

Neotropics as a Cradle for Adaptive Radiations

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Neotropical ecosystems are renowned for numerous examples of adaptive radiation in both plants and animals resulting in high levels of biodiversity and endemism. However, we still lack a comprehensive review of the abiotic and biotic factors that contribute to these adaptive radiations. To fill this gap, we delve into the geological history of the region, including the role

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of tectonic events such as the Andean uplift, the formation of the Isthmus of Panama, and the emergence of the Guiana and Brazilian Shields. We also explore the role of ecological opportunities created by the emergence of new habitats, as well as the role of key innovations, such as novel feeding strategies or reproductive mechanisms. We discuss different examples of adaptive radiation, including classic ones like Darwin's finches and *Anolis* lizards, and more recent ones like bromeliads and lupines. Finally, we propose new examples of adaptive radiations mediated by ecological interactions in their geological context. By doing so, we provide insights into the complex interplay of factors that contributed to the remarkable diversity of life in the Neotropics and highlight the importance of this region in understanding the origins of biodiversity.



“**A**daptive radiation,” which refers to the proliferation of species occupying diverse ecological niches, has been a well-studied pattern in evolutionary biology since the concept was first introduced by Osborn (1902). The Neotropics is home to several iconic radiations that have played a pivotal role in the development of the theory of evolution and the discovery of natural selection. However, there are several species-rich groups that are either not formally recognized as adaptive radiations or are ambiguously referred to as such (Fig. 1), which highlights the ongoing debate in the scientific community on the very definition of the concept (Givnish 2015). By examining these groups and the ecological and evolutionary factors driving their diversification, we can better understand the role that South American geography, and the neotropics as a whole, played in generating one of the most biologically diverse regions of the planet (Fig. 1).

In this review, we delve into some key patterns and evolutionary mechanisms that are associated with adaptive radiations in plant and animal groups in the Neotropics. Additionally, we identify some new putative examples of unrecognized adaptive radiations for both plants and animals in the region. By examining prominent examples of both adaptive and nonadaptive radiations in the Neotropics, we highlight the importance of geographic features, including geological dynamics, climate shifts, and other historic events, in shaping evolutionary radiations in this region.

While we acknowledge that nonadaptive radiation processes have undoubtedly contributed to the extraordinary neotropical diversity, the

processes underlying speciation in nonadaptive radiation do not involve ecologically based divergent natural selection. Moreover, understanding adaptive radiation hinges on two crucial factors: key innovations and ecological opportunities. Key innovations, representing evolutionary breakthroughs, drive diversification by conferring novel advantages. Simultaneously, ecological opportunities arise from untapped niches, fostering rapid speciation. The interplay between these factors propels species into divergent trajectories. By highlighting the roles of key innovations and ecological opportunities, scientists can gain profound insights into the dynamics and patterns of adaptive radiation across taxa and ecosystems.

In this review, we focus on investigating the ecological and evolutionary mechanisms underlying examples of adaptive radiation, shedding light on the diversity, adaptations, and coexistence of species across neotropical habitats.

WHAT DETERMINES ADAPTIVE AND NONADAPTIVE RADIATIONS?

The concept of adaptive radiation has played a fundamental role in understanding the ecological and evolutionary mechanisms underlying the proliferation of species and morphological diversification (Carlquist 1965; Mayr 1970; Stebbins 1974; Martin and Richards 2019; Gillespie et al. 2020). In a broad sense, adaptive radiation is defined as the evolution of ecological and phenotypic diversity in a rapidly diversifying lineage (Schluter 2000). This involves ecologically distinct species with high morphological diversity that allows adaptation to contrasting environ-

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Figure 1. The Neotropics and its geographic complexity is a cradle for adaptive radiations. Map of the Neotropical region, spanning from Mesoamerica to central Argentina, including all Caribbean Islands, the Galápagos archipelago, and Juan Fernández islands. The figure shows some of the most iconic examples of animal and plant Neotropical adaptive radiations: (A) *Liolemus*, (B) *Philisca*, (C) *Anolis*, (D) *Geospiza*, (E) *Lupinus*, (F) *Espeletia*, (G) *Scalesia*, (H) *Dendroseris*, (I) *Robinsonia*. (Photo credits by J. Chaves (D), J.E. Guevara-Andino (E), Phyllis Coley (F), Gonzalo Rivas-Torres (G), and D.D. Cotoras (A,B,C,H,I).)

ments or habitats. It also involves the diversification of multiple lineages from a single common ancestor.

Schlüter (2000) proposes four criteria that a group will have to meet to be classified as an example of adaptive radiation. The first criterion is common ancestry, which might or might not involve monophyletic groups. The second implies a correlation between phenotypic diversity and the occupation of novel environments (abiotic and biotic). The third criterion relates to the adaptive advantages of trait expression in their respective environments (trait utility) measured as the trait fitness values compared with the environment. The fourth criterion identifies rapid bursts of diversification for certain groups compared to sister lineages or clades as a characteristic of an adaptive radiation. Finally, some authors suggest a fifth criterion: the coexistence in sympatry of at least three successive sister species (Martin and Richards 2019).

However, it has proven difficult for most studies to fulfill all these criteria, and therefore there are few examples of “true” adaptive radiations (Carlquist 1965; Seehausen 2006; Grant and Grant 2008; Losos 2009; Brawand et al. 2014). Some authors have adopted broader perspectives in which not all of these criteria are met in order to define an adaptive radiation (Givnish 2015; Gillespie et al. 2020). For instance, studies for Darwin’s finches and Andean lupines have revealed the role of trait utility, phenotype–environment correlations, rapid bursts of speciation, and common ancestry. However, assuming a rapid burst of speciation as a defining characteristic of adaptive radiation could exclude some of the most iconic examples of adaptive radiations in South America such as the bromeliads of the genus *Brochinnia* (Givnish et al. 2011, 2014).

We do not intend to defend any definition or highlight which lineages qualify as a “true example” of adaptive radiation. Instead, we aim to



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draw attention to two fundamental elements of this process, ecological opportunity and key innovations (Gillespie et al. 2020). The concept of ecological opportunity as the driving factor in adaptive radiation stems from the early work of Simpson (1953). Ecological opportunity refers to the availability of ecological resources or niche space that were either unoccupied or previously used by competitors (Simpson 1953; Stroud and Losos 2016, De-Kayne et al. 2024). While Simpson (1953) suggested that ecological opportunities emerge from geographic, ecological, and evolutionary access to new niches that collectively determine a new “adaptive zone,” the existence of the space by itself does not generate adaptive radiation. Instead, a radiating lineage must gain access to new niches geographically through colonization, ecologically by using resources for which competition is reduced, and evolutionarily by having the adaptations to use such resources in the first place (i.e., enter a new “adaptive zone”; see also Donoghue and Sanderson 2015). Therefore, ecological opportunities are defined by the relationship between lineages and ecological space such that a lineage must gain access across all three dimensions (i.e., geographic, ecological, and evolutionary) to radiate (Simpson 1953; Stroud and Losos 2016). By conferring access to ecological opportunities, “key innovations”—traits that enable lineages to occupy a previously inaccessible ecological state that might promote diversification (Miller et al. 1949; Simpson 1953; Rabosky 2017)—become central to the concept of the adaptive zone. We explore how the evolution of key innovations is related to the occupation of novel niches (i.e., abiotic and biotic ecological opportunities) as the result of the dynamic neotropical geological history.

In contrast to adaptive radiations, nonadaptive radiations have been less studied, as ecological opportunity and adaptive divergence as drivers of diversification might be absent (Schlüter 2000; Rabosky 2017). Nonadaptive radiations may show morphological or physiological divergence unrelated to the environment or resource use patterns (Givnish 2015). For instance, the radiation of *Scytalopus* tapaculos among birds and *Phlegmariurus* fir mosses among plants are

considered as nonadaptive neotropical radiations (Testo et al. 2019; Cadena et al. 2020). Nonadaptive radiations presuppose rapid lineage diversification despite little-to-no ecological differentiation and usually as the result of allopatric or parapatric speciation (Rundell and Price 2009; Czekanski-Moir and Rundell 2019). Thus, the role of geographic barriers (e.g., lack of gene flow) promoting species formation among populations experiencing similar environments with no influence of divergent natural selection is a basic tenet of nonadaptive radiations. Because new alleles arising in different geographically isolated populations may become fixed or lost depending on population size and the selective advantage of the alleles, nonecological speciation may occur slowly in nonadaptive radiations (Nosil and Flaxman 2010; Czekanski-Moir and Rundell 2019). Nevertheless, nonadaptive radiations triggered by geographical isolation could predate ecological divergence (Losos and Ricklefs 2009; Rundell and Price 2009; Givnish 2015; Gillespie et al. 2020).

ECOLOGICAL OPPORTUNITY AND KEY INNOVATIONS IN THE CONTEXT OF NEOTROPICAL GEOLOGY

The geological history of the Neotropics has directly shaped biodiversity patterns across the continent (Fig. 2; see Box 1 for more details on neotropical geology). The tectonic and volcanic events that triggered the formation of the isthmus of Panamá, the Andean mountains uplift, the formation of the Caribbean islands, and the formation of the Pebas megalake system are just a few examples of the major geological events that created a myriad of habitats and ecosystems throughout the Neotropics (Hoorn et al. 2010; Gutiérrez-García and Vázquez-Domínguez 2013; Pérez-Escobar et al. 2022; Sanín et al. 2022). While the emergence of geographical barriers can promote nonadaptive radiation, we first focus on the role of geological dynamics in adaptive radiation. We outline how these events may shed light on ecological opportunity and ecological release as preconditions for subsequent adaptive radiations.

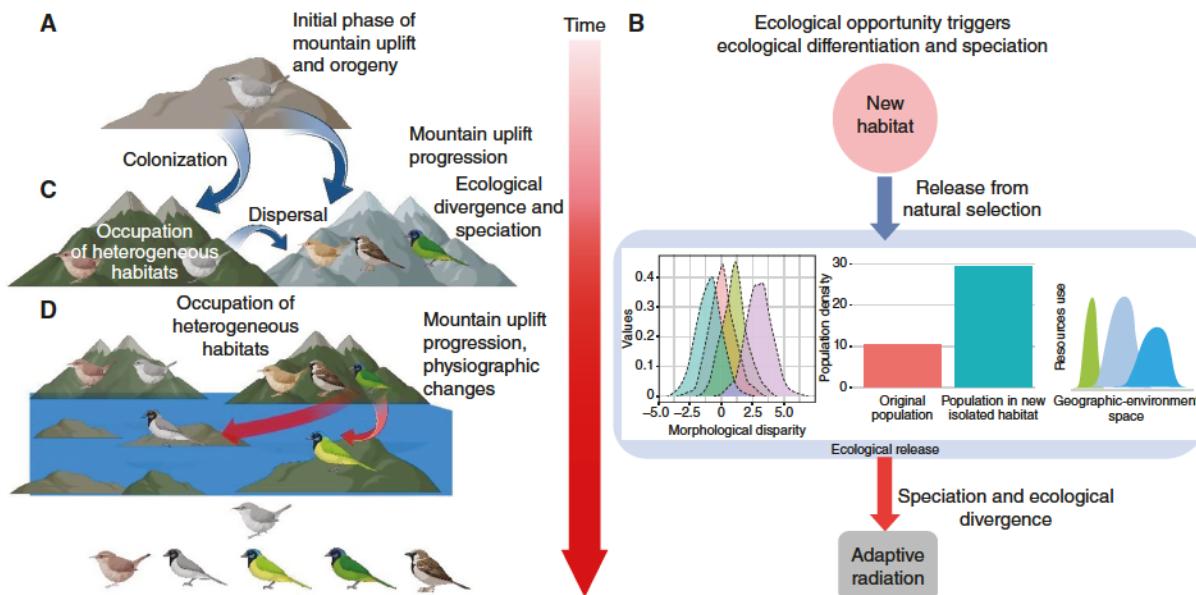


Figure 2. Geological events, in this case mountain orogeny, have a direct impact on adaptive evolution and ecological opportunity. Historical events have played a major role in the origin of biodiversity not just because geological changes can trigger evolution, but also because environmental conditions are the stage on which evolution occurs. However, few attempts have been made to understand the role of geomorphology on adaptive radiation. (A) Once plate tectonics is underway, mountain uplift creates the conditions for the formation of new habitats. Interrelatedly, ecological opportunity and ecological release occur after an ancestral lineage disperses to these new unoccupied environments. (B) Opportunity and release may act to relax negative selection, promoting lineage and morphological diversification (disparity) through habitat or resource use expansion, an increase in population density, and release from interspecific competition. (C) For instance, geomorphological changes during the Andean uplift may have created a myriad of novel environments where previously unused resources became available to new colonizers. Once an ancestral lineage colonizes this new environment, the lack of competition for resources should favor radiation in the multiple available adaptive zones (D).

BOX 1. A BRIEF GEOLOGICAL HISTORY OF THE NEOTROPICS**The Evolution of Western Amazonian Landscapes and the Pebas System**

The Andes are the longest mountain range in the world, and its emergence was a major historical event in the evolution of South America's landscape and particularly for Amazonian forests. Since the Cenozoic (~23 myr), the Andean uplift has led to highly heterogeneous habitats by modifying climatic patterns and generating physiographic changes (Hoorn et al. 2010, 2013). During the Paleogene and before the Andean uplift (65–30 myr), the major sources of sedimentary material for the Amazon basin were both the Guiana and the Brazilian Shields, which both formed during the Precambrian and were mainly characterized by poor nutrient content and quartzitic sandstone. The flow of sediments was in a westward direction with major depositional basins in the current Central and Western Amazonia. By this time, the proto-Amazon River drainage system was a reverse system completely dissimilar to the current drainage system. The environment of the pan-Amazonian basin during the Paleogene (65–30 myr) was mainly characterized by alternating fluvial conditions and marginal marine embayments (Roddaz et al. 2009; Hoorn et al. 2010). By the early mid-Miocene (23–16 myr), the central and northern portions of the Andes started to uplift creating the conditions for physiographic changes that contributed to the formation of the current Amazon River drainage (Hoorn et al. 2010).

