

Opinion piece



Cite this article: Zimmer C, Jimeno B, Martin LB. 2024 HPA flexibility and *FKBP5*: promising physiological targets for conservation. *Phil. Trans. R. Soc. B* **379**: 20220512. <https://doi.org/10.1098/rstb.2022.0512>

Received: 28 July 2023

Accepted: 22 October 2023

One contribution of 14 to a theme issue
'Endocrine responses to environmental
variation: conceptual approaches and recent
developments'.

Subject Areas:

physiology, evolution, ecology

Keywords:

stress, urbanization, wildlife, coping,
corticosterone, hormone

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HPA flexibility and *FKBP5*: promising physiological targets for conservation

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Hypothalamic–pituitary–adrenal axis (HPA) flexibility is an emerging concept recognizing that individuals that will cope best with stressors will probably be those using their hormones in the most adaptive way. The HPA flexibility concept considers glucocorticoids as molecules that convey information about the environment from the brain to the body so that the organismal phenotype comes to complement prevailing conditions. In this context, *FKBP5* protein appears to set the extent to which circulating glucocorticoid concentrations can vary within and across stressors. Thus, *FKBP5* expression, and the HPA flexibility it causes, seem to represent an individual's ability to regulate its hormones to orchestrate organismal responses to stressors. As *FKBP5* expression can also be easily measured in blood, it could be a worthy target of conservation-oriented research attention. We first review the known and likely roles of HPA flexibility and *FKBP5* in wildlife. We then describe putative genetic, environmental and epigenetic causes of variation in HPA flexibility and *FKBP5* expression among and within individuals. Finally, we hypothesize how HPA flexibility and *FKBP5* expression should affect organismal fitness and hence population viability in response to human-induced rapid environmental changes, particularly urbanization.

This article is part of the theme issue 'Endocrine responses to environmental variation: conceptual approaches and recent developments'.

1. What is HPA flexibility?

Organisms must cope with variety of unpredictable challenges in both natural and human-modified environments. Critical to responding to these challenges appropriately is the ability of an individual to adjust its phenotype to current or impending conditions [1,2]. In vertebrates, one hormonal system is exceptionally important to such adjustments, the hypothalamic–pituitary–adrenal (HPA) axis. The HPA axis enables organisms to respond to many factors but especially stressors (i.e. unpredictable or uncontrollable stimuli in the external and internal environments that threaten homeostasis), primarily via glucocorticoids (GCs) [3,4]. Circulating GCs mediate homeostasis and stress responses via a two-tiered receptor system [3,5]. Moderate daily and seasonally rhythmic variations in concentrations (i.e. baseline variation) are associated with changes in energy metabolism and behavioural activity and regulated largely by mineralocorticoid receptors (MRs). By contrast, rapid (i.e. within minutes) and larger increases in concentrations (i.e. stress responses), usually coincident with exposure to stressors, are regulated by glucocorticoid receptors (GRs) [3–5]. Surges like these are quickly (within minutes to hours) followed by decreases to pre-stressor concentrations (i.e. via negative feedback of the hormones on central GRs), an equally critical change, as sustained elevations of GCs can diminish health and fitness.

Whereas much attention has been devoted to understanding variation in GC concentrations in wildlife, no existing framework has yet proposed how

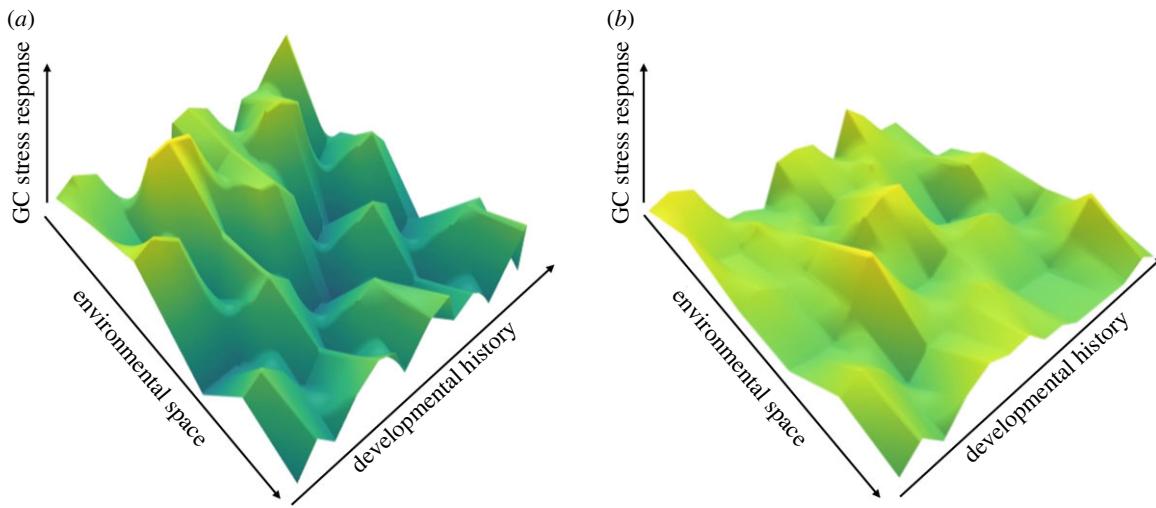


Figure 1. Examples of individuals with high (a) and low (b) HPA flexibility. The rugosity of the landscape depicts HPA flexibility, the variety of endocrine responses available to an organism over its life as a function of the prevailing environmental conditions, physiological state, and inherited factors (genotype and epigenetic effects). High HPA flexibility individuals should be better able to match their phenotypes to many more situations. By contrast, low HPA flexibility individuals should be less able to express an appropriate phenotype, at least in most contexts.

we might measure the trait central to the functional roles of glucocorticoids in the context of responses to stressors: the *ability of an individual to maintain and recover homeostasis*, what we and others have called endocrine or HPA flexibility. We know a lot about how glucocorticoids vary, but how this physiological variation *causes* variation in individual fitness is still obscure. Two ideas (i.e. allostasis and reactive scope) have generated a lot of research attention, but neither has yet resolved the roles of GCs in health nor conservation-related issues. Previously, we argued that these shortcomings derive from a lack of consideration of how GCs instantiate and convey *information* about stressors from the brain to the rest of the body [6,7]. Instead of trying to measure energy gains and losses and relating those changes to hormones (i.e. allostasis) or studying variation in mediator concentrations relative to presumably adaptive baselines, we could instead study hormones as difference-makers (i.e. the propensity of GCs to induce phenotypic change).

It is beyond the scope of this paper to detail the value of an information-based perspective for glucocorticoid regulation in the context of stress. However, our advocacy for HPA flexibility here is derived from this information-based mindset and is based on a few central tenets. First, our framework emphasizes that every organism will experience and need to resolve many kinds of stressors across its lifetime. Subsequently, one or a few concentration measures will be unlikely to capture the complex manner by which hormonal variation affects individual fitness. Sometimes one or a few plasma GC concentration measures predict variation in fitness, but examples are rare [8]. What we probably need to measure is the *ability of an individual animal to regulate its hormones*, in other words, HPA flexibility. Second, stressors vary in magnitude and type [6], so the relative fitness costs and benefits of GCs in stress responses will depend on the life-history stage, physiological state, and prior experience of an individual. The fittest individuals should be those that can most appropriately use their hormones to adjust the phenotype when the need arises, not those with high or low concentrations at various points in time. Third, circulating hormones are but one facet of how the phenotype is altered endocrinologically [1,9]. Salient information about stressors

cannot just reside in hormone concentrations; substantial information must also reside in receptors, metabolic enzymes, and other factors that determine the outcomes of glucocorticoid responses to stressors [10]. In this light, then, the body and brain should change together over the lifetime [7]; tissues should learn from experience (i.e. GC exposure) in such a way that historical information about the adversity of the environment is encoded into HPA axis regulatory components [6]. GCs are thus best understood as info-chemicals [11], factors that help individuals construct their phenotypes and conform to the environment as best they can. How individuals *regulate* GCs across stressors, then, represents their different propensities to instantiate information in their genomes and their cells [12]. It is this propensity that we call HPA flexibility.

For years, related ideas have been percolating in eco-evolutionary endocrinology [2,6,7,13–15] (and other papers in this theme issue). Most researchers, like us, have defined HPA flexibility as the *capacity of an individual to modify its HPA axis in response to stressors across multiple contexts* or something similar. Labs have measured HPA flexibility differently, but the consensus is that because GCs can only have phenotypic effects *after* they bind receptors and/or those hormone-receptor complexes bind genome-response elements [3,6], a few concentration measurements will be insufficient to describe HPA flexibility [9]. In figure 1, we depict two extremes of HPA flexibility that one might find in a natural population of vertebrates. Each landscape in each panel depicts HPA flexibility for an individual, the variety of GC responses to stressors possible for that individual over the course of its life. We expect that the rugosity of each landscape is set by its genetic and epigenetic makeup in the context of environmental conditions at any point in time (see below). One organism (figure 1a) has high HPA flexibility, a GC regulatory capacity suitable to many types of stress responses, whereas another organism (figure 1b) does not. Functionally, high HPA flexibility individuals should be able to achieve the most appropriate phenotype for the greatest diversity of environmental conditions; in having the most rugged GC landscapes (figure 1a), they should be able to recruit more adaptive endocrine response given the type and magnitude of the stressor in the context

of the current environment but also previous experience. Low HPA flexibility individuals, by contrast, should be able to realize fewer HPA phenotypes, limiting their ability to match their organismal phenotype to the environment (figure 1b). Such inflexibility could be wholly genetic, but it could also be environmentally induced, driven by stressors experienced during development. Nevertheless, as for GCs, fitness benefits of HPA flexibility are likely to be context-dependent. High HPA flexibility may not be advantageous in all environmental contexts because of the costs associated with high level of flexibility (e.g. information costs, search times for the optimal response in a complex landscape). Indeed, as GCs are pleiotropic hormones affecting multiple aspects of individuals' physiology and behaviour, in some environmental contexts it may better to be less flexible to avoid inappropriate responses. Thus, high HPA flexibility could be disadvantageous when living in a benign environment.

