

# Target and Nontarget Screening to Support Capacity Scaling for Substance Use Assessment through a Statewide Wastewater Surveillance Network in New York

Emily J. Vogel, Milagros Neyra, David A. Larsen, and Teng Zeng\*



Cite This: *Environ. Sci. Technol.* 2024, 58, 8518–8530



Read Online

ACCESS |

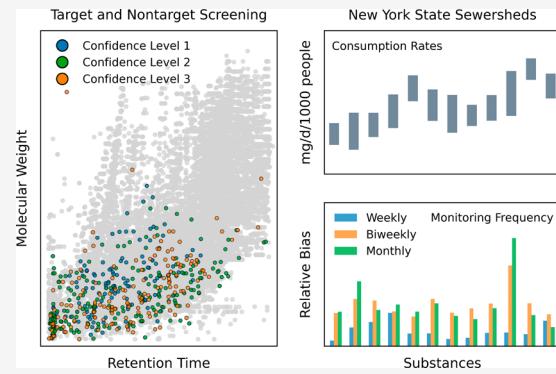
 Metrics & More

 Article Recommendations

 Supporting Information

**ABSTRACT:** Wastewater-based epidemiology (WBE) has been widely implemented around the world as a complementary tool to conventional surveillance techniques to inform and improve public health responses. Currently, wastewater surveillance programs in the U.S. are evaluating integrated approaches to address public health challenges across multiple domains, including substance abuse. In this work, we demonstrated the potential of online solid-phase extraction coupled with liquid chromatography–high-resolution mass spectrometry to support targeted quantification and nontargeted analysis of psychoactive and lifestyle substances as a step toward understanding the operational feasibility of a statewide wastewater surveillance program for substance use assessment in New York. Target screening confirmed 39 substances in influent samples collected from 10 wastewater treatment plants with varying sewershed characteristics and is anticipated to meet the throughput demands as the statewide program scales up to full capacity. Nontarget screening prioritized additional compounds for identification at three confidence levels, including psychoactive substances, such as opioid analgesics, phenethylamines, and cathinone derivatives. Consumption rates of 12 target substances detected in over 80% of wastewater samples were similar to those reported by previous U.S.-based WBE studies despite the uncertainty associated with back-calculations. For selected substances, the relative bias in consumption estimates was sensitive to variations in monitoring frequency, and factors beyond human excretion (e.g., as indicated by the parent-to-metabolite ratios) might also contribute to their prevalence at the sewershed scale. Overall, our study marks the initial phase of refining analytical workflows and data interpretation in preparation for the incorporation of substance use assessment into the statewide wastewater surveillance program in New York.

**KEYWORDS:** WBE, high-resolution mass spectrometry, nontargeted analysis, psychoactive substances, sewer



## INTRODUCTION

Wastewater-based epidemiology (WBE) involves the systematic analysis of untreated wastewater taken from sewer infrastructure to extract chemical signatures and/or biological markers and therefore represents a versatile tool for collecting community-wide information on various aspects of public health.<sup>1</sup> Many wastewater surveillance programs have been established or expanded in the U.S. and globally since the COVID-19 pandemic to track the emergence and circulation of SARS-CoV-2 variants.<sup>2</sup> For instance, the U.S. Centers for Disease Control and Prevention (CDC) launched the National Wastewater Surveillance System with partner agencies in September 2020 to monitor the spatiotemporal trends of SARS-CoV-2 viral activity levels and, more recently, the occurrence of monkeypox viral DNA, in wastewater.<sup>3</sup> In New York State (NYS), a collaborative wastewater surveillance network was also established to support pandemic management<sup>4</sup> and has expanded to cover 15.3 million residents (approximately 80% of the state's population).<sup>5</sup> With the COVID-19 pandemic subsiding, there is increasing momen-

tum to utilize the existing wastewater surveillance infrastructure in NYS and nationwide for assessing the impact of the opioid epidemic<sup>6</sup> and other substance use disorders given the rising number of fatal overdoses and nonfatal intoxications resulting from drug abuse in the U.S.<sup>7,8</sup>

Over the past decade, wastewater surveillance in Europe,<sup>9</sup> Australia,<sup>10</sup> and Canada<sup>11</sup> has proven successful in providing large-scale spatiotemporal data sets on population-level consumption of opioids, amphetamines, cocaine, cannabis, and other substances of concern. Wastewater surveillance in these regions has also demonstrated its potential to function as a warning system for detecting the emergence of new

Received: February 8, 2024

Revised: April 9, 2024

Accepted: April 16, 2024

Published: May 1, 2024



psychoactive substances despite their constantly changing profiles and lower prevalence of use compared to traditional drugs of abuse.<sup>12,13</sup> To date, most WBE studies in the U.S. have focused on assessing the mass loads and/or consumption rates of priority opioids and stimulants,<sup>14,15</sup> therapeutic drugs,<sup>16,17</sup> and lifestyle chemicals,<sup>18,19</sup> although recent efforts have sought to investigate the occurrence of emerging psychoactive substances.<sup>13,20</sup> Taken together, these studies have gathered important baseline data on substance consumption by diverse communities in the U.S. and underscored the opportunity to leverage ongoing or planned wastewater surveillance initiatives in supporting substance use assessment. From a practical standpoint, pivoting current wastewater surveillance initiatives, particularly those with broad spatio-temporal sampling regimes, to incorporate substance use assessment requires a phased approach to evaluate operational feasibility before the programs are scaled to reach their full potential.

Our primary objective of this study was to develop an integrated analytical framework that not only streamlines the routine, high-throughput quantification of target substances to ensure timely data sharing with stakeholders (e.g., health departments) and the public but also enables the qualitative screening of nontargeted analogs or derivatives that share similar structural cores with known or emerging substances of concern. To this end, we developed a target screening method based on online solid-phase extraction (SPE) coupled with liquid chromatography–high-resolution mass spectrometry (LC-HRMS) to quantify commonly targeted psychoactive and lifestyle substances (including their metabolites) in influent samples collected from 10 wastewater treatment plants (WWTPs) within the NYS wastewater surveillance network. Given the vast amount of chemical data embodied in untreated wastewater, we further developed a nontarget screening workflow to prioritize additional compounds, including new psychoactive substances, for identification at different confidence levels through mass spectral library searching complemented by the simultaneous filtering of diagnostic fragment ions and neutral losses extracted from the forensic toxicology literature. To place our work within the broader context of WBE, we also (i) estimated the population-level consumption rates of the 12 most frequently detected target substances for comparison with those previously measured for U.S. communities, (ii) evaluated the sensitivity of relative bias in substance consumption estimates to variations in monitoring frequency, and (iii) examined the parent-to-metabolite ratios of selected substances to assess the importance of human excretion relative to other sources in contributing to substance loads entering WWTPs. Our study did not aim to map substance consumption patterns in a spatiotemporally resolved manner or to establish a standardized protocol for substance use monitoring; instead, it served to explore the operational feasibility in anticipation of the growing capacity of wastewater surveillance for substance use assessment in the U.S.

## MATERIALS AND METHODS

**Chemicals and Materials.** Target substances of interest (Table S1) included compounds listed in the CDC's Opioid Polysubstance Mix Kit as well as additional psychoactive drugs and lifestyle chemicals frequently monitored in previous WBE studies. High-purity native standards of target substances ( $n = 51$ ) and isotope-labeled internal standards (ILIS;  $n = 42$ ) were

purchased from Sigma-Aldrich, Toronto Research Chemicals, and C/D/N Isotopes.

**Sample Collection.** Over the study period, 24 h flow-proportional composite influent samples were collected at mixed intervals (e.g., twice per week on weekdays) over multiple months between 2021 and 2022 from 10 WWTPs with an average design capacity of  $9.46 \times 10^3$  to  $3.19 \times 10^5$  m<sup>3</sup>/day (i.e., 2.5 to 84.2 million gallons per day), an estimated sewer transit time of  $0.6 \pm 1.3$  to  $4.1 \pm 2.8$  h, and a service population of 3076 to 242,377 (extracted from the U.S. Census Bureau's American Community Survey 2017–2021 5-Year Data<sup>21</sup>). Complete details of the sociodemographic attributes, health indicators, and opioid burdens of sewershed populations are summarized in Table S2. Samples were shipped overnight to SUNY Upstate Medical University for SARS-CoV-2 analysis and were stored at  $-80^{\circ}\text{C}$  until they were transferred to Syracuse University for substance analysis. General operational (e.g., flow rates) and hydrochemical parameters (e.g., 5-day biochemical oxygen demand (BOD<sub>5</sub>), 5-day carbonaceous BOD (CBOD<sub>5</sub>), total Kjeldahl nitrogen (TKN), and ammonia nitrogen (NH<sub>3</sub>-N)) were provided by the WWTPs.

**Sample Analysis.** Wastewater samples were spiked with a mixture of 42 ILIS (400 ng/L each), filtered by 0.22  $\mu\text{m}$  polyethersulfone syringe filters, and analyzed in duplicate by a Thermo Scientific TriPlus RSH autosampler and liquid handling system hyphenated with a Vanquish Horizon ultrahigh-performance liquid chromatograph and an Orbitrap Exploris 240 quadrupole-Orbitrap mass spectrometer. Briefly, 1 mL of filtered sample was loaded from a 5 mL stainless-steel sample loop onto a Hypersil GOLD aQ C18 trap column (20  $\times$  2.1 mm i.d., 12  $\mu\text{m}$ ) at 1 mL/min for preconcentration and extraction, and the trap column was subsequently washed with LC-MS grade water, followed by elution using the analytical pump gradient. Chromatographic separation was performed on a Hypersil GOLD aQ C18 analytical column (100  $\times$  2.1 mm, 1.9  $\mu\text{m}$ ; preceded with a 10  $\times$  2.1 mm guard cartridge) running LC-MS-grade water and methanol (acidified with 0.1% v/v formic acid) as the mobile phases at a flow rate of 200  $\mu\text{L}/\text{min}$  and a column temperature of  $35^{\circ}\text{C}$  for 32 min. Mass spectrometric analysis was conducted in positive and negative electrospray ionization modes. External mass calibration was performed using the Pierce FlexMix calibration solution. Internal mass calibration was activated through EASY-IC (fluoranthene) lock mass during data acquisition. Full-scan mass spectra were acquired from 100 to 1000 Da with a mass resolution of 120,000 at  $m/z$  200. Full-scan triggered data-dependent tandem mass (dd-MS2) spectra were acquired for targeted precursor ions in the inclusion list or for the five most intense precursor ions (excluding those registered in the exclusion list) with a mass resolution of 15,000 at  $m/z$  200 by higher energy collisional dissociation across five normalized collision energies ranging from 15 to 75%. Complete details of instrument settings and method parameters are provided in Tables S3–S5.

Calibration standards (i.e., prepared by spiking LC-MS grade water with 1–5000 or 500–25,000 ng/L of target substances and 400 ng/L of ILIS as a mixture) were run with each sample sequence. Continuous check standards (i.e., prepared by spiking deionized water with 400 ng/L of target substances and ILIS as a mixture) and procedural blanks were run every 10 samples to monitor any drift in instrument performance or carryover. Quality control samples (i.e., prepared by spiking pooled wastewater samples with 400 or

4000 ng/L of target substances and ILIS as a mixture) were also analyzed alongside nonspiked controls in triplicate within 1 day to evaluate intraday precision and accuracy and over 3 days to assess interday precision and accuracy. Complete details of the online SPE-LC-HRMS method performance (e.g., intraday/interday accuracy, intraday/interday precision, recoveries, and limits of quantification (LOQs) in wastewater) are summarized in **Table S6**.

**Target and Nontarget Screening.** Target screening was conducted using *TraceFinder* 5.2 SP1 (Thermo Scientific). Calibration curves for target substances were generated by  $1/x$ - or  $1/x^2$ -weighted linear or quadratic regression. Target substances were confirmed by verifying their chromatographic retention times and dd-MS2 spectra against those of the reference standards. Concentrations of substances detected in more than 50% of wastewater samples were quantified with reference to matching (i.e., structurally identical) ILIS or nonmatching ILIS with similar retention times. Complete details of *TraceFinder* method settings are provided in **Table S7**.

