

1 **Nemo knows: clownfishes differentiate cryptic host species across fine and broad**  
2 **geographic scales and reveal a potential adaptive radiation in the clownfish-hosting sea**  
3 **anemones.**

4

5 Tommaso Chiodo<sup>1,2</sup>, Aurélien De Jode<sup>1,2</sup>, Andrea Quattrini<sup>3</sup>, Miranda K. Gibson<sup>1,2</sup>, Catheline Y.  
6 M. Froehlich<sup>1,2</sup>, Danwei Huang<sup>4</sup>, Takuma Fujii<sup>5</sup>, Kensuke Yanagi<sup>6</sup>, James D. Reimer<sup>6,7,8</sup>, Anna  
7 Scott<sup>9</sup>, Estefanía Rodríguez<sup>10,†</sup>, Benjamin M. Titus<sup>1,2,†,\*</sup>

8

9 <sup>1</sup>Department of Biological Sciences, University of Alabama, Tuscaloosa, AL, USA 35487

10 <sup>2</sup>Dauphin Island Sea Lab, 101 Bienville Blvd., Dauphin Island, AL, USA 36528

11 <sup>3</sup>Department of Invertebrate Zoology, Smithsonian Institution's National Museum of Natural  
12 History, 10th and Constitution Ave NW, Washington, DC 20560, USA

13 <sup>4</sup>Lee Kong Chian Natural History Museum, National University of Singapore, Singapore 117377

14 <sup>5</sup>College of Bioresource Sciences, Nihon University, 1866 Kameino, Fujisawa, Kanagawa, Japan

15 <sup>6</sup>Coastal Branch of Natural History Museum and Institute, Chiba, Katsuura, Chiba, Japan

16 <sup>7</sup>Molecular Invertebrate Systematics and Ecology Laboratory, Department of Biology,  
17 Chemistry, and Marine Sciences, Faculty of Science, University of the Ryukyus, Nishihara,  
18 Okinawa, Japan

19 <sup>8</sup>Tropical Biosphere Research Center, University of the Ryukyus, Nishihara, Okinawa, Japan

20 <sup>9</sup>National Marine Science Centre, Faculty of Science and Engineering, Southern Cross  
21 University, PO Box 4321, Coffs Harbour, NSW 2450, Australia

22 <sup>10</sup>Division of Invertebrate Zoology, American Museum of Natural History, New York, NY, USA

23

24 <sup>†</sup>Denotes equal contributions and shared senior authorship

25 \*Corresponding author: bmtitus@ua.edu

26

27 **Abstract**

28 The symbiosis between clownfishes (or anemonefishes) and their host sea anemones  
29 ranks among the most recognizable animal interactions on the planet. Found on coral reef  
30 habitats across the Indian and Pacific Oceans, 28 recognized species of clownfishes adaptively  
31 radiated from a common ancestor to live obligately with only 10 nominal species of host sea  
32 anemones. Are the host sea anemones truly less diverse than clownfishes? Did the symbiosis  
33 with clownfishes trigger a reciprocal adaptive radiation in sea anemones, or minimally, a co-  
34 evolutionary response to the mutualism? To address these questions, we combined fine- and  
35 broad-scale biogeographic sampling with multiple independent genomic datasets for the bubble-  
36 tip sea anemone, *Entacmaea quadricolor*—the most common clownfish host anemone  
37 throughout the Indo-West Pacific. Fine-scale sampling and restriction site associated DNA  
38 sequencing (RADseq) throughout the Japanese Archipelago revealed three highly divergent  
39 cryptic species: two of which co-occur throughout the Ryukyu Islands and can be differentiated  
40 by the clownfish species they host. Remarkably, broader biogeographic sampling and bait-  
41 capture sequencing reveals that this pattern is not simply the result of local ecological processes  
42 unique to Japan, but part of a deeper evolutionary signal where some species of *E. quadricolor*  
43 serve as host to the generalist clownfish species *Amphiprion clarkii* and others serve as host to  
44 the specialist clownfish *A. frenatus*. In total, we delimit at least five cryptic species in *E.*  
45 *quadricolor* that have diversified within the last five million years. The rapid diversification of *E.*  
46 *quadricolor* combined with functional ecological and phenotypic differentiation supports the

47 hypothesis that this may represent an adaptive radiation in response to mutualism with  
48 clownfishes. Our data indicate that clownfishes are not merely settling in locally available hosts  
49 but recruiting to specialized host lineages with which they have co-evolved. These findings have  
50 important implications for understanding how the clownfish-sea anemone symbiosis has evolved  
51 and will shape future research agendas on this iconic model system.

52

### 53 **Introduction**

54 The importance of mutualism is underscored by its ubiquity- virtually all of life engages  
55 in complex multi-level mutualisms that critically impact the formation and distribution of  
56 biodiversity around the globe (Margulis and Fester 1991; Doebeli and Knowlton 1998; Herre et  
57 al., 1999; Moran 2007; Bronstein 2015). The degree to which mutualistic partners exchange, or  
58 provide, resources should lead to variation in the degree to which selection mediates the  
59 symbiosis over evolutionary timescales (Herre et al., 1999; Bronstein 2015). Ultimately, these  
60 processes manifest themselves at various levels of biological organization, impacting the species,  
61 population, and genomic diversity of the constituent partners (reviewed by Bronstein 2015).  
62 Mutualisms can trigger adaptive species radiations, lead to rapid population expansions, and  
63 radically shape the architecture of the genome (Herre et al., 1999; Moran 2007; Moya et al.,  
64 2008; Mueller et al 2011; Brucker and Bordenstein 2012; Joy 2013; Bronstein 2015; Rubin and  
65 Moreau 2016). Yet the role of mutualism in generating biodiversity within symbiotic systems  
66 can be difficult to discern, largely because of the various evolutionary, geographic, and  
67 ecological scales in which they operate (Herre et al., 1999).  
68 No mutualism is more representative of this puzzle than the iconic clownfish-sea  
69 anemone symbiosis, a model mutualism regularly used for exploring fundamental biological

70 processes (Ollerton et al 2007; Litsios et al., 2012; Marcionetti et al., 2019; Sahm et al. 2019;  
71 Roux et al. 2020, 2024; Klann et al. 2021; Laudet and Ravasi 2022; Marcionetti and Salamin  
72 2023 Moore et al. 2023), but one in which our understanding remains incomplete due to a lack of  
73 research into the evolution of the host sea anemones (De Jode et al., 2024; Titus et al., 2019a,  
74 2024). Broadly distributed on Indo-Pacific coral reefs, 28 described species of clownfishes form  
75 obligate mutualisms with just 10 species of sea anemones (Anthozoa: Actiniaria) (Fautin and  
76 Allen 1992; Titus et al., 2024). Mutualism with sea anemones is considered the key ecological  
77 innovation that triggered the adaptive radiation of clownfishes ~10-12 mya (Litsios et al., 2012),  
78 yet no evidence for host-driven patterns of diversification in clownfishes had been recovered in  
79 phylogenetic analyses until recently (De Jode et al., 2024; Gaboriau et al., 2024). Now, new  
80 evidence highlights that sea anemone host use can explain convergent phenotypic evolution in  
81 clownfish color patterns (Gaboriau et al., 2024), and further, that divergence times in the host sea  
82 anemones appear to be broadly coincident with the clownfish adaptive radiation (De Jode et al.,  
83 2024). The role of the host sea anemones in the evolution of the symbiosis is becoming  
84 increasingly established, yet key questions remain. Chief among these include: are the host sea  
85 anemones truly less diverse than the clownfishes? If not, given it is well established that the host  
86 anemones derive substantial benefits from hosting clownfishes, have the host sea anemones  
87 undergone their own hidden adaptive radiation?

88 One hypothesis is that the host sea anemones have undergone their own adaptive  
89 radiation, or are minimally, more diverse than currently recognized. Sea anemones have simple  
90 body plans, no hard parts, and few morphological characters that can reliably differentiate  
91 species (Titus et al. 2024). Undescribed cryptic species may thus be rampant within sea  
92 anemones (Titus et al. 2019a, b). Previous phylogenetic work using a suite of five traditional

93 Sanger loci could not resolve species level relationships for the two most specious host clades  
94 Stichodactylina and Heteractina but were able to partially resolve two nodes within the clade  
95 *Entacmaea* Titus et al. 2019a). The population-level signal present in *Entacmaea* when using  
96 traditional mitochondrial and nuclear markers is atypical across Anthozoa (Shearer et al., 2002;  
97 Huang et al. 2008; Daly et al. 2010; McFadden et al. 2011; Quattrini et al. 2023), which hints  
98 that this nominal species may be a species complex.

99 *Entacmaea* Ehrenberg, 1834 is a monotypic genus in the sea anemone superfamily  
100 Actinoidea. The sole species, *E. quadricolor* (Leuckart in Rüppell & Leuckart, 1828), is  
101 colloquially referred to as the bubble-tip sea anemone due to the characteristic bubbles/bulges  
102 that form at the tentacle tips (Titus et al. 2024). Of the host sea anemones, *E. quadricolor* has one  
103 of the largest biogeographic ranges, spanning from the Northern Red Sea, throughout the Indian  
104 Ocean, Coral Triangle, and into the Pacific (Titus et al. 2024). It has been documented to serve  
105 as a host to 20 of the 28 clownfish species, including both generalist and specialist clownfish  
106 species (Gaboriau et al., 2024). Where multiple clownfish species co-occur on the same reefs, *E.*  
107 *quadricolor* is regularly observed as hosts for multiple species (Fautin and Allen 1992).  
108 *Entacmaea quadricolor* also displays a great deal of intraspecific morphological variation in  
109 color, pattern, and tentacle shape (Titus et al. 2024). This has historically been attributed to  
110 phenotypic plasticity rather than species level differences (Fautin and Allen 1992; Richardson et  
111 al 1997; Titus et al. 2024). The degree of ecological and morphological variation within this  
112 single nominal species makes *E. quadricolor* an ideal candidate to test the degree to which the  
113 clownfish-hosting sea anemones are under described. The global importance of *E. quadricolor* as  
114 a host to >70% of all clownfish species further makes this putative species ideal for testing  
115 whether there are signatures of adaptive radiation within the host anemones.

116        Here we combine fine- and broad-scale biogeographic sampling of *E. quadricolor* with  
117    two independently derived genomic datasets to test for undescribed cryptic lineages and potential  
118    signatures of adaptive radiation linked to their mutualism with clownfishes. We couple our  
119    genomic datasets with an assessment of phenotypic diversity to tease apart any potential  
120    morphological differences among cryptic lineages. We discover that *E. quadricolor* is a diverse  
121    species complex that has rapidly diversified within the last five million years. Remarkably, we  
122    find co-occurring cryptic lineages of *E. quadricolor* not only host different species of  
123    clownfishes on the same reefs in the Japanese Archipelago, but are phenotypically different and  
124    derived from deeper evolutionary lineages linked to hosting specialist and generalist clownfish  
125    species. Our findings reveal far greater specialization and patterns of co-evolution in the  
126    clownfish-sea anemone symbiosis and the first potential evidence of a reciprocal adaptive  
127    radiation in the hosts. These findings will radically alter future research agendas within this  
128    iconic mutualism.

