



## COMPLIMENTARY/POSTER SESSION PAPER

# Drivers of Intraspecific Variation in Thermal Traits and Their Importance for Resilience to Global Change in Amphibians

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**Synopsis** Intraspecific variation can be as great as variation across species, but the role of intraspecific variation in driving local and large-scale patterns is often overlooked, particularly in the field of thermal biology. In amphibians, which depend on environmental conditions and behavior to regulate body temperature, recognizing intraspecific thermal trait variation is essential to comprehensively understanding how global change impacts populations. Here, we examine the drivers of micro- and macrogeographical intraspecific thermal trait variation in amphibians. At the local scale, intraspecific variation can arise via changes in ontogeny, body size, and between the sexes, and developmental plasticity, acclimation, and maternal effects may modulate predictions of amphibian performance under future climate scenarios. At the macrogeographic scale, local adaptation in thermal traits may occur along latitudinal and elevational gradients, with seasonality and range-edge dynamics likely playing important roles in patterns that may impact future persistence. We also discuss the importance of considering disease as a factor affecting intraspecific variation in thermal traits and population resilience to climate change, given the impact of pathogens on thermal preferences and critical thermal limits of hosts. Finally, we make recommendations for future work in this area. Ultimately, our goal is to demonstrate why it is important for researchers to consider intraspecific variation to determine the resilience of amphibians to global change.

## Introduction

Intraspecific trait variation (phenotypic trait variation within a species) can be more pronounced than variation across species (Albert et al. 2010; Des Roches et al. 2018). Yet, the mechanisms that drive intraspecific variation have historically received little attention in comparison to research that explores phenotypic differences between species (Des Roches et al. 2018). This is notably true for the field of thermal biology, where the majority of studies focus on interspecific variation in thermal traits—even though temperature is considered a significant abiotic factor driving the biology, ecology, and evolution of species and determining why a species lives where it does (Huey and Stevenson 1979; Overgaard et al. 2014; Nowakowski et al. 2018). In ectothermic organisms like amphibians, which rely on environmental conditions and behavior to regulate body temperature, understanding intraspecific variation in thermal traits

is crucial for gaining a comprehensive understanding of how global change impacts populations. While depending on environmental temperatures for thermoregulation may be energy efficient, it renders amphibians especially vulnerable to the effects of climate change, including rising temperatures and increased temperature variability (Deutsch et al. 2008; Bodensteiner et al. 2020).

Amphibians exhibit a variety of climate-relevant traits that researchers examine to identify temperatures that optimize or limit performance and can inform on the consequence of changing temperatures to amphibian physiology (Huey and Stevenson 1979; Llewellyn et al. 2016; see Table 1). Thermal tolerances, measured as critical thermal limits, are an indication of the maximum ( $CT_{max}$ ) and minimum temperature ( $CT_{min}$ ) an organism can survive within and can be a useful metric to understand distributional changes and extinction risk (Angilletta et al. 2002;

**Table 1** Climate-relevant thermal traits and their ecological relevance.

Thermal trait	Term	Abbreviation	Alternative terms or abbreviations used in literature	Description of term and ecological relevance	Example ref(s). showing how thermal trait can relate to species' distribution forecasting
Thermal tolerance	Critical thermal maximum	$CT_{max}$	Upper thermal limit (UTL)	Maximum temperature an individual can survive within. Can inform on distributional range when examined in correlation with environmental temperatures.	<a href="#">Diamond et al. (2012)</a> ; <a href="#">Kellermann et al. (2012)</a> ; <a href="#">Sunday et al. (2019)</a>
	Critical thermal minimum	$CT_{min}$	Lower thermal limit (LTL)	Minimum temperature an individual can survive within. Can inform on distributional range when examined in correlation with environmental temperatures.	<a href="#">Sunday et al. (2019)</a>
	Warming tolerance	WT		The temperature range between $CT_{max}$ and current or predicted environmental temperature. Can inform on the amount of warming that can occur before lethal temperatures are reached.	<a href="#">Deutsch et al. (2008)</a> ; <a href="#">Gerick et al. (2014)</a>
	Acclimation response ratio	ARR		The acclimation capacity of a thermal trait, predominantly calculated with thermal tolerances, and informs on the change in thermal tolerance detected with a change in acclimation temperature.	<a href="#">Barria and Bacigalupe (2017)</a> ; <a href="#">Enriquez-Urzelai et al. (2020)</a> ; <a href="#">Ruthsatz et al. (2022)</a> ; <a href="#">Cicchino et al. (2023a)</a>
Thermal performance	Thermal performance curve	TPC		Curve describing performance over a temperature gradient. Performance is ideally measured as a fitness trait, but often an indirect fitness trait is measured as a fitness proxy. Informs on how performance will change spatially and temporally with variable environmental temperatures.	<a href="#">Clusella-Trullas et al. (2011)</a> ; <a href="#">Ruiz-Aravena et al. (2014)</a>
	Thermal optimum	$T_{opt}$	$T_o$	The temperature at which a given trait reaches maximum performance ( $P_{max}$ ). Informs on the environmental temperature where the trait measured or fitness is at its highest.	<a href="#">Clusella-Trullas et al. (2011)</a>
	Thermal safety margin	TSM		The temperature range between $T_{opt}$ and the current or predicted environmental temperature. Informs on the amount of warming that can happen until a decrease in performance or fitness is seen (small TSM's means a slight increase in environmental temperature will decrease performance, large TSM's mean the species has more buffer from warming). It should be noted that some studies calculate this as $T_{opt} - T_e$ .	<a href="#">Deutsch et al. (2008)</a> ; <a href="#">Huey et al. 2009</a> ; <a href="#">Gerick et al. (2014)</a> ; <a href="#">Sunday et al. (2014)</a>

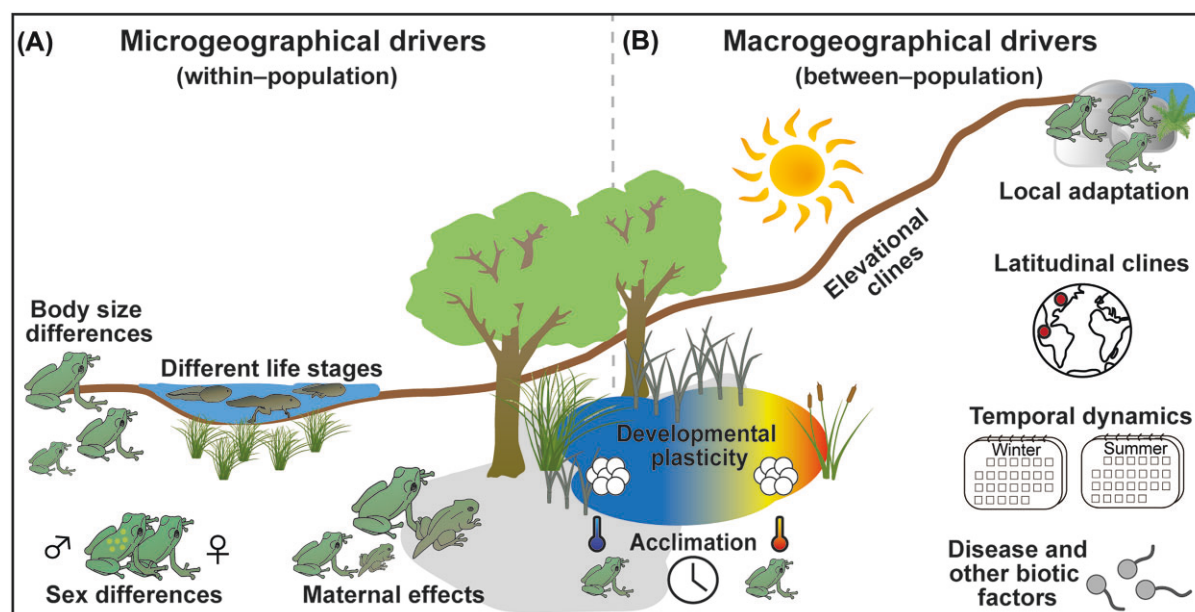
Table 1 Continued

Thermal trait	Term	Abbreviation	Alternative terms or abbreviations used in literature	Description of term and ecological relevance	Example ref(s). showing how thermal trait can relate to species' distribution forecasting
	Operative temperature	$T_e$	Often used as a proxy to body temperature ( $T_B$ )	The temperature(s) that an individual experiences within their natural environment. Can inform on the actual temperature's species will need in their environment when microhabitats are considered (and not solely based on average air temperature).	<a href="#">Camacho et al. (2015)</a>
	Optimal temperature range	$B_{80}$	$T_{br}$ , $P_{br}$ , $P_{80}$	The range of temperatures where performance is above 80% of the maximum performance. This range of optimal temperatures may be used in place of $T_{opt}$ .	
	Temperature preference	$T_{pref}$	$T_p$	The environmental temperature an individual will select to reside in. Informs on the thermoregulatory behavior of species in the wild.	<a href="#">Ruiz-Aravena et al. (2014)</a>
Thermal preference	Temperature selection	$T_{sel}$		A term to describe the temperature range an individual will reside in that is non-preferential but chosen due to experimental design (for example, an individual may choose to reside on one side of a thermal gradient to avoid extreme temperatures on the other side of the gradient, but this may not reflect a true preference). May inform on the thermoregulatory behavior of species in the wild when considering experimental methodology.	<a href="#">Navas et al. (2021)</a> ; <a href="#">Delgado-Suazo and Burrowes (2022)</a>
	Temperature preference range	$T_{set}$	$T_{set(80)}$	A range of environmental temperatures an individual will select to reside in. Informs on the thermoregulatory behavior of species in the wild.	

Climate-relevant thermal traits that researchers use to examine species' thermal niches, common abbreviations found throughout the literature, descriptions of terms and their ecological relevance, and examples of studies showing how thermal traits can inform on species responses to climate change.

[Kellermann et al. 2012](#); [Khaliq et al. 2023](#)). Thermal performance is measured by quantifying a relevant trait across temperatures and creating a thermal performance curve (TPC), and identifies at what temperature(s) fitness (or indirect fitness) is optimal ( $T_{opt}$ ) (see [Sinclair et al. 2016](#) for further discussion). Temperature preference ( $T_{pref}$ ) or temperature selection ( $T_{sel}$ ; [Navas et al. 2021](#)), which allow ectotherms to control body

temperature by moving to suitable microclimates, is often quantified to examine thermoregulatory behavior and can inform on how behavior will affect vulnerability to climate change. Temperature preference can aid or buffer natural selection because it modulates how often individuals are exposed to raw environmental conditions, and hence selective pressures ([Bogert 1949](#); [Muñoz and Bodensteiner 2019](#)). While broad trait



**Fig. 1** Micro- and macrogeographical drivers of intraspecific thermal trait variation in amphibians. Intraspecific variation in thermal traits can be found at **(A)** small, microgeographical scales (within-population) and across **(B)** large, macrogeographical scales (between populations) and may be caused by multiple drivers. For example, differences in body size, life stage, and sex can all contribute to variation found within a single population. Phenotypic plasticity produced from acclimation, developmental plasticity, or maternal effects may be important contributors to both within and between population variation. Adaptation to local environmental conditions, including differences driven by elevational and latitudinal clines, are important spatial drivers when considering macrogeographical patterns—as are large-scale temporal dynamics, including seasonal effects, and disease, which may play an important role in intraspecific thermal trait variation. Credits: Frog and tree symbols: Tracey Saxby, Integration and Application Network ([ian.umces.edu/media-library](http://ian.umces.edu/media-library)); White cattail: Jane Hawkey, Integration and Application Network ([ian.umces.edu/media-library](http://ian.umces.edu/media-library)); Grasses: Dylan Taillie, Integration and Application Network ([ian.umces.edu/media-library](http://ian.umces.edu/media-library)).

comparisons are often conducted between species (e.g., Hoffmann et al. 2013; May et al. 2019; Bennet et al. 2021; Pottier et al. 2022; DuBose et al. 2024; Pottier et al. 2024; Ruthsatz et al. 2024), ignoring variation within a species may result in inaccurate predictions of climate change impacts.

