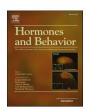
ELSEVIER

Contents lists available at ScienceDirect

### Hormones and Behavior

journal homepage: www.elsevier.com/locate/yhbeh





# Conditioned preferences: Gated by experience, context, and endocrine systems $^*$

Patrick K. Monari <sup>a,\*</sup>, Emma R. Hammond <sup>a</sup>, Xin Zhao <sup>a</sup>, Alyse N. Maksimoski <sup>b</sup>, Radmila Petric <sup>a,c</sup>, Candice L. Malone <sup>a</sup>, Lauren V. Riters <sup>b</sup>, Catherine A. Marler <sup>a,b,\*\*</sup>

- <sup>a</sup> University of Wisconsin-Madison, Department of Psychology, Madison, WI, USA
- <sup>b</sup> University of Wisconsin-Madison, Department of Integrative Biology, Madison, WI, USA
- <sup>c</sup> Institute for the Environment, University of North Carolina Chapel Hill, Chapel Hill, NC, USA

### ARTICLE INFO

# Keywords: Conditioned place preference Conditioned social preference Social conditioned place preference Naturalistic behavior Plasticity Testosterone Estradiol Neuropeptide Opioid Dopamine

### ABSTRACT

Central to the navigation of an ever-changing environment is the ability to form positive associations with places and conspecifics. The functions of location and social conditioned preferences are often studied independently, limiting our understanding of their interplay. Furthermore, a de-emphasis on natural functions of conditioned preferences has led to neurobiological interpretations separated from ecological context. By adopting a naturalistic and ethological perspective, we uncover complexities underlying the expression of conditioned preferences. Development of conditioned preferences is a combination of motivation, reward, associative learning, and context, including for social and spatial environments. Both social- and location-dependent reward-responsive behaviors and their conditioning rely on internal state-gating mechanisms that include neuroendocrine and hormone systems such as opioids, dopamine, testosterone, estradiol, and oxytocin. Such reinforced behavior emerges from mechanisms integrating past experience and current social and environmental conditions. Moreover, social context, environmental stimuli, and internal state gate and modulate motivation and learning via associative reward, shaping the conditioning process. We highlight research incorporating these concepts, focusing on the integration of social neuroendocrine mechanisms and behavioral conditioning. We explore three paradigms: 1) conditioned place preference, 2) conditioned social preference, and 3) social conditioned place preference. We highlight nonclassical species to emphasize the naturalistic applications of these conditioned preferences. To fully appreciate the complex integration of spatial and social information, future research must identify neural networks where endocrine systems exert influence on such behaviors. Such research promises to provide valuable insights into conditioned preferences within a broader naturalistic context.

### 1. Introduction

In navigating the complexities of natural environments, organisms face multifaceted challenges throughout their lives. Central to the navigation of such challenges is associative learning, enabling individuals to adapt to ever-changing environments that influence survivorship and reproductive success. This adaptability is informed by past experiences, incorporates current environmental factors, and facilitates predictions about future social and physical contexts. Researchers who study brains, hormones, and behavior within natural and semi-natural environments often examine the social dimensions and experiential history of an

individual, for example, the identities of social actors, the setting of the experience, and the focal individual's life and reproductive stages. Each of these elements modulates associative learning and gates the expression of behaviors in ecological contexts. However, the endocrine/neuroendocrine link between social and spatial environments, and the modulation of this connection by life and reproductive stages, represents a vast reservoir of untapped exploration. Here, we delve into essential components of associative learning leading to conditioned preferences that rely on social and locational factors in natural contexts. Our goal for this review is to link naturalistic and neuroendocrine perspectives on conditioned preferences (in particular conditioned place and social preferences) to explore the complexities of ecologically-relevant

<sup>\*</sup> Corresponding author.

<sup>\*\*</sup> Correspondence to: C.A. Marler, University of Wisconsin-Madison, Department of Psychology, Madison, WI, USA. *E-mail addresses*: monari@wisc.edu (P.K. Monari), catherine.marler@wisc.edu (C.A. Marler).

### Glossary

Conditioned place preference (CPP) An animal's preference for a particular location (i.e., conditioned stimulus, CS) in which it previously formed an association between said location and a *nonsocial* stimulus (i.e., unconditioned stimulus, US). The typical CPP experiment involves substances of abuse (Fig. 1B, top).

Social conditioned place preference (SCPP) A conditioned place preference formed by the association of a location (CS) with a *social* stimulus (US). The typical SCPP experiment involves sexual reward (Fig. 1B, middle).

Conditioned social preference (CSP) A non-location conditioned preference in which the CS and/or the US are social. For example, a conspecific (CS) is associated with a reinforcing substance (US). Or, for example, an odor (CS) is associated with a reinforcing social interaction (US) (Fig. 1B, bottom).

Social and environmental Context The classical conditioned preference is a laboratory construct. However, any conditioned preference can be modified by previous interactions with the social and ecological environment independently of a classical conditioned place preference.

Reward vs reinforcement In brief, reward is received from a stimulus that an animal interprets as positive (e.g., satisfying or pleasurable), whereas reinforcement is the process by which an experience increases the likelihood of a particular behavioral output. A rewarding experience often serves as a positive reinforcer, but by the same measure alleviation of an aversive experience serves as a negative reinforcer. In the example of a typical CPP the US (e.g., a substance of abuse) is rewarding, which reinforces the behavioral preference for the CS (e.g., the location in which the US was administered). It is important to mention that it is possible to have reinforcement through associative learning beyond the classical context of reward (for example, see Crawford and Domjan, 1993, also see Section 3.2. Motivation (wanting)).

associative learning. In doing so we hope to increase 1) appreciation by behavioral neuroscientists for the naturalistic factors that gate associative learning and 2) appreciation by behavioral ecologists for neuroendocrine advances that elucidate the underpinnings of complex naturalistic behavior. Our hope is that future studies will continue to elegantly bridge neuroendocrinology and behavioral ecology.

The fundamental basis of the conditioned preference is the formation of a reinforcing association between an animal and an appetitive stimulus, such as a specific location or social partner in the animal's environment. This association enhances the probability of the animal's return to (or prolonged interaction with) that appetitive stimulus in order to enhance its survival and reproductive success. A classic example of this is the preference developed by males of some species for the location in which they encountered sexual stimuli (reviews by Adkins-Regan, 2020; Domjan and Gutiérrez, 2019; Hollis, 1990). Because space use is a reliable method for assessing classical (i.e., Pavlovian, see Domjan, 2005) conditioning, the conditioned place preference (CPP) in particular has become a standard tool in basic and translational research. CPPs are evoked not only by ethologically relevant reinforcements, such as food and matings, but also by motivating and pleasurable substances, including psychostimulants and opiates. This has led to the widespread adoption of CPPs as a method for investigating

the underlying mechanisms of addiction and substance abuse in pharmacology and neuroscience. The present review shifts the focus towards an ethologically relevant approach to study the modulation of conditioned preference, integrating social and locational influences through the lens of behavioral neuroendocrinology. For example, what are the social and physiological factors that might lead a seasonally-breeding bird to return to a socially dense area at some times of the year but not others, or alter the locations to which paired and unpaired mice of a monogamous and territorial species will condition? We investigate how life history, reproductive stages, past and current social experiences, and the physical environment (e.g., familiarity) impact endocrine and neural systems, shaping the development and expression of conditioned preferences. This exploration addresses a key question: what are the natural adaptive functions of conditioned preferences from a neuroendocrine perspective? We highlight diverse species to enrich our understanding of these adaptive functions, showcasing innovative methods to investigate such preferences and answer ethologically grounded questions.

Pharmacologically-induced preferences are most often studied in social isolation. However, an increasing body of evidence is revealing the importance of social context and social endocrine systems in the regulation of reinforcement. For example, ethanol-induced social preference in female mice is enhanced by the intoxication of a peer, an effect facilitated by the presence of estradiol (Wood and Rice, 2013). Additionally, ethanol-induced social conditioning is abolished in oxytocin receptor/vasopressin receptor 1 A knockout female mice (Wood et al., 2015). Thus, social context serves as another important dimension of association and conditioning. Recent study designs have integrated concepts underlying place preference conditioning with those related to social reinforcement (e.g., the well-established partner preference test) to develop social context-dependent assessments of reinforcement, such as the conditioned social preference (CSP) and the social conditioned place preference (SCPP) (Fig. 1). This review explores 1) the conceptual overlap between the CPP, the CSP, and the SCPP; 2) a brief survey of their major neurobiological underpinnings; 3) the context-dependency of their establishment by endocrine examples especially gonadal hormones and neuropeptides. We also highlight two recent ethological examples of how the concept of a conditioned preference can be applied: 1) a set of laboratory studies with European starlings (Sturnus vulgaris) examining the use of CPPs to investigate changes in reward that lead to differences in reinforcement of social behavior and song across breeding and non-breeding seasons, and 2) field and laboratory studies using CPPs to investigate changes in the rewarding properties of testosterone pulses in bonded and non-bonded male monogamous California mice (Peromyscus californicus).

### 2. An overview of conditioned preferences

### 2.1. Conditioned preferences for natural experiences

Reinforcing experiences and behaviors are diverse and varied in natural settings, especially for social animals. Beyond reinforcement of resource acquisition and defense, such as foraging (Rudebeck and Izquierdo, 2022) and territoriality (X. Zhao et al., 2020), social interactions such as play (C. Zhao et al., 2020), mating (Pitchers et al., 2014), communication (Riters, 2012), bonding (Young and Wang, 2004), parental care (Mulligan et al., 2021), and even aggression (Farrell and Wilczynski, 2006; Legrand, 1970; Martínez et al., 1995; Tellegen et al., 1969; Tellegen and Horn, 1972) can be reinforcing. Associations between a place or another individual and a reinforcing experience present important opportunities for learning and adjusting to a changing environment; reinforcing stimuli likely evolved their reinforcing valence because they confer a fitness advantage. Conditioning to the context of a positive experience increases the likelihood of experiencing it again. The likelihood of engaging in a particular reinforcing experience, and thus forming a conditioned preference to its social or environmental context, is dependent on both reproductive and life

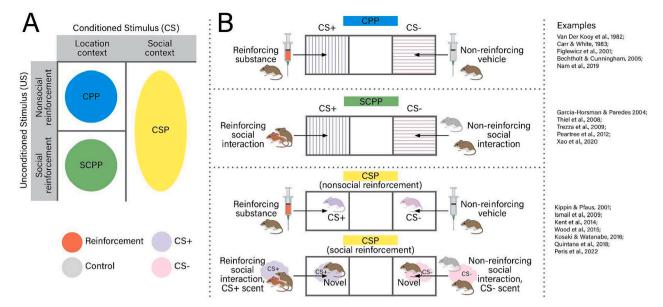


Fig. 1. Location and social context are two separate and overlapping dimensions for understanding context-dependent reinforcement conditioning. A. The conditioned place preference (CPP), conditioned social preference (CSP), and social conditioned place preference (SCPP) test preference with different combinations of social or nonsocial unconditioned stimuli (US) and conditioned stimuli (CS). B. Simplified diagrams of typical conditioned preference paradigms. For the CPP, the US is a nonsocial reinforcement and the CS+ is a location; the strength or reinforcement can be modulated by social context. For the SCPP, the US is social reinforcement (e.g. sexual experience) and the CS+ is a location. For the CSP, the US is either a social (e.g. scent of a conspecific) or nonsocial reinforcement and the CS+ is either a social stimulus (top) or a nonsocial stimulus with a social context (bottom) (CPP Examples: (Bechtholt and Cunningham, 2005; Carr and White, 1983; Figlewicz et al., 2001; Nam et al., 2019; Van Der Kooy et al., 1982); SCPP examples: (García Horsman and Paredes, 2004; Peartree et al., 2012; Thiel et al., 2008; Trezza et al., 2009; X. Zhao et al., 2020); CSP examples: (Ismail et al., 2009; Kent et al., 2014; Kippin and Pfaus, 2001; Kosaki and Watanabe, 2016; Peris et al., 2022; Quintana et al., 2019; Wood et al., 2015)).

history stages. For example, in rats, play is most reinforcing during the juvenile stage (Vanderschuren et al., 1997). Additionally, sexual behavior is reinforcing (Pfaus et al., 2001), but the strength of reinforcement may vary based on social context and the ovarian cycle phase (Corona, 2011; Paredes and Alonso, 1997). For example, in seasonally breeding animals, reinforcement changes seasonally such that sexual behavior is reinforcing during the breeding but nonsexual social interactions are rewarding during the non-breeding season (Riters and Stevenson, 2022). This evidence suggests that the capacity to either experience a stimulus as reinforcing or form an association between the stimulus and the social or environmental context is dependent on plasticity within neural systems involved in reward (Berridge and Dayan, 2021), social behavior (O'Connell and Hofmann, 2011), and spatial processing (Vann and Albasser, 2011). Moreover, as these neural networks are enriched for both gonadal hormone and neuropeptide receptors (ISH Data: Allen Brain Atlas: Mouse Brain, n.d.), and their signaling is regulated by reproductive and life history stages, evidence is emerging that such endocrine systems play an important role in the reinforcement of context-specific naturalistic experiences.

Pharmacologically-induced conditioned preference research has been an important component of understanding the underlying reinforcement neural circuitry co-opted by drugs of abuse, providing important insights for human health (Bardo and Bevins, 2000). However, these experiments are often done without consideration for social context. All brain and hormonal mechanisms evolved in natural environments; therefore, studies taking place in natural environments replete with relevant, competing, and multimodal stimuli, are essential for understanding the function of neural systems. Moreover, association of reinforcement with place and social stimuli is likely appropriate and beneficial to fitness in naturalistic conditions, and often only becomes inappropriate and maladaptive when selection pressures are lessened, such as in environments of abundance.

### 2.2. The conditioned place preference (CPP)

The CPP paradigm is a well-established method for investigating reinforcement learning (Tzschentke, 2007), typically in the absence of social context. The basic design of the task is to associate a particular location with the effects of a stimulus (typically a pharmacological substance) and associate a different environment with the absence of that stimulus. In brief, an animal will develop a preference for a previously neutral environment (the conditioned stimulus, CS+) if it becomes associated with a reinforcing stimulus (unconditioned stimulus, US, such as food or a drug of abuse) even after the reinforcing stimulus is no longer present. CPPs can also reflect the reward value of a stimulus, as the greater reward received from a stimulus, the greater the strength of the reinforcement (Tzschentke, 2007).

Commonly, this experiment consists of a three-chambered apparatus, with the outer chambers designed to have different features (e.g., textures, visual cues, etc.), while the central chamber is neutral (although variations exist, for a review see (Bardo et al., 1995)). A threechambered apparatus allows for some delineation between preference and avoidance. Training involves administering the potentially reinforcing drug (US) to an animal and confining their movement to one of the outer chambers (CS+) while the drug is active. The following day, the animal is placed in the opposite compartment (CS-) and is administered a control vehicle. This alternating procedure is typically repeated several times (often for 2-3 days for each bout of drug and vehicle administration). Following conditioning, the animal is tested for their location preference by allowing them to freely travel between all three chambers and recording the amount of time spent in each chamber. If the animal spends more time in the chamber where the drug was administered (CS+) than in the opposite chamber (CS-), as is often the case for drugs of abuse, it is concluded that a CPP was formed. However, if the opposite is observed, such as for drugs that induce pain or discomfort, then it is concluded that a conditioned place avoidance was formed. For examples of CPP, see Section 4.1. Testosterone.

### 2.2.1. Social context modulates CPPs

It is important to note that the reinforcing properties of some stimuli can be revealed or modulated by social context. Evidence is accumulating in both human and animal models that social context and social separation are capable of conferring resilience or susceptibility to the effects of pharmacologically-induced reinforcement. For example, adolescent social defeat stress strengthens adult CPP to amphetamines in male rats (Burke et al., 2011). Conversely, social interactions in male rats and mice inhibit the formation of cocaine-induced CPP (Zernig and Pinheiro, 2015).