129

## 130    **Materials and Methods**

### 131    *Sample collection*

132        To search for patterns of cryptic species-level diversity within *Entacmaea quadricolor*,  
133    we first conducted fine-scale phylogeographic surveys and sampling across the Japanese  
134    Archipelago. We surveyed and photographed N = 126 individuals and collected N = 93 tentacle  
135    clippings from 38 sample localities, including the remote Ogasawara Islands ~1000 km from  
136    Mainland Japan (Figure 1; Table S1). Samples were collected by hand using SCUBA between 1-  
137    29 m depth. *In situ* photos were taken of each anemone, clownfish symbionts were identified and  
138    quantified, and one or two tentacles were sampled using forceps and placed into individually

139 labeled collection bags. On shore, samples were preserved in 95% ethanol. To place our  
140 Japanese samples into a broader biogeographic context, we also collected *E. quadricolor* samples  
141 from adjacent biogeographic regions including Singapore, Maldives, Australia, and the  
142 Philippines (Table S3). Samples were collected and preserved as above.

143

144 *DNA Extraction, Library Preparation, and Sequencing*

145 Following collection, total genomic DNA was extracted using a modified DNeasy Blood  
146 and Tissue Kits (QIAGEN Inc.) and prepared for DNA sequencing (see Supplementary  
147 Methods). To test for undescribed cryptic diversity within *E. quadricolor* and place the resulting  
148 diversity into broader phylogeographic context, we generated two independent genomic datasets.  
149 First, for all samples we collected throughout the Japanese Archipelago, we used a double-digest  
150 restriction-site associated DNA sequencing (ddRADseq) approach for cryptic species discovery  
151 (Titus et al. 2019b; Titus et al., 2022; see Supplementary Methods). Next, we used bait-capture  
152 sequencing targeting ultra-conserved elements (UCEs) and exon loci to produce a second  
153 genomic dataset and place the resulting cryptic lineages recovered from Japan into hierarchical  
154 biogeographic context. Accordingly, we tested whether the *E. quadricolor* lineages in Japan  
155 were diversifying *in situ* or were the result of geographical range overlap of more distantly  
156 related species. We re-sequenced N = 64 individuals from Japan along with N = 46 additional  
157 samples from adjacent biogeographic regions including Singapore (Chan et al. 2020),  
158 Philippines, Australia, and Maldives (Table S2). We used this alternative genomic approach as  
159 *E. quadricolor* does not have a closely related sister species that would be suitable to use as an  
160 outgroup taxon to root a ddRADseq dataset (Titus et al. 2019a; De Jode et al. 2024) and  
161 previously published bait-capture data for sea anemones are publicly available on GenBank

162 (Table S3) (Quattrini et al. 2018, 2020). Bait-capture libraries were prepared and sequenced  
163 following the protocol developed by Quattrini et al. (2018; see Supplementary Methods).

164

165 *Dataset assembly*

166 Following sequencing, raw ddRADseq data were demultiplexed, aligned, and assembled  
167 *de novo* using ipyrad (Eaton and Overcast 2020). We set the clustering threshold (Wclust) to  
168 0.90 to assemble reads into loci and sequencing depth (e.g. mindepth\_statistical and  
169 mindepth\_majrule) parameters were set to 10 to reduce heterozygous calls due to sequencing  
170 error. The min\_samples\_locus was set to 76 as that would represent approximately 75%  
171 occupancy across samples for the locus to be retained in the ddRADseq dataset. One SNP per  
172 locus was randomly selected to build an unlinked SNP dataset. Raw ddRADseq data were  
173 deposited at the NCBI Sequence Read Archive under BioProject XXXXX (Table S1).

174 To assemble UCE and exon loci from raw bait-capture sequence data we used the  
175 program PHYLUCE (Faircloth 2016) and the hexa\_v2\_final bait set (Cowman et al. 2020; Glon  
176 et al. 2020). After demultiplexing, raw sequences for each individual sample were cleaned and  
177 trimmed using illumiprocessor (Bolger et al. 2014; Faircloth 2016, 71). Cleaned sequences were  
178 then assembled into contigs using SPAdes v3.14.1 (Bankevich et al. 2012) with the -careful and -  
179 cov-cutoff 2 parameters. We then used PHYLUCE, as described in online tutorials to extract  
180 exon and UCE loci and assemble the final dataset. UCE baits were matched and then extracted  
181 using phyluce\_assembly\_match\_contigs\_probes and phyluce\_assembly\_get\_match\_counts. We  
182 created a final dataset with 75% completeness (i.e. no more than 25% missing data at a single  
183 locus). Our dataset was aligned using MAFFT (Katoh et al. 2002) and internally trimmed using  
184 the version of Gblocks inside PHYLUCE

185 (phyluce\_align\_get\_gblocks\_trimmed\_alignments\_from\_untrimmed) with the default  
186 parameters. For downstream bait-capture analyses, *Epiactis georgiana* Carlgren, 1927 was used  
187 as an outgroup species as this is currently the most closely related known species to *E.*  
188 *quadricolor* (De Jode et al. 2024).

189

190 *Fine scale phylogeography and species delimitation in the Japanese Archipelago*

191 To test whether there is evidence that *E. quadricolor* is a cryptic species complex within  
192 the Japanese Archipelago we used our fine-scale ddRADseq dataset to conduct three species  
193 discovery analyses. We first conducted a principal coordinates analysis (PCA) as a preliminary  
194 investigation to visualize the genetic variation present in the Japanese Archipelago in our  
195 unlinked SNP dataset generated by ipyrad (see Supplementary Methods). We next performed a  
196 Discriminant Analysis of Principal Components (DAPC) on the ddRADseq data to assign  
197 individuals of *E. quadricolor* from Japan to genetic clusters using the R package adegenet  
198 (Jombart 2008). The optimal value of  $k$  was obtained using a  $k$ -means clustering algorithm  
199 (find.clusters()) and the  $k$  value with the lowest BIC value was selected. To determine the  
200 number of principal components to retain for the DAPC, an  $\lambda$ -score optimization run was done  
201 and cross validated by selecting the number of PCs with the highest average prediction success  
202 per group. A separate DAPC analysis was conducted for  $k = 2-5$  to compare group assignments  
203 of different  $k$  values to our biological lineages. Finally, we used the Bayesian genetic clustering  
204 approach implemented in the program fastStructure (Raj et al. 2014) to delimit major genetic  
205 clusters and assign individuals to putative species. Using default parameters, we tested  $k$  values  
206 from 2 to 5 to determine number of genetic clusters that maximized the marginal likelihood for  
207 our dataset. All clustering programs identified the same three major genetic clusters, and we

208 computed pairwise  $F_{ST}$  to determine the degree of genetic differentiation within and between  
209 clusters using the R package hierfstat (Goudet 2005).

210

211 *Entacmaea quadricolor phenotypic diversity*

212 To determine whether our newly delimited species in the Japanese Archipelago displayed  
213 consistent phenotypic characteristics that could be used to visually identify each lineage apart  
214 from hosting different clownfishes, we phenotyped  $N = 126$  *E. quadricolor* individuals from  
215 field survey photographs. For each individual, we recorded phenotypic information for tentacle  
216 shape, tentacle length, tentacle tip pattern, tentacle color, tentacle tip color, anemone group size,  
217 depth, and finally, clownfish symbiont species. Tentacle shape variables were categorized as  
218 “rounded bubble tips,” “bubble with extended tip,” or digitiform (uniformly shaped/finger-like  
219 tentacles with no bubble tip swelling; Figure 2). Tentacle length was categorized as short ( $< 5$   
220 cm) or long ( $> 5$  cm). Tentacle tip pattern was categorized into Dull/Matte (D/M), striated (ST),  
221 or striated and speckled (STSP; Figure 2). Tentacle color was categorized into bleached, brown,  
222 pale brown, green/brown, and green. Tentacle tip color was categorized as whether the tentacle  
223 tips had pink tips (PT) or not (noPT). Anemone group size was categorically defined as whether  
224 the anemone was a solitary individual (S) or part of a clonal group (G). To account for important  
225 differences in habitat, we included depth as a variable. Depth was binned into shallow ( $< 5$ m) or  
226 deep ( $> 5$ m). We recorded whether the clownfish present in the anemone was *A. clarkii*, *A.*  
227 *frenatus*, or if no fish were present. We tested for significant associations between each  
228 phenotypic character and our three newly delimited species using multinomial logistic regression  
229 models. Analyses were conducted with the vglm function in R-Studio using packages tidyverse

230 (Wickham et al. 2019), VGAM (Yee 2010), car (Fox and Weisberg 2018), and rcompanion  
231 (Mangiafico 2016).

232

233 *Phylogenetic reconstruction*

234 To place newly delimited *E. quadricolor* species from Japan into hierarchical and  
235 biogeographic context, we conducted phylogenetic analyses using our bait-capture sequencing  
236 dataset targeting UCE and exon loci. Using our 75% dataset, we constructed Maximum  
237 Likelihood (ML) phylogenetic analyses using IQTree2 (Minh et al 2020). IQTree2 analyses were  
238 conducted using a concatenated and partitioned dataset with nodal support assessed using 10,000  
239 ultrafast bootstraps (Hoang et al. 2018) and SH-like approximate likelihood ratio tests (Guindon  
240 et al. 2010). Partitioned IQTree2 analyses were generated using the loci positions output by  
241 PHYLUCE to delimit partitions and ModelFinder to select the most appropriate substitution  
242 model (Chernomer et al. 2016; Kalyaanamoorthy et al. 2017).

243 Next, we used a coalescent-based approach to reconstruct phylogenetic relationships in  
244 ASTRAL III (Zhang et al. 2018). ASTRAL III accounts for incomplete lineage sorting by  
245 estimating a species tree from many independent gene trees. Input gene trees were generated  
246 using IQTree2 for each locus in the 75% occupancy matrix using the best fit model of evolution  
247 selected with ModelFinder (Kalyaanamoorthy et al. 2017). The resulting trees were used as an  
248 input for ASTRAL-hybrid v1.15.2.3 analyses. Nodal support in ASTRAL III analyses were  
249 assessed using posterior probabilities. Lastly, we reconstructed *E. quadricolor* relationships  
250 using CASTER (Zhang et al. 2023). CASTER represents another coalescent-based species-tree  
251 approach but uses a multiple sequence alignment as input instead of gene trees. CASTER

252 analyses were run with the concatenated 75% multi-sequence alignment and nodal support  
253 evaluated using posterior probabilities.

254 To estimate divergence times for newly delimited *E. quadricolor* species recovered from  
255 our phylogenomic analysis, we constructed a large UCE dataset by combining our *E.*  
256 *quadricolor* UCE dataset with previously published bait-capture sequence data from Order  
257 Actiniaria (Table S3), including samples from all major sea anemone superfamilies. We  
258 conducted a full analysis of all Actiniaria to capture the root of all sea anemones as the root age  
259 for Actiniaria had been previously estimated in a broader fossil calibrated phylogenomic analysis  
260 of Class Anthozoa (Quattrini et al. 2020; McFadden et al. 2021) A time calibrated dataset was  
261 assembled following the same modified PHYLUCE pipeline to generate a 75% occupancy  
262 matrix and IQTree2 was used to generate an unpartitioned ML tree as the unpartitioned tree  
263 retained higher support values for deeper relationships. Using IQTree2, divergence times were  
264 estimated by converting substitutions per site in the unpartitioned ML tree to years by setting the  
265 root age of Actiniaria to the previously estimated divergence date between 424 and 608 million  
266 years ago (Ma) (Quattrini et al. 2020; McFadden et al 2021). The tip date was set to 0 and the  
267 confidence interval to 1,000 years.