Intraspecific variation is often discussed in terms of macrogeographical patterns, such as latitudinal or elevational gradients that drive temperature variation to which local populations are exposed (Phillips et al. 2016). These large-scale spatial and temporal dynamics (e.g., seasonal variation) may cause between-population differences in thermal traits (Fig. 1). Yet, thermal conditions found in and across microhabitats at a smaller, local scale may also drive intraspecific thermal trait variation within a population (Fig. 1). Microgeographical spatial and temporal thermal effects have been shown to result in adaptive differences in thermal performance within a single population (Blanckenhorn 1991; Kawecki and Ebert 2004; Blanquart et al. 2013; Richter-Boix et al. 2015), but receive little attention compared to large-scale patterns (Phillips et al. 2016). Here, we aim to highlight the drivers of micro- and macrogeographical intraspecific thermal trait variation in amphibians. We examine the drivers of intraspecific vari-

ation in thermal tolerance, thermal performance, and temperature preference because these traits can provide key information on how temperature may restrict an amphibian's current and predicted distribution, informing on species vulnerability, evolutionary trajectories, and resilience under future climate change scenarios (e.g., Sunday et al. 2011, 2019; Claunch et al. 2023; also see Table 1 for information on other climate-relevant metrics). Second, we make recommendations for future work in this area. Ultimately, our goal is to demonstrate why it is important for researchers to holistically consider intraspecific variation to determine the resilience of amphibians to global change.

## Methods

We surveyed the literature by searching Google Scholar with the following keywords: “intraspecific variation” or “within-species variation” AND “amphibian” AND either “thermal biology,” “thermal preference,” “temperature preference,” “thermal performance,” or “thermal tolerance.” For a more comprehensive review of references on ontogeny, please see Bodensteiner et al. (2020) that may contain additional literature on developmental plasticity. For macrogeographical drivers (i.e., local adaptation, elevational clines, latitudinal studies,

temporal dynamics, and range-edge dynamics), we only included studies where it was apparent that thermal traits from more than one population of a given species were examined. This means that some studies may have focused on interspecific variation but in doing so, had included two populations of a single-species, and thus, the study was included here. Please note that this was not intended to be a comprehensive review, but to highlight the importance of considering intraspecific thermal trait variation in amphibians.

### Microgeographic sources of intraspecific variation

Intraspecific variation within a population or across a small geographical area is important when considering that this is the scale relevant to gene flow and dispersal in amphibians. Although a common assumption is that high rates of gene flow in small areas will prevent local adaptation, fine-scale patterns in thermal environments are important for generating phenotypic variation for selection to act upon (see Richardson et al. 2014 for review and discussion on microgeographical adaptation).

Of the relatively few studies that examine thermal trait variation at fine spatial scales, work has focused on intraspecific variation in critical thermal limits and temperature preference in adult herpetofauna (e.g., Muñoz et al. 2014; Bestion et al. 2015; Gilbert and Miles 2017; Goodwin et al. 2019; Herrando-Pérez et al. 2019; Rivera-Ordóñez et al. 2019; von May et al. 2019; Percino-Daniel et al. 2021; Ohmer et al. 2023) (see Supplementary Table S1 for studies on amphibians). However, physiological differences between life stages and sexes, varying body sizes, or due to phenotypic plasticity can drive phenotypic differences that can contribute to resilience to changing conditions.

### Ontogeny

Thermal physiology can vary across life stages and sexes, and there are often strong patterns associated with body size, although the mechanisms driving these relationships are still debated (Bodensteiner et al. 2020; Verberk et al. 2021; Gunderson 2024). When considering ontogeny, ectothermic organisms may experience “critical windows” of heightened likelihood to phenotypic change that can drive intraspecific variation, including susceptibility to thermal stress during development (Mueller 2018). This may be the result of aerobic demands at different life stages due to morphological and/or physiological reorganization (Pörtner 2002; Leiva et al. 2019; Ruthsatz et al. 2024), and/or adaptation to different thermal environments individuals are exposed to (as in the transition from aquatic tadpole to

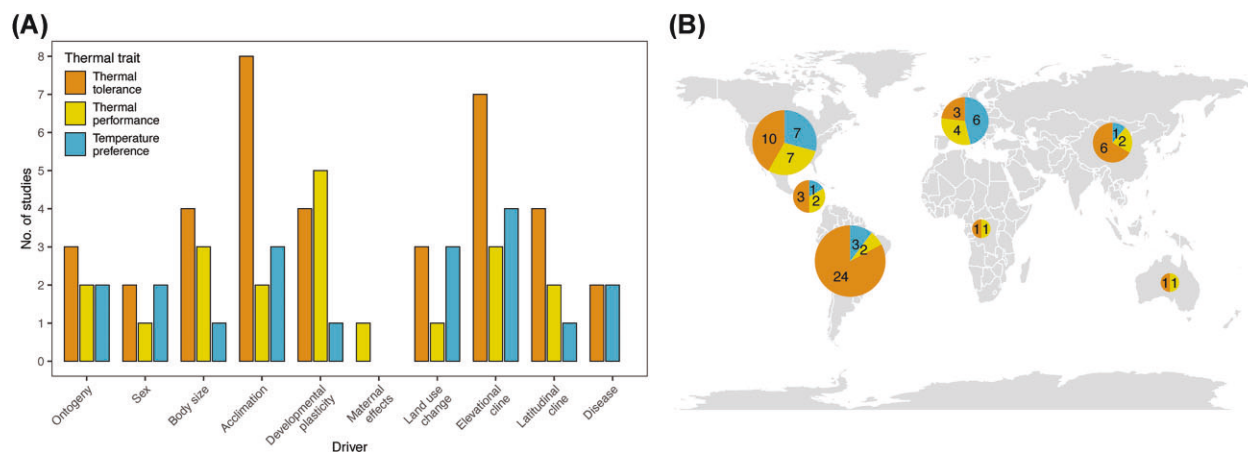
terrestrial juvenile, e.g., thermal adaptation hypothesis; Huey et al. 1999). For example, in Ruthsatz et al. (2022), the authors found that late-stage larvae had higher thermal tolerances than early stage larvae ( $CT_{max}$  increased 48%, from  $\sim 25^{\circ}\text{C}$  to  $\sim 37^{\circ}\text{C}$ ), but this declined slightly as amphibians reached metamorphic climax. This agrees with other work that has found that individuals near metamorphic climax have lower  $CT_{max}$  than preceding larval stages (Agudelo-Cantero and Navas 2019; for a review see Bodensteiner et al. 2020), and this period is the most energetically expensive in terms of oxygen consumption (Padilla et al. 2024). Furthermore, sessile eggs are unable to thermoregulate, and larvae confined to more thermally homogeneous aquatic environments may have fewer opportunities to do so, making them more vulnerable to changing conditions (Turriago et al. 2015). Interestingly, a recent meta-analysis found that  $CT_{max}$  in amphibians significantly differs between pre- and post-metamorphic life stages, with pre-metamorphic life stages often exhibiting a higher  $CT_{max}$  (e.g., Ruthsatz et al. 2024). This may be because it is advantageous for egg and aquatic larvae stages to exhibit higher thermal tolerances because they experience limited mobility and often have a smaller body size relative to adults (Ruthsatz et al. 2024).

### Sex differences

There may also be sex differences in thermal traits, in which males and females have divergent thermal optima, preferences, and/or critical temperatures. Both growth and development rates at different temperatures (Lambert et al. 2018), as well as the capacity of larvae to plastically respond to changing conditions (Brannelly et al. 2019), can vary between the sexes. A recent meta-analysis of sex differences in thermal acclimation capacity of ectotherms found only marginally greater heat and cold tolerance plasticity in females than males, which may be driven by sexual size dimorphism, but the overall pattern was weak (Pottier et al. 2021).

There is growing evidence that reproductive traits may covary with thermal physiology due to the reciprocal relationship between thermal ecology and mating system dynamics (Leith et al. 2022). For example, Rogers et al. (2007) found that male striped marsh frogs (*Limnodynastes peronii*) had greater acclimation capacity in metabolic enzymes in muscles associated with calling and amplexus than females, which may be linked to reproductive success. Conversely, other studies have found no sex differences in thermal traits in amphibians (Davies et al. 2015; Bókonyi et al. 2024). If sex differences in thermal traits exist, this may make one sex more vulnerable to heat stress than the other, which could impact reproductive success and the viability of future





**Fig. 2** Studies that examine intraspecific thermal trait variation in amphibians for micro- and macrogeographical drivers. **(A)** The number of studies since the year 2000 that examine within-species variation in thermal tolerance, thermal performance, and temperature preference for each driver discussed here. If a study examined more than one driver and/or thermal trait, it was included as an occurrence in each. **(B)** Global map showing where studies on intraspecific thermal trait variation have been performed since the year 2000. The location represents where the study species originates and was collected. The size of the pie chart correlates to the number of studies conducted on individual species in each geographic area, with the number of studies for each thermal trait also shown. This is species-level data and includes studies multiple times if they examined a thermal trait on multiple species in that geographical area. Studies included in these figures are found in [Supplementary Table S1](#).

populations ([Leith et al. 2022](#)). In a recent meta-analysis in invertebrates, sexual dimorphism in phenotypic plasticity of thermal traits was widespread, but there was a lack of clear systematic patterns (with the exception of female-biased plasticity in cold resistance; [Hangartner et al. 2022](#)). Overall, there is a paucity of studies on sex differences in thermal physiology in amphibians (see [Pottier et al. 2021](#); see Fig. 2A), and we encourage future work in this area.

### Body size

Body size is intricately linked with thermal physiology in wet-skinned ectotherms such as amphibians, which may be due to physiological constraints governing the relationship between surface area and body mass and corresponding impacts on both heating and water loss rates ([Tracy 1976](#)), or life-history optimization ([White and Marshall 2023](#)). Body size can be a strong driver of intra- and interspecific differences in critical thermal limits and thermal performance ([von May et al. 2019](#); [Ruthsatz et al. 2022](#); [Vidal et al. 2024](#)). This may be the result of inherent physiological differences in oxygen consumption and metabolism between large and small organisms (i.e., physiological differences), or that larger organisms warm or cool at a slower rate than smaller organisms, which can impact measured critical temperatures (i.e., physical differences; [Gunderson 2024](#)). Typically, within species, larger individuals demonstrate lower thermal tolerances, but experimental methods can confound large-scale trends, with smaller individ-

uals often being more sensitive to the duration of thermal assays that are used to measure tolerances ([Peralta-Maraver and Rezende 2021](#)). However, in amphibians, the opposite trend has also been found, in which larger individuals exhibit higher thermal limits, both within ([Barria and Bacigalupe 2017](#)) and across species ([von May et al. 2019](#)). Furthermore, thermally sensitive traits like metabolic rate typically scale hypoallometrically with body size ([White and Marshall 2023](#)), and this metabolic scaling can be difficult to disentangle from body size effects given that the rate of physiological processes increases with temperature, which can impact development and growth rate.

