

A revised terminology for male genitalia in Hymenoptera (Insecta), with a special emphasis on Ichneumonoidea

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ABSTRACT

Applying consistent terminology for morphological traits across different taxa is a highly pertinent task in the study of morphology and evolution. Different terminologies for the same traits can generate bias in phylogeny and prevent correct homology assessments. This situation is exacerbated in the male genitalia of Hymenoptera, and specifically in Ichneumonoidea, in which the terminology is not standardized and has not been fully aligned with the rest of Hymenoptera. In the current contribution, we review the terms used to describe the skeletal features of the male genitalia in Hymenoptera, and provide a list of authors associated with previously used terminology. We propose a unified terminology for the male genitalia that can be utilized across the order and a list of recommended terms. Further, we review and discuss the genital musculature for the superfamily Ichneumonoidea based on previous literature and novel observations and align the terms used for muscles across the literature.

Subjects Entomology, Evolutionary Studies, Taxonomy, Zoology

Keywords Confocal laser scanning microscopy, Male genitalia, Hymenoptera anatomy ontology, Ichneumonoidea, Braconidae, Ichneumonidae, Homology, Ontology, Unified terminology, Comparative anatomy

INTRODUCTION

The importance of a unified morphological terminology

The study of morphology entails the interpretation of anatomical structures shaped by evolutionary processes and their translation into rigorous and consistent data (Boudinot, 2019). This practice requires the application of terms and concepts for effectively identifying and describing the structures. However, terminologies employed in different groups of organisms can overlap and cause confusion. Homonyms, identical terms used for non-homologous structures, are widespread and employed to describe structures in unrelated taxa (e.g., Costa et al., 2013; Donkelaar et al., 2017). At the same time, homologous anatomical traits in related groups of organisms often have inconsistent terminologies (e.g., Schulmeister, 2001).

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Inconsistent terminologies are widespread within and among insect orders ([Yoder et al., 2010](#); [Costa et al., 2013](#); [Wirkner et al., 2017](#)). [Girón et al. \(2023\)](#) identified five main reasons for the emergence of inconsistent terminologies within insects, namely: (1) borrowing terms from the vertebrate anatomy (e.g., wings, head); (2) creating terms *de novo* (e.g., sclerite, sternite); (3) applying terms to different insect lineages to refer to similar structures located in similar areas of the body (e.g., *cercus* in Diplura and *cercus* in Hymenoptera); (4) changes in phylogenetic classification that caused a reassessment of the morphological terminology; and (5) deviation in the original application of a term due a subsequent misinterpretation (e.g., the concepts of *volsella*). As pointed out by [Yoder et al. \(2010\)](#), the consequences of having disparate terminologies can negatively impact the comparison of gene expression patterns, comparative morphological and phylogenetic studies, analyses of phenotype variability, integration of descriptive taxonomy and phenomics, and machine learning algorithms.

Recently, efforts to address terminological inconsistencies have been undertaken with the intent to unify the terminology and standardize morphological data, and establish a comparability and communicability framework (e.g., [Vogt, Bartolomaeus & Giribet, 2010](#)). The idea is to use ontology, the logical and linguistic machinery for interpreting physical observations, to relate conceptual objects defined by their general identities and their specific properties, and to use controlled vocabularies for communication among scientists via stabilization of terminologies ([Deans, Yoder & Balhoff, 2012](#); [Boudinot, 2019](#); [Girón et al., 2023](#)). Examples of successful attempts are the Drosophila Anatomy Ontology (DAO) ([Costa et al., 2013](#)), the Ontology of Arthropod Circulatory Systems (OArCS) ([Wirkner et al., 2017](#)), the Hymenoptera Anatomy Ontology (HAO) ([Yoder et al., 2010](#)), and more recently the Insect Anatomy Ontology ([Girón et al., 2023](#)). Of these, the HAO provides an essential tool for hymenopterists but still lacks terms and concepts employed in different taxonomic groups ([Boudinot, 2018](#); [Lanes et al., 2020](#); [de Brito, Lanes & Azevedo, 2021](#)). For instance, within the hyperdiverse superfamily Ichneumonoidea ontological alignments for the different morphological structures are still severely lacking.

Among hexapods, the study of male genitalia has long captivated entomologists due to their essential function, diverse morphology, and mechanical adaptations. Even though in some insect orders, male genitalia play a fundamental role in phylogenetic and taxonomic studies (e.g., [Song & Bucheli, 2010](#); [Van Dam, 2014](#); [Bollino, Uliana & Sabatinelli, 2018](#); [Lackner & Tarasov, 2019](#)), in Hymenoptera they have been relatively little explored (e.g., [Schulmeister, 2001](#); [Schulmeister, 2003](#); [Boudinot, 2013](#)), despite being recognized as a critical source of discrete and size-independent characters for both phylogenetic and taxonomic studies (e.g., [Tuxen, 1970](#); [Mikó et al., 2013](#); [Boudinot, 2015](#)). This is surprising given the high diversity and the great variety of forms and ecological roles of the order (more than 150,000 described species) ([Klopfstein et al., 2013](#); [Branstetter et al., 2018](#)). One of the possible causes for this is that these characters tend to suffer from rampant terminological inconsistency (e.g., [Tuxen, 1970](#)), making study difficult.

Hymenopteran male genitalia: a terminological nightmare

The external male genitalia can offer many potential characters for taxonomic and phylogenetic studies due to their complexity, variability, and accessibility (at least compared to their internal counterparts). However, there are terminological inconsistencies across studies, likely resulting from two different interpretations of homologies based on two competing theories ([Michener, 1956](#)). The first, the periphallalic origin theory, postulates that the male genitalia derived from true appendicular structures and are homologous across most insect orders (e.g., [Crampton, 1919](#); [Peck, 1937a](#); [Peck, 1937b](#); [Michener, 1944](#)). This theory was recently corroborated by [Boudinot \(2018\)](#) (discussed further below). The second, the phallic origin theory, postulates that at least in Hymenoptera, the male genitalia have arisen *de novo* (e.g., [Snodgrass, 1935](#); [Snodgrass, 1941](#); [Snodgrass, 1957](#)). Early studies (e.g., [Boulangé, 1924](#); [Beck, 1933](#); [Snodgrass, 1941](#); [Michener, 1956](#); [Smith, 1969](#); [Smith, 1970a](#); [Smith, 1970b](#); [Togashi, 1970](#); [Smith, 1972](#); [Birket-Smith, 1981](#); [Kopelke, 1981](#)) attempted to provide a list of synonymous terms, but suffered from mistakes and incongruities, leading to an increase, rather than a reduction, of the confusion.

It was only with [Schulmeister \(2001\)](#) that the first modern and comprehensive work was produced, combining a complete list of synonyms with an attempt to understand the organization of the male copulatory organs in the basal lineages of Hymenoptera. [Schulmeister \(2003\)](#) then extended the analysis to the other Hymenoptera, providing the first morphological matrix for the order, making extensive use of characters from the male genitalia. [Schulmeister's \(2001, 2003\)](#) analyses facilitated and bolstered further studies of genitalia within Hymenoptera, allowing subsequent refinement of terms (e.g., [Boudinot, 2013](#); [Mikó et al., 2013](#)). However, more recently, [Boudinot \(2018\)](#) rejected the phallic origin theory and provided a new genital terminology for basal Hymenoptera, generating more confusion.

To ameliorate this terminological quagmire, and to facilitate future taxonomic and evolutionary studies, a modern study of the hymenopteran male genitalia is hereby presented. The current contribution provides: (1) a thorough review of the literature; (2) a list of the preferred terminology for the different skeleto-muscular elements of Hymenoptera accompanied by a list of synonyms; (3) the first unified terminology for the skeleto-muscular element of Ichneumonoidea; (4) an alignment of the musculature across the order Hymenoptera; and (5) confirmation of the presence or absence of muscles within Ichneumonoidea.

MATERIALS AND METHODS

Sample preparations and imaging

Specimens used for dissection and imaging via confocal laser scanning microscope (CLSM) were collected in Manitoba (Canada), Florida (USA) and Arizona (USA) ([Table 1](#)), preserved in 70–90% ethanol and deposited at the University of Central Florida Collection (UCFC). Specimens collection at the Hal Scott Regional Preserve and Park was approved by St. Johns River Water Management District. Male genitalia were dissected by means of minuten pins under a dissecting stereomicroscope OPTIKA SZM-2 which was also used

for observations. Specimens used for CLSM imaging were bleached in 30% H₂O₂ for two hours, then placed in a droplet of glycerol and imaged with a ZEISS 710 CLSM at the microscope facility of the Burnett School of Biomedical Sciences (University of Central Florida) using 405 and 488nm lasers (following [Mikó & Deans \(2013\)](#)). Autofluorescence was collected using three channels with assigned contrasting pseudocolors (420–520 nm, blue; 490–520 nm, green; and 570–670 nm, red). Volume-rendered images and media files were generated using ImageJ ([Schindelin et al., 2015](#)). Male genitalia not used in CLSM imaging were left to dry and then glued to the tip of a minuten pin and imaged using a Canon Eos 7D camera with a Canon MP-E 65 mm f/2.8 1-5 × Macro and an M Plan Apo 10×Mitutoyo objective mounted onto the EF Telephoto 70–200 mm Canon zoom lens, and rendered using Zerene Stacker software v. 1.04. Images were enhanced using Photoshop 23.2.2.

Morphological nomenclature

Differently from previous authors who used letters (e.g., [Boulangé, 1924](#); [Schulmeister, 2001](#); [Schulmeister, 2003](#)) or numbers (e.g., [Snodgrass, 1941](#); [Alam, 1952](#)) to label the different muscle bundles, we follow [Daly \(1964\)](#), [Friedrich & Beutel \(2008\)](#), [Vilhelmsen \(1996\)](#); [Vilhelmsen \(2000a\)](#); [Vilhelmsen \(2000b\)](#), and [Mikó et al. \(2007\)](#); [Mikó et al. \(2013\)](#), referring to the muscles as follows: the first component of the name refers to the site of origin, while the second refers to the site of insertion of the muscle. For example, the *proximoventral gonostyle/volsella complex-penisvalva muscle* is the muscle that is attached to the *gonostyle/volsella complex* and to the *penisvalva*. The proximoventral location differentiates it from the other *gonostyle/volsella complex-penisvalva* muscles.

For deciding the preferred term among synonyms, we follow the criteria established by [de Brito, Lanes & Azevedo \(2021\)](#), with the integration of a new criterion (not in order of importance or priority): (1) the term that best represents the skeletal feature (shape and location of the body); (2) the term that is most widely used; (3) the term that was first introduced (oldest); (4) the term not employed also for other structures in other taxa (homonymy).

A list of the unified terminology employed here is provided in [Tables 1–4](#) along with associated abbreviations and definitions included into an ontological framework.

Morphological concepts

The insect cuticle is a continuous, acellular product of the single-layered outer epithelium (the *epidermis*) ([Hall, 1975](#); [Adler, 1975](#); [Denk-Lobnig & Martin, 2020](#)), consisting of comparatively rigid sclerites and comparatively flexible conjunctivae that alternate across the cuticle. The differences in flexibility of different cuticular regions allow the sclerites to change position relative to each other, enabling insects to achieve a wide range of motion types ([Mikó et al., 2013](#); [Girón et al., 2023](#)).

The number, shape, and pattern of the sclerite-conjunctiva system varies between taxa and has changed throughout the course of evolution. One notable difference among taxa is sclerite “fusion” or “division”; the former occurs when two or more sclerites merge due to the disappearance of the separating conjunctiva, the latter by splitting of a pre-existing

Table 1 Material examined with subfamily classification, locality information, repository, voucher number, and preservation method.

Subfamily	Taxon	Country	State/ Province	Locality	Collector(s)	Repository	Voucher number	Preservation method
Cremastinae	<i>Temelucha</i> sp.	USA	Florida	Hal Scott Regional Preserve, Pine Flatwoods	D. Dal Pos & A. Pandolfi	UCFC	UCFMG_0000006	Glycerol
Ichneumoninae	<i>Coelichneumon sassacus</i> (Viereck, 1917)	Canada	Manitoba	Whiteshell Prov. Pk., Pine Point Rapids Trail	Sharanowski lab	UCFC	UCFMG_0000001	Point
Ichneumoninae	<i>Melanichneumon lissorufus</i> Heinrich, 1962	Canada	Manitoba	Spruce Woods Prov. Pk.	Sharanowski lab	UCFC	UCFMG_0000002	Point
Labeninae	<i>Labena grallator</i> (Say, 1835)	USA	Florida	Highland Co., Archbold Biological Station	Y. M. Zhang	UCFC	UCFMG_0000003	Point
Labeninae	<i>Labena grallator</i> (Say, 1835)	USA	Florida	Hal Scott Regional Preserve, Cypress swamp	D. Dal Pos & A. Pandolfi	UCFC	UCFMG_0000016	Glycerol
Mesochorinae	<i>Mesochorus</i> sp.	Canada	Manitoba	University of Manitoba, Points	UW/ELZ	UCFC	UCFMG_0000011	Glycerol
Mesochorinae	<i>Mesochorus</i> sp.	Canada	Manitoba	University of Manitoba, Points	UW/ELZ	UCFC	UCFMG_0000014	Glycerol
Mesochorinae	<i>Mesochorus</i> sp.	Canada	Manitoba	University of Manitoba, Points	UW/ELZ	UCFC	UCFMG_0000015	Glycerol
Pimplinae	<i>Pimpla marginella</i> Brullé, 1846	USA	Florida	Hal Scott Regional Preserve, Pine Flatwoods	D. Dal Pos & A. Pandolfi	UCFC	UCFMG_0000004	Glycerol
Poemeninae	<i>Neoxorides pilosus</i> Townes, 1960	Canada	Manitoba	Agassiz Prov. Pk.	Sharanowski lab	UCFC	UCFMG_0000012	Glycerol
Tryphoninae	<i>Netelia</i> sp.	Canada	Manitoba	University of Manitoba, Points	UW/ELZ	UCFC	UCFMG_0000004	Point
Tryphoninae	<i>Netelia</i> sp.	USA	Florida	Martin Co., Seabranh Preserve SP, Baygall	D. Serrano	UCFC	UCFMG_0000007	Glycerol
Tryphoninae	<i>Netelia</i> sp.	USA	Arizona	Coconino Co., Tonto National Forest, 1 m W of Payson, 1440 m	D. Dal Pos & A. Pandolfi	UCFC	UCFMG_0000008	Glycerol
Tryphoninae	<i>Netelia</i> sp.	USA	Arizona	Coconino Co., Tonto National Forest, 1 m W of Payson, 1440 m	D. Dal Pos & A. Pandolfi	UCFC	UCFMG_0000009	Glycerol
Tryphoninae	<i>Netelia</i> sp.	Canada	Manitoba	Whiteshell Prov. Pk	Sharanowski lab	UCFC	UCFMG_0000017	Glycerol
Rhyssinae	<i>Rhyssa persuasoria</i> (Linnaeus, 1758)	Canada	Manitoba	Whiteshell Prov. Pk.	Sharanowski lab	UCFC	UCFMG_0000010	Glycerol
Xoridinae	<i>Xorides eastoni</i> (Rohwer, 1913)	Canada	Manitoba	Spruce Woods Prov. Pk	Sharanowski lab	UCFC	UCFMG_0000013	Glycerol

Table 2 Anatomical terms used for skeletal features, cross-referenced to an ontological (formal) definition in the Hymenoptera Anatomy Ontology (HAO), linked through the HAO Uniform Resource Identifier (URI).

Abbreviation	Label	Definition	URI
S9	Abdominal sclerite 9	The abdominal sternum that is located on abdominal segment 9.	http://purl.obolibrary.org/obo/HAO_0000047
aed	Aedeagus	The anatomical cluster that is composed of sclerites that are adjacent to the distal end of the ejaculatory duct	http://purl.obolibrary.org/obo/HAO_0000091
ag	Apex gonostipitis	The apodeme that is located medially on the proximoventral margin of the gonostipes and is the site of origin of the ventral gonotyle/volsella complex-penisvalval muscles	http://purl.obolibrary.org/obo/HAO_0000134
aps	Apiceps	The area that is the distal part of the gonossiculus and is connected to the parossiculus via membranous conjunctiva	http://purl.obolibrary.org/obo/HAO_0000141
bsr	Basiura	The area that is the proximal part of the gonossiculus and corresponds to the site of insertion of medial penisvalvo-gonossicula muscle	http://purl.obolibrary.org/obo/HAO_0000179
bs	Basivolsella	The area that is located on the parossiculus ventromedially of the cuspis	http://purl.obolibrary.org/obo/HAO_0001085
c	Cupula	The sclerite that is connected via conjunctiva and attached via muscles to abdominal tergum 9 and the gonostyle/volsella complex	http://purl.obolibrary.org/obo/HAO_0000238
cus	Cuspis	The projection that is located apicolaterally on the parossiculus and is adjacent to the digitus	http://purl.obolibrary.org/obo/HAO_0000239
ejd	Ejaculatory duct	The duct that connects the vas deferens with the endophallus and is ectodermal in origin.	http://purl.obolibrary.org/obo/HAO_0000283
end	Endophallus	The conjunctiva that connects the gonopore with the penisvalvae	http://purl.obolibrary.org/obo/HAO_0000291

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Table 2 (continued)

Abbreviation	Label	Definition	URI
erg	Ergot	The apodeme that is lateral, located medially on the penisvalva and corresponds to the sites of insertion of the lateral and distoventral gonostyle/volsella complex -penisvalval muscle and the parossiculo-penisvalval muscle	http://purl.obolibrary.org/obo/HAO_0000308
fd	Fibula ducti	The sclerite that is located in the proximal end of the unpaired part of the ductus ejaculatorius	http://purl.obolibrary.org/obo/HAO_0000328
fg	Foramen genitale	The anatomical space that is surrounded by the proximal margin of the cupula	http://purl.obolibrary.org/obo/HAO_0000346
gnm	Gonomacula	The conjunctiva that is located at the distal apex of the harpe	http://purl.obolibrary.org/obo/HAO_0000382
gss	Gonossiculus	The sclerite that is located on the distoventral part of the gonostyle/volsella complex, and is articulated with the more proximal sclerites of the gononstyle/volsella complex	http://purl.obolibrary.org/obo/HAO_0000385
gst	Gonostipes	The sclerite that is located dorsolaterally on the gonostyle/volsella complex, is connected to the distal margin of the cupula, to the proximal margin of the harpe, and to the lateral margin of the volsella	http://purl.obolibrary.org/obo/HAO_0000386
gsa	Gonostipital arm	The apodeme that is located proximally on the ventral part of the gonostipes	http://purl.obolibrary.org/obo/HAO_0000387
gs	Gonostyle	The anatomical cluster that is composed of sclerites located distally of the cupula dorsolaterally of the volsella, and that surround the aedeagus	http://purl.obolibrary.org/obo/HAO_0000389

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Table 2 (continued)

Abbreviation	Label	Definition	URI
hrp	Harpe	The sclerite that is located distally on the gonostyle/volsella complex and does not connect to the cupula, and does not connect to the volsella by conjunctiva or muscles	http://purl.obolibrary.org/obo/HAO_0000395
mss	Median sclerotized style	The sclerite that is located ventrally between the penisvalvae	http://purl.obolibrary.org/obo/HAO_0000531
prp	Parapenis	The area that is the dorsomedian part of the gonostipes and is the site of origin of the distodorsal and proximodorsal gonostyle/volsella complex-penisvalval muscles	http://purl.obolibrary.org/obo/HAO_0000692
pss	Parossiculus	The sclerite that is connected via conjunctiva distomedially to the gonostipes, and articulates with the gonossiculus	http://purl.obolibrary.org/obo/HAO_0000703
ph	Phallotrema	The anatomical space that is the distal opening of the endophallus	http://purl.obolibrary.org/obo/HAO_0000714
pv	Penisvalva	The sclerite that is in the middle of the external male genitalia, surrounds the distal part of the ductus ejaculatorius and the endophallus.	http://purl.obolibrary.org/obo/HAO_0000707
pgp	Primary gonopore	The anatomical space that is the transition from the ductus ejaculatorius to the endophallus and therefore the transition from the internal to the external male genitalia.	http://purl.obolibrary.org/obo/HAO_0000821
sp	Spatha	The sclerite that is unpaired and located just dorsally of the basal part of the aedeagus in some Aculeata.	http://purl.obolibrary.org/obo/HAO_0000942
sv	Seminal vesicle	The anatomical space that functions as storage of spermatozoa	http://purl.obolibrary.org/obo/HAO_0001081

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Table 2 (continued)

Abbreviation	Label	Definition	URI
spc	Spiculum	The apophysis that is located medially on the anterior margin of the abdominal sternum 9 and corresponds to the site of origin of the mediolateral S9-cupulal muscles	http://purl.obolibrary.org/obo/HAO_0000946
ts	Testis	The gonad that is consisting of testis follicles, is connected with the vas deferens and seminal vesicle	http://purl.obolibrary.org/obo/HAO_0001007
vd	Vas deferens	The duct that connect the testis with the ejaculatory duct	http://purl.obolibrary.org/obo/HAO_0001052
vvc	Valviceps	The area that is the distal part of the penisvalva dorsally of the ergot	http://purl.obolibrary.org/obo/HAO_0001047
vvr	Valvura	The area that is located proximally of the ergot on the penisvalva.	http://purl.obolibrary.org/obo/HAO_0001050
vol	Volsella	The anatomical cluster that is composed of the sclerites on the ventral part of the male genitalia that are not connected to the cupula via muscles	http://purl.obolibrary.org/obo/HAO_0001084

Table 3 Anatomical terms used for the muscles, cross-referenced to an ontological (formal) definition in the Hymenoptera Anatomy Ontology (HAO), linked through the HAO Uniform Resource Identifier (URI).