268

269 *Demographic modeling*

270 We conducted demographic modeling in *dadi* (diffusion approximations for demographic  
271 inference; Gutenkunst et al. 2009) to test competing diversification scenarios involving our  
272 newly delimited *E. quadricolor* from the Japanese Archipelago (*A. frenatus*-hosting, *A. clarkii*-  
273 hosting, and Ogasawara Islands). We tested whether ancestral diversification between *A. frenatus*  
274 and *A. clarkii* lineages occurred with or without geneflow, as well as tested whether co-occurring

275 lineages in Japan were reproductively isolated. We built a set of 17 demographic models (Figure  
276 4) following protocols and pipelines detailed by Portik et al. (2017). Each model was a three-  
277 population isolation-migration model that varied in the directionality and timing of gene flow  
278 between *E. quadricolor* species.

279 To model the evolutionary history of *E. quadricolor*, we built a three-dimensional Joint-  
280 folded Site Frequency Spectrum (JSFS) using our ddRADseq dataset from the Japanese  
281 Archipelago. Individual anemones were assigned to putative species based on species  
282 delimitation results. All loci not in Hardy-Weinberg Equilibrium ( $p > 0.05$ ) were removed using  
283 VCFTools (Danecek et al. 2011). Model simulations were conducted with consecutive rounds of  
284 optimization, where multiple replicates and previous parameter estimates from best scoring  
285 replicates were used to seed subsequent simulations. The default dadi\_pipeline settings were  
286 used for each round (replicates = 10, 20, 30, 40; maxiter = 3, 5, 10, 15; fold = 3, 2, 2, 1) and the  
287 parameter optimization followed the Nelder-Mead method (optimize\_log\_fmin). The optimized  
288 parameters of each replicate were used to simulate the 3D-JSFS and estimate the log-likelihood  
289 of the JSFS of the model. The best fit demographic model was selected using Akaike  
290 Information Criterion and model probabilities were calculated following Burnham and Anderson  
291 (2002).

292

## 293 **Results**

### 294 *Fine scale phylogeography and species delimitation in the Japanese Archipelago*

295 Fine-scale sampling and ddRADseq sequencing efforts resulted in 202 million raw  
296 sequence reads. After assembly and filtering in *ipyrad* our final dataset contained 3,516 unlinked  
297 loci across 82 individuals. All species discovery methods consistently recovered three distinct *E.*

298 *quadricolor* lineages (Figure 1). Principal Components Analysis (PCA), *k*-means clustering  
299 using discriminant analysis of principal components analysis (DAPC), and fastStructure all  
300 recovered two co-occurring *E. quadricolor* lineages in the Ryukyu Islands and a third allopatric  
301 lineage from the Ogasawara Islands (Figure 1; Figure S1 & S2).

302 Surprisingly, where the two putative *E. quadricolor* lineages co-occurred throughout the  
303 Ryukyu Islands they hosted different species of clownfishes with near 100% perfect habitat  
304 segregation. Our most heavily sampled *E. quadricolor* lineage harbored the host generalist  
305 Clark's clownfish *Amphiprion clarkii*. This species occurred from the Ryukyu Islands in  
306 southern Japan north into Mainland Japan. Our less frequently sampled *E. quadricolor* lineage  
307 harbored the host specialist Tomato clownfish *A. frenatus*, which is known to exclusively  
308 associate with *E. quadricolor* (Figure 1; Figure S1 & S2). We only sampled the *A. frenatus*  
309 hosting lineage in the Ryukyu Islands. In our most northern sampling localities from Mainland  
310 Japan, we collected some *E. quadricolor* individuals from shallow habitats (<1m depth) that did  
311 not host clownfishes. These clustered with the putative *A. clarkii* hosting lineage (Figure 1). In  
312 Ogasawara, our third lineage also served as host to *A. clarkii* although the Ogasawaran  
313 clownfishes exhibit a unique all-black phenotype and there has been speculation these represent  
314 a cryptic species within the *A. clarkii* complex. Genetic divergence between putative *E.*  
315 *quadricolor* lineages was high (Table 1). Pairwise Fst values were higher between co-occurring  
316 *E. quadricolor* lineages that hosted different clownfishes than between *E. quadricolor* lineages  
317 that hosted *A. clarkii* clownfishes but where allopatrically distributed between Mainland Japan  
318 and Ogasawara Islands (Table 1).

319

320 *Entacmaea quadricolor phenotypic diversity*

321 Phenotypic analyses revealed that tentacle shape, tentacle tip pattern, tentacle tip color,  
322 anemone group size, and tentacle color were all predictive of the three cryptic host species  
323 (Figure 2). In general, *E. quadricolor* anemones that hosted the Tomato clownfish *A. frenatus*  
324 had bubble-tip tentacle shapes with blunt-rounded tips and a dull-matte tentacle tip pattern  
325 (Figure 2a, f). These could be differentiated from co-occurring *E. quadricolor* anemones that  
326 hosted *A. clarkii*, which had bubble-tip tentacle shapes with a protruding/elongated tentacle tip  
327 extending from the bulbous swelling and had striated (i.e. striped) tentacle tip patterns with pink  
328 tips (Figure 2b, f). Ogasawaran *E. quadricolor* anemones had similar phenotypes as the *A. clarkii*  
329 hosting anemones in Mainland Japan with the addition of regularly having a speckled tentacle tip  
330 pattern and a pale-brown coloration (Figure 2c, f). The *A. frenatus* hosting *E. quadricolor* were  
331 also significantly more likely to be found in groups/aggregations of anemones and were less  
332 likely to have pink tipped tentacles, in contrast to both *A. clarkii* and Ogasawaran anemones,  
333 which were more frequently found solitarily and regularly had pink tentacle tips (Figure 2).

334

### 335 *Phylogenetic Reconstruction*

336 Bait-capture sequencing and dataset assembly resulted in an average of  $56,630 \pm 48,647$   
337 contigs per sample with a mean locus length of  $329 \pm 85$  basepairs across  $N = 103$  individuals  
338 from Japan, Singapore, Australia, Maldives, and the Philippines (Table S3). The total average  
339 base pairs per sample was  $18,603,577 \pm 12,788,845$ . After alignment and edge trimming, we  
340 recovered 1,542 UCE and exon loci. We assembled a final dataset requiring at least 75%  
341 completeness per locus, resulting in 1002 retained loci and 88,410 parsimoniously informative  
342 sites.

343                   Partitioned maximum likelihood phylogenetic analyses in IQtree2 resulted in a highly  
344 supported tree topology (Figure 3). We recovered the *E. quadricolor* lineage from the Ryukyu  
345 Islands and Mainland Japan that hosted *A. clarkii* as sister to the *A. clarkii* hosting *E. quadricolor*  
346 lineage from Ogasawara (Figure 3). Interestingly, we recovered *E. quadricolor* individuals from  
347 the Maldives, which also hosted *A. clarkii*, as sister to both Japanese *E. quadricolor* lineages that  
348 hosted *A. clarkii*. Additionally, we recovered the *E. quadricolor* lineage from the Ryukyu Islands  
349 that hosts *A. frenatus* to form a sister relationship with *E. quadricolor* from Singapore, which  
350 also hosts *A. frenatus*. Thus, the *E. quadricolor* species complex in our analyses formed two  
351 deep evolutionary clades associated with hosting generalist (*A. clarkii*) and specialist (*A.*  
352 *frenatus*) clownfishes (Figure 3). IQtree2 phylogenetic analyses placed all samples from  
353 Australia in a deeper monophyletic clade sister to all other samples (Figure 3). Individual *E.*  
354 *quadricolor* anemones from the Solitary Islands (Australia) hosted the Great Barrier Reef  
355 clownfish *A. akindynos*. The clownfish identity from the *E. quadricolor* anemones collected  
356 from the Great Barrier Reef were unknown as these were provided by Cairns Marine Inc. in the  
357 aquarium trade. In total, IQtree2 phylogenetic analyses recovered seven highly supported  
358 monophyletic *E. quadricolor* lineages that correspond to either clownfish host use or geography:  
359 1) Ryukyu Islands and Mainland Japan – *A. clarkii*, 2) Ogasawara Islands – *A. clarkii*, 3)  
360 Maldives – *A. clarkii*, 4) Ryukyu Islands Japan – *A. frenatus*, 5) Singapore – *A. frenatus*, 6)  
361 Great Barrier Reef Australia, 7) Solitary Islands Australia (Figure 3).

362                   Coalescent-based phylogenetic analyses in ASTRAL-III and CASTER recovered similar  
363 *E. quadricolor* lineages and tree topologies as IQtree2 but with some differences (Figure S3 &  
364 S4). Importantly, all analyses recovered the same three cryptic Japanese *E. quadricolor* lineages  
365 and demonstrated that co-occurring Japanese diversity was not the result of *in situ* endemic

366 diversification (Figure S3 & S4). Some ambiguity existed in the placement of the *E. quadricolor*  
367 lineage from the Maldives. In ASTRAL-III, the Maldives were placed as sister to Australia,  
368 Singapore, and the *A. frenatus* hosting *E. quadricolor* from Japan, rather than to the *A. clarkii*  
369 hosting anemones from Japan and the Ogasawaran Islands (Figure S3). Deep nodes, however,  
370 were less well supported in ASTRAL-III analyses than from IQtree2. CASTER phylogenetic  
371 analyses placed the Maldives back as sister to both *A. clarkii* hosting lineages from Japan with  
372 full support (Figure S4).

373 To estimate divergence times for newly delimited *E. quadricolor* species, we  
374 incorporated an additional  $N = 84$  previously sequenced sea anemone samples to capture the root  
375 of Order Actiniaria. We recovered 2496 UCE and exon loci with 185,085 parsimoniously  
376 informative sites. Using the previously estimated 95% confidence intervals for the root age for  
377 Order Actiniaria (424-608 Ma) our time calibrated analysis in IQtree2 dated divergence times for  
378 all members of the *E. quadricolor* species complex to be between 5.7 and 3 Ma (Figure S5).

379

380 *Demographic Modeling*

381 Among 17 unique demographic models, Akaike Information Criterion (AIC) model  
382 selection placed all model support on a three-population isolation-migration model (Figure 4,  
383 Table S3). The model included divergence with ancestral, bi-directional, symmetrical, migration  
384 at time 1 (T1), and divergence with contemporary unidirectional migration from Ryukyu  
385 Islands/Mainland *A. clarkii* hosting *E. quadricolor* to Ogasawaran *E. quadricolor* at T2 (Figure  
386 4; Table S3). Our best fit model did not include a gene flow parameter between co-occurring *E.*  
387 *quadricolor* anemones that host different clownfishes on reefs in the Ryukyu Islands during T2.  
388 No other simulated demographic model garnered appreciable model support (Table S3).

389

390 **Discussion**

391 Our results unequivocally identify the bubble-tip sea anemone *Entacmaea quadricolor* as  
392 a diverse species complex shaped by both its mutualism with clownfishes and geographic  
393 distribution. Additionally, our data support the hypothesis that *E. quadricolor* may have  
394 undergone its own adaptive radiation. Our discovery that clownfishes can differentiate and  
395 ecologically segregate by co-occurring cryptic *E. quadricolor* lineages in Japan, and that there is  
396 an evolutionary basis for clownfish host specialist and generalist species of *E. quadricolor* on  
397 broader biogeographic scales, points to the importance of co-evolutionary processes generating  
398 and maintaining biodiversity in both the clownfishes and host sea anemones.