Temperature also directly affects body size, and there is mounting evidence of organisms decreasing in size in response to climate warming ([Sheridan and Bickford 2011](#); [Verberk et al. 2021](#)). In ectotherms, this may be a result of the temperature-size rule (TSR), in which organisms develop quicker and at smaller sizes under warmer conditions ([Atkinson 1994](#)), but the physiological and evolutionary mechanisms driving this rule are still debated ([Verberk et al. 2021](#)). Indeed, there are examples of population-level reductions in average body size in response to a warming climate in amphibians ([Reading 2007](#); [Caruso et al. 2014](#); [Gunderson 2024](#)), but this pattern may also be precipitation-dependent, given the importance of water for amphibian life history and physiology ([Sheridan et al. 2022](#)). When considering that morphological traits generally have higher heritabilities (i.e., the potential for a trait to be passed

on between generations and hence, the greater response to selection) than behavioral, physiological, or fitness traits (Fisher 1930; Mousseau and Roff 1987), intraspecific variation in body size and its correlation to thermal tolerance has potential to contribute to long-term population resilience to climate change.

While most work has focused on thermal limits, there are comparatively fewer studies that have investigated how ontogeny, sex differences, and body size impact other thermal traits. For example, previous work has found that thermal preference increases with developmental stage in multiple amphibian species, peaking before or near metamorphic climax and then declining rapidly (Floyd 1984; Dupré and Petranks 1985; Wollmuth et al. 1987; Wollmuth and Crawshaw 1988). Similar results were found in newts, with late-stage larvae selecting higher temperatures than early stage larvae (Smolinsk'y and Gvoždík 2009). In addition, body size can impact thermal preference, in which larger amphibians select higher temperatures (Ohmer et al. 2023; Kirsch et al. 2021; Percino-Daniel et al. 2021), or thermoregulate more precisely (Mitchell and Bergmann 2015). Finally, there are only a few studies that we are aware of that have investigated sex differences in thermoregulatory behavior. In newts, gravid females thermoregulate more accurately, but this did not impact egg size (Toufarová and Gvoždík 2016), while male midwife toads carrying eggs prefer higher temperatures than males not carrying eggs (Lange et al. 2022). Overall, ontogeny, sex differences, and body size variation can all lead to widespread intraspecific variation in thermal physiological traits, which may impact a population's responses to changing conditions.

### Phenotypic plasticity

Phenotypic plasticity may enable species with complex life cycles, like amphibians, to survive under fluctuating environments (Pottier et al. 2022; Ruthsatz et al. 2022). For example, plasticity in thermal tolerance—where an individual's thermal history can alter thermal limits—has been suggested as a strategy to mitigate the effects of extreme heat events (Ruthsatz et al. 2022; Rohr et al. 2018). Yet, it may be insufficient to buffer organisms from the effects of climate change on its own (Gunderson and Stillman 2015; Morley et al. 2019; Pottier et al. 2024).

Whether variation driven by plasticity is examined across a micro- or macrogeographical scale, we contend that it is important to consider the timescale of thermal history. An individual may exhibit a thermal phenotype that has the potential to fluctuate over a short time frame and is reversible (i.e., acclimation), is affected by their developmental environment and

is generally non-reversible (i.e., developmental plasticity), or is determined by their parents and grandparents and is a transgenerational effect (i.e., maternal effects). Whether short- and/or long-term thermal history is considered will determine the process behind plasticity and, hence, the potential for the plastic trait to adapt and contribute to long-term species resilience (Bodensteiner et al. 2020). Here, we discuss phenotypic plasticity at fine spatial scales but want to emphasize that plasticity (particularly acclimation; see the temporal dynamics section) can also drive intraspecific variation more broadly and these processes are ultimately occurring across all spatial and temporal scales.

### Acclimation on the microgeographic scale

Reversible acclimation in response to temperature changes is common in ectothermic organisms, including amphibians (Gunderson and Stillman 2015; Seebacher et al. 2015). Acclimation can buffer organisms from temperature variability on a short timescale, and greater plasticity in thermal traits may be adaptive under a changing climate. Acclimation responses in amphibians have been demonstrated for many traits, including critical thermal limits (Fan et al. 2021; Carilo Filho et al. 2022), thermal preferences (Hadamová and Gvoždík 2011; but see Fan et al. 2021 for example of no effect on thermal preference), and the thermal sensitivity of water loss rates (Riddell et al. 2018), locomotor performance (Padilla et al. 2019), and metabolism (Gomes et al. 2004; Rogers et al. 2007; Messerman and Leal 2020). In addition, plasticity in thermal traits can vary both between and within species (for a review and meta-analysis, see Seebacher et al. 2015). Acclimation to one stressor may also impact resistance to another; amphibians acclimated to high salinity environments have shown a decreased  $CT_{max}$ , indicating a possible trade-off between tolerance of high temperatures and tolerance of stressful environments (Chuang et al. 2022). This bears significance when assessing the impact of anthropogenic pollutants, like road salts and heavy metals, on amphibians as global average air temperatures continue to increase.

One measure of plasticity, the acclimation response ratio (ARR), has been used extensively to compare thermal plasticity between species (Gunderson and Stillman 2015; Seebacher et al. 2015; Morley et al. 2019), but should be considered a useful measure for understanding the level of intraspecific variation in thermal plasticity as well (Barria and Bacigalupe 2017; Enriquez-Urzelai et al. 2020; Ruthsatz et al. 2022). For example, a study on the European common frog found that late-stage larvae exhibited a higher acclimation capacity in

$CT_{max}$  than early stage larvae (ARR of  $CT_{max}$  increased almost two-fold, from 0.333 to 0.635; [Ruthsatz et al. 2022](#)).

### Developmental plasticity

The thermal environment experienced during development can impact the thermal traits of amphibians during subsequent life stages. In amphibians, there is evidence that plasticity in response to temperature during development can lead to shifts in critical temperatures ([Mueller et al. 2019](#); [Pottier et al. 2022](#); [Ruthsatz et al. 2022](#); [Ohmer et al. 2023](#); but see [Enriquez-Urzelai et al. 2019](#)), thermal preferences ([Ohmer et al. 2023](#)), and the thermal-sensitivity of performance at later life stages ([Drakulić et al. 2016](#); [Ohmer et al. 2023](#)). Developmental plasticity in response to a changing larval environment can result in better fitness in the resulting metamorphic environment ([Beaman et al. 2016](#)). Thus, elevated levels of plasticity may be adaptive, particularly if that environment changes rapidly (e.g., along an elevational gradient), or may be maladaptive, depending on whether early life environments are predictive of later-life environments ([Beaman et al. 2016](#); [Zhang et al. 2023](#)). In a recent meta-analysis across ectotherms, the evidence for strong effects of the developmental environment on critical temperatures in subsequent life stages was weak ([Pottier et al. 2022](#)). However, we need additional studies that investigate long-term impacts (beyond the recently hatched/metamorphosed), as well as additional thermal trait metrics (e.g., thermal preference; see [Fig. 2A](#)).

### Maternal effects

Maternal effects reflect a type of phenotypic plasticity determined by ancestors—where the offspring's phenotype is determined by the phenotype and environment experienced by the parent (or grandparent; [Falconer and Mackay 1996](#)). The effects of transgenerational plasticity on thermal physiology in amphibians have seldom been examined, and there remain large gaps in this research area ([Fig. 2A](#)).

What studies have found is that the thermal history of parents can have a significant effect on offspring fitness. For example, in the fire-bellied toad (*Bombina orientalis*), mothers that experienced greater environmental temperature variability reduced maternal investment that decreased offspring fitness and thermal performance of sprint speed ([Kaplan and Phillips 2006](#)). In reptiles, the thermal environment the mother experienced during pregnancy significantly shifted temperature preferences by  $\pm 1^\circ\text{C}$  ([Paranjpe et al. 2013](#)) and reduced anti-predatory behavior ([Lorioux et al. 2013](#)). Even the pregnancy stage at which various thermal environments are experienced (e.g., early or late em-

bryonic growth, etc.) can differentially affect offspring behavior—with offspring exhibiting less exploratory behavior when their mother was exposed to cold conditions during early development compared to late development ([Lorioux et al. 2013](#)). Interestingly, thermal maternal effects may even influence the resilience of offspring to disease. In *Daphnia*, research has shown that mothers that experience warmer temperatures produce offspring more resistant to infection and this was consistent across two populations ([Garbutt et al. 2014](#)).

The little research that has been conducted in amphibians, and the corresponding research in other ectotherms, points to evidence that maternal effects can have a significant influence on progeny thermal phenotypes. Crucially, transgenerational plasticity can introduce diversity in thermal traits within a population and offer an alternative inheritance mechanism for selection to act upon ([Pfennig and Martin 2009](#)). Thus, this type of plasticity may notably facilitate rapid adaptive evolution because maternal effects can be conferred between generations ([Agrawal et al. 1999](#); [Plaistow et al. 2006](#); [Allen et al. 2008](#)). When thinking about resilience to climate change and other global stressors (like disease), understanding the role of transgenerational plasticity in determining intraspecific thermal trait variation is fundamental, and yet remains a largely unexplored topic in amphibians ([Fig. 2A](#)).

## Macrogeographic sources of intraspecific variation

Within-species thermal biology historically received attention at the macrogeographical scale in insect and reptile species and focused on critical thermal limits (see [Chown 2001](#); [Hoffmann et al. 2003](#); and [Angilletta 2009](#) for reviews). Yet, studies examining if large-scale spatial variation can lead to differences between amphibian populations have become more common (for example, see [Pottier et al. 2022](#) and [DuBose et al. 2024](#) for published databases on amphibian thermal limits; [Supplementary Table S1](#)).

### Local adaptation

Local adaptation occurs if contemporary selective pressures are strong enough to overcome the rate of gene flow and results in individuals with higher fitness in the local environment when compared to individuals from other populations ([Richardson et al. 2014](#)). Preliminary work in amphibians has indicated that local adaptation in thermal traits may exist. For example, a study on temperature preference in wood frog tadpoles (*Rana sylvatica*) found evidence for



counter-gradient variation, where tadpoles from cooler environments (closed canopy pools) preferred warmer temperatures (Freidenburg and Skelly 2004). Conversely, a study on two tropical species found that tadpoles from populations that experienced warmer environments preferred hotter temperatures and exhibited a higher  $CT_{max}$  (Sanabria et al. 2021). Land-use changes may also drive intraspecific thermal trait variation in amphibians (see Nowakowski et al. 2018 for work on how habitat modification drives interspecific variation in  $CT_{max}$  in amphibians). For example, populations in converted open-forest habitats exhibited a higher thermal tolerance ( $CT_{max}$  that was 1.8°C higher; Frishkoff et al. 2015), thermal performance of jumping speed ( $T_{opt}$  that was 2.3°C higher; Frishkoff et al. 2015), and temperature preference ( $T_{pref}$  that was 2.0°C higher; Rivera-Ordóñez et al. 2019) than those in closed forests. Yet, this is not always consistent across thermal traits—Rivera-Ordóñez et al. (2019) also found that a shift in thermal preference did not indicate a correlated shift in thermal tolerance.

Importantly, while the studies discussed here and in the sections below use methods that suggest that local adaptation most likely occurred (for example, Freidenburg and Skelly 2004 collected eggs from different ponds and reared in a common environment to remove acclimation and developmental plasticity effects), we were not able to find studies on intraspecific thermal trait variation in amphibians that clearly tested whether differences were due to local adaptation or plasticity. Although we realize the difficulty of such studies for relatively long-lived organisms like amphibians, for local adaptation to be confirmed across populations, reciprocal transplant studies (an experiment where individuals from two or more environments are swapped and fitness is measured to assess relative fitness of each population in their local versus foreign environment) or common garden experiments with maternal effects removed (an experiment where amphibians would be collected from the wild, reared in a common environment for at least two generations to remove transgenerational effects, and then thermal traits are measured) must be performed. Additionally, it would be an oversight to not highlight that researchers should also consider the growing field of bioinformatics as a tool to detect genomic changes linked to local environments (Meek et al. 2023). However, genomic differences detected between populations may be due to non-adaptive or neutral evolution, and we urge that genetic techniques should be combined with experimental approaches to conclude if local adaptation has occurred. Hence, we contend that reciprocal transplant or common garden studies investigating intraspecific thermal adaptation in amphibians should be a research priority. This research would

help to disentangle the relative contributions of evolutionary change and plasticity on thermal trait variation, ultimately enhancing our understanding of the mechanisms that can contribute to resilience under global change.

