

# PUTATIVE *CELTIS* LEAVES FROM EOCENE PATAGONIA ARE ALLIED WITH ASIAN ANACARDIACEAE

PETER WILF<sup>1</sup>, CYNTHIA C. GONZÁLEZ<sup>2</sup>, MARÍA A. GANDOLFO<sup>3</sup>, AND MARÍA C.

5 ZAMALOA<sup>4</sup>

<sup>1</sup>Department of Geosciences, Pennsylvania State University, University Park, Pennsylvania,  
16802, USA. *pwilf@psu.edu*

<sup>2</sup>Herbario Trelew, Facultad de Ciencias Naturales y Ciencias de la Salud, Universidad  
Nacional de la Patagonia San Juan Bosco – Sede Trelew, U9100GYO Trelew, Argentina.

10 *cynthiacgonzalez@yahoo.com.ar*

<sup>3</sup>L. H. Bailey Hortorium, Plant Biology Section, School of Integrative Plant Science, Cornell  
University, Ithaca, New York, 14853, USA. *magandolfo@cornell.edu*

<sup>4</sup>Museo Paleontológico Egidio Feruglio, U9100GYO Trelew, Argentina.  
*mzamaloa@gmail.com*

15

46 pages; 6 figures; 0 tables

Running header: WILF *ET AL.*: PATAGONIAN FOSSILS LINKED TO ASIAN  
ANACARDIACEAE

Short description: Abundant Eocene "*Celtis*" leaves from Laguna del Hunco in Patagonia are  
20 related to *Dobinea*, a mainland Asian genus of Anacardiaceae.

Corresponding author: Peter Wilf, *pwilf@psu.edu*

**Abstract.** The most common macrofossils in the highly diverse flora from Laguna del Hunco (early Eocene of Chubut, Argentina) are "*Celtis*" *ameghinoi* leaves, whose true affinities have remained enigmatic for a century. The species accounts for 14% of all plant fossils in unbiased field counts and bears diverse insect-feeding damage, suggesting its high biomass and paleoecological importance. The leaves have well-preserved architecture but lack cuticles or reproductive attachments. We find that the fossils only superficially resemble *Celtis* and comparable taxa in Cannabaceae, Ulmaceae, Rhamnaceae, Malvaceae, and many other families. However, the distinctive foliar morphology conforms in detail to *Dobinea* (Anacardiaceae), a genus with two species of shrubs and large herbs ranging from India's Far East and Tibet to Myanmar and central China, and we propose *Dobineaites ameghinoi* (E.W. Berry) gen et. comb. nov. for the fossils. This discovery strengthens the extensive biogeographic links between Eocene Patagonia and mainland Asia, provides the first fossil record related to *Dobinea*, and represents a rare Gondwanan macrofossil occurrence of Anacardiaceae, which was widespread and diversified in the Northern Hemisphere at the time. The diverse leaf architecture of Anacardiaceae includes several patterns usually associated with other taxa, and many other leaf fossils in this family may remain misidentified.

**Key words.** Anacardiaceae. Argentina. Chinese flora. *Dobinea*. Gondwana. Herbivory. Laguna del Hunco. Leaf architecture.

**Resumen.** HOJAS DE "*CELTIS*" DEL EOCENO DE PATAGONIA VINCULADAS CON ANACARDIACEAE ASIÁTICAS. Los macrofósiles más comunes en la flora altamente diversa de la Laguna del Hunco (Eoceno temprano de Chubut, Argentina) son hojas de "*Celtis*" *ameghinoi*, conocidas desde hace un siglo y cuyas afinidades han sido enigmáticas. La especie representa el 14% del total de las hojas fósiles en censos imparciales de campo, y los ejemplares exhiben diversos daños producidos por alimentación de insectos, lo que

sugiere que habrían tenido elevada biomasa e importancia ecológica. Las hojas presentan la arquitectura bien conservada, pero carecen de cutículas o conexiones con estructuras

50 reproductivas. Encontramos que los fósiles sólo se parecen superficialmente a *Celtis* y a taxones comparables de Cannabaceae, Ulmaceae, Rhamnaceae, Malvaceae, y muchas otras familias. Sin embargo, la morfología foliar distintiva se ajusta en detalle a *Dobinea* (Anacardiaceae), un género con dos especies de arbustos y grandes hierbas que se distribuye desde el Oriente de la India y el Tíbet hasta Myanmar y China central. Proponemos

55 *Dobineaites ameghinoi* (E.W. Berry) gen et. comb. nov. para las hojas fósiles. Este hallazgo refuerza los ya extensos vínculos biogeográficos entre el Eoceno de Patagonia y Asia continental, proporciona el primer registro fósil relacionado con *Dobinea* y representa una rara presencia macrofósil gondwánica de Anacardiaceae, una familia que estaba muy extendida y diversificada en el hemisferio norte en ese momento. La diversa arquitectura

60 foliar de las Anacardiaceae incluye varios patrones comúnmente asociados con otros grupos de plantas por lo que muchos fósiles de hojas de esta familia pueden permanecer aun mal identificados.

**Palabras clave.** Anacardiaceae. Argentina. Arquitectura foliar. *Dobinea*. Flora de China.

65 Gondwana. Herbivoría. Laguna del Hunco.

THE EARLY EOCENE FOSSIL-LAKE BEDS at Laguna del Hunco in the Piedra Parada Caldera of northwestern Chubut, Argentina, contain a well-preserved, highly diverse plant and animal biota that has been known for a century (Clark, 1923; Berry, 1925; Dolgopol de Sáez, 1941).

70 The pace of discovery has increased significantly over the past 25 years because of intensive, stratigraphically controlled collecting, resulting in over 8,000 specimens of more than 180

plant macrofossil species from more than 30 quarries in a 170 m stratigraphic section (*e.g.*, Wilf *et al.*, 2003, 2005a, 2023). More than 30 well-vetted plant families are present in the flora, including palynological occurrences (Barreda *et al.*, 2020).

75           Recent systematic work has vastly improved understanding of the floristic composition of the late-Gondwanan Laguna del Hunco flora and its biogeographic connections to living genera in South America, Africa, and especially Australasia and Southeast Asia (*e.g.*, Zamaloa *et al.*, 2006; Gandolfo *et al.*, 2011; Wilf *et al.*, 2013, 2019; Kooyman *et al.*, 2014). Most dominant plant species in the flora have been revised. For  
80   example, the second most abundant leaf species by field-census leaf counts, initially identified to the family Myrtaceae (Wilf *et al.*, 2005a), is now resolved in *Eucalyptus* and is associated with infructescences and flowers bearing in situ pollen of that genus (Gandolfo *et al.*, 2011; Hermsen *et al.*, 2012; Zamaloa *et al.*, 2020). The third most common, “*Tetracera*” *patagonica* leaves with putative affinities to Dilleniaceae (Berry, 1925), are fagaceous, now  
85   placed in *Castaneophyllum*, and associated with *Castanopsis* infructescences and dispersed castaneoid pollen (Wilf *et al.*, 2019; Barreda *et al.*, 2020). The seventh most abundant leaf type, the putative Cycadaceae “*Zamia tertiaria*,” actually represents the araucarian conifer *Agathis*, supported by co-occurring *Agathis* pollen cones, seed cones, and ovuliferous  
90   complexes bearing in situ seeds (Wilf *et al.*, 2014; quotation marks indicate uncertain botanical affinities for a published name or other nomenclatural issues).

          Despite these advances, the most common leaf species at Laguna del Hunco, “*Celtis*” *ameghinoi* E.W. Berry (1925), remains unrevised, leaving a gap in the fundamental knowledge of the assemblage and its paleoecology and biogeography. Notwithstanding its long collection history, the species is known only from leaf adpressions without cuticles, but  
95   its leaf architecture is well preserved.



Berry (1925) described "*Celtis*" "*ameghenoi*" based on three type specimens (Fig. 1) in the first report of plant fossils from Laguna del Hunco (also known then as El Mirador, Mirhoja, Laguna del Junco, and Laguna de los Huncos), from a small sample discovered by geologist Burton Clark (1923). The specific epithet honored this journal's namesake,  
 100 Florentino Ameghino (1853–1911), whom Berry (1925: 200) referred to as "my lamented friend." Later, Berry (1938: 41) credited Ameghino for his initial inspiration to study South American geology and referred to "long drawn out discussions" about the ages of South American fossils. Berry (1938) corrected the typographic error in the species epithet as "*ameghinoi*," which we follow here.

105 The "*Celtis*" *ameghinoi* syntypes (Fig. 1) include one small leaf with a narrow, lanceolate aspect (Fig. 1.1) and two that are larger, wider, and cordate (Fig. 1.2, 1.3). Specimens since collected (Figs. 2–4) display a more complete spectrum of variation that encompasses the morphologies of the three types, sharing characters such as a pinnate, unlobed blade; prominent, closely spaced, irregular to compound teeth that occur on nearly  
 110 all of the blade margin; secondary veins that increase in angle toward the base; widely spaced, weakly percurrent tertiary veins; and perpendicular vein branching from tooth principal veins to a vein knot just inside or at the tooth sinuses. Other common characteristics include a symmetrical ovate blade, cordate base (mostly on larger specimens), acute-to-acuminate apex, deflected major veins, and secondary veins that branch well inside the  
 115 margin. The overall feature combination makes the species easily recognizable. Berry (1925) noted that "*C.*" *ameghinoi* was the most abundant form in the assemblage, which remains true today (Wilf *et al.*, 2005a). He (Berry, 1925: 198–199) considered the fossils "very well marked and characteristic of *Celtis*" (Cannabaceae, formerly in Ulmaceae and Celtidaceae), and he also found them similar to the New World genera *Momisia* (now a synonym of *Celtis*),  
 120 *Trema* (Cannabaceae), and *Ampelocera* and *Phyllostylon* (Ulmaceae). Other genera that Berry

listed as similar to the fossils were *Gouania* (Rhamnaceae) as well as *Grewia* and *Triumfetta* (both Malvaceae).

In his classic monograph of the middle Eocene (47.7 Ma; Wilf, 2012) Río Pichileufú flora from Río Negro province, Argentina, Berry (1938) reported but did not illustrate

125 "*Celtis*" *ameghinoi* as a rare element. Rossetto-Harris (2023) re-examined the type and cohort Río Pichileufú collections that Berry studied (housed in the Division of Paleobotany, Smithsonian Institution, Washington, D.C.). She located four specimens labeled as "*Celtis* *ameghinoi*" in Berry's handwriting, finding that these specimens differ from the Laguna del Hunco type material and do not represent the same species.

130 Joaquín Frenguelli and Rodolfo Maldonado Bruzzone made significant early collections from Laguna del Hunco in the late 1930s and the early 1940s, housed at the Museo de La Plata (*e.g.*, Frenguelli, 1943a, 1943b). Traverso (1951) described ca. 60 specimens of "*Celtis*" *ameghinoi* from these collections in an unpublished thesis. Many years later, González (2008), also in an unpublished thesis, made the only other taxonomic

135 interpretations of "*Celtis*" *ameghinoi* to date. She studied the species using fossil collections housed at several institutions from Laguna del Hunco and Arroyo Chacay, an Eocene site in Río Negro Province (*e.g.*, Machado *et al.*, 2023). In preparing the present manuscript, we found the Arroyo Chacay material to represent another species; thus, we consider Laguna del Hunco to be the only known site where "*Celtis*" *ameghinoi* occurs. González (2008)

140 concluded that "*C.*" *ameghinoi* differed substantially from all the living genera that she compared, including extant *Celtis*, *Trema*, *Aphananthe*, and *Phyllostylon* (see Discussion for updated comparisons incorporating these observations).

"*Celtis*" *ameghinoi* is occasionally referenced as a true representative of *Celtis* (*e.g.*, Barreda & Palazzesi, 2007, 2010; Vento & Prámparo, 2018; Romero-Lebrón *et al.*, 2020).

145 More often, it serves as a morphotype in paleoecological analyses. From field-census data, "C." *ameghinoi* leaves were the most abundant fossils across all quarries at Laguna del Hunco, accounting for 597 of the 4,303 total specimens (13.9%; Wilf *et al.*, 2005a). The species also had the highest leaf counts at two of the four principal census quarries (21.7% at LH02 and 27.6% at LH04; *Eucalyptus frenguelliana* was dominant at the other two). Based on modern analog studies, ranked leaf counts correlate well with ranked source biomass by species (Burnham *et al.*, 1992; Burnham, 1997), implying a high biomass of the source plants. From the same field census, "C." *ameghinoi* showed insect damage in 12.5% of leaves (Wilf *et al.*, 2005b) and a substantial diversity of 26 damage types (DTs; Labandeira *et al.*, 2007) across all functional groups: external feeding, piercing-and-sucking, galling, mining, and oviposition. Two mining occurrences on "C." *ameghinoi* were illustrated as exemplar specimens of DT91 and DT93 in the standard guide for fossil insect damage (Labandeira *et al.*, 2007; Fig. 4.6–4.8). Some oviposition damage (Fig. 4.9) was subsequently described under the ichnospecies *Paleovoidus arcuatum* (Sarzetti *et al.*, 2009) and further analyzed morphometrically and behaviorally (Romero-Lebrón *et al.*, 2020, 2023). Its elevated abundance, diverse insect damage, and unknown biogeographic relationships all highlight the significance of understanding the botanical affinities of "*Celtis*" *ameghinoi*.

