



Data Article

Sea cucumber (*Holothuria glaberrima*) intestinal microbiome dataset from Puerto Rico, generated by shotgun sequencing



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ABSTRACT

The sea cucumber (*H. glaberrima*) is a species found in the shallow waters near coral reefs and seagrass beds in Puerto Rico. To characterize the microbial taxonomic composition and functional profiles present in the sea cucumber, total DNA was obtained from their intestinal system, fosmid libraries constructed, and subsequent sequencing was performed. The diversity profile displayed that the most predominant domain was Bacteria (76.56 %), followed by Viruses (23.24 %) and Archaea (0.04 %). Within the 11 phyla identified, the most abundant was Proteobacteria (73.16 %), followed by Terrabacteria group (3.20 %) and Fibrobacterota, Chlorobiota, Bacteroidota (FCB) superphylum (1.02 %). The most abundant species were *Porvindicencia rettgeri* (21.77 %), *Pseudomonas stutzeri* (14.78 %), and *Alcaligenes faecalis* (5.00 %). The functional profile revealed that the most abundant functions are related to transporters, MISC (miscellaneous information systems), organic nitrogen, energy, and carbon utilization. The data collected in this project on the diversity and functional profiles of the intestinal system of the *H.*

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glaberrima provided a detailed view of its microbial ecology. These findings may motivate comparative studies aimed at understanding the role of the microbiome in intestinal regeneration.

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Specifications Table

Subject	Microbiology
Specific subject area	Metagenomics
Data format	Raw data, Processed
Type of data	Fasta Qfile, Figures
Data collection	Sea cucumbers were collected from Piñones Beach in San Juan, Puerto Rico. Metagenomic DNA was extracted from three samples of the intestinal system of <i>Holothuria glaberrima</i> : the complete digestive system (DS), the washed intestine (WI), and the contents from the washed intestine (CW). The presented data resulted from the extraction of the fosmid pCC1FOS, shotgun sequencing and analysis in the National Microbiome Data Collaborative – Empowering the Development of Genomics Expertise (NMDC – EDGE) platform.
Data source location	Piñones Beach in San Juan, Puerto Rico (18.451141, -65.905634)
Data accessibility	Raw data and annotations of this metagenome are available in NCBI under BioProject PRJNA1061805 (https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1061805) and NMDC – EDGE under the project title “ <i>Holothuria glaberrima</i> - Eviscerated Gut Metagenome EDGE Bioinformatics” under link: https://nmdc-edge.org/public/project?code=uTDw1Jv0NGWhTOQV

1. Value of the Data

- This project represents the diversity and functional profiles of the intestinal system of the sea cucumber (*H. glaberrima*) in the Caribbean.
- The profiles generated can be used in comparative studies with the microbial flora present in other sea cucumbers.
- Diversity and functional profiles may prompt comparative bioprospecting studies to understand the potential role of the microbiome in processes such as intestinal regeneration.

2. Data Description

The sea cucumber (*H. glaberrima*) (Fig. 1) is a species found in the shallow waters near coral reefs and seagrass beds in Puerto Rico [1]. Members of the class Holothuroidea, including *H. glaberrima*, have the capability to regenerate their internal organs after the process of evisceration [2]. Here, we present descriptions of the diversity (Fig. 2) and functional profiles (Fig. 3) of the microbial communities within the intestinal system of the sea cucumber. These data were derived from a pool of three metagenomic libraries, each sourced from a specific anatomical region. From the evisceration process, the entire intestinal system, along with its internal contents were selected; this sample was identified as the complete digestive system (DS). Subsequently, only the intestine was selected and washed with a saline solution (0.85 % NaCl); this procedure generated two samples, the washed intestine (WI) and the contents from the wash (CW). To access, most of the microbial populations present in the anatomical regions of the sea cucumber



