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Review

Top ten hazards to avoid when modeling species distributions: a didactic guide of assumptions, problems, and recommendations

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Species distribution models, also known as ecological niche models or habitat suitability models, have become commonplace for addressing fundamental and applied biodiversity questions. Although the field has progressed rapidly regarding theory and implementation, key assumptions are still frequently violated and recommendations inadvertently overlooked. This leads to poor models being published and used in real-world applications. In a structured, didactic treatment, we summarize what in our view constitute the ten most problematic issues, or hazards, negatively affecting implementation of correlative approaches to species distribution modeling (specifically those that model suitability by comparing the environments of a species' occurrence records with those of a background or pseudoabsence sample). For each hazard, we state relevant assumptions, detail problems that arise when violating them, and convey straightforward existing recommendations. We also discuss five major outstanding questions of active current research. We hope this contribution will promote more rigorous implementation of these valuable models and stimulate further advancements.

Keywords: assumptions, ENM, guidelines, principles, SDM, theory

Modeling species distributions

Approaches termed species distribution models, ecological niche models, or habitat suitability models (hereafter, SDMs) encapsulate a set of theory and tools valuable in basic and applied biogeography, ecology, and evolutionary biology (Fig. 1) (Franklin 2010, Peterson et al. 2011, Guisan et al. 2017). Substantial literature exists regarding relationships between species ecological niches and geographic distributions, as well as correlative and mechanistic approaches to estimating them (Soberón 2007, Enriquez-Urzelai et al. 2019, Kearney and Porter 2020, Franklin 2023). Studies using these techniques generally aim to estimate environmental suitability for a species, mapping it onto geography to characterize spatial patterns (Fig. 1). These models



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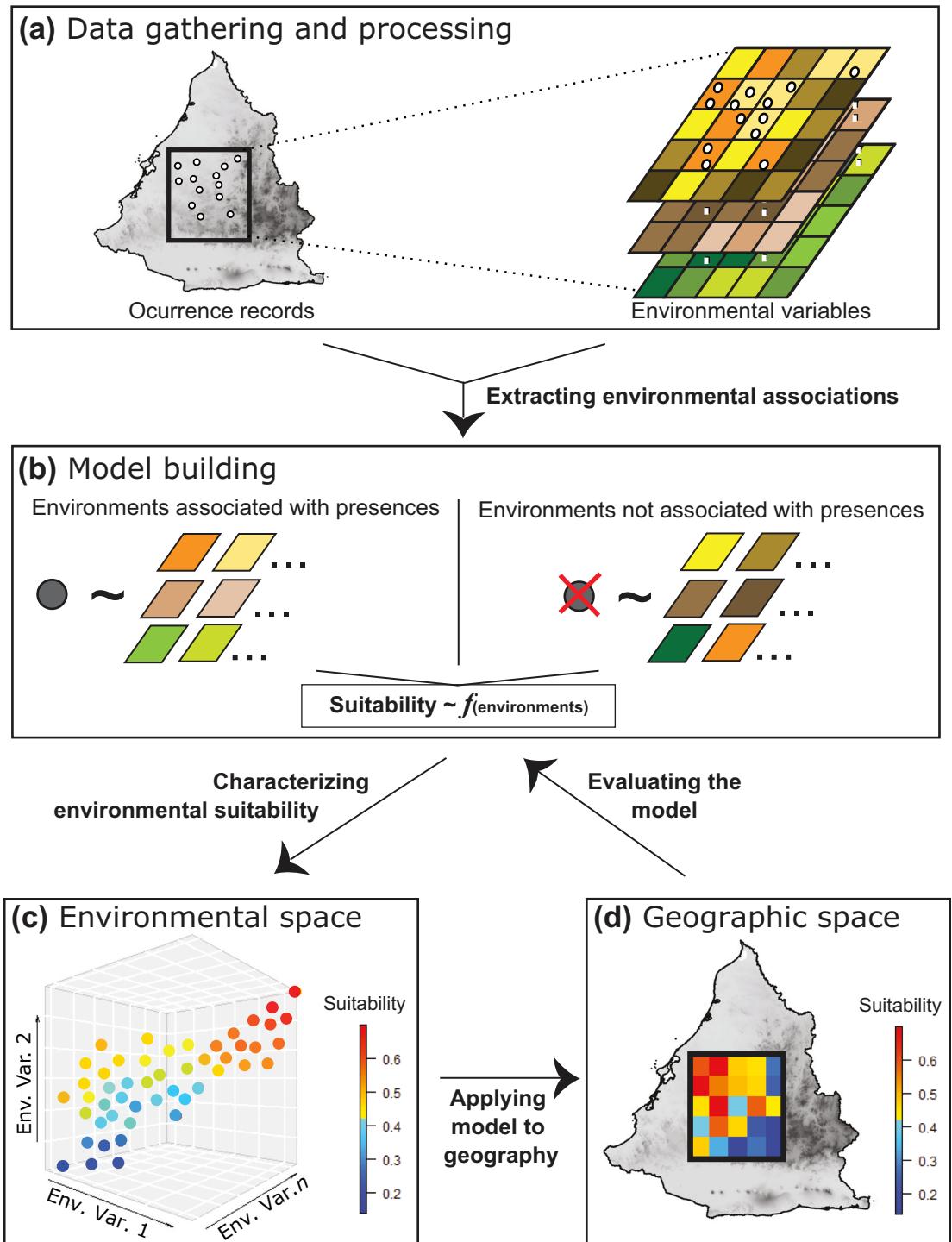


Figure 1. Schematic overview of how a species distribution model is built. (a) Records of a species' presence (white circles) within a study region (rectangle) are gathered, along with relevant environmental variables. Environmental variables typically consist of gridded GIS layers describing abiotic conditions of the study region (e.g. temperature, soil type; colors for each grid cell depict different values of the variable). (b) These data are used as input for any of the several available algorithms, which characterize suitability as a function of environmental variables. The model distinguishes the environmental conditions more frequently associated with presence of the species versus those from a comparison dataset (either absences or more commonly background or pseudoabsence information across the study region). (c) The model characterizes environmental suitability for the species in environmental space (warmer colors indicating increasingly suitable conditions). (d) Typically, the model is then applied to geographic space, indicating spatial patterns of suitability across the study region. Finally, the model is evaluated quantitatively, typically by assessing how well it predicts an evaluation dataset (arrow back to (b)).

are frequently applied to issues of importance to society, from agriculture and public health to biodiversity management and conservation, including the cross-cutting effects of climate change (Guisan et al. 2014, Willis et al. 2015, Johnson et al. 2019). However, key principles and procedures frequently remain underappreciated or misapplied (Morales et al. 2017, Araújo et al. 2019, Andrade et al. 2020). This can lead to unreliable models and erroneous interpretations being published or used as inputs for additional downstream analyses, which increasingly include valuable studies that address applied real-world problems (Brown et al. 2016, Fordham et al. 2018, Briscoe et al. 2019, Reid et al. 2019, Zurell et al. 2020a, Tuia et al. 2022).

This situation stems from several interrelated factors. First, SDMs have seen explosive use and development since around the year 2000, creating an enormous literature (Araújo et al. 2019, Feng et al. 2019a). Second, semi-automated software makes analyses quick and easy to implement, which can lure users into building models without considering the underlying principles carefully (Joppa et al. 2013, Merow et al. 2014, Escobar and Craft 2016, Sillero and Barbosa 2021). Especially in the era of 'big data,' this can be exacerbated in studies that model hundreds of species via workflows that employ automated code-based pipelines, which is often needed to achieve broad assessments of biodiversity (Brown et al. 2015, Gomes et al. 2019, Merow et al. 2022). Finally, SDMs commonly form part of interdisciplinary studies (e.g. combining demographic, molecular, or epidemiological approaches; Perktaş et al. 2017, Guevara et al. 2018a, Bonfim et al. 2019, González-Serna et al. 2019), where the research team may not include an SDM expert. As a consequence of these factors, researchers often struggle to follow recommendations from the massive, disparate literature that has accumulated for individual aspects of modeling. Indeed, a quantitative analysis of randomly selected SDM studies with an applied biodiversity focus published between 1995 and 2015 found that 46% of them were deficient regarding at least one key issue (Araújo et al. 2019). Perhaps most disturbingly, the evaluation of model assumptions saw a decrease in quality over that time period, suggesting that enormous growth in the use of these techniques unfortunately had been accompanied by lessened consideration of the field's conceptual foundations.

To help rectify this situation, here we highlight what in our view constitute the ten most problematic hazards negatively affecting SDM implementation – each with existing solutions. To varying degrees, these issues have been addressed elsewhere, although often independently (Araújo and Peterson 2012, Jiménez-Valverde et al. 2013, Cooper and Soberón 2018, Yates et al. 2018, Cobos et al. 2019a, Qiao et al. 2019, Warren et al. 2020). Fortunately, some synthetic books (Franklin 2010, Peterson et al. 2011, Guisan et al. 2017), shorter reviews (Elith et al. 2011, Anderson 2012, Merow et al. 2014, Jarnevich et al. 2015, Beery et al. 2021, Sillero et al. 2021, Franklin 2023), and proposed standards for modeling and reporting exist (Araújo et al. 2019, Feng et al. 2019a, Sofaer et al. 2019, Zurell et al. 2020b, Fitzpatrick et al.

2021). Nevertheless, the book-level syntheses are long and include mathematical and statistical formalizations, making it difficult for some readers to understand key information and deterring others from attempting. On the other hand, the shorter reviews each only consider a few key issues and sometimes are highly technical, requiring several papers to cover key topics and again limiting the audience. Finally, although comprehensive in breadth, the proposed standards do not go into great depth of explanation, emphasizing key principles and recommendations more than guiding the reader's understanding regarding problems that can occur if they are not followed. Complementary to these resources, here we present a concise, structured treatment of ten hazardous, frequently misunderstood issues (and ways that they interact), avoiding mathematical formulations to produce a didactic guide for readers with a wide range of exposure to these techniques. Additionally, we include a glossary of some important terms (Table 1), which appear in *italic* at first mention. We hope that this contribution will help a broad set of researchers reach a deeper understanding of these issues and prove useful to them as authors, reviewers, and editors. We also provide ample citations, constituting a rich resource for readers who want to delve deeper into the literature.

In this guide, we consider the most common implementation of correlative SDMs, where absence data do not exist and the researcher aims to use records of the species' presence (presence-only data) to model the environmental conditions (and geographic areas) suitable for it. Specifically, we focus on models that compare environmental conditions for records of the species' presence versus those of the background (the study region or a sample of it) or pseudoabsences (places within the study region that lack records of the species; Anderson 2012, Fig. 1). All the issues we cover hold true regardless of modeling algorithm; additionally, many are also applicable to implementations that exclusively use presence data (Booth et al. 2014), incorporate information regarding absences (Guisan et al. 2017), or model more than one species simultaneously (Pollock et al. 2014, Poggianto et al. 2021). Also, the issues we present (or an equivalent analog) are relevant to integrated modeling approaches that combine distinct data types (e.g. presence-only, presence–absence and abundance information; Fletcher et al. 2019, Miller et al. 2019, Isaac et al. 2020, Kays et al. 2022a).

We order the ten key hazards according to the step in which they arise (data gathering and processing, model building, model evaluation, and model interpretation; i.e. not according to importance or frequency in the literature; Table 2, Fig. 2). For each hazard, we state relevant assumptions, detail problems that arise when violating them, and summarize existing recommendations. In most entries, we explain one or more key interactions with other specific hazards, using the syntax 'interaction with Hazard X'; in contrast, we denote simple references to another hazard by the wording 'Hazard Y'. While we strive for general explanations, we also provide some examples for particular algorithms (especially the commonly used maximum entropy approach MaxEnt; Phillips et al. 2006). Given the increasing number of studies modeling many species

Table 1. Glossary of selected key terms.

Term	Definition
commission (Peterson et al. 2011) (= false positive rate; = 1 – sensitivity ; see also omission)	Error that represents the failure of a model to predict an absence of the species once a binary prediction of suitability is obtained (by applying a threshold above which a grid cell is considered suitable or predicted presence). In studies lacking true absence data, estimates of commission error are typically inflated.
evaluation	Process by which the model resulting from training is assessed. Ideally, the evaluation would be performed using fully independent testing data. However, because such information seldom exists, in most instances evaluation is conducted via a semi-independent validation dataset based on a partition not used in training (Hastie et al. 2009, Wenger and Olden 2012, Zurell et al. 2020b). We use evaluation as a blanket term that applies to either validation (via a partition of the same dataset used for training) or testing (based on independent data).
noise assumptions	The premise that within the study region, factors related to dispersal, establishment, and persistence of populations (e.g. dispersal barriers or small patch size); biotic interactions; and human modifications of the environment do not cause the species to occupy an environmentally biased subset of the areas abiotically suitable for it. Under these assumptions, even if the species does not inhabit all suitable areas in the study region (i.e. is not at spatial equilibrium), it occurs in environmental equilibrium, with such factors only adding statistical noise (Anderson 2013).
non-analog	Environments in which the conditions are different from those in which the model was trained. It usually refers to environmental values beyond the minimum and maximum present in the training data but can also refer to novel environmental combinations (Mesgaran et al. 2014, Guevara et al. 2018b).
omission (Peterson et al. 2011) (= false negative rate; = 1 – sensitivity ; see also commission)	Error that represents the failure of a model to predict a presence of the species once a binary prediction of suitability is obtained (by applying a threshold above which a grid cell is considered suitable or predicted presence).
overfit	A model that is fit too tightly to the training data. This reduces the generality and utility of the model, which will have high performance in predicting the sample with which it was trained but perform poorly on other datasets (e.g. those withheld for validation, deriving from additional sampling, or corresponding to transfer of the model to other areas or time periods; see transfer).
sampling bias	When known occurrences of the species reflect vagaries associated with patterns of biological sampling (e.g. greater efforts near roads or biodiverse regions). This sampling bias in geographic space usually translates into a bias in environmental space.
sensitivity	The ability of a model to predict a presence of the species (see omission).
specificity	The ability of a model to predict an absence of the species (see commission).
study region	The geographic region in which a species distribution model is trained, and from where the sample of presences and any environmental background (or pseudoabsence) information is obtained.
training (= fitting; referred to as calibration in some literature (Peterson et al. 2011))	The process by which a particular model is built (Hastie et al. 2009, Zurell et al. 2020b). Often this includes automated iterations with subsets of the data, when plausible environmental associations are explored and evaluated by the algorithm, aiming to achieve an optimal solution (e.g. maximizing likelihood in regression, or minimizing relative entropy in MaxEnt; Merow et al. 2013, Phillips et al. 2017).
transfer	Use of a model in a place or time period different from that in which it was trained. If non-analog environments are encountered (different from those in which the model was trained), extrapolation of the model is required (e.g. following response curves beyond the point of truncation; Guevara et al. 2018b).
tuning	Approximating optimal parameterization and model complexity for a given dataset. This is achieved by building multiple models differing in the underlying parameters (or settings that constrain their estimation from the data) and choosing the best one/s according to specific evaluation criteria (Elith et al. 2011, Merow et al. 2013).

individually via automated pipelines (Brown et al. 2015, Gomes et al. 2019), for each hazard we note the feasibility and challenges of implementing the recommendations in such approaches. In addition to the ten key hazards, we also discuss a few other important topics that researchers should be aware of, yet for which solutions and recommendations remain challenging and require additional research.