399

400 *Species delimitation and phylogeography of the Entacmaea quadricolor species complex*

401 With the aid of independent genomic datasets we recover seven monophyletic lineages.  
402 We choose to conservatively delimit five cryptic species of *E. quadricolor*: I) Ogasawara Islands  
403 II) Ryukyu Islands and Mainland Japan (*A. clarkii*-hosting), III) Maldives, IV) Ryukyu Islands  
404 Japan + Singapore (*A. frenatus*-hosting), and V) Australia (Figure 3). For consistency and with  
405 future research in mind we elect to designate newly delimited species using Roman numerals (I-  
406 V; Figure 3). If additional evidence emerges that justifies further splitting our species here, a  
407 lowercase letter can be added to create a consistent alphanumeric system to informally recognize  
408 cryptic *E. quadricolor* species (e.g. Ia, IIa) until a formal revision can be completed. We  
409 currently elect not to further delimit *E. quadricolor* sp. IV (*A. frenatus*-hosting anemones from  
410 Japan and Singapore) or *E. quadricolor* sp. V (Great Barrier Reef and Solitary Islands –  
411 Australia) although we recovered these sample localities as monophyletic. Genetic divergences

412 are not as deep as other splits in our phylogenetic reconstruction, and we do not have other  
413 additional lines of evidence to differentiate these groups (e.g. morphology/ecology), and so we  
414 elect not to delimit these as species for now (Figure 3).

415 The bubble-tip sea anemone has long been hypothesized to be a species complex, due  
416 largely to the highly variable phenotypic diversity encountered throughout its range (e.g. Titus et  
417 al. 2024). Traditional PCR-based sequencing hinted at the prospect of multiple species being  
418 present, even within Japan (Titus et al. 2019a) but the poorly resolving Sanger-loci were  
419 inconclusive. More recently, genomic approaches recovered similar genetic clusters within the  
420 Japanese Archipelago to ours here, yet lacked the biogeographic sampling needed to put this  
421 diversity into species-level context (Kashimoto et al. 2023). Our combined ddRADseq and UCE  
422 approach conclusively delimit three *E. quadricolor* species present within the Japanese  
423 Archipelago. We reveal that sympatric *E. quadricolor* diversity from the Ryukyu Islands are the  
424 result of geographic range overlap between two deeply diverged lineages (*E. quadricolor* sp. II  
425 and IV) that have evolved to host different clownfishes, rather than endemic *in situ*  
426 diversification within Japan. Subsequent allopatric speciation explains divergence between *E.*  
427 *quadricolor* sp. II (Ryukyu Islands and Mainland Japan) and *E. quadricolor* sp. I (Ogasawara  
428 Islands) that host *A. clarkii*. We make this interpretation as the *A. frenatus*-hosting *E.*  
429 *quadricolor* sp. IV from the Ryukyu Islands Japan is more closely related to individuals from  
430 Singapore than it is to the co-occurring *E. quadricolor* sp. II that hosts *A. clarkii*.

431 Demographic model selection in *dadi* demonstrates that ancestral divergence between the  
432 *A. frenatus* and *A. clarkii*-hosting species proceeded with bi-directional symmetric migration in  
433 T1. In T2, however, co-occurring *E. quadricolor* sp. II and IV lineages that host different species  
434 of clownfishes exist in complete genetic isolation with no signatures of contemporary

435 migration—reinforcing species delimitation results that these are true biological species. Our  
436 best fit model also indicates that *E. quadricolor* sp. I from Ogasawara Islands diverged from *E.*  
437 *quadricolor* sp. II (Ryukyu Island and Mainland Japan) with unidirectional gene flow to  
438 Ogasawara. The Ogasawara Islands are located >1,000 km from Mainland Japan and separated  
439 by the Kuroshio Current, which is a deep fast-moving ocean current that flows north. Spin-off  
440 eddies, rotating clockwise and travelling east could potentially provide periodic larval transport  
441 to Ogasawara from Mainland Japan and explain the directionality of gene flow during  
442 diversification. The Japanese Archipelago has long been recognized as an important marine  
443 biogeographic region as it sits just north of the Coral Triangle where the Pacific and Indian  
444 Oceans meet (Motomura et al. 2007; Bowen et al. 2016; Endo et al. 2022). Our comprehensive  
445 fine-scale sampling and species delimitation analyses reinforces Japan as a marine biodiversity  
446 hotspot and disentangles the complex evolutionary and biogeographic processes that can  
447 contribute to generating biodiversity within this region (e.g. Bowen et al. 2016; Reimer et al.  
448 2019).

449 Beyond Japan, hierarchical phylogenetic relationships among our newly delimited *E.*  
450 *quadricolor* species complex reveals the importance of broad-scale biogeographic processes that  
451 shapes diversity within this group. Strong geographic signal is evident within our bait-capture  
452 sequence data, and we recover most (but not all) sample localities as monophyletic. Even at the  
453 intraspecific levels we see signatures that demonstrate that the geographic scale of dispersal may  
454 be small for *E. quadricolor*. Previous work on *E. quadricolor* larval biology indicates planula  
455 larvae can metamorphose and settle as early as five days post fertilization, with most larvae  
456 settling within 10 days (Scott and Harrison 2008). Short larval durations could explain patterns  
457 in our data.

458                   We recover intraspecific phylogeographic lineages within the *A. frenatus*-hosting *E.*  
459    *quadricolor* sp. IV, separating the Ryukyu Islands Japan from Singapore, and *E. quadricolor* sp.  
460    V, separating tropical (Great Barrier Reef) and subtropical (Solitary Islands) populations in  
461    Eastern Australia. Interestingly, geography has had a similar impact on the diversification of  
462    clownfishes (Litsios et al. 2012, 2014; Gaboriau et al. 2024). Many clownfish species have small  
463    range sizes that are highly partitioned geographically and with minimal overlap with sister taxa,  
464    particularly within the Coral Triangle (Litsios et al. 2012, 2014; Gaboriau et al. 2024). Dispersal  
465    kernels that have been estimated for some clownfish species have been shown to be quite small  
466    (<30km) within the Coral Triangle (e.g. Pinsky et al. 2010, 2017) and more extensive in regions  
467    outside the center of marine biodiversity (e.g. Simpson et al. 2014). The range sizes for our  
468    newly delimited *E. quadricolor* species are unknown until further fine-scale sampling and  
469    sequencing can be conducted in the Coral Triangle, but our findings indicate that geographic  
470    processes will continue to be key to fully understand diversification of this species complex.

471

472    *The Entacmaea quadricolor adaptive radiation?*

473                   Although clearly important, geography alone cannot fully explain diversification within  
474    the *E. quadricolor* species complex. Our data indicate that mutualism with clownfishes is also a  
475    key part of the evolutionary history of this species complex. Within the Japanese Archipelago we  
476    find that different species of clownfishes ecologically segregate between co-occurring cryptic  
477    hosts species (as did Kashimoto et al. 2023), but that this pattern is not simply the result of local  
478    processes unique to Japan. Remarkably, our bait-capture data show differentiation by cryptic  
479    host species is part of a deeper evolutionary signal within the *E. quadricolor* complex where  
480    some lineages serve as hosts to the host generalist clownfish species *A. clarkii*, which lives

481 mutualistically on all 10 nominal clownfish-hosting anemone species, and others serve as host to  
482 the host specialist clownfish *A. frenatus*, which is only found with *E. quadricolor*.

483 Central to our interpretation of the evolutionary history of this species complex is the  
484 recognition that co-occurring *E. quadricolor* diversity within the Japanese Archipelago is not the  
485 result of endemic *in situ* diversification, as well as placement of the newly delimited species  
486 from the Maldives (*E. quadricolor* sp. III). Our phylogenetic analyses recover all *E. quadricolor*  
487 species that host *A. clarkii* belonging to a monophyletic clade. This clade includes *E. quadricolor*  
488 sp. I and II from Japan, which were recovered as allopatric sister species in all analyses, and *E.*  
489 *quadricolor* sp. III (Maldives), which also hosts *A. clarkii*. The hierarchical relationship linking  
490 Japan to the Maldives, to the exclusion of Singapore, runs counter to most phylogeographic  
491 patterns from the Indo-West Pacific. Typically, the deepest phylogeographic splits within  
492 tropical marine species complexes from this region partition Indian and Pacific Oceans into  
493 separate clades, with the Indo-Australian Archipelago (IAA) in the Coral Triangle serving as a  
494 well-resolved barrier (reviewed by Bowen et al. 2016). The presence of a deeper evolutionary  
495 signal within *E. quadricolor* connected to clownfish identity suggests the diversity of this species  
496 complex has an origin that is linked directly to the ecology of the mutualism. If so, our data may  
497 provide the first supporting evidence for the hypothesis that *E. quadricolor* has undergone its  
498 own adaptive radiation in response to mutualism with clownfishes.

499 Adaptive radiation has been defined as the “evolution of ecological and phenotypic  
500 diversity within a rapidly multiplying lineage” (Gavrilets and Vose 2005). Adaptive radiation  
501 requires a key innovation that provides a fitness advantage, the evolution of morphological and  
502 functional phenotypes that are linked to ecological niche space, and evidence of rapid speciation  
503 from a common ancestor (e.g Schluter 2020). To date, no key adaptive innovation has been

504 proposed or identified for the clownfish-hosting sea anemones. We propose that mutualism with  
505 clownfishes should be considered a key adaptive innovation for host sea anemones. The  
506 mutualistic benefits provided to sea anemones by hosting clownfishes are well established, and  
507 broadly, the presence of clownfishes is necessary for host anemone survival on tropical coral  
508 reefs. Clownfishes provide novel sources of nitrogen via waste byproducts that are taken up by  
509 the host anemones and their endosymbiotic algae— providing increased protein synthesis and  
510 algal density and allowing host anemones to thrive on oligotrophic coral reefs (Roopin et al.  
511 2008; Roopin and Chadwick 2009). Clownfishes also facilitate gas and nutrient transfer as they  
512 move through their host anemone tentacles (Szczebak et al. 2013; Herbert et al. 2017) and  
513 provide reciprocal protection by defending their sea anemone hosts from butterflyfishes, turtles,  
514 and other predators (Fautin 1991; Godwin and Fautin 1992). Sea anemones with clownfish  
515 symbionts have increased growth and survivorship over those that do not (Porat and Chadwick-  
516 Furman 2004, 2005). Finally, the clownfish-hosting anemones have historically been labeled  
517 “giant” tropical sea anemones because they attain large size classes—some reaching 1m in  
518 diameter (e.g. Fautin and Allen 1992). No other tropical sea anemones reach the same sizes and  
519 occur as free-living individuals on Indo-Pacific coral reefs without clownfish symbionts. Taken  
520 together, the host anemones are likely able to compete for, and maintain, habitat space on  
521 densely populated coral reef ecosystems because of their mutualism with clownfishes.

522 If mutualism with clownfishes is adaptive, our findings for *E. quadricolor* begin to fulfill  
523 other requirements of adaptive radiation. The discovery of an evolutionary signal linking  
524 generalist and specialist clownfish species to cryptic *E. quadricolor* speciation indicates  
525 functional and ecological differences between co-occurring host species that would limit niche  
526 overlap and competition for resources. Our phenotypic/morphological data reveals previously

527 unrecognized phenotypic differences between cryptic host species that are linked to clownfish  
528 symbiont identity. Historically, sea anemone color and pattern has been held as an uninformative  
529 taxonomic character. None of the phenotypic characters we explored here could be considered  
530 synapomorphic, yet we do find that tentacle tip shape and tentacle tip pattern are strongly  
531 predictive of cryptic host species in Japan. Tentacles with blunt-rounded bubble tips were  
532 exclusively found in *E. quadricolor* sp. IV which hosted *A. frenatus*, while tentacles with  
533 elongated tips extending from the bulbous swelling were nearly exclusively found in both *E.*  
534 *quadricolor* sp. I and II which hosted *A. clarkii*. Tentacle tip pattern also was strongly predictive  
535 of cryptic species, with *E. quadricolor* sp. IV anemones having a dull/matte tentacle tip pattern  
536 and with *E. quadricolor* sp. I and II anemones having striated patterns and pink tips.  
537 Interestingly, these phenotypes may have an evolutionary signal consistent across *E. quadricolor*  
538 lineages. While we did not have the images to conduct similar phenotypic analyses elsewhere,  
539 representative images from Singapore (*A. frenatus*-hosting *E. quadricolor* sp. IV) and the  
540 Maldives (*A. clarkii*-hosting *E. quadricolor* sp. III) align well with the phenotypes we recovered  
541 from Japan for both specialist and generalist *E. quadricolor* species (Figure 3d & 3e).