### Elevational and latitudinal clines

Macrogeographical variation in environmental temperature can be caused by climate patterns that change with elevation or latitudinal gradients. Much of the current work at this scale tends to focus on investigating if tropical or temperate species exhibit greater thermal breadth (i.e., the temperature range they can survive within). This literature often references Janzen's hypothesis, which posits that temperate species that experience greater seasonal fluctuations should have broader thermal tolerances than tropical species that are exposed to less environmental variability (Janzen 1967). Although initially proposed to explain differences in thermal breadth between species, Janzen's hypothesis has been extended to examine intraspecific variation along elevational clines in tropical amphibians that exhibit broad distributional ranges (i.e., Pintanel et al. 2019; Bovo et al. 2023). Importantly, tropical species may exhibit similar, if not more, intraspecific thermal trait variation than temperate species because spatial complexity in tropical ecosystems may be more multifaceted than temporal variation of temperate climates (Janzen 1967; Llewelyn et al. 2016).

Elevational cline studies in amphibians have largely focused on intraspecific variation in thermal tolerances and thermal breadth, calculated as the range between  $CT_{max}$  and  $CT_{min}$  (e.g., Delgado-Suazo and Burrowes 2022; Bovo et al. 2023; Cicchino et al. 2023a; Cicchino et al. 2023b; reviewed in Carilo Filho et al. 2022). General trends are consistent with Brett's heat-invariant hypothesis (Brett 1956), which posits that  $CT_{max}$  should be more conserved than  $CT_{min}$  over elevation and across thermal environments (e.g., Pintanel et al. 2019; Bovo et al. 2023). For example, research on five tropical frog species in Brazil and five tropical frog species in Ecuador found that  $CT_{max}$  was more conserved than  $CT_{min}$  along an elevational gradient (Pintanel et al. 2019; Bovo et al. 2023; Páez-Vacas and Funk 2024). Yet, evidence also exists for the opposite pattern (where  $CT_{max}$  is more variable across elevational clines; e.g., Percino-Daniel et al. 2021; Delgado-Suazo and Burrowes 2022). Research on two different tropical frogs found that  $CT_{min}$  was more conserved (Percino-Daniel et al. 2021; Delgado-Suazo and Burrowes 2022), as did a study on tadpoles of two temperate frog species (Cicchino et al. 2023b). Potentially because variation in thermal tolerances seems to be context-dependent and species-specific (as do plastic changes on  $CT_{max}$  at different

elevations; see [Cicchino et al. 2023a](#)), most of the research on intraspecific variation in thermal breadth is not able to conclude if environmental patterns associated with elevation directionally alter thermal breadth. The studies that are available on variation within species provide important data when examining the multitude of hypotheses on thermal tolerance variability across elevation (i.e., Janzen's hypothesis and the climate variability hypothesis, Brett's heat-invariant hypothesis, the cold-variability hypothesis; see [Pintanel et al. 2019](#) and [Bovo et al. 2023](#) for descriptions and context).

Recent literature has also explored if elevational changes can drive intraspecific variation in temperature preference and thermal performance ([Enriquez-Urzelai et al. 2018](#); [Trochet et al. 2018](#); [Percino-Daniel et al. 2021](#); [Delgado-Suazo and Burrowes 2022](#)). Counter-gradient variation in temperature preferences (when individuals from cooler environments prefer warmer temperatures compared to individuals originating from warmer environments) was found in tropical frogs ([Percino-Daniel et al. 2021](#); [Delgado-Suazo and Burrowes 2022](#)), and in one of the few studies on intraspecific thermal trait variation in a salamander species ([Trochet et al. 2018](#)). Overall trends suggest that maximum thermal performance (such as jumping distance or locomotor performance) may show a slight increase with elevation ([Navas 1996](#); [Enriquez-Urzelai et al. 2018](#); [Delgado-Suazo and Burrowes 2022](#)). However, the optimal temperature ( $T_{opt}$ ) for thermal performance appears to be commonly conserved along elevational gradients (e.g., locomotor performance; [Navas 1996](#); [Enriquez-Urzelai et al. 2018](#); and jumping performance; [Delgado-Suazo and Burrowes 2022](#)).

Latitudinal studies on intraspecific thermal trait variation are less common in amphibians (Fig. 2A), but also suggest species-specific results. Opposite patterns were found for  $CT_{max}$  in temperate ([Kim et al. 2022](#)) and tropical frog species ([Sanabria et al. 2014](#); [Barria and Bacigalupe 2017](#)). Thermal preference,  $CT_{min}$ , and  $T_{opt}$  have been shown to increase in populations located at warmer latitudes ( $T_{opt}$ : [Wilson 2001](#),  $T_{pref}$  and  $CT_{min}$ : [Barria and Bacigalupe 2017](#)), and while there is some evidence that tropical species demonstrate lower levels of thermal trait plasticity than temperate species ([Feder 1978](#); [Feder 1982](#); [Ghalambor et al. 2006](#)), there is also evidence indicating a lack of variation in thermal trait plasticity across latitudinal clines (e.g., [Gunderson and Stillman 2015](#); [Riquelme et al. 2016](#); [Simon et al. 2015](#); [Morley et al. 2019](#)).

### Temporal dynamics

Disentangling temporal patterns across large-scale elevational and latitudinal clines is difficult and remains relatively unexplored in thermal biology ([Giacometti](#)

[et al. 2024](#)). This is particularly true when examining seasonality as a driver of intraspecific variation in amphibians. Seasonality may be more intense at higher elevations and latitudes, and this may drive variation in climate-relevant traits across this scale. High elevations and latitudes result in shorter seasonal activity windows (i.e., the annual timeframe a species is active for reproduction and growth; [Dahl et al. 2012](#); [Mikolajewski et al. 2015](#); [Giacometti et al. 2024](#)). Individuals may optimize physiology and behavior within their seasonal activity window to increase foraging and reproduction, potentially resulting in higher-risk behaviors with fitness trade-offs ([Giacometti et al. 2024](#)). Such phenological shifts are commonly documented in amphibians ([Todd et al. 2011](#)), and changes have exhibited a strength two to four times greater than responses in other taxonomic groups ([Parmesan 2006](#); [Parmesan 2007](#); [Urban et al. 2014](#)). This seasonality may exert strong plastic responses and selective pressures on populations, resulting in adaptation in thermal performance and temperature preference ([Enriquez-Urzelai et al. 2018](#)). Additionally, in amphibians that experience multiple seasons across their lives, thermal traits may vary annually when considering plasticity effects and may contribute to long-term resilience differentially depending on seasonal selective pressures. Investigating this will be crucial for enhancing our comprehension of how amphibians will respond to climate change ([Giacometti et al. 2024](#)), and these temporal changes need to be considered when predicting species resilience.

### Range-edge dynamics

Individuals that reside at the edge of a species range—or when thinking at a microgeographical scale, at the edge of a population—may experience different environmental conditions and constraints than those found at the center of a species range. Although individual fitness is generally thought to be lower at range edges ([Kawecki 2008](#)), environmental conditions that drive variation in thermal traits at margins can be just as advantageous, if not more ([Granado-Yela et al. 2013](#); [Valladares et al. 2014](#)). Thus, intraspecific variation formed in peripheral populations can contribute to resilience to climate change, particularly if conditions increase plasticity in thermal traits or contribute to directional selection in thermal traits that is favorable under fluctuating conditions and rising temperatures ([Rehm et al. 2015](#)). For example, peripheral populations located at lower elevations may experience greater environmental heterogeneity and warmer conditions than those at higher elevations, generating higher within-species thermal tolerances (as found in some amphibian populations: [Percino-Daniel et al. 2021](#); [Delgado-Suazo and Burrowes 2022](#); [Cicchino et al. 2023b](#)).

Indeed, marginal populations can play an important role in species persistence under climate change by harboring advantageous phenotypic plasticity (Chevin and Lande 2011) and thermal alleles, and may indicate at what range-edge (trailing versus leading edge) future distributional changes and/or adaptation may occur (Valladares et al. 2014). Differences in thermal preferences between individuals at trailing and leading edges can also be an indicator of dispersal responses to global change. For example, a study in reptiles found that individuals that preferred cooler temperatures dispersed further in warming climates than those that preferred warmer temperatures (most likely to “chase” their optimal environment; Bestion et al. 2015). Hence, recording thermal trait variation at range edges (and at peripheral populations) in amphibians and incorporating these into spatial distribution and mechanistic niche models can inform on how populations may track future climate change.

### Considering other abiotic and biotic drivers

It would be unreasonable to ignore the influence that abiotic factors other than temperature can have on driving intraspecific thermal trait variation in amphibians. For example, as wet-skinned organisms, variation in water availability and changes to relative humidity across spatial and temporal scales will influence local phenotypes (Greenberg and Palen 2021; Delgado-Suazo and Burrowes 2022). Additionally, recent studies have found that anthropogenic impacts can be detrimental to intraspecific variation (Stockwell et al. 2003; Miraldo et al. 2016). For example, habitat degradation and landscape connectivity can influence available microhabitats and affect gene flow that may alter thermal traits (Nowakowski et al. 2018). Artificial selection and human-induced regional extinctions can cause declines in genetic diversity (Des Roches et al. 2018). Humans also tend to change environments in ways that make them more uniform, which may lead to strong directional selection that depletes phenotypic and genetic variation.

Biotic factors will most certainly play a role in shaping the thermal biology of populations. Population dynamics, community composition that affect species interactions and food availability, invasive species, and factors like disease can all potentially drive variation in thermal traits. Notably, introduction of invasive species may deplete intraspecific variation simply by affecting population dynamics and potentially reducing abundance. Importantly, because disease is a significant threat to amphibians (Luedtke et al. 2023), we discuss

its implications on intraspecific thermal trait variation below.

### Disease

In addition to generating population-specific patterns, species-specific thermal biology may moderate the susceptibility of organisms to other global change stressors, like pathogenic infections (Deutsch et al. 2008; Kearney et al. 2009; Rohr and Raffel 2010; Huey et al. 2012; Navas et al. 2013; Sunday et al. 2014; Nowakowski et al. 2018). In amphibians, the fungal pathogen *Batrachochytrium dendrobatidis* (Bd) has caused hundreds of population declines and extinctions worldwide (Skerratt et al. 2007; Scheele et al. 2019), and as both host and pathogen are ectothermic, temperature is a key factor driving susceptibility to disease (Woodhams et al. 2008; Sauer et al. 2020; Cohen et al. 2017). While Bd grows optimally between 15 and 25°C (Piotrowski et al. 2004), there is increasing evidence that Bd can more readily overcome host defenses at cooler temperatures. There are patterns of amphibians declining and pathogen prevalence increasing at the cold end of species distributions, at higher elevations, and at closed canopy sites (Muths et al. 2008; Becker et al. 2012; Sonn et al. 2019; for a review see Haver et al. 2022), and many documented cases of seasonal infection prevalence, with infections peaking during winter/cooler months (Kriger and Hero 2007; Phillot et al. 2013; Wilber et al. 2022). In addition, realistic heat pulses (Greenspan et al. 2017b), access to warm refugia (Waddle et al. 2024), and reduced canopy cover (Roznik et al. 2015) can increase amphibian survival with Bd, but this may only be the case in warm-adapted species.