Top ten hazards

Data gathering and processing

1. Overlooking errors in occurrence datasets

Online biodiversity portals (e.g. GBIF; eBird; Robertson et al. 2014) often aggregate information from many data

providers, affording invaluable access to species occurrence data. However, such data sources are often misused by researchers who employ records of insufficient quality (Newbold 2010, Maldonado et al. 2015).

Assumptions. Input data are free of substantial errors regarding taxonomic identification and georeference (e.g. latitude and longitude coordinates), with spatial and temporal uncertainty smaller than the resolution of the environmental predictors (Lozier et al. 2009, Aubry et al. 2017, Araújo et al. 2019).

Problem. Despite increasing attention to data quality and uncertainty in data portals (Anderson et al. 2020), errors of taxonomic identification and georeferencing remain

common, and high spatial uncertainty may always persist for many older records (Newbold 2010, Maldonado et al. 2015, Serra-Díaz et al. 2017). Depending on the type and magnitude of these errors, spatial resolution of the environmental predictors, and heterogeneity of the landscape, substantial distortions and inflations of inferred environmental associations and suitable areas may occur (Romero et al. 2014, Costa et al. 2015, Aubry et al. 2017, Gábor et al. 2020a). Unfortunately, species identifications may be incorrect, and aggregators seldom capture updates from recent taxonomic publications (Anderson et al. 2020). Additionally, the vast majority of records lack information regarding the uncertainty of identification and georeference (and unit conversions to decimal degrees often falsely implies high precision and accuracy).

Recommendations. Models should be built using occurrence data with correct taxonomic identifications and georeferences whose uncertainties do not affect the results greatly (e.g. only increase noise minimally and do not bias the model). Ideally, occurrence records would have been vetted by a specialist knowledgeable regarding the current taxonomy of the group at hand (e.g. by examining morphological, photographic, or audio vouchers to check the identification) and a person familiar with the region (e.g. by consulting maps and field notes to help determine an accurate and precise georeference; Maldonado et al. 2015, Soley-Guardia et al. 2019). For some databases, partial vetting has already been accomplished, with 'research grade' designations allowing the efficient use of only such data (e.g. by iNaturalist; inaturalist.org or Arctos; arctosdb.org; Kays et al. 2022b, Gaier and Resasco 2023). Similarly, some consortia of data providers have determined coordinate uncertainty, which percolates up to aggregators, so that researchers can use only those accurate enough for the aims of a given study (e.g. MaNIS; Stein and Wieczorek 2004). Focused endeavors to standardize and clean data for given taxonomic groups and regions also provide much better information than otherwise available (e.g. BIEN and BioModelos; Maitner et al. 2018, Velásquez-Tibatá et al. 2019). When these situations do not exist, automated methods for data cleaning can catch many but certainly not all errors (e.g. identifying unit conversion errors; spatial and environmental outliers; García-Roselló et al. 2014, Naimi et al. 2014, Robertson et al. 2016, Zizka et al. 2019). In cases where the uncertainties of records are not known, researchers can conduct qualitative or quantitative sensitivity analyses and discuss the likely effects on model output (Gábor et al. 2020a). Additionally, expert knowledge remains critical in interpreting the outputs of both the data-cleaning algorithms and resulting models (e.g. 'outliers' may represent rarely sampled sites and provide valuable information; Jiménez-Valverde et al. 2011, Soley-Guardia et al. 2014). All of these investments in data cleaning improve the quality of models, perhaps especially those generated via automated analyses for projects modeling many species (Velásquez-Tibatá et al. 2019).

2. Disregarding biases inherent to biological sampling

Aggregated biodiversity data combine records that generally were collected by opportunistic rather than stratified or

random sampling (Beck et al. 2014, Isaac and Pocock 2015, Daru et al. 2018).

Assumptions. Sampling effort has been homogeneous across the *study region*, so that no bias exists in the conditions inferred to represent the species' environmental associations (Castellanos et al. 2019, Vollering et al. 2019, Taylor et al. 2020).

Problem. Opportunistic sampling tends to be geographically biased towards accessible areas or habitats of particular interest (Daru et al. 2018, Tarli et al. 2018). Such geographic bias typically translates into environmental bias as well, with some conditions inhabited by the species being artificially overrepresented (Yackulic et al. 2013, Monserrat et al. 2019). If not corrected for, certain environmental combinations will erroneously be identified as indicating higher suitability for the species, yielding a biased model (Syfert et al. 2013, Ranc et al. 2017, Vollering et al. 2019). Importantly, inappropriate (yet commonly applied) *evaluation* approaches fail to detect this bias, which pervades both *training* and validation datasets (interaction with Hazard 8; Fig. 2).

Recommendations. Completely eliminating the false signal resulting from *sampling bias* remains challenging; however, several approaches may reduce its effects considerably (Franklin 2023). One theoretically sound approach corrects for sampling bias by contrasting the focal species' occurrences against a sample of the environment characterized by the same bias, via occurrence records of other species detected with the same techniques (i.e. 'target-group background'; Anderson 2003, Phillips et al. 2009, Merow et al. 2016). This approach can be modified by concentrating background sampling around occurrences of the target-group species in proportion to their density ('weighted locality approach'; Anderson 2003; sometimes called 'background thickening'; Ranc et al. 2017, Støa et al. 2018, Vollering et al. 2019, Barber et al. 2022). Alternatively, a practical approach that can improve performance is to apply a spatial or environmental filter that reduces the number of clustered occurrences ('thinning'), likely reducing the effects of sampling bias (Boria et al. 2014, Varela et al. 2014). Nevertheless, choosing an appropriate filtering distance that retains the niche signal while removing the bias remains challenging (Fourcade et al. 2014, Aiello-Lammens et al. 2015, Castellanos et al. 2019, Gábor et al. 2020b) but can be tuned empirically (Soley-Guardia et al. 2019). For projects implementing automated modeling of many species, either using target-group background correction (with experts defining suitable groupings based on relevant sampling techniques) or filtering approaches should be feasible.

3. Using spatially or temporally inconsistent proxy environmental variables

Predictor variables used in SDMs may affect species distributions directly (proximal/driving factors) or indirectly (via correlation with the former; Austin 2002, Anderson 2013, Title and Bemmels 2018). Whereas indirect proxy variables

Table 2. Overview of the ten most problematic hazards in species distribution modeling covered in this paper. The columns indicate: the corresponding step of modeling; the name of the hazard; the nature of the hazard; its relevance for predictions (whether it affects the model in the training region, under transferal, or both); interactions with other hazards (an asterisk indicates those where the interaction is explained in the main text only in the entry for the other hazard involved); and three selected references. All hazards correspond to important concepts in the field, but for their nature we note 'conceptual' for those with especially close ties to ecological theory. While Hazards 3 and 9 apply primarily to model transfer, they may also affect predictions within the time and space in which the model was trained (see main text).

Step of modeling	Hazard	Nature	Relevance (non-transfer and transfer)	Interaction with other hazards	References
Data gathering and processing	1. Overlooking errors in occurrence datasets	data quality	both	–	Maldonado et al. 2015, Velásquez-Tibatá et al. 2019, Anderson et al. 2020
	2. Disregarding biases inherent to biological sampling	data quality	both	4*, 6*, 8	Phillips et al. 2009, Boria et al. 2014, Monserrat et al. 2019
	3. Using spatially or temporally inconsistent environmental variables	conceptual	transfer (primarily)	6	Austin 2002, Dormann et al. 2013, Feng et al. 2019b
Model building	4. Relying on default settings	analytical	both	2, 5*, 8	Merow et al. 2014, Hallgren et al. 2019, Valavi et al. 2022
	5. Reducing predictor variables irrespective of their information content	analytical–conceptual	both	4	Breiman 2001, Dormann et al. 2013, Farrell et al. 2019
Model evaluation	6. Using an overly large study region	analytical–conceptual	both	2, 3*, 7, 9, 10*	Anderson and Raza 2010, Barve et al. 2011, Cooper and Soberón 2018
	7. Misinterpreting metrics of model performance	analytical	both	6	Araújo and Peterson 2012, Leroy et al. 2018, Bohl et al. 2019
Model interpretation	8. Evaluating models with random splits of the data	analytical	both	2, 4, 9	Veloz 2009, Radosavljevic and Anderson 2014, Roberts et al. 2017
	9. Misunderstanding or overlooking the effects of extrapolation	analytical–conceptual	transfer (primarily)	6*, 8*	Fitzpatrick and Hargrove 2009, Elith et al. 2010, Owens et al. 2013
	10. Comparing model outputs across different species or places	analytical–conceptual	both	6	Fithian and Hastie 2013, Phillips and Elith 2013, Guillera-Arroita et al. 2015

can provide strong predictive ability within a limited study region, their correlation with driving factors usually varies in other places or time periods (Dormann et al. 2013).

Assumptions. The variables (at a given spatial and temporal resolution; Hazard 1) incorporated into a model affect the species' distribution directly or are highly and consistently correlated with those that do (Anderson 2013, Lira-Noriega et al. 2013).

Problem. If this is not the case, predictions can be inaccurate – especially when occurrence datasets are small, study regions are overly large (interaction with Hazard 6; Fig. 2; because correlations between proxy and driving variables are less likely to be consistent across space), or models undergo *transfer* across space or time for the needs of the study. Elevation constitutes a classic example of a variable that frequently violates this assumption because its correlation with the presumed driving variable (temperature), differs across latitudes and over time periods (Austin 2002, Elith and Leathwick 2009). However, commonly used climatic variables also can suffer changes in correlation with

missing driving factors (e.g. soil type, groundwater availability; Dormann et al. 2013, Mesgarian et al. 2014, Soley-Guardia et al. 2014, Feng et al. 2019b).

Recommendations. Based on existing natural history or physiological information, researchers should strive to select environmental variables known or suspected to have a direct effect on suitability for the species, at relevant spatial and temporal resolutions (Hazard 1; Mod et al. 2016, Petitpierre et al. 2017, Reside et al. 2019, Morente-López et al. 2022). Fortunately, advances continue regarding variables with various spatial resolutions and potentially tighter links with species physiologies (e.g. 'WorldClim2'; Fick and Hijmans 2017; 'ENVIREM'; Title and Bemmels 2018; 'NicheMapR'; Kearney and Porter 2017, 2020, Enriquez-Urzelai et al. 2019). For projects modeling many species using automated pipelines, researchers can select environmental variables in bunches (e.g. for groups of species with similar natural histories) rather than using the same set for all of them, although that still may not achieve models as good as those possible with species-specific decisions.

Model building

4. Relying on default settings

SDMs characterize the relationships between occurrence data and environmental variables based on allowed mathematical options and algorithmic settings, including those that influence model complexity (e.g. feature classes and regularization multiplier in MaxEnt; Merow et al. 2013; lasso in general linear models (GLMs) and general additive models (GAMs); Dicko et al. 2014; tree depth in random forests; Valavi et al. 2021). Due to the availability of modeling software with defaults for key settings that affect model complexity, it is common for users to overlook the need to consider a variety of values for them, to approximate the optimal ones (a process often called *tuning* or *smoothing*; Morales et al. 2017, Feng et al. 2019a, Hallgren et al. 2019).

Assumptions. Models are fairly insensitive to settings that affect their complexity; or default settings result in an acceptable parameterization regardless of biases and nuances of each input dataset.

Problem. Estimates indeed can be highly sensitive to the algorithmic settings that control model building and complexity (Hallgren et al. 2019, Valavi et al. 2022). Default settings commonly result in models that are overly complex and *overfit* to the training data, producing distorted suitability estimates and poor transferability to other places or times (Moreno-Amat et al. 2015, Morales et al. 2017, Tracy et al. 2018). This is especially problematic when the occurrence data are biased (Merow et al. 2014, Warren et al. 2014, Galante et al. 2018; interaction with Hazard 2; Fig. 2) and for inappropriate model-evaluation schemes that cannot detect overfitting (interaction with Hazard 8; Fig. 2).

Recommendations. Instead of relying on default settings, tuning experiments allow researchers to explore multiple parameterization scenarios and evaluate them via approaches that can detect overfitting (e.g. spatial blocks for data partitioning; Merow et al. 2014, Tracy et al. 2018, Soley-Guardia et al. 2019; interaction with Hazard 8; Fig. 2). For example, with MaxEnt, multiple candidate models for each species can be run with different combinations of feature classes and regularization multipliers; with appropriate model-evaluation schemes, overfit models will show poorer performance on validation data than on training data (Radosavljevic and Anderson 2014). Recent software packages (e.g. ‘biomod2’; Thuiller et al. 2009; ‘ENMeval’; Kass et al. 2021; ‘SDMtoolbox’; Brown et al. 2017; ‘Wallace EcoMod’; Kass et al. 2018, 2023), have made this practical via automation, at least for certain algorithms. Despite increasing computational time, their ease of automation makes tuning exercises a feasible element for projects modeling many species (Valavi et al. 2022); nevertheless, as for all modeling efforts, expert inspection of the response curves and geographic prediction of the selected model to assess ecological

realism remains beneficial and wise (Hazard 9; Guevara et al. 2018b, Velásquez-Tibatá et al. 2019).

5. Reducing predictor variables irrespective of their information content

With the goal of minimizing variable correlations and/or overfitting, researchers often reduce the number of predictors prior to model building irrespective of their information content (i.e. their predictive power or ability to inform regarding a species’ distribution), for example via pre-determined cutoffs in correlation analyses (Breiman 2001, Feng et al. 2019b, Sillero et al. 2021). This approach stems from otherwise sound statistical practices in regression modeling, often also aimed at identifying the contributions of particular variables and obtaining simple, explanatory models that facilitate interpretation and hypothesis-testing (Dormann et al. 2013, Farrell et al. 2019, Feng et al. 2019b).

