

# Forest habitats and plant communities strongly predicts Megachilidae bee biodiversity

Lindsie M. McCabe<sup>1</sup>  , Paige Chesshire<sup>2</sup> and Neil S. Cobb<sup>3</sup>

<sup>1</sup> USDA-ARS Pollinating Insects Research Unit, Logan, Utah, United States

<sup>2</sup> Department of Biological Science, Northern Arizona University, Flagstaff, Arizona, United States

<sup>3</sup> Biodiversity Outreach Network, Flagstaff, AZ, United States

## ABSTRACT

Megachilidae is one of the United States' most diverse bee families, with 667 described species in 19 genera. Unlike other bee families, which are primarily ground nesters, most megachilid bees require biotic cavities for nesting (*i.e.*, wood, pithy stems, *etc.*). For this group, the availability of woody-plant species may be as important as nectar/pollen resources in maintaining populations. We studied Megachilidae biodiversity in the continental United States. We confirmed that the highest species richness of Megachilidae was in the southwestern United States. We examined the relationship between species richness and climate, land cover, tree species richness, and flowering plant diversity. When examining environmental predictors across the conterminous United States, we found that forested habitats, but not tree diversity, strongly predicted Megachilidae richness. Additionally, Megachilidae richness was highest in areas with high temperature and low precipitation, however this was not linearly correlated and strongly positively correlated with flowering plant diversity. Our research suggests that the availability of nesting substrate (forested habitats), and not only flowering plants, is particularly important for these cavity-nesting species. Since trees and forested areas are particularly susceptible to climate change, including effects of drought, fire, and infestations, nesting substrates could become a potential limiting resource for Megachilidae populations.

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Corresponding author

Lindsie M. McCabe,

[lindsie.mccabe@usda.gov](mailto:lindsie.mccabe@usda.gov)

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## INTRODUCTION

Insect pollinators provide key ecosystem services; over 86% of all flowering plant species depend on insects for pollination ([Klein et al., 2006](#); [Ollerton, Winfree & Tarrant, 2011](#)), and bees alone pollinate roughly one-third of the world's staple food crops ([Potts et al., 2010](#)). Reported declines in bee species abundance may affect both economically important crop yield and wild plant populations worldwide ([Brown & Paxton, 2009](#); [Potts et al., 2010](#); [Sahli & Conner, 2006](#)). Bee declines can be attributed to habitat loss, pesticides, invasive species, and climate change ([Klein et al., 2007](#); [Memmott et al., 2010](#); [Potts et al., 2016](#)). Despite the importance of bees, most bee species distributions are unknown, and even less is known about the factors that drive their distributions.

Bees are highly diverse and have a wide range of life history strategies including sociality, nesting strategies, parasitism and diets (Michener, 2007; Westrich, 2007; Danforth et al., 2019; Mikat, Matouskova & Straka, 2021; Bossert et al., 2022). Many of these traits have evolved and/or been lost over evolutionary history and geographic space, contributing to the diversification of bees (Danforth et al., 2013; Hettke, Patiny & Danforth, 2013). Climate, seasonality and shifting plants have all contributed to species and trait diversification for this group (Toussaint et al., 2012; Kerfoot et al., 2018). One unifying characteristic of bees, however, is that bees have strong biotic interactions, such as relying on pollen and nectar from plants for food (Bartomeus et al., 2013). Despite these strong ties, floral richness has not been well associated with bee species richness globally; the tropics are a hot bed of diversity for plant species but have the lowest bee species diversity (Michener, 2007; Orr et al., 2020). This suggests that other life history traits are strongly influencing geographic distribution for this taxa.

Megachilidae is globally distributed and comprises 35% of the known 4,000 bee species (Michener, 2007) in the United States. Megachilid bees have important roles in pollination of a diverse range of ecosystems, including natural, urban, and agricultural environments (Gonzalez et al., 2012). *Osmia* and *Megachile* are successfully managed for crop and wildflower pollination; and *Osmia lignaria* is the key pollinator reared for apple, cherries, and other fruit trees *Megachile rotundata* is reared as the primary pollinator of alfalfa (Bosch & Kemp, 2002; Boyle & Pitts-Singer, 2019; Pitts-Singer et al., 2018; Pitts-Singer & Cane, 2011; Sampson & Cane, 2000).

Many megachilid bees nest in pre-existing cavities in decaying wood, hollow plant stems, or hollow twigs, and use a range of materials to create their nest cells, including masticated leaves, mud, plant resins, or flower petals (Bosch & Kemp, 2002; Litman et al., 2011; Young et al., 2016). The wide use of foreign materials for nest building is a likely reason for evolutionary range expansion and high diversification in Megachilidae, (Gonzalez et al., 2012; Litman et al., 2011). About 75% of the genera for this family have above ground/cavity-nesting traits (Eickwort, Matthews & Carpenter, 1981), however primitive forms of nesting (*i.e.*, ground borrowing) still exist in eight of the 19 genera (Bosch, Maeta & Rust, 2001). It is likely that the above ground nesting behavior was repeatedly evolved and lost numerous times throughout the evolution of this group, since very closely related species can have drastically different nesting behaviors (Sedivy, Dorn & Muller, 2013).

Bees' habitat, shelter, and food may all be impacted by climate change events (Corbet et al., 1993; Tews et al., 2006; Bartomeus et al., 2011; McCabe, Asian & Cobb, 2022). When assessing how climate change and other anthropogenic disturbances will impact these species, research should not just look at how increasing temperatures will directly affect bees. Understanding where species are distributed can help to understand what the most limiting resources are for the success of a population.

Recent aggregation of databases and occurrence records have made it possible to assess spatial and temporal questions about insect species diversity (Cavender-Bares et al., 2009). This data can provide insight on how species and communities will respond to future disturbance (*i.e.*, climate, habitat loss, *etc.*), but it is first critical to understand what is

driving patterns of biodiversity and how these species respond to different environmental parameters ([Weiher et al., 2011](#)).

While it is widely accepted that Megachilidae have strong ties to biotic associations (e.g., host plants and cavity nesting resources: trees and pithy stems) in North America ([Martins & de Almeida, 1994](#); [Michener, 2007](#); [Bosch, Maeta & Rust, 2001](#); [Cane, Griswold & Parker, 2007](#)), no study has systematically assessed the relationship between species richness and nesting substrate. We reviewed the biogeographic patterns of United States Megachilidae species, with two primary questions. (1) How can biogeographic patterns be explained by climate, nesting resources, and floral resources? (2) Are there specific relationships between Megachilidae biodiversity hotspots and taxonomic and/or morphological structure of woody plants? We predicted that for Megachilidae in particular they would be distributed in areas where cavity-nesting resources were also abundant, and in areas with high plant diversity.