Just as pressing, and what has received little attention thus far, is understanding the impact of disease on intraspecific variation in climate-relevant thermal traits. Recent work has found that infection can change thermoregulatory behavior (Sherman 2008; Barrile et al. 2021) and restrict thermal tolerances in amphibians (Greenspan et al. 2017a; Siddons and Searle 2021)—potentially increasing risk of diseased populations to climate change by reducing the host’s ability to withstand thermal stress. Furthermore, amphibians that prefer warmer temperatures harbor lower Bd infection loads, which can reduce their risk of chytridiomycosis (Sauer et al. 2018). These changes may also result in different evolutionary trajectories of populations impacted by disease if environmental pressures drive selection on disease-determined phenotypical changes to thermal traits.

In some amphibian species, conservation efforts are underway to protect both naive populations and those that are persisting with Bd infections. These

populations may also represent the greatest chance of resilience to climate change if they harbor increased thermal tolerances and greater intraspecific thermal trait variation, and particularly if captive breeding programs reintroduce genetically diverse individuals with “hardy” thermal genotypes. Even with these endeavors, it is critical that we better understand population-level effects of disease on thermal biology and, conversely, how intraspecific variation in thermal traits may impact disease resilience. We encourage researchers to consider the importance of elucidating how different genotypes and/or pathogen strains interact with various populations of a given species. Until we can further clarify how disease and thermal biology interact with population-specific traits, we cannot fully comprehend the extent of how disease and climate change will interact and affect species resilience in a changing world.

### Expanding intraspecific thermal trait studies to assess resilience

Amphibians are considered one of the most threatened vertebrate groups (Luedtke et al. 2023) and understanding the factors underlying intraspecific thermal trait variation in amphibians and their role in enhancing species resilience to climate change is paramount for the conservation of these vulnerable taxa. By better understanding the drivers behind variation and collectively including them in predictions, we can more accurately plan and implement conservation measures. For example, examining local adaptation across elevational and latitudinal clines can help to isolate regional targets of selection that can help researchers identify the potential of a species to establish in new environments (Campbell-Staton et al. 2018), as well as the potential for evolution in the trait of interest.

A useful method to assess amphibian performance over space and time under different climate scenarios is mechanistic niche modeling (Kearney and Porter 2009; Briscoe et al. 2022; Riddell et al. 2023). We contend that researchers should use these models to include intraspecific thermal trait variation (since they are parameterized using physiological traits) across micro- and macrogeographical scales to assess resilience under global change. For example, incorporating acclimation responses into mechanistic niche models results in more optimistic predictions of amphibian performance under climate change (Riddell et al. 2018; Sinervo et al. 2024). We highlight the following specific areas as topics for future work to contribute to this goal:

- Future investigations should examine the potential implications of sex on amphibian thermal physiology and how it affects sex-biased mortality and population viability.
- Researchers interested in developmental plasticity should prioritize investigating the long-term impacts (beyond the recently hatched/metamorphosed life stage) of different developmental environments on thermal trait expression.
- Future work should focus on identifying how trans-generational plasticity affects progeny thermal phenotypes.
- When considering evolutionary potential and rescue as a means of resilience in amphibians, investigations into thermal trait heritability need to be conducted. Importantly, researchers should also consider how fluctuating environments across micro- and macrogeographical scales change heritability of thermal traits because heritability can be influenced by non-genetic environmental factors that make comparisons across populations, environments, and thermal traits invalid (Houle 1992).
- Reciprocal transplant or common garden studies investigating local adaptation in thermal traits should be a research priority and can help to clarify the respective roles of evolutionary change and plasticity on intraspecific thermal trait variation.
- While plasticity and evolutionary potential have been increasingly incorporated into niche models (Valladares et al. 2014; Duputié et al. 2015; Riddell et al. 2018; Riddell et al. 2023; Sinervo et al. 2024), there is a need to better understand and include the effects of seasonality and range-edge dynamics on within-species thermal trait variation in predictive models.
- Researchers should consider the importance of the interaction between a population and a particular genotype or strain of pathogen to fully comprehend how disease may impact intraspecific thermal trait variation and, subsequently, include the variability of these effects into resilience predictions.
- Lastly, we recognize that it is often unfeasible or impractical to measure all sources of intraspecific thermal trait variation for one species. While striving to gather such data remains a priority, in cases where it is unattainable, we contend that researchers should consider using data imputation to estimate thermal traits based on known relationships (for example, see Pottier et al. 2024) rather than omitting multivariate sources of intraspecific thermal trait variation.

### Conclusion

Environmental temperatures fluctuate both spatially and temporally, and discerning patterns within a species amidst these changes is crucial for predicting distributional changes and evolutionary capabilities of organisms responding to changing climates (Williams et al.



2008; Violle et al. 2012; Richter-Boix et al. 2015). Indeed, intraspecific thermal trait variation may be the key to determining a species' capacity for resilience to climate change (Llewellyn et al. 2016). Here, we aim to highlight the small- and large-scale drivers of intraspecific thermal trait variation in amphibians. In doing so, our goal is to underscore the significance of thoroughly realizing spatial and temporal variations in the thermal biology of a species, and to emphasize the importance of integrating these insights into mechanistic niche or species distribution models for a more comprehensive understanding of species resilience to global change.

## Author contributions

J.M.C. and M.E.B.O. conceptualized and wrote the manuscript, J.M.C. created the figures and tables, and both authors approved of the final version.

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## Supplementary data

Supplementary Data available at [ICB](#) online.

## Conflict of interest

The authors declare no conflict of interests.

## Data availability

No new data were generated or analysed in support of this research.

## References

- Agrawal AA, Laforsch C, Tollrian R. 1999. Transgenerational induction of defences in animals and plants. *Nature* 401:60–3.
- Agudelo-Cantero GA, Navas CA. 2019. Interactive effects of experimental heating rates, ontogeny and body mass on the upper thermal limits of anuran larvae. *J Therm Biol* 82:43–51.
- Albert CH, Thuiller W, Yoccoz NG, Douzet R, Aubert S, Lavorel S. 2010. A multi-trait approach reveals the structure and the relative importance of intra- vs. interspecific variability in plant traits. *Funct Ecol* 24:1192–201.
- Allen RM, Buckley YM, Marshall DJ. 2008. Offspring size plasticity in response to intraspecific competition: an adaptive maternal effect across life-history stages. *Am Nat* 171:225–37.
- Angilletta MJ, Jr. 2009. Thermal adaptation: a theoretical and empirical synthesis. Oxford: Oxford University Press.
- Angilletta MJ, Niewiarowski PH, Navas CA. 2002. The evolution of thermal physiology in ectotherms. *J Therm Biol* 27:249–68.
- Atkinson D. 1994. Temperature and organism size—a biological law for ectotherms? *Adv Ecol Res* 25:1–58.
- Barria AM, Bacigalupe LD. 2017. Intraspecific geographic variation in thermal limits and acclimatory capacity in a wide distributed endemic frog. *J Therm Biol* 69:254–60.
- Barrile GM, Chalfoun AD, Walters AW. 2021. Infection status as the basis for habitat choices in a wild amphibian. *Am Nat* 197:128–37.
- Beaman JE, White CR, Seebacher F. 2016. Evolution of plasticity: mechanistic link between development and reversible acclimation. *Trends Ecol Evol* 31:237–49.
- Becker CG, Rodriguez D, Longo AV, Talaba AL, Zamudio KR. 2012. Disease risk in temperate amphibian populations is higher at closed-canopy sites. *PLoS One* 7:e48205.
- Bennett JM, Sunday J, Calosi P, Villalobos F, Martínez B, Molina-Venegas R, Araújo MB, Algar AC, Clusella-Trullas S, Hawkins BA, et al. 2021. The evolution of critical thermal limits of life on Earth. *Nat Commun* 12:1198.
- Bestion E, Clobert J, Cote J. 2015. Dispersal response to climate change: scaling down to intraspecific variation. *Ecol Lett* 18:1226–33.
- Blanckenhorn WU. 1991. Life-history differences in adjacent water strider populations: phenotypic plasticity or heritable responses to stream temperature? *Evolution* 45:1520–5.
- Blanquart F, Kaltz O, Nuismer SL, Gandon S. 2013. A practical guide to measuring local adaptation. *Ecol Lett* 16:1195–205.
- Bodensteiner BL, Agudelo-Cantero GA, Arietta AZA, Gunderson AR, Muñoz MM, Refsnider JM, Gangloff EJ. 2020. Thermal adaptation revisited: how conserved are thermal traits of reptiles and amphibians? *J Exp Zool Pt A* 335:173–94.
- Bogert CM. 1949. Thermoregulation in reptiles, a factor in evolution. *Evolution* 3:195–211.
- Bókony V, Balogh E, Ujszegi J, Ujhegyi N, Szederkényi M, Hettyey A. 2024. Tadpoles develop elevated heat tolerance in urban heat islands regardless of sex. *Evol Biol* 51:209–16.
- Bovo RP, Simon MN, Provete DB, Lyra M, Navas CA, Andrade DV. 2023. Beyond Janzen's hypothesis: how amphibians that climb tropical mountains respond to climate variation. *Integr Org Biol* 5:obad009.
- Brannelly LA, Ohmer MEB, Saenz V, Richards-Zawacki CL. 2019. Effects of hydroperiod on growth, development, survival and immune defences in a temperate amphibian. *Funct Ecol* 33:1952–61.
- Brett JR. 1956. Some Principles in the Thermal Requirements of Fishes. *Q Rev Biol* 31:75–87.
- Briscoe NJ, Morris SD, Mathewson PD, Buckley LB, Jusup M, Levy O, Maclean IMD, Pincebourde S, Riddell EA, Roberts JA et al. 2023. Mechanistic forecasts of species responses to climate change: the promise of biophysical ecology. *Glob Chang Biol* 29:1451–70.
- Camacho A, Rodrigues MT, Navas C. 2015. Extreme operative temperatures are better descriptors of the thermal environment than mean temperature. *J Therm Biol* 49–50:106–11.
- Campbell-Staton SC, Bare A, Losos JB, Edwards SV, Cheviron ZA. 2018. Physiological and regulatory underpinnings of geo-

