

Diversity of intraspecific patterns of brain region size covariation in fish

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45 **Abstract**

46 Traits often do not evolve in isolation or vary independently of other traits. Instead, they can be
47 affected by covariation, both within and across species. However, the importance of within species
48 trait covariation and, critically, the degree to which it varies between species has yet to be
49 thoroughly studied. Brain morphology is a trait of great ecological and behavioral importance, with
50 regions that are hypothesized to vary in size based on behavioral and cognitive demands. Sizes of
51 brain regions have also been shown to covary with each other across various taxa. Here we test the
52 degree to which covariation in brain region sizes within species has been conserved across ten
53 teleost fish species. These ten species span five orders, allowing us to examine how phylogenetic
54 proximity influences similarities in intraspecific trait covariation. Our results showed a trend that
55 similar patterns of brain region size covariation occur in more closely related species. Interestingly,
56 there were certain brain region pairs that showed similar levels of covariation across all species
57 regardless of phylogenetic distance, such as the telencephalon and optic tectum, while others, such
58 as the olfactory bulb and the hypothalamus, varied more independently. Ultimately, the patterns of
59 brain region covariation shown here suggest that evolutionary mechanisms or constraints can act
60 on specific brain regions independently, and that these constraints can change over evolutionary
61 time.

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67 **Introduction**

68 *Trait Covariation*

69 Trait variation is ubiquitous between and within species, providing the substrate on which natural
70 selection can act. However, traits rarely vary independently and instead covary with other traits
71 due to functional, genetic, and developmental links (Armbruster et al., 2014; Peiman & Robinson,
72 2017). These effects can limit the degree of trait change because linked traits either need to change
73 together, or selection needs to be strong enough to break those links, as supported by theory
74 (Jones et al., 2014; Pigliucci, 2003), simulations (McGlothlin & Ketterson, 2008), and empirical
75 observations (Ungar & Hlusko, 2016). For example, Unger and Hlusko (2016) showed that
76 covariation in tooth morphology traits in early hominid species led to the evolution of suboptimal
77 tooth structure in these species, with teeth unable to evolve to optimally match the diet of
78 individual species.

79
80 Covariation between traits can be studied at multiple scales, including broad evolutionary to within
81 population ones (Atwell et al., 2014). However, the degree to which within species trait covariation
82 may be conserved or vary across species remains understudied (Peiman & Robinson, 2017). Similar
83 patterns of trait covariation across species would indicate that the level of interdependence of
84 traits may be evolutionarily conserved, while different patterns of trait covariation would suggest
85 that these links are evolutionarily flexible. An example of this can be seen in the integration of
86 marsupial skull morphology. Goswami (2007) showed that marsupial skulls show a phenotypic
87 integration of bone morphology within species, resulting in constraints on the phenotypes in skull
88 morphology that could evolve in these mammals. This paper also showed that closely related

89 species showed more similar levels of integrated changes in skull morphology across phylogenetic
90 distance. Constraints to trait covariation across species can limit the ability of individual species to
91 evolve optimal phenotypes in their local environments. Therefore, understanding the degree of
92 these constraints within and across species is critical to our ability to link form and function in the
93 evolution of organisms.

94

95 *Brain region variation*

96 Brain morphology is one trait that has been of interest to biologists for centuries⁷ and is critical to
97 animal performance across environments. It can influence both cognitive and behavioral abilities of
98 animals and thus affects animal performance and survival across environments (Axelrod et al.,
99 2022; Benson-Amram et al., 2016; Buechel et al., 2018; A. Kotrschal et al., 2013; MacLean et al.,
100 2014; Yang et al., [2024-n.d.](#)). Brains are divided into distinct regions, which are associated with
101 specific cognitive, perceptual, and behavioral functions (Healy & Rowe, 2007; Huber et al., 1997; K.
102 Kotrschal et al., 1998; Park & Bell, 2010; Pollen et al., 2007; Schumacher & Carlson, 2022; Striedter,
103 2005; Sukhum et al., 2018). Variation in the size of brain regions within and between species can be
104 indicative of differential selection due to environmental variation and other factors (Axelrod et al.,
105 2021; Gonzalez-Voyer & Kolm, 2010; Laberge & Hara, 2001). However, the sizes of different regions
106 are not independent of each other and can covary, as individuals with larger brains tend to also
107 have similar proportioned larger brain regions both within and across species (Striedter, 2005).
108 Here, we examine the degree to which variation in brain region size covaries within fish species,
109 and how these patterns may differ across species.

110

111 The degree of covariation in brain regions has been of interest to evolutionary biologists for
112 decades. The “concerted model” of brain evolution posits that the evolution of brain regions is
113 highly constrained, with evolutionary change in the size of brain regions resulting primarily from
114 shifts in overall brain size (Finlay & Darlington, 1995; Yopak et al., 2010). These studies found that
115 the proportional size of brain regions was highly predictable based on the size of the whole brain,
116 suggesting that the evolution of brain region size does not vary independently and is likely
117 constrained by brain region covariation. On the other hand, the “mosaic model” of brain evolution
118 suggests that specific brain regions are targeted to change in size, independent of other structures,
119 due to their association with certain behavioral or cognitive abilities (Barton & Harvey, 2000). Since
120 the development of these two alternative models, the evolution of brain morphology has been
121 studied as reflecting a degree of covariation between brain region size and whole brain size,
122 suggesting that the two hypotheses are not mutually exclusive. At broad evolutionary scales, brain
123 region scaling appears to be conserved, with occasional “mosaic” shifts occurring in some clades
124 (Hoops et al., 2017; Striedter, 2005; Sukhum et al., 2018; Yopak et al., 2010). For example, the
125 evolution of electrosensory abilities in fish is linked to a mosaic shift in the sizes of the cerebellum
126 and hindbrain (Schumacher & Carlson, 2022). However, at the intraspecific level, evidence of this
127 degree of brain region independence is less well established (Hager et al., 2012; A. Kotrschal et al.,
128 2017; Noreikiene et al., 2015).