### 2.3. The social conditioned place preference (SCPP)

Social interaction can be highly reinforcing in social species. Social reinforcement can serve as the US and location can serve as the CS, indicating place association with social experience. For example, paced mating produces a SCPP in female rats, indicated by a preference for the chamber in which they engaged in paced mating (Paredes and Alonso, 1997). Another instance is play; juvenile rats will prefer a location where they have previously played with other juveniles (Calcagnetti and Schechter, 1992).

Both affiliative behaviors (such as copulation and bonding behaviors) as well as aggressive behaviors (such as winning fights) are able to induce the formation of SCPPs. Regardless of the stimulus, the formation of SCPP is likely a function of classical conditioning whereby the repeated pairing of an unconditioned stimulus with a stimulus results in a new association formed with the previously conditioned stimulus. Importantly, the conditioned preferences rely on the transient effects of a substance beyond a baseline level (pulsatility), in order to associate the reinforcement with a particular context. For examples of SCPP, see Section 4.2. Estradiol.

### 2.4. The conditioned social preference (CSP)

By replacing the neutral environment CS+ with a neutral social context CS+ it is possible to determine if a substance increases the reinforcing nature of a particular social context, thus forming a conditioned social preference (Kent et al., 2013). The CSP paradigm combines elements of both the CPP and partner preference tests (Wood and Rice, 2013). Partner preference tests are typically used as indicators of sociosexual and pair-bond motivation, whereby the subject interacts with two tethered animals of the opposite sex and time spent near one or the other indicates preference (Paredes, 2009). However, other social preference tests, such as those for parent-offspring or same-sex affiliation are also used. In the CSP paradigm, however, the stimulus animals are instead the conditioned stimuli (CS+ and CS-) and a separate stimulus serves as the unconditioned stimulus (e.g. drugs, reinforcing social interactions).

# 2.4.1. It is important to note that a CSP can be formed with either a social or nonsocial US

For example, in female rats, multiple paced copulations with a male (US) scented with a particular odor (CS+) increased their solicitations of a novel male with the same odor, whereas nonpaced copulations did not produce the same effect (Coria-Avila et al., 2005). Similarly to females, male rats will prefer to ejaculate with an unfamiliar female with a particular scent (CS+) if that scent is associated with previous copulation and ejaculation (US), a particular type of CSP known as the conditioned ejaculatory preference (Kippin and Pfaus, 2001). In another, more complex example, male Japanese quail develop a CSP to another castrated male as a result of mating attempts, indicating that CSP can reveal plasticity in sexual preferences (Nash and Domjan, 1991). However, nonsocial reinforcing stimuli are also capable of producing CSPs. For example, ethanol consumption (US) in the presence of a particular rat (CS+) increases future time spent in the presence of that rat (Wood et al., 2015). Taken together, evidence suggests that the formation of a CSP can occur in a manner similar to the formation of a

CPP, suggesting that the same reinforcement system underlies both social and location associations. For examples of CSP, see Section 5. Oxytocin.

# 2.5. Ethologically relevant conditioned preferences and social behavior: an example research program in European starlings

What is reinforcing can change depending on ecological variables, social experience, and resource availability. The affective states induced by social stimuli adaptively reinforce behaviors that are critical for survival and reproduction and discourage others that may be maladaptive. Here, we briefly focus on research with European starlings to highlight the overlap of social- and place-based conditioned preferences.

For female European starlings, hearing male courtship song during the breeding season is rewarding, but only for females that possess the resources necessary for reproduction (i.e., nesting sites) (Riters et al., 2013; Spool and Riters, 2017). For male European starlings, the act of producing courtship song during the breeding season does not result in a CPP to the location associated with singing (Riters and Stevenson, 2012). Instead, it is thought that for breeding male songbirds, the reward comes from successful copulation, not singing itself (Riters and Stevenson, 2012).

A different scenario occurs in starlings during the non-breeding season, in which song is produced by both males and females and is a form of non-sexual flock cohesion rather than mate attraction (Riters et al., 2019). In this social and ecological context, the act of producing song itself then becomes rewarding, where starlings develop strong SCPPs for places paired with the production of their own flock songs (Hahn et al., 2017; Riters and Stevenson, 2012; Stevenson et al., 2020).

Taken together, these data demonstrate that social context is vitally important for the development of CPPs, especially in neurobiologically plastic and socially dynamic species. We do not know what components of a European starling's own song production lead to the associated preference, or how location and social context interact. Future research is needed to explore these interesting concepts.

Here, we describe an example of considerations for developing naturalistic conditioned preferences in nonmodel species. In both model and non-model species, there are certain behaviors that cannot be assessed within the confines of the most common CPP apparatus. For such behaviors, it becomes necessary to observe the behavior and then place the animal in the respective chamber for conditioning. Using this method, the physiological consequences of the behavior rather than the factors triggering the behavior are measured. This method has achieved robust results when determining copulation-associated reward in rats (Ågmo and Berenfeld, 1990; Paredes, 2009; Pfaus et al., 2001) and songassociated reward in birds (Riters, 2012; Riters et al., 2014; Riters and Stevenson, 2012; Stevenson et al., 2020). For the latter, non-sexual songs are often only produced when stress is low. Within the CPP apparatus, birds may not sing even when housed with familiar conspecifics. One solution is to capture a bird directly from the flock immediately after song is observed and place it into the designated CPP chamber for conditioning (Riters and Stevenson, 2012).

In non-model species, it is important to adjust the design of the CPP apparatus to suit the unique needs of that species. For instance, nesting European starlings are highly motivated to gather green nesting material (Rubalcaba et al., 2016), whereas female zebra finches avoid males with green leg bands (Hunt et al., 1997), making it ill-advised to use green in CPPs in these birds. There are additional concerns with using non-terrestrial animals that exist in three-dimensional space; for example, in birds, perches of varying yet consistent heights are used to achieve a semi-naturalistic testing chamber. In addition, wild-caught animals are often neophobic (e.g. (Kelly et al., 2020)), which may result in the birds confining themselves to a single chamber. For these species, extra caution is needed to include a habituation period prior to conditioning.

# 3. An overview of the neurobiology of associative learning through the lens of conditioned preferences

### 3.1. Neural dopamine and opioid underpinnings

The mechanisms of learned association underlying conditioned preferences are complex and can contribute to how (i.e. strength of) and whether conditioned preferences are formed. Neurobiologically, it becomes important to disentangle the three components of reward: "wanting", "liking", and "learning" (Berridge and Kringelbach, 2015; Berridge and Robinson, 2003; Dickinson and Balleine, 2002), further conceptualized as motivation, hedonics, and cognition (Trezza et al., 2010).

### 3.2. Motivation (wanting)

While a number of neurotransmitters are implicated in the subcomponents of motivation, hedonics, and cognition, mesolimbic dopaminergic signaling has typically been associated with the motivational aspects of reward, with many studies conducted in rodents. Appetitive motivation and the role of dopamine in "wanting" are well-reviewed. and well-debated, elsewhere (Rieger et al., 2022; Russo and Nestler, 2013). In brief, the reinforcing mechanism of the association formed in conditioned preferences relies heavily on the mesolimbic dopamine system. Numerous studies demonstrate that a CPP can be established by dopamine receptor agonists and blocked by receptor antagonists. Moreover, dopamine signaling drives both social reinforcement and drug reinforcement, suggesting a common underlying mechanism (Burkett and Young, 2012). Regions of the brain involved in reinforcement, in particular the nucleus accumbens (Dölen et al., 2013), amygdala (Hu et al., 2021), and preoptic area (Bayless et al., 2023; Riters and Stevenson, 2022), are likely highly overlapping across types of reinforcement: a common currency reinforcing network of interacting brain regions (Berridge and Kringelbach, 2015). Activation of the mesolimbic reinforcement system mediates motivation to experience social interaction (Borland et al., 2018). Thus, reinforcement for drugs of abuse and naturalistic reinforcement such as social interactions (affiliative behaviors, successful aggressive behaviors) likely emerge from similar processes.

Naturally rewarding behaviors (like fall birdsong or rodent social play) have an associated motivation to engage or seek out contexts in which those behaviors can be produced. For example, European starlings that produce fall song in flocks are more motivated to seek out conspecifics than birds that do not sing in flocks (Maksimoski et al., 2023), testifying to the positive affect associated with producing fall song and its link with social reward and motivation. In addition, fall song is strongly correlated with increased dopamine related mRNA expression in brain regions known to regulate appetitive and consummatory reinforcement (Maksimoski et al., 2023; Merullo et al., 2016).

### 3.3. Hedonics (liking)

The role of opioids in the regulation of subjective pleasure (i.e., "liking"), including that from social reward, has been theorized for several decades (Berridge, 1996; Hayward et al., 2002; Panksepp et al., 1980). Opioid activity within the nucleus accumbens modulates the hedonic properties of rewards such as food, and may indirectly enhance motivation in the ventral tegmental area by stimulating mesolimbic dopaminergic signaling (Barbano and Cador, 2007; Berridge and Kringelbach, 2008). Evidence for the role of the endogenous opioid system in mediating the hedonic components of reinforcement is evidenced by opioid involvement in many affective social behaviors including parenting, sexual behavior, and play (Depue and Morrone-Strupinsky, 2005).

SCPPs resulting from sexual reward likely require endogenous opioid release in both sexes (Miller and Baum, 1987; Paredes and Martínez,

2001). Females display mate-guarding behavior towards a male following paced mating (Holley et al., 2014). Accordingly, mate guarding behavior in female rats resulting from paced mating conditioning is eliminated if an opioid antagonist is administered during conditioning (Coria-Avila et al., 2008). In male rats, ejaculation leads to opioid release (Coolen et al., 2004), and copulation-induced CPP and conditioned ejaculatory preference depend on the reward state produced by opioids (Kippin and Pfaus, 2001; Ulloa et al., 2018). Both copulation-induced SCPP and conditioned ejaculatory preference are eliminated if an opioid antagonist is administered during conditioning sessions (Quintana et al., 2019). However, some conflicting evidence exists to suggest that opioids are involved in the expression of the conditioned preference, rather than its acquisition (Mehrara and Baum, 1990).

Activation of mu opioid receptors in a number of brain regions induces reward in rodents (Berridge and Kringelbach, 2013). For example, there is substantial evidence for the role of site-specific mu opioid activity in the avian nucleus accumbens and preoptic area in stimulating reward (Riters et al., 2017; Stevenson et al., 2020) and non-sexual fall song (Maksimoski et al., 2021; Polzin et al., 2022; Riters et al., 2014). Conversely, sexually motivated song is inhibited by mu opioid receptor stimulation in the preoptic area, perhaps by supplementing the external reward otherwise received from mating, thereby negating the motivation to sing in these contexts (Riters, 2012; Riters et al., 2019).

### 3.4. Cognition (learning)

While motivation and hedonics are crucial for the formation of conditioned preference, so too is the contextualization that situates the reinforcement within a particular location or social context. It is important to realize that conditioned preferences are a subset of associative learning and in the context of natural functions it is likely that several associative learning processes are occurring when a preference is formed. Thus, spatial perception and social memory are necessary to induce conditioned preferences.

One caveat is the difficulty of distinguishing between the cognitive learning and the hedonic/motivational components of a conditioned preference, as behavioral output requires an animal to remember the association (Huston et al., 2013). In the case of social reinforcement associated with location, there are two cognitive systems that interact with hedonic/motivation networks to impact the expression of a conditioned preference: the social behavioral network and the spatial memory network. In order to untangle the contribution of these components to conditioned preference, it is necessary to rely on a combination of behavioral assessments such as operant, partner preference, and memory tasks. These two cognitive systems in addition to the neural reward system (Fig. 3) are enriched for hormonal receptors, suggesting that they are regulated by endocrine systems (ISH Data: Allen Brain Atlas: Mouse Brain, n.d.). This is supported by an abundance of evidence demonstrating that hormones are capable of modulating both cognitive performance and drug induced CPP as reviewed elsewhere (Skuse and Gallagher, 2009; Williams and Meck, 1991). Cognitive properties of rewards are complex and varied and are likely modulated on multiple neural levels by an array of neurotransmitters and neurohormones.

### 4. Gonadal hormones

### 4.1. Testosterone

### 4.1.1. Testosterone and CPP in males

Studies across rodent species demonstrate that aggressive and sociosexual behavior can induce a CPP in males (Hughes et al., 1990; Mehrara and Baum, 1990; Miller and Baum, 1987). These effects are likely mediated by testosterone due to the role of testosterone in aggression and sexual behavior in males (Antunes and Oliveira, 2009; Hughes et al., 1990). Interestingly, conditioned cues are also able to

elicit hormone secretion, for example male rats have been shown to secrete testosterone in anticipation of sexual activity (Graham and Desjardins, 1980). Moreover, the propensity for the rapid action of testosterone to positively reinforce place associations is wellcharacterized in male mice, rats, and hamsters (Alexander et al., 1994; Arnedo et al., 2000; Wood et al., 2004; Zhao and Marler, 2014, 2016). Subcutaneous injections of testosterone are sufficient to establish a CPP in a dose-dependent fashion (Alexander et al., 1994; De Beun et al., 1992). Central injection of testosterone into the nucleus accumbens likewise establishes a CPP, suggesting that this reinforcement is driven by nucleus accumbens activity (Packard et al., 1997). Androgen reinforcement can act through membrane androgen receptors (Wood et al., 2004) and androgen metabolites (Frye, 2007; Rosellini et al., 2001). It is likely that sub-populations of androgen receptor neurons within the nucleus accumbens are responsible for the reinforcing properties of testosterone, as testosterone metabolites such as dihydrotestosterone and 3a-androstanediol induce CPP when administered to the nucleus accumbens shell but not the core (Frye et al., 2002).

The role of testosterone on CPP may be through the dopamine and opioid reward pathways, as treatments combining estrogens and androgens influence tyrosine hydroxylase (the rate limiting enzyme for the synthesis of both dopamine and norepinephrine) labeling density in birds (Appeltants et al., 2003; LeBlanc et al., 2007) and mammals (Kritzer, 2000, 2003), indicating that steroid hormones affect dopamine synthesis. Testosterone and its metabolites in mammals can also affect enkephalin opioid densities (Simerly et al., 1988; Watson et al., 1986). In male dark-eyed juncos, opioid receptor densities in the medial preoptic area and ventral tegmental area shift seasonally in association with gonadal volume (Woods et al., 2010), suggesting seasonal changes in testosterone in seasonally-breeding birds such as European starlings may alter opioid receptor activity in medial preoptic area and ventral tegmental area (Spool et al., 2016).

Dopamine signaling is central to the reinforcing effects of testosterone in the nucleus accumbens, as peripheral or intra-accumbens administration of dopamine receptor 1 or 2 antagonists prevent a testosterone-induced CPP (Becker and Marler, 2015; Packard et al., 1998; Schroeder and Packard, 2000). In addition to the nucleus accumbens, injections of testosterone into the medial preoptic area at low doses establish a CPP (King et al., 1999), suggesting a distributed network of regions may be responsible. For example, one brain area that is expected to be important for spatial and social memory processing is the hippocampus, which processes and integrates multimodal sensory information with emotional valence to consolidate memory engrams (Kitamura et al., 2017; Yu and Moss, 2022). While further research is necessary to uncover whether androgen signaling is capable of stimulating opioid release, there is evidence that blocking opioids can lead to pulsatile secretion of testosterone (Tenhola et al., 2012) and that the opioid antagonist naltroxone blocks self-administration of testosterone and the seeking of testosterone by increasing access via nose pokes in hamsters (Peters and Wood, 2005; Wood, 2008). Such results suggest a relationship between androgen and opioid systems and raise the intriguing possibility that opioid hotspots (Castro and Berridge, 2017) may be activated by testosterone.

