

1 **Diversity of intraspecific patterns of brain region size covariation in fish**  
2  
3 **Caleb J. Axelrod<sup>\*1</sup>, Ellen M. Urquhart<sup>\*2</sup>, Pria N. Mahabir<sup>3</sup>, Bruce A. Carlson<sup>\*\*2</sup>, and Swanne P.**  
4 **Gordon<sup>\*\*1</sup>**

5  
6 **\*These authors have contributed equally to this work**  
7 **\*\*These authors have contributed equally to this work**

8  
9 **\*Contact info:**  
10 [ca537@cornell.edu](mailto:ca537@cornell.edu)  
11 215 Tower Road  
12 Ithaca, NY, 14853  
13 314-348-6825

14  
15 <sup>1</sup>Department of Ecology and Evolution, Cornell University, Ithaca, NY, USA  
16 <sup>2</sup>Department of Biology, Washington University in St. Louis, St. Louis, MO, USA  
17 <sup>3</sup>Department of Integrative Biology, University of Guelph, Guelph, ON, Canada

18  
19 **Running Title:** Fish Brain Region Covariation

20  
21 **Word Count:** 5088

22  
23 **Acknowledgements:**

24 We thank A. Lopéz-Sepulcre for comments on a preliminary version of this manuscript. We also  
25 thank E. Kane for assistance with fish sampling. This work was supported by university funds from  
26 Cornell University (to S.P. Gordon), and the National Science Foundation (IOS-2203122 & IOS-  
27 2243230 to B.A. Carlson). Sampling was done in compliance with Institutional Animal Care and Use  
28 Committee protocols at Washington University in St. Louis (Protocol 22-0265) and Cornell  
29 University (Protocol 2022-0170).

30  
31 **Keywords:** Trait Covariation, Brain Morphology, Fish, Evolution, Constraint, Teleosts

32  
33 **Data Availability:** Upon acceptance, the underlying data will be made available on Dryad.

34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44

45 **Abstract**

46 Traits often do not evolve in isolation or vary independently of other traits. Instead, they can be  
47 affected by covariation, both within and across species. However, the importance of within species  
48 trait covariation and, critically, the degree to which it varies between species has yet to be  
49 thoroughly studied. Brain morphology is a trait of great ecological and behavioral importance, with  
50 regions that are hypothesized to vary in size based on behavioral and cognitive demands. Sizes of  
51 brain regions have also been shown to covary with each other across various taxa. Here we test the  
52 degree to which covariation in brain region sizes within species has been conserved across ten  
53 teleost fish species. These ten species span five orders, allowing us to examine how phylogenetic  
54 proximity influences similarities in intraspecific trait covariation. Our results showed a trend that  
55 similar patterns of brain region size covariation occur in more closely related species. Interestingly,  
56 there were certain brain region pairs that showed similar levels of covariation across all species  
57 regardless of phylogenetic distance, such as the telencephalon and optic tectum, while others, such  
58 as the olfactory bulb and the hypothalamus, varied more independently. Ultimately, the patterns of  
59 brain region covariation shown here suggest that evolutionary mechanisms or constraints can act  
60 on specific brain regions independently, and that these constraints can change over evolutionary  
61 time.

62

63

64

65

66

67 **Introduction**

68 *Trait Covariation*

69 Trait variation is ubiquitous between and within species, providing the substrate on which natural  
70 selection can act. However, traits rarely vary independently and instead covary with other traits  
71 due to functional, genetic, and developmental links (Armbruster et al., 2014; Peiman & Robinson,  
72 2017). These effects can limit the degree of trait change because linked traits either need to change  
73 together, or selection needs to be strong enough to break those links, as supported by theory  
74 (Jones et al., 2014; Pigliucci, 2003), simulations (McGlothlin & Ketterson, 2008), and empirical  
75 observations (Ungar & Hlusko, 2016). For example, Unger and Hlusko (2016) showed that  
76 covariation in tooth morphology traits in early hominid species led to the evolution of suboptimal  
77 tooth structure in these species, with teeth unable to evolve to optimally match the diet of  
78 individual species.

79

80 Covariation between traits can be studied at multiple scales, including broad evolutionary to within  
81 population ones (Atwell et al., 2014). However, the degree to which within species trait covariation  
82 may be conserved or vary across species remains understudied (Peiman & Robinson, 2017). Similar  
83 patterns of trait covariation across species would indicate that the level of interdependence of  
84 traits may be evolutionarily conserved, while different patterns of trait covariation would suggest  
85 that these links are evolutionarily flexible. An example of this can be seen in the integration of  
86 marsupial skull morphology. Goswami (2007) showed that marsupial skulls show a phenotypic  
87 integration of bone morphology within species, resulting in constraints on the phenotypes in skull  
88 morphology that could evolve in these mammals. This paper also showed that closely related

89 species showed more similar levels of integrated changes in skull morphology across phylogenetic  
90 distance. Constraints to trait covariation across species can limit the ability of individual species to  
91 evolve optimal phenotypes in their local environments. Therefore, understanding the degree of  
92 these constraints within and across species is critical to our ability to link form and function in the  
93 evolution of organisms.

94

95 *Brain region variation*

96 Brain morphology is one trait that has been of interest to biologists for centuries,<sup>7</sup> and is critical to  
97 animal performance across environments. It can influence both cognitive and behavioral abilities of  
98 animals and thus affects animal performance and survival across environments (Axelrod et al.,  
99 2022; Benson-Amram et al., 2016; Buechel et al., 2018; A. Kotrschal et al., 2013; MacLean et al.,  
100 2014; Yang et al., [2024-n.d.](#)). Brains are divided into distinct regions, which are associated with  
101 specific cognitive, perceptual, and behavioral functions (Healy & Rowe, 2007; Huber et al., 1997; K.  
102 Kotrschal et al., 1998; Park & Bell, 2010; Pollen et al., 2007; Schumacher & Carlson, 2022; Striedter,  
103 2005; Sukhum et al., 2018). Variation in the size of brain regions within and between species can be  
104 indicative of differential selection due to environmental variation and other factors (Axelrod et al.,  
105 2021; Gonzalez-Voyer & Kolm, 2010; Laberge & Hara, 2001). However, the sizes of different regions  
106 are not independent of each other and can covary, as individuals with larger brains tend to also  
107 have similar proportioned larger brain regions both within and across species (Striedter, 2005).  
108 Here, we examine the degree to which variation in brain region size covaries within fish species,  
109 and how these patterns may differ across species.

110

111 The degree of covariation in brain regions has been of interest to evolutionary biologists for  
112 decades. The “concerted model” of brain evolution posits that the evolution of brain regions is  
113 highly constrained, with evolutionary change in the size of brain regions resulting primarily from  
114 shifts in overall brain size (Finlay & Darlington, 1995; Yopak et al., 2010). These studies found that  
115 the proportional size of brain regions was highly predictable based on the size of the whole brain,  
116 suggesting that the evolution of brain region size does not vary independently and is likely  
117 constrained by brain region covariation. On the other hand, the “mosaic model” of brain evolution  
118 suggests that specific brain regions are targeted to change in size, independent of other structures,  
119 due to their association with certain behavioral or cognitive abilities (Barton & Harvey, 2000). Since  
120 the development of these two alternative models, the evolution of brain morphology has been  
121 studied as reflecting a degree of covariation between brain region size and whole brain size,  
122 suggesting that the two hypotheses are not mutually exclusive. At broad evolutionary scales, brain  
123 region scaling appears to be conserved, with occasional “mosaic” shifts occurring in some clades  
124 (Hoops et al., 2017; Striedter, 2005; Sukhum et al., 2018; Yopak et al., 2010). For example, the  
125 evolution of electrosensory abilities in fish is linked to a mosaic shift in the sizes of the cerebellum  
126 and hindbrain (Schumacher & Carlson, 2022). However, at the intraspecific level, evidence of this  
127 degree of brain region independence is less well established (Hager et al., 2012; A. Kotrschal et al.,  
128 2017; Noreikiene et al., 2015).

129

### 130 *Approach and Hypotheses*

131 Here, we test how much intraspecific brain region covariation is conserved across 10 teleost fish  
132 species. These 10 species are paired across 5 orders of fish, allowing us to test how evolutionary

133 proximity might impact the similarity of brain region covariation patterns. Similarities we find in  
134 brain region covariation patterns across species would indicate that covariation of brain region  
135 sizes is conserved across the teleost phylogeny. If the degree of brain region covariation is very  
136 flexible evolutionarily, we would expect covariation patterns to differ across species, and not be  
137 linked to evolutionary relatedness. Finally, a midpoint between these extremes could indicate that  
138 the degree of independence of within brain region variation can evolve, but more closely related  
139 species would be more likely to share these patterns of covariation. In this case we would expect  
140 species within orders to have more similar patterns of brain region covariation, with differences  
141 emerging at more distant phylogenetic comparisons.