542 Finally, our phylogenetic reconstruction and divergence time analyses demonstrate that  
543 the *E. quadricolor* species complex has diversified rapidly from a common ancestor to at least  
544 five unique species within the last 4-5 million years. Our species delimitation erred on being  
545 conservative to not over-delimit species, but our phylogenetic analyses delimited seven  
546 monophyletic lineages. Our study also focused on fine-scale sampling within the Japanese  
547 Archipelago and adjacent biogeographic regions. We have yet to conduct comprehensive  
548 sampling and sequencing to include the entire biogeographic range of the *E. quadricolor*  
549 complex, which extends from the Northern Red Sea and Arabian Peninsula, the Western Indian

550 Ocean, Western Australia, Coral Triangle, and well into the Central Pacific Ocean. Additional  
551 cryptic species undoubtedly remain to be delimited as the true scope of diversification within the  
552 *E. quadricolor* complex comes into focus. In sum, our data fail to reject the hypothesis that *E.*  
553 *quadricolor* has undergone an adaptive radiation in response to its mutualism with clownfishes.  
554 Whether mutualism with clownfishes is a true key innovation, and the *E. quadricolor* complex  
555 exhibits enough ecological niche partitioning and character displacement to be considered an  
556 adaptive radiation, is unclear but will remain an important future hypothesis to test.

557

558 *Emerging patterns of co-diversification within the clownfish-sea anemone symbiosis.*

559 Mutualism establishes a strong *a priori* hypothesis for co-evolutionary patterns between  
560 partner symbionts. Yet within the clownfish-sea anemone symbiosis co-evolutionary patterns  
561 have remained elusive. Unlike the clownfishes, which have descended from a common ancestor  
562 and belong to the same genus, the 10 nominal species of host sea anemones belong to five genera  
563 within three clades that have evolved symbiosis with clownfishes independently. Thus, no  
564 taxonomic justification for classic co-diversification/cladogenesis existed based on traditional  
565 sea anemone systematics (Fautin and Allen 1992; Titus et al. 2019, 2024; De Jode et al. 2024),  
566 and previous efforts with traditional PCR-based genetic markers have been unsuccessful  
567 (Ngyuyen et al. 2020). Recently, however, the first divergence time estimates for the clownfish-  
568 hosting sea anemones using genomic approaches revealed broadly coincident diversification  
569 times between sea anemones and the clownfish-radiation (De Jode et al. 2024), providing the  
570 first evidence that both symbiotic partners were diversifying around the same time. Here, our  
571 data reveal an evolutionary basis for specialist and generalist lineages of host sea anemones  
572 within just one host species complex. Our data indicate that clownfishes are not merely settling

573 in locally available hosts but recruiting to specialized host lineages with which they have co-  
574 evolved. The discovery of what appears to be clownfish host generalist and clownfish host  
575 specialist *E. quadricolor* lineages represents a major insight that furthers our evolutionary  
576 understanding of the clownfish-sea anemone symbiosis and underscores the importance of  
577 disentangling the systematics and diversity of the host sea anemones for a comprehensive  
578 understanding of this charismatic mutualism.

579

580 **Acknowledgements:**

581 We thank Laura Simmons and Cairns Marine Inc. for providing tissue samples of *Entacmaea*  
582 *quadricolor* from the Great Barrier Reef (Australia). Dr. Christopher Meyer (National Museum  
583 of Natural History) and Small Island Lodge are thanked for fieldwork and logistics in the  
584 Maldives. Charlotte Benedict and Robert Laroche assisted with DNA extractions for all samples  
585 at the American Museum of Natural History and were funded by a National Science Foundation  
586 (NSF) Research Experience for Undergraduate award DBI-1358465 to M. Siddall. Field research  
587 and genomic sequencing were funded by NSF awards DEB-1934274 to BMT and ER, and DEB-  
588 145781 to ER. Field research in Japan was funded by the Japan Society for the Promotion of  
589 Science (JSPS) Kakenhi Grants (JP255440221 to KY, and JP17K15198, JP17H01913, and  
590 23K21774 grants to TF), and Kagoshima University (Establishment of Research and Education  
591 Network of Biodiversity and its Conservation in the Satsunan Islands project to TF). Further  
592 funding to BMT was provided by University of Alabama start-up research funds, and through a  
593 Gerstner Scholar Postdoctoral Fellowship and Gerstner Family Foundation, The Lerner Gray  
594 Fund for Marine Research, and Richard Gilder Graduate School at the American Museum of  
595 Natural History.

596

597 **References:**

598 Bankevich, A., Nurk, S., Antipov, D., Gurevich, A.A., Dvorkin, M., Kulikov, A.S., Lesin, V.M.,  
599 Nikolenko, S.I., Pham, S., Prjibelski, A.D. and Pyshkin, A.V., 2012. SPAdes: a new  
600 genome assembly algorithm and its applications to single-cell sequencing. *Journal of*  
601 *Computational Biology*, 19(5), pp.455-477.

602 Bolger, A.M., Lohse, M. and Usadel, B., 2014. Trimmomatic: a flexible trimmer for Illumina  
603 sequence data. *Bioinformatics*, 30(15), pp.2114-2120.

604 Bowen, B.W., Gaither, M.R., DiBattista, J.D., Iacchei, M., Andrews, K.R., Grant, W.S.,  
605 Toonen, R.J. and Briggs, J.C., 2016. Comparative phylogeography of the ocean  
606 planet. *Proceedings of the National Academy of Sciences*, 113(29), pp.7962-7969.

607 Bronstein, J.L. ed., 2015. *Mutualism*. Oxford University Press.

608 Brucker, R.M. and Bordenstein, S.R., 2012. Speciation by symbiosis. *Trends in Ecology &*  
609 *Evolution*, 27(8), pp.443-451.

610 Burnham, K.P. and Anderson, D.R., 2002. *Model selection and multimodal inference: a*  
611 *practical information theoretic approach*. 2nd ed. NY: Springer.

612 Chan, W.W.R., Tay, Y.C., Ang, H.P., Tun, K., Chou, L.M., Huang, D. and Meier, R., 2020.  
613 Reproduction in urbanised coastal waters: shallow-water sea anemones (*Entacmaea*  
614 *quadricolor* and *Stichodactyla haddoni*) maintain high genetic diversity and  
615 panmixia. *Diversity*, 12(12), p.467.

616 Cowman, P.F., Quattrini, A.M., Bridge, T.C., Watkins-Colwell, G.J., Fadli, N., Grinblat, M.,  
617 Roberts, T.E., McFadden, C.S., Miller, D.J. and Baird, A.H., 2020. An enhanced target-  
618 enrichment bait set for Hexacorallia provides phylogenomic resolution of the staghorn  
619 corals (Acroporidae) and close relatives. *Molecular Phylogenetics and Evolution*, 153,  
620 p.106944.

621 Daly, M., Gusmao, L.C., Reft, A.J. and Rodríguez, E., 2010. Phylogenetic signal in  
622 mitochondrial and nuclear markers in sea anemones (Cnidaria, Actiniaria). *Integrative  
623 and Comparative Biology*, 50(3):371–88.

624 Danecek, P., Auton, A., Abecasis, G., Albers, C.A., Banks, E., DePristo, M.A., Handsaker,  
625 R.E., Lunter, G., Marth, G.T., Sherry, S.T. and McVean, G., 2011. The variant call  
626 format and VCFtools. *Bioinformatics*, 27(15), pp.2156-2158.

627 De Jode, A., Quattrini, AM, Chiodo, T., Daly, M., McFadden, CS, Berumen ML, Meyer, CP,  
628 Mills, S., Beldade, R., Bartholomew, A., Scott, A., Reimer, JD, Yanagi, K., Fujii, T.,  
629 Rodriguez, E. Titus, BM. 2024. Phylogenomics reveals coincident divergence between  
630 giant host sea anemones and the clownfish adaptive radiation. *bioRxiv* pp. 2024-01.

631 Doebeli, M. and Knowlton, N., 1998. The evolution of interspecific mutualisms. *Proceedings of  
632 the National Academy of Sciences*, 95(15), pp.8676-8680.

633 Eaton, D.A. and Overcast, I., 2020. ipyrad: Interactive assembly and analysis of RADseq  
634 datasets. *Bioinformatics*, 36(8), pp.2592-2594.

635 Endo H, Matsuura K. 2022. Geography, Currents, and Fish Diversity of Japan. In: Kai Y,  
636 Motomura H, Matsuura K, editors. *Fish Diversity of Japan: Evolution, Zoogeography,*  
637 *and Conservation*. Singapore: Springer Nature, p. 7–18.

638 Faircloth, B.C., 2016. PHYLUCE is a software package for the analysis of conserved genomic  
639 loci. *Bioinformatics*, 32(5), pp.786-788.

640 Fautin, D.G., 1991. Review article The Anemonefish Symbiosis: What is Known and What is  
641 Not. *Symbiosis*.

642 Fautin, D.G., Allen, G.R. 1992. *Field guide to anemonefishes and their host sea anemones* (p.  
643 78). Perth: Western Australian Museum.

644 Fox, J. and Weisberg, S., 2018. *An R companion to applied regression*. Sage publications.

645 Gaboriau, T., Marcionetti, A., Garcia Jimenez, A., Schmid, S., Fitzgerald, L.M., Micheli, B.,  
646 Titus, B. and Salamin, N., 2024. Host-use drives convergent evolution in clownfish and  
647 disentangles the mystery of an iconic adaptive radiation. *bioRxiv*, pp.2024-07.

648 Gavrillets, S. and Vose, A., 2005. Dynamic patterns of adaptive radiation. *Proceedings of the*  
649 *National Academy of Sciences*, 102(50), pp.18040-18045.

650 Glon, H., Quattrini, A., Rodríguez, E., Titus, B.M. and Daly, M., 2021. Comparison of  
651 sequence-capture and ddRAD approaches in resolving species and populations in  
652 hexacorallian anthozoans. *Molecular Phylogenetics and Evolution*, 163, p.107233.

653 Godwin, J. and Fautin, D.G., 1992. Defense of host actinians by  
654 anemonefishes. *Copeia*, 1992(3), pp.902-908.

655 Goudet, J., 2005. Hierfstat, a package for R to compute and test hierarchical  
656 F-statistics. *Molecular Ecology Notes*, 5(1), pp.184-186.

657 Guindon, S., Dufayard, J.F., Lefort, V., Anisimova, M., Hordijk, W. and Gascuel, O., 2010.  
658 New algorithms and methods to estimate maximum-likelihood phylogenies: assessing  
659 the performance of PhyML 3.0. *Systematic Biology*, 59(3), pp.307-321.

660 Gutenkunst, R.N., Hernandez, R.D., Williamson, S.H. and Bustamante, C.D., 2009. Inferring  
661 the joint demographic history of multiple populations from multidimensional SNP  
662 frequency data. *PLoS Genetics*, 5(10), p.e1000695.

663 Herbert, N.A., Bröhl, S., Springer, K. and Kunzmann, A., 2017. Clownfish in hypoxic  
664 anemones replenish host O<sub>2</sub> at only localised scales. *Scientific Reports*, 7(1), p.6547.