## METHODS

### Data extraction

Bee data was collected from three aggregate databases: iDigBio, Symbiota Collection of Arthropod Network (SCAN) ([SCAN, 2019](#)), and Global Biodiversity Information Facility (GBIF) ([Global Biodiversity Information Facility, 2020](#)). For all three databases, all records from the family "Megachilidae" occurring in the "United States", "Canada" and "Mexico" were extracted. These records were thoroughly cleaned to only include those with georeferenced information, identified to species (if they record was lacking scientific name or classified as a morphospecies they were not included), and were determined to be specimen records; observation records were not used in this data set (records such as iNaturalist observations, BugGuide and/or Xerces society records that appeared on GBIF or SCAN were not used). All duplicate records were removed by filtering each dataset using the Darwin Core Archive (DwC) field "catalog number" and "institution code". Records were then checked for taxonomic (including synonyms) and geographical errors. We resolved all spelling, male/female suffix discrepancies and all additional information that may have been included in this field. All species were vetted for location accuracy and probability that the species occurred in the area sampled by taxonomic experts of this family (T. Griswold & J. Ascher, 2021, personal communication). Data were cleaned in the same manner as in [Chessire et al. \(2023\)](#) and as in [Dorey et al. \(2023\)](#). Great care was taken to make sure all records were as accurate as possible. There was significant lack of occurrence records for north and central Canada, and patches of minimal records throughout Montana and the Dakotas in the United States, which is likely due to sampling biases, however data was restricted only to the United States due to insufficient land cover data available in Canada and/or Mexico. In total we used 248,367 cleaned georeferenced records that comprised 667 Megachilidae species for the United States and represented 19 genera ([Table 1](#)). All Megachilidae species were used in these analyses regardless of nesting preference since nesting preference is so wildly variable between related species and species level nesting preferences were available for less than 20% of all Megachilidae species.

**Table 1** List of genera and their known nesting substrates, number of occurrence records used in this analysis, and average number of plants visited based on known literature and DiscoverLife.

Genus	# of species	Number of occurrence records
Anthidiellum	5	2,660
Anthidium	33	13,278
Ashmeadiella	51	23,752
Atoposmia	23	1,837
Chelostoma	10	3,604
Coelioxys	37	7,601
Dianthidium	22	8,664
Dioxys	4	383
Heriades	11	9,325
Hoplitis	55	16,111
Lithurgopsis	6	1,519
Lithurgus		288
Megachile	116	66,676
Osmia	129	83,445
Paranthidium		783
Protosmia		2,965
Pseudoanthidium		55
Stelis	40	3,333
Trachusa	15	2,088

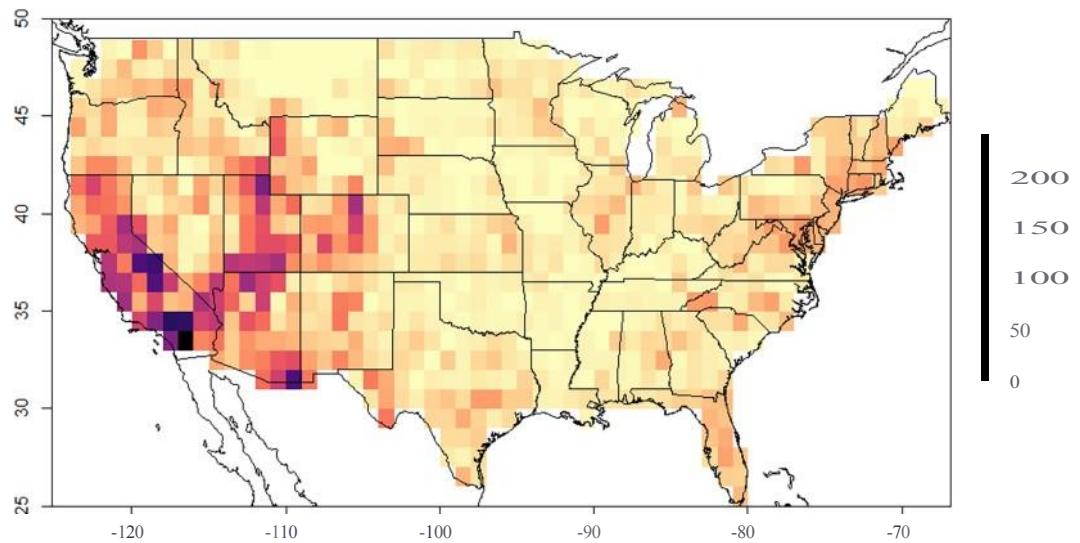
Climatic variables were extracted from WorldClim ([Fick & Hijmans, 2017](#)). All 19 climatic variables were downloaded and used in our initial analysis. Vegetation and Canopy cover were extracted from LandFire ([Rollins, 2009](#)). The LF EVT 14 (LandFire Existing Vegetation Type 2014) dataset was used to distinguish vegetation habitat classification effects on richness. Locality of the bee occurrence records determined the extraction point information from LandFire and WorldClim. All data was upscaled to a 60 x 60 km (12,100 km<sup>2</sup>) resolution; For WorldClim data we took the average among cells and for LandFire data we took the majority values for each cell to scale up to a coarser resolution, using methods developed by [Gann \(2019\)](#) LandFire land cover classifications were used in two resolutions; firstly, we used the EVT\_LF group, which splits landcover into eight distinct land cover categories, such as tree, agriculture, shrubland. Secondly, for those points that occurred in the "tree" land cover type, we used the EVT\_PHY land cover category, which has finer land cover classifications. This group included 31 total land cover types, seven of which occurred in the tree cover type ([Supplemental 1](#)). Flowering plant richness was roughly assigned to regions based on [Barthlott et al. \(2007\)](#) data, and since no large-scale data for the United States exists, methods for assigning flower diversity to regions were based on [Bystríková et al. \(2018\)](#). Additionally, tree diversity was derived from [Wilson, Lister & Riemann \(2012\)](#) data sets and upscaled to the same resolutions as the other factors.

