

Research



Cite this article: Alaasam VJ, Behnke TL, Grant AR, Ouyang JQ. 2024 Glucocorticoids and land cover: a largescale comparative approach to assess a physiological biomarker for avian conservation. *Phil. Trans. R. Soc. B* **379**: 20220508.
<https://doi.org/10.1098/rstb.2022.0508>

Received: 3 July 2023

Accepted: 19 October 2023

One contribution of 14 to a theme issue 'Endocrine responses to environmental variation: conceptual approaches and recent developments'.

Subject Areas:

physiology

Keywords:

corticosterone, birds, HormoneBase, habitat, National Land Cover Database

Author for correspondence:

Valentina J. Alaasam

e-mail: valentina@nevada.unr.edu

[†]Shared first authorship.

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.6996932>.

Glucocorticoids and land cover: a largescale comparative approach to assess a physiological biomarker for avian conservation

Valentina J. Alaasam^{1,†}, Tessa L. Behnke^{2,†}, Avery R. Grant¹ and Jenny Q. Ouyang¹

¹Department of Biology, and ²Department of Natural Resources and Environmental Science, University of Nevada, Reno, NV 89557, USA

VJA, 0000-0003-1670-6780

As humans alter landscapes worldwide, land and wildlife managers need reliable tools to assess and monitor responses of wildlife populations. Glucocorticoid (GC) hormone levels are one common physiological metric used to quantify how populations are coping in the context of their environments. Understanding whether GC levels can reflect broad landscape characteristics, using data that are free and commonplace to diverse stakeholders, is an important step towards physiological biomarkers having practical application in management and conservation. We conducted a phylogenetic comparative analysis using publicly available datasets to test the efficacy of GCs as a biomarker for large spatial-scale avian population monitoring. We used hormone data from HormoneBase (51 species), natural history information and US national land cover data to determine if baseline or stress-induced corticosterone varies with the amount of usable land cover types within each species' home range. We found that stress-induced levels, but not baseline, positively correlated with per cent usable land cover both within and across species. Our results indicate that GC concentrations may be a useful biomarker for characterizing populations across a range of habitat availability, and we advocate for more physiological studies on non-traditional species in less studied populations to build on this framework.

This article is part of the theme issue 'Endocrine responses to environmental variation: conceptual approaches and recent developments'.

1. Introduction

Habitat loss is one of the greatest threats to biodiversity, threatening 85% of all species on the IUCN's Red List [1]. Degradation of natural habitats has been caused by expansion of agricultural land, timber harvesting, overgrazing and other anthropogenic development in response to a growing human population, resulting in half of the world's non-glacial land being converted for human use [2,3]. Reduction in habitat quality or availability can cause rapid and lasting physiological responses in organisms. In particular, it is well documented that habitat alteration can cause physiological responses to stress [4–9], which can lead to decreased reproductive success, pathology and increased mortality rates in vertebrates [10,11]. As a result, a surge of research in the recent past decades has focused on refining the use of physiological metrics to assess animal responses to environmental change and inform conservation and management decisions [12–15].

Glucocorticoid hormones (GCs) have long been regarded as a candidate biomarker of physiological stress across vertebrates [16,17] and a valuable tool in conservation physiology [18]. This is due to their well-conserved role in mediating vertebrate behaviour and physiology in response to both long-term environmental changes and acute perturbations [19]. GCs are released from

the adrenal gland via the hypothalamic–pituitary–adrenal (HPA) axis and regulate blood glucose levels and mobilize energy reserves. At baseline levels, GCs fluctuate daily and seasonally, reflecting the cumulative energetic demands required to maintain internal homeostasis across changing conditions [20]. For example, baseline GCs can vary during energetically demanding life-history stages such as breeding and migration [21,22], weather events [23,24] or periods of resource limitation [25]. In response to acute or unpredicted challenges, GCs rise quickly and promote immediate survival, e.g. by facilitating predator escape behaviour, territory defense behaviour or mounting of an immune response [26,27]. A fast and robust GC stress response has traditionally been considered critical to an individual's ability to maintain homeostasis through changing environmental conditions (i.e. allostasis) [19,28]. However, absolute levels of GCs and the extent to which they mediate responses to an individual's environment are highly context-dependent [17,29]. Both baseline and stress-induced GC levels can vary across life-history stage [30–33], age [34], sex [35] and species [36,37]. Whether an environmental challenge elicits a GC response, and whether the response is appropriate or harmful, also depends on the nature, frequency and context of the challenge [24,29]. Moreover, recent studies assert that proper interpretation of GC measures requires some knowledge of other components of the HPA axis (e.g. endocrine receptors and binding agents, or the efficiency of recovery from maximum levels) [29,38,39].

Despite this variability of GCs, large-scale comparative studies have been successful at detecting broad trends in GC profiles within and across species [17,40]. Indeed, while empirical studies are vital to our biological understanding of GCs, complementary meta-analyses provide support for using GCs as a physiological indicator of organismal response to environmental variation. For example, several reviews in the past decade have linked GC variation to latitude [41,42], ambient temperature [43,44], primary productivity [42] and food availability [45], suggesting that similar GC phenotypes may be selected for across species in similar environments. As a result, GCs have become one of the most common physiological biomarkers measured in the context of conservation [17,18]. However, there is a deficit in our understanding of how GCs relate to broad landscape characteristics, specifically those used by land and wildlife managers to remotely identify suitable habitat for species. Advancements in global satellite imaging, now able to accurately classify land cover types remotely [46], have become a widespread and common practice for assessing landscape and habitat characteristics [47–49]. Although they are still relatively crude estimates of habitat composition and overall quality compared to on-the-ground surveys [46,50], medium resolution satellite data offer a cost-effective way to assess land cover on very large scales and are accessible to organizations with limited time or resources [14,51,52]. As GCs have potential to be a valuable tool in conservation physiology across taxa [17,18,53], bridging the rich GC literature with broadscale, open-access data sources is of great value to ongoing management initiatives and to broadening global participation in conservation.

In this study, we asked whether broad trends in GC profiles exist across avian species in relation to landscape characteristics, using free, publicly available land cover data and natural history databases. We gathered GC data from HormoneBase, a comprehensive database containing records of GCs collected from adult, free-living vertebrates over the past six

decades [54]. Since its publication, HormoneBase has facilitated the investigation of many similar comparative physiological questions at broad spatial and temporal scales, such as relating GCs to life-history traits [55], metabolic rate [56], energetic costs of thermoregulation [57] and conservation status [58], but no studies have assessed the utility of GCs in predicting organismal responses to available land cover types. We focused our study on birds due to their broad distributions, utilization of diverse habitats, extensively studied natural history and well-documented variation in GC (corticosterone in birds) concentrations across life-history stages. We hypothesized that if GC levels reflect the status of an organism's interaction with its surrounding environment, then populations will differ in their GC characteristics along a gradient of appropriate landscape type availability. Specifically, we predicted that populations inhabiting areas characterized by more usable land cover types may have lower baseline GC levels and a robust stress-induced response. Alternatively, baseline levels could be higher if populations inhabiting more suitable habitat exist at greater densities and experience more intraspecific competition, as predicted by ideal free distribution [59].

2. Material and methods

(a) Data extraction

We gathered data on plasma GC concentration, species natural history and land cover from three databases for our analysis. These databases include (1) HormoneBase, a database containing extensive published and unpublished records of circulating androgens and GCs collected from free-living adult vertebrates [54]; (2) Birds of North America (BNA), a Cornell Lab of Ornithology database providing detailed species accounts of North American bird biology written by recognized experts [60], and (3) the National Land Cover Database (NLCD) [61–64], a US Geological Survey database characterizing the US land surface at a medium resolution of 30 m².

We used HormoneBase to obtain records of plasma corticosterone (cort) sampled from free-living adult bird populations in the USA from 1969 through 2015. The HormoneBase dataset was compiled via searches through online academic databases for published (and some unpublished) data of free-living, unmanipulated, adult populations where plasma hormone levels were measured (see [46] for a detailed description of HormoneBase inclusion criteria). We limited our study to the USA due to the availability of high-resolution land cover data for the entire country (i.e. Landsat products). We initially selected all records that included baseline circulating GC measurements. Because the goal of this study was to test for habitat effects, we limited our dataset to records with accurate reported coordinates and excluded records with estimated study locations. To account for sampling variance across studies, we also limited our dataset to studies with reported standard errors. These criteria resulted in the inclusion of 281 records of 51 species collected between 1982 and 2012. A subset of these records included measurements of both baseline and stress-induced cort concentrations, but the presence of a stress-induced cort measurement was not required for inclusion in our dataset. Stress-induced cort values were available in 216 records, representing 44 species. From HormoneBase, we also noted whether records were obtained during 'breeding' or 'non-breeding' periods to account for variance in life-history stage. Although more specific information about the timing within the annual cycle (e.g. moulting, migration) was not consistently available, we assumed breeding periods to be separate from migration.

