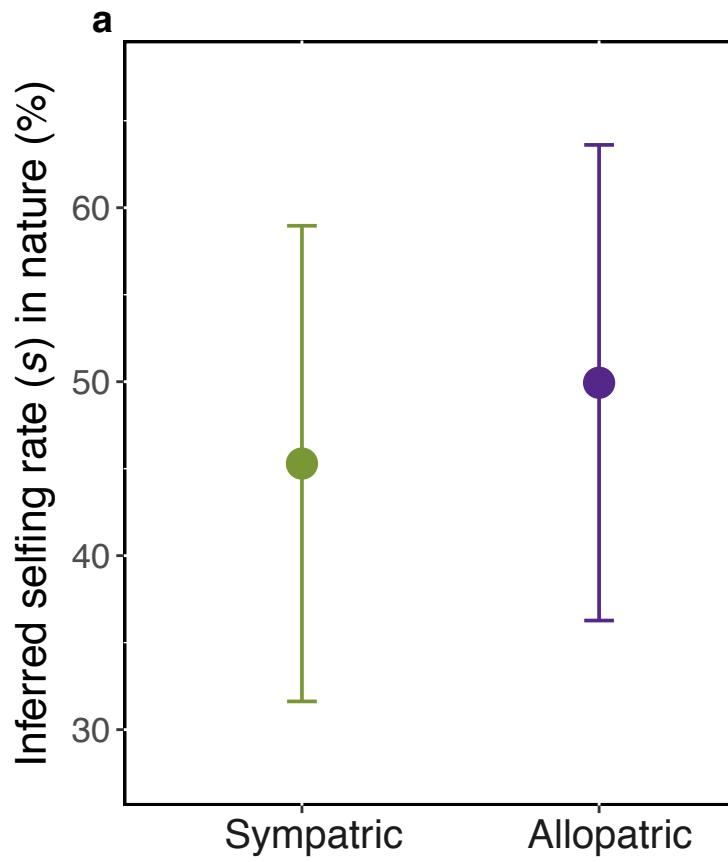
648 Figure 2: Floral development of *Phlox cuspidata* including cross sections of the floral tube (top and  
649 bottom panels). Five stages of floral and anther-stigma development (left to right) where Stage 0 =  
650 Pigmented bud a day before opening with no anther dehiscence (slight damage occurred to anthers  
651 during dissection), Stage 1 = Day 1 of a fully open flower with dehisced anthers, Stage 2 = Day 2 of  
652 open flower with some pollen on stigma, Stage 3 = Day 3 of open flower starting to wilt with most  
653 pollen around and on stigma, and Stage 4 = Day 4 showing flower completely wilted with extensive  
654 pollen having reached stigma.

655

656 Figure 3: Variation in traits between sympatric (green) and allopatric (purple) *Phlox cuspidata*  
657 individuals including (A) selfing rate inferred from genetic data (B) autonomous self-fruit set across  
658 flower developmental stage (C) total autonomous selfing rate in greenhouse conditions (D) seed count  
659 per fruit from autonomous self-set seeds in the greenhouse (E) flower length (F) flower diameter (G)  
660 internode length between 2<sup>nd</sup> and 3<sup>rd</sup> leaf pair (H) minimum distance between anther and stigma. The  
661 mean of the seven traits measured are plotted as points. 95% confidence intervals are indicated as  
662 bars. Asterisks (\*) indicate significant differences in flower length and diameter between sympatric  
663 and allopatric populations.







Appendices for manuscript: Experimental and genetic analysis of selfing reveal no reinforcement in *Phlox cuspidata*