## Richness map

Occurrence data records were aggregated to a 60 x 60 km resolution raster. Duplicate species data points that fell into the same 60 x 60 km squares were aggregated to one point (keeping just richness values). Rasters were generated by rasterizing the occurrence records using the raster package in R ([Hijmans & van Etten, 2014](#)). This meant that for each grid cell, an occupied value was only included if one or more occurrence records were found within that cell. Rasters were then summed together via RasterStack to create a map of overall observed richness for the entire Megachilidae family. Due to sampling biases around roads, developed areas and areas that are home to large museum collections ([Cobb et al., 2019](#)), a 60 x 60 km spatial resolution was selected. Finer resolutions increased the number of grid cells that have zero points and coarser resolutions compromised the habitat preferences. Even while using a coarse resolution, roughly half of the grid cells had no species recorded. However, despite this resolution selection there were still a number of pixels that have less than 10 species. Any pixel that had less than ten species (presumably due to sampling bias) were removed from the data set. Unfortunately, due to sampling incompleteness, many of these zero values occurred in the Midwest and Nevada. To account for low sampling in these areas, we broke up the United States into regions, and samples the North East, Southern East, Midwest, Northwest, and Southwest to analyze richness and landcover effects only.

## Statistical analyses

To test the relationship between climate variables and richness we ran a generalized linear model with a negative binomial distribution (GLMnb) that included all eight of the selected climate variables. Prior to this analysis we ran a correlation test to assess the relationships between the 19 WorldClim variables ([Supplemental 2](#)). If one of the bioclimatic variables was highly correlated with another one, we selected the variable we believed would have the greatest ecological impact on these taxa. Additionally, we ran another GLMnb to test the relationship between vegetation cover and Megachilidae richness. We then ran a least squares mean *post hoc* test to determine the relationship between each of the vegetation types. Regional-level centroid averages should not show autocorrelation (verified using Morans I) the models above could be used for assessing variable importance and species richness. In addition to the GLMnb test that was performed on the entire United States, we ran an additional analysis where both landcover (vegetation type) and region were compared to species richness. We then ran a least squares mean *post hoc* test to determine the relationship between each of the vegetation types for each region. We did not compare regions to one another, only vegetation type within a region. We performed a non-metric multidimensional scaling ordination (NMDS) using a Bray-Curtis distance matrix to visualize differences in species among land cover types. All analyses were done in R 4.0.3 ([R Core Team, 2013](#)) using packages MASS for GLMnb ([Ripley et al., 2013](#)), package Corrplot for correlation analysis ([Wei et al., 2011](#)) and Vegan for NMDS ([Dixon, 2003](#)).



**Figure 1** 60 x 60 km resolution (3,600 km<sup>2</sup>) of Megachilidae species diversity across the lower 48 United States. Areas with high diversity are indicated by purple to black, while those areas with low diversity are indicated by light yellow or orange colors. [Full-size figure](https://doi.org/10.7717/peerj.16145/fig-1) DOI: [10.7717/peerj.16145/fig-1](https://doi.org/10.7717/peerj.16145/fig-1)

## RESULTS

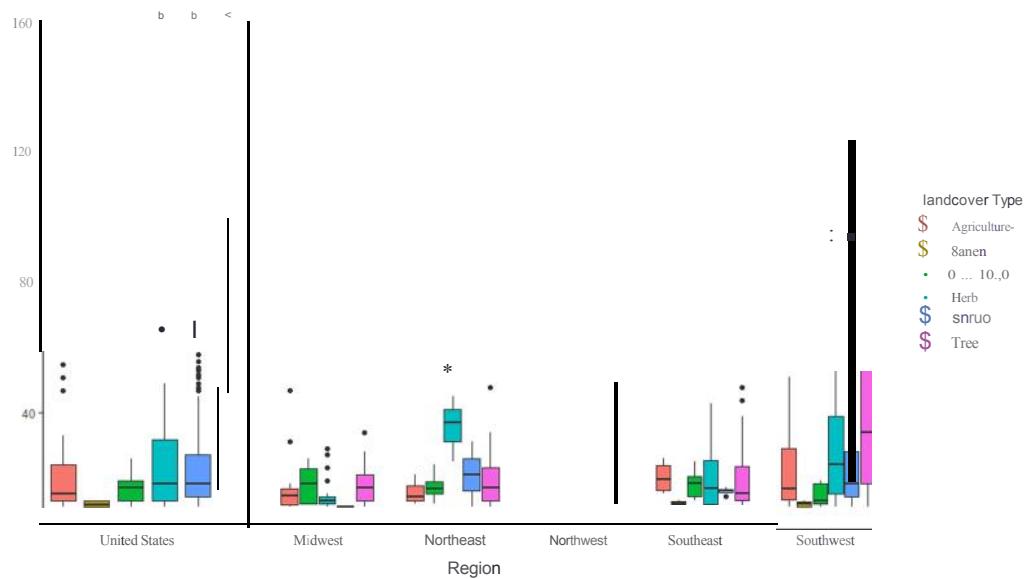
There was one hotspot of biodiversity identified in the United States for Megachilidae bees. The southwestern United States by far had the greatest species richness over any other area in the US (Fig. 1). Average richness in the southwest was  $43.5 \pm 26.12$  species, while in the southeastern US it was  $19.03 \pm 8.64$  species, northeast was  $19.36 \pm 8.12$ , Midwest was  $16.81 \pm 7.1$  species and northwest was  $21.59 \pm 12.43$  species per 60 x 60 km area. The greatest species richness was in southern California near Los Angeles with 253 species for the 60 x 60 km area. Higher elevation mountain regions, including the Sierra Nevada Mountain Range, the Colorado Plateau and the Wasatch Mountains were the most diverse areas within the western region and overall United States.

Richness was predicted by three factors: land cover (*i.e.*, tree cover), climate, and flowering plants (Table 2). Land cover type was the most correlated environmental factor in explaining bee richness ( $F = 28.57$ ,  $p < 0.001$ ). Areas that were classified by forested habitats (trees) had twice as much diversity than any other habitat with  $44.75 \pm 25.53$  species per given pixel (Fig. 2). Shrub and herb dominated habitats had the second highest number of species rich pixels with roughly  $25.03 \pm 17.56$  and  $27.10 \pm 23.48$  average species per pixel, respectively. Herb and shrub habitats were not significantly different from one another ( $p = 0.185$ ). Agriculture, developed, barren and sparse habitats had averages between 10-15 species per pixel; none of these land cover types were significantly different from one another (Table S2). Despite land cover type being highly correlated with Megachilidae richness, overall tree diversity did not have a significant effect in predicting Megachilidae richness ( $z = 1.321$ ,  $p = 0.187$ ).

The southwestern United States drove the pattern for having the most species in treed/forested habitats. In the midwest, the northwest and southeast none of the landcover types were different from one another (Table S2, Fig. 2). All habitats had an equal number

**Table 2** Key environmental predictors of Megachilidae richness and general linear models fitted to the species richness of Megachilidae to the United States extent.

