

1 **Editor summary:**

2 In this Perspective, the authors develop a risk assessment framework for forest microbiomes
3 under climate change that unites microbial and forest ecology. They define processes that
4 amplify or buffer microbial sensitivity and exposure risk and feedbacks that mediate impacts on
5 microbial communities.

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9

Title: A Risk Assessment Framework for the Future of Forest Microbiomes in a Changing Climate

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Abstract: Microbes inhabiting the above and belowground tissues of forest trees and soils play a critical role in the response of forest ecosystems to global climate change. However, generalizations about the vulnerability of the forest microbiome to climate change have been challenging due to often context-dependent responses. Here we apply a risk assessment framework to evaluate microbial community vulnerability to climate change across forest ecosystems. We define factors that determine exposure risk and processes that amplify or buffer sensitivity to change and describe feedback mechanisms that will modulate this exposure and sensitivity as climatic change progresses. This risk assessment approach unites microbial ecology and forest ecology to develop a more comprehensive understanding of forest vulnerability in the 21st century.

Resumen (Español): Los microorganismos que habitan en los tejidos aéreos y subterráneos de los árboles y suelos forestales desempeñan un papel fundamental en la respuesta de los ecosistemas forestales al cambio climático. Sin embargo, las generalizaciones sobre la respuesta del microbioma forestal al cambio climático ha sido un reto debido a que las respuestas son contexto-dependientes. Aquí aplicamos un marco de evaluación de riesgos para evaluar la vulnerabilidad de la comunidad microbiana al cambio climático en los ecosistemas forestales. Definimos los factores que determinan el riesgo a la exposición y los procesos que amplifican o amortiguan la sensibilidad al cambio climático, y describimos los mecanismos de retroalimentación que modularán la exposición y sensibilidad a medida que avance el cambio climático. Esta aproximación de evaluación de riesgos une a la ecología microbiana y la ecología forestal para desarrollar una comprensión más completa de la vulnerabilidad de los bosques en el siglo XXI.

Resumo (Português): Os microrganismos que habitam o solo e os tecidos das árvores acima e abaixo do solo desempenham um papel crítico na resposta dos ecossistemas florestais às mudanças climáticas globais. No entanto, generalizações sobre a resposta do microbioma florestal às alterações climáticas têm sido um desafio devido às frequentes respostas dependentes do contexto. Aqui, nós aplicamos uma estrutura de avaliação de risco para avaliar a vulnerabilidade da comunidade microbiana às mudanças climáticas em ecossistemas florestais. Nós definimos os fatores que determinam o risco de exposição e os processos que amplificam ou atenuam a sensibilidade à mudança, e descrevemos os mecanismos de retroalimentação que modularão essa exposição e sensibilidade à medida que as alterações climáticas progridem. Essa abordagem de avaliação de risco une a ecologia microbiana e a ecologia florestal para desenvolver uma compreensão mais abrangente da vulnerabilidade das florestas no século XXI.

11 Introduction

12 Forest ecosystems cover approximately one-third of the Earth's land surface, harboring immense
13 biodiversity¹ and supporting diverse human cultural needs and values². Forests sequester over
14 three-quarters of terrestrial carbon globally³, however, due to climate change and land use practices
15 in the "Capitalocene"⁴, the functioning and persistence of forest ecosystems represents an area of
16 critical scientific uncertainty^{5,6}. Investigations of forest response to climate change have largely
17 focused on tree demography and ecophysiology, satellite measurements, and restructuring of forest
18 community structure and function. Collectively, these studies indicate that fire, drought, increasing
19 CO₂ concentrations, and rising temperatures imperil the health and carbon sequestration potential
20 of forest ecosystems^{5,6}. Improving understanding of forest responses to climate change is critical
21 for effective policy and climate adaptation initiatives.

22 The forest microbiome mediates forest responses to a number of climatic stressors
23 including drought⁷, rising temperatures⁸, and elevated CO₂⁹. Microbiomes also play an important
24 role in forest recovery from fire^{10,11} and hurricanes¹², and microbes shape soil methane production
25 and consumption¹³, as well as the ability of soils to sequester CO₂¹⁴. There is now also unequivocal
26 evidence that forest microbial communities are vulnerable to climate change^{15–18}, highlighting the
27 need for their explicit consideration in efforts to predict the fate of forest ecosystems.
28 Observational studies across local to global-scale environmental gradients document how
29 microbial community composition and function are locally adapted, and highly sensitive to
30 prevailing climatological conditions^{17,19–22}. In addition to observational inferences, experimental
31 studies of microbial communities across diverse forest environments demonstrate dynamic
32 compositional and functional responses to climatic disturbances¹⁸. Because rates of microbial
33 responses to disturbance may differ from forest trees, mismatches between plant communities,
34 local-climates, and microbial communities can occur, with important implications for forest
35 function^{23,24}.

36 Forest microbes often exhibit strong context- and scale-dependent responses to
37 disturbance^{25–27}, challenging generalizations of microbiome response to climate change.
38 Accordingly, these context dependencies have stymied the effective incorporation of microbial
39 dynamics into predictive models and understanding of forest ecosystems^{5,6,27,28}. To address this
40 knowledge gap and better predict and integrate microbiomes into a more holistic understanding of

forest response to climate change, we adapt a risk assessment framework for forest microbiomes in the Anthropocene.

Defining forests and their microbiomes

The Kyoto Protocol (1997) categorizes forests as areas greater than 0.5-1.0 hectares where the minimum “tree” crown cover ranges from 10-30%²⁹. From the boreal to the Amazon, forest ecosystems span broad climatic regions⁶. Throughout these regions, trees scaffold and shape forest systems, giving rise to distinct habitats and microclimates for understory species³⁰. Trees are generally defined as plants capable of growing at least two meters tall²⁹, though their stature and architecture varies greatly by species³¹. Lifespans of trees are also highly variable across taxa, ranging from 25 to over 5000 years³². The architectural, demographic, and ecophysiological characteristics of trees are predicted to play important roles in forest responses to climate change^{5,6}.

The forest microbiome – a rich diversity of bacteria, archaea, fungi, lichens, and viruses – supports plant productivity and forest function across the globe^{33–35} (Fig. 1a). Forest ecosystems are among the most biodiverse microbial habitats on Earth. For example, individual leaves can host hundreds of bacterial and fungal taxa³⁶ and tens of thousands of microbial species inhabit forest soils³⁷. Forest microbes span a range of interaction types, including pathogens and mutualists. These microbes cumulatively facilitate nutrient cycling^{35,38} and impact plant community structure by mediating plant coexistence and diversity^{39–41}. Key microbial groups in soil include saprotrophic fungi and bacteria, which drive decomposition and soil carbon accumulation⁴². Saprotrophic microbes mediate soil nutrient availability by mineralizing organic nitrogen into plant-available inorganic forms^{42,43}. Other key groups include ancient interactions between tree roots and mutualistic mycorrhizal fungi, such as arbuscular (AMF) and ectomycorrhizal fungi (EMF), which shape host environmental tolerances⁸ and represent an important component of nutrient acquisition strategies for trees¹⁹ (Fig. 1c). Similarly, mutualistic relationships between roots of certain tree species and N-fixing bacteria influence biogeochemical cycling in forest systems³⁵. Fungal and bacterial endophytes in roots and leaves can protect trees against pathogens³⁶, promote drought tolerance⁴⁴, influence host thermal tolerance³⁶, and contribute to host N supply through N-fixation⁴⁵ (Figure 1a). N-fixing bacteria have also been found in forest soils, leaves, and leaf litter, as well as in association with mosses and lichens, contributing to forest N-availability and biogeochemical cycling⁴⁶. In this review, we especially

focus on the bacterial and fungal components of the forest microbiome given available knowledge and their pivotal role in forest function³⁴.

A risk assessment framework for forest microbiomes

Risk assessments are used to predict and manage the vulnerability of diverse systems by providing insights into where exposure may be reduced or sensitivity might be mitigated^{6,47}. Because risk frameworks have predominately been used in macroecological contexts⁴⁷, to utilize this framework, we first define microbially-relevant scales of exposure and attributes of microbial communities and individuals that govern their sensitivity to climatic disturbance (Fig. 2)^{15,47,48}. Then, we define important feedbacks that can inform the trajectory of forest microbiome responses to ongoing climate change. This framework intends to synthesize forest microbiome response to climate change in order to unite forest microbial ecology with macroecological understanding of forest responses to climate change⁶. While we specifically apply the framework in a forest ecosystem context, the principles should also be broadly applicable, and the framework is intended for diverse terrestrial ecosystems.

Exposure

The effects of warming, drought, and other climatic disturbances vary across geographic and temporal scales⁶. We define exposure risk as the probability that climatic disturbance will extend beyond physical baseline conditions enough to impact forest microbiome community structure or function. Accordingly, quantifying climatic exposure risk for the forest microbiome depends on the integration of three different components of climatic disturbance: disturbance type, spatiotemporal scales of disturbance, and the capacity for physical buffering of disturbance across microbial habitats.

Categorizing climatic disturbances

The persistence of forest systems is threatened by the increasing frequency and severity of climatic disturbance⁶. Different types of perturbations associated with a changing climate can be categorized in a pulse-pressure framework to understand their potential impact across spatiotemporal scales and intensity of disturbance^{15,49}. Disturbances like elevated CO₂ (eCO₂) and rising air

temperatures represent press events, disturbances which are ongoing and relatively slow⁴⁹. Press disturbances are predicted to select for microbial traits associated with greater physiological plasticity and niche breadth (tolerance), and strong acclimation or rapid adaptation will be required to maintain community composition as a result of continuing disturbance^{15,16,50}. Conversely, pulse events are discrete disturbances, including droughts, hurricanes, floods, or wildfires⁴⁹. Pulse disturbances tend to favor microbial community traits associated with rapid recovery (often referred to as resilience)¹⁵. Though pulse disturbances like forest fires can initially reduce soil fungal and bacterial biomass and diversity^{51,52}, pulse disturbances have a defined end period, in some cases allowing microbial communities to eventually recover parts of their original structure and function¹⁵. However, recovery of microbiome function following pulse events is variable and can take as long as several decades¹⁰. While the pulse-press framework is conceptually useful for comparing and synthesizing the physical impacts of diverse climatic stressors on organisms and communities, it is important to note that both press and pulse disturbances increasingly act in tandem⁵³.

Consideration of microbial lifespan and life-history strategies are essential to contextualizing exposure risk¹⁵. For instance, some long-lived soil fungi can persist for years to decades⁵⁴, whereas many co-occurring soil bacteria turnover on the order of hours or days in response to drought⁵⁵. As a result, while a multi-month drought may represent a pulse disturbance for a tree or long-lived fungus, droughts typically impact many generations of more ephemeral fungi or bacteria¹⁵. Consequently, bacteria and some fungi may undergo rapid evolution in response to pulse disturbances, whereas evolution for some fungi may occur over longer timescales⁵⁶.

Climatic disturbances can act either directly on microbial cells, or through indirect pathways. For example, drought and rising temperatures can directly stress microbial cells^{55,57}. In contrast, eCO₂ indirectly impacts soil communities, primarily via shifts in substrate-availability^{58,59}. Greater C allocation of trees belowground in response to eCO₂ can increase colonization of roots by both EMF and AMF fungi, and result in greater rhizodeposition, increasing availability of carbon-substrates to microbes^{60,61}. Tracing the various pathways whereby climatic stressors directly or indirectly impact microbial communities will be critical to accurately projecting the magnitude of community responses.