Nontarget screening was conducted using *Compound Discoverer* 3.3 SP2 (Thermo Scientific) with a node-based workflow consisting of spectrum processing (e.g., align retention times), compound detection (e.g., group compounds and fill gaps), peak area refinement (e.g., apply QC correction and mark background compounds), compound identification (e.g., search *mzCloud*, search *mzVault*, search *ChemSpider*, and search mass lists), and compound scoring (e.g., compound class scoring and search neutral losses). To extend library search beyond *mzCloud* and *MassBank*,<sup>22,23</sup> the search *mzVault* node was set to import the *High-Resolution Mass Spectral Libraries for Opioid Analysis* curated by the CDC<sup>24</sup> and the HighResNPS consensus library (October 2023 version).<sup>25</sup> Given that psychoactive substances typically contain characteristic structural cores,<sup>26</sup> the compound class scoring node was implemented to enable the filtering of diagnostic fragment ions for fentanyl analogs, synthetic cannabinoids, synthetic cathinones, phenyl-substituted phenethylamines, arylcyclohexylamines, and indolealkylamines (e.g., lysergamides and tryptamines).<sup>27</sup> To complement class coverage scoring, the search neutral loss node was also configured to enable the simultaneous filtering of neutral losses commonly observed for these substance classes.<sup>27</sup> With this workflow, mass spectral features that met all the following criteria were prioritized for inspection: a peak rating of  $>5$ , a mass accuracy tolerance of 5 ppm, a *mzCloud* and/or *mzVault* best match score of  $>60$ , and a retention time within the 95% confidence interval predicted by the LogP-retention time relationship established via the analysis of 432 compounds (e.g., pharmaceuticals, pesticides, personal care and household chemicals, industrial additives, and their transformation products) covering a range of polarities (**Figure S1**). Mass spectral features of interest were either confirmed by reference standards (i.e., confidence level 1), identified as probable structures (i.e., level 2), or assigned as tentative candidates (i.e., level 3) when applicable.<sup>28</sup> Complete details of *Compound Discoverer* node settings, the list of substances in the CDC and HighResNPS libraries, and fragment ions for selected classes of psychoactive substances are provided in **Tables S8–S10**.

**Substance Consumption Estimation.** For target substances ( $n = 12$ ) detected in over 80% of wastewater samples, the population-normalized mass loads (PNMLs) and con-

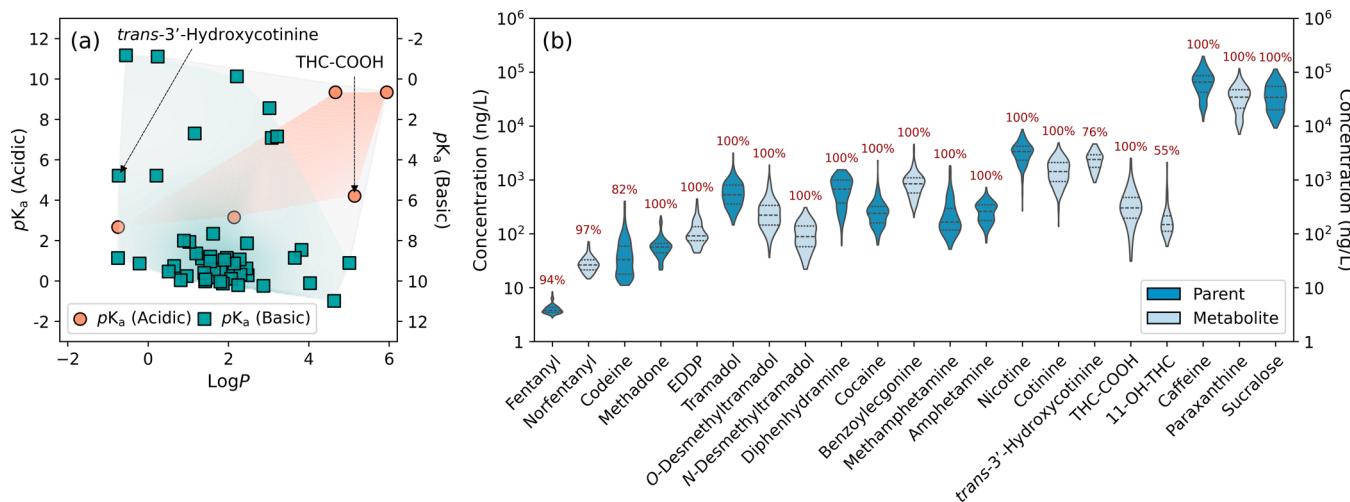
sumption rates (CRs) were back-calculated<sup>29,30</sup> using corresponding drug target residues (DTRs):

$$\text{PNML}_{i,j} = C_i \times Q_j \times \left( \frac{100}{100 + \text{stability}_i} \right) \times \left( \frac{100}{100 - \text{sorption}_i} \right) \times \left( \frac{1000}{\text{population}_j} \right) \quad (1)$$

$$\text{CR}_{i,j} = \text{PNML}_{i,j} \times \left( \frac{1}{\text{excretion}_i} \right) \times \left( \frac{\text{MW}_{i,\text{parent}}}{\text{MW}_{i,\text{DTR}}} \right) \quad (2)$$

where  $C_i$  is the aqueous concentration of substance  $i$  in each WWTP  $j$  influent sample averaged from duplicate measurements by online SPE-LC-HRMS (ng/L),  $Q_j$  is the average daily influent flow rate recorded by WWTP  $j$  for each sampling date (m<sup>3</sup>/day),  $\text{stability}_i$  is the percentage in-sample and/or in-sewer stability change of substance  $i$  derived from literature data (**Table S11**),  $\text{sorption}_i$  is the percentage of substance  $i$  sorbed to suspended particulate matter derived from literature data (**Table S12**),  $\text{PNML}_{i,j}$  is the mass load of substance  $i$  entering WWTP  $j$  normalized by the population in sewershed  $j$  (mg/day/1000 people) estimated for each sampling date,  $\text{population}_j$  is the *de facto* population in sewershed  $j$  estimated using the concentration of NH<sub>3</sub>-N measured in each WWTP  $j$  influent sample,  $\text{CR}_{i,j}$  is the consumption rate of substance  $i$  in sewershed  $j$  (mg/day/1000 people) estimated for each sampling date,  $\text{excretion}_i$  is the excretion rate of substance  $i$  derived from literature data (**Table S13**),  $\text{MW}_{i,\text{parent}}$  is the molecular weight of the parent compound of substance  $i$ , and  $\text{MW}_{i,\text{DTR}}$  is the molecular weight of the DTR of substance  $i$  (i.e., either the parent compound or its metabolite(s); **Table S14**). For each parameter (except for  $\text{MW}_{i,\text{parent}}$  and  $\text{MW}_{i,\text{DTR}}$ ), the uncertainty was characterized by a probability distribution (e.g., normal or beta distribution) as proposed by Jones *et al.*<sup>30</sup>

To propagate uncertainty (i.e., standard errors associated with parameter estimates) in **eqs 1** and **2**, Monte Carlo simulations were performed using *Colab Pro* (Google) to estimate the PNMLs and CRs of substances over 50,000 iterations.<sup>30</sup> Monte Carlo-simulated PNMLs and CRs were exported as the means and 95% confidence intervals and aggregated by substance. To contextualize the impact of the monitoring frequency on CR estimates, Monte Carlo simulations were also performed for bias analysis by subsampling CR data sets for three sampling intervals of practical relevance (i.e., weekly, biweekly, and monthly). Missing CR estimates were imputed using a random forest regressor following hyperparameter optimization and 10-fold cross-validation, and the distribution similarity between raw and imputed data was assessed by the Kolmogorov–Smirnov statistic. For each substance, the relative bias in CRs calculated from reduced sample sets was averaged across WWTPs to evaluate statistical differences in deviations from baseline values (i.e., calculated based on the complete CR data sets) for the three monitoring scenarios.<sup>31</sup> For a subset of target substances, the parent-to-metabolite (P:M) ratios were also calculated by dividing the PNMLs estimated using parent compounds by those estimated using corresponding metabolites to assess the relative importance of human excretion versus other sources in contributing to substance loads entering WWTPs.<sup>32</sup>



**Figure 1.** Target screening of substances in wastewater samples. (a) Target substances ( $n = 51$ ) validated by the online SPE-LC-HRMS method developed in this work. Shaded areas highlight the analytical space defined by target substances. Predicted LogP and  $pK_a$  of target substances are summarized in Table S15. Strongest acidic  $pK_a$  values are plotted on the left y-axis, whereas strongest basic  $pK_a$  values are plotted on the right y-axis (reversed). Sucralose is not plotted because it lacks ionizable atoms within the range of a minimal basic  $pK_a$  of  $-2$  and a maximum acidic  $pK_a$  of  $12$  as defined by MarvinSketch 23.10.0 (ChemAxon Ltd.). (b) Target substances ( $n = 21$ ) detected in over  $50\%$  of wastewater samples. On each violin plot, the dashed centerline marks the median, and the dotted lines bracket the interquartile range of concentrations. The percentage above each violin represents the detection frequency of each substance. Concentration ranges and detection frequencies of target substances are summarized in Table S16.

## RESULTS AND DISCUSSION

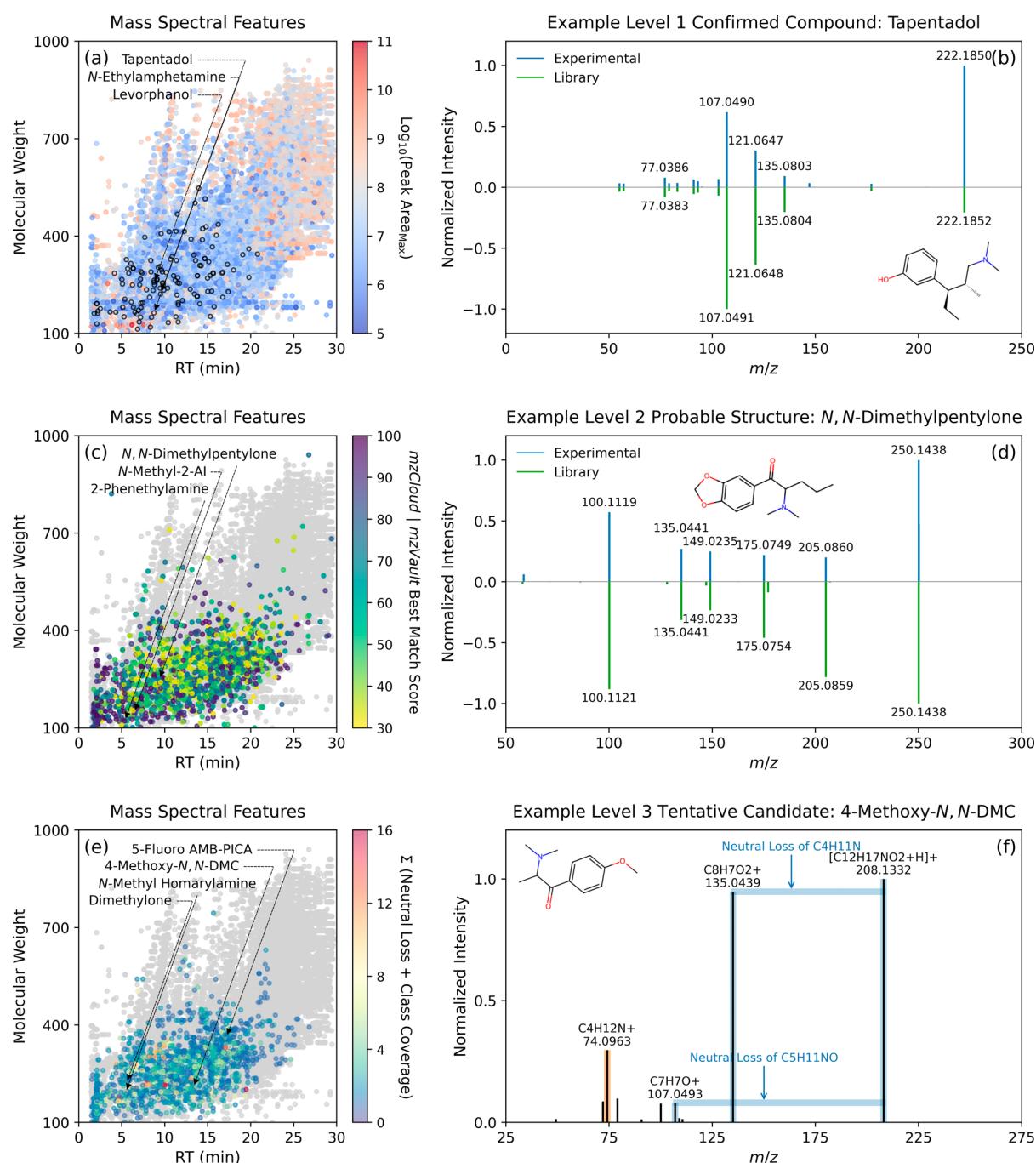
**Occurrence Patterns of Target Substances.** Our online SPE-LC-HRMS method enabled the high-throughput quantification of 51 substances, including 28 opioids and their metabolites, four benzodiazepines and their metabolites, and four amphetamines, as well as multiple lifestyle substances (i.e., cocaine, nicotine, cannabis, caffeine, and their metabolites). Compared to similar techniques developed by prior WBE studies (Figure S2),<sup>33–38</sup> our method achieved satisfactory sensitivity (LOQs in wastewater ranging from  $1.1$  to  $31$  ng/L) and captured both basic, hydrophilic compounds such as *trans*-3'-hydroxycotinine (with a predicted strongest basic  $pK_a$  of  $4.79$  and a predicted LogP of  $-0.73$ ; Figure 1a) and acidic, lipophilic compounds such as 11-nor-9-carboxy- $\Delta^9$ -tetrahydrocannabinol (THC-COOH; with a predicted strongest acidic  $pK_a$  of  $4.21$  and a predicted LogP of  $5.14$ ; Figure 1a). On average, the recoveries of target substances ranged from  $69 \pm 6\%$  for morphine-3-glucuronide to  $95 \pm 3\%$  for norfentanyl, the intraday or interday precision fell within  $17 \pm 10\%$ , and the intraday or interday accuracy varied between  $91 \pm 11\%$  and  $125 \pm 20\%$ , respectively (Figure S3). With a method runtime of  $32$  min, it was feasible to analyze up to  $40$  samples per day with continuous check standards and procedural blanks, which should provide the throughput needed to support fixed-interval (e.g., weekly to monthly) monitoring for WWTPs participating in the NYS wastewater surveillance network (Figure S4), assuming minimal supply chain disruptions and instrument downtime.