Taxonomic descriptions of isolated leaf taxa without cuticles have well-known caveats, especially when based on superficial comparisons with living genera, as historically practiced (see Dilcher, 1971; Hill, 1982). However, leaf fossils that are described using a full suite of well-defined leaf-architectural characters (Ellis *et al.*, 2009) and compared carefully with living analogs advance our understanding much more than historical names in scare quotes or informal, often unillustrated morphotypes. Several angiosperm leaf taxa with no associated reproductive organs have been described from Laguna del Hunco, including species of *Akania* (Akaniaceae), *Ripogonum* (Ripogonaceae), Proteaceae,

170 Atherospermataceae, and Monimiaceae (Romero & Hickey, 1976; González *et al.*, 2007;  
 Knight & Wilf, 2013; Carpenter *et al.*, 2014). Leaves from the site that are associated with,  
 but not attached to, related reproductive organs include species of *Eucalyptus* (Myrtaceae),  
*Macaranga* (Euphorbiaceae), castaneoid Fagaceae, and Menispermaceae (Gandolfo *et al.*,  
 2011; Jud *et al.*, 2018; Wilf *et al.*, 2019, 2023). Here, we present compelling similarities of  
 175 the "*C. ameghinoi*" fossils to an unexpected group, the Anacardiaceae, particularly the living  
 Chinese species *Dobinea delavayi*. We consider the implications of our results for the  
 paleoecology of Laguna del Hunco, biogeographic connections of late-Gondwanan  
 Patagonia, and the fossil history of Anacardiaceae.

**Institutional abbreviations.** Acronyms for repositories and their corresponding institutions  
 180 (when distinct) denote individual specimens or institutions, respectively. **USNM**, National  
 Museum of Natural History, Smithsonian Institution, Washington, D.C., USA (institutional  
 acronym **NMNH**); **MPEF-Pb**, Paleobotanical Collection of Museo Paleontológico Egidio  
 Feruglio, Trelew, Argentina (**MEF**); **LPPB**, Paleobotanical Collection of Museo de La Plata,  
 La Plata, Argentina (**MLP**); **BA-Pb**, Paleobotanical Collection of Museo Argentino de  
 185 Ciencias Naturales Bernardino Rivadavia, Buenos Aires, Argentina (**MACN**); **MJHG**,  
 Museo Jorge H. Gerhold, Ingeniero Jacobacci, Argentina; **LILPB**, Paleobotanical Collection  
 of Fundación Miguel Lillo, Tucumán, Argentina (**LIL**); **K**, Royal Botanic Gardens, Kew, UK;  
**BAB**, Instituto Nacional de Tecnología Agropecuaria, Castelar, Argentina; **SI**, Instituto de  
 Botánica Darwinion, San Isidro, Argentina; **CTES**, Instituto de Botánica del Nordeste,  
 190 Corrientes, Argentina; **BH**, L.H. Bailey Hortorium, Cornell University, Ithaca, USA; **L**,  
 Naturalis Biodiversity Center, Leiden, The Netherlands; **US**, United States National  
 Herbarium of the Smithsonian Institution, Washington, D.C., USA; **P**, Muséum National  
 d'Histoire Naturelle (**MNHN**), Paris Herbarium, France; **NY**, New York Botanical Garden,

New York, USA. Several additional herbaria mentioned in the text are cited by their standard  
 195 acronyms according to the Index Herbariorum, <https://sweetgum.nybg.org/science/ih>.

## MATERIALS AND METHODS

All fossils discussed here came from the Tufolitas Laguna del Hunco, a unit of mixed  
 siliciclastic-volcaniclastic fossil-lake beds of the Eocene Huitrera Formation in the middle  
 200 Río Chubut region of northwest Chubut, Argentina (Petersen, 1946; Aragón & Mazzoni,  
 1997; Gosses *et al.*, 2021). The Tufolitas record rapid infilling of the subsiding Piedra Parada  
 Caldera following a series of massive eruptions (estimated >300 km<sup>3</sup> emplacement) recorded  
 in the underlying Ignimbrita Barda Colorada (IBC; Aragón *et al.*, 2018). The Tufolitas are  
 exposed throughout the extensive caldera (ca. 30 km in diameter). The strata are visually  
 205 spectacular; they weather a brilliant white and are underlain by thick ignimbrites, intruded by  
 extensive dikes and laccoliths, and capped with basalt flows (Petersen, 1946; Aragón &  
 Mazzoni, 1997; Aragón *et al.*, 2001, 2004).

Fossil preservation in the Tufolitas is outstanding along a 1.3-km long escarpment in  
 its northeastern exposures near Cerro Mirador at Laguna del Hunco ("Lake of Rushes," a  
 210 small playa lake), possibly because of reduced igneous disturbance of the strata in the area.  
 Jason Hicks (in Wilf *et al.*, 2003) established a 170 m composite stratigraphic section of the  
 Tufolitas at Laguna del Hunco, incorporating 25 fossil quarries (LH01–LH25), two  
 paleomagnetic reversals, and three <sup>40</sup>Ar/<sup>39</sup>Ar dated primary airfall tuffs. Additionally, the  
 youngest ignimbrites in the IBC have an <sup>40</sup>Ar/<sup>39</sup>Ar age of 52.54 ± 0.17 Ma, which provides a  
 215 maximum age for all fossils found in the superposed Tufolitas (Gosses *et al.*, 2021). As most  
 recently summarized elsewhere from these and other constraints (Gosses *et al.*, 2021; Wilf *et al.*,  
 2023), all fossils at Laguna del Hunco date to the interval 52.2–52.0 Ma, coinciding with

the early Eocene climatic optimum and the final stages of Gondwana. The paleoenvironment is well established as an everwet (perhumid), mesic, lake-margin rainforest, based on the documented drought tolerances and climatic ranges of the living relatives of the fossil plants (e.g., Wilf, 2012; Merkhofer *et al.*, 2015) and the preservation of extraordinarily drought-sensitive accessory transfusion tissue in the leaves of the fossil podocarp conifer *Acmopyle grayae* (Andruchow-Colombo *et al.*, 2023).

Several collections have been made at Laguna del Hunco over the past century; however, until comparatively recently (Wilf *et al.*, 2003), no precise locality data have been recorded. Many older collections appear to have the characteristic lithology of quarry LH04 of Wilf *et al.* (2003), the most accessible site and the only location we have seen showing definite evidence of prior excavations (Wilf, 2020). The three syntypes of "*Celtis*" *ameghinoi* (Fig. 1) are now housed in the Laguna del Hunco type collection (Berry, 1925) at NMNH. Subsequent collections were made for several Argentine institutions, including MLP, the University of Buenos Aires (made by Edgardo Romero, currently uncatalogued), MACN (E. Romero), MJHG (made by Rodolfo Casamiquela, see Wilf *et al.* 2023), LIL, and MEF.

Since the late 1990s, international expeditions to Laguna de Hunco have been launched from the MEF, leading to extensive collections tied to precise stratigraphy and age control that have supported most subsequent research on the site (Wilf *et al.*, 2003, 2005a). Field censuses of 4,303 total specimens from 25 individual quarries (LH01–LH25) in 1999 and 2002 (Wilf *et al.*, 2003, 2005a) established the high abundance of "*C.*" *ameghinoi* fossils (see Introduction). Additional material of the species was collected during several subsequent MEF field expeditions, including two newer quarries, LH27 and LH29 (see Gandolfo *et al.*, 2011; Deanna *et al.*, 2020).

A total of 280 specimens of "*C. ameghinoi*" were studied at NMNH, MEF, MLP, LIL, MACN, and (by A. Iglesias, see Acknowledgments; Fig. 3.12) MJHG. At least 50 additional specimens that remain uncatalogued and unavailable for validation are held at the University of Buenos Aires. Photography began in the late 1990s and involved a series of film and

245 digital cameras. More recently, Nikon D90, D700, and D850 DSLRs with 60 mm and 105 mm macro lenses and polarizing filters have been used. Due to the lack of cuticular or other micromorphological preservation, little microscope photography was required. A library containing original-resolution images of the fossils, a specimen list, and high-resolution versions of Figures 1–4 is deposited on Figshare at

250 <https://doi.org/10.6084/m9.figshare.24451249>.

The extant material consulted included the recent open-access digital compilation of more than 26,000 cleared and x-rayed leaves and many of the corresponding physical slides, representing more than 350 families and 4,500 genera in total (Wilf *et al.*, 2021; Fig. 5).

Herbarium surveys by CCG at BAB, SI, CTES, BH, and US extensively covered the

255 comparable genera of Cannabaceae and Ulmaceae. We also used several online herbaria to examine relevant taxa (Fig. 6), including those of L, <https://bioportal.naturalis.nl>, US,

<https://collections.nmnh.si.edu/search/botany>, P,

<https://science.mnhn.fr/institution/mnhn/collection/p/item/search/form>, and NY via the C. V.

Starr Virtual Herbarium, <http://sweetgum.nybg.org/science/vh>. Aggregator sites included the

260 Chinese Virtual Herbarium, <https://www.cvh.ac.cn>, and JSTOR Global Plants,

<https://plants.jstor.org>. For Anacardiaceae, additional sources included the literature

(especially Gentry, 1993; Martínez-Millán & Cevallos-Ferriz, 2005; Ming & Barfod, 2008;

Andrés-Hernández & Terrazas, 2009; Pell *et al.*, 2010; Zich *et al.*, 2020; Mitchell *et al.*,

2022) and inspection of *Dobinea* species by T. Wang at K (see Acknowledgments; Fig. 6.2).

265 For increased readability, taxonomic authorities are only provided in the text where  
 necessary to support new taxonomic descriptions. However, all nomenclature and use of  
 authorities for extant taxa follow standard compilations, including Tropicos,  
<https://www.tropicos.org>, World Flora Online, <https://www.worldfloraonline.org>, and Kew's  
 Plants of the World Online, <https://powo.science.kew.org>. Authorities for fossil taxa can be  
 270 found in the corresponding cited literature. Leaf-architectural terminology follows Ellis et al.  
 (2009).

## SYSTEMATIC PALEONTOLOGY

275 Family ANACARDIACEAE R. Brown, 1818: 431

**Genus *Dobineaites*** Wilf, C. González, Gandolfo & Zamaloa gen. nov.

**Generic diagnosis.** Leaves opposite, simple, petiolate. Blade ovate, unlobed, usually  
 symmetrical; primary venation pinnate, deflected. Secondary veins craspedodromous,  
 280 dichotomizing laterally; secondaries increase in angle and decrease in spacing basally.  
 Agrophic veins compound, the first set expressed only along the basal margin. Tertiary veins  
 thick, moderately and irregularly spaced, weakly to strongly opposite percurrent, angle  
 increasing basally. Margin serrate; teeth prominent, closely spaced, continuous nearly to the  
 blade base. Teeth simple or compound, apex simple and apiculate, sinuses angular, principal  
 285 vein medial and prominent with near-perpendicular branches to vein junctions near or at the  
 sinuses.



**Type species.** *Dobineaites ameghinoi* (E.W. Berry) Wilf, C. González, Gandolfo & Zamaloea  
comb. nov.

Figures 1–4

290

**Basionym.** *Celtis ameghenoi* E.W. Berry, 1925, *Johns Hopkins University Studies in Geology*  
6:198.

**Lectotype here designated.** USNM 222678 (Fig. 1.1). Tufolitas Laguna del Hunco, Huitrera  
Formation, early Eocene (Ypresian), collected in the early 1920s by Burton Clark, exact date  
295 and collection site unknown.

**Syntypes.** USNM 222679 (Fig. 1.2), USNM 222680 (Fig. 1.3); provenance as for the  
lectotype.

**Referred material (277 specimens).** From quarry LH01: MPEF-Pb 7700, 7701. From LH02:  
MPEF-Pb 1053, 3168, 7702–7725, 7794–7818. From LH03: MPEF-Pb 7726, 7727. From  
300 LH04: MPEF-Pb 995, 2329, 3169, 7728–7762, 7819–7839, 7858–7864, 7870, 7871, 7881–  
7895. From LH06: MPEF-Pb 7763–7769, 7840–7846. From LH08: MPEF-Pb 7770. From  
LH13: MPEF-Pb 3171, 7771–7776, 7847–7854, 7896, 7897. From LH15: MPEF-Pb 7777,  
7865. From LH16: MPEF-Pb 7778, 7779, 7907. From LH17: MPEF-Pb 7780–7787. 7855–  
7857. From LH18: MPEF-Pb 7788. From LH20: MPEF-Pb 7789. From LH22: MPEF-Pb  
305 7790. From LH23: MPEF-Pb 7791, 7792. From LH25: MPEF-Pb 7866–7872. From LH27:  
MPEF-Pb 7869, 7873–7879, 7898. From LH29: MPEF-Pb 7899–7901. From float, unknown,  
or out-of-section locations (at MEF): MPEF-Pb 1449, 1450, 7793, 7880, 7902–7906. Exact  
collection site unknown, other: BA-Pb 12633; MJHG 45Pb–53Pb; Lil-Pb 5815, 5825, 5913,  
5920, 5931; LPPB 739, 741–746, 749, 759–762, 765–767, 773, 775, 781, 782, 898, 920, 943,

310 2470–2472, 2475–2477, 2483–2485, 2487, 2490, 2492, 2497–2502, 2504, 2506, 20417,  
20418, 20420, 20437.