665 Herre, E.A., Knowlton, N., Mueller, U.G. and Rehner, S.A., 1999. The evolution of mutualisms:  
666 exploring the paths between conflict and cooperation. *Trends in Ecology &*  
667 *Evolution*, 14(2), pp.49-53.

668 Hoang, D.T., Chernomor, O., Von Haeseler, A., Minh, B.Q. and Vinh, L.S., 2018. UFBoot2:  
669 improving the ultrafast bootstrap approximation. *Molecular Biology and*  
670 *Evolution*, 35(2), pp.518-522.

671 Huang, D., Meier, R., Todd, P.A. and Chou, L.M., 2008. Slow mitochondrial COI sequence  
672 evolution at the base of the metazoan tree and its implications for DNA  
673 barcoding. *Journal of Molecular Evolution*, 66, pp.167-174.

674 Jombart, T., 2008. adegenet: a R package for the multivariate analysis of genetic  
675 markers. *Bioinformatics*, 24(11), pp.1403-1405.

676 Joy, J.B., 2013. Symbiosis catalyses niche expansion and diversification. *Proceedings of the*  
677 *Royal Society B: Biological Sciences*, 280, p.20122820.

678 Kalyaanamoorthy, S., Minh, B.Q., Wong, T.K., Von Haeseler, A. and Jermiin, L.S., 2017.

679 ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature*  
680 *Methods*, 14(6), pp.587-589.

681 Kashimoto, R., Mercader, M., Zwahlen, J., Miura, S., Tanimoto, M., Yanagi, K., Reimer, J.D.,  
682 Khalturin, K. and Laudet, V., 2024. Anemonefish are better taxonomists than  
683 humans. *Current Biology*, 34(5), pp.R193-R194.

684 Katoh, K., Misawa, K., Kuma, K.I. and Miyata, T., 2002. MAFFT: a novel method for rapid  
685 multiple sequence alignment based on fast Fourier transform. *Nucleic Acids*  
686 *Research*, 30(14), pp.3059-3066.

687 Klann, M., Mercader, M., Salis, P., Reynaud, M., Roux, N., Laudet, V. and Besseau, L., 2021.

688 Anemonefishes. In *Handbook of Marine Model Organisms in Experimental Biology* (pp.  
689 443-464). CRC Press.

690 Laudet, V. and Ravasi, T. eds., 2022. *Evolution, development and ecology of anemonefishes:*  
691 *model organisms for marine science*. CRC Press.

692 Listios G., Sims, C.A., Wüest, R.O., Pearman, P.B., Zimmermann, N.E. and Salamin, N., 2012.

693 Mutualism with sea anemones triggered the adaptive radiation of clownfishes. *BMC*  
694 *evolutionary biology*, 12, pp.1-15.

695 Litsios, G., Pearman, P.B., Lanterbecq, D., Tolou, N. and Salamin, N., 2014. The radiation of  
696 the clownfishes has two geographical replicates. *Journal of Biogeography*, 41(11),  
697 pp.2140-2149.

698 Mangiafico, S.S., 2016. Summary and analysis of extension program evaluation in R. *Rutgers*  
699 *Cooperative Extension: New Brunswick, NJ, USA*, 125, pp.16-22.

700 Marcionetti, A. and Salamin, N., 2023. Insights into the genomics of clownfish adaptive  
701 radiation: the genomic substrate of the diversification. *Genome Biology and*  
702 *Evolution*, 15(7), p.evad088.

703 Marcionetti, A., Rossier, V., Roux, N., Salis, P., Laudet, V. and Salamin, N., 2019. Insights into  
704 the genomics of clownfish adaptive radiation: genetic basis of the mutualism with sea  
705 anemones. *Genome Biology and Evolution*, 11(3), pp.869-882.

706 Margulis, L. and Fester, R. eds., 1991. *Symbiosis as a source of evolutionary innovation:*  
707 *speciation and morphogenesis*. MIT press.

708 McFadden CS, Benayahu, Y., Pante, E., Thoma, J.N., Nevarez, P.A. and France, S.C., 2011.  
709 Limitations of mitochondrial gene barcoding in Octocorallia. *Molecular Ecology*  
710 *Resources*, 11(1), pp.19-31.

711 McFadden, C.S., Quattrini, A.M., Brugler, M.R., Cowman, P.F., Dueñas, L.F., Kitahara, M.V.,  
712 Paz-García, D.A., Reimer, J.D. and Rodríguez, E., 2021. Phylogenomics, origin, and  
713 diversification of Anthozoans (Phylum Cnidaria). *Systematic Biology*, 70(4), pp.635-  
714 647.

715 Minh, B.Q., Schmidt, H.A., Chernomor, O., Schrempf, D., Woodhams, M.D., Von Haeseler, A.  
716 and IQ-TREE, R.L., 2020. 2: New models and efficient methods for phylogenetic  
717 inference in the genomic era., 2020, 37. DOI: <https://doi.org/10.1093/molbev/msaa015>,  
718 pp.1530-1534.

719 Moore, B., Herrera, M., Gairin, E., Li, C., Miura, S., Jolly, J., Mercader, M., Izumiya, M.,  
720 Kawai, E., Ravasi, T. and Laudet, V., 2023. The chromosome-scale genome assembly of  
721 the yellowtail clownfish *Amphiprion clarkii* provides insights into the melanic  
722 pigmentation of anemonefish. *G3: Genes, Genomes, Genetics*, 13(3), p.jkad002.

723 Moran, N.A., 2007. Symbiosis as an adaptive process and source of phenotypic  
724 complexity. *Proceedings of the National Academy of Sciences*, 104, pp.8627-8633.

725 Motomura H, Kimura S, Haraguchi Y. 2007. Two Carangid Fishes (Actinopterygii:  
726 Perciformes), *Caranx heberi* and *Ulua mentalis*, from Kagoshima: The First Records  
727 from Japan and Northernmost Records for the Species. *Species Diversity*, 12(4):223–35.

728 Moya, A., Peretó, J., Gil, R. and Latorre, A., 2008. Learning how to live together: genomic  
729 insights into prokaryote–animal symbioses. *Nature Reviews Genetics*, 9, pp.218-229.

730 Mueller, U.G., Mikheyev, A.S., Solomon, S.E. and Cooper, M., 2011. Frontier mutualism:  
731 coevolutionary patterns at the northern range limit of the leaf-cutter ant–fungus  
732 symbiosis. *Proceedings of the Royal Society B: Biological Sciences*, 278, pp.3050-3059.

733 Nguyen, H.T.T., Dang, B.T., Glenner, H. and Geffen, A.J., 2020. Cophylogenetic analysis of  
734 the relationship between anemonefish *Amphiprion* (Perciformes: Pomacentridae) and

735 their symbiotic host anemones (Anthozoa: Actiniaria). *Marine Biology Research*, 16(2),  
736 pp.117-133.

737 Ollerton, J., McCollin, D., Fautin, D.G. and Allen, G.R., 2007. Finding NEMO: nestedness  
738 engendered by mutualistic organization in anemonefish and their hosts. *Proceedings of  
739 the Royal Society B: Biological Sciences*, 274(1609), pp.591-598.

740 Pinsky, M.L., Montes, Jr, H.R. and Palumbi, S.R., 2010. Using isolation by distance and  
741 effective density to estimate dispersal scales in anemonefish. *Evolution*, 64(9), pp.2688-  
742 2700.

743 Pinsky, M.L., Saenz-Agudelo, P., Salles, O.C., Almany, G.R., Bode, M., Berumen, M.L.,  
744 Andr fou t, S., Thorrold, S.R., Jones, G.P. and Planes, S., 2017. Marine dispersal scales  
745 are congruent over evolutionary and ecological time. *Current Biology*, 27(1), pp.149-  
746 154.

747 Porat D, Chadwick-Furman NE. 2004. Effects of anemonefish on giant sea anemones:  
748 expansion behavior, growth, and survival. *Hydrobiologia*, 530(531):513-520.

749 Porat, D. and Chadwick-Furman, N.E., 2005. Effects of anemonefish on giant sea anemones:  
750 ammonium uptake, zooxanthella content and tissue regeneration. *Marine and  
751 Freshwater Behaviour and Physiology*, 38(1), pp.43-51.

752 Portik, D.M., Leach , A.D., Rivera, D., Barej, M.F., Burger, M., Hirschfeld, M., R del, M.O.,  
753 Blackburn, D.C. and Fujita, M.K., 2017. Evaluating mechanisms of diversification in a  
754 Guineo-Congolian tropical forest frog using demographic model selection. *Molecular  
755 Ecology*, 26(19), pp.5245-5263.

756 Quattrini, A.M., Faircloth, B.C., Dueñas, L.F., Bridge, T.C., Brugler, M.R., Calixto-Botía, I.F.,  
757 DeLeo, D.M., Forêt, S., Herrera, S., Lee, S.M. and Miller, D.J., 2018. Universal  
758 target-enrichment baits for anthozoan (Cnidaria) phylogenomics: New approaches to  
759 long-standing problems. *Molecular Ecology Resources*, 18(2), pp.281-295.

760 Quattrini, A.M., Rodríguez, E., Faircloth, B.C., Cowman, P.F., Brugler, M.R., Farfan, G.A.,  
761 Hellberg, M.E., Kitahara, M.V., Morrison, C.L., Paz-García, D.A. and Reimer, J.D.,  
762 2020. Palaeoclimate ocean conditions shaped the evolution of corals and their skeletons  
763 through deep time. *Nature Ecology & Evolution*, 4(11), pp.1531-1538.

764 Quattrini, A.M., Snyder, K.E., Purow-Ruderman, R., Seiblitz, I.G., Hoang, J., Floerke, N.,  
765 Ramos, N.I., Wirshing, H.H., Rodriguez, E. and McFadden, C.S., 2023. Mito-nuclear  
766 discordance within Anthozoa, with notes on unique properties of their mitochondrial  
767 genomes. *Scientific Reports*, 13(1), p.7443.

768 R Core Team. R: A Language and Environment for Statistical Computing. Vienna, Austria: R  
769 Foundation for Statistical Computing; 2023. Available from: <https://www.R-project.org/>

770 Raj, A., Stephens, M. and Pritchard, J.K., 2014. fastSTRUCTURE: variational inference of  
771 population structure in large SNP data sets. *Genetics*, 197(2), pp.573-589.

772 Reimer, J.D., Biondi, P., Lau, Y.W., Masucci, G.D., Nguyen, X.H., Santos, M.E. and Wee,  
773 H.B., 2019. Marine biodiversity research in the Ryukyu Islands, Japan: current status  
774 and trends. *PeerJ*, 7, p.e6532.

775 Richardson DL, Harriott VJ, Harrison PL. 1997. Distribution and abundance of giant sea  
776 anemones (Actiniaria) in subtropical eastern Australian waters. *Marine and Freshwater*  
777 *Research*, 48(1):59–66.

778 Roopin, M. and Chadwick, N.E., 2009. Benefits to host sea anemones from ammonia  
779 contributions of resident anemonefish. *Journal of Experimental Marine Biology and*  
780 *Ecology*, 370(1-2), pp.27-34.

781 Roopin, M., Henry, R.P. and Chadwick, N.E., 2008. Nutrient transfer in a marine mutualism:  
782 patterns of ammonia excretion by anemonefish and uptake by giant sea  
783 anemones. *Marine Biology*, 154, pp.547-556.

784 Roux, N., Delannoy, C., Yu, S.Y., Miura, S., Carlu, L., Besseau, L., Nakagawa, T., Sato, C.,  
785 Kitajima, K., Guerardel, Y. and Laudet, V., 2024. Anemonefish use sialic acid  
786 metabolism as Trojan horse to avoid giant sea anemone stinging. *bioRxiv*, pp.2024-04.