Fig. 1. *Holothuria glaberrima* before and after evisceration. **(A)** *H. glaberrima* exhibits an average length ranging from 10 to 15 cm (4 to 6 inches). Its predominant coloration is characterized by shades of black and dark brown [3]. **(B)** Evisceration process of *H. glaberrima* after induction with 2 mL of KCl [0.35 M]. Credit: Edwin Omar Rivera-Lopez and Carlos Ríos-Velázquez

at the genetic level, especially those in low abundance, independent DNA extractions from each region were performed and then combined to further be sequenced. Furthermore, the overarching goal is to understand the microbiome in each section of the sea cucumber's intestine. A total of 266,235 large insert (40kb) metagenomic clones were generated. After sequencing, a total of 9,259 sequences were obtained, with a median length of 653 bp, containing a cumulative length of 9,039,785 nucleotides. Furthermore, Table 1 provides a summary of sequence assembly statistics, including the total number of outputs read after filtering and trimming, the number of scaffolds, gaps percentage, the contig N50 value, and the number of genes per 1Mbp. Table 2 offers a summary of the metagenome the gene calls for the assembled scaffolds. Additionally, the Supplementary Data contains annotated genes associated with Fig. 3.

Table 1
Summary of sequence assembly statistics.

Total number of outputs read after filters and trimming	1,018,378
Scaffolds	9,259
Gaps PCT	0
Ctg_N50	2,035
Genes per 1Mbp	1,852.04

Table 2
Summary of metagenome gene calls.

Feature type	Prediction method	Number of seqs	Number of bps	Median length	Average length	Length shortest seq	Length longest seq	Standard deviation	Number of predicted features
CDS	Prodigal v2.6.3_patched	8,911	7,224,504	438	496.666	75	6,081	355.181	14,546
CDS	GeneMark.hmm-2 v1.25_lic	1,614	602,838	207	322.546	90	3,447	307.198	1,869
misc_feature	INFERNAL 1.1.3 (Nov 2019)	3	177	59	59	59	59	0	3
regulatory	INFERNAL 1.1.3 (Nov 2019)	3	177	59	59	59	59	0	3
rRNA	INFERNAL 1.1.3 (Nov 2019)	3	177	59	59	59	59	0	3
tmRNA	INFERNAL 1.1.3 (Nov 2019)	3	177	59	59	59	59	0	3
ncRNA	INFERNAL 1.1.3 (Nov 2019)	3	177	59	59	59	59	0	3
misc_binding	INFERNAL 1.1.3 (Nov 2019)	3	177	59	59	59	59	0	3