The spatial and temporal nature of climatic disturbances

The type, intensity, frequency, and timing of disturbances varies dramatically across forest biomes^{6,18} (Fig. 3). Catastrophic droughts, flooding, and severe hurricanes affect tropical forests, while reduced snowpack and permafrost melt threaten boreal and high-elevation forest systems⁶². In many temperate forests, drought and rising temperatures compound with settler-colonial fire suppression to create fires of increasing frequency and intensity⁶³. Increasing frequency and spatial extent of disturbance threaten the capacity of microbial communities to withstand and/or recover their pre-disturbance community structure⁶⁴ and shifts in pathogen ranges, including bark beetle and invasive microbial diseases, can result in mass tree mortality, jeopardizing forest persistence in some systems^{7,65}. Importantly, the intensity of disturbance will also vary across forest biomes. The rate of warming, for instance, differs across the globe with boreal forest temperatures projected to increase at approximately twice the rate of other forests⁶⁶ (Fig. 3a-b). As a result, microbiome exposure to warming is likely greater in boreal compared to temperate and tropical forest systems⁶².

For pulse-type disturbances, the timing of a disturbance also influences microbiome exposure risk, especially for forests with strong seasonality (Fig. 3c). For example, spring fires result in greater reductions in fungal saprotrophic diversity compared to fires of similar severity which occur in autumn⁶⁷. One hypothesis is that spring fires are more damaging for microbial growth and reproduction than fires that occur during periods of higher microbial dormancy⁶⁷. Climate change will continue to advance the onset of summer and spring, while autumn and winter seasons are expected to be shortened and delayed⁶⁸. Thus, longer periods of microbial activity may increase the probability of greater impact of pulse disturbances on microbial community structure and function. Additionally, autumn is an especially critical ecological period, especially for many fungi, as this is when they transition from high periods of activity to reproduction⁶⁹. Scarce datasets in Western science track long-term shifts in microbial reproduction, making generalizations about shifts in fruiting phenology associated with climate change challenging^{70,71}. However, many Indigenous communities have been observing patterns and processes of fungal phenology for thousands of years⁷². For example, the Karuk tribe (Northern California, USA) has demonstrated how delays in autumn (warmer temperatures continuing into October/November and later rainfall) lead to declines in ectomycorrhizal fruiting, with important consequences for the communities and ecosystems these fungi help to sustain⁷³ (Box 1). Indigenous science has a critical lens into how

climate change is shaping forest ecosystems as its period of observation extends back tens of thousands of years; the centering of Indigenous-led science will be critical in better understanding and managing forest systems in a changing climate⁷⁴.

Microclimates, microhabitats, and microbe-specific niches

The physical structure of forest ecosystems plays an underappreciated role in determining the exposure risk of different microbial communities to climatic disturbance (Fig. 3d). Due to their immense structural complexity relative to other terrestrial ecosystems, different locations in forests confer varying degrees of physical buffering from climatic disturbance³⁰. Throughout the forest canopy, light, temperature, and moisture conditions vary dramatically^{75,76}. Microbes associated with different forest compartments (e.g. leaves versus roots) experience varying degrees of physical buffering (Fig. 3d-e) and have different inherent exposure risks to climatic disturbances. To distinguish these from the physical structures that hosts can evolve to spatially separate symbionts (compartmentalization), herein, we refer to these different microbially habitats (leaves, roots, soil, etc.) as microbial-specific niches (MSNs)³⁴.

Plant hosts can also actively and passively modulate microbiome exposure. For example, bacterial communities associated with roots of *Sequoia sempervirens* are less sensitive to water-deficit relative to those in soil, potentially due to consistent water availability in roots²². In *Eucalyptus*, latent cooling through evapotranspiration reduced leaf temperatures by up to 7.5°C relative to air temperatures during a heatwave⁷⁷, helping to buffer heat exposure for phyllosphere and endophyte microbial communities. Active cooling via evapotranspiration may also help to explain how foliar endophyte communities associated with *Pinus ponderosa* can maintain community structure whereas root-associated microbial communities are reshaped by warmer temperatures⁷⁸. The soil matrix can also buffer microbiome exposure. For example, the composition and diversity of bacterial and fungal communities in the soil surface (0-5 cm depth) are often strongly impacted by fire, but heat from surface fires attenuates rapidly with soil depth, suggesting that deeper soils may act as a potential refugia for microbial communities⁷⁹ (Fig. 3e).

Canopy gaps also create distinct microclimates (Fig. 3d). These gaps are relatively warmer and drier, and consequentially, distinct wood decay fungal communities inhabiting them exhibit reduced rates of decomposition⁸⁰; similarly, canopy gaps may reduce the local diversity of EMF communities⁸¹. In xeric systems, removal and management of understory plants through practices

such as cultural burning can increase soil water content and nutrient availability⁸², providing important habitat for plant and microbial communities by reducing understory plant competition⁸³. However, extensive habitat fragmentation (due to logging or large-scale climatic disturbances like stand-replacing fires or hurricanes) may exacerbate forest warming through reduced shading and evapotranspiration, further threatening forest communities. Altogether, the unique buffering capacity of distinct MSNs, suggests that microbiome community structure and function could shift at different rates across leaf, root, and soil communities potentially causing dysbiosis for tree hosts⁸⁴. Exploring how different MSN combine to collectively influence plant health remains an intriguing area of future research.

Sensitivity

In the broadest terms, sensitivity describes the intrinsic factors that determine how a community, species, or individual is impacted by climatic disturbance⁴⁷. At the community scale, sensitivity describes the degree to which microbial community composition or function responds to disturbance and their capacity to recover composition or function post-disturbance (Fig. 4a)⁸⁵. At the species and individual scales, low sensitivity is the persistence of a taxon or individual or their capacity to recover physiological function following a short-term disturbance⁸⁵. Here, we identify and describe evolutionary and ecological processes that modulate the sensitivity of microbial populations and communities.

Global variation in the sensitivity of microbial communities

Surveys across continental and global scales have identified strong biogeographic patterns in microbial communities, with high degrees of endemism (for some taxonomic groups) and unique functional trait profiles across the major tropical, temperate, and boreal forest biomes²¹. This suggests that climatic disturbances may result in divergent responses for microbial communities inhabiting different biomes or habitats. The biogeographic patterns in microbial communities result from prevailing climatic regimes (e.g. mean annual temperature; MAT)²¹, geochemical attributes (e.g. parent material, pH, Ca)⁸⁶, different evolutionary origins of microbial taxa⁸⁷, and the composition of forest tree species^{88,89}. Additionally, limitations to microbial dispersal can drive distinct community⁹⁰ and population structures⁹¹.

In the face of climatic change, varying sensitivity of microbial communities can result in several response scenarios: tolerance (resistance to climate disturbance⁹²), recovery (initial change followed by return to original abundance or community structure⁹³), and transition (i.e. to an alternative state^{15,50} (Fig 4a). There is growing consensus that communities which have experienced greater historical environmental fluctuations will be more likely to tolerate future disturbance^{50,94}. For example, forest biomes in which temperature fluctuations are relatively large, such as in high-latitude forests, exhibit relatively small compositional changes when exposed to warming compared with lower latitude forests^{95,96}. Similar patterns of tolerance have also been observed for short-lived disturbances like drought. In temperate forests, regions with lower historical precipitation select for microbial communities with higher tolerance to acute drought⁹², including Actinobacteria, Firmicutes, and Thaumarchaeota^{22,97}. Historical legacies of water-availability also correspond to drought tolerance for microbes in tropical forests; in forests where drought treatments had been experimentally imposed, tropical microbial communities experienced less pronounced shifts in community composition compared to the controls with no prior drought exposure⁹⁸.

Community recovery potential also appears to vary across biomes with different disturbance regimes. In forests where fire is historically common, microbial taxa and even whole microbial communities recover rapidly, even from severe disturbance^{99–102}. Microbial community recovery is facilitated by an initial increase in fire-specialized taxa⁵¹. For example, spores of a root-associated EMF species (*Rhizopogon olivaceotinctus*), nearly double their colonization potential following heating¹⁰³, and soil-dwelling *Arthrobacter* bacteria experience post-fire population booms⁵¹. These pyrophilous microbes may facilitate recovery of the original community by breaking down hydrophobic or pyrolyzed organic matter⁹⁹. Importantly, early successional microbes are generally poor competitors that do not appear to prevent re-establishment of the initial community¹⁰⁴. In contrast, for forest ecosystems adapted to less frequent fire regimes and having fewer pyrophilous community members, populations of both saprotrophic and mutualistic microbes are heavily reduced after fire and appear less likely to recover their original state^{52,100} (Fig. 4a).

The impacts of increasing frequency of disturbance and compounding disturbance types might be exceptionally stressful for microbial communities, potentially impeding community recovery and leading to a transition to alternative states¹⁰⁵. More frequent disturbances are

especially likely to result in functional collapse and transition in microbial ecosystems⁶⁴. For example, while microbial recovery potential appears high in fire adapted landscapes, fire regimes are shifting rapidly⁶. In fire-adapted *Eucalyptus pilularis* forests of Australia and coniferous forests in Western USA, fire adapted microbial and lichen communities appear less likely to recover with increases in fire frequency¹⁰⁶, especially when combined with periods of intensifying drought stress¹⁰¹ and emerging infectious disease¹⁰⁷. Microbial community transitions are also predicted where disturbance results in novel environmental conditions. For example, rising temperatures decrease climatic controls on rates of decomposition, leading to predicated transitions from EMF to AMF-dominated forests, especially at boreal-temperate ecotones¹⁹.

Studies documenting microbial community tolerance, recovery, or transition may not necessarily correspond with identical shifts in microbiome functioning. This decoupling could arise due to processes of microbial acclimation to warming via physiological shifts in cellular processes¹⁰⁸. Similarly, high degrees of functional redundancy in microbial communities could result in modest or non-linear functional responses despite strong community shifts. Relative decoupling of composition-function relationships could also vary across forest biomes if historical environmental fluctuations exert a strong impact on community sensitivity and a more variable impact on functional sensitivity. Accordingly, in boreal forest soil communities, prolonged warming may trigger moderate community shifts (tolerance) and a temporarily increased but progressively weakening effect on soil respiration due to microbial acclimation¹⁰⁹. This response contrasts with observations of accelerating community and respiration responses in experimentally warmed tropical forests⁹⁵. Sufficient temporal inference is lacking to fully describe these relationships, and further work is required. We stress that community tolerance, recovery, and transitions represent operational definitions; the linkages between community composition and microbiome function remains one of the most critical areas of ongoing investigation¹¹⁰.

Variation in sensitivity across microbe-specific niches

Microbes are adapted to distinct environmental fluctuations and resource availabilities across microbial-specific niches (MSNs) and therefore exhibit unique sensitivities among these habitats^{34,111,112}. For example, wood decay fungi such as *Phlebiopsis* and *Phellinus* exhibit optimal growth temperatures that vary across biogeographic ranges but generally fall between 20-28°C, with growth declining rapidly above 30°C¹¹³. By contrast, the surface of sunlit leaves can

frequently exceed 30°C¹¹⁴ and leaf-inhabiting microbial endophytes are adapted to warmer temperatures; many of these endophytes, including *Burkholderia*, *Acinetobacter*, and yeasts such as *Rhodotorula*, are regularly cultured at 30°C¹¹⁵. For highly diverse MSNs such as soils, there is evidence that microbes found in these MSNs may be more tolerant as compared to leaf or root communities¹¹⁶. Accordingly, the potential for microbial communities to respond asymmetrically across MSNs, especially above and belowground, is an important area of future investigation³. Studies of potential dysbiosis among MSN and functional consequences for hosts are critical in elucidating forest sensitivity to climate change¹¹⁷.

In addition to inhabiting unique physical niches in trees and forests (e.g. in roots versus leaves), microbial host-dependence is another important dimension of MSNs. While some microbes are free-living, others are facultatively or obligately host-associated and exhibit varying degrees of host specificity³⁴. For obligately host-associated microbes or those with high host-specificity, the persistence of host plants is a minimum criterion for the presence of suitable MSNs^{118,119}. For some obligate host associates, an effective tolerance strategy may be to improve host survival of the same disturbance¹²⁰. By contrast, where microbial associations increase mortality risks of their hosts, a decline in available hosts to infect may render these microbes more sensitive to climate change. For example, white pine blister rust (*Cronatium ribicola*) increases host mortality at low elevations during drought, thereby decreasing the population of its available hosts (alongside rates of infection)⁷.