Abbreviation	Label	Definition	URI
c-gsdl	Dorsolateral cupulo-gonostyle/volsella complex	The cupulo-gonostyle/volsella complex muscle that arises from the dorsolateral part of the cupula, just laterally of the site of origin of the dorsomedian cupulo-gonostipal muscle, and inserts on the dorsolateral part of the gonostipes	http://purl.obolibrary.org/obo/HAO_0000278
c-gsdm	Dorsomedial cupulo-gonostyle/volsella complex muscle	The cupulo-gonostyle/volsella complex muscle that inserts medially on the dorsal region of the gonostyle/volsella complex	http://purl.obolibrary.org/obo/HAO_0000279
c-gsvl	Ventrolateral cupulo-gonostyle/volsella complex muscle	The cupulo-gonostyle/volsella complex muscle that inserts ventrolaterally on the gonostyle/volsella complex between the site of insertion of the ventromedial and dorsolateral cupulo-gonostyle/volsella complex muscles.	http://purl.obolibrary.org/obo/HAO_0001074
c-gsvm	Ventromedial cupulo-gonostyle/volsella complex muscle	The cupulo-gonostyle/volsella complex muscle that inserts medially on the ventral region of the gonostyle/volsella complex.	http://purl.obolibrary.org/obo/HAO_0001075
gn-pssd	Distal gonostipo-parossicular muscle	The gonostyle/parossicular muscle that arises distally of the lateral part of the gonostipes and inserts on the distal part of the parossiculus distally of the site of origin of the proximal gonostipo-parossicular muscle	http://purl.obolibrary.org/obo/HAO_0000247
gn-pssp	Proximal gonostipo-parossicular muscle	The gonostyle-parossicular muscle that arises proximally from the lateral part of the gonostipes and inserts on the proximal part of the parossiculus	http://purl.obolibrary.org/obo/HAO_0000876
gs-gs	Intragonostyle muscle	The muscle that connects the ventral and dorsal walls of the gonostyle basally	http://purl.obolibrary.org/obo/HAO_0002581
gs-pss	Gonostyle/volsella complex-parossicular muscle	The muscle that arises ventromedially from the gonostyle, is proximomedially oriented, and inserts on the proximalmost sclerite of the volsella.	http://purl.obolibrary.org/obo/HAO_0002041
gs-pvdd	Distodorsal gonostyle/volsella complex - penisvalval muscles	The dorsal gonostyle/volsella complex-penisvalval muscle that arises distodorsally from the gonostyle volsella complex and inserts on the proximal region of the penisvalva	http://purl.obolibrary.org/obo/HAO_0000250
gs-pvdv	Distoventral gonostyle/volsella complex - penisvalval muscle	The ventral gonostyle/volsella complex-penisvalval muscle that arises from the proximoventral part of the gonostyle/volsella complex, inserts medially on the penisvalva and is oriented distodorsally	http://purl.obolibrary.org/obo/HAO_0000251

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Table 3 (continued)

Abbreviation	Label	Definition	URI
gs-pvl	Lateral gonostyle/volsella complex-penisvalval muscle	The gonostyle/volsella complex-penisvalval muscle that arises anterolaterally of the site of origin of the distodorsal gonostyle/volsella complex-penisvalval muscle and inserts laterally on the penisvalva	http://purl.obolibrary.org/obo/HAO_0000472
gs-pvpd	Proximodorsal gonostyle/volsella complex - penisvalva muscle	The dorsal gonostyle/volsella complex penisvalval muscle that arises proximodorsally from the gonostyle/volsella complex and inserts on the penisvalva distally of the site of insertion of the distodorsal gonostyle/volsella complex-penisvalva muscle.	http://purl.obolibrary.org/obo/HAO_0000877
gs-pvpv	Proximoventral gonostyle/volsella complex - penisvalval muscle	The ventral gonostyle/volsella complex-penisvalval muscle that arises ventromedially from the gonostyle/volsella complex, inserts on the proximal end of the penisvalva and is oriented proximodorsally	http://purl.obolibrary.org/obo/HAO_0000879
gss-ph	Gonossiculo-phallotremal muscle	The muscle that arises from the gonossiculus and inserts on the phallotrema.	http://purl.obolibrary.org/obo/HAO_0002577
ha-gon	Harpo-gonomaculal muscle	The male genitalia muscle that arises form the harpe and inserts on the gonomacula	http://purl.obolibrary.org/obo/HAO_0000396
gs-hra	Apical gonostyle/volsella complex - harpal muscles	The gonostyle/volsella complex-harpal muscle that arises from the distolateral margin of the gonostyle/volsella complex and inserts on the lateral wall of the harpe	http://purl.obolibrary.org/obo/HAO_0000246
ga-hrd	Distal gonostyle/volsella complex-harpal muscle	The gonostyle/volsella complex-harpal muscle that inserts on the median wall of the harpe and arises distally of the site of origin of the proximal gonostyle/volsella complex-harpal muscle	http://purl.obolibrary.org/obo/HAO_0000336
gs-hrp	Proximal gonostyle/volsella complex-harpal muscle	The gonostyle/volsella complex-harpal muscle that inserts on the median wall of the harpe and arises proximally of the site of origin of the distal gonostyle/volsella complex-harpal muscle	http://purl.obolibrary.org/obo/HAO_0000926
imvl	Median gonostyle/volsella complex-volsella muscle	The gonostyle/volsella complex-volsella muscle that arises medially of the submedian conjunctiva on the distoventral margin of gonostyle/volsella complex	http://purl.obolibrary.org/obo/HAO_0000473
imvll	Lateral gonostyle/volsella complex-volsella muscle	The gonostyle/volsella complex-volsella muscle that arises laterally of the submedian conjunctiva on the distoventral margin of gonostyle/volsella complex	http://purl.obolibrary.org/obo/HAO_0002580

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Table 3 (continued)

Abbreviation	Label	Definition	URI
imvm	Gonostyle/volsella complex-gonossiculus muscle	The gonossiculus muscle that arises ventromedially from the gonostyle/volsella complex and inserts laterally on the gonossiculus	http://purl.obolibrary.org/obo/HAO_0000517
pss-ph	Parossiculo-phallotremal muscle	The male genitalia muscle that originates from the parossiculus and inserts on the endophallic membrane around the phallotrema	http://purl.obolibrary.org/obo/HAO_0000702
pss-pv	Parossiculo-penisvalval muscle	The male genitalia muscle that arises from the proximal apex of the parossiculus and inserts medially on the penisvalva. The muscle inserts on the ergot if present.	http://purl.obolibrary.org/obo/HAO_0000701
pv-gssl	Lateral penisvalvo-gonossiculus muscle	The penisvalvo gonossiculus muscle that is lateral to the medial penisvalvo-gonossiculus muscle and attaches to the apiceps	http://purl.obolibrary.org/obo/HAO_0002579
pv-gssm	Medial penisvalvo-gonossiculus muscle	The penisvalvo-gonossiculus muscle that is medial to the lateral penisvalvo-gonossiculus muscle and attaches to the basiura.	http://purl.obolibrary.org/obo/HAO_0002578
pv-mss	Penisvalvo-median sclerotized style muscle	The muscle that attaches to the median sclerotized style and to the valvura.	http://purl.obolibrary.org/obo/HAO_0002582
pv-ph	Penisvalvo-phallotremal muscle	The male genitalia muscle that arises from the medial surface of the proximal part of the penisvalva and inserts on the endophallus just around the phallotrema.	http://purl.obolibrary.org/obo/HAO_0000710
pv-pv	Interpenisvalval muscle	The male genitalia muscle that connects the valvurae.	http://purl.obolibrary.org/obo/HAO_0000433
S9-cl	Lateral S9-cupulal muscle	The S9-cupulal muscle that arises sublaterally from S9 and inserts medioventrally on the cupula	http://purl.obolibrary.org/obo/HAO_0000464
S9-cm	Medial S9-cupulal muscle	The male genitalia muscle that arises from the spiculum and inserts on the gonocondyle	http://purl.obolibrary.org/obo/HAO_0000516
S9-cml	Mediolateral S9-cupulal muscle	The cupulal muscle that arises medially from abdominal sternum 9 and inserts ventrolaterally on the cupula	http://purl.obolibrary.org/obo/HAO_0000533
vl-vl	Intervolsellal muscle	The male genitalia muscle that connects the proximal part of parossiculi.	http://purl.obolibrary.org/obo/HAO_0000441

Table 4 Anatomical terms used for the muscles and cross-referenced with terms (including numbers and letters) used by other authors. An en-dash symbol (–) identified that that specific muscle has not been treated by the author in the given work.

Abbreviation	Label	Boulangé (1924)	Peck (1937a); Peck (1937b)	Snodgrass (1941)	Alam (1952)	Schulmeister (2001); Schulmeister (2003)	Mikó et al. (2013)
S9-cm	Medial S9-cupulal muscle	a	A	1	2	a	–
S9-cml	Mediolateral S9-cupulal muscle	b	B	2	3	b	Mediolateral S9-cupulal muscle
S9-cl	Lateral S9-cupulal muscle	c	C	3	1	c	Lateral S9-cupulal muscle
c-gsvm	Ventromedial cupulo-gonostyle/volsella complex muscle	d	D	4	7	d	Ventromedial cupulo-gonostyle/volsella complex muscle
c-gsvl	Ventrolateral cupulo-gonostyle/volsella complex muscle	e	E	5	4, 5	e	Ventrolateral cupulo-gonostyle/volsella complex muscle
c-gsdl	Dorsolateral cupulo-gonostyle/volsella complex	f	F	7	–	f	Dorsolateral cupulo-gonostyle/volsella complex
c-gsdm	Dorsomedial cupulo-gonostyle/volsella complex muscle	g	G	6	6	g	Dorsomedian cupulo-gonostyle/volsella complex muscle
gs-pvpv	Proximoventral gonostyle/volsella complex - penisvalval muscle	h	H	8	14	h	Proximoventral gonostyle/volsella complex - penisvalval muscle
gs-pvdv	Distoventral gonostyle/volsella complex - penisvalval muscle	i	I	9	15	i	Distoventral gonostyle/volsella complex - penisvalval muscle
gs-pvdd	Distodorsal gonostyle/volsella complex - penisvalval muscles	j	J	10	13	j	Distodorsal gonostyle/volsella complex - penisvalval muscles
gs-pvpd	Proximodorsal gonostyle/volsella complex - penisvalval muscle	k	K	11	16	k	Proximodorsal gonostyle/volsella complex - penisvalval muscle
gs-pvl	Lateral gonostyle/volsella complex-penisvalval muscle	l	L	12	18	l	Lateral gonostyle/volsella complex-penisvalval muscle
pv-gssl	Lateral penisvalvo-gonossicula muscle	m	M	22	12	m	–
pv-gssm	Medial penisvalvo-gonossicula muscle				–	n	–
pv-ph	Penisvalvo-phallotremal muscle	n	N	24	17	nb	–
gss-ph	Gonossiculo-phallotremal muscle				–	nd	–
pss-ph	Parossiculo-phallotremal muscle				–	nl	–
gs-pss	Gonostyle/volsella complex-parossicula muscle			–	–	o	Gonostyle/volsella complex-parossicula muscle
gn-pssp	Proximal gonostipo-parossicula muscle	o	O	18	8	o'	–
gn-pssd	Distal gonostipo-parossicula muscle			20	–	o''	–
imvll	Lateral gonostyle/volsella complex-volsella muscle	p	P	19	10	p	Lateral gonostyle/volsella complex-volsella muscle
imvl	Median gonostyle/volsella complex-volsella muscle	q	Q				
		r	R	21	9	qr	Medial gonostyle/volsella complex-volsella muscle
imvm	Gonostyle/volsella complex-gonossiculus muscle	s	S	23	11	s	Gonostyle/volsella complex-gonossiculus muscle
pss-pv	Parossiculo-penisvalval muscle	si	–	–	–	si	Parossiculo-penisvalval muscle
gs-hrd	Distal gonostyle/volsella complex-harpal muscle	t	T	16	–	t'	Distal gonostyle/volsella complex-harpal muscle
gs-hrp	Proximal gonostyle/volsella complex-harpal muscle			15	–	t''	Proximal gonostyle/volsella complex-harpal muscle

(continued on next page)

Table 4 (continued)

Abbreviation	Label	Boulangé (1924)	Peck (1937a); Peck (1937b)	Snodgrass (1941)	Alam (1952)	Schulmeister (2001); Schulmeister (2003)	Mikó et al. (2013)
ga-hra	Apical gonostyle/volsella complex - harpal muscles	u	U	–	–	u	Distal gonostipes/volsella complex-harpal muscle
ha-gon	Harpo-gonomaculal muscle	v	V	17	–	v	–
gs-gs	Intragonostyle muscle	–	–	–	–	w	–
pv-pv	Interpenisvalval muscle	x	–	13	–	x	–
vl-vl	Intervolsellal muscle	–	–	–	–	y	–
pv-mss	Penisvalvo-median sclerotized style muscle	z	–	14	–	z	–

single sclerite by the development of a conjunctiva across it. For instance, the single sclerite connected to the *cupula* in the male genitalia in Ichneumonidae (=gonostyle) has been interpreted as the result of the fusion of the *harpe* and the *gonostipes*, forming one single continuous structure, named *gonoforceps* by different authors (Peck, 1937a).

The appearance or disappearance of conjunctivae is why the term “complex” (e.g., *gonostyle/volsella complex*) was introduced by Ronquist & Nordlander (1989) and has been widely used for describing anatomical ontologies (Mikó et al., 2013; Aibekova et al., 2022). The term “anatomical complex” refers to a sclerite present in a particular taxon, which occupies a region that in other taxa is occupied by multiple sclerites. For instance, the *volsella* and the *gonostyle* are two different completely separate sclerites in the subfamily Labeninae (Ichneumonidae), but they are partially or entirely continuous in Mesochorinae.

Description format

To help researchers navigate the different terminologies in standard taxonomic descriptions and future evolutionary studies, we provide a detailed morphological treatment of Hymenoptera male genitalia elements using the following structure:

- (1) **Labels** –A list of synonymous labels employed by various authors is provided following the preferred term. The first author listed after the term is either the person who coined the term or the one who applied it for the first time in Hymenoptera, followed by authors who employed the term afterward. Newly proposed synonyms are marked with an asterisk (*).
- (2) **Concept** –A general, homology-free diagnosis of the element and its components with special emphasis on their connectedness and structural properties, *i.e.*, epistemological recognition criteria.
- (3) **Definition** –The Aristotelian definition of the element. Aristotelian definitions are used to build ontologies as they represent universal statements (see Vogt, Bartolomaeus & Giribet (2022) for more discussion).
- (4) **Discussion of terminology** –The review of the usage of terms referring to the element within Hymenoptera with special emphasis on Ichneumonoidea.
- (5) **Preferred term** –the label that has been selected as preferred, using the above criteria.
- (6) **Morphological variation in Ichneumonoidea** –overview of the variation in the elements as observed from dissected specimens or as described in previous literature.
- (7) **Comments** –general comments on the anatomical elements.

Each of the main elements (*abdominal sternum 9*, *cupula*, *gonostyle*, *volsella*, and *penisvalva*) of the male genitalia can be composed of a single sclerite or divided into multiple sclerites. Sclerites can be further divided into regions (also called areas) with more or less well-defined boundaries. For the best modeling of this complex system, we listed these structures nested within each other. This should help the reader navigate across the different terms.

Main elements of the male genitalia are identified by a Roman numeral (e.g., GONOSTYLE=III); followed by another number referring to individual sclerites of the element (e.g., GONOSTIPES=III.1), and finally, a Latin letter identifies regions of the

sclerite (e.g., PARAPENIS=III.1.a). For example, the *parapenis* (III.1.a) is an area of the *gonostipes* (III.1) which is one of the sclerites that compose the *gonostyle* (III).

A note of caution: in some dry specimens, regions can appear more definable and can be possibly misidentified as separate sclerites, instead of being simply areas of certain sclerites. Thus, wet specimens are critical for understanding where sclerites start and end and thus ontological alignment of terms.

RESULTS

A review of Ichneumonoid male genitalia

With more than 48,000 described species, Ichneumonoidea is one of the largest superfamilies of Hymenoptera ([Branstetter et al., 2018](#)) and comprises roughly a third of all recognized species of Hymenoptera ([Sharanowski et al., 2021](#)). It is divided into two families, Braconidae (>21,000 spp.) and Ichneumonidae (>25,000 spp.) ([Quicke, 2015](#); [Yu, van Achterberg & Horstmann, 2016](#); [Klopfstein et al., 2019](#); [Sharanowski et al., 2021](#)).

Many lineages of the superfamily show incredible external variation in the male genitalia. However, the genital morphology in Ichneumonoidea remains substantially undescribed for almost all of the 48,000 species, with the exception of the genus *Netelia* Gray (Ichneumonidae, Tryphoninae) (e.g., [Townes, 1939](#); [Konishi, 1991](#); [Konishi, 1992](#); [Konishi, 1996](#); [Konishi, 2010](#); [Bennett, 2015](#); [Konishi, Chen & Pham, 2022](#)). Some authors have included descriptions of the genitalia in occasional single species descriptions (e.g., [Loan, 1974](#); [Walker, 1994](#); [Watanabe & Matsumoto, 2010](#); [Watanabe, Taniwaki & Kasparyan, 2015](#); [Sobczak et al., 2019](#); [Brajković et al., 2010](#)).

The first description of the male genitalia of Ichneumonidae was provided by [Bordas \(1893\)](#), who analyzed the internal and external genital organs of five taxa, while [Peck \(1937a\)](#) and [Peck \(1937b\)](#) provided the first, and so far only, extensive study of Ichneumonidae male genitalia, with 96 taxa analyzed and comments on muscles, sclerite movements, and homology statements. In Braconidae, [Seurat \(1898\)](#) and [Seurat \(1899\)](#) was the first to mention the genitalia, while [Snodgrass \(1941\)](#) briefly analyzed and compared them with those of Ichneumonidae. Subsequently, [Alam \(1952\)](#) provided a more detailed study on the skeleto-musculature for one species of the subfamily Braconinae, while [Karlsson & Ronquist \(2012\)](#) provided the first modern description of the male external genitalia of two braconid species in the subfamily Opiinae.

Despite the fact that [Schulmeister \(2003\)](#) demonstrated that male genitalia characters can be informative for the higher-level classification of basal Hymenoptera and that other authors have successfully applied these structures in genus-level phylogenetic studies within sawflies ([Malagon-Aldana et al., 2021](#)) and Apocrita ([Andena et al., 2007](#); [Owen et al., 2007](#); [Mikó et al., 2013](#)), external male genitalia are rarely employed in phylogenetic studies on Ichneumonoidea. Slightly more research has been devoted to the genital organs of Braconidae, but most, if not all, of these studies show inconsistent terminologies. Some cite synonymous terms that are no longer valid (as discussed by [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#)), and others associate a valid term with referring to a different sclerite.

The first suprageneric classification of Ichneumonidae using copulatory organs was proposed by [Peck \(1937b\)](#) (based on his previous work ([Peck, 1937a](#))), who emphasized

the importance of the *abdominal sternum 9* (= subgenital plate). Likewise, [Pratt \(1939\)](#) discussed the genital characters of a subset of Ichneumonidae, providing the first (and only) key to the tribes based solely on male genitalia.

Various genera of Braconidae were analyzed by [Tobias \(1967\)](#), who was the first to explore differences between subfamilies, while the subfamily Aphidiinae was extensively studied by [Tremblay \(1979\)](#), [Tremblay \(1981\)](#) and [Tremblay \(1983\)](#). [Quicke \(1988\)](#) surveyed Braconinae, concluding that the characters could be potentially useful for higher-level classification in the subfamily. Later, [Maetô \(1996\)](#) assessed the inter-generic variation of external male genitalia in Microgastrinae, while more recently [Bratkovič et al. \(2010\)](#) and [Žikić et al. \(2011\)](#) did the same in Agathidinae.

Over the years, other authors have included male genitalia in their phylogenies of Ichneumonoidea or one of its two families (e.g., [Wahl & Gauld, 1998](#)) but the degree to which these characters have been employed is minimal. For instance, the recent morphological phylogenetic analyses of Ichneumonidae subfamilies by [Bennett et al. \(2019\)](#) included only four characters of the male terminalia, of which only two belong to the genital capsule. Reasons for the lack of use of male genitalic characters in Ichneumonoidea phylogenetics are unclear but possibly are due to: (1) the Ichneumonoidea classification system is mostly based on females and the association with males has been proven to be challenging; (2) males are rarely dissected, and genitalia characters have never been thoroughly assessed, nor they have been employed in taxonomic studies; and (3) rampant terminological inconsistencies, coupled with the overall complexity of the male genitalia, have discouraged researchers from exploring the male genitalia in Ichneumonoidea. This is why it becomes paramount to overcome at least one of these impediments, providing for the first time a complete assessment of the terminology.

Towards a unified terminology

The male reproductive organs of Hymenoptera are composed of internal and external structures, in continuity with each other ([Schulmeister, 2001](#)). The inner reproductive system is composed of the *testis*, *vas deferens*, *seminal vesicle*, *accessory gland*, and *ductus ejaculatorius*. The external male genitalia consist of five elements: (1) *abdominal sternum 9*; (2) *cupula*; (3) *gonostyle*; (4) *volsella*; and (5) *penisvalva* ([Figs. 1, 2](#)). Note that all structures of the external male genitalia and their historical terms are described in depth below (see ‘Methods and Results’). According to [Schulmeister \(2001\)](#) and [Schulmeister \(2003\)](#), there are also two other sclerites: (6) the *median sclerotized style*, which is a thin sclerite that lies on the median axis of the ventral side of the external genitalia between the two *penisvalvae*, and (7) the *fibula ducti*, which is a sclerite present in the proximal end of the *ductus ejaculatorius*.

The elements of the male genitalia are interconnected through a network of muscles that move the sclerites individually or in conjunction ([Duncan, 1939](#); [Snodgrass, 1941](#); [Snodgrass, 1957](#); [Michener, 1956](#); [Schulmeister, 2001](#); [Schulmeister, 2003](#); [Mikó et al., 2013](#)). Four major muscle groups have been identified within Hymenoptera: (1) *abdominal sternum 9* to *cupula*, which are usually three muscles that control the movement of the entire genital capsule; (2) *cupula* to *gonostyle*, which are usually four distinct bundles that

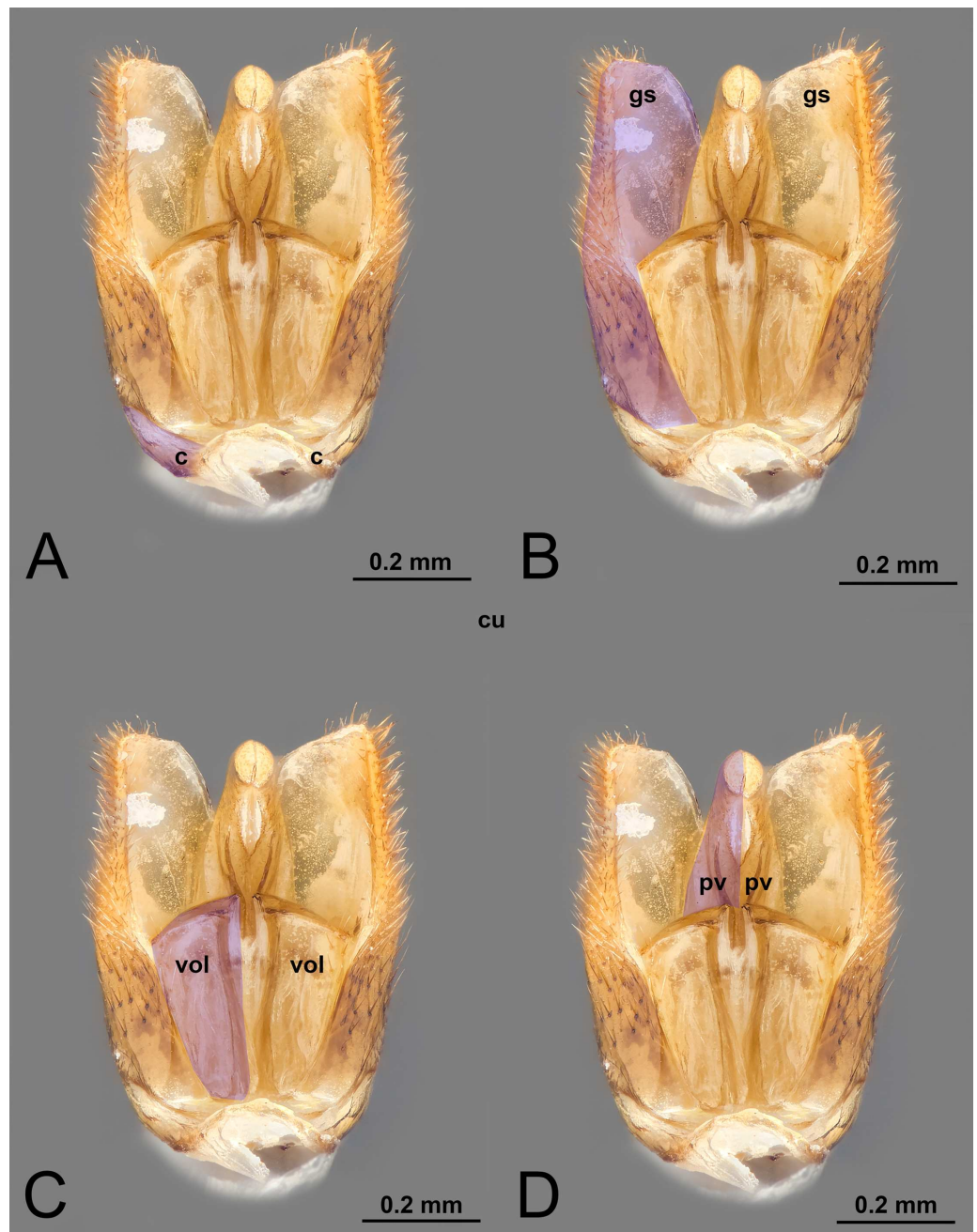


Figure 1 Ventral view of male genitalia of *Melanichneumon lissorufus* (Ichneumonidae: Ichneumoninae) with different elements highlighted as follow: (A) Cupula (c). (B) Gonostyle (gs). (C) Volsella (vol). (D) Penisvalva (pv).

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control the movement of the *gonostyles*; (3) *gonostyle* to *volsella*, which can be up to three distinct muscles that generally control the lateral motion of the *volsella* and the opening and closing of the apical clasp structure; (4) *gonostyle* to *penisvalva*, which usually consist of five distinct bundles, and control the motion of the *penisvalvae*.

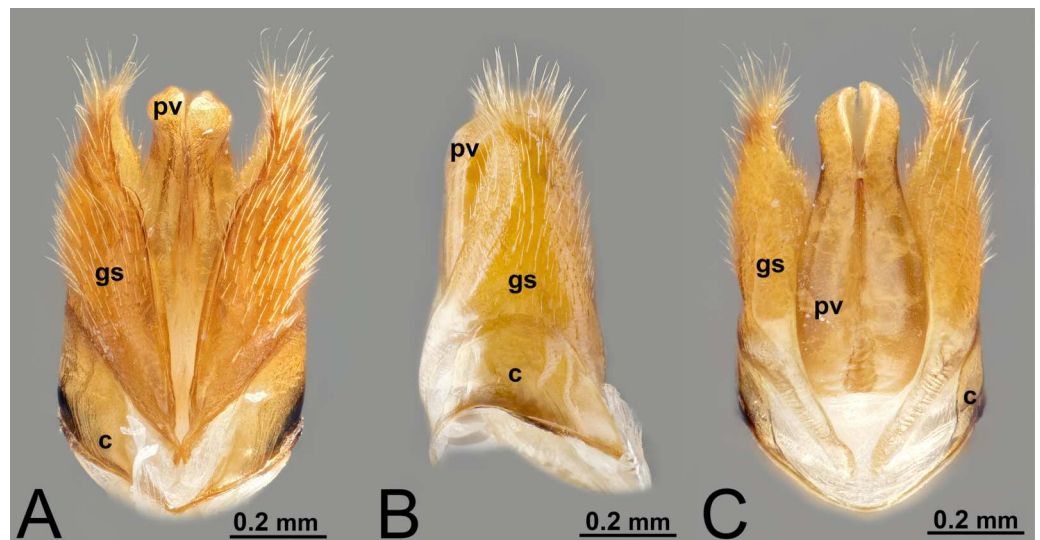


Figure 2 Male genitalia of *Labena grallator* (Ichneumonidae: Labeninae). (A) Ventral view. (B) Lateral view. (C) Dorsal view.