### 4.1.2. Testosterone, CPP, and social context

The CPP test is used frequently in non-social conditions in laboratory studies for measuring the reinforcing and addictive nature of drugs and neurochemicals. However, we understand little about the natural functions and social relevance of location preferences, including those produced by testosterone. Pulses of testosterone are evoked naturally in males during aggressive male-male and affiliative female-male encounters, suggesting the role of testosterone on SCPPs is driven by territorial and sexual behavior (Camacho et al., 2004; Gleason et al., 2009; Goymann et al., 2019). Recent studies by Zhao, Marler, and colleagues highlight a function for testosterone in ethologically relevant CPPs and social-context dependent CPPs in the monogamous and highly territorial

California mice in the lab. In these California mice, Zhao and Marler (2014) investigated variation in testosterone-driven space use by examining the formation of CPP in sexually naive and pair-bonded males. Subcutaneous administration of physiologically relevant levels of testosterone induced a place preference to a novel environment in unpaired adult males (Zhao and Marler, 2014). In paired males, testosterone strengthened a CPP for the location containing the nesting area in which they may have naturally conditioned to via cues such as the scents of their mate. The effect of testosterone on place preference was shown to be socially context dependent, as testosterone did not lead to a CPP when given to males in the location containing the nesting area with other male cagemates (Zhao and Marler, 2016). Thus, testosterone is likely important for territory development in locations in which the male has had successful sexual encounters and where the male has formed a bond with the female mate.

The social context dependence of testosterone-induced CPPs is reflected in the neural plasticity of androgen receptors. Singly housed males exhibit upregulated androgen receptor expression in the preoptic area relative to same-sex group housed males, which positively correlated with the strength of the CPP (X. Zhao et al., 2020), implicating a connection of the preoptic area to CPPs. This change in androgen sensitivity in the preoptic area may be characteristic of males that have dispersed to establish their own territory, making them more sensitive to testosterone signaling. Winning and testosterone pulses can result in an increase in AR receptors in the nucleus accumbens, ventral tegmental area, and the bed nucleus of the stria terminalis when experienced in a familiar environment, which may be associated with territorial establishment, maintenance, or expansion (Fuxjager et al., 2010). Collectively, these findings support that the reinforcing nature of testosterone is dependent on the familiarity of the environment, pair bond status, and sexual experience (Fig. 2).

# 4.1.3. Ethologically relevant conditioned preferences and hormones: an example research program in California mice

There are classic studies relating testosterone CPP conditioning to the context of sexual behavior in the lab (reviews by Adkins-Regan, 2020; Domjan and Gutiérrez, 2019; Hollis, 1990). In the first test of CPP in the wild, Petric, Kalcounis-Rupell and Marler (Petric et al., 2022) confirmed the classical view of T-induced CPPs as being site specific even in a complex natural environment. Among California mice, territorial defense, parental care, and resource acquisition are carried out by both sexes, with considerable behavioral plasticity both within and between pairs (Monari et al., 2021; Rieger et al., 2019, 2021) and compensation can occur between pair members such as time at the nest (Trainor and Marler, 2001). One mechanism for adjusting time between locations (i.e. nest versus territorial boundary) is through conditioned preferences. As described above, testosterone is rewarding and reinforcing and males can experience a pulse in testosterone after both malemale and male-female encounters (Fig. 2). The male testosterone response to female California mice represent laboratory data that are presented for the first time in this review and are critical for demonstrating male testosterone responsiveness to females in California mice (Fig. 2 and Supplement 1). Three experimentally-induced pulses of testosterone mimicking natural levels were administered to paired males in the field over five days in their nests (Petric et al., 2022). Males treated with testosterone while in traps at the nest developed an altered spatial preference, accompanied by changes in social interactions and call production. Males treated with testosterone spent more time in the nest, which likely increased parental care both because of being in the nest and because testosterone implants alone increase paternal behavior (Trainor and Marler, 2001). As described earlier for laboratory research, males also spent more time in the nest chamber previously associated with his mate when paired with testosterone pulses (Zhao and Marler, 2016). We therefore see consistency between the laboratory and field studies. Untreated female mates rapidly respond to changes in male spatial preference by spending more time away from the nest. In

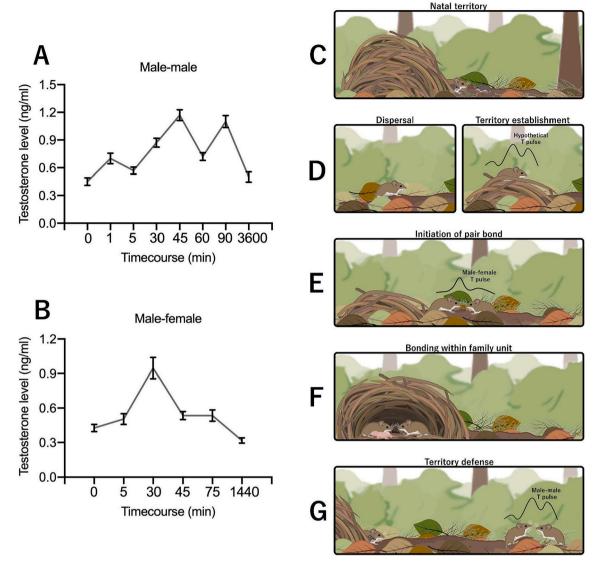


Fig. 2. Male California mouse testosterone responses to A. A male encounter (based on Marler et al., 2005) and B. A female encounter (new data; Zhao and Marler). Detailed methods for measurements of testosterone levels (as also described in Bester-Meredith and Marler, 2001; Davis and Marler, 2003; Trainor and Marler, 2001) and male-female encounters are provided in Supplement 1. A one-way ANOVA demonstrated a statistically significant overall impact (F(5,39) = 2.64, P = 0.03). Subsequent Dunnett's multiple comparisons test against the 0 min control group indicated a significant rise in T levels at 30 min (Adjusted P = 0.0446). Testosterone therefore peaked 30 min after a male-female encounter. C–G. Testosterone pulses and their potential contextual place preference functions. Natural conditioned place preferences (CPPs) are dependent on location, social context, and hormone signaling. C. Juveniles/adults within their natal territory do not establish a CPP to testosterone pulses. D. Testosterone pulses following maturation and dispersal from the natal territory may induce a CPP as they establish and defend a new territory. E. Sociosexual behavior induces testosterone release that elicits CPP. F. Conditioned preferences related to family unit bonding likely depend on additional hormonal signaling such as oxytocin. G. Testosterone pulses resulting from territorial defense induce a CPP that adjusts time spent at the territory boundary versus the nest based on location and social interactions with neighboring competitors.

response to the female mate's absence, more vocalizations were produced (Petric et al., 2022; Timonin et al., 2018), specifically hypothesized to be associated with long distance communication in the field (Briggs and Kalcounis-Rueppell, 2011). Such vocalizations are increased when the mates are far apart but not close together, and this selectivity is likely a combination of the change in location and direct and indirect effects of testosterone. The above results with California mice illustrate that a place preference can be induced through testosterone pulses in a similar manner in the laboratory and field and that behavioral changes may occur because of both the conditioning altering location preferences and, potentially, through other direct and indirect (such as through location changes) changes induced by testosterone. A potential scenario for the functions of testosterone pulses at different stages of the life history of California mice is shown in Fig. 2.

### 4.1.4. Testosterone influences spatial context learning in males

While the above studies suggest that testosterone is capable of providing a *reinforcing* signal, it is also likely that it alters the capability of *learning* the reinforcing association. Brain regions central to learning and memory, such as the hippocampus, are highly enriched for androgen receptors (Naghdi et al., 2001), suggesting a link between the experience of testosterone-induced reinforcement and the cognitive aspects of reinforcement learning. Androgens regulate spatial memory in male rodents, although there is little consensus of the direction of this effect across studies. In some cases, androgens have been observed to improve spatial memory in tasks such as the Y-maze and the Morris water maze (Hawley et al., 2013; Khalil et al., 2005), while in other cases androgen treatment was associated with impaired spatial memory or no effect (Goudsmit et al., 1990; Naghdi et al., 2001). Testosterone

administered directly into the hippocampus improved spatial navigation performance (Roof and Havens, 1992), whereas flutamide, an androgen receptor antagonist, decreased spatial learning (Naghdi et al., 2001). Moreover, testosterone supplementation may improve spatial memory in men (Cherrier et al., 2001; Driscoll and Resnick, 2007; Janowsky, 2006; Sherwin, 2003). At a structural level, androgens in the hippocampus produce changes in spine density and neurogenesis in a sexually differentiated way (Atwi et al., 2016). For example, testosterone enhances the survival of immature dentate gyrus neurons in adult males via an androgen receptor mechanism (Hamson et al., 2013). Androgens likely result in complex effects on spatial memory due to their conversion to various metabolites, such as estradiol and dihydrotestosterone, in several regions including the hippocampus (Tabatadze et al., 2014; Yague et al., 2010). When considering the propensity for androgens to establish place preferences, it is important to integrate reinforcement with spatial learning.

### 4.1.5. Testosterone and CPP in females

Research is limited on the role of testosterone on CPP establishment in females. Females do not exhibit the same testosterone pulse associated with the reward of an aggressive encounter in males (Davis and Marler, 2003). Additionally, there are mixed results regarding the impact of testosterone on reward in females in general, with some studies showing that testosterone does not elicit a CPP in female rodents, while others showing that the testosterone metabolite, 3αDIOL, does induce a CPP in female rodents (De Beun et al., 1992; Jorge et al., 2005). The influence of testosterone on reward in females may be more closely related to sexual behavior rather than aggression, as one study found a prenatal blockade of androgen receptors reduced the number of intromissions necessary for paced mating to induce a CPP, highlighting a potential role of androgen receptors in the rewarding value of female sexual behavior (Domínguez-Salazar et al., 2005). Future studies should explore the complex interactions of hormonal systems, including testosterone, on reinforcement in females (Smiley et al., 2022).

### 4.2. Estradiol

### 4.2.1. Estradiol and CPP/SCPP formation in females

17β-estradiol is the most bioactive estrogen in reproductive age females (Salole, 1986; "The Smoking Estrogens – a Potential Synergy between Estradiol and Benzo(a)Pyrene," 2021) and is an important contributor to the development of conditioned preferences. There is a close link between estradiol levels and sexual receptivity in females, as well as motivated sexual behavior, or sexual proceptivity (Johnson and Phoenix, 1976; Spiteri et al., 2009). Reproductive behavior may therefore be the evolutionary basis for the role of estradiol on motivation and reward and its impact on CPP formation (Corona, 2011; Frye and Rhodes, 2006; Richard et al., 2017). Female rodents exclusively engage in copulation in the estrus phase, immediately following the estradiol peak which occurs during the proestrus phase (Hardy, 1972; Jennings and de Lecea, 2020). Estradiol treatment that mimics this peak in ovariectomized female rodents is known to induce a CPP (Frye and Rhodes, 2006)

Sexual interaction in female rodents leads to conditioned preferences in a context-dependent manner. Females exposed to a male form a SCPP in the compartment where copulation occurred (Meisel and Joppa, 1994; Oldenburger et al., 1992). However, paced mating, where females have control over the sexual interaction, is necessary for the SCPP to form (Paredes and Alonso, 1997; Paredes and Martínez, 2001). Furthermore, estradiol, rather than copulation alone, may be necessary for this SCPP to occur. In a study evaluating the dose-dependent impact of estradiol on CPPs, ovariectomized female rats were treated with both high and low levels of estradiol and all females engaged in paced mating (Corona, 2011). The females treated with higher levels of estradiol preferred the chamber in which mating had occurred relative to females treated with low levels of estradiol (Corona, 2011). This highlights the

importance of estradiol concentration in a SCPP formation and reward more broadly, as it suggests that male stimulus in addition to copulation was only rewarding when estradiol levels were high. The internal environment in the form of the background hormonal milieu is therefore also critical for forming conditioned preferences.

Not only does estradiol administration lead directly to a CPP, but estradiol levels also influence the ability of other rewarding substances to induce a CPP. For example, estradiol can both upregulate and downregulate non-sexual reward (Hilderbrand and Lasek, 2018; Richard et al., 2017). Motivation for sucrose decreased during phases associated with high estradiol in naturally cycling female rats, and estradiol injection in ovariectomized rats also decreased motivation for sucrose, measured via lever pressing (Richard et al., 2017). It is possible that estradiol may alter motivation from food-seeking behavior to sexual behavior through the reward system, an adaptive shift to prioritize reproductive behavior during ovulation (E. Yoest et al., 2014).

Estradiol can also influence CPP by increasing the strength of reinforcement of a reward. Estradiol treatment enhances ethanol and cocaine induced CPP in ovariectomized rodents ("Estradiol," 2010; Hilderbrand and Lasek, 2018). In women, subjective hedonic responses to amphetamines are increased during the late follicular phase, when estradiol levels are rising, compared to phases when estradiol levels are lower (Justice and de Wit, 1999), which may contribute to the increased severity of and vulnerability to substance use disorders in females relative to males (McHugh et al., 2018; Tonn Eisinger et al., 2018).

It is likely that estradiol influences CPP in non-rodent species as well. The link between estradiol and sexual motivation is not limited to rodents, suggesting certain types of behavior or social encounters may be more rewarding depending on estradiol concentration, impacting the likelihood of the formation of a CPP. Little is known about the formation of CPPs in non-human primates, but there is evidence to support an important role for estradiol in sexual motivation and reward (Bardo and Bevins, 2000). Estradiol is not necessary for copulation to occur in primates, but females exhibit greater motivation to engage in sexual behavior in menstrual phases where estradiol levels are high. Rhesus macaques are more likely to seek out males to copulate during the follicular phase ("Sex and Context," 2001). Estradiol may regulate CPP formation in birds as well, as female Japanese quail have demonstrated a cocaine-induced CPP with high estradiol, but this preference can be erased when estradiol levels are low (Gill et al., 2016). This, of course, highlights the importance of female reproductive state for the reinforcing effects of drugs.

The coupling of sexual behavior with a specific location may have reproductive advantages. In European starlings, breeding condition females develop strong natural place preferences for nesting sites. Female European starlings that are allowed to explore a nest site have higher concentrations of circulating estradiol and express higher levels of the endogenous opioid preproenkephalin in the medial preoptic area; a region in which estrogens and opioids modulate sexual motivation (Spool et al., 2018). This demonstrates that not only can estradiol given in a specific location under the context of sexual behavior lead to the formation of a CPP, but locations associated with reproduction, such as a nesting site, can also lead to release of estradiol which may both induce a preference for a nesting site and stimulate reproductive behavior (Spool et al., 2018).

To summarize, hormonal state must be considered when determining whether or not a stimulus will or will not be rewarding and lead to the formation of a CPP. Under levels of high estradiol, a sexual stimulus may be highly rewarding and lead to the formation of a CPP, while a food reward will not (Corona, 2011; Richard et al., 2017). Under conditions of low estradiol, copulation may not lead to a CPP, but a food reward will (Corona, 2011; Richard et al., 2017).

### 4.2.2. Mechanisms by which estradiol influences CPP

We have demonstrated that estrogenic activity can modulate rewards leading to a CPP in females; the mechanisms through which estradiol

may influence CPPs likely function via estrogen receptors on neurons interacting within the reward system, such as the nucleus accumbens and ventral tegmental area. Both estrogen receptor alpha (ER $\alpha$ ) and estrogen receptor beta (ER $\beta$ ) play a role in the formation of a CPP, as ethanol in conjunction with an ER $\alpha$  or ER $\beta$  agonist alone was not sufficient to enhance an ethanol-induced a CPP, but activation of both enhances an ethanol-induced CPP (Hilderbrand and Lasek, 2018). However, amphetamine-induced CPPs are enhanced by activation of ER $\beta$  alone, with no effect of ER $\alpha$  (Silverman and Koenig, 2007).