Variable	Acronym	Units	Source	z-value	p-value
Annual mean temperature	BIOi	°C	BIOCLIM ( <i>Hijmans et al., 2005</i> )	4.559	<0.001
Mean diurnal range	BI02	°C	BIOCLIM ( <i>Hijmans et al., 2005</i> )	10.45	<0.001
Max temperature of the warmest month	BIOS	°C	BIOCLIM ( <i>Hijmans et al., 2005</i> )	0.883	0.377
Mean temperature of the wettest month	BIOS	°C	BIOCLIM ( <i>Hijmans et al., 2005</i> )	-11.13	<0.001
Mean temperature of the driest quarter	BI09	°C	BIOCLIM ( <i>Hijmans et al., 2005</i> )	23.15	<0.001
Annual mean precipitation	BIO12	mm	BIOCLIM ( <i>Hijmans et al., 2005</i> )	-12.57	<0.001
Precipitation seasonality	BIOIS	mm	BIOCLIM ( <i>Hijmans et al., 2005</i> )	8.774	<0.001
Precipitation of the warmest quarter	BIOIS	mm	BIOCLIM ( <i>Hijmans et al., 2005</i> )	-16.65	<0.001
LandFire existing vegetation type	EVT_LF	Categorical classification	LandFire 2016	28.57	<0.001
LandFire existing vegetation type subclass forest types	EVT_SBCLS	Categorical classification	LandFire 2016	13.68	<0.001
United States tree diversity	TreeDiv	Number of species per grid cell	adapted from <i>Wilson, Lister &amp; Riemann (2012)</i>	1.321	0.187
United States herbaceous flowering plant diversity	PlantDiv	Number of species per grid cell	adapted from <i>Barthlott et al (2007)</i>	4.742	<0.001

**Figure 2** The average Megachilidae richness for each LandFire classification type (EVT\_LF). Letters and/or asterisk denote significance. [Full-size](#) DOI: [10.7717/peerj.16145/fig-2](https://doi.org/10.7717/peerj.16145/fig-2)

of species per pixel. However, the southwestern United States had at least double the species in treed habitats than it did in agriculture, barren, developed, and shrub habitat ( $F = 30.702, p < 0.001$ , Fig. 2). Additionally, shrub and herbaceous habitats had more richness than agriculture, barren and developed lands.

All eight climatic variables selected showed a significant effect in predicting richness except for maximum temperature of the warmest quarter ( $z = 0.883, p = 0.337$ ). Mean temperature of the driest quarter explained 4x as much variance than any other climatic variable ( $z = 23.15, p < 0.001$ ). While mean temperature of the warmest quarter and precipitation seasonality were significant in the model, they explained less than 1% of the variance combined ( $z = -11.33, p < 0.001$  &  $z = 8.774, p < 0.001$  respectively). Herbaceous flowering plant diversity strongly predicted Megachilidae richness; as flowering plant diversity increased, so did Megachilidae diversity ( $z = 4.742, p < 0.001$ ). Flowering plant richness also peaked in the southwestern United States. We did not examine specific associations for this analysis.

## DISCUSSION

Our results indicate that Megachilidae biodiversity has its hotspot in the southwestern United States, this is consistent with geographic patterns found by [Michener \(2007\)](#). Additionally, this is true for many bee species outside of Megachilidae, as well, since the southwestern United States is also a hotspot for bee biodiversity on a global scale ([Griswold et al., 2018](#); [Orr et al., 2020](#)). The southwest is typically associated with dry air climates that are more suitable for ground nesting bees and thus hypothesized to harbor the greatest bee diversity globally, with around 80% of those being ground nesters ([Danforth et al., 2019](#)). However for Megachilidae, where 70% of the genera are cavity nesters, the same environmental traits may not be as crucial ([Cane, Griswold & Parker, 2007](#)). Importantly, the majority of cavity nesting species in tribe Osmiini are concentrated in North America ([Praz et al., 2008](#)), while many highly related *Megachile* species in the Palearctic still nest in the ground or have diversified nesting strategies ([Danforth et al., 2004](#); [Patiny, Michez & Danforth, 2008](#)). It is hypothesized that cavity nesters were better able to survive trips across long water barriers, potentially explaining why it is mostly these Osmiini species inhabiting the new world ([Michener, 2007](#)). Therefore, some of the climate preferences and/or floral dependencies may be carried over from this lineage of soil nesting bees.

What makes United States Megachilidae so unique, however, is their strong ties to biotic nesting resources. Contrary to what [Orr et al. \(2020\)](#) found for all bee species, here we show that Megachilidae biodiversity was highly correlated with forested habitat, particularly in the southwestern United States, suggesting that these areas of more heterogeneous landscapes could be driving species diversification for this group ([Hedtke, Patiny & Danforth, 2013](#)). Groups such as Colletidae have shown stronger correlations with floral resources and climate than they have nesting requirements ([Bystrakova et al., 2018](#)). Other groups like *Bombus* also seem to be more strongly tied to floral resources for driving biogeographical patterns ([Williams, Lonsdorf & Ward, 2014](#)). Our study is not the first to show that bees may be restricted in distribution due to nesting resources or habitats ([Gordon, 2000](#); [Romero, Ornosa & Vargas, 2020](#)). Additionally, traditional range models only factor in climate analyses however it is likely that landscape/ habitat play a large role in determining species geographic ranges ([Graham & Maclean, 2018](#); [Palma et al., 2016](#)). [McCabe et al. \(2020\)](#) found that, along an elevation gradient in the southwest US Megachilidae were distributed in higher cooler elevations. These higher elevation

environments also correspond with habitats that are forested. This pattern has only been documented in one other bee group, *Bombus* (Williams *et al.*, 2014). However, *Bombus* is a social ground nesting species and has much lower diversity than the Megachilidae family.

Megachilidae showed a strong preference to floral diversity, as would be expected. Increased plant richness and diversity in an area, which likely also equates to increased floral abundance, can lead to greater floral resource availability (Blaauw & Isaacs, 2014; Crone & Williams, 2016). However, it was impossible to link specific flora types with drivers of Megachilidae biodiversity since many host plant associations are unknown. Even where host plant specialization has been well studied, it is unclear how the distribution of those plants plays a role in the bee species distributions (Michener, 2007). Although there are limitations for plant-insect interactions, open forest habitat with understory vegetation or open meadows with floral resources promotes bee diversity (Mola *et al.*, 2021). Likewise, areas of increasing canopy cover in the southwest have been shown to decrease bee species richness, but Megachilidae species richness increases (McCabe *et al.*, 2019).