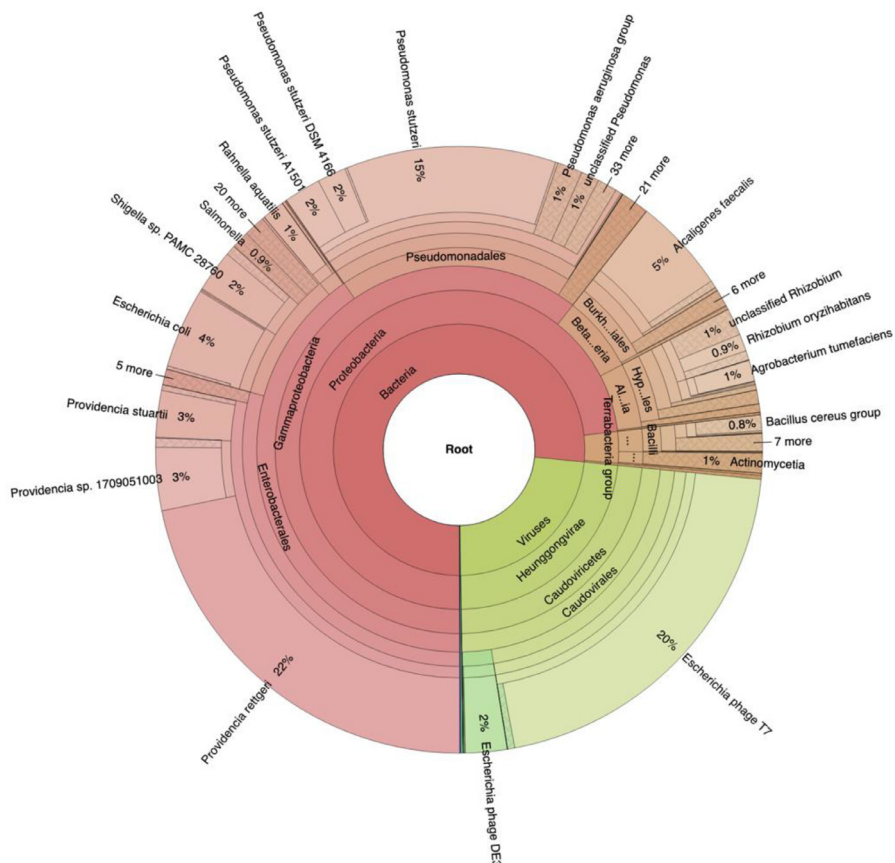


Fig. 2. Taxonomic diversity of the sea cucumber (*Holothuria glaberrima*) intestinal metagenome. Utilizing the Centrifuge Metagenomic Classification Tool through NMDC-EDGE showed that there were 5,557,577 classified reads and 314,616 species reads. The most abundant domain was Bacteria (76.56 %), followed by Virus (23.24 %) and Archaea (0.04 %). The most common Phyla out of the 11 found in Bacteria was Proteobacteria (73.16 %), followed by those included in the Terrabacteria group (3.20%) and Fibrobacterota, Chlorobiota, Bacteroidota (FCB) superphylum (1.02 %). The most abundant species were *Porvindicia rettgeri* (21.77 %), *Pseudomonas stutzeri* (14.78 %), and *Alcaligenes faecalis* (5.00 %).

3. Experimental Design, Materials and Methods

3.1. Sample Collection

The specimens were collected by Dr. Jose Garcia-Arraras' Laboratory from Piñones Beach in San Juan, Puerto Rico. A total of 13 specimens were collected and transported to the laboratory, where they were kept in a marine water aquarium (extracted from the natural environment) for 24 hours prior to the induction of evisceration (Fig. 1B) [4–6]. Subsequently, the samples DS, WI, and CW were collected in Falcon tubes and transported on dry ice to the Laboratory of Microbial Biotechnology and Bioprospecting, at the University of Puerto Rico at Mayagüez for genetic material extraction, processing and metagenomic libraries generation.

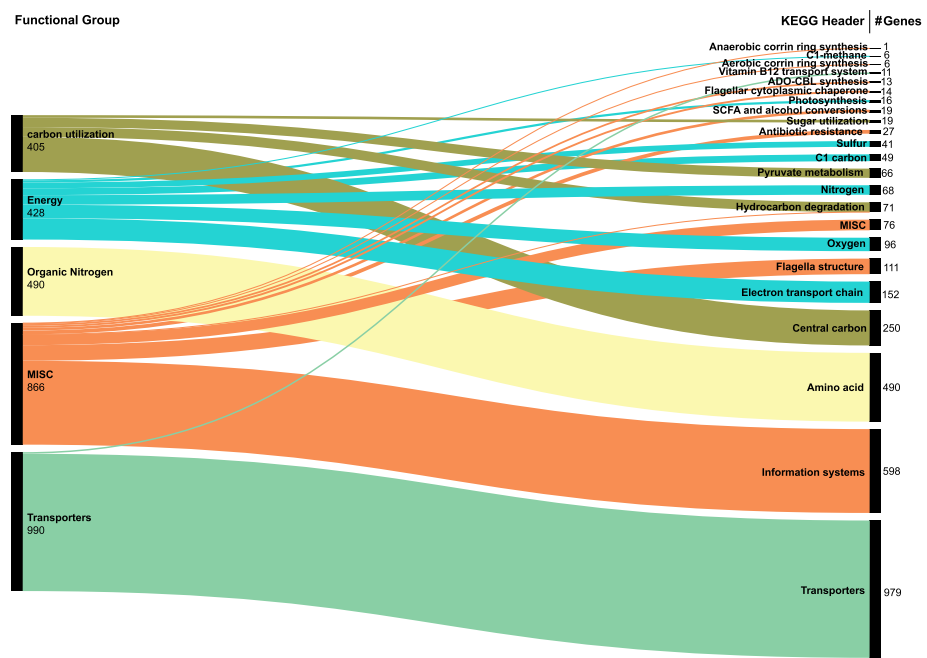


Fig. 3. Functional annotation of the *Holothuria glaberrima* anatomic regions metagenomic genes. Alluvial plot exhibits the total genes that were annotated and assigned to a KEGG category by DRAM metabolism sheet. The colors represent different types of functional groups. The numbers represent the total number of genes identified per category. Full functional annotation of metagenomes is publicly available via the NMDC EDGE platform and NCBI. From the assembly scaffolds, gene calls resulted in 16,615 total genes. Of those, 9,092 contained a KO number, used to categorize them metabolically, obtaining 2,480 genes.