Assumptions. Modeling with fewer or less correlated variables yields simpler and better models (Breiman 2001, Elith et al. 2008, Evans et al. 2013) that are not fitted to nuances in the data.

Problem. Removing predictor variables a priori without consideration of their information content may arbitrarily discard informative data (reducing the predictive ability of the model) and does not directly address the problem of overfitting (Breiman 2001, Olden et al. 2008, Braunisch et al. 2013).

Recommendations. After identifying candidate variables based on their biological relevance (Hazard 3), approaches that consider information content to limit complexity (instead of via correlations a priori) can reduce overfitting and lead to improved models (Cobos et al. 2019a, Farrell et al. 2019). Many algorithms have options for controlling the degree of fitting to the sample by penalizing complex models (e.g. those with higher numbers of variables and more complex responses to them). Approaches for limiting complexity include regularization procedures like lasso and ridge regression in GLMs and MaxEnt and pruning in regression trees (Hastie et al. 2009, Dicko et al. 2014, Guisan et al. 2017, Valavi et al. 2022). Values of the penalties for higher complexity (e.g. the regularization multiplier in MaxEnt; interaction with Hazard 4; Fig. 2) can be tuned using evaluation procedures that detect overfitting (Hazard 8). Increasing the penalization for complexity tends to lead to zero contribution for variables with low or no information content (by themselves or in combination with correlated variables that are more informative), in effect excluding them from the final model (interaction with Hazard 4; Fig. 2; Phillips et al. 2017, Farrell et al. 2019, Valavi et al. 2022). Because of this, algorithms that implement variable selection indirectly through penalties for complexity tend to yield models without highly correlated variables, which also simplifies interpretation of the response curves for those retained by the algorithm (Feng et al. 2019b, Morente-López et al. 2022). Such approaches dovetail with projects that automate model-building for many species,

as long as appropriate evaluation procedures are employed (Hazard 8).

6. Using an overly large study region

Often, researchers use an overly large study region for model building and/or evaluation (Araújo and Peterson 2012, Liang et al. 2018). Sometimes such a region is selected with the misconception that it will yield an accurate depiction of the species' range (rather than the areas suitable for it) but probably most often because a map of suitability is desired in that full area. For example, researchers often seek to find suitable areas with sparse sampling, identify likely areas of spread for invasive species, or characterize potential overlap where related species may occur (uses that all require estimates of suitability).

Assumptions. Within the study region for model training, the species occurs in spatial (or at least environmental) equilibrium. In the first scenario, the species inhabits all suitable areas (i.e. spatial equilibrium; Peterson et al. 2011, Guisan et al. 2017). Because of this, its distribution also corresponds to environmental equilibrium: the places occupied by the species accurately reflect the environments suitable for it. In the second scenario, although the species does not occur in all suitable areas, it inhabits an environmentally representative (unbiased) subset of them (i.e. environmental equilibrium). Hence, it fulfills the *noise assumptions* (Anderson 2013), whereby factors related to dispersal, establishment, and persistence of populations; biotic interactions; and human actions do not cause the species to occupy an environmentally biased subset of the areas suitable for it, but rather only add statistical noise.

Problem. The chances of violating the spatial-equilibrium or environmental-equilibrium (noise) assumptions increase with the extent of the study region (Saupe et al. 2012, Cooper and Soberón 2018). Strong departures from these assumptions occur especially because of barriers to dispersal and for species not yet at equilibrium with suitability (e.g. invasive species or those not quickly tracking a changing climate; Elith et al. 2010, Anderson 2013). Environmental information extracted from areas that are suitable yet unoccupied can bias the inferred environmental associations, underestimating suitability in areas both within and beyond the range (Jiménez-Valverde et al. 2013, Cooper and Soberón 2018, Liang et al. 2018); the same is true for occupied regions not yet documented because of biased sampling; Hazard 2). Unfortunately, such violations often lead to inflated measures of performance when using evaluation statistics that include overestimates of *commission* error (a pervasive problem with background or pseudoabsence data; Jiménez-Valverde 2012, Radosavljevic and Anderson 2014; interaction with Hazard 7; Fig. 2). This constitutes a particularly dangerous interaction between hazards because it gives a false impression of good model performance.

Recommendations. An appropriate study region approximating the assumption of spatial equilibrium (or the noise

assumptions) should be defined using available information regarding the species' natural history and the configuration of the landscape (e.g. excluding regions beyond likely barriers to dispersal; Anderson and Raza 2010, Barve et al. 2011, Saupe et al. 2012, Guisan et al. 2017; or with particularly low sampling effort; Hazard 2). Doing so has the effect of masking out areas where the lack of occurrence records derives from factors other than the environmental variables considered. Buffers around occurrences or established biogeographic regions may better approximate assumptions, although the buffering distances employed generally remain subjective (Brown et al. 2016, Mammola and Isaia 2017, Soley-Guardia et al. 2019, Andrade et al. 2020). Buffering known occurrence records also can have the benefit of excluding consideration of areas with particularly low sampling effort, where such a bias can negatively affect model building by sending a false negative signal (interaction with Hazard 2; Fig. 2). If needed, models can then be transferred to larger spatial extents to assess suitability beyond the known range, minding necessary caveats regarding any *non-analog* conditions that require extrapolation in environmental space (interaction with Hazard 9; Fig. 2). Complementarily, these models can be post-processed to obtain actual ranges (e.g. masking predictions with land-use layers; Heap 2016, Calixto-Pérez et al. 2018, Merow et al. 2022), if such estimates are needed (instead of suitability). Projects modeling many species likely will employ automated decision-making (e.g. buffering occurrence records by a given distance or using biogeographic regions). Although species-specific choices likely would approximate assumptions more closely and produce superior models, making these decisions by bunches (e.g. for species with similar natural histories and inhabiting regions with comparable levels of environmental heterogeneity) may prove more reasonable than a single choice for all species.

Model evaluation

7. Misinterpreting metrics of model performance

A particularly widespread hazard in studies employing presence-background or presence-pseudoabsence data is evaluating performance using metrics designed for analyses with reliable absence information (e.g. AUC, Kappa, true skill statistic), under the misconception that they represent statistically unbiased, absolute measures of performance (Lobo et al. 2008, Leroy et al. 2018). Unfortunately, with great frequency researchers interpret these metrics in the same manner as when true absence data are used for their calculation (e.g. predictions with AUC values > 0.5 being better than random and those approaching 1.0 being a requisite for or indicative of a good model); or as comparable among predictions for different species or study regions (Jiménez-Valverde et al. 2013, Liang et al. 2018, Fernandes et al. 2019).

Assumptions. Error rates for *omission* (false negatives) and *commission* (false positives) are both accurately calculated (Lobo et al. 2008, Jiménez-Valverde 2012, Leroy et al. 2018; see also their respective complements, *sensitivity* and

specificity). For commission error and metrics that include it, comparisons across species or regions assume that the ratio of suitable to unsuitable environments remains constant (Jiménez-Valverde et al. 2013, Bohl et al. 2019).

Problem. When absence data are not available, commission errors estimated from a background or pseudoabsence sample suffer from (an unknown) statistical bias. Great numbers of map pixels typically exist within areas that are suitable for the species, but the vast majority of them are not documented by the limited sample of presence records (a problem exacerbated by overly large study regions, where factors other than environmental suitability preclude the species' presence; interaction with Hazard 6; Fig. 2). Because of this, the estimate of commission error is strongly inflated (Anderson et al. 2003, Araújo and Peterson 2012, Leroy et al. 2018, Saupe et al. 2018). Therefore, when calculated using background or pseudoabsence data, commission error and metrics derived from it are biased indicators of performance (Radosavljevic and Anderson 2014, Fourcade et al. 2018, Liang et al. 2018), penalizing models that correctly predict suitability beyond the species' documented occurrences and favoring those that do not (Araújo and Peterson 2012, Jiménez-Valverde 2012). Critically, it follows that because of this bias, such metrics do not represent values of absolute performance, but instead relative ones valid for comparisons only with other analyses for the same species and study region (Lobo et al. 2008, VanDerWal et al. 2009, Jiménez-Valverde et al. 2013). Additionally, this bias undermines the use of these metrics in identifying an optimal threshold for converting model output into a binary prediction of suitable vs unsuitable conditions (e.g. via the sum of sensitivity and specificity; Liu et al. 2013).

Recommendations. Adjustments for differential weighting of omission and commission errors have been proposed (Peterson et al. 2008), but appropriate weights for given study systems remain elusive. Instead, metrics that gauge performance without estimating commission error are valid and more appropriate for this type of data, both for assessing model performance and for use in threshold selection. For example, rates of omission at a given threshold are commonly used and comparable across species (e.g. validation omission rate based on a threshold corresponding to the minimum training presence value; Peterson et al. 2011), at least when the effects of sampling bias have been ameliorated (Hazard 2). In situations where metrics based on both omission and commission can provide useful information (e.g. assessing discrimination from the background for the same species and study region in tuning exercises; Soley-Guardia et al. 2016, Galante et al. 2018), they are best interpreted against a null expectation specific to the system. Fortunately, null-model approaches now exist to determine statistical significance and effect size (Raes and ter Steege 2007, Bohl et al. 2019; for example including implementation in 'ENMeval'; Kass et al. 2021). These approaches increase computational time greatly but do not represent analytical barriers for projects that automate modeling of many species.

8. Evaluating models with random splits of the data

Most SDM studies lack truly independent occurrence data for testing (e.g. independently collected from a random or stratified survey design). Because of this, researchers commonly conduct model evaluation by partitioning available presence (and background or pseudoabsence) data into subsets, with some being used for model training and the rest for validation (Hastie et al. 2009, Zurell et al. 2020b). Over the past two decades, a particularly widespread partitioning approach has employed random division of the data (e.g. cross validation with random division into k folds, or groups, one of which is excluded from training in each round to serve instead for evaluation; Naimi and Araújo 2016, Roberts et al. 2017).

Assumptions. Training and validation datasets created using random splits are independent from one another.

Problem. Random splits of occurrence data do not provide statistical independence; instead, data points of the training and validation subsets often end up lying in geographic proximity (especially if sampling is spatially clustered; interaction with Hazard 2; Fig. 2). Due to pervasive spatial autocorrelation of the environment, geographically proximal records share similar or identical conditions; furthermore, because training and validation subsets are highly non-independent, they both reflect the same biases of the overall sample (Jiménez-Valverde et al. 2011, Wenger and Olden 2012, Roberts et al. 2017). Hence, undesired fitting to biases in the training data (interaction with Hazard 2; Fig. 2), which is more likely in overly complex models (interaction with Hazard 4), will go undetected – yielding inflated measures of performance via an especially misleading interaction among these three hazards (Veloz 2009, Anderson 2012, Hijmans 2012, Fourcade et al. 2018).

Recommendations. Splitting data non-randomly increases independence and reduces spatial correlation among subsets (e.g. using spatial or environmental blocks; Radosavljevic and Anderson 2014, Roberts et al. 2017). Such data-partitioning schemes make it possible to detect fitting to biases in the sample (interaction with Hazard 2; Fig. 2). Hence, they can be applied to estimate optimal model complexity appropriately in tuning exercises, rather than relying on default settings that often lead to overfitting (interaction with Hazard 4; Fig. 2). Fortunately, evaluation procedures with automated non-random (e.g. spatial) partitioning exist ('BlockCV'; Valavi et al. 2019; 'ENMeval'; Kass et al. 2021; 'ENMTML'; Andrade et al. 2020; 'kuenm'; Cobos et al. 2019b; 'SDMtoolbox'; Brown et al. 2017; 'Wallace EcoMod'; Kass et al. 2018, 2023). Notably, this approach often faces the additional challenge of requiring extrapolation of response curves into non-analog environments (entailing additional caveats, including changes in the correlation structure of the variables; interaction with Hazard 9; Fig. 2). However, this can be regarded as an advantage when the ultimate use will require model transfer to another place or time (because spatial blocks allow assessment of the model under transfer;

Roberts et al. 2017, Soley-Guardia et al. 2019). Non-random splits for cross-validation exercises are feasible for automated analyses modeling many species, but they increase the need for expert inspection of the response curves and geographic prediction of the selected model.

Model interpretation

9. Misunderstanding or overlooking the effects of extrapolation

When the ultimate use of a model requires its transfer to another place or time, the new conditions typically contain at least some non-analog environments (beyond those found in the training study region). This makes extrapolation of the modeled response curves necessary to make a prediction (Anderson 2013; see slightly different use of terminology in Owens et al. 2013, Qiao et al. 2019). Frequently, however, researchers confuse this need to extrapolate with the particular manner in which extrapolation is accomplished.

Assumptions. Two fundamental methods exist for extrapolating beyond the minimum and maximum environmental values found for any variable within the training dataset (composed of the sample of presences and the background or pseudoabsence information). One assumes that the modeled response to a given variable continues its trend unconstrained (e.g. achieved by including cubic and quadratic terms in GLMs, or disabling ‘clamping’ in MaxEnt); the other assumes that the response remains fixed at the last value of the training dataset (e.g. using linear or constant splines in GLMs, ‘clamping’ in MaxEnt, or in all uses of classification and regression trees including random forests and boosted regression trees; Elith and Graham 2009, Elith et al. 2010, Anderson 2013). Which method is more appropriate, and how much the corresponding estimates of suitability differ, both depend on several factors. These include the degree of environmental novelty in the new area or time (i.e. how different the conditions are compared with those used for model training) and the tendency and height of the response curve at its point of truncation (e.g. increasing or decreasing; nearing suitability limits or not; Fitzpatrick and Hargrove 2009, Anderson 2013, Guevara et al. 2018b).

Problem. Confusing the need to extrapolate with the manner in which it is accomplished can result in not only incorrectly documenting this important aspect of modeling but also ignoring its effects on predicted suitability and biological inferences (Guevara et al. 2018a, Feng et al. 2019a). While this applies primarily to model transfer, it also can affect predictions within the time and space where the model was built (when the background or pseudoabsence sample does not contain the full set of environments of the study region (Guevara et al. 2018b).