142

#### 143 **Methods**

##### 144 **Specimen collection**

145 To compare patterns of brain region covariation, we collected data from 10 teleost fish species  
146 spanning 5 orders (Perciformes, Scorpaeniformes, Cyprinodontiformes, Cypriniformes, and  
147 Osteoglossiformes) (figure 1). We collected the data for the Trinidadian guppy (*Poecilia reticulata*),  
148 Western mosquitofish (*Gambusia affinis*), Creek chub (*Semotilus atromaculatus*), Blacknose dace  
149 (*Rhinichthys atratulus*), African butterfly fish (*Pantodon buchholzi*), and Black baby whale fish  
150 (*Brienomyrus brachyistius*). Of these, the guppy fish were lab reared descendants of wild collected  
151 fish from Trinidad. *G. affinis*, *S. atromaculatus*, and *R. atratulus*, and *S. atromaculatus* were wild  
152 collected samples. *P. buchholzi* and *B. brachyistius* were sourced from a tropical fish supplier. We  
153 sourced data for the Pumpkinseed sunfish (*Lepomis gibbosus*), Bluegill sunfish (*Lepomis*  
154 *macrochirus*), Threespine stickleback (*Gasterosteus aculeatus*), and Ninespine stickleback (*Pungitius*

Formatted: Font: Italic

Formatted: Font: Italic

155 *pungitius*) from previously published studies that employed the same data collection method. All  
156 species contain sufficient variation in body size to allow measure region covariation. The sources of  
157 data, sample size, and coefficients of variation in body size for each species can be seen in ~~Tables~~ 1  
158 and 2. After euthanasia, all fish samples were fixed by being completely submerged in 10%  
159 formalin, and stored until processing.

160

161 *Brain region size estimation*

162 To measure brain region size, we estimated the volumes of five regions of the brain: cerebellum,  
163 optic tectum, telencephalon, olfactory bulb, and hypothalamus. For all species, we (or the authors  
164 of the relevant studies) extracted brains via dissection, ensuring that all regions were extracted  
165 intact and that external nerves were trimmed consistently. We then took photographs of the brains  
166 from dorsal, ventral, and lateral orientations. From these photographs, we measured the length (L),  
167 width (W), and height (H) of each brain region in ImageJ, and then calculated region volumes (V)  
168 using the ellipsoid formula ( $V=(L \times W \times H) \pi / 6$ ) (White & Brown, 2015) (supplementary figure 1). We  
169 estimated each region's length as a straight line from its most rostral plane to its most caudal plan.  
170 Its width was perpendicular, or 90 degrees, to the length, at the widest point of that ovoid. We  
171 estimated the height of each region by generating a straight line perpendicular to the horizontal  
172 axis of the brain, at the widest part of that lobe from the lateral view. To estimate total volume for  
173 regions with bilaterally symmetrical lobes, we summed both sides together. For example,  
174 estimating the telencephalon involved measurements of two bilaterally symmetrical lobes, and the  
175 volumes of these lobes were added together to obtain the total telencephalon volume. One  
176 exception was the olfactory bulb of *S. atromaculatus*, for which we could not reliably measure

177 olfactory bulb height. As such, we calculated the area of this region in this species ( $A=L/2 \times W/2 \times \pi$ ).

178 These data are still usable as area and volume of brain regions are highly correlated (C.A. personal  
179 observation).

180

181 Since the cerebellum of *B. brachystius* drastically varies morphologically from other teleost fish  
182 species (Sukhum et al., 2018), we had to deviate from the volume estimation methods used in the  
183 other species. Seven ovoid regions were estimated to encompass the shape and size of the  
184 enlarged *B. brachystius* cerebellum (supplementary figure 2). Two heights, for both the rostral and  
185 caudal ovoids, were taken for the *B. brachystius* cerebellum, corresponding to its rostral and  
186 caudal ovoid lengths and widths, respectively. The volumes of each of the seven cerebellum ovoids  
187 were summed together to estimate total cerebellum volume.

188

189 *Brain region principal component variation*

190 To determine how brain region volumes vary within species and observe trends in their covariation,  
191 we ran separate principal component analyses in each species. Percentage of variation explained  
192 by PC1 correlated with total brain volume, which is expected in allometric relationships  
193 (Klingenberg, 1996). These percentages were compared across species, but all regions in all species  
194 loaded in the same direction along PC1. Additionally, the direction of where the brain regions  
195 loaded along PC2 were qualitatively compared across species. PCAs were performed using the R  
196 package “FactoMineR” (Lê et al., 2008), which generated PC1-5 for each species. The majority of  
197 the variation was explained by PC1 and PC2 (ranging from ~79-98% across the 10 species), so we  
198 focused on those two axes for comparisons across species.

199

200 *Within species brain region correlations*

201 To establish the level of independence of brain region variation within each species, we calculated  
202 Pearson correlations of each pairwise comparison of brain regions in each species. We use this  
203 approach because Pearson correlations allow for differences in scale between regions being  
204 compared, which is important as regions differ in size within species. This approach also allowed for  
205 the area of the olfactory bulb in *S. atromaculatus* to be effectively compared to the volume of the  
206 other regions in that species. From each pairwise correlation of brain regions, we can establish a  
207 separate correlation matrix for each species. Lower levels of correlation between brain regions  
208 indicate higher levels of independent variation.

209

210 *Species comparisons of brain region correlation patterns*

211 To compare the patterns of within species brain region correlation we estimated the similarity of  
212 species correlation matrices. We used a Mantel test (Vegan R package; Oksanen et al. 2020) to  
213 estimate the pairwise correlation of each species' correlation matrix. This test examines the  
214 correlation of two matrices using a Pearson correlation method, and then employs a permutation  
215 approach using 10,000 permutations to estimate the significance of that correlation. We chose this  
216 approach as our goal was to get a broad sense of the similarity of within species brain region  
217 correlation patterns and how these vary between species, rather than to establish specific  
218 mechanisms of brain region covariation. Understanding the mechanisms of how and why brain  
219 regions covary and how these differ among species would require additional data including genetic  
220 data or the use of selection or breeding experiments, though this was not the focus of this study.

221

222 *Phylogenetic distance comparison*

223 Finally, to test whether more closely related species have more similar patterns of brain region

224 correlation, we compared phylogenetic distance between species to the similarity of their

225 correlation matrices. We used the correlations estimated from the Mantel test comparing

226 individual species to construct a species matrix including all pairwise species comparisons (figure 4).

227 We then generated a phylogenetic distance matrix showing time (in millions of years) since the

228 most recent common ancestor for each pairwise species comparison (supplementary figure 3).

229 Time since most recent common ancestor for each species comparison was gathered from

230 Timetree.org. We then compared these two species level matrices using the same Mantel test

231 approach as we used to compare brain region correlation matrices. In this case, if more closely

232 related species share more similar brain region correlation patterns, we expect a negative

233 correlation between the species level matrices.

234

235 **Results**

236 *Intraspecific brain region principal component variation*

237 All brain regions covaried in the same direction along PC1, which reflected total brain volume

238 (figure 2), but the percent variation explained by this axis varied across the 5 different fish orders

239 (supplementary figure 4). The Cypriniformes had the highest amount of variation explained by PC1

240 (90.06% for *S. atromaculatus* and 95.03% for the *R. atratulus*), whereas the Scorpaeniformes and

241 Osteoglossiformes had lower amounts of brain region variation explained by total brain volume

242 (~65-70% across the four species in these orders). The Perciformes and Cyprinodontiformes had  
243 more intermediate levels of variation explained by PC1 (~75-85%).

244

245 There was a noticeable difference in spread along the PC2 axis (for the five brain regions),  
246 indicating the differing strengths of independent contributions separate regions have on brain  
247 variation (figure 2). There was a split between species where the olfactory bulbs contributed  
248 disproportionately to most of the percent variation described by PC2, (~6-10% for  
249 Cyprinodontiformes, 14-20% for Scorpaeniformes, ~10-13% for Perciformes), and those where  
250 multiple regions contributed to the variation along PC2. These include the Cypriniformes, which  
251 had relatively little variation and spread explained by PC2 in any brain region (~2-4%), and the  
252 Osteoglossiformes, for which different brain regions contributed to varying degrees to PC2 in the  
253 two species studied (~13-14%). In *P. buchholzi*, the hypothalamus contributed the most to PC2,  
254 followed by telencephalon, optic tectum, olfactory bulb, and finally cerebellum. By contrast, the  
255 hypothalamus and olfactory bulb contributed the most to PC2 in *B. brachystius*, followed by the  
256 optic tectum, cerebellum, and finally telencephalon. Interestingly, the cerebellum and optic tectum  
257 loaded in the same direction for all species except *B. brachystius*.