Despite having low levels of estrogen receptors, there are rapid effects of estradiol on activity in the nucleus accumbens that have been associated with estradiol-associated CPP formation (Frye and Rhodes, 2006; Yoest et al., 2014). This may be due to the presence of estrogen receptors on extranuclear sites surrounding the nucleus accumbens core and shell, influencing GABAergic and catecholaminergic systems at the presynaptic level, and ultimately impacting dopamine transmission (Almey et al., 2022). The coupling of estradiol activity during reproductive periods and sexual motivation and reward has been linked to the action of estrogen receptors on metabotropic glutamate receptors on the nucleus accumbens (Tonn Eisinger et al., 2018). This is consistent with the finding that the formation of a CPP during paced mating is associated with estradiol levels in the nucleus accumbens (Frye and Rhodes, 2006). Estradiol also reduces expression of Regulator G-protein Signaling 9-2 protein in the nucleus accumbens shell, an important protein for the behavioral responses to rewarding substances such as amphetamines and cocaine (Silverman and Koenig, 2007).

Estrogenic activity in the ventral tegmental area also plays an important role in both social and non-social CPP formation. Activation of the ventral tegmental area is crucial for female sexual receptivity in rodents (Sumida et al., 2005), and may also play a major role in female sexual interest (Yin and Lin, 2023). Ovariectomy decreases the preference in a cocaine-induced CPP test, and decreases the amount of dopamine and serotonin present in the ventral tegmental area (Russo et al., 2003), two neurochemicals that are central to the motivation and reward signaling pathway. Dopamine neuron sensitivity in the ventral tegmental area fluctuates throughout the estrous cycle, and direct treatment with estradiol in ovariectomized mice enhances the response of dopamine neurons in the ventral tegmental area to both dopamine and ethanol (Vandegrift et al., 2017). This may help to explain how estradiol enhances ethanol-induced CPPs (Hilderbrand and Lasek, 2018). The ventral tegmental area may also be critical for the estradiolinduced suppression of food motivation during reproduction, as estradiol injected directly into the ventral tegmental area reduces sucroseinduced CPPs, suggesting an important regulatory role of the ventral tegmental area (Richard et al., 2017).

### 4.2.3. Estradiol and spatial context learning in females

It is well-established that estrogens play an important role in spatial cognition, a critical component to the formation of CPPs (Genazzani et al., 2007). Estrogen receptors are found within the hippocampus as well as in prefrontal cortices (Genazzani et al., 2007). Estradiol-induced synaptic plasticity during seasonal changes and ovarian cycling in brain regions important for cognition (García-Segura et al., 1994) suggest that estradiol modulates the response to environmental and social context in a top-down fashion. Indeed, estradiol regulates spatial memory and hippocampal plasticity (Gibbs et al., 2004; Li et al., 2004; Luine et al., 1998). Ovariectomized female rats display memory and spatial learning impairments that can be rescued via estradiol treatment (Rashidy-Pour et al., 2019). Contrary to this, naturally cycling female rhesus macaques demonstrate reduced spatial memory performance during ovulation, when estradiol concentrations are high (Lacreuse et al., 2001). To form a CPP, the animal must also learn and remember the location. Therefore, the role of estradiol on spatial learning may play a critical role in the formation of CPPs, beyond enhancing the reward.

### 4.2.4. Estradiol and CPP in males

Little is known about the direct role of estradiol and CPPs in males. However, many of the rewarding effects of testosterone may come from the conversion of testosterone into estradiol via aromatase, as aromatase knockout male mice do not form SCPPs exposed to females in estrus (Pierman et al., 2006). This demonstrates that estradiol may be similarly rewarding in both males and females.

### 5. Oxytocin

Oxytocin is an evolutionarily conserved neuropeptide, synthesized in the paraventricular nucleus and the supraoptic nucleus of the hypothalamus, that is critical for the regulation of social behavior in mammals (Lim and Young, 2006). Oxytocin receptors are distributed widely in the rodent brain (Tribollet et al., 1988, 1992), and oxytocin is a critical modulator of affiliative and aggressive behaviors. Campbell (2008) and others suggest that it may increase social salience (Shamay-Tsoory and Abu-Akel, 2016). The impact of oxytocin on social recognition and reinforcement is well-established and its release may temporally link the social context of mating to reward (Burkett and Young, 2012). Additionally, oxytocin receptor expression is highly linked to partner preference formation in female prairie voles (Ross et al., 2009), and social recognition is absent in oxytocin knockout mice (Carter et al., 2008).

### 5.1. Oxytocin and conditioned preference in females

Beyond partner and sexual preference, the ability for oxytocin to induce conditioned preferences has been primarily studied in female-female affiliative contexts (Wood et al., 2015). Ventral tegmental area oxytocin receptor activation is important for the reinforcing properties of social interactions in Syrian hamsters, and same-sex social interactions are more reinforcing in females than in males (Borland et al., 2018). Although activation of oxytocin receptors can be reinforcing in the absence of a social stimulus and subcutaneous oxytocin administration is able to produce a CPP, intracerebroventricular infusion of oxytocin dose-dependently induces a CSP in ovariectomized female mice given estradiol implants, suggesting that it is capable of reinforcement in both solitary and social conditions (Kent et al., 2013). Other research has found that intranasal oxytocin induced a conditioned same-sex social preference (CSP) in female mice but not a CPP (Kosaki and Watanabe, 2016).

Ethanol can modify the development of conditioned preferences. If female mice are intoxicated when exposed to a stimulus female, they will form a CSP (Wood and Rice, 2013). Oxytocin receptor and vasopressin receptor 1a knockout female mice do not form a CSP following ethanol administration, and in fact developed conditioned social avoidance, suggesting that these receptors are required for conditioning an association between a reward and a social stimulus (Wood et al., 2015).

The ability of oxytocin to lead to a CSP is an inverted U-shaped response curve, with both high and low doses showing no effect and medium doses leading to a CSP (Kent et al., 2013). This inverted U may be influenced by the presence of estradiol; estradiol upregulates oxytocin and oxytocin receptor expression, which may contribute both to sex differences in the ability of oxytocin to induce a conditioned place or social preference, as well as changes across the hormonal cycle (Borland et al., 2019; Jirikowski et al., 2018). To summarize, oxytocin appears to be an important contributor to female-female CSPs with variable results for CCPs.

### 5.2. Oxytocin and conditioned preference in males

For oxytocin-induced conditioned preferences in males, there is more information available for the brain regions involved. Infusion of oxytocin into the central amygdala induces a CPP in male rats (László

et al., 2016), and both the reinforcing and anxiolytic effects of this treatment can be abolished with a dopamine receptor 2 antagonist (László et al., 2020). Additionally, oxytocin receptors are present on dopamine neurons in the ventral tegmental area that project to limbic regions (Peris et al., 2017), and mesolimbic dopamine release is modulated by exogenous oxytocin (Estes et al., 2019). These findings support the close interaction between oxytocin and the mesolimbic reinforcement system (Love, 2014). The generalizability of the reinforcing effects of oxytocin requires further investigation, as there is conflicting evidence of the ability for exogenous oxytocin to form a nonsocial CPP in males (Kosaki and Watanabe, 2016). For example, while subcutaneous injections of oxytocin can produce a CPP in male rats (Liberzon et al., 1997), other studies did not observe a nonsocial CPP when oxytocin was infused intraperitoneally in male rats (Ramos et al., 2015), intracerebroventricularly in male mice (Qi et al., 2009), or into the nucleus accumbens in male rats (Baracz et al., 2012). This continues to emphasize the importance of the social component necessary for oxytocin effects on the formation of condition place preferences.

Whether or not oxytocin is reinforcing in nonsocial contexts (rarely conducted in laboratory settings, as rodents are most often grouphoused), the social environment is an important modulator of the reinforcement. Accordingly, intraperitoneal injection of oxytocin can induce a robust CPP only if another rat is present during the conditioning sessions (Ramos et al., 2015). Another caveat is that effects are likely dependent on dose and route of administration, as other experiments failed to observe either a conditioned same sex social preference or a CPP in male rats (Kosaki and Watanabe, 2016). Sex differences in the reinforcing properties of oxytocin are consistent with prior research demonstrating that exogenous oxytocin in the absence of mating can induce an opposite-sex partner preference in female but not male prairie voles (Cushing and Carter, 2000). The different susceptibilities of male and female subjects to exogenous oxytocin are in accordance with previous studies focusing on the effect of oxytocin on different aspects of social cognition and behavior. For example, administration of oxytocin either centrally (via intracerebroventricular injection; (Insel and Hulihan, 1995)) or peripherally (via subcutaneous injection; (Cushing and Carter, 2000)) facilitates the formation of opposite sex partner preference without mating experience in female but not male prairie voles, whereas administration of vasopressin to male prairie voles enhances partner preference for a familiar female over a novel female prior to cohabitation (Cho et al., 1999). Oxytocin therefore plays a significant role in CSPs, but more information is needed regarding its effects on nonsocial conditioning.

### 5.3. Oxytocin and spatial context learning

Oxytocin can modulate memory processes, especially social memory (Albers, 2012; Gabor et al., 2012). Therefore, in studies of social conditioned preferences, it is necessary to disentangle the possibility that activation of oxytocin receptors influences memory rather than reward itself (Bardo and Bevins, 2000). An interesting method to dissociate memory and reward in the reinforcing effects of oxytocin was recently described (Borland et al., 2018, 2019), in which a novel operant social preference task was used to assess behavioral effort in male Syrian hamsters. Hamsters were placed in a three-chambered apparatus and given access to a social chamber containing a conspecific or a nonsocial chamber, with each chamber separated by a vertical swinging door able to change the degree of behavioral effort required to gain access to the adjacent chamber. Animals demonstrate a preference for the social chamber, and motivation to access the social chamber can be studied by the frequency of chamber entries and by increasing the behavioral effort required to access the chamber. Infusion of oxytocin into the ventral tegmental area decreased the motivation to seek social interactions, whereas the opposite was observed for an oxytocin antagonist (Borland et al., 2019), providing evidence for the hypothesis that oxytocin receptors in the ventral tegmental area are directly involved in social reinforcement.

### 5.4. Oxytocin-vasopressin interplay and conditioned preference

Oxytocin and vasopressin are highly similar in structure (Acher and Chauvet, 1995), as are their receptors (Gimpl and Fahrenholz, 2001). Promiscuity between these ligands and receptors has been observed (Song et al., 2016; Song and Albers, 2018), and it is therefore challenging to separate the functions of oxytocin and vasopressin signaling. However, carefully designed studies have been able to tease apart the role of vasopressin, oxytocin, and their interplay on conditioned preferences. For example, male rats given a subcutaneous injection of oxytocin developed a conditioned ejaculatory preference, but not rats given vasopressin or saline, indicating that oxytocin is able to enhance the association of odor with sexual reinforcement (Ménard et al., 2019). Additionally, through its action on oxytocin receptors rather than vasopressin receptors, vasopressin can produce a CPP when injected into the ventral tegmental area (Song et al., 2014). This evidence suggests that oxytocin receptor activity drives conditioned preference formation.

Oxytocin and vasopressin may differ in their interaction with nonsocial reward induced CPPs. For example, intracerebroventricular and nucleus accumbens shell oxytocin administration enhances the expression of a morphine-induced CPP in male rats (Moaddab et al., 2015), while lateral septum vasopressin administration decreases the expression of amphetamine-induced CPP in male rats (Gárate-Pérez et al., 2021). Moreover, vasopressin may contribute to the impact of social context on the ability of drugs of abuse to result in conditioned preferences. Both vasopressin antagonists and housing with drug-naive animals reduces the expression of a morphine-induced CPP in male mice, suggesting that the protective effect of housing on the formation of opioid-dependent CPPs may be mediated by vasopressin (Bates et al., 2018). Additionally, ethanol-induced CSPs to same-sex cagemates are reduced in oxytocin receptor/vasopressin receptor 1 A knockout female mice (Wood et al., 2015). Taken together, the present research emphasizes the complexity of the interplay between oxytocin and vasopressin and emphasizes the need for future experiments to disentangle the differential impacts of neuropeptides on conditioned preferences.

### 6. Conclusion and future directions

Associative learning is a fundamental process that can lead to location and social environment-dependent conditioned preferences in both the lab and field. These preferences are mediated by endocrine signals that interact with pathways of reward, reflecting an internal stategating. To fully grasp both the naturalistic functions of conditioned preferences and the emergence of pathological conditioned preferences, a comprehensive approach that encompasses sex differences, life history, reproductive, social, and environmental factors is essential. The studies involving European starlings and California mice serve as exemplary models, demonstrating the effective application of these holistic approaches in ethological research. Conditioned preferences, in their simplicity, offer a versatile methodology applicable in both laboratory and field settings, allowing for an integrated examination of location and social preferences at behavioral and neural levels.

We advocate for the crucial role of hormonal and neuroendocrine system signaling in mediating both top-down influences (such as life history and reproductive stage) and bottom-up factors (like social and environmental contexts) in the formation of conditioned preferences. While not discussed in-depth in this review, future research directions should better elucidate the brain networks that interact to orchestrate complex naturalistic associative preferences. In Fig. 3 we provide a broad conceptual framework for guiding such future investigations.

Given that brain networks related to social behavior, reward processing, and spatial cognition are rich in endocrine receptors, often in a sex-dependent manner, a deeper understanding of their individual contributions to the hedonic, motivational, and learning aspects of

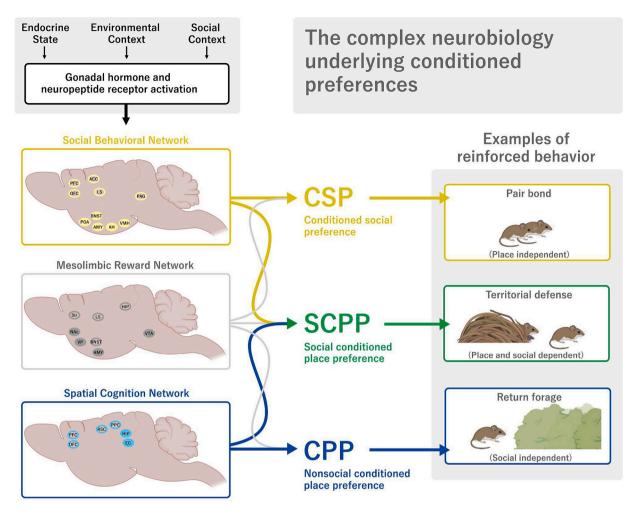


Fig. 3. A "future directions" conceptual framework for understanding how social and location-dependent conditioned preferences may emerge from associations formed between three overlapping neural networks enriched for sex hormone and neuropeptide receptors: social, reward, and spatial networks. Top left: Internal and external factors lead to context-dependent gonadal hormone and neuropeptide signaling in the brain. Bottom left: Simplified diagrams of the overlapping networks involved in conditioned social preferences (CSP), and social and nonsocial conditioned place preferences (SCPP and CPP, respectively). Key regions are color-filled, other important regions are color-outlined. Center: Network activity interactions lead to learned associations and conditioning. Right: Plasticity results in the expression of conditioned preferences in social and nonsocial contexts.

conditioned preference is invaluable. Future research into the endocrine regulation of conditioned preferences should delve into the comparative effects of endocrine signaling on the release of neurotransmitters, such as dopamine and opioids. For instance, while it is known that testosterone can trigger dopamine release, the specifics of how testosterone influences opioid signaling—and its impact on conditioned preferences via this pathway—remain largely unexplored. Investigating these mechanisms will not only enhance our understanding of the neurobiological basis of conditioned preferences in situations that are ethologically relevant but also aid in developing context-specific treatments for addiction, offering new perspectives and therapeutic avenues.

Supplementary data to this article can be found online at https://doi.org/10.1016/j.yhbeh.2024.105529.