Recommendations. Several analytical approaches now exist to provide a clearer understanding and documentation of extrapolation. Together, they allow for flagging areas with

high uncertainty due to extrapolation, which can help researchers identify regions where drawing any inference from the suitability values may be unadvisable (Owens et al. 2013, Franklin 2023). For example, MaxEnt can calculate the degree of environmental novelty across space (‘MESS’ and ‘MoD’ analyses) as well as how much the prediction depends on the particular extrapolation method (‘clamping analysis’; Elith et al. 2010). Such analyses are best interpreted in conjunction with inspection of the response curve for each environmental variable included in the final model, to assess its plausibility in non-analog conditions (Guevara et al. 2018b). Additionally, the model’s robustness to extrapolation can be estimated with evaluation schemes that include transfer, such as spatial blocks (Roberts et al. 2017, Soley-Guardia et al. 2019) (Hazard 8). Fortunately, tools for tackling aspects of extrapolation continue to be developed (Mesgaran et al. 2014, Bartley et al. 2019, Cobos et al. 2019b, Andrade et al. 2020). These now include the ability to make separate decisions (whether or not to constrain the response) for each tail of every environmental variable (for example, depending on whether the response curve is increasing or decreasing at the point of truncation; Anderson 2013, Guevara et al. 2018b, Kass et al. 2021). Additionally, researchers can compare results among techniques that extrapolate only via a fixed response (e.g. classification and regression trees), others that do so unconstrained (e.g. some implementations of GLMs), and those with the ability to do either (e.g. MaxEnt and contrasting ways to employ GLMs) – and then characterize the associated uncertainty (Elith and Graham 2009, Araújo et al. 2019). Decisions regarding environmental extrapolation can be automated but represent a challenge for analyses modeling many species, where expert inspection of response curves, maps of predicted suitability, and uncertainty due to extrapolation remain advisable.

10. Comparing model outputs across different species or places

Without additional data or assumptions, algorithms that use presence data and a background or pseudoabsence sample (i.e. without true absence information) do not provide probability of presence but rather an output that represents relative suitability across map pixels for a given species (Phillips and Elith 2013, Yackulic et al. 2013). Nevertheless, direct comparisons of such outputs across species or geographic regions is frequently desired (e.g. to assess competition or niche overlap; Peterson 2011, Gutiérrez et al. 2014). Only within a study region where the species’ distribution is in equilibrium with environmental suitability (interaction with Hazard 6; Fig. 2), can relative suitability values be interpreted as related to probability of presence (by an unknown function, which is presumably monotonic although not necessarily linear). For techniques with theoretical links to population ecology (e.g. MaxEnt, GLMs, and GAMs; Hastie and Fithian 2013, Phillips and Elith 2013), such outputs of relative suitability can be transformed to yield absolute probability of presence, allowing direct comparisons across species and geographic regions. This requires an explicit rescaling function, obtained

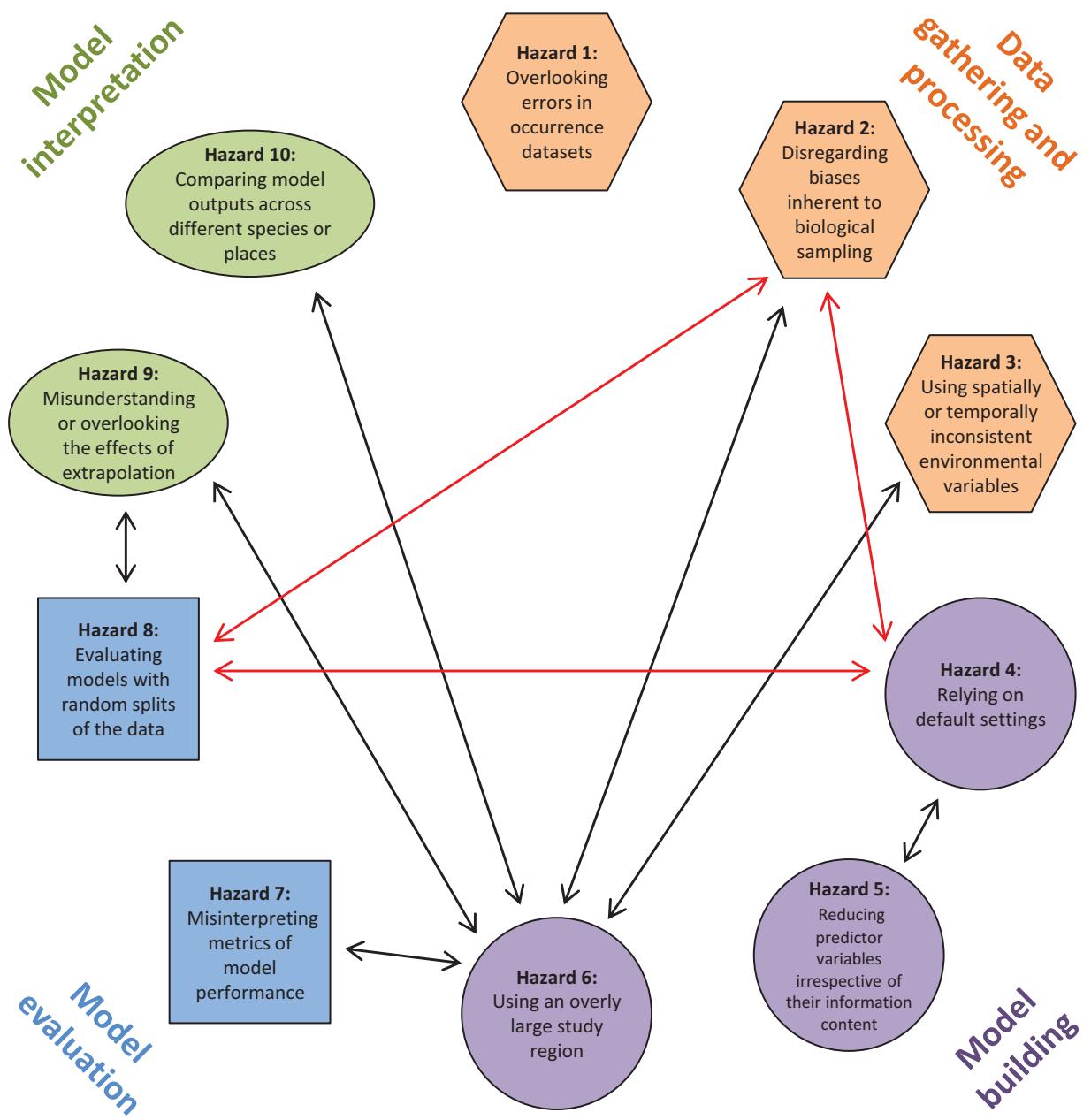


Figure 2. Illustration of interactions between pairs of the ten most problematic hazards in species distribution modeling covered in this paper (arrows; Table 2). The colored shapes denote the corresponding step of modeling (orange hexagons: data gathering and processing; purple circles: model building; blue squares: model evaluation; green ovals: model interpretation). Red arrows highlight an especially misleading interaction among hazards from three different steps of modeling that together can trick researchers into making overfit models that falsely appear to have excellent performance (Hazards 2, 4 and 8). Other interactions appear in black. Notably, Hazard 6 shows interactions with over half of the other individual hazards, emphasizing the far-reaching repercussions of violating its associated assumptions. In addition to these direct interactions explained in the text, the various hazards also have diffuse effects on each other (not shown), including the cascading impacts of Hazard 1 (overlooking errors in occurrence datasets).

from additional data or assumptions (Guillera-Arroita *et al.* 2015, Renner *et al.* 2015). For instance, MaxEnt's 'raw' output can be transformed to a probability of presence (0–1) via a scaling parameter representing either prevalence (fraction of the study region occupied; Guillera-Arroita *et al.* 2014) or local abundance ('logistic' and 'cloglog' outputs, respectively; Phillips and Elith 2013, Phillips *et al.* 2017). Estimates of

overall prevalence or abundance at particular sites can be provided by the user, but because such information rarely exists, default values typically are used (Guillera-Arroita *et al.* 2014).

Assumptions. Values given to scaling parameters are appropriate (whether default or user-defined). Alternatively, it can be assumed that even if mis-specified, the rescaling parameters

are still equivalent among the entities involved (e.g. for ecologically similar species), rendering output values that are consistently incorrect yet comparable in absolute terms.

Problem. If these assumptions are violated (or no theoretical rescaling to probability of presence exists for the algorithm being used), suitability can only be interpreted as relative across the study area of a given analysis, and the values of the predictions of different models should not be directly compared across species or regions (Fithian and Hastie 2013, Merow and Silander 2014, Guillera-Arroita et al. 2015).

Recommendations. In modeling techniques with theoretical links to population ecology (e.g. MaxEnt, GLMs, and GAMs), available estimates of occupancy or abundance can be used to set the scaling parameters rather than relying on default values (Guillera-Arroita et al. 2014, Phillips et al. 2017). By extension, if scaling parameters are unknown but can be justified as likely to be similar across species, the resulting values can be interpreted across models in absolute terms (but as comparable suitability scores, not as probability of presence; see also a modification of logistic regression aimed at allowing comparisons across species; Real et al. 2006). In all other cases, model output should be interpreted as only relative estimates of suitability across map pixels of the area of analysis (specific to that given species and region), whether or not values have been rescaled (Soley-Guardia et al. 2016). For any of these three situations, researchers can enhance interpretations by calculating the species' prevalence separately across different levels of suitability (although necessary information regarding detectability often remains lacking; Anderson 2023). In projects automating the production of models for many species, researchers likely will make assumptions regarding scaling parameters for sets of species with similar natural histories (to yield values for comparison across species or regions as suitability scores, not probability of presence).

Outstanding questions

In addition to the ten commonly misunderstood hazards discussed above (each with existing recommendations), other topics remain challenging and require additional research. Various such issues exist (e.g. spatial and temporal resolutions and correlations among environmental variables; standardization of methods to increase comparability of models across species and studies; emerging approaches for modeling with big data; Araújo et al. 2019, Franklin 2023). Below, we cover five of them, important topics currently being investigated and each related in various ways to the ten hazards covered above: model uncertainty; model complexity; biotic interactors; interactions among suitability drivers; and intraspecific niche variation. Although definitive resolutions for these issues do not yet exist, researchers should be able to appreciate them and comment upon how they may affect conclusions (Araújo et al. 2019, Sofaer et al. 2019, Zurell et al. 2020b).

Quantifying model uncertainty

Although most SDM studies rely on conclusions from a single final estimate of suitability, the field lacks a unified approach to quantify, partition, and map the uncertainty that arises from the many factors that can affect predictions (Beale and Lennon 2012, Peterson et al. 2018, Yates et al. 2018, Araújo et al. 2019). To characterize such uncertainty, researchers usually have focused on different sources by combining models built with various splits of the data (Soley-Guardia et al. 2019), under different parameterizations of the same algorithm (Breiner et al. 2015, Boria et al. 2017), and using different algorithms ('ensemble' or 'consensus' approach; Araújo and New 2007, Meller et al. 2014, Hao et al. 2020). The latter involves difficult challenges, including how to combine different output formats (Marmion et al. 2009, Sillero 2011, Andrade et al. 2020) and ensuring an equally sound implementation of each algorithm (Barry and Elith 2006, Jarnevich et al. 2015, Hao et al. 2019, 2020). Additionally, the degree of uncertainty (or outright bias) introduced by errors in taxonomy and georeferences almost always remains unknown, and the answer surely differs among taxonomic groups and geographic regions (Beale and Lennon 2012, Costa et al. 2015, Gábor et al. 2020a). Simulation studies with virtual species can be informative for determining how sensitive different approaches are to particular aspects of the modeling process (Meynard et al. 2019), but resolution of this issue and corresponding recommendations likely will depend on aspects of the system.

Reaching consensus on optimal model complexity

No consensus exists on the best criteria for approximating optimal model complexity (Peterson et al. 2011, Merow et al. 2014, Warren et al. 2014, 2020). A commonly used metric that penalizes complexity via the number of parameters incorporated into the model (AICc) remains statistically imperfect for some algorithms (e.g. MaxEnt, because the degrees of freedom cannot be calculated exactly; Warren and Seifert 2011, Warren et al. 2014, Galante et al. 2018, Velasco and González-Salazar 2019). Additionally, for measures of performance calculated using partitioned validation data or fully withheld testing data, the particular metric employed can lead to selection of different models as optimal, and no single one has been demonstrated as reliable and sufficient on its own (Hirzel et al. 2006, Boria et al. 2017, Bohl et al. 2019, Jiménez and Soberón 2020). Disagreement even remains about whether simpler models are always desirable (García-Callejas and Araújo 2016, Yates et al. 2018, Coelho et al. 2019), and few studies have investigated how complexity correlates with predictive ability (Galante et al. 2018, Velasco and González-Salazar 2019). Future studies comparing model performance according to diverse criteria (Norberg et al. 2019) should shed light on this issue and are now facilitated by several software options (e.g. 'biomod2'; Thuiller et al. 2009; 'ENMeval'; Kass et al. 2021; 'sdm'; Naimi and Araújo 2016; 'Wallace EcoMod'; Kass et al. 2018, 2023).

Integrating biotic interactors

Most SDM implementations rely on the Eltonian noise hypothesis that biotic interactions occurring at a fine spatial resolution in local communities do not alter the abiotic signal retrieved from the coarser-grain environmental data used across large spatial extents (Soberón and Nakamura 2009, Lira-Noriega et al. 2013). Nevertheless, key biotic interactions often carry their effects to resolutions and extents relevant for SDMs (Wisz et al. 2013). In these cases, SDMs can benefit from incorporating data regarding biotic interactors (assuming that the nature of their effects remains stationary across the study region; Sanín and Anderson 2018, Fern et al. 2019, Kass et al. 2020). However, for the correlative models considered here (comparing presence data with background or pseudoabsence information), only biotic interactors with unidirectional effects on the focal species should be considered as predictor variables (the interactor affects the distribution of the focal species but not vice versa; Soberón 2007, 2010, Anderson 2017). Other interactors can be informative during post-processing of model output (Peers et al. 2013, Gutiérrez et al. 2014) or via classes of models that incorporate population demography (Zurell 2017). Even joint species distribution models that can fit environmental responses of multiple species simultaneously still assume stationarity of the effects of relevant biotic interactions (Pollock et al. 2014, Poggiato et al. 2021). Clearly, incorporating biotic interactions remains difficult, and some unresolved challenges include: accounting for their statistical interaction with abiotic predictors and historical contingencies such as past extinctions and dispersal barriers (Warton et al. 2015, Dormann et al. 2018, Brown and Carnaval 2019, Early and Keith 2019, Franklin 2023); computational limitations for considering inputs from adjacent populations in local population-dynamic models (Zurell et al. 2020a); and extrapolating to non-analog biotic contexts (Williams and Jackson 2007, Jaeschke et al. 2012).