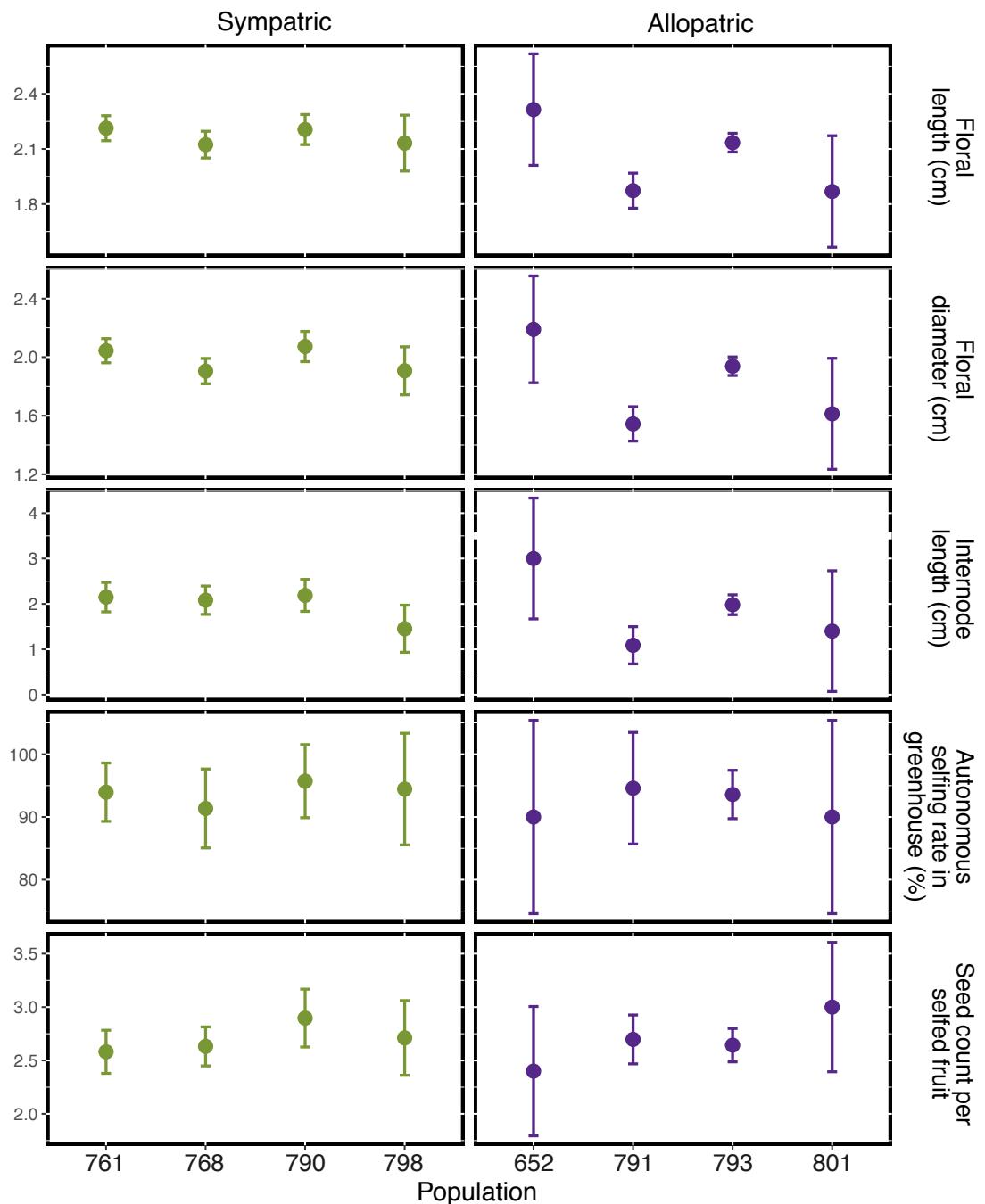
Authors: Bushra M. Shahid\*, Grace A. Burgin\*, Robin Hopkins

\*authors contributed equally

**Appendix Figure 1.** Floral size was measured by scanning six flowers per individual plant. **(A)** Scans included three face-down flowers to measure floral diameter as indicated by the dotted black line. The diameter was calculated by summing both dotted lines. **(B)** An additional three flowers were scanned on their side to measure flower height as indicated by the dotted black line. Flower height was then calculated by summing both dotted lines. All measurements were calculated using ImageJ.



**Appendix Figure 2.** Floral trait variation by population in sympatry versus allopatry. Sympatric populations are shown in the left panel in green. Allopatric populations are shown in the right panel in purple. Predicted population means are plotted as points and 95% confidence intervals are shown as lines. Floral traits plotted from top to bottom include floral length, flower diameter, internode length (2<sup>nd</sup>-3<sup>rd</sup> internode), autonomous selfing rate under greenhouse conditions and seed count per selfed fruit.



**Appendix Table 1.** Summary of linear models to detect the effect of zone (sympatry vs. allopatry) on  $F_{IS}$  and selfing rate ( $s$ ) inferred from genetic variation.

	<b>F-statistic</b>	<b>df</b>	<b>p-value</b>
<b><math>F_{IS}</math></b>	0.2276	10	0.644
<b>Selfing rate (<math>s</math>) in nature</b>	0.2869	10	0.604

**Appendix Table 2.** Per population estimate of  $F_{IS}$  and selfing rate ( $s$ ) inferred from genetic variation.

Population	Zone	$F_{IS}$	$s$	No. of individuals
659	sympatry	0.331	0.4973704	6
668	sympatry	0.0376	0.07247494	8
761	sympatry	0.3349	0.50176043	20
768	sympatry	0.3992	0.57061178	21
790	sympatry	0.3637	0.53340177	17
798	sympatry	0.3717	0.54195524	14
	<b>Mean</b>	<b>0.30635</b>	<b>0.45292909</b>	
				<b>86</b>
651	allopatry	0.2643	0.41809697	8
652	allopatry	0.4216	0.5931345	25
653	allopatry	0.3681	0.53811856	9
666	allopatry	0.2789	0.43615607	4
791	allopatry	0.2408	0.38813669	4
793	allopatry	0.4522	0.62277923	18
	<b>Mean</b>	<b>0.33765</b>	<b>0.49940367</b>	
				<b>68</b>
	<b>Total</b>			<b>154</b>