These physiographic changes set the conditions for the simultaneous formation of the Pebas lacustrine system during the middle-late Miocene 29–9 myr (Wesselingh et al. 2006; Roddaz et al. 2009). A drastic reorganization of these landscapes occurred in the early late Miocene, some 8–9 myr. In a short time, fluvial landscapes led by the uplifting Andean hinterland to the west replaced the former Pebasian wetlands and the modern easterly course of the Amazon became established. Fossil evidence of freshwater fauna (e.g., reptiles, fishes) corroborates the evolution of a heterogeneous landscape including freshwater swamps and islands of terra firme-white sand forests in the current Western Amazonia. Marine incursions from the Caribbean Sea through the Llanos system during the Miocene also contributed to the landscape heterogeneity (Alvim et al. 2021).

The Guiana Shield

The geomorphological dynamics of the Guiana Shield is related to the topographic, geological, and hydrological dynamics of the so-called Amazon platform of Proterozoic origin (2.5–1.8 bya). In some regions such as the northern portion of the Amazon platform, Proterozoic outcrops are visible as is the case of Roraima mountain (2810 m) or Pico de Neblina mountain (3014 m). These are the major tepui-like topographic formations in a vast region that comprises ~2,280,000 km² (Lujan et al. 2011).

The Brazilian Shield

The Brazilian Shield covers most of the Brazilian territory expanding toward the center of the Andean chain to the west and the Patagonian massif to the south (Hartmann and Delgado 2001). The three major tectonic units are older than 900 myr, and there is evidence of Proterozoic parental material in the eastern portion of Brazil (Hartmann and Delgado 2001). Most of the Brazilian Shield also includes the highlands extending between the central-north portion of the Amazon toward the Rio de la Plata estuary in the south. Little is known about the geomorphological history of the Brazilian Shield and it has been argued that long-term stability tectonics might be responsible for this lack of information. Nonetheless, the eastern margin of the shield has suffered significant tectonic and physiographic changes since the Cenozoic. Some of the most important geological events that shaped the eastern portion of the Brazilian Shield are related to the uplift of mountain ranges along the Atlantic coast of Brazil that have produced a sharp geomorphological contrast between the narrow Atlantic coastal slope and the broad inland (Harrington 1962; Buckup 2011). Two major geological basins characterize the lowland eastern portion of the shield, the Sao Francisco and the Parana River basins. Together these geomorphological characteristics have produced an extremely heterogeneous landscape with several habitats and climatic conditions favoring the diversification of several plant and animal groups.

Continued

Andean Orogeny and the Evolution of the Two “Arid Diagonals” Páramo–Puna–Altiplano Systems

The Andean orogeny is complex and there is still ongoing debate about the pace and time of the evolution of this system. The Andes-Altiplano is one of the largest mountain belt systems in the world extending from ~10°N to 50°S as a consequence of subduction of oceanic lithosphere beneath an initially flat continental margin. The subduction along the Pacific margin originated during the Paleogene (65–34 myr), causing the initial uplift of the Northern and Central Andes (Hoorn et al. 2010; Armijo et al. 2015). However, this is disputed as it has been suggested that the initial stage of the uplift of the northern Andes occurred at ~80 myr (Horton 2018). The limit between the northern and central Andes is defined by the subduction of the Carnegie ridge under the South American plate in Ecuador producing the geomorphological phenomenon known as the Huancabamba depression (Pérez-Escobar et al. 2022). In the northern portion of the Andes, one of the major geological events was the initial uplift of the eastern cordillera that provoked subsequent orographic and climatic changes including the separation of the Orinoco and Magdalena rivers basins (Hoorn et al. 1995; Horton et al. 2010). This process initiated ~26–23 myr and was one of the major physiographic changes that promoted the evolution of habitat heterogeneity at basin scale. The central portion of the Andes, which extends along a 2000 km belt in a NW–SE direction and 3200 km in a N–S direction, began orogenic deformation in the Cretaceous ~70 myr (Horton and DeCelles 1997; Pérez-Escobar et al. 2022). Associated with the Altiplano formation is the existence of a series of high-elevation lakes, of which the one with the current largest size is Lago Titicaca. Those lakes have been strongly affected by changes in precipitation regime during the Pleistocene glaciations, which has resulted in expansions, connections, and droughts of many of them at different times (Horton and DeCelles 1997). In the case of the southern portion of the Andes geological and physiographic changes started ~100 myr, but extreme uplift events that shaped the current conformation began only ~15 myr (Hervé et al. 2000).

Together these orogenic events changed dramatically the climate and physiography of the entire continent creating two “arid diagonals” (Luebert 2021). In the northern and central Andes, uplift increased rainfall in the east and a rain shadow effect with dry conditions in the west of the mountain range, this created the conditions for the evolution of landscapes and habitats in the eastern dry diagonal including the Cerrado, Caatinga, and Chaco biomes (Pérez-Escobar et al. 2022). In the central and southern Andes, the western arid diagonal connects the Atacama desert, Monte, Prepuna, dry Puna, Pampas, and eastern Patagonia. These two diagonals effectively isolate three major humid forest formations in the continent: (1) Amazonia + tropical Andes + Chocó, (2) Mata Atlántica, and (3) Valdivian Jungle.

Continental and Volcanic Islands

South America is surrounded by several volcanic archipelagos and continental islands. Along the Pacific coast from north to south, it is possible to find: Isla Malpelo, the Galápagos archipelago, the Desventuradas islands, and the Juan Fernández archipelago. All of them are over the Nazca plate, which creates a general time chronosequence where the older islands tend to be closer to the continent (Harpp and Geist 2018; Lara et al. 2018). Along the Atlantic coast, also from north to south, it is possible to find: the São Pedro and São Paulo archipelago, the Fernando de Noronha archipelago, and the Trindade e Martim Vaz archipelago. There are also several near-shore continental islands and island formations on the delta of large rivers (e.g., Amazon and Río de la Plata). The area with the highest concentration of them corresponds to the fjords area in southern Chile. Most of them are of glacial origin.

Major geological changes through time in the continental Neotropics often involve mountain uplift and plate tectonics. Uplift during mountain growth is directly related to physiographic modifications including changes in drainage patterns, which in turn could lead to

bridges or barriers for species dispersal. Mountain uplift also generates modifications in atmospheric circulation patterns, leading to changes in precipitation regimes. In South America, the Andean orogeny and the emergence of the tepuis formation in Brazilian and Guiana Shield cra-



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tons dramatically changed the flow and the direction of sediment deposition for more than 100 myr (Hoorn et al. 2010, 2013; Pérez-Escobar et al. 2022). These changes, along with widespread erosional processes, changes in orographic precipitation, and marine incursions, have resulted in the formation of a highly heterogeneous landscape with multiple opportunities for radiation (Fig. 2).

Although many of the best-documented examples of adaptive radiation in the Neotropics are associated with geological events, we do not suggest that geology alone triggers bursts of speciation and ecological divergence. Instead, we propose that the environmental settings for evolution are necessarily related to historic events involving natural selection. Therefore, geological events have played a major role in the origin of neotropical biodiversity, especially because extrinsic abiotic factors (i.e., geology) such as the emergence or collapse of geological barriers, island formation, or climate variation, can trigger evolutionary processes. Despite several attempts to correlate neotropical geological history to ecological opportunity in the context of adaptive radiation, few examples have explicitly tested the role of the paleoenvironment in diversification and ecological divergence. Thus, we link geological dynamics with ecological opportunity, and key innovations to explain particular plant and animal neotropical adaptive radiations (Fig. 2). In the next sections, we will summarize the most iconic examples of adaptive radiation in both plants and animals and the link between ecological opportunity and neotropical geological history.

Plants

In this section, we describe a few proposed cases of adaptive radiations in neotropical plant groups. However, the paucity of literature on plant adaptive radiations in South America creates a major gap in understanding how geomorphology and climate shape these radiations in the Neotropics. Although multiple studies have focused on documenting bursts of diversification (Meseguer et al. 2022) or the evolution of “key innovations” to enable the colonization and

spur the diversification of various plant lineages in new ecological settings (Kadereit and von Hagen 2003; von Hagen and Kadereit 2003), few studies have examined explicitly and experimentally how trait utility and environment to phenotype correlations influence organismal fitness and diversification. Further work is critically needed to link ecological opportunity and adaptive divergence from comparative, populational, and optimality perspectives in the neotropical flora (Kadereit and von Hagen 2003; von Hagen and Kadereit 2003; Olson and Arroyo-Santos 2015).

The most common feature of plant adaptive radiations in the neotropics is the role of South American geological history. From the emergence of the Galapagos archipelago to the intricate geology of the Andes, the Guiana Shield, and the Brazilian Shield, these geological events have created a mosaic of diverse habitats and climatic niches. These factors have significantly contributed to generating ecological opportunities for the adaptive radiations of plants. We synthesize cases of adaptive radiation in neotropical plant groups and summarize the characteristics of the most well-documented cases, including the *Espeletia* complex, lupines, and bromeliads (Table 1). In addition, we present evidence for a new case of a plant adaptive radiation in the genus *Scalesia*, which has not been treated as such in the literature.

Owing to their remarkable ecological and phenotypic diversity, the *Scalesia* radiation is often likened to the “Darwin’s finches of the plant world.” Endemic to the Galápagos Islands, *Scalesia* comprises approximately 15 species of trees and shrubs occupying various climates and habitats across 11 islands (Schilling et al. 1994; Itow 1995; Fernández-Mazuecos et al. 2020). Geological dynamics have created extreme habitat heterogeneity, promoting ecological opportunities for speciation and adaptation in *Scalesia* (Geist et al. 2014). Leaf and inflorescence morphology have evolved repeatedly, leading to differentiation in climatic niches and plant–pollinator interactions (Fernández-Mazuecos et al. 2020). Leaf variation in *Scalesia* reflects adaptation to within-island climatic gradients, with smaller-leaved species occupying dry lowland habitats

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and larger-leaved species thriving in humid uplands (Perez et al. 2023). While the *Scalesia* adaptive radiation is well-established, the genomic basis of adaptive traits remains largely unexplored (Cerca et al. 2023), evidence indicates that the selection of genes associated with climatic niches may drive the adaptive radiation in *Scalesia* (Eliasson 1974; Walter et al. 2016; Fernández-Mazuecos et al. 2020; Cercá et al. 2023; Perez et al. 2023).

Animals

In this section, we describe the better-documented cases of adaptive radiation in neotropical vertebrate groups highlighting recent evidence to propose new cases of adaptive radiation (Table 2). The most iconic examples of vertebrate radiation come from the studies of Caribbean *Anolis* and Darwin's finches. Nonetheless, mounting evidence from phylogenomics, anatomical, paleontological, and functional analyses have shed light in terms of documenting additional examples including neotropical cichlids, as well as multiple mammals and bird lineages (Grant 1986; Grant and Grant 2008; Tebbich et al. 2010; Lamichhaney et al. 2015; Jiménez-Ortega et al. 2023).

The neotropical cichlids (subfamily Cichlinae) include more than 600 species distributed in riverine and lacustrine systems of Central and South America (López-Fernández et al. 2010, 2013; Arbour and López-Fernández 2016). This fish radiation is both morphologically and ecologically diverse, with rampant repeated evolution of specialized feeding strategies (e.g., detritivory, piscivory, substrate sifting) in Central and South America clades (Winemiller et al. 1995; Arbour and López-Fernández 2014, 2016). Variation in rates of evolution of functional feeding morphology is consistent with a scenario of changing ecological opportunity and ecological release operating at different spatial and phylogenetic scales. As an example, colonization of new habitats facilitated by the Panamá Isthmus formation ~3–10 myr (Hulsey et al. 2010) and release from South American competitors promoted bursts of diversification and morphological disparity in Central American lineages (e.g.,

Geophagus, *Acarychthis*, *Herychthis*) (Hulsey et al. 2010; Arbour and López-Fernández 2016).

Numerous instances of diversification and adaptive radiation in tropical birds have been extensively studied, with hummingbirds (family Trochilidae) serving as a prominent example. These nonpasserine birds, comprising approximately 340 species diverging from swifts ~42 myr, exhibit specialization in nectar consumption, leading to intense interspecific competition (Temeles et al. 2009; Martín González et al. 2015). Rapid and variable diversification rates underscore the complexity of hummingbird evolution (McGuire et al. 2014). Hummingbirds have undergone adaptive radiation driven by their expansion into new habitats, particularly in the Andes, recent evidence suggests the adaptive value of genes associated with high-elevation physiological tolerance both between and within species level (Lim et al. 2019, 2021; Barreto et al. 2023). The timing of Andean uplifts, ~6–10 myr and 2–5 myr, facilitated habitat expansion and access to new food resources, contributing to the evolutionary success of hummingbirds (Gregory-Wodzicki 2000; Garzoni et al. 2008; Bershad et al. 2010; McGuire et al. 2014; Lim et al. 2019). Overall, hummingbirds represent one of the most comprehensively studied neotropical adaptive radiations.