### CRediT authorship contribution statement

Patrick K. Monari: Writing – review & editing, Writing – original draft, Visualization, Supervision, Funding acquisition, Conceptualization. Emma R. Hammond: Writing – review & editing, Writing – original draft. Xin Zhao: Writing – original draft, Methodology, Data curation, Conceptualization. Alyse N. Maksimoski: Writing – review & editing, Writing – original draft. Radmila Petric: Writing – original draft, Conceptualization. Candice L. Malone: Writing – original draft.

**Lauren V. Riters:** Writing – review & editing, Writing – original draft. **Catherine A. Marler:** Writing – review & editing, Writing – original draft, Supervision, Funding acquisition, Conceptualization.

### Declaration of competing interest

None.

### Acknowledgements

We thank the editors of this special issue for the invitation to write this article. Writing of this manuscript was supported by the National Science Foundation (IOS 1946613 and DGE-1747503).

### References

Acher, R., Chauvet, J., 1995. The neurohypophysial endocrine regulatory cascade: precursors, mediators, receptors, and effectors. Front. Neuroendocrinol. 16 (3), 237–289. https://doi.org/10.1006/frne.1995.1009.

Adkins-Regan, E., 2020. Sexual and pairing partner preference in birds and other animals. Horm. Behav. 118, 104646 https://doi.org/10.1016/j.yhbeh.2019.104646. Ågmo, A., Berenfeld, R., 1990. Reinforcing properties of ejaculation in the male rat: role of opioids and dopamine. Behav. Neurosci. 104 (1), 177–182. https://doi.org/ 10.1037/0735-7044.104.1.177.

- Albers, H.E., 2012. The regulation of social recognition, social communication and aggression: vasopressin in the social behavior neural network. Horm. Behav. 61 (3), 283–292. https://doi.org/10.1016/j.yhbeh.2011.10.007.
- Alexander, G.M., Packard, M.G., Hines, M., 1994. Testosterone has rewarding affective properties in male rats: implications for the biological basis of sexual motivation. Behav. Neurosci. 108 (2), 424–428. https://doi.org/10.1037/0735-7044.108.2.424.
- Almey, A., Milner, T.A., Brake, W.G., 2022. Estrogen receptors observed at extranuclear neuronal sites and in glia in the nucleus accumbens core and shell of the female rat: evidence for localization to catecholaminergic and GABAergic neurons. J. Comp. Neurol. 530 (11), 2056–2072. https://doi.org/10.1002/cne.25320.
- Antunes, R.A., Oliveira, R.F., 2009. Hormonal anticipation of territorial challenges in cichlid fish. Proc. Natl. Acad. Sci. USA 106 (37), 15985–15989. https://doi.org/ 10.1073/pnas.0900817106.
- Appeltants, D., Ball, G.F., Balthazart, J., 2003. Song activation by testosterone is associated with an increased catecholaminergic innervation of the song control system in female canaries. Neuroscience 121 (3), 801–814. https://doi.org/ 10.1016/S0306-4522(03)00496-2.
- Arnedo, M.T., Salvador, A., Martinez-Sanchis, S., Gonzalez-Bono, E., 2000. Rewarding properties of testosterone in intact male mice: a pilot study. Pharmacol. Biochem. Behav. 65 (2), 327–332. https://doi.org/10.1016/S0091-3057(99)00189-6.
- Atwi, S., McMahon, D., Scharfman, H., MacLusky, N.J., 2016. Androgen modulation of hippocampal structure and function. Neuroscientist 22 (1), 46–60. https://doi.org/ 10.1177/1073858414558065.
- Baracz, S.J., Rourke, P.I., Pardey, M.C., Hunt, G.E., McGregor, I.S., Cornish, J.L., 2012. Oxytocin directly administered into the nucleus accumbens core or subthalamic nucleus attenuates methamphetamine-induced conditioned place preference. Behav. Brain Res. 228 (1), 185–193. https://doi.org/10.1016/j.bbr.2011.11.038.
- Barbano, M.F., Cador, M., 2007. Opioids for hedonic experience and dopamine to get ready for it. Psychopharmacology 191 (3), 497–506. https://doi.org/10.1007/ s00213-006-0521-1.
- Bardo, M.T., Bevins, R.A., 2000. Conditioned place preference: what does it add to our preclinical understanding of drug reward? Psychopharmacology 153 (1), 31–43. https://doi.org/10.1007/s002130000569.
- Bardo, M.T., Rowlett, J.K., Harris, M.J., 1995. Conditioned place preference using opiate and stimulant drugs: a meta-analysis. Neurosci. Biobehav. Rev. 19 (1), 39–51. https://doi.org/10.1016/0149-7634(94)00021-R.
- Bates, M.L.S., Hofford, R.S., Emery, M.A., Wellman, P.J., Eitan, S., 2018. The role of the vasopressin system and dopamine D1 receptors in the effects of social housing condition on morphine reward. Drug Alcohol Depend. 188, 113–118. https://doi. org/10.1016/j.drugalcdep.2018.03.021.
- Bayless, D.W., Davis, C.-H.O., Yang, R., Wei, Y., de Andrade Carvalho, V.M., Knoedler, J. R., Yang, T., Livingston, O., Lomvardas, A., Martins, G.J., Vicente, A.M., Ding, J.B., Luo, L., Shah, N.M., 2023. A neural circuit for male sexual behavior and reward. Cell 186 (18), 3862–3881.e28. https://doi.org/10.1016/j.cell.2023.07.021.
- Bechtholt, A.J., Cunningham, C.L., 2005. Ethanol-induced conditioned place preference is expressed through a ventral tegmental area dependent mechanism. Behav. Neurosci, 119 (1), 213–223. https://doi.org/10.1037/0735-7044.119.1.213.
- Becker, E.A., Marler, C.A., 2015. Postcontest blockade of dopamine receptors inhibits development of the winner effect in the California mouse (*Peromyscus californicus*). Behav. Neurosci. 129 (2), 205–213. https://doi.org/10.1037/bne0000043.
- Berridge, K.C., 1996. Food reward: brain substrates of wanting and liking. Neurosci. Biobehav. Rev. 20 (1), 1–25. https://doi.org/10.1016/0149-7634(95)00033-B.
- Berridge, K.C., Dayan, P., 2021. Liking. Curr. Biol. 31 (24), R1555–R1557. https://doi. org/10.1016/j.cub.2021.09.069.
- Berridge, K.C., Kringelbach, M.L., 2008. Affective neuroscience of pleasure: reward in humans and animals. Psychopharmacology 199 (3), 457–480. https://doi.org/ 10.1007/s00213-008-1099-6.
- Berridge, K.C., Kringelbach, M.L., 2013. Neuroscience of affect: brain mechanisms of pleasure and displeasure. Curr. Opin. Neurobiol. 23 (3), 294–303. https://doi.org/ 10.1016/j.conb.2013.01.017.
- Berridge, K.C., Kringelbach, M.L., 2015. Pleasure systems in the brain. Neuron 86 (3), 646–664. https://doi.org/10.1016/j.neuron.2015.02.018.
- Berridge, K.C., Robinson, T.E., 2003. Parsing reward. Trends Neurosci. 26 (9), 507–513. https://doi.org/10.1016/S0166-2236(03)00233-9.
- Bester-Meredith, J.K., Marler, C.A., 2001. Vasopressin and aggression in cross-fostered California mice (*Peromyscus californicus*) and white-footed mice (*Peromyscus leucopus*). Horm. Behav. 40 (1), 51–64. https://doi.org/10.1006/hbeh.2001.1666.
- The smoking estrogens a potential synergy between estradiol and benzo(a)pyrene. Biomed. Pharmacother. 139, 2021, 111658. https://doi.org/10.1016/j. biophs. 2021.111658.
- Borland, J.M., Grantham, K.N., Aiani, L.M., Frantz, K.J., Albers, H.E., 2018. Role of oxytocin in the ventral tegmental area in social reinforcement. Psychoneuroendocrinology 95, 128–137. https://doi.org/10.1016/j. psynguen.2018.05.028.
- Borland, J.M., Rilling, J.K., Frantz, K.J., Albers, H.E., 2019. Sex-dependent regulation of social reward by oxytocin: an inverted U hypothesis. Neuropsychopharmacology 44 (1), 1. https://doi.org/10.1038/s41386-018-0129-2.
- Briggs, J.R., Kalcounis-Rueppell, M.C., 2011. Similar acoustic structure and behavioural context of vocalizations produced by male and female California mice in the wild. Anim. Behav. 82 (6), 1263–1273. https://doi.org/10.1016/j.anbehav.2011.09.003.
- Burke, A.R., Watt, M.J., Forster, G.L., 2011. Adolescent social defeat increases adult amphetamine conditioned place preference and alters D2 dopamine receptor expression. Neuroscience 197, 269–279. https://doi.org/10.1016/j. neuroscience.2011.09.008.

- Burkett, J.P., Young, L.J., 2012. The behavioral, anatomical and pharmacological parallels between social attachment, love and addiction. Psychopharmacology 224 (1), 1–26. https://doi.org/10.1007/s00213-012-2794-x.
- Calcagnetti, D.J., Schechter, M.D., 1992. Place conditioning reveals the rewarding aspect of social interaction in juvenile rats. Physiol. Behav. 51 (4), 667–672. https://doi. org/10.1016/0031-9384(92)90101-7.
- Camacho, F., Sandoval, C., Paredes, R.G., 2004. Sexual experience and conditioned place preference in male rats. Pharmacol. Biochem. Behav. 78 (3), 419–425. https://doi. org/10.1016/j.pbb.2004.04.015.
- Campbell, A., 2008. Attachment, aggression and affiliation: the role of oxytocin in female social behavior. Biol. Psychol. 77 (1), 1–10. https://doi.org/10.1016/j. biopsycho.2007.09.001.
- Carr, G.D., White, N.M., 1983. Conditioned place preference from intra-accumbens but not intra-caudate amphetamine injections. Life Sci. 33 (25), 2551–2557. https://doi. org/10.1016/0024-3205(83)90165-0.
- Carter, C.S., Grippo, A.J., Pournajafi-Nazarloo, H., Ruscio, M.G., Porges, S.W., 2008. Oxytocin, vasopressin and sociality. In: Neumann, I.D., Landgraf, R. (Eds.), Progress in Brain Research, vol. 170. Elsevier, pp. 331–336. https://doi.org/10.1016/S0079-6123(08)00427-5.
- Castro, D.C., Berridge, K.C., 2017. Opioid and orexin hedonic hotspots in rat orbitofrontal cortex and insula. Proc. Natl. Acad. Sci. 114 (43), E9125–E9134. https://doi.org/10.1073/pnas.1705753114.
- Cherrier, M.M., Asthana, S., Plymate, S., Baker, L., Matsumoto, A.M., Peskind, E., Raskind, M.A., Brodkin, K., Bremner, W., Petrova, A., LaTendresse, S., Craft, S., 2001. Testosterone supplementation improves spatial and verbal memory in healthy older men. Neurology 57 (1), 80–88. https://doi.org/10.1212/WNL.57.1.80.
- Cho, M.M., DeVries, A.C., Williams, J.R., Carter, C.S., 1999. The effects of oxytocin and vasopressin on partner preferences in male and female prairie voles (*Microtus ochrogaster*). Behav. Neurosci. 113 (5), 1071–1079. https://doi.org/10.1037/0735-7044.113.5.1071.
- Coolen, L.M., Fitzgerald, M.E., Yu, L., Lehman, M.N., 2004. Activation of μ opioid receptors in the medial preoptic area following copulation in male rats. Neuroscience 124 (1), 11–21. https://doi.org/10.1016/j.neuroscience.2003.10.045.
- Coria-Avila, G.A., Ouimet, A.J., Pacheco, P., Manzo, J., Pfaus, J.G., 2005. Olfactory conditioned partner preference in the female rat. Behav. Neurosci. 119 (3), 716–725. https://doi.org/10.1037/0735-7044.119.3.716.
- Coria-Avila, G.A., Solomon, C.E., Vargas, E.B., Lemme, I., Ryan, R., Ménard, S., Gavrila, A.M., Pfaus, J.G., 2008. Neurochemical basis of conditioned partner preference in the female rat: I. Disruption by naloxone. Behavioral Neuroscience 122 (2), 385–395. https://doi.org/10.1037/0735-7044.122.2.385.
- Corona, R., 2011. Different doses of estradiol benzoate induce conditioned place preference after paced mating. Horm. Behav. 60 (3), 264–268. https://doi.org/ 10.1016/j.yhbeh.2011.05.013.
- Crawford, L.L., Domjan, M., 1993. Sexual approach conditioning: omission contingency tests. Anim. Learn. Behav. 21 (1), 42–50.
- Cushing, B.S., Carter, C.S., 2000. Peripheral pulses of oxytocin increase partner preferences in female, but not male, prairie voles. Horm. Behav. 37 (1), 49–56. https://doi.org/10.1006/hbeh.1999.1558.
- Davis, E.S., Marler, C.A., 2003. The progesterone challenge: steroid hormone changes following a simulated territorial intrusion in female *Peromyscus californicus*. Horm. Behav. 44 (3), 185–198. https://doi.org/10.1016/S0018-506X(03)00128-4.
- De Beun, R., Jansen, E., Slangen, J.L., Van de Poll, N.E., 1992. Testosterone as appetitive and discriminative stimulus in rats: sex- and dose-dependent effects. Physiol. Behav. 52 (4), 629–634. https://doi.org/10.1016/0031-9384(92)90389-J.
- Depue, R.A., Morrone-Strupinsky, J.V., 2005. A neurobehavioral model of affiliative bonding: implications for conceptualizing a human trait of affiliation. Behav. Brain Sci. 28 (3), 313–350. https://doi.org/10.1017/S0140525X05000063 (discussion 350-395).
- Dickinson, A., Balleine, B., 2002. The role of learning in the operation of motivational systems. In: Steven's Handbook of Experimental Psychology: Learning, Motivation, and Emotion, , 3rd edvol. 3. John Wiley & Sons, Inc., pp. 497–533. https://doi.org/10.1002/0471214426.pas0312
- Dölen, G., Darvishzadeh, A., Huang, K.W., Malenka, R.C., 2013. Social reward requires coordinated activity of nucleus accumbens oxytocin and serotonin. Nature 501 (7466), 7466. https://doi.org/10.1038/nature12518.
- Domínguez-Salazar, E., Camacho, F.J., Paredes, R.G., 2005. Prenatal blockade of androgen receptors reduces the number of intromissions needed to induce conditioned place preference after paced mating in female rats. Pharmacol. Biochem. Behav. 81 (4), 871–878. https://doi.org/10.1016/j.pbb.2005.06.011.
- Domjan, M., 2005. Pavlovian conditioning: a functional perspective. Annu. Rev. Psychol. 56, 179–206.
- Domjan, M., Gutiérrez, G., 2019. The behavior system for sexual learning. Behav. Process. 162, 184–196. https://doi.org/10.1016/j.beproc.2019.01.013.
- Driscoll, I., Resnick, S.M., 2007. Testosterone and cognition in normal aging and Alzheimer's disease: an update. Curr. Alzheimer Res. 4 (1), 33–45. https://doi.org/ 10.2174/156720507779939878.
- Estes, M.K., Freels, T.G., Prater, W.T., Lester, D.B., 2019. Systemic oxytocin administration alters mesolimbic dopamine release in mice. Neuroscience 408, 226–238. https://doi.org/10.1016/j.neuroscience.2019.04.006.
- Farrell, W.J., Wilczynski, W., 2006. Aggressive experience alters place preference in green anole lizards. Anolis carolinensis. Animal Behaviour 71 (5), 1155–1164. https://doi.org/10.1016/j.anbehav.2005.10.006.
- Figlewicz, D.P., Higgins, M.S., Ng-Evans, S.B., Havel, P.J., 2001. Leptin reverses sucroseconditioned place preference in food-restricted rats. Physiol. Behav. 73 (1), 229–234. https://doi.org/10.1016/S0031-9384(01)00486-3.