We obtained species' diet and average adult mass data from BNA because these variables have been shown to play a role in

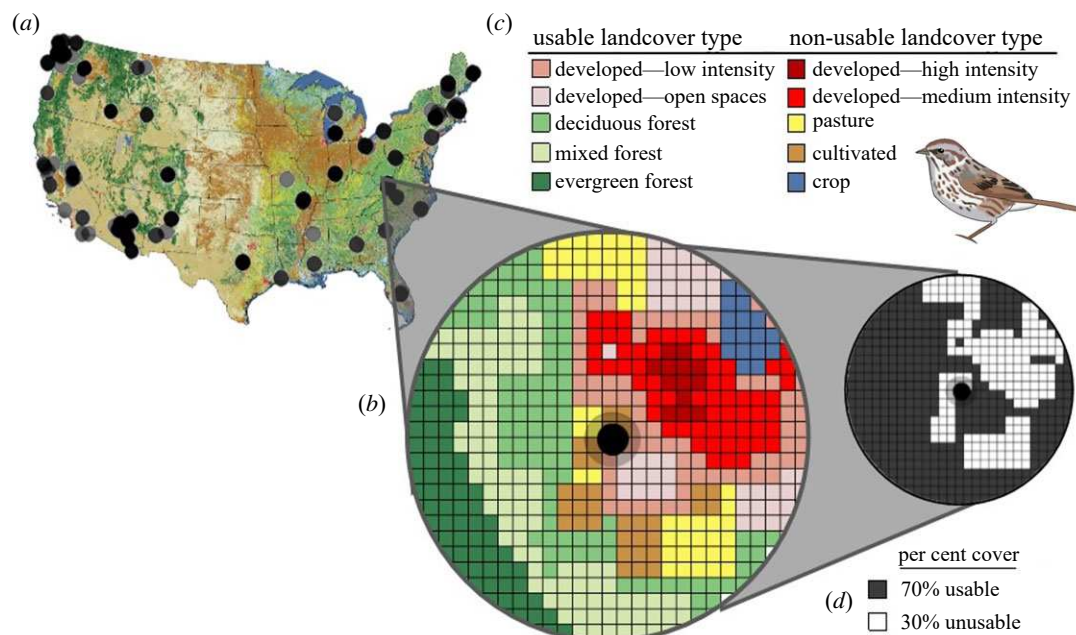


Figure 1. Methods for extracting per cent availability of appropriate landcover cover using the National Land Cover Database (NLCD) and Birds of North America (BNA) natural history information, demonstrated using a song sparrow (*Melospiza melodia*). For each HormoneBase record, we assigned a binned home range area (less than 1, 1–10, 10–100, greater than 100 ha). Centring each home range area on the coordinates of the HormoneBase cort sample (a), we layered each home range area on the NLCD raster (b). Using natural history information from the Birds of North America database (c), we extracted per cent cover of usable habitat for each HormoneBase record in our study based on each record's binned home range area (d). Map courtesy of NLCD. (Online version in colour.)

the variation of GC levels, as well as in responses to anthropogenic change [65,66]. We selected the species' diet (carnivore, herbivore or omnivore) and average mass from BNA that most closely matched the sex, life-history stage (i.e. breeding or non-breeding), geographical location and time of sample collection for each HormoneBase record. For example, for widespread species with different specific diets across its range, we selected the diet representative of populations in the same region or same ecosystem type of the HormoneBase record. Similarly, when diet was reported to vary seasonally or across life-history stages, we selected a relevant time of sample collection as the same month, season, or life-history stage. See electronic supplementary material, table S1 for all designations.

We used BNA to identify each species' approximate home range and the types of land cover that each species is reported to use (figure 1). Again, we extracted the most relevant information corresponding to the sex, life-history stage, geographical location and season of each HormoneBase record. For example, if a HormoneBase record was obtained during breeding, we extracted the home range and usable land types as specified on BNA for breeding populations specifically. In the absence of a reported home range, we used reported distances travelled from the nest, defining this distance as the home range radius. We categorized appropriate land cover types for each species using the 18 land surface classes defined in the NLCD [62]. Habitat classes include forested (coniferous, deciduous, mixed), wetland (woody wetlands, herbaceous wetlands), short vegetated (shrub/scrub, grassland/herbaceous), northern latitude-associated (dwarf scrub, sedge/herbaceous), unvegetated (open water, perennial snow and ice, barren land) and developed habitats (developed open spaces; low-, medium- and high-intensity development; cultivated crops; hay/pasture; figure 1). For species that use edge habitats, we included land cover categories for both types of land cover forming the edge.

We then used the 'raster' package [67] in R v.3.4.3 [68] to extract the percentage of cover of each NLCD land cover class within a species' home range, centred on the geographical coordinate of each HormoneBase record (figure 1). We pooled home ranges into four bins (1 ha, 10 ha, 100 ha and 1000 ha), resulting in similar numbers of species in each bin (1 ha = 10 species,

10 ha = 26 species, 100 ha = 8 species, 1000 ha = 9 species). Because NLCD produced land cover datasets in 1992, 2001, 2006 and 2011 and land cover can change over time, we extracted land cover data from the NLCD dataset closest in year to each HormoneBase record. We then calculated the percentage of usable land cover available by summing the per cent cover of land classes that species have been reported to use (figure 1).

(b) Analyses

All analyses were conducted in R v.3.5.2 [68]. We used phylogenetic generalized linear mixed models (PGLMM) to assess the impact of life history and per cent usable land cover on cort concentrations [55,69]. We used the package 'MCMCglmm' and incorporated phylogeny using the 'pedigree' argument within the 'MCMCglmm' function [69]. For our phylogeny, we constructed a consensus tree using 1000 Hackett *et al.* [70] backbone trees downloaded from BirdTree.org [71]. Specifically, we built a 50% majority-rule consensus tree [72] using the Python program 'SumTrees' in 'DendroPy' [73] following the methods of Rubolini *et al.* [74]. We trimmed trees to include species in the baseline and stress-induced data subsets using the R package 'APE' [75].

We created separate models for baseline and stress-induced cort. We applied a $\log_{10}$ transformation to both baseline and stress-induced cort population means to improve fit to a normal distribution. Our fixed effects included latitude, sex, mean mass ($\log_{10}$ -transformed), home range size, diet, life-history stage (breeding or non-breeding), per cent usable land cover type, and the times at which baseline and stress-induced cort were measured post-capture [76]. Most (92%) of baseline levels were collected < 3 min, and all were collected within 7.5 min, post-capture. Similarly, most (80%) of stress-induced levels were collected at 30 ± 5 min, with only eight records collected in less than 25 min and all collected within 60 min post-capture. We included species as a random effect, as well as laboratory ID to control for inter-laboratory variation [77]. We included standard error reported with each record as a random effect, with parameter-expanded priors to control for potential measurement biases. We defined non-informative priors for both R and G as inverse-Wishart distributions



Figure 2. Phylogeny of species included in our analyses. We selected records from HormoneBase that reported mean population baseline cort, standard error and study site geographical coordinates for bird populations in North America (281 records of 51 species). Records that lacked stress-induced cort values are noted with asterisks. Within-species analyses were conducted for five species with more than 2 population records (denoted by +).

(expected variance, $V = 1$; degree of belief, $\nu = 0.002$). For all models, we ran three chains of 200 000 iterations, with a burn-in of 50 000 and thinning interval of 100. We diagnosed convergence visually and with Gelman-Rubin statistics [78].