258

259 *Intraspecific brain region correlation*

260 Across all species, brain region comparisons showed a positive correlation, though the strength of  
261 these correlations varied within and among species (figure 3). All these positive correlations were  
262 statistically significant, other than the olfactory bulb and cerebellum comparison, and the olfactory  
263 bulb and optic tectum comparison, in *P. pungitius*. To get a more general sense of within species

264 brain region independence, for every species we calculated the average correlation for each brain  
265 region with the other regions of the brain. This revealed that species vary in their overall level of  
266 brain region covariation, with average correlations ranging from 0.56 in *G. aculeatus* and *P.*  
267 *pungitius* to 0.94 in *R. atratulus* (table 3). The Cypriniformes showed very high levels of region  
268 correlations, with all regions correlating with each other by at least 0.82 in *S. atromaculatus* and  
269 0.88 in *R. atratulus*. Further, in all fish species other than the two Osteoglossiform species (*P.*  
270 *buchholzi* and *B. brachystius*), the olfactory bulb showed the lowest average correlation with the  
271 other brain regions, though in the Cypriniformes this was a very small difference (0.03 or less  
272 compared to other region averages) (table 3). In both Osteoglossiform species, the hypothalamus  
273 showed the lowest average correlation with the rest of the brain regions (0.51 for *P. buchholzi* and  
274 0.52 for *B. brachystius*). The correlation between optic tectum size and telencephalon size showed  
275 the highest correlation of any region comparison in every species other than *P. buchholzi*, which  
276 showed the highest correlation between the optic tectum and cerebellum (table 3).  
277

#### 278 *Species comparison*

279 Within species brain region covariation patterns correlated across certain fish species (figure 4). *P.*  
280 *reticulata*, *G. affinis*, *L. gibbosus*, *L. macrochirus*, *G. aculeatus*, and *P. pungitius* all showed relatively  
281 high correlations of their brain region covariation patterns (at least 0.751), with all pairwise  
282 correlations being significant other than *P. pungitius* with *P. reticulata*, *P. pungitius* with *L.*  
283 *macrochirus*, and *P. pungitius* with *G. aculeatus* (figure 4). This indicates that these groups all share  
284 a similar pattern of brain region correlations. *S. atromaculatus* and *R. atratulus* also share some  
285 similar patterns with this broad group, though the correlations were weaker (figure 4). *S.*

286 *atromaculatus* showed a significant correlation with the brain region covariation patterns of *P.*  
287 *reticulata* ( $r=0.784$ ), *L. macrochirus* ( $r=0.681$ ), and *G. aculeatus* ( $r=0.581$ ). *R. atratulus* correlated  
288 with *P. reticulata* ( $r=0.773$ ) and *L. macrochirus* ( $r=0.770$ ). *S. atromaculatus* and *R. atratulus* also  
289 showed a strong significant correlation with each other, at 0.833. Both Osteoglossiform species  
290 showed divergence in their brain region covariation patterns, with the only significant correlation  
291 occurring between *B. brachystius* and *R. atratulus* (0.839). The covariation pattern of *P. buchholzi*  
292 was not significantly correlated with any other species.

293

294 ~~We note that differences in average brain region correlation levels among species observed here~~  
295 ~~may be related to differences in body size variation, as greater body size variation could establish~~  
296 ~~the possibility for stronger correlations. However, this effect would not impact our ability to~~  
297 ~~compare the degree of similarity of correlation matrices, as the Mantel test comparison is not~~  
298 ~~affected by differences in average matrix values. To support this assumption, we compared the~~  
299 ~~similarity of the *S. atromaculatus* matrix and a correlation matrix calculated from a truncated *S.*~~  
300 ~~*atromaculatus* data set with the largest (>20g) and smallest (<2g) individuals removed, resulting in~~  
301 ~~a lower coefficient of body mass variation of 0.55 (supplementary figure 5). These matrices were~~  
302 ~~highly correlated with each other ( $r=0.96$ ,  $p=0.0083$ ), indicating that differences in average levels of~~  
303 ~~trait correlations do not impact the outcomes of our Mantel test. As such, we will only consider~~  
304 ~~results related to differences related to comparisons among regions within species, and how these~~  
305 ~~vary across species, and not discuss overall average levels of brain region correlation within~~  
306 ~~species.~~

307 We note that differences in average brain region correlation levels among species observed here  
308 may be related to differences in body size variation (table 1), as greater body size variation could  
309 establish the possibility for stronger correlations. However, this effect would not impact our ability  
310 to compare the degree of similarity of correlation matrices, as the Mantel test comparison is not  
311 affected by differences in average matrix values. To support this assumption, we used an additional  
312 analysis of the species in our data set with the highest body mass coefficient of variation, *S.  
313 atromaculatus*. We compared the similarity of the *S. atromaculatus* matrix and a correlation matrix  
314 calculated from a truncated *S. atromaculatus* data set with the largest (>20g) and smallest (<2g)  
315 individuals removed, resulting in a lower coefficient of body mass variation of 0.55 (supplementary  
316 figure 5). These matrices were highly correlated with each other ( $r=0.96$ ,  $p=0.0083$ ), indicating that  
317 differences in average levels of trait correlations do not impact the outcomes of our Mantel test. As  
318 such, we will only consider results related to differences related to comparisons among regions  
319 within species, and how these vary across species, and not discuss overall average levels of brain  
320 region correlation within species.

Formatted: Font: Italic

321  
322 Brain region covariation patterns appeared to be influenced by phylogenetic distance. We found  
323 that the similarity in brain region covariation patterns was inversely correlated with phylogenetic  
324 distance (figure 5). This trend was not significant ( $r=-0.671$ ,  $p=0.999$ ).  
325

## 326 Discussion

327 We tested the degree of covariation of brain region sizes within 10 teleost fish species to examine  
328 how brain region covariation patterns vary across species. Our results showed that brain regions

329 varied in the strength of their covariation within and across species, with the telencephalon and  
330 optic tectum usually showing very high levels of covariation, and the olfactory bulb showing lower  
331 covariation with the rest of the brain. Further, we found an insignificant trend that patterns of  
332 brain region covariation are shared across more closely related species. Specifically, this trend  
333 indicates that the similarity of within species brain region correlation patterns decreases as  
334 phylogenetic distance between species increases.

335

336 *Within species patterns of brain region covariation*

337 Patterns of brain region volume covariation within species indicate the potential for independent  
338 change in brain region size. These patterns can arise through various, non-mutually exclusive  
339 mechanisms, including differences in the strength of selection on different brain regions, or from  
340 genetic and developmental constraints (Davidowitz et al., 2012; Peiman & Robinson, 2017).

341

342 Covariation between the sizes of brain regions can occur due to patterns of selection acting on  
343 brain regions independently. This pattern can occur when selection consistently acts in similar ways  
344 on different traits (Sinervo & Svensson, 2002). For example, we observed that the telencephalon  
345 and the optic tectum showed a high level of correlation in most species. If selection consistently  
346 acts in the same direction on these two regions, it could result in this pattern. Similarly, the high  
347 level of independence of the olfactory bulb in most of the species we studied could result from  
348 selection on this specific region acting in a different direction to other regions of the brain. We  
349 consider this explanation to be unlikely to explain most of the variation we see in brain region  
350 covariation for two reasons. First, we found similar patterns in brain region covariation across

351 several fish species. Selection would have needed to be consistent in its effect on all brain regions  
352 across all these species regardless of population or environment, which decreases its likelihood as  
353 an explanation. Second, prior research suggests that selection can act differently across brain  
354 regions, which would result in less covariation across regions and is unlikely to then lead to very  
355 consistent patterns in brain region covariation between species. For example, Gonzalez-Voyer &  
356 Kolm (2010) found that sexual selection in Lake Tanganyikan Cichlid *species* resulted in divergence  
357 in the size of the hypothalamus and cerebellum, with other regions of the brain diverging in the  
358 opposite direction. Further, Kotrschal et al. (2017) found that Trinidadian guppy brain regions  
359 differed in how they diverged in response to predation levels across populations. Although we  
360 consider other explanations of the patterns we observed to be more likely, we cannot rule out  
361 correlated patterns of selection. Further research on the consistency of how selection acts on  
362 different brain regions within and across species would be needed to fully evaluate this possibility.