- Frye, C.A., 2007. Some rewarding effects of androgens may be mediated by actions of its  $5\alpha$ -reduced metabolite  $3\alpha$ -androstanediol. Pharmacol. Biochem. Behav. 86 (2), 354–367. https://doi.org/10.1016/j.pbb.2006.10.003.
- Frye, C.A., Rhodes, M.E., 2006. Administration of estrogen to ovariectomized rats promotes conditioned place preference and produces moderate levels of estrogen in the nucleus accumbens. Brain Res. 1067 (1), 209–215. https://doi.org/10.1016/j. brainres.2005.10.038.
- Frye, C.A., Rhodes, M.E., Rosellini, R., Svare, B., 2002. The nucleus accumbens as a site of action for rewarding properties of testosterone and its 5α-reduced metabolites. Pharmacol. Biochem. Behav. 74 (1), 119–127. https://doi.org/10.1016/S0091-3057 (02)00968-1.
- Fuxjager, M.J., Forbes-Lorman, R.M., Coss, D.J., Auger, C.J., Auger, A.P., Marler, C.A., 2010. Winning territorial disputes selectively enhances androgen sensitivity in neural pathways related to motivation and social aggression. Proc. Natl. Acad. Sci. 107 (27), 12393–12398. https://doi.org/10.1073/pnas.1001394107.
- Gabor, C.S., Phan, A., Clipperton-Allen, A.E., Kavaliers, M., Choleris, E., 2012. Interplay of oxytocin, vasopressin, and sex hormones in the regulation of social recognition. Behav. Neurosci. 126 (1), 97–109. https://doi.org/10.1037/a0026464.
- Gárate-Pérez, M.F., Méndez, A., Bahamondes, C., Sanhueza, C., Guzmán, F., Reyes-Parada, M., Sotomayor-Zárate, R., Renard, G.M., 2021. Vasopressin in the lateral septum decreases conditioned place preference to amphetamine and nucleus accumbens dopamine release. Addict. Biol. 26 (1), e12851 https://doi.org/10.1111/adb.12851.
- García Horsman, P., Paredes, R.G., 2004. Dopamine antagonists do not block conditioned place preference induced by paced mating behavior in female rats. Behav. Neurosci. 118 (2), 356–364. https://doi.org/10.1037/0735-7044.118.2.356.
- García-Segura, L.M., Chowen, J.A., Párducz, A., Naftolin, F., 1994. Gonadal hormones as promoters of structural synaptic plasticity: cellular mechanisms. Prog. Neurobiol. 44 (3), 279–307. https://doi.org/10.1016/0301-0082(94)90042-6.
- Genazzani, A.R., Pluchino, N., Luisi, S., Luisi, M., 2007. Estrogen, cognition and female aging. Hum. Reprod. Update 13 (2), 175–187. https://doi.org/10.1093/humupd/
- Gibbs, R.B., Gabor, R., Cox, T., Johnson, D.A., 2004. Effects of raloxifene and estradiol on hippocampal acetylcholine release and spatial learning in the rat. Psychoneuroendocrinology 29 (6), 741–748. https://doi.org/10.1016/S0306-4530 (03)00118-5.
- Gill, K.E., Reynolds, A.R., Prendergast, M.A., Akins, C.K., 2016. Female Japanese quail with high levels of estradiol demonstrate cocaine-induced conditioned place preference. Exp. Clin. Psychopharmacol. 24 (3), 185–192. https://doi.org/10.1037/ pha0000070.
- Gimpl, G., Fahrenholz, F., 2001. The oxytocin receptor system: structure, function, and regulation. Physiol. Rev. 81 (2), 629–683. https://doi.org/10.1152/ physrev.2001.81.2.629.
- Gleason, E.D., Fuxjager, M.J., Oyegbile, T.O., Marler, C.A., 2009. Testosterone release and social context: when it occurs and why. Front. Neuroendocrinol. 30 (4), 460–469. https://doi.org/10.1016/j.yfrne.2009.04.009.
- Goudsmit, E., Van De Poll, N.E., Swaab, D.F., 1990. Testosterone fails to reverse spatial memory decline in aged rats and impairs retention in young and middle-aged animals. Behav. Neural Biol. 53 (1), 6–20. https://doi.org/10.1016/0163-1047(90) 00720.
- Goymann, W., Moore, I.T., Oliveira, R.F., 2019. Challenge Hypothesis 2.0: a fresh look at an established idea. BioScience 69 (6), 432–442. https://doi.org/10.1093/biosci/big041
- Graham, J.M., Desjardins, C., 1980. Classical conditioning: induction of luteinizing hormone and testosterone secretion in anticipation of sexual activity. Science 210 (4473), 1039–1041. https://doi.org/10.1126/science.7434016.
- Hahn, A.H., Merullo, D.P., Spool, J.A., Angyal, C.S., Stevenson, S.A., Riters, L.V., 2017. Song-associated reward correlates with endocannabinoid-related gene expression in male European starlings (Sturnus vulgaris). Neuroscience 346, 255–266. https://doi. org/10.1016/j.neuroscience.2017.01.028.
- Hamson, D.K., Wainwright, S.R., Taylor, J.R., Jones, B.A., Watson, N.V., Galea, L.A.M., 2013. Androgens increase survival of adult-born neurons in the dentate gyrus by an androgen receptor-dependent mechanism in male rats. Endocrinology 154 (9), 3294–3304. https://doi.org/10.1210/en.2013-1129.
- Hardy, D.F., 1972. Sexual behavior in continuously cycling rats. Behaviour 41 (3–4), 288–297. https://doi.org/10.1163/156853972X00068.
- Hawley, W.R., Grissom, E.M., Martin, R.C., Halmos, M.B., Bart, C.L.S., Dohanich, G.P., 2013. Testosterone modulates spatial recognition memory in male rats. Horm. Behav. 63 (4), 559–565. https://doi.org/10.1016/j.yhbeh.2013.02.007.
- Hayward, M.D., Pintar, J.E., Low, M.J., 2002. Selective reward deficit in mice lacking β-Endorphin and enkephalin. J. Neurosci. 22 (18), 8251–8258. https://doi.org/ 10.1523/JNEUROSCI.22-18-08251.2002.
- Hilderbrand, E.R., Lasek, A.W., 2018. Estradiol enhances ethanol reward in female mice through activation of ER $\alpha$  and ER $\beta$ . Horm. Behav. 98, 159–164. https://doi.org/10.1016/j.yhbeh.2018.01.001.
- Holley, A., Shalev, S., Bellevue, S., Pfaus, J.G., 2014. Conditioned mate-guarding behavior in the female rat. Physiol. Behav. 131, 136–141. https://doi.org/10.1016/ j.physbeh.2014.04.034.
- Hollis, K.L., 1990. The role of Pavlovian conditioning in territorial aggression and reproduction. In: Contemporary Issues in Comparative Psychology. Sinauer Associates, pp. 197–219. https://doi.org/10.1037/11525-009.
- Sex and context: hormones and primate sexual motivation. Horm. Behav. 40 (2), 2001, 339–357. https://doi.org/10.1006/hbeh.2001.1696.
- Estradiol: a key biological substrate mediating the response to cocaine in female rats. Horm. Behav. 58 (1), 2010, 33–43. https://doi.org/10.1016/j.yhbeh.2009.12.003.

- ISH Data: Allen Brain Atlas: mouse brain. Retrieved September 8, 2023, from. https://mouse.brain-map.org/ (n.d.).
- Hu, R.K., Zuo, Y., Ly, T., Wang, J., Meera, P., Wu, Y.E., Hong, W., 2021. An amygdala-to-hypothalamus circuit for social reward. Nat. Neurosci. 24 (6), 6 https://doi.org/10.1038/s41593-021-00828-2.
- Hughes, A.M., Everitt, B.J., Herbert, J., 1990. Comparative effects of preoptic area infusions of opioid peptides, lesions and castration on sexual behaviour in male rats: studies of instrumental behaviour, conditioned place preference and partner preference. Psychopharmacology 102 (2), 243–256. https://doi.org/10.1007/ BF02245929.
- Hunt, S., Cuthill, I.C., Swaddle, J.P., Bennett, A.T.D., 1997. Ultraviolet vision and bandcolour preferences in female zebra finches. Taeniopygia guttata. Animal Behaviour 54 (6), 1383–1392. https://doi.org/10.1006/anbe.1997.0540.
- Huston, J.P., Silva, M.A. de S., Topic, B., Müller, C.P., 2013. What's conditioned in conditioned place preference? Trends Pharmacol. Sci. 34 (3), 162–166. https://doi. org/10.1016/j.tips.2013.01.004.
- Insel, T.R., Hulihan, T.J., 1995. A gender-specific mechanism for pair bonding: oxytocin and partner preference formation in monogamous voles. Behav. Neurosci. 109 (4), 782–789. https://doi.org/10.1037/0735-7044.109.4.782.
- Ismail, N., Girard-Bériault, F., Nakanishi, S., Pfaus, J.G., 2009. Naloxone, but not flupenthixol, disrupts the development of conditioned ejaculatory preference in the male rat. Behav. Neurosci. 123 (5), 992–999. https://doi.org/10.1037/a0017096.
- Janowsky, J.S., 2006. Thinking with your gonads: testosterone and cognition. Trends Cogn. Sci. 10 (2), 77–82. https://doi.org/10.1016/j.tics.2005.12.010.
- Jennings, K.J., de Lecea, L., 2020. Neural and hormonal control of sexual behavior. Endocrinology 161 (10), bqaa150. https://doi.org/10.1210/endocr/bqaa150.
- Jirikowski, G.F., Ochs, S.D., Caldwell, J.D., 2018. Oxytocin and steroid actions. In: Hurlemann, R., Grinevich, V. (Eds.), Behavioral Pharmacology of Neuropeptides: Oxytocin. Springer International Publishing, pp. 77–95. https://doi.org/10.1007/7854-2017.9
- Johnson, D.F., Phoenix, C.H., 1976. Hormonal control of female sexual attractiveness, proceptivity, and receptivity in rhesus monkeys. J. Comp. Physiol. Psychol. 90 (5), 473–483. https://doi.org/10.1037/h0077216.
- Jorge, J.C., Velázquez, K.T., Ramos-Ortolaza, D.L., Lorenzini, I., Marrero, J., Maldonado-Vlaar, C.S., 2005. A testosterone metabolite is rewarding to ovariectomized female rats. Behav. Neurosci. 119 (5), 1222–1226. https://doi.org/10.1037/0735-7044119.5.1222
- Justice, A.J.H., de Wit, H., 1999. Acute effects of d-amphetamine during the follicular and luteal phases of the menstrual cycle in women. Psychopharmacology 145 (1), 67–75. https://doi.org/10.1007/s002130051033.
- Kelly, T.R., Kimball, M.G., Stansberry, K.R., Lattin, C.R., 2020. No, you go first: phenotype and social context affect house sparrow neophobia. Biol. Lett. 16 (9), 20200286 https://doi.org/10.1098/rsbl.2020.0286.
- Kent, K., Arientyl, V., Khachatryan, M.M., Wood, R.I., 2013. Oxytocin induces a conditioned social preference in female mice. J. Neuroendocrinol. 25 (9), 803–810. https://doi.org/10.1111/ine.12075.
- Kent, K., Butler, K., Wood, R.I., 2014. Ethanol induces conditioned social preference in male mice. Alcohol. Clin. Exp. Res. 38 (4), 1184–1192. https://doi.org/10.1111/ acer.12342
- Khalil, R., King, M.A., Soliman, M.R.I., 2005. Testosterone reverses ethanol-induced deficit in spatial reference memory in castrated rats. Pharmacology 75 (2), 87–92. https://doi.org/10.1159/000087188.
- King, B.E., Packard, M.G., Alexander, G.M., 1999. Affective properties of intra-medial preoptic area injections of testosterone in male rats. Neurosci. Lett. 269 (3), 149–152. https://doi.org/10.1016/S0304-3940(99)00440-1.
- Kippin, T.E., Pfaus, J.G., 2001. The development of olfactory conditioned ejaculatory preferences in the male rat: I. Nature of the unconditioned stimulus. Physiol. Behav. 73 (4), 457–469. https://doi.org/10.1016/S0031-9384(01)00484-X.
- Kitamura, T., Ogawa, S.K., Roy, D.S., Okuyama, T., Morrissey, M.D., Smith, L.M., Redondo, R.L., Tonegawa, S., 2017. Engrams and circuits crucial for systems consolidation of a memory. Science 356 (6333), 73–78. https://doi.org/10.1126/ science.aam6808.
- Kosaki, Y., Watanabe, S., 2016. Conditioned social preference, but not place preference, produced by intranasal oxytocin in female mice. Behav. Neurosci. 130 (2), 182–195. https://doi.org/10.1037/bne0000139.
- Kritzer, M.F., 2000. Effects of acute and chronic gonadectomy on the catecholamine innervation of the cerebral cortex in adult male rats: insensitivity of axons immunoreactive for dopamine-β-hydroxylase to gonadal steroids, and differential sensitivity of axons immunoreactive for tyrosine hydroxylase to ovarian and testicular hormones. J. Comp. Neurol. 427 (4), 617–633. https://doi.org/10.1002/ 1096-9861(20001127)427:4<617::AID-CNE9>3.0.CO:2-#.
- Kritzer, M.F., 2003. Long-term gonadectomy affects the density of tyrosine hydroxylasebut not dopamine-β-hydroxylase-, choline acetyltransferase- or serotoninimmunoreactive axons in the medial prefrontal cortices of adult male rats. Cereb. Cortex 13 (3), 282–296. https://doi.org/10.1093/cercor/13.3.282.
- Lacreuse, A., Verreault, M., Herndon, J.G., 2001. Fluctuations in spatial recognition memory across the menstrual cycle in female rhesus monkeys. Psychoneuroendocrinology 26 (6), 623–639. https://doi.org/10.1016/S0306-4530 (01)00017-8.
- László, K., Kovács, A., Zagoracz, O., Ollmann, T., Péczely, L., Kertes, E., Lacy, D.G., Lénárd, L., 2016. Positive reinforcing effect of oxytocin microinjection in the rat central nucleus of amygdala. Behav. Brain Res. 296, 279–285. https://doi.org/ 10.1016/j.bbr.2015.09.021.
- László, K., Péczely, L., Géczi, F., Kovács, A., Zagoracz, O., Ollmann, T., Kertes, E., Kállai, V., László, B., Berta, B., Karádi, Z., Lénárd, L., 2020. The role of D2 dopamine