Traits and tradeoffs that structure microbial sensitivity

Particular microbial traits predispose microbes to heightened climatic sensitivity because of how trait-tradeoffs interact with climatic perturbation^{22,51,121,122}. Trait-tradeoffs have been observed for bacterial⁴⁸ and fungal taxa, such as EMF¹²³, as well as in response to diverse disturbances, including water deficit²², warming¹²⁴ and fire^{51,102}. For example, trade-offs in bacterial suitability to post-fire habitability and competitive abilities have been observed¹⁰². We expect that investigations exploring the identity and function of specific traits influencing microbial sensitivity at both the taxon and community scale will be an especially active area of future research.

Climatic disturbances could significantly impact microbial physiology, as well as community composition and function. Trait tradeoffs between microbial growth yield (Y), resource acquisition (A), and stress tolerance (S) broadly define microbial sensitivity to climatic

perturbation (the Y-A-S framework; Fig. 4b)⁴⁸. When climatic disturbance drives resource limitation, such as during drought, microbes exhibit tradeoffs between traits associated with microbial resource acquisition and growth yields^{16,48}. This has been observed for soil bacteria across a natural gradient of water deficit, where filamentous *Streptomyces* bacteria associated with *Sequoia sempervirens* root communities dominate in dry soils but unicellular flagellate bacteria, such as *Bradyrhizobium*, are more dominant in wetter soils (Fig. 4b)²². Filamentous lifestyles excel at resource acquisition by scavenging a larger soil volume, whereas flagellate bacteria have higher growth yields, but are dependent on saturated soils for motility and access to soil resources⁵⁵.

Traits associated with stress tolerance are also important to microbial persistence during climatic perturbation^{48,55,124}. For example, exposure to acute drought resulted in an upregulation of genes associated with cell wall and membrane maintenance, but an overall reduction in gene expression and resource acquisition in foraging hyphae for the EMF fungus, *Suillus pungens*¹²³, likely impacting the cellular tolerance of stress and resource-acquisition of this fungus. Similar cellular tradeoffs also appear to apply for rising temperatures. A study of long-term soil warming in the Alaskan boreal forest found that genes associated with cellular maintenance were upregulated compared to genes associated with microbial resource acquisition and growth¹²⁴. Cellular dormancy is another important facet of microbial stress tolerance and could be a common strategy for members of the forest microbiome^{51,125,126}. Due to larger intracellular investment costs, dormancy appears to have important tradeoffs with capacity for vegetative growth rates^{48,51,102}. While little is known about cross forest biome variation in the distribution of cellular dormancy, dormancy and sporulation could be particularly common in forests which experience large seasonal climatic fluctuations or frequent pulse disturbances^{15,94,125}. Soils experiencing frequent fire are enriched in endospore-forming Firmicute bacteria^{10,102} and mycorrhizal fungi which form resistant spore banks may possess an early competitive advantage post-fire¹²⁶.

338 **Feedback mechanisms that amplify or buffer vulnerability**

339 Feedback processes can either amplify or buffer microbiome sensitivity and exposure to future
340 climatic disturbance⁴⁷. Feedbacks within forest systems play an underrecognized role in
341 determining the vulnerability of the microbiome to future climate change. Here, we outline several
342 pathways where biogeochemical, climatological, and biotic feedbacks could modify forest
343 microbiome vulnerability to future climatic disturbance.

344 At the largest scales, shifts in forest microbiome structure and function will influence
345 critical biogeochemical cycles. For example, trees that associate with N-fixing microbial
346 symbionts are favored under increasingly dry forest conditions¹²⁷. However, the dominance of N-
347 fixation in arid soils has also been linked to the release of NO_x, a potent greenhouse gas¹²⁸. As a
348 result of NO_x release, warming and drying are likely to be amplified, increasing the exposure of
349 forests and their microbiomes to further disturbance. Similarly, warming temperatures are poised
350 to stimulate the decay of soil organic matter (SOM) by microbial communities, particularly in
351 high-latitude soils¹⁴. The substantial release of CO₂ from increasing rates of microbial activity will
352 result in a positive climate feedback loop, further exposing forest systems to higher temperatures
353 in the future. Increased rainfall in certain tropical regions can also substantially shift
354 biogeochemical cycles by stimulating microbial methane production, the release of which can
355 further accelerate rising temperatures and other climatic disturbances; this is especially
356 pronounced after deforestation has occurred¹³. Elevated CO₂ (eCO₂) has significantly increased
357 the strength of the terrestrial C sink. This sink is contingent on the sustained nutrient-foraging
358 attributes of mycorrhizal symbionts and their ability to stimulate plant growth^{3,9}. However, for
359 trees associated with EMF, enhanced plant investment in fungal mutualists under eCO₂ could
360 reduce soil C stocks, with uncertain consequences on ecosystem C balance. Moreover, increased
361 EMF foraging under eCO₂ could alter substrate quality for free-living bacteria and fungi³, thereby
362 influencing nutrient mobilization and potentially plant nutrition. Long-term observations of shifts
363 in leaf and root-litter stoichiometry potentially support altered nutrient availability resultant from
364 shifted microbial functioning¹²⁹. In addition to eCO₂ effects, long-term shifts in soil substrate
365 availability and quality could also occur due to drought stress or increased fire frequency¹³⁰, with
366 similar consequences for microbial functioning, plant productivity and draw down of atmospheric
367 CO₂.

Climatic disturbances that restructure plant communities or impact plant ecophysiology will also create feedbacks that impact future microbiome exposure and sensitivity. Warming, drying, and increased fire frequency and severity are hypothesized to increase the activity, abundance, and pathogenicity of certain bacteria and fungi^{101,131}. Increased pathogenicity partially results from climatic release where higher winter temperatures increase winter survivorship for pathogens¹³². Increased pathogen abundance can hasten forest compositional turnover for trees already stressed due to changing climate. As climatic niches for EMF and AMF hosts shift with climate change, AMF-associated hosts are predicted to become more dominant in temperate systems¹³³. Because EMF-associated trees are thought to have higher resistance to root pathogens due to physical protection conferred by hyphal mantles surrounding the host roots³⁹, increased root pathogen abundance may interact with other microbial guilds such as AMF or EMF, to generate plant compositional feedbacks³⁹. The potential for transitions between plant communities dominated by different mycorrhizal associates will have consequences for nutrient cycling¹⁹, soil water retention¹³⁴, and forest structure and diversity¹³⁵.

Finally, as microbial responses to climate change will influence plant community coexistence and competition dynamics^{8,41}, shifts in forest community structure may generate additional feedbacks for microbial communities. Similarly climatic processes which alter plant community diversity or function could have knock-on effects that mediate the vulnerability of the microbiome to future disturbance. For example, drought can destabilize plant community coexistence in forests through modification of plant-soil-feedbacks¹³⁶. Drought or warming that result in reduced plant richness or plant trait diversity, could also reduce microbial network complexity¹³⁷, microbial multi-functionality, and potentially render communities less resilient to future disturbance¹³⁸. As microbes play important roles in dynamics of plant coexistence and competition^{8,41}, investigating these relationships under different climatic conditions or with novel microbial consortia will be important in determining forest vulnerability to climate change.

Future Directions: Microbiome research in the Anthropocene

Synthesizing microbial responses to climatic change remains an ongoing challenge due to the context-dependency of these responses and lack of baseline measurements¹⁸. Moreover, there are relatively few studies which directly compare standardized microbial responses to climatic disturbance across biomes or MSNs. Large-scale manipulative experiments can provide

standardized manipulations across a range of biomes (e.g. Free Air CO₂ Enrichment), and such initiatives provide important insights into the role of the microbiome in plant growth dynamics³. Similarly, sampling networks, such as the National Ecological Observatory Network (NEON), enable tracking microbiome and tree responses across natural gradients, over time, or across disturbance events^{139,140}. Importantly, studies that standardize sequencing approaches¹⁴¹, genetic markers^{142,143}, and downstream bioinformatic processing can ensure effective comparisons among studies¹⁴⁴.

Another important future direction for forest microbiome research is the incorporation of microbial processes into ecosystem and Earth system models²⁶. One primary challenge involves modeling how rates of nutrient cycling, decomposition, pathogen spread, or tree health will be impacted by future microbiome responses to disturbance (Box 2). This is particularly complex as multiple types of climatic disturbance occur simultaneously, sometimes buffering and other times exacerbating the effects of these disturbances on one another¹⁴⁵. Although microbial processes are often implicitly incorporated into models focusing on projecting future forest dynamics⁶, a specific challenge revolves around understanding the contexts in which microbially-explicit models outperform traditional first-order process-based models¹⁴⁶. Incorporating projections of microbiome functioning could potentially increase the uncertainty of some model projections¹⁴⁷, at least in the near term. Efforts to constrain the relative effect size of microbially-mediated processes in models remains a critical area of research²⁶. Trait-based approaches, which delineate microbial communities based on gene or enzymatic traits, are a promising opportunity to integrate microbes into ecosystem models¹⁴⁸. Additionally, properly measuring trait responses to climatic disturbances at model-relevant spatial and temporal scales will be essential to incorporating microbes into ecosystem models¹⁴⁷.

Last, there is bias towards North American and European authors cited in our literature, as well as in other recent reviews on the topic¹⁸; this stems in large part from our own language limitations as well as many structural and systemic biases in research and publication. Research investigating forest microbiome response to climate change and functional consequences for forest ecosystems is especially poorly represented for Latin America, Africa, and Southeast Asia. Decolonial approaches to research will be critical in better understanding and predicting a truly global response of forest microbiomes to climate change.

Conclusions

Forest microbiomes play a key role in the structure and function of global forests, but they are threatened by rapid and ongoing climate change. In this review, we adapt a risk assessment framework to identify the vulnerabilities of forest microbiomes to climate change, as well as contextualize potential impacts of microbiomes on forest function. Integrating the forest microbiome into conceptual and empirical models of forest functioning will advance understanding of forest health in the 21st century. There remains significant uncertainty in the cascade of microbiome-mediated feedbacks that will influence forest ecosystem function under climate change, and this represents an important area of future study. Overall, the forest microbiome remains an underappreciated target of conservation initiatives in Western science. We emphasize the importance of Indigenous science and sovereignty in managing Native ecosystems⁷³, as well as the need for more holistic studies linking macroecology with microbial ecology in determining responses of forests to climate change.

Box 1 | chí xáyviish nushtúkkareesh (Let's go pick mushrooms!)

Due to limitations in the extent of microbial natural history records in Western science, the study of microbial response to climate change presents many challenges. Consequently, studies of fungal fruiting phenology, for instance, have largely been restricted to temperate forests in Europe over the past several decades using herbaria collections^{70,71}. By contrast, Indigenous communities, including the Karuk Tribe in Northern California, USA have been documenting patterns of fungal fruiting phenology for thousands of years. For example, *Tricholoma magnivelare*, known as matsutake or xáyviish in the Karuk language, is an important EMF species used not only as a food and medicine, but as an indicator species of forest health. Until recently, xáyviish has been accessible and abundant for the Karuk people. However, Karuk Cultural Practitioners describe how while substantial early rains in September stimulate xáyviish development, when seasonal rainfall is delayed as is happening more frequently with climate change (first substantial rain occurring in November), mushroom development ceases for the year.

While the impacts of a long-term decline in xáyviish fruiting are not fully known, declines in fungal fruiting may have consequences not just for the fungal taxa themselves, but also for the establishment of seedlings and persistence of forests and the many organisms which they support⁴⁰.

For instance, deer and elk rely on xáyviish in the fall, during their own reproductive periods⁷³. For Karuk people, “Karuk traditional knowledge cannot be separated from either the practices that generated the knowledge, or the practices that emerge from it. These practices, known as ‘traditional management’ are, in turn, central to Karuk culture, identity, spirituality and mental and physical health”⁷³. The centering of Indigenous voices and acknowledgement of Indigenous sovereignty will be critical towards a more in-depth understanding of forest (including forest microbiome) response to climate change.

