

Metatranscriptomes of two biological soil crust types from the Mojave desert in response to wetting

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ABSTRACT We present eight metatranscriptomic datasets of light algal and cyanolichen biological soil crusts from the Mojave Desert in response to wetting. These data will help us understand gene expression patterns in desert biocrust microbial communities after they have been reactivated by the addition of water.

KEYWORDS biocrust, RNA, transcriptome, wetting, desert, soil

Biological soil crusts comprise diverse microbial communities that carry out vital ecological functions in dryland ecosystems (1). Under dry conditions, biocrust microbes primarily persist in dormancy (2–4). When water becomes available, they quickly respond by exploiting moisture to repair cell damage and synthesize new biomass (5, 6). Nevertheless, the specific gene expression and metabolic processes underlying these responses remain poorly understood.

We sought to compare two kinds of biocrust commonly found in the Sheephole Valley Wilderness (Mojave Desert): light algal crust (LAC) and cyanolichen crust (CLC). In all, 10 biocrust samples, each measuring 5 cm², were collected at GPS location 34.1736 N, 115.3888 W. Each sample was placed in a 10 cm petri dish with 2 mL of sterile ultrapure water added on top, covered with a petri dish cover, and incubated at ambient laboratory conditions. After 0.5, 6, 18, 30, and 50 h time points, an entire biocrust sample was transferred and stored at –80°C for subsequent total RNA extraction using a NucleoBond RNA Soil Midi kit (740140.20, Macherey-Nagel, Nordrhein-Westfalen, Germany). We pursued rRNA depletion of 100 ng of total RNA using a QIAseq FastSelect 5S/16S/23S kit for bacteria and FastSelect rRNA yeast and plant depletion for eukaryotes (335921, 334219, and 334319, QIAGEN, Germantown, MD) following the manufacturer's instructions. The resulting RNA was reverse transcribed to create first-strand cDNA using a TruSeq Stranded mRNA Library prep kit (20020594, Illumina Inc., San Diego, CA). To synthesize second-strand cDNA, deoxyuridine triphosphate was incorporated in place of deoxythymidine triphosphate to quench the second strand during amplification and achieve strand specificity. Double-stranded cDNA fragments were A-tailed and ligated to JGI dual-indexed Y-adapters, followed by 10 cycles of PCR. The prepared libraries were quantified using KAPA Biosystems' next-generation sequencing library qPCR kit and run on a LightCycler 480 real-time PCR instrument (Roche Diagnostics Corporation, Indianapolis, IN). NovaSeq sequencing (Illumina Inc., San Diego, CA) was performed using NovaSeq XP V1 reagent kits and an S4 flowcell following a 2 × 151 bp indexed run recipe. BBDuk version 38.87 (<https://jgi.doe.gov/data-and-tools/bbtools/>) was used to remove contaminants, trim adapters from Illumina raw sequencing reads, remove any reads that contained "N" bases, and were shorter than 51 bp. Filtered reads were assembled with MEGAHIT version v1.2.9 (7) and mapped back to the final transcriptome assembly and coverage determined using BBMap version 38.86 (8).

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TABLE 1 Accession numbers and characteristics of metatranscriptomes from two types of biological soil crusts, light algal crust (LAC) and cyanolichen crust (CLC), over the course of a re-wetting experiment (times shown indicate sample harvest time post-wetting; CLC samples at 0.5 and 30 h time points did not generate sufficient high-quality RNA yields for sequencing). All contigs are ≥ 0.1 kb

Meta-transcriptome	NCBI BioSample ID	NCBI BioProject ID	No. of raw reads	No. of filtered reads	Assembly BioSample ID	No. of Contigs	No. of assembled (150 bp) reads	Assembly length (bp)	Transcriptome coverage	N50 (bp)	Max contig length (KB)
LAC 0.5 h	SAMIN17674635	PRJNA697426	378,329,084	15,399,682	GKPO000000000	58,795	12,494,595	31,311,788	59.9×	18,350	7.034
LAC 6 h	SAMIN18245122	PRJNA710733	406,275,950	19,607,874	GKPP000000000	88,036	16,171,069	50,130,842	48.4×	25,380	20.259
LAC 18 h	SAMIN17675269	PRJNA697427	437,433,136	20,442,408	GKPN000000000	72,020	16,932,941	38,371,519	66.2×	22,289	7.537
LAC 30 h	SAMIN17675483	PRJNA697428	500,168,512	20,768,548	GKPP000000000	86,683	17,426,116	50,104,316	52.2×	24,532	14.942
LAC 50 h	SAMIN17674330	PRJNA697429	670,916,034	38,911,978	GKPP000000000	109,448	32,668,699	61,798,533	79.3×	31,386	18.369
CLC 6 h	SAMIN17674629	PRJNA697430	590,894,720	32,744,316	GKPS000000000	88,422	27,681,580	50,698,865	81.9×	24,701	23.151
CLC 18 h	SAMIN18247024	PRJNA710734	528,673,374	28,175,474	GKPT000000000	60,086	23,018,914	35,379,485	97.6×	15,771	19.855
CLC 50 h	SAMIN18245957	PRJNA710735	682,130,280	29,262,602	GKPU000000000	94,375	23,172,351	51,949,060	66.9×	27,333	27.808

Nearly 95% of reads aligned to ribosomal reference sequences in the SILVA database (9) using BBDuk (version 38.87, default settings), suggesting that experimental rRNA depletion was not effective. Nevertheless, these rRNA reads could be assembled and used to comprehensively survey the taxonomic diversity contained within these biocrusts (10). We obtained at least 25 million mRNA reads per sample, of which 80% could be assembled into contigs; this represents an average transcriptome coverage of ~69× and should be sufficient depth for functional analyses of wetting the reanimation process.

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DATA AVAILABILITY

Raw sequencing data and assemblies are accessible at the NCBI using the BioSample and BioProject IDs listed in Table 1. The data are also available from JGI's genome portal (<https://genome.jgi.doe.gov/portal/ProMicSoilCrusts/ProMicSoilCrusts.info.html>) or GOLD database (<https://gold.jgi.doe.gov/study?id=Gs0142145>).

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