Overall, 39 out of 51 target substances were detected and confirmed in one or more WWTP influent samples (Table S16), of which 21 occurred in over  $50\%$  of samples at concentrations ranging from low ng/L to tens of  $\mu$ g/L (Figure 1b). Four opioids (i.e., fentanyl, codeine, methadone, and tramadol) and their metabolites (i.e., norfentanyl, 2-ethylidene-1,5-dimethyl-3,3-diphenylpyrrolidine (EDDP), O-desmethyltramadol, and N-desmethyltramadol) were detected in 82–100% of the samples. Furthermore, the concentrations of these

four opioids and their metabolites were positively correlated with the concentration of diphenhydramine (Spearman's  $\rho = 0.433$ – $0.894$ ;  $p < 0.0001$ ), which is an over-the-counter H1 antihistamine commonly combined with opioids as an adulterant.<sup>39</sup> Two amphetamines (i.e., amphetamine and methamphetamine) as well as cocaine and its metabolite benzoyllecgonine were detected in 100% of the samples. Amphetamine might be excreted as a minor metabolite of methamphetamine,<sup>40</sup> although their concentrations exhibited a much weaker correlation (Spearman's  $\rho = 0.418$ ;  $p < 0.0001$ ) than those observed for the other six parent-metabolite pairs (Spearman's  $\rho = 0.786$ – $0.974$ ;  $p < 0.0001$ ). Nicotine and its two metabolites (i.e., cotinine and *trans*-3'-hydroxycotinine) were detected in 76–100% of the samples, whereas the two metabolites of cannabis (i.e., THC-COOH and 11-hydroxy- $\Delta^9$ -tetrahydrocannabinol (11-OH-THC)) were detected in 100 and 55% of the samples, respectively. Caffeine, its metabolite paraxanthine, and sucralose were detected in every sample at the highest median concentrations of  $64.9$ ,  $34.6$ , and  $33.8$   $\mu$ g/L, respectively, as expected from their widespread consumption among the U.S. population.<sup>41,42</sup> Eighteen other target substances (i.e., meperidine, normeperidine, norcodeine, hydrocodone, morphine, oxycodone, noroxycodone, dihydromorphine, buprenorphine, norbuprenorphine, naloxone, alprazolam,  $\alpha$ -hydroxyalprazolam, diazepam, nordiazepam, norcocaine, 3,4-methylenedioxymethamphetamine, and 3,4-methylenedioxymphetamine) were also confirmed in wastewater samples but not quantified due to their low detection frequencies (i.e.,  $0.4$ – $10.9\%$ ).

### Nontarget Screening beyond Target Substances.

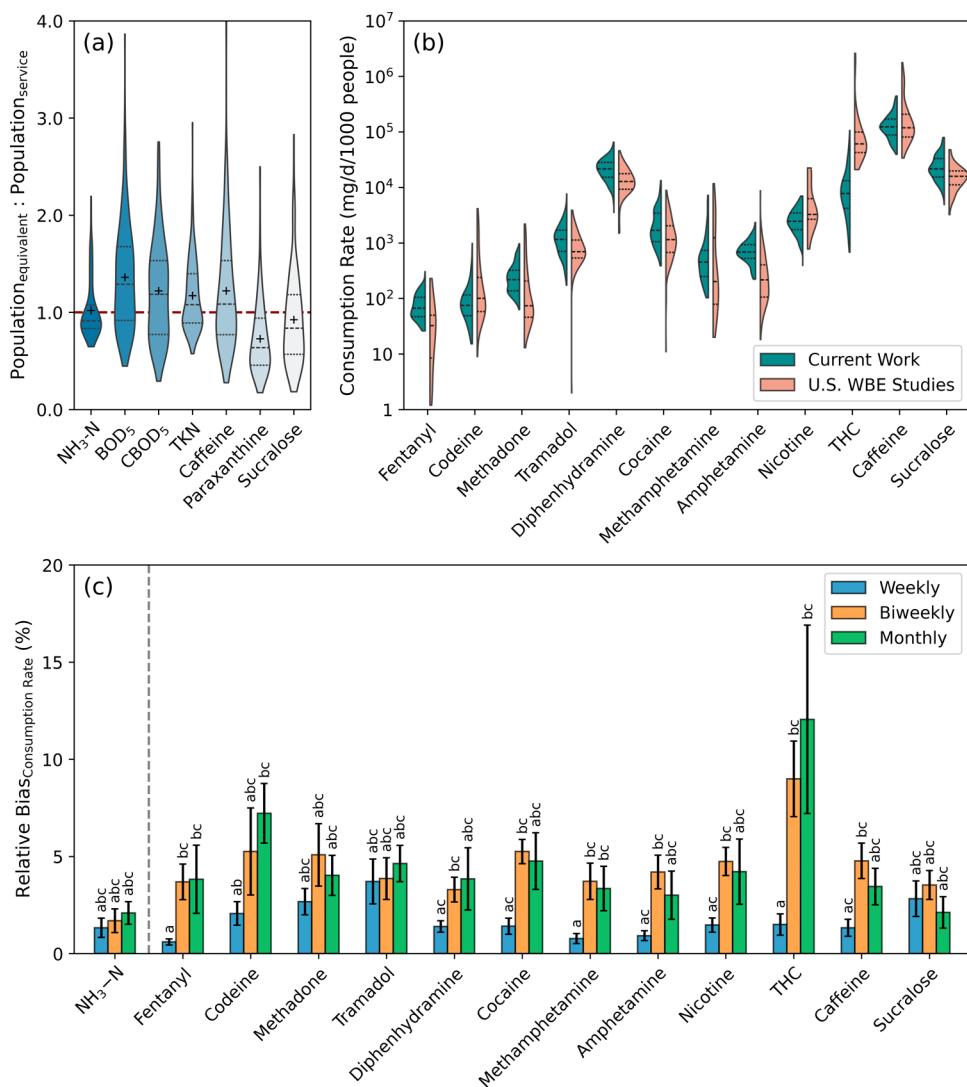
Nontargeted analysis of wastewater samples led to the detection of nonredundant mass spectral features<sup>43</sup> spanning a wide range of molecular weights, polarities, and peak intensities (Figure 2a). Of these mass spectral features, 86 were further confirmed at level 1 by reference standards (Table S17; beyond those detected by target screening), 196 were identified at level 2 as probable structures by library matching



**Figure 2.** Nontarget screening of substances in wastewater samples. (a) Mass spectral features (nonredundant) with a range of molecular weights, retention times, and peak intensities. The color bar measures the maximum peak areas of features on a logarithmic scale. Circles with black outline represent mass spectral features ( $n = 125$ , including 39 confirmed by target screening) confirmed at confidence level 1 by reference standards (Table S17). (b) Head-to-tail plot of experimental (top) and library (bottom) dd-MS2 spectra of tapentadol with additional fragmentation information provided in Table S20. (c) Mass spectral features with a range of *mzCloud* and/or *mzVault* best match scores. The color bar indicates the *mzCloud* or *mzVault* library best match score. (d) Head-to-tail plot of experimental (top) and library (bottom) dd-MS2 spectra of *N,N*-dimethylpentylone with additional fragmentation information provided in Table S23. (e) Mass spectral features with a range of diagnostic fragment ions and neutral losses present in the dd-MS2 spectra. The color bar measures the total number of diagnostic fragment ions and neutral losses. (f) Experimental dd-MS2 spectrum of mass spectral feature  $C_{12}H_{17}NO_2$  assigned tentatively as 4-methoxy-*N,N*-dimethylcathinone or its isomer based on the diagnostic fragment ions and neutral losses as detailed in Table S26.

(Table S18) and 158 were assigned level 3 as tentative candidates (Table S19), respectively. Confirmed compounds mostly consisted of over-the-counter and prescription pharmaceuticals such as anti-inflammatories (e.g., diclofenac and naproxen), antiarrhythmics (e.g., flecainide and propafenone), antiepileptics (e.g., carbamazepine and gabapentin),

antidepressants (e.g., bupropion and venlafaxine), antidiabetics (e.g., metformin and sitagliptin), antihypertensives (e.g., diltiazem and valsartan), antivirals (e.g., abacavir and darunavir), and contraceptives (e.g., norgestrel) as well as insect repellents (e.g., DEET and icaridin), cosmetic ingredients (e.g., benzophenones), pesticides (e.g., triazines),



**Figure 3.** Consumption estimates of target substances detected in over 80% of wastewater samples. (a) Comparison of sewershed populations (i.e., population<sub>equivalent</sub>) estimated based on hydrochemical parameters (i.e., NH<sub>3</sub>-N, 5-day biochemical oxygen demand, 5-day carbonaceous BOD, and total Kjeldahl nitrogen) or high-consumption substances such as caffeine (and its metabolite paraxanthine) and sucralose with service populations (i.e., population<sub>service</sub>). On each violin plot, the dashed centerline marks the median, the “+” sign marks the mean, and the dotted lines bracket the interquartile range of population<sub>equivalent</sub> to population<sub>service</sub> ratios. The maroon dashed line marks a ratio of 1.0. (b) Consumption rates of substances (in mg/day/1000 people) estimated in this work compared with those reported by 15 WBE studies conducted in the U.S. between 2014 and 2024 (Table S31). On each grouped violin plot, the dashed centerline marks the median, and the dotted lines bracket the interquartile range of consumption rates. For this work, the consumption rates of six substances were estimated via their respective metabolites: methadone via EDDP, tramadol via *O*-desmethyltramadol, cocaine via benzoylecgonine, nicotine via cotinine, THC via THC-COOH, and caffeine via paraxanthine. Sewershed-specific substance consumption rates are summarized in Table S32. (c) Comparison of the relative bias for estimating the consumption rates of 12 target substances and the mass load of NH<sub>3</sub>-N for weekly, biweekly, and monthly monitoring scenarios, where bars sharing the same letters are not statistically different (Mann-Whitney *U* test *p* ≥ 0.05). Error bars represent the standard deviation of the relative bias.

and rubber-derived chemicals (e.g., benzothiazoles and benzotriazoles), again highlighting the chemical diversity of substances in sewer systems.