Accounting for interactions among suitability drivers

Although SDM predictions often represent the combination of independent responses estimated for each predictor variable, suitability is likely driven by non-additive effects (Merow et al. 2014, Golding and Purse 2016). Accounting for such effects can be particularly important when extrapolating into novel environmental combinations (Zurell et al. 2012, Mesgarian et al. 2014, Feng et al. 2019b). Realistic inclusion of such statistical interactions is currently possible for only a few modeling algorithms (e.g. multivariate adaptive regression splines, 'MARS'; Leathwick et al. 2006; Gaussian processes; Golding and Purse 2016). Others provide partial solutions to the problem, for instance allowing simple pairwise fixed interactions (e.g. MaxEnt's product features; Merow et al. 2013, Phillips et al. 2017), or providing great flexibility but without much specification control by the user (random forests and boosted regression trees; Elith et al.

2008, Merow et al. 2014). Simulation studies are needed to assess the importance of accounting for such interactions and determine which particular approaches are most suited to the task under what circumstances (Golding and Purse 2016).

Incorporating intraspecific niche variation

SDMs assume equal environmental associations for individuals across populations (i.e. niche conservatism over space), which may closely approximate reality for many taxa and commonly used predictor variables (Peterson 2011). However, in an increasing number of studies, genetically determined differences have been documented for factors related to distributional limits among populations (Pelini et al. 2009, Fournier-Level et al. 2011, Morente-López et al. 2022, Franklin 2023). Whereas the problem of violating this assumption of stationarity in SDMs has been addressed in studies of invasive species (van Boheemen et al. 2019, Pili et al. 2020), phylogeography (Costa et al. 2002), and climate change (Fitzpatrick and Keller 2015, Moran and Ormond 2015, Martin et al. 2020), resolving it fully involves multiple conceptual and practical challenges (Smith et al. 2019). These challenges include: 1) detecting deviations from niche homogeneity when data regarding adaptation and plasticity are not available (i.e. poor prediction in spatial transfers could stem from niche variability or from various methodological factors, including overfit models; Peterson and Holt 2003, Brown and Carnaval 2019); 2) building models with population-specific data (i.e. separate models for evolutionarily distinct populations vs single models integrating data on functional differences across the range; Fitzpatrick and Keller 2015, Hällfors et al. 2016, Thorson et al. 2016); and 3) identifying appropriate means for transferring such models across space and time (e.g. involving estimation of distributional limits for particular populations and the effect of any future intermixing; Prates et al. 2016, Martin et al. 2020).

Closing remarks

With many topics in species distribution modeling now well understood and others advancing rapidly, researchers can take advantage of over two decades of impressive progress to make defensible and useful models (Araújo et al. 2019, Sofaer et al. 2019, Zurell et al. 2020b). In contrast, not addressing adequately any of the ten hazards detailed here can hinder progress in basic science – including the understanding of fundamental biological processes – as well as lead to inefficient or ineffective use of resources and counterproductive decision-making for important applications in areas as varied as invasive agricultural pests, human zoonotic diseases, and the effects of climate change on biodiversity (Guillera-Arroita et al. 2015, Morales et al. 2017, Tuia et al. 2022). Attention to key principles holds particular relevance as researchers increasingly embark on projects modeling species en masse via automated pipelines, where quality can suffer

without expert supervision and insights. Importantly, we note several direct interactions between pairs of hazards, and in one case among three of them (Fig. 2, Table 2). Therefore, because of both their individual and collective effects, we encourage researchers to follow – and reviewers and editors to call for heeding – existing recommendations for these and other critical issues. As we mention, comprehensive treatments exist to guide research and promote quality (and replicability) across all aspects of modeling species distributions (Araújo et al. 2019, Zurell et al. 2020b). In closing, we hope that researchers will think about all of these topics carefully for successful implementations and continue to develop novel approaches that better approximate reality.

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Mariano Soley-Guardia: Conceptualization (equal); Project administration (lead); Visualization (supporting); Writing – original draft (lead); Writing – review and editing (supporting). **Diego F. Alvarado-Serrano:** Conceptualization (equal); Project administration (supporting); Visualization (lead); Writing – original draft (supporting); Writing – review and editing (supporting). **Robert P. Anderson:** Conceptualization (equal); Funding acquisition (lead); Project administration (supporting); Visualization (supporting); Writing – original draft (supporting); Writing – review and editing (lead).

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References

Aiello-Lammens, M. E., Boria, R. A., Radosavljevic, A., Vilela, B. and Anderson, R. P. 2015. *spThin*: an R package for spatial thinning of species occurrence records for use in ecological niche models. – *Ecography* 38: 541–545.

Anderson, R. P. 2003. Real vs artefactual absences in species distributions: tests for *Oryzomys albigularis* (Rodentia: Muridae) in Venezuela. – *J. Biogeogr.* 30: 591–605.

Anderson, R. P. 2012. Harnessing the world's biodiversity data: promise and peril in ecological niche modeling of species distributions. – *Ann. N. Y. Acad. Sci.* 1260: 66–80.

Anderson, R. P. 2013. A framework for using niche models to estimate impacts of climate change on species distributions. – *Ann. N. Y. Acad. Sci.* 1297: 8–28.

Anderson, R. P. 2017. When and how should biotic interactions be considered in models of species niches and distributions? – *J. Biogeogr.* 44: 8–17.

Anderson, R. P. 2023. Integrating habitat-masked range maps with quantifications of prevalence to estimate area of occupancy in IUCN assessments. – *Conserv. Biol.* 37: e14019.

Anderson, R. P. and Raza, A. 2010. The effect of the extent of the study region on GIS models of species geographic distributions and estimates of niche evolution: preliminary tests with montane rodents (genus *Nephelomys*) in Venezuela. – *J. Biogeogr.* 37: 1378–1393.

Anderson, R. P., Lew, D. and Peterson, A. T. 2003. Evaluating predictive models of species' distributions: criteria for selecting optimal models. – *Ecol. Modell.* 162: 211–232.

Anderson, R. P., Araújo, M. B., Guisan, A., Lobo, J. M., Martínez-Meyer, E., Peterson, A. T. and Soberón, J. M. 2020. Optimizing biodiversity informatics to improve information flow, data quality, and utility for science and society. – *Front. Biogeogr.* 12: e47839.

Andrade, A. F. A. de, Velazco, S. J. E. and Júnior, P. D. M. 2020. ENMTML: an R package for a straightforward construction of complex ecological niche models. – *Environ. Modell. Softw.* 125: 104615.

Araújo, M. B. and New, M. 2007. Ensemble forecasting of species distributions. – *Trends Ecol. Evol.* 22: 42–47.

Araújo, M. B. and Peterson, A. T. 2012. Uses and misuses of bioclimatic envelope modeling. – *Ecology* 93: 1527–1539.

Araújo, M. B., Anderson, R. P., Barbosa, A. M., Beale, C. M., Dormann, C. F., Early, R., Garcia, R. A., Guisan, A., Maiorano, L., Naimi, B., O'Hara, R. B., Zimmerman, N. E. and Rahbek, C. 2019. Standards for distribution models in biodiversity assessments. – *Sci. Adv.* 5: eaat4858.

Aubry, K. B., Raley, C. M. and McKelvey, K. S. 2017. The importance of data quality for generating reliable distribution models for rare, elusive, and cryptic species. – *PLoS One* 12: e0179152.

Austin, M. P. 2002. Spatial prediction of species distribution: an interface between ecological theory and statistical modelling. – *Ecol. Modell.* 157: 101–118.

Barber, R. A., Ball, S. G., Morris, R. K. A. and Gilbert, F. 2022. Target-group backgrounds prove effective at correcting sampling bias in Maxent models. – *Divers. Distrib.* 28: 128–141.

Barry, S. and Elith, J. 2006. Error and uncertainty in habitat models. – *J. Appl. Ecol.* 43: 413–423.

Bartley, M. L., Hanks, E. M., Schliep, E. M., Soranno, P. A. and Wagner, T. 2019. Identifying and characterizing extrapolation in multivariate response data. – *PLoS One* 14: e0225715.

Barve, N., Barve, V., Jiménez-Valverde, A., Lira-Noriega, A., Maher, S. P., Peterson, A. T., Soberón, J. and Villalobos, F. 2011. The crucial role of the accessible area in ecological niche modeling and species distribution modeling. – *Ecol. Modell.* 222: 1810–1819.

Beale, C. M. and Lennon, J. J. 2012. Incorporating uncertainty in predictive species distribution modelling. – *Phil. Trans. R. Soc. B* 367: 247–258.

Beck, J., Böller, M., Erhardt, A. and Schwanghart, W. 2014. Spatial bias in the GBIF database and its effect on modeling species' geographic distributions. – *Ecol. Inform.* 19: 10–15.

Beery, S., Cole, E., Parker, J., Perona, P. and Winner, K. 2021. Species distribution modeling for machine learning practitioners: a review. – *COMPASS '21: Proc. of the 2021 4th ACM SIGCAS Conf. on Computing and Sustainable Societies*. pp. 329–348.

Bohl, C. L., Kass, J. M. and Anderson, R. P. 2019. A new null model approach to quantify performance and significance for ecological niche models of species distributions. – *J. Biogeogr.* 46: 1101–1111.

Bonfim, F. C. G., Cordeiro, P. H. C., Peres, C. A., Canale, G. R. and Bernardo, C. S. S. 2019. Combining modeling tools to identify conservation priority areas: a case study of the last large-bodied avian frugivore in the Atlantic Forest. – *Global Ecol. Conserv.* 17: e00426.

Booth, T. H., Nix, H. A., Busby, J. R. and Hutchinson, M. F. 2014. BIOCLIM: the first species distribution modelling package, its early applications and relevance to most current MaxEnt studies. – *Divers. Distrib.* 20: 1–9.

Boria, R. A., Olson, L. E., Goodman, S. M. and Anderson, R. P. 2014. Spatial filtering to reduce sampling bias can improve the performance of ecological niche models. – *Ecol. Modell.* 275: 73–77.

Boria, R. A., Olson, L. E., Goodman, S. M. and Anderson, R. P. 2017. A single-algorithm ensemble approach to estimating suitability and uncertainty: cross-time projections for four Malagasy tenrecs. – *Divers. Distrib.* 23: 196–208.

Braunisch, V., Coppes, J., Arlettaz, R., Suchant, R., Schmid, H. and Bollmann, K. 2013. Selecting from correlated climate variables: a major source of uncertainty for predicting species distributions under climate change. – *Ecography* 36: 971–983.

Breiman, L. 2001. Statistical modeling: the two cultures. – *Stat. Sci.* 16: 199–215.

Breiner, F. T., Guisan, A., Bergamini, A. and Nobis, M. P. 2015. Overcoming limitations of modelling rare species by using ensembles of small models. – *Methods Ecol. Evol.* 6: 1210–1218.

Briscoe, N. J., Elith, J., Salguero-Gómez, R., Lahoz-Monfort, J. J., Camac, J. S., Giljohann, K. M., Holden, M. H., Hradsky, B. A., Kearney, M. R., McMahon, S. M., Phillips, B. L., Regan, T. J., Rhodes, J. R., Vesk, P. A., Wintle, B. A., Yen, J. D. L. and Guillera-Arroita, G. 2019. Forecasting species range dynamics with process-explicit models: matching methods to applications. – *Ecol. Lett.* 22: 1940–1956.

Brown, J. L. and Carnaval, A. C. 2019. A tale of two niches: methods, concepts, and evolution. – *Front. Biogeogr.* 11: e44158.

Brown, K. A., Parks, K. E., Bethell, C. A., Johnson, S. E. and Mulligan, M. 2015. Predicting plant diversity patterns in Madagascar: understanding the effects of climate and land cover change in a biodiversity hotspot. – *PLoS One* 10: e0122721.

Brown, J. L., Weber, J. J., Alvarado-Serrano, D. F., Hickerson, M. J., Franks, S. J. and Carnaval, A. C. 2016. Predicting the genetic consequences of future climate change: the power of coupling spatial demography, the coalescent, and historical landscape changes. – *Am. J. Bot.* 103: 153–163.

Brown, J. L., Bennett, J. R. and French, C. M. 2017. SDMtoolbox 2.0: the next generation Python-based GIS toolkit for landscape genetic, biogeographic and species distribution model analyses. – *PeerJ* 5: e4095.

Calixto-Pérez, E., Alarcón-Guerrero, J., Ramos-Fernández, G., Dias, P. A. D., Rangel-Negrín, A., Améndola-Pimenta, M., Domingo, C., Arroyo-Rodríguez, V., Pozo-Montuy, G., Pinacho-Guendulain, B., Urquiza-Haas, T., Koleff, P. and Martínez-Meyer, E. 2018. Integrating expert knowledge and ecological niche models to estimate Mexican primates' distribution. – *Primates* 59: 451–467.

Castellanos, A. A., Huntley, J. W., Voelker, G. and Lawing, A. M. 2019. Environmental filtering improves ecological niche models across multiple scales. – *Methods Ecol. Evol.* 10: 481–492.

Cobos, M. E., Peterson, A. T., Osorio-Olvera, L. and Jiménez-García, D. 2019a. An exhaustive analysis of heuristic methods for variable selection in ecological niche modeling and species distribution modeling. – *Ecol. Inform.* 53: 100983.

Cobos, M. E., Peterson, A. T., Barve, N. and Osorio-Olvera, L. 2019b. kuenm: an R package for detailed development of ecological niche models using Maxent. – *PeerJ* 7: e6281.

Coelho, M. T. P., Diniz-Filho, J. A. and Rangel, T. F. 2019. A parsimonious view of the parsimony principle in ecology and evolution. – *Ecography* 42: 968–976.

Cooper, J. C. and Soberón, J. 2018. Creating individual accessible area hypotheses improves stacked species distribution model performance. – *Global Ecol. Biogeogr.* 27: 156–165.

Costa, J., Peterson, A. T. and Beard, C. B. 2002. Ecologic niche modeling and differentiation of populations of *Triatoma brasiliensis* Neiva, 1911, the most important Chagas' disease vector in northeastern Brazil (Hemiptera, Reduviidae, Triatominae). – *Am. J. Trop. Med. Hyg.* 67: 516–520.

Costa, H., Foody, G. M., Jiménez, S. and Silva, L. 2015. Impacts of species misidentification on species distribution modeling with presence-only data. – *ISPRS Int. J. Geo-Inf.* 4: 2496–2518.

Daru, B. H., Park, D. S., Primack, R. B., Willis, C. G., Barrington, D. S., Whitfeld, T. J. S., Seidler, T. G., Sweeney, P. W., Foster, D. R., Ellison, A. M. and Davis, C. C. 2018. Widespread sampling biases in herbaria revealed from large-scale digitization. – *New Phytol.* 217: 939–955.