**Emended specific description.** Leaves (Figs. 1–4) are opposite (Fig. 4.1–4.4), simple, and petiolate with marginal insertion. The lamina is unlobed, and the margin is strongly serrate. Petioles are eight to >23 mm in length and 0.8–2.0 mm wide. Laminar size (from Merkhofer  
315 *et al.* 2015,  $n = 101$ ) is 75–15,000 mm<sup>2</sup> (nanophyll to mesophyll), with a mean area of 1133 mm<sup>2</sup> (based on means of natural logs; microphyll). Laminar length is 2.5–14.5 cm, width is 1.0–10.2 cm, and length-width ratio is 1.1–2.9:1. Small axillary leaves are present (Fig. 4.1–4.4). The blade shape is ovate to lanceolate (Fig. 3.11), sometimes elliptic (Fig. 4.5), and symmetrical or slightly asymmetrical basally (*e.g.*, Fig. 3.1). The base is convex to slightly  
320 cordate (*e.g.*, Fig. 2.2) but may be cuneate (Fig. 3.12); the base angle is variably acute or obtuse. The apex shape is usually straight to slightly acuminate, sometimes convex, and the angle is acute. The overall leaf form varies with size (Figs. 1–3). Smaller leaves, such as the lectotype (Fig. 1.1), more often have a non-cordate base, ovate-lanceolate shape, and higher aspect ratios; larger leaves, such as the syntypes (Fig. 1.2, 1.3) tend to have broad-ovate  
325 blades with cordate bases and lower aspect ratios (*e.g.*, Fig. 2.1).

Primary venation is pinnate, and the midvein course is often deflected at secondary junctions. Secondary veins are decurrent in ca. 5–10 offset pairs, craspedodromous, and dichotomize up to 3–4 times. The secondary course is weak, moderately recurved, and often slightly deflected at junctions with minor secondary and tertiary veins. Secondary spacing  
330 decreases, and the secondary angle markedly increases basally (from ca. 35–55° to 70–100°). Most leaves have one or more abruptly thinner, high-angled or obtuse basal secondary pairs that diverge near the petiole insertion (*e.g.*, Fig. 2.2, Fig. 3.3). Agrophic veins are usually compound, and the first agrophic complex is restricted to a small area near the basal margin of the blade because of the high-angled basal secondaries that limit its potential extent. The

335 major and minor secondaries terminate at the tooth apex, often with perpendicular tertiary branches to vein junctions (vein knots) near or at the nadirs of the sinuses. A thin fimbrial vein is present (Fig. 2.3). Intersecondary veins are absent.

Tertiary venation (*e.g.*, Figs. 2.2, 3.4) is moderately spaced, thick, and usually weakly opposite percurrent but may be mixed percurrent or reticulate. The tertiary spacing and angle  
 340 are irregular. The departure of percurrent tertiaries from the midvein and major secondaries is generally perpendicular but inconsistently obtuse or acute. The tertiary angle to the midvein markedly increases toward the base. The tertiary course is usually convex but may be straight or sinuous or form a concentric pattern near the base. The tertiary distal course is basiflexed, and the junction with the subjacent secondaries is generally perpendicular. Tertiaries that  
 345 reach the margin terminate in the tooth apices or vein knots located at or near the sinuses (Fig. 2.3). The quaternary venation is weakly mixed percurrent to irregular reticulate, and the quinternary venation is regular to irregular reticulate. Areolation is well developed, with freely ending veinlets mostly one-branched at the sixth order.

The teeth (*e.g.*, Fig. 2.3) are irregular in size and always prominent, projecting from  
 350 the margin up to 4.4 mm (measured medially from the tooth apex to a line projected orthogonally from the basal sinus nadir). Teeth are present continuously over nearly the full blade margin, and the only untoothed area is the immediate vicinity of the petiole insertion. The teeth are simple or once compound, closely spaced (ca. 2–7 major teeth per cm), with angular sinuses. The tooth shape is generally flexuous with an apiculate apex; flank shapes  
 355 include flexuous/flexuous, flexuous/convex, straight/flexuous, convex/convex, straight/straight (triangular aspect), or straight/convex. The tooth apex is apiculate and simple (non-glandular; Fig. 2.3). The principal vein is a major secondary, minor secondary, or exterior tertiary vein that is usually deflected by vein junctions before entering the tooth. The principal vein is medial and prominent, and its course is slightly curved, basally deflected, or

nearly straight. The accessory veins emerge roughly perpendicular to the principal vein and form irregular loops, reticulate, or run toward the adjacent sinus nadir, joining a knot with other minor veins.

The insect damage types (Figs. 2.5, 3, 4.5–4.8) previously recorded (Wilf *et al.*, 2005b; Labandeira *et al.*, 2007; Sarzetti *et al.*, 2009) include external feeding (DTs 1–5, 7, 12–17, 22, 26, 29, 57), piercing-and-sucking (DT46), galling (DTs 32–34), mining (DTs 41, 90, 91), and oviposition (DTs 54, 76), to which we add an occurrence of mining DT93 on a specimen here identified to the species (Fig. 4.6, 4.7).

## DISCUSSION

### Affinities of *Dobineaites ameghinoi* comb. nov.

The characters of the fossils observed in the three type specimens (Fig. 1) are consistent with hundreds of subsequently collected fossils (Figs. 2–4), which fill the morphological continuum between the types (Fig. 1) and support Berry's (1925) hypothesis of a single species. Despite the elevated diversity of the Laguna del Hunco leaf flora, *Dobineaites ameghinoi* is rapidly identifiable based on the combination of obtuse basal secondary veins and prominent, densely spaced, apiculate, often flexuous, frequently compound teeth with non-glandular apices. The fossils can often be confidently identified even without the margin or base preserved (*e.g.*, Fig. 4.6) because of their distinctive deflected major veins, branching secondaries, and thick, usually weakly opposite percurrent, irregularly spaced, and angled tertiaries.

Our leaf architectural survey indicates that all groups previously considered for the fossils (*e.g.*, Berry, 1925) have only superficial similarities, such as the presence of an ovate blade and large or compound teeth in some species. Due to the substantial number of

compared taxa and the low similarity of nearly all of them to the fossils, we expedite the  
 385 discussion below using a set of cleared leaf exemplars as visual references (Fig. 5).

Despite their basionym, the fossils only superficially resemble *Celtis* (Fig. 5.1, 5.2).  
*Celtis* leaves are alternate and often have markedly asymmetrical petiole insertions and  
 laminae, unlike the fossils; the primaries may be pinnate like the fossils or palmate, usually  
 with three primaries. The basal pair of *Celtis* lateral primaries or secondaries is robust, often  
 390 runs on (naked basal veins) or close to the margin near the petiole insertion, and is usually  
 abruptly acute, set well below the next set of secondaries and generating prominent agrophic  
 veins that extend well up the blade. This architecture contrasts sharply with the secondaries  
 that basally decrease in spacing and increase in angle in the fossils (Figs. 2–4), leaving little  
 space for the basalmost set of agrophic veins to develop. *Celtis* secondary veins usually loop  
 395 inside the margin (Manchester *et al.*, 2002), unlike the craspedodromous secondary veins of  
 the fossils. *Celtis* leaves may be toothed or entire; however, when present, *Celtis* teeth are not  
 usually compound and never markedly compound like the fossils. This discussion also  
 applies to other Cannabaceae genera with comparable leaves (Fig. 5.3–5.5), such as  
*Aphananthe*, *Lozanella*, *Pteroceltis*, and *Trema*. Thus, we can confidently eliminate affinities  
 400 of the fossils to Cannabaceae. The only remaining South American *Celtis* fossils that are still  
 valid (*e.g.*, Manchester *et al.*, 2002) appear to be well-preserved *C. santosi* endocarps from  
 the Paleogene Itaboraí beds in Rio de Janeiro State, Brazil (Beurlen & Sommer, 1954).

Regarding Ulmaceae, Berry (1925) mentioned *Ampelocera* (Fig. 5.8) and  
*Phyllostylon*, but neither has many comparable features. Some Moraceae (*Morus*,  
 405 *Broussonetia*) and Urticaceae (*Boehmeria*, *Leucosyke*) have species with similar leaves to the  
 fossils, such as large, compound, or densely spaced teeth; however, they also have markedly  
 acute, robust basal secondaries and several other differences (Fig. 5.6, 5.7). In Rhamnaceae,  
 leaves most like the fossils typically have markedly acute, robust basal secondaries (or

acrodromous lateral primaries) that run close to or on the margin, set back from the next pair,  
 410 and generate well-developed agrophic veins over much of the blade, unlike the high-angled,  
 crowded, weak basal secondaries and reduced agrophic veins in the fossils. Rhamnaceae  
 secondary veins are strong, regular, and smoothly recurved apically rather than deflected like  
 the fossils; when present, the teeth are often reduced and glandular and rarely compound like  
 the fossils. Examples include the ziziphoid genera *Gouania* (Fig. 5.9), *Ceanothus*, and  
 415 *Paliurus*, which also have distinctive interior secondary venation not seen in these fossils (see  
 Wilf et al. 2022), as well as *Colubrina*, *Hovenia*, and *Ziziphus*.

In Rosaceae, many genera have species with large, densely spaced teeth superficially  
 similar to the fossils but few other shared features. *Rhodotypos scandens* (the sole species of  
 its genus, with opposite leaves) and *Rubus* species (Fig. 5.10, 5.11) have obtuse basal  
 420 secondaries and compound teeth; however, the secondary veins are more numerous and  
 regular and less deflected, and the often-glandular teeth do not resemble the fossils.  
 Moreover, *Rubus* species frequently have compound leaves and lobed leaf blades. Other  
 families of Rosales (as defined in Chase *et al.*, 2016) show no significant similarities to the  
 fossils, which also lack the characteristic glandular teeth that are widespread in the order  
 425 (Hickey & Wolfe, 1975). We conclude that the fossils do not belong to Rosales as long  
 suggested (Berry, 1925).

Outside Rosales, the leaves of some Euphorbiaceae species (Malpighiales; Fig. 5.12)  
 show only a superficial resemblance to the fossils. Malvaceae (Malvales) is also not suitable  
 for this material. The family tends to have alternate leaves as well as higher-ranked leaf  
 430 organization and other features that are unlike the fossils, such as palmate venation with  
 strong, markedly acute lateral primary veins or basal secondaries that emerge from the base;  
 well-developed and organized agrophic veins; and thin, closely spaced, regularly opposite  
 percurrent tertiaries (Hickey & Wolfe, 1975; Carvalho *et al.*, 2011; Wilf *et al.*, 2022). This

syndrome includes the taxa mentioned by Berry (1925), *Triumfetta* and *Grewia* (Fig. 5.14,  
 435 5.15), and many others such as *Melochia* (Fig. 5.13).

In Sapindales, entire-margined, compound-leaved taxa are prevalent, and very few  
 species remotely resemble the toothed, simple-leaved fossils. Some maples have some  
 similarities, such as *Acer tataricum* (Sapindaceae; Fig. 5.16), which has simple, lobed or  
 unlobed, cordate-ovate leaves and compound teeth. However, among other differences,  
 440 analogs of the frequent shallow to more incised lobes would be apparent in the large sample  
 size of the fossil species. Partial similarities to the fossils occur in some species of *Bursera*  
 (Burseraceae, e.g., *B. epinnata*) and *Brucea* (Simaroubaceae).

Like the other sapindalean families, the Anacardiaceae usually display entirely  
 dissimilar leaf architecture to these fossils, as described in surveys (Martínez-Millán &  
 445 Cevallos-Ferriz, 2005; Andrés-Hernández & Terrazas, 2009; Mitchell *et al.*, 2022). Many of  
 the more than 870 Anacardiaceae species have alternate, compound leaves with a  
 characteristic general appearance of the leaflets that is unlike the fossils, including various  
 combinations of asymmetrical blades (usually due to compound leaf organization), irregular  
 cladodromous or craspedodromous secondaries, secondaries and intersecondaries that  
 450 terminate in tooth sinuses, ramified to elaborately ramified tertiary veins, and irregularly  
 sized teeth when present. There are seven genera (*Schinus*, *Lithraea*, *Mauria*, *Astronium*,  
*Loxopterygium*, *Myracrodruon*, and *Schinopsis*) of the family in Argentina, none of which  
 resembles the fossils (but see Passalia *et al.*, 2019 for Miocene *Lithraea*). However, the leaf  
 architecture of Anacardiaceae encompasses diverse forms similar to other plant groups, and  
 455 many fossils of the family probably remain unrecognized for this reason (e.g., Passalia *et al.*,  
 2019). These include clusioid venation in species of *Ozoroa* and *Abrahamia* (which usually  
 have some degree of cladodromous secondary branching and ramified tertiaries); Myrtaceae-  
 like architecture in several *Spondias* species (which have asymmetrical blades and some

ramifications of the tertiaries); and several architectural similarities with the closely related families Sapindaceae and Burseraceae as well as Lauraceae, in the case of simple-leaved Anacardiaceae (Gentry, 1993). Only a few genera of Anacardiaceae have species with simple, toothed leaves, such as *Cotinus*, *Rhus*, and *Schinus* (the range of *S. roigii* includes the Laguna del Hunco site), none of which resembles the fossils, and *Dobinea* Buchanan-Hamilton (1825) ex D. Don (Figs. 5.17, 6), whose leaves provide unexpected and compelling similarities.