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ABDOMINAL STERNUM 9

I. ABDOMINAL STERNUM 9 (S9, Figs. 3A, 4A, 4C)

ninth abdominal sternite by [Worthley \(1924\)](#); [Snodgrass \(1935\)](#).

subgenital plate by [Snodgrass \(1935\)](#); [Watanabe & Matsumoto \(2010\)](#).

*nono urotergite by [Tremblay \(1979\)](#); [Tremblay \(1981\)](#); [Tremblay \(1983\)](#).

hypopygium by [Pratt \(1939\)](#); [Nichols \(1989\)](#); [Karlsson & Ronquist \(2012\)](#); [Broad, Shaw & Fitton \(2018\)](#); [Bennett et al. \(2019\)](#).

hypopygidium by [Nichols \(1989\)](#); [Schulmeister \(2001\)](#).

hypandrium by [Nichols \(1989\)](#); [Peck \(1937a\)](#); [Peck \(1937b\)](#); [Karlsson & Ronquist \(2012\)](#).

annular lamina by [Nichols \(1989\)](#).

hypotome by [Nichols \(1989\)](#).

ninth sternal lobe by [Nichols \(1989\)](#).

poculus by [Nichols \(1989\)](#).

postgenital plate by [Nichols \(1989\)](#).

metasomal sternum viii by [Brothers & Carpenter \(1993\)](#).

abdominal sternum 9 by [Karlsson & Ronquist \(2012\)](#).

Concept. The abdominal sternum 9 is the ventral part of the ninth abdominal segment and connects the abdominal segments to the genital sclerites. Although not a direct component of the male genitalia, the abdominal sternum 9 has a strong association with the cupula, by means of three major muscles: the medial (S9-cml), the mediolateral (S9-cm), and the lateral S9-cupulal (S9-cl) muscle (Figs. 3A–3B, 4A–4B, 5A–5B; Tables 3–4). These muscles allow the protraction and retraction of the entire male genitalia. The abdominal sternum 9 is usually produced proximo-medially into a process called the spiculum (spc,

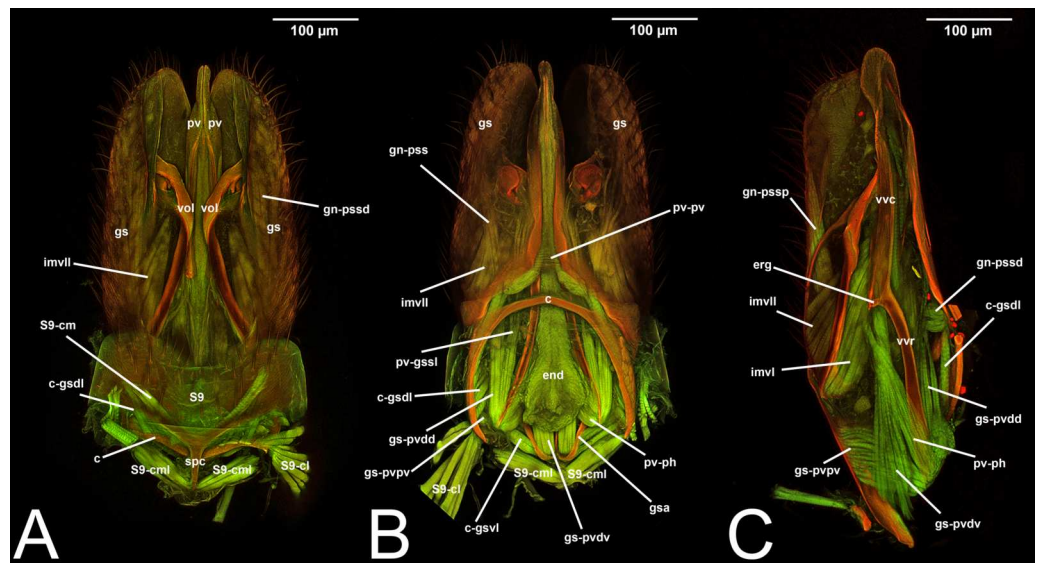


Figure 3 CLSM volume rendered images of male genitalia of *Temelucha* sp. (Ichneumonidae: Cremastinae). (A) Ventral view. (B) Dorsal view. (C) Median view.

Full-size [DOI: 10.7717/peerj.15874/fig-3](https://doi.org/10.7717/peerj.15874/fig-3)

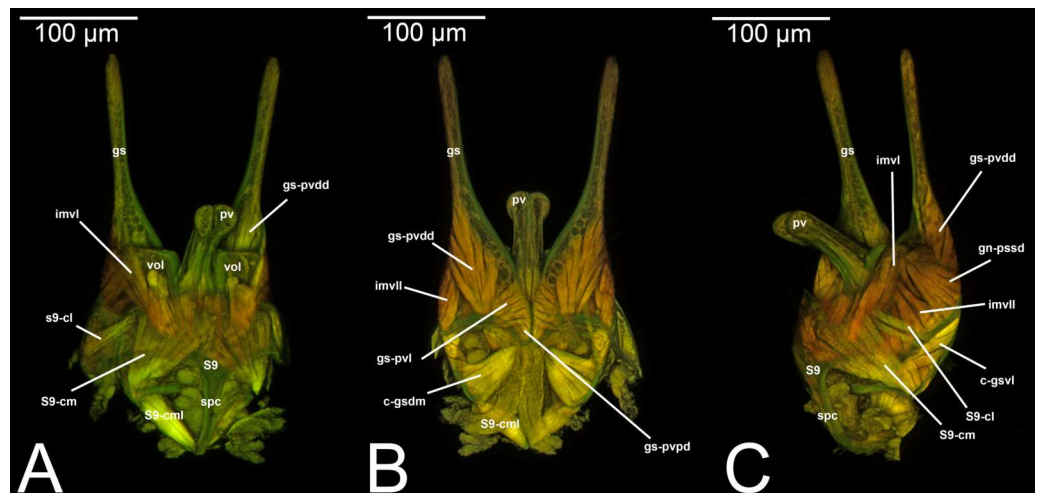


Figure 4 CLSM volume rendered images of male genitalia of *Mesochorus* sp. (Ichneumonidae: Mesochorinae). (A) Ventral view. (B) Dorsal view. (C) Lateral view.

Full-size [DOI: 10.7717/peerj.15874/fig-4](https://doi.org/10.7717/peerj.15874/fig-4)

Figs. 3A, 4A, 4C), which is an apophysis that corresponds to the site of origin of the mediolateral (S9-cml) and medial S9-cupulal (S9-cm) muscles (Figs. 3A–3B, Figs. 4A–4B).

Definition. As defined by the HAO, the *abdominal sternum 9* is the abdominal sternum that is located on *abdominal segment 9* (Table 2).

Discussion of terminology. Many of the terms employed are either variations of *abdominal sternum 9* (e.g., *ninth sternal segment*) or have been rarely employed (e.g., *poculus*). However, one of the most popular terms employed is *hypopygium*, which has

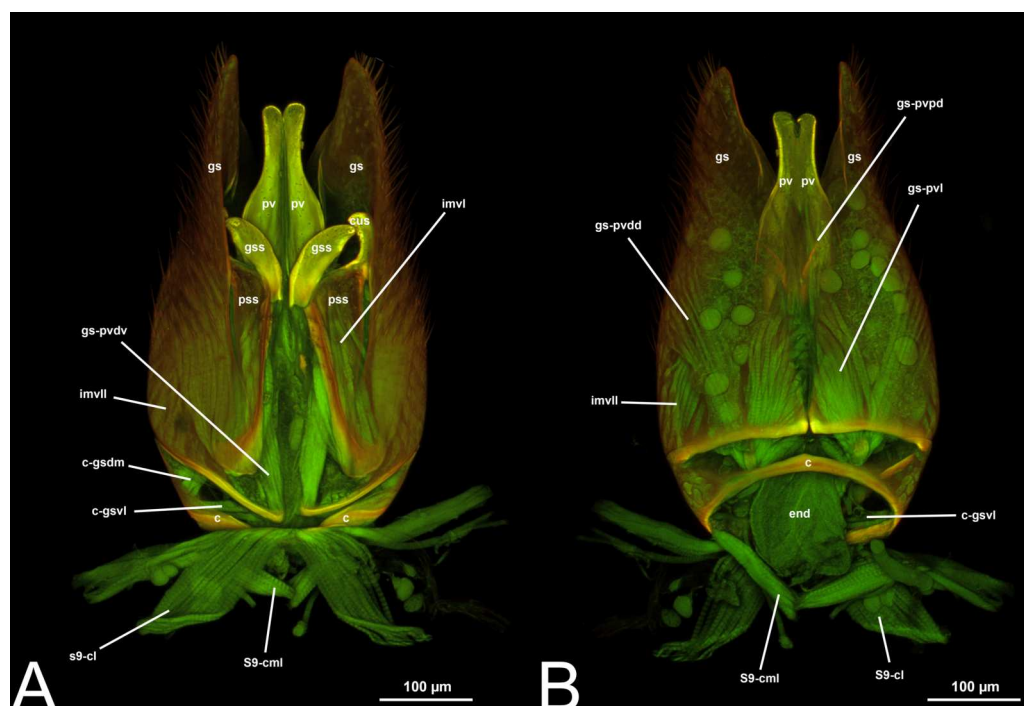


Figure 5 CLSM volume rendered images of male genitalia of *Xorides eastoni* (Ichneumonidae: Xoridinae). (A) Ventral view. (B) Dorsal view.

Full-size [DOI: 10.7717/peerj.15874/fig-5](https://doi.org/10.7717/peerj.15874/fig-5)

been employed to refer to the most posterior sternite in different insect orders. However, from a morphological perspective, the *hypopygium* can be confusing. In fact, it has been used within Hymenoptera to refer to the *abdominal sternum 7* in females (the last observable sternite in females) or to the *abdominal sternum 9* in males (the last observable sternite in males) (e.g., [Karlsson & Ronquist, 2012](#); [Broad, Shaw & Fitton, 2018](#); [Bennett et al., 2019](#)). Moreover, *hypopygium* can also refer to different structures in different orders: in Diptera it refers to the terminalia, in Lepidoptera to the multiple sclerites fused together, while in Cicadomorpha (Hemiptera) it is used for the fused tergal and pleural parts of segment 9 ([Tuxen, 1956](#)). For these reasons, we strongly encourage using abdominal sternum 9, which fulfills criterion 1 and 4.

Within Ichneumonoidea, *hypopygium* has been widely employed (e.g., [Pratt, 1939](#); [Karlsson & Ronquist, 2012](#); [Broad, Shaw & Fitton, 2018](#); [Bennett et al., 2019](#)), while *hypandrium* was employed only by [Peck \(1937a\)](#); [Peck \(1937b\)](#) and by [Karlsson & Ronquist \(2012\)](#). Many other studies tend to exclude this sclerite as being part of the male genitalia (e.g., [Quicke & van Achterberg, 1990](#)).

Preferred term. Abdominal sternum 9.

Morphological variation in Ichneumonoidea. According to several authors, the *abdominal sternum 9* varies across the Ichneumonoidea. It has been used for differentiating genera (e.g., [Heinrich, 1961](#)) and employed in phylogenetic reconstruction (e.g., [Bennett et al., 2019](#)). There are several areas of variation: (1) the distal margin, which can be elongated,

flat, or concave ([Heinrich, 1961](#); [Bennett et al., 2019](#)); (2) the shape of the *spiculum*, which can be extremely elongated (spc, [Figs. 3A, 4A, 4C](#)) or reduced, wide or thin ([Peck, 1937a](#)); (3) in the overall shape of the sclerite (see the image in [Peck, 1937a](#), p. 246).

II. CUPULA

II. CUPULA (c, [Figs. 1A, 2A–2C, 3A–3B, 5A–5B, 6B](#))

cupule by [Audouin \(1821\)](#).

hornringe by [Hartig \(1837\)](#).

horny ring by [Nichols \(1989\)](#).

pièce basilaire by [Dufour \(1841\)](#).

kapsel by [Schenck \(1851\)](#).

cardo by [Thomson \(1872\)](#).

écailles by [André \(1879\)](#).

lame basilaire by [Bordas \(1893\)](#).

ringstück by [Verhoeff \(1832\)](#).

plaque basilaire by [Seurat \(1898\)](#).

sclerite accessorio by [Berlese \(1909\)](#).

annular lamina by [Wheeler \(1910\)](#).

pièce annulaire by [Boulangé \(1914\)](#).

basalstück by [Enslin \(1918\)](#).

gonocardo by [Crampton \(1919\)](#); [Peck \(1937a\)](#); [Peck \(1937b\)](#); [Pratt \(1939\)](#).

lamina annularis by [Snodgrass \(1941\)](#).

gonobase by [Michener \(1944\)](#); [Michener \(1956\)](#); [Delrio \(1975\)](#); [Tremblay \(1979\)](#);

[Tremblay \(1981\)](#); [Tremblay \(1983\)](#); [Quicke & van Achterberg \(1990\)](#);

[Belokobylskij, Zaldivar-Riveron & Quicke \(2004\)](#).

basal ring by [Crampton \(1919\)](#); [Snodgrass \(1935\)](#); [Snodgrass \(1941\)](#); [Alam \(1952\)](#);

[Snodgrass \(1957\)](#); [Tobias \(1967\)](#); [Olmi \(1984a\)](#); [Olmi \(1984b\)](#); [Johnson \(1984\)](#);

[Quicke & van Achterberg \(1990\)](#); [Konishi \(1991\)](#); [Konishi \(1992\)](#); [Konishi \(1996\)](#);

[Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#); [Belokobylskij, Zaldivar-Riveron & Quicke \(2004\)](#);

[Konishi \(2005\)](#); [Brajković et al. \(2010\)](#).

section 1 by [Smith \(1969\)](#).

**guaina basale* by [Tremblay \(1979\)](#); [Tremblay \(1981\)](#); [Tremblay \(1983\)](#).

cupula by [Birket-Smith \(1981\)](#); [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#);

[Karlsson & Ronquist \(2012\)](#); [Mikó et al. \(2013\)](#).

Concept. The *cupula* is an unpaired sclerite located at the proximal basis of the male genitalia. Different authors (see below in the Discussion of terminology) have consistently identified (with different names) an unpaired sclerite surrounding the proximal basis of the male genitalia. The *cupula* delimits a basal opening called the *foramen genitale* ([Snodgrass, 1941](#); [Schulmeister, 2001](#)), and is connected via three muscles to the *abdominal sternum 9* and via four muscles to the *gonostyle* ([Table 4](#)).

Definition. As defined by the HAO, the *cupula* is the sclerite that is connected via conjunctiva and attached via muscle to the abdominal tergum 9 and the *gonostyle/volsella complex* ([Table 2](#)).

Discussion of terminology. At least 21 terms have been introduced to refer to the *cupula*. Of these, only four (*basal ring*, *cupula*, *gonobase*, *gonocardo*) are worth discussing as they have been employed more than once after their introduction. All the other terms (see above), are rejected as they do not conform to criterion 2.

The first author to identify the basal sclerites of the genital organs in Hymenoptera was [Audouin \(1821\)](#), who proposed the term *cupule* (later modified to *cupula* by [Birket-Smith \(1981\)](#)) to refer to an unpaired sclerite, which the author called the “*support commun* [= general support]”.

In 1872, Thomson introduced *cardo* as a new term for the same structure after he studied the genus “*Bombis* [sic] [= *Bombus*]” and identified a basal capsule divided into two halves ([Thomson, 1872](#)). However, [Crampton \(1919\)](#) realized that *cardo* was also used for the basal sclerite of the maxillae in insects (rejected by criterion 4), and he proposed to replace *cardo* with *gonocardo* without realizing that another available term (*cupule*) has been already proposed. In the same work, [Crampton \(1919\)](#) introduced an additional term for the same sclerite, *basal ring*. This latter term, however, refers also to two different structures in two other orders: in Protura, it is used to refer to a basal structure of the male genitalia, while in Diptera, it is used for the combination between tergite and sternum IX ([Tuxen, 1970](#)). Since the term was first introduced in Diptera, and it is not equivalent to the one in Hymenoptera, *basal ring* should not be considered the preferred term (rejected by criterion 4).

Later on, [Michener \(1944\)](#), following the periphallalic theory, introduced *gonobase*, and subsequently, [Michener \(1956\)](#) formally synonymized *gonocardo* under *gonobase*, using the same rationale. Unfortunately, both authors failed to notice that the term *gonobase* corresponds to the non-equivalent *gonobasis* in basal insects (e.g., [Willman, 1998](#); [Schulmeister, 2001](#)), and, therefore, it should also not be used as the preferred term (rejected by criterion 4).

It must be noticed that there is a *cupula* also in Lepidoptera that was introduced by [Field \(1950\)](#) for a pair of pouches located on the 7th abdominal sternite of females ([Tuxen, 1956](#)). These two structures are not analogous to the *cupula* of Hymenoptera (HAO:0000238, see comments). Due to the date of introduction (the *cupula* of [Audouin \(1821\)](#) has been introduced before the *cupula* of [Field \(1950\)](#)) and due to the fact that the term has been specifically introduced in Hymenoptera, the *cupula* of [Audouin \(1821\)](#) can be retained and used as the preferred label for this sclerite (approved by criterion 3).

Within Ichneumonoidea, a number of authors used the term *gonobase* referring to the *cupula* ([Tremblay, 1979](#); [Tremblay, 1981](#); [Tremblay, 1983](#); [Quicke & van Achterberg, 1990](#); [Belokobylskij, Zaldivar-Riveron & Quicke, 2004](#)), and only two used *gonocardo* ([Peck, 1937a](#); [Peck, 1937b](#); [Pratt, 1939](#)) and *basal ring* ([Alam, 1952](#); [Konishi, 2005](#)). As far as we know, the term *cupula* was applied to Ichneumonoidea only by [Schulmeister \(2003\)](#).

Preferred term. *Cupula*.

Morphological variation in Ichneumonoidea. Across Ichneumonoidea, the *cupula* has undergone repeated reduction, sometimes within a subfamily. In *Labena grillator* (Say, 1845) (Ichneumonidae, Labeninae), *Xorides eastoni* (Rohwer, 1913) (Ichneumonidae, Xoridinae), and *Coelichneumon* Thomson, 1896 (Ichneumonidae, Ichneumoninae), the

cupula is well developed, especially on the lateral side (Figs. 2A–2B, 6A), while in *Mesochorus* Gravenhorst, 1829 (Ichneumonidae, Mesochorinae) and *Melanichneumon* Thomson, 1893 (Ichneumonidae, Ichneumoninae) (Fig. 1A) it is significantly reduced. Similar variations have also been observed by previous authors. *Peck (1937b)* described a ventrolaterally unusually broad *cupula* in *Megarhyssa macrura* Linnaeus, 1771 (Ichneumonidae, Rhyssinae) and *Banchus falcatorius* (Fabricius, 1775) (Hymenoptera, Banchinae), while *Pratt (1939)* noticed variations between members of the same subfamily, with *Delomerista* Förster, 1869, and *Perithous* Holmgren, 1859 (Ichneumonidae, Pimplinae) showing a well-developed and an extremely reduced *cupula*, respectively. The same pattern was observed in Braconidae, with some subfamilies (e.g., Braconinae and Doryctinae) bearing a very elongated *cupula* (Figs. 77. 78 in *Quicke & van Achterberg, 1990*, p. 62), while in others (e.g., Histomerinae) the same sclerite is reduced (Fig. 73 in *Quicke & van Achterberg, 1990*, p. 61).

Comments on cupula. *Vilhelmsen (1997)* identified the *cupula* as a hymenopteran synapomorphy, but this result will likely require further investigation. In fact, other insect groups have a sclerite located basally to the rest of the sclerites and connected to the abdominal sternum 9 (e.g., the *phallobase* in Coleoptera, see *Snodgrass, 1957*, p. 30, fig. C).

A final note concerns *Boudinot (2018)*, while he rejected the phallic theory, he did not introduce a new term for the *cupula*, but considered the structure a part of the fragmented base of genital appendages.

III. GONOSTYLE

III. GONOSTYLE (gs, Figs. 1–7)

gonoforceps (part.) by *Peck (1937a)*; *Peck (1937b)*; *Michener (1944)*; *Ross (1945)*; *Michener (1956)*; *Olmi (1984a)*; *Olmi (1984b)*; *Schulmeister (2001)*; *Schulmeister (2003)*; *Karlsson & Ronquist (2012)*; *Bennett et al. (2019)*.

**squama* by *Townes (1939)*.

paramere (part.) by *Snodgrass (1957)*; *Pratt (1939)*; *Tobias (1967)*; *Olmi (1984a)*; *Olmi (1984b)*; *Quicke & van Achterberg (1990)*; *Belokobylskij, Zaldivar-Riveron & Quicke (2004)*; *Konishi (2005)*; *Watanabe & Matsumoto (2010)*; *Žikić et al. (2011)*; *Boudinot (2013)*; *Watanabe, Taniwaki & Kasparyan (2015)*; *Broad, Shaw & Fitton (2018)*; *Brajković et al. (2010)*.

outer or central arm by *Arnold (1951)*.

gonopoden des 9 Segmentes by *Haupt (1962)*.

paramere exteriores by *Priesner (1966)*.

**claspers* by *Townes (1969a)*; *Townes (1969b)*; *Townes (1969c)*; *Dasch (1971)*; *Townes (1971)*; *Lee (1991)*.

gonostyle by *Bohart & Menke (1976)*; *Mikó et al. (2013)*.

stipes by *Birket-Smith (1981)*.

**gonostipite* by *Tremblay (1983)*.

latimere by *Schulmeister (2001)*; *Schulmeister (2003)*.

**gonopods* by *Boudinot (2018)*.

Concept. The *gonostyle* and the *volsella*, constitute the *gonostyle/volsella complex*. The *gonostyle* is the outermost structure of the male genitalia that is connected proximally with the *cupula* and medially with the *volsella*. When two clearly separated sclerites are present (the proximal *gonostipes* and the distal *harpe*), the entire structure has been named in different ways, mainly *paramere*, *gonostyle*, *latimere*, and *stipes* (see the list of synonymous terms above). However, when only one sclerite is present, only one term, *gonoforceps*, has been employed. Below we give an extensive explanation of why this has happened and why we propose *gonostyle* as the preferred term. Proximally, the *gonostyle* is elongated into two brace-like apodemes, called the *gonostipital arms* (gsa, Fig. 3B). At the tip of these, another apodeme, called *apex gonostipitis*, allows the insertion of two muscles, the *proximoventral* (gs-pvpv) and the *distoventral gonostyle/volsella complex-penisvalval muscle* (gs-pvdv) (Figs. 3B–3C, 5A; Tables 3–4). Historically a dorsomedial area has been identified, called *parapenis* (see below for an extensive treatment) that functions as a site of origin for the *proximodorsal* (gs-pvpd) and *distodorsal gonostyle/volsella complex-penisvalval muscle* (gs-pvdd) (Figs. 3B–3C, 4A–4C, 6B–6C, 7B; Tables 3–4).

Definition. As defined by the HAO, the *gonostyle* is the anatomical cluster that is composed of sclerites located distally of the *cupula*, dorsoventrally of the *volsella*, and that surrounds the *aedeagus* (Table 2).

Discussion of terminology. At least 12 terms have been introduced to refer to the *gonostyle*. Of these, five (*paramere*, *gonostyle*, *latimere*, *gonoforceps*, *claspers*) are worth discussing as they have been employed more than once after their introduction. The other terms (see above), are rejected as they do not conform to criterion 2.

Among the terms mentioned above, *paramere* has been the most widely used to refer to this cluster of sclerites of the male genitalia. The term has been applied across different insect orders (e.g., Coleoptera), and within Hymenoptera is a renowned case of homonymy (Yoder et al., 2010). Applied for the first time in Hymenoptera by Verhoeff (1832) to refer to the *gonostipes+harpe+volsella*, the concept of *paramere* changed to refer only to the *penisvalva* (Beck, 1933; Peck, 1937b), then to the entire male genitalia (*cupula* excluded) (Wheeler, 1910), then only to the *harpe* (Snodgrass, 1941; Königsmann, 1976), and finally to *gonostipe+harpe* (Snodgrass, 1957). According to this already rampant confusion, Peck (1937a) coined a new term, *gonoforceps*, to refer to those specific cases in which the *harpe* and *gonostipes* are not distinguishable, and only a single continuous sclerite is present. *Gonoforceps* has been introduced specifically in Hymenoptera and has been very successfully applied in Ichneumonidae. Later on, probably for the need to refer generally to a structure that could either be composed of two sclerites or just one, Bohart & Menke (1976) introduced the term *gonostyle* defining it as the “outermost paired appendages of male genitalia, sometimes divided into basistyle and dististyle” (see below for these last two terms).

Schulmeister (2001) first identified the confusion regarding homonyms of the term *paramere* and chose to reject its use. However, at the same time, she also introduced a new term, *latimere*, to identify the anatomical cluster formed by the *gonostipes+harpe*, maintained *gonoforceps* when the two sclerites were not distinguishable, and ignored the term *gonostyle*.