Below (§§4c,d), we discuss the promise of an HPA flexibility-based framework for understanding how individuals and hence populations of wild vertebrates will cope with human-induced rapid environmental change (HIREC) [16]. We first summarize how one could effectively measure endocrine flexibility (EF) in any vertebrate species. We then discuss the role of *FKBP5*, the gene encoding FK506 binding protein 51, as a promising and much more simply measured proxy for HPA flexibility [17]. We close by proposing hypotheses and a research plan to capitalize on HPA flexibility and especially *FKBP5* to mitigate effects of anthropogenic change. We predict that high HPA flexibility taxa are more apt to exploit and adjust to human-modified conditions, especially cities.

2. Measuring HPA flexibility

There is as yet no consensus approach to measure HPA flexibility [2,18], but the most popular methods involve descriptions of hormonal reaction norms. Typically, GC reaction norms are measured as the slopes of the relationships between hormone concentrations and environmental context for an individual animal [19–21]. While this approach has been insightful [22], we are sceptical of its suitability to describe HPA flexibility. GC concentrations measured prior to (i.e. baseline), during (i.e. post-stressor), and after (i.e. negative feedback) exposure to stressors will probably have co-evolved in such a way that they should be studied as a unit, a single physiological response. The current practice of estimating reaction norms for baseline, post-stressor, and post negative feedback concentrations, separately, unjustifiably analyses these measurements independently [7]. Individuals would be unlikely to have evolved to release excessive GCs into circulation if they lacked the ability to engage a robust negative feedback response [23–25]. HPA flexibility described as concentration reaction norms therefore does focus on the trait on which natural selection has acted, the *ability to regulate* the hormone [26]. As above, HPA flexibility is most sensibly understood as a landscape of GC responses available to an individual at any given time, a kind of endocrine hypervolume (i.e. the rugosity of the landscape in figure 1). Reaction norms for concentrations across temperature, social context or some other environmental gradient might resemble HPA flexibility as in figure 1, but this assumption must be substantiated empirically.

We recently proposed a fairly simple method to describe HPA flexibility, the square root of the mean squared differences (RMSSD) of sequential glucocorticoid stress responses measured in one animal [27]. Our rationale was that the more distinct sequential stress responses (and resolutions thereof) were in an individual across contexts, the higher HPA flexibility that individual must have. To test this idea, we first quantified RMSSD, developed initially to describe heart rate flexibility, using the mean glucocorticoid concentration across four stress responses of house sparrows (*Passer domesticus*); each stress response was described according to convention (i.e. baseline, post-stressor and after negative feedback activation concentrations for each individual) in stress responses measured a week apart [17]. We found that birds varied quite extensively in HPA flexibility; some had very high values (RMSSD = 24) whereas others were comparatively inflexible (RMSSD = 4). More importantly with respect to the presumed adaptiveness of HPA flexibility, birds with high RMSSD values were also more *behaviourally* flexible than birds with low RMSSD values [17]. More sophisticated and perhaps more accurate forms of descriptors of stress responses could have been used (e.g. area under the concentration curve, AUC) as well as more powerful statistical efforts (i.e. double-hierarchical general linear models, DH-GLMs) [28], but for our purposes, the simplistic approach was effective. Individuals varied quite a bit in HPA flexibility, and HPA flexibility was related in the expected direction to a presumably adaptive behaviour.

3. *FKBP5*: a simple-to-describe proxy of HPA flexibility

Accurately quantifying HPA flexibility as RMSSD will always require several, repeated hormone measurements in the same individuals in different contexts. For many wild species, especially from threatened populations, those of small body size, or those particularly difficult to maintain in captivity, such data will be hard to collect. Consequently, we advocate that, instead, *FKBP5*, a co-chaperone in the GR complex regulating GR function and activity [1], be the focus of study. Extensive biomedical research shows that central *FKBP5* expression in the few vertebrates studied so far increases within about 1 h in response to elevated GC concentrations. These elevated local levels then create an intracellular, ultra-short negative feedback loop, regulating GR affinity for GCs [29,30]. At organismal level then, the more *FKBP5* that an individual expresses, the more its GR resistance is increased, lowering negative feedback efficacy of GCs on brain regions, and hence compromising adaptive GC regulation [29,31]. In the context of figure 1, then, *FKBP5* probably sets the rugosity of the landscape, giving some individuals but not others a propensity to manifest diverse endocrine responses contingent on historical and current context.

To date, almost all data supporting this possibility have come from humans and laboratory rodents [29,31,32]. Nevertheless, there is no obvious reason that these relationships would not apply to most vertebrate wildlife. In domesticated mice, low *FKBP5* expression underlies an attenuated stress response and increased negative feedback efficacy associated with enhanced stress-coping behaviour (i.e. exploration) [33,34]. In the above study of house sparrows, HPA flexibility (measured as RMSSD) was inversely correlated to *FKBP5*

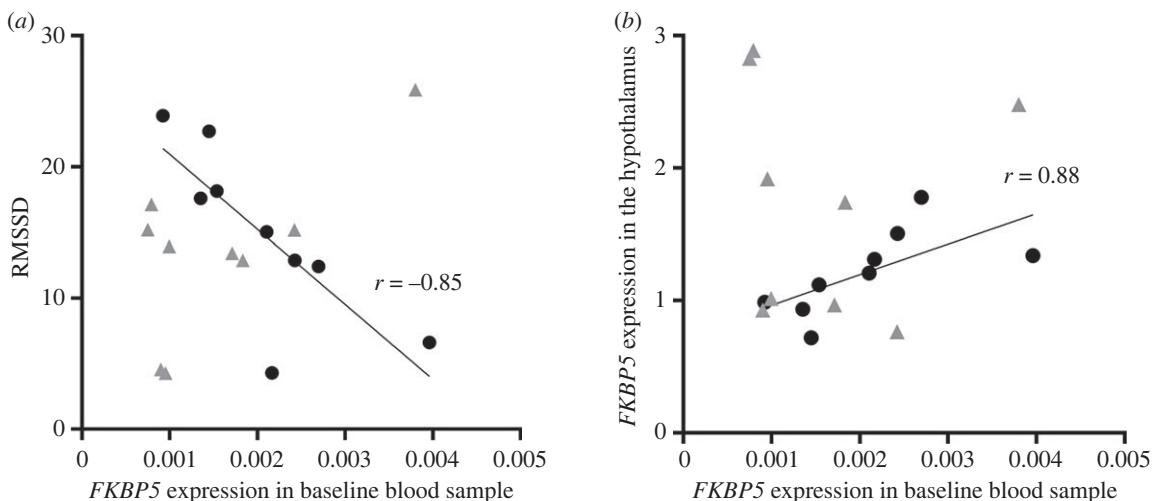


Figure 2. *FKBP5* expression as a biomarker of HPA flexibility in house sparrows (*Passer domesticus*). (a) Relationships between the square root of the mean squared differences of successive stress series (RMSSD) and *FKBP5* relative expression in baseline blood samples at capture in adult (black circle) and juvenile birds (grey triangle). (b) Relationship between *FKBP5* expression in the hypothalamus and the baseline blood samples at capture in adults (black circles) and juveniles (grey triangles). Regression lines were only calculated for adults. Reprinted with permission from [17].

expression in the hypothalamus [17]. In the same birds, low *FKBP5* expression in the hippocampus was also associated with higher level of exploration in a novel environment. Perhaps most promising to the utility of *FKBP5* in wildlife research, *FKBP5* expression in baseline blood samples from house sparrows was correlated with hypothalamic expression (figure 2). As this relationship also seems to exist in laboratory mice [31], scientists interested in studying HPA flexibility in threatened, small, or hard to keep captive wildlife might need only measure *FKBP5* in blood samples once. If such blood–brain relationships are confirmed in other species, they could become a very valuable marker of stress resilience in wildlife. It is possible that the level of *FKBP5* expression will not directly relate between blood and brain, but even in these cases, plasticity in *FKBP5* expression in response to external and internal factors might [9]. Indeed, in laboratory mice and house sparrows, *FKBP5* expression increased similarly in brain and peripheral tissues in response to experimental stressors and GC treatments [27,31,35,36]. As establishing this blood–brain correlation will not be possible in a threatened species, a closely related species of no or little conservation concern could be used as a proxy.

4. Causes of variation in endocrine flexibility and *FKBP5*

Differences in HPA flexibility may manifest through a variety of mechanisms besides *FKBP5*, including hormone levels, receptor abundance and affinity [37], or carrier proteins [38,39]. We expect *FKBP5* to be central to HPA flexibility, but much research lies ahead. If *FKBP5* largely determines HPA flexibility, it should be a major focus of natural selection [40]. It should therefore tend to be repeatable and heritable, and indeed, in house sparrows, *FKBP5* expression was repeatable ($R = 0.45 \pm 0.18$ [0.04–0.71], $p = 0.003$) in measurements in blood made weeks apart [17].

(a) Among-individuals variation

Studies of *FKBP5* expression in wild animal populations are presently rare, but existing research in domesticated rodents

and humans gives quite a lot of perspective about the causes and extent of individual variation. For instance, functional genetic polymorphisms for *FKBP5* (as well as MR and GR) exist in both humans and laboratory rodents, and partly explain variation among individuals in their sensitivity to GCs [41]. In humans, impaired GC receptor function also appears to underlie the development of many metabolic diseases [41,42] and psychiatric disorders [43]. *FKBP5* seems a key mechanism orchestrating these outcomes; upregulation of *FKBP5* results in stronger GR resistance, weaker HPA axis negative feedback, and sustained and high circulating GC concentrations [29,44,45]. A haplotype of *FKBP5*, characterized by high *FKBP5* expression in response to GR activation, has also been associated with differences in the risk of post-traumatic stress disorder [46–48], bipolar disorder [49,50] and high anxiety [34,51].