Dicko, A. H., Lancelot, R., Seck, M. T., Guerrini, L., Sall, B., Lo, M., Vreyen, M. J. B., Lefrançois, T., Fonta, W. M., Peck, S. L. and Bouyer, J. 2014. Using species distribution models to optimize vector control in the framework of the tsetse eradication campaign in Senegal. – *Proc. Natl Acad. Sci. USA* 111: 10149–10154.

Dormann, C. F., Elith, J., Bacher, S., Buchmann, C., Carl, G., Carré, G., Marquéz, J. R. G., Gruber, B., Lafourcade, B., Leitão, P. J., Münkemüller, T., McClean, C., Osborne, P. E., Reineking, B., Schröder, B., Skidmore, A. K., Zurell, D. and Lautenbach, S. 2013. Collinearity: a review of methods to deal with it and a simulation study evaluating their performance. – *Ecography* 36: 27–46.

Dormann, C. F., Bobrowski, M., Dehling, D. M., Harris, D. J., Hartig, F., Lischke, H., Moretti, M. D., Pagel, J., Pinkert, S., Schleuning, M., Schmidt, S. I., Sheppard, C. S., Steinbauer, M. J., Zeuss, D. and Kraan, C. 2018. Biotic interactions in species distribution modelling: 10 questions to guide interpretation and avoid false conclusions. – *Global Ecol. Biogeogr.* 27: 1004–1016.

Early, R. and Keith, S. A. 2019. Geographically variable biotic interactions and implications for species ranges. – *Global Ecol. Biogeogr.* 28: 42–53.

Elith, J. and Graham, C. H. 2009. Do they? How do they? WHY do they differ? On finding reasons for differing performances of species distribution models. – *Ecography* 32: 66–77.

Elith, J. and Leathwick, J. R. 2009. Species distribution models: ecological explanation and prediction across space and time. – *Annu. Rev. Ecol. Evol. Syst.* 40: 677–697.

Elith, J., Leathwick, J. R. and Hastie, T. 2008. A working guide to boosted regression trees. – *J. Anim. Ecol.* 77: 802–813.

Elith, J., Kearney, M. and Phillips, S. 2010. The art of modelling range-shifting species. – *Methods Ecol. Evol.* 1: 330–342.

Elith, J., Phillips, S. J., Hastie, T., Dudík, M., Chee, Y. E. and Yates, C. J. 2011. A statistical explanation of MaxEnt for ecologists. – *Divers. Distrib.* 17: 43–57.

Enriquez-Urzelai, U., Kearney, M. R., Nicieza, A. G. and Tingley, R. 2019. Integrating mechanistic and correlative niche models to unravel range-limiting processes in a temperate amphibian. – *Global Change Biol.* 25: 2633–2647.

Escobar, L. E. and Craft, M. E. 2016. Advances and limitations of disease biogeography using ecological niche modeling. – *Front. Microbiol.* 7: 1174.

Evans, M. R., Grimm, V., Johst, K., Knuuttila, T., de Langhe, R., Lessells, C. M., Merz, M., O’Malley, M. A., Orzack, S. H., Weisberg, M., Wilkinson, D. J., Wolkenhauer, O. and Benton, T. G. 2013. Do simple models lead to generality in ecology? – *Trends Ecol. Evol.* 28: 578–583.

Farrell, A., Wang, G., Rush, S. A., Martin, J. A., Belant, J. L., Butler, A. B. and Godwin, D. 2019. Machine learning of large-scale spatial distributions of wild turkeys with high-dimensional environmental data. – *Ecol. Evol.* 9: 5938–5949.

Feng, X., Park, D. S., Walker, C., Peterson, A. T., Merow, C. and Papeš, M. 2019a. A checklist for maximizing reproducibility of ecological niche models. – *Nat. Ecol. Evol.* 3: 1382–1395.

Feng, X., Park, D. S., Liang, Y., Pandey, R. and Papeš, M. 2019b. Collinearity in ecological niche modeling: confusions and challenges. – *Ecol. Evol.* 9: 10365–10376.

Fern, R. R., Morrison, M. L., Wang, H. H., Grant, W. E. and Campbell, T. A. 2019. Incorporating biotic relationships improves species distribution models: modeling the temporal influence of competition in conspecific nesting birds. – *Ecol. Modell.* 408: 108743.

Fernandes, R. F., Scherrer, D. and Guisan, A. 2019. Effects of simulated observation errors on the performance of species distribution models. – *Divers. Distrib.* 25: 400–413.

Fick, S. E. and Hijmans, R. J. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. – *Int. J. Climatol.* 37: 4302–4315.

Fithian, W. and Hastie, T. 2013. Finite-sample equivalence in statistical models for presence-only data. – *Ann. Appl. Stat.* 7: 1917–1939.

Fitzpatrick, M. C. and Hargrove, W. W. 2009. The projection of species distribution models and the problem of non-analog climate. – *Biodivers. Conserv.* 18: 2255–2261.

Fitzpatrick, M. C. and Keller, S. R. 2015. Ecological genomics meets community-level modelling of biodiversity: mapping the genomic landscape of current and future environmental adaptation. – *Ecol. Lett.* 18: 1–16.

Fitzpatrick, M. C., Lachmuth, S. and Haydt, N. T. 2021. The ODMAP protocol: a new tool for standardized reporting that could revolutionize species distribution modeling. – *Ecography* 44: 1067–1070.

Fletcher, R. J., Hefley, T. J., Robertson, E. P., Zuckerberg, B., McCleery, R. A. and Dorazio, R. M. 2019. A practical guide for combining data to model species distributions. – *Ecology* 100: e02710.

Fordham, D. A., Bertelsmeier, C., Brook, B. W., Early, R., Neto, D., Brown, S. C., Ollier, S. and Araújo, M. B. 2018. How complex should models be? Comparing correlative and mechanistic range dynamics models. – *Global Change Biol.* 24: 1357–1370.

Fourcade, Y., Engler, J. O., Rödder, D. and Secondi, J. 2014. Mapping species distributions with MAXENT using a geographically biased sample of presence data: a performance assessment of methods for correcting sampling bias. – *PLoS One* 9: e97122.

Fourcade, Y., Besnard, A. G. and Secondi, J. 2018. Paintings predict the distribution of species, or the challenge of selecting environmental predictors and evaluation statistics. – *Global Ecol. Biogeogr.* 27: 245–256.

Fournier-Level, A., Korte, A., Cooper, M. D., Nordborg, M., Schmitt, J. and Wilczek, A. M. 2011. A map of local adaptation in *Arabidopsis thaliana*. – *Science* 334: 86–89.

Franklin, J. 2010. Mapping species distributions: spatial inference and prediction. – Cambridge Univ. Press.

Franklin, J. 2023. Species distribution modelling supports the study of past, present and future biogeographies. – *J. Biogeogr.* 50: 1533–1545.

Gábor, L., Moudrý, V., Lecours, V., Malavasi, M., Barták, V., Fogl, M., Šimová, P., Rocchini, D. and Václavík, T. 2020a. The effect of positional error on fine scale species distribution models increases for specialist species. – *Ecography* 43: 256–269.

Gábor, L., Moudrý, V., Barták, V. and Lecours, V. 2020b. How do species and data characteristics affect species distribution models and when to use environmental filtering? – *Int. J. Geogr. Inf. Sci.* 34: 1567–1584.

Gaier, A. G. and Resasco, J. 2023. Does adding community science observations to museum records improve distribution modeling of a rare endemic plant? – *Ecosphere* 14: e4419.

Galante, P. J., Alade, B., Muscarella, R., Jansa, S. A., Goodman, S. M. and Anderson, R. P. 2018. The challenge of modeling niches and distributions for data-poor species: a comprehensive approach to model complexity. – *Ecography* 41: 726–736.

García-Callejas, D. and Araújo, M. B. 2016. The effects of model and data complexity on predictions from species distributions models. – *Ecol. Modell.* 326: 4–12.

García-Roselló, E., Guisande, C., Heine, J., Pelayo-Villamil, P., Manjarrés-Hernández, A., González Vilas, L., González-Dacosta, J., Vaamonde, A. and Granado-Lorencio, C. 2014. Using MODESTR to download, import and clean species distribution records. – *Methods Ecol. Evol.* 5: 708–713.

Golding, N. and Purse, B. V. 2016. Fast and flexible Bayesian species distribution modelling using Gaussian processes. – *Methods Ecol. Evol.* 7: 598–608.

Gomes, V. H. F., Vieira, I. C. G., Salomão, R. P. and ter Steege, H. 2019. Amazonian tree species threatened by deforestation and climate change. – *Nat. Clim. Change* 9: 547–553.

González-Serna, M. J., Cordero, P. J. and Ortego, J. 2019. Spatiotemporally explicit demographic modelling supports a joint effect of historical barriers to dispersal and contemporary landscape composition on structuring genomic variation in a red-listed grasshopper. – *Mol. Ecol.* 28: 2155–2172.

Guevara, L., León-Paniagua, L., Ríos, J. and Anderson, R. P. 2018a. Variación entre modelos de circulación global para reconstrucciones de distribuciones geográficas del último máximo glacial: relevancia para la filogeografía. – *Ecosistemas* 27: 62–76.

Guevara, L., Gerstner, B. E., Kass, J. M. and Anderson, R. P. 2018b. Toward ecologically realistic predictions of species distributions: a cross-time example from tropical montane cloud forests. – *Global Change Biol.* 24: 1511–1522.

Guillera-Arroita, G., Lahoz-Monfort, J. J. and Elith, J. 2014. Maxent is not a presence-absence method: a comment on Thibaud et al. – *Methods Ecol. Evol.* 5: 1192–1197.

Guillera-Arroita, G., Lahoz-Monfort, J. J., Elith, J., Gordon, A., Kujala, H., Lentini, P. E., McCarthy, M. A., Tingley, R. and Wintle, B. A. 2015. Is my species distribution model fit for purpose? Matching data and models to applications. – *Global Ecol. Biogeogr.* 24: 276–292.

Guisan, A., Petitpierre, B., Broennimann, O., Daehler, C. and Kueffer, C. 2014. Unifying niche shift studies: insights from biological invasions. – *Trends Ecol. Evol.* 29: 260–269.

Guisan, A., Thuiller, W. and Zimmermann, N. E. 2017. Habitat suitability and distribution models: with applications in R. – Cambridge Univ. Press.

Gutiérrez, E. E., Boria, R. A. and Anderson, R. P. 2014. Can biotic interactions cause allopatry? Niche models, competition, and distributions of South American mouse opossums. – *Ecography* 37: 741–753.

Hälfors, M. H., Liao, J., Dzurisin, J., Grundel, R., Hyvärinen, M., Towle, K., Wu, G. C. and Hellmann, J. J. 2016. Addressing potential local adaptation in species distribution models: implications for conservation under climate change. – *Ecol. Appl.* 26: 1154–1169.

Hallgren, W., Santana, F., Low-Choy, S., Zhao, Y. and Mackey, B. 2019. Species distribution models can be highly sensitive to algorithm configuration. – *Ecol. Modell.* 408: 108719.

Hao, T., Elith, J., Guillera-Arroita, G. and Lahoz-Monfort, J. J. 2019. A review of evidence about use and performance of species distribution modelling ensembles like BIOMOD. – *Divers. Distrib.* 25: 839–852.

Hao, T., Elith, J., Lahoz-Monfort, J. J. and Guillera-Arroita, G. 2020. Testing whether ensemble modelling is advantageous for maximising predictive performance of species distribution models. – *Ecography* 43: 549–558.

Hastie, T. and Fithian, W. 2013. Inference from presence-only data: the ongoing controversy. – *Ecography* 36: 864–867.

Hastie, T., Tibshirani, R. and Friedman, J. 2009. The elements of statistical learning: data mining, inference, and prediction. 2nd edn. – Springer.

Heap, M. J. 2016. Introducing BioSARN – an ecological niche model refinement tool. – *Ecol. Evol.* 6: 5808–5816.

Hijmans, R. J. 2012. Cross-validation of species distribution models: removing spatial sorting bias and calibration with a null model. – *Ecology* 93: 679–688.

Hirzel, A. H., Le Lay, G., Helfer, V., Randin, C. and Guisan, A. 2006. Evaluating the ability of habitat suitability models to predict species presences. – *Ecol. Modell.* 199: 142–152.

Isaac, N. J. B. and Pocock, M. J. O. 2015. Bias and information in biological records. – *Biol. J. Linn. Soc. Lond.* 115: 522–531.

Isaac, N. J. B., Jarzyna, M. A., Keil, P., Damblay, L. I., Boersch-Supan, P. H., Browning, E., Freeman, S. N., Golding, N., Guillera-Arroita, G., Henrys, P. A., Jarvis, S., Lahoz-Monfort, J., Pagel, J., Pescott, O. L., Schmucki, R., Simmonds, E. G. and O'Hara, R. B. 2020. Data integration for large-scale models of species distributions. – *Trends Ecol. Evol.* 35: 56–67.

Jaeschke, A., Bittner, T., Jentsch, A., Reineking, B., Schlumprecht, H. and Beierkuhnlein, C. 2012. Biotic interactions in the face of climate change: a comparison of three modelling approaches. – *PLoS One* 7: e51472.

Jarnevich, C. S., Stohlgren, T. J., Kumar, S., Morissette, J. T. and Holcombe, T. R. 2015. Caveats for correlative species distribution modeling. – *Ecol. Inform.* 29: 6–15.

Jiménez, L. and Soberón, J. 2020. Leaving the area under the receiving operating characteristic curve behind: an evaluation method for species distribution modelling applications based on presence-only data. – *Methods Ecol. Evol.* 11: 1571–1586.

Jiménez-Valverde, A. 2012. Insights into the area under the receiver operating characteristic curve (AUC) as a discrimination measure in species distribution modelling. – *Global Ecol. Biogeogr.* 21: 498–507.

Jiménez-Valverde, A., Peterson, A. T., Soberón, J., Overton, J. M., Aragón, P. and Lobo, J. M. 2011. Use of niche models in invasive species risk assessments. – *Biol. Invas.* 13: 2785–2797.

Jiménez-Valverde, A., Acevedo, P., Barbosa, A. M., Lobo, J. M. and Real, R. 2013. Discrimination capacity in species distribution models depends on the representativeness of the environmental domain. – *Global Ecol. Biogeogr.* 22: 508–516.

Johnson, E. E., Escobar, L. E. and Zambrana-Torrelío, C. 2019. An ecological framework for modeling the geography of disease transmission. – *Trends Ecol. Evol.* 34: 655–668.