Nationwide, climate analysis showed a trend towards warmer and drier environments. Megachilidae richness increased with increasing mean annual temperatures and decreasing mean annual precipitation, which is consistent with trends found in other bee species (Michener, 2007; Orr *et al.*, 2020). However, analyses with the more nuanced climate variables revealed even more informative patterns for Megachilid species. Mean temperature of the wettest quarter was important in predicting richness; as mean temperatures decreased, Megachilidae richness increased. This is likely because species in the Megachilidae family often require a minimum number of cold days in order to complete diapause (Bosch, Sgolastra & Kemp, 2010) and the number of cool days during overwintering can directly influence their emergence timing (Sgolastra & Kemp, 2010). Unfortunately, future warming will likely change the distribution patterns of many Megachilidae species. McCabe, Asian & Cobb (2022) found that for a community of cavity nesting species in Ariwna, warming overwintering temperatures negatively affected emergence rates. Although the species were able to withstand cooler temperatures in higher elevations, there were no nesting resources available in those environments.

Megachilidae may have more restrictions when adapting to climate change due to their associations with cavity nest availability (*e.g.*, dead and down wood, pithy stems, *etc.*) and floral resources availability (Cane, Griswold & Parker, 2007). This dependency on nesting cavities adds an additional level of complexity when asking the question of how they will respond to climate change. In fact, it is estimated that 98% of Megachilidae depend on some type of non-floral, biotic resource (Requier & Leonhardt, 2020). Climate change is likely to only exacerbate the local levels of tree die-off in forested habitat, and in some instances, severe drought events can lead to over 90% tree die-off (Breshears *et al.*, 2005). Additionally, an increase in wildfire frequency can lead to complete change in forest stands (Flannigan, Stocks & Wotton, 2000). It is unknown how these changes in forest structure will lead to subsequent changes in pollinator communities, especially within Megachilidae. Although little is known about climate change and soil composition, it is unlikely that ground nesting bees will have the same dramatic loss of nesting habitat under future climate scenarios (Allen, Singh & Dalal, 2011; Rinot *et al.*, 2019). This uncertainty furthers

the need to quantify the amount of nesting resources needed for Megachilidae within local communities.

Understanding what factors influence the distribution and biogeography of this taxa is important for future conservation efforts. With megachilids in particular, it may not only be the direct effect of warming temperature that causes shifts in species distributions. These species have strong ties to their host plants which could cause phenological mismatch in the future if the bees and plants are not tracking climate in the same manner (*Bartomeus et al., 2018; Biesmeijer et al., 2006*). Future conservation efforts for this family should consider altogether the direct effects of warming temperature, availability of floral resources, and availability of nesting resources.

## ADDITIONAL INFORMATION AND DECLARATIONS

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### Competing Interests

The authors declare that they have no competing interests.

### Author Contributions

- Lindsie M. McCabe conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Paige Chesshire performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Neil S. Cobb conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.

### Data Availability

The following information was supplied regarding data availability:

The data is available at figshare: McCabe, Lindsie (2023). Megachilidae specimen records. figshare. Dataset. [https://figshare.com/articles/dataset/Megachilidae\\_specimen\\_records/24084942/1](https://figshare.com/articles/dataset/Megachilidae_specimen_records/24084942/1).

### Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.1614S#supplemental-information>.

## REFERENCES

Allen DE, Singh BP, Dalal RC. 2011. Soil health indicators under climate change: a review of current knowledge. *Soil Health and Climate Change* 29:25-45 DOI [10.1007/978-3-642-20256-8](https://doi.org/10.1007/978-3-642-20256-8).

**Barthlott W, Hostert A, Kier G, Kueper W, Kreft H, Mutke J, Rafiqpoor MD, Sommer JH. 2007.** Geographic patterns of vascular plant diversity at continental to global scales (Geographische Muster der Gefäßpflanzenvielfalt im kontinentalen und globalen Maßstab). *Erdkunde* 305-315.

**Bartomeus I, Ascher JS, Wagner D, Danforth BN, Colla S, Kornbluth S, Winfree R. 2011.** Climate-associated phenological advances in bee pollinators and bee-pollinated plants. *Proceedings of the National Academy of Sciences of the United States of America* 108(51):20645-20649 DOI [10.1073/pnas.1115559108](https://doi.org/10.1073/pnas.1115559108).

**Bartomeus I, Park MG, Gibbs J, Danforth BN, Lakso AN, Winfree R. 2013.** Biodiversity ensures plant-pollinator phenological synchrony against climate change. *Ecology Letters* 16(11):1331-1338 DOI [10.1111/ele.12170](https://doi.org/10.1111/ele.12170).

Bartomeus I, Stavert J, Ward D, Aguado O. 2018. Historical collections as a tool for assessing the global pollination crisis. *Philosophical Transactions of the Royal Society B* 374(1763):20170389 DOI [10.1098/rstb.2017.0389](https://doi.org/10.1098/rstb.2017.0389).

**Biesmeijer JC, Roberts S, Reemer M, Ohlemüller R, Edwards M, Peeters T, Schaffers A, Potts S, Kleukers R, Thomas C. 2006.** Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands. *Science* 313(5785):351-354 DOI [10.1126/science.1127863](https://doi.org/10.1126/science.1127863).

Blaauw BR, Isaacs R. 2014. Larger patches of diverse floral resources increase insect pollinator density, diversity, and their pollination of native wildflowers. *Basic and Applied Ecology* 15(8):701-711 DOI [10.1016/j.baae.2014.10.001](https://doi.org/10.1016/j.baae.2014.10.001).

Bosch J, Kemp WP. 2002. Developing and establishing bee species as crop pollinators: the example of Osmia spp. (Hymenoptera: Megachilidae) and fruit trees. *Bulletin of Entomological Research* 92(1):3-16 DOI [10.1079/BER2001139](https://doi.org/10.1079/BER2001139).

Bosch J, Maeta Y, Rust R. 2001. A phylogenetic analysis of nesting behavior in the genus Osmia (Hymenoptera: Megachilidae). *Annals of the Entomological Society of America* 94(4):617-627 DOI [10.1603/0013-8746\(2001\)094\(0617:APAONB\)2.0.CO;2](https://doi.org/10.1603/0013-8746(2001)094(0617:APAONB)2.0.CO;2).

Bosch J, Sgolastra F, Kemp WP. 2010. Timing of eclosion affects diapause development, fat body consumption and longevity in Osmia lignaria, a univoltine, adult-wintering solitary bee. *Journal of Insect Physiology* 56(12):1949-1957 DOI [10.1016/j.jinsphys.2010.08.017](https://doi.org/10.1016/j.jinsphys.2010.08.017).

**Bossert S, Wood TJ, Patiny S, Michez D, Almeida EA, Minckley RL, Murray EA. 2022.** Phylogeny, biogeography and diversification of the mining bee family Andrenidae. *Systematic Entomology* 47(2):283-302 DOI [10.1111/syen.12530](https://doi.org/10.1111/syen.12530).