363

364 The other possible mechanism that can generate covariation between traits is some type of  
365 constraint, including genetic constraints and developmental constraints. Genetic constraints occur  
366 through pleiotropy (a single gene being associated with multiple phenotypic traits) or gene linkage  
367 (genes for different traits occurring close together on the same chromosome), limiting the ability of  
368 traits to evolve independently of each other (Ott et al., 2015; Solovieff et al., 2013). Although the  
369 degree of pleiotropy in determining brain region size has not been specifically measured, prior  
370 work has suggested that aspects of brain connectivity and structure are influenced by pleiotropic or  
371 linked genes (Moreau et al., 2022; van der Meer et al., 2020; Zhao et al., 2021). It is therefore likely  
372 that the evolution of the size of brain regions may be similarly influenced by genetic architecture

373 constraints. The strength of these genetic links between brain regions could thus result in the  
374 patterns of brain region covariation that we observed across species.

375

376 Developmental constraints are limitations to the variability of traits caused by the structure or  
377 dynamics of organism ontogeny (Smith et al., 1985). This was the form of constraint proposed to  
378 explain the covariation of brain regions among mammals under the concerted brain evolution  
379 hypothesis, where the evolution of larger brain size was linked to increases in the relative size of  
380 later developing brain regions (Finlay & Darlington, 1995). As fish have indeterminate growth and  
381 maintain widespread neurogenesis of the brain into adulthood (Zupanc, 2006), this specific process  
382 is unlikely to explain our results. However, a similar kind of developmental limitation may. The  
383 growth of different brain regions may be influenced by overlapping developmental mechanisms  
384 such as the ability to generate new neurons in specific areas of the brain (Ganz & Brand, 2016;  
385 Kaslin et al., 2008). We are not able to determine specific mechanisms generating the covariation  
386 patterns we observed, and future work examining the genetic architecture and developmental  
387 processes of brain region evolution will be needed to fully elucidate these.

388

389 *Evolution of brain region covariation patterns*

390 We found a trend that more closely related species tend to share similar patterns of brain region  
391 covariation, while these patterns diverge as phylogenetic distance increases. Our results indicate  
392 that Perciformes and Cyprinodontiformes show strong evidence of a shared pattern of brain region  
393 covariation. Scorpaeniformes also show a similar pattern to these orders, but with **higher overall**  
394 **levels of brain region independence and** less consistency, as *P. pungitius* appear to be less similar to

395 the rest of these species. These six species are closely related in evolutionary terms compared to  
396 the rest of the species we included, with a common ancestor ~112 million years ago, though they  
397 differ in terms of ecology, particularly between orders (Lowe-Mcconnell, 2012; Magalhaes et al.,  
398 2016). This similarity suggests that the constraints limiting independent brain region change are  
399 evolutionarily conserved across closely related fish species, even when ecology, and likely selection  
400 pressures, differ.

401  
402 Our two Cypriniform species (*S. atromaculatus* and *R. atratulus*) show similar patterns to each  
403 other, particularly in terms of overall level of brain region independence, as both show very little  
404 independence in brain regions. The Cypriniform brain differs morphologically from other fish as  
405 they are characterized by extended and somewhat enlarged olfactory bulbs (Brandstätter &  
406 Ketschach, 1990; Evans, 1952). Beyond this morphological difference, it is unclear why these species  
407 have such strong correlations between their brain regions. Broadly, this result indicates that  
408 selection would need to be very strong in order to shift individual brain regions in these species  
409 independent of the rest of the brain, as evolution would need to break this strong covariation  
410 pattern, potentially limiting the ability of these species to respond to selective pressures.

411  
412 Our results indicate that patterns of intraspecific brain region covariation can evolve, and these  
413 patterns are more similar between more closely related species. The link between phylogenetic  
414 distance and within species brain region covariation is supported by two pieces of evidence here.  
415 First, qualitatively, our results support this conclusion as we found high similarity in brain region  
416 covariation patterns among closely related species, and the most distantly related species in our

417 sample, the two Osteoglossiform species, show very little similarity to each other and to the rest of  
418 the species in their brain region covariation patterns. This order diverged from the rest of the  
419 species in our samples approximately 263 million years ago. *B. brachystomus* and *P. buchholzi*  
420 diverged from each other approximately 200 million years ago, much greater than the divergence  
421 time between the species in the other orders we tested. Second, our quantitative comparison  
422 suggests that phylogenetic distance between the species is negatively correlated with the similarity  
423 of their brain region covariation patterns, though this pattern was not statistically significant. This is  
424 likely due to the limited variation in time since the most recent common ancestor, which restricts  
425 the statistical power of permutation tests. Permutation tests rely on variability in the data to allow  
426 for estimating a random distribution of potential results. Many species comparisons here share the  
427 same time since most recent common ancestor, resulting in extremely limited variation from which  
428 the permutation test can derive a results distribution. Our evidence of a negative correlation  
429 between brain region correlation patterns and phylogenetic relatedness is therefore limited in its  
430 reliability. Together these indicate that evolutionary time is required to break the constraints that  
431 cause these patterns of brain region covariation, potentially through selection acting strongly in an  
432 opposite direction on correlated regions. Understanding the specific selection forces that lead to  
433 divergence in brain region covariation patterns will require further study focusing on mechanisms  
434 and evolution of constraints in brain region correlation.

435

436 Beyond more ancient divergence times between each other and the remainder of the species, the  
437 two Osteoglossiform species possess phenotypic distinctiveness in comparison to the other teleosts  
438 studied here. For example, *P. buchholzi*, a surface-oriented fish, exhibits the slowest rate of

439 morphological divergence among other studied Osteoglossiformes, despite tens of millions of years  
440 of genetic divergence between populations (Lavoué et al., 2011). This high level of phenotypic  
441 stability over such a long timescale in this fish lineage is thought to surpass all other known  
442 examples of morphological stasis in extant vertebrates. The other osteoglossomorph we studied, *B.*  
443 *brachyistius*, is in the superfamily Mormyroidea, a group of African freshwater fishes that have  
444 electromotor and electrosensory systems (Carlson & Arnegard, 2011; Crampton, 2019). In addition,  
445 mormyroids also possess extremely large brains (Nilsson, 1996; Sukhum et al., 2016, 2018), a suite  
446 of diverse craniofacial morphologies (Peterson et al., 2022), large intra- and interspecific variety in  
447 communication signals and behaviors (Hopkins 1986), and are the only known vertebrates whose  
448 sperm lack flagella (Saunders & Gallant, 2024). These novel phenotypic features in both *B.*  
449 *brachyistius* and *P. buchholzi*, along with their ancient divergence times, could potentially be  
450 related to breaks in genetic and/or developmental constraints maintained within other teleost  
451 species. However, further research is needed comparing more Osteoglossiform species across a  
452 greater variety of phylogenetic distances to examine how covariation in brain region sizes has been  
453 maintained over time in this specific group.

454

#### 455 *Limitations*

456 There are key limitations to our study that must be considered. First, due to logistical reasons the  
457 number of species we sampled here was low. Our focus was on building enough within species  
458 samples to confidently measure within species brain region covariation, which made sampling a  
459 larger number of species difficult. This lower representation of species particularly limited our  
460 ability to adequately test the within and across species trait covariation patterns in a

461 phylogenetically controlled way. Second, we built our database using a combination of pre-  
462 collected data and fish samples and newly sampled species, resulting in variation in sampling  
463 techniques, including fish that were reared in labs and wild collected fish. This benefits our study in  
464 some ways, notably by increasing the possible variability between species, making our observation  
465 of similar patterns across species stronger. However, by doing this we are unable to determine how  
466 differences in selection or environmental variation may be impacting our results, limiting our ability  
467 to elucidate the mechanisms generating the covariation patterns we see. Finally, we do not have  
468 any genetic information about the patterns of covariation we observed. Previous work on  
469 evolutionary constraints caused by the covariation of traits has focused on genetic covariation  
470 (Davidowitz et al., 2012; Lande, 1984; Olson & Miller, 1958; Ott et al., 2015), with the goal of linking  
471 trait covariation to the evolutionary mechanism of constraint. We also are unable to determine the  
472 heritability of patterns of trait covariation we observed. The genetic data needed for this type of  
473 analysis were not available to us. Future work should examine both the heritability of brain region  
474 covariation patterns as well as the genetic architecture causing these patterns of covariation.

475

476 *Conclusion*

477 Here, using large, within-species data sets, we examined the degree of covariation of brain region  
478 sizes within 10 fish species, and tested whether those patterns vary across species. Our results  
479 indicate that brain regions vary in their level of covariation, and that these patterns are shared  
480 between closely related species of fish. Broadly, these results could indicate that the size of  
481 individual brain regions is constrained in its ability to evolve within species, and that it takes a great  
482 deal of evolutionary time to break or change these constraints. Future work will be needed to

483 elucidate the specific nature of these constraints, as well as the evolutionary processes that can  
484 break these patterns.