More recently, [Mikó et al. \(2013\)](#) employed the term *gonoforceps* in Ceraphronoidea when only one sclerite was discernible, but when the *harpe* and the *gonostipes* were present (as it is the case for the majority of the species), they employed the term *gonostyle*. On the other hand, [Boudinot \(2013\)](#) used *paramere*, instead of *gonoforceps*, even though a clear distally delimited sclerite (the *harpe*) is not present.

The term *claspers* were applied by [Townes \(1969a\)](#) to Ichneumonidae to refer to a special case in which the *gonostyle* is elongated, forming a rod (Figs. 4A–4C).

Among these many terms and concepts, it is not easy to identify which are preferred. However, some conclusions can be drawn. Following [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#), we reject the term *paramere* based on criterion 1 as it does not best represent the skeletal structure. At the same time, *claspers* should be rejected because, as pointed out by [Tuxen \(1956\)](#), the term is widely used in many different other insect orders to refer to widely different structures, some of which are not homologous to the *gonostyle* (rejected by criterion 4). The term *latimere* also can be rejected as it has not been employed in any other work after [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#) (rejected by criterion 2). Between *gonoforceps* and *gonostyle*, we suggest the latter as it better represents this skeletal area (fulfilling criterion 1). In fact, if the presence of the single sclerite has evolved through a fusion of *gonostipes* and *harpe*, it would be equivalent to the entire *gonostyle*, while if evolution resulted in a loss of the *harpe*, the *gonoforceps* would be equivalent to the *gonostipes* alone ([Schulmeister, 2001](#)). In both cases, the use of *gonoforceps* is unwarranted as there is no need for the employment of different terms to refer to a structure that is equivalent to either one or two sclerites. We certainly understand the “tradition” in using *gonoforceps*, but we also believe that to be able to move Hymenoptera and Ichneumonoidea into the phenomic era and allow future data mining for morphological features, it is better to avoid multiple terms for the same structure or group of sclerites, and employ accurate anatomical concepts ([Girón et al., 2023](#)). In this framework, the original definition of *gonostyle* proposed by [Bohart & Menke \(1976\)](#) allows the employment of the term in multiple cases: (1) when a clear delimitation between *harpe* and *gonostipes* is present; (2) when a clear delimitation between *harpe* and *gonostipes* is absent (the *gonoforceps*; see below for more details); and (3) when a delimitation between *harpe* and *gonostipes* is present, but it is not complete (see [Peck \(1937a\)](#)). Therefore, we strongly encourage the employment of the term *gonostyle* and use it as the preferred label.

As a side note, there is one term that was recently introduced by [Mikó et al. \(2013\)](#): the *gonostyle/volsella complex*. It refers to the anatomical cluster composed of the sclerites located distally of the *cupula* and surrounding the *aedeagus*. We encourage the employment of this term especially when there is no clear delimitation between the *volsella* and the *gonostyles*.

[Peck \(1937a\)](#) and [Peck \(1937b\)](#) introduced the term *gonosquama* within the Ichneumonoidea. However, the term has not been used since then. On the other hand, the term *paramere* has been employed several times. Depending on the authors, *paramere* was used either to refer to the single sclerite that we now identify as a *gonostyle* ([Pratt, 1939](#); [Tobias, 1967](#); [Quicke & van Achterberg, 1990](#); [Belokobylskij, Zaldivar-Riveron & Quicke, 2004](#); [Žikić et al., 2011](#); [Broad, Shaw & Fitton, 2018](#); [Brajković et al., 2010](#)) or to indicate

just its distal part (Alam, 1952; Tremblay, 1979; Tremblay, 1981; Tremblay, 1983). The term *gonoforceps*, even though introduced purposely for Ichneumonidae, has only been used by three authors (Peck, 1937a; Peck, 1937b; Karlsson & Ronquist, 2012; Bennett et al., 2019). Finally, *claspers*, has been widely used in Ichneumonidae (e.g., Townes, 1969a; Townes, 1969b; Townes, 1969c; Townes, 1971), especially when referring to the *gonostyle* of members in the subfamily Mesochorinae (e.g., Dasch, 1971; Lee, 1991).

Preferred term. *Gonostyle*.

Morphological variation in Ichneumonoidea. Among the Ichneumonoidea herein surveyed and analyzed in the literature, the *gonostyle* show a small degree of variability. Within Ichneumonidae, the entire subfamily Mesochorinae (Figs. 4A–4C), and the genera *Lusius* Tosquinet, 1903 (Ichneumoninae) (DDP personal observation, 2023) and *Nematopodius* Granvenhorst, 1829 (Cryptinae) (G. Broad, personal communication, 2023) show a strong reduction of the apical part of the *gonostyle*, while Pratt (1939) found an apical knob in Rhyssinae (Ichneumonidae), hereby confirmed in *Rhyssa persuasoria* (Fig. 6C). Clear *gonostipital arms* are present in *Temelucha* (Fig. 3B), while they are inconsistent in *Mesochorus* (Fig. 4). In some Braconidae subfamilies (e.g., Exothecinae), these sclerites likely underwent a shortening, exposing the *penisvalvae* and part of the *volsella* (Figs. 75, 76 in Quicke & van Achterberg, 1990, p. 61).

III.1. GONOSTIPES (Fig. 9A in Mikó et al., 2013, p. 275)

stipes by Thomson (1872); Zander (1900); Wheeler (1910).

gonostipes by Crampton (1919); Peck (1937a); Peck (1937b); Ross (1937); Ross (1945);

Schulmeister (2001); Schulmeister (2003); Mikó et al. (2013).

pièce principale by Boulangé (1924).

coxopodite by Beck (1933).

basal part of forceps by Snodgrass (1941).

basiparamere by Snodgrass (1941).

lamina parameralis by Snodgrass (1941).

**parameral plate* by Snodgrass (1941).

gonocoxite by Michener (1944); Michener (1956).

basimere by Snodgrass (1941); Boudinot (2013).

basal part of stipes by Birket-Smith (1981).

section 2 by Smith (1969).

basistyle by Bohart & Menke (1976).

**basiparamere* by Brajković et al. (2010).

**gonocoxa* by Boudinot (2013).

Concept. The *gonostipes* is the proximal sclerite composing the *gonostyle*. It has received several names, of which only one was applied to refer to two distinct concepts. When firstly introduced, the term *gonostipes* was applied to the *gonostipes+volsella* (Crampton, 1919), and only subsequently (e.g., Peck, 1937a; Peck, 1937b; Ross, 1937; Ross, 1945; Königsmann, 1976) it was used to refer only to the *gonostipes*, excluding the *volsella*. The latter is the concept that has been more broadly and consistently employed (Schulmeister, 2001).

Definition. As defined by the HAO, the *gonostipes* is the sclerite that is located dorsolaterally on the *gonostyle/volsella* complex, and is connected to the distal margin of the *cupula*, to the proximal margin of the *harpe*, and to the lateral margin of the *volsella* (Table 2).

Discussion of terminology. At least 15 terms have been introduced to refer to the *gonostipes*. Of these, only three (*stipes*, *gonostipes*, *basimere*) are worth discussing as they have been employed more than once after their introduction. All the other terms (see above), are rejected as they do not conform to criterion 2.

The term *stipes* was introduced by Thomson (1872) to an area or sclerite of the male genitalia of *Bombus* Latreille (Hymenoptera: Apidae). As noted by Schulmeister (2001), it is not clear if he referred to the *volsella* (which is extremely reduced in *Bombus*) or to the proximal sclerite of the *gonostyle*. Crampton (1919) replaced Thomson's (1872) *stipes* with *gonostipes* acknowledging the fact that *stipes* is also employed to describe the appendages that bear the maxillary palps (rejected by criterion 4).

Later, Michener (1944), following the periphallallic theory, introduced the term *gonocoxite*, and, in 1957, Snodgrass employed the term *basimere* (firstly introduced by Crampton (1942) in Diptera) to refer to the *gonostipes* (Snodgrass, 1957), a term that was also preferred by Boudinot (2013).

Among these terms, we strongly encourage the use of *gonostipes*, which is not only the oldest (fulfilling criterion 3) but also better represents the skeletal structure (fulfilling criterion 1). In fact, *gonocoxite* has never been employed after Michener (1956), and, as already explained by Schulmeister (2001), the term implies a homology with the coxa, and thus should be avoided (rejected by criteria 2 & 4). The term *basimere* is strongly connected to the concept of *paramere* (being its proximal sclerite), which, as we argued above, should be avoided (see under Discussion of terminology of the *Gonostyle*) (rejected by criteria 1). As already noticed by Schulmeister (2001), *gonostipes* is a very well-known term and even though at its introduction, it was applied to the *gonostipes+volsella* complex, the modern concept of the term refers to the proximal sclerite of the *gonostyle*.

For the above reasons, the term *gonostipes* should be considered the preferred term.

The application of the term *gonostipes* is minimal in Ichneumonoidea due to the lack of a clear delimitation between the proximal sclerite and the *harpe* in most of the taxa. Only two authors used *gonostipes* (Peck, 1937a; Peck, 1937b; Tremblay, 1979; Tremblay, 1981; Tremblay, 1983).

Preferred term. *Gonostipes*.

Morphological variation in Ichneumonoidea. From our dissections, a clear delimitation of sclerites of the *gonostyle* is not evident in Ichneumonidae. However, Peck (1937a) acknowledged the presence of an articulated appendage in few ichneumonids. Further analyses, incorporating Peck's (1937a) taxa, will be required to better understand the issue.

III.1.a. PARAPENIS (prp, Figs. 6B, 7B)

parapenis by Crampton (1919); Boulangé (1924); Schulmeister (2001);

Schulmeister (2003); Boudinot (2013); Koch & Liston (2017).

manubrium by Crampton (1919).

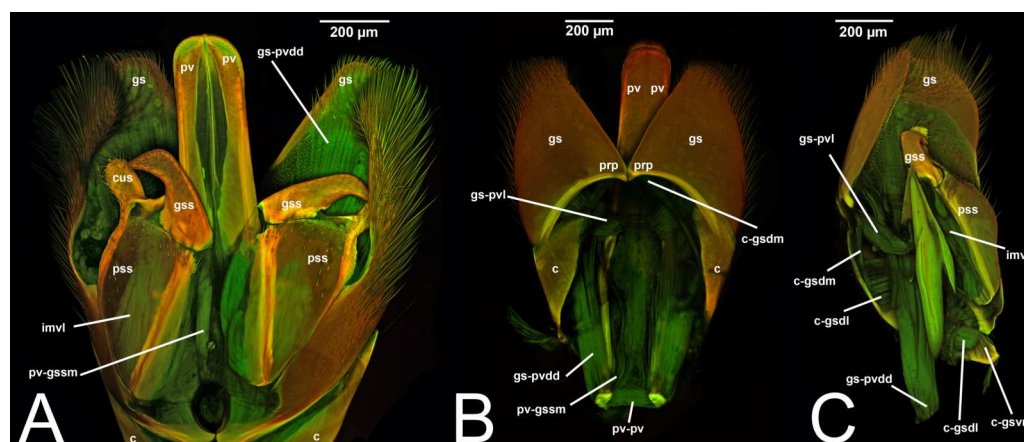


Figure 6 CLSM volume rendered images of male genitalia of *Rhyssa persuasoria* (Ichneumonidae: Rhyssinae). (A) Ventral view. (B) Dorsal view. (C) Median view.

Full-size [DOI: 10.7717/peerj.15874/fig-6](https://doi.org/10.7717/peerj.15874/fig-6)

**praeputium* by [Crampton \(1919\)](#).

**parapenis plate* by [Crampton \(1919\)](#).

parapenial lobe by [Evans \(1950\)](#).

lobi parapenialis by [Priesner \(1966\)](#).

Concept. The *parapenis* is an area of the *gonostipes* located dorsomedially that has been mostly identified in basal Hymenoptera and very rarely in Apocrita (e.g., [Schulmeister, 2003](#)). The area functions as a site of origin for *proximodorsal* (gs-pvpd) and *distodorsal* gonostyle/volsella complex-penisvalval muscles (gs-pvdd) (Tables 3–4). Only one concept (the original) has been applied to this area.

Definition. As defined by the HAO, the *parapenis* is the area that is the dorsomedial part of the *gonostipes* and is the site of origin of the *distodorsal* and *proximodorsal* gonostyle/volsella complex-penisvalval muscles (Table 2).

Discussion of terminology. [Crampton \(1919\)](#) introduced four terms to identify the area of *gonostipes*. Of these, only one, *parapenis*, has been employed more than once after its introduction by [Boulangé \(1924\)](#) and [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#). The other two terms, *parapenial lobe*, and *lobi parapenialis*, have not been employed further (rejected by criterion 2). Overall, the term *parapenis* has not been widely used, probably due to low variation across Apocrita, and to the lack of clear boundaries that delimit it. [Boudinot \(2013\)](#) decided not to use the term in Formicidae due to uncertain homology.

Within Ichneumonoidea, the term has not been employed, and the area has never been identified.

Preferred term. *Parapenis*.

Morphological variation in Ichneumonoidea. According to [Schulmeister \(2003\)](#), the *parapenis* of Ichneumonidae is not set off from the rest of the *gonostipes*. In our observations, this is true for *Temelucha* (Fig. 3B), *Labena grallator* (Fig. 2A), *Mesochorus* (Fig. 4B), and

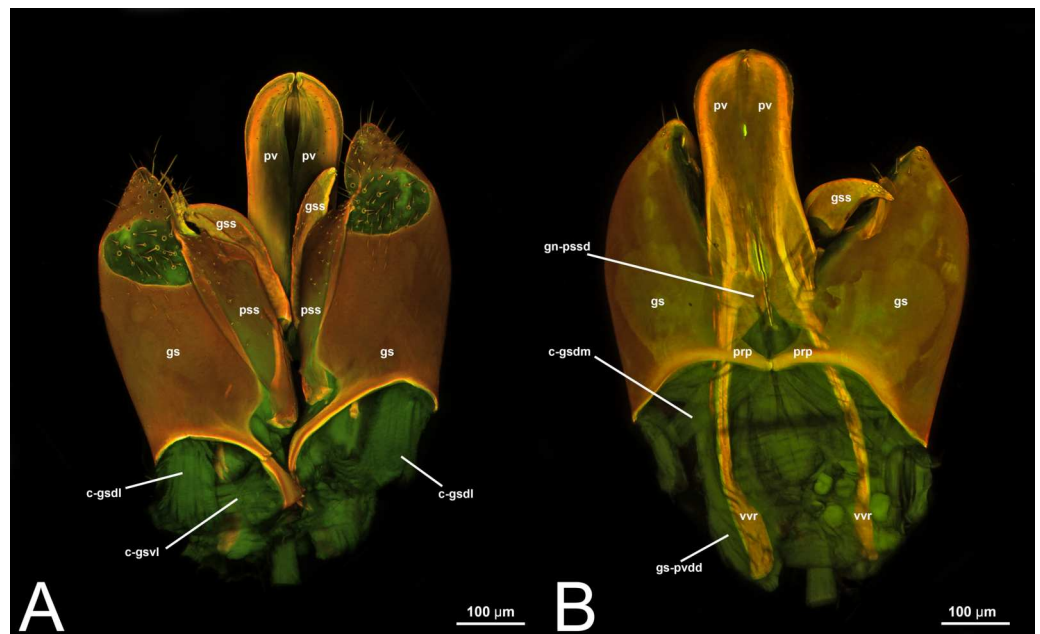


Figure 7 CLSM volume rendered images of male genitalia of *Neoxorides pilosus* (Ichneumonidae: Pomeniinae). (A) Ventral view. (B) Dorsal view.

Full-size [DOI: 10.7717/peerj.15874/fig-7](https://doi.org/10.7717/peerj.15874/fig-7)

Netelia (Fig. 10B), but it is produced in the middle in Pomeniinae (Fig. 7B) and Rhyssinae (Fig. 6B).

III.2. HARPE (Fig. 9A in Mikó et al., 2013, p. 275)

harpe by Crampton (1919); Schulmeister (2001); Schulmeister (2003); Mikó et al. (2013).

distal segment of gonopod by Crampton (1919).

palette by Boulangé (1924).

gonosquama by Peck (1937a); Peck (1937b).

paramere (part.) by Snodgrass (1941); Michener (1944); Alam (1952); Michener (1956); Königsmann (1976).

gonostylus by Michener (1944); Michener (1956).

harpes [sic] by Ross (1945); Wong (1963); Königsmann (1976).

telomere by Snodgrass (1957); Boudinot (2013).

harpago by Snodgrass (1957); Yoshimura & Fisher (2011).

squama by Townes (1957).

disistyle by Bohart & Menke (1976).

harpide by Audouin (1821); Birket-Smith (1981).

*paramero by Tremblay (1979); Tremblay (1981).

*lobo paramerale by Tremblay (1981); Tremblay (1983).

ventral paramere by Gobbi & Azevedo (2016).

*stylus by Boudinot (2018).

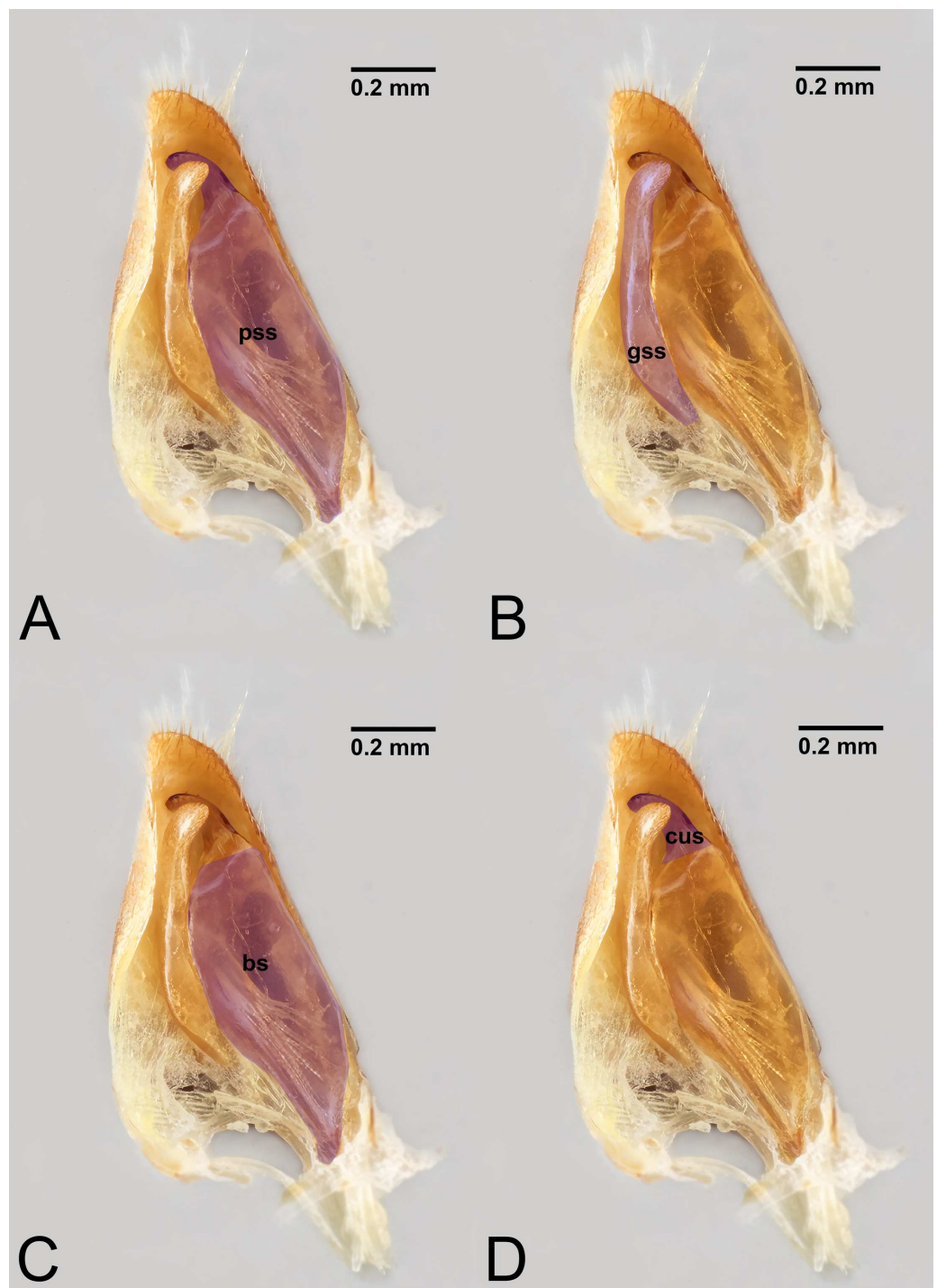


Figure 8 Median view of volsella of *Labena grallator* (Ichneumonidae: Labeninae) with different elements highlighted as follow: (A) Parossiculus (pss). (B) Gonossiculus (gss). (C) Basivolsella (bas). (D) Cuspis (cus).

Full-size [DOI: 10.7717/peerj.15874/fig-8](https://doi.org/10.7717/peerj.15874/fig-8)

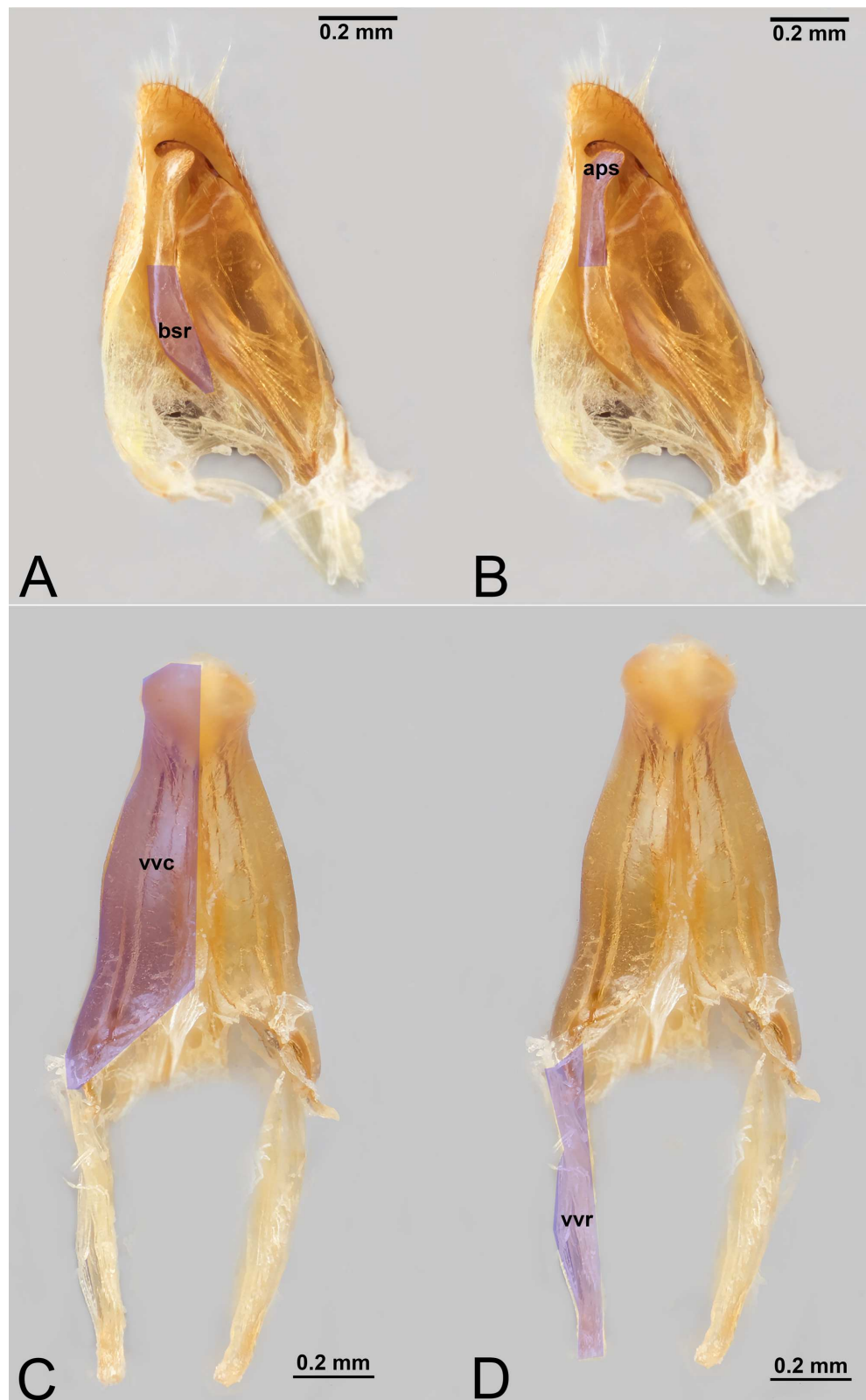


Figure 9 Male genitalia of *Labena grallator* (Ichneumonidae: Labeninae) with different elements highlighted as follow: (A–B) Volsella, median view. (A) Basiura (bsr). (B) Apiceps (aps). (C–D) Penis-valva, ventral view. (C) Valviceps (vvc). (D) Valvura (vvr).