*Dobinea* is a dioecious Asian genus containing two species. The type species, *D. vulgaris* Buchanan-Hamilton (1825) ex D. Don., is opposite-leaved like the fossils but entirely dissimilar in leaf architecture (Fig. 6.4), whereas *D. delavayi* (Baillon) Baillon (1890), is alternate-leaved, like most Anacardiaceae species, but has leaf architecture identical to that of the fossils (Figs. 5.17, 6.1–6.3; Wu, 1979, 1986; Ming & Barfod, 2008). *Dobinea vulgaris* is a hill-forest dwarf shrub (elevation 1300–1400 m) with a broad, generally subtropical range, including Bangladesh, Assam, and the eastern Himalaya to south-central China and Myanmar, whereas *D. delavayi* is a perennial shrub-like herb with a more temperate, restricted distribution in grasslands and woodlands at 1100–2300 m elevation in southwestern Sichuan and central northwestern Yunnan (Ming, 1980; Ming & Barfod, 2008). Ming (1980) considered south-central China the likely area of origin for several anacardiaceous genera, including *Dobinea*. The most conspicuous feature of this genus, not yet observed in the fossil record, is its terminal female inflorescence containing accrescent, foliose, papery bracts. The bracts have venation and teeth similar to the regular leaves but include several sets of obtuse basal secondary veins. The pedicel is adnate to the bract midvein and bears unicarpellate, perianth-free flowers and subsequent small fruits (Fig. 6.1; Baillon, 1887; Engler, 1896; Ming & Barfod, 2008; Pell *et al.*, 2010). These elm-like winged



fruits enable wind dispersal, and the genus is wind-pollinated, which is unusual in the Anacardiaceae (Pell *et al.*, 2010).

485           The systematic placement of *Dobinea* has varied historically among a separate family (Podoaceae, along with *Campylopetalum*, a monotypic herb endemic to Thailand), Sapindaceae (as a member of the former Aceraceae), and Anacardiaceae. Molecular analyses have resolved the two species of *Dobinea* as a monophyletic group in Anacardiaceae, estimated the species divergence at ca. 10.8 Ma, and estimated the genus divergence as 40–50  
490 Ma, similar to the age of these fossils (Pan *et al.*, 2008; Pell *et al.*, 2010; Weeks *et al.*, 2014; Liu *et al.*, 2021; Joyce *et al.*, 2023). *Campylopetalum* has resolved as a sister genus to *Dobinea* (Weeks *et al.*, 2014; Joyce *et al.*, 2023), and the two genera together resolved as sister to the large A2 clade of Joyce *et al.* (2023; the A2 clade is equivalent to the Anacardioideae with *Campnosperma* removed). *Campylopetalum* and *Dobinea* share winged  
495 fruits, chromosome number ( $n = 7$ ), and several pollen characters, such as a small overall size of ca. 10–15  $\mu\text{m}$ , suboblate-oblate spheroidal shape, three (four) compound apertures, and exine with reticulate sculpture (Erdtman, 1952; Forman, 1953; Pell *et al.*, 2010).

Comparisons of vegetative characters show close similarities between the fossils and *Dobinea* (Figs. 5.17, 6), especially *D. delavayi* (羊角天麻, roughly translated as Goat's Horn  
500 Weed). Stipules are absent in *Dobinea*, as for Anacardiaceae as a whole (Pell *et al.*, 2010); however, we observed stipule-like small or immature leaves that are often present on axils or axillary shoots in both extant species (Fig. 6.3, 6.4) and the fossils (Fig. 4.1–4.4). *Dobinea* leaves are simple, petiolate, unlobed, toothed, and either opposite, with rounded to cuneate bases, oblong to lanceolate blade shape, and a regularly serrulate margin (*D. vulgaris*), or  
505 alternate, with cordate to convex bases, ovate to lanceolate blade shape, acuminate apex, and an irregularly serrated margin, the leaf size decreasing markedly along the shoot (*D. delavayi*; Fig. 6.1, 6.3; Baillon, 1887; Wu, 1986, 1988; Ming & Barfod, 2008). Interestingly, in the

basionym description of the species (as *Podoon delavayi* Baillon), Baillon (1887: 682) noted,  
 “*Ses organes de végétation sont ceux d'un Morus ou d'un Celtis*” (its vegetative organs are  
 510 those of a *Morus* or a *Celtis*).

Like *D. delavayi*, the fossil leaves are petiolate, symmetrical, or slightly asymmetrical  
 at the base. In both the fossil and living species, the primary and secondary venation is weak  
 and frequently deflected; there is usually a thin, basal pair of secondaries diverging near the  
 petiole insertion, the secondaries are craspedodromous and often dichotomize inside the  
 515 margin, intersecondaries are absent, and the secondary vein angle increases markedly towards  
 the base. Agrophic veins are compound and slightly irregular, and the high-angled basal  
 secondaries restrict the basalmost set to a small portion of the basal margin. Tertiary veins are  
 weakly percurrent, moderately and irregularly spaced, and become more obtuse basally.  
 Higher-order venation is reticulate, and mostly once-branched freely ending veinlets are  
 520 present. The tooth features are also the same (*e.g.*, Fig. 6.2), including coverage nearly to the  
 blade base, close spacing, irregular sizes, angular sinuses, simple or compound organization,  
 flexuous and other flank shapes, and non-glandular, apiculate apices (which appear thickened  
 due to abrupt apical narrowing around the principal vein but have no glandular tissue). The  
 tooth venation includes a prominent medial vein that generates perpendicular branches to  
 525 vein knots inside or at the sinuses.

Additional features of the fossils are also consistent with *Dobinea delavayi*. With  
 decreasing blade size, the fossils generally show increasing aspect ratios and a gradation from  
 cordate to ovate-lanceolate blade shapes (*e.g.*, Fig. 3), as seen distally along single branches  
 of *D. delavayi* (Fig. 6.1). Moreover, the discovery of a single fossil specimen with attached  
 530 leaves (Fig. 4.1–4.4) confirms that the fossil species had simple leaves and shows small  
 axillary leaves, a feature often seen in both living *Dobinea* species (Fig. 6.3, 6.4). Perhaps the  
 only difference between the fossils and *D. delavayi* is that the fossil leaves are opposite (Fig.

4.1), a rare trait in Anacardiaceae that nonetheless occurs in the other living species in the genus (*D. vulgaris*). We conclude that the fossil leaves of *D. ameghinoi* are strikingly similar to those of extant *D. delavayi* and only superficially comparable with the other taxa examined (Fig. 5), which nearly all have strong, acute lateral primaries or secondaries emerging from the petiole insertion (vs. obtuse, weak basal secondaries in the fossils) and robust major-vein courses (vs. deflected major veins and bifurcating secondaries in the fossils).

Although we consider the leaf fossils to be closely related to living *Dobinea*, we maintain caution by placing them in the new extinct genus *Dobineaites* due to the absence to date of other evidence for *Dobinea* at the site, including the distinctive winged fruits (Fig. 6.1). *Dobinea*-like pollen (Radlkofer, 1888; Erdtman, 1952; Forman, 1953) was also not found in the dispersed associated palynoflora (Barreda *et al.*, 2020), although Anacardiaceae grains are rare in the assemblage overall, and many other abundant macrofossil taxa in the flora are not yet matched with associated palynotaxa (Barreda *et al.*, 2020). More circumstantially, the impressive abundance of the fossil leaves, suggesting high biomass, as discussed earlier, appears to be inconsistent with the herb and shrub habits of the living *Dobinea* species.

## Contribution to the Anacardiaceae fossil record

*Dobineaites ameghinoi* fossils from Laguna del Hunco represent a rare macrofossil occurrence of Anacardiaceae in Gondwana, the first report of a fossil allied with *Dobinea*, and one of the few fossil occurrences affiliated with living Asian-endemic Anacardiaceae. The Anacardiaceae have a rich macrofossil record, especially from the Eocene onwards in North America and Europe (e.g., Manchester, 1999; Manchester *et al.*, 2009; Herrera *et al.*, 2012; Mitchell *et al.*, 2022; Del Rio *et al.*, 2023; Joyce *et al.*, 2023), but they have little

representation in Gondwana. Examples of numerous Laurasian Paleogene occurrences based on reproductive materials include fossil cashew nuts (*Anacardium*) with characteristic inflated hypocarps from the middle Eocene Messel locality (Germany; Manchester *et al.*, 2007), the extinct fruit genus *Pentoperculum* (Spondaeae) from the early Eocene London Clay and the middle Eocene Clarno flora of Oregon (Reid & Chandler, 1933; Manchester, 1994), and *Rhus* fruits from Clarno (Manchester & Judd, 2022). *Rhus* has a rich record of fossil leaves since the Eocene, although much of the foliage may belong to extinct genera (Manchester & Judd, 2022). Paleogene leaf records attributed to *Rhus* and other genera mostly come from several Eocene sites in the western USA and the Oligocene of Puebla, Mexico (*e.g.*, MacGinitie, 1953, 1969; Wolfe & Wehr, 1987; Meyer & Manchester, 1997; Manchester, 2001; Ramírez & Cevallos-Ferriz, 2002; Flynn *et al.*, 2019; Mitchell *et al.*, 2022). Reliable fossil evidence for living endemic genera of Asian Anacardiaceae includes *Choerospondias* endocarps from the London Clay and several Oligocene and younger sites in Eastern Europe, China, and Japan (summarized by Manchester *et al.*, 2009; Wang *et al.*, 2020), along with *Dracontomelon* endocarps from the late Eocene of Panamá (Herrera *et al.*, 2012).

There are almost no macrofossil records of Anacardiaceae in the Southern Hemisphere outside of South America. One important exception is fossil wood consistent with the Anacardioideae subfamily (*Anacardioxylon semecarpoides*; Wheeler *et al.*, 2017) from the ca. 66 Ma Deccan sequence in Maharashtra, India, which was located in the Southern Hemisphere tropics at the time, long after India had separated from Gondwana. Several palynological occurrences have been resolved at the family level in Australia (*e.g.*, compilation in Kooyman *et al.*, 2014).

In Gondwanan South America, putative occurrences of *Anacardium* nuts from Perú, Ecuador, and Colombia, also reported by Berry in the 1920s (Berry, 1924a, 1924b, 1929a,

1929b), are doubtful because of the lack of attached hypocarps (Manchester *et al.*, 2007). As reviewed by Burnham and Carranco (2004), there are numerous historical reports of putative fossil anacardiaceous foliage from several South American countries that should be  
 585 considered unconfirmed, including "*Schinopsis*" *patagonica* from Laguna del Hunco (Berry, 1925) and several species from Río Pichileufú (Berry, 1938). Post-Gondwanan occurrences in northern South America include fossil wood with features of *Anacardium* and *Mangifera* from the late Eocene Piedra Chamana Fossil Forest of Perú (Woodcock *et al.*, 2017) and a winged fruit of *Loxopterygium* from Ecuador (Burnham & Carranco, 2004).

590 Nevertheless, there is growing evidence for the presence of Anacardiaceae in southern South America since the early Paleocene. At Laguna del Hunco, supporting data include pollen occurrences of the family mentioned earlier (Barreda *et al.*, 2020) and several other leaf morphotypes that appear to belong to Sapindales and potentially Anacardiaceae (Wilf *et al.*, 2005a). An older record, and potentially the oldest macrofossil occurrence worldwide,  
 595 consists of isolated foliage with characteristic admedially ramified venation from the early Paleocene (early Danian) Salamanca Formation in southern Chubut (Iglesias *et al.*, 2021: morphotype SA050), which was validated in a recent specialist review (Mitchell *et al.*, 2022). The Salamanca Formation has also yielded anacardiaceous pollen, as has the Danian portion of the Lefipán Formation in northwest Chubut (see compilation in Kooyman *et al.*, 2014).

600 Post-Gondwanan Anacardiaceae in the Southern Cone include diverse fossil-wood occurrences (none related to *Dobinea*) from Oligocene to Pleistocene strata of Argentina (e.g., Lutz, 1979; Franco, 2009; Pujana, 2009; reviewed by Pujana, 2022). Additional leaf records attributed to Anacardiaceae come from the ?Paleogene of Chile (Troncoso, 1992) and several Neogene sites in Chile and Argentina (e.g., Troncoso and Encinas 2006; Anzótegui *et al.*, 2007). However, per Burnham and Carranco (2004), additional documentation is needed  
 605 to confirm some of these occurrences as Anacardiaceae. One notable revision is *Lithraea*

*australis* from the Miocene Ñirihuau Formation in Río Negro, Argentina, which Passalia *et al.* (2019) recently transferred from its historical assignment as foliage of the cycad *Zamia* (Berry, 1928).

610

### Paleoecology and biogeography

The diverse Eocene Laguna del Hunco rainforest had multiple strata, from a rich understory with, among many other taxa, ferns, Asteraceae, Solanaceae, and Rubiaceae, to large angiosperm trees of Myrtaceae and Fagaceae and probable coniferous emergents in the Cupressaceae, Araucariaceae, and Podocarpaceae (Gandolfo *et al.*, 2011; Wilf *et al.*, 2014, 2019; Barreda *et al.*, 2020; Deanna *et al.*, 2020; Pujana *et al.*, 2020). The dominance by leaf count of *Dobineaites ameghinoi* suggests high original biomass near the depocenter (*e.g.*, Burnham *et al.*, 1992; Burnham, 1997), comparable to the conifers in the assemblage and *Eucalyptus*, and contrasts sharply with the shrubby or perennial herbaceous habit of extant *Dobinea*, which inhabits more open and seasonally dry environments. These observations indicate that the ancient species had different ecological roles from its living relatives. To the best of our knowledge, there are no published data on *Dobinea* folivore damage for comparison with the fossils. An herbarium survey could be productive for testing the idea of host tracking through time (*e.g.*, Donovan *et al.*, 2023); however, insect mines appear to be very rare on hundreds of *Dobinea* herbarium sheets examined, suggesting that a field study would be necessary.