485

486

487

488

489

490 Tables and Figures:

491

492 Table 1. List of fish species used, including the source of the data and the sample size.

Common Name	Scientific Name	Order	Source	N	Coefficient of variation in body mass
Pumpkinseed Sunfish	<i>Lepomis gibbosus</i>	Perciformes	Axelrod et al. 2021	113	0.59
Bluegill Sunfish	<i>Lepomis macrochirus</i>	Perciformes	Axelrod et al. 2021	94	0.52
Threespine Stickleback	<i>Gasterosteus aculeatus</i>	Scorpaeniformes	Herczeg et al. 2015	231	0.24
Ninespine Stickleback	<i>Pungitius pungitius</i>	Scorpaeniformes	Gonda et al. 2009	120	0.37
Trinidadian Guppy	<i>Poecilia reticulata</i>	Cyprinodontiformes	Authors	296	0.46
Mosquitofish	<i>Gambusia affinis</i>	Cyprinodontiformes	Authors	133	0.68
Creek Chub	<i>Semotilus atromaculatus</i>	Cypriniformes	Authors	194	0.91
Black Nose Dace	<i>Rhinichthys atratulus</i>	Cypriniformes	Authors	82	0.8
African Butterflyfish	<i>Pantodon buchholzi</i>	Osteoglossiformes	Authors	61	0.39
Black baby whale	<i>Brienomyrus brachyistius</i>	Osteoglossiformes	Authors	56	0.57

493

494

495 Table 2. Collection details for each fish species used.

Species	Collection Details
<i>Lepomis gibbosus</i>	Wild collected fish, collected via angling from the wild in summer 2016 and 2017 from four pelagic sites and four littoral sites of Ashby Lake, Ontario.
<i>Lepomis macrochirus</i>	Wild collected fish, collected via angling from the wild in summer 2017 from one pelagic site and one littoral site of Holcomb Lake, Michigan.
<i>Gasterosteus aculeatus</i>	F1 offspring of wild collected fish from the Baltic Sea in 2011. F1 fish were reared until adulthood in either an enriched environment or a simple environment.
<i>Pungitius pungitius</i>	F1 offspring of wild collected fish from four sites in the Baltic Sea in 2011. F1 fish were reared until adulthood either alone or in groups of 20 fish.
<i>Poecilia reticulata</i>	F2 offspring of wild collected fish from two populations in Trinidad, Aripo high predation (HP) and Aripo low predation (LP). F2 fish from both populations were reared until sexual maturity under two predator environments (pred+ and pred-) and three social environments (Solo, with HP conspecifics, and with LP conspecifics).
<i>Gambusia affinis</i>	Wild collected fish from 4 freshwater sites in southern Louisiana in the summer of 2022. Fish were collected with dip nets from one high salinity site and one low salinity site in each of two streams.
<i>Semotilus atromaculatus</i>	Wild collected via electrofishing in the summer of 2018 and 2019 from 17 freshwater stream representing a gradient of agricultural intensity within 6 major watersheds in Southwest Ontario, Canada.
<i>Rhinichthys atratulus</i>	Wild collected fish from Fall Creek, NY. Fish were collected in November of 2023 using seine nets.
<i>Pantodon buchholzi</i>	Sourced from tropical fish supplier
<i>Brienomyrus brachyistius</i>	Sourced from tropical fish supplier

496

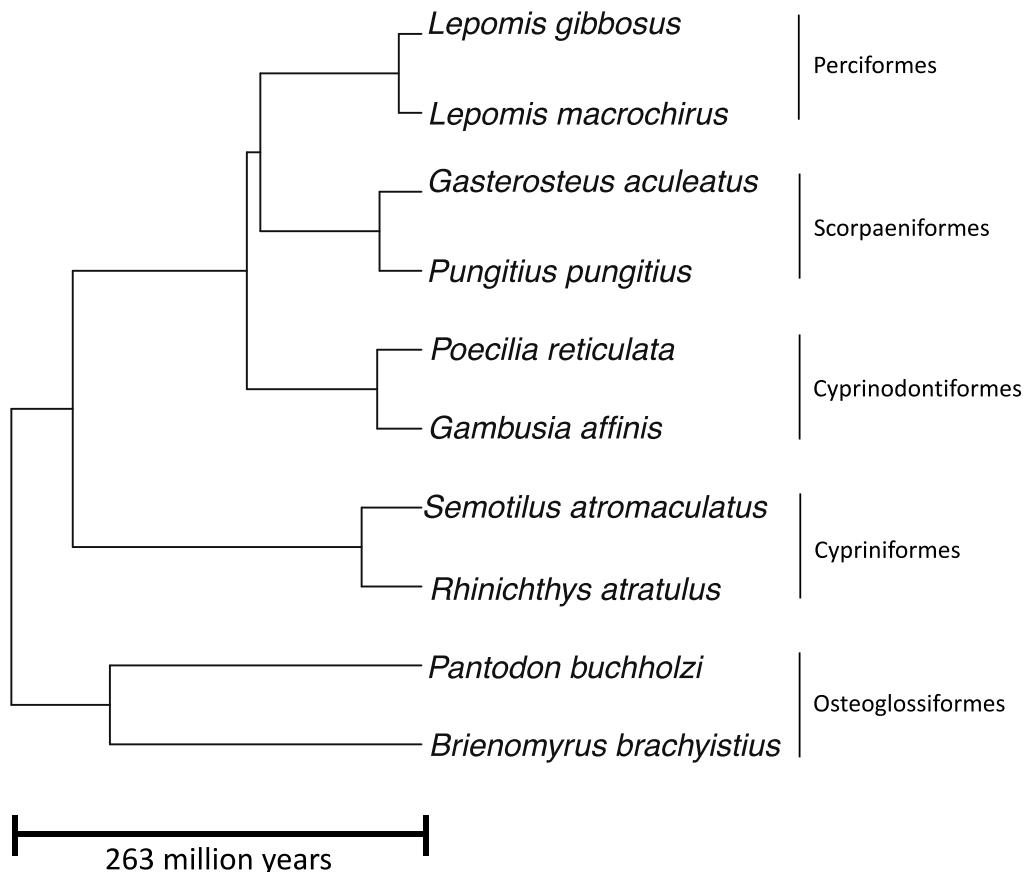
497

498 Table 3. Average correlation coefficients for brain regions across species. Averages are calculated  
499 from the correlations of each region with every other region in each species.

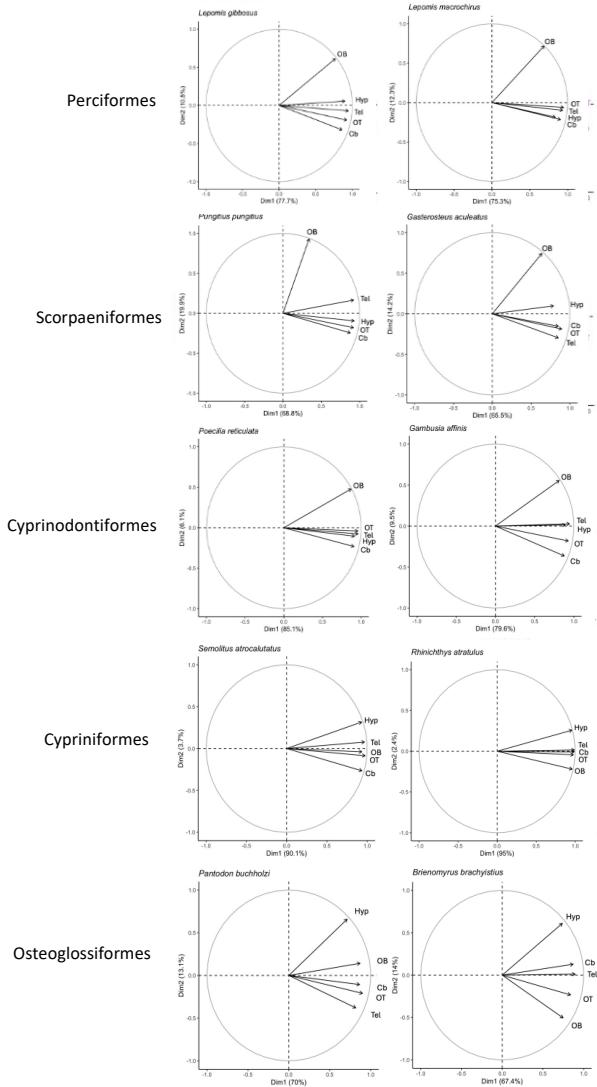
Species	Cb	OT	Tel	OB	Hyp	Total
<i>Poecilia reticulata</i>	0.72	0.78	0.79	0.64	0.72	0.73
<i>Gambusia affinis</i>	0.73	0.78	0.80	0.67	0.74	0.75
<i>Lepomis gibbosus</i>	0.70	0.76	0.79	0.61	0.74	0.72
<i>Lepomis macrochirus</i>	0.72	0.76	0.75	0.53	0.66	0.68
<i>Gasterosteus aculeatus</i>	0.60	0.64	0.59	0.43	0.55	0.56
<i>Pungitius pungitius</i>	0.59	0.64	0.64	0.23	0.66	0.56
<i>Semotilus atromaculatus</i>	0.86	0.91	0.90	0.86	0.86	0.88
<i>Rhinichthys atratulus</i>	0.94	0.95	0.96	0.92	0.92	0.94
<i>Pantodon buchholzi</i>	0.65	0.69	0.62	0.66	0.51	0.63
<i>Brienomyrus brachyistius</i>	0.64	0.61	0.67	0.53	0.52	0.59