Three of the compounds prioritized by nontarget screening (Tables S20–S22; detected in 5.3–27% of wastewater samples) were confirmed as psychoactive substances with abuse potential. Tapentadol (Figure 2b) is a synthetic opioid analgesic structurally similar to tramadol,<sup>44</sup> and its occurrence in untreated wastewater has been reported by WBE studies conducted in the Western U.S.,<sup>45</sup> Australia,<sup>46</sup> and Greece.<sup>47</sup> Levorphanol (Figure S6) is another synthetic opioid analgesic with properties similar to those of morphine<sup>44</sup> and has been identified in primary sludge extracts collected from WWTPs in

Connecticut during the early months of the COVID-19 pandemic.<sup>48</sup> Tapentadol and levorphanol were also detected at moderate frequencies in urban wastewater samples collected from multiple countries in a recent international collaborative study.<sup>49</sup> *N*-Ethylamphetamine (Figure S7) is an *N*-substituted derivative of amphetamine with lower potency and prevalence<sup>50</sup> and was previously identified as a synthesis impurity of amphetamine in WWTP influent samples from a Lithuanian city following a suspected dumping event.<sup>51</sup>

Three of the level 2 mass spectral features (Tables S23–S25; detected in 2.9–29% of wastewater samples) were identified as probable structures of psychoactive substances through mass spectral library matching. *N,N*-Dimethylpentylone (also known

as dipentylylone; **Figure 2d**) is a synthetic cathinone increasingly being identified in forensic toxicology samples in the U.S.<sup>52</sup> and has been detected at low ng/L levels in untreated wastewater collected on weekends or during special events in Spain.<sup>53,54</sup> *N*-Methyl-2-aminoindane (also known as *N*-methyl-2-*AI*; **Figure S9**) is a cyclic analog of methamphetamine, which has been tentatively identified in untreated wastewater from Poland<sup>56</sup> and Greece<sup>57</sup> and, more recently, quantified at a high frequency in South Korean WWTP influent samples.<sup>58</sup> 2-Phenethylamine (**Figure S10**) is a structural motif widely presented in endogenous catecholamines and naturally occurring alkaloids;<sup>59</sup> however, it is also a central nervous system stimulant and was first detected in WWTP influent samples collected from urban areas in Poland and Slovenia<sup>56</sup> as part of a European-wide study and later in pooled urine and untreated wastewater collected during music festivals in Norway and Portugal, respectively.<sup>60</sup>

Four level 3 mass spectral features (**Tables S26–S29**; detected in 0.4–37% of wastewater samples) were assigned as tentative candidates of psychoactive substances following the examination of diagnostic fragment ions and neutral losses present in their dd-MS2 spectra.  $C_{12}H_{17}NO_2$  (*m/z* 208.1332 for  $[M + H]^+$ ;  $\Delta$ Mass = −0.32 ppm) was tentatively identified as 4-methoxy-*N,N*-dimethylcathinone (also known as 4-methoxy-*N,N*-DMC; **Figure 2f**) or its isomer based on the characteristic neutral losses of  $C_4H_{11}N$  (e.g., *N,N*-dimethyl-*l*-alanine) and  $C_5H_{11}NO$  (e.g., 2-(dimethylamino)propanal) from the precursor ion to form the diagnostic (4-formylphenyl)(methylene)oxonium ion  $C_8H_7O_2^+$  (i.e., *m/z* 135.0439;  $\Delta$ Mass = −1.11 ppm) found for methoxy cathinones<sup>27</sup> and the 4-methoxybenzene-1-ylium ion  $C_7H_7O^+$  (i.e., *m/z* 107.0493;  $\Delta$ Mass = 1.40 ppm), respectively.  $C_{12}H_{15}NO_3$  (*m/z* 222.1124 for  $[M + H]^+$ ;  $\Delta$ Mass = −0.41 ppm) was tentatively identified as dimethyllylone (also known as bk-MDDMA; **Figure S12**) or its isomer based on the characteristic neutral losses of  $C_4H_{11}N$  and  $C_5H_{11}NO$  from the precursor ion to form the diagnostic (benzo[*d*][1,3]dioxol-5-ylmethyldiene)oxonium ion  $C_8H_5O_3^+$  (i.e., *m/z* 149.0230;  $\Delta$ Mass = −2.28 ppm) found for methylenedioxy cathinones<sup>27</sup> and the benzo[*d*][1,3]dioxol-5-ylium ion  $C_7H_5O_2^+$  (i.e., *m/z* 121.0284;  $\Delta$ Mass = 0.08 ppm), respectively.  $C_{11}H_{15}NO_2$  (*m/z* 194.1174 for  $[M + H]^+$ ;  $\Delta$ Mass = −0.57 ppm) was tentatively identified as *N*-methyl homarylamine (**Figure S13**) or its isomer based on the characteristic neutral losses of  $C_2H_7N$  (e.g., dimethylamine) and  $C_2H_7N$  plus CO (i.e.,  $C_3H_7NO$ ) from the precursor ion that led to the formation of two diagnostic ions, 1-(benzo[*d*][1,3]dioxol-5-yl)ethan-1-ylium ion  $C_9H_9O_2^+$  (i.e., *m/z* 149.0595;  $\Delta$ Mass = −1.68 ppm) and benzo[*d*][1,3]dioxol-5-ylmethylium ion  $C_8H_7O_2^+$  (i.e., *m/z* 135.0442;  $\Delta$ Mass = 0.81 ppm), as typically observed for methylenedioxy phenethylamines.<sup>27</sup>  $C_{20}H_{27}FN_2O_3$  (*m/z* 363.2078 for  $[M + H]^+$ ;  $\Delta$ Mass = 0.01 ppm) was tentatively identified as 5-fluoro AMB-PICA (also known as MMB-2201; **Figure S14**) or its isomer based on the detection of the diagnostic (1*H*-indol-3-yl)(oxo)methylium ion  $C_9H_8NO^+$  (i.e., *m/z* 144.0440;  $\Delta$ Mass = −2.71 ppm) found for indole-3-carboxamide-based synthetic cannabinoids<sup>27</sup> and the neutral loss of  $C_6H_{13}NO_2$  (e.g., methyl 2-amino-3-methylbutanoate) from the precursor ion to form the (1-(5-fluoropentyl)-1*H*-indol-3-yl)(oxo)methylium ion  $C_{14}H_{15}FNO^+$  (i.e., *m/z* 232.1130;  $\Delta$ Mass = −1.42 ppm). Overall, nontargeted analysis complemented target screening by prioritizing additional psychoactive substances for identification at confidence level

3 or higher, although reference standards are necessary to confirm probable structures and tentative candidates before quantitative analysis. Given the occurrence of tapentadol and levorphanol in multiple sewersheds, further analytical efforts are warranted to incorporate the monitoring of these two substances and their metabolites into the surveillance program.

**Consumption Estimates of Target Substances.** Comparing the CRs of substances targeted in this work with literature data is challenging due to differences in sampling design, analytical methods, and assumptions made for back-calculations, each of which can introduce varying degrees of uncertainty into the final interpretation of results.<sup>61</sup> For example, converting substance concentrations measured in WWTP influent samples into CRs requires estimating sewersheds populations, which has long been perceived as a significant source of uncertainty among other parameters for back-calculations (e.g., excretion rates).<sup>61</sup> Our analysis applied a normalization factor of  $8.8 \pm 1.3$  g  $NH_3N$ /day/person to account for population dynamics in the sewersheds despite the known limitations of  $NH_3N$  loading for population normalization in the absence of more refined proxies (e.g., mobile network signals<sup>62</sup> or concurrent census estimates<sup>63</sup>). Our normalization factor resembled those measured for sewersheds in New York City (i.e.,  $7.2 \pm 0.7$  g  $NH_3N$ /day/person)<sup>64</sup> as well as those reported by WBE studies in Switzerland (i.e.,  $8.1 \pm 0.4$  g  $NH_3N$ /day/person)<sup>65</sup> and China (i.e., 6.0–9.7 g  $NH_3N$ /day/person).<sup>66,67</sup> On average, the ratio of  $NH_3N$  equivalent sewersheds populations to the service populations of WWTPs was  $1.02 \pm 0.32$  (**Figure 3a**), and the PNMLs and CRs of substances estimated based on  $NH_3N$  equivalent populations were not statistically different from those calculated using service populations (paired *t* test two-tailed *p* = 0.2905–0.4841). Three additional normalization factors were derived from  $BOD_5$  (i.e.,  $67 \pm 10$  g  $BOD_5$ /day/person),  $CBOD_5$  (i.e.,  $60 \pm 9$  g  $CBOD_5$ /day/person), and  $TKN$  (i.e.,  $12.2 \pm 2.1$  g  $TKN$ /day/person), but the ratios of  $BOD_5$ ,  $CBOD_5$ , and  $TKN$  equivalent populations to service populations were more variable (i.e.,  $1.36 \pm 0.57$ ,  $1.22 \pm 0.53$ , and  $1.17 \pm 0.39$ , respectively) than those for  $NH_3N$  equivalent populations. Still, these normalization factors were comparable to those applied by previous WBE studies (e.g., 51–60 g of  $BOD_5$ /day/person,<sup>68,69</sup> 64–71 g of  $CBOD_5$ /day/person,<sup>64</sup> and 10.6–13.4 g of  $TKN$ /day/person;<sup>64,67,69</sup> **Table S30**). Given the consistent detection of caffeine, paraxanthine, and sucralose at elevated concentrations in samples, the applicability of these substances as population indicators was also evaluated for comparison to hydrochemical markers. With an average beverage caffeine intake of 165 mg/day/person<sup>70</sup> and an estimated average consumption from all sources of 224 mg/day/person<sup>41</sup> for the U.S. population, the ratios of caffeine equivalent populations to service populations were  $0.90 \pm 0.45$  and  $1.22 \pm 0.62$ , respectively, which exceeded those (i.e.,  $0.54 \pm 0.28$  to  $0.73 \pm 0.38$ ) calculated based on its metabolite paraxanthine (i.e., 139–188 mg/day/person assuming an 84% metabolic conversion from caffeine<sup>71</sup>). With an average consumption of sucralose at 18.5 to 26 mg/day/person,<sup>19,72</sup> the ratios of sucralose equivalent populations to service populations ranged from  $0.92 \pm 0.48$  to  $1.30 \pm 0.67$ . Taken together, while caffeine and sucralose loadings may serve as complementary metrics for population estimation, the analysis of  $NH_3N$  and other hydrochemical parameters is an integral component of WWTP operations for regulatory compliance and can readily be incorporated into the surveillance program.

