

1 **Editor summary:**

2 In this Perspective, the authors develop a risk assessment framework for forest microbiomes
3 under climate change that unites microbial and forest ecology. They define processes that
4 amplify or buffer microbial sensitivity and exposure risk and feedbacks that mediate impacts on
5 microbial communities.

6 **Peer Review Information:**

7 Nature Climate Change thanks Eleonora Egidi, Kevin Newsham and the other, anonymous,
8 reviewer(s) for their contribution to the peer review of this work.

9

Title: A Risk Assessment Framework for the Future of Forest Microbiomes in a Changing Climate

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Abstract: Microbes inhabiting the above and belowground tissues of forest trees and soils play a critical role in the response of forest ecosystems to global climate change. However, generalizations about the vulnerability of the forest microbiome to climate change have been challenging due to often context-dependent responses. Here we apply a risk assessment framework to evaluate microbial community vulnerability to climate change across forest ecosystems. We define factors that determine exposure risk and processes that amplify or buffer sensitivity to change and describe feedback mechanisms that will modulate this exposure and sensitivity as climatic change progresses. This risk assessment approach unites microbial ecology and forest ecology to develop a more comprehensive understanding of forest vulnerability in the 21st century.

Resumen (Español): Los microorganismos que habitan en los tejidos aéreos y subterráneos de los árboles y suelos forestales desempeñan un papel fundamental en la respuesta de los ecosistemas forestales al cambio climático. Sin embargo, las generalizaciones sobre la respuesta del microbioma forestal al cambio climático ha sido un reto debido a que las respuestas son contexto-dependientes. Aquí aplicamos un marco de evaluación de riesgos para evaluar la vulnerabilidad de la comunidad microbiana al cambio climático en los ecosistemas forestales. Definimos los factores que determinan el riesgo a la exposición y los procesos que amplifican o amortiguan la sensibilidad al cambio climático, y describimos los mecanismos de retroalimentación que modularán la exposición y sensibilidad a medida que avance el cambio climático. Esta aproximación de evaluación de riesgos une a la ecología microbiana y la ecología forestal para desarrollar una comprensión más completa de la vulnerabilidad de los bosques en el siglo XXI.

Resumo (Português): Os microrganismos que habitam o solo e os tecidos das árvores acima e abaixo do solo desempenham um papel crítico na resposta dos ecossistemas florestais às mudanças climáticas globais. No entanto, generalizações sobre a resposta do microbioma florestal às alterações climáticas têm sido um desafio devido às frequentes respostas dependentes do contexto. Aqui, nós aplicamos uma estrutura de avaliação de risco para avaliar a vulnerabilidade da comunidade microbiana às mudanças climáticas em ecossistemas florestais. Nós definimos os fatores que determinam o risco de exposição e os processos que amplificam ou atenuam a sensibilidade à mudança, e descrevemos os mecanismos de retroalimentação que modularão essa exposição e sensibilidade à medida que as alterações climáticas progridem. Essa abordagem de avaliação de risco une a ecologia microbiana e a ecologia florestal para desenvolver uma compreensão mais abrangente da vulnerabilidade das florestas no século XXI.

11 **Introduction**

12 Forest ecosystems cover approximately one-third of the Earth's land surface, harboring immense
13 biodiversity¹ and supporting diverse human cultural needs and values². Forests sequester over
14 three-quarters of terrestrial carbon globally³, however, due to climate change and land use practices
15 in the "Capitalocene"⁴, the functioning and persistence of forest ecosystems represents an area of
16 critical scientific uncertainty^{5,6}. Investigations of forest response to climate change have largely
17 focused on tree demography and ecophysiology, satellite measurements, and restructuring of forest
18 community structure and function. Collectively, these studies indicate that fire, drought, increasing
19 CO₂ concentrations, and rising temperatures imperil the health and carbon sequestration potential
20 of forest ecosystems^{5,6}. Improving understanding of forest responses to climate change is critical
21 for effective policy and climate adaptation initiatives.

22 The forest microbiome mediates forest responses to a number of climatic stressors
23 including drought⁷, rising temperatures⁸, and elevated CO₂⁹. Microbiomes also play an important
24 role in forest recovery from fire^{10,11} and hurricanes¹², and microbes shape soil methane production
25 and consumption¹³, as well as the ability of soils to sequester CO₂¹⁴. There is now also unequivocal
26 evidence that forest microbial communities are vulnerable to climate change^{15–18}, highlighting the
27 need for their explicit consideration in efforts to predict the fate of forest ecosystems.
28 Observational studies across local to global-scale environmental gradients document how
29 microbial community composition and function are locally adapted, and highly sensitive to
30 prevailing climatological conditions^{17,19–22}. In addition to observational inferences, experimental
31 studies of microbial communities across diverse forest environments demonstrate dynamic
32 compositional and functional responses to climatic disturbances¹⁸. Because rates of microbial
33 responses to disturbance may differ from forest trees, mismatches between plant communities,
34 local-climates, and microbial communities can occur, with important implications for forest
35 function^{23,24}.

36 Forest microbes often exhibit strong context- and scale-dependent responses to
37 disturbance^{25–27}, challenging generalizations of microbiome response to climate change.
38 Accordingly, these context dependencies have stymied the effective incorporation of microbial
39 dynamics into predictive models and understanding of forest ecosystems^{5,6,27,28}. To address this
40 knowledge gap and better predict and integrate microbiomes into a more holistic understanding of

41 forest response to climate change, we adapt a risk assessment framework for forest microbiomes
42 in the Anthropocene.

43

44 **Defining forests and their microbiomes**

45 The Kyoto Protocol (1997) categorizes forests as areas greater than 0.5-1.0 hectares where the
46 minimum “tree” crown cover ranges from 10-30%²⁹. From the boreal to the Amazon, forest
47 ecosystems span broad climatic regions⁶. Throughout these regions, trees scaffold and shape forest
48 systems, giving rise to distinct habitats and microclimates for understory species³⁰. Trees are
49 generally defined as plants capable of growing at least two meters tall²⁹, though their stature and
50 architecture varies greatly by species³¹. Lifespans of trees are also highly variable across taxa,
51 ranging from 25 to over 5000 years³². The architectural, demographic, and ecophysiological
52 characteristics of trees are predicted to play important roles in forest responses to climate change^{5,6}.

53 The forest microbiome – a rich diversity of bacteria, archaea, fungi, lichens, and viruses –
54 supports plant productivity and forest function across the globe^{33–35} (Fig. 1a). Forest ecosystems
55 are among the most biodiverse microbial habitats on Earth. For example, individual leaves can
56 host hundreds of bacterial and fungal taxa³⁶ and tens of thousands of microbial species inhabit
57 forest soils³⁷. Forest microbes span a range of interaction types, including pathogens and
58 mutualists. These microbes cumulatively facilitate nutrient cycling^{35,38} and impact plant
59 community structure by mediating plant coexistence and diversity^{39–41}. Key microbial groups in
60 soil include saprotrophic fungi and bacteria, which drive decomposition and soil carbon
61 accumulation⁴². Saprotrophic microbes mediate soil nutrient availability by mineralizing organic
62 nitrogen into plant-available inorganic forms^{42,43}. Other key groups include ancient interactions
63 between tree roots and mutualistic mycorrhizal fungi, such as arbuscular (AMF) and
64 ectomycorrhizal fungi (EMF), which shape host environmental tolerances⁸ and represent an
65 important component of nutrient acquisition strategies for trees¹⁹ (Fig. 1c). Similarly, mutualistic
66 relationships between roots of certain tree species and N-fixing bacteria influence biogeochemical
67 cycling in forest systems³⁵. Fungal and bacterial endophytes in roots and leaves can protect trees
68 against pathogens³⁶, promote drought tolerance⁴⁴, influence host thermal tolerance³⁶, and
69 contribute to host N supply through N-fixation⁴⁵ (Figure 1a). N-fixing bacteria have also been
70 found in forest soils, leaves, and leaf litter, as well as in association with mosses and lichens,
71 contributing to forest N-availability and biogeochemical cycling⁴⁶. In this review, we especially

72 focus on the bacterial and fungal components of the forest microbiome given available knowledge
73 and their pivotal role in forest function³⁴.

74

75 **A risk assessment framework for forest microbiomes**

76 Risk assessments are used to predict and manage the vulnerability of diverse systems by providing
77 insights into where exposure may be reduced or sensitivity might be mitigated^{6,47}. Because risk
78 frameworks have predominately been used in macroecological contexts⁴⁷, to utilize this
79 framework, we first define microbially-relevant scales of exposure and attributes of microbial
80 communities and individuals that govern their sensitivity to climatic disturbance (Fig. 2)^{15,47,48}.
81 Then, we define important feedbacks that can inform the trajectory of forest microbiome responses
82 to ongoing climate change. This framework intends to synthesize forest microbiome response to
83 climate change in order to unite forest microbial ecology with macroecological understanding of
84 forest responses to climate change⁶. While we specifically apply the framework in a forest
85 ecosystem context, the principles should also be broadly applicable, and the framework is intended
86 for diverse terrestrial ecosystems.