Full-size DOI: [10.7717/peerj.15874/fig-9](https://doi.org/10.7717/peerj.15874/fig-9)

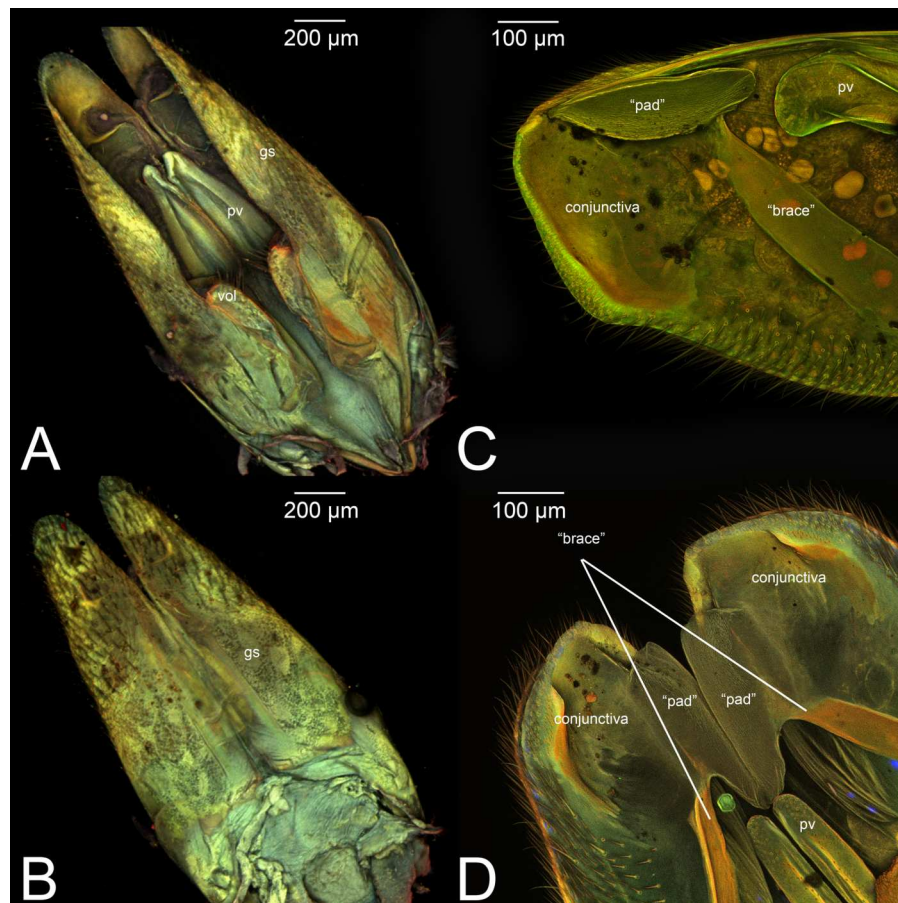


Figure 10 CLSM volume rendered images of male genitalia of *Netelia* sp. (Ichneumonidae: Tryphoninae). (A) Ventral view. (B) Dorsal view. (C) Gonostyle, right apical view. (D) Gonostyle, apical view.

Full-size [DOI: 10.7717/peerj.15874/fig-10](https://doi.org/10.7717/peerj.15874/fig-10)

Concept. The *harpe* is the distally located sclerite of the *gonostyle* and is articulated via muscle with the *gonostipes*. On the *harpe* of Xyelidae, Pamphiliidae, Megalodontesidae, Siricidae, and Xiphydriidae, there is a conjunctiva called the *gonomacula* which seems to function as a suction cup (Schulmeister, 2001). Despite the many different terms applied to this sclerite, there has not been confusion regarding its identification and concept.

Definition. As defined by the HAO, the *harpe* is the sclerite that is located distally on the *gonostyle/volsella* complex and does not connect to the *cupula* nor to the *volsella* by conjunctiva or muscles (Table 2).

Discussion of terminology. At least 16 terms have been introduced to refer to the *harpe*. Of these, only five (*harpe*, *harpago*, *harpide*, *paramere*, *telomere*) are worth discussing as they have been employed more than once after their introduction. All the other terms (see above) are rejected as they do not conform to criterion 2.

The first to name the distal sclerite of the *gonostyle* was Crampton (1919), who introduced the term *harpe*. Subsequently, Peck (1937a) acknowledged the presence of an articulated appendage in a few ichneumonids, which he called *gonosquama*, a term that has not been

used since then (rejected by criterion 2). [Snodgrass \(1941\)](#) decided to use *paramere* to refer only to the *harpe*, while [Ross \(1945\)](#), followed by subsequent authors (e.g., [Wong, 1963](#); [Königsmann, 1976](#)) used the plural of *harpe* (*harpes*) to refer to the singular sclerite (and not to two sclerites). This is unwarranted and should be treated as a misspelling of *harpe*. In 1957, Snodgrass employed the term *telomere* (replacement for *distamire* introduced by [Crampton \(1942\)](#) in Diptera) to refer to the *harpe* ([Snodgrass, 1957](#)), a term that was also preferred by [Boudinot \(2013\)](#). In the same work in which he introduced *telomere*, [Snodgrass \(1957\)](#) introduced another synonym, *harpago*, for the same structure. As far as we know, the latter term has been used only once after its introduction and can be rejected following criterion 2.

An interesting case is the term *harpide*, which was introduced by [Audouin \(1821\)](#) and then employed by [Birket-Smith \(1981\)](#) to replace *harpe*. However, the term, as explained by [Schulmeister \(2001\)](#), was initially introduced by [Audouin \(1821\)](#) to identify a structure (not clear which one as there is no description nor images of it) of the male genitalia of bumblebees. Since bumblebees do not have a distally articulated sclerite, the *harpide* cannot refer to the *harpe*. Therefore, not only is *harpide* a synonym of *harpe* but it has been improperly applied. Of all these terms, we recommend the use of *harpe*, which is the oldest (fulfilling criterion 3) and that best represents the skeletal structure (fulfilling criterion 1). In fact, *paramere*, as we already discussed previously (see under Discussion of terminology of the *gonostyle*), suffers from extensive homonymy, and we strongly discourage its use (see also discussion in [Schulmeister \(2001\)](#)). At the same time, the term *telomere* is connected to the concept of *paramere* (being its distal sclerite), and therefore we also discourage its use.

As far as we know, the term *harpe* has never been used in Ichneumonoidea, and only [Peck \(1937a\)](#) employed the term *gonosquama* within Ichneumonidae (see below for more details).

Preferred term. *Harpe*.

Morphological variation in Ichneumonoidea. From our dissections, a clear delimitation of two individual sclerites is not evident in Ichneumonidae. However, [Peck \(1937a\)](#) acknowledged the presence of an articulated appendage in a few ichneumonids. Further analyses, incorporating [Peck's \(1937a\)](#) taxa, will be required to better understand the issue.

Comments on gonostyle and its associated elements. [Schulmeister \(2001\)](#) considered the absence of a *harpe* as a synapomorphy of the Vespina (Orussidae + Apocrita) and concluded that even when a distally delimited sclerite on the *gonostyle* is present in the Apocrita, it is probably not homologous with the *harpe* because there would be no associated musculature. However, [Mikó et al. \(2013\)](#) found a muscled *harpe* in some Ceraphronoidea and Trigonalioidea (both Apocrita) even though the arrangements of these muscles are different from that of the lower Hymenoptera. Their discovery was not enough to deduce homology of the *harpe* between Apocrita and lower Hymenoptera. [Boudinot \(2013\)](#) also found intrinsic muscles in approximately the same position in Formicidae.

It must be noted that [Boudinot \(2018\)](#) rejected the term *paramere sensu Snodgrass (1957)* because it was explicitly introduced within the phallic theory framework, and he instead employed the term *gonopods* for the same structures following the conclusion from the

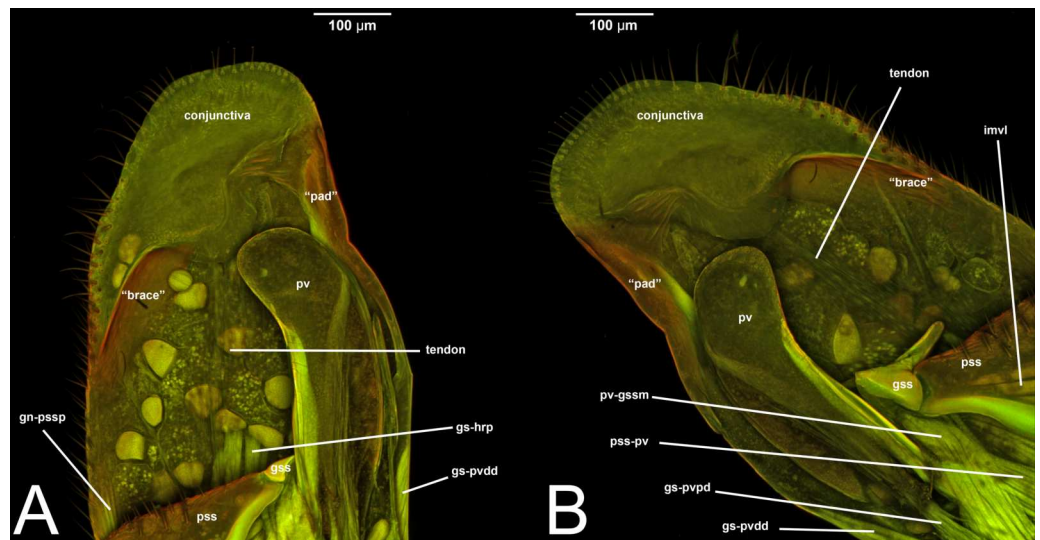


Figure 11 CLSM volume rendered images of male genitalia of *Netelia* sp. (Ichneumonidae: Tryphoninae). (A) Right gonostyle, median view. (B) Left gonostyle, median view.

Full-size [DOI: 10.7717/peerj.15874/fig-11](https://doi.org/10.7717/peerj.15874/fig-11)

coxopod theory. Moreover, according to the same author, the *gonocoxa* (=gonostipes in the phallic theory), which is a fragment of the apical part of the sternum IX, fragmented a second time, forming the *parossiculus* (see below, under Volsella). According to this view, the *parossiculus* is just a secondary fragmentation of the *gonostipes* rather than part of the *volsella*. Boudinot (2018) also homologized the *harpe* with *stylus*, which he considered to have separated from the phallic apparatus (=aedeagus, see under Penisvalvae). This makes the *stylus* a sclerite with a completely different origin than the gonostyle. Boudinot (2018) did not provide any evidence or comments for the formation of a single sclerite.

IV. VOLSELLA

IV. VOLSELLA (vol, Figs. 1C, 3A, 4A, 5A, 6A, 6C, 7A 8A–8B, 9A–9B, 10A, 11)

volsella by Dufour (1841); Peck (1937a); Peck (1937b); Pratt (1939); Michener (1944); Ross (1945); Alam (1952); Michener (1956); Snodgrass (1941); Snodgrass (1957); Scobiola (1963); Tobias (1967); Königsmann (1976); Johnson (1984); Olmi (1984a); Olmi (1984b); Quicke & van Achterberg (1990); Schulmeister (2001); Schulmeister (2003); Konishi (2005); Žikić et al. (2011); Karlsson & Ronquist (2012); Boudinot (2013); Mikó et al. (2013); Broad, Shaw & Fitton (2018); Brajković et al. (2010)

innere Haltzange by Enslin (1918).

tenette by Snodgrass (1941).

ossicle by Ross (1945).

section 3 by Smith (1969).

section 3 of gonocoxite by Schulmeister (2001).

Concept. The *volsella*, together with the *gonostyle*, constitute the *gonostyle/volsella* complex, and it lies between the *gonostipes* and the *penisvalva*. Overall, the *volsella* consists

of two sclerites: (1) the *gonossiculus*, which is located distoventrally (gss, Fig. 8B); and (2) the *parossiculus*, located proximally (pss, Fig. 8A). The *parossiculus* is further divided into a basal and distal area: the *basivolsella* (bs, Fig. 8C) and the *cusps* (cus, Fig. 8D), respectively.

The term *volsella* was introduced by *Dufour (1841)*, who did not clearly define which sclerite he was referring to. Over the years, it was employed for two main concepts, making it another case of homonymy (see the term *paramere*). For some authors (e.g., *Crampton, 1919*; *Birket-Smith, 1981*), the *volsella* referred only to the sclerite connected with the *gonostipes*, and therefore it was identified with the *parossiculus*. For other authors (e.g., *Snodgrass, 1941*; *Michener, 1944*; *Michener, 1956*), the *volsella* referred to the anatomical cluster composed of its proximal and its distal articulated sclerite; therefore, it included both the *parossiculus*+*gonossiculus*. This latter concept was employed more often after the redefinition by *Snodgrass (1941)*.

Definition. As defined by the HAO, the *volsella* is the anatomical cluster composed of the sclerite on the ventral part of the male genitalia that is connected to the *cupula* via muscles (Table 2).

Discussion of terminology. Few terms have been introduced to identify this group of sclerites, and all of them, except *volsella*, have never been used since their introduction. As *Schulmeister (2001)* pointed out, there is no suitable alternative to *volsella sensu Snodgrass (1941)* (see above). Therefore, given the priority (criterion 3) and the wide usage (criterion 2), *volsella* should be considered the preferred term.

Within Ichneumonoidea, many authors employed the term *volsella* to identify the entire anatomical cluster which lies between the *gonostyles* and the *penisvalva* and does not articulate with the *cupula* (*Peck, 1937a*; *Peck, 1937b*; *Alam, 1952*; *Tobias, 1967*; *Konishi, 2005*; *Žikić et al., 2011*; *Karlsson & Ronquist, 2012*; *Bratković et al., 2010*). Only five identified it only with the *parossiculus* (*Pratt, 1939*; *Townes, 1939*; *Tremblay, 1979*; *Tremblay, 1981*; *Tremblay, 1983*; *Quicke & van Achterberg, 1990*; *Belokobylskij, Zaldivar-Riveron & Quicke, 2004*), while the two most recent studies that employed genitalia do not mention the term (*Broad, Shaw & Fitton, 2018*; *Bennett et al., 2019*).

Preferred term. *Volsella*.

Morphological variation in Ichneumonoidea. For the *volsella* we provided a brief account of the variability below each treatment of the sclerites or areas (see below), rather than a general observation of the cluster. This should guide future researchers to further analyses of the different parts.

IV.1. PAROSSICULUS (pss, Figs. 5A, 6A, 6C, 7A, 8A, 11)

parossiculus by *Crampton (1919)*; *Schulmeister (2001)*; *Schulmeister (2003)*;

Mikó & Deans (2009); *Karlsson & Ronquist (2012)*; *Mikó et al. (2013)*.

volsella (part.) by *Wheeler (1910)*; *Crampton (1919)*; *Ross (1937)*; *Pratt (1939)*; *Tremblay (1979)*;

Birket-Smith (1981); *Tremblay (1981)*; *Tremblay (1983)*; *Quicke & van Achterberg (1990)*.

pièce complémentaire by *Boulangé (1924)*.

**cups* by *Tobias (1967)*.

plaques volsellaires by *Dessart & Gärdenfors (1985)*.

Concept. The *parossiculus* is the proximal sclerite of the *volsella* and is composed of two areas. (1) the *basivolsella* (bs, Fig. 8C), located basally; and (2) the *cuspis* (cus, Fig. 8D), which is a distally produced ‘hook’.

Definition. As defined by the HAO, the *parossiculus* is the sclerite that is connected via conjunctiva distomedially to the *gonostipes* and articulates with the *gonossiculus* (Table 2).

Discussion of terminology. The term *parossiculus* was introduced by Crampton (1919) to refer to the proximal sclerite of the *volsella*. Similar to *volsella*, no better alternative to the term *parossiculus* has been proposed and employed. Therefore, given the priority (criterion 3) and the wide usage (criterion 2), *parossiculus* should be considered the preferred term.

Within Ichneumonoidea, the term *parossiculus* has been either wrongly identified with the entire *volsella* (Pratt, 1939; Townes, 1939; Quicke & van Achterberg, 1990; Belokobylskij, Zaldivar-Riveron & Quicke, 2004) or simply not recognized (e.g., Peck, 1937a; Peck, 1937b). To our knowledge, only Karlsson & Ronquist (2012) employed *parossiculus* correctly to define the basal part of the *volsella*.

Preferred term. *Parossiculus*.

Morphological variation in Ichneumonoidea. According to Peck (1937b), the *parossiculus* and the *gonostyle* are indistinguishable in some Cryptinae, Ichneumoninae, Ophioninae, and Tryphoninae, forming, therefore, a clear *gonostyle/volsella complex*. This is true also for the Mesochorinae (Fig. 4), but not for Labeninae (Figs. 8, 9A–9B), and Rhyssinae (Fig. 6A) in which the two are clearly distinguishable. The majority variability relies more on the presence/absence of the *cuspis*, which seems to vary according to the different subfamilies (see under *Cuspis*).

IV.1.a. BASIVOLSELLA (bas, Fig. 8C)

basivolsella by Peck (1937a); Peck (1937b); Ross (1945); Alam (1952); Olmi (1984a); Olmi (1984b); Schulmeister (2001); Schulmeister (2003).

lamina volsellaris by Snodgrass (1941); Johnson (1984); Maetô (1996); Konishi (2005).

**lamina volsellare* by Tremblay (1979); Tremblay (1981); Tremblay (1983).

volsellar plate by Snodgrass (1941); Snodgrass (1957).

Concept. The *basivolsella* is the proximal area of the *parossiculus*. There has not been much confusion regarding the identification of this area, even though its fiat boundaries led many authors (e.g., Karlsson & Ronquist, 2012) to not discuss it in their work.

Definition. As defined by the HAO, the *basivolsella* is the area that is located on the *parossiculus* ventromedially to the *cuspis* (Table 2).

Discussion of terminology. There have been few issues regarding what term to apply to the proximal area of the *parossiculus*. In fact, *basivolsella* was the first term introduced (Peck, 1937a) and, by far, has been the most employed by different authors. Subsequently, Snodgrass (1941) referred to this area with the general term of *volsellar plate* and *lamina volsellaris*, which was not commonly employed afterwards. Therefore, given the priority (criterion 3) and the wide usage (criterion 2), *basivolsella* should be considered the preferred term.

Within Ichneumonidae, the term *basivolsella*, despite being introduced specifically for the superfamily by [Peck \(1937a\)](#), was subsequently applied only by [Peck \(1937b\)](#) and [Alam \(1952\)](#). Other authors either did not recognize the structure (e.g., [Karlsson & Ronquist, 2012](#)) or used the synonym *lamina volsellaris* ([Tremblay, 1979](#); [Tremblay, 1981](#); [Tremblay, 1983](#); [Maetô, 1996](#); [Konishi, 2005](#)).

Preferred term. *Basivolsella*.

Morphological variation in Ichneumonidae. According to [Peck \(1937b\)](#), the *basivolsella* can be used for delimiting subfamilies and various genera, and its dimension can be employed to easily separate several species groups of Ichneumoninae. From our observations, it is clear that the *basivolsellae* in *Labena grallator* and *Rhyssa persuasoria* are notably expanded ([Figs. 6A, 8C](#)), while it seems less so in *Temelucha* sp. ([Fig. 3A](#)). Further studies are needed to assess the accuracy of [Peck \(1937b\)](#).

IV.1.b. CUSPIS (cus, [Figs. 6A, 8D](#))

distivolsella by [Peck \(1937a\)](#); [Peck \(1937b\)](#); [Pratt \(1939\)](#); [Olmi \(1984a\)](#); [Olmi \(1984b\)](#); [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#).

**apical lobe of volsella* by [Townes \(1939\)](#).

cuspis by [Snodgrass \(1941\)](#); [Michener \(1944\)](#); [Alam \(1952\)](#); [Michener \(1956\)](#);

[Snodgrass \(1957\)](#); [Delrio \(1975\)](#); [Tremblay \(1979\)](#); [Birket-Smith \(1981\)](#);

[Tremblay \(1981\)](#); [Tremblay \(1983\)](#); [Maetô \(1996\)](#); [Schulmeister \(2001\)](#);

[Schulmeister \(2003\)](#); [Belokobylskij, Zaldivar-Riveron & Quicke \(2004\)](#); [Konishi \(2005\)](#);

[Žikić et al. \(2011\)](#); [Brajković et al. \(2010\)](#).

cuspis volsellaris by [Snodgrass \(1941\)](#).

**cuspidal processes* by [Tobias \(1967\)](#); [Quicke & van Achterberg \(1990\)](#).

**cuspid* by [Tremblay \(1979\)](#); [Tremblay \(1981\)](#); [Tremblay \(1983\)](#).

**cupides* by [Tremblay \(1979\)](#); [Tremblay \(1981\)](#); [Tremblay \(1983\)](#).

**lobo volsellare* by [Tremblay \(1979\)](#); [Tremblay \(1981\)](#); [Tremblay \(1983\)](#).

Concept. The *cuspis* is the distally located, elongated area of the *parossiculus*. There has been little confusion regarding the identification of this area, even though its arbitrary boundaries led many authors (e.g., [Karlsson & Ronquist, 2012](#)) to not discuss it in their work.

Definition. As defined by the HAO, the *cuspis* is the projection that is located apicolaterally on the *parossiculus* and is adjacent to the *gonossiculus* ([Table 2](#)).

Discussion of terminology. There are two competing terms that could be applied to the distal area of the *parossiculus*, namely the *cuspis* and the *distivolsella*. The latter was introduced by [Peck \(1937a\)](#), and it was employed in four subsequent papers ([Peck, 1937b](#); [Pratt, 1939](#); [Schulmeister, 2001](#); [Schulmeister, 2003](#)). On the other hand, *cuspis* was introduced later on by [Snodgrass \(1941\)](#), and it became more widely used (at least 16 papers).

[Schulmeister \(2001\)](#) noticed that there is no objective reason to prefer either *cuspis* or *distivolsella*. However, she discouraged the use of *distivolsella*, stating that when *basivolsella* and *distivolsella* are used together, they could imply that the two alone (without the

gonossiculus) actually compose the entire *volsella*. This misconception could lead to two consequences: (1) identifying the *volsella* only with the *parossiculus* (without the *gonossiculus*); and (2) identifying the *gonossiculus* as part of the *cuspis*. Therefore, she proposed the employment of *cuspis* to avoid such conceptual misunderstanding, a decision that has also been followed by Mikó *et al.* (2013) and Boudinot (2013).

The reasoning adopted by Schulmeister (2001) seems to fulfill criterion 1 and 2. Therefore, *cuspis* should be considered the preferred term.

Within Ichneumonoidea, the term *cuspis* has been used regularly (Alam, 1952; Tremblay, 1979; Tremblay, 1981; Tremblay, 1983; Maetô, 1996; Belokobylskij, Zaldivar-Riveron & Quicke, 2004; Konishi, 2005; Žikić *et al.*, 2011; Brajković *et al.*, 2010), while *distivolsella* has been used by only two authors (Peck, 1937a; Peck, 1937b; Pratt, 1939). Townes (1939) used the general term apical lobe of the *volsella*, while Quicke & van Achterberg (1990), following Tobias (1967), used *cuspidal processes*, but these terms have never been used outside these works.

Preferred term. *Cuspis*.

Morphological variation in Ichneumonoidea. The presence of the *cuspis* varies within the different subfamilies. In fact, it is clearly present in Labeninae (Fig. 8D), Rhyssinae (Fig. 6) and Xoridinae (Fig. 5A) but is indistinguishable in *Melanichneumon* (Ichneumoninae) (Fig. 1C). Pratt (1939) also found differences among Acaenitini (*cuspis* absent) and Xoridinae (*cuspis* present). We confirmed the presence of a *cuspis* in Xoridinae.

From our dissections, the *cuspis* is absent in *Netelia* (Figs. 11A–11B), but Delrio (1975, p. 9), stated that “*les parties apicoventrales (section 3) sont bifurquées et forment la volsella, avec un digitus médian et une cuspis latérale (=the apicoventral parts (section 3) are bifurcated and form the volsella, with a median digitus and a lateral cuspis)*”, which seems to provide evidence for the presence of a *cuspis* in *Netelia*. However, it is unclear if Delrio refers to the entire *parossiculus*, to its apical region (=the actual *cuspis*), or to the *gonossiculus*, and therefore misidentified the sclerites. Further studies are required to fully understand the variation of this area in *Netelia*.

IV.2. GONOSSICULUS (gss, Figs. 6A, 6C, 7A, 8B)

squama by Thomson (1872).

valva interna by Zander (1900).

lacinia by Wheeler (1910).

gonossiculus by Crampton (1919); Schulmeister (2001); Schulmeister (2003); Mikó et al. (2013).

copulatory ossicle by Crampton (1919).

pièce en trébuchet by Boulangé (1924).

sagitta by Ross (1937); Pratt (1939); Townes (1939).

gonolacinia by Peck (1937a); Peck (1937b).

digitus by Snodgrass (1941); Michener (1944); Alam (1952); Michener (1956); Snodgrass (1957); Delrio (1975); Tremblay (1979); Birket-Smith (1981); Tremblay (1981); Tremblay (1983); Johnson (1984); Maetô (1996); Schulmeister (2001); Schulmeister (2003); Belokobylskij, Zaldivar-Riveron & Quicke (2004); Konishi (2005); Brajković et al. (2010); Žikić et al. (2011); Karlsson & Ronquist (2012).

digitus volsellaris by Snodgrass (1941).

inner paramere by Arnold (1951).

forcipes exteriores by Haupt (1962).

parameres interiores by Priesner (1966).