The proposed affinities of the *D. ameghinoi* fossils to an Asian endemic genus are striking but not surprising because numerous other lineages with extant ranges on the Asian mainland and in maritime Southeast Asia co-occur with *D. ameghinoi* in the same fossil beds at Laguna del Hunco, including *Agathis* (Araucariaceae), *Dacrycarpus* (Podocarpaceae),

630

*Castanopsis* (Fagaceae), *Macaranga* (Euphorbiaceae), and engelhardioid Juglandaceae (Wilf, 2012; Wilf *et al.*, 2014, 2019, 2023; Hermsen & Gandolfo, 2016). Many other Laguna del Hunco genera reach Australasia and Malesia but not the Asian mainland, as summarized elsewhere (*e.g.*, Wilf *et al.*, 2013, 2023; Kooyman *et al.*, 2014). Similarly, Weeks *et al.* (2014) found from molecular data that long-distance movements characterize the entire evolutionary history of Anacardiaceae. The floristic connections from ancient Patagonia to modern Asia are thought to represent post-Gondwanan lineage survival on the Australian plate (Sahul), followed by exchange with Asia during the Neogene Sahul-Sunda collision (Wilf *et al.*, 2013; Kooyman *et al.*, 2014, 2019; Wilf & Kooyman, 2023). No reliable macrofossils of Laguna del Hunco genera have yet been found in Paleogene (*i.e.*, pre-collision) Asia that would contradict this idea.

## CONCLUSIONS

Fossil leaves known since the 1920s as "*Celtis*" *ameghinoi* are the most abundant plant fossils at the extraordinarily diverse Laguna del Hunco locality, early Eocene of Patagonian Argentina. Based on their well-preserved leaf architecture, we could not place these fossils in any previously considered taxa, including the Cannabaceae or any Rosales family. However, we found compelling similarities with the Asian genus *Dobinea* (Anacardiaceae), particularly *D. delavayi*, which is endemic to south-central China. Given its high abundance, *Dobineaites ameghinoi* comb. nov. probably had high biomass, unlike the small-statured living *Dobinea* species, and its diverse insect damage also indicates its paleoecological significance. Our analysis presents a new floristic link from West Gondwana to mainland Asia, as seen in several angiosperm and conifer genera at the same fossil site, and a rare Gondwanan macrofossil record of Anacardiaceae.

655

## ACKNOWLEDGMENTS

We thank scientists and staff at many institutions for assistance with fossils and extant material, including MEF: Rubén Cúneo, Pablo Puerta, Mariano Caffa, Eduardo Ruigomez, Leandro Canessa, Laura Reiner, Ignacio Escapa, Magalí Cárdenas; NMNH: Jon Wingerath, 660 Scott Wing; LIL: Hugo Carrizo; MLP: Eduardo Morel, Analía Artabe, Ari Iglesias, Patricio Knight, the late Alba B. Zamuner; BAB: Renée Fortunato; BH: Kevin Nixon, Jennifer Svitko; SI: Norma B. Deginani; and MJHG: J. Heredia. We also thank Kirk Johnson, Scott Wing, and numerous other participants in field trips; Secretaría de Ciencia y Cultura del Chubut and the Nahueltripay Family for land access; Lisa Merkhofer for assistance with the descriptions; 665 Conrad Labandeira for early involvement with the fossil insect damage (as cited); Ignacio Escapa, Lisa Merkhofer, Bárbara Cariglino, and Rebecca Horwitt for assistance with photography at MEF; Tengxiang Wang for photography at K (Fig. 6.2) and guidance with the Chinese literature; Ari Iglesias for photography at MJHG (Fig. 3.12); Steven Manchester, John Mitchell, Douglas Daly, and Editor Camila Martínez for helpful reviews; Viviana 670 Barreda, Douglas Daly, John Mitchell, and Edgardo Romero for discussions; and the support staff of virtual herbaria around the world (see Methods). Recent financial support was provided by National Science Foundation (NSF) grants EAR-1925755, EAR-1925552, DEB-1556666, and DEB-1556136. Earlier support came from NSF grants DEB-0345750, DEB-0919071, DEB-0918932, and National Geographic Society grant 7337-02.



## 675 REFERENCES

- Andrés-Hernández, A. R., & Terrazas, T. (2009). Leaf architecture of *Rhus* s.str. (Anacardiaceae). *Feddes Repertorium*, 120(5–6), 293–306.
- Andruchow-Colombo, A., Rossetto-Harris, G., Brodribb, T. J., Gandolfo, M. A., & Wilf, P. (2023). A new fossil *Acmopyle* with accessory transfusion tissue and potential reproductive buds: direct evidence for ever-wet rainforests in Eocene Patagonia. *American Journal of Botany*, 110(8), e16221.
- 680 Anzótegui, L. M., Horn, Y., & Herbst, R. (2007). Paleoflora (Fabaceae y Anacardiaceae) de la Formación Andalhuala (Plioceno Inferior), provincia de Catamarca, Argentina. *Ameghiniana*, 44(3), 525–535.
- 685 Aragón, E., Aguilera, Y. E., González, P. D., Gómez Peral, L., Cavarozzi, C. E., & Ribot, A. (2001). El Intrusivo Florentina del Complejo Volcánico Piroclástico del río Chubut medio (Paleoceno-Eoceno medio): un ejemplo de etmolito o embudo. *Revista de la Asociación Geológica Argentina*, 56(2), 161–172.
- Aragón, E., Castro, A., Diaz-Alvarado, J., Pinotti, L., D'Eramo, F., Demartis, M., Coniglio, J., Hernando, I., & Rodríguez, C. (2018). Mantle derived crystal-poor rhyolitic ignimbrites: eruptive mechanism from geochemical and geochronological data of the Piedra Parada caldera, southern Argentina. *Geoscience Frontiers*, 9(5), 1529–1553.
- 690 Aragón, E., González, P. D., Aguilera, Y., Marquett, C., Cavarozzi, C., & Ribot, A. (2004). El domo vitrofírico Escuela Piedra Parada del Complejo Volcánico Piroclástico del Río Chubut medio. *Revista de la Asociación Geológica Argentina*, 59(4), 634–642.
- 695 Aragón, E., & Mazzoni, M. M. (1997). Geología y estratigrafía del complejo volcánico piroclástico del río Chubut medio (Eoceno), Chubut, Argentina. *Revista de la Asociación Geológica Argentina*, 52(3), 243–256.

- Baillon, H. E. (1887). Un nouveau type apérianthé. *Bulletin Mensuel de la Société Linnéenne de Paris*, 1(86), 681–683.
- Baillon, H. E. (1890). Les rapports du *Podoon* et du *Dobinea*. *Bulletin Mensuel de la Société Linnéenne de Paris*, 2(105), 834–835.
- Barreda, V., & Palazzesi, L. (2007). Patagonian vegetation turnovers during the Paleogene-early Neogene: origin of arid-adapted floras. *Botanical Review*, 73(1), 31–50.
- Barreda, V., & Palazzesi, L. (2010). Vegetation during the Eocene-Miocene interval in central Patagonia: a context of mammal evolution. In R. H. Madden, A. A. Carlini, M. G. Vucetich, & R. F. Kay (Eds.), *The Paleontology of Gran Barranca* (pp. 375–382). Cambridge University Press.
- Barreda, V. D., Zamaloa, M. C., Gandolfo, M. A., Jaramillo, C., & Wilf, P. (2020). Early Eocene spore and pollen assemblages from the Laguna del Hunco fossil lake beds, Patagonia, Argentina. *International Journal of Plant Sciences*, 181(6), 594–615.
- Berry, E. W. (1924a). New Tertiary species of *Anacardium* and *Vantanea* from Colombia. *Pan American Geologist*, 42, 259–262.
- Berry, E. W. (1924b). An Oligocene cashew nut from South America. *American Journal of Science*, 8(44), 123–126.
- Berry, E. W. (1925). A Miocene flora from Patagonia. *Johns Hopkins University Studies in Geology*, 6, 183–251.
- Berry, E. W. (1928). Tertiary fossil plants from the Argentine Republic. *Proceedings of the United States National Museum*, 73(22), 1–27.
- Berry, E. W. (1929a). Early Tertiary fruits and seeds from Belén, Peru. *Johns Hopkins University Studies in Geology*, 10, 137–172.
- Berry, E. W. (1929b). Fossil fruits in the Ancon sandstone of Ecuador. *Journal of Paleontology*, 3(3), 298–301.

- Berry, E. W. (1938). Tertiary flora from the Río Pichileufú, Argentina. *Geological Society of America Special Papers*, 12, 1–149.
- Beurlen, K., & Sommer, F. W. (1954). Restos vegetais fósseis e tectônica da Bacia Calcárea de Itaboraí: Estado do Rio de Janeiro. *Boletim do Departamento Nacional de Produção Mineral, Divisão de Geologia e Mineralogia*, 149, 1–27.
- Brown, R. (1818). Appendix No. V. In J. H. Tuckey & S. Smith (Eds.), *Narrative of an Expedition to Explore the River Zaire* (pp. 420–485). John Murray.
- Buchanan-Hamilton, F. (1825). *Prodromus Florae Nepalensis*. J. Gale.
- Burnham, R. J. (1997). Stand characteristics and leaf litter composition of a dry forest hectare in Santa Rosa National Park, Costa Rica. *Biotropica*, 29(4), 384–395.
- Burnham, R. J., & Carranco, N. L. (2004). Miocene winged fruits of *Loxopterygium* (Anacardiaceae) from the Ecuadorian Andes. *American Journal of Botany*, 91(11), 1767–1773.
- Burnham, R. J., Wing, S. L., & Parker, G. G. (1992). The reflection of deciduous forest communities in leaf litter: implications for autochthonous litter assemblages from the fossil record. *Paleobiology*, 18(1), 30–49.
- Carpenter, R. J., Wilf, P., Conran, J. G., & Cúneo, N. R. (2014). A Paleogene trans-Antarctic distribution for *Ripogonum* (Ripogonaceae: Liliales)? *Palaeontologia Electronica*, 17(3), 39A.
- Carvalho, M. R., Herrera, F. A., Jaramillo, C. A., Wing, S. L., & Callejas, R. (2011). Paleocene Malvaceae from northern South America and their biogeographical implications. *American Journal of Botany*, 98(8), 1337–1355.
- Chase, M. W., Christenhusz, M. J. M., Fay, M. F., Byng, J. W., Judd, W. S., Soltis, D. E., Mabberley, D. J., Sennikov, A. N., Soltis, P. S., Stevens, P. F., & The Angiosperm Phylogeny Group (2016). An update of the Angiosperm Phylogeny Group

- classification for the orders and families of flowering plants: APG IV. *Botanical*  
750 *Journal of the Linnean Society*, 181(1), 1–20.
- Clark, B. (1923). *The Economic Deduction from an Incidental General Reconnaissance of*  
*Patagonia between East Central Santa Cruz and Northwestern Chubut*. ESSO. Field  
Office Argentine No. 35, Office Argentine No. 70.
- Deanna, R., Wilf, P., & Gandolfo, M. A. (2020). New physaloid fruit-fossil species from early  
755 Eocene South America. *American Journal of Botany*, 107(12), 1749–1762.
- Del Rio, C., Tosal, A., Kara, E., Manchester, S. R., Herrera, F., Collinson, M. E., & De  
Franceschi, D. (2023). Fruits of Anacardiaceae from the Paleogene of the Paris Basin,  
France. *International Journal of Plant Sciences*, 184(3), 164–176.
- Dilcher, D. L. (1971). A revision of the Eocene flora of southeastern North America. *The*  
760 *Palaeobotanist*, 20(1), 7–18.
- Dolgopol de Sáez, M. (1941). Noticias sobre peces fósiles Argentinos. Siluroideos Terciarios  
del Chubut. *Notas del Museo de La Plata, Paleontologia*, 6, 451–457.
- Donovan, M. P., Wilf, P., Iglesias, A., Cúneo, N. R., & Labandeira, C. C. (2023). Insect  
herbivore and fungal communities on *Agathis* (Araucariaceae) from the latest  
765 Cretaceous to Recent. *PhytoKeys*, 226, 109–158.
- Ellis, B., Daly, D. C., Hickey, L. J., Johnson, K. R., Mitchell, J. D., Wilf, P., & Wing, S. L.  
(2009). *Manual of Leaf Architecture*. Cornell University Press.
- Engler, A. (1896). Anacardiaceae. In A. Engler & K. Prantl (Eds.), *Die Natürlichen*  
*Pflanzenfamilien III*, 5 (pp. 138–178). Engelmann.
- 770 Erdtman, G. (1952). *Pollen Morphology and Plant Taxonomy: Angiosperms*. Chronica  
Botanica.