87

88 **Exposure**

89 The effects of warming, drought, and other climatic disturbances vary across geographic and
90 temporal scales⁶. We define exposure risk as the probability that climatic disturbance will extend
91 beyond physical baseline conditions enough to impact forest microbiome community structure or
92 function. Accordingly, quantifying climatic exposure risk for the forest microbiome depends on
93 the integration of three different components of climatic disturbance: disturbance type,
94 spatiotemporal scales of disturbance, and the capacity for physical buffering of disturbance across
95 microbial habitats.

96 **Categorizing climatic disturbances**

97 The persistence of forest systems is threatened by the increasing frequency and severity of climatic
98 disturbance⁶. Different types of perturbations associated with a changing climate can be
99 categorized in a pulse-press framework to understand their potential impact across spatiotemporal
100 scales and intensity of disturbance^{15,49}. Disturbances like elevated CO₂ (eCO₂) and rising air

101 temperatures represent press events, disturbances which are ongoing and relatively slow⁴⁹. Press
102 disturbances are predicted to select for microbial traits associated with greater physiological
103 plasticity and niche breadth (tolerance), and strong acclimation or rapid adaptation will be required
104 to maintain community composition as a result of continuing disturbance^{15,16,50}. Conversely, pulse
105 events are discrete disturbances, including droughts, hurricanes, floods, or wildfires⁴⁹. Pulse
106 disturbances tend to favor microbial community traits associated with rapid recovery (often
107 referred to as resilience)¹⁵. Though pulse disturbances like forest fires can initially reduce soil
108 fungal and bacterial biomass and diversity^{51,52}, pulse disturbances have a defined end period, in
109 some cases allowing microbial communities to eventually recover parts of their original structure
110 and function¹⁵. However, recovery of microbiome function following pulse events is variable and
111 can take as long as several decades¹⁰. While the pulse-press framework is conceptually useful for
112 comparing and synthesizing the physical impacts of diverse climatic stressors on organisms and
113 communities, it is important to note that both press and pulse disturbances increasingly act in
114 tandem⁵³.

115 Consideration of microbial lifespan and life-history strategies are essential to
116 contextualizing exposure risk¹⁵. For instance, some long-lived soil fungi can persist for years to
117 decades⁵⁴, whereas many co-occurring soil bacteria turnover on the order of hours or days in
118 response to drought⁵⁵. As a result, while a multi-month drought may represent a pulse disturbance
119 for a tree or long-lived fungus, droughts typically impact many generations of more ephemeral
120 fungi or bacteria¹⁵. Consequently, bacteria and some fungi may undergo rapid evolution in
121 response to pulse disturbances, whereas evolution for some fungi may occur over longer
122 timescales⁵⁶.

123 Climatic disturbances can act either directly on microbial cells, or through indirect
124 pathways. For example, drought and rising temperatures can directly stress microbial cells^{55,57}. In
125 contrast, eCO₂ indirectly impacts soil communities, primarily via shifts in substrate-
126 availability^{58,59}. Greater C allocation of trees belowground in response to eCO₂ can increase
127 colonization of roots by both EMF and AMF fungi, and result in greater rhizodeposition,
128 increasing availability of carbon-substrates to microbes^{60,61}. Tracing the various pathways
129 whereby climatic stressors directly or indirectly impact microbial communities will be critical to
130 accurately projecting the magnitude of community responses.

131 **The spatial and temporal nature of climatic disturbances**

132 The type, intensity, frequency, and timing of disturbances varies dramatically across forest
133 biomes^{6,18} (Fig. 3). Catastrophic droughts, flooding, and severe hurricanes affect tropical forests,
134 while reduced snowpack and permafrost melt threaten boreal and high-elevation forest systems⁶².
135 In many temperate forests, drought and rising temperatures compound with settler-colonial fire
136 suppression to create fires of increasing frequency and intensity⁶³. Increasing frequency and spatial
137 extent of disturbance threaten the capacity of microbial communities to withstand and/or recover
138 their pre-disturbance community structure⁶⁴ and shifts in pathogen ranges, including bark beetle
139 and invasive microbial diseases, can result in mass tree mortality, jeopardizing forest persistence
140 in some systems^{7,65}. Importantly, the intensity of disturbance will also vary across forest biomes.
141 The rate of warming, for instance, differs across the globe with boreal forest temperatures projected
142 to increase at approximately twice the rate of other forests⁶⁶ (Fig. 3a-b). As a result, microbiome
143 exposure to warming is likely greater in boreal compared to temperate and tropical forest
144 systems⁶².

145 For pulse-type disturbances, the timing of a disturbance also influences microbiome
146 exposure risk, especially for forests with strong seasonality (Fig. 3c). For example, spring fires
147 result in greater reductions in fungal saprotrophic diversity compared to fires of similar severity
148 which occur in autumn⁶⁷. One hypothesis is that spring fires are more damaging for microbial
149 growth and reproduction than fires that occur during periods of higher microbial dormancy⁶⁷.
150 Climate change will continue to advance the onset of summer and spring, while autumn and winter
151 seasons are expected to be shortened and delayed⁶⁸. Thus, longer periods of microbial activity may
152 increase the probability of greater impact of pulse disturbances on microbial community structure
153 and function. Additionally, autumn is an especially critical ecological period, especially for many
154 fungi, as this is when they transition from high periods of activity to reproduction⁶⁹. Scarce datasets
155 in Western science track long-term shifts in microbial reproduction, making generalizations about
156 shifts in fruiting phenology associated with climate change challenging^{70,71}. However, many
157 Indigenous communities have been observing patterns and processes of fungal phenology for
158 thousands of years⁷². For example, the Karuk tribe (Northern California, USA) has demonstrated
159 how delays in autumn (warmer temperatures continuing into October/November and later rainfall)
160 lead to declines in ectomycorrhizal fruiting, with important consequences for the communities and
161 ecosystems these fungi help to sustain⁷³ (Box 1). Indigenous science has a critical lens into how

162 climate change is shaping forest ecosystems as its period of observation extends back tens of
163 thousands of years; the centering of Indigenous-led science will be critical in better understanding
164 and managing forest systems in a changing climate⁷⁴.

165

166 **Microclimates, microhabitats, and microbe-specific niches**

167 The physical structure of forest ecosystems plays an underappreciated role in determining the
168 exposure risk of different microbial communities to climatic disturbance (Fig. 3d). Due to their
169 immense structural complexity relative to other terrestrial ecosystems, different locations in forests
170 confer varying degrees of physical buffering from climatic disturbance³⁰. Throughout the forest
171 canopy, light, temperature, and moisture conditions vary dramatically^{75,76}. Microbes associated
172 with different forest compartments (e.g. leaves versus roots) experience varying degrees of
173 physical buffering (Fig. 3d-e) and have different inherent exposure risks to climatic disturbances.
174 To distinguish these from the physical structures that hosts can evolve to spatially separate
175 symbionts (compartmentalization), herein, we refer to these different microbially habitats (leaves,
176 roots, soil, etc.) as microbial-specific niches (MSNs)³⁴.

177 Plant hosts can also actively and passively modulate microbiome exposure. For example,
178 bacterial communities associated with roots of *Sequoia sempervirens* are less sensitive to water-
179 deficit relative to those in soil, potentially due to consistent water availability in roots²². In
180 *Eucalyptus*, latent cooling through evapotranspiration reduced leaf temperatures by up to 7.5°C
181 relative to air temperatures during a heatwave⁷⁷, helping to buffer heat exposure for phyllosphere
182 and endophyte microbial communities. Active cooling via evapotranspiration may also help to
183 explain how foliar endophyte communities associated with *Pinus ponderosa* can maintain
184 community structure whereas root-associated microbial communities are reshaped by warmer
185 temperatures⁷⁸. The soil matrix can also buffer microbiome exposure. For example, the
186 composition and diversity of bacterial and fungal communities in the soil surface (0-5 cm depth)
187 are often strongly impacted by fire, but heat from surface fires attenuates rapidly with soil depth,
188 suggesting that deeper soils may act as a potential refugia for microbial communities⁷⁹ (Fig. 3e).