In many species, molecular epigenetic processes will probably also affect individual variation in HPA flexibility. DNA methylation, which largely occurs at CpG motifs in gene regulatory regions, allows the integration of environmental signals into the genome to affect subsequent gene expression [52]. These changes alter the accessibility of transcription factors to binding factors, including *FKBP5* [53], probably sculpting the chromatin such that its structure affects future endocrine responses. In the adaptive sense, this sculpting could better mould endocrine responses to prevailing and/or past conditions; in the maladaptive sense, the epigenetic marks could instead prevent the organismal phenotype from tracking salient environmental change [54]. As of now, we know little about how methylation and other marks on the DNA affect HPA flexibility, much less whether any are heritable [55]. Further, whereas there are many examples of enduring effects of early-life stressors on later-life GC regulation, whether these effects apply to HPA flexibility or involve *FKBP5* remain obscure. Early-life environmental conditions alter specific regulatory elements of HPA flexibility [56,57], and some such effects are epigenetically mediated [58,59]. Previous research in vertebrates also suggests that GR is a primary target for long-term epigenetic programming of the HPA axis. Epigenetic changes to GR can enduringly modify HPA axis regulation depending on the natal environments experienced by individuals

[60–65]. Again, research investigating these patterns in wildlife are scarce, although recent work has reported comparable effects of early-life adversity on GR methylation and expression in both captive and free-living birds [66,67].

Given the function of *FKBP5* and the importance of its epigenetic regulation in biomedical contexts, molecular epigenetic factors are likely to be major drivers of within-individual variation in HPA flexibility in nature, too. In humans, DNA methylation of *FKBP5* affects individual risk of developing psychiatric disorders; exposure to adverse conditions during childhood, but not during adulthood, was associated with low methylation at specific CpG sites in the *FKBP5* gene in individuals with a certain genetic polymorphism (i.e. the risk T allele [68]). Long-lasting, lower methylation in *FKBP5* in response to adverse conditions during early life also seems to result from sustained GR activation in early life [29,69]. Based on these two studies, *FKBP5* methylation status could amplify or limit HPA flexibility depending on *actual* environmental adversity experienced by an individual [1]. Transgenerational effects of adverse conditions on DNA methylation of *FKBP5* have also been reported in humans [70] and rats [59]. In the latter study, rats with low *FKBP5* DNA methylation had high *FKBP5* expression and sustained HPA activation [59].

(b) Within-individual variation

Within-individual variation in HPA flexibility and *FKBP5* expression could be extensive and driven by a variety of exogenous and endogenous environmental forces. Below, we outline four important contexts expected to affect HPA flexibility and *FKBP5* and make some predictions about both to motivate research efforts. We also recognize that by reviewing what is known or expected about within-individual variation in HPA flexibility, we emphasize that the landscapes in figure 1 could be in fact quite plastic [71,72]. If so, the study of these landscapes will be quite challenging (i.e. in terms of statistical power) [73]. Still, this complexity should not stop our investigations of them. We must instead apply creative methods (e.g. response-surface regression) to describing and understanding how landscape rugosity shapes and is shaped by experience [74].

(i) Seasonal

Seasonal variation in HPA regulatory components is well known in wildlife and domesticated organisms, and seasonal differences in HPA flexibility and *FKBP5* expression are obviously expected. In free-living birds, multiple components of the HPA axis change over the year (i.e. membrane and cytosolic receptor levels, plasma GC levels, corticosteroid binding globulins; [75]). This variation has been linked to variation in behavioural and physiological responses of individuals to environmental challenges, consistent with expected functions of GCs for various seasonal [76] and daily [77] activities. One would expect HPA flexibility to be highest and *FKBP5* to be lowest in seasons where stressors are least predictable and/or most consequential for fitness.

(ii) Resource availability

Very few studies have addressed how resource availability might affect HPA flexibility, although there is reason to expect that both food quantity and quality will be important. For instance, birds facing high foraging costs expressed less

GR compared with birds facing low foraging costs; these patterns were also associated with changes in GC responses to a standardized stressor (i.e. higher baseline levels and weaker negative feedback response [67]). Consistently, too, food-restricted house sparrows increased baseline GC levels as their body masses decreased, but individuals differed in GC responses to food restriction [78]. In laboratory mice, after 24 h food deprivation, *FKBP5* expression was dramatically increased in many brain regions [79]. Altogether, higher foraging costs and/or lower food availability should lead to high *FKBP5* expression and low HPA flexibility.

(iii) Social context

In both social and non-social taxa, the frequency and type of interactions with conspecifics can affect HPA axis activity [80]. Similar social conditions potentially also drive differences in HPA flexibility and *FKBP5* expression. To date, research in this area has almost exclusively focused on plasma GCs and responses to single stressors, yielding contrasting results [81–83]. To our knowledge, only one study has measured gene expression in a social stressor context, but GR was measured, not *FKBP5* [84]. In this study, social information from food-restricted individuals reduced GR expression in HPA tissues of red crossbills (*Loxia curvirostra*). These changes could reflect low GR sensitivity and potentially lower EF, but no specific data yet exist. One would expect that individuals with high HPA flexibility (and low *FKBP5*) would cope most effectively in novel social contexts. However, given the multitude of diverse costs and benefits of social interactions, the social roles of *FKBP5* and HPA flexibility could be quite complex.

(iv) Age

GC secretion often increases with age, but it remains unknown whether these changes are indicative of adaptive HPA flexibility or simply senescence. In humans, older individuals tend to have weak negative feedback and high baseline GCs [85,86], the latter condition being associated with higher risk for many non-infectious diseases (e.g. diabetes, hypertension, cardiovascular diseases) as well as sleep deterioration and depression [85,87]. Interestingly, Blair *et al.* [88] showed that *FKBP5* expression and protein levels increased with age, which promoted the pathogenesis of Alzheimer's disease. Patients with Alzheimer's disease also had higher levels of *FKBP5* in the brain as compared with controls. Although further research is needed to confirm the consistency and generality of these patterns, the above evidence suggests that all increases in *FKBP5* might not reflect adaptive change. We do not know much about changes in GC secretion in aged wildlife. In birds, baseline and stress-induced GCs tend to decrease with age [89–93], but we know almost nothing about HPA axis regulation and flexibility in old individuals. Studies directed at age-related changes up to and around the time of reproductive maturation would be most likely to reveal adaptive, age-dependent variation in HPA flexibility and *FKBP5* in wildlife.

(c) Eco-evolutionary implications of HPA flexibility

Although we presently know very little about HPA flexibility and *FKBP5* in wild animals, enough relevant literature exists to propose a promising research programme in the context of anthropogenic change. Whether GC regulation affects the

ability of wildlife to cope well with anthropogenic change is still unclear [94–99], but a focus on HPA flexibility, instead of GC concentrations, reframes the scope of the problem in two hopefully productive ways. First, it shifts efforts to describe regulatory control of the hormones [6,7,18]. It is the ability to regulate GCs that should affect fitness [100]. Second, it views GCs as physiological sculptors [101,102], info-molecules that help the organism become what environmental signals convey that it should be [6,103–105], not simple proxies of stress. GCs are among the most pleiotropic molecules circulating in vertebrates [4,105–108], so traits that describe the propensity of individuals to regulate them are more apt to illuminate how particular taxa will endure or suffer from anthropogenic change [6,96,108,109].

Such a framework for understanding physiological responses to stressors in wildlife, based on HPA flexibility, resonates well with the ‘morphology, performance, and fitness’ paradigm so powerful in other subdisciplines of organismal biology [110]. We have previously focused on hormone concentrations, assuming them to be indicative of stress because it is comparatively easy to measure hormones, even in fur, feathers, faeces or scales. However, what we always needed to understand was the ability of an animal to use its hormones adaptively. In this light, we summarize below what is known and expected about HPA flexibility in animals occurring in cities, areas where HIREC is particularly concentrated [111]. We focus on birds because GCs in this taxon are so well studied and are also common exploiters and victims of HIREC [112]. However, our rationale likely applies to all vertebrates including fish, amphibians, and other aquatic species. Further, although we focus on urbanization, we expect that our ideas apply to other forms of HIREC, including climate-driven and more directly human-caused geographical range shifts, but also local forms of habitat degradation, including light, noise and toxicant pollution [113].

(d) Life in the city

For well over a decade, extensive efforts have been made to understand urbanization and GCs in wildlife [114]. Whereas many studies have revealed differences in GC concentrations between urban and non-urban organisms (reviewed in [98]), strong support for GC dysregulation as a causative force driving conservation concerns in urban wildlife has been lacking [96]. Iglesias-Carrasco *et al.* [115], for instance, found no effects of urbanization on circulating GC concentrations among 27 avian species across 34 studies, even when accounting for many putative modifiers of urbanization effects (e.g. sex, season, life stage, taxon, size of the city, etc). By contrast, Injaian *et al.* [98] discovered that only one aspect of urbanization (i.e. noise pollution but not light pollution of an urbanization index) was related to GC concentrations in birds and reptiles. Even, noise pollution effects on GCs were revealed only after an urban adaptability score for species was included as a predictive factor in models. Because this latter project involved HormoneBase [116], a very large compilation of wildlife endocrine data, the absence of urbanization effects on GCs in this particular study suggests that either GCs truly play no role in adaptation or adjustment to cities or, more likely, efforts to discern how GCs enable or prevent populations from mitigating urban stressors will require more sophisticated approaches. A few studies have moved in this more sophisticated direction,

focusing on specific facets of urbanization that might affect GCs. For example, chronic traffic noise was revealed to alter GC responses to physical restraint stressors in tree swallows (*Tachycineta bicolor*) [117]. In another study, artificial (white) light at night exposure was found to change GC concentrations in wild great tits (*Parus major*) [118]. Although these studies and others imply that GC regulation probably affects successful coping (or not) with particular urban stressors, generalities are very few [119]. We propose that a shift to HPA flexibility (and/or *FKBP5*) will be useful and might even reveal some actionable, broad patterns.

Before we propose such a study plan, some simplifying assumptions are necessary. First, we agree with Deviche and colleagues [95] that attending better to the dimensions of cities apt to be acting as stressors to wildlife will augment progress. Not all cities will have the same stressors, just as not all urban stressors will have the same implications for GC regulation. Likewise, non-urban sites are not necessarily an appropriate foil for urban sites, as natural places are so heterogeneous that the type and extent of environmental variation in non-urban sites would make few non-urban sites appropriate comparators. We also agree with Injaian *et al.* [98] that the observations that some species thrive whereas others avoid or even suffer in cities [120] will be important to consider. Perhaps because some species evolved in environments resembling cities, or simply evolved to be generalists, a few taxa will do quite well in urban contexts. The roles of GCs in coping with urban conditions might be general, but outcomes could vary depending on the species studied, at least whether the species is an urban avoider or urban exploiter.