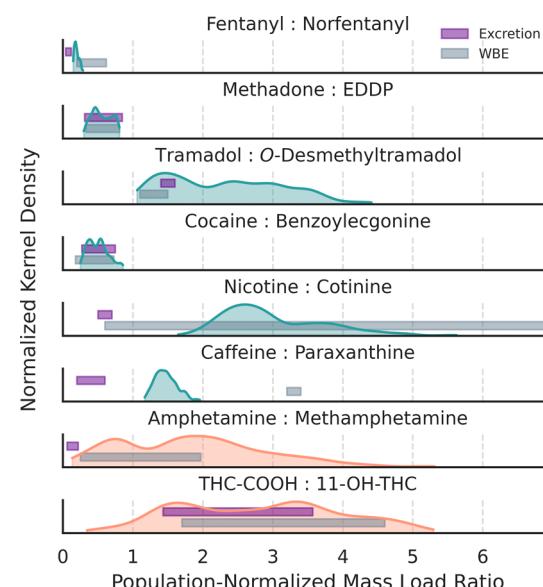
Overall, the median CRs of the 12 most frequently detected target substances ranged from 66 to  $1.20 \times 10^5$  mg/day/1000 people (Figure 3b) and followed patterns compiled from 15 WBE studies conducted in the U.S. between 2014 and 2024 (Table S31). For codeine, tramadol, nicotine, caffeine, and sucralose, the median CRs were within  $\pm 30\%$  of those measured for communities in other states (e.g., Arizona, Kentucky, Massachusetts, Nevada, and Washington). For fentanyl, methadone, diphenhydramine, cocaine, amphetamine, and methamphetamine, the CRs fell on the higher end of the ranges reported in the literature. Furthermore, the CR of fentanyl showed moderate to strong positive correlations with those of cocaine and methamphetamine (Spearman's  $\rho = 0.552$ – $0.830$ ;  $p < 0.0001$ ), which qualitatively agreed with the increasing co-consumption of fentanyl with these substances as revealed by nationwide urine drug testing.<sup>73</sup> Compared to data from the literature, the median CR of THC was an order of magnitude lower, which might be attributed to nonstandardized sample preservation and extraction techniques,<sup>74</sup> inconsistent excretion rates applied during back-calculations (e.g., ranging from 0.5% for THC-COOH via urinary excretion to 6.7% based on models that account for fecal excretion),<sup>75–78</sup> or uncertainty resulting from the lack of measurements for particulate-bound metabolites.<sup>40,79–81</sup>

Over the study period, the CRs of substances varied across sewersheds (Table S32) as one might expect from shifts in the sociodemographic status, health conditions, and behavioral factors of contributing populations. For instance, the CRs of fentanyl, codeine, cocaine, methamphetamine, and THC were positively correlated with the percentages of low-income households and individuals in their early adulthood who have lower educational attainment and are unemployed but negatively correlated with the percentages of high-income households and married individuals with employment and higher educational attainment (Figure S15). No further attempts were made to interpret the covariations of CRs, if any, with the health indicators or opioid burdens of sewersheds given the inconsistency in spatial resolutions among different data sets (e.g., sewersheds level versus ZIP code level or county level). Concerted efforts to integrate wastewater-derived data with epidemiological modeling at the sewersheds scale and other spatially comparable observations may overcome such limitations to identify meaningful relationships within the full-scale surveillance framework.

To investigate the effect of monitoring frequency on the relative bias in consumption estimates, the CRs of 12 target substances were calculated using reduced sample sets for three scenarios assuming weekly, biweekly, or monthly intervals and evaluated against baseline estimates derived from the complete sample sets. For eight out of the 12 substances (i.e., methadone, tramadol, diphenhydramine, cocaine, amphetamine, nicotine, caffeine, and sucralose), the relative bias in CRs derived from monthly monitoring (Figure 3c) and showed no significant difference compared to that derived from weekly and biweekly monitoring (Mann-Whitney  $U$  test  $p = 0.0539$ – $0.9698$ ), a pattern similar to that observed for NH<sub>3</sub>-N (Mann-Whitney  $U$  test  $p = 0.1859$ – $0.7337$ ). For fentanyl, codeine, methamphetamine, and THC, the relative bias in CRs derived from monthly monitoring was  $80 \pm 7\%$  higher than that derived from weekly monitoring (Mann-Whitney  $U$  test  $p = 0.0028$ – $0.0376$ ) but did not differ significantly from those derived from biweekly monitoring

(Mann-Whitney  $U$  test  $p = 0.2413$ – $0.7913$ ). Collectively, our simulations illustrated that for most of the 12 substances, the degree of deviation in CR estimates from baseline estimates was similar for weekly to monthly monitoring; however, the relative bias in CRs for fentanyl, codeine, methamphetamine, and THC exhibited a higher sensitivity to changes in monitoring frequency. Our approach did not seek to define an acceptable threshold of uncertainty or the minimum number of samples needed for representative CR estimates because quantifying the *true* loads of substances entering any WWTP requires high-frequency, longitudinal sampling.<sup>14,82,83</sup> To maximize the information gained relative to the resources allocated for the surveillance program, future assessments should further quantify the effects of additional factors (e.g., weekdays versus weekends or special events, combined versus separate sewers under dry and wet weather conditions, or WWTP inlet sampling versus sewer network node sampling) on consumption estimates.

**PNML Ratios for Substances.** Concurrent measurements of PNMLs for six parent–metabolite pairs of substances (i.e., fentanyl, methadone, tramadol, cocaine, nicotine, and caffeine) enabled the comparison of the P:M ratios measured in wastewater samples to those obtained from pharmacokinetic studies (Figure 4).<sup>32</sup> On average, the P:M ratios for



**Figure 4.** Comparison of population-normalized mass load (PNML) ratios for structurally related target substances. On each ridge plot, the indigo bar and the gray bar highlight the ranges of excretion ratios and the PNML ratios reported by previous WBE studies, respectively. Parent-to-metabolite (P:M) ratios were calculated for fentanyl (with its metabolite norfentanyl), methadone (with its metabolite EDDP), tramadol (with its metabolite *O*-desmethyltramadol), cocaine (with its metabolite benzoylecgonine), nicotine (with its metabolite cotinine), and caffeine (with its metabolite paraxanthine), respectively. Sewersheds-specific ratios are summarized in Table S33.

fentanyl:norfentanyl were relatively stable across sewersheds (Table S33) with a mean of  $0.20 \pm 0.04$ , which fell on the lower end of the range (i.e.,  $0.20$ – $0.62$ ) reported by a longitudinal WBE study conducted in the Midwestern U.S. communities<sup>14</sup> but was higher than the urinary excretion ratio (i.e.,  $0.08 \pm 0.04$ ).<sup>29</sup> The P:M ratios for methadone:EDDP converged at  $0.54 \pm 0.13$  and matched the value (i.e.,  $0.57 \pm$

0.24) averaged from 24 WBE studies conducted in Europe, Australia, China, and U.S.A. as well as the excretion ratio (i.e.,  $0.58 \pm 0.27$ ) derived from urinary measurements.<sup>84</sup> The P:M ratios for tramadol:*O*-desmethyltramadol ranged from  $1.3 \pm 0.2$  to  $2.9 \pm 0.7$ , which encompassed the urinary excretion ratio (i.e.,  $1.5 \pm 0.1$ )<sup>29</sup> and the ratio (i.e.,  $1.3 \pm 0.2$ ) documented by a WBE study conducted in Leuven, Belgium.<sup>85</sup> The P:M ratios for cocaine:benzoyllecgonine varied from  $0.41 \pm 0.08$  to  $0.64 \pm 0.10$ , consistent with those (i.e., 0.18–0.73) reported by WBE studies conducted in a Southwestern U.S. university campus<sup>86</sup> and communities in Kentucky<sup>16,87</sup> as well as the range of urinary excretion ratios (i.e., 0.27–0.75).<sup>88,89</sup> The P:M ratios for nicotine:cotinine (i.e.,  $2.4 \pm 0.4$  to  $4.2 \pm 0.7$ ) were less variable than the range (i.e., 0.6–9.2) observed by a WBE study conducted in communities with both separate and combined sewers in the Northeastern and Western U.S.A.<sup>18</sup> but far exceeded the urinary excretion ratio (i.e.,  $0.6 \pm 0.1$ ).<sup>66</sup> The P:M ratios for caffeine:paraxanthine (i.e.,  $1.5 \pm 0.2$ ) fell between the ratio (i.e.,  $3.3 \pm 0.1$ ) measured by a SARS-CoV-2 WBE study conducted in Missouri<sup>90</sup> and the urinary excretion ratio (i.e.,  $0.4 \pm 0.2$ ).<sup>91</sup> Overall, the P:M ratios for fentanyl, tramadol, nicotine, and caffeine provided an initial sign of nonconsumed parent compounds entering sewer systems through additional inputs; in contrast, the P:M ratios for methadone and cocaine suggested human excretion as the primary factor contributing to their prevalence at the sewershed level but in-sewer fate modeling would be required to assess the impacts of dynamic sewer conditions on P:M ratios.

Considering the possibility of amphetamine being excreted into sewer systems following methamphetamine consumption,<sup>92</sup> the PNML ratios for amphetamine and methamphetamine were also calculated even though the amphetamine:methamphetamine ratio did not necessarily differentiate between excretion and other pathways like the P:M ratios above. On average, the amphetamine:methamphetamine ratios for five of the sewersheds ranged from  $1.6 \pm 0.2$  to  $4.1 \pm 0.5$ , indicating a higher consumption of amphetamine than methamphetamine with reference to ratios (i.e., 0.20–3.4)<sup>93</sup> observed in 17 WBE studies across five continents. Conversely, the ratios for the remaining half of sewersheds were within the range of  $0.27 \pm 0.03$  to  $0.83 \pm 0.10$ , pointing to comparable consumption of amphetamine and methamphetamine.<sup>93</sup> Taking into account the solid–liquid partitioning of cannabis biomarkers,<sup>79,81</sup> the PNML ratios for THC-COOH and 11-OH-THC ranged from  $1.7 \pm 0.7$  to  $2.8 \pm 0.6$  and approached the values calculated for urine and feces (i.e.,  $2.5 \pm 1.1$ )<sup>81</sup> and unfiltered wastewater (i.e., 1.7–4.6).<sup>80</sup> To what extent this ratio may serve as a measure of the primary route by which THC enters sewer systems requires more clinical research to refine the excretion profiles of THC metabolites (e.g., the amount of THC-COOH excreted across a range of product types, consumption methods and frequencies, and co-consumption effects),<sup>92</sup> along with additional field and experimental investigation into their transformations and partitioning (e.g., the fraction of fecally excreted THC-COOH dissolved in wastewater) during in-sewer transit, sampling, and storage.<sup>80</sup>

**Implications and Limitations.** This work demonstrated the potential of online SPE-LC-HRMS for high-throughput quantification and nontargeted analysis in support of substance use assessment through a statewide wastewater surveillance network. Our target screening method covered a panel of

acidic, lipophilic, and basic hydrophilic compounds and is anticipated to meet the throughput requirements for weekly-to-monthly monitoring of influent samples from WWTPs participating in the NYS wastewater surveillance network. Going forward, targeted method development might consider combining mixed-bed multilayer online SPE<sup>94</sup> or less-selective enrichment techniques (e.g., vacuum-assisted evaporative concentration<sup>95</sup>) with alternative chromatographic modes (e.g., hydrophilic interaction<sup>96</sup> or mixed-mode<sup>97</sup> liquid chromatography) to broaden the analytical coverage. Our nontargeted analysis applied filtering for diagnostic fragment ions and characteristic neutral losses to prioritize the identification of additional psychoactive substances of concern, and a logical next step would be to explore the use of mass defect filtering for the selective profiling of specific substance classes (e.g., synthetic cannabinoids<sup>98</sup> and fentanyl analogs<sup>99</sup>) as well as the practicality of *in silico* mass spectral prediction models (e.g., domain-specific CFM-ID<sup>100</sup>) for high-confidence structural annotation of newly emerging or unknown psychoactive substances absent from mass spectral libraries. Complementary workflows, particularly those incorporating ion mobility separation, should also be implemented to remove mass spectral interferences and enhance structural elucidation when LC-HRMS alone cannot definitively resolve isobaric or isomeric substances in wastewater matrices.<sup>101</sup> Our consumption estimates relied on back-calculations that were highly sensitive to uncertainties associated with the in-sample stability, in-sewer transformation, partitioning behavior, and the excretion profiles of substances, as well as the choice of population biomarkers.<sup>61</sup> Furthermore, generating such estimates would likely be impractical for substances without known metabolic pathways and excretion rates, or those not excreted in detectable quantities in wastewater.<sup>101</sup> Given the variability in parent-to-metabolite ratios, the application of diagnostic tools like enantiomeric analysis<sup>102</sup> is warranted to differentiate the relative importance of human excretion versus other contributing sources to the presence of substances in sewer systems before performing consumption estimates. Overall, our study supports the operational feasibility of a statewide wastewater surveillance program for substance use assessment in New York and identifies several limitations and opportunities that could inform the implementation of similar initiatives in other regions of the U.S.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.est.4c01251>.