189 Canopy gaps also create distinct microclimates (Fig. 3d). These gaps are relatively warmer
190 and drier, and consequentially, distinct wood decay fungal communities inhabiting them exhibit
191 reduced rates of decomposition⁸⁰; similarly, canopy gaps may reduce the local diversity of EMF
192 communities⁸¹. In xeric systems, removal and management of understory plants through practices

193 such as cultural burning can increase soil water content and nutrient availability⁸², providing
194 important habitat for plant and microbial communities by reducing understory plant competition⁸³.
195 However, extensive habitat fragmentation (due to logging or large-scale climatic disturbances like
196 stand-replacing fires or hurricanes) may exacerbate forest warming through reduced shading and
197 evapotranspiration, further threatening forest communities. Altogether, the unique buffering
198 capacity of distinct MSNs, suggests that microbiome community structure and function could shift
199 at different rates across leaf, root, and soil communities potentially causing dysbiosis for tree
200 hosts⁸⁴. Exploring how different MSN combine to collectively influence plant health remains an
201 intriguing area of future research.

202 **Sensitivity**

203 In the broadest terms, sensitivity describes the intrinsic factors that determine how a community,
204 species, or individual is impacted by climatic disturbance⁴⁷. At the community scale, sensitivity
205 describes the degree to which microbial community composition or function responds to
206 disturbance and their capacity to recover composition or function post-disturbance (Fig. 4a)⁸⁵. At
207 the species and individual scales, low sensitivity is the persistence of a taxon or individual or their
208 capacity to recover physiological function following a short-term disturbance⁸⁵. Here, we identify
209 and describe evolutionary and ecological processes that modulate the sensitivity of microbial
210 populations and communities.

211

212 **Global variation in the sensitivity of microbial communities**

213 Surveys across continental and global scales have identified strong biogeographic patterns in
214 microbial communities, with high degrees of endemism (for some taxonomic groups) and unique
215 functional trait profiles across the major tropical, temperate, and boreal forest biomes²¹. This
216 suggests that climatic disturbances may result in divergent responses for microbial communities
217 inhabiting different biomes or habitats. The biogeographic patterns in microbial communities
218 result from prevailing climatic regimes (e.g. mean annual temperature; MAT)²¹, geochemical
219 attributes (e.g. parent material, pH, Ca)⁸⁶, different evolutionary origins of microbial taxa⁸⁷, and
220 the composition of forest tree species^{88,89}. Additionally, limitations to microbial dispersal can drive
221 distinct community⁹⁰ and population structures⁹¹.

222 In the face of climatic change, varying sensitivity of microbial communities can result in
223 several response scenarios: tolerance (resistance to climate disturbance⁹²), recovery (initial change
224 followed by return to original abundance or community structure⁹³), and transition (i.e. to an
225 alternative state^{15,50} (Fig. 4a). There is growing consensus that communities which have
226 experienced greater historical environmental fluctuations will be more likely to tolerate future
227 disturbance^{50,94}. For example, forest biomes in which temperature fluctuations are relatively large,
228 such as in high-latitude forests, exhibit relatively small compositional changes when exposed to
229 warming compared with lower latitude forests^{95,96}. Similar patterns of tolerance have also been
230 observed for short-lived disturbances like drought. In temperate forests, regions with lower
231 historical precipitation select for microbial communities with higher tolerance to acute drought⁹²,
232 including Actinobacteria, Firmicutes, and Thaumarchaeota^{22,97}. Historical legacies of water-
233 availability also correspond to drought tolerance for microbes in tropical forests; in forests where
234 drought treatments had been experimentally imposed, tropical microbial communities experienced
235 less pronounced shifts in community composition compared to the controls with no prior drought
236 exposure⁹⁸.

237 Community recovery potential also appears to vary across biomes with different
238 disturbance regimes. In forests where fire is historically common, microbial taxa and even whole
239 microbial communities recover rapidly, even from severe disturbance^{99–102}. Microbial community
240 recovery is facilitated by an initial increase in fire-specialized taxa⁵¹. For example, spores of a
241 root-associated EMF species (*Rhizopogon olivaceotinctus*), nearly double their colonization
242 potential following heating¹⁰³, and soil-dwelling *Arthrobacter* bacteria experience post-fire
243 population booms⁵¹. These pyrophilous microbes may facilitate recovery of the original
244 community by breaking down hydrophobic or pyrolyzed organic matter⁹⁹. Importantly, early
245 successional microbes are generally poor competitors that do not appear to prevent re-
246 establishment of the initial community¹⁰⁴. In contrast, for forest ecosystems adapted to less
247 frequent fire regimes and having fewer pyrophilous community members, populations of both
248 saprotrophic and mutualistic microbes are heavily reduced after fire and appear less likely to
249 recover their original state^{52,100} (Fig. 4a).

250 The impacts of increasing frequency of disturbance and compounding disturbance types
251 might be exceptionally stressful for microbial communities, potentially impeding community
252 recovery and leading to a transition to alternative states¹⁰⁵. More frequent disturbances are

especially likely to result in functional collapse and transition in microbial ecosystems⁶⁴. For example, while microbial recovery potential appears high in fire adapted landscapes, fire regimes are shifting rapidly⁶. In fire-adapted *Eucalyptus pilularis* forests of Australia and coniferous forests in Western USA, fire adapted microbial and lichen communities appear less likely to recover with increases in fire frequency¹⁰⁶, especially when combined with periods of intensifying drought stress¹⁰¹ and emerging infectious disease¹⁰⁷. Microbial community transitions are also predicted where disturbance results in novel environmental conditions. For example, rising temperatures decrease climatic controls on rates of decomposition, leading to predicated transitions from EMF to AMF-dominated forests, especially at boreal-temperate ecotones¹⁹.

Studies documenting microbial community tolerance, recovery, or transition may not necessarily correspond with identical shifts in microbiome functioning. This decoupling could arise due to processes of microbial acclimation to warming via physiological shifts in cellular processes¹⁰⁸. Similarly, high degrees of functional redundancy in microbial communities could result in modest or non-linear functional responses despite strong community shifts. Relative decoupling of composition-function relationships could also vary across forest biomes if historical environmental fluctuations exert a strong impact on community sensitivity and a more variable impact on functional sensitivity. Accordingly, in boreal forest soil communities, prolonged warming may trigger moderate community shifts (tolerance) and a temporarily increased but progressively weakening effect on soil respiration due to microbial acclimation¹⁰⁹. This response contrasts with observations of accelerating community and respiration responses in experimentally warmed tropical forests⁹⁵. Sufficient temporal inference is lacking to fully describe these relationships, and further work is required. We stress that community tolerance, recovery, and transitions represent operational definitions; the linkages between community composition and microbiome function remains one of the most critical areas of ongoing investigation¹¹⁰.

277 Variation in sensitivity across microbe-specific niches

Microbes are adapted to distinct environmental fluctuations and resource availabilities across microbial-specific niches (MSNs) and therefore exhibit unique sensitivities among these habitats^{34,111,112}. For example, wood decay fungi such as *Phlebiopsis* and *Phellinus* exhibit optimal growth temperatures that vary across biogeographic ranges but generally fall between 20-28°C, with growth declining rapidly above 30°C¹¹³. By contrast, the surface of sunlit leaves can

283 frequently exceed 30°C¹¹⁴ and leaf-inhabiting microbial endophytes are adapted to warmer
284 temperatures; many of these endophytes, including *Burkholderia*, *Acinetobacter*, and yeasts such
285 as *Rhodotorula*, are regularly cultured at 30°C¹¹⁵. For highly diverse MSNs such as soils, there is
286 evidence that microbes found in these MSNs may be more tolerant as compared to leaf or root
287 communities¹¹⁶. Accordingly, the potential for microbial communities to respond asymmetrically
288 across MSNs, especially above and belowground, is an important area of future investigation³.
289 Studies of potential dysbiosis among MSN and functional consequences for hosts are critical in
290 elucidating forest sensitivity to climate change¹¹⁷.

291 In addition to inhabiting unique physical niches in trees and forests (e.g. in roots versus
292 leaves), microbial host-dependence is another important dimension of MSNs. While some
293 microbes are free-living, others are facultatively or obligately host-associated and exhibit varying
294 degrees of host specificity³⁴. For obligately host-associated microbes or those with high host-
295 specificity, the persistence of host plants is a minimum criterion for the presence of suitable
296 MSNs^{118,119}. For some obligate host associates, an effective tolerance strategy may be to improve
297 host survival of the same disturbance¹²⁰. By contrast, where microbial associations increase
298 mortality risks of their hosts, a decline in available hosts to infect may render these microbes more
299 sensitive to climate change. For example, white pine blister rust (*Cronartium ribicola*) increases
300 host mortality at low elevations during drought, thereby decreasing the population of its available
301 hosts (alongside rates of infection)⁷.