Boyle NK, Pitts-Singer TL. 2019. Assessing blue orchard bee (Osmia lignaria) propagation and pollination services in the presence of honey bees (Apis mellifera) in Utah tart cherries. *PeerJ* 7(6):e7639 DOI [10.7717/peerj.7639](https://doi.org/10.7717/peerj.7639).

**Breshears DD, Cobb NS, Rich PM, Price KP, Allen CD, Bailee RG, Romme WH, Kastens JH, Floyd ML, Belnap J, Anderson JJ, Myers OB, Meyer CW. 2005.** Regional vegetation die-off in response to global-change-type drought. *Proceedings of the National Academy of Sciences of the United States of America* 102(42):15144-15148 DOI [10.1073/pnas.0505734102](https://doi.org/10.1073/pnas.0505734102).

**Brown MJ, Paxton RJ. 2009.** The conservation of bees: a global perspective. *Apidologie* 40(3):410-416 DOI [10.1051/apido/2009019](https://doi.org/10.1051/apido/2009019).

**Bystríková N, Griswold T, Ascher JS, Kuhlmann M. 2018.** Key environmental determinants of global and regional richness and endemism patterns for a wild bee subfamily. *Biodiversity and Conservation* 27(2):287-309 DOI [10.1007/s10531-017-1432-7](https://doi.org/10.1007/s10531-017-1432-7).

Cane JH, Griswold T, Parker FD. 2007. Substrates and materials used for nesting by North American Osmia bees (Hymenoptera: Apiformes: Megachilidae). *Annals of the Entomological Society of America* 100(3):350-358 DOI [10.1603/0013-8746\(2007\)100\[350:SAMUFN\]2.0.CO;2](https://doi.org/10.1603/0013-8746(2007)100[350:SAMUFN]2.0.CO;2).

**Cavender-Bares J, Kozak KH, Fine PV, Kembel SW. 2009.** The merging of community ecology and phylogenetic biology. *Ecology Letters* 12(7):693-715 DOI [10.1111/j.1461-0248.2009.01314.x](https://doi.org/10.1111/j.1461-0248.2009.01314.x).

Chessire PR, Fischer EE, Dowdy NJ, Griswold TL, Hughes AC, Orr MC, Ascher JS, Guzman LM, Hung KLJ, Cobb NS. 2023. Completeness analysis for over 3000 United States bee species identifies persistent data gap. *Ecography* 2023(5):138 DOI [10.1111/ecog.06584](https://doi.org/10.1111/ecog.06584).

Cobb NS, Gall LF, Zaspel JM, Dowdy NJ, McCabe LM, Kawahara AY. 2019. Assessment of North American arthropod collections: prospects and challenges for addressing biodiversity research. *PeerJ* 7:e8086 DOI [10.7717/peerj.8086](https://doi.org/10.7717/peerj.8086).

**Corbet SA, Fussell M, Ake R, Fraser A, Gunson C, Savage A, Smith K. 1993.** Temperature and the pollinating activity of social bees. *Ecological Entomology* 18(1):17-30 DOI [10.1111/j.1365-2311.1993.tb01075.x](https://doi.org/10.1111/j.1365-2311.1993.tb01075.x).

**Crone EE, Williams NM. 2016.** Bumble bee colony dynamics: quantifying the importance of land use and floral resources for colony growth and queen production. *Ecology Letters* 19(4):460-468 DOI [10.1111/ele.12581](https://doi.org/10.1111/ele.12581).

Danforth BN, Brady SG, Sipes SD, Pearson A. 2004. Single-copy nuclear genes recover cretaceous-age divergences in bees. *Systematic Biology* 309-326 DOI [10.1080/10635150490423737](https://doi.org/10.1080/10635150490423737).

Danforth BN, Cardinal S, Praz C, Almeida EA, Michez D. 2013. The impact of molecular data on our understanding of bee phylogeny and evolution. *Annual Review of Entomology* 58(1):57-78 DOI [10.1146/annurev-ento-120811-153633](https://doi.org/10.1146/annurev-ento-120811-153633).

Danforth BN, Minckley RL, Neff JL, Fawcett F. 2019. *The solitary bees: biology, Evolution, Conservation*: Princeton University Press.

**Dixon P.** 2003. VEGAN, a package of R functions for community ecology. *Journal of Vegetation Science* 14(6):927-930 DOI [10.1111/j.1654-1103.2003.tb02228.x](https://doi.org/10.1111/j.1654-1103.2003.tb02228.x).

**Dorey JB, Fischer EE, Chessire PR, Nava-Bolanos A, O'Reilly RL, Bossert S, Collins SM. 2023.** BeeDC: an R package and globally synthesised and flagged bee occurrence dataset. *bioRxiv* 9: e70289 DOI [10.1101/2023.06.30.547152](https://doi.org/10.1101/2023.06.30.547152).

Eickwort GC, Matthews RW, Carpenter J. 1981. Observations on the nesting behavior of Megachile rubi and M. texana with a discussion of the significance of soil nesting in the evolution of megachilid bees (Hymenoptera: Megachilidae). *Journal of the Kansas Entomological Society* 54:557-570.

Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* 37(12):4302-4315 DOI [10.1002/joc.5086](https://doi.org/10.1002/joc.5086).

**Flannigan MD, Stocks BJ, Wotton BM. 2000.** Climate change and forest fires. *Science of the Total Environment* 262(3):221-229 DOI [10.1016/S0048-9697\(00\)00524-6](https://doi.org/10.1016/S0048-9697(00)00524-6).

**Gann D. 2019.** Quantitative spatial upscaling of categorical information: the multi-dimensional grid-point scaling algorithm. *Methods in Ecology and Evolution* 10(12):2090-2104.

Global Biodiversity Information Facility. 2020. *Occurrence Download*. The Global Biodiversity Information Facility.

Gonzalez VH, Griswold T, Praz CJ, Danforth BN. 2012. Phylogeny of the bee family Megachilidae (Hymenoptera: Apoidea) based on adult morphology. *Systematic Entomology* 37(2):261-286 DOI [10.1111/j.1365-3113.2012.00620.x](https://doi.org/10.1111/j.1365-3113.2012.00620.x).

Gordon DM. 2000. Plants as indicators of leafcutter bee (Hymenoptera: Megachilidae) nest habitat in coastal dunes. *Pan-Pacific Entomologist* 76(4):219-233.

**Graham KK, Maclean MG. 2018.** Presence-only modeling is ill-suited for a recent generalist invader, *Anthidium manicatum*. *Ecological Indicators* 89(2):56-62  
[DOI 10.1016/j.ecolind.2018.02.002](https://doi.org/10.1016/j.ecolind.2018.02.002).