Characteristics of WWTPs and sewershed populations; online SPE-LC-HRMS method parameters for screening and quantification of substances; map of municipal WWTPs with an average design hydraulic flow of  $\geq 1$  MGD in NYS; stability factors, sorption data, and excretion rates of target substances from the literature; concentration ranges and detection frequencies of target substances; fragmentation information for substances identified at confidence levels 1, 2, and 3 through nontarget screening; population normalization factors; consumption rates of substances; and population-normalized mass load ratios for substances (PDF)

## AUTHOR INFORMATION

### Corresponding Author

Teng Zeng – Department of Civil and Environmental Engineering, Syracuse University, Syracuse, New York 13244, United States; [orcid.org/0000-0002-0374-9549](https://orcid.org/0000-0002-0374-9549); Phone: +1-315-443-1099; Email: [tezeng@syr.edu](mailto:tezeng@syr.edu)

### Authors

Emily J. Vogel – Department of Civil and Environmental Engineering, Syracuse University, Syracuse, New York 13244, United States

Milagros Neyra – Department of Public Health, Syracuse University, Syracuse, New York 13244, United States

David A. Larsen – Department of Public Health, Syracuse University, Syracuse, New York 13244, United States

Complete contact information is available at:

<https://pubs.acs.org/10.1021/acs.est.4c01251>

### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

We gratefully acknowledge the WWTP operators for their sample collection efforts. We thank Dr. Shiru Wang (Syracuse University) for her contribution to sample analysis and Dr. Qian Du (Quadrant Biosciences Inc.) for her assistance with sample handling. We also thank Chemaxon Ltd. for providing the academic research license for *MarvinSketch* (Version 23.10.0). We further thank the editor and five anonymous reviewers for their constructive feedback. This material is based upon work supported by the National Science Foundation under Grant No. 2018497.

## REFERENCES

- O'Keeffe, J. Wastewater-based epidemiology: Current uses and future opportunities as a public health surveillance tool. *Environ. Health Rev.* **2021**, *64* (3), 44–52.
- Diamond, M. B.; Keshaviah, A.; Bento, A. I.; Conroy-Ben, O.; Driver, E. M.; Ensor, K. B.; Halden, R. U.; Hopkins, L. P.; Kuhn, K. G.; Moe, C. L.; Rouchka, E. C.; Smith, T.; Stevenson, B. S.; Susswein, Z.; Vogel, J. R.; Wolfe, M. K.; Stadler, L. B.; Scarpino, S. V. Wastewater surveillance of pathogens can inform public health responses. *Nat. Med.* **2022**, *28* (10), 1992–1995.
- Centers for Disease Control and Prevention. *National Wastewater Surveillance System (NWSS)*. Centers for Disease Control and Prevention, Atlanta, GA, 2023. <https://www.cdc.gov/nwss/wastewater-surveillance.html> (accessed March 31, 2024).
- Neyra, M.; Hill, D. T.; Bennett, L. J.; Dunham, C. N.; Larsen, D. A. Establishing a statewide wastewater surveillance system in response to the COVID-19 Pandemic: A reliable model for continuous and emerging public health threats. *J. Public Health Manage.* **2023**, *29* (6), 854–862.
- New York State Department of Health. *COVID-19 Wastewater Surveillance*. New York State Department of Health, Albany, NY, 2024. <https://coronavirus.health.ny.gov/covid-19-wastewater-surveillance> (accessed March 31, 2024).
- Ahmed, F.; O'Brien, J. W.; Keshaviah, A.; Hall, W. D.; Bade, R.; Tscharke, B.; Matus, M.; Mueller, J. F.; Thomas, K. V.; Burgard, D. A. Wastewater-based monitoring could help guide responses to the USA opioid epidemic. *Nat. Water* **2023**, *1* (5), 401–404.
- Office of the New York State Comptroller. *Continuing Crisis: Drug Overdose Deaths in New York*. Office of the New York State Comptroller, Albany, NY, 2022. <https://www.osc.ny.gov/reports/continuing-crisis-drug-overdose-deaths-new-york> (accessed March 31, 2024).
- Centers for Disease Control and Prevention. *Understanding Drug Overdoses and Deaths*. Centers for Disease Control and Prevention, Atlanta, GA, 2023. <https://www.cdc.gov/drugoverdose/epidemic/index.html> (accessed March 31, 2024).
- European Monitoring Centre for Drugs and Drug Addiction. *Wastewater analysis and drugs - a European multi-city study*. European Monitoring Centre for Drugs and Drug Addiction, Lisbon, Portugal, 2023. [https://www.emcdda.europa.eu/publications/html/pods/waste-water-analysis\\_en](https://www.emcdda.europa.eu/publications/html/pods/waste-water-analysis_en) (accessed March 31, 2024).
- Australian Criminal Intelligence Commission. *National Wastewater Drug Monitoring Program*. Australian Criminal Intelligence Commission, Sydney, Australia, 2023. <https://www.acic.gov.au/publications/national-wastewater-drug-monitoring-program-reports/report-20-national-wastewater-drug-monitoring-program> (accessed March 31, 2024).
- Werschler, T.; Brennan, A. *Wastewater-based Estimates of Cannabis and Drug Use in Canada: Pilot test Detailed Results*. Statistics Canada, Ottawa, ON, Canada, 2019. <https://www150.statcan.gc.ca/n1/pub/11-621-m/11-621-m2019004-eng.htm> (accessed March 31, 2024).
- Castiglioni, S.; Salgueiro-González, N.; Bijlsma, L.; Celma, A.; Gracia-Lor, E.; Beldean-Galea, M. S.; Mackul'ak, T.; Emke, E.; Heath, E.; Kasprzyk-Hordern, B.; Petkovic, A.; Poretti, F.; Rangelov, J.; Santos, M. M.; Sremački, M.; Styszko, K.; Hernández, F.; Zuccato, E. New psychoactive substances in several European populations assessed by wastewater-based epidemiology. *Water Res.* **2021**, *195*, No. 116983.
- Bade, R.; Rousis, N.; Adhikari, S.; Baduel, C.; Bijlsma, L.; Bizani, E.; Boogaerts, T.; Burgard, D. A.; Castiglioni, S.; Chappell, A.; Covaci, A.; Driver, E. M.; Sodre, F. F.; Fatta-Kassinos, D.; Galani, A.; Gerber, C.; Gracia-Lor, E.; Gracia-Marín, E.; Halden, R. U.; Heath, E.; Hernandez, F.; Jaunay, E.; Lai, F. Y.; Lee, H.-J.; Laimou-Geraniou, M.; Oh, J.-E.; Olafsdottir, K.; Phung, K.; Castro, M. P.; Psichoudaki, M.; Shao, X.; Salgueiro-Gonzalez, N.; Feitosa, R. S.; Gomes, C. S.; Subedi, B.; Löve, A. S. C.; Thomaidis, N.; Tran, D.; van Nuijs, A.; Verovšek, T.; Wang, D.; White, J. M.; Yargeau, V.; Zuccato, E.; Mueller, J. F. Three years of wastewater surveillance for new psychoactive substances from 16 countries. *Water Res.: X* **2023**, *19*, No. 100179.
- Gushgari, A. J.; Venkatesan, A. K.; Chen, J.; Steele, J. C.; Halden, R. U. Long-term tracking of opioid consumption in two United States cities using wastewater-based epidemiology approach. *Water Res.* **2019**, *161*, 171–180.
- Luo, J.; Bello, D.; Pagsuyoin, S. Long-term wastewater-based surveillance and impacts of the COVID-19 pandemic on drug use trends in a U.S. Northeast rural town. *Sci. Total Environ.* **2023**, *877*, No. 162806.
- Croft, T. L.; Huffines, R. A.; Pathak, M.; Subedi, B. Prevalence of illicit and prescribed neuropsychiatric drugs in three communities in Kentucky using wastewater-based epidemiology and Monte Carlo simulation for the estimation of associated uncertainties. *J. Hazard. Mater.* **2020**, *384*, No. 121306.
- Halwatura, L. M.; McLellan, I. S.; Weglarski, D. L.; Ahmed, Z. U.; Ye, Y.; Bradley, I. M.; Aga, D. S. Complementing RNA detection with pharmaceutical monitoring for early warning of viral outbreaks through wastewater-based epidemiology. *Environ. Sci. Technol. Lett.* **2022**, *9* (6), 567–574.
- Chen, J.; Venkatesan, A. K.; Halden, R. U. Alcohol and nicotine consumption trends in three U.S. communities determined by wastewater-based epidemiology. *Sci. Total Environ.* **2019**, *656*, 174–183.
- Gerrity, D.; Crank, K.; Oh, E. C.; Quinones, O.; Trenholm, R. A.; Vanderford, B. J. Wastewater surveillance of high risk substances in Southern Nevada: Sucralose normalization to translate data for potential public health action. *Sci. Total Environ.* **2024**, *908*, No. 168369.
- O'Rourke, C. E.; Subedi, B. Occurrence and mass loading of synthetic opioids, synthetic cathinones, and synthetic cannabinoids in wastewater treatment plants in four U.S. communities. *Environ. Sci. Technol.* **2020**, *54* (11), 6661–6670.

(21) U.S. Census Bureau. *American Community Survey 2017–2021 5-Year Data*. U.S. Census Bureau, Washington, D.C., <https://www.census.gov/programs-surveys/acs> (accessed March 31, 2024).

(22) MassBank-consortium and its contributors. *MassBank/MassBank-data: Release version 2023, 09, 2023*, (accessed September 1, 2023) DOI: 10.5281/zenodo.8308157.

(23) MassBank of North America. *MassBank of North America (MoNA)*. 2023. <https://mona.fiehnlab.ucdavis.edu/> (accessed September 1, 2023).

(24) Centers for Disease Control and Prevention. *High-Resolution Mass Spectral Libraries for Opioid Analysis*. Centers for Disease Control and Prevention, Atlanta, GA, 2023. [https://www.cdc.gov/nceh/dls/erb\\_hrms\\_libraries.html](https://www.cdc.gov/nceh/dls/erb_hrms_libraries.html) (accessed September 1, 2023).

(25) Mardal, M.; Andreasen, M. F.; Mollerup, C. B.; Stockham, P.; Telving, R.; Thomaidis, N. S.; Diamanti, K. S.; Linnet, K.; Dalsgaard, P. W. HighResNPS.com: An online crowd-sourced HR-MS database for suspect and non-targeted screening of new psychoactive substances. *J. Anal. Toxicol.* 2019, 43 (7), S20–S27.

(26) Zapata, F.; Matey, J. M.; Montalvo, G.; García-Ruiz, C. Chemical classification of new psychoactive substances (NPS). *Microchem. J.* 2021, 163, No. 105877.

(27) Matey, J. M.; Zapata, F.; Menéndez-Quintanal, L. M.; Montalvo, G.; García-Ruiz, C. Identification of new psychoactive substances and their metabolites using non-targeted detection with high-resolution mass spectrometry through diagnosing fragment ions/neutral loss analysis. *Talanta* 2023, 265, No. 124816.

(28) Schymanski, E. L.; Jeon, J.; Gulde, R.; Fenner, K.; Ruff, M.; Singer, H. P.; Hollender, J. Identifying small molecules via high resolution mass spectrometry: Communicating confidence. *Environ. Sci. Technol.* 2014, 48 (4), 2097–2098.

(29) Baker, D. R.; Barron, L.; Kasprzyk-Hordern, B. Illicit and pharmaceutical drug consumption estimated via wastewater analysis. Part A: Chemical analysis and drug use estimates. *Sci. Total Environ.* 2014, 487, 629–641.

(30) Jones, H. E.; Hickman, M.; Kasprzyk-Hordern, B.; Welton, N. J.; Baker, D. R.; Ades, A. E. Illicit and pharmaceutical drug consumption estimated via wastewater analysis. Part B: Placing back-calculations in a formal statistical framework. *Sci. Total Environ.* 2014, 487, 642–650.

(31) Gehring, T.; Deineko, E.; Hobus, I.; Kolisch, G.; Lübben, M.; Wichern, M. Effect of sewage sampling frequency on determination of design parameters for municipal wastewater treatment plants. *Water Sci. Technol.* 2021, 84 (2), 284–292.

(32) Quireyns, M.; Boogaerts, T.; Van Wichelen, N.; Covaci, A.; van Nuijs, A. L. N. State-of-the-art analytical approaches and strategies to assess disposal of drugs for wastewater-based epidemiology. *WIREs Forensic Sci.* 2023, 5 (1), No. e1469.