Finally, cities vary extensively on several continua (i.e. size, age, proximity to natural areas, greenspace, etc.), but some broad trends in stressors very likely exist. Compared with non-urban areas, for instance, urban ones will often be more stable because of deliberate, ongoing habitat modifications by humans [95,121]. From this perspective, predictability of stressors should be higher in most urban versus non-urban areas (figure 3a). The times and places that organisms are exposed to stressors, on both short (daily) and long (seasonal) time scales, should be more knowable than for non-urban areas. City-dwelling animals will not always have the capacity to mitigate such conditions, but some organisms might avoid some stress simply by learning when and where stressors are likely to occur. Relatedly, in cities, the evolutionary novelty of stressors will typically be higher (figure 3a). Adverse factors such as noise, light, endocrine-disrupting chemicals, and other stressors with which ancestral populations will have had little to no experience will abound in urban areas. Of course, these factors might be common in some non-urban areas, too, but on balance, the collective of novel stressors should be higher inside than outside cities [122]. Even novel degrees of natural stressors such as food availability (i.e. more low-quality but more predictable food [123]) and social conditions (i.e. higher conspecific densities over longer portions of the year) will differ between urban and non-urban places [124]. In cities, too, interactions among novel and natural stressors should be common [94], perhaps expanding combinatorially the scope of stressors that wildlife will encounter. From this perspective, diverse endocrine responses would presumably foster diverse organismal phenotypic responses. Cities should thus tend to contain individuals with high HPA

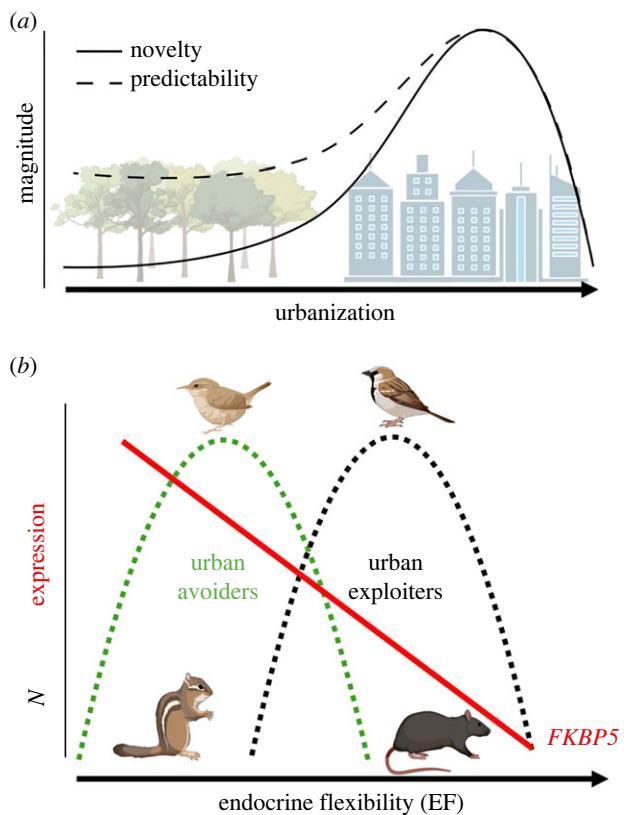


Figure 3. Stressors and organismal coping capacities for stressors along urban gradients. (a) Expected variation in stressor novelty and predictability along an urbanization gradient. Stressor novelty (in an evolutionary sense) should be higher in cities relative to surrounding natural areas. Stressor predictability, too, should be higher in cities given human amelioration of natural environmental change. However, even in non-urban areas, stressor predictability should not fall to zero but instead depend on local climate. (b) Predicted variation in HPA flexibility and *FKBP5* expression in city-dwelling organisms. Generally, individuals, populations or species with high HPA flexibility should fare best in cities, if the assumptions in (a) are valid. However, urban avoiders and exploiters should differ such that HPA flexibility should be low and *FKBP5* high in avoiders compared with exploiters. Created with Biorender.com.

flexibility and low *FKBP5*. On the other hand, if stressor predictability is more consequential than stressor novelty, fewer forms of GC responses (i.e. lower HPA flexibility) could instead be favoured in cities. These possibilities as yet await testing, but all things considered, we presently expect that high HPA flexibility and low *FKBP5* will largely be favoured in cities, even if stressors are relatively predictable there (figure 3b).

These propositions are offered solely as a reasonable starting place for investigation, and any productive future research will attempt to consider, empirically, stressor gradients in cities and organismal and endocrine responses to them. All such studies will also benefit by recognizing that species vary in how they cope with cities demographically; some species endure city conditions well (tolerators), others do not (avoiders), and still others seemingly thrive in their presence (i.e. preferers, exploiters and adapters) [120,125,126]. In figure 3b, we intermingle our expectations about urban stressors in figure 3a with known variation in ecological responses of species to anthropogenic effects. We favour the urban tolerance framework of Callaghan and colleagues [120] because it is a continuous form of responsiveness to city conditions and

derived from more than 100 million observations of 338 avian species. Among that subset of birds, 75% of species had negative urban tolerance and 25% had positive tolerance, categories the authors named urban *avoiders* and *exploiters*, respectively [120]. The take-home message from that work is: most species do not fare well in cities, but quite a few do. We expect that HPA flexibility differs between these two categories of animals, with the highest forms of HPA flexibility and the lowest levels of *FKBP5* expression found in the exploiters (figure 3b). Of course, we should also moderate the above predictions with a few caveats. Some urban exploiters can also thrive outside cities, and those populations might have appreciably lower HPA flexibility than urban ones, especially if stressors in a specific city do not match patterns described in figure 3a. Parsimony also suggests that *FKBP5* expression should track HPA flexibility consistently for each species, with higher *FKBP5* expression related in the same manner to EF across species (figure 3b). This proposition warrants investigation, though, as the relationship between HPA flexibility and *FKBP5* has as yet been studied in very few species. Further, the predictions in figure 3 are not intended to capture possible allelic variants of or epigenetic effects on *FKBP5*, partly because we presently know nothing about them in wildlife.

Despite the above open issues, there are many insightful opportunities implicit in figure 3. For instance, one could assess relationships between HPA flexibility and fitness over time in urban populations to resolve how populations adapt or at least cope endocrinologically with urban conditions. One might also measure *FKBP5* (and/or HPA flexibility) in several cities or parts of large cities to implicate the specific aspects of urbanization most consequential to colonization and/or persistence. Finally, one could survey HPA flexibility or *FKBP5* broadly across taxa, seeking to identify organisms most likely to act as exploiters. Extensive efforts and abundant funds have been devoted to identifying pest biomarkers; perhaps organisms with low *FKBP5* are the ones resource managers most need to find and control.

Before closing, we must briefly mention that urbanization is not the only dimension of HIREC for which HPA flexibility warrants study [127]. Climate-driven and human-facilitated range expansions, too, should be affected by HPA flexibility and *FKBP5*. Just as with urbanization, GC regulation is justifiably expected to be involved in range expansions [128], and some data support such relationships. House sparrows at the vanguard of range expansions across Kenya [129,130] and Senegal [131,132], for instance, regulated GCs quite differently from birds from the core of populations. A similar pattern was revealed in a southward-expanding tree swallow population relative to resident populations [133]. In cane toads (*Rhinella marina*) [134–136] and Egyptian mongooses (*Herpestes ichneumon*) [137], GCs varied with range expansion but in a manner different from that in the above passerines. Broadly, across more than 100 bird and reptile species, variation in GC concentrations was unrelated to where samples were collected in a species' range [97]. Another comparative study on *Peromyscus* mice likewise found no intelligible patterns when comparing GC concentrations between a few broadly and narrowly distributed species [138].

Whereas the role of GCs in range expansions and geographic distributions will probably be nuanced, future studies focused specifically on HPA flexibility might be quite insightful. Colonizers or individuals enduring

suboptimal abiotic or biotic conditions at range margins might be more active, bolder and/or more exploratory (and therefore potentially more likely to disperse), traits that all are related to GC regulation [139]. Just how GCs cause this behavioural variation (i.e. how GCs encode information) might differ among species [108], and this possibility should be investigated. Still, we expect that HPA flexibility will be quite high and *FKBP5* expression will be low at expanding range edges or indeed in any environments where stressors are novel and numerous.

5. Looking forward

Right now, because HIREC is such a problem for our health and wildlife welfare, we need new measurable targets, and these two factors, HPA flexibility and *FKBP5*, could be as valuable to conservation as they have been to medicine. Experts have long agreed that investigations of how wildlife populations respond physiologically to HIREC are important to management [140,141]. Likewise, scientists have long understood that organisms experiencing HIREC might provide valuable basic perspective into evolutionary change [142–144] and ecological impact (e.g. zoonosis spillover, extirpation of native populations by pests, etc.) [124,145–148]. We agree in both senses, but we also argue strongly that we must reduce our reliance on simplistic approaches (i.e. using one or a few measures of GCs, especially in inert tissues or faeces), and direct attention instead to traits like HPA flexibility and *FKBP5* [149]. These traits capture better how hormones encode information and hence enable the phenotype to be adjusted to the environment. Whether HIREC takes the form of urbanization, climate change, or range shifts, the organisms most adept at enduring these challenges will

tend to be the flexible ones [150]. Sometimes, flexibility will be mediated by *FKBP5*, but often other molecular capacitors of adaptive variation will be important, too [151,152]. Regardless of the specific mechanisms, as Callaghan *et al.* wrote [120, p. 411]: ‘a species’ adaptive capacity, caused by individual, population or species-level attributes, may be important for conservation since it is one component that can make a species vulnerable to environmental change.’ We think that HPA flexibility and *FKBP5* are such attributes, and not only will they be important for conservation purposes, but also they will help us comprehend better how endocrine systems function and evolve.

Data accessibility. This article has no additional data.