ECOLOGICAL OPPORTUNITY AND KEY INNOVATIONS IN THE CONTEXT OF BIOTIC INTERACTIONS

Plant–Herbivore Interactions

It has been long recognized that interspecific competition for resources could drive divergence during adaptive radiations. The classical ecological theory of adaptive radiation states that the last stage in the diversification process should be related to the increased phenotypic divergence of closely related species coexisting in sympatry (Lack 1983; Schlüter 2000). This process has been linked to divergence mediated by resource partitioning and increased specialization after speciation, and most of the literature on adaptive radiations has been focused on the role of competition and ecological opportunity





Table 1. Summary of neotropical plant adaptive radiations highlighting accepted examples and proposing new cases based on current evidence about the role of geology

Adaptive radiation	Clade	Age (myr)	Number of species	Associated geological event	Ecological opportunity	Key innovation(s)	Accepted as adaptive radiation following Schlüter (2000)	Key references
Lupines	<i>Lupinus</i>	0.4–1.93	85–90	Final phase of the northern Andean uplift	Appearance of high-altitude cold-humid habitats (paramos, montane forests)	Variation in growth form and perenniarity	Yes, but contested by Givnish (2015)	Hughes and Eastwood 2006; Drummond et al. 2012; Hughes and Atchison 2015; Nevado et al. 2016
Espeletia complex	<i>Espeletia</i>	3–5	~140	Final phase of the Andean uplift	Appearance of high-altitude cold habitats (super paramos, high-altitude swamps)	Rosette growth form evolution, reproductive syndromes associated with the type of inflorescence	Yes	Monasterio and Sarmiento 1991; Cuatrecasas 2013; Diazgranados and Barber 2017; Pouchon et al. 2018
Bromeliads	<i>Tillandsioideae</i>	15.1–16.9	~1256	Evolution of the Andes and the Atlantic forests	Creation of new open multiple edaphic and climatic niches	Epiphytism, tank habit, water and soil nutrient absorptive trichomes, the CAM photosynthetic pathway, and avian pollination	Yes	Givnish et al. 2011, 2014; Silvestro et al. 2014; Males 2018
	<i>Bromelioideae</i> (Brazilian Shield clade, <i>Brochinia</i>)	7.5–9.4	753	Emergence of the Brazilian and Guiana Shields	Heterogeneous landscape spanning climatic and edaphic gradients	Epiphytism, tank habit, water and soil nutrient absorptive trichomes, the CAM photosynthetic pathway, and avian pollination	Yes	Givnish et al. 2011, 2014; Silvestro et al. 2014; Males 2018

Continued

**Table 1.** *Continued*

Adaptive radiation	Clade	Age (myr)	Number of species	Associated geological event	Ecological opportunity	Key innovation(s)	Accepted as adaptive radiation following Schlüter (2000)	Key references
	Bromelioideae (Tank clade)	9.4–10.7	629	Emergence of the Brazilian and Guyana Shields and evolution of the Andes	New open multiple edaphic and climatic niche	Epiphytism, tank habit, water and soil nutrient absorptive trichomes, the CAM photosynthetic pathway, and avian pollination	Yes	Givnish et al. 2011, 2014, Silvestro et al. 2014; Males 2018
Darwin giant daisies	Scalesia	0.63–0.74	~16	Galapagos archipelago geology	New open climatic niches and available pollinators	Leaf dissection, pubescence, and inflorescence morphology	Recently proposed	Itow 1995; Fernández-Mazuecos et al. 2020; Cerca et al. 2023; Perez et al. 2023

Further details of these systems are provided in the text and the Supplemental Information.



Table 2. Summary of neotropical animal adaptive radiations highlighting accepted examples and proposing new cases based on current evidence about the role of geology

Adaptive radiation	Clade	Age (myr)	Number of species	Associated geological event	Ecological opportunity	Key innovation(s)	Accepted as adaptive radiation following Schlüter (2000)	Key references
Hummingbirds	Trochilidae	42	363	Andean uplift	Flower nectar	Variability in bill size and shape, body size	Yes	McGuire et al. 2014; Barreto et al. 2023
Tyrant flycatchers	Tyrannidae	~25	>441	Expansion of semi-open and open habitats from mid-Miocene onward	Insects in diverse levels of vegetation clutter	Foraging behavior	Yes, but not tested with recent macroevolutionary techniques	Fitzpatrick 1985; Ohlson et al. 2008
Ovenbirds and woodcreepers	Furnariidae	~25	315	Andean uplift, expansion of semi-open and open habitats, river basin reconfiguration	Insects on trunks and vegetation	Ecomorphological adaptability, nest architecture	Yes	Claramunt 2010; Derryberry et al. 2011
Tanagers	Thraupidae	~12	384	Andean uplift, expansion of semi-open and open habitats	Dietary and habitat niches	Rapid bill shape evolution	Yes	Sedano and Burns 2010; Vinciguerra and Burns 2021
Darwin finches	Geospiza	1.5	14	N/A	New and unused food resources	Beak size and behavioral innovation	Yes	Grant 1986; Grant and Grant 2008, 2016; Tebbich et al. 2010; Lamichhaney et al. 2015; Burress et al. 2021
<i>Liolaemus</i> lizards	<i>Liolaemus</i>	~39	>250	Andean uplift	Cold mountaintops	Viviparity	Yes	Esquerre et al. 2019

Continued

**Table 2. Continued**

Adaptive radiation	Clade	Age (myr)	Number of species	Associated geological event	Ecological opportunity	Key innovation(s)	Accepted as adaptive radiation following Schlüter (2000)	Key references
Caribbean Anolis	<i>Anolis</i>	~43–51	>400	Great Antilles and northern Lesser Antilles geology	Microhabitat differentiation promoted by Caribbean islands emergence	Adhesive toe pads, sexual dimorphism	Yes	Losos et al. 1998; Losos and Ricklefs 2009; Mahler et al. 2010; Huie et al. 2021
Rain frogs	<i>Eleutherodactylus</i>	~20	>167	N/A	Caribbean islands	Diversity of habitat use	Yes	Jiménez-Ortega et al. 2023
Caribbean pupfishes	<i>Cyprinodon</i>	0.01	~55	Caribbean islands complex geology, islands formation, and sea level changes	Availability of diverse freshwater habitats	Exceptional craniofacial divergence	Yes	Martin and Wainwright 2013; Martin 2016; Martin et al. 2017
Neotropical cichlids	Cichlinae	~60	600	N/A	New lacustrine and riverine habitats (e.g., Central America)	Jaw and body size variability	Yes	Arbour and López-Fernández 2016
Platyrrhine monkeys	Platyrrhini	~25	160	N/A	Diet	Brain shape, face, neurocranium, and body/cranial size	Yes	Aristide et al. 2018
Neotropical leaf-nosed bats	Phyllostomidae	~30	227	N/A	Nocturnal noninsect diets	Variation in skull shape	Yes	Dumont et al. 2012; Rojas et al. 2018

Further details of these systems are provided in the text and the Supplemental Information.

as drivers of adaptive diversification (Schluter and McPhail 1992; Grant and Grant 2006). However, other types of ecological interactions including exploitative interactions might be promoting divergence during adaptive radiation (Schluter 2000). Understanding how some types of exploitative interactions, such as predation, could lead to diversification, is fundamental to defining the role of ecological interactions as drivers of adaptive radiations.

One of the most ubiquitous examples of exploitative interactions is the association between plants and their insect herbivore enemies. More than 50 years ago, Ehrlich and Raven's foundational paper proposed that selection by herbivores was central for plant adaptive radiation (Ehrlich and Raven 1964). Specifically, they predicted that after the evolution of a key innovation, in this case, a "new defense" in response to herbivory, plant species would be able to escape and radiate. Similarly, selection would favor counter-adaptations from herbivores to this new defense and ultimately adaptive radiation onto a set of host plants (Ehrlich and Raven 1964). Ehrlich and Raven argued these coevolutionary arms race may explain a substantial fraction of plant and insect diversities. Although enemy-driven adaptive radiation via the evolution of plant defenses was a key prediction from Ehrlich and Raven's (1964) coevolutionary hypothesis, it has remained largely understudied.

Recent advancements in phylogenetic and metabolomic tools are shedding light on the role that plant–herbivore interactions may play in the rapid and adaptive diversification of species-rich groups in Amazon tree lineages. The tree genus *Inga* (Fabaceae) exhibits the highest diversification rates in the Amazonian tree flora with more than 300 species (Richardson et al. 2001). *Inga* has rapidly radiated over the past 4–8 myr, displaying remarkable levels of sympatry with more than 40 species coexisting at a single site (Richardson et al. 2001; Valencia et al. 2004; Nicholls et al. 2015). Despite similar resource use, pollination, and dispersal traits (Koptur 1983; Pennington et al. 1997), divergence in defenses enables high congeneric coexistence, with co-occurring species being more dissimilar in defenses than expected by chance (Kursar et al. 2009; Forrister et al. 2019;

Endara et al. 2022). This is consistent with the idea that rapid evolutionary change is likely due to selection by herbivores (Kursar et al. 2009; Forrister et al. 2023).

Despite the consequences that multiple defensive niche dimensions have on fitness and adaptation, these differences have not been tested experimentally in *Inga*. Evidence suggests that different defensive traits (i.e., chemistry, developmental defenses, physical defenses) have diverse functions and evolve independently providing numerous niche dimensions, potentially driving speciation and adaptation in *Inga* (Coley and Kursar 2014; Endara et al. 2017). Empirical observations suggest that divergence in chemical defensive traits in populations of *Inga leiocalycina* in contrasting environments might be related to increased fitness (MJ Endara et al. unpubl.). Furthermore, feeding choice experiments and field surveys of sawfly larvae among populations of the *Inga capitata* complex in Peru showed that differences in chemical defenses are related to fitness when combined with asymmetries in other defensive axes (e.g., developmental, phenological) (Endara et al. 2015). Current evidence suggests that there might be two specific mechanisms operating on the evolution of adaptive chemical defensive traits in *Inga* (Endara et al. 2023; Forrister et al. 2023). The evolution of novel chemicals to escape from herbivores may be the result of gene duplication and neofunctionalization of sets of genes previously involved in a specific metabolic pathway to produce a chemical compound and compound classes (Lego chemistry scenario). These key innovations may open new adaptive zones for those lineages that have evolved such chemical defenses (Fig. 3).

Comparative evidence suggests the evolution of L-tyrosine, a primary metabolite overexpressed in the expanding leaves of *Inga*, might be a key innovation. This chemical compound, highly toxic for generalist herbivores at concentrations from 5% to 20%, evolved once in *Inga* ~4.7 myr in a clade comprising 21 species (Lokvam et al. 2006; Coley et al. 2019). Thus, the overexpression of this chemical compound class may have allowed the ancestor of the L-tyrosine clade to enter a new enemy-free space adaptive



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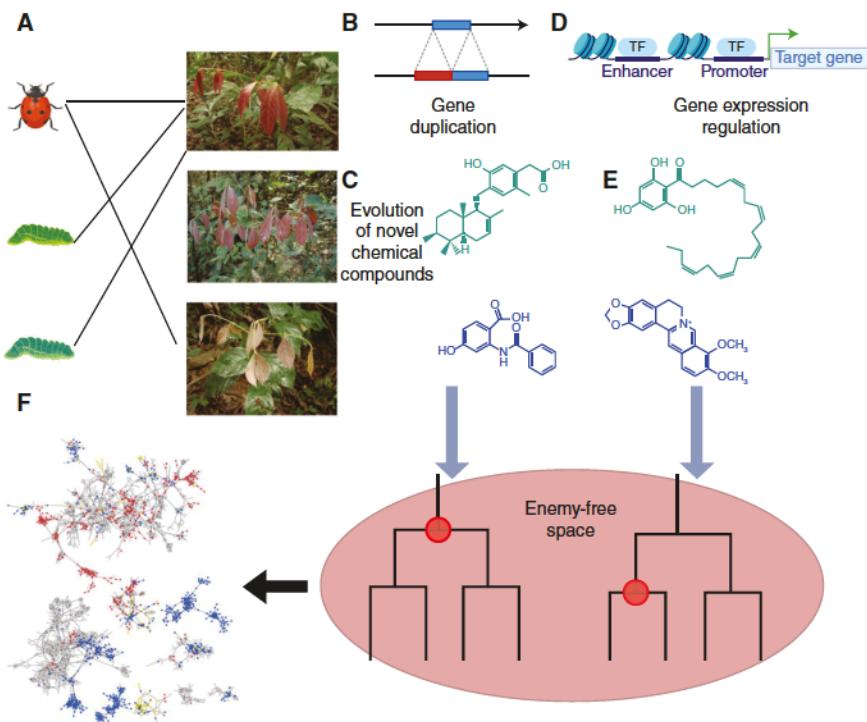


Figure 3. Conceptual framework for plant-herbivore mediated adaptive radiation in the neotropical tree genus *Inga*. (A) Selective pressure from herbivores promotes the evolution of chemical defenses to counterbalance the effects of herbivory on plant fitness. In the case of the neotropical genus *Inga* two main genetic mechanisms are proposed to be involved in the evolution of chemical defenses and subsequent radiation of host plants. (B) Duplication and subsequent neofunctionalization of specific genes involved in the expression of chemical structures used to deter herbivores should produce novel structures. Yet, just a small portion results in gene paralogs that can acquire potentially adaptive mutations under relaxed selection. These adaptive mutations eventually become fixed in the populations resulting in novel enzymatic functions. (C) Because the majority of gene duplications that result in novel defensive chemical structures are rare evolutionary events, chemical “key innovations” are conserved at deeper nodes of the phylogeny. (D) The second mechanism suggests that changes to the expression of individual or biosynthetically related metabolites result in the evolution of novel combinations of existing defense compounds leading to divergent chemical profiles among close relatives. (E) These changes might occur rapidly over evolutionary time leading to fixation on shallower nodes of the phylogeny. (F) The result is sympatric species that are highly divergent in their chemical profiles, as has been shown in the genus *Inga*.

zone. Several species within this clade have evolved more biologically derived compounds, including tyrosine and tyramine depsides, providing evidence for the “escalation of defense” through gradual modifications of core chemical structures. However, the mechanisms linking evolutionary changes driven by herbivores to reproductive isolation and subsequent speciation remain unclear. Marquis et al. (2016) proposed that coupled evolution of herbivore defense and pollinator attraction could act as a prezygotic isolation mechanism (Marquis et al. 2016;

Maron et al. 2019) and that parapatric and allopatric speciation mediated by selection against hybrids and spatial variation in herbivores could trigger divergent selection along environmental gradients (Marquis et al. 2016). In yet another mechanism, recent research suggests hybridization may have played a significant role in the rapid diversification of sympatric *Inga* species (MJ Endara, unpubl.), providing genetic fuel for combinatorial speciation and reshuffling of existing genomic variation related to chemical defenses (Marques et al. 2019; Schley et al.

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2020; Forrister et al. 2023). However, further theoretical and empirical work is required to integrate the evolution of chemical defenses with increased diversification in *Inga*.