302

303 **Traits and tradeoffs that structure microbial sensitivity**

304 Particular microbial traits predispose microbes to heightened climatic sensitivity because of how
305 trait-tradeoffs interact with climatic perturbation^{22,51,121,122}. Trait-tradeoffs have been observed for
306 bacterial⁴⁸ and fungal taxa, such as EMF¹²³, as well as in response to diverse disturbances,
307 including water deficit²², warming¹²⁴ and fire^{51,102}. For example, trade-offs in bacterial suitability
308 to post-fire habitability and competitive abilities have been observed¹⁰². We expect that
309 investigations exploring the identity and function of specific traits influencing microbial sensitivity
310 at both the taxon and community scale will be an especially active area of future research.

311 Climatic disturbances could significantly impact microbial physiology, as well as
312 community composition and function. Trait tradeoffs between microbial growth yield (Y),
313 resource acquisition (A), and stress tolerance (S) broadly define microbial sensitivity to climatic

314 perturbation (the Y-A-S framework; Fig. 4b)⁴⁸. When climatic disturbance drives resource
315 limitation, such as during drought, microbes exhibit tradeoffs between traits associated with
316 microbial resource acquisition and growth yields^{16,48}. This has been observed for soil bacteria
317 across a natural gradient of water deficit, where filamentous *Streptomyces* bacteria associated with
318 *Sequoia sempervirens* root communities dominate in dry soils but unicellular flagellate bacteria,
319 such as *Bradyrhizobium*, are more dominant in wetter soils (Fig. 4b)²². Filamentous lifestyles excel
320 at resource acquisition by scavenging a larger soil volume, whereas flagellate bacteria have higher
321 growth yields, but are dependent on saturated soils for motility and access to soil resources⁵⁵.

322 Traits associated with stress tolerance are also important to microbial persistence during
323 climatic perturbation^{48,55,124}. For example, exposure to acute drought resulted in an upregulation
324 of genes associated with cell wall and membrane maintenance, but an overall reduction in gene
325 expression and resource acquisition in foraging hyphae for the EMF fungus, *Suillus pungens*¹²³,
326 likely impacting the cellular tolerance of stress and resource-acquisition of this fungus. Similar
327 cellular tradeoffs also appear to apply for rising temperatures. A study of long-term soil warming
328 in the Alaskan boreal forest found that genes associated with cellular maintenance were
329 upregulated compared to genes associated with microbial resource acquisition and growth¹²⁴.
330 Cellular dormancy is another important facet of microbial stress tolerance and could be a common
331 strategy for members of the forest microbiome^{51,125,126}. Due to larger intracellular investment costs,
332 dormancy appears to have important tradeoffs with capacity for vegetative growth rates^{48,51,102}.
333 While little is known about cross forest biome variation in the distribution of cellular dormancy,
334 dormancy and sporulation could be particularly common in forests which experience large
335 seasonal climatic fluctuations or frequent pulse disturbances^{15,94,125}. Soils experiencing frequent
336 fire are enriched in endospore-forming Firmicute bacteria^{10,102} and mycorrhizal fungi which form
337 resistant spore banks may possess an early competitive advantage post-fire¹²⁶.

338 **Feedback mechanisms that amplify or buffer vulnerability**

339 Feedback processes can either amplify or buffer microbiome sensitivity and exposure to future
340 climatic disturbance⁴⁷. Feedbacks within forest systems play an underrecognized role in
341 determining the vulnerability of the microbiome to future climate change. Here, we outline several
342 pathways where biogeochemical, climatological, and biotic feedbacks could modify forest
343 microbiome vulnerability to future climatic disturbance.

344 At the largest scales, shifts in forest microbiome structure and function will influence
345 critical biogeochemical cycles. For example, trees that associate with N-fixing microbial
346 symbionts are favored under increasingly dry forest conditions¹²⁷. However, the dominance of N-
347 fixation in arid soils has also been linked to the release of NO_x, a potent greenhouse gas¹²⁸. As a
348 result of NO_x release, warming and drying are likely to be amplified, increasing the exposure of
349 forests and their microbiomes to further disturbance. Similarly, warming temperatures are poised
350 to stimulate the decay of soil organic matter (SOM) by microbial communities, particularly in
351 high-latitude soils¹⁴. The substantial release of CO₂ from increasing rates of microbial activity will
352 result in a positive climate feedback loop, further exposing forest systems to higher temperatures
353 in the future. Increased rainfall in certain tropical regions can also substantially shift
354 biogeochemical cycles by stimulating microbial methane production, the release of which can
355 further accelerate rising temperatures and other climatic disturbances; this is especially
356 pronounced after deforestation has occurred¹³. Elevated CO₂ (eCO₂) has significantly increased
357 the strength of the terrestrial C sink. This sink is contingent on the sustained nutrient-foraging
358 attributes of mycorrhizal symbionts and their ability to stimulate plant growth^{3,9}. However, for
359 trees associated with EMF, enhanced plant investment in fungal mutualists under eCO₂ could
360 reduce soil C stocks, with uncertain consequences on ecosystem C balance. Moreover, increased
361 EMF foraging under eCO₂ could alter substrate quality for free-living bacteria and fungi³, thereby
362 influencing nutrient mobilization and potentially plant nutrition. Long-term observations of shifts
363 in leaf and root-litter stoichiometry potentially support altered nutrient availability resultant from
364 shifted microbial functioning¹²⁹. In addition to eCO₂ effects, long-term shifts in soil substrate
365 availability and quality could also occur due to drought stress or increased fire frequency¹³⁰, with
366 similar consequences for microbial functioning, plant productivity and draw down of atmospheric
367 CO₂.

368 Climatic disturbances that restructure plant communities or impact plant ecophysiology
369 will also create feedbacks that impact future microbiome exposure and sensitivity. Warming,
370 drying, and increased fire frequency and severity are hypothesized to increase the activity,
371 abundance, and pathogenicity of certain bacteria and fungi^{101,131}. Increased pathogenicity partially
372 results from climatic release where higher winter temperatures increase winter survivorship for
373 pathogens¹³². Increased pathogen abundance can hasten forest compositional turnover for trees
374 already stressed due to changing climate. As climatic niches for EMF and AMF hosts shift with
375 climate change, AMF-associated hosts are predicted to become more dominant in temperate
376 systems¹³³. Because EMF-associated trees are thought to have higher resistance to root pathogens
377 due to physical protection conferred by hyphal mantles surrounding the host roots³⁹, increased root
378 pathogen abundance may interact with other microbial guilds such as AMF or EMF, to generate
379 plant compositional feedbacks³⁹. The potential for transitions between plant communities
380 dominated by different mycorrhizal associates will have consequences for nutrient cycling¹⁹, soil
381 water retention¹³⁴, and forest structure and diversity¹³⁵.

382 Finally, as microbial responses to climate change will influence plant community
383 coexistence and competition dynamics^{8,41}, shifts in forest community structure may generate
384 additional feedbacks for microbial communities. Similarly climatic processes which alter plant
385 community diversity or function could have knock-on effects that mediate the vulnerability of the
386 microbiome to future disturbance. For example, drought can destabilize plant community
387 coexistence in forests through modification of plant-soil-feedbacks¹³⁶. Drought or warming that
388 result in reduced plant richness or plant trait diversity, could also reduce microbial network
389 complexity¹³⁷, microbial multi-functionality, and potentially render communities less resilient to
390 future disturbance¹³⁸. As microbes play important roles in dynamics of plant coexistence and
391 competition^{8,41}, investigating these relationships under different climatic conditions or with novel
392 microbial consortia will be important in determining forest vulnerability to climate change.

393

394 **Future Directions: Microbiome research in the Anthropocene**

395 Synthesizing microbial responses to climatic change remains an ongoing challenge due to the
396 context-dependency of these responses and lack of baseline measurements¹⁸. Moreover, there are
397 relatively few studies which directly compare standardized microbial responses to climatic
398 disturbance across biomes or MSNs. Large-scale manipulative experiments can provide

399 standardized manipulations across a range of biomes (e.g. Free Air CO₂ Enrichment), and such
400 initiatives provide important insights into the role of the microbiome in plant growth dynamics³.
401 Similarly, sampling networks, such as the National Ecological Observatory Network (NEON),
402 enable tracking microbiome and tree responses across natural gradients, over time, or across
403 disturbance events^{139,140}. Importantly, studies that standardize sequencing approaches¹⁴¹, genetic
404 markers^{142,143}, and downstream bioinformatic processing can ensure effective comparisons among
405 studies¹⁴⁴.