For species with multiple (more than 2) populations represented in HormoneBase, we also ran individual within-species linear models. These species were the house sparrow (*Passer domesticus*), semi-palmated sandpiper (*Calidris pusilla*), dark-eyed junco (*Junco hyemalis*), song sparrow (*Melospiza melodia*) and white-crowned sparrow (*Zonotrichia leucophrys*). For each species, we ran separate models for baseline and stress-induced cort, including sex, life-history stage (breeding or non-breeding) and per cent usable land cover type as fixed effects. All models met assumptions of a normal distribution. These models are redundant with the main model (which included species as a random effect) but were included to demonstrate

the variability in trends across species, and to provide an example of how our methods can be applied to a single focal species (figure 2).

3. Results

(a) Baseline cort

Baseline cort correlated with sex and species mean mass, but not with per cent usable land cover type. Males had higher baseline cort compared to females, with the 95% credible interval not overlapping zero (table 1). Baseline cort decreased with increasing mean mass, with the 95% credible interval not overlapping zero (table 1). There was no support for an effect of latitude, diet type, home range size, life-history stage, late sample time

Table 1. Results from the PGLMMs for baseline and stress-induced corticosterone models. In the p_{MCMC} columns, values in bold are significant for the 95% credible interval (CI); those in parentheses are significant for the 90% credible interval.

	baseline				stress-induced			
	post. mean	lower 95% CI	upper 95% CI	p_{MCMC}	post. mean	lower 95% CI	upper 95% CI	p_{MCMC}
usable landcover (%)	0.102	−0.034	0.245	0.171	0.144	0.048	0.253	0.004
latitude	0.003	−0.003	0.008	0.345	<0.001	−0.004	0.004	0.845
late cort (sample time)	0.028	−0.002	0.006	0.073	0.002	−0.003	0.007	0.351
diet (carnivore)	0.089	−0.211	0.392	0.524	0.145	−0.039	0.347	0.143
diet (herbivore)	0.015	−0.231	0.303	0.913	−0.075	−0.304	0.137	0.467
breeding cycle (non-breeding)	−0.056	−0.127	0.023	0.156	−0.112	−0.173	−0.035	<0.001
mass	−0.434	−0.872	−0.011	0.044	−0.204	−0.458	0.040	(0.103)
sex (male)	0.056	−0.003	0.110	0.045	0.067	0.023	0.110	0.005
home range size	< 0.001	< −0.001	< 0.001	0.544	< 0.001	< −0.001	< 0.001	0.966

or per cent usable land cover type (table 1). The random effect of species, which incorporates the underlying phylogeny, was important (posterior mean = 0.437, lower credible interval (LCI) = 0.149, upper credible interval (UCI) = 0.790), whereas laboratory identity (posterior mean = 0.062, LCI = 0.006, UCI = 0.140) explained a much smaller proportion of the variance and the effect of study standard error was negligible (posterior mean = 0.0005, LCI = 0.0001, UCI = 0.001).

(b) Stress-induced cort

Stress-induced cort significantly correlated with per cent usable land cover type, sex, life-history stage and species mean mass. Stress-induced cort levels increased with increasing percentage of usable habitat, with the 95% credible interval not overlapping zero (table 1). Males had higher average stress-induced cort than females, with the 95% credible interval not overlapping zero (table 1). Breeding populations had higher stress-induced cort levels than non-breeding, with the 95% credible intervals not overlapping zero (table 1). There was also support for a negative effect of mean mass, with the 90% credible intervals not overlapping zero (table 1). There was no support for an effect of latitude, late sample time, diet type or home range size. The random effect of species was important (posterior mean = 0.104, 95% LCI = 0.036, UCI = 0.183), though less so than in the baseline cort model. Again, laboratory identity explained much less of the variance (posterior mean = 0.017, 95% LCI = 0.0003, UCI = 0.048) and the effect of study standard error was negligible (posterior mean = 0.0001, 95% LCI = 0.00008, UCI = 0.0002).

(c) Within-species relationships between cort and per cent usable land cover type

Within-species analysis of five taxa all showed similar trending relationships between cort and per cent usable land cover type as the PGLMM. There was no relationship between baseline cort and land cover availability for any species, and the direction of the slopes differed among species (table 2). For stress-induced cort, all species showed positive trending relationships

with per cent usable land cover type (figure 3), although significance level and effect sizes varied (table 2). House sparrow ($n = 25$), semi-palmated sandpiper ($n = 6$) and dark-eyed junco ($n = 5$) models showed significant effects of per cent usable land cover on stress-induced cort, while song sparrow ($n = 11$) and white-crowned sparrow ($n = 22$) models did not (table 2).

4. Discussion

As landscape alteration and habitat loss increase globally, land managers and conservation practitioners need reliable tools to assess and monitor wildlife populations. Within-individual physiological responses can scale up to population-level responses and have great potential to increase the effectiveness of conservation decisions [40,79,80], yet identifying practical physiological biomarkers is challenging. We sought to evaluate the efficacy of GCs as a physiological biomarker that may help assess how populations are coping with their surrounding environment. Specifically, we tested whether the availability of appropriate landcover types within a population's generalized home range correlates with GC profiles, across 51 species of birds. We intentionally used free, accessible environmental data from the National Land Cover Database, and life-history information from the Birds of North America dataset, to generate insights that are relevant and accessible to any land and wildlife manager. We found that stress-induced cort, but not baseline cort, correlated with differences in usable habitat, where populations surrounded by higher percentages of usable habitat had higher stress-induced cort levels. We discuss our results in light of what is known about avian GC responses and recommend how this framework may be applied to conservation in the future.

As in other comparative studies, sex and mass explained variance in both baseline and stress-induced cort levels across species. Males had higher baseline and stress-induced cort than females [55,81], and larger species (by average mass) had lower baseline and stress-induced levels of cort, consistent with other avian comparative analyses [28,66,82]. Here, mass represents a mean per sex per species because

Table 2. Within-species linear models testing the effect of per cent useable landcover type on baseline and stress-induced corticosterone levels. Corticosterone values were obtained from HormoneBase and filtered for records that with accurate GPS coordinates and standard errors recorded. Species were selected for individual analysis if records from more than 2 locations existed in HormoneBase. Models include life-history stage, sex and per cent useable landcover type, except for the semi-palmated sandpiper and song sparrow. All stress-induced corticosterone records of the song sparrow came from males during breeding, so sex and life-history stage were excluded from this model. All records of the semi-palmated sandpiper came from males during breeding, so sex and life-history stage were excluded from both models. Bold values indicate $p < 0.05$.

	baseline				stress-induced			
	estimate	s.e.	t	p-value	estimate	s.e.	t	p-value
<i>dark-eyed junco</i>								
usable landcover (%)	−0.35	0.47	−0.74	0.487	−0.25	0.002	−122.94	0.005
breeding cycle (non-breeding)	−0.54	0.23	−2.30	0.061	−0.60	0.001	−535.77	0.001
sex	−0.15	0.24	−0.61	0.562	−0.08	0.001	−82.96	0.007
<i>house sparrow</i>								
usable landcover (%)	−1.77	1.01	−1.76	0.122	0.26	0.089	2.91	0.008
breeding cycle (non-breeding)	0.24	0.21	1.18	0.278	−0.05	0.061	−0.83	0.414
sex	0.23	0.21	1.18	0.122	0.03	0.064	0.53	0.603
<i>song sparrow</i>								
usable landcover (%)	0.21	0.16	1.4	0.202	0.09	0.118	0.81	0.440
breeding cycle (non-breeding)	−0.30	0.13	−2.3	0.040	—	—	—	—
sex	0.18	0.12	1.48	0.165	—	—	—	—
<i>white-crowned sparrow</i>								
usable landcover (%)	−0.15	0.21	−0.71	0.486	0.16	0.194	0.82	0.425
breeding cycle (non-breeding)	−0.34	0.26	−1.28	0.212	−0.19	0.220	−0.86	0.403
sex	0.17	0.13	1.40	0.176	0.20	0.098	2.05	0.056
<i>semi-palmated sandpiper</i>								
usable landcover (%)	−0.14	0.24	−0.57	0.600	3.97	0.524	7.58	0.002

we did not have information about variation in mass across specific HormoneBase records. Thus, the relationship between GCs and mass that we detected likely indicates an evolutionary life-history phenomenon, rather than a reflection of individual populations being smaller or of poor body condition. It is also important to note that one other study assessing avian data (separate from HormoneBase) found that mass was not an important predictor of GCs when accounting for latitude [42], which also covaries with body size among species, but we did not find the same result here.