129

130 *Approach and Hypotheses*

131 Here, we test how much intraspecific brain region covariation is conserved across 10 teleost fish
132 species. These 10 species are paired across 5 orders of fish, allowing us to test how evolutionary

133 proximity might impact the similarity of brain region covariation patterns. Similarities we find in
134 brain region covariation patterns across species would indicate that covariation of brain region
135 sizes is conserved across the teleost phylogeny. If the degree of brain region covariation is very
136 flexible evolutionarily, we would expect covariation patterns to differ across species, and not be
137 linked to evolutionary relatedness. Finally, a midpoint between these extremes could indicate that
138 the degree of independence of within brain region variation can evolve, but more closely related
139 species would be more likely to share these patterns of covariation. In this case we would expect
140 species within orders to have more similar patterns of brain region covariation, with differences
141 emerging at more distant phylogenetic comparisons.

142

143 **Methods**

144 **Specimen collection**

145 To compare patterns of brain region covariation, we collected data from 10 teleost fish species
146 spanning 5 orders (Perciformes, Scorpaeniformes, Cyprinodontiformes, Cypriniformes, and
147 Osteoglossiformes) (figure 1). We collected the data for the Trinidadian guppy (*Poecilia reticulata*),
148 Western mosquitofish (*Gambusia affinis*), Creek chub (*Semotilus atromaculatus*), Blacknose dace
149 (*Rhinichthys atratulus*), African butterfly fish (*Pantodon buchholzi*), and Black baby whale fish
150 (*Brienomyrus brachyistius*). Of these, the guppy fish were lab reared descendants of wild collected
151 fish from Trinidad. *G. affinis*, *S. atromaculatus*, and *R. atratulus* were wild
152 collected samples. *P. buchholzi* and *B. brachyistius* were sourced from a tropical fish supplier. We
153 sourced data for the Pumpkinseed sunfish (*Lepomis gibbosus*), Bluegill sunfish (*Lepomis*
154 *macrochirus*), Threespine stickleback (*Gasterosteus aculeatus*), and Ninespine stickleback (*Pungitius*

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155 *pungitius*) from previously published studies that employed the same data collection method. All
156 species contain sufficient variation in body size to allow measure region covariation. The sources of
157 data, sample size, and coefficients of variation in body size for each species can be seen in [Tables 1](#)
158 and 2. After euthanasia, all fish samples were fixed by being completely submerged in 10%
159 formalin, and stored until processing.

160

161 *Brain region size estimation*

162 To measure brain region size, we estimated the volumes of five regions of the brain: cerebellum,
163 optic tectum, telencephalon, olfactory bulb, and hypothalamus. For all species, we (or the authors
164 of the relevant studies) extracted brains via dissection, ensuring that all regions were extracted
165 intact and that external nerves were trimmed consistently. We then took photographs of the brains
166 from dorsal, ventral, and lateral orientations. From these photographs, we measured the length (L),
167 width (W), and height (H) of each brain region in ImageJ, and then calculated region volumes (V)
168 using the ellipsoid formula ($V = (L \times W \times H) \pi / 6$) (White & Brown, 2015) (supplementary figure 1). We
169 estimated each region's length as a straight line from its most rostral plane to its most caudal plan.
170 Its width was perpendicular, or 90 degrees, to the length, at the widest point of that ovoid. We
171 estimated the height of each region by generating a straight line perpendicular to the horizontal
172 axis of the brain, at the widest part of that lobe from the lateral view. To estimate total volume for
173 regions with bilaterally symmetrical lobes, we summed both sides together. For example,
174 estimating the telencephalon involved measurements of two bilaterally symmetrical lobes, and the
175 volumes of these lobes were added together to obtain the total telencephalon volume. One
176 exception was the olfactory bulb of *S. atromaculatus*, for which we could not reliably measure

177 olfactory bulb height. As such, we calculated the area of this region in this species ($A=L/2 \times W/2 \times \pi$).
178 These data are still usable as area and volume of brain regions are highly correlated (C.A. personal
179 observation).

180

181 Since the cerebellum of *B. brachyistius* drastically varies morphologically from other teleost fish
182 species (Sukhum et al., 2018), we had to deviate from the volume estimation methods used in the
183 other species. Seven ovoid regions were estimated to encompass the shape and size of the
184 enlarged *B. brachyistius* cerebellum (supplementary figure 2). Two heights, for both the rostral and
185 caudal ovoids, were taken for the *B. brachyistius* cerebellum, corresponding to its rostral and
186 caudal ovoid lengths and widths, respectively. The volumes of each of the seven cerebellum ovoids
187 were summed together to estimate total cerebellum volume.

188

189 *Brain region principal component variation*

190 To determine how brain region volumes vary within species and observe trends in their covariation,
191 we ran separate principal component analyses in each species. Percentage of variation explained
192 by PC1 correlated with total brain volume, which is expected in allometric relationships
193 (Klingenberg, 1996). These percentages were compared across species, but all regions in all species
194 loaded in the same direction along PC1. Additionally, the direction of where the brain regions
195 loaded along PC2 were qualitatively compared across species. PCAs were performed using the R
196 package “FactoMineR” (Lê et al., 2008), which generated PC1-5 for each species. The majority of
197 the variation was explained by PC1 and PC2 (ranging from ~79-98% across the 10 species), so we
198 focused on those two axes for comparisons across species.

199

200 *Within species brain region correlations*

201 To establish the level of independence of brain region variation within each species, we calculated
202 Pearson correlations of each pairwise comparison of brain regions in each species. We use this
203 approach because Pearson correlations allow for differences in scale between regions being
204 compared, which is important as regions differ in size within species. This approach also allowed for
205 the area of the olfactory bulb in *S. atromaculatus* to be effectively compared to the volume of the
206 other regions in that species. From each pairwise correlation of brain regions, we can establish a
207 separate correlation matrix for each species. Lower levels of correlation between brain regions
208 indicate higher levels of independent variation.

209

210 *Species comparisons of brain region correlation patterns*

211 To compare the patterns of within species brain region correlation we estimated the similarity of
212 species correlation matrices. We used a Mantel test (Vegan R package; Oksanen et al. 2020) to
213 estimate the pairwise correlation of each species' correlation matrix. This test examines the
214 correlation of two matrices using a Pearson correlation method, and then employs a permutation
215 approach using 10,000 permutations to estimate the significance of that correlation. We chose this
216 approach as our goal was to get a broad sense of the similarity of within species brain region
217 correlation patterns and how these vary between species, rather than to establish specific
218 mechanisms of brain region covariation. Understanding the mechanisms of how and why brain
219 regions covary and how these differ among species would require additional data including genetic
220 data or the use of selection or breeding experiments, though this was not the focus of this study.