Joppa, L. N., McInerny, G., Harper, R., Salido, L., Takeda, K., O'Hara, K., Gavaghan, D. and Emmott, S. 2013. Troubling trends in scientific software use. – *Science* 340: 814–815.

Kass, J. M., Vilela, B., Aiello-Lammens, M. E., Muscarella, R., Merow, C. and Anderson, R. P. 2018. Wallace: a flexible platform for reproducible modeling of species niches and distributions built for community expansion. – *Methods Ecol. Evol.* 9: 1151–1156.

Kass, J. M., Anderson, R. P., Espinosa-Lucas, A., Juárez-Jaimes, V., Martínez-Salas, E., Botello, F., Tavera, G., Flores-Martínez, J. J. and Sánchez-Cordero, V. 2020. Biotic predictors with phenological information improve range estimates for migrating monarch butterflies in Mexico. – *Ecography* 43: 341–352.

Kass, J. M., Muscarella, R., Galante, P. J., Bohl, C. L., Pinilla-Buitrago, G. E., Boria, R. A., Soley-Guardia, M. and Anderson, R. P. 2021. ENMeval 2.0: redesigned for customizable and reproducible modeling of species' niches and distributions. – *Methods Ecol. Evol.* 12: 1602–1608.

Kass, J. M., Pinilla-Buitrago, G. E., Paz, A., Johnson, B. A., Grisales-Betancur, V., Meenan, S. I., Attali, D., Broennimann, O., Galante, P. J., Maitner, B. S., Owens, H. L., Varela, S., Aiello-Lammens, M. E., Merow, C., Blair, M. E. and Anderson, R. P. 2023. wallace 2: a shiny app for modeling species niches and distributions redesigned to facilitate expansion via module contributions. – *Ecography* 2023: e06547.

Kays, R. et al. 2022a. The Movebank system for studying global animal movement and demography. – *Methods Ecol. Evol.* 13: 419–431.

Kays, R., Lasky, M., Allen, M. L., Dowler, R. C., Hawkins, M. T. R., Hope, A. G., Kohli, B. A., Mathis, V. L., McLean, B., Olson, L. E., Thompson, C. W., Thornton, D., Widness, J. and Cove, M. V. 2022b. Which mammals can be identified from camera traps and crowdsourced photographs? – *J. Mammal.* 103: 767–775.

Kearney, M. R. and Porter, W. P. 2017. NicheMapR – an R package for biophysical modelling: the microclimate model. – *Ecography* 40: 664–674.

Kearney, M. R. and Porter, W. P. 2020. NicheMapR – an R package for biophysical modelling: the ectotherm and dynamic energy budget models. – *Ecography* 43: 85–96.

Leathwick, J. R., Elith, J. and Hastie, T. 2006. Comparative performance of generalized additive models and multivariate adaptive regression splines for statistical modelling of species distributions. – *Ecol. Modell.* 199: 188–196.

Leroy, B., Delsol, R., Hugueny, B., Meynard, C. N., Barhoumi, C., Barbet-Massin, M. and Bellard, C. 2018. Without quality pres-

ence-absence data, discrimination metrics such as TSS can be misleading measures of model performance. – *J. Biogeogr.* 45: 1994–2002.

Liang, W., Papeş, M., Tran, L., Grant, J., Washington-Allen, R., Stewart, S. and Wiggins, G. 2018. The effect of pseudo-absence selection method on transferability of species distribution models in the context of non-adaptive niche shift. – *Ecol. Modell.* 388: 1–9.

Lira-Noriega, A., Soberón, J. and Miller, C. P. 2013. Process-based and correlative modeling of desert mistletoe distribution: a multiscalar approach. – *Ecosphere* 4: 1–23.

Liu, C., White, M. and Newell, G. 2013. Selecting thresholds for the prediction of species occurrence with presence-only data. – *J. Biogeogr.* 40: 778–789.

Lobo, J. M., Jiménez-Valverde, A. and Real, R. 2008. AUC: a misleading measure of the performance of predictive distribution models. – *Global Ecol. Biogeogr.* 17: 145–151.

Lozier, J. D., Aniello, P. and Hickerson, M. J. 2009. Predicting the distribution of Sasquatch in western North America: anything goes with ecological niche modelling. – *J. Biogeogr.* 36: 1623–1627.

Maitner, B. S. et al. 2018. The BIEN R package: a tool to access the Botanical Information and Ecology Network (BIEN) database. – *Methods Ecol. Evol.* 9: 373–379.

Maldonado, C., Molina, C. I., Zizka, A., Persson, C., Taylor, C. M., Albán, J., Chilquillo, E., Rønsted, N. and Antonelli, A. 2015. Estimating species diversity and distribution in the era of big data: to what extent can we trust public databases? – *Global Ecol. Biogeogr.* 24: 973–984.

Mammola, S. and Isaia, M. 2017. Rapid poleward distributional shifts in the European cave-dwelling *Meta* spiders under the influence of competition dynamics. – *J. Biogeogr.* 44: 2789–2797.

Marmion, M., Parviainen, M., Luoto, M., Heikkinen, R. K. and Thuiller, W. 2009. Evaluation of consensus methods in predictive species distribution modelling. – *Divers. Distrib.* 15: 59–69.

Martin, Y., Van Dyck, H., Legendre, P., Settele, J., Schweiger, O., Harpke, A., Wiemers, M., Ameztegui, A. and Titeux, N. 2020. A novel tool to assess the effect of intraspecific spatial niche variation on species distribution shifts under climate change. – *Global Ecol. Biogeogr.* 29: 590–602.

Meller, L., Cabeza, M., Pironon, S., Barbet-Massin, M., Maiorano, L., Georges, D. and Thuiller, W. 2014. Ensemble distribution models in conservation prioritization: from consensus predictions to consensus reserve networks. – *Divers. Distrib.* 20: 309–321.

Merow, C. and Silander, J. A. 2014. A comparison of Maxlike and Maxent for modelling species distributions. – *Methods Ecol. Evol.* 5: 215–225.

Merow, C., Smith, M. J. and Silander, J. A. 2013. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. – *Ecography* 36: 1058–1069.

Merow, C., Smith, M. J., Edwards, T. C., Guisan, A., McMahon, S. M., Normand, S., Thuiller, W., Wüest, R. O., Zimmermann, N. E. and Elith, J. 2014. What do we gain from simplicity versus complexity in species distribution models? – *Ecography* 37: 1267–1281.

Merow, C., Allen, J. M., Aiello-Lammens, M. and Silander, J. A. 2016. Improving niche and range estimates with Maxent and point process models by integrating spatially explicit information. – *Global Ecol. Biogeogr.* 25: 1022–1036.

Merow, C., Galante, P. J., Kass, J. M., Aiello-Lammens, M. E., Babich Morrow, C. B., Gerstner, B. E., Grisales Betancur, V., Moore, A. C., Noguera-Urbano, E. A., Pinilla-Buitrago, G. E., Velásquez-Tibatá, J., Anderson, R. P. and Blair, M. E. 2022. Operationalizing expert knowledge in species' range estimates using diverse data types. – *Front. Biogeogr.* 14: e53589.

Mesgaran, M. B., Cousens, R. D. and Webber, B. L. 2014. Here be dragons: a tool for quantifying novelty due to covariate range and correlation change when projecting species distribution models. – *Divers. Distrib.* 20: 1147–1159.

Meynard, C. N., Leroy, B. and Kaplan, D. M. 2019. Testing methods in species distribution modelling using virtual species: what have we learnt and what are we missing? – *Ecography* 42: 2021–2036.

Miller, D. A. W., Pacifici, K., Sanderlin, J. S. and Reich, B. J. 2019. The recent past and promising future for data integration methods to estimate species' distributions. – *Methods Ecol. Evol.* 10: 22–37.

Mod, H. K., Scherrer, D., Luoto, M. and Guisan, A. 2016. What we use is not what we know: environmental predictors in plant distribution models. – *J. Veg. Sci.* 27: 1308–1322.

Monserrat, S., Boshoff, A. F. and Kerley, G. I. H. 2019. Accessibility maps as a tool to predict sampling bias in historical biodiversity occurrence records. – *Ecography* 42: 125–136.

Morales, N. S., Fernández, I. C. and Baca-González, V. 2017. MaxEnt's parameter configuration and small samples: are we paying attention to recommendations? A systematic review. – *PeerJ* 5: e3093.

Moran, E. V. and Ormond, R. A. 2015. Simulating the interacting effects of intraspecific variation, disturbance, and competition on climate-driven range shifts in trees. – *PLoS One* 10: e0142369.

Moreno-Amat, E., Mateo, R. G., Nieto-Lugilde, D., Morueta-Holme, N., Svenning, J. C. and García-Amorena, I. 2015. Impact of model complexity on cross-temporal transferability in Maxent species distribution models: an assessment using paleobotanical data. – *Ecol. Modell.* 312: 308–317.

Morente-López, J., Kass, J. M., Lara-Romero, C., Serra-Díaz, J. M., Soto-Correa, J. C., Anderson, R. P. and Iriondo, J. M. 2022. Linking ecological niche models and common garden experiments to predict phenotypic differentiation in stressful environments: assessing the adaptive value of marginal populations in an alpine plant. – *Global Change Biol.* 28: 4143–4162.

Naimi, B. and Araújo, M. B. 2016. sdm: a reproducible and extensible R platform for species distribution modelling. – *Ecography* 39: 368–375.

Naimi, B., Hamm, N. A. S., Groen, T. A., Skidmore, A. K. and Toxopeus, A. G. 2014. Where is positional uncertainty a problem for species distribution modelling? – *Ecography* 37: 191–203.

Newbold, T. 2010. Applications and limitations of museum data for conservation and ecology, with particular attention to species distribution models. – *Prog. Phys. Geogr.* 34: 3–22.

Norberg, A. et al. 2019. A comprehensive evaluation of predictive performance of 33 species distribution models at species and community levels. – *Ecol. Monogr.* 89: e01317.

Olden, J. D., Lawler, J. J. and Poff, N. L. 2008. Machine learning methods without tears: a primer for ecologists. – *Q. Rev. Biol.* 83: 171–193.

Owens, H. L., Campbell, L. P., Dornak, L. L., Saupe, E. E., Barve, N., Soberón, J., Ingenloff, K., Lira-Noriega, A., Hensz, C. M.,

Myers, C. E. and Peterson, A. T. 2013. Constraints on interpretation of ecological niche models by limited environmental ranges on calibration areas. – *Ecol. Modell.* 263: 10–18.

Peers, M. J. L., Thornton, D. H. and Murray, D. L. 2013. Evidence for large-scale effects of competition: niche displacement in Canada lynx and bobcat. – *Proc. R. Soc. B* 280: 20132495.

Pelini, S. L., Dzurisin, J. D. K., Prior, K. M., Williams, C. M., Marsico, T. D., Sinclair, B. J. and Hellmann, J. J. 2009. Translocation experiments with butterflies reveal limits to enhancement of poleward populations under climate change. – *Proc. Natl Acad. Sci. USA* 106: 11160–11165.

Perktaş, U., Peterson, A. T. and Dyer, D. 2017. Integrating morphology, phylogeography, and ecological niche modeling to explore population differentiation in North African common chaffinches. – *J. Ornithol.* 158: 1–13.

Peterson, A. T. 2011. Ecological niche conservatism: a time-structured review of evidence. – *J. Biogeogr.* 38: 817–827.

Peterson, A. T. and Holt, R. D. 2003. Niche differentiation in Mexican birds: using point occurrences to detect ecological innovation. – *Ecol. Lett.* 6: 774–782.

Peterson, A. T., Papeş, M. and Soberón, J. 2008. Rethinking receiver operating characteristic analysis applications in ecological niche modeling. – *Ecol. Modell.* 213: 63–72.

Peterson, A. T., Soberón, J., Pearson, R. G., Anderson, R. P., Martínez-Meyer, E., Nakamura, M. and Araújo, M. B. 2011. Ecological niches and geographic distributions (Monographs in Population Biology-49). – Princeton Univ. Press.

Peterson, A. T., Cobos, M. E. and Jiménez-García, D. 2018. Major challenges for correlational ecological niche model projections to future climate conditions. – *Ann. N. Y. Acad. Sci.* 1429: 66–77.

Petitpierre, B., Broennimann, O., Kueffer, C., Daehler, C. and Guisan, A. 2017. Selecting predictors to maximize the transferability of species distribution models: lessons from cross-continental plant invasions. – *Global Ecol. Biogeogr.* 26: 275–287.

Phillips, S. J., Anderson, R. P. and Schapire, R. E. 2006. Maximum entropy modeling of species geographic distributions. – *Ecol. Modell.* 190: 231–259.

Phillips, S. J. and Elith, J. 2013. On estimating probability of presence from use-availability or presence-background data. – *Ecology* 94: 1409–1419.

Phillips, S. J., Dudík, M., Elith, J., Graham, C. H., Lehmann, A., Leathwick, J. and Ferrier, S. 2009. Sample selection bias and presence-only distribution models: implications for background and pseudo-absence data. – *Ecol. Appl.* 19: 181–197.

Phillips, S. J., Anderson, R. P., Dudík, M., Schapire, R. E. and Blair, M. E. 2017. Opening the black box: an open-source release of Maxent. – *Ecography* 40: 887–893.

Pili, A. N., Tingley, R., Sy, E. Y., Diesmos, M. L. L. and Diesmos, A. C. 2020. Niche shifts and environmental non-equilibrium undermine the usefulness of ecological niche models for invasion risk assessments. – *Sci. Rep.* 10: 7972.

Poggianti, G., Münkemüller, T., Bystrova, D., Arbel, J., Clark, J. S. and Thuiller, W. 2021. On the interpretations of joint modeling in community ecology. – *Trends Ecol. Evol.* 36: 391–401.

Pollock, L. J., Tingley, R., Morris, W. K., Golding, N., O'Hara, R. B., Parris, K. M., Vesk, P. A. and McCarthy, M. A. 2014. Understanding co-occurrence by modelling species simultaneously with a joint species distribution model (JSDM). – *Methods Ecol. Evol.* 5: 397–406.

Prates, I., Xue, A. T., Brown, J. L., Alvarado-Serrano, D. F., Rodrigues, M. T., Hickerson, M. J. and Carnaval, A. C. 2016. Inferring responses to climate dynamics from historical demography in Neotropical forest lizards. – *Proc. Natl Acad. Sci. USA* 113: 7978–7985.

Qiao, H., Feng, X., Escobar, L. E., Peterson, A. T., Soberón, J., Zhu, G. and Papeş, M. 2019. An evaluation of transferability of ecological niche models. – *Ecography* 42: 521–534.