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors’ contributions. C.Z.: conceptualization, visualization, writing—original draft, writing—review and editing; B.J.: conceptualization, writing—original draft, writing—review and editing; L.B.M.: conceptualization, visualization, writing—original draft, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed herein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. L.B.M. acknowledges support from the National Science Foundation (grant nos 2027040 and 2110070). B.J. was funded by the European Union’s Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement no. 101027784.

Acknowledgements. L.B.M. thanks Vania Assis, Kailey McCain, Gabby Mansilla and Arryson Martin for feedback on an earlier draft. L.B.M. and C.Z. also thank Art Woods, Karl Friston, Michael Romero, Michaela Hau, Conor Taff, John Wingfield, Maren Vitousek, and many other colleagues for their constructive criticism and thoughts about endocrine flexibility. Lastly, all authors thank the special issue editors for the chance to be involved in this project.

References

1. Zimmer C, Hanson HE, Wildman DE, Uddin M, Martin LB. 2020 *FKBP5*: a key mediator of how vertebrates flexibly cope with adversity. *Bioscience* **70**, 1127–1138. (doi:10.1093/biosci/biaa114)
2. Taff CC, Vitousek MN. 2016 Endocrine flexibility: optimizing phenotypes in a dynamic world? *Trends Ecol. Evol.* **31**, 476–488. (doi:10.1016/j.tree.2016.03.005)
3. Sapolsky RM, Romero LM, Munck AU. 2000 How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr. Rev.* **21**, 55–89. (doi:10.1210/edrv.21.1.0389)
4. Hau M, Casagrande S, Ouyang JQ, Baugh AT. 2016 Glucocorticoid-mediated phenotypes in vertebrates: multilevel variation and evolution. *Adv. Stud. Behav.* **48**, 41–115. (doi:10.1016/bs.asb.2016.01.002)
5. Wingfield JC, Maney DL, Breuner CW, Jacobs JD, Lynn S, Ramenofsky M, Richardson RD. 1998 Ecological bases of hormone–behavior interactions: the ‘emergency life history stage’. *Am. Zool.* **38**, 191–206. (doi:10.1093/icb/38.1.191)
6. Zimmer C, Woods HA, Martin LB. 2022 Information theory in vertebrate stress physiology. *Trends Endocrinol. Metab.* **33**, 8–17. (doi:10.1016/j.tem.2021.10001)
7. Martin LB, Zimmer C. 2022 Endocrine flexibility. *J. Exp. Biol.* **225**, jeb.244646. (doi:10.1242/jeb.244646)
8. Schoenle LA, Zimmer C, Miller ET, Vitousek MN. 2021 Does variation in glucocorticoid concentrations predict fitness? A phylogenetic meta-analysis. *Gen. Comp. Endocrin.* **300**, 113611. (doi:10.1016/j.ygenc.2020.113611)
9. Jimeno B, Zimmer C. 2022 Glucocorticoid receptor expression as an integrative measure to assess glucocorticoid plasticity and efficiency in evolutionary endocrinology: a perspective. *Horm. Behav.* **145**, 105240. (doi:10.1016/j.yhbeh.2022.105240)
10. Romero LM, Beattie UK. 2022 Common myths of glucocorticoid function in ecology and conservation. *J. Exp. Zool. A* **337**, 7–14. (doi:10.1002/jez.2459)
11. Muller C, Caspers BA, Gadau J, Kaiser S. 2020 The power of infochemicals in mediating individualized niches. *Trends Ecol. Evol.* **35**, 981–989. (doi:10.1016/j.tree.2020.07.001)
12. Kultz D *et al.* 2013 New frontiers for organismal biology. *Bioscience* **63**, 464–471. (doi:10.1525/bio.2013.63.6.8)
13. Grindstaff JL, Beatty LF, Ambardar M, Luttbeg B. 2022 Integrating theoretical and empirical approaches for a robust understanding of endocrine flexibility. *J. Exp. Biol.* **225**, jeb.243408. (doi:10.1242/jeb.243408)
14. Bonier F, Martin PR, Moore IT, Wingfield JC. 2009 Do baseline glucocorticoids predict fitness? *Trends Ecol. Evol.* **24**, 634–642. (doi:10.1016/j.tree.2009.04.013)
15. Breuner CW, Patterson SH, Hahn TP. 2008 In search of relationships between the acute adrenocortical response and fitness. *Gen. Comp. Endocrin.* **157**, 288–295. (doi:10.1016/j.ygenc.2008.05.017)
16. Sih A. 2013 Understanding variation in behavioural responses to human-induced rapid environmental change: a conceptual overview. *Anim. Behav.* **85**, 1077–1088. (doi:10.1016/j.anbehav.2013.02.017)
17. Zimmer C, Hanson HE, Martin LB. 2021 *FKBP5* expression is related to HPA flexibility and the capacity to cope with stressors in female and male house sparrows. *Horm. Behav.* **135**, 105038. (doi:10.1016/j.yhbeh.2021.105038)
18. Lema SC, Kitano J. 2013 Hormones and phenotypic plasticity: implications for the evolution of

integrated adaptive phenotypes. *Curr. Zool.* **59**, 506–525. (doi:10.1093/czoolo/59.4.506)

19. Baldan D, Negash M, Ouyang JQ. 2021 Are individuals consistent? Endocrine reaction norms under different ecological challenges. *J. Exp. Biol.* **224**, jeb.240499. (doi:10.1242/jeb.240499)

20. Hau M, Deimel C, Moiron M. 2022 Great tits differ in glucocorticoid plasticity in response to spring temperature. *Proc. R. Soc. B* **289**, 20221235. (doi:10.1098/rspb.2022.1235)

21. Malkoc K, Mentesana L, Casagrande S, Hau M. 2022 Quantifying glucocorticoid plasticity using reaction norm approaches: there still is so much to discover! *Integr. Comp. Biol.* **62**, 58–70. (doi:10.1093/icb/icab196)

22. Taff C, Baldan D, Mentesana L, Ouyang J, Vitousek M, Hau M. 2024 Endocrine flexibility can facilitate or constrain the ability to cope with global change. *Phil. Trans. R. Soc. B* **379**, 20220502. (doi:10.1098/rstb.2022.0502)

23. Liebl AL, Shimizu T, Martin LB. 2013 Covariation among glucocorticoid regulatory elements varies seasonally in house sparrows. *Gen. Comp. Endocr.* **183**, 32–37. (doi:10.1016/j.ygenc.2012.11.021)

24. Zimmer C, Taff CC, Ardia DR, Ryan TA, Winkler DW, Vitousek MN. 2019 On again, off again: acute stress response and negative feedback together predict resilience to experimental challenges. *Funct. Ecol.* **33**, 619–628. (doi:10.1111/1365-2435.13281)

25. Zimmer C, Taff CC, Ardia DR, Rose AP, Aborn DA, Johnson LS, Vitousek MN. 2020 Environmental unpredictability shapes glucocorticoid regulation across populations of tree swallows. *Scient. Rep.* **10**, 13682. (doi:10.1038/s41598-020-70161-4)

26. McGlothlin JW, Whittaker DJ, Schrock SE, Gerlach NM, Jawor JM, Snajdr EA, Ketterson ED. 2010 Natural selection on testosterone production in a wild songbird population. *Am. Nat.* **175**, 687–701. (doi:10.1086/652469)

27. Park JE, Lee JY, Kang SH, Choi JH, Kim TY, So HS, Yoon IY. 2017 Heart rate variability of chronic posttraumatic stress disorder in the Korean veterans. *Psychiat. Res.* **255**, 72–77. (doi:10.1016/j.psychres.2017.05.011)

28. Cleasby IR, Nakagawa S, Schielzeth H. 2015 Quantifying the predictability of behaviour: statistical approaches for the study of between-individual variation in the within-individual variance. *Methods Ecol. Evol.* **6**, 27–37. (doi:10.1111/2041-210x.12281)

29. Zannas AS, Wiechmann T, Gassen NC, Binder EB. 2016 Gene–stress–epigenetic regulation of *FKBP5*: clinical and translational Implications. *Neuropsychopharmacology* **41**, 261–274. (doi:10.1038/npp.2015.235)

30. Rein T. 2016 FK506 binding protein 51 integrates pathways of adaptation: FKBP51 shapes the reactivity to environmental change. *Bioessays* **38**, 894–902. (doi:10.1002/bies.201600050)

31. Lee RS. 2016 Glucocorticoid-dependent epigenetic regulation of *Fkbp5*. In *Epigenetics and neuroendocrinology: clinical focus on psychiatry* (eds D Spengler, E Binder), pp. 97–114. Cham, Switzerland: Springer International Publishing. (doi:10.1007/978-3-319-24493-8_4)

32. Menke A *et al.* 2012 Dexamethasone stimulated gene expression in peripheral blood is a sensitive marker for glucocorticoid receptor resistance in depressed patients. *Neuropsychopharmacology* **37**, 1455–1464. (doi:10.1038/npp.2011.331)

33. Haušl AS *et al.* 2021 The co-chaperone Fkbp5 shapes the acute stress response in the paraventricular nucleus of the hypothalamus of male mice. *Mol. Psychiatr.* **26**, 3060–3076. (doi:10.1038/s41380-021-01044-x)

34. Touma C *et al.* 2011 FK506 Binding protein 5 shapes stress responsiveness: modulation of neuroendocrine reactivity and coping behavior. *Biol. Psychiat.* **70**, 928–936. (doi:10.1016/j.biopsych.2011.07.023)

35. Lotvedt P, Fallahshahroudi A, Bektic I, Altimiras J, Jensen P. 2017 Chicken domestication changes expression of stress-related genes in brain, pituitary and adrenals. *Neurobiol. Stress* **7**, 113–121. (doi:10.1016/j.ynstr.2017.08.002)

36. Rensel MA, Schlinger BA. 2020 The stressed brain: regional and stress-related corticosterone and stress-regulated gene expression in the adult zebra finch (*Taeniopygia guttata*). *J. Neuroendocr.* **32**, e12852. (doi:10.1111/jne.12852)