Box 2 | Predicting the assembly of non-analog microbial communities

New methods in biodiversity modeling provide opportunities to forecast the composition and function of non-analog forest microbiome communities under future climates. Non-analog communities are comprised of new combinations of taxa with no contemporaneous parallels. Due to expectations of heterogeneous responses to climatic change (outlined above), non-analog communities are expected to proliferate. To predict the composition and function of these communities, species distribution models (SDMs), sometimes called ecological niche models, use contemporaneous geographic occurrence data along with local environmental variables to generate species range maps. SDMs can then be used with future climatic data to forecast shifts in single microbial taxa or functional groups¹³³. However, concurrently modeling hundreds to thousands of species present in the forest microbiome is challenging using stacked SDM approaches. Joint Species Distribution Models (JSDMs), a relatively new class of statistical models, are well suited to the multivariate and sparse data structures generated by the molecular meta-barcoding studies used to study the forest microbiome¹⁴⁹. These modeling approaches offer insight into the structure and potential function of non-analog communities by accounting for species tradeoffs and spatially explicit species-environment linkages¹⁵⁰. JSDM provides opportunities for non-analog and shifting communities to be incorporated into temporal biogeochemical models seeking to predict forest soil respiration or CO₂ fertilization responses. One of the first studies to apply JSDM to the forest microbiome demonstrated that fungal saprotrophs with strong co-occurrence patterns were especially sensitive to forest management practices, indicating that biotic interactions can impact microbial community responses to disturbance and climate change more broadly¹⁵⁰.

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Figure captions

Figure 1 | The forest microbiome in a changing climate. A | The exceptionally diverse forest microbiome colonizes nearly every surface and plant tissue of the forest. Distinct microbial specific niches (MSN) occur throughout the forests, depicted with dashed yellow circles. Major microbial groups and associated MSN include: 1). Foliar endophytes and epiphytes (bacteria and fungi) within the 2). Forest Canopy. 3). Nitrogen fixing free-living bacteria occur in soil. 4). Forest gaps are unique microhabitats within forests. 5). Arbuscular mycorrhizal fungi (AMF); Ectomycorrhizal fungi (EMF), and the associated myco-rhizosphere dominate in soils; **B|** Heat stress and drying associated with climate change will shift the community structure and function of the forest microbiome. A reduction in available water is depicted by the reduction in the blue speckles throughout both panels 6). Wood and litter-decomposers in soil (bacteria, archaea, and fungi). Free-living soil microbes (bacteria, archaea, and fungi occur in soil and in decaying wood). 7). N-fixing bacteria may be free-living, root or leaf associated; 8). Forest canopy gaps can be particularly deleterious for microbial communities under climate change 9). Filamentous soil bacteria. 10). N-fixing bacterium associated with plant-roots. **C|** Distinct mycorrhizal symbioses are associated with major forest biomes where they influence biogeochemical cycles and forest

responses to climate change. Boreal forest ecosystems are dominated by ectomycorrhizal fungi (EMF), some EMF may enzymatically degrade soil organic matter to obtain organic forms of nitrogen, which influences host nutrition. Contrastingly, in tropical forest ecosystems, the majority of plants are associated with arbuscular mycorrhizal fungi (AMF), which efficiently scavenge nitrate (NO_3^-) and ammonium (PO_4^{3-}) from soil solution. The relative abundance of AMF and EMF hosts is more variable in temperate forest ecosystems.

Figure 2 | An organizational framework for studying forest microbiomes in a changing climate. Together, the sensitivity and exposure risk of microbial communities determine their overall vulnerability to climatic disturbance. Subsequent feedbacks may exacerbate or buffer subsequent microbial responses to climate change. We define climatic exposure as the cumulative severity of disturbance. Microbial community exposure is broadly shaped by the disturbance type, spatiotemporal scale, and attributes of the microbial microhabitat. Microbial sensitivity describes the degree to which microbial fitness or community structure and function is impacted by climatic disturbance. Broadly, microbial traits, niche attributes, and the climatic history of a forest can impact the sensitivity of a taxon or community. Finally, climatic feedbacks represent an array of processes, spanning macro-scale biogeochemical processes, to plant soil feedbacks that modulate future microbial exposure and sensitivity (dashed arrows).

Figure 3 | Scales of forest microbiome exposure to climate change. A | Examples of three forest biomes (boreal, temperate broadleaf, and tropical) and the relative distributions of different forest types across the globe. Inset photos are representative examples of major forest biomes. **B |** Exposure risk varies across these three broad forest biomes. Here we compare the mean temperature in the warmest quarter from 1970-2000 to the values predicted for 2061-2080 based on the Coupled Model Intercomparison Projects 5 (CMIP5) from the Intergovernmental Panel on Climate Change (IPCC) Fifth Assessment Report (AR5) in 2014. Three representative forest sites are reported. The predicted warming is based on the intermediate scenario where the concentrations of greenhouse gases (based on the Representative Concentration Pathways; RCP) assumes that emissions will peak in 2040 (RCP 4.5). Below, the line plot denotes the mean monthly temperature fluctuations throughout the year for these same plots from 1970-2000, indicating that microbes from these habitats are adapted to different temperature fluctuations and are liable to

demonstrate varying sensitivity to climate change as a result. Data are taken from WorldClim 2 (Fick, S.E. and R.J. Hijmans, 2017). **C** | The deviation of monthly mean minimum and maximum temperatures (seasonality) across these three representative forest sites compared to the annual mean annual temperature. Data are taken from WorldClim 2 (Fick, S.E. and R.J. Hijmans, 2017). **D and E** | Temperature fluxes in microbial microhabitats (MSN) are poorly captured in measurements of air temperature. **D** | Temperatures across the different forest microclimates for a representative forest in a Costa Rican Lowland Tropical Forest. **E** | Soil temperatures across different depths for a hypothetical soil, indicating unique microclimates which influence microbiome exposure. Data for **D** | temperatures in different forest microhabitats, taken from Fetcher et al. (1985). Data for **E** | mean temperature of soil at different depths taken from Blume et al. (2016).

Figure 4 | Microbial sensitivity and underlying traits and tradeoffs. **A** | Microbial communities exhibit temporally variable responses to climatic disturbance (dashed white line in the grey bar), minimal shift in community structure (tolerance), recovery to pre-disturbance conditions after some recovery period (recovery), or a permanent shift in community structure (transition). Notably, these dynamics could vary across forest biomes (different colored lines). Underlying each response dynamic is a specific example shown below for different disturbance types and microbial communities. **B** | Trait-tradeoffs mediate microbial sensitivity to hypothetical drought stress for rhizosphere bacterial taxa (*Bradyrhizobium*, *Bacillus*, *Streptomyces*). Under ambient conditions, resource abundance may select for taxa with high growth yields (**Y**), and reduced investment in stress tolerance. In contrast, in periods of resource limitation, taxa that can rapidly acquire resources, using extracellular enzymes (**A**) may dominate. Under drought conditions (or other climatic disturbances), stress tolerance (**S**), or stress-tolerant-resource-acquisitive strategies may dominate (Adopted from Malik et al., 2020). We also display an example of how bacterial life-history trait trade-offs can be geographically structured. Trade-offs related to high-growth yield (orange) and resource acquisition (green) are expected to vary across a gradient of water availability; traits associated with resource acquisition may be favored in drier conditions. Hypothetical microbial communities are plotted across the coast redwood range of California, USA where the northern part of the redwood range receives twice as much annual rainfall as the southern extent (Johnson and Dawson, 2011). Consequently, microbes that tolerate water deficit

are more abundant in the southern extent of the redwood range whereas microbes with high resource acquisition are more abundant at the wetter part of the range (Willing et al., 2020).

Photo associated with Box 1 | ***Tricholoma magnivelare*, also known as matsutake, tanoak mushroom, or xáyviish in Karuk language.** Photo courtesy of Monika Fischer, University of California, Berkeley.

Citations

1. Strassburg, B. B. N. *et al.* Global priority areas for ecosystem restoration. *Nature* **586**, 724–729 (2020).
2. Miner, K. *et al.* The co-production of knowledge for climate science. *Nature Climate Change* **13**, 307–308 (2023).
3. Terrer, C. *et al.* A trade-off between plant and soil carbon storage under elevated CO₂. *Nature* **591**, 599–603 (2021).
4. Haraway, D. Anthropocene, Capitalocene, Plantationocene, Chthulucene: Making Kin. *Environmental Humanities* **6**, 159–165 (2015).
5. Brodribb, T. J., Powers, J., Cochard, H. & Choat, B. Hanging by a thread? Forests and drought. *Science* **368**, 261–266 (2020).
6. Anderegg, W. R. L. *et al.* A climate risk analysis of Earth’s forests in the 21st century. *Science* **377**, 1099–1103 (2022).
7. Dudley, J. *et al.* Nonlinear shifts in infectious rust disease due to climate change. *Nat Commun* **12**, 5102 (2021).
8. Allsup, C. M., George, I. & Lankau, R. A. Shifting microbial communities can enhance tree tolerance to changing climates. *Science* **380**, 835–840 (2023).
9. Pellitier, P. T., Ibáñez, I., Zak, D. R., Argiroff, W. A. & Acharya, K. Ectomycorrhizal access to organic nitrogen mediates CO₂ fertilization response in a dominant temperate tree. *Nat Commun* **12**, 5403 (2021).
10. Pérez-Valera, E., Verdú, M., Navarro-Cano, J. A. & Goberna, M. Soil microbiome drives the recovery of ecosystem functions after fire. *Soil Biology and Biochemistry* **149**, 107948 (2020).

11. Filialuna, O. & Cripps, C. Evidence that pyrophilous fungi aggregate soil after forest fire. *Forest Ecology and Management* **498**, 119579 (2021).
12. Liu, X. *et al.* Responses of Soil Labile Organic Carbon to a Simulated Hurricane Disturbance in a Tropical Wet Forest. *Forests* **9**, 420 (2018).
13. Venturini, A. M. *et al.* Increased soil moisture intensifies the impacts of forest-to-pasture conversion on methane emissions and methane-cycling communities in the Eastern Amazon. *Environmental Research* **212**, 113139 (2022).
14. García-Palacios, P. *et al.* Evidence for large microbial-mediated losses of soil carbon under anthropogenic warming. *Nature Reviews Earth & Environment* **2**, 507–517 (2021).
15. Shade, A. *et al.* Fundamentals of Microbial Community Resistance and Resilience. *Front. Microbiol.* **3**, 417 (2012).
16. Kearns, P. J. & Shade, A. Trait-based patterns of microbial dynamics in dormancy potential and heterotrophic strategy: case studies of resource-based and post-fire succession. *ISME J* **12**, 2575–2581 (2018).
17. Jansson, J. K. & Hofmockel, K. S. Soil microbiomes and climate change. *Nat Rev Microbiol* **18**, 35–46 (2020).
18. Baldrian, P., López-Mondéjar, R. & Kohout, P. Forest microbiome and global change. *Nat Rev Microbiol* 1–15 (2023) doi:10.1038/s41579-023-00876-4.
19. Steidinger, B. S. *et al.* Climatic controls of decomposition drive the global biogeography of forest-tree symbioses. *Nature* **569**, 404–408 (2019).
20. Bui, A. *et al.* Soil fungal community composition and functional similarity shift across distinct climatic conditions. *FEMS Microbiology Ecology* fiae193 (2020) doi:10.1093/femsec/fiae193.
21. Bahram, M. *et al.* Structure and function of the global topsoil microbiome. *Nature* **560**, 233–237 (2018).
22. Willing, C. E., Pierroz, G., Coleman-Derr, D. & Dawson, T. E. The generalizability of water-deficit on bacterial community composition; Site-specific water-availability predicts the bacterial community associated with coast redwood roots. *Molecular Ecology* **29**, 4721–4734 (2020).
23. Cavicchioli, R. *et al.* Scientists’ warning to humanity: microorganisms and climate change. *Nat Rev Microbiol* **17**, 569–586 (2019).
24. Rudgers, J. A. *et al.* Climate Disruption of Plant-Microbe Interactions. *Annu. Rev. Ecol. Evol. Syst.* **51**, annurev-ecolsys-011720-090819 (2020).