787 Roux, N., Salis, P., Lee, S.H., Besseau, L. and Laudet, V., 2020. Anemonefish, a model for eco-  
788 evo-devo. *EvoDevo*, 11(1), p.20.

789 Rubin, B.E. and Moreau, C.S., 2016. Comparative genomics reveals convergent rates of  
790 evolution in ant–plant mutualisms. *Nature Communications*, 7(1), p.12679.

791 Sahm, A., Almaida-Pagán, P., Bens, M., Mutualipassi, M., Lucas-Sánchez, A., de Costa Ruiz, J.,  
792 Görlach, M. and Cellerino, A., 2019. Analysis of the coding sequences of clownfish  
793 reveals molecular convergence in the evolution of lifespan. *BMC Evolutionary*  
794 *Biology*, 19, pp.1-12.

795 Schluter D. 2020. *The Ecology of Adaptive Radiation*. OUP Oxford.

796 Scott, A. and Harrison, P.L., 2008. Larval settlement and juvenile development of sea anemones  
797 that provide habitat for anemonefish. *Marine Biology*, 154(5), pp.833-839.

798 Shearer, T.L., Van Oppen, M.J.H., Romano, S.L. and Wörheide, G., 2002. Slow mitochondrial  
799 DNA sequence evolution in the Anthozoa (Cnidaria). *Molecular Ecology*, 11(12),  
800 pp.2475-2487.

801 Simpson, S.D., Harrison, H.B., Claereboudt, M.R. and Planes, S., 2014. Long-distance dispersal  
802 via ocean currents connects Omani clownfish populations throughout entire species  
803 range. *PLoS One*, 9(9), p.e107610.

804 Szczebak, J.T., Henry, R.P., Al-Horani, F.A. and Chadwick, N.E., 2013. Anemonefish  
805 oxygenate their anemone hosts at night. *Journal of Experimental Biology*, 216(6),  
806 pp.970-976.

807 Titus, B.M. and Daly, M., 2022. Population genomics for symbiotic anthozoans: can reduced  
808 representation approaches be used for taxa without reference  
809 genomes? *Heredity*, 128(5), pp.338-351.

810 Titus, B.M. Bennett-Smith M., Chiodo, T., Rodriguez, E. 2024. The clownfish-hosting sea  
811 anemones (Anthozoa: Actiniaria): updated nomenclature, biogeography, and practical  
812 field guide. *Zootaxa*, 5506, 1-34.

813 Titus, B.M., Benedict, C., Laroche, R., Gusmão, L.C., Van Deusen, V., Chiodo, T., Meyer, C.P.,  
814 Berumen, M.L., Bartholomew, A., Yanagi, K. and Reimer, J.D., 2019a. Phylogenetic  
815 relationships among the clownfish-hosting sea anemones. *Molecular Phylogenetics and*  
816 *Evolution*, 139, p.106526.

817 Titus, B.M., Blischak, P.D. and Daly, M., 2019b. Genomic signatures of sympatric speciation  
818 with historical and contemporary gene flow in a tropical anthozoan (Hexacorallia:  
819 Actiniaria). *Molecular Ecology*, 28(15), pp.3572-3586.

820 Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L.D.A., François, R., Grolemund,  
821 G., Hayes, A., Henry, L., Hester, J. and Kuhn, M., 2019. Welcome to the  
822 Tidyverse. *Journal of Open Source Software*, 4(43), p.1686.

823 Yee, T.W., 2010. The VGAM package for categorical data analysis. *Journal of Statistical*  
824 *Software*, 32, pp.1-34.

825 Zhang, C., Nielsen, R. and Mirarab, S., 2023. CASTER: Direct species tree inference from  
826 whole-genome alignments. *bioRxiv*, pp.2023-10.

827 Zhang, C., Rabiee, M., Sayyari, E. and Mirarab, S., 2018. ASTRAL-III: polynomial time  
828 species tree reconstruction from partially resolved gene trees. *BMC bioinformatics*, 19,  
829 pp.15-30.

830

831

832

833

834

835

836

837

838

839 **Table 1.** Pairwise  $F_{ST}$  values between newly delimited cryptic species of *Entacmaea quadricolor*  
840 (EQ) in the Japanese Archipelago that host different clownfishes (*Amphiprion frenatus* and *A.*  
841 *clarkii*).  $F_{ST}$  values were calculated using double-digest restriction site associated DNA  
842 sequencing (ddRADseq).

843

	EQ- <i>A. frenatus</i>	EQ- <i>A. clarkii</i>
EQ- <i>A. frenatus</i>	-	
EQ- <i>A. clarkii</i>	0.27	-
EQ-Ogasawara	0.32	0.15

844

845

846

847

848

849

850

851

852

853

854

855

856

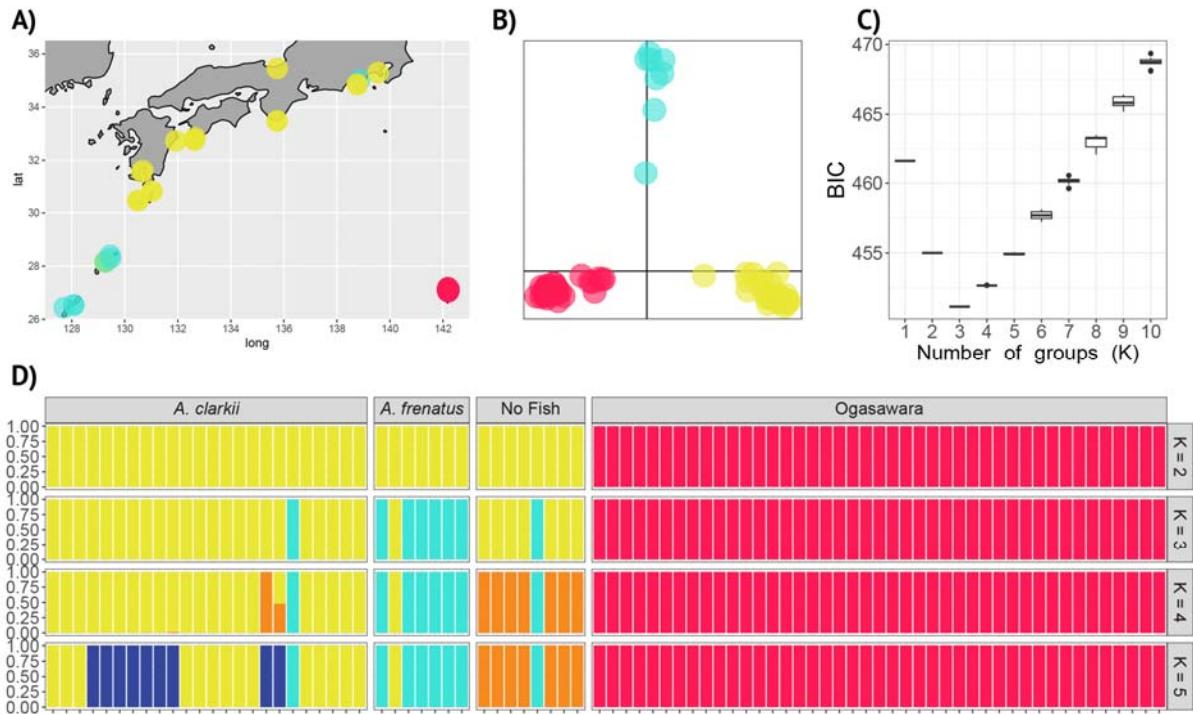
857

858

859

860

861

862 **Figures**

863

864 **Figure 1.** Species delimitation results for the bubble-tip sea anemone *Entacmaea quadricolor*  
 865 using double-digest restriction site associated DNA sequencing (ddRADseq) demonstrating  
 866 cryptic lineages of *E. quadricolor* co-occur throughout Japan and host different species of  
 867 clownfishes. A) Field sampling localities across the Japanese Archipelago and the Ogasawara  
 868 Islands (Red). Sampling localities are color coded by the results of discriminant analysis of  
 869 principal components (DAPC) genetic clustering results for each individual *E. quadricolor* sea  
 870 anemone in the dataset (panels B and D) and reflect newly delimited cryptic *E. quadricolor*  
 871 species. B) DAPC scatter plot of axis one and two for  $k = 3$  genetic cluster assignments. C)  
 872 Results of Bayesian Information Criterion model selection results demonstrating  $k = 3$  as the  
 873 best genetic clustering model for *E. quadricolor*. D) Posterior probabilities of individual group  
 874 assignments for  $k = 2$ -5 for the ddRADseq dataset. For co-occurring cryptic *E. quadricolor*  
 875 lineages in the Ryukyu Islands and Mainland Japan (yellow and teal) the identity of the  
 876 clownfish symbiont present in each anemone is provided.