406 Another important future direction for forest microbiome research is the incorporation of
407 microbial processes into ecosystem and Earth system models²⁶. One primary challenge involves
408 modeling how rates of nutrient cycling, decomposition, pathogen spread, or tree health will be
409 impacted by future microbiome responses to disturbance (Box 2). This is particularly complex as
410 multiple types of climatic disturbance occur simultaneously, sometimes buffering and other times
411 exacerbating the effects of these disturbances on one another¹⁴⁵. Although microbial processes are
412 often implicitly incorporated into models focusing on projecting future forest dynamics⁶, a specific
413 challenge revolves around understanding the contexts in which microbially-explicit models
414 outperform traditional first-order process-based models¹⁴⁶. Incorporating projections of
415 microbiome functioning could potentially increase the uncertainty of some model projections¹⁴⁷,
416 at least in the near term. Efforts to constrain the relative effect size of microbially-mediated
417 processes in models remains a critical area of research²⁶. Trait-based approaches, which delineate
418 microbial communities based on gene or enzymatic traits, are a promising opportunity to integrate
419 microbes into ecosystem models¹⁴⁸. Additionally, properly measuring trait responses to climatic
420 disturbances at model-relevant spatial and temporal scales will be essential to incorporating
421 microbes into ecosystem models¹⁴⁷.

422 Last, there is bias towards North American and European authors cited in our literature, as
423 well as in other recent reviews on the topic¹⁸; this stems in large part from our own language
424 limitations as well as many structural and systemic biases in research and publication. Research
425 investigating forest microbiome response to climate change and functional consequences for forest
426 ecosystems is especially poorly represented for Latin America, Africa, and Southeast Asia.
427 Decolonial approaches to research will be critical in better understanding and predicting a truly
428 global response of forest microbiomes to climate change.

429

430 **Conclusions**

431 Forest microbiomes play a key role in the structure and function of global forests, but they are
432 threatened by rapid and ongoing climate change. In this review, we adapt a risk assessment
433 framework to identify the vulnerabilities of forest microbiomes to climate change, as well as
434 contextualize potential impacts of microbiomes on forest function. Integrating the forest
435 microbiome into conceptual and empirical models of forest functioning will advance
436 understanding of forest health in the 21st century. There remains significant uncertainty in the
437 cascade of microbiome-mediated feedbacks that will influence forest ecosystem function under
438 climate change, and this represents an important area of future study. Overall, the forest
439 microbiome remains an underappreciated target of conservation initiatives in Western science. We
440 emphasize the importance of Indigenous science and sovereignty in managing Native
441 ecosystems⁷³, as well as the need for more holistic studies linking macroecology with microbial
442 ecology in determining responses of forests to climate change.

443

444 **Box 1 | chí xáyviish nushtúkkareesh (Let's go pick mushrooms!)**

445 Due to limitations in the extent of microbial natural history records in Western science, the study
446 of microbial response to climate change presents many challenges. Consequently, studies of fungal
447 fruiting phenology, for instance, have largely been restricted to temperate forests in Europe over
448 the past several decades using herbaria collections^{70,71}. By contrast, Indigenous communities,
449 including the Karuk Tribe in Northern California, USA have been documenting patterns of fungal
450 fruiting phenology for thousands of years . For example, *Tricholoma magnivelare*, known as
451 matsutake or xáyviish in the Karuk language, is an important EMF species used not only as a food
452 and medicine, but as an indicator species of forest health. Until recently, xáyviish has been
453 accessible and abundant for the Karuk people. However, Karuk Cultural Practitioners describe
454 how while substantial early rains in September stimulate xáyviish development, when seasonal
455 rainfall is delayed as is happening more frequently with climate change (first substantial rain
456 occurring in November), mushroom development ceases for the year.

457 While the impacts of a long-term decline in xáyviish fruiting are not fully known, declines
458 in fungal fruiting may have consequences not just for the fungal taxa themselves, but also for the
459 establishment of seedlings and persistence of forests and the many organisms which they support⁴⁰.

460 For instance, deer and elk rely on xáyyiish in the fall, during their own reproductive periods⁷³. For
461 Karuk people, “Karuk traditional knowledge cannot be separated from either the practices that
462 generated the knowledge, or the practices that emerge from it. These practices, known as
463 ‘traditional management’ are, in turn, central to Karuk culture, identity, spirituality and mental and
464 physical health”⁷³. The centering of Indigenous voices and acknowledgement of Indigenous
465 sovereignty will be critical towards a more in-depth understanding of forest (including forest
466 microbiome) response to climate change.

467

468 **Box 2 | Predicting the assembly of non-analog microbial communities**

469 New methods in biodiversity modeling provide opportunities to forecast the composition and
470 function of non-analog forest microbiome communities under future climates. Non-analog
471 communities are comprised of new combinations of taxa with no contemporaneous parallels. Due
472 to expectations of heterogeneous responses to climatic change (outlined above), non-analog
473 communities are expected to proliferate. To predict the composition and function of these
474 communities, species distribution models (SDMs), sometimes called ecological niche models, use
475 contemporaneous geographic occurrence data along with local environmental variables to generate
476 species range maps. SDMs can then be used with future climatic data to forecast shifts in single
477 microbial taxa or functional groups¹³³. However, concurrently modeling hundreds to thousands of
478 species present in the forest microbiome is challenging using stacked SDM approaches. Joint
479 Species Distribution Models (JSDMs), a relatively new class of statistical models, are well suited
480 to the multivariate and sparse data structures generated by the molecular meta-barcoding studies
481 used to study the forest microbiome¹⁴⁹. These modeling approaches offer insight into the structure
482 and potential function of non-analog communities by accounting for species tradeoffs and spatially
483 explicit species-environment linkages¹⁵⁰. JSDM provides opportunities for non-analog and shifting
484 communities to be incorporated into temporal biogeochemical models seeking to predict forest
485 soil respiration or CO₂ fertilization responses. One of the first studies to apply JSDM to the forest
486 microbiome demonstrated that fungal saprotrophs with strong co-occurrence patterns were
487 especially sensitive to forest management practices, indicating that biotic interactions can impact
488 microbial community responses to disturbance and climate change more broadly¹⁵⁰.

489

490 **Acknowledgments:** The authors wish to thank Joan Dudney, Roxy Cruz de Hoyos, Aidee
491 Guzman, and Rob Jackson for helpful feedback on this manuscript, and Andressa Venturini for
492 help with translating our abstract to Portuguese for our readership. Additionally, we would like to
493 thank Deniss Martinez for her input on potential synergies between Indigenous and Western
494 Science in the context of this article.

495

496 **Author Contributions:** CEW, PTP, and KGP jointly conceived of the manuscript. CEW and PTP
497 jointly wrote the manuscript with input from all co-authors; CEW and PTP jointly share in first
498 authorship. CEW, LMV, SDB, and WT conceived of and wrote Box 1 of the manuscript. VL
499 designed Figure 2 with input from CEW and PTP. CEW designed Figure 2 with input from MVN
500 and PTP. Figure 3 was designed by MVN, CEW, and JAM. Figure 4 was designed by CEW and
501 PTP. All authors have read and approved the final version of the manuscript.

502

503 **Conflict of Interest Statement:** The authors declare no conflicts of interest.

504

505 *Figure captions*

506 **Figure 1 | The forest microbiome in a changing climate.** **A** | The exceptionally diverse forest
507 microbiome colonizes nearly every surface and plant tissue of the forest. Distinct microbial
508 specific niches (MSN) occur throughout the forests, depicted with dashed yellow circles. Major
509 microbial groups and associated MSN include: 1). Foliar endophytes and epiphytes (bacteria and
510 fungi) within the 2). Forest Canopy. 3). Nitrogen fixing free-living bacteria occur in soil. 4). Forest
511 gaps are unique microhabitats within forests. 5). Arbuscular mycorrhizal fungi (AMF);
512 Ectomycorrhizal fungi (EMF), and the associated myco-rhizosphere dominate in soils; **B** | Heat
513 stress and drying associated with climate change will shift the community structure and function
514 of the forest microbiome. A reduction in available water is depicted by the reduction in the blue
515 speckles throughout both panels 6). Wood and litter-decomposers in soil (bacteria, archaea, and
516 fungi). Free-living soil microbes (bacteria, archaea, and fungi occur in soil and in decaying wood).
517 7). N-fixing bacteria may be free-living, root or leaf associated; 8). Forest canopy gaps can be
518 particularly deleterious for microbial communities under climate change 9). Filamentous soil
519 bacteria. 10). N-fixing bacterium associated with plant-roots. **C** | Distinct mycorrhizal symbioses
520 are associated with major forest biomes where they influence biogeochemical cycles and forest

521 responses to climate change. Boreal forest ecosystems are dominated by ectomycorrhizal fungi
522 (EMF), some EMF may enzymatically degrade soil organic matter to obtain organic forms of
523 nitrogen, which influences host nutrition. Contrastingly, in tropical forest ecosystems, the majority
524 of plants are associated with arbuscular mycorrhizal fungi (AMF), which efficiently scavenge
525 nitrate (NO_3^-) and ammonium (PO_4^{3-}) from soil solution. The relative abundance of AMF and EMF
526 hosts is more variable in temperate forest ecosystems.