Mutualistic Interactions

The diversification of the campanuloid clade in the bellflowers (family Campanulaceae) is one of the most striking examples of how ecological opportunity related to previously unoccupied habitats and unused resources triggers adaptive radiation in plants (Lagomarsino et al. 2014, 2016). In addition to orogeny and climate change, mutualistic relationships associated with seed dispersal and pollination syndromes contribute significantly to diversification rates in this clade (Lagomarsino et al. 2016). Bird-dispersed species with fleshy berries exhibit a 3.5-fold increase in diversification rates compared to species with abiotic dispersal (e.g., wind dispersal). A similar result arises when comparing species with vertebrate-pollinated flowers which exhibit an approximately sixfold increase in diversification rate relative to invertebrate-pollinated species (Lagomarsino et al. 2016). In the campanuloid clade, the increased speciation rates may be the result of floral isolation mediated by the interplay of floral morphology and pollinator behavior favoring the development of prezygotic reproductive isolation. In this case, the interplay between biotic and abiotic factors including floral morphology, pollinator behavior, and paleoelevation dynamics have contributed to the extreme diversification in these Andean bellflowers (Muchhal 2007; Lagomarsino et al. 2014, 2016).

Predator–Prey Relationships

Local adaptation in response to novel ecological conditions is not the only driver of adaptive radiation. Divergence in visual signals such as aposematic (warning) coloration has similarly produced a number of radiations in diverse animal taxa such as chemically defended butterflies (Kozak et al. 2015) and frogs (Symula et al. 2001) where differences in predation risk across geographic landscapes can drive phenotypic di-

vergence. In the Neotropics, multiple radiations of mimicry rings can be found, perhaps best exemplified by *Heliconius* butterflies (Kozak et al. 2015; Merrill et al. 2015) and *Ranitomeya* poison frogs (Symula et al. 2001).

Heliconius butterflies appear to be much more complex than a simple radiation in warning colorations, as recent studies have revealed a much more intricate reticulated evolutionary history comprising multiple mimicry rings including members of various genera (Mallet and Gilbert 1995; Merrill et al. 2015). Diversification can be the outcome of a suite of different mechanisms, including host plant specialization (Merrill et al. 2013), predator avoidance (Langham 2004), and differences in visual sensitivities (Briscoe et al. 2010; Finkbeiner et al. 2014). Despite these adaptations and degree of specialization, highly reticulate evolutionary histories between divergent taxa have been documented (Mallet et al. 2007; Nadeau et al. 2012; Kronforst et al. 2013). As the greatest *Heliconius* diversity is found in contiguous regions of Amazonia, speciation rates do not seem to be entirely driven by diversification in allopatry. Evidence suggests that isolation and divergence with secondary contact and speciation among parapatric species is plausible in some instances (Mallet and Turner 1997; Rosser et al. 2014). However, diversification (or speciation) seems to have occurred at different pace within and between lineages in this group (Kozak et al. 2015).

Similarly, mimicry drove a well-studied adaptive radiation in the poison frog *Ranitomeya imitator*. This species is the mimic of four congeners throughout its geographic range. Across this distribution, genotypic and phenotypic variation between *R. imitator* and the other species diverges (Twomey et al. 2013). This pattern corresponds to radiations of mimetic phenotypes driven by divergent selection on color and pattern elements (Yeager et al. 2012). Geographic transitions between mimetic phenotypes result in narrow clinal transitions which are maintained by both natural and sexual selection (Chouteau and Angers 2012; Twomey et al. 2014, 2016). Divergent phenotypes are maintained for mimetic accuracy (Yeager et al. 2012; Twomey et al. 2013, 2014), such that predators

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recognize and avoid local phenotypes (Chouteau and Angers 2012).

Resource Uses and Ecological Release

Phyllostomid bats, known as Neotropical leaf-nosed bats, comprise remarkable ecomorphological diversity (Freeman 2000). Their extensive adaptations, including occupation of diverse dietary niches ranging from ancestral insectivory to carnivory, nectarivory, frugivory, and even dedicated sanguivory, make them an exemplary adaptive radiation (Martin and Richards 2019). South America hosts the greatest extant species diversity of phyllostomids, with diversification and disparification occurring together (Rojas et al. 2016). Notably, the fig-eating phyllostomid subfamily Stenodermatinae experienced a significant increase in diversification rates because of a reorganized skull architecture, enabling access to figs and entered a new dietary adaptive zone with lower trophic level and almost tripled diversification rates (Dumont et al. 2012; Shi and Rabosky 2015; Rojas et al. 2018). Other adaptive peaks are associated with omnivory, nectarivory, and highly advantageous short-faced skulls (Dumont et al. 2014). Recent analyses suggest that only some phyllostomid lineages capitalized on ancient preadaptations within the family or even the neotropical superfamily Noctilionoidea (Davies et al. 2020; Hall et al. 2021; Potter et al. 2021), which only some phyllostomid lineages capitalized on.

HYBRIDIZATION, INTROGRESSION, AND ADMIXTURE AS DRIVERS OF SPECIATION AND ADAPTIVE RESPONSE TO ECOLOGICAL OPPORTUNITY

Speciation, the formation of reproductive barriers among populations, can arise from divergent selection, such as ecological or sexual selection, or genetic incompatibilities due to genetic drift or genomic conflict (Seehausen et al. 2014). In addition, hybridization and polyploidization can also contribute to speciation (Feder et al. 2012). In particular, recent evidence suggests hybridization among closely related species can lead to extensive adaptive responses to divergent natural selection, challenging the necessity of re-

productive isolation (Mayr 1947; Dobzhansky 1950; Seehausen et al. 2014; Grant and Grant 2019; Meier et al. 2019; Peñalba et al. 2024).

The arrival of a single lineage colonizer to a new habitat or ecological space where unused resources or no competitors (ecological opportunity) are present is a common scenario for an adaptive radiation (Schluter 2000; De-Kayne et al. 2024). In the presence of ample ecological opportunity, lineages radiate to occupy multiple vacant niches producing swarms of species with contrasting ecology. Multiple lines of evidence suggest that hybridization should trigger adaptive radiation in the presence of ecological opportunity (Meier et al. 2017, 2019; Meyer et al. 2017; Edelman et al. 2019). Whether hybridization predates adaptive radiation or closely related lineages arriving to a geographic area hybridize to successfully fill vacant niches is a matter of debate. Because hybridization of closely related lineages should not produce much genetic novelty and hybrids from distantly related lineages should be characterized by intrinsic incompatibilities (e.g., infertile, nonviable), one would expect that intermediate genetic distances between lineages maximize the role of hybridization in adaptive divergence (Meier et al. 2019).

Some lineages, like *Anolis* lizards, undergo rapid speciation and adaptive radiations, whereas others remain species-poor despite ecological opportunities (Losos and Thorpe 2004). The variance in speciation and adaptation rates is influenced by lineage-specific traits such as sexual selection, dispersal ability, phenotypic evolvability, and ecological versatility, all of which rely on standing genetic variation (Claramunt et al. 2012; Wagner et al. 2012; Rabosky et al. 2013; Stroud and Losos 2016; De-Kayne et al. 2024). High genetic variation facilitates speciation by increasing the potential for reproductive isolation and phenotypic evolution. It arises from new mutations or recombination of old genetic variants, forming genomic divergence islands resistant to gene flow (Han et al. 2017; Marques et al. 2019). Recent evidence also suggests that ancestral haplotypes act as genetic modules driving phenotypic diversity essential for species adaptation to changing environments (Rubin et al. 2022).



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Recent genomic studies on neotropical fauna have started to untangle the genomic architecture of speciation and reproductive isolation during adaptive radiation. These studies have particularly highlighted the importance of the reassembly of old genetic variants into novel combinations for rapid speciation. For instance, genomic divergence islands should exhibit higher absolute differentiation (d_{XY}) relative to the genetic background in both sympatric and allopatric pairs of species. This has been found in recent studies of the genetic architecture of speciation and adaptation in Darwin's finches, suggesting that gene flow plays a minor role in the evolution of genomic divergence islands and supporting the hypothesis that haplotypes at genomic islands became genetically isolated before the rest of the genomes (Lamichhaney et al. 2015; Han et al. 2017). Further, adaptive introgression of ancient genetic variants at the *ALX1* and *HMGA2* genes underlying beak traits has played a key role in the adaptive radiation of Darwin's finches in the Galápagos islands (Han et al. 2017). Because beak morphology is associated not only with adaptation to food resources but also to species discrimination and mate choice, it is plausible that these loci are also involved in reproductive isolation. This has been corroborated in a study showing evidence for rapid hybrid speciation involving a founder male with standing genetic variation related to large and blunt beaks for the *HMGA2 L* and the *ALX1 B* alleles (Lamichhaney et al. 2015). Thus, ancient polymorphisms and hybridization via introgression may have facilitated the diversification of beak morphology in terms of ecological opportunity.

Similarly, introgression of the large-effect ancient allele *optix* in New World *Heliconius* butterflies has enabled rapid diversification and the generation of novel phenotypes (Reed et al. 2011). The evolution of novel wing scale patterns through hybridization represents the evolution of key innovations that facilitated the occupation of multiple different ecological spaces mediated by ecological opportunity. *Optix* is one of the major genes responsible for mimetic wing pattern evolution, controlling extreme red wing pattern variation in this rapidly radiating butterfly

genus. The co-option of this pleiotropic gene contributed to the explosive diversification of wing patterns and adaptive radiation (Martin et al. 2014). Moreover, cross-species introgression of *optix* alleles is responsible for the evolution of multiple mimetic convergences across *Heliconius*. While introgression among *Heliconius* species is not directly involved in the formation of novel key traits, it is related to the propagation of adaptive variants across gene pools. In this way, hybrids may exhibit the effects of novelty through epistasis between wing pattern loci (Martin et al. 2014).

Speciation patterns in the Galápagos Islands endemic plant *Scalesia* are in agreement with an ancestral colonization in multiple islands followed by within-island speciation (Fernández-Mazuecos et al. 2020). In addition, signatures of hybridization in *Scalesia* radiation might be related to the evolution of partially heterogamous flowers via introgression (Lindhardt et al. 2009; Fernández-Mazuecos et al. 2020; Cerca et al. 2023). Strong signatures of hybridization during within-island speciation in *Scalesia* come from the evidence of introgression between four species with partially heterogamous capitula *S. retroflexa*, *S. incisa*, *S. hellerii*, *S. baurii*, and the widespread species with fully heterogamous capitula *S. affinis* (Fernández-Mazuecos et al. 2020). The occurrence of partially heterogamous capitula in *S. retroflexa* and *S. baurii* is the result of introgression with *S. affinis*. These introgression events are possible because the three species coexist in a single island in the Galápagos archipelago (Lindhardt et al. 2009; Fernández-Mazuecos et al. 2020).

Ecological opportunity related to novel trophic specialization is a key signature of the Caribbean pupfishes' adaptive radiation (Martin and Wainwright 2013; Martin 2016; McGirr and Martin 2017; Richards and Martin 2017). Accelerated rates of trophic diversification and shifts to new adaptive zones in *Cyprinodon* species are the product of fixation of de novo mutations during adaptive radiation toward distant phenotypic optima (Martin 2016; Martin et al. 2017; McGirr and Martin 2017). These adaptive peaks are related to the evolution of novel ecological traits allowing the occupation of new tro-

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phic niches in the specialist scale-eaters and snail-eaters compared to their single ancestral generalist species (Martin et al. 2017; McGirr and Martin 2017). Adaptive introgression of candidate genes associated with jaw development has been detected to be the result of selective sweeps favoring the speciation process of trophic specialists from a single ancestor from the Bahamas in the case of the *Cyprinodon* species in San Salvador (Richards and Martin 2017). However, speciation rates in San Salvador line-

ages are fivefold higher among populations within-island species compared to the role of adaptive introgression from Caribbean genetic diversity.

In sum, biased hybridization between some individuals from one species with heterospecifics should be related to adaptive introgression of ancient genetic variation from the donor species to a recipient species (Fig. 4). This process may fuel adaptive responses in the form of novel traits (key innovations) in response to new open

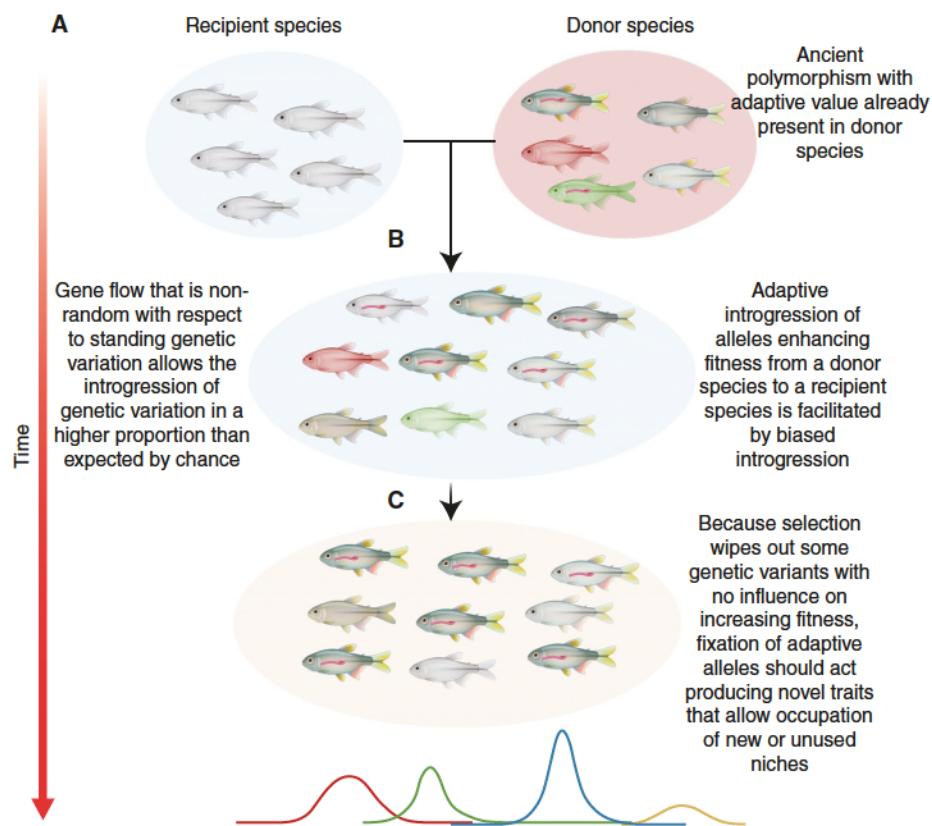


Figure 4. Hybridization determines which alleles initially move from one population to another. (A) Biased hybridization should facilitate nonrandom sampling of genetic variation to move from a donor to a recipient species. Because this genetic variation is the result of nonrandom hybridization with respect to segregating variation in a population there is an overrepresentation of genetic variation from donor species in the recipient species. If these variants are adaptive in the recipient species, such biases can enhance the likelihood of adaptive introgression. (B) Individuals carrying novel variants in the hybrid species are subject to selection in a similar ecological space to the recipient species (light blue background). (C) Over time these novel adaptive alleles are selectively favored if hybrids colonize a novel environment (light red background) or unused resources are available in such a way that hybrids descendants occupy these novel trophic niches. Deleterious variants (individuals in red and green color) are eventually lost. Colored waves represent niche partitioning among coexisting species in sympatry.