500

501  
502  
503  
504  
505



506  
507 Figure 1. Phylogenetic relationships between the 10 teleost fish species in our study.  
508



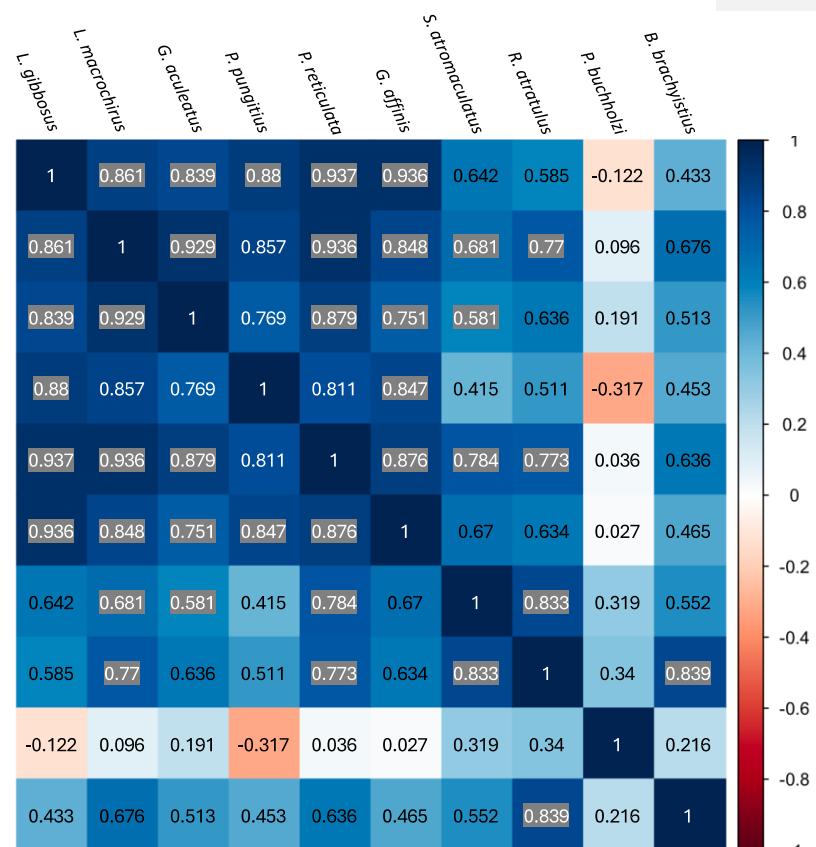
509  
510  
511  
512  
513  
514

Figure 2: PC loading vectors of each brain region for each species, for PC1 (Dim1, correlated with total brain volume) and PC2 (Dim2, correlated with covariation in brain regions not related to total brain size). Cb, cerebellum; Hyp, hypothalamus; OB, olfactory bulbs; OT, optic tectum; Tel, telencephalon.

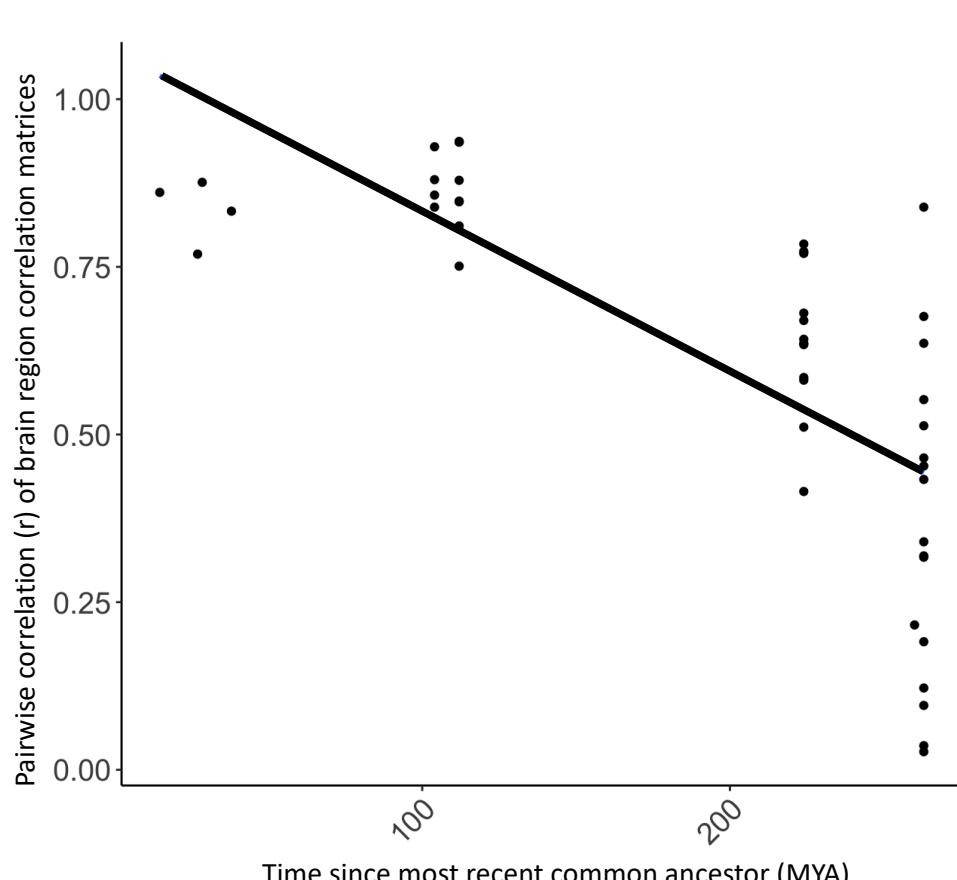
		Cb	OT	Tel	OB	Hyp	
<b>Perciformes</b>	<i>Lepomis gibbosus</i>	1	0.79	0.78	0.52	0.71	
		OT	0.79	1	0.9	0.6	0.76
		Tel	0.76	0.9	1	0.66	0.82
		OB	0.52	0.6	0.66	1	0.66
		Hyp	0.71	0.76	0.82	0.66	1
<b>Scorpaeniformes</b>	<i>Lepomis macrochirus</i>	1	0.86	0.83	0.48	0.7	
		OT	0.86	1	0.88	0.59	0.72
		Tel	0.83	0.88	1	0.56	0.73
		OB	0.48	0.59	0.56	1	0.47
		Hyp	0.7	0.72	0.73	0.47	1
<b>Cyprinodontiformes</b>	<i>Gasterosteus aculeatus</i>	1	0.74	0.65	0.43	0.56	
		OT	0.74	1	0.76	0.45	0.59
		Tel	0.65	0.76	1	0.37	0.59
		OB	0.43	0.45	0.37	1	0.46
		Hyp	0.56	0.59	0.59	0.46	1
<b>Cypriniformes</b>	<i>Pungitius pungitius</i>	1	0.78	0.72	0.1	0.79	
		OT	0.78	1	0.8	0.15	0.83
		Tel	0.72	0.8	1	0.43	0.79
		OB	0.1	0.15	0.43	1	0.23
		Hyp	0.79	0.83	0.79	0.23	1
<b>Osteoglossiformes</b>	<i>Poecilia reticulata</i>	1	0.79	0.78	0.6	0.71	
		OT	0.79	1	0.9	0.78	0.75
		Tel	0.78	0.9	1	0.67	0.8
		OB	0.6	0.68	0.67	1	0.62
		Hyp	0.71	0.75	0.8	0.62	1
<b>Pantodontidae</b>	<i>Gambusia affinis</i>	1	0.82	0.8	0.56	0.73	
		OT	0.82	1	0.87	0.65	0.78
		Tel	0.8	0.87	1	0.77	0.77
		OB	0.56	0.65	0.77	1	0.69
		Hyp	0.73	0.78	0.77	0.69	1
<b>Sisoridae</b>	<i>Semotilus atromaculatus</i>	1	0.92	0.87	0.83	0.82	
		OT	0.92	1	0.93	0.9	0.88
		Tel	0.87	0.93	1	0.88	0.9
		OB	0.83	0.9	0.88	1	0.83
		Hyp	0.82	0.88	0.9	0.83	1
<b>Rhinichthys atratulus</b>	<i>Pantodon buchholzi</i>	1	0.97	0.95	0.93	0.92	
		OT	0.97	1	0.97	0.93	0.93
		Tel	0.95	0.97	1	0.95	0.95
		OB	0.93	0.93	0.85	1	0.88
		Hyp	0.92	0.93	0.95	0.88	1
<b>Brienomyrus brachystomus</b>	<i>Rhinichthys atratulus</i>	1	0.97	0.95	0.93	0.92	
		OT	0.97	1	0.97	0.93	0.93
		Tel	0.95	0.97	1	0.95	0.95
		OB	0.93	0.93	0.85	1	0.88
		Hyp	0.92	0.93	0.95	0.88	1