527

528 **Figure 2 | An organizational framework for studying forest microbiomes in a changing**
529 **climate.** Together, the sensitivity and exposure risk of microbial communities determine their
530 overall vulnerability to climatic disturbance. Subsequent feedbacks may exacerbate or buffer
531 subsequent microbial responses to climate change. We define climatic exposure as the cumulative
532 severity of disturbance. Microbial community exposure is broadly shaped by the disturbance type,
533 spatiotemporal scale, and attributes of the microbial microhabitat. Microbial sensitivity describes
534 the degree to which microbial fitness or community structure and function is impacted by climatic
535 disturbance. Broadly, microbial traits, niche attributes, and the climatic history of a forest can
536 impact the sensitivity of a taxon or community. Finally, climatic feedbacks represent an array of
537 processes, spanning macro-scale biogeochemical processes, to plant soil feedbacks that modulate
538 future microbial exposure and sensitivity (dashed arrows).

539

540 **Figure 3 | Scales of forest microbiome exposure to climate change. A |** Examples of three forest
541 biomes (boreal, temperate broadleaf, and tropical) and the relative distributions of different forest
542 types across the globe. Inset photos are representative examples of major forest biomes. **B |**
543 Exposure risk varies across these three broad forest biomes. Here we compare the mean
544 temperature in the warmest quarter from 1970-2000 to the values predicted for 2061-2080 based
545 on the Coupled Model Intercomparison Projects 5 (CMIP5) from the Intergovernmental Panel on
546 Climate Change (IPCC) Fifth Assessment Report (AR5) in 2014. Three representative forest sites
547 are reported. The predicted warming is based on the intermediate scenario where the
548 concentrations of greenhouse gases (based on the Representative Concentration Pathways; RCP)
549 assumes that emissions will peak in 2040 (RCP 4.5). Below, the line plot denotes the mean monthly
550 temperature fluctuations throughout the year for these same plots from 1970-2000, indicating that
551 microbes from these habitats are adapted to different temperature fluctuations and are liable to

552 demonstrate varying sensitivity to climate change as a result. Data are taken from WorldClim 2
553 (Fick, S.E. and R.J. Hijmans, 2017). **C** | The deviation of monthly mean minimum and maximum
554 temperatures (seasonality) across these three representative forest sites compared to the annual
555 mean annual temperature. Data are taken from WorldClim 2 (Fick, S.E. and R.J. Hijmans, 2017).
556 **D and E** | Temperature fluxes in microbial microhabitats (MSN) are poorly captured in
557 measurements of air temperature. **D** | Temperatures across the different forest microclimates for a
558 representative forest in a Costa Rican Lowland Tropical Forest. **E** | Soil temperatures across
559 different depths for a hypothetical soil, indicating unique microclimates which influence
560 microbiome exposure. Data for D | temperatures in different forest microhabitats, taken from
561 Fletcher et al. (1985). Data for E | mean temperature of soil at different depths taken from Blume
562 et al. (2016).

563

564 **Figure 4 | Microbial sensitivity and underlying traits and tradeoffs.** **A** | Microbial communities
565 exhibit temporally variable responses to climatic disturbance (dashed white line in the grey bar),
566 minimal shift in community structure (tolerance), recovery to pre-disturbance conditions after
567 some recovery period (recovery), or a permanent shift in community structure (transition).
568 Notably, these dynamics could vary across forest biomes (different colored lines). Underlying each
569 response dynamic is a specific example shown below for different disturbance types and microbial
570 communities. **B** | Trait-tradeoffs mediate microbial sensitivity to hypothetical drought stress for
571 rhizosphere bacterial taxa (*Bradyrhizobium*, *Bacillus*, *Streptomyces*). Under ambient conditions,
572 resource abundance may select for taxa with high growth yields (**Y**), and reduced investment in
573 stress tolerance. In contrast, in periods of resource limitation, taxa that can rapidly acquire
574 resources, using extracellular enzymes (**A**) may dominate. Under drought conditions (or other
575 climatic disturbances), stress tolerance (**S**), or stress-tolerant-resource-acquisitive strategies may
576 dominate (Adopted from Malik et al., 2020). We also display an example of how bacterial life-
577 history trait trade-offs can be geographically structured. Trade-offs related to high-growth yield
578 (orange) and resource acquisition (green) are expected to vary across a gradient of water
579 availability; traits associated with resource acquisition may be favored in drier conditions.
580 Hypothetical microbial communities are plotted across the coast redwood range of California,
581 USA where the northern part of the redwood range receives twice as much annual rainfall as the
582 southern extent (Johnson and Dawson, 2011). Consequently, microbes that tolerate water deficit

583 are more abundant in the southern extent of the redwood range whereas microbes with high
584 resource acquisition are more abundant at the wetter part of the range (Willing et al., 2020).

585

586 Photo associated with Box 1 | ***Tricholoma magnivelare*, also known as matsutake, tanoak**
587 **mushroom, or xáyviish in Karuk language.** Photo courtesy of Monika Fischer, University of
588 California, Berkeley.