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niches or unused resources (ecological opportunity). However, the prevalence of this process in producing adaptive radiation is still a matter of debate and needs to be further investigated.

FUTURE DIRECTIONS

The advent of metabolomics, genomics has opened the door to new questions and tests as some of the most recent studies in adaptive radiation have demonstrated. For instance, metabolomic analyses in the genus *Espeletia* showed that there is strong biogeographic differentiation in the secondary metabolites fingerprint of lineages distributed across the five main biogeographic regions in the paramos of Colombia and Venezuela (Padilla-González et al. 2017). These results support the hypothesis that diversification has resulted in sets of different groups of secondary metabolites, which might be responsible for widespread chemical innovations to deter different herbivores in the adaptive radiation of *Espeletia*. Further studies combining genomics and metabolomics in *Espeletia* and other systems might shed light on this finding. Identifying the set of genes responsible for the evolution of major secondary metabolites might enable us to detect differential selection in populations along spatial and environmental gradients, as has been found in hummingbirds.

Determining how genomic mechanisms facilitate or constrain adaptive radiation once a lineage encounters a new adaptive zone is a long-standing goal in evolutionary biology. Recent advances in genomic sequencing technologies have provided researchers with a powerful tool to study the genetic mechanisms underlying adaptation and speciation, as well as the timing and order of evolutionary events. For instance, when comparing two sympatric species, a positive relationship between speciation rates and the number of fixed indels relative to the time of their last most recent common ancestor (MRCA) should be a signature of divergent selection (McGee et al. 2020). This suggests that variation in speciation rates can be associated with whether a lineage has unusually many large indels for its age in ecologically associated genome regions (McGee et al. 2020). This highlights the need to

analyze indels and single-nucleotide polymorphisms at the genome level in studies of adaptive radiation and speciation (McGee et al. 2020). Moreover, genomic data can provide valuable information about which indels are associated with ecological variables, capturing the major ecological dimensions of adaptive radiation. Combining genomically informed speciation research with macroevolutionary analyses of diversification enables discovering why some clades produce spectacular radiations, whereas others do not (Gillespie et al. 2020). Furthermore, genomic analyses can reveal how often repeated evolutionary transitions to new ecological niches are associated with the repeated genetic changes across independent lineages. In addition, genomics can provide insights into the molecular basis of morphological and physiological adaptations, and how genomic architecture and the source of genetic variation may promote or constrain adaptive radiation into a complex, multidimensional niche space (Marques et al. 2022).

Studying the adaptive landscape is crucial for understanding adaptive radiation as it provides insights into the evolutionary trajectories of species diversification. By examining how environmental factors impact the distribution of traits within a population, researchers can unravel the mechanisms driving adaptive radiations, shedding light on the origins and maintenance of biodiversity.

The adaptive landscape, originally defined by Wright (1932) as a multidimensional space in which each dimension corresponds to allele frequencies and mean fitness for a particular allele frequency to the height of this surface, has been fundamental to understanding how natural selection affects population survival and reproductive success. Simpson (1944, 1953) proposed a modification of this idea arguing that to better understand the role of divergent natural selection on survival and reproductive success in a particular environment we should account for phenotype differences in the adaptive landscape (Schluter 2000). Since then, the study of the adaptive landscapes has been focused on determining how morphological adaptation and ecological performance relate to ecological di-

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vergence (Grant and Grant 2002; Losos and Mahler 2010; Mahler et al. 2010; Brawand et al. 2014). Adaptive landscape topologies may represent the effects of stabilizing selection as a unimodal distribution in trait space, while disruptive selection should be related to multimodal distributions in trait space. When no selection is involved, the distribution should be uniform (Pfaender et al. 2016). Comparisons of variation-related fitness in a single phenotypic trait have proven effective in estimating the adaptive landscape for Darwin's finches (Schluter and Grant 1984; Lawson and Petren 2017). However, natural selection may affect multiple traits simultaneously and therefore adaptive landscapes could be used to analyze multivariate axes of phenotypic trait variation.

Studies performed in the neotropical cichlids, Neotropical leaf-nosed bats, and Darwin's finches have provided compelling evidence on how morphological divergence and ecological differentiation lead to differential fitness as reflected in adaptive peaks (Arbour and López-Fernández 2014; Lawson and Petren 2017; Beausoleil et al. 2023). Using Ornstein-Uhlenbeck models coupled with ancestral trait estimation and phylogenetic multidimensional analysis to define morphospace disparity, Arbour and López-Fernández (2014) detected seven adaptive peaks in the adaptive radiation of neotropical cichlids (Cichlinae). Three adaptive peak shifts were detected, related to the occupation of different regions of the biomechanics and feeding functional morphospace by different lineages, demonstrating functional disparity compatible with different selective regimes (Arbour and López-Fernández 2014). Likewise, Ornstein-

Uhlenbeck analyses coupled with finite-element engineering models of the phyllostomid skull reveal four adaptive peaks associated with divergent diets, and corresponding functional performance within each adaptive zone (Dumont et al. 2014), with subsequent quantitative genetics and comparative analyses expanding the number of peaks while accounting for population variation (Rossoni et al. 2019). These examples demonstrate the power of trait comparative analyses to identify adaptive zones, and their links to functional performance in adaptive radiations.

Despite the availability of options for combining phylogenetics and genetics with phenotypic traits, there are few examples of adaptive landscapes in neotropical adaptive radiations (Beausoleil et al. 2023). However, novel technologies including genomics, proteomics, or metabolomics could enhance the analyses of adaptive landscapes or adaptive peak shifts. For instance, one could combine character displacement experiments related to pollinator access together with genomic and metabolomic analysis to shed light on the gene expression modules responsible for floral, ethological, and mechanical isolation in plant species coexisting in sympatry (Hodges and Dierig 2009; Moreira-Hernández and Muchhal 2019).

Trait utility and phenotype–environment correlations are key characteristics of adaptive radiations. Hence, to demonstrate the adaptive component of a particular radiation, it is necessary to test how well different phenotypes of descendant lineages fit into the divergent ecological spaces they occupy. Additionally, complementary evidence that confirms the differential performance of traits in contrasting environments is necessary.

BOX 2. OUTSTANDING QUESTIONS

1. How can adaptive introgression influence processes such as range expansion, exploring novel niches in response to global change, and avoiding extirpation/extinction?
2. What is the prevalence of adaptive hybridization in adaptive radiations?
3. How can we use genomic, metabolomic, and phylogenomic data in a standardized way in order to determine general patterns of ecology and evolution across systems in adaptive radiations?
4. How can we integrate geomorphological, environmental, and experimental data to fill gaps in our understanding of adaptive radiations?

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Reciprocal transplant experiments in contrasting environments, physiological performance along environmental gradients, or experimental interspecific pollen transfer might be incorporated as sine qua nonanalyses for trait utility.

CONCLUDING REMARKS

The unique and complex geography of the Neotropics, from its splendid isolation to the Andean uplift, is one of the main factors contributing to evolutionary radiations. Whereas many geological events have shaped the emergence of novel and complex habitats throughout South America, there is considerable variation in how different lineages have responded. Geomorphological dynamics of the Andean system, the Brazilian and Guiana Shield cratons as well as the historic events that shaped the Mata Atlantica are fundamental for the evolution of many iconic examples of adaptive radiation in South America, including among others *Espeletia*, bromeliads, primates, and *Heliconius* butterflies. Despite great advances, a better understanding of the processes underlying plant and animal adaptive radiations in the Neotropics requires integrating phylogenies with traits, traits with functional performance in particular niches, and, in some cases, traits to specific genes under natural selection (Box 2).

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REFERENCES

*Reference is also in this subject collection.

Alvim AMV, Santos RV, Roddaz M, Antoine PO, Ramos MIF, do Carmo DA, Linhares AP, Negri FR. 2021. Fossil

isotopic constraints (C, O and 87Sr/86Sr) on Miocene shallow-marine incursions in Amazonia. *Palaeogeogr Palaeoclimatol Palaeoecol* 573: 110422. doi:10.1016/j.palaeo.2021.110422

Arbour JH, López-Fernández H. 2014. Adaptive landscape and functional diversity of Neotropical cichlids: implications for the ecology and evolution of Cichlinae (Cichlidae; Cichliformes). *J Evol Biol* 27: 2431–2442. doi:10.1111/jeb.12486

Arbour JH, López-Fernández H. 2016. Continental cichlid radiations: functional diversity reveals the role of changing ecological opportunity in the Neotropics. *Proc Biol Sci* 283: 20160556. doi:10.1098/rspb.2016.0556

Aristide L, Bastide P, dos Reis SF, Pires dos Santos TM, Lopes RT, Perez SI. 2018. Multiple factors behind early diversification of skull morphology in the continental radiation of New World monkeys. *Evolution (NY)* 72: 2697–2711. doi:10.1111/evol.13609

Armijo R, Lacassin R, Coudurier-Curveur A, Carrizo D. 2015. Coupled tectonic evolution of Andean orogeny and global climate. *Earth Sci Rev* 143: 1–35. doi:10.1016/j.earscirev.2015.01.005

Barreto E, Lim MCW, Rojas D, Dávalos LM, Wüest RO, Machac A, Graham CH. 2023. Morphology and niche evolution influence hummingbird speciation rates. *Proc Biol Sci* 290: 20221793. doi:10.1098/rspb.2022.1793

Beausoleil MO, Lorena Carrión-Avilés P, Podos J, Camacho C, Rabadán-González J, Richard R, Lalla K, Raeymaekers JAM, Knutie SA, De León LF, et al. 2023. The fitness landscape of a community of Darwin's finches. *Evolution (NY)* 77: 2533–2546. doi:10.1093/evolut/qpad160

Bershaw J, Garzione CN, Higgins P, MacFadden BJ, Anaya F, Alvarenga H. 2010. Spatial-temporal changes in Andean plateau climate and elevation from stable isotopes of mammal teeth. *Earth Planet Sci Lett* 289: 530–538. doi:10.1016/j.epsl.2009.11.047

Brawand D, Wagner CE, Li YI, Malinsky M, Keller I, Fan S, Simakov O, Ng AY, Lim ZW, Bezault E, et al. 2014. The genomic substrate for adaptive radiations in African cichlid fish. *Nature* 513: 375–381. doi:10.1038/nature13726

Briscoe AD, Bybee SM, Bernard GD, Yuan F, Sison-Mangus MP, Reed RD, Warren AD, Llorente-Bousquets J, Chiao CC. 2010. Positive selection of a duplicated UV-sensitive visual pigment coincides with wing pigment evolution in *Heliconius* butterflies. *Proc Natl Acad Sci* 107: 3628–3633. doi:10.1073/pnas.0910085107

Buckup PA. 2011. The eastern Brazilian shield. In *Historical biogeography of neotropical freshwater fishes* (ed. Albert J). University of California Press, Berkeley, CA.

Burress ED, Muñoz MM. 2021. Ecological opportunity from innovation, not islands, drove the anole lizard adaptive radiation. *Syst Biol* 71: 93–104. doi:10.1093/sysbio/syab031

Cadena CD, Cuervo AM, Céspedes LN, Bravo GA, Krabbe N, Schulenberg TS, Derryberry GE, Silveira LF, Derryberry EP, Brumfield RT, et al. 2020. Systematics, biogeography, and diversification of *Scytalopus* tapaculos (Rhinocryptidae), an enigmatic radiation of Neotropical montane birds. *Auk* 137: ukz077. doi:10.1093/auk/ukz077

Carlquist S. 1930–1965. *Island life; a natural history of the islands of the world* (Sherwin Carlquist, Illus. by Sherwin

Neotropics and Adaptive Radiations

Carlquist, Jeanne R. Janish, and Charles S. Papp). American Museum of Natural History [by] the Natural History Press, Garden City, New York.

Cerca J, Cotoras D, Santander C, Bieker V, Hutchins L, Morin-Lagos J, Prada C, Kennedy S, Krehenwinkel H, Rominger A, et al. 2023. Multiple paths towards repeated phenotypic evolution in the spiny-leg adaptive radiation (Tetragnatha; Hawaii). *Mol Ecol* **32**: 4971–4985. doi:10.1111/mec.17082

Chouteau M, Angers B. 2012. Wright's shifting balance theory and the diversification of aposematic signals. *PLoS ONE* **7**: e34028. doi:10.1371/journal.pone.0034028

Claramunt S. 2010. Discovering exceptional diversifications are continental scales: the case of the endemic families of neotropical suboscine passerines. *Evolution (NY)* **64**: 2004–2019. doi:10.1111/j.1558-5646.2010.00971.x

Claramunt S, Derryberry EP, Brumfield RT, Remsen JV. 2012. Ecological opportunity and diversification in a continental radiation of birds: climbing adaptations and cladogenesis in the Furnariidae. *Am Nat* **179**: 649–666. doi:10.1086/664998

Coley PD, Kursar TA. 2014. On tropical forests and their pests. *Science* **343**: 35–36. doi:10.1126/science.1248110

Coley PD, Endara MJ, Ghabash G, Kidner CA, Nicholls JA, Pennington RT, Mills AG, Soule AJ, Lemes MR, Stone GN, et al. 2019. Macroevolutionary patterns in overexpression of tyrosine: an anti-herbivore defence in a speciose tropical tree genus, *Inga* (Fabaceae). *J Ecol* **107**: 1620–1632. doi:10.1111/1365-2745.13208

Cuatrecasas J. 2013. A systematic study of the subtribe Espeletiinae (Heliantheae, Asteraceae). In *Memoirs of the New York Botanical Garden*, Vol. 107, pp. i–xii, 1–689. New York Botanical Garden, Bronx, New York.