221

222 *Phylogenetic distance comparison*

223 Finally, to test whether more closely related species have more similar patterns of brain region
224 correlation, we compared phylogenetic distance between species to the similarity of their
225 correlation matrices. We used the correlations estimated from the [M](#)antel test comparing
226 individual species to construct a species matrix including all pairwise species comparisons (figure 4).
227 We then generated a phylogenetic distance matrix showing time (in millions of years) since the
228 most recent common ancestor for each pairwise species comparison (supplementary figure 3).
229 Time since most recent common ancestor for each species comparison was gathered from
230 Timetree.org. We then compared these two species level matrices using the same Mantel test
231 approach as we used to compare brain region correlation matrices. In this case, if more closely
232 related species share more similar brain region correlation patterns, we expect a negative
233 correlation between the species level matrices.

234

235 **Results**

236 *Intraspecific brain region principal component variation*

237 All brain regions covaried in the same direction along PC1, which reflected total brain volume
238 (figure 2), but the percent variation explained by this axis varied across the 5 different fish orders
239 (supplementary figure 4). The Cypriniformes had the highest amount of variation explained by PC1
240 (90.06% for *S. atromaculatus* and 95.03% for the *R. atratulus*), whereas the Scorpaeniformes and
241 Osteoglossiformes had lower amounts of brain region variation explained by total brain volume

242 (~65-70% across the four species in these orders). The Perciformes and Cyprinodontiformes had
 243 more intermediate levels of variation explained by PC1 (~75-85%).
 244
 245 There was a noticeable difference in spread along the PC2 axis (for the five brain regions),
 246 indicating the differing strengths of independent contributions separate regions have on brain
 247 variation (figure 2). There was a split between species where the olfactory bulbs contributed
 248 disproportionately to most of the percent variation described by PC2, (~6-10% for
 249 Cyprinodontiformes, 14-20% for Scorpaeniformes, ~10-13% for Perciformes), and those where
 250 multiple regions contributed to the variation along PC2. These include the Cypriniformes, which
 251 had relatively little variation and spread explained by PC2 in any brain region (~2-4%), and the
 252 Osteoglossiformes, for which different brain regions contributed to varying degrees to PC2 in the
 253 two species studied (~13-14%). In *P. buchholzi*, the hypothalamus contributed the most to PC2,
 254 followed by telencephalon, optic tectum, olfactory bulb, and finally cerebellum. By contrast, the
 255 hypothalamus and olfactory bulb contributed the most to PC2 in *B. brachyistius*, followed by the
 256 optic tectum, cerebellum, and finally telencephalon. Interestingly, the cerebellum and optic tectum
 257 loaded in the same direction for all species except *B. brachyistius*.
 258
 259 *Intraspecific brain region correlation*
 260 Across all species, brain region comparisons showed a positive correlation, though the strength of
 261 these correlations varied within and among species (figure 3). All these positive correlations were
 262 statistically significant, other than the olfactory bulb and cerebellum comparison, and the olfactory
 263 bulb and optic tectum comparison, in *P. pungitius*. To get a more general sense of within species

264 brain region independence, for every species we calculated the average correlation for each brain
265 region with the other regions of the brain. This revealed that species vary in their overall level of
266 brain region covariation, with average correlations ranging from 0.56 in *G. aculeatus* and *P.*
267 *pungitius* to 0.94 in *R. atratulus* (table 3). The Cypriniformes showed very high levels of region
268 correlations, with all regions correlating with each other by at least 0.82 in *S. atromaculatus* and
269 0.88 in *R. atratulus*. Further, in all fish species other than the two Osteoglossiform species (*P.*
270 *buchholzi* and *B. brachyistius*), the olfactory bulb showed the lowest average correlation with the
271 other brain regions, though in the Cypriniformes this was a very small difference (0.03 or less
272 compared to other region averages) (table 3). In both Osteoglossiform species, the hypothalamus
273 showed the lowest average correlation with the rest of the brain regions (0.51 for *P. buchholzi* and
274 0.52 for *B. brachyistius*). The correlation between optic tectum size and telencephalon size showed
275 the highest correlation of any region comparison in every species other than *P. buchholzi*, which
276 showed the highest correlation between the optic tectum and cerebellum (table 3).

277

278 *Species comparison*

279 Within species brain region covariation patterns correlated across certain fish species (figure 4). *P.*
280 *reticulata*, *G. affinis*, *L. gibbosus*, *L. macrochirus*, *G. aculeatus*, and *P. pungitius* all showed relatively
281 high correlations of their brain region covariation patterns (at least 0.751), with all pairwise
282 correlations being significant other than *P. pungitius* with *P. reticulata*, *P. pungitius* with *L.*
283 *macrochirus*, and *P. pungitius* with *G. aculeatus* (figure 4). This indicates that these groups all share
284 a similar pattern of brain region correlations. *S. atromaculatus* and *R. atratulus* also share some
285 similar patterns with this broad group, though the correlations were weaker (figure 4). *S.*

286 *atromaculatus* showed a significant correlation with the brain region covariation patterns of *P.*
287 *reticulata* ($r=0.784$), *L. macrochirus* ($r=0.681$), and *G. aculeatus* ($r=0.581$). *R. atratulus* correlated
288 with *P. reticulata* ($r=0.773$) and *L. macrochirus* ($r=0.770$). *S. atromaculatus* and *R. atratulus* also
289 showed a strong significant correlation with each other, at 0.833. Both Osteoglossiform species
290 showed divergence in their brain region covariation patterns, with the only significant correlation
291 occurring between *B. brachyistius* and *R. atratulus* (0.839). The covariation pattern of *P. buchholzi*
292 was not significantly correlated with any other species.