(33) Chiaia, A. C.; Banta-Green, C.; Field, J. Eliminating solid phase extraction with large-volume injection LC/MS/MS: Analysis of illicit and legal drugs and human urine indicators in US wastewaters. *Environ. Sci. Technol.* 2008, 42 (23), 8841–8848.

(34) Postigo, C.; Lopez de Alda, M. J.; Barceló, D. Fully automated determination in the low nanogram per liter level of different classes of drugs of abuse in sewage water by on-line solid-phase extraction-liquid chromatography-electrospray-tandem mass spectrometry. *Anal. Chem.* 2008, 80 (9), 3123–3134.

(35) Heuett, N. V.; Ramirez, C. E.; Fernandez, A.; Gardinali, P. R. Analysis of drugs of abuse by online SPE-LC high resolution mass spectrometry: Communal assessment of consumption. *Sci. Total Environ.* 2015, 511, 319–330.

(36) López-García, E.; Mastroianni, N.; Postigo, C.; Barceló, D.; López de Alda, M. A fully automated approach for the analysis of 37 psychoactive substances in raw wastewater based on on-line solid phase extraction-liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A* 2018, 1576, 80–89.

(37) Wang, S.; Green, H. C.; Wilder, M. L.; Du, Q.; Kmush, B. L.; Collins, M. B.; Larsen, D. A.; Zeng, T. High-throughput wastewater analysis for substance use assessment in central New York during the COVID-19 pandemic. *Environ. Sci.: Processes Impacts* 2020, 22 (11), 2147–2161.

(38) Senta, I.; Rodríguez-Mozaz, S.; Corominas, L.; Covaci, A.; Petrovic, M. Applicability of an on-line solid-phase extraction liquid chromatography – tandem mass spectrometry for the wastewater-based assessment of human exposure to chemicals from personal care and household products. *Sci. Total Environ.* 2022, 845, No. 157309.

(39) Singh, V. M.; Browne, T.; Montgomery, J. The emerging role of toxic adulterants in street drugs in the US illicit opioid crisis. *Public Health Rep.* 2020, 135 (1), 6–10.

(40) Khan, U.; Nicell, J. A. Sewer epidemiology mass balances for assessing the illicit use of methamphetamine, amphetamine and tetrahydrocannabinol. *Sci. Total Environ.* 2012, 421–422, 144–162.

(41) Rodak, K.; Kokot, I.; Kratz, E. M. Caffeine as a factor influencing the functioning of the human body - Friend or foe? *Nutrients* 2021, 13 (9), 3088.

(42) Lenighan, Y. M.; Metreto, J.; Martyn, D. M.; Darch, M.; Gwenter, L. S.; Thornton, E.; Jack, M. M. Low- and no-calorie sweetener intakes from beverages - An up-to-date assessment in four regions: Brazil, Canada, Mexico and the United States. *Food Addit. Contam.: Part A* 2023, 40 (1), 26–42.

(43) Overdahl, K. E.; Sutton, R.; Sun, J.; DeStefano, N. J.; Getzinger, G. J.; Ferguson, P. L. Assessment of emerging polar organic pollutants linked to contaminant pathways within an urban estuary using non-targeted analysis. *Environ. Sci.: Processes Impacts* 2021, 23 (3), 429–445.

(44) Depriest, A. Z.; Puet, B. L.; Holt, A. C.; Roberts, A.; Cone, E. J. Metabolism and disposition of prescription opioids: A review. *Forensic Sci. Rev.* 2015, 27, 115–145.

(45) Bishop, N.; Jones-Lepp, T.; Margetts, M.; Sykes, J.; Alvarez, D.; Keil, D. E. Wastewater-based epidemiology pilot study to examine drug use in the Western United States. *Sci. Total Environ.* 2020, 745, No. 140697.

(46) Ahmed, F.; Tscharke, B.; O'Brien, J. W.; Thompson, J.; Zheng, Q.; Mueller, J. F.; Thomas, K. V. Quantification of selected analgesics and their metabolites in influent wastewater by liquid chromatography tandem mass spectrometry. *Talanta* 2021, 234, No. 122627.

(47) Galani, A.; Alygizakis, N.; Aalizadeh, R.; Kastritis, E.; Dimopoulos, M.-A.; Thomaidis, N. S. Patterns of pharmaceuticals use during the first wave of COVID-19 pandemic in Athens, Greece as revealed by wastewater-based epidemiology. *Sci. Total Environ.* 2021, 798, No. 149014.

(48) Nason, S. L.; Lin, E.; Eitzer, B.; Koelmel, J.; Peccia, J. Changes in sewage sludge chemical signatures during a COVID-19 community lockdown, Part 1: Traffic, drugs, mental health, and disinfectants. *Environ. Toxicol. Chem.* 2022, 41 (5), 1179–1192.

(49) Bade, R.; van Herwerden, D.; Rousis, N.; Adhikari, S.; Allen, D.; Baduel, C.; Bijlsma, L.; Boogaerts, T.; Burgard, D.; Chappell, A.; Driver, E. M.; Sodre, F. F.; Fatta-Kassinos, D.; Gracia-Lor, E.; Gracia-Marín, E.; Halden, R. U.; Heath, E.; Jaunay, E.; Krotulski, A.; Lai, F. Y.; Löve, A. S. C.; O'Brien, J. W.; Oh, J.-E.; Pasin, D.; Castro, M. P.; Psichoudaki, M.; Salgueiro-Gonzalez, N.; Gomes, C. S.; Subedi, B.; Thomas, K. V.; Thomaidis, N.; Wang, D.; Yargeau, V.; Samanipour, S.; Mueller, J. Workflow to facilitate the detection of new psychoactive substances and drugs of abuse in influent urban wastewater. *J. Hazard. Mater.* 2024, 469, No. 133955.

(50) Niebel, A.; Westendorf, L.; Krumbiegel, F.; Hartwig, S.; Parr, M. K.; Tsokos, M. Prevalence and concentrations of new designer stimulants, synthetic opioids, benzodiazepines, and hallucinogens in postmortem hair samples: A 13-year retrospective study. *Drug Test. Anal.* 2022, 14 (1), 110–121.

(51) Boogaerts, T.; Jurgelaitiene, L.; Dumitrescu, C.; Kasprzyk-Hordern, B.; Kannan, A.; Been, F.; Emke, E.; de Voogt, P.; Covaci, A.; van Nuijs, A. L. N. Application of wastewater-based epidemiology to investigate stimulant drug, alcohol and tobacco use in Lithuanian communities. *Sci. Total Environ.* 2021, 777, No. 145914.

(52) Walton, S.; Fogarty, M.; Papsun, D.; Lamb, M.; Logan, B.; Krotulski, A. N. N-Dimethylpentylone—an emerging NPS stimulant of

concern in the United States. *Ann. Toxicol. Anal.* **2022**, *34* (3), S67–S68.

(53) Celma, A.; Sancho, J. V.; Salgueiro-González, N.; Castiglioni, S.; Zuccato, E.; Hernández, F.; Bijlsma, L. Simultaneous determination of new psychoactive substances and illicit drugs in sewage: Potential of micro-liquid chromatography tandem mass spectrometry in wastewater-based epidemiology. *J. Chromatogr. A* **2019**, *1602*, 300–309.

(54) Rousis, N.; Bade, R.; Romero-Sánchez, I.; Mueller, J. F.; Thomaidis, N. S.; Thomas, K. V.; Gracia-Lor, E. Festivals following the easing of COVID-19 restrictions: Prevalence of new psychoactive substances and illicit drugs. *Environ. Int.* **2023**, *178*, No. 108075.

(55) Mestria, S.; Odoardi, S.; Federici, S.; Bilel, S.; Tirri, M.; Marti, M.; Strano Rossi, S. Metabolism study of N-methyl 2-aminoindane (NM2AI) and determination of metabolites in biological samples by LC–HRMS. *J. Anal. Toxicol.* **2021**, *45* (5), 475–483.

(56) Salgueiro-González, N.; Castiglioni, S.; Gracia-Lor, E.; Bijlsma, L.; Celma, A.; Bagnati, R.; Hernández, F.; Zuccato, E. Flexible high resolution-mass spectrometry approach for screening new psychoactive substances in urban wastewater. *Sci. Total Environ.* **2019**, *689*, 679–690.

(57) Alygizakis, N.; Galani, A.; Rousis, N. I.; Aalizadeh, R.; Dimopoulos, M.-A.; Thomaidis, N. S. Change in the chemical content of untreated wastewater of Athens, Greece under COVID-19 pandemic. *Sci. Total Environ.* **2021**, *799*, No. 149230.

(58) Lee, H.-J.; Oh, J.-E. Target and suspect screening of (new) psychoactive substances in South Korean wastewater by LC-HRMS. *Sci. Total Environ.* **2023**, *875*, No. 162613.

(59) Nieto, C. T.; Manchado, A.; Belda, L.; Diez, D.; Garrido, N. M. 2-Phenethylamines in medicinal chemistry: A review. *Molecules* **2023**, *28* (2), 855.

(60) Bijlsma, L.; Celma, A.; Castiglioni, S.; Salgueiro-González, N.; Bou-Iserte, L.; Baz-Lomba, J. A.; Reid, M. J.; Dias, M. J.; Lopes, A.; Matias, J.; Pastor-Alcañiz, L.; Radonić, J.; Turk Sekulic, M.; Shine, T.; van Nuijs, A. L. N.; Hernandez, F.; Zuccato, E. Monitoring psychoactive substance use at six European festivals through wastewater and pooled urine analysis. *Sci. Total Environ.* **2020**, *725*, No. 138376.

(61) Castiglioni, S.; Bijlsma, L.; Covaci, A.; Emke, E.; Hernández, F.; Reid, M.; Ort, C.; Thomas, K. V.; van Nuijs, A. L. N.; de Voogt, P.; Zuccato, E. Evaluation of uncertainties associated with the determination of community drug use through the measurement of sewage drug biomarkers. *Environ. Sci. Technol.* **2013**, *47* (3), 1452–1460.

(62) Thomas, K. V.; Amador, A.; Baz-Lomba, J. A.; Reid, M. Use of mobile device data to better estimate dynamic population size for wastewater-based epidemiology. *Environ. Sci. Technol.* **2017**, *51* (19), 11363–11370.

(63) O'Brien, J. W.; Thai, P. K.; Eaglesham, G.; Ort, C.; Scheidegger, A.; Carter, S.; Lai, F. Y.; Mueller, J. F. A model to estimate the population contributing to the wastewater using samples collected on census day. *Environ. Sci. Technol.* **2014**, *48* (1), 517–525.

(64) Hoar, C.; Li, Y.; Silverman, A. I. Assessment of commonly measured wastewater parameters to estimate sewershed populations for use in wastewater-based epidemiology: Insights into population dynamics in New York City during the COVID-19 pandemic. *ACS ES&T Water* **2022**, *2* (11), 2014–2024.

(65) Been, F.; Rossi, L.; Ort, C.; Rudaz, S.; Delémont, O.; Esseiva, P. Population normalization with ammonium in wastewater-based epidemiology: Application to illicit drug monitoring. *Environ. Sci. Technol.* **2014**, *48* (14), 8162–8169.

(66) Zheng, Q.-D.; Lin, J.-G.; Pei, W.; Guo, M.-X.; Wang, Z.; Wang, D.-G. Estimating nicotine consumption in eight cities using sewage epidemiology based on ammonia nitrogen equivalent population. *Sci. Total Environ.* **2017**, *590–591*, 226–232.

(67) Duan, L.; Zhang, Y.; Wang, B.; Yu, G.; Gao, J.; Cagnetta, G.; Huang, C.; Zhai, N. Wastewater surveillance for 168 pharmaceuticals and metabolites in a WWTP: Occurrence, temporal variations and feasibility of metabolic biomarkers for intake estimation. *Water Res.* **2022**, *216*, No. 118321.

(68) Baker, D. R.; Očenášková, V.; Kvicalova, M.; Kasprzyk-Hordern, B. Drugs of abuse in wastewater and suspended particulate matter — Further developments in sewage epidemiology. *Environ. Int.* **2012**, *48*, 28–38.