3.2. DNA Extraction and Library Preparation

For DNA extraction, a direct method was employed, combining mechanical (freezing and thawing), enzymatic (lysozyme), and chemical approaches (SDS and GITC as a chaotropic agent) [7]. Fragments of 40 kb were chosen from the agarose gel. Subsequently, the purified DNA was ligated into the fosmid pCC1FOS and packed into Lambda phages (MaxPlax™ Lambda), followed by the transduction of the packed DNA into *Escherichia coli* EPI300-T1R. The clones were combined into a masterpool, which was subsequently stored at -80°C. The procedure was carried out with DS, WI, and CW parts of the sea cucumber's intestinal system.

3.3. Metagenome Sequencing

Fosmid extraction (QIAGEN Plasmid Midi) was carried out from a masterpool culture of each sample (DS, WI, and CW), incubated for 5 hours at 37°C. The resulting metagenomic DNA was sent to Mr. DNA's laboratory (<http://www.mrdnalab.com>) for short-read sequencing (Illumina). In this process, the sample was fragmented, and the adapter sequences were incorporated. Subsequently, the library concentration was reduced to 4.0 nM and sequencing was performed with 600 cycles using the Illumina MiSeq system.

3.4. Metagenomic Data Processing

Metagenomic data based on the pooled fosmids was processed through the National Microbiome Data Collaborative (NMDC) open informatics platform, EDGE [8] using their

standardized bioinformatic workflows. Docker images and full commands for each of the processes can be found on their GitHub (<https://github.com/microbiomedata>). Briefly, raw reads files were uploaded to the NMDC EDGE platform via their graphic user interface (GUI) (<https://nmdc-edge.org/home>) and the full metagenomics workflow was employed. Read quality control was performed by *rqfilter2* from BBTools [9] as described on the NMDC GitHub (<https://github.com/microbiomedata/ReadsQC>). Read-based taxonomy was assigned using three different tools: Kraken2 [10], GOTCHA [11], and Centrifuge [12] (<https://github.com/microbiomedata/ReadbasedAnalysis>). With the resulting KRONA plot from Centrifuge shown in Figure 2. Metagenome assembly was performed with MetaSpades [13] (<https://github.com/microbiomedata/metaAssembly>). Assembled scaffolds were then annotated with tRNAscan_SE [14], RFAM [15], CRT [16], Prodigal [17], and GeneMarkS [18] as described in the NMDC workflow (https://github.com/microbiomedata/mg_annotation). The resulting KOs were then assigned to KEGG functional groups using the DRAM metabolism hierarchy [19] and plotted in Figure 3. Additionally, metagenome assembled genomes (MAGs) were as described in the NMDC GitHub (<https://github.com/microbiomedata/metaMAGs>).

Limitations

Not applicable.

Ethics Statement

This article is an original work of the authors. *Holothuria glaberrima* is found in large numbers along the coast of Puerto Rico. This species is not threatened with extinction nor subject to protective measures. Since they are invertebrates, there is no need for specific permits when collecting them.

Data Availability

[Holothuria glaberrima Eviscerated Gut Metagenome Raw sequence reads \(Original data\)](#) (NCBI).

CRedit Author Statement

Edwin Omar Rivera-Lopez: Investigation, Writing – original draft; **Rene Nieves-Morales:** Investigation, Writing – original draft; **Gabriela Melendez-Martinez:** Investigation, Writing – original draft, Formal analysis; **Jessica Alejandra Paez-Diaz:** Investigation, Writing – original draft; **Sofia Marie Rodriguez-Carrio:** Investigation, Writing – original draft; **Josue Rodriguez-Ramos:** Formal analysis; **Luis Morales-Valle:** Formal analysis; **Carlos Rios-Velazquez:** Supervision, Writing – review & editing.

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Declaration of Competing Interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Supplementary Materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.dib.2024.110421.

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