516 Figure 3. Matrices showing each pairwise Pearson correlation among five brain regions for all 10  
 517 fish species: cerebellum (Cb), optic tectum (OT), telencephalon (Tel), olfactory bulb (OB), and  
 518 hypothalamus (Hyp). Box color shows the strength of the correlations as indicated by the scale bar  
 519 on the right, and numbers show the correlation coefficient. Statistically significant correlations are  
 520 shown as white numbers, and non-significant correlations are shown as black numbers: all  
 521 correlations are significant other than the Cb/OB and OT/OB correlations in *P. pungitius*.  
 522



523  
 524 Figure 4. Matrix showing the pairwise Pearson correlations of within species brain region size  
 525 correlation matrices for each of the 10 fish species in our study. Box color shows the strength of the  
 526 correlations as indicated by the scale bar on the right. Numbers show the correlation coefficient (r)  
 527 for each comparison. Gray boxes indicate significant correlations. Numbers are colored for optimal  
 528 visibility.  
 529





568 Davidowitz, G., Nijhout, H. F., & Roff, D. A. (2012). Predicting the response to simultaneous  
569 selection: Genetic architecture and physiological constraints. *Evolution; International  
570 Journal of Organic Evolution*, 66(9), 2916–2928. [https://doi.org/10.1111/j.1558-  
571 5646.2012.01644.x](https://doi.org/10.1111/j.1558-<br/>571 5646.2012.01644.x)

572 Finlay, B., & Darlington, R. (1995). Linked regularities in the development and evolution of  
573 mammalian brains. *Science*, 268(5217), 1578–1584.  
574 <https://doi.org/10.1126/science.7777856>

575 Ganz, J., & Brand, M. (2016). Adult Neurogenesis in Fish. *Cold Spring Harbor Perspectives in Biology*,  
576 8(7), a019018. <https://doi.org/10.1101/cshperspect.a019018>

577 Gonzalez-Voyer, A., & Kolm, N. (2010). Sex, Ecology and the Brain: Evolutionary Correlates of Brain  
578 Structure Volumes in Tanganyikan Cichlids. *PLoS ONE*, 5(12), e14355.  
579 <https://doi.org/10.1371/journal.pone.0014355>

580 Hager, R., Lu, L., Rosen, G. D., & Williams, R. W. (2012). Genetic architecture supports mosaic brain  
581 evolution and independent brain–body size regulation. *Nature Communications*, 3, 1079.  
582 <https://doi.org/10.1038/ncomms2086>

583 Healy, S. D., & Rowe, C. (2007). A critique of comparative studies of brain size. *Proceedings of the  
584 Royal Society B: Biological Sciences*, 274(1609), 453–464.  
585 <https://doi.org/10.1098/rspb.2006.3748>

586 Hoops, D., Vidal-García, M., Ullmann, J. F. P., Janke, A. L., Stait-Gardner, T., Duchêne, D. A., Price,  
587 W. S., Whiting, M. J., & Keogh, J. S. (2017). Evidence for Concerted and Mosaic Brain  
588 Evolution in Dragon Lizards. *Brain, Behavior and Evolution*, 90(3), 211–223.  
589 <https://doi.org/10.1159/000478738>

590 Huber, R., van Staaden, M. J., Kaufman, L. S., & Liem, K. F. (1997). Microhabitat use, trophic  
591 patterns, and the evolution of brain structure in African cichlids. *Brain, Behavior and  
592 Evolution*, 50(3), 167–182. <https://doi.org/10.1159/000113330>

593 Jones, A. G., Bürger, R., & Arnold, S. J. (2014). Epistasis and natural selection shape the mutational  
594 architecture of complex traits. *Nature Communications*, 5(1), 3709.  
595 <https://doi.org/10.1038/ncomms4709>

596 Kaslin, J., Ganz, J., & Brand, M. (2008). Proliferation, neurogenesis and regeneration in the non-  
597 mammalian vertebrate brain. *Philosophical Transactions of the Royal Society B: Biological  
598 Sciences*, 363(1489), 101–122. <https://doi.org/10.1098/rstb.2006.2015>

599 Klingenberg, C. P. (1996). Individual Variation of Ontogenies: A Longitudinal Study of Growth and  
600 Timing. *Evolution*, 50(6), 2412–2428. <https://doi.org/10.1111/j.1558-5646.1996.tb03628.x>

601 Kotrschal, A., Rogell, B., Bundsen, A., Svensson, B., Zajitschek, S., Bränström, I., Immler, S.,  
602 Maklakov, A. A., & Kolm, N. (2013). Artificial Selection on Relative Brain Size in the Guppy  
603 Reveals Costs and Benefits of Evolving a Larger Brain. *Current Biology*, 23(2), 168–171.  
604 <https://doi.org/10.1016/j.cub.2012.11.058>

605 Kotrschal, A., Zeng, H.-L., Bijl, W. van der, Öhman-Mägi, C., Kotrschal, K., Pelckmans, K., & Kolm, N.  
606 (2017). Evolution of brain region volumes during artificial selection for relative brain size.  
607 *Evolution*, 71(12), 2942–2951. <https://doi.org/10.1111/evo.13373>

608 Kotrschal, K., Van Staaden, M. J., & Huber, R. (1998). Fish brains: Evolution and environmental  
609 relationships. *Reviews in Fish Biology and Fisheries*, 8(4), 373–408.  
610 <https://doi.org/10.1023/A:1008839605380>

611 Laberge, F., & Hara, T. J. (2001). Neurobiology of fish olfaction: A review. *Brain Research. Brain*  
612 *Research Reviews*, 36(1), 46–59. [https://doi.org/10.1016/s0165-0173\(01\)00064-9](https://doi.org/10.1016/s0165-0173(01)00064-9)

613 Lande, R. (1984). The genetic correlation between characters maintained by selection, linkage and  
614 inbreeding. *Genetical Research*, 44(3), 309–320.  
615 <https://doi.org/10.1017/s0016672300026549>

616 Lavoué, S., Miya, M., Arnegard, M. E., McIntyre, P. B., Mamonekene, V., & Nishida, M. (2010).  
617 Remarkable morphological stasis in an extant vertebrate despite tens of millions of years of  
618 divergence. *Proceedings of the Royal Society B: Biological Sciences*, 278(1708), 1003–1008.  
619 <https://doi.org/10.1098/rspb.2010.1639>

620 Lowe-Mcconnell, R. (2012). Ecology and Evolution of Poeciliid Fishes. *Zoological Journal of the*  
621 *Linnean Society*, 166(3), 688–688. <https://doi.org/10.1111/j.1096-3642.2012.00843.x>

622 MacLean, E. L., Hare, B., Nunn, C. L., Addessi, E., Amici, F., Anderson, R. C., Aureli, F., Baker, J. M.,  
623 Bania, A. E., Barnard, A. M., Boogert, N. J., Brannon, E. M., Bray, E. E., Bray, J., Brent, L. J. N.,  
624 Burkart, J. M., Call, J., Cantlon, J. F., Cheke, L. G., ... Zhao, Y. (2014). The evolution of self-  
625 control. *Proceedings of the National Academy of Sciences*, 111(20), E2140–E2148.  
626 <https://doi.org/10.1073/pnas.1323533111>

627 Magalhaes, I. S., D'Agostino, D., Hohenlohe, P. A., & MacColl, A. D. C. (2016). The ecology of an  
628 adaptive radiation of three-spined stickleback from North Uist, Scotland. *Molecular Ecology*,  
629 25(17), 4319. <https://doi.org/10.1111/mec.13746>