- graphic variation in reptilian cold tolerance across a latitudinal cline. *Mol Ecol* 27:2243–55.
- Carilo Filho LM, Gomes L, Katzenberger M, Solé M, Orrico VGD. 2022. There and back again: a meta-analytical approach on the influence of acclimation and altitude in the upper thermal tolerance of amphibians and reptiles. *Front Ecol Evol* 10, 1017255.
- Caruso NM, Sears MW, Adams DC, Lips KR. 2014. Widespread rapid reductions in body size of adult salamanders in response to climate change. *Glob Change Biol* 20:1751–9.
- Chevin L-M, Lande R. 2011. Adaptation to marginal habitats by evolution of increased phenotypic plasticity. *J Evol Biol* 24:1462–76.
- Chown SL. 2001. Physiological variation in insects: hierarchical levels and implications. *J Insect Physiol* 47:649–60.
- Chuang M-F, Cheng Y-J, Andersen D, Borzée A, Wu C-S, Chang Y-M, Yang Y-J, Jang Y, Kam Y-C. 2022. Increasing salinity stress decreases the thermal tolerance of amphibian tadpoles in coastal areas of. *Sci Rep* 12:9014.
- Cicchino AS, Shah AA, Forester BR, Dunham JB, Ghalambor CK, Funk WC. 2023b. Multi-scale relationships in thermal limits within and between two cold-water frog species uncover different trends in physiological vulnerability. *Freshw Biol* 68:1267–78.
- Cicchino AS, Shah AA, Forester BR, Dunham JB, Poff NL, Ghalambor CK, Funk WC. 2023a. Acclimation capacity of critical thermal maximum varies among populations: consequences for estimates of vulnerability. *Ecosphere* 14:e4691.
- Claunch NM, Goodman CM, Kluever BM, Barve N, Guralnick RP, Romagosa CM. 2023. Commonly collected thermal performance data can inform species distributions in a data-limited invader. *Sci Rep* 13, 15880.
- Clusella-Trullas S, Blackburn TM, Chown SL. 2011. Climatic predictors or temperature performance curve parameters in ectotherms imply complex responses to climate change. *Am Nat* 177:738–51.
- Cohen JM, Venesky MD, Sauer EL, Civitello DJ, McMahon TA, Roznik EA, Rohr JR. 2017. The thermal mismatch hypothesis explains host susceptibility to an emerging infectious disease. *Ecol Lett* 20:184–93.
- Dahl E, Orizaola G, Nicieza AG, Laurila A. 2012. Time constraints and flexibility of growth strategies: geographic variation in catch-up growth responses in amphibian larvae. *J Anim Ecol* 81:1233–43.
- Davies SJ, McGeoch MA, Clusella-Trullas S. 2015. Plasticity of thermal tolerance and metabolism but not water loss in an invasive reed frog. *Comp Biochem Physiol A Mol Integr Physiol* 189:11–20.
- Delgado-Suazo P, Burrowes PA. 2022. Response to thermal and hydric regimes point to differential inter- and intraspecific vulnerability of tropical amphibians to climate warming. *J Therm Biol* 103:103148.
- Des Roches S, David P, Turley N, Bailey J, Hendry A, Kinnison M, Schweitzer J, Palkovacs E. 2018. The ecological importance of intraspecific variation. *Nat Ecol Evol* 2:57–64.
- Deutsch CA, Tewksbury JJ, Huey RB, Sheldon KS, Ghalambor CK, Haak DC, Martin PR. 2008. Impacts of climate warming on terrestrial ectotherms across latitude. *Proc Natl Acad Sci USA* 105:6668–72.
- Diamond SE, Nichols LM, McCoy N, Hirsch C, Pelini SL, Sanders NJ, Ellison AM, Gotelli NJ, Dunn RR. 2012. A physiological trait-based approach to predicting the responses of species to experimental climate warming. *Ecology* 93:2305–12.
- Drakulić S, Feldhaar H, Lisičić D, Mioč M, Cizelj I, Seiler M, Spatz T, Rödel M-O. 2016. Population-specific effects of developmental temperature on body condition and jumping performance of a widespread European frog. *Ecol Evol* 6:3115–28.
- Dubose TP, Catalan V, Moore CE, Farallo VR, Benson AL, Dade JL, Hopkins WA, Mims MC. 2024. Thermal traits of Anurans database for the Southeastern United States (TRAD): a database of thermal trait values for 40 Anuran species. *Ichthyol Herpetol* 112:21–30.
- Dupré RK, Petranksa JW. 1985. Ontogeny of temperature selection in larval amphibians. *Copeia* 1985:462–7.
- Duputié A, Rutschmann A, Ronce O, Chuine I. 2015. Phenological plasticity will not help all species adapt to climate change. *Glob Chang Biol* 21:3062–73.
- Enriquez-Urzelai U, Palacio AS, Merino NM, Sacco M, Nicieza AG. 2018. Hindered and constrained: limited potential for thermal adaptation in post-metamorphic and adult *Rana temporaria* along elevational gradients. *J Evol Biol* 31:1852–62.
- Enriquez-Urzelai U, Sacco M, Palacio AS, Pintanel P, Tejedo M, Nicieza AG. 2019. Ontogenetic reduction in thermal tolerance is not alleviated by earlier developmental acclimation in *Rana temporaria*. *Oecologia* 189:385–94.
- Enriquez-Urzelai U, Tingley R, Kearney MR, Sacco M, Palacio AS, Tejedo M, Nicieza AG. 2020. The roles of acclimation and behaviour in buffering climate change impacts along elevational gradients. *J Anim Ecol* 89:1722–34.
- Falconer DS, Mackay TFC. 1996. Introduction to quantitative genetics. 4th ed. New York (NY): John Wiley & Sons, Inc.
- Fan XL, Lin ZH, Scheffers BR. 2021. Physiological, developmental, and behavioral plasticity in response to thermal acclimation. *J Therm Bio* 97:102866.
- Feder ME. 1978. Environmental variability and thermal acclimation in neotropical and temperate zone salamanders. *Physiol Zool* 51:7–16.
- Feder ME. 1982. Environmental variability and thermal acclimation of metabolism in tropical anurans. *J Therm Biol* 7:23–8.
- Fisher RA. 1930. The genetical theory of natural selection, the genetical theory of natural selection Oxford: Clarendon Press.
- Floyd RB. 1984. Variation in temperature preference with stage of development of *Bufo marinus* larvae. *J Herpetol* 18:153–8.
- Freidenburg KL, Skelly DK. 2004. Microgeographical variation in thermal preference by an amphibian. *Ecol Lett* 7:369–73.
- Frishkoff LO, Hadly EA, Daily GC. 2015. Thermal niche predicts tolerance to habitat conversion in tropical amphibians and reptiles. *Global Change Biol* 21:3901–16.
- Garbutt JS, Scholefield JA, Vale PF, Little TJ. 2014. Elevated maternal temperature enhances offspring disease resistance in *Daphnia magna*. *Funct Ecol* 28:424–31.
- Gerick AA, Munshaw RG, Palen WJ, Combes SA, O'Regan SM. 2014. Thermal physiology and species distribution models reveal climate vulnerability of temperate amphibians. *J Biogeogr* 41:713–23.
- Ghalambor CK, Huey RB, Martin PR, Tewksbury JJ, Wang G. 2006. Are mountain passes higher in the tropics? Janzen's hypothesis revisited. *Integr Comp Biol* 46:5–17.
- Giacometti D, Palaoro AV, Leal LC, de Barros FC. 2024. How seasonality influences the thermal biology of lizards with different thermoregulatory strategies: a meta-analysis. *Biol Rev* 99: 409–29.

- Gilbert AL, Miles DB. 2017. Natural selection on thermal preference, critical thermal maxima and locomotor performance. *Proc Biol Sci* 284:20170536.
- Gomes FR, Guilherme Chauí-Berlinck J, Bicudo JEPW, Navas CA. 2004. Intraspecific relationships between resting and activity metabolism in Anuran amphibians: influence of ecology and behavior. *Physiol Biochem Zool* 77:197–208.
- Goodwin KJA, Kissel AM, Palen WJ. 2019. Individual variation in thermal performance of a temperate, montane amphibian (*Rana cascadae*). *Herpetol Conserv Biol* 14:420–8.
- Granado-Yela C, Balaguer L, García-Verdugo C, Carrillo K, Méndez M. 2013. Thriving at the limit: differential reproductive performance in range-edge populations of a Mediterranean sclerophyll (*Olea europaea*). *Acta Oecologica* 52:29–37.
- Greenberg DA, Palen WJ. 2021. Hydrothermal physiology and climate vulnerability in amphibians. *Proc Biol Sci* 288:20202273.
- Greenspan SE, Bower DS, Roznik EA, Pike DA, Marantelli G, Alford RA, Schwarzkopf L, Scheffers BR. 2017a. Infection increases vulnerability to climate change via effects on host thermal tolerance. *Sci Rep* 7:9349.
- Greenspan SE, Bower DS, Webb RJ, Roznik EA, Stevenson LA, Berger L, Marantelli G, Pike DA, Schwarzkopf L, Alford RA. 2017. Realistic heat pulses protect frogs from disease under simulated rainforest frog thermal regimes. *Funct Ecol* 31:2274–86.
- Gunderson AR, Stillman JH. 2015. Plasticity in thermal tolerance has limited potential to buffer ectotherms from global warming. *Proc Biol Sci* 282:20150401.
- Gunderson AR. 2024. Disentangling physiological and physical explanations for body size-dependent thermal tolerance. *J Exp Biol* 227:jeb245645.
- Hadamová M, Gvoždík L. 2011. Seasonal acclimation of preferred body temperatures improves the opportunity for thermoregulation in newts. *Physiol Biochem Zool* 84:166–74.
- Hangartner S, Sgrò CM, Connallon T, Booksmythe I. 2022. Sexual dimorphism in phenotypic plasticity and persistence under environmental change: an extension of theory and meta-analysis of current data. *Ecol Lett* 25:1550–65. Available from: <https://doi.org/10.1111/ele.14005>.
- Haver M, Le Roux G, Friesen J, Loyau A, Vredenburg VT, Schmeller DS. 2022. The role of abiotic variables in an emerging global amphibian fungal disease in mountains. *Sci Total Environ* 815:152735.
- Herrando-Pérez S, Ferri-Yáñez F, Monasterio C, Beukema W, Gomes V, Belliure J, Chown SL, Vieites DR, Araújo MB. 2019. Intraspecific variation in lizard heat tolerance alters estimates of climate impact. *J Anim Ecol* 88:247–57.
- Hoffmann AA, Chown SL, Clusella-Trullas S. 2013. Upper thermal limits in terrestrial ectotherms: how constrained are they?. *Funct Ecol* 27:934–49.
- Hoffmann AA, Sørensen JG, Loeschcke V. 2003. Adaptation of *Drosophila* to temperature extremes: bringing together quantitative and molecular approaches. *J Therm Biol* 28:175–216.
- Houle D. 1992. Comparing evolvability and variability of quantitative traits. *Genetics* 130:195–204.
- Huey RB, Berrigan D, Gilchrist GW, Herron JC. 1999. Testing the adaptive significance of acclimation: a strong inference Approach. *Am Zool* 39:323–36.
- Huey RB, Deutsch CA, Tewksbury JJ, Vitt LJ, Hertz PE, Álvarez Pérez JH, Garland T. 2009. Why tropical forest lizards are vulnerable to climate warming. *Proc R Soc B* 276:1939–1948.
- Huey RB, Kearney MR, Krockenberger A, Holtum JAM, Jess M, Williams SE. 2012. Predicting organismal vulnerability to climate warming: roles of behaviour, physiology and adaptation. *Phil Trans R Soc B* 367:1665–79.
- Huey RB, Stevenson RD. 1979. Integrating thermal physiology and ecology of ectotherms: a discussion of approaches. *Integr Comp Biol* 19:357–66.
- Janzen DH. 1967. Why mountain passes are higher in the tropics. *Am Nat* 101:233–49.
- Kaplan RH, Phillips PC. 2006. Ecological and developmental context of natural selection: maternal effects and thermally induced plasticity in the from *Bombina orientalis*. *Evolution* 60:142–56.
- Kawecki T, Ebert D. 2004. Conceptual issues in local adaptation. *Ecol Lett* 7:1225–41.
- Kawecki TJ. 2008. Adaptation to marginal habitats. *Annu Rev Ecol Syst* 39:321–42.
- Kearney M, Porter WP. 2009. Mechanistic niche modelling: combining physiological and spatial data to predict species' ranges. *Ecol Lett* 12:334–50.
- Kearney M, Shine R, Porter WP. 2009. The potential for behavioral thermoregulation to buffer “cold-blooded” animals against climate warming. *Proc Natl Acad Sci USA* 106:3835–40.
- Kellermann V, Overgaard J, Hoffmann AA, Flojgaard C, Svenning JC, Loeschcke V. 2012. Upper thermal limits of *Drosophila* are linked to species distributions and strongly constrained phylogenetically. *Proc Natl Acad Sci USA* 109:16228–33.
- Khalique I, Shahid MJ, Kamran H, Sheraz M, Awais M, Shabir M, Asghar M, Rehman A, Riaz M, Braschler B, et al. 2023. The role of thermal tolerance in determining elevational distributions of four arthropod taxa in mountain ranges of southern Asia. *J Anim Ecol* 92:2052–66.
- Kim YI, Chuang M-F, Borzée A, Kwon S, Jang Y. 2022. Latitude-induced and behaviorally thermoregulated variations in upper thermal tolerance of two Anuran species. *Biology (Basel)* 11:1506.
- Kirsch DR, Fix S, Davenport JM, Cecala KK, Ennen JR. 2021. Body Size Is Related to Temperature Preference in *Hyla chrysoscelis* Tadpoles. *J Herpetol* 55:21–5.
- Kriger KM, Hero JM. 2007. Large-scale seasonal variation in the prevalence and severity of chytridiomycosis. *J Zool* 271:352–9.
- Lambert MR, Smylie MS, Roman AJ, Freidenburg LK, Skelly DK. 2018. Sexual and somatic development of wood frog tadpoles along a thermal gradient. *J Exp Zool Pt A* 329:72–9.
- Lange L, Brischoux F, Lourdaïs O. 2022. Benefits of paternal thermoregulation: male midwife toads select warmer temperature to shorten embryonic development. *Behav Ecol Sociobiol* 76:48.
- Leith NT, Fowler-Finn KD, Moore MP. 2022. Evolutionary interactions between thermal ecology and sexual selection. *Ecol Lett* 25:1919–36.
- Leiva FP, Calosi P, Verberk WC. 2019. Scaling of thermal tolerance with body mass and genome size in ectotherms: a comparison between water-and air-breathers. *Phil Trans R Soc B* 374:20190035.