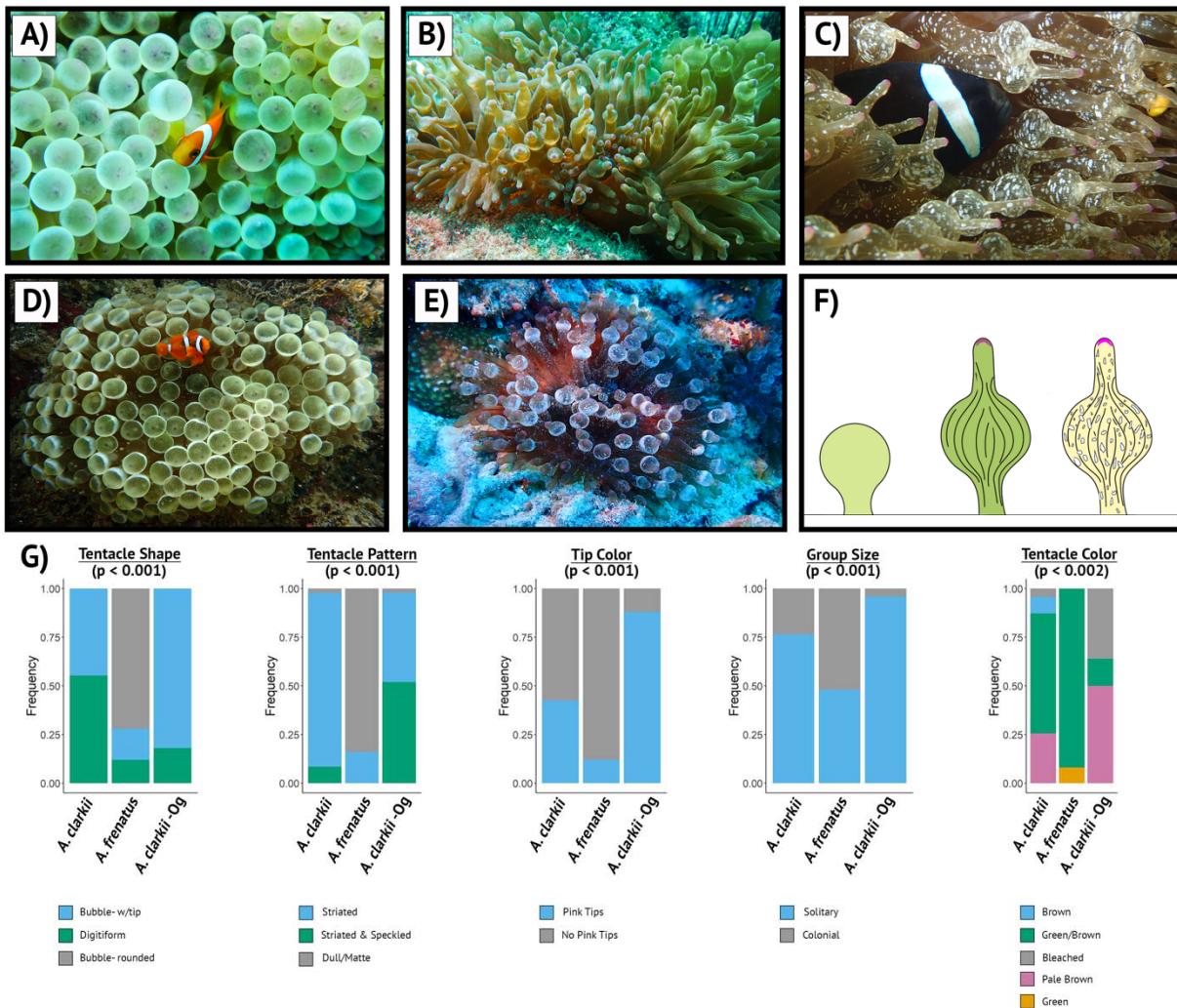
877

878

879

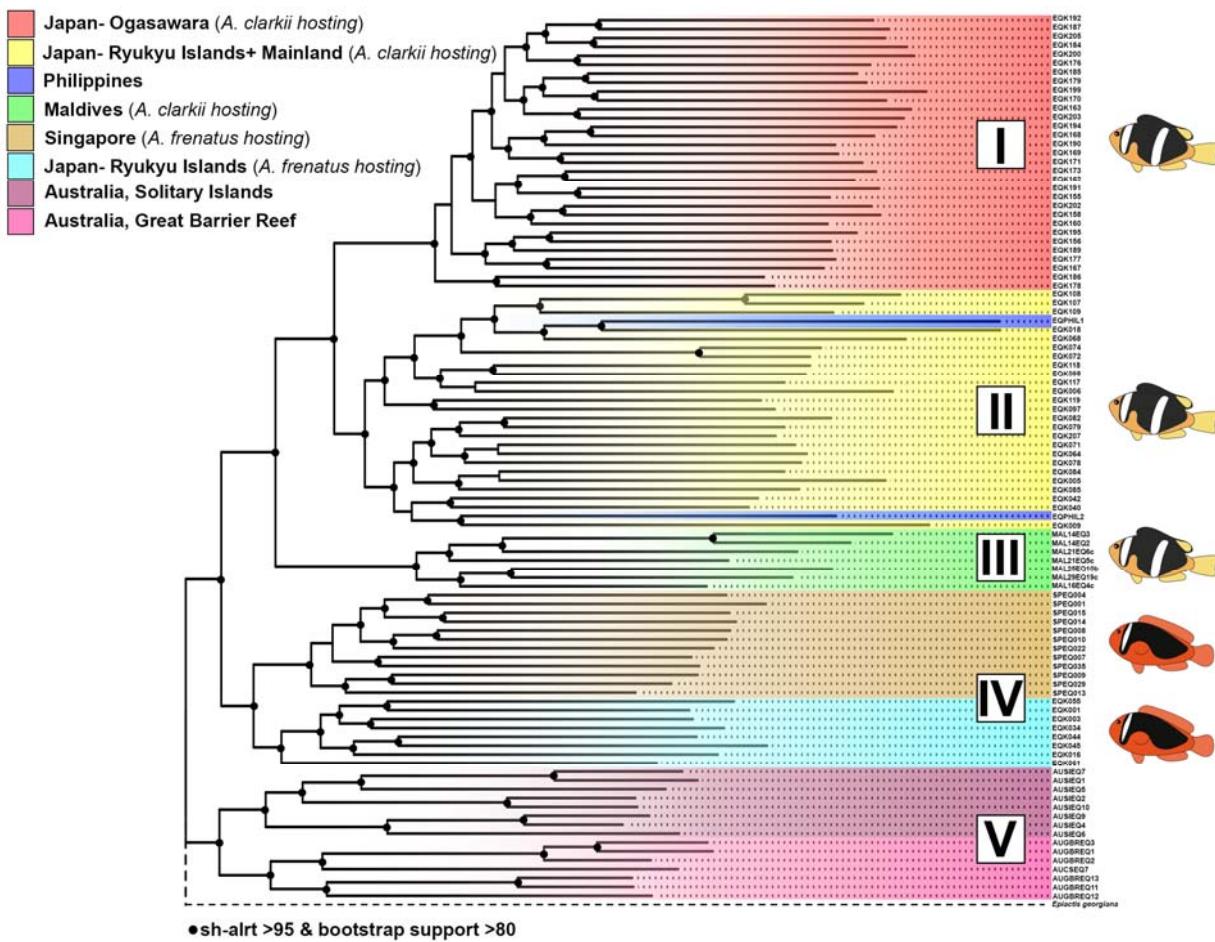
880

881

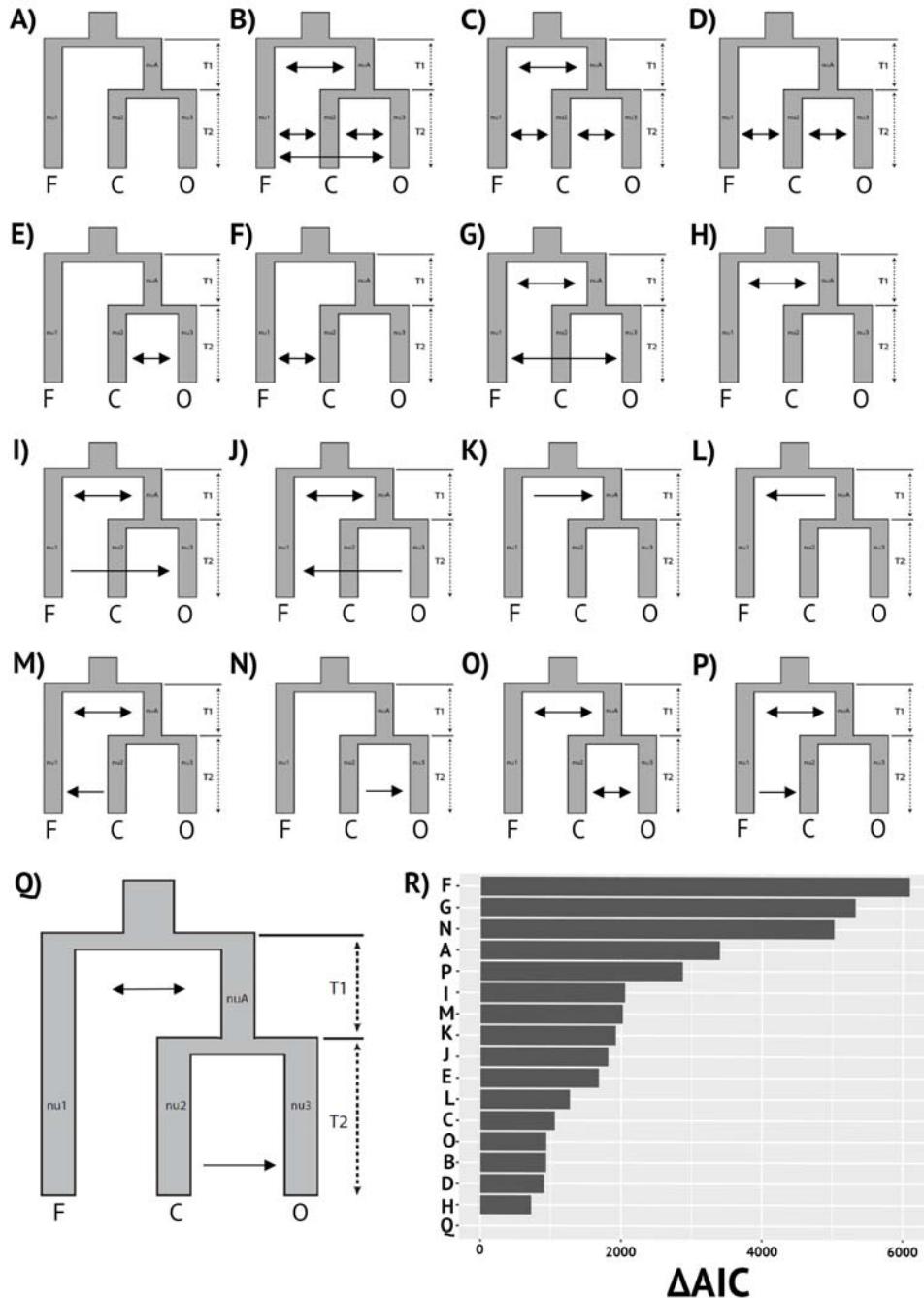


882  
883  
884  
885  
886  
887  
888  
889  
890  
891  
892  
893  
894  
895  
896  
897  
898  
899  
900

**Figure 2.** Consistent phenotypic differences among cryptic *Entacmaea quadricolor* species that host different clownfishes in the Japanese Archipelago. A) *Amphiprion frenatus* hosting sea anemones from the Ryukyu Islands are characterized by rounded bubble tips, a dull/matte tentacle tip pattern, and no significant terminal tentacle tip color. B) *A. clarkii* hosting sea anemones from the Ryukyu Islands and Mainland Japan have bubble-tip tentacles with an extended tip protruding from the bulb and a striated tentacle tip pattern. C) *A. clarkii* hosting sea anemones from the Ogasawara Islands have bubble-tip tentacles with an extended tip protruding from the bulb, a striated and speckled tentacle tip pattern, and are frequently a pale brown color. D) Representative photograph (by Y.C. Tay) of a bubble-tip sea anemone from Singapore that hosts *A. frenatus* and has a qualitatively similar phenotype to the *A. frenatus* hosting anemones from Japan. E) Representative photograph of a bubble-tip sea anemone from the Maldives that hosts *A. clarkii* and has a qualitatively similar phenotype to the *A. clarkii* hosting anemones from Mainland Japan. F) Representative tentacle shape, color, and pattern that align with the three cryptic *E. quadricolor* species in panels A, B, and C, respectively.



903 **Figure 3.** Maximum Likelihood (ML) phylogenetic reconstruction of the bubble-tip sea  
 904 anemone *Entacmaea quadricolor* reveals at least five cryptic species (I-V) and that generalist  
 905 and specialist clownfishes segregate by cryptic host sea anemone species over broad geographic  
 906 scales. ML analyses conducted using partitioned phylogenetic analysis in IQtree2 and bait-  
 907 capture dataset targeting ultra-conserved element and exon loci (75% data occupancy matrix  
 908 and 1002 loci). Results demonstrate that co-occurring *E. quadricolor* lineages in Japan are not  
 909 the result of *in situ* diversification in Japan, but rather geographic range overlap of more deeply  
 910 divergent lineages linked to the clownfish species they host.



921  
922  
923  
924  
925  
926  
927  
928  
929  
930

**Figure 4.** Schematic model summaries (A-Q) and results of demographic model selection analyses in *dadi* testing diversification scenarios in three cryptic species of *Entacmaea quadricolor* in the Japanese Archipelago. Each model is a three-population isolation-migration model with variation in the directionality (arrows) and timing (T1 & T2) of gene flow between co-occurring cryptic species. F = *Amphiprion frenatus* hosting *E. quadriolor* species in Mainland Japan, C = *A. clarkii* hosting *E. quadriolor* species in Mainland Japan, and O = *A. clarkii* hosting *E. quadricolor* species in the Ogasawara Islands (see Fig. 1). Model Q represents the best fit demographic model as determined by Akaike Information Criterion (AIC). R) ΔAIC results, which is the difference between the AIC scores of each model and the best fit model.