37. Banerjee SB, Arterberry AS, Fergus DJ, Adkins-Regan E. 2012 Deprivation of maternal care has long-lasting consequences for the hypothalamic–pituitary–adrenal axis of zebra finches. *Proc. R. Soc. B* **279**, 759–766. (doi:10.1098/rspb.2011.1265)

38. Breuner CW, Delehanty B, Boonstra R. 2013 Evaluating stress in natural populations of vertebrates: total CORT is not good enough. *Funct. Ecol.* **27**, 24–36. (doi:10.1111/1365-2435.12016)

39. Schoech SJ, Romero LM, Moore IT, Bonier F. 2013 Constraints, concerns and considerations about the necessity of estimating free glucocorticoid concentrations for field endocrine studies. *Funct. Ecol.* **27**, 1100–1106. (doi:10.1111/1365-2435.12142)

40. Bonier F, Martin PR. 2016 How can we estimate natural selection on endocrine traits? Lessons from evolutionary biology. *Proc. R. Soc. B* **283**, 20161887. (doi:10.1098/rspb.2016.1887)

41. Stevens A, Donn R, Ray D. 2004 Regulation of glucocorticoid receptor gamma (GR γ) by glucocorticoid receptor haplotype and glucocorticoid. *Clin. Endocr.* **61**, 327–331. (doi:10.1111/j.1365-2265.2004.02097.x)

42. Draper N *et al.* 2002 Association studies between microsatellite markers within the gene encoding human 11 β -hydroxysteroid dehydrogenase type 1 and body mass index, waist to hip ratio, and glucocorticoid metabolism. *J. Clin. Endocr. Metab.* **87**, 4984–4990. (doi:10.1210/jc.2001-011375)

43. Hartman RJG, Mokry M, Pasterkamp G, den Ruijter HM. 2021 Sex-dependent gene co-expression in the human body. *Scient. Rep.* **11**, 18758. (doi:10.1038/s41598-021-98059-9)

44. Zannas AS, Binder EB. 2014 Gene–environment interactions at the *FKBP5* locus: sensitive periods, mechanisms and pleiotropism. *Genes Brain Behav.* **13**, 25–37. (doi:10.1111/gbb.12104)

45. Matosin N, Halldorsdottir T, Binder EB. 2018 Understanding the molecular mechanisms underpinning gene by environment interactions in psychiatric disorders: the *FKBP5* model. *Biol. Psychiat.* **83**, 821–830. (doi:10.1016/j.biopsych.2018.01.021)

46. Binder EB *et al.* 2008 Association of *FKBP5* polymorphisms and childhood abuse with risk of posttraumatic stress disorder symptoms in adults. *J. Am. Med. Assoc.* **299**, 1291–1305. (doi:10.1001/jama.299.11.1291)

47. Yehuda R, Bierer LA. 2009 The relevance of epigenetics to PTSD: implications for the *DSM-V*. *J. Trauma. Stress* **22**, 427–434. (doi:10.1002/jts.20448)

48. Hawn SE *et al.* 2019 Examination of the effects of impulsivity and risk-taking propensity on alcohol use in OEF/OIF/OND veterans. *J. Milit. Veteran Family Health* **5**, 88–99. (doi:10.3138/jmvfh.2018-0002)

49. Willour VL *et al.* 2009 Family-based association of *FKBP5* in bipolar disorder. *Mol. Psychiatr.* **14**, 261–268. (doi:10.1038/sj.mp.4002141)

50. Seifuddin F, Pirooznia M, Judy JT, Goes FS, Potash JB, Zandi PP. 2013 Systematic review of genome-wide gene expression studies of bipolar disorder. *BMC Psychiatr.* **13**, 213. (doi:10.1186/1471-244x-13-213)

51. Criado-Marrero M, Gebru NT, Gould LA, Smith TM, Kim S, Blackburn RJ, Dickey CA, Blair LJ. 2019 Early life stress and high *FKBP5* interact to increase anxiety-like symptoms through altered AKT signaling in the dorsal hippocampus. *Int. J. Mol. Sci.* **20**, 2738. (doi:10.3390/ijms20112738)

52. Kilvits HJ, Hanson H, Schrey AW, Martin LB. 2017 Epigenetic potential as a mechanism of phenotypic plasticity in vertebrate range expansions. *Integr. Comp. Biol.* **57**, 385–395. (doi:10.1093/icb/icx082)

53. Jaenisch R, Bird A. 2003 Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nat. Genet.* **33**, 245–254. (doi:10.1038/ng1089)

54. Husby A. 2022 Wild epigenetics: insights from epigenetic studies on natural populations. *Proc. R. Soc. B* **289**, 20211633. (doi:10.1098/rspb.2021.1633)

55. Sepers B, Mateman AC, Gaweens F, Verhoeven KJF, van Oers K. 2023 Developmental stress does not induce genome-wide DNA methylation changes in wild great tit (*Parus major*) nestlings. *Mol. Ecol.* **32**, 3960–3974. (doi:10.1111/mec.16973)

56. McEwen BS, Eiland L, Hunter RG, Miller MM. 2012 Stress and anxiety: structural plasticity and epigenetic regulation as a consequence of stress. *Neuropharmacology* **62**, 3–12. (doi:10.1016/j.neuropharm.2011.07.014)

57. Rodgers AB, Morgan CP, Leu NA, Bale TL. 2015 Transgenerational epigenetic programming via sperm microRNA recapitulates effects of paternal

stress. *Proc. Natl Acad. Sci. USA* **112**, 13 699–13 704. (doi:10.1073/pnas.1508347112)

58. Klengel T *et al.* 2013 Allele-specific *FKBP5* DNA demethylation mediates gene–childhood trauma interactions. *Nat. Neurosci.* **16**, 33–41. (doi:10.1038/nn.3275)

59. St-Cyr S, Abuash S, Sivanathan S, McGowan PO. 2017 Maternal programming of sex-specific responses to predator odor stress in adult rats. *Horm. Behav.* **94**, 1–12. (doi:10.1016/j.yhbeh.2017.06.005)

60. Meaney MJ, Szyf M, Seckl JR. 2007 Epigenetic mechanisms of perinatal programming of hypothalamic–pituitary–adrenal function and health. *Trends Mol. Med.* **13**, 269–277. (doi:10.1016/j.molmed.2007.05.003)

61. Zimmer C, Larriva M, Boogert NJ, Spencer KA. 2017 Transgenerational transmission of a stress-coping phenotype programmed by early-life stress in the Japanese quail. *Scient. Rep.* **7**, 46125. (doi:10.1038/srep46125)

62. Zimmer C, Spencer KA. 2014 Modifications of glucocorticoid receptors mRNA expression in the hypothalamic–pituitary–adrenal axis in response to early-life stress in female Japanese quail. *J. Neuroendocr.* **26**, 853–860. (doi:10.1111/jne.12228)

63. Bartlett AA, Lapp HE, Hunter RG. 2019 Epigenetic mechanisms of the glucocorticoid receptor. *Trends Endocr. Metab.* **30**, 807–818. (doi:10.1016/j.tem.2019.07.003)

64. Weaver ICG, Cervoni N, Champagne FA, D’Alessio AC, Sharma S, Seckl JR, Dymov S, Szyf M, Meaney MJ. 2004 Epigenetic programming by maternal behavior. *Nat. Neurosci.* **7**, 847–854. (doi:10.1038/nn1276)

65. Yehuda R, Daskalakis NP, Lehrner A, Desarnaud F, Bader HN, Makotkine I, Flory JD, Bierer LM, Meaney MJ. 2014 Influences of maternal and paternal PTSD on epigenetic regulation of the glucocorticoid receptor gene in Holocaust survivor offspring. *Am. J. Psychiat.* **171**, 872–880. (doi:10.1176/appi.ajp.2014.13121571)

66. Ruiz-Raya F, Noguera JC, Velando A. 2023 Covariation between glucocorticoid levels and receptor expression modulates embryo development and postnatal phenotypes in gulls. *Horm. Behav.* **149**, 105316. (doi:10.1016/j.yhbeh.2023.105316)

67. Jimeno B, Hau M, Gomez-Diaz E, Verhulst S. 2019 Developmental conditions modulate DNA methylation at the glucocorticoid receptor gene with cascading effects on expression and corticosterone levels in zebra finches. *Scient. Rep.* **9**, 15869. (doi:10.1038/s41598-019-52203-8)

68. Klengel T, Binder EB. 2013 Allele-specific epigenetic modification: a molecular mechanism for gene–environment interactions in stress-related psychiatric disorders? *Epigenomics* **5**, 109–112. (doi:10.2217/Epi.13.11)

69. Wiechmann T *et al.* 2019 Identification of dynamic glucocorticoid-induced methylation changes at the *FKBP5* locus. *Clin. Epigenet.* **11**, 83. (doi:10.1186/s13148-019-0682-5)

70. Yehuda R. 2015 Is the glucocorticoid receptor a therapeutic target for the treatment of PTSD. *Psychoneuroendocrinology* **61**, 2. (doi:10.1016/j.psyneuen.2015.07.391)

71. Martin LB, Liebl AL. 2014 Physiological flexibility in an avian range expansion. *Gen. Comp. Endocr.* **206**, 227–234. (doi:10.1016/j.ygenc.2014.07.016)

72. Nussey DH, Wilson AJ, Brommer JE. 2007 The evolutionary ecology of individual phenotypic plasticity in wild populations. *J. Evol. Biol.* **20**, 831–844. (doi:10.1111/j.1420-9101.2007.01300.x)

73. Dingemanse NJ, Kazem AJN, Reale D, Wright J. 2010 Behavioural reaction norms: animal personality meets individual plasticity. *Trends Ecol. Evol.* **25**, 81–89. (doi:10.1016/j.tree.2009.07.013)

74. McCoy MW, Bolker BM. 2008 Trait-mediated interactions: influence of prey size, density and experience. *J. Anim. Ecol.* **77**, 478–486. (doi:10.1111/j.1365-2656.2008.01372.x)

75. Breuner CW, Orchinik M. 2001 Seasonal regulation of membrane and intracellular corticosteroid receptors in the house sparrow brain. *J. Neuroendocr.* **13**, 412–420. (doi:10.1046/j.1365-2826.2001.00646.x)