25. Lladó, S., López-Mondéjar, R. & Baldrian, P. Forest Soil Bacteria: Diversity, Involvement in Ecosystem Processes, and Response to Global Change. *Microbiology and Molecular Biology Reviews* **81**, e00063-16 (2017).
26. Wan, J. & Crowther, T. W. Uniting the scales of microbial biogeochemistry with trait-based modelling. *Functional Ecology* **36**, 1457–1472 (2022).
27. Mishra, A., Singh, L. & Singh, D. Unboxing the black box—one step forward to understand the soil microbiome: A systematic review. *Microbial Ecology* **85**, 669–683 (2023).
28. Mitchard, E. T. A. The tropical forest carbon cycle and climate change. *Nature* **559**, 527–534 (2018).
29. *Kyoto Protocol to the United Nations Framework Convention on Climate Change*. 2303 U.N.T.S. 162. (1997).
30. De Frenne, P. *et al.* Forest microclimates and climate change: Importance, drivers and future research agenda. *Global Change Biology* **27**, 2279–2297 (2021).
31. Poorter, L., Bongers, L. & Bongers, F. Architecture of 54 Moist-Forest Tree Species: Traits, Trade-Offs, and Functional Groups. *Ecology* **87**, 1289–1301 (2006).
32. Piovesan, G. & Biondi, F. On tree longevity. *New Phytologist* **231**, 1318–1337 (2021).
33. Averill, C., Turner, B. L. & Finzi, A. C. Mycorrhiza-mediated competition between plants and decomposers drives soil carbon storage. *Nature* **505**, 543–545 (2014).
34. Baldrian, P. Forest microbiome: diversity, complexity and dynamics. *FEMS Microbiol Rev* **41**, 109–130 (2017).
35. Epihov, D. Z. *et al.* Legume–microbiome interactions unlock mineral nutrients in regrowing tropical forests. *Proceedings of the National Academy of Sciences* **118**, (2021).
36. Rodriguez, R. J., Jr, J. F. W., Arnold, A. E. & Redman, R. S. Fungal endophytes: diversity and functional roles. *New Phytologist* **182**, 314–330 (2009).
37. Fierer, N. Embracing the unknown: disentangling the complexities of the soil microbiome. *Nat Rev Microbiol* **15**, 579–590 (2017).
38. Pérez-Izquierdo, L. *et al.* Plant intraspecific variation modulates nutrient cycling through its below ground rhizospheric microbiome. *Journal of Ecology* **107**, 1594–1605 (2019).
39. Bennett, J. A. *et al.* Plant-soil feedbacks and mycorrhizal type influence temperate forest population dynamics. *Science* **355**, 181–184 (2017).

40. Peay, K. G. Timing of mutualist arrival has a greater effect on *Pinus muricata* seedling growth than interspecific competition. *Journal of Ecology* **106**, 514–523 (2018).
41. Van Nuland, M. E., Ke, P.-J., Wan, J. & Peay, K. G. Mycorrhizal nutrient acquisition strategies shape tree competition and coexistence dynamics. *Journal of Ecology* **111**, 564–577 (2023).
42. Lehmann, J. *et al.* Persistence of soil organic carbon caused by functional complexity. *Nature Geoscience* **13**, 529–534 (2020).
43. Fernandez, C. W., Heckman, K., Kolka, R. & Kennedy, P. G. Melanin mitigates the accelerated decay of mycorrhizal necromass with peatland warming. *Ecology Letters* **22**, 498–505 (2019).
44. Arnold, A. E. & Engelbrecht, B. M. J. Fungal Endophytes Nearly Double Minimum Leaf Conductance in Seedlings of a Neotropical Tree Species. *Journal of Tropical Ecology* **23**, 369–372 (2007).
45. Moyes, A. B. *et al.* Evidence for foliar endophytic nitrogen fixation in a widely distributed subalpine conifer. *New Phytologist* **210**, 657–668 (2016).
46. Zheng, M. *et al.* Effects of human disturbance activities and environmental change factors on terrestrial nitrogen fixation. *Global Change Biology* **26**, 6203–6217 (2020).
47. Williams, S. E., Shoo, L. P., Isaac, J. L., Hoffmann, A. A. & Langham, G. Towards an Integrated Framework for Assessing the Vulnerability of Species to Climate Change. *PLoS Biol* **6**, e325 (2008).
48. Malik, A. A. *et al.* Defining trait-based microbial strategies with consequences for soil carbon cycling under climate change. *ISME J* **14**, 1–9 (2020).
49. Bender, E. A., Case, T. J. & Gilpin, M. E. Perturbation Experiments in Community Ecology: Theory and Practice. *Ecology* **65**, 1–13 (1984).
50. Allison, S. D. & Martiny, J. B. H. Resistance, resilience, and redundancy in microbial communities. *Proceedings of the National Academy of Sciences* **105**, 11512–11519 (2008).
51. Whitman, T. *et al.* Soil bacterial and fungal response to wildfires in the Canadian boreal forest across a burn severity gradient. *Soil Biology and Biochemistry* **138**, 107571 (2019).
52. Pulido-Chavez, M. F., Alvarado, E. C., DeLuca, T. H., Edmonds, R. L. & Glassman, S. I. High-severity wildfire reduces richness and alters composition of ectomycorrhizal fungi in low-severity adapted ponderosa pine forests. *Forest Ecology and Management* **485**, 118923 (2021).

693 53. Scheffer, M., Carpenter, S., Foley, J. A., Folke, C. & Walker, B. Catastrophic shifts in ecosystems. *Nature* **413**,
694 591–596 (2001).

695 54. Smith, S. E. & Read, D. J. *Mycorrhizal Symbiosis*. (Academic Press, 2010).

696 55. Schimel, J. P. Life in Dry Soils: Effects of Drought on Soil Microbial Communities and Processes. *Annual*
697 *Review of Ecology, Evolution, and Systematics* **49**, 409–432 (2018).

698 56. Martiny, J. B. H. *et al.* Investigating the eco-evolutionary response of microbiomes to environmental change.
699 *Ecology Letters* (2023).

700 57. Zhou, J. *et al.* Temperature mediates continental-scale diversity of microbes in forest soils. *Nat Commun* **7**,
701 12083 (2016).

702 58. Iversen, C. M., Ledford, J. & Norby, R. J. CO₂ enrichment increases carbon and nitrogen input from fine roots
703 in a deciduous forest. *New Phytologist* **179**, 837–847 (2008).

704 59. Norby, R. J. & Zak, D. R. Ecological lessons from free-air CO₂ enrichment (FACE) experiments. *Annual*
705 *Review of Ecology, Evolution, and Systematics* **42**, (2011).

706 60. Heinemeyer, A., Ineson, P., Ostle, N. & Fitter, A. H. Respiration of the external mycelium in the arbuscular
707 mycorrhizal symbiosis shows strong dependence on recent photosynthates and acclimation to temperature.
708 *New Phytologist* **171**, 159–170 (2006).

709 61. Olsrud, M., Carlsson, B. Å., Svensson, B. M., Michelsen, A. & Melillo, J. M. Responses of fungal root
710 colonization, plant cover and leaf nutrients to long-term exposure to elevated atmospheric CO₂ and warming
711 in a subarctic birch forest understory. *Global Change Biology* **16**, 1820–1829 (2010).

712 62. Ikeda, K. *et al.* Snowfall and snowpack in the Western U.S. as captured by convection permitting climate
713 simulations: current climate and pseudo global warming future climate. *Clim Dyn* **57**, 2191–2215 (2021).

714 63. Millar, C. I. & Stephenson, N. L. Temperate forest health in an era of emerging megadisturbance. *Science* **349**,
715 823–826 (2015).

716 64. König, S. *et al.* Spatiotemporal disturbance characteristics determine functional stability and collapse risk of
717 simulated microbial ecosystems. *Sci Rep* **8**, 9488 (2018).

718 65. Barnes, I. *et al.* New *Ceratocystis* species associated with rapid death of *Metrosideros polymorpha* in Hawai'i.
719 *Persoonia - Molecular Phylogeny and Evolution of Fungi* **40**, 154–181 (2018).

66. Reich, P. B. *et al.* Even modest climate change may lead to major transitions in boreal forests. *Nature* **608**, 540–545 (2022).
67. Livne-Luzon, S. *et al.* High resilience of the mycorrhizal community to prescribed seasonal burnings in eastern Mediterranean woodlands. *Mycorrhiza* **31**, 203–216 (2021).
68. Wang, J. *et al.* Changing Lengths of the Four Seasons by Global Warming. *Geophysical Research Letters* **48**, e2020GL091753 (2021).
69. Gallinat, A. S., Primack, R. B. & Wagner, D. L. Autumn, the neglected season in climate change research. *Trends in Ecology & Evolution* **30**, 169–176 (2015).
70. Gange, A. C., Gange, E. G., Sparks, T. H. & Boddy, L. Rapid and Recent Changes in Fungal Fruiting Patterns. *Science* **316**, 71–71 (2007).
71. Kausrud, H. *et al.* Warming-induced shift in European mushroom fruiting phenology. *Proceedings of the National Academy of Sciences* **109**, 14488–14493 (2012).
72. Anderson, M. K. & Lake, F. K. California Indian Ethnomycology and Associated Forest Management. *Journal of Ethnobiology* **33**, 33–85 (2013).
73. Karuk Tribe. Karuk Climate Adaptation Plan. (2019).
74. Hernandez, J., Meisner, J., Jacobs, L. A. & Rabinowitz, P. M. Re-Centering Indigenous Knowledge in climate change discourse. *PLOS Climate* **1**, e0000032 (2022).
75. Gervers, K. A., Thomas, D. C., Roy, B. A., Spatafora, J. W. & Busby, P. E. Crown closure affects endophytic leaf mycobiome compositional dynamics over time in *Pseudotsuga menziesii* var. *menziesii*. *Fungal Ecology* **57–58**, 101155 (2022).
76. Gora, E. M., Lucas, J. M. & Yanoviak, S. P. Microbial Composition and Wood Decomposition Rates Vary with Microclimate From the Ground to the Canopy in a Tropical Forest. *Ecosystems* **22**, 1206–1219 (2019).
77. Drake, J. E. *et al.* Trees tolerate an extreme heatwave via sustained transpirational cooling and increased leaf thermal tolerance. *Global Change Biology* **24**, 2390–2402 (2018).
78. Bowman, E. A. & Arnold, A. E. Distributions of ectomycorrhizal and foliar endophytic fungal communities associated with *Pinus ponderosa* along a spatially constrained elevation gradient. *American Journal of Botany* **105**, 687–699 (2018).

747 79. Nelson, A. R. *et al.* Wildfire-dependent changes in soil microbiome diversity and function. *Nat Microbiol* **7**,
748 1419–1430 (2022).

749 80. Krah, F.-S. *et al.* Independent effects of host and environment on the diversity of wood-inhabiting fungi.
750 *Journal of Ecology* **106**, 1428–1442 (2018).

751 81. Van Nuland, M. E. *et al.* Warming and disturbance alter soil microbiome diversity and function in a northern
752 forest ecotone. *FEMS Microbiology Ecology* **96**, fiae108 (2020).

753 82. Giuggiola, A. *et al.* Competition for water in a xeric forest ecosystem – Effects of understory removal on soil
754 micro-climate, growth and physiology of dominant Scots pine trees. *Forest Ecology and Management* **409**,
755 241–249 (2018).

756 83. Long, J. W., Lake, F. K. & Goode, R. W. The importance of Indigenous cultural burning in forested regions of
757 the Pacific West, USA. *Forest Ecology and Management* **500**, 119597 (2021).