Czekanski-Moir JE, Rundell RJ. 2019. The ecology of non-ecological speciation and nonadaptive radiations. *Trends Ecol Evol* **34**: 400–415. doi:10.1016/j.tree.2019.01.012

Davies KTJ, Yohe LR, Almonte J, Sánchez MKR, Rengifo EM, Dumont ER, Sears KE, Dávalos LM, Rossiter SJ. 2020. Foraging shifts and visual preadaptation in ecologically diverse bats. *Mol Ecol* **29**: 1839–1859. doi:10.1111/mec.15445

* De-Kayne R, Schley R, Barth JMI, Campillo LC, Chaparro-Pedraza C, Joshi J, Salzburger W, Van Bocxlaer B, Cotoras DD, Fruciano C, et al. 2024. Why do some lineages radiate while others do not? Perspectives for future research on adaptive radiations. *Cold Spring Harb Perspect Biol* doi:10.1101/cshperspect.a041448

Derryberry EP, Claramunt S, Derryberry G, Chesser RT, Cracraft J, Aleixo A, Pérez-Emán J, Remsen JV Jr, Brumfield RT. 2011. Lineage diversification and morphological evolution in a large-scale continental radiation: the neotropical ovenbirds and woodcreepers (Aves: Furnariidae). *Evolution (NY)* **65**: 2973–2986. doi:10.1111/j.1558-5646.2011.01374.x

Diazgranados M, Barber JC. 2017. Geography shapes the phylogeny of frailejones (Espeletiinae Cuatrec., Asteraceae): a remarkable example of recent rapid radiation in sky islands. *PeerJ* **5**: e2968. doi:10.7717/peerj.2968

Dobzhansky T. 1950. Evolution in the tropics. *Am Sci* **38**: 209–221.

Donoghue MJ, Sanderson MJ. 2015. Confluence, synnovation, and depauperation in plant diversification. *New Phytol* **207**: 260–274. doi:10.1111/nph.13367

Drummond CS, Eastwood RJ, Miotto STS, Hughes CE. 2012. Multiple continental radiations and correlates of diversification in *Lupinus* (Leguminosae): testing for key innovation with incomplete taxon sampling. *Syst Biol* **61**: 443–460. doi:10.1093/sysbio/syr126

Dumont ER, Dávalos LM, Goldberg A, Santana SE, Rex K, Voigt CC. 2012. Morphological innovation, diversification and invasion of a new adaptive zone. *Proc R Soc B* **279**: 1797–1805. doi:10.1098/rspb.2011.2005

Dumont ER, Samadévam K, Grosse I, Warsi OM, Baird B, Dávalos LM. 2014. Selection for mechanical advantage underlies multiple cranial optima in new world leaf-nosed bats. *Evolution (NY)* **68**: 1436–1449. doi:10.1111/evol.12358

Edelman NB, Frandsen PB, Miyagi M, Clavijo B, Davey J, Dikow RB, García-Accinelli G, Van Belleghem SM, Patterson N, Neafsey DE, et al. 2019. Genomic architecture and introgression shape a butterfly radiation. *Science* **366**: 594–599. doi:10.1126/science.aaw2090

Ehrlich PR, Raven PH. 1964. Butterflies and plants: a study in coevolution. *Evolution (NY)* **18**: 586–608. doi:10.2307/2406212

Eliasson U. 1974. Studies in Galapagos plants. XIV: The genus *Scalesia Arn.* *Opera Bot* **36**: 1–117.

Endara MJ, Weinhold A, Cox JE, Wiggins NL, Coley PD, Kursar TA. 2015. Divergent evolution in antiherbivore defences within species complexes at a single Amazonian site. *J Ecol* **103**: 1107–1118. doi:10.1111/1365-2745.12431

Endara MJ, Coley P, Ghabash G, Nicholls J, Dexter K, Donoso D, Stone G, Pennington R, Kursar T. 2017. Coevolutionary arms race versus host defense chase in a tropical herbivore–plant system. *Proc Natl Acad Sci* **114**: 201707727. doi:10.1073/pnas.1707727114

Endara M, Soule AJ, Forrister DL, Dexter KG, Pennington RT, Nicholls JA, Loiseau O, Kursar TA, Coley PD. 2022. The role of plant secondary metabolites in shaping regional and local plant community assembly. *J Ecol* **110**: 34–45. doi:10.1111/1365-2745.13646

Endara MJ, Forrister D, Coley P. 2023. The evolutionary ecology of plant chemical defenses: from molecules to communities. *Annu Rev Ecol Evol Syst* **54**: 107–127. doi:10.1146/annurev-ecolsys-102221-045254

Esquerre D, Brennan IG, Catullo RA, Torres-Pérez F, Keogh JS. 2019. How mountains shape biodiversity: the role of the Andes in biogeography, diversification, and reproductive biology in South America's most species-rich lizard radiation (Squamata: Liolaemidae). *Evolution (NY)* **73**: 214–230. doi:10.1111/evo.13657

Feder JL, Egan SP, Nosil P. 2012. The genomics of speciation-with-gene-flow. *Trends Genet* **28**: 342–350. doi:10.1016/j.tig.2012.03.009

Fernández-Mazuecos M, Vargas P, McCauley RA, Monjas D, Otero A, Chaves JA, Guevara Andino JE, Rivas-Torres G. 2020. The radiation of Darwin's giant daisies in the Galápagos Islands. *Curr Biol* **30**: 4989–4998.e7. doi:10.1016/j.cub.2020.09.019

Finkbeiner SD, Briscoe AD, Reed RD. 2014. Warning signals are seductive: relative contributions of color and pattern to predator avoidance and mate attraction in *Heliconius*

J.E. Guevara-Andino et al.

butterflies. *Evolution (NY)* **68**: 3410–3420. doi:10.1111/evo.12524

Fitzpatrick JW. 1985. Form, foraging behavior, and adaptive radiation in the Tyrannidae. *Ornithol Monographs* **36**: 447–470. doi:10.2307/40168298

Forrister DL, Endara MJ, Younkin GC, Coley PD, Kursar TA. 2019. Herbivores as drivers of negative density dependence in tropical forest saplings. *Science* **363**: 1213–1216. doi:10.1126/science.aau9460

Forrister DL, Endara MJ, Soule AJ, Younkin GC, Mills AG, Lokvam J, Dexter KG, Pennington RT, Kidner CA, Nicholls JA, et al. 2023. Diversity and divergence: evolution of secondary metabolism in the tropical tree genus *Inga*. *New Phytol* **237**: 631–642. doi:10.1111/nph.18554

Freeman P. 2000. Macroevolution in Microchiroptera: recoupling morphology and ecology with phylogeny. *Evol Ecol Res* **2**: 317–335.

Garzione CN, Hoke GD, Libarkin JC, Withers S, MacFadden B, Eiler J, Ghosh P, Mulch A. 2008. Rise of the Andes. *Science* **320**: 1304–1307. doi:10.1126/science.1148615

Geist DJ, Snell H, Snell H, Goddard C, Kurz MD. 2014. A paleogeographic model of the Galápagos Islands and biogeographical and evolutionary implications. In *The Galápagos, geophysical monograph series*, pp. 145–166. American Geophysical Union, Washington, DC. doi:10.1002/9781118852538.ch8

Gillespie RG, Bennett GM, De Meester L, Feder JL, Fleischer RC, Harmon LJ, Hendry AP, Knope ML, Mallet J, Martin C, et al. 2020. Comparing adaptive radiations across space, time, and taxa. *J Hered* **111**: 1–20. doi:10.1093/jhered/esz064

Givnish TJ. 2015. Adaptive radiation versus “radiation” and “explosive diversification”: why conceptual distinctions are fundamental to understanding evolution. *New Phytol* **207**: 297–303. doi:10.1111/nph.13482

Givnish TJ, Barfuss MHJ, Van Ee B, Riina R, Schulte K, Horres R, Gonsiska PA, Jabaily RS, Crayn DM, Smith JAC, et al. 2011. Phylogeny, adaptive radiation, and historical biogeography in Bromeliaceae: insights from an eight-locus plastid phylogeny. *Am J Bot* **98**: 872–895. doi:10.3732/ajb.1000059

Givnish TJ, Barfuss MHJ, Ee BV, Riina R, Schulte K, Horres R, Gonsiska PA, Jabaily RS, Crayn DM, Smith JAC, et al. 2014. Adaptive radiation, correlated and contingent evolution, and net species diversification in Bromeliaceae. *Mol Phylogenet Evol* **71**: 55–78. doi:10.1016/j.ympev.2013.10.010

Grant PR. 1986. *Ecology and evolution of Darwin's finches* (Princeton Science Library Edition). Princeton University Press, Princeton, NJ.

Grant PR, Grant BR. 2002. Adaptive radiation of Darwin's finches: recent data help explain how this famous group of Galápagos birds evolved, although gaps in our understanding remain. *Am Sci* **90**: 130–139. doi:10.1511/2002.10.130

Grant PR, Grant BR. 2006. Evolution of character displacement in Darwin's finches. *Science* **313**: 224–226. doi:10.1126/science.1128374

Grant BR, Grant PR. 2008. Fission and fusion of Darwin's finches populations. *Philos Trans R Soc Lond B Biol Sci* **363**: 2821–2829. doi:10.1098/rstb.2008.0051

Grant PR, Grant RB. 2016. Introgressive hybridization and natural selection in Darwin's finches. *Biol J Linnean Soc* **117**: 812–822. doi:10.1111/bij.12702

Grant PR, Grant BR. 2019. Hybridization increases population variation during adaptive radiation. *Proc Natl Acad Sci* **116**: 23216–23224. doi:10.1073/pnas.1913534116

Gregory-Wodzicki KM. 2000. Uplift history of the Central and Northern Andes: a review. *Geol Soc Am Bull* **112**: 1091–1105. doi:10.1130/0016-7606(2000)112<1091:UHOTCA>2.0.CO;2

Gutiérrez-García TA, Vázquez-Domínguez E. 2013. Consensus between genes and stones in the biogeographic and evolutionary history of Central America. *Quat Res* **79**: 311–324. doi:10.1016/j.yqres.2012.12.007

Hall RP, Mutumi GL, Hedrick BP, Yohe LR, Sadier A, Davies KTJ, Rossiter SJ, Sears K, Dávalos LM, Dumont ER. 2021. Find the food first: an omnivorous sensory morphotype predates biomechanical specialization for plant based diets in phyllostomid bats. *Evolution (NY)* **75**: 2791–2801. doi:10.1111/evo.14270

Han F, Lamichhaney S, Grant BR, Andersson L, Webster MT. 2017. Gene flow, ancient polymorphism, and ecological adaptation shape the genomic landscape of divergence among Darwin's finches. *Genome Res* **27**: 1004–1015. doi:10.1101/gr.212522.116

Harpp K, Geist D. 2018. The evolution of Galápagos volcanoes: an alternative perspective. *Front Earth Sci* **6**: 50. doi:10.3389/feart.2018.00050

Harrington HJ. 1962. Paleogeographic development of South America. *Am Assoc Pet Geol Bull* **46**: 1773–1814. doi:10.1306/BC7438F1-16BE-11D7-8645000102C1865D

Hartmann LA, Delgado I. 2001. Cratons and orogenic belts of the Brazilian Shield and their contained gold deposits. *Mineralium Deposita* **36**: 207–217. doi:10.1007/s001260100175

Hervé F, Demant A, Ramos V, Pankhurst R, Suárez M. 2000. The southern Andes. Tectonic evolution of South America. *Int Geol Congr* 605–634.

Hodges SA, Dierieg NJ. 2009. Adaptive radiations: from field to genomic studies. *Proc Natl Acad Sci* **106**: 9947–9954. doi:10.1073/pnas.0901594106

Hoorn C, Guerrero J, Sarmiento-Pérez G, Lorente M. 1995. Andean tectonics as a cause for changing drainage patterns in Miocene northern South America. *Geology* **23**: 237–240. doi:10.1130/0091-7613(1995)023<0237:ATAACF>2.3.CO;2

Hoorn C, Wesselingh FP, ter Steege H, Bermudez MA, Mora A, Sevink J, Sanmartín I, Sanchez-Meseguer A, Anderson CL, Figueiredo JP, et al. 2010. Amazonia through time: Andean uplift, climate change, landscape evolution, and biodiversity. *Science* **330**: 927–931. doi:10.1126/science.1194585

Hoorn C, Mosbrugger V, Mulch A, Antonelli A. 2013. Biodiversity from mountain building. *Nat Geosci* **6**: 154–154. doi:10.1038/ngeo1742

Horton BK. 2018. Sedimentary record of Andean mountain building. *Earth Sci Rev* **178**: 279–309. doi:10.1016/j.earscirev.2017.11.025

Horton BK, DeCelles PG. 1997. The modern foreland basin system adjacent to the Central Andes. *Geology* **25**: 895–

Neotropics and Adaptive Radiations

898. doi:10.1130/0091-7613(1997)025<0895:TMFBSA>2.3.CO;2

Horton B, Parra M, Saylor J, Nie J, Mora A, Torres V, Stockli D, Strecker M. 2010. Resolving uplift of the Northern Andes using detrital zircon age signatures. *GSA Today* **20**: 4–9. doi:10.1130/GSATG76A.1

Hughes CE, Atchison GW. 2015. The ubiquity of alpine plant radiations: from the Andes to the Hengduan Mountains. *New Phytol* **207**: 275–282. doi:10.1111/nph.13230

Hughes C, Eastwood R. 2006. Island radiation on a continental scale: exceptional rates of plant diversification after uplift of the Andes. *Proc Natl Acad Sci* **103**: 10334–10339. doi:10.1073/pnas.0601928103

Huie JM, Prates I, Bell RC, de Queiroz K. 2021. Convergent patterns of adaptive radiation between island and mainland *Anolis* lizards. *Biol J Linnear Soc* **134**: 85–110. doi:10.1093/biolinnean/blab072