76. Romero LM. 2002 Seasonal changes in plasma glucocorticoid concentrations in free-living vertebrates. *Gen. Comp. Endocr.* **128**, 1–24. (doi:10.1016/S0016-6480(02)00064-3)

77. Sonnweber R *et al.* 2018 Circadian rhythms of urinary cortisol levels vary between individuals in wild male chimpanzees: a reaction norm approach. *Front. Ecol. Evol.* **6**, 85. (doi:10.3389/fevo.2018.00085)

78. Lendvai AZ, Ouyang JQ, Schoenle LA, Fasanella V, Haussmann MF, Bonier F, Moore IT. 2014 Experimental Food Restriction Reveals Individual Differences in Corticosterone Reaction Norms with No Oxidative Costs. *PLoS ONE* **9**, e0110564. (doi:10.1371/journal.pone.0110564)

79. Scharf SH, Liebl C, Binder EB, Schmidt MV, Muller MB. 2011 Expression and regulation of the *Fkbp5* gene in the adult mouse brain. *Plos One* **6**, e16883. (doi:10.1371/journal.pone.0016883)

80. Creel S, Dantzer B, Goymann W, Rubenstein DR. 2013 The ecology of stress: effects of the social environment. *Funct. Ecol.* **27**, 66–80. (doi:10.1111/j.1365-2435.2012.02029.x)

81. Mooring MS, Patton ML, Lance VA, Hall BM, Schaad EW, Fetter GA, Fortin SS, McPeak KM. 2006 Glucocorticoids of bison bulls in relation to social status. *Horm. Behav.* **49**, 369–375. (doi:10.1016/j.yhbeh.2005.08.008)

82. Sapolsky RM, Altmann J. 1991 Incidence of hypercortisolism and dexamethasone resistance increases with age among wild baboons. *Biol. Psychiat.* **30**, 1008–1016. (doi:10.1016/0006-3223(91)90121-2)

83. Sapolsky RM. 1993 Potential behavioral-modification of glucocorticoid damage to the hippocampus. *Behav. Brain Res.* **57**, 175–182. (doi:10.1016/0166-4328(93)90133-B)

84. Cornelius JM, Perreau G, Bishop VR, Krause JS, Smith R, Hahn TP, Meddle SL. 2018 Social information changes stress hormone receptor expression in the songbird brain. *Horm. Behav.* **97**, 31–38. (doi:10.1016/j.yhbeh.2017.10.002)

85. Otte C, Jahn H, Yassouridis A, Arlt J, Stober N, Maass P, Wiedemann K, Kellner M. 2003 The mineralocorticoid receptor agonist, fludrocortisone, inhibits pituitary-adrenal activity in humans after pre-treatment with metyrapone. *Life Sci.* **73**, 1835–1845. (doi:10.1016/S0024-3205(03)00513-7)

86. Dodt C, Theine KJ, Uthgenannt D, Born J, Fehm HL. 1994 Basal secretory activity of the hypothalamo-pituitary–adrenocortical axis is enhanced in healthy elderly. An assessment during undisturbed nighttime sleep. *Eur. J. Endocr.* **131**, 443–450. (doi:10.1530/eje.0.1310443)

87. Lupien SJ, de Leon M, de Santi S, Convit A, Tarshish C, Thakur M, McEwen BS, Hauger RL, Meaney MJ. 1998 Cortisol levels during human aging predict hippocampal atrophy and memory deficits. *Nat. Neurosci.* **1**, 69–73. (doi:10.1038/271)

88. Blair LJ *et al.* 2013 Accelerated neurodegeneration through chaperone-mediated oligomerization of tau. *J. Clin. Invest.* **123**, 4158–4169. (doi:10.1172/JCI69003)

89. Angelier F, Chastel O, Lendvai AZ, Parenteau C, Weimerskirch H, Wingfield JC. 2020 When do older birds better resist stress? A study of the corticosterone stress response in snow petrels. *Biol. Lett.* **16**, 20190733. (doi:10.1098/rsbl.2019.0733)

90. Angelier F, Shaffer SA, Weimerskirch H, Chastel O. 2006 Effect of age, breeding experience and senescence on corticosterone and prolactin levels in a long-lived seabird: the wandering albatross. *Gen. Comp. Endocr.* **149**, 1–9. (doi:10.1016/j.ygenc.2006.04.006)

91. Heidinger BJ, Chastel O, Nisbet ICT, Ketterson ED. 2010 Mellowing with age: older parents are less responsive to a stressor in a long-lived seabird. *Funct. Ecol.* **24**, 1037–1044. (doi:10.1111/j.1365-2435.2010.01733.x)

92. Heidinger BJ, Nisbet ICT, Ketterson ED. 2006 Older parents are less responsive to a stressor in a long-lived seabird: a mechanism for increased reproductive performance with age? *Proc. R. Soc. B* **273**, 2227–2231. (doi:10.1098/rspb.2006.3557)

93. Lendvai ÁZ, Giraudeau M, Bókony V, Angelier F, Chastel O. 2015 Within-individual plasticity explains age-related decrease in stress response in a short-lived bird. *Biol. Lett.* **11**, 20150272. (doi:10.1098/rsbl.2015.0272)

94. Seebacher F. 2022 Interactive effects of anthropogenic environmental drivers on endocrine responses in wildlife. *Mol. Cell. Endocr.* **556**, 111737. (doi:10.1016/j.mce.2022.111737)

95. Deviche P, Sweazea K, Angelier F. 2023 Past and future: urbanization and the avian endocrine system. *Gen. Comp. Endocr.* **332**, 114159. (doi:10.1016/j.ygenc.2022.114159)

96. Bonier F. 2023 Future directions in urban endocrinology – the effects of endocrine plasticity on urban tolerance. *Mol. Cell. Endocr.* **565**, 111886. (doi:10.1016/j.mce.2023.111886)

97. Martin LB *et al.* 2018 IUCN conservation status does not predict glucocorticoid concentrations in reptiles and birds. *Integr. Comp. Biol.* **58**, 800–813. (doi:10.1093/icb/icy102)

98. Injaian AS *et al.* 2020 Baseline and stress-induced corticosterone levels across birds and reptiles do not reflect urbanization levels. *Conserv. Physiol.* **8**, coz110. (doi:10.1093/conphys/coz110)

99. Sinclair ECC, Martin PR, Bonier F. 2022 Among-species variation in hormone concentrations is associated with urban tolerance in birds. *Proc. R. Soc. B* **289**, 20221600. (doi:10.1098/rspb.2022.1600)

100. McEwen BS, Wingfield JC. 2010 What's in a name? Integrating homeostasis, allostasis and stress. *Horm. Behav.* **57**, 105. (doi:10.1016/j.yhbeh.2009.09.011)

101. Cohen AA, Martin LB, Wingfield JC, McWilliams SR, Dunne JA. 2012 Physiological regulatory networks: ecological roles and evolutionary constraints. *Trends Ecol. Evol.* **27**, 428–435. (doi:10.1016/j.tree.2012.04.008)

102. Martin LB, Cohen A. 2014 Physiological regulatory networks: the orchestra of life? In *Integrative organismal biology* (eds LB Martin, CK Ghalambor, HA Woods), pp. 137–152. Hoboken, NJ: Wiley Blackwell.

103. Ketterson ED, Atwell JW, McGlothlin JW. 2009 Phenotypic integration and independence: hormones, performance, and response to environmental change. *Integr. Comp. Biol.* **49**, 365–379. (doi:10.1093/icb/icp057)

104. McGlothlin JW, Ketterson ED. 2008 Hormone-mediated suites as adaptations and evolutionary constraints. *Phil. Trans. R. Soc. B* **363**, 1611–1620. (doi:10.1098/rstb.2007.0002)

105. Martin LB, Liebl AL, Trotter JH, Richards CL, McCoy K, McCoy MW. 2011 Integrator networks: illuminating the black box linking genotype and phenotype. *Integr. Comp. Biol.* **51**, 514–527. (doi:10.1093/icb/icr049)

106. Ketterson ED, Nolan Jr V. 1992 Hormones and life histories: an integrative approach. *Am. Nat.* **140**, S33–S62. (doi:10.1086/285396)

107. Duffy AM, Clobert J, Moller AP. 2002 Hormones, developmental plasticity and adaptation. *Trends Ecol. Evol.* **17**, 190–196. (doi:10.1016/S0169-5347(02)02498-9)

108. Dantzer B. 2023 Frank Beach Award Winner: The centrality of the hypothalamic-pituitary-adrenal axis in dealing with environmental change across temporal scales. *Horm. Behav.* **150**, 105311. (doi:10.1016/j.yhbeh.2023.105311)

109. Taborsky B, English S, Fawcett TW, Kuijper B, Leimar O, McNamara JM, Ruuskanen S, Sandi C. 2021 Towards an Evolutionary Theory of Stress Responses. *Trends Ecol. Evol.* **36**, 39–48. (doi:10.1016/j.tree.2020.09.003)

110. Arnold SJ. 1983 Morphology, Performance and Fitness. *Am. Zool.* **23**, 347–361. (doi:10.1093/icb/23.2.347)

111. Shanahan DF, Strohbach MW, Warren PS, Fuller RA. 2014 The challenges of urban living. In *Avian urban ecology: behavioural and physical adaptations* (eds D Gil, H Brumm), pp. 3–20. Oxford, UK: Oxford University Press.

112. Gil D, Brumm H. 2014 Avian urban ecology: behavioural and physiological adaptations introduction. In *Avian urban ecology: behavioural and physiological adaptations* (eds D Gil, H Brumm), pp. xiii–xv. Oxford, UK: Oxford University Press.