**lateropenite* by Boudinot (2018).

Concept. The *gonossiculus* is the distal sclerite of the *volsella*, articulated with the *parossiculus*. It consists of two areas: (1) the *basiura* (bsr, Figs. 6A), which is the proximal part of the sclerite and is the site of insertion of the *parossiculo-gonossicula* muscles, and (2) the *apiceps* (aps, Fig. 6B), which is the distal part of the sclerite. Despite the many different terms applied to this sclerite, there has never been confusion regarding its identification.

Definition. As defined by the HAO, the *gonossiculus* is the sclerite that is located on the distoventral part of the *gonostyle/volsella* complex, and is articulated with the more proximal sclerites of the *gonostyle/volsella* complex (Table 2).

Discussion of terminology. At least 14 terms have been introduced to refer to the *gonossiculus*. Of these, four (*gonossiculus*, *sagitta*, *gonolacinia*, *digitus*) are worth discussing as they have been employed more than once after their introduction. All of the other terms (see above) are rejected as they do not conform to criterion 2.

Crampton (1919) introduced the term *gonossiculus* to refer to a distal sclerite articulated with the *parossiculus*, while Ross (1937) introduced the term *sagitta* for the same sclerite. However, Peck (1937a) noticed that this latter term was already in use for the *penisvalva* in *Bombus* (Apidae), and he replaced it with *gonolacinia* (following criterion 4). Later on, Snodgrass (1941) introduced the term *digitus*, which became popular among hymenopterists.

Schulmeister (2001), surveying the terminology, rejected *sagitta*, for the same reasons advanced by Peck (1937a), and *gonolacinia*, since the term had not been employed after Peck (1937a); Peck (1937b) (rejected by criterion 2). However, Schulmeister (2001) could not decide whether to use *gonossiculus* and *digitus*, and she opted to use both of them interchangeably. This latter action, however, did not help build a unified terminology. In

fact, different authors, preferred different terms. For example, [Mikó et al. \(2013\)](#) opted for *gonossiculus*, while [Boudinot \(2013\)](#) chose *digitus*.

Choosing between *gonossiculus* and *digitus* is not easy. If we consider the oldest name, then *gonossiculus* should be the preferred term. Still, if we consider common usage, then the preferred term would be *digitus*. However, it must be noted that *digitus* is also applied to a structure in Lepidoptera, which is not homologous to the *digitus* in Hymenoptera ([Tuxen, 1970](#)), and potentially it has been used in other insect orders as well, even though not reported in the glossary. Therefore, to avoid future confusion, we recommend the use of *gonossiculus* because it is the oldest name (fulfilling criterion 3), and the one that does not correspond to any other structure in other orders (fulfilling criterion 4).

The term *gonossiculus* has never been used in Ichneumonoidea, preferring the synonym *digitus* (e.g., [Konishi, 2005](#); [Karlsson & Ronquist, 2012](#)).

Preferred terms. *Gonossiculus*.

Morphological variation in Ichneumonoidea. The major feature of the *gonossiculus* seems to be its presence or absence. In fact, Ichneumonidae shows variability in the sclerite, which is present and articulated in *Labena grallator* (Labeninae) ([Figs. 8B, 9A–9B](#)), Poemeninae ([Fig. 7](#)), Rhyssinae ([Figs. 6A & 6C](#)), Tryphoninae ([Figs. 11A–11B](#)), and Xoridinae ([Fig. 5A](#)) but absent in *Melanichneumon lissorufus* (Ichneumoninae) ([Figs. 1C](#)) and Mesochorinae ([Fig. 4A](#)). The length of the *gonossiculus* is another important character. [Pratt \(1939\)](#) realized that its length varies according to the different taxa (e.g., shorter in Ichneumoninae and longer in Pimplinae, Labeninae, and some Tryphoninae). This is confirmed in this paper as well: *Labena grallator* (Labeninae) ([Figs. 8B, 9A–9B](#)), *Neoxorides* (Poemeninae) ([Fig. 7](#)), *Rhyssa* (Rhyssinae) ([Figs. 6A, 6C](#)) and *Xorides* (Xoridinae) ([Fig. 5A](#)) have a very elongated *gonossiculus*, while in *Netelia* (Tryphoninae) ([Fig. 11A–11B](#)) and Mesochorinae ([Fig. 4A](#)) the *gonossiculus* is extremely reduced. In Braconidae, the literature is unclear. However, according to [Quicke & van Achterberg \(1990\)](#), there is a range of variability depending on the subfamily.

IV.2.a. BASIURA (bsr, [Fig. 9A](#))

basiura by [Ross \(1945\)](#).

digiura by [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#).

Concept. The *basiura* is the proximal area of the *gonossiculus*. There has never been much confusion regarding the identification of this area, even though its unclear boundaries led many authors to not discuss it in their work (e.g., [Karlsson & Ronquist, 2012](#)).

Definition. As defined by the HAO, the *basiura* is the area that is the proximal part of the *gonossiculus* and corresponds to the insertion of the *medial penisvalvo-gonossicula muscle* ([Table 2](#)).

Discussion of terminology. The term *basiura* was introduced by [Ross \(1945\)](#) to identify the basal portion of the *gonossiculus*. After that, no other terms were introduced for the same area, with the exception of *digiura* by [Schulmeister \(2001\)](#), who did so because she realized that [Ross \(1945\)](#) did not clearly define the limit of this area.

However, due to the paucity of the usage of these two terms (as far as we know, there are no authors who identified the different areas of the *gonossiculus* except for the two above and never outside the sawflies), we can base the decision of the preferred term based solely on the date of introduction (following criterion 3). Therefore, the preferred name should be *basiura*.

The term *basiura* has never been used in Ichneumonoidea.

Preferred terms. *Basiura*.

Morphological variation in Ichneumonoidea. It is extremely difficult to observe the delimitation of different areas of the *gonossiculus* in Ichneumonoidea. Several previous researchers did not discuss or identify the two areas (e.g., [Peck, 1937a](#); [Peck, 1937b](#); [Pratt, 1939](#); [Karlsson & Ronquist, 2012](#)).

IV.2.b. APICEPS (aps, [Fig. 9B](#))

apiceps by [Ross \(1945\)](#).

digiceps by [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#).

Concept. The *apiceps* is the distal area of the *gonossiculus*. There has never been much confusion regarding the identification of this area, even though its unclear boundaries led many authors to not discuss it in their work (e.g., [Karlsson & Ronquist, 2012](#)).

Definition. As defined by the HAO, the *apiceps* is the area that is the distal part of the *gonossiculus* and is connected to the *parossiculus* via membranous conjunctiva ([Table 2](#)).

Discussion of terminology. The term *apiceps* was introduced by [Ross \(1945\)](#) to identify the basal portion of the *gonossiculus*. After that, no other terms were introduced for the same area, with the exception of *digiceps* by [Schulmeister \(2001\)](#), who did so because she realized that [Ross \(1945\)](#) did not clearly define the limit of this area.

However, due to the paucity of the usage of any of these two terms (as far as we know there are no authors who identified the different areas of the *gonossiculus* except for the two above, and never outside the sawflies), we can base the decision of the preferred term based solely on the date of introduction (following criterion 3). Therefore, the preferred name should be *apiceps*.

The term *apiceps* has never been used in Ichneumonoidea.

Preferred terms. *Apiceps*.

Morphological variation in Ichneumonoidea. It is extremely difficult to observe the delimitation of different areas of the *gonossiculus* in Ichneumonoidea. Several previous researchers (e.g., [Peck, 1937a](#); [Peck, 1937b](#); [Pratt, 1939](#); [Karlsson & Ronquist, 2012](#)) did not discuss or identify the two areas.

Comments on volsella and its associated elements. [Vilhelmsen \(1997\)](#) identified the entire *volsella* as a hymenopteran synapomorphy. According to [Boudinot \(2018\)](#), only the *parossiculus* is an actual synapomorphy among Hymenoptera, not the entire *volsella*. This reassessment was based on the fact that only the *parossiculus* bears intrinsic musculature,

absent in other orders. Further studies are required to corroborate Boudinot's (2018) hypothesis.

V. PENISVALVA

V. PENISVALVA (pv, Figs. 1D, 2, 3A–3B, 4, 5, 6A–6B, 7, 9C–9D, 10A, 10C–10D, 11)
thyrses by Audouin (1821).
baguette du fourreau by Dufour (1841).
sagitta by Thomson (1872).
crochet by Seurat (1898).
valve du pénis by Boulangé (1914).
penisvalva by Crampton (1919); Townes (1939); Snodgrass (1941); Michener (1944); Alam (1952); Michener (1956); Snodgrass (1957); Olmi (1984a); Olmi (1984b); Schulmeister (2001); Schulmeister (2003); Žikić et al. (2011); Karlsson & Ronquist (2012); Boudinot (2013); Mikó et al. (2013).
penis rod by Crampton (1919).
penis valve by Crampton (1919); Snodgrass (1941); Ross (1945); Johnson (1984).
valve composing penis by Crampton (1919).
paramere (part.) by Beck (1933); Peck (1937a); Peck (1937b).
lamina aedeagalis by Snodgrass (1941).
aedeagal sclerite by Snodgrass (1941).
aedeagus (part.) by Alam (1952); Peck (1937a); Peck (1937b); Pratt (1939); Snodgrass (1941); Snodgrass (1957); Wahl & Gauld (1998); Konishi (2005); Watanabe & Matsumoto (2010); Žikić et al. (2011); Watanabe, Taniwaki & Kasparyan (2015); Broad, Shaw & Fitton (2018); Bennett et al. (2019); Brajković et al. (2010).
**edeago* by Tremblay (1979); Tremblay (1981); Tremblay (1983).
inner or dorsal arm of the stipes by Arnold (1951).
aedeagal rod by Snodgrass (1957).
thyrsos by Birket-Smith (1981).
**valva* by Brajković et al. (2010).
**penial sclerite* by Boudinot (2018).
**penis* by Boudinot (2018).

Concept. The *penisvalva* is a paired sclerite located in the middle of the external male genitalia. Each of the two *penisvalvae* consists of two areas: (1) the *valviceps* (vvc, Fig. 9C) located distally; and (2) the *valvura* (vvr, Fig. 9D), located proximally. Where the two areas meet, there is the *ergot*, an apodeme which is the site of insertion of the *lateral* (gs-pvl) and *distoventral gonostyle/volsella complex-penisvalval muscle* (gs-pvdv) (Figs. 3B–3C, 4B & 5A–5B), and the *parossiculo-penisvalval muscle* (pss-pv).

The majority of the misconceptions regarding this sclerite are based on two terms: *aedeagus* and *penisvalva* (see below).

Definition. As defined by the HAO, the *penisvalva* is the sclerite that is in the middle of the external male genitalia, and surrounds the distal part of the *ductus ejaculatorius* and the *endophallus* (Table 2).

Discussion of terminology. There has been little confusion regarding the term *penisvalva*. It was introduced by [Crampton \(1919\)](#), referring to the two sclerites in the middle of the male external genitalia, and only [Birket-Smith \(1981\)](#) tried to replace the term with *thyrsos*, based on *thyrses* ([Audouin, 1821](#)). However, this latter term was not applied any further within Hymenoptera, preferring the simpler *penisvalva*. Only [Boudinot \(2018\)](#) decided to use the term *penial sclerite*.

As already mentioned above, the majority of the problems involve the terms *aedeagus* and *penisvalva*. In fact, the former has been used interchangeably with *penisvalva* (e.g., [Brajković et al., 2010](#); [Žikić et al., 2011](#); [Broad, Shaw & Fitton, 2018](#); [Bennett et al., 2019](#)). However, despite being regularly used in recent years, *aedeagus* is correctly applied to the *penisvalvae*+*endophallus*, and not just to one sclerite. Therefore, *penisvalva*, which is restricted to Hymenoptera, is the preferred term over *aedeagus* when referring just to the sclerites, while *aedeagus* is the preferred term when used to refer to both the two *penisvalvae* and the *endophallus*.

Except for [Karlsson & Ronquist \(2012\)](#), all the recent works on the external genitalia in Ichneumonoidea refer to the two *penisvalvae* as the *aedeagus* ([Peck, 1937a](#); [Peck, 1937b](#); [Pratt, 1939](#); [Alam, 1952](#); [Konishi, 2005](#); [Žikić et al., 2011](#); [Broad, Shaw & Fitton, 2018](#); [Bennett et al., 2019](#); [Brajković et al., 2010](#)).

Preferred term. *Penisvalva*.

Morphological variation in Ichneumonoidea. Within Ichneumonoidea, there seems to be variability of the *penisvalva*. In *Labena grallator*, the separation of the *penisvalvae* is not discernible dorsomedially, and the *penisvalvae* are dorsoventrally flattened ([Figs. 2A–2C](#)), preventing, or at least reducing, the ability of the *penisvalvae* to open. This condition also seems to be present in Ichneumoninae ([Fig. 1D](#)), while *Netelia* (Tryphoninae) and *Temelucha* (Cremastinae) appear to have the two *penisvalvae* completely divided ([Fig. 3A, 10A](#)). In the latter taxa, the *penisvalvae* seems not to be separated ventromedially from the gonostyle ([Fig. 3A](#)). The size of the *penisvalva* also varies. In *Netelia* ([Figs. 10C–10D](#)) and *Mesochorus* ([Figs. 4A–4C](#)) the structure is roughly half the length of the gonostyle, while in all the other Ichneumonidae (e.g., [Figs. 3, 6](#)), it is as long as the gonostyle. [Karlsson & Ronquist \(2012\)](#) observed that in both the species of Braconidae studied by them, the *penisvalva* is longer than the *volsella* and the gonostyle.

V.a. VALVURA (vvr, Figs. 7B, 9D)

apophyse péniale by [Boulangé \(1924\)](#).

valvura by [Ross \(1945\)](#); [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#); [Karlsson & Ronquist \(2012\)](#); [Boudinot \(2013\)](#).

aedeagal apodeme by [Snodgrass \(1941\)](#); [Alam \(1952\)](#); [Snodgrass \(1957\)](#); [Konishi \(2005\)](#); [Brajković et al. \(2010\)](#).

apodeme of penis valve by [Michener \(1965\)](#).

**apodema edeagale* by [Tremblay \(1979\)](#); [Tremblay \(1981\)](#); [Tremblay \(1983\)](#).

apodema thyrsos by [Birket-Smith \(1981\)](#).

**penial apodeme* by [Boudinot \(2018\)](#).

Concept. The *valvura* is the proximally located area of the *penisvalva*. Other than the original, no other concepts have been applied to this area.

Definition. As defined by the HAO, the *valvura* is the area that is located proximally to the *ergot* on the *penisvalva* ([Table 2](#)).

Discussion of terminology. Only two terms have been employed by more than one author since their introduction: *valvura* and *aedeagal apodeme*.

The term *valvura* was introduced by [Ross \(1945\)](#) and was applied to the long basal structure of the *penisvalva*. Later on, *aedeagal apodeme* was introduced by [Snodgrass \(1941\)](#) to refer to the same area. However, *aedeagal apodeme* is also used for a non-homologous structure in Siphonaptera ([Tuxen, 1970](#)). Therefore, *valvura* is not only the first term to be introduced (fulfilling criterion 3), but it is also the name that does not correspond to any other structure in other orders (fulfilling criterion 4). *Valvura* should be considered the preferred term.

Except for [Karlsson & Ronquist \(2012\)](#), who applied the term *valvura* to the basal part of the *penisvalva*, the other authors either did not mention the term or they used *aedeagal apodeme* ([Alam, 1952](#); [Konishi, 2005](#); [Brajković et al., 2010](#)).

Preferred term. *Valvura*.

Morphological variation in Ichneumonoidea. The overall morphology of the *valvura* in Ichneumonoidea, seems to vary according to certain taxa. [Peck \(1937a\)](#) observed that the *valvura* of some Anomaloninae (*Agrypon flaveolatum* (Gravenhorst, 1807) and *Therion circumflexum* (Linnaeus, 1758)) is visibly medio-laterally expanded (Figs. 129, 141 in [Peck, 1937a](#), p. 251), as it is in many Banchinae ([Pratt, 1939](#)). Also, the shape of the *ergot* varies significantly, being extremely produced in Pimplinae (e.g., *Dolichomitus tuberculatus* (Geoffrey, 1875)) (Figs. 147, 141 in [Peck, 1937a](#), p. 252) but extremely reduced in *Megharyssa macrura* (Figs. 134, 141 in [Peck, 1937a](#), p. 251). In *Temelucha* (Cremastinae), there is also a well-defined, pointed *ergot* (erg, [Fig. 3C](#)).

V.b. VALVICEPS (vvc, Fig. 9C)

valviceps by Ross (1945); Schulmeister (2001); Schulmeister (2003).

**penisvalva* (part.) by Konishi (2005).

**distal portion* by Boudinot (2018).

Concept. The *valviceps* is the proximally located area of the *penisvalva*. Other than the original, no other concepts have been applied to this area.

Definition. As defined by the HAO, the *valviceps* is the area that is the distal part of the *penisvalva*, distally of the *ergot* (Table 2).

Discussion of terminology. The term *valviceps* was introduced by Ross (1945) and was applied to the apical part of the *penisvalva*. No other term has been introduced to refer to this area, with the exception of Konishi (2005), who applied the term *penisvalva* for this area (instead of referring to the entire sclerite). This makes *penisvalva* a case of homonymy.

The term *valviceps* has never been used in Ichneumonoidea.

Preferred term. *Valviceps*.

Morphological variation in Ichneumonoidea. Very little information is available for the *valviceps* in Ichneumonoidea. From what can be observed in Peck's (1937a, p. 251–252) drawings, the majority of the variability for this area relies on the overall curvature of the part. For instance, *Virgichneumon maculicauda* (Perkins, 1953) (Ichneumoninae), in lateral view, has a distinct sinuosity of the *valviceps*, possibly functioning as muscle attachments (Fig. 135 in Peck, 1937a, p. 251). More exploration is needed.

Comments on penisvalva and its associated elements. Boudinot (2018) proposed the term *penial sclerite* (for one sclerite) and *penis* (for both sclerites), believing that the *penisvalva* of the Hexapoda derived from the medially fused primary gonopods (gonopore-bearing limbs).

VI. MEDIAN SCLEROTIZED STYLE

VI. MEDIAN SCLEROTIZED STYLE (Figs. 8A, 9A in Schulmeister, 2003, p. 261–262)

median sclerotized style by Ross (1937); Schulmeister (2001); Schulmeister (2003).

ventral rod of aedeagus by Snodgrass (1941).

detached rhachies by Smith (1969); Smith (1970a); Smith (1970b).

Concept. The *median sclerotized style* is a long, thin sclerite located between the two *penisvalvae*. One muscle, the *penisvalvo-median sclerotized style muscle*, connects the *valvura* to the *median sclerotized style*.

Definition. As defined by the HAO, the *median sclerotized style* is the sclerite that is located ventrally between the *penisvalvae* (Table 2).

Discussion of terminology. The term *median sclerotized style* was introduced by Ross (1937) to identify a sclerite present in Siricidae and Cephidae. Later on, Smith (1969) introduced *detached rhachies* for the same structure, while Snodgrass (1941) used the term

ventral rod of aedeagus. This latter term has not been used after its introduction and can be rejected following criterion 2. There are no major differences in the use of *median sclerotized style* and *detached rhachies* as both were employed in three papers. Therefore the preferred between the two is *median sclerotized style* because it was the first to be introduced (fulfilling criterion 3).

The term *median sclerotized style* has never been used in Ichneumonoidea.

Preferred term. *Median sclerotized style*.

Morphological variation in Ichneumonoidea. A sclerite located between the two *penisvalvae* has never been located in Ichneumonoidea (e.g., [Schulmeister, 2003](#)).

Comments on the median sclerotized style. According to [Schulmeister \(2001\)](#), the *median sclerotized style* has been recorded only in Cephidae and Siricidae, and is located in the same place where the *phallotrema* is located in other hymenopteran taxa (e.g., Ichneumonoidea). According to [Smith \(1970a\)](#), the *median sclerotized style* is a sclerite originally separated from the *penisvalvae*.

In Apoidea, other types of sclerotization of the membrane are present, but are not homologous to the *median sclerotized style* ([Schulmeister, 2001](#)). One example is the *spatha* which is an unpaired sclerite located dorsally of the basal section of the *aedeagus* of some Aculeata.

VII. FIBULA DUCTI

VII. FIBULA DUCTI (Fig. 3B in [Schulmeister, 2003](#), p. 256)

**sperrkeil* by [Clausen \(1938\)](#).

**wedge* by [Forbes & Do-Van-Quy \(1965\)](#).

fibula ducti by [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#).

**wedge sclerite* by [Boudinot \(2013\)](#).

Concept. The *fibula ducti* is a small sclerite located in the proximal section of the *ductus ejaculatorius*. It is considered part of the internal male genitalia.

Definition. As defined by the HAO, the *fibula ducti* is the sclerite that is located in the proximal end of the unpaired part of the *ductus ejaculatorius* ([Table 2](#)).

Discussion of terminology. The first author to identify a sclerotized area on the *ductus ejaculatorius* was [Clausen \(1938\)](#) who coined the term *sperrkeil* in *Formica rufa* Linnaeus, 1761 (Formicidae). Later on, [Forbes & Do-Van-Quy \(1965\)](#) identified a similar structure in *Neavamymex* Borgmeier, 1940 (Formicidae), referring to it with the general term *wedge*, while ([Smith, 1990](#), p. 46) depicted a sclerite in the same position in Pergidae without naming it. [Schulmeister \(2001\)](#) acknowledged [Smith's \(1990\)](#) image, and decided to name the sclerite *fibula ducti*. More recently, [Boudinot \(2013\)](#), while referring to the *wedge* of [Forbes & Do-Van-Quy \(1965\)](#), named the same structure *wedge sclerite*.

Among these terms, only one best fulfills criteria 2 and 4, and therefore should be considered the preferred term: *fibula ducti*. In fact, the others have not been used after the

original introduction (e.g., *sperrkeil*) or generally refer to the shape of the sclerite rather than provide a specific term for it (e.g., *wedge*).

Preferred term. *Fibula ducti*.

Morphological variation in Ichneumonoidea. A sclerite located proximally on the *ductus ejaculatorius* has never been located in Ichneumonoidea (e.g., [Schulmeister, 2003](#)).

Comments on the fibula ducti. According to [Schulmeister \(2001\)](#), more investigation is needed to understand if the *sperrkeil* of [Clausen \(1938\)](#) is homologous to the *fibula ducti*. However, the similar location (proximal to the *ductus ejaculatorius*) and the similar shape are good indicators that the two are very likely the same structure and thus are synonymized here.

So far, the *fibula ducti* was identified only in three families of Hymenoptera: Formicidae ([Clausen, 1938](#); [Forbes & Do-Van-Quy, 1965](#); [Boudinot, 2013](#)), Pergidae ([Smith, 1990](#); [Schulmeister, 2001](#)), and Argidae ([Schulmeister, 2003](#)).

The *Netelia* case

As mentioned above, the external male genitalia of Ichneumonoidea have been very little explored. The exception is the genus, *Netelia* Gray, belonging to the subfamily Tryphoninae (Ichneumonidae). With more than 330 valid species worldwide ([Yu, van Achterberg & Horstmann, 2016](#)), the genus *Netelia* has a long tradition of male genitalia description, so much that some terms have been coined and used only within the genus.

Pad and *brace* refer to structures that are difficult to interpret ([Figs. 10C–10D, 11](#)). Both are located in the inner part of the *gonostyle* and are continuous with each other ([Fig. 10D](#)). [Townes \(1939\)](#) defined *brace* as a sclerome located basally to the *pad*, which is a “*flap or vesicle that sticks out*”. Since then, these two terms have been employed and are still in use (e.g., [Broad, 2012](#); [Konishi, Chen & Pham, 2022](#)). It is very difficult to understand what those two structures are homologous with since these two terms have not been utilized in other insect groups ([Tuxen, 1956](#); [Tuxen, 1970](#)), but we draw some possible conclusions.

The two *pads* are located medially, between the two *gonostyles* and distal to the two *penisvalvae*. They are connected between each other along their proximo-medial internal margin, and are in continuity with a conjunctiva for the entire length of their external margins. This conjunctiva covers the internal distal portion of the *gonostyle*, and is delimited proximally by the *brace*, a more sclerotized area that runs proximo-distally and that is in connection with the *pad* in its distal part ([Figs. 10C–10D](#)). Being composed of conjunctiva, the two *pads* are flexible, and once they are disconnected from each other, they fold towards the internal part of the *gonostyle* ([Figs. 10C, 11A–11B](#)). The proximal section of this entire structure (*brace* + *conjunctiva* + *pad*) is the site of insertion of the tendon belonging to a muscle that inserts into the proximal margin of the *gonostyle* ([Fig. 11](#)).