**Griswold T, Ascher JS, Ikerd H, Sagot P, Vandame R, Orr MC, Portman Z, Wilson JS. 2018.** The native bees of the Americas: patterns across the western hemisphere. In: *Entomology 2018*. ESA.

Hedtke SM, Patiny S, Danforth BN. 2013. The bee tree of life: a supermatrix approach to apoid phylogeny and biogeography. *BMC Evolutionary Biology* 13(1):1-13  
[DOI 10.1186/1471-2148-13-138](https://doi.org/10.1186/1471-2148-13-138).

Hijmans RJ, van Etten J. 2014. Raster: geographic data analysis and modeling. R package version 2.

Hijmans RJ, Cameron S, Parra J, Jones P, Jarvis A, Richardson K. 2005. WorldClim-global climate data. *Very High Resolution Interpolated Climate Surfaces for Global Land Areas*.

Kergoat GJ, Condamine FL, Toussaint EFA, Capdeville-Dulac C, Clamens A-L, Barbut J, Goldstein PZ, Ru BL. 2018. Opposite macroevolutionary responses to environmental changes in grasses and insects during the Neogene grassland expansion. *Nature Communications* 9(1):5089 [DOI 10.1038/s41467-018-07537-8](https://doi.org/10.1038/s41467-018-07537-8).

Klein A-M, Vaissiere BE, Cane JH, Steffan-Dewenter I, Cunningham SA, Kremen C, Tscharntke T. 2006. Importance of pollinators in changing landscapes for world crops. *Proceedings of the Royal Society B: Biological Sciences* 274(1608):303-313  
[DOI 10.1098/rspb.2006.3721](https://doi.org/10.1098/rspb.2006.3721).

Klein A-M, Vaissiere BE, Cane JH, Steffan-Dewenter I, Cunningham SA, Kremen C, Tscharntke T. 2007. Importance of pollinators in changing landscapes for world crops. *Proceedings of the Royal Society B: Biological Sciences* 274(1608):303-313  
[DOI 10.1098/rspb.2006.3721](https://doi.org/10.1098/rspb.2006.3721).

**Litman JR, Danforth BN, Eardley CD, Praz CJ. 2011.** Why do leafcutter bees cut leaves? New insights into the early evolution of bees. *Proceedings of the Royal Society B: Biological Sciences* 278:3593-3600 [DOI 10.1098/rspb.2011.0365](https://doi.org/10.1098/rspb.2011.0365).

**Martins RP, de Almeida DA. 1994.** Is the bee, *Megachile assumptionis* (Hymenoptera: Megachilidae), a cavity-nesting specialist? *Journal of Insect Behavior* 7:759-765  
[DOI 10.1007/BF01997444](https://doi.org/10.1007/BF01997444).

McCabe LM, Asian CE, Cobb NS. 2022. Decreased bee emergence along an elevation gradient: implications for climate change revealed by a transplant experiment. *Ecology* 103:e03598  
[DOI 10.1002/ecy.3598](https://doi.org/10.1002/ecy.3598).

**McCabe LM, Chesshire PR, Smith DR, Wolf A, Gibbs J, Griswold TL, Wright KW, Cobb NS. 2020.** Bee species checklist of the San Francisco Peaks, Arizona. *Biodiversity Data Journal* 8: e49285 [DOI 10.3897/BDJ.8.e49285](https://doi.org/10.3897/BDJ.8.e49285).

McCabe LM, Colella E, Chesshire P, Smith D, Cobb NS. 2019. The transition from bee-to-fly dominated communities with increasing elevation and greater forest canopy cover. *PLOS ONE* 14(6):e0217198 [DOI 10.1371/journal.pone.0217198](https://doi.org/10.1371/journal.pone.0217198).

Memmott J, Carvell C, Pywell RF, Craze PG. 2010. The potential impact of global warming on the efficacy of field margins sown for the conservation of bumble-bees. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* 365(1549):2071-2079  
[DOI 10.1098/rstb.2010.0015](https://doi.org/10.1098/rstb.2010.0015).

Michener CD. 2007. *The bees of the world*. Baltimore, Maryland: John Hopkins University Press.

Mikat M, Matouskova E, Straka J. 2021. Nesting of *Ceratina nigrolabiata*, a biparental bee. *Scientific Reports* 11(1):5026 DOI [10.1038/s41598-021-83940-4](https://doi.org/10.1038/s41598-021-83940-4).

Mola JM, Hemberger J, Kochanski J, Richardson LL, Pearse IS. 2021. The importance of forests in bumble bee biology and conservation. *Bioscience* 71(12):1234-1248.

Ollerton J, Winfree R, Tarrant S. 2011. How many flowering plants are pollinated by animals? *Oikos* 120(3):321-326 DOI [10.1111/j.1600-0706.2010.18644.x](https://doi.org/10.1111/j.1600-0706.2010.18644.x).

Orr MC, Hughes AC, Chesters D, Pickering J, Zhu C-D, Ascher JS. 2020. Global patterns and drivers of bee distribution. *Current Biology* 31(3):451-458 DOI [10.1016/j.cub.2020.10.053](https://doi.org/10.1016/j.cub.2020.10.053).

**Palma DA, Abrahamczyk S, Marcelo AA, Albrecht M, Basset Y, Bates A, Blake RJ. 2016.**  
Predicting bee community responses to land-use changes: effects of geographic and taxonomic biases. *Scientific Reports* 6:31153 DOI [10.1038/srep31153](https://doi.org/10.1038/srep31153).

**Patiny S, Michez D, Danforth B. 2008.** Phylogenetic relationships and host-plant evolution within the basal clade of Halictidae (Hymenoptera, Apoidea). *Cladistics* 24(3):255-269 DOI [10.1111/j.1096-0031.2007.00182.x](https://doi.org/10.1111/j.1096-0031.2007.00182.x).

**Pitts-Singer TL, Artz DR, Peterson SS, Boyle NK, Wardell GI. 2018.** Examination of a managed pollinator strategy for almond production using *Apis mellifera* (Hymenoptera: Apidae) and *Osmia lignaria* (Hymenoptera: Megachilidae). *Environmental Entomology* 47(2):364-377 DOI [10.1093/ee/nvy009](https://doi.org/10.1093/ee/nvy009).

Pitts-Singer TL, Cane JH. 2011. The alfalfa leafcutting bee, *Megachile rotundata*: the world's most intensively managed solitary bee. *Annual Review of Entomology* 56(1):221-237 DOI [10.1146/annurev-ento-120709-144836](https://doi.org/10.1146/annurev-ento-120709-144836).

Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE. 2010. Global pollinator declines: trends, impacts and drivers. *Trends in Ecology & Evolution* 25(6):345-353 DOI [10.1016/j.tree.2010.01.007](https://doi.org/10.1016/j.tree.2010.01.007).

Potts SG, Ngo HT, Biesmeijer JC, Breeze TD, Dicks LV, Garibaldi LA, Hill R, Settele J, Vanbergen A. 2016. The assessment report of the intergovernmental science-policy platform on biodiversity and ecosystem services on pollinators, pollination and food production. Available at [https://nora.nerc.ac.uk/id/eprint/5192271/ll/individual\\_chapters\\_pollination\\_20170305.pdf](https://nora.nerc.ac.uk/id/eprint/5192271/ll/individual_chapters_pollination_20170305.pdf)

Praz CJ, Millier A, Danforth BN, Griswold TL, Widmer A, Dom S. 2008. Phylogeny and biogeography of bees of the tribe Osmiini (Hymenoptera: Megachilidae). *Molecular Phylogenetics and Evolution* 49(1):185-197 DOI [10.1016/j.ympev.2008.07.005](https://doi.org/10.1016/j.ympev.2008.07.005).

R Core Team. 2013. *R: a language and environment for statistical computing*. Vienna: The R Foundation for Statistical Computing. Available at <http://www.R-project.org/>.

Requier F, Leonhardt SD. 2020. Beyond flowers: including non-floral resources in bee conservation schemes. *Journal of Insect Conservation* 24(1):5-16 DOI [10.1007/s10841-019-00206-1](https://doi.org/10.1007/s10841-019-00206-1).

Rinot O, Levy GJ, Steinberger Y, Svoray T, Eshel G. 2019. Soil health assessment: a critical review of current methodologies and a proposed new approach. *Science of the Total Environment* 648(6):1484-1491 DOI [10.1016/j.scitotenv.2018.08.259](https://doi.org/10.1016/j.scitotenv.2018.08.259).

Ripley B, Venables B, Bates DM, Hornik K, Gebhardt A, Firth D, Ripley MB. 2013. Package 'mass'. *Cran r* 538:113-120.

Rollins MG. 2009. LANDFIRE: a nationally consistent vegetation, wildland fire, and fuel assessment. *International Journal of Wildland Fire* 18(3):235-249 DOI [10.1071/WF08088](https://doi.org/10.1071/WF08088).

Romero D, Omosa C, Vargas P. 2020. Where and why? Bees, snail shells and climate: distribution of rhodanthidium (Hymenoptera: Megachilidae) in the Iberian Peninsula. *Entomological Science* 23(3):256-270 DOI [10.1111/ens.12420](https://doi.org/10.1111/ens.12420).

Sahli HF, Conner JK. 2006. Characterizing ecological generalization in plant-pollinationsystems. *Oecologia* 148(3):365-372 DOI [10.1007/s00442-006-0396-l](https://doi.org/10.1007/s00442-006-0396-l).

Sampson BJ, Cane JH. 2000. Pollination efficiencies of three bee (Hymenoptera: Apoidea) species visiting rabbiteye blueberry. *Journal of Economic Entomology* 93(6):1726-1731 DOI [10.1603/0022-0493-93.6.1726](https://doi.org/10.1603/0022-0493-93.6.1726).

**SCAN. 2019.** The Symbiota collections of Arthropods Network (SCAN) serves specimen occurrence records and images from North American arthropod collections. Available at <https://scan-bugs.org/>.

**Sedivy C, Dorn S, Müller A. 2013.** Evolution of nesting behaviour and kleptoparasitism in a selected group of osmiine bees (Hymenoptera: Megachilidae). *Biological Journal of the Linnean Society* 108(2):349-360 DOI [10.1111/j.1095-8312.2012.02024.x](https://doi.org/10.1111/j.1095-8312.2012.02024.x).

**Sgolastra F, Kemp WP. 2010.** Timing of eclosion affects diapause development, fat body consumption and longevity in *Osmia lignaria*, a univoltine, adult-wintering solitary bee. *Journal of Insect Physiology* 56(12):1949-1957 DOI [10.1016/j.jinsphys.2010.08.017](https://doi.org/10.1016/j.jinsphys.2010.08.017).

Tews J, Esther A, Milton SJ, Jeltsch F. 2006. Linking population model with an ecosystem model: assessing the impact of land use and climate change on savanna shrub cover dynamics. *Ecological Modelling* 195(3-4):219-228 DOI [10.1016/j.ecolmodel.2005.11.025](https://doi.org/10.1016/j.ecolmodel.2005.11.025).

**Toussaint EFA, Condamine FL, Kergoat GJ, Capdevielle-Dulac C, Barbut J, Silvain J-F, Le Ru BP. 2012.** Palaeoenvironmental shifts drove the adaptive radiation of a noctuid stemborer tribe (Lepidoptera, Noctuidae, Apameini) in the Miocene. *PLOS ONE* 7(7):e41377 DOI [10.1371/journal.pone.0041377](https://doi.org/10.1371/journal.pone.0041377).

Wei T, Simko V, Levy M, Xie Y, Jin Y, Zemla J. 2017. Package 'corrplot'. *Statistician* 56:e24.

Weiher E, Freund D, Bunton T, Stefanski A, Lee T, Bentivenga S. 2011. Advances, challenges and a developing synthesis of ecological community assembly theory. *Philosophical Transactions of the Royal Society B: Biological Sciences* 366(1576):2403-2413 DOI [10.1098/rstb.2011.0056](https://doi.org/10.1098/rstb.2011.0056).

**Westrich P. 2007.** Ein weiterer Beleg fir den Bivoltinismus und das Wirt Parasit-Verhältnis von *Andrena bicolor* (Fabricius 1804) und *Nomada fabriciana* (Kirby 1802) (Hym. Apidae). *Linzer Biologische Beiträge* 38:919-923.

Williams NM, Lonsdorf EV, Ward K. 2014. Selection and testing of plant species for pollinator habitat, from decision analysis to validation. In: *Entomological Society of America Annual Meeting*. vol. 201: Portland, OR.

Williams PH, Thorp RW, Richardson LL, Colla SR. 2014. *Bumble bees of North America: an identification guide*. Princeton, New Jersey: Princeton University Press.

Wilson BT, Lister AJ, Riemann RI. 2012. A nearest-neighbor imputation approach to mapping tree species over large areas using forest inventory plots and moderate resolution raster data. *Forest Ecology and Management* 271(May):182-198 DOI [10.1016/j.foreco.2012.02.002](https://doi.org/10.1016/j.foreco.2012.02.002).

Young BE, Schweitzer DF, Hammerson GA, Sears NA, Ormes MF, Tomaino AO. 2016. North American leafcutter bees.