- Flynn, S., DeVore, M. L., & Pigg, K. B. (2019). Morphological features of sumac leaves (*Rhus*, Anacardiaceae), from the latest early Eocene flora of Republic, Washington. *International Journal of Plant Sciences*, 180(6), 464–478.
- 775 Forman, L. L. (1953). A new genus from Thailand. *Kew Bulletin*, 8(4), 555–564.
- Franco, M. J. (2009). Leños fósiles de Anacardiaceae en la Formación Ituzaingó (Plioceno-Pleistoceno), Toma Vieja, Paraná, Entre Ríos, Argentina. *Ameghiniana* 46(4), 587–604.
- Frenguelli, J. (1943a). Proteáceas del Cenozoico de Patagonia. *Notas del Museo de La Plata*, 780 8, 201–213.
- Frenguelli, J. (1943b). Restos de *Casuarina* en el Mioceno de El Mirador, Patagonia central. *Notas del Museo de La Plata*, 8(56), 349–354.
- Gandolfo, M. A., Hermsen, E. J., Zamaloa, M. C., Nixon, K. C., González, C. C., Wilf, P., Cúneo, N. R., & Johnson, K. R. (2011). Oldest known *Eucalyptus* macrofossils are 785 from South America. *PLOS ONE*, 6(6), e21084.
- Gentry, A. H. (1993). *A Field Guide to the Families and Genera of Woody Plants of Northwest South America (Colombia, Ecuador, Peru)*. Conservation International.
- González, C. C. (2008). *Revisión taxonómica y biogeográfica de las familias de angiospermas dominantes de la "Flora del Hunco" (Eoceno temprano), Chubut*, 790 *Argentina* (Doctoral thesis, Universidad de Buenos Aires, Buenos Aires).
- González, C. C., Gandolfo, M. A., Zamaloa, M. C., Cúneo, N. R., Wilf, P., & Johnson, K. R. (2007). Revision of the Proteaceae macrofossil record from Patagonia, Argentina. *Botanical Review*, 73(3), 235–266.
- Gosses, J., Carroll, A. R., Bruck, B. T., Singer, B. S., Jicha, B. R., Aragón, E., Walters, A. P., 795 & Wilf, P. (2021). Facies interpretation and geochronology of diverse Eocene floras

and faunas, northwest Chubut Province, Patagonia, Argentina. *Geological Society of America Bulletin*, 133(3–4), 740–752.

Hermesen, E. J., & Gandolfo, M. A. (2016). Fruits of Juglandaceae from the Eocene of South America. *Systematic Botany*, 41(2), 316–328.

800 Hermesen, E. J., Gandolfo, M. A., & Zamaloa, M. C. (2012). The fossil record of *Eucalyptus* in Patagonia. *American Journal of Botany*, 99(8), 1356–1374.

Herrera, F., Manchester, S. R., & Jaramillo, C. (2012). Permineralized fruits from the late Eocene of Panama give clues of the composition of forests established early in the uplift of Central America. *Review of Palaeobotany and Palynology*, 175, 10–24.

805 Hickey, L. J., & Wolfe, J. A. (1975). The bases of angiosperm phylogeny: vegetative morphology. *Annals of the Missouri Botanical Garden*, 62(3), 538–589.

Hill, R. S. (1982). The Eocene megafossil flora of Nerriga, New South Wales, Australia. *Palaeontographica Abt. B*, 181(1–3), 44–77.

Iglesias, A., Wilf, P., Stiles, E., & Wilf, R. (2021). Patagonia's diverse but homogeneous early  
810 Paleocene forests: angiosperm leaves from the Danian Salamanca and Peñas  
Coloradas formations, San Jorge Basin, Chubut, Argentina. *Palaeontologia  
Electronica*, 24(1), a02.

Joyce, E. M., Appelhans, M. S., Buerki, S., Cheek, M., de Vos, J. M., Pirani, J. R., Zuntini, A.  
R., Bachelier, J. B., Bayly, M. J., Callmander, M. W., Devecchi, M. F., Pell, S. K.,  
815 Groppo, M., Lowry, P. P., II, Mitchell, J., Siniscalchi, C. M., Munzinger, J., Orel, H.  
K., Pannell, C. M., Nauheimer, L., Sauquet, H., Weeks, A. Muellner-Riehl, A. N.,  
Leitch, I. J., Maurin, O., Forest, F., Nargar, K., Thiele, K. R., Baker, W. J., & Crayn,  
D. M. (2023). Phylogenomic analyses of Sapindales support new family relationships,  
rapid Mid-Cretaceous Hothouse diversification, and heterogeneous histories of gene  
820 duplication. *Frontiers in Plant Science*, 14, 1063174.

- Jud, N. A., Iglesias, A., Wilf, P., & Gandolfo, M. A. (2018). Fossil moonseeds from the Paleogene of West Gondwana (Patagonia, Argentina). *American Journal of Botany*, 105(5), 927–942.
- 825 Knight, C. L., & Wilf, P. (2013). Rare leaf fossils of Monimiaceae and Atherospermataceae (Laurales) from Eocene Patagonian rainforests and their biogeographic significance. *Palaeontologia Electronica*, 16(3), 26A.
- Kooyman, R. M., Morley, R. J., Crayn, D. M., Joyce, E. M., Rossetto, M., Slik, J. W. F., Strijk, J. S., Su, T., Yap, J.-Y. S., & Wilf, P. (2019). Origins and assembly of Malesian rainforests. *Annual Review of Ecology, Evolution, and Systematics*, 50, 119–143.
- 830 Kooyman, R. M., Wilf, P., Barreda, V. D., Carpenter, R. J., Jordan, G. J., Sniderman, J. M. K., Allen, A., Brodribb, T. J., Crayn, D., Feild, T. S., Laffan, S. W., Lusk, C. H., Rossetto, M., & Weston, P. H. (2014). Paleo-Antarctic rainforest into the modern Old World Tropics: the rich past and threatened future of the "southern wet forest survivors." *American Journal of Botany*, 101(12), 2121–2135.
- 835 Labandeira, C. C., Wilf, P., Johnson, K. R., & Marsh, F. (2007). *Guide to Insect (and other) Damage Types on Compressed Plant Fossils. Version 3.0*. Smithsonian Institution. Available at <https://doi.org/10.6084/m9.figshare.16571441.v1>
- Liu, C., Yang, J., Jin, L., Wang, S., Yang, Z., & Ji, Y. (2021). Plastome phylogenomics of the East Asian endemic genus *Dobinea*. *Plant Diversity*, 43(1), 35–42.
- 840 Lutz, A. I. (1979). Maderas de angiospermas (Anacardiaceae y Leguminosae) del Plioceno de la provincia de Entre Ríos, Argentina. *FACENA*, 3, 39–63.
- MacGinitie, H. D. (1953). Fossil plants of the Florissant beds, Colorado. *Carnegie Institution of Washington Publications*, 599, 1–198.

- 845 MacGinitie, H. D. (1969). The Eocene Green River flora of northwestern Colorado and  
northeastern Utah. *University of California Publications in Geological Sciences*, 83,  
1–203.
- Machado, M. A., Passalia, M. G., Vera, E. I., & Yañez, A. (2023). Ferns from the Arroyo  
Chacay flora (Huitrera Formation, Eocene) Río Negro Province, Argentina. *Review of*  
850 *Palaeobotany and Palynology*, 313, 104892.
- Manchester, S. R. (1994). Fruits and seeds of the middle Eocene Nut Beds flora, Clarno  
Formation, Oregon. *Palaeontographica Americana*, 58, 1–205.
- Manchester, S. R. (1999). Biogeographical relationships of North American Tertiary floras.  
*Annals of the Missouri Botanical Garden*, 86(2), 472–522.
- 855 Manchester, S. R. (2001). Update on the megafossil flora of Florissant, Colorado.  
*Proceedings of the Denver Museum of Nature & Science, Series 4, 1*, 137–161.
- Manchester, S. R., Akhmetiev, M. A., & Kodrul, T. M. (2002). Leaves and fruits of *Celtis*  
*aspera* (Newberry) comb. nov. (Celtidaceae) from the Paleocene of North America  
and eastern Asia. *International Journal of Plant Sciences*, 163(5), 725–736.
- 860 Manchester, S. R., Chen, Z.-D., Lu, A.-M., & Uemura, K. (2009). Eastern Asian endemic  
seed plant genera and their paleogeographic history throughout the Northern  
Hemisphere. *Journal of Systematics and Evolution*, 47(1), 1–42.
- Manchester, S. R., & Judd, W. S. (2022). Extinct anacardiaceous samaras and sumac-like  
leaves from the Eocene of western North America. *International Journal of Plant*  
865 *Sciences*, 183(5), 357–366.
- Manchester, S. R., Wilde, V., & Collinson, M. E. (2007). Fossil cashew nuts from the Eocene  
of Europe: biogeographic links between Africa and South America. *International*  
*Journal of Plant Sciences*, 168(8), 1199–1206.



- Martínez-Millán, M., & Cevallos-Ferriz, S. R. S. (2005). Arquitectura foliar de  
 870       Anacardiaceae. *Revista Mexicana de Biodiversidad*, 76(2), 137–190.
- Merkhofer, L., Wilf, P., Haas, M. T., Kooyman, R. M., Sack, L., Scoffoni, C., & Cúneo, N. R.  
 (2015). Resolving Australian analogs for an Eocene Patagonian paleorainforest using  
 leaf size and floristics. *American Journal of Botany*, 102(7), 1160–1173.
- Meyer, H. W., & Manchester, S. R. (1997). The Oligocene Bridge Creek flora of the John  
 875       Day Formation, Oregon. *University of California Publications in Geological  
 Sciences*, 141, 1–195.
- Ming, T.-L. (1980). The geographic distribution and floristic character of Chinese  
 Anacardiaceae. *Acta Botanica Yunnanica*, 2(4), 390–401.
- Ming, T.-L., & Barfod, A. (2008). Anacardiaceae. *Flora of China*, 11, 335–357.
- 880       Mitchell, J. D., Pell, S. K., Bachelier, J. B., Warschefsky, E. J., Joyce, E. M., Canadell, L. C.,  
 da Silva-Luz, C. L., & Coiffard, C. (2022). Neotropical Anacardiaceae (cashew  
 family). *Brazilian Journal of Botany*, 45(1), 139–180.
- Pan, Y.-Z., Gong, X., & Yang, Y. (2008). Phylogenetic position of the genus *Dobinea*:  
 Evidence from nucleotide sequences of the chloroplast gene *rbcL* and the nuclear  
 885       ribosomal ITS region. *Journal of Systematics and Evolution*, 46(4), 586–594.
- Passalia, M. G., Caviglia, N., & Vera, E. I. (2019). *Lithraea australis* (Berry) comb. nov.  
 (Anacardiaceae) from the upper section of Ñirihuau Formation (middle Miocene),  
 Patagonia. *Review of Palaeobotany and Palynology*, 266, 1–11.
- Pell, S. K., Mitchell, J. D., Miller, A. J., & Lobova, T. A. (2010). Anacardiaceae. In K.  
 890       Kubitzki (Ed.), *Flowering Plants. Eudicots. The Families and Genera of Vascular  
 Plants, Volume 10* (pp. 7–50). Springer.
- Petersen, C. S. (1946). Estudios geológicos en la región del Río Chubut medio. *Dirección de  
 Minas y Geología Boletín*, 59, 1–137.

- Pujana, R. R. (2009). Fossil woods from the Oligocene of southwestern Patagonia (Río Leona Formation). *Atherospermataceae, Myrtaceae, Leguminosae and Anacardiaceae. Ameghiniana*, 46(3), 523–535.
- Pujana, R. R. (2022). Fossil woods from Argentina (1884–2021). *Revista del Museo Argentino de Ciencias Naturales n.s.*, 24(2), 217–240.
- Pujana, R. R., Wilf, P., & Gandolfo, M. A. (2020). Conifer wood assemblage dominated by Podocarpaceae, early Eocene of Laguna del Hunco, central Argentinean Patagonia. *PhytoKeys*, 156, 81–102.
- Radlkofer, L. (1888). Ueber die Versetzung der Gattung *Dobinea* von den Acerineen zu den Anacardiaceen. *Sitzungsberichte der Mathematisch-Physikalischen Classe der Bayerischen K. B. Akademie der Wissenschaften zu München*, 18, 385–395.
- Ramírez, J. L., & Cevallos-Ferriz, S. R. S. (2002). A diverse assemblage of Anacardiaceae from Oligocene sediments, Tepexi de Rodríguez, Puebla, Mexico. *American Journal of Botany*, 89(3), 535–545.
- Reid, E. M., & Chandler, M. E. J. (1933). *The London Clay Flora*. The British Museum (Natural History).
- Romero, E. J., & Hickey, L. J. (1976). A fossil leaf of Akaniaceae from Paleocene beds in Argentina. *Bulletin of the Torrey Botanical Club*, 103(3), 126–131.
- Romero-Lebrón, E., Fernández-Monescillo, M., Matushkina, N., Delclòs, X., & Gleiser, R. M. (2023). Damselflies (Coenagrionidae) have been avoiding leaf veins during oviposition for at least 52 million years. *iScience*, 26(6), 106865.
- Romero-Lebrón, E., Gleiser, R. M., & Petrusevičius, J. F. (2020). Geometric morphometrics of endophytic oviposition traces of Odonata (Eocene, Argentina). *Royal Society Open Science*, 7(12), 201126.

- Rossetto-Harris, G. (2023). *Origins of icehouse vegetation in southern South America: climate change and turnover in Eocene and Oligocene Patagonian fossil floras* (Doctoral thesis, Pennsylvania State University, University Park).
- 920 Sarzetti, L. C., Labandeira, C. C., Muzón, J., Wilf, P., Cúneo, N. R., Johnson, K. R., & Genise, J. F. (2009). Odonatan endophytic oviposition from the Eocene of Patagonia: the ichnogenus *Paleoovoidus* and implications for behavioral stasis. *Journal of Paleontology*, 83(3), 431–447.
- 925 Traverso, N. E. (1951). *La flora fósil de El Mirador (Chubut) conservada en el Museo de la Plata* (Doctoral thesis, Universidad Nacional de La Plata, La Plata).
- Troncoso, A. (1992). La taoflora Terciaria de Quinamávida (VII Region, Chile). *Boletín del Museo Nacional de Historia Natural, Chile*, 43, 155–178.
- Troncoso, A., & Encinas, A. (2006). La taoflora de cerro Centinela (Chile, VI Región):  
 930 vegetación y clima de Chile central a fines del Mioceno-comienzos del Plioceno. *Ameghiniana*, 43(1), 171–180.
- Vento, B., & Prámparo, M. B. (2018). Angiosperm association from the Río Turbio Formation (Eocene–?Oligocene) Santa Cruz, Argentina: revision of Hünicken's (1955) fossil leaves collection. *Alcheringa*, 42(1), 125–153.
- 935 Wang, Z., Herrera, F., Shu, J., Yin, S., & Shi, G. (2020). A new *Choerospondias* (Anacardiaceae) endocarp from the middle Miocene of Southeast China and its paleoecological implications. *Review of Palaeobotany and Palynology*, 283, 104312.
- Weeks, A., Zapata, F., Pell, S. K., Daly, D. C., Mitchell, J. D., & Fine, P. V. (2014). To move or to evolve: contrasting patterns of intercontinental connectivity and climatic niche  
 940 evolution in "Terebinthaceae" (Anacardiaceae and Burseraceae). *Frontiers in Genetics*, 5, 409.