- receptors in oxytocin induced place preference and anxiolytic effect. Horm. Behav. 124, 104777 https://doi.org/10.1016/j.yhbeh.2020.104777.
- LeBlanc, M.M., Goode, C.T., MacDougall-Shackleton, E.A., Maney, D.L., 2007. Estradiol modulates brainstem catecholaminergic cell groups and projections to the auditory forebrain in a female songbird. Brain Res. 1171, 93–103. https://doi.org/10.1016/j.brainres.2007.06.086
- Legrand, R., 1970. Successful aggression as the reinforcer for runway behavior of mice. Psychon. Sci. 20 (5), 303–305. https://doi.org/10.3758/BF03329080.
- Li, C., Brake, W.G., Romeo, R.D., Dunlop, J.C., Gordon, M., Buzescu, R., Magarinos, A.M., Allen, P.B., Greengard, P., Luine, V., McEwen, B.S., 2004. Estrogen alters hippocampal dendritic spine shape and enhances synaptic protein immunoreactivity and spatial memory in female mice. Proc. Natl. Acad. Sci. 101 (7), 2185–2190. https://doi.org/10.1073/pnas.0307313101.
- Liberzon, I., Trujillo, K.A., Akil, H., Young, E.A., 1997. Motivational properties of oxytocin in the conditioned place preference paradigm. Neuropsychopharmacology 17 (6), 6. https://doi.org/10.1016/S0893-133X(97)00070-5.
- Lim, M.M., Young, L.J., 2006. Neuropeptidergic regulation of affiliative behavior and social bonding in animals. Horm. Behav. 50 (4), 506–517. https://doi.org/10.1016/ i.vhbeh.2006.06.028.
- Love, T.M., 2014. Oxytocin, motivation and the role of dopamine. Pharmacol. Biochem. Behav. 119, 49-60. https://doi.org/10.1016/j.pbb.2013.06.011.
- Luine, V.N., Richards, S.T., Wu, V.Y., Beck, K.D., 1998. Estradiol enhances learning and memory in a spatial memory task and effects levels of monoaminergic neurotransmitters. Horm. Behav. 34 (2), 149–162. https://doi.org/10.1006/ hbeh.1998.1473.
- Maksimoski, A.N., Polzin, B.J., Stevenson, S.A., Zhao, C., Riters, L.V., 2021. μ-Opioid receptor stimulation in the nucleus accumbens increases vocal–social interactions in flocking European starlings. Sturnus Vulgaris. eNeuro 8 (5). https://doi.org/10.1523/ENEURO.0219-21.2021.
- Maksimoski, A.N., Stevenson, S.A., Polzin, B.J., Zhao, C., Luebke, E.M., Riters, L.V., 2023. The motivation to flock correlates with vocal-social behaviors and dopaminerelated gene expression in male European starlings. Horm. Behav. 153, 105374 https://doi.org/10.1016/j.yhbeh.2023.105374.
- Marler, C.A., Oyegbile, T.O., Plavicki, J., Trainor, B.C., 2005. Response to Wingfield's commentary on "A continuing saga: The role of testosterone in aggression. Hormones and Behavior 48 (3), 256–258.
- Martínez, M., Guillén-Salazar, F., Salvador, A., Simón, V.M., 1995. Successful intermale aggression and conditioned place preference in mice. Physiol. Behav. 58 (2), 323–328. https://doi.org/10.1016/0031-9384(95)00061-M.
- McHugh, R.K., Votaw, V.R., Sugarman, D.E., Greenfield, S.F., 2018. Sex and gender differences in substance use disorders. Clin. Psychol. Rev. 66, 12–23. https://doi. org/10.1016/j.cpr.2017.10.012.
- Mehrara, B.J., Baum, M.J., 1990. Naloxone disrupts the expression but not the acquisition by male rats of a conditioned place preference response for an oestrous female. Psychopharmacology 101 (1), 118–125. https://doi.org/10.1007/ RE02253728
- Meisel, R.L., Joppa, M.A., 1994. Conditioned place preference in female hamsters following aggressive or sexual encounters. Physiol. Behav. 56 (5), 1115–1118. https://doi.org/10.1016/0031-9384(94)90352-2.
- Ménard, S., Gelez, H., Girard-Bériault, F., Coria-Avila, G., Pfaus, J.G., 2019. Differential role of oxytocin and vasopressin in the conditioned ejaculatory preference of the male rat. Physiol. Behav. 208, 112577 https://doi.org/10.1016/j. physbeb 2019 112577
- Merullo, D.P., Angyal, C.S., Stevenson, S.A., Riters, L.V., 2016. Song in an affiliative context relates to the neural expression of dopamine- and neurotensin-related genes in male European starlings. Brain Behav. Evol. 88 (2), 81–92. https://doi.org/ 10.1157/000048111
- Miller, R.L., Baum, M.J., 1987. Naloxone inhibits mating and conditioned place preference for an estrous female in male rats soon after castration. Pharmacol. Biochem. Behav. 26 (4), 781–789. https://doi.org/10.1016/0091-3057(87)90611-3.
- Moaddab, M., Hyland, B.I., Brown, C.H., 2015. Oxytocin enhances the expression of morphine-induced conditioned place preference in rats. Psychoneuroendocrinology 53, 159–169. https://doi.org/10.1016/j.psyneuen.2015.01.003.
- Monari, P.K., Rieger, N.S., Schefelker, J., Marler, C.A., 2021. Intranasal oxytocin drives coordinated social approach. Sci. Rep. 11 (1), 1 https://doi.org/10.1038/s41598-021-97468-0.
- Mulligan, E.M., Lowe, M., Flynn, H., Hajcak, G., 2021. The rewards of motherhood: neural response to reward in pregnancy prospectively predicts maternal bonding with the infant in the postpartum period. Biol. Psychol. 163, 108148 https://doi. org/10.1016/j.biopsycho.2021.108148.
- Naghdi, N., Nafisy, N., Majlessi, N., 2001. The effects of intrahippocampal testosterone and flutamide on spatial localization in the Morris water maze. Brain Res. 897 (1), 44–51. https://doi.org/10.1016/S0006-8993(00)03261-3.
- Nam, M.-H., Han, K.-S., Lee, J., Won, W., Koh, W., Bae, J.Y., Woo, J., Kim, J., Kwong, E., Choi, T.-Y., Chun, H., Lee, S.E., Kim, S.-B., Park, K.D., Choi, S.-Y., Bae, Y.C., Lee, C.J., 2019. Activation of astrocytic μ-opioid receptor causes conditioned place preference. Cell Rep. 28 (5), 1154–1166.e5. https://doi.org/10.1016/j.celrep.2019.06.071.
- Nash, S., Domjan, M., 1991. Learning to discriminate the sex of conspecifics in male Japanese quail (Coturnix coturnix japonica): tests of "biological constraints". J. Exp. Psychol. Anim. Behav. Process. 17 (3), 342.
- O'Connell, L.A., Hofmann, H.A., 2011. The vertebrate mesolimbic reward system and social behavior network: a comparative synthesis. J. Comp. Neurol. 519 (18), 3599–3639. https://doi.org/10.1002/cne.22735.
- Oldenburger, W.P., Everitt, B.J., De Jonge, F.H., 1992. Conditioned place preference induced by sexual interaction in female rats. Horm. Behav. 26 (2), 214–228. https://doi.org/10.1016/0018-506X(92)90043-U.

- Packard, M.G., Cornell, A.H., Alexander, G.M., 1997. Rewarding affective properties of intra-nucleus accumbens injections of testosterone. Behav. Neurosci. 111 (1), 219–224. https://doi.org/10.1037/0735-7044.111.1.219.
- Packard, M.G., Schroeder, J.P., Alexander, G.M., 1998. Expression of testosterone conditioned place preference is blocked by peripheral or intra-accumbens injection of α-Flupenthixol. Horm. Behav. 34 (1), 39–47. https://doi.org/10.1006/ bbsh.1008.1461.
- Panksepp, J., Herman, B.H., Vilberg, T., Bishop, P., DeEskinazi, F.G., 1980. Endogenous opioids and social behavior. Neurosci. Biobehav. Rev. 4 (4), 473–487. https://doi.org/10.1016/0149-7634(80)90036-6.
- Paredes, R.G., 2009. Evaluating the neurobiology of sexual reward. ILAR J. 50 (1), 15-27. https://doi.org/10.1093/ilar.50.1.15.
- Paredes, R.G., Alonso, A., 1997. Sexual behavior regulated (paced) by the female induces conditioned place preference. Behav. Neurosci. 111 (1), 123–128. https://doi.org/ 10.1037/0735-7044.111.1.123
- Paredes, R.G., Martínez, I., 2001. Naloxone blocks place preference conditioning after paced mating in female rats. Behav. Neurosci. 115 (6), 1363–1367. https://doi.org/ 10.1037/0735-7044.115.6.1363.
- Peartree, N.A., Hood, L.E., Thiel, K.J., Sanabria, F., Pentkowski, N.S., Chandler, K.N., Neisewander, J.L., 2012. Limited physical contact through a mesh barrier is sufficient for social reward-conditioned place preference in adolescent male rats. Physiol. Behav. 105 (3), 749–756. https://doi.org/10.1016/j.physbeh.2011.10.001.
- Peris, J., MacFadyen, K., Smith, J.A., de Kloet, A.D., Wang, L., Krause, E.G., 2017. Oxytocin receptors are expressed on dopamine and glutamate neurons in the mouse ventral tegmental area that project to nucleus accumbens and other mesolimbic targets. J. Comp. Neurol. 525 (5), 1094–1108. https://doi.org/10.1002/cne.24116.
- Peris, J., Totten, K., Montgomery, D., Lester, H., Weatherington, A., Piotrowski, B., Sowell, S., Doyle, K., Scott, K., Tan, Y., MacFadyen, K.A., Engle, H., de Kloet, A.D., Krause, E.G., 2022. Conditioned social preference and reward value of activating oxytocin-receptor-expressing ventral tegmental area neurons following repeated daily binge ethanol intake. Alcohol. Clin. Exp. Res. 46 (2), 194–206. https://doi.org/10.1111/acer.14769.
- Peters, K.D., Wood, R.I., 2005. Androgen dependence in hamsters: overdose, tolerance, and potential opioidergic mechanisms. Neuroscience 130 (4), 971–981. https://doi.org/10.1016/j.neuroscience.2004.09.063.
- Petric, R., Kalcounis-Rueppell, M., Marler, C.A., 2022. Testosterone pulses paired with a location induce a place preference to the nest of a monogamous mouse under field conditions. eLife 11, e65820. https://doi.org/10.7554/eLife.65820.
- Pfaus, J.G., Kippin, T.E., Centeno, S., 2001. Conditioning and sexual behavior: a review. Horm. Behav. 40 (2), 291–321. https://doi.org/10.1006/hbeh.2001.1686.
- Pierman, S., Tirelli, E., Douhard, Q., Baum, M.J., Bakker, J., 2006. Male aromatase knockout mice acquire a conditioned place preference for cocaine but not for contact with an estrous female. Behav. Brain Res. 174 (1), 64–69. https://doi.org/10.1016/j.bbr.2006.07.002.
- Pitchers, K.K., Coppens, C.M., Beloate, L.N., Fuller, J., Van, S., Frohmader, K.S., Laviolette, S.R., Lehman, M.N., Coolen, L.M., 2014. Endogenous opioid-induced neuroplasticity of dopaminergic neurons in the ventral tegmental area influences natural and opiate reward. J. Neurosci. 34 (26), 8825–8836. https://doi.org/ 10.1523/JNEUROSCI.0133-14.2014.
- Polzin, B.J., Maksimoski, A.N., Stevenson, S.A., Zhao, C., Riters, L.V., 2022. Mu opioid receptor stimulation in the medial preoptic area or nucleus accumbens facilitates song and reward in flocking European starlings. Front. Physiol. 13 https://doi.org/ 10/3389/fphys/2022/970920
- Qi, J., Yang, J.-Y., Wang, F., Zhao, Y.-N., Song, M., Wu, C.-F., 2009. Effects of oxytocin on methamphetamine-induced conditioned place preference and the possible role of glutamatergic neurotransmission in the medial prefrontal cortex of mice in reinstatement. Neuropharmacology 56 (5), 856–865. https://doi.org/10.1016/j. neuropharm.2009.01.010.
- Quintana, G.R., Birrel, M., Marceau, S., Kalantari, N., Bowden, J., Bachoura, Y., Borduas, E., Lemay, V., Payne, J.W., Cionnaith, C.M., Pfaus, J.G., 2019. Differential disruption of conditioned ejaculatory preference in the male rat based on different sensory modalities by micro-infusions of naloxone to the medial preoptic area or ventral tegmental area. Psychopharmacology 236 (12), 3613–3623. https://doi.org/ 10.1007/s00213-019-05334-9.
- Ramos, L., Hicks, C., Caminer, A., Goodwin, J., McGregor, I.S., 2015. Oxytocin and MDMA ('Ecstasy') enhance social reward in rats. Psychopharmacology 232 (14), 2631–2641. https://doi.org/10.1007/s00213-015-3899-9.
- Rashidy-Pour, A., Bavarsad, K., Miladi-Gorji, H., Seraj, Z., Vafaei, A.A., 2019. Voluntary exercise and estradiol reverse ovariectomy-induced spatial learning and memory deficits and reduction in hippocampal brain-derived neurotrophic factor in rats. Pharmacol. Biochem. Behav. 187, 172819 https://doi.org/10.1016/j.pbb.2019.172819.
- Richard, J.E., López-Ferreras, L., Anderberg, R.H., Olandersson, K., Skibicka, K.P., 2017. Estradiol is a critical regulator of food-reward behavior. Psychoneuroendocrinology 78, 193–202. https://doi.org/10.1016/j.psyneuen.2017.01.014.
- Rieger, N.S., Stanton, E.H., Marler, C.A., 2019. Division of labour in territorial defence and pup retrieval by pair-bonded California mice, Peromyscus californicus. Anim. Behav. 156, 67–78. https://doi.org/10.1016/j.anbehav.2019.05.023.
- Rieger, N.S., Monari, P.K., Hartfield, K., Schefelker, J., Marler, C.A., 2021. Pair-bonding leads to convergence in approach behavior to conspecific vocalizations in California mice (*Peromyscus californicus*). PLoS One 16 (8), e0255295. https://doi.org/ 10.1371/journal.pone.0255295.
- Rieger, N.S., Guoynes, C.D., Monari, P.K., Hammond, E.R., Malone, C.L., Marler, C.A., 2022. Neuroendocrine mechanisms of aggression in rodents. Motiv. Sci. 8 (2), 81–105. https://doi.org/10.1037/mot0000260.