630 McGlothlin, J. W., & Ketterson, E. D. (2008). Hormone-mediated suites as adaptations and  
631 evolutionary constraints. *Philosophical Transactions of the Royal Society B: Biological*  
632 *Sciences*, 363(1497), 1611–1620. <https://doi.org/10.1098/rstb.2007.0002>

633 Moreau, C. A., Kumar, K., Harvey, A., Huguet, G., Urchs, S. G. W., Schultz, L. M., Sharmarke, H., Jizi,  
634 K., Martin, C.-O., Younis, N., Tamer, P., Martineau, J.-L., Orban, P., Silva, A. I., Hall, J., van  
635 den Bree, M. B. M., Owen, M. J., Linden, D. E. J., Lippé, S., ... Jacquemont, S. (2022). Brain  
636 functional connectivity mirrors genetic pleiotropy in psychiatric conditions. *Brain*, 146(4),  
637 1686–1696. <https://doi.org/10.1093/brain/awac315>

638 Nilsson, G. E. (1996). Brain and Body Oxygen Requirements of *Gnathonemus Petersii*, a Fish with an  
639 Exceptionally Large Brain. *Journal of Experimental Biology*, 199(3), 603–607.  
640 <https://doi.org/10.1242/jeb.199.3.603>

641 Noreikiene, K., Herczeg, G., Gonda, A., Balázs, G., Husby, A., & Merilä, J. (2015). Quantitative  
642 genetic analysis of brain size variation in sticklebacks: Support for the mosaic model of brain  
643 evolution. *Proceedings of the Royal Society B: Biological Sciences*, 282(1810), 20151008.  
644 <https://doi.org/10.1098/rspb.2015.1008>

645 Olson, E. C., & Miller, R. L. (1999). *Morphological Integration*. University of Chicago Press.  
646 <https://press.uchicago.edu/ucp/books/book/chicago/M/bo3620375.html>

647 Ott, J., Wang, J., & Leal, S. M. (2015). Genetic linkage analysis in the age of whole-genome  
648 sequencing. *Nature Reviews Genetics*, 16(5), 275–284. <https://doi.org/10.1038/nrg3908>

649 Park, P. J., & Bell, M. A. (2010). Variation of telencephalon morphology of the threespine  
650 stickleback (*Gasterosteus aculeatus*) in relation to inferred ecology. *Journal of Evolutionary  
651 Biology*, 23(6), 1261–1277. <https://doi.org/10.1111/j.1420-9101.2010.01987.x>

652 Peiman, K. S., & Robinson, B. W. (2017). Comparative Analyses of Phenotypic Trait Covariation  
653 within and among Populations. *The American Naturalist*, 190(4), 451–468.  
654 <https://doi.org/10.1086/693482>

655 Peterson, R. D., Sullivan, J. P., Hopkins, C. D., Santaquiteria, A., Dillman, C. B., Pirro, S., Betancur-R,  
656 R., Arcila, D., Hughes, L. C., & Ortí, G. (2022). Phylogenomics of Bony-Tongue Fishes  
657 (Osteoglossomorpha) Shed Light on the Craniofacial Evolution and Biogeography of the  
658 Weakly Electric Clade (Mormyridae). *Systematic Biology*, 71(5), 1032–1044.  
659 <https://doi.org/10.1093/sysbio/syac001>

660 Pigliucci, M. (2003). Phenotypic integration: Studying the ecology and evolution of complex  
661 phenotypes. *Ecology Letters*, 6(3), 265–272. <https://doi.org/10.1046/j.1461-0248.2003.00428.x>

663 Pollen, A. A., Dobberfuhl, A. P., Scace, J., Igulu, M. M., Renn, S. C. P., Shumway, C. A., & Hofmann,  
664 H. A. (2007). Environmental Complexity and Social Organization Sculpt the Brain in Lake  
665 Tanganyikan Cichlid Fish. *Brain, Behavior and Evolution*, 70(1), 21–39.  
666 <https://doi.org/10.1159/000101067>

667 Saunders, A. N., & Gallant, J. R. (n.d.). A review of the reproductive biology of mormyroid fishes: An  
668 emerging model for biomedical research. *Journal of Experimental Zoology Part B: Molecular  
669 and Developmental Evolution*, n/a(n/a). <https://doi.org/10.1002/jez.b.23242>

670 Schumacher, E. L., & Carlson, B. A. (2022). Convergent mosaic brain evolution is associated with the  
671 evolution of novel electrosensory systems in teleost fishes. *eLife*, 11, e74159.  
672 <https://doi.org/10.7554/eLife.74159>

673 Sinervo, B., & Svensson, E. (2002). Correlational selection and the evolution of genomic  
674 architecture. *Heredity*, 89(5), 329–338. <https://doi.org/10.1038/sj.hdy.6800148>

675 Smith, J. M., Burian, R., Kauffman, S., Alberch, P., Campbell, J., Goodwin, B., Lande, R., Raup, D., &  
676 Wolpert, L. (1985). Developmental Constraints and Evolution: A Perspective from the

677        Mountain Lake Conference on Development and Evolution. *The Quarterly Review of Biology*,  
678        60(3), 265–287.

679        Solovieff, N., Cotsapas, C., Lee, P. H., Purcell, S. M., & Smoller, J. W. (2013). Pleiotropy in complex  
680        traits: Challenges and strategies. *Nature Reviews Genetics*, 14(7), 483–495.  
681        <https://doi.org/10.1038/nrg3461>

682        Striedter, G. F. (2005). *Principles of brain evolution* (pp. xii, 436). Sinauer Associates.

683        Sukhum, K. V., Freiler, M. K., Wang, R., & Carlson, B. A. (2016). The costs of a big brain: Extreme  
684        encephalization results in higher energetic demand and reduced hypoxia tolerance in  
685        weakly electric African fishes. *Proceedings of the Royal Society B: Biological Sciences*,  
686        283(1845), 20162157. <https://doi.org/10.1098/rspb.2016.2157>

687        Sukhum, K. V., Shen, J., & Carlson, B. A. (2018). Extreme Enlargement of the Cerebellum in a Clade  
688        of Teleost Fishes that Evolved a Novel Active Sensory System. *Current Biology*, 28(23), 3857–  
689        3863.e3. <https://doi.org/10.1016/j.cub.2018.10.038>

690        Ungar, P. S., & Hlusko, L. J. (2016). The evolutionary path of least resistance. *Science*, 353(6294),  
691        29–30. <https://doi.org/10.1126/science.aaf8398>

692        van der Meer, D., Frei, O., Kaufmann, T., Shadrin, A. A., Devor, A., Smeland, O. B., Thompson, W. K.,  
693        Fan, C. C., Holland, D., Westlye, L. T., Andreassen, O. A., & Dale, A. M. (2020). Understanding  
694        the genetic determinants of the brain with MOSTest. *Nature Communications*, 11(1), 3512.  
695        <https://doi.org/10.1038/s41467-020-17368-1>

696        White, G. E., & Brown, C. (2015). Variation in Brain Morphology of Intertidal Gobies: A Comparison  
697        of Methodologies Used to Quantitatively Assess Brain Volumes in Fish. *Brain, Behavior and  
698        Evolution*, 85(4), 245–256. <https://doi.org/10.1159/000398781>

699 Yang, Y., Axelrod, C. J., Grant, E., Earl, S. R., Urquhart, E. M., Talbert, K., Johnson, L. E., Walker, Z.,  
700 Hsiao, K., Stone, I., Carlson, B. A., López-Sepulcre, A., & Gordon, S. P. (2024n.d.).  
701 Evolutionary divergence of developmental plasticity and learning of mating tactics in  
702 Trinidadian guppies. *Journal of Animal Ecology*, *n/a*:15(n/a).  
703 <https://doi.org/10.1111/1365-2656.14043>

704 Yopak, K. E., Lisney, T. J., Darlington, R. B., Collin, S. P., Montgomery, J. C., & Finlay, B. L. (2010). A  
705 conserved pattern of brain scaling from sharks to primates. *Proceedings of the National  
706 Academy of Sciences*, *107*(29), 12946–12951. <https://doi.org/10.1073/pnas.1002195107>

707 Zhao, B., Shan, Y., Yang, Y., Yu, Z., Li, T., Wang, X., Luo, T., Zhu, Z., Sullivan, P., Zhao, H., Li, Y., & Zhu,  
708 H. (2021). Transcriptome-wide association analysis of brain structures yields insights into  
709 pleiotropy with complex neuropsychiatric traits. *Nature Communications*, *12*(1), 2878.  
710 <https://doi.org/10.1038/s41467-021-23130-y>

711 Zupanc, G. K. H. (2006). Neurogenesis and neuronal regeneration in the adult fish brain. *Journal of  
712 Comparative Physiology A*, *192*(6), 649. <https://doi.org/10.1007/s00359-006-0104-y>

713

714