758 84. Vandenkoornhuyse, P., Quaiser, A., Duhamel, M., Le Van, A. & Dufresne, A. The importance of the
759 microbiome of the plant holobiont. *New Phytologist* **206**, 1196–1206 (2015).

760 85. Dawson, T. P., Jackson, S. T., House, J. I., Prentice, I. C. & Mace, G. M. Beyond Predictions: Biodiversity
761 Conservation in a Changing Climate. *Science* **332**, 53–58 (2011).

762 86. Talbot, J. M. *et al.* Endemism and functional convergence across the North American soil mycobiome.
763 *Proceedings of the National Academy of Sciences* **111**, 6341–6346 (2014).

764 87. Shu, W.-S. & Huang, L.-N. Microbial diversity in extreme environments. *Nat Rev Microbiol* **20**, 219–235
765 (2022).

766 88. Moeller, H. V., Dickie, I. A., Peltzer, D. A. & Fukami, T. Mycorrhizal co-invasion and novel interactions
767 depend on neighborhood context. *Ecology* **96**, 2336–2347 (2015).

768 89. U'Ren, J. M. *et al.* Host availability drives distributions of fungal endophytes in the imperilled boreal realm.
769 *Nature Ecology & Evolution* **3**, 1430–1437 (2019).

770 90. Peay, K. G., Schubert, M. G., Nguyen, N. H. & Bruns, T. D. Measuring ectomycorrhizal fungal dispersal:
771 macroecological patterns driven by microscopic propagules. *Molecular Ecology* **21**, 4122–4136 (2012).

772 91. Branco, S. *et al.* Genetic isolation between two recently diverged populations of a symbiotic fungus. *Molecular*
773 *Ecology* **24**, 2747–2758 (2015).

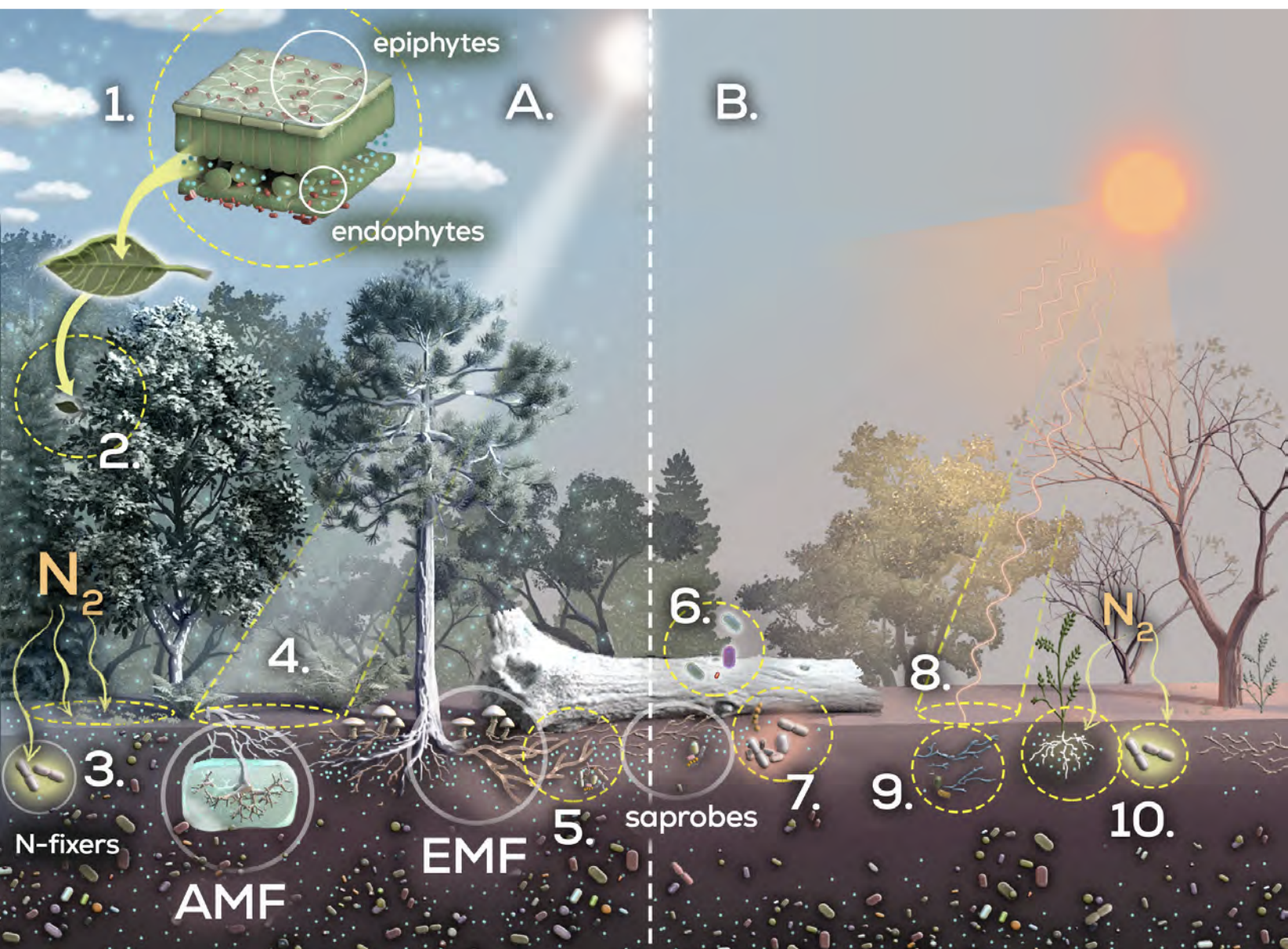
92. Evans, S. E., Allison, S. D. & Hawkes, C. V. Microbes, memory and moisture: Predicting microbial moisture responses and their impact on carbon cycling. *Functional Ecology* **36**, 1430–1441 (2022).
93. Dove, N. C., Taş, N. & Hart, S. C. Ecological and genomic responses of soil microbiomes to high-severity wildfire: linking community assembly to functional potential. *ISME J* **16**, 1853–1863 (2022).
94. Hawkes, C. V. & Keitt, T. H. Resilience vs. historical contingency in microbial responses to environmental change. *Ecology Letters* **18**, 612–625 (2015).
95. Nottingham, A. T., Meir, P., Velasquez, E. & Turner, B. L. Soil carbon loss by experimental warming in a tropical forest. *Nature* **584**, 234–237 (2020).
96. Wang, C. *et al.* The temperature sensitivity of soil: microbial biodiversity, growth, and carbon mineralization. *ISME J* **15**, 2738–2747 (2021).
97. Meisner, A., Jacquiod, S., Snoek, B. L., ten Hooven, F. C. & van der Putten, W. H. Drought Legacy Effects on the Composition of Soil Fungal and Prokaryote Communities. *Frontiers in Microbiology* **9**, (2018).
98. Bouskill, N. J. *et al.* Pre-exposure to drought increases the resistance of tropical forest soil bacterial communities to extended drought. *ISME J* **7**, 384–394 (2013).
99. Fischer, M. S. *et al.* Pyrolyzed Substrates Induce Aromatic Compound Metabolism in the Post-fire Fungus, *Pyronema domesticum*. *Frontiers in Microbiology* **12**, 3085 (2021).
100. Smith, G. R., Edy, L. C. & Peay, K. G. Contrasting fungal responses to wildfire across different ecosystem types. *Molecular Ecology* **30**, 844–854 (2021).
101. Bowd, E. J. *et al.* Direct and indirect effects of fire on microbial communities in a pyrodiverse dry-sclerophyll forest. *Journal of Ecology* **110**, 1687–1703 (2022).
102. Enright, D. J., Frangioso, K. M., Isobe, K., Rizzo, D. M. & Glassman, S. I. Mega-fire in redwood tanoak forest reduces bacterial and fungal richness and selects for pyrophilous taxa that are phylogenetically conserved. *Molecular Ecology* **n/a**, (2022).
103. Bruns, T. D., Hale, M. L. & Nguyen, N. H. *Rhizopogon olivaceotinctus* increases its inoculum potential in heated soil independent of competitive release from other ectomycorrhizal fungi. *Mycologia* **111**, 936–941 (2019).

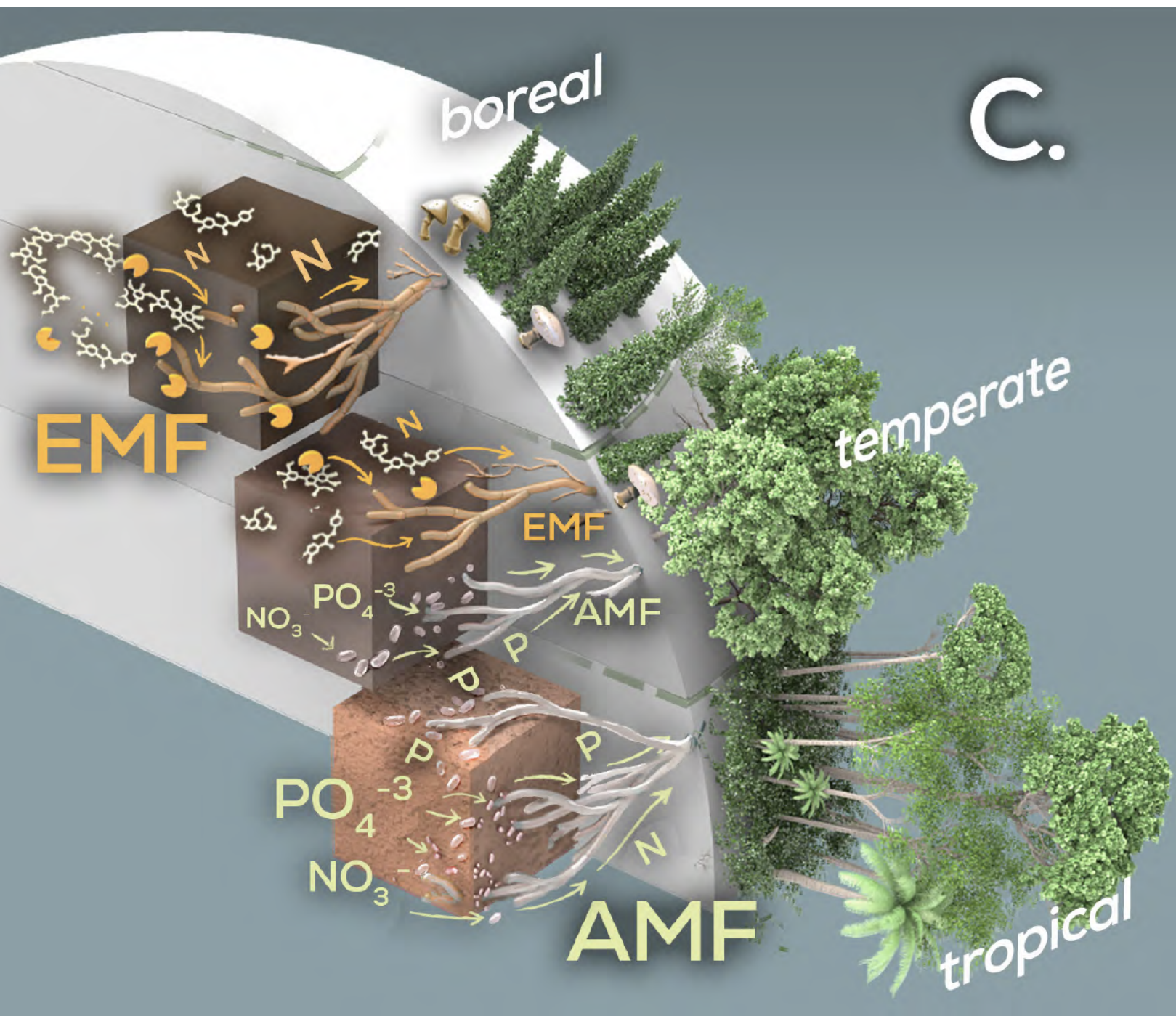
104. Kennedy, P. G., Higgins, L. M., Rogers, R. H. & Weber, M. G. Colonization-Competition Tradeoffs as a Mechanism Driving Successional Dynamics in Ectomycorrhizal Fungal Communities. *PLoS ONE* **6**, e25126 (2011).
105. Bardgett, R. D. & Caruso, T. Soil microbial community responses to climate extremes: resistance, resilience and transitions to alternative states. *Philosophical Transactions of the Royal Society B: Biological Sciences* **375**, 20190112 (2020).
106. Miller, J. E. D., Root, H. T. & Safford, H. D. Altered fire regimes cause long-term lichen diversity losses. *Global Change Biology* **24**, 4909–4918 (2018).
107. Metz, M. R., Varner, J. M., Frangioso, K. M., Meentemeyer, R. K. & Rizzo, D. M. Unexpected redwood mortality from synergies between wildfire and an emerging infectious disease. *Ecology* **94**, 2152–2159 (2013).
108. Robinson, J. M. *et al.* Rapid laboratory measurement of the temperature dependence of soil respiration and application to changes in three diverse soils through the year. *Biogeochemistry* **133**, 101–112 (2017).
109. Bradford, M. A. *et al.* Cross-biome patterns in soil microbial respiration predictable from evolutionary theory on thermal adaptation. *Nat Ecol Evol* **3**, 223–231 (2019).
110. Malik, A. A. & Bouskill, N. J. Drought impacts on microbial trait distribution and feedback to soil carbon cycling. *Functional Ecology* **36**, 1442–1456 (2022).
111. Chomicki, G., Werner, G. D. A., West, S. A. & Kiers, E. T. Compartmentalization drives the evolution of symbiotic cooperation. *Phil. Trans. R. Soc. B* **375**, 20190602 (2020).
112. Willing, C. E. *et al.* Keep your friends close: Host compartmentalisation of microbial communities facilitates decoupling from effects of habitat fragmentation. *Ecology Letters* **24**, 2674–2686 (2021).
113. Maynard, D. S. *et al.* Consistent trade-offs in fungal trait expression across broad spatial scales. *Nature Microbiology* **4**, 846–853 (2019).
114. Pounds, J. A. & Puschendorf, R. Clouded futures. *Nature* **427**, 107–109 (2004).
115. Khan, Z. *et al.* Growth enhancement and drought tolerance of hybrid poplar upon inoculation with endophyte consortia. *Current Plant Biology* **6**, 38–47 (2016).
116. Nimmo, D. G., Mac Nally, R., Cunningham, S. C., Haslem, A. & Bennett, A. F. Vive la résistance: reviving resistance for 21st century conservation. *Trends in Ecology & Evolution* **30**, 516–523 (2015).