(69) Sim, W.; Park, S.; Ha, J.; Kim, D.; Oh, J.-E. Evaluation of population estimation methods for wastewater-based epidemiology in a metropolitan city. *Sci. Total Environ.* **2023**, *857*, No. 159154.

(70) Mitchell, D. C.; Knight, C. A.; Hockenberry, J.; Teplansky, R.; Hartman, T. J. Beverage caffeine intakes in the U.S. *Food Chem. Toxicol.* **2014**, *63*, 136–142.

(71) Nehlig, A. Interindividual differences in caffeine metabolism and factors driving caffeine consumption. *Pharmacol. Rev.* **2018**, *70* (2), 384–411.

(72) Subedi, B.; Kannan, K. Fate of artificial sweeteners in wastewater treatment plants in New York State, U.S.A. *Environ. Sci. Technol.* **2014**, *48* (23), 13668–13674.

(73) Krock, K.; Nickley, J.; Tran, K.; Ackerman, R. T. G.; Pesce, A. Correlation of fentanyl positive drug screens with other medications in patients from pain, rehabilitation and behavioral programs. *Ann. Clin. Lab. Sci.* **2020**, *50* (2), 260–265.

(74) Causanilles, A.; Baz-Lomba, J. A.; Burgard, D. A.; Emke, E.; González-Mariño, I.; Krizman-Matasic, I.; Li, A.; Löve, A. S. C.; McCall, A. K.; Montes, R.; van Nuijs, A. L. N.; Ort, C.; Quintana, J. B.; Senta, I.; Terzic, S.; Hernandez, F.; de Voogt, P.; Bijlsma, L. Improving wastewater-based epidemiology to estimate cannabis use: Focus on the initial aspects of the analytical procedure. *Anal. Chim. Acta* **2017**, *988*, 27–33.

(75) Postigo, C.; Lopez de Alda, M. J.; Barceló, D. Analysis of drugs of abuse and their human metabolites in water by LC-MS2: A non-intrusive tool for drug abuse estimation at the community level. *TrAC, Trends Anal. Chem.* **2008**, *27* (11), 1053–1069.

(76) Postigo, C.; de Alda, M. L.; Barceló, D. Evaluation of drugs of abuse use and trends in a prison through wastewater analysis. *Environ. Int.* **2011**, *37* (1), 49–55.

(77) Been, F.; Schneider, C.; Zobel, F.; Delémont, O.; Esseiva, P. Integrating environmental and self-report data to refine cannabis prevalence estimates in a major urban area of Switzerland. *Int. J. Drug Policy* **2016**, *36*, 33–42.

(78) Burgard, D. A.; Williams, J.; Westerman, D.; Rushing, R.; Carpenter, R.; LaRock, A.; Sadetsky, J.; Clarke, J.; Fryhle, H.; Pellman, M.; Banta-Green, C. J. Using wastewater-based analysis to monitor the effects of legalized retail sales on cannabis consumption in Washington State, USA. *Addiction* **2019**, *114* (9), 1582–1590.

(79) Senta, I.; Krizman, I.; Ahel, M.; Terzic, S. Integrated procedure for multiresidue analysis of dissolved and particulate drugs in municipal wastewater by liquid chromatography–tandem mass spectrometry. *Anal. Bioanal. Chem.* **2013**, *405* (10), 3255–3268.

(80) Campos-Mañas, M. C.; Van Wichelen, N.; Covaci, A.; van Nuijs, A. L. N.; Ort, C.; Béen, F.; Castiglioni, S.; Hernández, F.; Bijlsma, L. Analytical investigation of cannabis biomarkers in raw urban wastewater to refine consumption estimates. *Water Res.* **2022**, *223*, No. 119020.

(81) Pandopoulos, A. J.; Simpson, B. S.; White, J. M.; Bade, R.; Gerber, C. Partitioning of phytocannabinoids between faeces and water – Implications for wastewater-based epidemiology. *Sci. Total Environ.* **2022**, *805*, No. 150269.

(82) Ort, C.; Eppler, J. M.; Scheidegger, A.; Rieckermann, J.; Kinzig, M.; Sörgel, F. Challenges of surveying wastewater drug loads of small populations and generalizable aspects on optimizing monitoring design. *Addiction* **2014**, *109* (3), 472–481.

(83) Humphries, M. A.; Bruno, R.; Lai, F. Y.; Thai, P. K.; Holland, B. R.; O'Brien, J. W.; Ort, C.; Mueller, J. F. Evaluation of monitoring schemes for wastewater-based epidemiology to identify drug use trends using cocaine, methamphetamine, MDMA and methadone. *Environ. Sci. Technol.* **2016**, *50* (9), 4760–4768.

(84) Thai, P. K.; Lai, F. Y.; Bruno, R.; van Dyken, E.; Hall, W.; O'Brien, J.; Prichard, J.; Mueller, J. F. Refining the excretion factors of

methadone and codeine for wastewater analysis - Combining data from pharmacokinetic and wastewater studies. *Environ. Int.* **2016**, *94*, 307–314.

(85) Boogaerts, T.; Quireyns, M.; De Loof, H.; Bertels, X.; Van Wichelen, N.; Pussig, B.; Saevels, J.; Lahousse, L.; Bonmariage, P.; Hamelinck, W.; Aertgeerts, B.; Covaci, A.; van Nuijs, A. L. N. Do the lockdown-imposed changes in a wastewater treatment plant catchment's socio-demographics impact longitudinal temporal trends in psychoactive pharmaceutical use? *Sci. Total Environ.* **2023**, *876*, No. 162342.

(86) Gushgari, A. J.; Driver, E. M.; Steele, J. C.; Halden, R. U. Tracking narcotics consumption at a Southwestern U.S. university campus by wastewater-based epidemiology. *J. Hazard. Mater.* **2018**, *359*, 437–444.

(87) Skees, A. J.; Foppe, K. S.; Loganathan, B.; Subedi, B. Contamination profiles, mass loadings, and sewage epidemiology of neuropsychiatric and illicit drugs in wastewater and river waters from a community in the Midwestern United States. *Sci. Total Environ.* **2018**, *631*–632, 1457–1464.

(88) Van Nuijs, A. L. N.; Peccue, B.; Theunis, L.; Dubois, N.; Charlier, C.; Jorens, P. G.; Bervoets, L.; Blust, R.; Meulemans, H.; Neels, H.; Covaci, A. Can cocaine use be evaluated through analysis of wastewater? A nation-wide approach conducted in Belgium. *Addiction* **2009**, *104* (5), 734–741.

(89) Postigo, C.; López de Alda, M. J.; Barceló, D. Drugs of abuse and their metabolites in the Ebro River basin: Occurrence in sewage and surface water, sewage treatment plants removal efficiency, and collective drug usage estimation. *Environ. Int.* **2010**, *36* (1), 75–84.

(90) Hsu, S.-Y.; Bayati, M.; Li, C.; Hsieh, H.-Y.; Belenchia, A.; Klutts, J.; Zemmer, S. A.; Reynolds, M.; Semkiw, E.; Johnson, H.-Y.; Foley, T.; Wieberg, C. G.; Wenzel, J.; Johnson, M. C.; Lin, C.-H. Biomarkers selection for population normalization in SARS-CoV-2 wastewater-based epidemiology. *Water Res.* **2022**, *223*, No. 118985.

(91) Gracia-Lor, E.; Rousis, N. I.; Zuccato, E.; Bade, R.; Baz-Lomba, J. A.; Castrignanò, E.; Causanilles, A.; Hernández, F.; Kasprzyk-Hordern, B.; Kinyua, J.; McCall, A.-K.; van Nuijs, A. L. N.; Plósz, B. G.; Ramin, P.; Ryu, Y.; Santos, M. M.; Thomas, K.; de Voogt, P.; Yang, Z.; Castiglioni, S. Estimation of caffeine intake from analysis of caffeine metabolites in wastewater. *Sci. Total Environ.* **2017**, *609*, 1582–1588.

(92) Gracia-Lor, E.; Zuccato, E.; Castiglioni, S. Refining correction factors for back-calculation of illicit drug use. *Sci. Total Environ.* **2016**, *573*, 1648–1659.

(93) Gao, J.; Burgard, D. A.; Tscharke, B. J.; Lai, F. Y.; O'Brien, J. W.; Nguyen, H. D.; Zheng, Q.; Li, J.; Du, P.; Li, X.; Wang, D.; Castiglioni, S.; Cruz-Cruz, C.; Baz-Lomba, J. A.; Yargeau, V.; Emke, E.; Thomas, K. V.; Mueller, J. F.; Thai, P. K. Refining the estimation of amphetamine consumption by wastewater-based epidemiology. *Water Res.* **2022**, *225*, No. 119182.

(94) Huntscha, S.; Singer, H. P.; McArdell, C. S.; Frank, C. E.; Hollender, J. Multiresidue analysis of 88 polar organic micropollutants in ground, surface and wastewater using online mixed-bed multilayer solid-phase extraction coupled to high performance liquid chromatography–tandem mass spectrometry. *J. Chromatogr. A* **2012**, *1268*, 74–83.

(95) Mechelke, J.; Longrée, P.; Singer, H.; Hollender, J. Vacuum-assisted evaporative concentration combined with LC-HRMS/MS for ultra-trace-level screening of organic micropollutants in environmental water samples. *Anal. Bioanal. Chem.* **2019**, *411* (12), 2555–2567.

(96) Köke, N.; Zahn, D.; Knepper, T. P.; Frömel, T. Multi-layer solid-phase extraction and evaporation—enrichment methods for polar organic chemicals from aqueous matrices. *Anal. Bioanal. Chem.* **2018**, *410* (9), 2403–2411.

(97) Montes, R.; Aguirre, J.; Vidal, X.; Rodil, R.; Cela, R.; Quintana, J. B. Screening for polar chemicals in water by trifunctional mixed-mode liquid chromatography–high resolution mass spectrometry. *Environ. Sci. Technol.* **2017**, *51* (11), 6250–6259.

(98) Grabenauer, M.; Krol, W. L.; Wiley, J. L.; Thomas, B. F. Analysis of synthetic cannabinoids using high-resolution mass spectrometry and mass defect filtering: Implications for nontargeted screening of designer drugs. *Anal. Chem.* **2012**, *84* (13), 5574–5581.

(99) Swanson, D.; Stickle, D.; Evans-Nguyen, T. Analysis of unknown fentanyl analogs using high resolution mass spectrometry with mass defect filtering. *Int. J. Mass Spectrom.* **2023**, *485*, No. 116992.

(100) Wang, F.; Pasin, D.; Skinnider, M. A.; Liigand, J.; Kleis, J.-N.; Brown, D.; Oler, E.; Sajed, T.; Gautam, V.; Harrison, S.; Greiner, R.; Foster, L. J.; Dalsgaard, P. W.; Wishart, D. S. Deep learning-enabled MS/MS spectrum prediction facilitates automated identification of novel psychoactive substances. *Anal. Chem.* **2023**, *95* (50), 18326–18334.

(101) Bijlsma, L.; Bade, R.; Been, F.; Celma, A.; Castiglioni, S. Perspectives and challenges associated with the determination of new psychoactive substances in urine and wastewater – A tutorial. *Anal. Chim. Acta* **2021**, *1145*, 132–147.

(102) Castrignanò, E.; Yang, Z.; Bade, R.; Baz-Lomba, J. A.; Castiglioni, S.; Causanilles, A.; Covaci, A.; Gracia-Lor, E.; Hernandez, F.; Kinyua, J.; McCall, A.-K.; van Nuijs, A. L. N.; Ort, C.; Plósz, B. G.; Ramin, P.; Rousis, N. I.; Ryu, Y.; Thomas, K. V.; de Voogt, P.; Zuccato, E.; Kasprzyk-Hordern, B. Enantiomeric profiling of chiral illicit drugs in a pan-European study. *Water Res.* **2018**, *130*, 151–160.

## ■ NOTE ADDED AFTER ASAP PUBLICATION

This paper was published ASAP on May 1, 2024, with an incorrect version of the Supporting Information. The corrected version was reposted on May 2, 2024.