Wheeler, E. A., Srivastava, R., Manchester, S. R., & Baas, P. (2017). Surprisingly modern latest Cretaceous–earliest Paleocene woods of India. *IAWA Journal*, 38(4), 456–542.

Wilf, P. (2012). Rainforest conifers of Eocene Patagonia: attached cones and foliage of the extant Southeast Asian and Australasian genus *Dacrycarpus* (Podocarpaceae). *American Journal of Botany*, 99(3), 562–584.

Wilf, P. (2020). Eocene “*Chusquea*” fossil from Patagonia is a conifer, not a bamboo. *PhytoKeys*, 139, 77–89.

Wilf, P., Cúneo, N. R., Escapa, I. H., Pol, D., & Woodburne, M. O. (2013). Splendid and seldom isolated: the paleobiogeography of Patagonia. *Annual Review of Earth and Planetary Sciences*, 41, 561–603.

Wilf, P., Cúneo, N. R., Johnson, K. R., Hicks, J. F., Wing, S. L., & Obradovich, J. D. (2003). High plant diversity in Eocene South America: evidence from Patagonia. *Science*, 300(5616), 122–125.

Wilf, P., Escapa, I. H., Cúneo, N. R., Kooyman, R. M., Johnson, K. R., & Iglesias, A. (2014). First South American *Agathis* (Araucariaceae), Eocene of Patagonia. *American Journal of Botany*, 101(1), 156–179.

Wilf, P., Iglesias, A., & Gandolfo, M. A. (2023). The first Gondwanan Euphorbiaceae fossils reset the biogeographic history of the *Macaranga-Mallotus* clade. *American Journal of Botany*, 110(5), e16169.

Wilf, P., Johnson, K. R., Cúneo, N. R., Smith, M. E., Singer, B. S., & Gandolfo, M. A. (2005a). Eocene plant diversity at Laguna del Hunco and Río Pichileufú, Patagonia, Argentina. *American Naturalist*, 165(6), 634–650.

Wilf, P., & Kooyman, R. M. (2023). Do Southeast Asia's paleo-Antarctic trees cool the planet? *New Phytologist*, 239(5), 1556–1566.

- Wilf, P., Labandeira, C. C., Johnson, K. R., & Cúneo, N. R. (2005b). Richness of plant-insect associations in Eocene Patagonia: a legacy for South American biodiversity. *Proceedings of the National Academy of Sciences USA*, 102(25), 8944–8948.
- 970 Wilf, P., Nixon, K. C., Gandolfo, M. A., & Cúneo, N. R. (2019). Eocene Fagaceae from Patagonia and Gondwanan legacy in Asian rainforests. *Science*, 364(6444), eaaw5139.
- 975 Wilf, P., Wing, S. L., Meyer, H. W., Rose, J. A., Saha, R., Serre, T., Cúneo, N. R., Donovan, M. P., Erwin, D. M., Gandolfo, M. A., González-Akre, E., Herrera, F., Hu, S., Iglesias, A., Johnson, K. R., Karim, T. S., & Zou, X. (2021). An image dataset of cleared, x-rayed, and fossil leaves vetted to plant family for human and machine learning. *PhytoKeys*, 187, 93–128.
- Wilf, P., Zou, X., Donovan, M. P., Kocsis, L., Briguglio, A., Shaw, D., Slik, J. W. F., & Lambiase, J. J. (2022). First fossil-leaf floras from Brunei Darussalam show dipterocarp dominance in Borneo by the Pliocene. *PeerJ*, 10, e12949.
- 980 Wolfe, J. A., & Wehr, W. C. (1987). Middle Eocene dicotyledonous plants from Republic, northeastern Washington. *U. S. Geological Survey Bulletin*, 1597, 1–25.
- Woodcock, D. W., Meyer, H. W., & Prado, Y. (2017). The Piedra Chamana fossil woods (Eocene, Peru). *IAWA Journal*, 38(3), 313–365.
- Wu, C. Y. (Ed.). (1979). *Flora Yunnanica, Volume 2*. Science Press.
- 985 Wu, C. Y. (Ed.). (1986). *Flora Xizangica, Volume 3*. Science Press.
- Wu, Y.-T. (1988). Anacardiaceae. In C.-Y. Chang (Ed.), *Flora Sichuanica. 4. Spermatophyta* (pp. 116–153). Sichuan Science and Technology Press.
- Zamaloa, M. C., Gandolfo, M. A., González, C. C., Romero, E. J., Cúneo, N. R., & Wilf, P. (2006). Casuarinaceae from the Eocene of Patagonia, Argentina. *International*
- 990 *Journal of Plant Sciences*, 167(6), 1279–1289.

Zamaloa, M. C., Gandolfo, M. A., & Nixon, K. C. (2020). 52 million years old *Eucalyptus* flower sheds more than pollen grains. *American Journal of Botany*, 107(12), 1763–1771.

995 Zich, F. A., Hyland, B. P. M., Whiffin, T., & Kerrigan, R. A., (2020). *Australian Tropical Rainforest Plants, Edition 8*. <https://apps.lucidcentral.org/rainforest>

**Figure 1.** Type specimens of *Dobineaites ameghinoides* comb. nov. **1**, Lectotype, USNM

1000 222678 (drawn in Berry, 1925: pl. 3, fig. 1). **2**, Syntype, USNM 222679 (Berry, 1925: pl. 3, fig. 2). **3**, Syntype, USNM 22680 (Berry, 1925: pl. 3, fig. 3). Scale bars: 1 cm.

**Figure 2.** *Dobineaites ameghinoides* comb. nov., selected larger leaves similar to the two

1005 syntypes (Fig. 1.2, 1.3). **1-3**, MPEF-Pb 7839, with details of base and margin. Note the pair of thin, obtuse secondary veins emerging near the petiole insertion. **4**, MPEF-Pb 7833. **5**, MPEF-Pb 7857a, with a curvilinear, frass-filled mine (DT90) oviposited adjacent to the midvein (upper arrow), separate oviposition sites on the midvein (DT76, lower arrow), hole feeding (DT2), and margin feeding (DTs 12, 15). Scale bars: 5 cm (1), 1 cm (2–5).

1010 **Figure 3.** *Dobineaites ameghinoides* comb. nov., selected specimens showing general variation with decreasing size and shape, from often larger, broad-cordate forms similar to the two syntypes (Fig. 1.2, 1.3; see also Fig. 2) to smaller, narrow-aspect forms similar to the lectotype (Fig. 1.1). The same variation occurs along single stems in extant *D. delavayi* (Fig. 6.1). Colorization of the leaves results from replacement by secondary minerals. **1**, MPEF-Pb 1015 7778, compressed with an unidentified angiosperm leaf(let) at upper right. **2**, MPEF-Pb 7819. **3**, MPEF-Pb 7763, with numerous galls (DT32), hole feeding (DTs 2, 3), and a pair of thin, obtuse basal secondaries emerging near the petiole insertion (arrow), as found on most of the fossils. **4**, MPEF-Pb 7803, with hole feeding (DTs 1, 3) and skeletonization (DT16). **5**, MPEF-Pb 7890. **6**, MPEF-Pb 7787. **7**, MPEF-Pb 7892 **8**, MPEF-Pb 7754b. **9**, MPEF-Pb 1020 7806. **10**, MPEF-Pb 7797, with a small, probably aborted curvilinear mine oviposited adjacent to the midvein (DT90, arrow). **11**, MPEF-Pb 7829, with margin feeding (DT12). **12**, MJHG 49Pb. **13**, MPEF-Pb 1450, with margin feeding (DT12). Scale bars: 1 cm.

**Figure 4.** *Dobineaites ameghinoides* comb. nov., branch segment with wide-cordate, attached

1025 opposite leaves (4.1–4) and selected insect damage (4.5–9). **1**, MPEF-Pb 7901, showing pair  
of opposite, petiolate leaves. **2**, detail from (4.1) showing pair of small axillary leaves (lower  
arrows) and a second node with opposite leaf scars (upper arrows) subtending one remaining  
axillary leaf. **3**, Detail from (4.2, see upper arrows) after preparation, showing the distal leaf  
node with opposite scars and the remaining axillary leaf, which has marginal teeth and other  
1030 standard leaf architecture of *D. ameghinoides*, further validating the attached large leaves (1, 2).  
**4**, Detail from (4.2, see lower arrows) of the paired small, axillary leaves. **5**, MPEF-Pb 7751a,  
with extensive hole feeding (DTs 1, 2, 3). **6**, MPEF-Pb 7749b, with several serpentine mines  
preserving sinusoidal, then particulate frass trails (the DT93 exemplar in Labandeira *et al.*,  
2007). **7**, detail of one mine in (6), oviposited adjacent to the midvein. **8**, MPEF-Pb 7772,  
1035 with a wide, possibly aborted mine containing medial particulate frass (the DT91 exemplar in  
Labandeira *et al.*, 2007). **9**, MPEF-Pb 1053b, with more than 90 oviposition scars (DT54, the  
scars separately cataloged as MPEF-IC 1370) arrayed in several arcs, previously described as  
the ichnospecies *Paleovoidus arcuatum* (Sarzetti *et al.*, 2009). Scale bars: 2 cm (1), 5 mm (2),  
2 mm (3, 4, 7), 1 cm (5, 6, 8, 9).

1040

**Figure 5.** Selected cleared leaves representing taxa compared with the fossils (see

Discussion) from the Jack A. Wolfe and Leo J. Hickey components of the National Cleared  
Leaf Collection, housed in the Division of Paleobotany, NMNH, and the Daniel I. Axelrod  
Cleared Leaf Collection, housed at the University of California Museum of Paleontology,

1045 Berkeley. The images are available at <https://doi.org/10.25452/figshare.plus.14980698.v1>

(Wilf *et al.*, 2021). Names are updated here using World Flora Online. Parentheses denote the



cleared leaf collection (Wolfe, Hickey, or Axelrod), the respective catalog number, and the source herbarium voucher, where applicable. (5.1–5.5), Cannabaceae. **1**, *Celtis bungeana* (Axelrod 162). **2**, *Celtis tetrandra* (Wolfe 9002; A s/n, December 1952, Taiwan). **3**,  
1050 *Aphananthe aspera* (Wolfe 153; UCH M008603, Panamá). **4**, *Trema orientalis* (Hickey 2834; FM Hueber s/n, Fiji). **5**, *Lozanella enantiophylla* (Wolfe 7036; CAS 422782, Mexico). **6**, *Broussonetia kazinoki* (Moraceae; Hickey 6448; YU, Li Hao-Min 13038, China). **7**, *Leucosyke alba* (Urticaceae; Hickey 6784; YU, Ahern 88, Philippines). **8**, *Ampelocera hottlei* (Ulmaceae; Wolfe 5557; F 1599211, Belize). **9**, *Gouania longispicata* (Rhamnaceae,  
1055 Wolfe 4863; MO 2053220, Burundi). **10**, *Rubus idaeus* (Rosaceae; Wolfe 1468b; UCH 1092750, Alaska). **11**, *Rhodotypos scandens* (Rosaceae; Hickey 3705; US 03703771, Japan). **12**, *Croton hircinus* (Euphorbiaceae; Hickey 6223; US 01229157, Panamá. (5.13–5.15), Malvaceae. **13**, *Melochia lupulina* (Hickey 5555; US 01092271, US Virgin Islands). **14**, *Triumfetta columnaris* (Hickey 5444; US 00535720, Mexico). **15**, *Grewia vitiensis* (Wolfe  
1060 11369; UC 1016242, Fiji). **16**, *Acer tataricum* (Sapindaceae; Wolfe 8576; A, Muroi 4232, Japan). **17**, *Dobinea delavayi* (Anacardiaceae; Wolfe 8206; A, Rock 6026, Yunnan, China).  
Scale bars: 1 cm.