293

294 ~~We note that differences in average brain region correlation levels among species observed here~~
295 ~~may be related to differences in body size variation, as greater body size variation could establish~~
296 ~~the possibility for stronger correlations. However, this effect would not impact our ability to~~
297 ~~compare the degree of similarity of correlation matrices, as the Mantel test comparison is not~~
298 ~~affected by differences in average matrix values. To support this assumption, we compared the~~
299 ~~similarity of the *S. atromaculatus* matrix and a correlation matrix calculated from a truncated *S.*~~
300 ~~*atromaculatus* data set with the largest ($>20g$) and smallest ($<2g$) individuals removed, resulting in~~
301 ~~a lower coefficient of body mass variation of 0.55 (supplementary figure 5). These matrices were~~
302 ~~highly correlated with each other ($r=0.96$, $p=0.0083$), indicating that differences in average levels of~~
303 ~~trait correlations do not impact the outcomes of our Mantel test. As such, we will only consider~~
304 ~~results related to differences related to comparisons among regions within species, and how these~~
305 ~~vary across species, and not discuss overall average levels of brain region correlation within~~
306 ~~species.~~

307 We note that differences in average brain region correlation levels among species observed here
308 may be related to differences in body size variation (table 1), as greater body size variation could
309 establish the possibility for stronger correlations. However, this effect would not impact our ability
310 to compare the degree of similarity of correlation matrices, as the Mantel test comparison is not
311 affected by differences in average matrix values. To support this assumption, we used an additional
312 analysis of the species in our data set with the highest body mass coefficient of variation, *S.*
313 *atromaculatus*. We compared the similarity of the *S. atromaculatus* matrix and a correlation matrix
314 calculated from a truncated *S. atromaculatus* data set with the largest (>20g) and smallest (<2g)
315 individuals removed, resulting in a lower coefficient of body mass variation of 0.55 (supplementary
316 figure 5). These matrices were highly correlated with each other ($r=0.96$, $p=0.0083$), indicating that
317 differences in average levels of trait correlations do not impact the outcomes of our Mantel test. As
318 such, we will only consider results related to differences related to comparisons among regions
319 within species, and how these vary across species, and not discuss overall average levels of brain
320 region correlation within species.

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322 Brain region covariation patterns appeared to be influenced by phylogenetic distance. We found
323 that the similarity in brain region covariation patterns was inversely correlated with phylogenetic
324 distance (figure 5). This trend was not significant ($r=-0.671$, $p=0.999$).

326 Discussion

327 We tested the degree of covariation of brain region sizes within 10 teleost fish species to examine
328 how brain region covariation patterns vary across species. Our results showed that brain regions

varied in the strength of their covariation within and across species, with the telencephalon and optic tectum usually showing very high levels of covariation, and the olfactory bulb showing lower covariation with the rest of the brain. Further, we found an insignificant trend that patterns of brain region covariation are shared across more closely related species. Specifically, this trend indicates that the similarity of within species brain region correlation patterns decreases as phylogenetic distance between species increases.

335

Within species patterns of brain region covariation

Patterns of brain region volume covariation within species indicate the potential for independent change in brain region size. These patterns can arise through various, non-mutually exclusive mechanisms, including differences in the strength of selection on different brain regions, or from genetic and developmental constraints (Davidowitz et al., 2012; Peiman & Robinson, 2017).

341

Covariation between the sizes of brain regions can occur due to patterns of selection acting on brain regions independently. This pattern can occur when selection consistently acts in similar ways on different traits (Sinervo & Svensson, 2002). For example, we observed that the telencephalon and the optic tectum showed a high level of correlation in most species. If selection consistently acts in the same direction on these two regions, it could result in this pattern. Similarly, the high level of independence of the olfactory bulb in most of the species we studied could result from selection on this specific region acting in a different direction to other regions of the brain. We consider this explanation to be unlikely to explain most of the variation we see in brain region covariation for two reasons. First, we found similar patterns in brain region covariation across

several fish species. Selection would have needed to be consistent in its effect on all brain regions across all these species regardless of population or environment, which decreases its likelihood as an explanation. Second, prior research suggests that selection can act differently across brain regions, which would result in less covariation across regions and is unlikely to then lead to very consistent patterns in brain region covariation between species. For example, Gonzalez-Voyer & Kolm (2010) found that sexual selection in Lake Tanganyikan Cichlid species resulted in divergence in the size of the hypothalamus and cerebellum, with other regions of the brain diverging in the opposite direction. Further, Kotrschal et al. (2017) found that Trinidadian guppy brain regions differed in how they diverged in response to predation levels across populations. Although we consider other explanations of the patterns we observed to be more likely, we cannot rule out correlated patterns of selection. Further research on the consistency of how selection acts on different brain regions within and across species would be needed to fully evaluate this possibility.

363

The other possible mechanism that can generate covariation between traits is some type of constraint, including genetic constraints and developmental constraints. Genetic constraints occur through pleiotropy (a single gene being associated with multiple phenotypic traits) or gene linkage (genes for different traits occurring close together on the same chromosome), limiting the ability of traits to evolve independently of each other (Ott et al., 2015; Solovieff et al., 2013). Although the degree of pleiotropy in determining brain region size has not been specifically measured, prior work has suggested that aspects of brain connectivity and structure are influenced by pleiotropic or linked genes (Moreau et al., 2022; van der Meer et al., 2020; Zhao et al., 2021). It is therefore likely that the evolution of the size of brain regions may be similarly influenced by genetic architecture

373 constraints. The strength of these genetic links between brain regions could thus result in the
374 patterns of brain region covariation that we observed across species.

375

376 Developmental constraints are limitations to the variability of traits caused by the structure or
377 dynamics of organism ontogeny (Smith et al., 1985). This was the form of constraint proposed to
378 explain the covariation of brain regions among mammals under the concerted brain evolution
379 hypothesis, where the evolution of larger brain size was linked to increases in the relative size of
380 later developing brain regions (Finlay & Darlington, 1995). As fish have indeterminate growth and
381 maintain widespread neurogenesis of the brain into adulthood (Zupanc, 2006), this specific process
382 is unlikely to explain our results. However, a similar kind of developmental limitation may. The
383 growth of different brain regions may be influenced by overlapping developmental mechanisms
384 such as the ability to generate new neurons in specific areas of the brain (Ganz & Brand, 2016;
385 Kaslin et al., 2008). We are not able to determine specific mechanisms generating the covariation
386 patterns we observed, and future work examining the genetic architecture and developmental
387 processes of brain region evolution will be needed to fully elucidate these.

388

389 *Evolution of brain region covariation patterns*

390 We found a trend that more closely related species tend to share similar patterns of brain region
391 covariation, while these patterns diverge as phylogenetic distance increases. Our results indicate
392 that Perciformes and Cyprinodontiformes show strong evidence of a shared pattern of brain region
393 covariation. Scorpaeniformes also show a similar pattern to these orders, but with higher overall
394 levels of brain region independence and less consistency, as *P. pungitius* appear to be less similar to

395 the rest of these species. These six species are closely related in evolutionary terms compared to
396 the rest of the species we included, with a common ancestor ~112 million years ago, though they
397 differ in terms of ecology, particularly between orders (Lowe-McConnell, 2012; Magalhaes et al.,
398 2016). This similarity suggests that the constraints limiting independent brain region change are
399 evolutionarily conserved across closely related fish species, even when ecology, and likely selection
400 pressures, differ.