Hulsey CD, Hollingsworth PR, Fordyce JA. 2010. Temporal diversification of Central American cichlids. *BMC Evol Biol* **10**: 279. doi:10.1186/1471-2148-10-279

Itow S. 1995. Phytogeography and ecology of *Scalesia* (Compositae) endemic to the Galapagos Islands. <http://hdl.handle.net/10125/2271>

Jiménez-Ortega D, Valente L, Dugo-Cota Á, Rabosky DL, Vilà C, Gonzalez-Voyer A. 2023. Diversification dynamics in Caribbean rain frogs (*Eleutherodactylus*) are uncoupled from the anuran community and consistent with adaptive radiation. *Proc R Soc B Biol Sci* **290**: 20222171. doi:10.1098/rspb.2022.2171

Kadereit JW, von Hagen KB. 2003. The evolution of flower morphology in Gentianaceae-Swertiinae and the roles of key innovations and niche width for the diversification of *Gentianella* and *Halenia* in South America. *Int J Plant Sci* **164**: S441–S452. doi:10.1086/376880

Koptur S. 1983. Flowering phenology and floral biology of *Inga* (Fabaceae: Mimosoideae). *Syst Bot* **8**: 354–368. doi:10.2307/2418355

Kozak KM, Wahlberg N, Neild AFE, Dasmahapatra KK, Mallet J, Jiggins CD. 2015. Multilocus species trees show the recent adaptive radiation of the mimetic *Heliconius* butterflies. *Syst Biol* **64**: 505–524. doi:10.1093/sysbio/syv007

Kronforst MR, Hansen MEB, Crawford NG, Gallant JR, Zhang W, Kulathinal RJ, Kapan DD, Mullen SP. 2013. Hybridization reveals the evolving genomic architecture of speciation. *Cell Rep* **5**: 666–677. doi:10.1016/j.celrep.2013.09.042

Kursar TA, Dexter KG, Lokvam J, Pennington RT, Richardson JE, Weber MG, Murakami ET, Drake C, McGregor R, Coley PD. 2009. The evolution of antiherbivore defenses and their contribution to species coexistence in the tropical tree genus *Inga*. *Proc Natl Acad Sci* **106**: 18073–18078. doi:10.1073/pnas.0904786106

Lack D. 1983. *Darwin's finches*. Cambridge University Press, Cambridge.

Lagomarsino LP, Antonelli A, Muchhal N, Timmermann A, Mathews S, Davis CC. 2014. Phylogeny, classification, and fruit evolution of the species-rich Neotropical bellflowers (Campanulaceae: Lobelioidae). *Am J Bot* **101**: 2097–2112. doi:10.3732/ajb.1400339

Lagomarsino LP, Condamine FL, Antonelli A, Mulch A, Davis CC. 2016. The abiotic and biotic drivers of rapid diversification in Andean bellflowers (Campanulaceae). *New Phytol* **210**: 1430–1442. doi:10.1111/nph.13920

Lamichhaney S, Berglund J, Almén MS, Maqbool K, Grabherr M, Martinez-Barrio A, Promerová M, Rubin C-J, Wang C, Zamani N, et al. 2015. Evolution of Darwin's finches and their beaks revealed by genome sequencing. *Nature* **518**: 371–375. doi:10.1038/nature14181

Langham GM. 2004. Specialized avian predators repeatedly attack novel color morphs of *Heliconius* butterflies. *Evolution (NY)* **58**: 2783–2787. doi:10.1111/j.0014-3820.2004.tb01629.x

Lara LE, Reyes J, Jicha BR, Díaz-Naveas J. 2018. $^{40}\text{Ar}/^{39}\text{Ar}$ geochronological constraints on the age progression along the Juan Fernández Ridge, SE Pacific. *Front Earth Sci* **6**: 194. doi:10.3389/feart.2018.00194

Lawson LP, Petren K. 2017. The adaptive genomic landscape of beak morphology in Darwin's finches. *Mol Ecol* **26**: 4978–4989. doi:10.1111/mec.14166

Lim MCW, Witt CC, Graham CH, Dávalos LM. 2019. Parallel molecular evolution in pathways, genes, and sites in high-elevation hummingbirds revealed by comparative transcriptomics. *Genome Biol Evol* **11**: 1573–1585. doi:10.1093/gbe/evz101

Lim MCW, Bi K, Witt CC, Graham CH, Dávalos LM. 2021. Pervasive genomic signatures of local adaptation to altitude across highland specialist Andean hummingbird populations. *J Hered* **112**: 229–240. doi:10.1093/jhered/esab008

Lindhardt MS, Philipp M, Tye A, Nielsen LR. 2009. Molecular, morphological, and experimental evidence for hybridization between threatened species of the Galapagos endemic genus *Scalesia* (Asteraceae). *Int J Plant Sci* **170**: 1019–1030. doi:10.1086/605113

Lokvam J, Brenes-Arguedas T, Lee JS, Coley PD, Kursar TA. 2006. Allelochemical function for a primary metabolite: the case of L-tyrosine hyper-production in *Inga umbellifera* (Fabaceae). *Am J Bot* **93**: 1109–1115. doi:10.3732/ajb.93.8.1109

López-Fernández H, Winemiller KO, Honeycutt RL. 2010. Multilocus phylogeny and rapid radiations in Neotropical cichlid fishes (Perciformes: Cichlidae: Cichlinae). *Mol Phylogenet Evol* **55**: 1070–1086. doi:10.1016/j.ympev.2010.02.020

López-Fernández H, Arbour J, Winemiller K, Honeycutt R. 2013. Testing for ancient adaptive radiations in Neotropical cichlid fishes. *Evolution (NY)* **67**: 1321–1337. doi:10.1111/evol.12038

Losos JB. 2009. *Lizards in an evolutionary tree*, 1st ed. University of California Press, Berkeley, CA.

Losos JB, Mahler DL. 2010. Adaptive radiation: the interaction of ecological opportunity, adaptation, and speciation. *Evol Darwin First* **150**: 381–420.

Losos JB, Ricklefs RE. 2009. Adaptation and diversification on islands. *Nature* **457**: 830–836. doi:10.1038/nature07893

Losos J, Thorpe R. 2004. Evolutionary diversification of *Anolis* lizards: introduction. In *Adaptive speciation* (ed. Diekmann U, et al.), pp. 343–344. Cambridge University Press, Cambridge.

Losos JB, Jackman TR, Larson A, Queiroz K, Rodriguez-Schettino L. 1998. Contingency and determinism in replicated adaptive radiations of island lizards. *Science* **279**: 2115–2118. doi:10.1126/science.279.5359.2115

Luebert F. 2021. The two South American dry diagonals. *Front Biogeogr* **13**. doi:10.21425/F5FBG51267

Lujan NK, Armbruster JW, Albert J, Reis R. 2011. The guiana shield. *Hist Biogeogr Neotrop Freshw Fishes* **21**: 224.

Mahler DL, Revell LJ, Glor RE, Losos JB. 2010. Ecological opportunity and the rate of morphological evolution in the diversification of Greater Antillean Anoles. *Evolution (NY)* **64**: 2731–2745. doi:10.1111/j.1558-5646.2010.01026.x

Males J. 2018. Geography, environment and organismal traits in the diversification of a major tropical herbaceous angiosperm radiation. *AoB Plants* **10**: ply008. doi:10.1093/aobpla/ply008

Mallet J, Gilbert LE Jr. 1995. Why are there so many mimicry rings? Correlations between habitat, behaviour and mimicry in *Heliconius* butterflies. *Biol J Linn Soc* **55**: 159–180. doi:10.1111/j.1095-8312.1995.tb01057.x

Mallet J, Turner J. 1997. Biotic drift or the shifting balance—did forest islands drive the diversity of warningly coloured butterflies? In *Evolution on islands*, pp. 262–280. Oxford University Press, Oxford.

Mallet J, Beltrán M, Neukirchen W, Linares M. 2007. Natural hybridization in heliconiine butterflies: the species boundary as a continuum. *BMC Evol Biol* **7**: 28. doi:10.1186/1471-2148-7-28

Maron JL, Agrawal AA, Schemske DW. 2019. Plant–herbivore coevolution and plant speciation. *Ecology* **100**: e02704. doi:10.1002/ecy.2704

Marques DA, Meier JI, Seehausen O. 2019. A combinatorial view on speciation and adaptive radiation. *Trends Ecol Evol* **34**: 531–544. doi:10.1016/j.tree.2019.02.008

Marques DA, Jones FC, Di Palma F, Kingsley DM, Reimchen TE. 2022. Genomic changes underlying repeated niche shifts in an adaptive radiation. *Evolution (NY)* **76**: 1301–1319. doi:10.1111/evo.14490

Marquis RJ, Salazar D, Baer C, Reinhardt J, Priest G, Barnett K. 2016. Ode to Ehrlich and Raven or how herbivorous insects might drive plant speciation. *Ecology* **97**: 2939–2951. doi:10.1002/ecy.1534

Martin CH. 2016. The cryptic origins of evolutionary novelty: 1000-fold faster trophic diversification rates without increased ecological opportunity or hybrid swarm. *Evolution (NY)* **70**: 2504–2519. doi:10.1111/evo.13046

Martin CH, Richards EJ. 2019. The paradox behind the pattern of rapid adaptive radiation: how can the speciation process sustain itself through an early burst? *Annu Rev Ecol Evol Syst* **50**: 569–593. doi:10.1146/annurev-ecolsys-110617-062443

Martin C, Wainwright P. 2013. Multiple fitness peaks on the adaptive landscape drive adaptive radiation in the wild. *Science* **339**: 208–211. doi:10.1126/science.1227710

Martin A, McCulloch KJ, Patel NH, Briscoe AD, Gilbert LE, Reed RD. 2014. Multiple recent co-options of Optix associated with novel traits in adaptive butterfly wing radiations. *Evodevo* **5**: 7. doi:10.1186/2041-9139-5-7

Martin CH, Erickson PA, Miller CT. 2017. The genetic architecture of novel trophic specialists: larger effect sizes are associated with exceptional oral jaw diversification in a pupfish adaptive radiation. *Mol Ecol* **26**: 624–638. doi:10.1111/mec.13935

Martín González AM, Dalsgaard B, Nogués-Bravo D, Graham CH, Schleuning M, Maruyama PK, Abrahamczyk S, Alarcón R, Araújo AC, Araújo FP, et al. 2015. The macroecology of phylogenetically structured hummingbird–plant networks. *Glob Ecol Biogeogr* **24**: 1212–1224. doi:10.1111/geb.12355

Mayr E. 1947. Ecological factors in speciation. *Evolution (NY)* **1**: 263–288. doi:10.2307/2405327

Mayr E. 1970. *Populations, species, and evolution: an abridgement of animal species and evolution*. Belknap, Cambridge, MA.

McGee MD, Borstein SR, Meier JI, Marques DA, Mwaiko S, Taabu A, Kishe MA, O'Meara B, Bruggmann R, Excoffier L, et al. 2020. The ecological and genomic basis of explosive adaptive radiation. *Nature* **586**: 75–79. doi:10.1038/s41586-020-2652-7

McGirr JA, Martin CH. 2017. Novel candidate genes underlying extreme trophic specialization in Caribbean pupfishes. *Mol Biol Evol* **34**: 873–888. doi:10.1093/molbev/msw286

McGuire JA, Witt CC, Remsen JV Jr, Corl A, Rabosky DL, Altshuler DL, Dudley R. 2014. Molecular phylogenetics and the diversification of hummingbirds. *Curr Biol* **24**: 910–916. doi:10.1016/j.cub.2014.03.016

Meier JI, Marques DA, Mwaiko S, Wagner CE, Excoffier L, Seehausen O. 2017. Ancient hybridization fuels rapid cichlid fish adaptive radiations. *Nat Commun* **8**: 14363. doi:10.1038/ncomms14363

Meier JI, Stelkens RB, Joyce DA, Mwaiko S, Phiri N, Schlieben UK, Selz OM, Wagner CE, Katongo C, Seehausen O. 2019. The coincidence of ecological opportunity with hybridization explains rapid adaptive radiation in Lake Mweru cichlid fishes. *Nat Commun* **10**: 5391. doi:10.1038/s41467-019-13278-z

Merrill RM, Naisbit RE, Mallet J, Jiggins CD. 2013. Ecological and genetic factors influencing the transition between host-use strategies in sympatric *Heliconius* butterflies. *J Evol Biol* **26**: 1959–1967. doi:10.1111/jeb.12194

Merrill RM, Dasmahapatra KK, Davey JW, Dell'Aglio DD, Hanly JJ, Huber B, Jiggins CD, Joron M, Kozak KM, Llaurens V, et al. 2015. The diversification of *Heliconius* butterflies: what have we learned in 150 years? *J Evol Biol* **28**: 1417–1438. doi:10.1111/jeb.12672

Meseguer AS, Michel A, Fabre PH, Pérez Escobar OA, Chomicki G, Riina R, Antonelli A, Antoine PO, Delsuc F, Condamine FL. 2022. Diversification dynamics in the Neotropics through time, clades, and biogeographic region. *eLife* **11**: e74503. doi:10.7554/eLife.74503

Meyer BS, Matschiner M, Salzburger W. 2017. Disentangling incomplete lineage sorting and introgression to refine species-tree estimates for Lake Tanganyika cichlid fishes. *Syst Biol* **66**: 531–550. doi:10.1093/sysbio/syw069

Miller A, Mayr E, Schüz E. 1949. *Ornithologie als biologische Wissenschaft*. Vlg C. Winter, Heidelberg.