- Llewelyn J, Macdonald SL, Hatcher A, Moritz C, Phillips BL. 2016. Intraspecific variation in climate-relevant traits in a tropical rainforest lizard. *Divers Distrib* 22:1000–12.
- Lorion S, Vaugoyeau M, Denardo D, Clobert J, Guillon M, Lourdaux O. 2013. Stage dependence of phenotypical and phenological maternal effects: insight into squamate reptile reproductive strategies. *Am Nat* 182:223–33.
- Luedtke JA, Chanson J, Neam K, Hobin L, Maciel AO, Catenazzi A, Borzée A, Hamidy A, Aowphol A, Jean A et al. 2023. Ongoing declines for the world's amphibians in the face of emerging threats. *Nature* 622:308–14.
- Meek MH, Beever EA, Barbosa S, Fitzpatrick SW, Fletcher NK, Mittan-Moreau CS, Reid BN, Campbell-Staton SC, Green NF, Hellmann JJ. 2023. Understanding local adaptation to prepare populations for climate change. *Bioscience* 73:36–47.
- Messerman AF, Leal M. 2020. Inter- and intraspecific variation in juvenile metabolism and water loss among five biphasic amphibian species. *Oecologia* 194:371–82.
- Mikolajewski D, De Block M, Stoks R. 2015. The interplay of adult and larval time constraints shapes species differences in larval life history. *Ecology* 96:1128–38.
- Miraldo A, Li S, Borregaard MK, Flórez-Rodríguez A, Gopalakrishnan S, Rizvanovic M, Wang Z, Rahbek C, Marske KA, Nogués-Bravo D. 2016. An Anthropocene map of genetic diversity. *Science* 353:1532–5.
- Mitchell A, Bergmann PJ. 2016. Thermal and moisture habitat preferences do not maximize jumping performance in frogs. *Funct Ecol* 30:733–42.
- Morley SA, Peck LS, Sunday JM, Heiser S, Bates AE. 2019. Physiological acclimation and persistence of ectothermic species under extreme heat events. *Global Ecol Biogeogr* 28:1018–37.
- Mousseau T, Roff D. 1987. Natural selection and the heritability of fitness components. *Heredity* 59:181–97.
- Mueller CA, Bucsky J, Korito L, Manzanares S. 2019. Immediate and persistent effects of temperature on oxygen consumption and thermal tolerance in embryos and larvae of the Baja California chorus frog, *Pseudacris hypochondriaca*. *Front Physiol* 10:754.
- Mueller CA. 2018. Critical windows in animal development: interactions between environment, phenotype, and time. In: Burggren W, Dubinsky B, eds. *Development and environment*. Cham, Switzerland: Springer International Publishing, p.41–72.
- Muñoz MM, Bodensteiner BL. 2019. Janzen's hypothesis meets the Bogert effect: connecting climate variation, thermoregulatory behavior, and rates of physiological evolution. *Integr Org Biol* 1:oby002.
- Muñoz MM, Stimola MA, Algar AC, Conover A, Rodriguez AJ, Landestoy MA, Bakken GS, Losos JB. 2014. Evolutionary stasis and lability in thermal physiology in a group of tropical lizards. *Proc Biol Sci* 281:20132433.
- Muths E, Pilliod DS, Livo LJ. 2008. Distribution and environmental limitations of an amphibian pathogen in the Rocky Mountains, USA. *Biol Conserv* 141:1484–92.
- Navas CA, Carvajalino-Fernández JM, Saboyá-Acosta LP, Rueda-Solano LA, Carvajalino-Fernández MA. 2013. The body temperature of active amphibians along a tropical elevation gradient: patterns of mean and variance and inference from environmental data. *Funct Ecol* 27:1145–54.
- Navas CA, Gouveia SF, Solano-Iguarán JJ, Vidal MA, Bacigalupe LD. 2021. Amphibian responses in experimental thermal gradients: concepts and limits for inference. *Comp Biochem Physiol B: Biochem Mol Biol* 254:110576.
- Navas CA. 1996. Metabolic physiology, locomotor performance, and thermal niche breadth in neotropical Anurans. *Physiol Zool* 69:1481–501.
- Nowakowski AJ, Watling JJ, Thompson ME, Brusch IVGA, Catenazzi A, Whitfield SM, Kurz DJ, Suárez-Mayorga Á, Aponte-Gutiérrez A, Donnelly MA et al. 2018. Thermal biology mediates responses of amphibians and reptiles to habitat modification. *Ecol Lett* 21:345–55.
- Ohmer MEB, Hammond TT, Switzer S, Wantman T, Bednark JG, Paciotta E, Coscia J, Richards-Zawacki CL. 2023. Developmental environment has lasting effects on amphibian post-metamorphic behavior and thermal physiology. *J Exp Biol* 226:jeb244883.
- Overgaard J, Kearney MR, Hoffmann AA. 2014. Sensitivity to thermal extremes in Australian *Drosophila* implies similar impacts of climate change on the distribution of widespread and tropical species. *Glob Change Biol* 20:1738–50.
- Padilla P, Ducret V, Bonneaud C, Courant J, Herrel A. 2019. Acclimation temperature effects on locomotor traits in adult aquatic anurans (*X. tropicalis* and *X. laevis*) from different latitudes: possible implications for climate change. *Conserv Physiol* 7:coz019.
- Padilla P, Herrel A, Denoël M. 2024. Invading new climates at what cost? Ontogenetic differences in the thermal dependence of metabolic rate in an invasive amphibian. *J Therm Biol* 121:103836.
- Páez-Vacas MI, Funk WC. 2024. Thermal limits along tropical elevational gradients: Poison frog tadpoles show plasticity but maintain divergence across elevation. *J Therm Bio* 120:103815.
- Paranjpe D, Bastiaans E, Patten A, Cooper R, Sinervo B. 2013. Evidence of maternal effects on temperature preference in side-blotched lizards: implications for evolutionary response to climate change. *Ecol Evol* 3:1977–91.
- Parnes C. 2006. Ecological and evolutionary responses to recent climate change. *Annu Rev Ecol Syst* 37:637–69.
- Parnes C. 2007. Influences of species, latitudes and methodologies on estimates of phenological response to global warming. *Glob Chang Biol* 13:1860–72.
- Peralta-Maraver I, Rezende EL. 2021. Heat tolerance in ectotherms scales predictably with body size. *Nat Clim Chang* 11:58–63.
- Percino-Daniel R, Contreras López JM, Téllez-Valdés O, Méndez de la Cruz FR, Gonzalez-Voyer A, Piñero D. 2021. Environmental heterogeneity shapes physiological traits in tropical direct-developing frogs. *Ecol Evol* 11:6688–702.
- Pfennig DW, Martin RA. 2009. A maternal effect mediates rapid population divergence and character displacement in spadefoot toads. *Evolution* 63:898–909.
- Phillips BL, Muñoz MM, Hatcher A, Macdonald SL, Llewelyn J, Lucy V, Moritz C. 2016. Heat hardening in a tropical lizard: geographic variation explained by the predictability and variance in environmental temperatures. *Funct Ecol* 30:1161–8.
- Phillott AD, Grogan LF, Cashins SD, McDonald KR, Berger LEE, Skerratt LF. 2013. Chytridiomycosis and seasonal mortality of tropical stream-associated frogs 15 years after introduction of *Batrachochytrium dendrobatidis*. *Conserv Biol* 27:1058–68.
- Pintanel P, Tejedo M, Ron SR, Llorente GA, Merino-Viteri A. 2019. Elevational and microclimatic drivers of thermal tolerance in Andean *Pristimantis* frogs. *J Biogeogr* 46:1664–75.