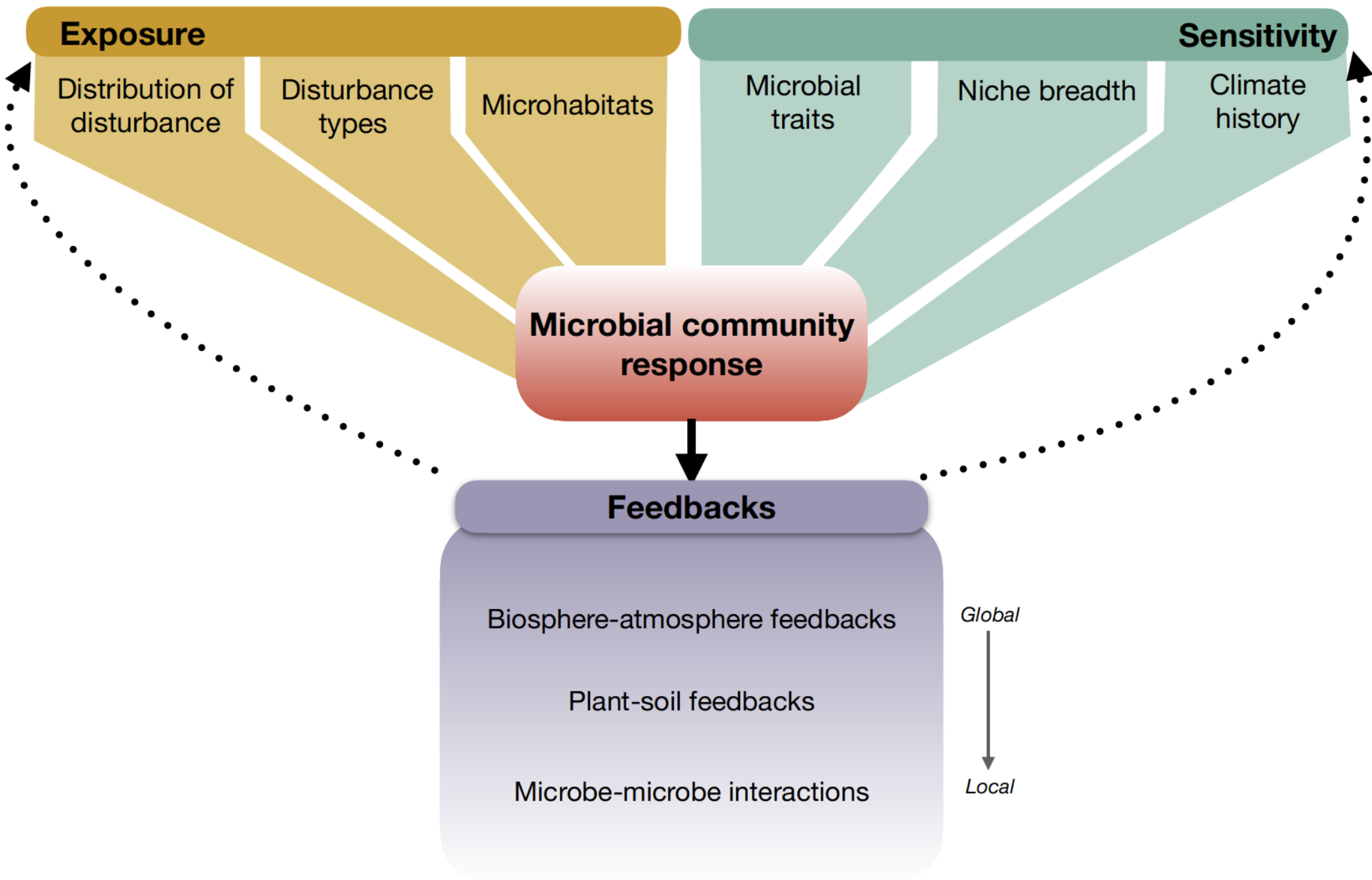
117. Cregger, M. A. *et al.* The *Populus* holobiont: dissecting the effects of plant niches and genotype on the microbiome. *Microbiome* **6**, 31 (2018).
118. Rigling, D. & Prospero, S. *Cryphonectria parasitica*, the causal agent of chestnut blight: invasion history, population biology and disease control. *Mol Plant Pathol* **19**, 7–20 (2017).
119. Agan, A. *et al.* The Relationship between Fungal Diversity and Invasibility of a Foliar Niche—The Case of Ash Dieback. *Journal of Fungi* **6**, 150 (2020).
120. Gehring, C. A., Sthultz, C. M., Flores-Rentería, L., Whipple, A. V. & Whitham, T. G. Tree genetics defines fungal partner communities that may confer drought tolerance. *Proc Natl Acad Sci USA* **114**, 11169–11174 (2017).
121. Lustenhouwer, N. *et al.* A trait-based understanding of wood decomposition by fungi. *PNAS* **117**, 11551–11558 (2020).
122. Alvarez-Manjarrez, J. & Garibay-Orijel, R. Resilience of soil fungal community to hurricane Patricia (category 4). *Forest Ecology and Management* **498**, 119550 (2021).
123. Erlandson, S. R. *et al.* Transcriptional acclimation and spatial differentiation characterize drought response by the ectomycorrhizal fungus *Suillus pungens*. *New Phytologist* **243**, 1910–1913 (2021).
124. Romero-Olivares, A. L., Meléndrez-Carballo, G., Lago-Lestón, A. & Treseder, K. K. Soil Metatranscriptomes Under Long-Term Experimental Warming and Drying: Fungi Allocate Resources to Cell Metabolic Maintenance Rather Than Decay. *Frontiers in Microbiology* **10**, (2019).
125. Lennon, J. T. & Jones, S. E. Microbial seed banks: the ecological and evolutionary implications of dormancy. *Nat Rev Microbiol* **9**, 119–130 (2011).
126. Glassman, S. I., Levine, C. R., DiRocco, A. M., Battles, J. J. & Bruns, T. D. Ectomycorrhizal fungal spore bank recovery after a severe forest fire: some like it hot. *ISME J* **10**, 1228–1239 (2016).
127. Gei, M. *et al.* Legume abundance along successional and rainfall gradients in Neotropical forests. *Nat Ecol Evol* **2**, 1104–1111 (2018).
128. Kou-Giesbrecht, S. & Menge, D. Nitrogen-fixing trees could exacerbate climate change under elevated nitrogen deposition. *Nat Commun* **10**, 1493 (2019).
129. Mason, R. E. *et al.* Evidence, causes, and consequences of declining nitrogen availability in terrestrial ecosystems. *Science* **376**, (2022).