589

590

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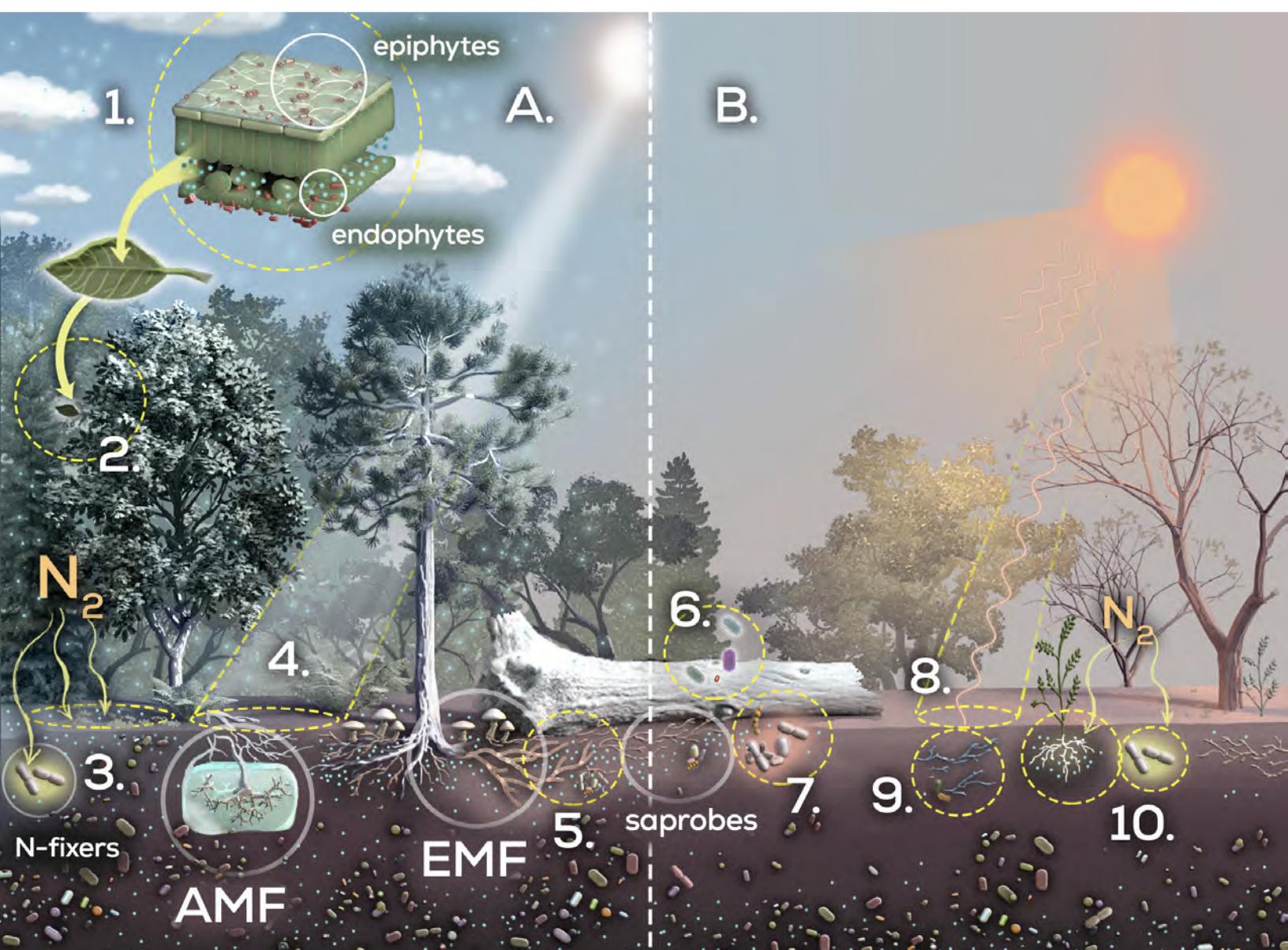
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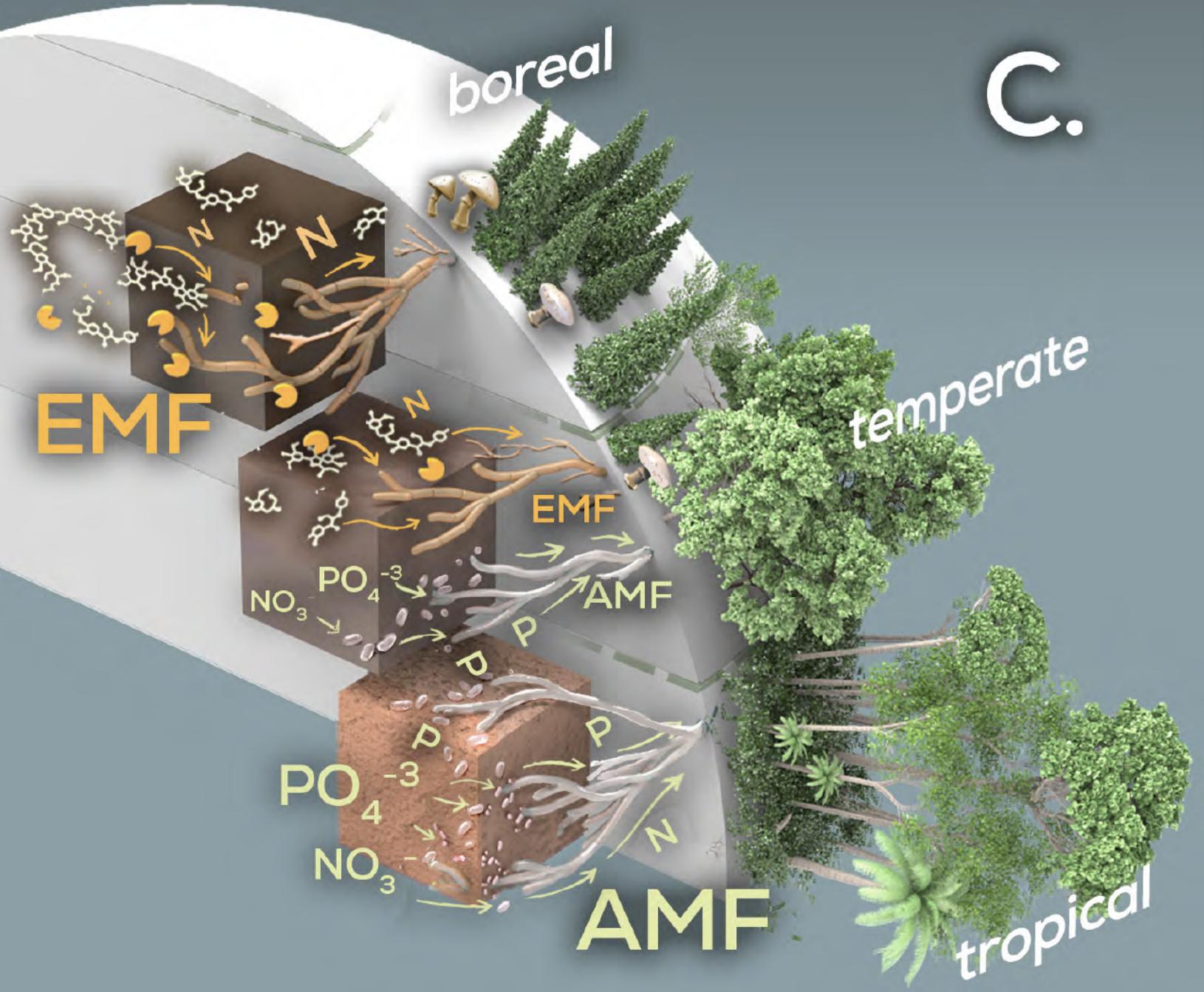
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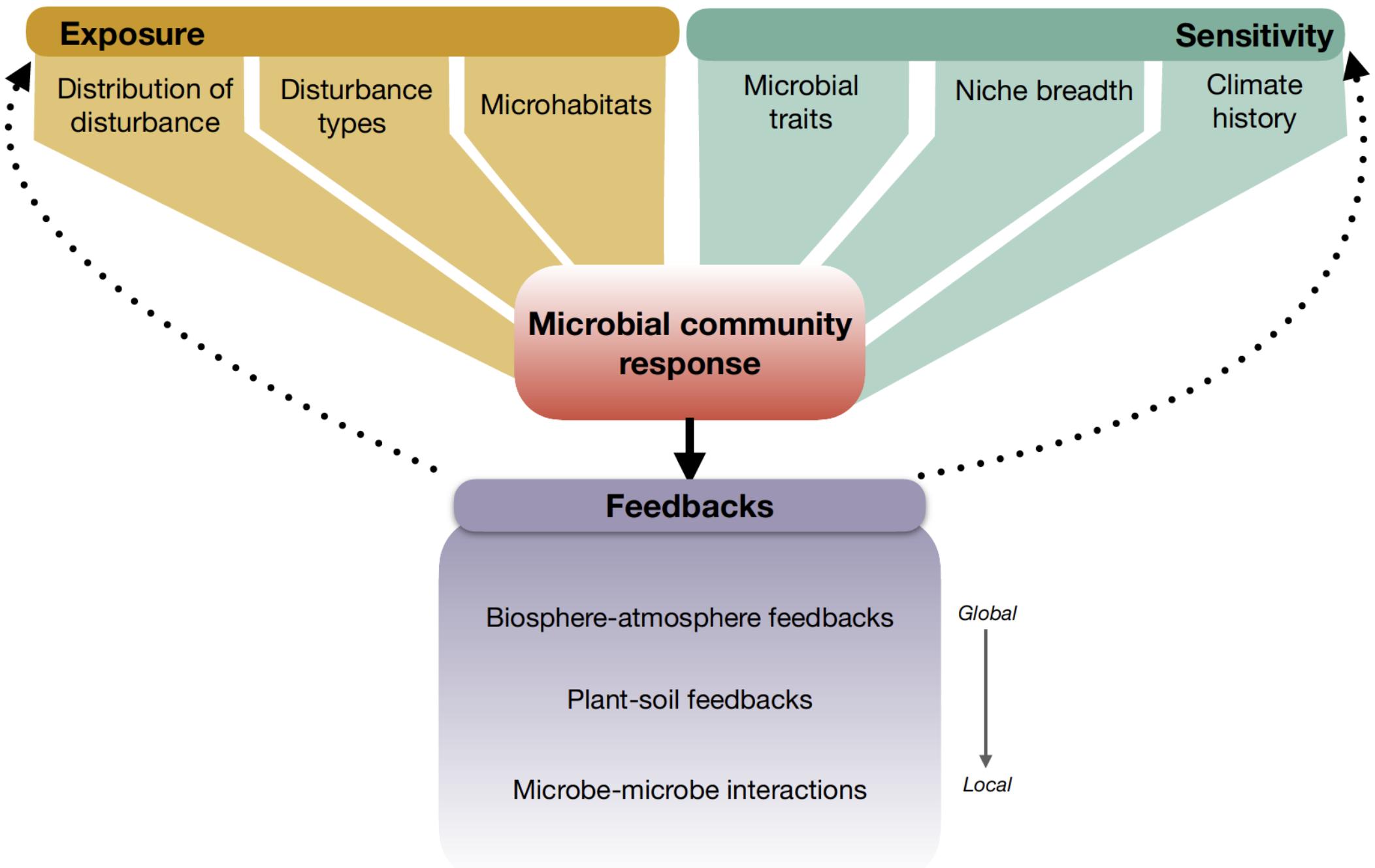