401
402 Our two Cypriniform species (*S. atromaculatus* and *R. atratulus*) show similar patterns to each
403 other, particularly in terms of overall level of brain region independence, as both show very little
404 independence in brain regions. The Cypriniform brain differs morphologically from other fish as
405 they are characterized by extended and somewhat enlarged olfactory bulbs (Brandstätter &
406 Kotrschal, 1990; Evans, 1952). Beyond this morphological difference, it is unclear why these species
407 have such strong correlations between their brain regions. Broadly, this result indicates that
408 selection would need to be very strong in order to shift individual brain regions in these species
409 independent of the rest of the brain, as evolution would need to break this strong covariation
410 pattern, potentially limiting the ability of these species to respond to selective pressures.

411
412 Our results indicate that patterns of intraspecific brain region covariation can evolve, and these
413 patterns are more similar between more closely related species. The link between phylogenetic
414 distance and within species brain region covariation is supported by two pieces of evidence here.
415 First, qualitatively, our results support this conclusion as we found high similarity in brain region
416 covariation patterns among closely related species, and the most distantly related species in our

sample, the two Osteoglossiform species, show very little similarity to each other and to the rest of the species in their brain region covariation patterns. This order diverged from the rest of the species in our samples approximately 263 million years ago. *B. brachyistius* and *P. buchholzi* diverged from each other approximately 200 million years ago, much greater than the divergence time between the species in the other orders we tested. Second, our quantitative comparison suggests that phylogenetic distance between the species is negatively correlated with the similarity of their brain region covariation patterns, though this pattern was not statistically significant. This is likely due to the limited variation in time since the most recent common ancestor, which restricts the statistical power of permutation tests. Permutation tests rely on variability in the data to allow for estimating a random distribution of potential results. Many species comparisons here share the same time since most recent common ancestor, resulting in extremely limited variation from which the permutation test can derive a results distribution. Our evidence of a negative correlation between brain region correlation patterns and phylogenetic relatedness is therefore limited in its reliability. Together these indicate that evolutionary time is required to break the constraints that cause these patterns of brain region covariation, potentially through selection acting strongly in an opposite direction on correlated regions. Understanding the specific selection forces that lead to divergence in brain region covariation patterns will require further study focusing on mechanisms and evolution of constraints in brain region correlation.

Beyond more ancient divergence times between each other and the remainder of the species, the two Osteoglossiform species possess phenotypic distinctiveness in comparison to the other teleosts studied here. For example, *P. buchholzi*, a surface-oriented fish, exhibits the slowest rate of

439 morphological divergence among other studied Osteoglossiformes, despite tens of millions of years
440 of genetic divergence between populations (Lavoué et al., 2011). This high level of phenotypic
441 stability over such a long timescale in this fish lineage is thought to surpass all other known
442 examples of morphological stasis in extant vertebrates. The other osteoglossomorph we studied, *B.*
443 *brachyistius*, is in the superfamily Mormyroidea, a group of African freshwater fishes that have
444 electromotor and electrosensory systems (Carlson & Arnegard, 2011; Crampton, 2019). In addition,
445 mormyroids also possess extremely large brains (Nilsson, 1996; Sukhum et al., 2016, 2018), a suite
446 of diverse craniofacial morphologies (Peterson et al., 2022), large intra- and interspecific variety in
447 communication signals and behaviors (Hopkins 1986), and are the only known vertebrates whose
448 sperm lack flagella (Saunders & Gallant, 2024). These novel phenotypic features in both *B.*
449 *brachyistius* and *P. buchholzi*, along with their ancient divergence times, could potentially be
450 related to breaks in genetic and/or developmental constraints maintained within other teleost
451 species. However, further research is needed comparing more Osteoglossiform species across a
452 greater variety of phylogenetic distances to examine how covariation in brain region sizes has been
453 maintained over time in this specific group.

454

455 *Limitations*

456 There are key limitations to our study that must be considered. First, due to logistical reasons the
457 number of species we sampled here was low. Our focus was on building enough within species
458 samples to confidently measure within species brain region covariation, which made sampling a
459 larger number of species difficult. This lower representation of species particularly limited our
460 ability to adequately test the within and across species trait covariation patterns in a

461 phylogenetically controlled way. Second, we built our database using a combination of pre-
462 collected data and fish samples and newly sampled species, resulting in variation in sampling
463 techniques, including fish that were reared in labs and wild collected fish. This benefits our study in
464 some ways, notably by increasing the possible variability between species, making our observation
465 of similar patterns across species stronger. However, by doing this we are unable to determine how
466 differences in selection or environmental variation may be impacting our results, limiting our ability
467 to elucidate the mechanisms generating the covariation patterns we see. Finally, we do not have
468 any genetic information about the patterns of covariation we observed. Previous work on
469 evolutionary constraints caused by the covariation of traits has focused on genetic covariation
470 (Davidowitz et al., 2012; Lande, 1984; Olson & Miller, 1958; Ott et al., 2015), with the goal of linking
471 trait covariation to the evolutionary mechanism of constraint. We also are unable to determine the
472 heritability of patterns of trait covariation we observed. The genetic data needed for this type of
473 analysis were not available to us. Future work should examine both the heritability of brain region
474 covariation patterns as well as the genetic architecture causing these patterns of covariation.

475

476 *Conclusion*

477 Here, using large, within-species data sets, we examined the degree of covariation of brain region
478 sizes within 10 fish species, and tested whether those patterns vary across species. Our results
479 indicate that brain regions vary in their level of covariation, and that these patterns are shared
480 between closely related species of fish. Broadly, these results could indicate that the size of
481 individual brain regions is constrained in its ability to evolve within species, and that it takes a great
482 deal of evolutionary time to break or change these constraints. Future work will be needed to

483 elucidate the specific nature of these constraints, as well as the evolutionary processes that can

484 break these patterns.

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490 Tables and Figures:

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492 Table 1. List of fish species used, including the source of the data and the sample size.