Within Hymenoptera, only one muscle shares the same insertion of this muscle as in *Netelia*: the *proximal gonostyle/volsella complex-harpal muscle* (muscle t’ of [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#)). This muscle, together with the *distal gonostyle/volsella complex-harpal muscle* (muscle t’’ of [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#)), is tightly

linked to the presence of the *harpe* (Schulmeister, 2003). In Ichneumonoidea, the *harpe* is not discernible as a distinct sclerite, nor is it in *Netelia*, and thus, the *proximal gonostyle/volsella complex-harpal muscle* should be absent. However, their presence (even though not recognized as two different muscles) was discovered in *Megarhyssa lunator* by Peck (1937a), who associated them with the presence of a *harpe* that was only partially distinct (“incompletely fused” for the author). Mikó et al. (2013) also reported the same muscle in Ceraphronoidea without a *harpe* and proposed a change of muscle function for the *proximal gonostyle/volsella complex-harpal muscle* that went from moving the *harpe* to laterally bending the *gonostyle*. In *Netelia* the *proximal gonostyle/volsella complex-harpal muscles* (gs-hrp, Fig. 11A) could also have changed its function, and it is used to bend the entire complex formed by the *brace*, the *pad* and the rest of the conjunctiva.

Regarding the conjunctiva, it is not clear what it might be homologous with. Schulmeister (2001), reported the presence of the conjunctiva located on the distal portion of the *harpe* in certain families of basal Hymenoptera. The structure was firstly called *gonomacula* by Crampton (1919) and several authors advanced the hypothesis that the *gonomacula* could serve as a suction cup to hold the female during copulation, due to the presence of a muscle, the *harpo-gonomaculal muscle* (ha-gon, Table 3). Even though the conjunctiva and the pad in *Netelia* share the same composition (conjunctiva) and a similar position of the *gonomacula*, we refrain from treating them as homologous because a muscle in the same position of the *harpo-gonomaculal muscle* has not been retrieved.

However, it is possible that the conjunctiva in *Netelia* evolved independently from the *gonomacula* in basal Hymenoptera to perform a similar function. In fact, the *penisvalva* in *Netelia* is extremely short, barely reaching half of the length of the *gonostyle*. Therefore, the conjunctiva could help in holding the female during copulation (functioning as a suction cup), while the *pad* helps the transfer of the sperm (functioning as a receptacle) to the female genital organs. Further comparative analyses including members of the subfamily Tryphoninae and a broader representation of Ichneumonidae will be needed to fully understand the function and identity of these structures. Investigations on the internal copulatory organ of females will likely be informative.

Musculature of Ichneumonoidea male genitalia

Musculature is essential for a comparative framework to understand relations among skeletal features (e.g., Vilhelmsen, 2010; Mikó et al., 2012). Even though a thorough study of the musculature of the male genitalia in Ichneumonoidea is beyond the scope of the current study, we are providing the following resources to facilitate future researchers in this field: (1) definitions and abbreviations of the different muscles according to an ontological framework (Table 3); (2) an alignment of the terminology for the musculature of the entire order Hymenoptera (Table 4); (3) a summary of the observations and analyses of the musculature of Ichneumonoidea (Table 5).

The musculature of the Ichneumonoidea male genitalia remains mostly unexplored, with only a handful of authors studying a very limited number of species. The first to fully analyze the muscles and their possible functions in Ichneumonoidea were Peck (1937a) for *Megarhyssa lunator* (Rhyssinae), and Alam (1952) for *Stenobracon deesae* (Braconinae).

Table 5 Muscle presence or absence per family within Ichneumonoidea, according to different authors. 0, absent; 1, present; –, missing data.

Abbreviation	Muscle	Ichneumonidae (Schulmeister, 2003)	Ichneumonidae (Peck, 1937b)	Braconidae (Alam, 1952)	Confirmed in this paper
S9-cm	Medial S9-cupulal muscle	0	1	1	1
S9-cml	Mediolateral S9-cupulal muscle	1	1	1	1
S9-cl	Lateral S9-cupulal muscle	1	1	1	1
c-gsvm	Ventromedial cupulo-gonostyle/volsella complex muscle	1	1	1	1
c-gsvl	Ventrolateral cupulo-gonostyle/volsella complex muscle	1	1	1	1
c-gsdl	Dorsolateral cupulo-gonostyle/volsella complex	1	1	–	1
c-gsdm	Dorsomedial cupulo-gonostyle/volsella complex muscle	1	1	1	1
gs-pvpv	Proximoventral gonostyle/volsella complex - penisvalval muscle	1	1	1	1
gs-pvdv	Distoventral gonostyle/volsella complex - penisvalval muscle	1	1	1	1
gs-pvdd	Distodorsal gonostyle/volsella complex - penisvalval muscles	1	1	1	1
gs-pvpd	Proximodorsal gonostyle/volsella complex - penisvalval muscle	1	1	–	1
gs-pvl	Lateral gonostyle/volsella complex-penisvalval muscle	0	1	1	1
pv-gssl	Lateral penisvalvo-gonossicula muscle	0	1	–	1
pv-gssm	Medial penisvalvo-gonossicula muscle	0	1	1	1
pv-ph	Penisvalvo-phallotremal muscle	0	1	1	1
gss-ph	Gonossiculo-phallotremal muscle	0	1	1	–
pss-ph	Parossiculo-phallotremal muscle	1	1	1	–
gs-pss	Gonostyle/volsella complex-parossicula muscle	1	1	1	1
gn-pssp	Proximal gonostipo-parossicula muscle	–	–	1	1
gn-pssd	Distal gonostipo-parossicula muscle	–	–	–	1
imvll	Lateral gonostyle/volsella complex-volsella muscle	1	1	1	1
imvl	Median gonostyle/volsella complex-volsella muscle	1	1	1	1
imvm	Gonostyle/volsella complex-gonossiculus muscle	0	1	1	1
pss-pv	Parossiculo-penisvalval muscle	1	–	–	1

(continued on next page)

Table 5 (continued)

Abbreviation	Muscle	Ichneumonidae (Schulmeister, 2003)	Ichneumonidae (Peck, 1937b)	Braconidae (Alam, 1952)	Confirmed in this paper
gs-hrd	Distal gonostyle/volsella complex-harpal muscle	0	1	—	0
gs-hrp	Proximal gonostyle/volsella complex-harpal muscle	0	1	—	1 (only in <i>Netelia</i>)
gs-hra	Apical gonostyle/volsella complex - harpal muscles	0	0	—	0
ha-gon	harpo-gonomaculal muscle	0	0	—	0
gs-gs	intragonostyle muscle	0	—	—	—
pv-pv	interpenisvalval muscle	1	—	—	1
vl-vl	intervolsellal muscle	—	—	—	—
pv-mss	penisvalvo-median sclerotized style muscle	0	—	—	0

While [Snodgrass \(1941\)](#) also provided some comments on the musculature of the male genitalia, his assessment was not comprehensive and focused only on the muscles of the *volsella*. The most recent study on the musculature for the superfamily was provided by [Schulmeister \(2003\)](#), who studied two specimens, generally referred to as “Ichneumonidae sp.1” and “Ichneumonidae sp.2.” By comparing the results of these studies ([Table 5](#)), we can see some inconsistencies. [Peck \(1937a\)](#) identified 20 muscles, [Alam \(1952\)](#) only 18, and [Schulmeister \(2003\)](#) only 16. It is unclear if this difference is due to variability within and across the families or simply a different interpretation of muscles between authors. In the current studies we have found 24 muscles, confirming all the muscles already reported by previous authors.

There is inconsistency in the presence or absence of certain muscles (see [Table 5](#)). According to [Schulmeister \(2003\)](#), only two of the three muscles connecting the *cupula* to the *abdominal sternum 9* are present in Ichneumonidae, namely the *mediolateral* and the *lateral S9-cupulal muscles* (muscle b and c of [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#)) ([Table 4](#)), but [Peck \(1937a\)](#) claimed to have retrieved the *medial S9-cupulal muscle* (muscle a of [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#)) in the subfamily Rhyssinae, and [Alam \(1952\)](#) retrieved the same muscles in Braconidae. This is unusual, as [Schulmeister \(2003\)](#) did not observe the muscle within the Apocrita, only in the basal Hymenoptera. In this paper, we were able to retrieve the *medial S9-cupulal muscle* (S9-cm) in Cremastinae ([Fig. 3A](#)) and Mesochorinae ([Figs. 4A, 4C](#)), confirming [Peck's \(1937a\)](#) observations. Moreover, the position of this muscle is very similar to the one reported by [Alam \(1952, p. 5\)](#) running almost parallel by inserting laterally on the *abdominal sternum 9* and laterally on the *cupula*.

An interesting muscle recorded in Ichneumonidae only by [Schulmeister \(2003\)](#) is the *interpenisvalval muscle* (muscle x in [Schulmeister \(2001\)](#); [Schulmeister \(2003\)](#)) which was first described by [Boulangé \(1924\)](#) in basal Hymenoptera (*Abia lonicerae* (Linnaeus, 1758), Cimbicidae). [Schulmeister \(2003\)](#) disproved the presence of the muscle in the genus *Abia* but pointed out that only Ichneumonidae (across the Hymenoptera surveyed) bear the *interpenisvalval muscle*. It is a single muscle running transversely, connecting the two *penisvalvae*, and opening and closing them during copulation. We corroborated the

presence of this muscle within the Ichneumonidae, reporting it within the genus *Temelucha* (Ichneumonidae, Cremastinae) (Fig. 3B), and in the species *Rhyssa persuasoria* (Rhyssinae) (Fig. 6B). This is also the first time that this muscle has been imaged. The position of the *interpenisvalval muscle*, however, seems to vary between the two subfamilies. In fact, in Rhyssinae it is located proximally, connecting the two *valvurae* (as reported also by Schulmeister (2001)), while in Cremastinae, is located more apically, connecting the two *valviceps*. The latter position has not been previously reported. A possible explanation for this variation is based on the correlation of the *interpenisvalval muscle* with the level of division of the *penisvalvae*. In Cremastinae, the conjunctiva that connects the two sclerites disappears proximally, allowing only a distal opening of the two *penisvalvae*. In Rhyssinae, the conjunctiva seems to be present across the entire length of the *penisvalvae*, and by being located proximally, the *interpenisvalval muscle*, allows a far greater range of opening of the two sclerites. Future studies are required to assess if this muscle is relegated to Ichneumonidae (and to what extent) or present also in Braconidae and other hymenopteran families.

Another confirmation of a muscle within Ichneumonidae is the *proxima gonostyle/volsella complex-harpal muscle* (gs-hrp, Fig. 11A), reported here in *Netelia* (Tryphoninae) for the first time, and only reported within Ichneumonidae by Peck (1937b). For a complete treatment, please read above, under “The *Netelia* case.”

Alam (1952) described and imaged a muscle that he named “17” (see Fig. 3A in Alam, 1952, p. 625), running antero-laterally from the “median plate” of the *penisvalva* to the *valvura*. The author believed the muscle to be an adductor of the *valvura*. Alam’s (1952) “median plate” is described as a dorsal extension of the wall surrounding the *aedagus*, which is connected via conjunctiva to the *gonostyle*. As far as we know, “median plate” was employed only by Alam (1952), and muscle “17” was not discussed by subsequent authors. In our interpretation, the “median plate” is either the *ergot* or the entire *valviceps* that in Ichneumonoidea is laterally expanded and proximally produced, while muscle “17”, could be related to a similar muscle already identified by Schulmeister (2003) in Hymenoptera, muscle “nb”. This muscle runs from the *valvura* to the *phallotrema* and Schulmeister (2003) interpreted it as a subdivision, occurring in certain taxa, of muscle “n” which, when undivided, runs from the *valvura* to the *basiura*. In our survey, we discovered a paired muscle in *Temelucha* sp. (Cremastinae) that originates from the *valvura* and attaches to a membranous area adjacent to the *ergot*. Alam (1952) possibly misinterpreted the insertion of his muscle “17”, which actually runs from the *valvura* to the *phallotrema*, and we treat muscle “17” as the *penisvalvo-phallotremal muscle* (pv-ph, Figs. 3B–3C). In Rhyssinae, this muscle is undivided, and only the *medial penisvalvo-gonossicula muscle* (muscle n of Schulmeister (2001)) is present (pv-gssm, Fig. 6B).

Alam (1952) interpreted muscles “4” and “5” as two different muscles, when they have the same insertion but slightly different attachment (two different, but close, locations, typical of a fan-shaped muscle) and therefore can be considered two bundles of the same muscle (see Fig. 6 in Alam, 1952, p. 627).

CONCLUSION

From the current study, it is clear that the male genitalia in Ichneumonoidea has suffered from terminological inconsistencies that have prevented the exploration of characters in taxonomic studies and establishing correct homology statements that are needed to conduct evolutionary studies. In this regard, the most problematic group of sclerites is the *gonostyle-volsella complex* which has generated several synonyms. Reasons for these synonyms include its overall complexity (e.g., several parts organized differently according to the taxon) and the reduced number of comparative anatomy studies across Hymenoptera (as recently discussed also by [Lanes et al. \(2020\)](#) for Bethyridae) that have led different authors to propose new names for the same parts.

Implementation of a larger sample size is required for a more comprehensive anatomical study to advance our understanding of the skeleto-musculature of male genitalia in Ichneumonoidea. This is important for resolving some of the inconsistencies found in the musculature and laying the foundation for testing the phylogenetic signal of male genitalia. The hope is that the tools provided in this contribution will guide and foster further studies on the sclerites and musculature of ichneumonoid male genitalia.

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Author Contributions

- Davide Dal Pos conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- István Mikó conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Elijah J. Talamas analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
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REFERENCES

- Adler PN. 1975.** Gene expression and morphogenesis during the deposition of *Drosophila* wing cuticle. *Fly* **11**:194–199.
- Aibekova L, Boudinot BE, Beutel RG, Richter A, Keller R, Hita-Garcia F, Economo EP. 2022.** The skeletomuscular system of the mesosoma of *Formica rufa* workers (Hymenoptera: Formicidae). *Insect Systematics and Diversity* **6**(2):1–26 DOI [10.1093/isd/ixac002](https://doi.org/10.1093/isd/ixac002).
- Alam SM. 1952.** Studies on “skeleto-muscular mechanism” of the male genitalia in *Stenobracon deesae* Cam. (Hymenoptera: Braconidae). *Beiträge Zur Entomologie* **2**:620–634.
- Andena SR, Noll FB, Carpenter JM, Zucchi R. 2007.** Phylogenetic analysis of the neotropical *Pseudopolybia* de Saussure, 1863, with description of the male genitalia of *Pseudopolybia vespiceps* (Hymenoptera: Vespidae, Epiponini). *American Museum Novitates* **3586**:1–11 DOI [10.1206/0003-0082\(2007\)3586\[1:PAOTNP\]2.0.CO;2](https://doi.org/10.1206/0003-0082(2007)3586[1:PAOTNP]2.0.CO;2).
- André E. 1879.** *Species des Hymenopteres d'Europe et d'Algerie*. Beaune: Hachette Livre, 1–11.

- Arnold G. 1951.** Sphecidae and Pompilidae (Hymenoptera) collected by Mr. K. M. Guichard in West Africa and Ethiopia 1941–48. *Bulletin of the British Museum (Natural History) Entomology* 2:95–183 DOI [10.5962/bhl.part.27752](https://doi.org/10.5962/bhl.part.27752).
- Audouin M. 1821.** Observations sur les organes copulateurs mâles des Bourdons. Rapport lit par Latreille en lundi 9 avril. *Annales Générales des Sciences Physiques* 8:285–289.
- Beck DE. 1933.** A morphological study of the male genitalia of various genera of bees. PhD thesis, Iowa State College, Ames.
- Belokobylskij SA, Zaldivar-Riveron A, Quicke DLJ. 2004.** Phylogeny of the genera of the parasitic wasps subfamily Doryctinae (Hymenoptera: Braconidae) based on morphological evidence. *Zoological Journal of the Linnean Society* 142:369–404 DOI [10.1111/j.1096-3642.2004.00133.x](https://doi.org/10.1111/j.1096-3642.2004.00133.x).
- Bennett AMR. 2015.** Revision of the world genera of Tryphoninae (Hymenoptera: Ichneumonidae). *Memoirs of the American Entomological Institute* 86:1–385.
- Bennett AMR, Cardinal S, Gauld ID, Wahl DB. 2019.** Phylogeny of the subfamilies of Ichneumonidae (Hymenoptera). *Journal of Hymenoptera Research* 71:1–156 DOI [10.3897/jhr.71.32375](https://doi.org/10.3897/jhr.71.32375).
- Berlese A. 1909.** *Gli insetti: loro organizzazione, sviluppo, abitudini e rapporti con l'uomo. Volume primo embriologia e morfologia.* Milano: Società Editrice libraria, 1044.
- Birket-Smith S. 1981.** The male genitalia of Hymenoptera—a review based on morphology in Dorylidae (Formicoidea). *Entomologica Scandinavica Supplement* 15:377–397.
- Bohart RM, Menke AS. 1976.** *Sphecids wasps of the world: a generic revision.* Berkeley: University California Press, 695.
- Bollino M, Uliana M, Sabatinelli G. 2018.** The *Pygopleurus* Motschulsky (Coleoptera: Scarabaeoidea: Glaphyridae) of mainland Greece: taxonomy, faunistics and ecological notes. *Zootaxa* 4674(2):151–202 DOI [10.11646/zootaxa.4674.2.1](https://doi.org/10.11646/zootaxa.4674.2.1).
- Bordas L. 1893.** Sur l'appareil génital mâle des Hyménoptères. *Annales des Sciences Naturelles* 20:103–184.
- Boudinot BE. 2013.** The male genitalia of ants: musculature, homology, and functional morphology (Hymenoptera, Aculeata, Formicidae). *Journal of Hymenoptera Research* 30:29–49 DOI [10.3897/jhr.30.3535](https://doi.org/10.3897/jhr.30.3535).
- Boudinot BE. 2015.** Contributions to the knowledge of Formicidae (Hymenoptera, Aculeata): a new diagnosis of the family, the first global male-based key to subfamilies, and a treatment of early branching lineages. *European Journal of Taxonomy* 120:1–62 DOI [10.5852/ejt.2015.120](https://doi.org/10.5852/ejt.2015.120).
- Boudinot BE. 2018.** A general theory of genital homologies for the Hexapoda (Pancrustacea) derived from skeletomuscular correspondences, with emphasis on the Endopterygota. *Arthropod Structure & Development* 47:563–613 DOI [10.1016/j.asd.2018.11.001](https://doi.org/10.1016/j.asd.2018.11.001).
- Boudinot BE. 2019.** Toward phylomics in entomology: current systematic and evolutionary morphology. *Insect Systematics and Diversity* 3:1–4 DOI [10.1093/isd/ixz019](https://doi.org/10.1093/isd/ixz019).

- Boulangé H. 1914.** Sur les appareils copulateurs de *Dolerus gonager* F et de *Dolerus nigratus* MUII [Hym Tenthredinidae]. *Bulletin de la Société Entomologique de France* 1914:416–418 DOI [10.5962/bhl.part.26812](https://doi.org/10.5962/bhl.part.26812).
- Boulangé H. 1924.** Recherches sur l'appareil copulateur des Hyménoptères et spécialement des Chalastogastres. *Mémoires et Travaux de la Faculté Catholique de Lille* 1:1–444.
- Brajković M, Stanisavljević L, Nikolić Z, Ćurčić SB, Zivić I, Stojanović D. 2010.** Male genitalia of the species of the subfamily Agathidinae (Hymenoptera: Braconidae) and their importance in taxonomy. *Archives of Biological Sciences* 62:455–467 DOI [10.2298/ABS1002455B](https://doi.org/10.2298/ABS1002455B).
- Branstetter MG, Childers AK, Cox-Foster D, Hopper KR, Kapheim KM, Toth AL, Worley KC. 2018.** Genomes of the Hymenoptera. *Current Opinion in Insect Science* 25:65–75 DOI [10.1016/j.cois.2017.11.008](https://doi.org/10.1016/j.cois.2017.11.008).
- Broad GR. 2012.** *Keys for the identification of British and Irish nocturnal Ichneumonidae*. London: Department of Entomology. Natural History Museum, 38.
- Broad GR, Shaw MR, Fitton MG. 2018.** Ichneumonid wasps (Hymenoptera: Ichneumonidae): their classification and biology. *Handbooks for the Identification of British Insects* 7(12):1–418.
- Brothers DJ, Carpenter JM. 1993.** Phylogeny of Aculeata: Chrysidoidea and Vespoidea. *Journal of Hymenoptera Research* 2:227–304.
- Clausen R. 1938.** Untersuchungen über den männlichen Copulationsapparat der Ameisen, speziell der Formicinae. *Mitteilungen Der Schweizerischen Entomologischen Gesellschaft* 17:233–246.
- Costa M, Reeve S, Grumbling G, Osumi-Sutherland D. 2013.** The Drosophila anatomy ontology. *Journal of Biomedical Semantics* 4:32 DOI [10.1186/2041-1480-4-32](https://doi.org/10.1186/2041-1480-4-32).
- Crampton GC. 1919.** The genitalia and terminal abdominal structures of males, and the terminal abdominal structures of the larvae of “chalastogastrous” Hymenoptera. *Proceedings of the Entomological Society of Washington* 21:129–151.
- Crampton GC. 1942.** *The external morphology of the Diptera. Guide to the insects of Connecticut. Part VI. The Diptera or true flies of Connecticut*. Hartford: State of Connecticut, 10–16.
- Daly HV. 1964.** *Skeleto-muscular morphogenesis of the thorax and wings of the Honey Bee Apis mellifera (Hymenoptera: Apidae)*. Vol. 39. Berkeley and Los Angeles: University of California Press, 77.
- Dasch CE. 1971.** Ichneumon-flies of America North of Mexico: 6. Subfamily Mesochorinae. *Memoirs of the American Entomological Institute* 16:1–376.
- Deans AR, Yoder MJ, Balhoff JP. 2012.** Time to change how we describe biodiversity. *Trends in Ecology & Evolution* 27:78–84 DOI [10.1016/j.tree.2011.11.007](https://doi.org/10.1016/j.tree.2011.11.007).
- de Brito CD, Lanes GdO, Azevedo CO. 2021.** Anatomic glossary of mesopleural structures in Bethyridae (Hymenoptera: Chrysidoidea). *Papéis Avulsos de Zoologia* 61:e20216152 DOI [10.11606/1807-0205/2021.61.52](https://doi.org/10.11606/1807-0205/2021.61.52).
- Delrio G. 1975.** Révision des espèces ouest-paléarctiques du genre *Netelia* Gray (Hym., Ichneumonidae). *Annali della Facoltà di Agraria dell'Università di Sassari* 23:1–126.

- Denk-Lobnig M, Martin AC. 2020.** Divergent and combinatorial mechanical strategies that promote epithelial folding during morphogenesis. *Current Opinion in Genetics & Development* **63**:24–29 DOI [10.1016/j.gde.2020.02.014](https://doi.org/10.1016/j.gde.2020.02.014).
- Dessart P, Gärdenfors U. 1985.** *Dendrocerus paradoxus* n. sp. et *D. ulmicola* n. sp. (Hym. Ceraphronoidea Megaspilidae), deux nouveaux hyperparasites paléarctiques de pucerons. *Bulletin et Annales de la Société Royale Belge d'Entomologie* **121**:197–211.
- Donkelaar HJT, Broman J, Neumann PE, Puellas L, Riva A, Tubbs RS, Kachlik D. 2017.** Towards a terminologia neuroanatomica. *Clinical Anatomy* **30**:145–155 DOI [10.1002/ca.22809](https://doi.org/10.1002/ca.22809).
- Dufour L. 1841.** *Recherches anatomiques et physiologiques sur les Orthoptères, les Hyménoptères et les Névroptères*. Paris: Impr. royale, 265–647.
- Duncan CD. 1939.** *A contribution to the biology of North American vespine wasps*. California: Stanford University Press, 272.
- Enslin E. 1918.** Die Tenthredinoidea Mitteleuropas. Berlin: Friedländer & Sohn, 790.
- Evans HE. 1950.** A taxonomic study of the Nearctic spider wasps belonging to the tribe Pompilini (Hymenoptera: Pompilidae). Part I. *Transactions of the American Entomological Society* **75**:133–270.
- Field WD. 1950.** Moths of the genus *Cincia* and three new and closely related genera. *Proceedings of the United States National Museum* **100**:311–326 DOI [10.5479/si.00963801.100-3264.311](https://doi.org/10.5479/si.00963801.100-3264.311).
- Forbes J, Do-Van-Quy DR. 1965.** The anatomy and histology of the male reproductive system of the legionary ant, *Neivamyrmex harrisi* (Haldeman) (Hymenoptera: Formicidae). *Journal of the New York Entomological Society* **73**(2):95–111.
- Friedrich F, Beutel R. 2008.** The thorax of *Zorotypus* (Hexapoda, Zoraptera) and a new nomenclature for the musculature of Neoptera. *Arthropod Structure & Development* **37**:29–54 DOI [10.1016/j.asd.2007.04.003](https://doi.org/10.1016/j.asd.2007.04.003).
- Girón JC, Tarasov S, González Montaña LA, Matentzoglou N, Smith AD, Koch M, Boudinot BE, Bouchard P, Burks R, Vogt L, Yoder MJ, Osumi-Sutherland D, Friedrich F, Beutel R, Miko I. 2023.** Formalizing invertebrate morphological data: A descriptive model for cuticle-based skeleto-muscular systems, an ontology for insect anatomy, and their potential applications in biodiversity research and informatics. *Systematic Biology* syad025 DOI [10.1093/sysbio/syad025](https://doi.org/10.1093/sysbio/syad025).
- Gobbi FT, Azevedo CO. 2016.** Revision of *Calobrachium* Gobbi & Azevedo, gen. nov. (Hymenoptera, Bethyridae), with description of seven new Oriental species. *Zootaxa* **4083**:221–238 DOI [10.11646/zootaxa.4083.2.3](https://doi.org/10.11646/zootaxa.4083.2.3).
- Hall BK. 1975.** Evolutionary consequences of skeletal differentiation. *American Zoologist* **15**:329–350 DOI [10.1093/icb/15.2.329](https://doi.org/10.1093/icb/15.2.329).
- Hartig T. 1837.** *Die Aderflugler Deutschlands mit besonderer Berücksichtigung ihres Larvenzustandes und ihres Wirkens in Waldern und Garten. Bd1. Die Familien der Blattwespen und Holzwespen, nebst einer allgemeinen Einleitung zur Naturgeschichte der Hymenopteren*. Haude und Spenersche Buchhandlung. Berlin: Neue Ausgabe, 329–350.