We found that stress-induced cort was higher for breeding populations. This is contrary to some previous avian empirical studies and reviews that show stress-induced cort is lowered during breeding [66,83]. Higher levels of stress-induced cort are sometimes associated with reduced breeding success [84,85]. However, cort phenotypes also vary temporally within the breeding season [22,55] and depend on brood value [66], which can reflect a combination of many variables such as relative clutch size, age, mate choice etc. It could be that the coarse dichotomy of ‘breeding’ and ‘non-breeding’ available on HormoneBase does not capture the complexity of circannual cort variation. There is not a consistent correlation between breeding and stress-induced cort in birds according to the literature, to date [55,82,86].

No correlation was found between baseline cort levels and per cent useable land cover type, in agreement with

several other meta-analyses that suggest baseline cort may not be a reliable biomarker on broad comparative scales [22,30,45]. This is likely because individual variation in baseline cort is high, due to its role in metabolism, energy expenditure and reflection of daily and seasonally fluctuating conditions (e.g. weather or breeding/migration status, respectively [23,87,88]). Our results support other recent literature advocating for caution when interpreting single measurements of baseline GCs [38,89]. Measurements of baseline circulating cort may be a valuable metric for individual health on a case-by-case basis if investigators take repeated measurements and incorporate species biology into sampling design [45,90–92].

We did find a relationship between population-level stress-induced cort profiles and per cent useable land cover type. Populations living in areas with more useable land cover had higher stress-induced plasma cort levels. In contrast to baseline levels, stress-induced cort levels represent an acute physiological response that is relatively more repeatable over an individual's lifetime [93–96] and is heritable [97–99]. The positive relationship between stress-induced cort and land cover availability is consistent with the traditional view that a robust GC stress response is an adaptive mechanism critical to the maintenance of homeostasis and may be indicative of population health [11,100–102]. However, it is becoming increasingly evident that the benefits of a robust response may depend on the efficacy of recovery

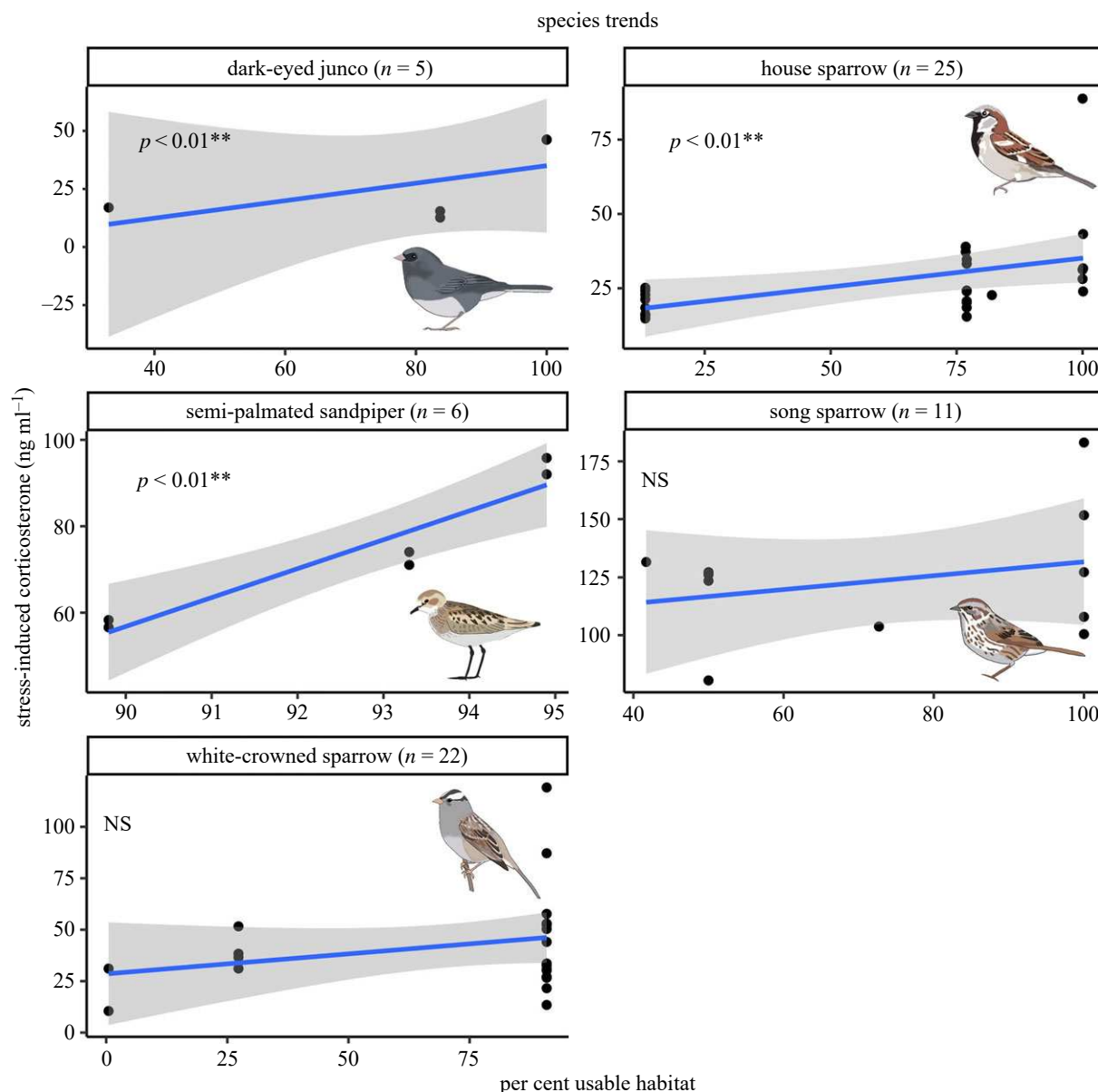


Figure 3. Individual species trends for the relationship between stress-induced corticosterone and per cent availability of appropriate landcover; p -values indicate significance based on linear models. Positive trends within species models support findings from interspecific models. (Online version in colour.)

to baseline levels, i.e. the rate at which individual GC levels return to baseline after a disturbance [39,44].

While our results show that land use can affect physiology on a population level, the magnitude and direction of the effect should be carefully interpreted on a case-by-case basis. For example, the positive relationship between stress-induced cort levels and per cent usable land cover could reflect a dampened stress response in populations inhabiting sub-optimal, habitat-limited areas. There are several possible mechanisms that explain a reduced stress response when habitat availability is reduced. First, dampening of the stress response may be adaptive for populations in poor-quality environments that invest more in reproduction than immediate survival, because their likelihood of surviving to the next breeding season is relatively low [66]. Second, populations surrounded by lower amounts of usable land cover types could exhibit lower stress-induced cort as a result of habituation or maternal effects, as might be expected in a chronically stressful environment [103]. For example, nutritionally restricted maternal environments can lead to decreased responsiveness of the hypothalamic-pituitary-adrenal axis in fledglings [104],

leading to long-term changes in populations. Decreased GC responsiveness may not be maladaptive, as some populations in urban environments exhibit dampened stress responses while survivorship remains high [105–107], which is important to consider when interpreting GC variation at the population level. Finally, areas with less usable habitat might be disproportionately occupied by subordinate or younger/inexperienced individuals [108], with more dominant/older individuals controlling more optimal habitats. Results may reflect differences in population structure that can vary across temporal and spatial scales and that we are unable to deduce from the HormoneBase database (although all samples here represent adult populations).

It is also important to acknowledge that the trend we detected might be driven by species' land-use specificity. We classified surrounding usable land cover types based on information about the habitat types used by each species. So, species that are more generalist in regard to habitat have, by default, more 'options' of usable land cover types. As a result, more generalist species should show a higher probability of being surrounded by a high percentage of

suitable land-cover types, regardless of the specific mosaic of land-cover classifications.