Common Name	Scientific Name	Order	Source	N	Coefficient of variation in body mass
Pumpkinseed Sunfish	<i>Lepomis gibbosus</i>	Perciformes	Axelrod et al. 2021	113	0.59
Bluegill Sunfish	<i>Lepomis macrochirus</i>	Perciformes	Axelrod et al. 2021	94	0.52
Threespine Stickleback	<i>Gasterosteus aculeatus</i>	Scorpaeniformes	Herczeg et al. 2015	231	0.24
Ninespine Stickleback	<i>Pungitius pungitius</i>	Scorpaeniformes	Gonda et al. 2009	120	0.37
Trinidadian Guppy	<i>Poecilia reticulata</i>	Cyprinodontiformes	Authors	296	0.46
Mosquitofish	<i>Gambusia affinis</i>	Cyprinodontiformes	Authors	133	0.68
Creek Chub	<i>Semotilus atromaculatus</i>	Cypriniformes	Authors	194	0.91
Black Nose Dace	<i>Rhinichthys atratulus</i>	Cypriniformes	Authors	82	0.8
African Butterflyfish	<i>Pantodon buchholzi</i>	Osteoglossiformes	Authors	61	0.39
Black baby whale	<i>Brienomyrus brachyistius</i>	Osteoglossiformes	Authors	56	0.57

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495 Table 2. Collection details for each fish species used.

Species	Collection Details
<i>Lepomis gibbosus</i>	Wild collected fish, collected via angling from the wild in summer 2016 and 2017 from four pelagic sites and four littoral sites of Ashby Lake, Ontario.
<i>Lepomis macrochirus</i>	Wild collected fish, collected via angling from the wild in summer 2017 from one pelagic site and one littoral site of Holcomb Lake, Michigan.
<i>Gasterosteus aculeatus</i>	F1 offspring of wild collected fish from the Baltic Sea in 2011. F1 fish were reared until adulthood in either an enriched environment or a simple environment.
<i>Pungitius pungitius</i>	F1 offspring of wild collected fish from four sites in the Baltic Sea in 2011. F1 fish were reared until adulthood either alone or in groups of 20 fish.
<i>Poecilia reticulata</i>	F2 offspring of wild collected fish from two populations in Trinidad, Aripo high predation (HP) and Aripo low predation (LP). F2 fish from both populations were reared until sexual maturity under two predator environments (pred+ and pred-) and three social environments (Solo, with HP conspecifics, and with LP conspecifics).
<i>Gambusia affinis</i>	Wild collected fish from 4 freshwater sites in southern Louisiana in the summer of 2022. Fish were collected with dip nets from one high salinity site and one low salinity site in each of two streams.
<i>Semotilus atromaculatus</i>	Wild collected via electrofishing in the summer of 2018 and 2019 from 17 freshwater stream representing a gradient of agricultural intensity within 6 major watersheds in Southwest Ontario, Canada.
<i>Rhinichthys atratulus</i>	Wild collected fish from Fall Creek, NY. Fish were collected in November of 2023 using seine nets.
<i>Pantodon buchholzi</i>	Sourced from tropical fish supplier
<i>Brienyomys brachyistius</i>	Sourced from tropical fish supplier

Table 3. Average correlation coefficients for brain regions across species. Averages are calculated from the correlations of each region with every other region in each species.

Species	Cb	OT	Tel	OB	Hyp	Total
<i>Poecilia reticulata</i>	0.72	0.78	0.79	0.64	0.72	0.73
<i>Gambusia affinis</i>	0.73	0.78	0.80	0.67	0.74	0.75
<i>Lepomis gibbosus</i>	0.70	0.76	0.79	0.61	0.74	0.72
<i>Lepomis macrochirus</i>	0.72	0.76	0.75	0.53	0.66	0.68
<i>Gasterosteus aculeatus</i>	0.60	0.64	0.59	0.43	0.55	0.56
<i>Pungitius pungitius</i>	0.59	0.64	0.64	0.23	0.66	0.56
<i>Semotilus atromaculatus</i>	0.86	0.91	0.90	0.86	0.86	0.88
<i>Rhinichthys atratulus</i>	0.94	0.95	0.96	0.92	0.92	0.94
<i>Pantodon buchholzi</i>	0.65	0.69	0.62	0.66	0.51	0.63
<i>Brienyomys brachyistius</i>	0.64	0.61	0.67	0.53	0.52	0.59

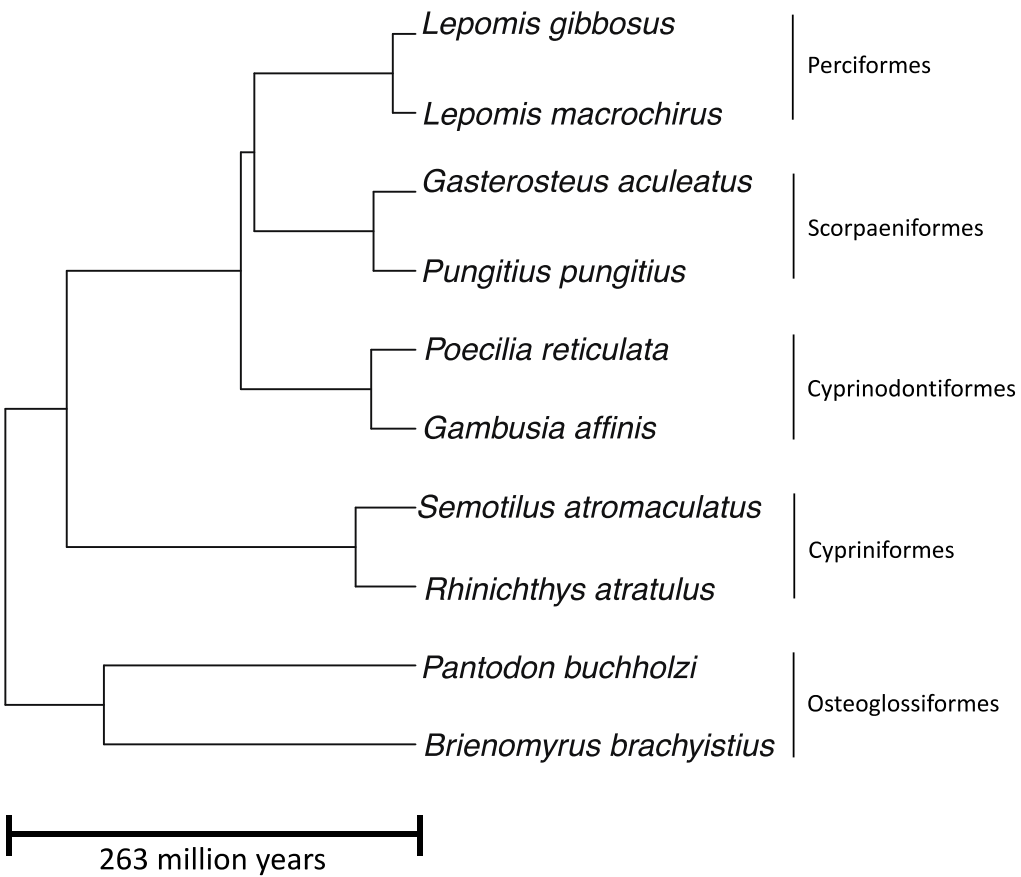


Figure 1. Phylogenetic relationships between the 10 teleost fish species in our study.