**Figure 6.** Herbarium vouchers of (6.1–6.3) *Dobinea delavayi* and (6.4) *Dobinea vulgaris*. **1**,  
1065 US 03349094 (Yunnan), showing leaf size and shape variation along a single axis from larger broad-cordate to smaller narrow-lanceolate leaf forms, encompassing much of the variation seen in the fossils (Figs. 1–4), and fertile axes of elm-like fruits. Image courtesy of US via <https://collections.nmnh.si.edu/search/botany>. **2**, K, G Forrest 15318 (Yunnan), detail of marginal teeth and venation, including thin fimbrial vein (compare, *e.g.*, Fig. 2.3). **3**, P  
1070 04860032 (Yunnan), showing wide-cordate leaf morphology and miniature axillary leaves similar to the fossils (Fig. 4.1–4.4). Image courtesy of P via

<http://coldb.mnhn.fr/catalognumber/mnhn/p/p04860032>. 4, NY 2592273 (cultivated),

showing bases of the opposite elliptical leaves of *D. vulgaris*, with miniature axillary leaf-

bearing branches similar to the fossils (Fig. 4.1–4.4). Image courtesy of the C. V. Starr Virtual

1075 Herbarium, <http://sweetgum.nybg.org/science/vh>. Scale bars: 5 cm (1, 3), 1 cm (2, 4).

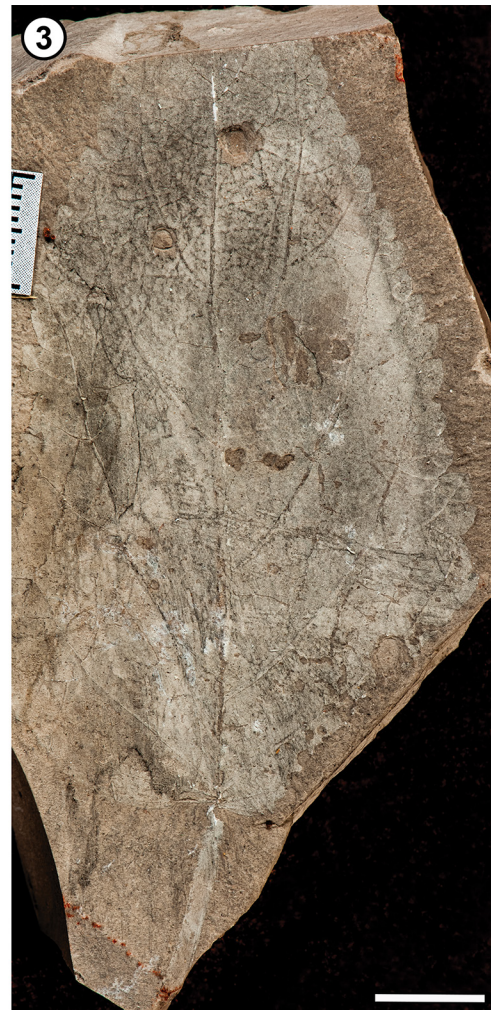


Fig. 1 R1





Fig. 2 R1



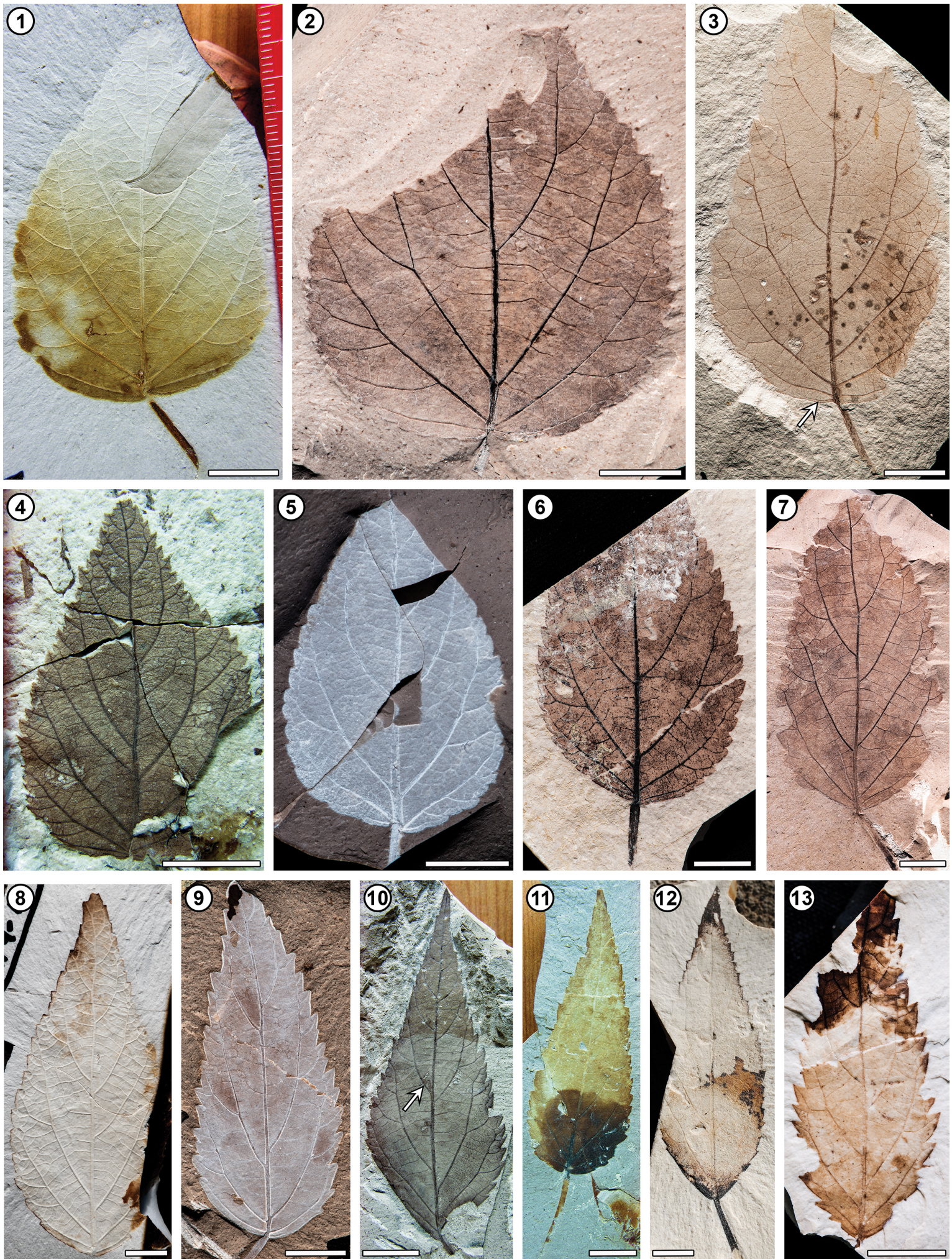


Fig. 3 R1



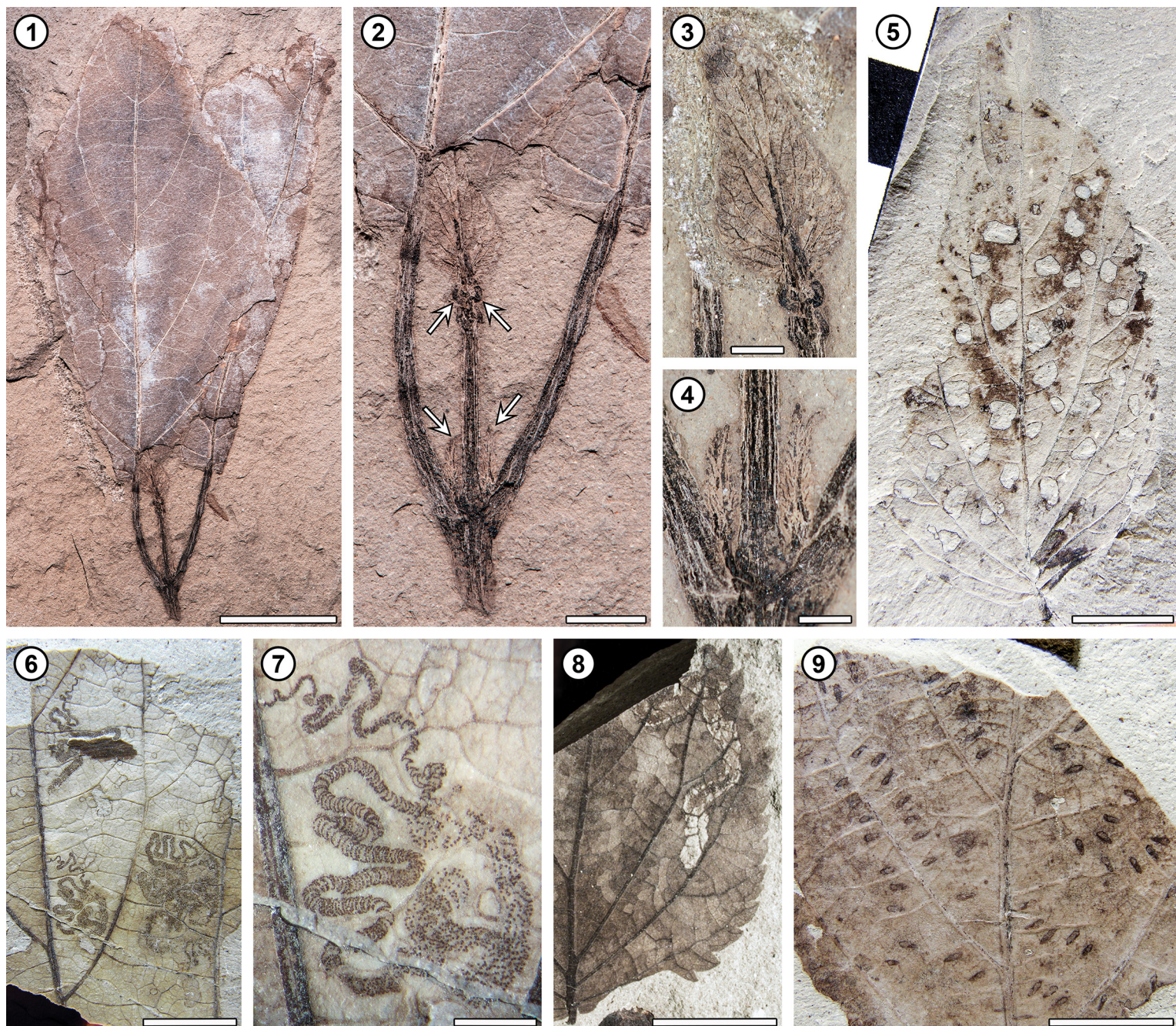


Fig. 4



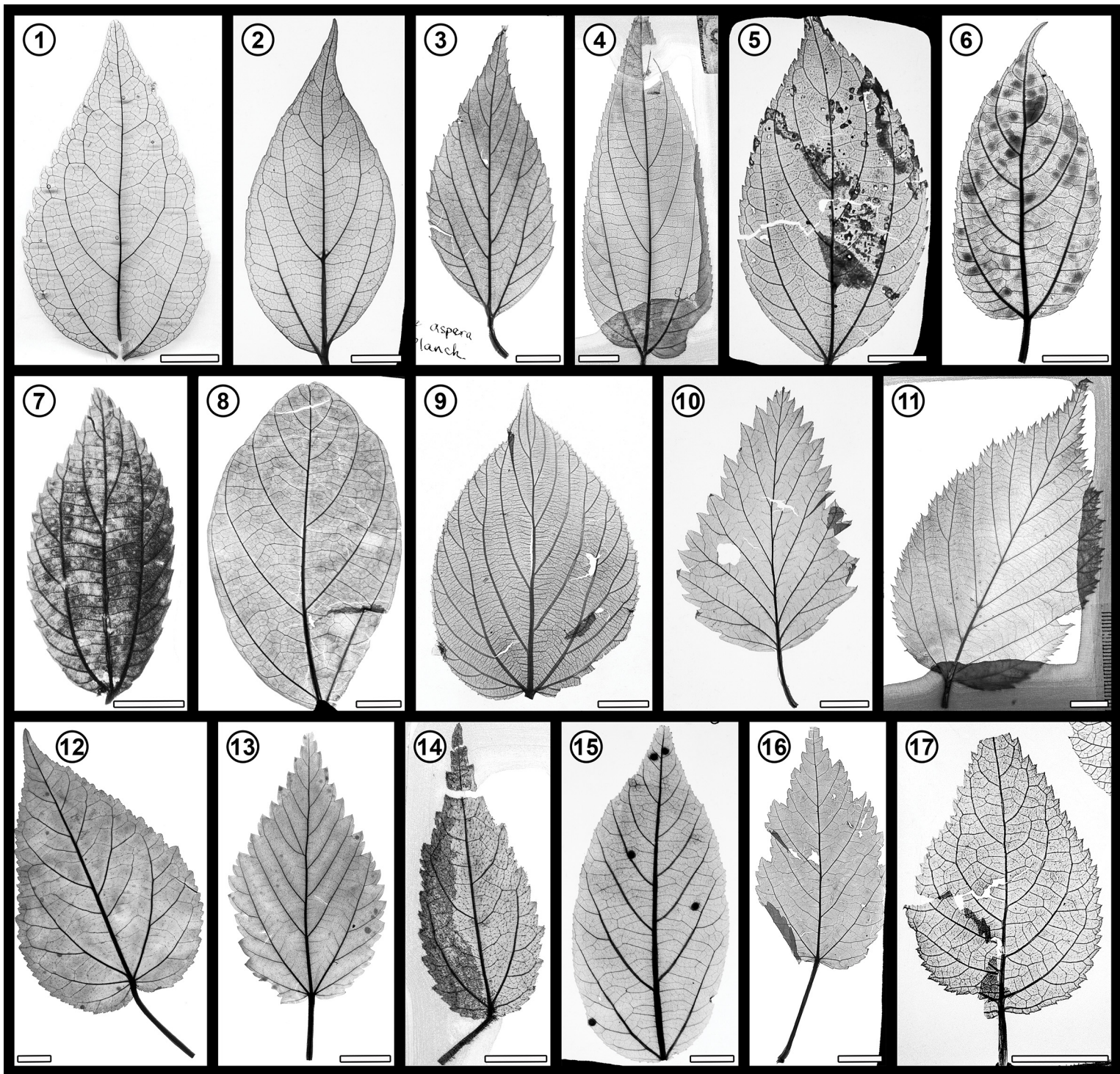


Fig. 5 R1





Fig. 6