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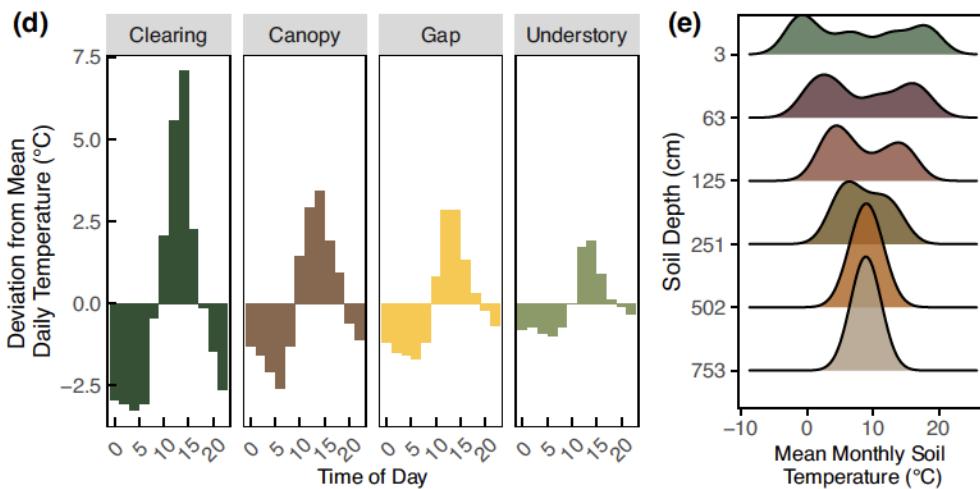
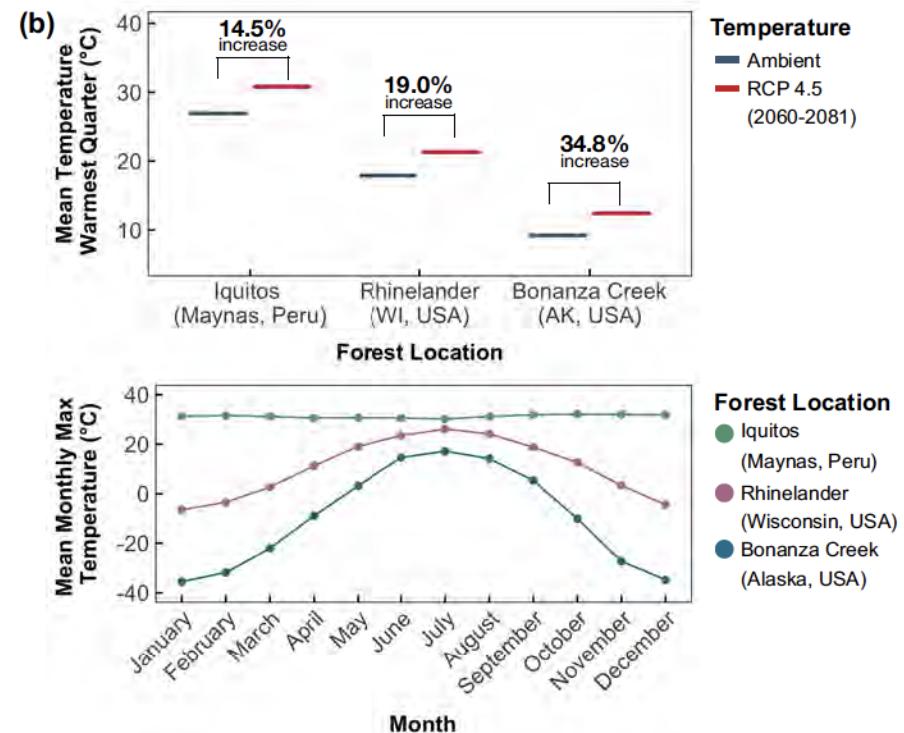
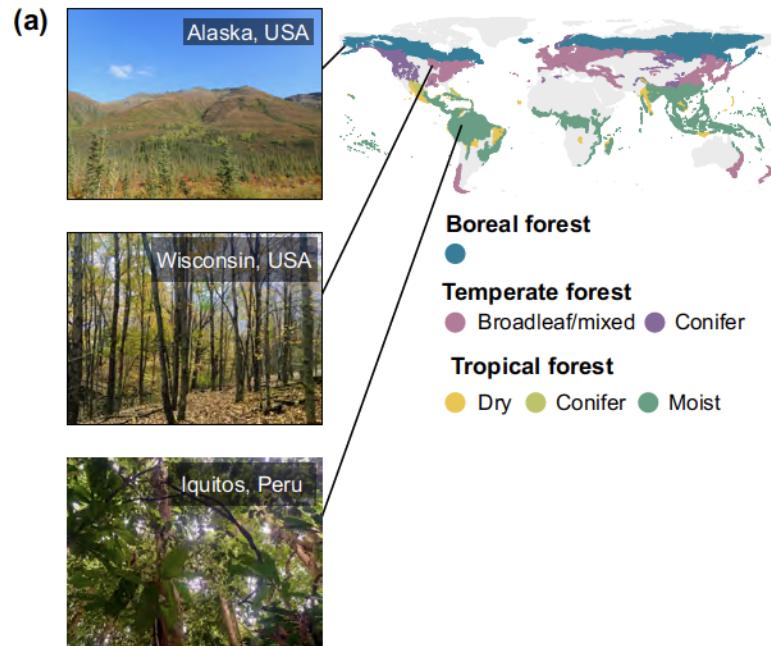
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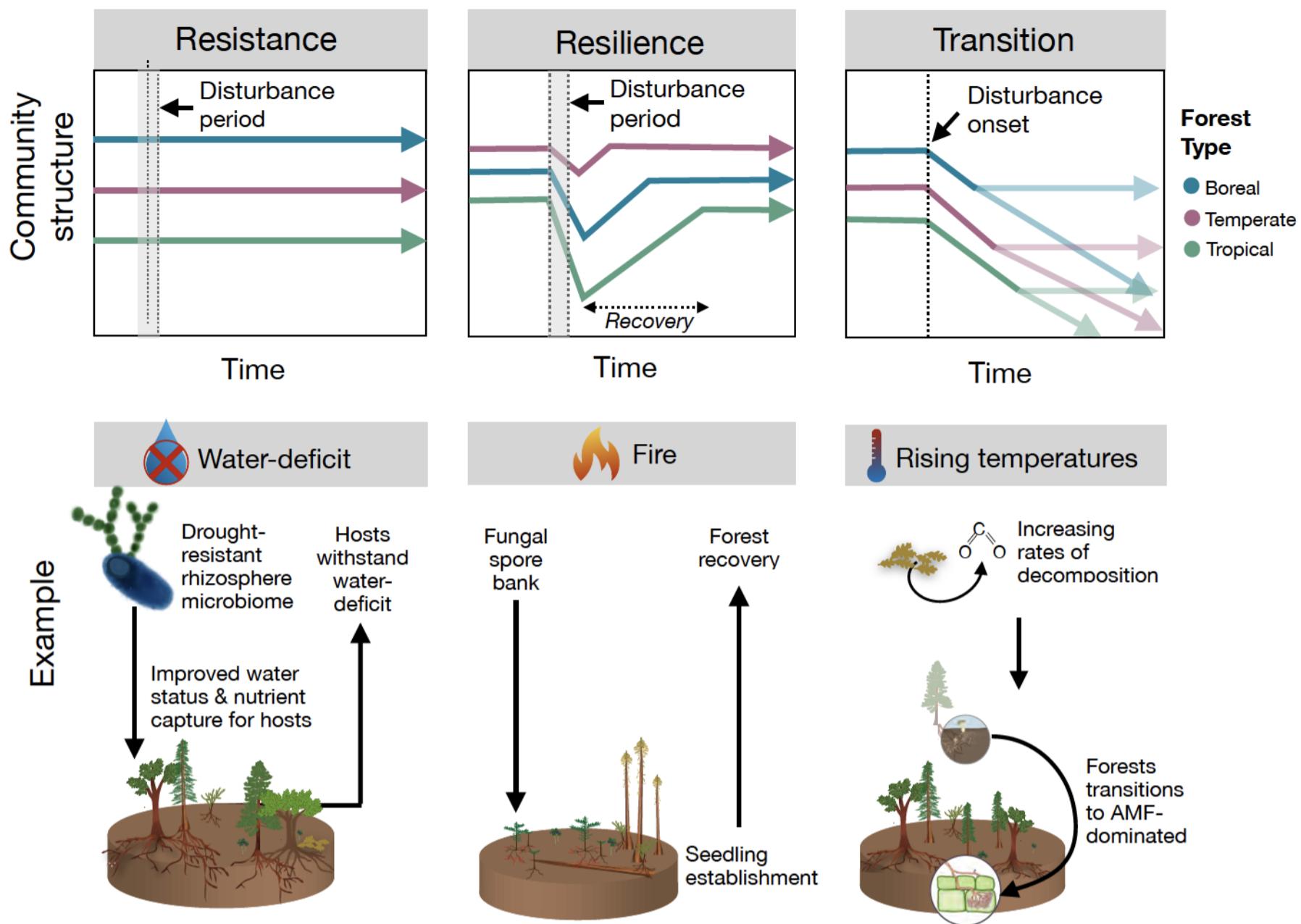






(a)

Community sensitivity

**(b)**

Organismal sensitivity