- Piotrowski JS, Annis SL, Longcore JE. 2004. Physiology of *Batrachochytrium dendrobatidis*, a chytrid pathogen of amphibians. *Mycologia* 96:9–15.
- Plaistow S, Lapsley C, Benton T. 2006. Context-dependent intergenerational effects: the interaction between past and present environments and its effect on population dynamics. *Am Nat* 167:206–15.
- Pörtner HO. 2002. Climate variations and the physiological basis of temperature dependent biogeography: systemic to molecular hierarchy of thermal tolerance in animals. *Comp Biochem Physiol A Mol Integr Physiol* 13:739–61.
- Pottier P, Burke S, Drobniak SM, Lagisz M, Nakagawa S. 2021. Sexual (in)equality? A meta-analysis of sex differences in thermal acclimation capacity across ectotherms. *Funct Ecol* 35:2663–78.
- Pottier P, Kearney MR, Wu NC, Gunderson AR, Rej JE, Rivera-Villanueva AN, Pollo P, Burke S, Drobniak SM, Nakagawa S. 2024. Vulnerability of amphibians to global warming. *Eco-EvoRxiv*. <https://ecoevovxiv.org/repository/view/6521/> (August 2024, date last accessed).
- Pottier P, Lin H-Y, Oh RRY, Pollo P, Rivera-Villanueva AN, Valdebenito JO, Yang Y, Amano T, Burke S, Drobniak SM et al. 2022. A comprehensive database of amphibian heat tolerance. *Sci Data* 9:600.
- Reading CJ. 2007. Linking global warming to amphibian declines through its effects on female body condition and survivorship. *Oecologia* 151:125–31.
- Rehm EM, Olivas P, Stroud J, Feeley KJ. 2015. Losing your edge: climate change and the conservation value of range-edge populations. *Ecol Evol* 5:4315–26.
- Richardson JL, Urban MC, Bolnick DI, Skelly DK. 2014. Microgeographic adaptation and the spatial scale of evolution. *Trends Ecol Evol* 29:165–76.
- Richter-Boix A, Katzenberger M, Duarte H, Quintela M, Tejedo M, Laurila A. 2015. Local divergence of thermal reaction norms among amphibian populations is affected by pond temperature variation. *Evolution* 69:2210–26.
- Riddell EA, Burger IJ, Tyner-Swanson TL, Biggerstaff J, Muñoz MM, Levy O, Porter CK. 2023. Parameterizing mechanistic niche models in biophysical ecology: a review of empirical approaches. *J Exp Biol* 226:jeb245543.
- Riddell EA, Odom JP, Damm JD, Sears MW. 2018. Plasticity reveals hidden resistance to extinction under climate change in the global hotspot of salamander diversity. *Sci Adv* 4:eaar5471.
- Riquelme NA, Díaz-Páez H, Ortiz JC. 2016. Thermal tolerance in the Andean toad *Rhinella spinulosa* (Anura: Bufonidae) at three sites located along a latitudinal gradient in Chile. *J Therm Biol* 60:237–245.
- Rivera-Ordóñez JM, Justin Nowakowski A, Manansala A, Thompson ME, Todd BD. 2019. Thermal niche variation among individuals of the poison frog, *Oophaga pumilio*, in forest and converted habitats. *Biotropica* 51:747–56.
- Rogers KD, Thompson MB, Seebacher F. 2007. Beneficial acclimation: sex specific thermal acclimation of metabolic capacity in the striped marsh frog (*Limnodynastes peronii*). *J Exp Biol* 210:2932–8.
- Rohr JR, Civitello DJ, Cohen JM, Roznik EA, Sinervo B, Dell AI. 2018. The complex drivers of thermal acclimation and breadth in ectotherms. *Ecol Lett* 21:1425–39.
- Rohr JR, Raffel TR. 2010. Linking global climate and temperature variability to widespread amphibian declines putatively caused by disease. *Proc Natl Acad Sci USA* 107:8269–74.
- Roznik E, Sapsford S, Pike DA, Schwarzkopf L, Alford RA. 2015. Natural disturbance reduces disease risk in endangered rainforest frog populations. *Sci Rep* 5:13472.
- Ruiz-Aravena M, Gonzalez-Mendez A, Estay SA, Gaitán-Espitia JD, Barria-Oyarzo I, Bartheld JL, Bacigalupe LD. 2014. Impact of global warming at the range margins: phenotypic plasticity and behavioral thermoregulation will buffer an endemic amphibian. *Ecol Evol* 4:4467–75.
- Ruthsatz K, Dahlke F, Alter K, Wohlrab S, Eterovick PC, Lyra ML, Gippner S, Cooke SJ, Peck MA. 2024. Acclimation capacity to global warming of amphibians and freshwater fishes: drivers, patterns, and data limitations. *Glob Chang Biol* 30:e17318.
- Ruthsatz K, Dausmann KH, Peck MA, Glos J. 2022. Thermal tolerance and acclimation capacity in the European common frog (*Rana temporaria*) change throughout ontogeny. *J Exp Zool Pt A* 337:477–90.
- Sanabria EA, González E, Quiroga LB, Tejedo M. 2021. Vulnerability to warming in a desert amphibian tadpole community: the role of interpopulational variation. *J Zool* 313:283–96.
- Sanabria EA, Vaira M, Quiroga LB, Akmentins MS, Pereyra LC. 2014. Variation of thermal parameters in two different color morphs of a diurnal poison toad, *Melanophryniscus rubriventris* (Anura: Bufonidae). *J Therm Biol* 41:1–5.
- Sauer EL, Cohen JM, Lajeunesse MJ, McMahon TA, Civitello DJ, Knutie SA, Nguyen K, Roznik EA, Sears BF, Bessler S et al. 2020. A meta-analysis reveals temperature, dose, life stage, and taxonomy influence host susceptibility to a fungal parasite. *Ecology* 101:e02979.
- Sauer EL, Fuller RC, Richards-Zawacki CL, Sonn J, Sperry JH, Rohr JR. 2018. Variation in individual temperature preferences, not behavioural fever, affects susceptibility to chytridiomycosis in amphibians. *Proc R Soc B* 285:20181111.
- Scheele BC, Pasmans F, Skerratt LF, Berger L, Martel A, Beukema W, Acevedo AA, Burrows PA, Carvalho T, Catenazzi A et al. 2019. Amphibian fungal panzootic causes catastrophic and ongoing loss of biodiversity. *Science* 363:1459–63.
- Seebacher F, White CR, Franklin CE. 2015. Physiological plasticity increases resilience of ectothermic animals to climate change. *Nat Clim Chang* 5:61–6.
- Sheridan JA, Bickford D. 2011. Shrinking body size as an ecological response to climate change. *Nat Clim Chang* 1:401–6.
- Sheridan JA, Mendenhall CD, Yambun P. 2022. Frog body size responses to precipitation shift from resource-driven to desiccation-resistant as temperatures warm. *Ecol Evol* 12:e9589.
- Sherman E. 2008. Thermal biology of newts (*Notophthalmus viridescens*) chronically infected with a naturally occurring pathogen. *J Therm Biol* 33:27–31.
- Siddons SR, Searle CL. 2021. Exposure to a fungal pathogen increases the critical thermal minimum of two frog species. *Ecol Evol* 11:9589–98.
- Simon MN, Ribeiro PL, Navas CA. 2015. Upper thermal tolerance plasticity in tropical amphibian species from contrasting habitats: implications for warming impact prediction. *J Therm Biol* 48:36–44.
- Sinclair BJ, Marshall KE, Sewell MA, Levesque DL, Willett CS, Slotsbo S, Dong Y, Harley CDG, Marshall DJ, Helmuth BS et al. 2016. Can we predict ectotherm responses to climate change

- using thermal performance curves and body temperatures? *Ecol Lett* 19:1372–85.
- Sinervo B**, Lara Reséndiz RA, Miles DB, Lovich JE, Rosen PC, Gadsden H, Gaytán GC, Tessaro PG, Luja VH, Huey RB et al. 2024. Climate change and collapsing thermal niches of desert reptiles and amphibians: assisted migration and acclimation rescue from extirpation. *Sci Total Environ* 908:168431.
- Skerratt LF**, Berger L, Speare R, Cashins S, McDonald KR, Phillott AD, Hines HB, Kenyon N. 2007. Spread of chytridiomycosis has caused the rapid global decline and extinction of frogs. *EcoHealth* 4:125–34.
- Smolinský R**, Gvoždík L. 2009. The ontogenetic shift in thermoregulatory behaviour of newt larvae: testing the ‘enemy-free temperatures’ hypothesis. *J Zool* 279:180–6.
- Sonn JM**, Utz RM, Richards-Zawacki CL. 2019. Effects of latitudinal, seasonal, and daily temperature variations on chytrid fungal infections in a North American frog. *Ecosphere* 10:e02892.
- Stockwell CA**, Hendry AP, Kinnison MT. 2003. Contemporary evolution meets conservation biology. *Trends Ecol Evol* 18:94–101.
- Sunday JM**, Bates AE, Dulvy NK. 2011. Global analysis of thermal tolerance and latitude in ectotherms. *Proc R Soc B* 278:1823–1830.
- Sunday JM**, Bates AE, Kearney MR, Colwell RK, Dulvy NK, Longino JT, Huey RB. 2014. Thermal-safety margins and the necessity of thermoregulatory behavior across latitude and elevation. *Proc Natl Acad Sci USA* 111:5610–5.
- Sunday JM**, Bennet JM, Calosi P, Clusella-Trullas S, Gravel S, Hargreaves AL, Leiva FP, Verberk WCEP, Olalla-Tárraga MA, Morales-Castilla I. 2019 Thermal tolerance patterns across latitude and elevation. *Phil. Trans. R. Soc. B* 374:20190036.
- Todd BD**, Scott DE, Pechmann JHK, Gibbons JW. 2011. Climate change correlates with rapid delays and advancements in reproductive timing in an amphibian community. *Proc Natl Acad Sci B* 278:2191–7.
- Toufarová E**, Gvoždík L. 2016. Do female newts modify thermoregulatory behavior to manipulate egg size? *J Therm Biol* 57:72–7.
- Tracy CR**. 1976. A model of the dynamic exchanges of water and energy between a terrestrial amphibian and its environment. *Ecol Monogr* 46:293–326.
- Trochet A**, Dupoué A, Souchet J, Bertrand R, Deluen M, Murarasu S, Calvez O, Martinez-Silvestre A, Verdaguer-Foz I, Darnet E et al. 2018. Variation of preferred body temperatures along an altitudinal gradient: a multi-species study. *J Therm Biol* 77:38–44.
- Turriago JL**, Parra CA, Bernal MH. 2015. Upper thermal tolerance in anuran embryos and tadpoles at constant and variable peak temperatures. *Can J Zool* 93:267–72.
- Urban MC**, Richardson JL, Freidenfelds NA. 2014. Plasticity and genetic adaptation mediate amphibian and reptile responses to climate change. *Evol Appl* 7:88–103.
- Valladares F**, Matesanz S, Guilhaumon F, Araújo MB, Balaguer L, Benito-Garzón M, Cornwell W, Gianoli E, van Kleunen M, Naya DE et al. 2014. The effects of phenotypic plasticity and local adaptation on forecasts of species range shifts under climate change. *Ecol Lett* 17:1351–64.
- Verberk WCEP**, Atkinson D, Hoefnagel KN, Hirst AG, Horne CR, Siepel H. 2021. Shrinking body sizes in response to warming: explanations for the temperature–size rule with special emphasis on the role of oxygen. *Biol Rev* 96:247–68.
- Vidal MA**, Rezende EL, Bacigalupe LD. 2024. Intra and interspecific variation in thermal performance and critical limits in anurans from southern Chile. *J Therm Bio* 121:103851.
- Violle C**, Enquist BJ, McGill BJ, Jiang L, Albert CH, Hulshof C, Jung V, Messier J. 2012. The return of the variance: intraspecific variability in community ecology. *Trends Ecol Evol* 27:244–52.
- von May R**, Catenazzi A, Santa-Cruz R, Gutierrez AS, Moritz C, Rabosky DL. 2019. Thermal physiological traits in tropical lowland amphibians: vulnerability to climate warming and cooling. *PLoS One* 14:e0219759.
- Waddle AW**, Clulow S, Aquilina A, Sauer EL, Kaiser SW, Miller C, Flegg JA, Campbell PT, Gallagher H, Dimovski I et al. 2024. Hotspot shelters stimulate frog resistance to chytridiomycosis. *Nature* 631:344–9.
- White CR**, Marshall DJ. 2023. How and why does metabolism scale with body mass? *Physiology* 38:266–74.
- Wilber MQ**, Ohmer ME, Altman KA, Brannnelly LA, LaBumbard BC, Le Sage EH et al. 2022. Once a reservoir, always a reservoir? Seasonality affects the pathogen maintenance potential of amphibian hosts. *Ecology* 103:e3759.
- Williams SE**, Shoo LP, Isaac JL, Hoffmann AA, Langham G 2008. Towards an integrated framework for assessing the vulnerability of species to climate change. *PLoS Biol* 6:e325.
- Wilson RS**. 2001. Geographic variation in thermal sensitivity of jumping performance in the frog *Limnodynastes peronii*. *J Exp Biol* 204:4227–4236.
- Wollmuth LP**, Crawshaw LI 1988. The effect of development and season on temperature selection in bullfrog tadpoles. *Physiol Zool* 61:461–9.
- Wollmuth LP**, Crawshaw LI, Forbes RB, Grahn DA 1987. Temperature selection during development in a montane anuran species. *Physiol Zool* 60:472–80.
- Woodhams DC**, Alford RA, Briggs CJ, Johnson M, Rollins-Smith LA 2008. Life-history trade-offs influence disease in changing climates: strategies of an amphibian pathogen. *Ecology* 89:1627–39.
- Zhang RY**, Wild KH, Pottier P, Carrasco MI, Nakagawa S, Noble DWA 2023. Developmental environments do not affect thermal physiological traits in reptiles: an experimental test and meta-analysis. *Biol Lett* 19:20230019.