5. Future directions for research

Overall, we interpret the significant positive relationship between appropriate land cover availability and stress-induced cort levels as notable, especially given the resolution of our land cover data and inclusivity of our criteria for identifying usable habitat for a population. Using BNA life-history information, we identified land cover types as 'usable' for a population if the species had any record of inhabiting the type of land cover at the appropriate time of year/life-history stage, without attempting to estimate differences in the quality or preference, likely overestimating the area that a population could realistically exploit. Still, examining the limitations of our study provides an opportunity to recommend some ways in which researchers can increase the utility of GC measures in conservation and management in the future. First, while HormoneBase offers the most comprehensive hormone database in existence to date, data were strongly biased toward passerine bird species and some species lack records from multiple populations or are limited to data from only one sex or life-history stage. Even for the five species highlighted here with records for more than 3 populations, there were sometimes only one or two representative populations per sex, per life-history stage (see results for song sparrow and semi-palmated sandpiper), limiting the power of these analyses. More empirical data are needed to test whether our findings are biologically meaningful for all taxa and to examine interactive effects (e.g. whether the effect of surrounding land cover varies between age classes or between breeding and non-breeding individuals of a population). Assessing interactions could be important for managers to interpret variation within and across populations and identify particularly vulnerable groups. Additionally, many of the GPS points recorded for populations in HormoneBase are approximate, (e.g. representing the research station rather than exact capture points). Remote sensing technology and satellite data are currently advancing at an exciting rate. As high-resolution land cover data becomes more accurate and available, efforts to record precise GPS coordinates will be especially worthwhile, allowing future studies to consider the existence of microrefugia within an otherwise unusable landscape, or degree of connectivity with other usable habitat areas.

6. Conservation application

What application do GCs have in management and conservation? We show that, despite the known individual variability and complexity of the GC response, access to suitable land cover types can affect GC phenotypes at the population level. This is an important contribution, demonstrating a utility for GCs at the scales that managers are most concerned with (populations) and a framework for predicting how changes in land-use will affect populations. For example, it may be possible to calculate a threshold of per cent usable land cover at which populations will have a physiological response when making decisions about converting landscapes (e.g. for agriculture, urbanization, or restoration etc.). Contrary to occupancy and demographic surveys, GC measurements may offer a way to monitor and assess populations before declines occur.

Below we offer several suggestions for conservation practitioners and wildlife managers applying this framework to monitor species in their jurisdictions.

First, when using GC data in an applied context, we strongly suggest incorporating more specific natural history information into investigations than we were able to here. Our method of quantifying the per cent of surrounding habitat variability could be much improved by wildlife managers who have more accurate information about their species of interest. For example, a more comprehensive definition of usable 'habitat' might be more closely related to food abundance, foraging efficiency, competitor densities, or other abiotic and biotic factors unrelated to land cover [109]. Even within a species, we found that the per cent of usable land cover sometimes varies depending on the life-history stage or sex of individuals in the same population. When possible, it would be informative to incorporate the extent to which species depend on specific resources in their environment, and how this dependency may vary across life history or ontogeny.

Second, it is important to consider the context in which GC measures were collected before making judgements about populations. The conditions under which GC responsiveness is adaptive depend on the nature of the stressor [29]. For example, factors such as the abruptness of a landscape change, history of a landscape and concomitant responses of competitors or predators will determine the extent to which a change in land cover imposes 'stress' on a population [89].

Additionally, whether a GC response is considered adaptive or harmful in a particular context may vary between species or life-history stages within species. For example, studies have shown that the relationship between GC levels and fitness may be highly variable across breeding stages [22,31,33,110], but also can depend on the lifetime reproductive opportunity of a species [82,111]. Pairing GC measures with information about survivorship and recruitment will help better understand the relationship between GC levels and fitness for different species or populations.

Finally, conservation practitioners should be cautious in using one-time plasma samples of GCs to make long-term judgements about populations. While it is possible to detect broad trends using metrics as crude as population means of one-time measures, it is becoming increasingly evident that other aspects of the HPA axis are equally (or more) important for wholistically interpreting the GC response. As new data emerge, we hope that it becomes possible to start incorporating more integrative measures into wildlife assessments. Our results represent a meaningful starting point, but more repeated measures of various endocrine traits within species and across geographically distinct populations will strengthening the utility of GC as a conservation tool.

Ethics. This work did not require ethical approval from a human subject or animal welfare committee.

Data accessibility. No primary data were generated in the production of this manuscript. Datasets curated and used in analyses are available in electronic supplementary material [112], table S1 and from the GitHub digital repository: https://github.com/valaasam/GC_and_landcover_2023 [113].

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. V.J.A.: conceptualization, data curation, formal analysis, methodology, visualization, writing—original draft, writing—review and editing; T.L.B.: conceptualization, data curation, formal analysis, methodology, writing—original draft, writing—review and editing; A.R.G.: conceptualization, data curation,

writing—review and editing; J.Q.O.: conceptualization, data curation, project administration, supervision, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. The authors declare that we have no competing interests.

Funding. J.Q.O. is funded by NSF IOS-2141693 and V.J.A. funded by NSF GRFP (grant no. 1937966).

Acknowledgements. We thank A. Lindauer for significant contributions to data curation and analysis. We also thank K. Shoemaker, K. Wagner and J. Jahner for advice, and A. Gopal for our bird illustrations.