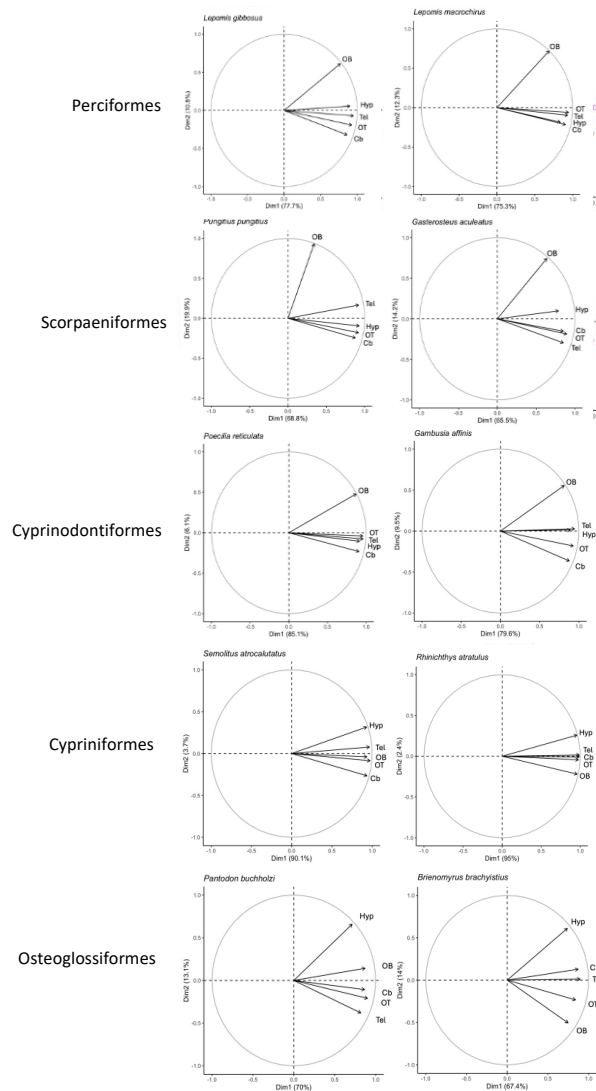
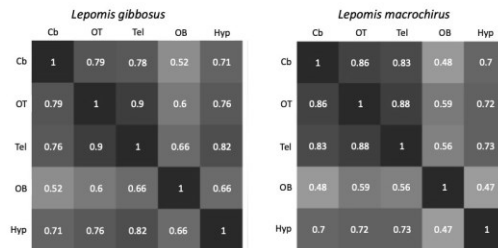
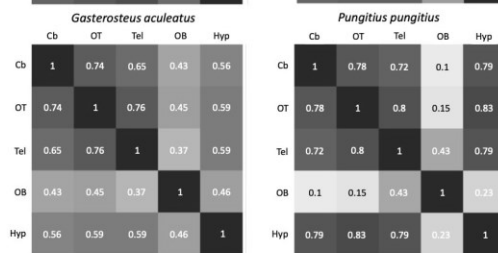


Figure 2: PC loading vectors of each brain region for each species, for PC1 (Dim1, correlated with total brain volume) and PC2 (Dim2, correlated with covariation in brain regions not related to total brain size). Cb, cerebellum; Hyp, hypothalamus; OB, olfactory bulbs; OT, optic tectum; Tel, telencephalon.

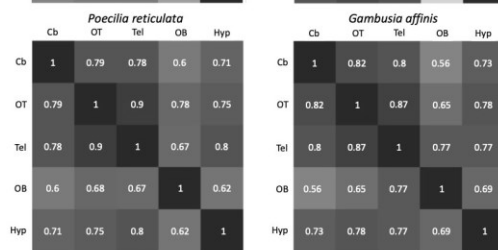
Perciformes



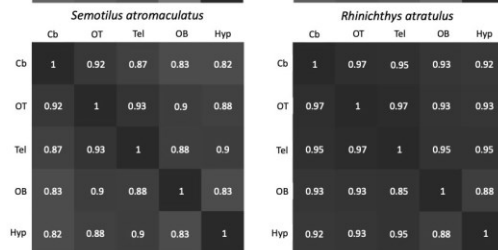
Scorpaeniformes



Cyprinodontiformes



Cypriniformes



Osteoglossiformes

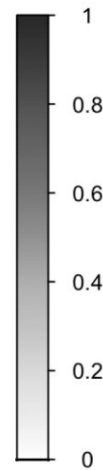
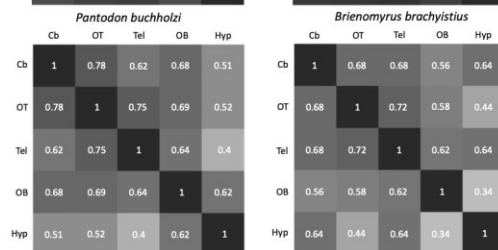


Figure 3. Matrices showing each pairwise Pearson correlation among five brain regions for all 10 fish species: cerebellum (Cb), optic tectum (OT), telencephalon (Tel), olfactory bulb (OB), and hypothalamus (Hyp). Box color shows the strength of the correlations as indicated by the scale bar on the right, and numbers show the correlation coefficient. Statistically significant correlations are shown as white numbers, and non-significant correlations are shown as black numbers: all correlations are significant other than the Cb/OB and OT/OB correlations in *P. pungitius*.