- Haupt H. 1962.** The Pompilidae of Israel. *Bulletin of the Research Council of Israel* 11(B):1–70.
- Heinrich G. 1961.** Synopsis of Nearctic Ichneumoninae Stenopneusticae with particular reference to the Northeastern Region (Hymenoptera). Part I. Introduction, Key to Nearctic Genera of Ichneumoninae Stenopneusticae, and synopsis of the Protichneumonini North of Mexico. *The Canadian Entomologist* 92:1–88.
- Johnson NF. 1984.** Systematics of Nearctic *Telenomus*: classification and revisions of the *podisi* and *phymatae* species groups (Hymenoptera: Scelionidae). *Bulletin of the Ohio Biological Survey* 6:1–113.
- Karlsson D, Ronquist F. 2012.** Skeletal morphology of *Opius dissitus* and *Biosteres carbonarius* (Hymenoptera: Braconidae), with a discussion of terminology. *PLOS ONE* 7:e32573 DOI 10.1371/journal.pone.0032573.
- Klopfstein S, Santos BF, Shaw MR, Alvarado M, Bennett AMR, Dal Pos D, Giannotta M, Herrera ADF, Karlsson D, Khalaim AI, Lima AR, Mikó I, Sääksjärvi IE, Shimizu S, Spasojevic T, van Noort S, Vilhelmsen L, R BG. 2019.** Darwin wasps: a new name heralds renewed efforts to unravel the evolutionary history of Ichneumonidae. *Entomological Communications* 1:ec010 DOI 10.37486/2675-1305.ec01006.
- Klopfstein S, Vilhelmsen L, Heraty JM, Sharkey M, Ronquist F. 2013.** The Hymenopteran tree of life: evidence from protein-coding genes and objectively aligned ribosomal data. *PLOS ONE* 8:e69344 DOI 10.1371/journal.pone.0069344.
- Koch F, Liston AD. 2017.** Revision of the Afrotropical genus *Distega* Konow, 1904 (Hymenoptera: Symphyta: Tenthredinidae: Blennocampinae). *Mitteilungen der Münchner Entomologischen Gesellschaft* 107:43–112.
- Konishi K. 1991.** A revision of the subgenus *Prosthodocis* Enderlein of the genus *Netelia* Gray of Japan (Hymenoptera, Ichneumonidae), I. *Japanese Journal of Entomology* 59:775–788.
- Konishi K. 1992.** A revision of the subgenus *Prosthodocis* Enderlein of the genus *Netelia* Gray of Japan (Hymenoptera, Ichneumonidae), II. *Japanese Journal of Entomology* 60:39–53.
- Konishi K. 1996.** A revision of the subgenus *Paropheltes* Cameron of the genus *Netelia* Gray (Hymenoptera, Ichneumonidae) of Japan. *Japanese Journal of Entomology* 64:163–188.
- Konishi K. 2005.** A preliminary revision of the subgenus *Netelia* of the genus *Netelia* from Japan (Hymenoptera, Ichneumonidae, Tryphoninae). *Insecta Matsumurana* 62:45–121.
- Konishi K. 2010.** Taxonomic status of *Amebachia* Uchida in the genus *Netelia* Gray (Hymenoptera; Ichneumonidae; Tryphoninae) with descriptions of four new species from Japan. *Entomological Science* 13:216–225 DOI 10.1111/j.1479-8298.2010.00376.x.
- Konishi K, Chen H-P, Pham NT. 2022.** A taxonomic review of the genus *Netelia*, subgenus *Monomacrodon* (Hymenoptera: Ichneumonidae: Tryphoninae), with description of a new species. *Raffles Bulletin of Zoology* 13:216–225.

- Kopelke J-P. 1981.** Funktion der Genitalstrukturen bei *Bombus*-Arten am Beispiel von *B. lapidarius* (Linnaeus 1758) und deren Bedeutung für die Systematik (Insecta: Hymenoptera: Apidae). *Senckenbergiana Biologica* **62**:267–286.
- Königsmann E. 1976.** Das phylogenetische System der Hymenoptera. Teil 1: Einführung, Grundplanmerkmale, Schwestergruppe und Fossilfunde. *Deutsche Entomologische Zeitschrift* **23**:253–279.
- Lackner T, Tarasov S. 2019.** Female genitalia are moderately informative for phylogenetic inference and not concerted with male genitalia in Sapriniinae beetles (Coleoptera: Histeridae). *Systematic Entomology* **4**:667–685 DOI [10.1111/syen.12346](https://doi.org/10.1111/syen.12346).
- Lanes GO, Kawada R, Azevedo CO, Brothers DJ. 2020.** Revisited morphology applied for systematics of flat wasps (Hymenoptera, Bethyridae). *Zootaxa* **4752**(1):1–127 DOI [10.11646/zootaxa.4752.1.1](https://doi.org/10.11646/zootaxa.4752.1.1).
- Lee JW. 1991.** A revision of the genus *Cidaphus* (Hymenoptera: Ichneumonidae: Mesochorinae). *Contributions of the American Entomological Institute* **26**:1–48.
- Loan CC. 1974.** The European species of *Leiophron* Nees and *Peristenus* Foerster (Hymenoptera: Braconidae, Euphorinae). *Transactions of the Royal Entomological Society of London* **126**:207–238.
- Maetô K. 1996.** Inter-generic variation in the external male genitalia of the subfamily Microgastrinae (Hymenoptera, Braconidae), with a reassessment of Mason's tribal system. *Journal of Hymenoptera Research* **5**:38–52.
- Malagon-Aldana LA, Smith DR, Shinohara A, Vilhelmsen L. 2021.** From *Arge* to *Zenarge*: adult morphology and phylogenetics of argid sawflies (Hymenoptera: Argidae). *Zoological Journal of the Linnean Society* **193**(3):880–938 DOI [10.1093/zoolinnean/zlaa170](https://doi.org/10.1093/zoolinnean/zlaa170).
- Michener CD. 1944.** A comparative study of the appendages of the eighth and ninth abdominal segments of insects. *Annals of the Entomological Society of America* **37**:336–351 DOI [10.1093/aesa/37.3.336](https://doi.org/10.1093/aesa/37.3.336).
- Michener CD. 1956.** Hymenoptera. In: Tuxen SL, ed. *Taxonomist's glossary of genitalia in insects*. Copenhagen: Munksgaard, 131–140.
- Michener CD. 1965.** A classification of the bees of the Australian and South Pacific regions. *Bulletin of the American Museum of Natural History* **130**:1–394.
- Mikó I, Deans AR. 2009.** *Masner*, a new genus of Ceraphronidae (Hymenoptera, Ceraphronoidea) described using controlled vocabularies. *Zookeys* **20**:127–153 DOI [10.3897/zookeys.20.119](https://doi.org/10.3897/zookeys.20.119).
- Mikó I, Deans AR. 2013.** What is fluorescing? *Hamuli* **4**:19–22.
- Mikó I, Friedrich F, Yoder MJ, Hines HM, Deitz LL, Bertone MA, Seltmann KC, Wallace MS, Deans AR. 2012.** On dorsal prothoracic appendages in treehoppers (Hemiptera: Membracidae) and the nature of morphological evidence. *PLOS ONE* **7**:e30137 DOI [10.1371/journal.pone.0030137](https://doi.org/10.1371/journal.pone.0030137).
- Mikó I, Masner L, Johannes E, Yoder MJ, Deans AR. 2013.** Male terminalia of Ceraphronoidea: morphological diversity in an otherwise monotonous taxon. *Insect Systematics & Evolution* **44**:261–347.

- Mikó I, Vilhelmsen L, Johnson NF, Masner L, Penzes Z. 2007. Skeletomusculature of Scelionidae (Hymenoptera: Platygastroidea): head and mesosoma. *Zootaxa* 1571:1–78 DOI 10.11646/zootaxa.1571.1.1.
- Nichols SW. 1989. *The Torre-Bueno glossary of entomology*. New York: New York Entomological Society and the American Museum of Natural History, 261–347.
- Olmi M. 1984a. A revision of the Dryinidae (Hymenoptera). Part I. *Memoirs of the American Entomological Insitute* 37:1–946.
- Olmi M. 1984b. A revision of the Dryinidae (Hymenoptera). Part II. *Memoirs of the American Entomological Insitute* 37:947–1913.
- Owen AK, George J, Pinto JD, Heraty JM. 2007. A molecular phylogeny of the Trichogrammatidae (Hymenoptera: Chalcidoidea), with an evaluation of the utility of their male genitalia for higher level classification. *Systematic Entomology* 32:227–251 DOI 10.1111/j.1365-3113.2006.00361.x.
- Peck O. 1937a. The male genitalia in the Hymenoptera (Insecta), especially the family Ichneumonidae: I. Comparative morphology. *Canadian Journal of Research* 15:221–252.
- Peck O. 1937b. The male genitalia in the Hymenoptera (Insecta), especially the family Ichneumonidae: II. Taxonomy of the Ichneumonid genitalia. *Canadian Journal of Research* 15:253–274.
- Pratt HD. 1939. Studies on the Ichneumonidae of New England. Part II. Male Genitalia of the subfamily Ichneumoninae. *Annals of the Entomological Society of America* 32:727–742 DOI 10.1093/aesa/32.4.727.
- Priesner H. 1966. Zur Kenntnis der Gattung *Episyron* (Hymenoptera, Pompilidae). *Bollettino Dell'Istituto Di Entomologia Della Universita Degli Studi Di Bologna* 28:29–55.
- Quicke DLJ. 1988. Inter-generic variation in the male genitalia of the Braconinae (Insecta, Hymenoptera, Braconidae). *Zoologica Scripta* 17:399–409 DOI 10.1111/j.1463-6409.1988.tb00115.x.
- Quicke DLJ. 2015. *The Braconid and Ichneumonid parasitoid wasps: biology, systematics, evolution and ecology*. Oxford: John Wiley & Sons, 681.
- Quicke DLJ, van Achterberg C. 1990. Phylogeny of the subfamilies of the family Braconidae (Hymenoptera: Ichneumonoidea). *Zoologische Verhandelingen* 258:1–95.
- Ronquist F, Nordlander G. 1989. Skeletal morphology of an archaic cynipoid, *Ibalia rufipes* (Hymenoptera: Ibalidae). *Entomologica Scandinavica—Supplementum* 33:1–60.
- Ross H. 1937. A generic classification of the Nearctic sawflies (Hymenoptera, Symphyta). *Illinois Biological Monographs* 15:1–137.
- Ross H. 1945. Sawfly genitalia: terminology and study techniques. *Entomological News* 61:261–268.
- Schenck A. 1851. Beschreibung nassauischer Bienenarten. *JahrbüCher Des Nassauischen Vereins für Naturkunde* 7:1–105.

- Schindelin J, Rueden CT, Hiner MC, Eliceiri KW. 2015.** The ImageJ ecosystem: and open platform for biomedical image analysis. *Molecular Reproduction and Development* **82**(7–8):518–529 DOI [10.1002/mrd.22489](https://doi.org/10.1002/mrd.22489).
- Schulmeister S. 2001.** Functional morphology of the male genitalia and copulation in lower Hymenoptera, with special emphasis on the Tenthredinoidea s. str. (Insecta, Hymenoptera, ‘Symphyta’). *Acta Zoologica* **82**:331–349 DOI [10.1046/j.1463-6395.2001.00094.x](https://doi.org/10.1046/j.1463-6395.2001.00094.x).
- Schulmeister S. 2003.** Genitalia and terminal abdominal segments of male basal Hymenoptera (Insecta): morphology and evolution. *Organisms Diversity & Evolution* **3**(4):253–279 DOI [10.1078/1439-6092-00078](https://doi.org/10.1078/1439-6092-00078).
- Scobiola X. 1963.** Sur quelques Pompiloides, nouveaux ou rares, de la R.P. Roumaine: 11e nôte. *Travaux Du Museum D’Histoire Naturelle ‘Grigore Antipa’* **4**:269–281.
- Seurat LG. 1898.** Observations sur les organes génitaux des Braconides. *Annales Des Sciences Naturelles* **8**:293–303.
- Seurat LG. 1899.** *Contributions à l’étude des Hyménoptères entomophages*. Paris: Masson, 159.
- Sharanowski BJ, Ridenbaugh RD, Piekarski PK, Broad GR, Burke GR, Deans AR, Lemmon AR, Lemmon ECM, Diehl GJ, Whitfield JB, Hines HM. 2021.** Phylogenomics of Ichneumonoidea (Hymenoptera) and implications for evolution of mode of parasitism and viral endogenization. *Molecular Phylogenetic and Evolution* **156**:107023 DOI [10.1016/j.ympev.2020.107023](https://doi.org/10.1016/j.ympev.2020.107023).
- Smith DR. 1990.** A synopsis of the sawflies (Hymenoptera, Symphyta) of America south of the United States: Pergidae. *Revista Brasileira de Entomologia* **34**:7–200.
- Smith EL. 1969.** Evolutionary morphology of external insect genitalia. 1. Origin and relationships to other appendages. *Annals of the Entomological Society of America* **62**:1051–1079 DOI [10.1093/aesa/62.5.1051](https://doi.org/10.1093/aesa/62.5.1051).
- Smith EL. 1970a.** Evolutionary morphology of the external insect genitalia. 2. Hymenoptera. *Annals of the Entomological Society of America* **63**:1–27 DOI [10.1093/aesa/63.1.1](https://doi.org/10.1093/aesa/63.1.1).
- Smith EL. 1970b.** Hymenoptera. In: Tuxen SL, ed. *Taxonomist’s glossary of genitalia in insects*. Copenhagen: Munksgaard, 156–170.
- Smith EL. 1972.** Biosystematics and morphology of Symphyta—III. External genitalia of *Euura* (Hymenoptera: Tenthredinidae): sclerites, sensilla, musculature, development and oviposition behavior. *International Journal of Insect Morphology and Embryology* **1**(4):321–365 DOI [10.1016/0020-7322\(72\)90016-5](https://doi.org/10.1016/0020-7322(72)90016-5).
- Snodgrass RE. 1935.** *Principles of insect morphology*. New York: McGraw-Hill, 667.
- Snodgrass RE. 1941.** The male genitalia of Hymenoptera. *Smithsonian Miscellaneous Collections* **135**:1–86.
- Snodgrass RE. 1957.** Revised interpretation of the external reproductive organs of male insects. *Smithsonian Miscellaneous Collections* **135**:1–60.

- Sobczak JF, Loffredo APdS, Penteado-Dias A, Messas Y, Pádua DG. 2019.** Description of the male of *Hymenoepimecis bicolor* (Brullé, 1846) (Hymenoptera, Ichneumonidae, Pimplinae). *Brazilian Journal of Biology* **79**:154–157 DOI [10.1590/1519-6984.178889](https://doi.org/10.1590/1519-6984.178889).
- Song H, Bucheli SR. 2010.** Comparison of phylogenetic signal between male genitalia and non-genital characters in insect systematics. *Cladistics* **26**:23–35 DOI [10.1111/j.1096-0031.2009.00273.x](https://doi.org/10.1111/j.1096-0031.2009.00273.x).
- Thomson CG. 1872.** *Hymenoptera Scandinaviae vol. II*. Lundae: H. Ohlsson, 387–399.
- Tobias V. 1967.** A review of the classification, phylogeny and evolution of the family Braconidae (Hymenoptera). *Entomological Review* **46**:387–399.
- Togashi I. 1970.** The comparative morphology of the internal reproductive organs of the Symphyta (Hymenoptera). *Mushi* **66**:1–114.
- Townes HK. 1939.** The Nearctic species of *Netelia* (*Paniscus* of authors) and a revision of the genera of Netelini. *Lloydia* **1**:168–231.
- Townes HK. 1957.** Nearctic wasps of the subfamilies Pepsinae and Ceropalinae. *Bulletin of the United States National Museum* **209**:1–286.
- Townes HK. 1969a.** The genera of Ichneumonidae, Part 1. *Memoirs of the American Entomological Institute* **11**:1–300.
- Townes HK. 1969b.** The genera of Ichneumonidae, Part 2. *Memoirs of the American Entomological Institute* **12**:1–537.
- Townes HK. 1969c.** The genera of Ichneumonidae, Part 3. *Memoirs of the American Entomological Institute* **13**:1–307.
- Townes HK. 1971.** The genera of Ichneumonidae, Part 4. *Memoirs of the American Entomological Institute* **13**:1–372.
- Tremblay E. 1979.** Le armature genitali maschili nei generi degli Aphidiinae (Hymenoptera Braconidae). I. *Bollettino del Laboratorio di Entomologia Agraria “Filippo Silvestri”* **38**:221–233.
- Tremblay E. 1981.** Le armature genitali maschili nei generi degli Aphidiinae (Hymenoptera Braconidae). II. *Bollettino del Laboratorio di Entomologia Agraria “Filippo Silvestri”* **38**:251–263.
- Tremblay E. 1983.** Le armature genitali maschili nei generi degli Aphidiinae (Hymenoptera Braconidae). III. *Bollettino del Laboratorio di Entomologia Agraria “Filippo Silvestri”* **40**:147–158.
- Tuxen SL. 1956.** *Taxonomist’s glossary of genitalia in insects*. Copenhagen: Munksgaard, 283.
- Tuxen SL. 1970.** *Taxonomist’s glossary of genitalia in insects*. Copenhagen: Munksgaard, 359.
- Van Dam MH. 2014.** A simple, rapid technique for the inflation of the endophallus, with particular focus on the Curculionoidea (Coleoptera). *The Coleopterists Bulletin* **68**:263–268 DOI [10.1649/0010-065X-68.2.263](https://doi.org/10.1649/0010-065X-68.2.263).
- Verhoeff C. 1832.** Finden sich für die laminae basales der männlichen Coleopteren Homologa bei den Hymenopteren. *Zoologischer Anzeiger* **16**:407–412.

- Vilhelmsen L. 1996.** The preoral cavity of lower Hymenoptera (Insecta): comparative morphology and phylogenetic significance. *Zoologica Scripta* **25**:143–170 DOI [10.1111/j.1463-6409.1996.tb00156.x](https://doi.org/10.1111/j.1463-6409.1996.tb00156.x).
- Vilhelmsen L. 1997.** The phylogeny of lower Hymenoptera (Insecta), with a summary of the early evolutionary history of the order. *Journal of Zoological Systematics and Evolutionary Research* **35**:49–70 DOI [10.1111/j.1439-0469.1997.tb00404.x](https://doi.org/10.1111/j.1439-0469.1997.tb00404.x).
- Vilhelmsen L. 2000a.** Before the wasp-waist: comparative anatomy and phylogenetic implications of the skeleto-musculature of the thoraco-abdominal boundary region in basal Hymenoptera (Insecta). *Zoomorphology* **119**:185–221 DOI [10.1007/PL00008493](https://doi.org/10.1007/PL00008493).
- Vilhelmsen L. 2000b.** Cervical and prothoracic skeleto-musculature in the basal Hymenoptera (Insecta): comparative anatomy and phylogenetic implications. *Zoologischer Anzeiger* **239**:105–138.
- Vilhelmsen L. 2010.** Beyond the wasp-waist: structural diversity and phylogenetic significance of the mesosoma in apocritan wasps (Insecta: Hymenoptera). *Zoological Journal of the Linnean Society* **159**:22–194 DOI [10.1111/j.1096-3642.2009.00576.x](https://doi.org/10.1111/j.1096-3642.2009.00576.x).
- Vogt L, Bartolomaeus T, Giribet G. 2010.** The linguistic problem of morphology: structure versus homology and the standardization of morphological data. *Cladistics* **26**:301–325 DOI [10.1111/j.1096-0031.2009.00286.x](https://doi.org/10.1111/j.1096-0031.2009.00286.x).
- Vogt L, Bartolomaeus T, Giribet G. 2022.** Anatomy and the type concept in biology show that ontologies must be adapted to the diagnostic needs of research. *Journal of Biomedical Semantics* **13**(1):18 DOI [10.1186/s13326-022-00268-2](https://doi.org/10.1186/s13326-022-00268-2).
- Wahl DB, Gauld ID. 1998.** The cladistics and higher classification of the Pimpliformes (Hymenoptera: Ichneumonidae). *Systematic Entomology* **23**:265–298 DOI [10.1046/j.1365-3113.1998.00057.x](https://doi.org/10.1046/j.1365-3113.1998.00057.x).
- Walker AK. 1994.** Species of Microgastrinae (Hymenoptera: Braconidae) parasitizing lepidopterous cereal stem borers in Africa. *Bulletin of Entomological Research* **84**:421–434 DOI [10.1017/S0007485300032557](https://doi.org/10.1017/S0007485300032557).
- Watanabe K, Matsumoto R. 2010.** Disjunctive distribution of the basal genus *Aplomerus* (Hymenoptera: Ichneumonidae: Xoridinae) in East Asia and North America, with a new species from Japan. *Entomological Science* **13**(4):375–380 DOI [10.1111/j.1479-8298.2010.00397.x](https://doi.org/10.1111/j.1479-8298.2010.00397.x).
- Watanabe K, Taniwaki T, Kasparyan DR. 2015.** *Tanzawana flavomaculata* (Hymenoptera, Ichneumonidae, Ctenopelmatinae), a new genus and species of parasitoid of *Fagineura crenativora* (Tenthredinidae, Nematinae), a serious pest of beech tree. *Zootaxa* **4040**(2):236–242 DOI [10.11646/zootaxa.4040.2.9](https://doi.org/10.11646/zootaxa.4040.2.9).
- Wheeler WM. 1910.** *Ants: Their structure, development and behaviour*. New York: Columbia Press, 663.
- Willman. 1998.** Advances and problems in insect phylogeny. In: Fortey TR, ed. *Arthropod relationships*. London: Chapman & Hall, 269–279.
- Wirkner CS, Göpel T, Runge J, Keiler J, Klusmann-Fricke B-J, Huckstorf K, Scholz S, Mikó I, Yoder MJ, Richter S. 2017.** The first organ-based ontology for arthropods (Ontology of Arthropod Circulatory Systems-OArCS) and its integration into a

- novel formalization scheme for morphological descriptions. *Systematic Biology* **66**:754–768.
- Wong HR. 1963.** The external morphology of the adults and ultimate larval instar of the larch sawfly, *Pristiphora erichsonii* (Htg.) (Hymenoptera: Tenthredinidae). *The Canadian Entomologist* **95**:897–921 DOI [10.4039/Ent95897-9](https://doi.org/10.4039/Ent95897-9).
- Worthley EN. 1924.** The biology of *Trichopoda pennipes* Fab (Diptera, Tachinidae), a parasite of the common squash bug. *Psyche* **31**:57–77 DOI [10.1155/1924/96797](https://doi.org/10.1155/1924/96797).
- Yoder MJ, Mikó I, Seltmann KC, Bertone MA, Deans AR. 2010.** A gross anatomy ontology for Hymenoptera. *PLOS ONE* **5**:e15991 DOI [10.1371/journal.pone.0015991](https://doi.org/10.1371/journal.pone.0015991).
- Yoshimura M, Fisher BL. 2011.** A revision of male ants of the Malagasy region (Hymenoptera: Formicidae): Key to genera of the subfamily Dolichoderinae. *Zootaxa* **2794**:1–34 DOI [10.11646/zootaxa.2794.1.1](https://doi.org/10.11646/zootaxa.2794.1.1).
- Yu DSK, van Achterberg C, Horstmann K. 2016.** *Taxapad 2016, Ichneumonoidea 2015*. Nepean, Ontario, Canada: Database on flash-drive.
- Zander E. 1900.** Beiträge zur Morphologie der männlichen Geschlechtsanhänge der Hymenopteren. *Zeitschrift für Wissenschaftliche Zoologie* **76**:431–489.
- Žikić V, van Achterberg C, Stanković SS, Ilić M. 2011.** The male genitalia in the subfamily Agathidinae (Hymenoptera: Braconidae): morphological information of species on generic level. *Zoologischer Anzeiger* **250**:246–257 DOI [10.1016/j.jcz.2011.04.006](https://doi.org/10.1016/j.jcz.2011.04.006).