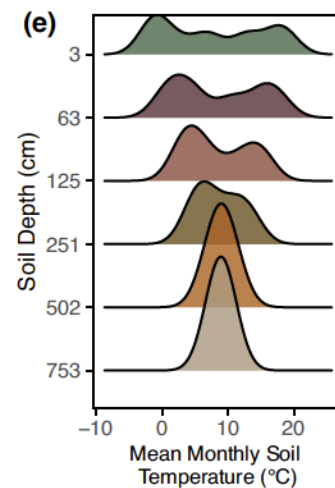
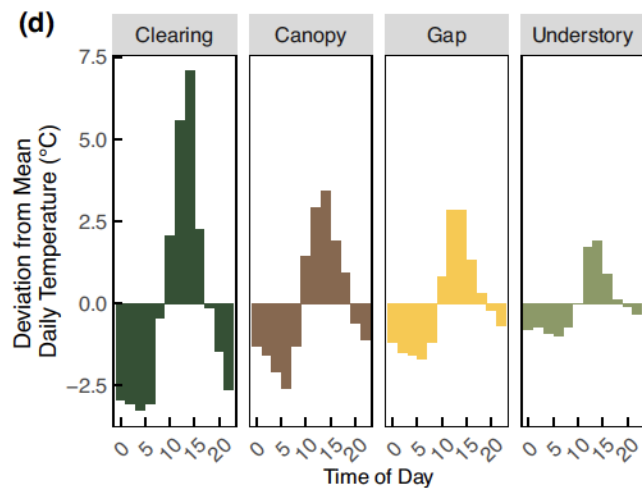
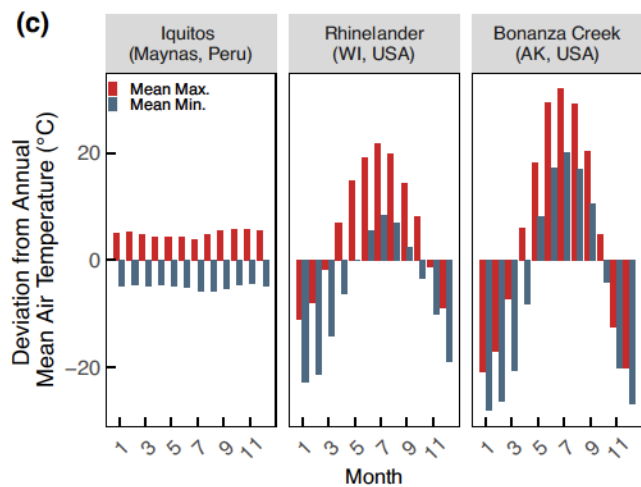
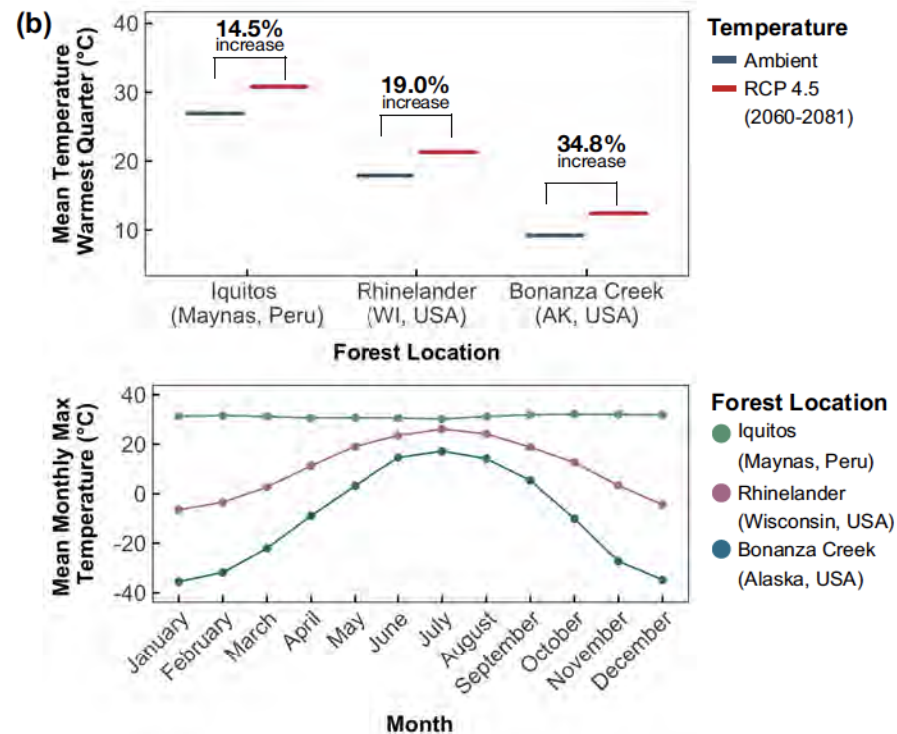
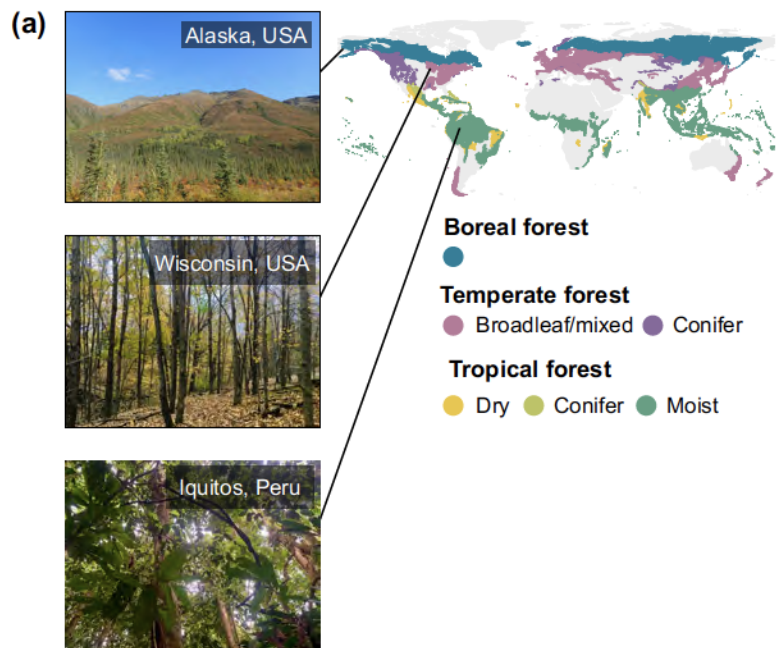
130. Pellegrini, A. F. A. *et al.* Fire effects on the persistence of soil organic matter and long-term carbon storage. *Nat. Geosci.* **15**, 5–13 (2022).
131. Delgado-Baquerizo, M. *et al.* The proportion of soil-borne pathogens increases with warming at the global scale. *Nat. Clim. Chang.* **10**, 550–554 (2020).
132. Pfender, W. F. & Vollmer, S. S. Freezing Temperature Effect on Survival of *Puccinia graminis* subsp. *graminicola* in *Festuca arundinacea* and *Lolium perenne*. *Plant Disease* **83**, 1058–1062 (1999).
133. Steidinger, B. S. *et al.* Ectomycorrhizal fungal diversity predicted to substantially decline due to climate changes in North American Pinaceae forests. *Journal of Biogeography* **47**, 772–782 (2020).
134. Yuan, Z. *et al.* Divergent above- and below-ground biodiversity pathways mediate disturbance impacts on temperate forest multifunctionality. *Global Change Biology* **27**, 2883–2894 (2021).
135. Corrales, A., Mangan, S. A., Turner, B. L. & Dalling, J. W. An ectomycorrhizal nitrogen economy facilitates monodominance in a neotropical forest. *Ecology Letters* **19**, 383–392 (2016).
136. Dudenhöffer, J.-H., Luecke, N. C. & Crawford, K. M. Changes in precipitation patterns can destabilize plant species coexistence via changes in plant–soil feedback. *Nat Ecol Evol* **6**, 546–554 (2022).
137. Konopka, A., Lindemann, S. & Fredrickson, J. Dynamics in microbial communities: unraveling mechanisms to identify principles. *ISME J* **9**, 1488–1495 (2015).
138. Gao, C. *et al.* Co-occurrence networks reveal more complexity than community composition in resistance and resilience of microbial communities. *Nat Commun* **13**, 3867 (2022).
139. Qin, C., Pellitier, P. T., Van Nuland, M. E., Peay, K. G. & Zhu, K. Niche modelling predicts that soil fungi occupy a precarious climate in boreal forests. *Global Ecology and Biogeography* **32**, 1127–1139 (2023).
140. Walkup, J. *et al.* The predictive power of phylogeny on growth rates in soil bacterial communities. *ISME COMMUN.* **3**, 1–8 (2023).
141. Salipante, S. J. *et al.* Performance Comparison of Illumina and Ion Torrent Next-Generation Sequencing Platforms for 16S rRNA-Based Bacterial Community Profiling. *Applied and Environmental Microbiology* **80**, 7583–7591 (2014).
142. Bruns, T. D. & Taylor, J. W. Comment on ‘Global assessment of arbuscular mycorrhizal fungus diversity reveals very low endemism’. *Science* **351**, 826–826 (2016).

143. Johnson, J. S. *et al.* Evaluation of 16S rRNA gene sequencing for species and strain-level microbiome analysis. *Nat Commun* **10**, 5029 (2019).
144. Tedersoo, L. *et al.* Best practices in metabarcoding of fungi: From experimental design to results. *Molecular Ecology* **31**, 2769–2795 (2022).
145. Philippot, L., Griffiths, B. S. & Langenheder, S. Microbial Community Resilience across Ecosystems and Multiple Disturbances. *Microbiology and Molecular Biology Reviews* **85**, 10.1128/mmbr.00026-20 (2021).
146. Nevison, C., Hess, P., Goodale, C., Zhu, Q. & Vira, J. Nitrification, denitrification, and competition for soil N: Evaluation of two Earth System Models against observations. *Ecological Applications* **32**, e2528 (2022).
147. Bradford, M. A. *et al.* Quantifying microbial control of soil organic matter dynamics at macrosystem scales. *Biogeochemistry* **156**, 19–40 (2021).
148. Baskaran, P. *et al.* Modelling the influence of ectomycorrhizal decomposition on plant nutrition and soil carbon sequestration in boreal forest ecosystems. *New Phytologist* **213**, 1452–1465 (2017).
149. Ovaskainen, O. & Abrego, N. *Joint Species Distribution Modelling: With Applications in R (Ecology, Biodiversity and Conservation)*. (Cambridge University Press, Cambridge, 2020).
150. Abrego, N., Dunson, D., Halme, P., Salcedo, I. & Ovaskainen, O. Wood-inhabiting fungi with tight associations with other species have declined as a response to forest management. *Oikos* **126**, (2017).



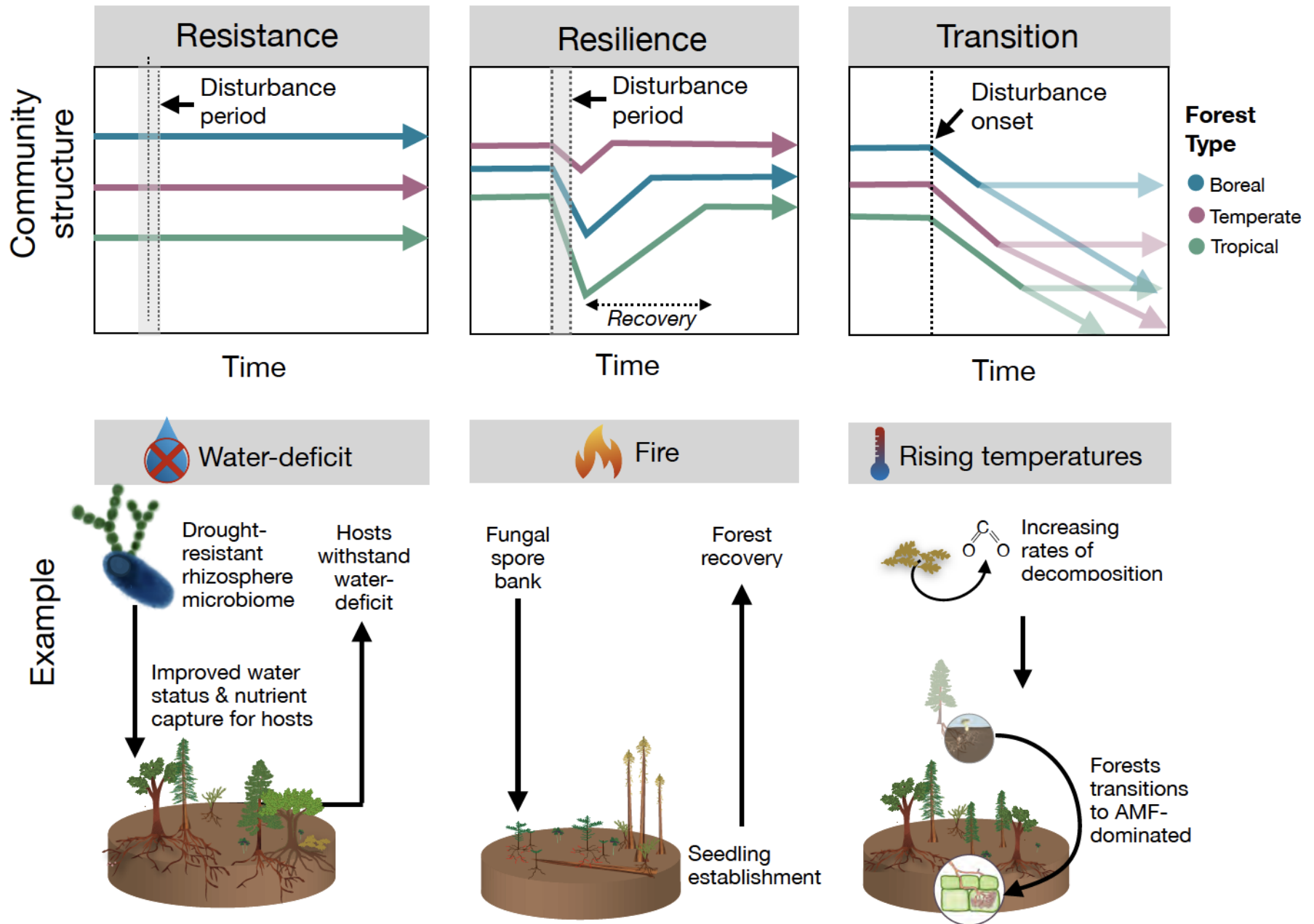






(a)

Community sensitivity



(b)

Organismal sensitivity