References

- IUCN. 2020 IUCN Red List quadrennial report 2017–2020. See https://nc.iucnredlist.org/redlist/resources/files/1630480997-IUCN_RED_LIST_QUADRENNIAL_REPORT_2017-2020.pdf (accessed on 28 May 2023).
- WWF. 2018 *Living Planet Report - 2018: Aiming Higher* (eds M Grooten, REA Almond). Gland, Switzerland: WWF.
- Hooke RLeB, Martin-Duque JF. 2012 Land transformation by humans: a review. *GSA Today* **12**, 4–10. (doi:10.1130/GSAT151A.1)
- Balestri M, Barresi M, Campera M, Serra V, Ramanamanjato JB, Heistermann M, Donati G. 2014 Habitat degradation and seasonality affect physiological stress levels of *Eulemur collaris* in littoral forest fragments. *PLoS ONE* **9**, e107698. (doi:10.1371/journal.pone.0107698)
- Newcomb HR, Regosin JV, Rodrigues DM, Reed JM, Windmiller BS, Romero LM. 2003 Impacts of varying habitat quality on the physiological stress of spotted salamanders (*Ambystoma maculatum*). *Anim. Conserv.* **6**, 11–18. (doi:10.1017/S1367943003003032)
- Suorsa P, Huhta E, Nikula A, Nikinmaa M, Jäntti A, Helle H, Hakkarainen H. 2003 Forest management is associated with physiological stress in an old-growth forest passerine. *Proc. R. Soc. B* **270**, 963–969. (doi:10.1098/rspb.2002.2326)
- Janin A, Léna J-P, Joly P. 2011 Beyond occurrence: body condition and stress hormone as integrative indicators of habitat availability and fragmentation in the common toad. *Biol. Conserv.* **144**, 1008–1016. (doi:10.1016/j.biocon.2010.12.009)
- Hinam HL, St Clair CC. 2008 High levels of habitat loss and fragmentation limit reproductive success by reducing home range size and provisioning rates of Northern saw-whet owls. *Biol. Conserv.* **141**, 524–535. (doi:10.1016/j.biocon.2007.11.011)
- Johnstone CP, Lill A, Reina RD. 2014 Habitat loss, fragmentation and degradation effects on small mammals: analysis with conditional inference tree statistical modelling. *Biol. Conserv.* **176**, 80–98. (doi:10.1016/j.biocon.2014.04.025)
- Sapolsky RM, Romero LM, Munck A U. 2000 How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr. Rev.* **21**, 55–89. (doi:10.1210/edrv.21.1.0389)
- Romero LM, Dickens MJ, Cyr NE. 2009 The reactive scope model—a new model integrating homeostasis, allostasis, and stress. *Horm. Behav.* **55**, 375–389. (doi:10.1016/j.yhbeh.2008.12.009)
- Wikelski M, Cooke SJ. 2006 Conservation physiology. *Trends Ecol. Evol.* **21**, 38–46. (doi:10.1016/j.tree.2005.10.018)
- Ellis RD, McWhorter TJ. 2012 Integrating landscape ecology and conservation physiology. *Landscape Ecol. Rev.* **27**, 1–12. (doi:10.1007/s10980-011-9671-6)
- Cooke SJ *et al.* 2020 Reframing conservation physiology to be more inclusive, integrative, relevant and forward-looking: reflections and a horizon scan. *Conserv. Physiol.* **8**, 16. (doi:10.1093/conphys/coaa016)
- Dantzer B, Fletcher QE, Boonstra R, Sheriff MJ. 2014 Measures of physiological stress: a transparent or opaque window into the status, management and conservation of species? *Conserv. Physiol.* **2**, cou023. (doi:10.1093/conphys/cou023)
- Davis AK, Maney DL. 2018 The use of glucocorticoid hormones or leucocyte profiles to measure stress in vertebrates: what's the difference? *Methods Ecol. Evol.* **9**, 1556–1568. (doi:10.1111/2041-210X.13020)
- Guindre-Parker S. 2018 The evolutionary endocrinology of circulating glucocorticoids in free-living vertebrates: recent advances and future directions across scales of study. *Integr. Comp. Biol.* **58**, 814–825. (doi:10.1093/icb/icy048)
- Madliger CL, Love OP, Hultine KR, Cooke SJ. 2018 The conservation physiology toolbox: status and opportunities. *Conserv. Physiol.* **6**, 29. (doi:10.1093/conphys/coy029)
- Romero LM. 2004 Physiological stress in ecology: lessons from biomedical research. *Trends Ecol. Evol.* **19**, 249–255. (doi:10.1016/j.tree.2004.03.008)
- McEwen BS, Wingfield JC. 2003 The concept of allostasis in biology and biomedicine. *Horm. Behav.* **43**, 2–15. (doi:10.1016/S0018-506X(02)00024-7)
- Eikenaar C, Mu F, Leutgeb C, Hessler S, Lebus K, Taylor PD, Schmaljohann H. 2017 Corticosterone and timing of migratory departure in a songbird. *Proc. R. Soc. B* **284**, 9–14. (doi:10.1098/rspb.2016.2300)
- Ouyang JQ, Hau M, Bonier F. 2011 Within seasons and among years: when are corticosterone levels repeatable? *Horm. Behav.* **60**, 559–564. (doi:10.1016/j.yhbeh.2011.08.004)
- Crino OL, Driscoll SC, Brandl HB, Buchanan KL, Griffith SC. 2020 Under the weather: corticosterone levels in wild nestlings are associated with ambient temperature and wind. *Gen. Comp. Endocrinol.* **285**, 113247. (doi:10.1016/j.ygcen.2019.113247)
- Guindre-Parker S, Rubenstein DR. 2021 Long-term measures of climate unpredictability shape the avian endocrine stress axis. *Am. Nat.* **198**, 394–405. (doi:10.1086/715628)
- Jenni-Eiermann S, Glaus E, Gruebler M, Schwabl H, Jenni L. 2008 Glucocorticoid response to food availability in breeding barn swallows (*Hirundo rustica*). *Gen. Comp. Endocrinol.* **155**, 558–565. (doi:10.1016/j.ygcen.2007.08.011)
- Cockrem JF. 2013 Individual variation in glucocorticoid stress responses in animals. *Gen. Comp. Endocrinol.* **181**, 45–58. (doi:10.1016/j.ygcen.2012.11.025)
- Wingfield JC, Maney DL, Breuner CW, Jacobs JD, Lynn S, Ramenofsky M, Richardson RD. 1998 Ecological bases of hormone–behavior interactions: the 'Emergency Life History Stage'. *Integr. Comp. Biol.* **38**, 191–206. (doi:10.1093/icb/38.1.191)
- Hau M, Ricklefs RE, Wikelski M, Lee KA, Brawn JD. 2010 Corticosterone, testosterone and life-history strategies of birds. *Proc. R. Soc. B* **277**, 3203–3212. (doi:10.1098/rspb.2010.0673)
- Schoenle LA, Zimmer C, Vitousek MN. 2018 Understanding context dependence in glucocorticoid–fitness relationships: the role of the nature of the challenge, the intensity and frequency of stressors, and life history. *Integr. Comp. Biol.* **58**, 777–789. (doi:10.1093/icb/icy046)
- Bonier F, Martin PR, Moore IT, Wingfield JC. 2009 Do baseline glucocorticoids predict fitness? *Trends Ecol. Evol.* **24**, 634–642. (doi:10.1016/j.tree.2009.04.013)
- Ouyang JQ, Sharp PJ, Dawson A, Quetting M, Hau M. 2011 Hormone levels predict individual differences in reproductive success in a passerine bird. *Proc. R. Soc. B* **278**, 2537–2545. (doi:10.1098/rspb.2010.2490)
- Magee SE, Neff BD, Knapp R. 2006 Plasma levels of androgens and cortisol in relation to breeding behavior in parental male bluegill sunfish, *Lepomis macrochirus*. *Horm. Behav.* **49**, 598–609. (doi:10.1016/j.yhbeh.2005.12.003)
- Vitousek MN, Taff CC, Hallinger KK, Zimmer C, Winkler DW. 2018 Hormones and fitness: evidence for trade-offs in glucocorticoid regulation across contexts. *Front. Ecol. Evol.* **6**, 42. (doi:10.3389/fevo.2018.00042)
- Sims CG, Holberton RL. 2000 Development of the corticosterone stress response in young Northern Mockingbirds (*Mimus polyglottos*). *Gen. Comp. Endocrinol.* **119**, 193–201. (doi:10.1006/gcen.2000.7506)
- Bonier F, Martin PR, Sheldon KS, Jensen JP, Foltz SL, Wingfield JC. 2007 Sex-specific consequences of life in the city. *Behav. Ecol.* **18**, 121–129. (doi:10.1093/beheco/arl050)
- Crespi EJ, Williams TD, Jessop TS, Delehanty B. 2013 Life history and the ecology of stress: how do