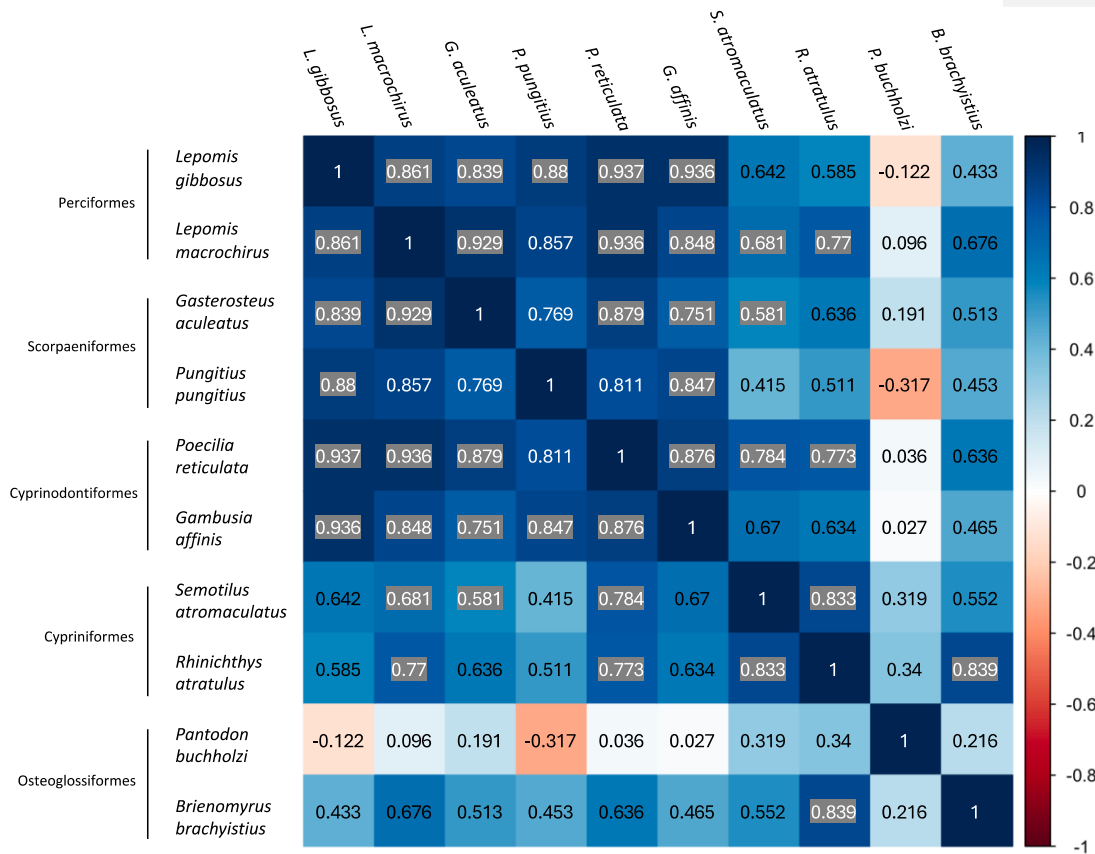
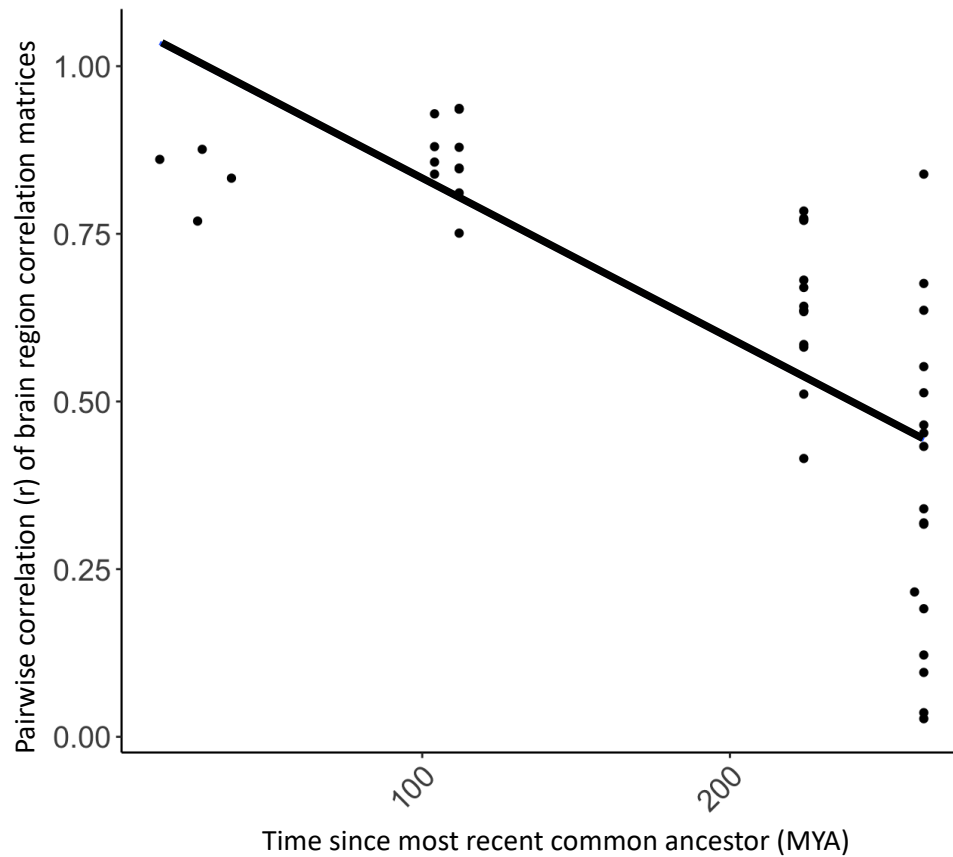


Figure 4. Matrix showing the pairwise Pearson correlations of within species brain region size correlation matrices for each of the 10 fish species in our study. Box color shows the strength of the correlations as indicated by the scale bar on the right. Numbers show the correlation coefficient (r) for each comparison. Gray boxes indicate significant correlations. Numbers are colored for optimal visibility.

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Figure 5. Linear relationship between time since most recent common ancestor and correlation of within species brain region size correlation matrices (see figure 4). Dots show individual species comparisons, and the line shows the linear regression. Mantel test $r^2=0.671$, $p=0.999$. Line of best fit equation: $r=1.07-0.0024(\text{MYA})$.

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