Radosavljevic, A. and Anderson, R. P. 2014. Making better Maxent models of species distributions: complexity, overfitting and evaluation. – *J. Biogeogr.* 41: 629–643.

Raes, N. and ter Steege, H. 2007. A null-model for significance testing of presence-only species distribution models. – *Ecography* 30: 727–736.

Ranc, N., Santini, L., Rondinini, C., Boitani, L., Poitevin, F., Angerbjörn, A. and Maiorano, L. 2017. Performance tradeoffs in target-group bias correction for species distribution models. – *Ecography* 40: 1076–1087.

Real, R., Barbosa, A. M. and Vargas, J. M. 2006. Obtaining environmental favourability functions from logistic regression. – *Environ. Ecol. Stat.* 13: 237–245.

Reid, B. N., Kass, J. M., Wollney, S., Jensen, E. L., Russello, M. A., Viola, E. M., Pantophlet, J., Iverson, J. B., Peery, M. Z., Raxworthy, C. J. and Naro-Maciel, E. 2019. Disentangling the genetic effects of refugial isolation and range expansion in a trans-continentially distributed species. – *Heredity* 122: 441–457.

Renner, I. W., Elith, J., Baddeley, A., Fithian, W., Hastie, T., Phillips, S. J., Popovic, G. and Warton, D. I. 2015. Point process models for presence-only analysis. – *Methods Ecol. Evol.* 6: 366–379.

Reside, A. E., Critchell, K., Crayn, D. M., Goosem, M., Goosem, S., Hoskin, C. J., Sydes, T., Vanderduys, E. P. and Pressey, R. L. 2019. Beyond the model: expert knowledge improves predictions of species' fates under climate change. – *Ecol. Appl.* 29: e01824.

Roberts, D. R., Bahn, V., Ciuti, S., Boyce, M. S., Elith, J., Guillera-Arroita, G., Hauenstein, S., Lahoz-Monfort, J. J., Schröder, B., Thuiller, W., Warton, D. I., Wintle, B. A., Hartig, F. and Dornmann, C. F. 2017. Cross-validation strategies for data with temporal, spatial, hierarchical, or phylogenetic structure. – *Ecography* 40: 913–929.

Robertson, T., Döring, M., Guralnick, R., Bloom, D., Wieczorek, J., Braak, K., Otegui, J., Russell, L. and Desmet, P. 2014. The GBIF integrated publishing toolkit: facilitating the efficient publishing of biodiversity data on the internet. – *PLoS One* 9: e102623.

Robertson, M. P., Visser, V. and Hui, C. 2016. Bioge: an R package for assessing and improving data quality of occurrence record datasets. – *Ecography* 39: 394–401.

Romero, D., Olivero, J., Márquez, A. L., Báez, J. C. and Real, R. 2014. Uncertainty in distribution forecasts caused by taxonomic ambiguity under climate change scenarios: a case study with two newt species in mainland Spain. – *J. Biogeogr.* 41: 111–121.

Sanín, C. and Anderson, R. P. 2018. A framework for simultaneous tests of abiotic, biotic, and historical drivers of species distributions: empirical tests for North American wood warblers based on climate and pollen. – *Am. Nat.* 192: E48–E61.

Saupe, E. E., Barve, V., Myers, C. E., Soberón, J., Barve, N., Hensz, C. M., Peterson, A. T., Owens, H. L. and Lira-Noriega, A. 2012. Variation in niche and distribution model performance: the need for a priori assessment of key causal factors. – *Ecol. Modell.* 237–238: 11–22.

Saupe, E. E., Barve, N., Owens, H. L., Cooper, J. C., Hosner, P. A. and Peterson, A. T. 2018. Reconstructing ecological niche evolution when niches are incompletely characterized. – *Syst. Biol.* 67: 428–438.

Serra-Diaz, J. M., Enquist, B. J., Maitner, B., Merow, C. and Svenning, J. C. 2017. Big data of tree species distributions: how big and how good? – *For. Ecosyst.* 4: 30.

Sillero, N. 2011. What does ecological modelling model? A proposed classification of ecological niche models based on their underlying methods. – *Ecol. Modell.* 222: 1343–1346.

Sillero, N. and Barbosa, A. M. 2021. Common mistakes in ecological niche models. – *Int. J. Geogr. Inf. Sci.* 35: 213–226.

Sillero, N., Arenas-Castro, S., Enriquez-Urzelai, U., Vale, C. G., Sousa-Guedes, D., Martínez-Freiría, F., Real, R. and Barbosa, A. M. 2021. Want to model a species niche? A step-by-step guideline on correlative ecological niche modelling. – *Ecol. Modell.* 456: 109671.

Smith, A. B., Godsoe, W., Rodríguez-Sánchez, F., Wang, H. H. and Warren, D. 2019. Niche estimation above and below the species level. – *Trends Ecol. Evol.* 34: 260–273.

Soberón, J. 2007. Grinnellian and Eltonian niches and geographic distributions of species. – *Ecol. Lett.* 10: 1115–1123.

Soberón, J. M. 2010. Niche and area of distribution modeling: a population ecology perspective. – *Ecography* 33: 159–167.

Soberón, J. and Nakamura, M. 2009. Niches and distributional areas: concepts, methods, and assumptions. – *Proc. Natl Acad. Sci. USA* 106: 19644–19650.

Sofaer, H. R., Jarnevich, C. S., Pearse, I. S., Smyth, R. L., Auer, S., Cook, G. L., Edwards, T. C., Guala, G. F., Howard, T. G., Morissette, J. T. and Hamilton, H. 2019. Development and delivery of species distribution models to inform decision-making. – *BioScience* 69: 544–557.

Soley-Guardia, M., Radosavljevic, A., Rivera, J. L. and Anderson, R. P. 2014. The effect of spatially marginal localities in modelling species niches and distributions. – *J. Biogeogr.* 41: 1390–1401.

Soley-Guardia, M., Gutiérrez, E. E., Thomas, D. M., Ochoa-G, J., Aguilera, M. and Anderson, R. P. 2016. Are we overestimating the niche? Removing marginal localities helps ecological niche models detect environmental barriers. – *Ecol. Evol.* 6: 1267–1279.

Soley-Guardia, M., Carnaval, A. C. and Anderson, R. P. 2019. Sufficient versus optimal climatic stability during the Late Quaternary: using environmental quality to guide phylogeographic inferences in a Neotropical montane system. – *J. Mammal.* 100: 1783–1807.

Stein, B. R. and Wieczorek, J. R. 2004. Mammals of the World: MaNIS as an example of data integration in a distributed network environment. – *Biodiv. Inform.* 1: 14–22.

Støa, B., Halvorsen, R., Mazzoni, S. and Gusarov, V. I. 2018. Sampling bias in presence-only data used for species distribution modelling: theory and methods for detecting sample bias and its effects on models. – *Sommerfeltia* 38: 1–53.

Syfert, M. M., Smith, M. J. and Coomes, D. A. 2013. The effects of sampling bias and model complexity on the predictive performance of MaxEnt species distribution models. – *PLoS One* 8: e55158.

Tarli, V. D., Grandcolas, P. and Pellens, R. 2018. The informative value of museum collections for ecology and conservation: a comparison with target sampling in the Brazilian Atlantic forest. – *PLoS One* 13: e0205710.

Taylor, A. T., Hafen, T., Holley, C. T., González, A. and Long, J. M. 2020. Spatial sampling bias and model complexity in stream-based species distribution models: a case study of paddlefish (*Polyodon spathula*) in the Arkansas River basin, USA. – *Ecol. Evol.* 10: 705–717.

Thorson, J. T., Ianelli, J. N., Larsen, E. A., Ries, L., Scheuerell, M. D., Szuwalski, C. and Zipkin, E. F. 2016. Joint dynamic species distribution models: a tool for community ordination and spatio-temporal monitoring. – *Global Ecol. Biogeogr.* 25: 1144–1158.

Thuiller, W., Lafourcade, B., Engler, R. and Araújo, M. B. 2009. BIOMOD – a platform for ensemble forecasting of species distributions. – *Ecography* 32: 369–373.

Title, P. O. and Bemmel, J. B. 2018. ENVIREM: an expanded set of bioclimatic and topographic variables increases flexibility and improves performance of ecological niche modeling. – *Ecography* 41: 291–307.

Tracy, J. L., Trabucco, A., Lawing, A. M., Giermakowski, J. T., Tchakerian, M., Drus, G. M. and Coulson, R. N. 2018. Random subset feature selection for ecological niche models of wildfire activity in western North America. – *Ecol. Modell.* 383: 52–68.

Tuia, D., Kellenberger, B., Beery, S., Costelloe, B. R., Zuffi, S., Risse, B., Mathis, A., Mathis, M. W., van Langevelde, F., Burghardt, T., Kays, R., Klinck, H., Wikelski, M., Couzin, I. D., van Horn, G., Crofoot, M. C., Stewart, C. V. and Berger-Wolf, T. 2022. Perspectives in machine learning for wildlife conservation. – *Nat. Commun.* 13: 792.

Valavi, R., Elith, J., Lahoz-Monfort, J. J. and Guillera-Arroita, G. 2019. blockCV: an R package for generating spatially or environmentally separated folds for k-fold cross-validation of species distribution models. – *Methods Ecol. Evol.* 10: 225–232.

Valavi, R., Elith, J., Lahoz-Monfort, J. J. and Guillera-Arroita, G. 2021. Modelling species presence-only data with random forests. – *Ecography* 44: 1731–1742.

Valavi, R., Guillera-Arroita, G., Lahoz-Monfort, J. J. and Elith, J. 2022. Predictive performance of presence-only species distribution models: a benchmark study with reproducible code. – *Ecol. Monogr.* 92: e01486.

van Boheemen, L. A., Atwater, D. Z. and Hodgins, K. A. 2019. Rapid and repeated local adaptation to climate in an invasive plant. – *New Phytol.* 222: 614–627.

VanDerWal, J., Shoo, L. P., Graham, C. and Williams, S. E. 2009. Selecting pseudo-absence data for presence-only distribution modeling: how far should you stray from what you know? – *Ecol. Modell.* 220: 589–594.

Varela, S., Anderson, R. P., García-Valdés, R. and Fernández-González, F. 2014. Environmental filters reduce the effects of sampling bias and improve predictions of ecological niche models. – *Ecography* 37: 1084–1091.

Velasco, J. A. and González-Salazar, C. 2019. Akaike information criterion should not be a 'test' of geographical prediction accuracy in ecological niche modelling. – *Ecol. Inform.* 51: 25–32.

Velásquez-Tibatá, J., Olaya-Rodríguez, M. H., López-Lozano, D., Gutiérrez, C., González, I. and Londoño-Murcia, M. C. 2019. BioModelos: a collaborative online system to map species distributions. – *PLoS One* 14: e0214522.

Veloz, S. D. 2009. Spatially autocorrelated sampling falsely inflates measures of accuracy for presence-only niche models. – *J. Biogeogr.* 36: 2290–2299.

Vollering, J., Halvorsen, R., Auestad, I. and Rydgren, K. 2019. Bunching up the background betters bias in species distribution models. – *Ecography* 42: 1717–1727.

Warren, D. L. and Seifert, S. N. 2011. Ecological niche modeling in Maxent: the importance of model complexity and the performance of model selection criteria. – *Ecol. Appl.* 21: 335–342.

Warren, D. L., Wright, A. N., Seifert, S. N. and Shaffer, H. B. 2014. Incorporating model complexity and spatial sampling bias into ecological niche models of climate change risks faced by 90 California vertebrate species of concern. – *Divers. Distrib.* 20: 334–343.

Warren, D. L., Matzke, N. J. and Iglesias, T. L. 2020. Evaluating presence-only species distribution models with discrimination accuracy is uninformative for many applications. – *J. Biogeogr.* 47: 167–180.

Warton, D. I., Blanchet, F. G., O'Hara, R. B., Ovaskainen, O., Taskinen, S., Walker, S. C. and Hui, F. K. C. 2015. So many variables: joint modeling in community ecology. – *Trends Ecol. Evol.* 30: 766–779.

Wenger, S. J. and Olden, J. D. 2012. Assessing transferability of ecological models: an underappreciated aspect of statistical validation. – *Methods Ecol. Evol.* 3: 260–267.

Williams, J. W. and Jackson, S. T. 2007. Novel climates, no-analog communities, and ecological surprises. – *Front. Ecol. Environ.* 5: 475–482.

Willis, S. G., Foden, W., Baker, D. J., Belle, E., Burgess, N. D., Carr, J. A., Doswald, N., Garcia, R. A., Hartley, A., Hof, C., Newbold, T., Rahbek, C., Smith, R. J., Visconti, P., Young, B. E. and Butchart, S. H. M. 2015. Integrating climate change vulnerability assessments from species distribution models and trait-based approaches. – *Biol. Conserv.* 190: 167–178.

Wisz, M. S. et al. 2013. The role of biotic interactions in shaping distributions and realised assemblages of species: implications for species distribution modelling. – *Biol. Rev.* 88: 15–30.

Yackulic, C. B., Chandler, R., Zipkin, E. F., Royle, J. A., Nichols, J. D., Campbell Grant, E. H. and Veran, S. 2013. Presence-only modelling using MAXENT: when can we trust the inferences? – *Methods Ecol. Evol.* 4: 236–243.

Yates, K. L. et al. 2018. Outstanding challenges in the transferability of ecological models. – *Trends Ecol. Evol.* 33: 790–802.

Zizka, A., Silvestro, D., Andermann, T., Azevedo, J., Duarte Ritter, C., Edler, D., Farooq, H., Herdean, A., Ariza, M., Scharn, R., Svantesson, S., Wengström, N., Zizka, V. and Antonelli, A. 2019. CoordinateCleaner: standardized cleaning of occurrence records from biological collection databases. – *Methods Ecol. Evol.* 10: 744–751.

Zurell, D. 2017. Integrating demography, dispersal and interspecific interactions into bird distribution models. – *J. Avian Biol.* 48: 1505–1516.

Zurell, D., Elith, J. and Schröder, B. 2012. Predicting to new environments: tools for visualizing model behaviour and impacts on mapped distributions. – *Divers. Distrib.* 18: 628–634.

Zurell, D., Zimmermann, N. E., Gross, H., Baltensweiler, A., Satler, T. and Wüest, R. O. 2020a. Testing species assemblage predictions from stacked and joint species distribution models. – *J. Biogeogr.* 47: 101–113.

Zurell, D. et al. 2020b. A standard protocol for reporting species distribution models. – *Ecography* 43: 1261–1277.