- glucocorticoid hormones influence life-history variation in animals? *Funct. Ecol.* **27**, 93–106. (doi:10.1111/1365-2435.12009)
37. Dickens MJ, Romero LM. 2013 A consensus endocrine profile for chronically stressed wild animals does not exist. *Gen. Comp. Endocrinol.* **191**, 177–189. (doi:10.1016/j.ygcen.2013.06.014)
 38. Romero LM, Gormally BMG. 2019 How truly conserved is the 'well-conserved' vertebrate stress response? *Integr. Comp. Biol.* **59**, 273–281. (doi:10.1093/icb/icz011)
 39. Taff CC, Zimmer C, Vitousek MN. 2018 Efficacy of negative feedback in the HPA axis predicts recovery from acute challenges. *Biol. Lett.* **14**, 20180131. (doi:10.1098/rsbl.2018.0131)
 40. Madliger CL *et al.* 2016 Success stories and emerging themes in conservation physiology. **4**, 1–17. (doi:10.1093/conphys/cov057)
 41. Eikenaar C, Husak J, Escallón C, Moore IT. 2012 Variation in testosterone and corticosterone in amphibians and reptiles: relationships with latitude, elevation, and breeding season length. *Am. Nat.* **180**, 642–654. (doi:10.1086/667891)
 42. Jessop TS, Woodford R, Symonds MRE. 2013 Macrostress: do large-scale ecological patterns exist in the glucocorticoid stress response of vertebrates? *Funct. Ecol.* **27**, 120–130. (doi:10.1111/j.1365-2435.2012.02057.x)
 43. Jessop TS, Lane ML, Teasdale L, Stuart-Fox D, Wilson RS, Careau V, Moore IT. 2016 Multiscale evaluation of thermal dependence in the glucocorticoid response of vertebrates. *Am. Nat.* **188**, 342–356. (doi:10.1086/687588)
 44. Taff CC, Wingfield JC, Vitousek MN. 2022 The relative speed of the glucocorticoid stress response varies independently of scope and is predicted by environmental variability and longevity across birds. *Horm. Behav.* **144**, 105226. (doi:10.1016/j.yhbeh.2022.105226)
 45. Sorenson GH, Dey CJ, Madliger CL, Love OP. 2017 Effectiveness of baseline corticosterone as a monitoring tool for fitness: a meta analysis in seabirds. *Oecologia* **183**, 353–365. (doi:10.1007/s00442-016-3774-3)
 46. Pengra BW, Stehman SV, Horton JA, Dockter DJ, Schroeder TA, Yang Z, Cohen WB, Healey SP, Loveland TR. 2020 Quality control and assessment of interpreter consistency of annual land cover reference data in an operational national monitoring program. *Remote Sens. Environ.* **238**, 111261. (doi:10.1016/j.rse.2019.111261)
 47. Leimgruber P, Christen CA, Laborde A. 2005 The impact of Landsat satellite monitoring on conservation biology. *Environ. Monit. Assess.* **106**, 81–101. (doi:10.1007/s10661-005-0763-0)
 48. Hoekstra JM, Boucher TM, Ricketts TH, Roberts C. 2005 Confronting a biome crisis: global disparities of habitat loss and protection. *Ecol. Lett.* **8**, 23–29. (doi:10.1111/j.1461-0248.2004.00686.x)
 49. Turner BL, Lambin EF, Reenberg A. 2007 The emergence of land change science for global environmental change and sustainability. *Proc. Natl. Acad. Sci. USA* **104**, 20 666–20 671. (doi:10.1073/pnas.0704119104)
 50. Foody GM. 2010 Assessing the accuracy of land cover change with imperfect ground reference data. *Remote Sens. Environ.* **114**, 2271–2285. (doi:10.1016/j.rse.2010.05.003)
 51. Saddique N, Mahmood T, Bernhofer C. 2020 Quantifying the impacts of land use/land cover change on the water balance in the afforested River Basin, Pakistan. *Environ. Earth Sci.* **79**, 448. (doi:10.1007/s12665-020-09206-w)
 52. Mengist W, Soromessa T, Feyisa GL. 2021 Monitoring afro-montane forest cover loss and the associated socio-ecological drivers in Kaffa biosphere reserve, Ethiopia. *Trees Forests People* **6**, 100161. (doi:10.1016/j.tfp.2021.100161)
 53. Lennox R, Cooke SJ. 2014 State of the interface between conservation and physiology: a bibliometric analysis. *Conserv. Physiol.* **2**, cou003. (doi:10.1093/conphys/cou003)
 54. Vitousek MN *et al.* 2018 HormoneBase, a population-level database of steroid hormone levels across vertebrates. *Sci. Data* **5**, 180097. (doi:10.1038/sdata.2018.97)
 55. Casagrande S *et al.* 2018 Do seasonal glucocorticoid changes depend on reproductive investment? A comparative approach in birds. *Integr. Comp. Biol.* **58**, 739–750. (doi:10.1093/icb/icy022)
 56. Francis CD *et al.* 2018 Metabolic scaling of stress hormones in vertebrates. *Integr. Comp. Biol.* **58**, 729–738. (doi:10.1093/icb/icy063)
 57. Rubalcaba JG, Jimeno B. 2022 Biophysical models unravel associations between glucocorticoids and thermoregulatory costs across avian species. *Funct. Ecol.* **36**, 64–72. (doi:10.1111/1365-2435.13946)
 58. Martin LB *et al.* 2018 IUCN conservation status does not predict glucocorticoid concentrations in reptiles and birds. *Integr. Comp. Biol.* **58**, 800–813. (doi:10.1093/icb/icy102)
 59. Avgar T, Betini GS, Fryxell JM. 2020 Habitat selection patterns are density dependent under the ideal free distribution. *J. Anim. Ecol.* **89**, 2777–2787. (doi:10.1111/1365-2656.13352)
 60. Rodewald P. (ed.). 2015 *The birds of North America*. Ithaca, NY: Cornell Laboratory of Ornithology.
 61. Vogelmann JE, Howard SM, Yang L, Larson CR, Wylie BK, Van Driel JN. 2001 Completion of the 1990s National Land Cover Data set for the conterminous United States from Landsat Thematic Mapper data and ancillary data sources. *Photogramm. Eng. Remote Sensing* **67**, 650–662. (doi:10.1007/978-94-011-4976-1_32)
 62. Homer C, Dewitz L, Yang S, Danielson P, Xian G, Coulston J, Herold N, Wickham J, Megown K. 2015 Completion of the 2011 National Land Cover Database for the conterminous United States – representing a Decade of Land Cover Change Information. *Photogramm. Eng. Remote Sensing* **81**, 345–354. (doi:10.1016/S0099-1112(15)30100-2)
 63. Homer CG *et al.* 2007 Completion of the 2001 National Land Cover Database for the conterminous United States. *Photogramm. Eng. Remote Sensing* **73**, 337–341. (doi:10.1016/j.rse.2010.02.018)
 64. Fry J, Xian G, Jin S, Dewitz J, Homer CG, Yang L, Barnes CA, Herold ND, Wickham JD. 2011 Completion of the 2006 national land cover database for the conterminous United States. *Photogramm. Eng. Remote Sensing* **77**, 858–864.
 65. Senzaki M, Barber JR, Phillips JN, Carter NH, Cooper CB, Dittmer MA, Frisrup KM, McClure CJW, Mennitt DJ. 2020 Sensory pollutants alter bird phenology and fitness across a continent. *Nature* **587**, 605–609. (doi:10.1038/s41586-020-2903-7)
 66. Bókony V *et al.* 2009 Stress response and the value of reproduction: are birds prudent parents? *Am. Nat.* **173**, 589–598. (doi:10.1086/597610)
 67. Hijmans RJ. 2015 Raster: Geographic Data Analysis and Modeling. R Package v.2.4-15. (See <http://CRAN.R-project.org/package=raster>.)
 68. R Core Team. 2021 *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. (See <https://www.R-project.org/>.)
 69. Hadfield JD, Nakagawa S. 2010 General quantitative genetic methods for comparative biology: phylogenies, taxonomies and multi-trait models for continuous and categorical characters. *J. Evol. Biol.* **23**, 494–508. (doi:10.1111/j.1420-9101.2009.01915.x)
 70. Hackett SJ *et al.* 2008 A phylogenomic study of birds reveals their evolutionary history. *Science* **320**, 1763–1768. (doi:10.1126/science.1157704)
 71. Jetz W, Thomas GH, Joy JB, Hartmann K, Mooers AO. 2012 The global diversity of birds in space and time. *Nature* **491**, 444–448. (doi:10.1038/nature11631)
 72. Holder MT, Sukumaran J, Lewis PO. 2008 A justification for reporting the majority-rule consensus tree in Bayesian phylogenetics. *Syst. Biol.* **57**, 814–821. (doi:10.1080/10635150802422308)
 73. Sukumaran J, Holder MT. 2010 DendroPy: a Python library for phylogenetic computing. *Bioinformatics* **26**, 1569–1571. (doi:10.1093/bioinformatics/btq228)
 74. Rubolini D, Liker A, Garamszegi LZ, Møller AP, Saino N. 2015 Using the BirdTree.org website to obtain robust phylogenies for avian comparative studies: a primer. *Curr. Zool.* **61**, 959–965. (doi:10.1093/czoolo/61.6.959)
 75. Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
 76. Romero LM, Reed JM. 2005 Collecting baseline corticosterone samples in the field: is under 3 min good enough? *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **140**, 73–79. (doi:10.1016/j.cbpb.2004.11.004)
 77. Fanson KV, Németh Z, Ramenofsky M, Wingfield JC, Buchanan KL. 2017 Inter-laboratory variation in corticosterone measurement: implications for comparative ecological and evolutionary studies. *Methods Ecol. Evol.* **8**, 1745–1754. (doi:10.1111/2041-210X.12821)