Monasterio M, Sarmiento L. 1991. Adaptive radiation of Espeletia in the cold Andean tropics. *Trends Ecol Evol* **6**: 387–391. doi:10.1016/0169-5347(91)90159-U



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Moreira-Hernández JI, Muchhal N. 2019. Importance of pollinator-mediated interspecific pollen transfer for angiosperm evolution. *Annu Rev Ecol Evol Syst* **50**: 191–217. doi:10.1146/annurev-ecolsys-110218-024804

Muchhal N. 2007. Adaptive trade-off in floral morphology mediates specialization for flowers pollinated by bats and hummingbirds. *Am Nat* **169**: 494–504. doi:10.1086/512047

Nadeau NJ, Whibley A, Jones RT, Davey JW, Dasmahapatra KK, Baxter SW, Quail MA, Joron M, ffrench-Constant RH, Blaxter ML, et al. 2012. Genomic islands of divergence in hybridizing *Heliconius* butterflies identified by large-scale targeted sequencing. *Philos Trans R Soc Lond B Biol Sci* **367**: 343–353. doi:10.1098/rstb.2011.0198

Nevado B, Atchison GW, Hughes CE, Filatov DA. 2016. Widespread adaptive evolution during repeated evolutionary radiations in New World lupins. *Nat Commun* **7**: 12384. doi:10.1038/ncomms12384

Nicholls J, Pennington R, Koenen E, Hughes C, Hearn J, Bunnefeld L, Dexter K, Stone G, Kidner C. 2015. Using targeted enrichment of nuclear genes to increase phylogenetic resolution in the neotropical rain forest genus *Inga* (Leguminosae: Mimosoideae). *Front Plant Sci* **6**: 710. doi:10.3389/fpls.2015.00710

Nosil P, Flaxman SM. 2010. Conditions for mutation-order speciation. *Proc Biol Sci* **278**: 399–407. doi:10.1098/rspb.2010.1215

Ohlson J, Fjeldså J, Ericson PGP. 2008. Tyrant flycatchers coming out in the open: phylogeny and ecological radiation of Tyrannidae (Aves, Passeriformes). *Zool Scr* **37**: 315–335. doi:10.1111/j.1463-6409.2008.00325.x

Olson ME, Arroyo-Santos A. 2015. How to study adaptation (and why to do it that way). *Q Rev Biol* **90**: 167–191. doi:10.1086/681438

Osborn HF. 1902. The law of adaptive radiation. *Am Nat* **36**: 353–363. doi:10.1086/278137

Padilla-González GF, Diazgranados M, Da Costa FB. 2017. Biogeography shaped the metabolome of the genus *Espeletia*: a phytochemical perspective on an Andean adaptive radiation. *Sci Rep* **7**: 8835. doi:10.1038/s41598-017-0943-1-7

* Peñalba JV, Runemark A, Meier JI, Singh P, Wogan GOU, Sánchez-Guillén R, Mallet J, Rometsch SJ, Menon M, Seehausen O, et al. 2024. The role of hybridization in species formation and persistence. *Cold Spring Harb Perspect Biol* doi:10.1101/cshperspect.a041445

Pennington TD, Wise R, Royal Botanic Gardens K. 1997. *The genus Inga* botany. Royal Botanic Gardens, Kew, Richmond, UK. <https://books.google.com.ec/books?id=ZxXHMgEACAAJ>

Perez TM, Andino JEG, Rivas-Torres G, Feeley KJ. 2023. Climate constrains photosynthetic strategies in Darwin's Daisies: a test of the climatic variability and jack-of-all-trades hypotheses. *Am Nat* **201**: 78–90. doi:10.1086/721957

Pérez-Escobar O, Zizka A, Bermúdez M, Meseguer AS, Condamine F, Hoorn C, Hooghiemstra H, Pu Y, Bogarín D, Boschman L, et al. 2022. The Andes through time: evolution and distribution of Andean floras. *Trends Plant Sci* **27**: 364–378. doi:10.1016/j.tplants.2021.09.010

Pfaender J, Hadiaty RK, Schliewen UK, Herder F. 2016. Rugged adaptive landscapes shape a complex, sympatric radiation. *Proc Biol Sci* **283**: 20152342. doi:10.1098/rspb.2015.2342

Potter JHT, Davies KJ, Yohe LR, Sanchez MKR, Rengifo EM, Struebig M, Warren K, Tsagkogeorga G, Lim BK, dos Reis M, et al. 2021. Dietary diversification and specialization in neotropical bats facilitated by early molecular evolution. *Mol Biol Evol* **38**: 3864–3883. doi:10.1093/molbev/msab028

Pouchon C, Fernández A, Nassar JM, Boyer F, Aubert S, Lavergne S, Mavárez J. 2018. Phylogenomic analysis of the explosive adaptive radiation of the *Espeletia* complex (Asteraceae) in the tropical Andes. *Syst Biol* **67**: 1041–1060. doi:10.1093/sysbio/syy022

Rabosky DL. 2017. Phylogenetic tests for evolutionary innovation: the problematic link between key innovations and exceptional diversification. *Philos Trans R Soc Lond B Biol Sci* **372**: 20160417. doi:10.1098/rstb.2016.0417

Rabosky DL, Santini F, Eastman J, Smith SA, Sidlauskas B, Chang J, Alfaro ME. 2013. Rates of speciation and morphological evolution are correlated across the largest vertebrate radiation. *Nat Commun* **4**: 1958. doi:10.1038/ncomms2958

Reed RD, Papa R, Martin A, Hines HM, Counterman BA, Pardo-Díaz C, Jiggins CD, Chamberlain NL, Kronforst MR, Chen R, et al. 2011. Optix drives the repeated convergent evolution of butterfly wing pattern mimicry. *Science* **333**: 1137–1141. doi:10.1126/science.1208227

Richards EJ, Martin CH. 2017. Adaptive introgression from distant Caribbean islands contributed to the diversification of a microendemic adaptive radiation of trophic specialist pupfishes. *PLoS Genet* **13**: e1006919. doi:10.1371/journal.pgen.1006919

Richardson JE, Pennington RT, Pennington TD, Hollingsworth PM. 2001. Rapid diversification of a species-rich genus of neotropical rain forest trees. *Science* **293**: 2242–2245. doi:10.1126/science.1061421

Roddaz M, Hermoza W, Mora A, Baby P, Parra M, Christophoul F, Brusset S, Espurt N. 2009. Cenozoic sedimentary evolution of the Amazonian foreland basin system. In *Amazonia: landscape and species evolution*, pp. 61–88. Wiley, Hoboken, NJ.

Rojas D, Warsi OM, Dávalos LM. 2016. Bats (Chiroptera: Noctilionoidea) challenge a recent origin of extant neotropical diversity. *Syst Biol* **65**: 432–448. doi:10.1093/sysbio/syw011

Rojas D, Ramos Pereira MJ, Fonseca C, Dávalos LM. 2018. Eating down the food chain: generalism is not an evolutionary dead end for herbivores. *Ecol Lett* **21**: 402–410. doi:10.1111/ele.12911

Rosser N, Dasmahapatra KK, Mallet J. 2014. Stable *Heliconius* butterfly hybrid zones are correlated with a local rainfall peak at the edge of the Amazon basin. *Evolution (NY)* **68**: 3470–3484. doi:10.1111/evo.12539

Rossoni DM, Costa BMA, Giannini NP, Marroig G. 2019. A multiple peak adaptive landscape based on feeding strategies and roosting ecology shaped the evolution of cranial covariance structure and morphological differentiation in phyllostomid bats. *Evolution (NY)* **73**: 961–981. doi:10.1111/evo.13715

Rubin CJ, Enbody ED, Dobrev MP, Abzhanov A, Davis BW, Lamichhaney S, Pettersson M, Sendell-Price AT, Sprehn CG, Valle CA, et al. 2022. Rapid adaptive radia-

tion of Darwin's finches depends on ancestral genetic modules. *Sci Adv* **8**: eabm5982. doi:10.1126/sciadv.eabm5982

Rundell RJ, Price TD. 2009. Adaptive radiation, nonadaptive radiation, ecological speciation and nonecological speciation. *Trends Ecol Evol* **24**: 394–399. doi:10.1016/j.tree.2009.02.007

Sanín MJ, Cardona A, Valencia-Montoya WA, Jiménez MFT, Carvalho-Madrigal S, Gómez AC, Bacon CD, Tangarife TR, Jaramillo JS, Zapata S, et al. 2022. Volcanic events coincide with plant dispersal across the Northern Andes. *Glob Planet Change* **210**: 103757. doi:10.1016/j.gloplacha.2022.103757

Schilling EE, Panero JL, Eliasson UH. 1994. Evidence from chloroplast DNA restriction site analysis on the relationships of *Scalesia* (Asteraceae: Heliantheae). *Am J Bot* **81**: 248–254. doi:10.1002/j.1537-2197.1994.tb15436.x

Schley RJ, Pennington RT, Pérez-Escobar OA, Helmstetter AJ, de la Estrella M, Larridon I, Sabino Kikuchi IAB, Barracough TG, Forest F, Klitgård B. 2020. Introgression across evolutionary scales suggests reticulation contributes to Amazonian tree diversity. *Mol Ecol* **29**: 4170–4185. doi:10.1111/mec.15616

Schlüter D. 2000. *The ecology of adaptive radiation*. Oxford University Press, Oxford.

Schlüter D, Grant PR. 1984. Determinants of morphological patterns in communities of Darwin's finches. *Am Nat* **123**: 175–196. doi:10.1086/284196

Schlüter D, McPhail JD. 1992. Ecological character displacement and speciation in sticklebacks. *Am Nat* **140**: 85–108. doi:10.1086/285404

Sedano RE, Burns KJ. 2010. Are the Northern Andes a species pump for Neotropical birds? Phylogenetics and biogeography of a clade of Neotropical tanagers (Aves: Thraupini). *J Biogeogr* **37**: 325–343. doi:10.1111/j.1365-2699.2009.02200.x

Seehausen O. 2006. African cichlid fish: a model system in adaptive radiation research. *Proc Biol Sci* **273**: 1987–1998. doi:10.1098/rspb.2006.3539

Seehausen O, Butlin RK, Keller I, Wagner CE, Boughman JW, Hohenlohe PA, Peichel CL, Saetre GP, Bank C, Bränström Å, et al. 2014. Genomics and the origin of species. *Nat Rev Genet* **15**: 176–192. doi:10.1038/nrg3644

Shi JJ, Rabosky DL. 2015. Speciation dynamics during the global radiation of extant bats. *Evolution (NY)* **69**: 1528–1545. doi:10.1111/evol.12681

Silvestro D, Zizka G, Schulte K. 2014. Disentangling the effects of key innovations on the diversification of Bromeliidae (Bromeliaceae). *Evolution (NY)* **68**: 163–175. doi:10.1111/evol.12236

Simpson GG. 1944. *Tempo and mode in evolution*. Columbia University Press, New York.

Simpson GG. 1953. *The major features of evolution*. Columbia University Press, New York.

Stebbins GL. 1974. *Flowering plants*. Harvard University Press, Cambridge.

Stroud JT, Losos JB. 2016. Ecological opportunity and adaptive radiation. *Annu Rev Ecol Evol Syst* **47**: 507–532. doi:10.1146/annurev-ecolys-121415-032254

Symula R, Schulte R, Summers K. 2001. Molecular phylogenetic evidence for a mimetic radiation in Peruvian poison frogs supports a Müllerian mimicry hypothesis. *Proc Biol Sci* **268**: 2415–2421. doi:10.1098/rspb.2001.1812

Tebbich S, Sterelny K, Teschke I. 2010. The tale of the finch: adaptive radiation and behavioural flexibility. *Philos Trans R Soc B: Biol Sci* **365**: 1099–1109.

Temeles EJ, Koulouris CR, Sander SE, Kress WJ. 2009. Effect of flower shape and size on foraging performance and trade-offs in a tropical hummingbird. *Ecology* **90**: 1147–1161.

Testo WL, Sessa E, Barrington DS. 2019. The rise of the Andes promoted rapid diversification in Neotropical *Phlegmariurus* (Lycopodiaceae). *New Phytol* **222**: 604–613.

Twomey E, Yeager J, Brown JL, Morales V, Cummings M, Summers K. 2013. Phenotypic and genetic divergence among poison frog populations in a mimetic radiation. *PLoS ONE* **8**: e55443.

Twomey E, Vestergaard JS, Summers K. 2014. Reproductive isolation related to mimetic divergence in the poison frog *Ranitomeya imitator*. *Nat Commun* **5**: 4749.

Twomey E, Vestergaard JS, Venegas PJ, Summers K. 2016. Mimetic divergence and the speciation continuum in the mimic poison frog *Ranitomeya imitator*. *Am Naturalist* **187**: 205–224.

Valencia R, Condit R, Foster RB, Romoleroux K, Villa Muñoz G, Svenning J-C, Magård E, Bass M, Losos E, Balslev H. 2004. Yasuni forest dynamics plot, Ecuador. In *Tropical forest diversity and dynamism: findings from a large-scale plot network* (ed. Losos JB, Leigh EC Jr), p. 620. University of Chicago Press, Chicago.

Vinciguerra NT, Burns KJ. 2021. Species diversification and ecomorphological evolution in the radiation of tanagers (Passeriformes: Thraupidae). *Biol J Linnean Soc* **133**: 920–930.

von Hagen KB, Kadereit JW. 2003. The diversification of *Halenia* (Gentianaceae): ecological opportunity versus key innovation. *Evolution (NY)* **57**: 2507–2518. doi:10.1554/02-742

Wagner CE, Harmon LJ, Seehausen O. 2012. Ecological opportunity and sexual selection together predict adaptive radiation. *Nature* **487**: 366–369.

Walter GM, Wilkinson MJ, James ME, Richards TJ, Aguirre JD, Ortiz-Barrientos D. 2016. Diversification across a heterogeneous landscape. *Evolution* **70**: 1979–1992.

Wesselingh F, Anderson L, Kadolsky D. 2006. Molluscs from the Miocene Pebas formation of Peruvian and Colombian Amazonia. *Scripta Geologica*. <https://www.semanticscholar.org/paper/Molluscs-from-the-Miocene-Pebas-Formation-of-and-Wesselingh-Anderson/1697f935965fb6cd2e35eb5eefbd316d2c65b6b6> (Accessed April 1, 2023).

Winemiller KO, Kelso-Winemiller LC, Brenkert AL. 1995. Ecomorphological diversification and convergence in fluvial cichlid fishes. *Environ Biol Fishes* **44**: 235–261.

Wright S. 1932. The roles of mutation, inbreeding, cross-breeding, and selection in evolution. In *Proceedings of the VI International Congress of Genetics*, pp. 356–366. Blackwell, London.

Yeager J, Brown JL, Morales V, Cummings M, Summers K. 2012. Testing for selection on color and pattern in a mimetic radiation. *Curr Zool* **58**: 668–676.