- Riters, L.V., 2012. The role of motivation and reward neural systems in vocal communication in songbirds. Front. Neuroendocrinol. 33 (2), 194–209. https://doi. org/10.1016/j.yfrne.2012.04.002.
- Riters, L.V., Stevenson, S.A., 2012. Reward and vocal production: song-associated place preference in songbirds. Physiol. Behav. 106 (2), 87–94. https://doi.org/10.1016/j. physbeb.2012.01.010.
- Riters, L.V., Stevenson, S.A., 2022. Using seasonality and birdsong to understand mechanisms underlying context-appropriate shifts in social motivation and reward. Horm. Behav. 142, 105156 https://doi.org/10.1016/j.yhbeh.2022.105156.
- Riters, L.V., Ellis, J.M.S., Angyal, C.S., Borkowski, V.J., Cordes, M.A., Stevenson, S.A., 2013. Links between breeding readiness, opioid immunolabeling, and the affective state induced by hearing male courtship song in female European starlings (Sturnus vulgaris). Behav. Brain Res. 247, 117–124. https://doi.org/10.1016/j.bbr.2013.02.041.
- Riters, L.V., Stevenson, S.A., DeVries, M.S., Cordes, M.A., 2014. Reward associated with singing behavior correlates with opioid-related gene expression in the medial preoptic nucleus in male European starlings. PLoS One 9 (12), e115285. https://doi. org/10.1371/journal.pone.0115285.
- Riters, L.V., Cordes, M.A., Stevenson, S.A., 2017. Prodynorphin and kappa opioid receptor mRNA expression in the brain relates to social status and behavior in male European starlings. Behav. Brain Res. 320, 37–47. https://doi.org/10.1016/j. bbr.2016.11.056
- Riters, L.V., Kelm-Nelson, C.A., Spool, J.A., 2019. Why do birds flock? A role for opioids in the reinforcement of gregarious social interactions. Front. Physiol. 10 https://doi. org/10.3389/fphys.2019.00421.
- Roof, R.L., Havens, M.D., 1992. Testosterone improves maze performance and induces development of a male hippocampus in females. Brain Res. 572 (1), 310–313. https://doi.org/10.1016/0006-8993(92)90491-0.
- Rosellini, R.A., Svare, B.B., Rhodes, M.E., Frye, C.A., 2001. The testosterone metabolite and neurosteroid 3α-androstanediol may mediate the effects of testosterone on conditioned place preference. Brain Res. Rev. 37 (1), 162–171. https://doi.org/10.1016/S0165-0173(01)00116-3.
- Ross, H.E., Cole, C.D., Smith, Y., Neumann, I.D., Landgraf, R., Murphy, A.Z., Young, L.J., 2009. Characterization of the oxytocin system regulating affiliative behavior in female prairie voles. Neuroscience 162 (4), 892–903. https://doi.org/10.1016/j. neuroscience.2009.05.055.
- Rubalcaba, J.G., Polo, V., Maia, R., Rubenstein, D.R., Veiga, J.P., 2016. Sexual and natural selection in the evolution of extended phenotypes: the use of green nesting material in starlings. J. Evol. Biol. 29 (8), 1585–1592. https://doi.org/10.1111/ ieb.12803
- Rudebeck, P.H., Izquierdo, A., 2022. Foraging with the frontal cortex: a cross-species evaluation of reward-guided behavior. Neuropsychopharmacology 47 (1), 1. https://doi.org/10.1038/s41386-021-01140-0.
- Russo, S.J., Nestler, E.J., 2013. The brain reward circuitry in mood disorders. Nat. Rev. Neurosci. 14 (9), 9 https://doi.org/10.1038/nrn3381.
- Russo, S.J., Festa, E.D., Fabian, S.J., Gazi, F.M., Kraish, M., Jenab, S., Quiñones-Jenab, V., 2003. Gonadal hormones differentially modulate cocaine-induced conditioned place preference in male and female rats. Neuroscience 120 (2), 523–533. https://doi.org/10.1016/S0306-4522(03)00317-8.
- Salole, E.G., 1986. Estradiol. In: Florey, K. (Ed.), Analytical Profiles of Drug Substances, vol. 15. Academic Press, pp. 283–318. https://doi.org/10.1016/S0099-5428(08) 60416-5.
- Schroeder, J.P., Packard, M.G., 2000. Role of dopamine receptor subtypes in the acquisition of a testosterone conditioned place preference in rats. Neurosci. Lett. 282 (1), 17–20. https://doi.org/10.1016/S0304-3940(00)00839-9.
- Shamay-Tsoory, S.G., Abu-Akel, A., 2016. The social salience hypothesis of oxytocin. Biol. Psychiatry 79 (3), 194–202. https://doi.org/10.1016/j.biopsych.2015.07.020.
- Sherwin, B.B., 2003. Steroid hormones and cognitive functioning in aging men. J. Mol. Neurosci. 20 (3), 385–393. https://doi.org/10.1385/JMN:20:3:385.
- Silverman, J., Koenig, J., 2007. Evidence for the involvement of ER $\beta$  and RGS9-2 in 17- $\beta$  estradiol enhancement of amphetamine-induced place preference behavior. Horm. Behav. 52 (2), 146–155. https://doi.org/10.1016/j.yhbeh.2007.03.017.
- Simerly, R.B., McCall, L.D., Watson, S.J., 1988. Distribution of opioid peptides in the preoptic region: immunohistochemical evidence for a steroid-sensitive enkephalin sexual dimorphism. J. Comp. Neurol. 276 (3), 442–459. https://doi.org/10.1002/ cne 902760309
- Skuse, D.H., Gallagher, L., 2009. Dopaminergic-neuropeptide interactions in the social brain. Trends Cogn. Sci. 13 (1), 27–35. https://doi.org/10.1016/j.tics.2008.09.007.
- Smiley, K.O., Lipshutz, S.E., Kimmitt, A.A., DeVries, M.S., Cain, K.E., George, E.M., Covino, Kristen, M., 2022. Beyond a biased binary: a perspective on the misconceptions, challenges, and implications of studying females in avian behavioral endocrinology. Front. Physiol. 13 https://doi.org/10.3389/fphys.2022.970603.
- Song, Z., Albers, H.E., 2018. Cross-talk among oxytocin and arginine-vasopressin receptors: relevance for basic and clinical studies of the brain and periphery. Front. Neuroendocrinol. 51, 14–24. https://doi.org/10.1016/j.yfrne.2017.10.004.
- Song, Z., McCann, K.E., McNeill, J.K., Larkin, T.E., Huhman, K.L., Albers, H.E., 2014. Oxytocin induces social communication by activating arginine-vasopressin V1a receptors and not oxytocin receptors. Psychoneuroendocrinology 50, 14–19. https://doi.org/10.1016/j.psyneuen.2014.08.005.
- Song, Z., Borland, J.M., Larkin, T.E., O'Malley, M., Albers, H.E., 2016. Activation of oxytocin receptors, but not arginine-vasopressin V1a receptors, in the ventral tegmental area of male Syrian hamsters is essential for the reward-like properties of social interactions. Psychoneuroendocrinology 74, 164–172. https://doi.org/ 10.1016/j.psyneuen.2016.09.001.
- Spiteri, T., Musatov, S., Ogawa, S., Ribeiro, A., Pfaff, D.W., Ågmo, A., 2009. Estrogeninduced sexual incentive motivation, proceptivity and receptivity depend on a

- functional estrogen receptor  $\alpha$  in the ventromedial nucleus of the hypothalamus but not in the amygdala. Neuroendocrinology 91 (2), 142–154. https://doi.org/
- Spool, J.A., Riters, L.V., 2017. Associations between environmental resources and the "Wanting" and "Liking" of male song in female songbirds. Integr. Comp. Biol. 57 (4), 835–845. https://doi.org/10.1093/icb/icx117.
- Spool, J.A., Stevenson, S.A., Angyal, C.S., Riters, L.V., 2016. Contributions of testosterone and territory ownership to sexually-motivated behaviors and mRNA expression in the medial preoptic area of male European starlings. Horm. Behav. 86, 36–44. https://doi.org/10.1016/j.yhbeh.2016.09.004.
- Spool, J.A., Jay, M.D., Riters, L.V., 2018. Nest box exploration may stimulate breeding physiology and alter mRNA expression in the medial preoptic area of female European starlings. J. Exp. Biol. 221 (11), jeb174441 https://doi.org/10.1242/ jeb.174441.
- Stevenson, S.A., Piepenburg, A., Spool, J.A., Angyal, C.S., Hahn, A.H., Zhao, C., Riters, L. V., 2020. Endogenous opioids facilitate intrinsically-rewarded birdsong. Sci. Rep. 10 (1), 1 https://doi.org/10.1038/s41598-020-67684-1.
- Sumida, K., Walf, A.A., Frye, C.A., 2005. Progestin-facilitated lordosis of hamsters may involve dopamine-like type 1 receptors in the ventral tegmental area. Behav. Brain Res. 161 (1), 1–7. https://doi.org/10.1016/j.bbr.2005.02.013.
- Tabatadze, N., Sato, S.M., Woolley, C.S., 2014. Quantitative analysis of long-form aromatase mRNA in the male and female rat brain. PLoS One 9 (7), e100628. https://doi.org/10.1371/journal.pone.0100628.
- Tellegen, A., Horn, J.M., 1972. Primary aggressive motivation in three inbred strains of mice. J. Comp. Physiol. Psychol. 78 (2), 297–304. https://doi.org/10.1037/b0033192
- Tellegen, A., Horn, J.M., Legrand, R.G., 1969. Opportunity for aggression as a reinforcer in mice. Psychon. Sci. 14 (3), 104–105. https://doi.org/10.3758/BF03332727.
- Tenhola, H., Sinclair, D., Alho, H., Lahti, T., 2012. Effect of opioid antagonists on sex hormone secretion. J. Endocrinol. Investig. 35 (2), 227–230. https://doi.org/ 10.3275/8181.
- Thiel, K.J., Okun, A.C., Neisewander, J.L., 2008. Social reward-conditioned place preference: a model revealing an interaction between cocaine and social context rewards in rats. Drug Alcohol Depend. 96 (3), 202–212. https://doi.org/10.1016/j. drugalcdep.2008.02.013.
- Timonin, M.E., Kalcounis-Rueppell, M.C., Marler, C.A., 2018. Testosterone pulses at the nest site modify ultrasonic vocalization types in a monogamous and territorial mouse. Ethology 124 (11), 804–815. https://doi.org/10.1111/eth.12812.
- Tonn Eisinger, K.R., Larson, E.B., Boulware, M.I., Thomas, M.J., Mermelstein, P.G., 2018.
  Membrane estrogen receptor signaling impacts the reward circuitry of the female brain to influence motivated behaviors. Steroids 133, 53–59. https://doi.org/10.1016/j.steroids.2017.11.013.
- Trainor, B.C., Marler, C.A., 2001. Testosterone, paternal behavior, and aggression in the monogamous California mouse (*Peromyscus californicus*). Horm. Behav. 40 (1), 32–42. https://doi.org/10.1006/hbeh.2001.1652.
- Trezza, V., Damsteegt, R., Vanderschuren, L.J.M.J., 2009. Conditioned place preference induced by social play behavior: parametrics, extinction, reinstatement and disruption by methylphenidate. Eur. Neuropsychopharmacol. 19 (9), 659–669. https://doi.org/10.1016/j.euroneuro.2009.03.006.
- Trezza, V., Baarendse, P.J.J., Vanderschuren, L.J.M.J., 2010. The pleasures of play: pharmacological insights into social reward mechanisms. Trends Pharmacol. Sci. 31 (10), 463–469. https://doi.org/10.1016/j.tips.2010.06.008.
- Tribollet, E., Barberis, C., Jard, S., Dubois-Dauphin, M., Dreifuss, J.J., 1988. Localization and pharmacological characterization of high affinity binding sites for vasopressin and oxytocin in the rat brain by light microscopic autoradiography. Brain Res. 442 (1), 105–118. https://doi.org/10.1016/0006-8993(88)91437-0.
- Tribollet, E., Barberis, C., Dubois-Dauphin, M., Dreifuss, J.J., 1992. Localization and characterization of binding sites for vasopressin and oxytocin in the brain of the guinea pig. Brain Res. 589 (1), 15–23. https://doi.org/10.1016/0006-8993(92) 01156.0
- Tzschentke, T.M., 2007. REVIEW ON CPP: measuring reward with the conditioned place preference (CPP) paradigm: update of the last decade. Addict. Biol. 12 (3–4), 227–462. https://doi.org/10.1111/j.1369-1600.2007.00070.x.
- Ulloa, M., Portillo, W., Díaz, N.F., Young, L.J., Camacho, F.J., Rodríguez, V.M., Paredes, R.G., 2018. Mating and social exposure induces an opioid-dependent conditioned place preference in male but not in female prairie voles (*Microtus ochrogaster*). Horm. Behav. 97, 47–55. https://doi.org/10.1016/j. yhbeh.2017.10.015.
- Van Der Kooy, D., Mucha, R.F., O'Shaughnessy, M., Bucenieks, P., 1982. Reinforcing effects of brain microinjections of morphine revealed by conditioned place preference. Brain Res. 243 (1), 107–117. https://doi.org/10.1016/0006-8993(82) 91124-6
- Vandegrift, B.J., You, C., Satta, R., Brodie, M.S., Lasek, A.W., 2017. Estradiol increases the sensitivity of ventral tegmental area dopamine neurons to dopamine and ethanol. PLoS One 12 (11), e0187698. https://doi.org/10.1371/journal. pope 0187698
- Vanderschuren, L.J.M.J., Niesink, R.J.M., Van Pee, J.M., 1997. The neurobiology of social play behavior in rats. Neurosci. Biobehav. Rev. 21 (3), 309–326. https://doi.org/10.1016/S0149-7634(96)00020-6.
- Vann, S.D., Albasser, M.M., 2011. Hippocampus and neocortex: recognition and spatial memory. Curr. Opin. Neurobiol. 21 (3), 440–445. https://doi.org/10.1016/j. conb.2011.02.002.
- Watson, R.E., Hoffmann, G.E., Wiegand, S.J., 1986. Sexually dimorphic opioid distribution in the preoptic area: manipulation by gonadal steroids. Brain Res. 398 (1), 157–163. https://doi.org/10.1016/0006-8993(86)91261-8.

- Williams, C.L., Meck, W.H., 1991. The organizational effects of gonadal steroids on sexually dimorphic spatial ability. Psychoneuroendocrinology 16 (1), 155–176. https://doi.org/10.1016/0306-4530(91)90076-6.
- Wood, R.I., 2008. Anabolic–androgenic steroid dependence? Insights from animals and humans. Front. Neuroendocrinol. 29 (4), 490–506. https://doi.org/10.1016/j. yfrne.2007.12.002.
- Wood, R.I., Rice, R., 2013. Ethanol-induced conditioned partner preference in female mice. Behav. Brain Res. 243, 273–277. https://doi.org/10.1016/j.bbr.2013.01.021.
- Wood, R.I., Johnson, L.R., Chu, L., Schad, C., Self, D.W., 2004. Testosterone reinforcement: intravenous and intracerebroventricular self-administration in male rats and hamsters. Psychopharmacology 171 (3), 298–305. https://doi.org/ 10.1007/s00213-003-1587-7.
- Wood, R.I., Knoll, A.T., Levitt, P., 2015. Social housing conditions and oxytocin and vasopressin receptors contribute to ethanol conditioned social preference in female mice. Physiol. Behav. 151, 469–477. https://doi.org/10.1016/j. physbeh.2015.08.018.
- Woods, J.K., Deviche, P., Corbitt, C., 2010. Opioid receptor densities analyzed across seasons in the POM and VTA of the dark-eyed junco, *Junco hyemalis*. J. Chem. Neuroanat. 40 (2), 123–129. https://doi.org/10.1016/j.jchemneu.2010.05.002.
- Yague, J.G., Azcoitia, I., DeFelipe, J., Garcia-Segura, L.M., Muñoz, A., 2010. Aromatase expression in the normal and epileptic human hippocampus. Brain Res. 1315, 41–52. https://doi.org/10.1016/j.brainres.2009.09.111.
- Yin, L., Lin, D., 2023. Neural control of female sexual behaviors. Horm. Behav. 151, 105339 https://doi.org/10.1016/j.yhbeh.2023.105339.

- Yoest, K.E., Cummings, J.A., Becker, J.B., 2014. Estradiol, dopamine and motivation. Central Nervous System Agents in Medicinal ChemistryChemistry - Central Nervous System Agents 14 (2), 83–89.
- Young, L.J., Wang, Z., 2004. The neurobiology of pair bonding. Nat. Neurosci. 7 (10), 10 https://doi.org/10.1038/nn1327.
- Yu, C., Moss, C.F., 2022. Natural acoustic stimuli evoke selective responses in the hippocampus of passive listening bats. Hippocampus 32 (4), 298–309. https://doi. org/10.1002/hipp.23407.
- Zernig, G., Pinheiro, B.S., 2015. Dyadic social interaction inhibits cocaine-conditioned place preference and the associated activation of the accumbens corridor. Behav. Pharmacol. 26 (6), 580–594. https://doi.org/10.1097/FBP.00000000000000167.
- Zhao, X., Marler, C.A., 2014. Pair bonding prevents reinforcing effects of testosterone in male California mice in an unfamiliar environment. Proc. R. Soc. B Biol. Sci. 281 (1788), 20140985 https://doi.org/10.1098/rspb.2014.0985.
- Zhao, X., Marler, C.A., 2016. Social and physical environments as a source of individual variation in the rewarding effects of testosterone in male California mice (*Peromyscus californicus*). Horm. Behav. 85, 30–35. https://doi.org/10.1016/j. physby 2016.07.007
- Zhao, X., Castelli, F.R., Wang, R., Auger, A.P., Marler, C.A., 2020a. Testosterone-related behavioral and neural mechanisms associated with location preferences: a model for territorial establishment. Horm. Behav. 121, 104709 https://doi.org/10.1016/j. vhbeh.2020.104709.
- Zhao, C., Chang, L., Auger, A.P., Gammie, S.C., Riters, L.V., 2020b. Mu opioid receptors in the medial preoptic area govern social play behavior in adolescent male rats. Genes Brain Behav. 19 (7), e12662 https://doi.org/10.1111/gbb.12662.