78. Gelman A, Rubin DB. 1992 Inference from iterative simulation using multiple sequences. *Stat. Sci.* **7**, 457–472. (doi:10.1214/SS/1177011136)
79. Cooke SJ, Sack L, Franklin CE, Farrell AP, Beardall J, Chown SL, Wikelski M. 2013 What is conservation physiology? Perspectives on an increasingly integrated and essential science. *Conserv. Physiol.* **1**, 1. (doi:10.1093/conphys/cot001)
80. Congdon JD, Dunham AE, Hopkins WA, Rowe CL, Hinton TG. 2001 Resource allocation-based life histories: a conceptual basis for studies of ecological toxicology. *Environ. Toxicol. Chem. Int. J.* **20**, 1698–1703. (doi:10.1002/etc.5620200811)
81. Injaian AS *et al.* 2020 Baseline and stress-induced corticosterone levels across birds and reptiles do not reflect urbanization levels. *Conserv. Physiol.* **8**, coz110. (doi:10.1093/conphys/coz110)
82. Vitousek MN *et al.* 2019 Macroevolutionary patterning in glucocorticoids suggests different selective pressures shape baseline and stress-induced levels. *Am. Nat.* **193**, 866–880. (doi:10.1086/703112)
83. Ouyang JQ, Quetting M, Hau M. 2012 Corticosterone and brood abandonment in a passerine bird. *Anim. Behav.* **84**, 261–268. (doi:10.1016/j.anbehav.2012.05.006)
84. Spée M, Marchal L, Lazin D, Le Maho Y, Chastel O, Beaulieu M, Raclot T. 2011 Exogenous corticosterone and nest abandonment: a study in a long-lived bird, the Adélie penguin. *Horm. Behav.* **60**, 362–370. (doi:10.1016/j.yhbeh.2011.07.003)
85. Angelier F, Clement-Chastel C, Welcker J, Gabrielsen GW, Chastel O. 2009 How does corticosterone affect parental behaviour and reproductive success? A study of prolactin in black-legged kittiwakes. *Funct. Ecol.* **23**, 784–793. (doi:10.1111/j.1365-2435.2009.01545.x)
86. Sinclair ECC, Martin PR, Bonier F. 2022 Among-species variation in hormone concentrations is associated with urban tolerance in birds. *Proc. R. Soc. B* **289**, 20221600. (doi:10.1098/rspb.2022.1600)
87. Landys MM, Ramenofsky M, Wingfield JC. 2006 Actions of glucocorticoids at a seasonal baseline as compared to stress-related levels in the regulation of periodic life processes. *Gen. Comp. Endocrinol.* **148**, 132–149. (doi:10.1016/j.ygcen.2006.02.013)
88. Marra PP, Holberton RL. 1998 Corticosterone levels as indicators of habitat quality: effects of habitat segregation in a migratory bird during the non-breeding season. *Oecologia* **116**, 284–292. (doi:10.1007/s004420050590)
89. MacDougall-Shackleton SA, Bonier F, Romero LM, Moore IT. 2019 Glucocorticoids and ‘stress’ are not synonymous. *Integr. Organ. Biol.* **1**, obz017. (doi:10.1093/iob/obz017)
90. Wada H, Sewall KB. 2014 Introduction to the Symposium—uniting evolutionary and physiological approaches to understanding phenotypic plasticity. *Integr. Comp. Biol.* **54**, 774–782. (doi:10.1093/icb/icu097)
91. Fanson KV, Biro PA. 2019 Meta-analytic insights into factors influencing the repeatability of hormone levels in agricultural, ecological, and medical fields. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **316**, R101–R109. (doi:10.1152/ajpregu.00006.2018)
92. Taff CC, Vitousek MN. 2016 Endocrine flexibility: optimizing phenotypes in a dynamic world? *Trends Ecol. Evol.* **31**, 476–488. (doi:10.1016/j.tree.2016.03.005)
93. Grace JK, Anderson DJ. 2014 Corticosterone stress response shows long-term repeatability and links to personality in free-living Nazca boobies. *Gen. Comp. Endocrinol.* **208**, 39–48. (doi:10.1016/j.ygcen.2014.08.020)
94. Small TW, Schoech SJ. 2015 Sex differences in the long-term repeatability of the acute stress response in long-lived, free-living Florida scrub jays (*Aphelocoma coerulescens*). *J. Comp. Physiol. B* **185**, 119–133. (doi:10.1007/s00360-014-0866-4)
95. Grant AR, Baldan D, Kimball MG, Malisch JL, Ouyang JQ. 2020 Across time and space: hormonal variation across temporal and spatial scales in relation to nesting success. *Gen. Comp. Endocrinol.* **292**, 113462. (doi:10.1016/j.ygcen.2020.113462)
96. Cockrem JF, Candy EJ, Barrett DP, Agnew P, Potter MA. 2017 Individual variation and repeatability of corticosterone responses of little penguins (*Eudyptula minor*) sampled in two successive years at Oamaru, New Zealand. *Gen. Comp. Endocrinol.* **244**, 86–92. (doi:10.1016/j.ygcen.2016.01.010)
97. Jenkins BR, Vitousek MN, Hubbard JK, Safran RJ. 2014 An experimental analysis of the heritability of variation in glucocorticoid concentrations in a wild avian population. *Proc. R. Soc. B* **281**, 20141302. (doi:10.1098/rspb.2014.1302)
98. Pottinger TG, Carrick TR. 1999 Modification of the plasma cortisol response to stress in rainbow trout by selective breeding. *Gen. Comp. Endocrinol.* **116**, 122–132. (doi:10.1006/gcen.1999.7355)
99. Evans MR, Roberts ML, Buchanan KL, Goldsmith AR. 2006 Heritability of corticosterone response and changes in life history traits during selection in the zebra finch. *J. Evol. Biol.* **19**, 343–352. (doi:10.1111/j.1420-9101.2005.01034.x)
100. Romero LM, Wikelski M. 2002 Severe effects of low-level oil contamination on wildlife predicted by the corticosterone-stress response: preliminary data and a research agenda. *Spill Sci. Technol. Bull.* **7**, 309–313. (doi:10.1016/S1353-2561(02)00067-1)
101. Angelier F, Wingfield JC. 2013 Importance of the glucocorticoid stress response in a changing world: theory, hypotheses and perspectives. *Gen. Comp. Endocrinol.* **190**, 118–128. (doi:10.1016/j.ygcen.2013.05.022)
102. Fefferman NH, Romero LM. 2013 Can physiological stress alter population persistence? A model with conservation implications. *Conserv. Physiol.* **1**, cot012. (doi:10.1093/conphys/cot012)
103. Atwell JW, Cardoso GC, Whittaker DJ, Campbell-Nelson S, Robertson KW, Ketterson ED. 2012 Boldness behavior and stress physiology in a novel urban environment suggest rapid correlated evolutionary adaptation. *Behav. Ecol.* **23**, 960–969. (doi:10.1093/beheco/ars059)
104. Love OP, Williams TD. 2008 Plasticity in the adrenocortical response of a free-living vertebrate: the role of pre- and post-natal developmental stress. *Horm. Behav.* **54**, 496–505. (doi:10.1016/j.yhbeh.2008.01.006)
105. Sepp T, McGraw KJ, Kaasik A, Giraudeau M. 2018 A review of urban impacts on avian life-history evolution: does city living lead to slower pace of life? *Global Chang. Biol.* **24**, 1452–1469. (doi:10.1111/gcb.13969)
106. Grunst ML, Rotenberry JT, Grunst AS. 2014 Variation in adrenocortical stress physiology and condition metrics within a heterogeneous urban environment in the song sparrow *Melospiza melodia*. *J. Avian Biol.* **45**, 574–583. (doi:10.1111/jav.00459)
107. Ibáñez-Álamo JD, Jimeno B, Gil D, Thomson RL, Aguirre JI, Díez-Fernández A, Fairve B, Tieleman BI, Figuerola J. 2020 Physiological stress does not increase with urbanization in European blackbirds: evidence from hormonal, immunological and cellular indicators. *Sci. Total Environ.* **721**, 137332. (doi:10.1016/j.scitotenv.2020.137332)
108. Marra PP. 1999 The role of behavioral dominance in structuring patterns of habitat occupancy in a migrant bird during the nonbreeding season. *Behav. Ecol.* **11**, 299–308. (doi:10.1016/j.parkreldis.2014.09.008)
109. Johnson MD. 2007 Measuring habitat quality: a review. *Condor* **109**, 489. (doi:10.1650/8347.1)
110. Bonier F, Moore IT, Martin PR, Robertson RJ. 2009 The relationship between fitness and baseline glucocorticoids in a passerine bird. *Gen. Comp. Endocrinol.* **163**, 208–213. (doi:10.1016/j.ygcen.2008.12.013)
111. Schoenle LA, Zimmer C, Miller ET, Vitousek MN. 2021 Does variation in glucocorticoid concentrations predict fitness? A phylogenetic meta-analysis. *Gen. Comp. Endocrinol.* **300**, 113611. (doi:10.1016/j.ygcen.2020.113611)
112. Alaasam VJ, Behnke TL, Grant AR, Ouyang JQ. 2024 Glucocorticoids and land cover: a largescale comparative approach to assess a physiological biomarker for avian conservation. Figshare. (doi:10.6084/m9.figshare.c.6996932)
113. Alaasam VJ, Behnke TL, Grant AR, Ouyang JQ. 2024 Glucocorticoids and land cover: a largescale comparative approach to assess a physiological biomarker for avian conservation. GitHub digital repository. (https://github.com/valaasam/GC_and_landcover_2023)