113. Martin LB, Hopkins WA, Mydlarz L, Rohr JR. 2010 The effects of anthropogenic global changes on immune functions and disease resistance. *Ann. NY Acad. Sci.* **1195**, 129–148. (doi:10.1111/j.1749-6632.2010.05454.x)

114. Bonier F. 2012 Hormones in the city: endocrine ecology of urban birds. *Horm. Behav.* **61**, 763–772. (doi:10.1016/j.yhbeh.2012.03.016)

115. Iglesias-Carrasco M, Aich U, Jennions MD, Head ML. 2020 Stress in the city: meta-analysis indicates no overall evidence for stress in urban vertebrates. *Proc. R. Soc. B* **287**, 20201754. (doi:10.1098/rspb.2020.1754)

116. Vitousek MN *et al.* 2018 HormoneBase, a population-level database of steroid hormone levels across vertebrates. *Sci. Data* **5**, 180097. (doi:10.1038/sdata.2018.97)

117. Injaian AS, Uehling JJ, Taff CC, Vitousek MN. 2021 Effects of artificial light at night on avian provisioning, corticosterone, and reproductive success. *Integr. Comp. Biol.* **61**, 1147–1159. (doi:10.1093/icb/icab055)

118. Ouyang JQ, de Jong M, Hau M, Visser ME, van Grunsven RH, Spoelstra K. 2015 Stressful colours: corticosterone concentrations in a free-living songbird vary with the spectral composition of experimental illumination. *Biol. Lett.* **11**, 20150517. (doi:10.1098/rsbl.2015.0517)

119. Lane SJ, Emmerson MG, VanDiest IJ, Hucul C, Beck ML, Davies S, Gilbert ER, Sewall KB. 2021 Hypothalamic-pituitary-adrenal axis regulation and organization in urban and rural song sparrows. *Gen. Comp. Endocrin.* **310**, 113809. (doi:10.1016/j.ygenc.2021.113809)

120. Callaghan CT, Palacio FX, Benedetti Y, Morelli F, Bowler DE. 2023 Large-scale spatial variability in urban tolerance of birds. *J. Anim. Ecol.* **92**, 403–416. (doi:10.1111/1365-2656.13862)

121. Ibanez-Alamo JD, Jimeno B, Gil D, Thomson RL, Aguirre JL, Diez-Fernandez A, Faivre B, Tielemans BL, Figuerola J. 2020 Physiological stress does not increase with urbanization in European blackbirds: evidence from hormonal, immunological and cellular indicators. *Sci. Total Environ.* **721**, 137332. (doi:10.1016/j.scitotenv.2020.137332)

122. Shochat E, Warren PS, Faeth SH, McIntyre NE, Hope D. 2006 From patterns to emerging processes in mechanistic urban ecology. *Trends Ecol. Evol.* **21**, 186–191. (doi:10.1016/j.tree.2005.11.019)

123. Shochat E. 2004 Credit or debit? Resource input changes population dynamics of city-slicker birds. *Oikos* **106**, 622–626. (doi:10.1111/j.0030-1299.2004.13159.x)

124. Martin LB, Boruta M. 2014 The impacts of urbanization on avian disease transmission and emergence. In *Avian urban ecology: behavioural and physical adaptations* (eds D Gil, H Brumm), pp. 116–128. Oxford, UK: Oxford University Press.

125. Callaghan CT, Sayol F, Benedetti Y, Morelli F, Sol D. 2021 Validation of a globally-applicable method to measure urban tolerance of birds using citizen science data. *Ecol. Indic.* **120**, 106905. (doi:10.1016/j.ecolind.2020.106905)

126. Callaghan CT, Cornwell WK, Poore AGB, Benedetti Y, Morelli F. 2021 Urban tolerance of birds changes throughout the full annual cycle. *J. Biogeogr.* **48**, 1503–1517. (doi:10.1111/jbi.14093)

127. Martin LB, Brace AJ, Gervasi SS, Kilvits HJ. 2016 Invader endocrinology: how to regulate a pesky phenotype. In *Biological invasions and animal behaviour* (eds JS Weis, D Sol), pp. 47–62. Cambridge, UK: Cambridge University Press.

128. Wingfield JC. 2013 The comparative biology of environmental stress: behavioural endocrinology and variation in ability to cope with novel, changing environments. *Anim. Behav.* **85**, 1127–1133. (doi:10.1016/j.anbehav.2013.02.018)

129. Liebl AL, Martin LB. 2012 Exploratory behaviour and stressor hyper-responsiveness facilitate range expansion of an introduced songbird. *Proc. R. Soc. B* **279**, 4375–4381. (doi:10.1098/rspb.2012.1606)

130. Liebl AL, Martin LB. 2013 Stress hormone receptors change as range expansion progresses in house sparrows. *Biol. Lett.* **9**, 20130181. (doi:10.1098/rsbl.2013.0181)

131. Kilvits HJ, Ardia DR, Thiam M, Martin LB. 2018 Corticosterone is correlated to mediators of neural plasticity and epigenetic potential in the hippocampus of Senegalese house sparrows (*Passer domesticus*). *Gen. Comp. Endocr.* **269**, 177–183. (doi:10.1016/j.ygenc.2018.09.014)

132. Martin LB, Kilvits HJ, Thiam M, Ardia DR. 2017 Corticosterone regulation in house sparrows invading Senegal. *Gen. Comp. Endocr.* **250**, 15–20. (doi:10.1016/j.ygenc.2017.05.018)

133. Siefferman L, Bentz AB, Rosvall KA. 2023 Decoupling pioneering traits from latitudinal patterns in a North American bird experiencing a southward range shift. *J. Anim. Ecol.* **92**, 1149–1160. (doi:10.1111/1365-2656.13907)

134. Assis VR, Gardner ST, Smith KM, Gomes FR, Mendonca MT. 2020 Stress and immunity: field comparisons among populations of invasive cane toads in Florida. *J. Exp. Zool. A* **333**, 779–791. (doi:10.1002/jez.2389)

135. Brown GP, Kelehear C, Shilton CM, Phillips BL, Shine R. 2015 Stress and immunity at the invasion front: a comparison across cane toad (*Rhinella marina*) populations. *Biol. J. Linn. Soc.* **116**, 748–760. (doi:10.1111/bij.12623)

136. Jessop TS, Letnic M, Webb JK, Dempster T. 2013 Adrenocortical stress responses influence an invasive vertebrate's fitness in an extreme environment. *Proc. R. Soc. B* **280**, 20131444. (doi:10.1098/rspb.2013.1444)

137. Azevedo A, Bailey L, Bandeira V, Fonseca C, Wauters J, Jewgenow K. 2021 Decreasing glucocorticoid levels towards the expansion front suggest ongoing

expansion in a terrestrial mammal. *Conserv. Physiol.* **9**, coab050. (doi:10.1093/conphys/coab050)

138. Martin LB, Trainor BC, Finy MS, Nelson RJ. 2007 HPA activity and neotic and anxiety-like behavior vary among *Peromyscus* species. *Gen. Comp. Endocr.* **141**, 342–350. (doi:10.1016/j.ygcen.2007.02.001)

139. Cote J, Clobert J, Brodin T, Fogarty S, Sih A. 2010 Personality-dependent dispersal: characterization, ontogeny and consequences for spatially structured populations. *Phil. Trans. R. Soc. B* **365**, 4065–4076. (doi:10.1098/rstb.2010.0176)

140. Dickens MJ, Delehanty DJ, Romero LM. 2009 Stress and translocation: alterations in the stress physiology of translocated birds. *Proc. R. Soc. B* **276**, 2051–2056. (doi:10.1098/rspb.2008.1778)

141. Cooke SJ, Sack L, Franklin CE, Farrell AP, Beardall J, Wikelski M, Chown SL. 2013 What is conservation physiology? Perspectives on an increasingly integrated and essential science. *Conserv. Physiol.* **1**, cot001. (doi:10.1093/conphys/cot001)

142. Love A, Wagner GP. 2022 Co-option of stress mechanisms in the origin of evolutionary novelties. *Evolution* **76**, 394–413. (doi:10.1111/evo.14421)

143. Lema SC. 2020 The adaptive value of hormones: endocrine systems as outcomes and initiators of evolution. *Mol. Cell. Endocr.* **517**, 110983. (doi:10.1016/j.mce.2020.110983)

144. Houslay TM, Earley RL, White SJ, Lammers W, Grimmer AJ, Travers LM, Johnson EL, Young AJ, Wilson A. 2022 Genetic integration of behavioural and endocrine components of the stress response. *eLife* **11**, e67126. (doi:10.7554/eLife.67126)

145. Martin LB *et al.* 2019 Extreme competence: keystone hosts of infections. *Trends Ecol. Evol.* **34**, 303–314. (doi:10.1016/j.tree.2018.12.009)

146. Beldomenico PM, Begon M. 2016 Stress-host-parasite interactions: a vicious triangle? *Rev. FAVE Cienc. Vet.* **14**, 6–19. (doi:10.14409/favecv.v14i1/2.5160)

147. Gervasi SS, Burkett-Cadena N, Burgan SC, Schrey AW, Hassan HK, Unnasch TR, Martin LB. 2016 Host stress hormones alter vector feeding preferences, success, and productivity. *Proc. R. Soc. B* **283**, 20161278. (doi:10.1098/rspb.2016.1278)

148. Gervasi SS, Burgan SC, Hofmeister EK, Unnasch TR, Martin LB. 2017 Stress hormones predict a host superspread phenotype in the West Nile virus system. *Proc. R. Soc. B* **284**, 20171090. (doi:10.1098/rspb.2017.1090)

149. Donelan SC, Hellmann JK, Bell AM, Luttbeg B, Orrock JL, Sheriff MJ, Sih A. 2020 Transgenerational plasticity in human-altered environments. *Trends Ecol. Evol.* **35**, 115–124. (doi:10.1016/j.tree.2019.09.003)

150. Snell-Rood EC. 2013 An overview of the evolutionary causes and consequences of behavioural plasticity. *Anim. Behav.* **85**, 1004–1011. (doi:10.1016/j.anbehav.2012.12.031)

151. Hanson HE, Wang CQ, Schrey AW, Liebl AL, Ravinet M, Jiang RHY, Martin LB. 2022 Epigenetic potential and DNA methylation in an ongoing house sparrow (*Passer domesticus*) range expansion. *Am. Nat.* **200**, 662–674. (doi:10.1086/720950)

152. Smallegange IM. 2022 Integrating developmental plasticity into eco-evolutionary population dynamics. *Trends Ecol. Evol.* **37**, 129–137. (doi:10.1016/j.tree.2021.09.005)